

UNIVERSITI PUTRA MALAYSIA

CARDIOPROTECTIVE EFFECTS AND NUTRIGENOMIC STUDY OF EDIBLE BIRD'S NEST IN VITRO AND IN VIVO

ZHANG YIDA

IB 2016 12



CARDIOPROTECTIVE EFFECTS AND NUTRIGENOMIC STUDY OF EDIBLE BIRD'S NEST IN VITRO AND IN VIVO



Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Doctor of Philosophy

CARDIOPROTECTIVE EFFECTS AND NUTRIGENOMIC STUDY OF EDIBLE BIRD'S NEST IN VITRO AND IN VIVO

By

ZHANG YIDA

March 2016

Chairman: Maznah Ismail, PhD

Institute : Bioscience

Cardiovascular disease (CVD) is one of the major cause of morbidity and mortality globally, which is contributed by multiple risk factors including hyperlipidemia, insulin resistance, hypercoagulation, inflammation and oxidative stress. Current therapies have several limitations and are not able to tackle all metabolic perturbations related to CVD. Alternative therapies like edible birds nest (EBN) are therefore receiving closer attention. EBN has been used for thousands of years to improve wellbeing in Asia but there is lack of scientific evidence to back up its use. The present work focused on the cardioprotective effects of edible bird's nest and the nutrigenomic basis for such effect, in vitro and in vivo. In vitro antioxidant potentials of EBN extracts were determined by 2,2' -azinobis- (3-ethylbenzothiazoline-6- sulfonic acid) (ABTS) and oxygen radical absorbance capacity (ORAC) assays, while their effects against oxidative stress were tested in HepG2 cell line. The results showed that EBN extracts possessed potent antioxidant potentials especially after simulated gastrointestinal digestion; at 1000 µg/mL, there was up to 50% radical scavenging without signs of toxicity. This was followed by in vivo testing of the EBN using a high fat diet induced hyperlipidemic Sprague Dawley rat model, in which high fat diet (HFD)+2.5% EBN, HFD+20%EBN, HFD+10mg/kg/day simvastatin, HFD alone and normal pellet were fed to different rat groups for 12 weeks. The results showed that the EBN groups improved HFD-induced hyperlipidemia (total cholesterol = 6.04 mmol/L (19.14%), triglyceride = 0.54 mmol/L (55.37%), and low density lipoprotein = 4.52 mmol/L (9.24%) for HFD+2.5%EBN, and 4.17 mmol/l (44.18%), 0.44 mmol/L (63.64%), 2.98 mmol/L (40.16%) for HFD+20%EBN, respectively), similar to simvastatin (4.99 mmol/L (33.20%), 0.63 mmol/L (47.93%) and 3.6 mmol/L (27.71%), respectively) in comparison with the HFD alone group (7.47 mmol/L, 1.21 mmol/L and 4.98 mmol/L, respectively). EBN also lowered the risk of HFD-induced insulin resistance unlike simvastatin which increased such risk (homeostatic model of insulin resistance was 1.63±0.71, 2.83±0.79 and 2.46±0.22 for HFD+20%EBN, HFD+simvastatin and HFD alone, respectively). EBN also improved HFD-induced inflammation, oxidative stress and coagulation as evidenced by attenuation of HFD-induced alterations of Tumor necrosis factor-alpha (TNF-α), Interleukin-6 (IL-6), C-reactive protein (CRP), ABTS, Thiobarbituric acid reactive substances (TBARS), Activated partial thromboplastin time (APTT), Prothrombin time (PT), Bleeding time (BT), Platelet count (PC),

Oxidized low density lipoprotein (OxLDL), platelet aggregation, leptin, adiponectin and Nitric oxide synthase 3 (NOS3). Furthermore, HFD-induced transcriptional changes were attenuated by EBN; HFD+20% EBN group showed upregulation of the insulin receptor substrate 2 (IRS2), phosphatidyl inositol-3-kinase (PI3K), glucokinase (GCK), gluthatione reductase (Gsr), superoxide dismutase (SOD) and gluthatione peroxidase (Gpx) genes, and downregulation of the inhibitor of nuclear factor kappa-B kinase subunit beta (IKBKB), mitogen-activated protein kinase 1 (MAPK1), Chemokine (C-C motif) ligand 2 (Ccl2), C-reactive protein (CRP), nuclear factor kappa beta1 (Nfkb1), tumor necrosis factor (TNF), von willibrand factor (vWF) and plasminogen activator inhibitor-1 (PAI-1) genes which were the possible basis for the improved insulin resistance, inflammation, oxidative stress and coagulation. EBN was also found to be predominantly protein (57%) of EBN, and sialic acid was the major protein constituent (11%) of EBN. Sialic acid was therefore tested to determine if it was a major contributor to the effects of EBN, with results showing that it contributed significantly as evidenced by similar effects it produced in comparison with the EBN treatment. The findings thus far suggests that EBN and sialic acid may be good candidates for cardioprotection through regulation of hyperlipidemia, insulin resistance, inflammation, oxidative stress and hypercoagulation which are all linked with cardiovascular disease.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

KAJIAN KESAN PERLINDUNGAN KARDIOVASKULAR DAN NUTRIGENOMIKS SARANG BURUNG WALIT *IN VITRO* DAN *IN VIVO*

Oleh

ZHANG YIDA

Mac 2016

Pengerusi: Maznah Ismail, PhD

Institut : Biosains

Penyakit kardiovaskular (CVD) merupakan salah satu punca utama morbiditi dan kematian di seluruh dunia yang disumbangkan oleh pelbagai faktor risiko termasuk hiperlipidemia, rintangan in<mark>sulin, pembek</mark>uan lampau, keradangan dan tekanan oksidatif. Terapi semasa masih tidak dapat menangani semua gangguan metabolik yang berkaitan dengan penyakit kardiovaskular dan ini menimbulkan batasan dalam bidang perubatan. Sehingga kini, pengambilan sarang burung walit sebagai terapi alternatif untuk menangani penyakit kardioyaskular semakin mendapat perhatian. Pemakanan sarang burung walit telah diamalkan sejak beribu-ribu tahun dahulu untuk meningkatkan tahap kesihatan di Asia, namun bukti saintifik terhadap kesan sarang burung walit kepada kesihatan didapati terhad. Oleh yang demikian, kajian ini berfokus kepada kesan perlindungan kardiovaskular dan ciri-ciri nutrigenomik sarang burung walit secara in vitro dan in vivo. Aktiviti anti-pengoksidaan ekstrak sarang burung walit ditentukan melalui ujian ABTS dan ORAC secara in vitro. Di samping itu, aktiviti ekstrak sarang burung walit terhadap tekanan oksidatif dikaji dengan menggunakan sel HepG2. Hasil kajian menunjukkan bahawa ekstrak sarang burung walit berpontensi tinggi sebagai bahan antioksidan terutamanya selepas simulasi pencernaan gastrousus. Pada kepekatan 1000 µg/mL, ekstrak sarang burung walit mampu menghapuskan 50% radikal bebas tanpa memberi kesan ketoksikan. Ini diikuti dengan kajian ekstrak sarang burung walit secara in vivo dengan menggunakan tikus 'Sprague Dawley' yang diaruh diet tinggi lemak selama 12 minggu, di mana pembahagian kumpulan tikus adalah seperti berikut: diet tinggi lemak + 2.5% ekstrak sarang burung walit, diet tinggi lemak + 20% ekstrak sarang burung walit, diet tinggi lemak + 10 mg/kg/hari simvastatin, diet tinggi lemak + kawalan, pelet normal. Hasil kajian menunjukkan bahawa ekstrak sarang burung walit dan simvastatin menambahbaik semua kumpulan tikus yang diberi diet tinggi lemak berbanding dengan kumpulan kawalan (kumpulan 2.5% ekstrak sarang burung walit: jumlah kolesterol = 6.04 mmol/L (19.14%), trigliserida = 0.54 mmol/L (55.37)%, lipoprotein ketumpatan rendah = 4.52 mmol/L (9.24%); kumpulan 20% ekstrak sarang burung: jumlah kolesterol = 4.17 mmol/L (44.18%), trigliserida = 0.44 mmol/L (63.64%), lipoprotein ketumpatan rendah = 2.98 mmol/L (40.16%); kumpulan simvastatin: jumlah kolesterol = 4.99 mmol/L (33.20%), trigliserida = 0.63 mmol/L (47.93%), lipoprotein ketumpatan rendah = 3.6 mmol/L (27.71%); kumpulan kawalan: jumlah kolesterol = 7.47 mmol/L, trigliserida = 1.21 mmol/L, lipoprotein ketumpatan rendah = 4.98 mmol/L). Sarang burung walit juga mengurangkan risiko kerintangan insulin disebabkan oleh diet tinggi lemak dan simvastatin mempunyai kesan yang sebaliknya (kerintangan insulin untuk kumpulan diet tinggi lemak + 20% ekstrak sarang burung walit, kumpulan diet tinggi lemak + simvastatin dan kumpulan diet tinggi lemak kawalan adalah 1.63 ± 0.71 , 2.83 ± 0.79 dan 2.46 ± 0.22 masing-masing). Ekstrak sarang burung walit menambahbaik keradangan, tekanan oksidatif dan pembekuan darah yang disebabkan oleh diet tinggi lemak dan ini boleh dibuktikan dengan hasil kajian menglibatkan TNF-α, IL-6, CRP, ABTS, TBARS, APTT, PT, BT, PC, OxLDL, pengagregatan platelet, leptin, adiponektin dan NOS3. Sebagai tambahan, ekstrak sarang burung walit telah dibuktikan berjaya mengurangkan perubahan transkripsi gen yang disebabkan oleh diet tinggi lemak. Kumpulan diet tinggi lemak + 20% ekstrak sarang burung walit didapati meningkatkan regulasi gen IRS2, PI3K, GCK, Gsr, SOD, GPX1 sementara mengurangkan regulasi gen IKBKB, MAPK1, Ccl2, CRP, Nfkb1, TNF, vWF dan PAI-1. Ini mungkin merupakan asas kepada penambahbaikan rintangan insulin, keradangan, tekanan oksidatif, dan pembekuan darah. Protein merupakan unsur dominan dalam sarang burung walit (57% daripada sarang burung walit) dan asid sialik merupakan konstituen protein utama (11% daripada sarang burung walit). Dengan itu, asid sialik turut dikaji untuk memastikan sumbangannya dalam ekstrak sarang burung walit yang digunakan. Hasil kajian membuktikan bahawa rawatan asid sialik memberi kesan yang sama berbanding dengan rawatan ekstrak sarang burung walit. Penemuan daripada kajian ini mencadangkan bahawa sarang burung walit dan asid sialik adalah sumber yang bagus untuk perlindungan kardio melalui pengawalan hiperlipidemia, rintangan insulin, keradangan, tekanan oksidatif dan hiperkoagulasi yang mana berkait dengan penyakit kardiovaskular.

ACKNOWLEDGEMENTS

I wish to give special thanks to Universiti Putra Malaysia, Chengde Medical University and the Affiliated Hospital of Chengde Medical University for giving me the opportunity for this academic pursuit. I would like to express my deepest gratitude to Prof. Dr. Maznah Ismail for her guidance, encouragement and mentorship throughout the period of my studentship. To my co-supervisors, Prof. Dr. Aini Ideris and Dr. Maizaton Atmadini Abdullah, I also express sincere gratitude for their invaluable support.

To my senior colleagues who helped me, including Mustapha Umar Imam, Norsharina Ismail, Kim Wei Chan, Mastura Ab Ghani, Nurul Nazira, Norhayati Yusuf, Nor Asma Ab Razak, Zainura Mat Hussain and others too numerous to mention, I am very grateful. I am also indebted to Zhiping Hou, Der-Jiun Ooi, Nur Hanisah Azmi, Sarega Nadarajan, Waiteng Wong, Lian Chee Foong, Hadiza Altine Adamu, Nur Diyana Md Zamri, Maryam Murtala Nyako, Siti Farhana Fathy and other peers at the Laboratory of Molecular Biomedicine. My family was very valuable, understanding and supportive during my entire studentship and I wouldn't have made it without them. I also acknowledge the financial support for this project from the Malaysian Ministry of Science, Technology and Innovation (MOSTI) and Universiti Putra Malaysia.

I certify that a Thesis Examination Committee has met on 23 March 2016 to conduct the final examination of Zhang Yida on his thesis entitled "Cardioprotective Effects and Nutrigenomic Study of Edible Bird's Nest *In Vitro* and *In Vivo*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Loh Su Peng, PhD

Associate Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Chairman)

Norhaizan binti Mohd Esa, PhD

Associate Professor
Faculty of Medicine and Health Science
Universiti Putra Malaysia
(Internal Examiner)

Noorjahan Banu binti Mohammed Alitheen, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal Examiner)

Zaheed Husain, PhD

Instructor in Medicine
Harvard Medical School
United States
(External Examiner)

33

ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 21 April 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Maznah Ismail, PhD

Professor Institute of Bioscience Universiti Putra Malaysia (Chairman)

Aini Ideris, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Maizaton Atmadini Abdullah, PhD

Senior Medical Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:	
Name and Matric	No.: Zhang Yida, GS37381	

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision; Guide to Thesis Preparation
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	
Name of	
Chairman of	
Supervisory	
Committee: _	Prof. Dr. Maznah Ismail
Signature:	
Name of	
Member of	
Supervisory	
Committee:	Prof. Dr. Aini Ideris
V	
Signature:	
Name of	
Member of	
Supervisory	
Committee:	Dr Maizaton Atmadini Abdullah

TABLE OF CONTENTS

		Page
ABSTRA	CT	i
ABSTRAI	K	iii
ACKNOV	VLEDGEMENTS	v
APPROV	AL	vi
DECLAR	ATION	viii
LIST OF	TABLES	xiii
	FIGURES	xiv
LIST OF	ABBREVIATIONS	xvii
CHAPTE	R	
1	INTRODUCTION	1
2	LITERATURE REVIEW	4
	Cardiovascular disease	4
	Introduction	4
	Epidemiology	4
	Pathogenesis of CVD	5
	CVD and lipid metabolism	7
	CVD and glucose metabolism	9
	CVD and inflammation	13
	CVD and oxidative stress	14
	CVD and hypercoagulation	16
	Management of CVD	18
	Edible bird's nest (EBN)	23
	Introduction	23
	Main components and main bioactives of EBN	24
	EBN and CVD	25
	Nutrigenomics	26
3	IN VITRO BIOACCESSIBILITY AND ANTIOXIDANT	28
	PROPERTIES OF EDIBLE BIRD'S NEST FOLLOWING	
	SIMULATED HUMAN GASTRO-INTESTINAL	
	DIGESTION	
	Abstract	28
	Background	29
	Materials and methods	29
	Results and discussion	32
	Conclusion	39
	Acknowledgments	39
	References	39
	Copyright/permission	41

4	EDIBLE BIRD'S NEST PREVENTS HIGH FAT DIET-INDUCED INSULIN RESISTANCE IN RATS	42
	Abstract	42
	Introduction	42
	Materials and methods	43
	Results and discussion	49
	Conclusion	58
	Acknowledgments	58
	References	59
	Copyright/permission	62
5	EDIBLE BIRD'S NEST ATTENUATES HIGH FAT	63
	DIET-INDUCED OXIDATIVE STRESS AND	
	INFLAMMATION VIA REGULATION OF HEPATIC	
	ANTIOXIDANT AND INFLAMMATORY GENES	
	Abstract	63
	Background	64
	Materials and methods	64
	Results and discussion	69
	Conclusion	74
	Acknowledgments	74
	References	74
	Copyright/permission	76
6	EDIBLE BIRD'S NEST ATTENUATES	77
	PRO-COAGULATION EFFECTS OF HIGH FAT DIET	
	IN RATS	
	Abstract	77
	Introduction	77
	Materials and methods	78
	Results and discussion	84
	Conclusion	91
	Acknowledgments	92
	References	92
	Copyright/permission	94
7	N-ACETYLNEURAMINICACID SUPPLEMENTATION	95
	PREVENTS HIGH FAT DIET-INDUCED INSULIN	
	RESISTANCE IN RATS THROUGH	
	TRANSCRIPTIONAL AND NON-TRANSCRIPTIONAL	
	MECHANISM	
	Abstract	95
	Introduction	95
	Materials and methods	96
	Results	100
	Discussion	105
	Conclusion	103
	Acknowledgments Pafarances	108
	References	108
	Copyright/permission	110

8	HIGH FAT DIET-INDUCED INFLAMMATION AND OXIDATIVE STRESS ARE ATTENUATED BY N-ACETYLNEURAMINIC ACID IN RATS	111
	Abstract	111
	Introduction	112
	Materials and methods	112
	Results and discussion	118
	Conclusion	127
	Acknowledgments	128
	References	128
	Copyright/permission	131
	Copyright permission	131
9	N-ACETYLNEURAMINIC ACID ATTENUATES	132
	HYPERCOAGULATION ON HIGH FAT	
	DIET-INDUCED HYPERLIPIDEMIC RATS	
	Abstract	132
	Introduction	133
	Materials and methods	133
	Results and discussion	139
	Conclusion	146
	Acknowledgments	146
	References	147
	Copyright/permission	149
10	GENERAL DISCUSSION	150
11	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	152
	Summary and general conclusion	152
	Recommendation for future research	152
DEFER	THOUGH.	150
REFERI		153
APPENI		167
	A OF STUDENT	171
LIST OF	PUBLICATIONS	172

LIST OF TABLES

Table		Page
2.1	The risk factors of cardiovascular disease	5
2.2	Pharmaceutical drugs used in managing cardiovascular disease	19
2.3	Selected foods with cardioprotective properties	20
2.4	Selected herbs with cardioprotective properties	22
2.5	Studies on bioactivity of edible birds' nest	24
2.6	Nutrigenomic mechanisms underlying the functional effects of some foods	27
4.1	Food composition and animal groups	44
4.2	Names, accession number and primer sequences used in the study	47
4.3	Proximate analyses and lactoferrin, ovotransferrin and sialic acid concentrations of edible birds' nest	49
4.4	Food intake and biochemical parameters	52
5.1	Food composition of rat groups, food intake and body weight	66
5.2	Names, accession number and primer sequences used in the study	68
6.1	Food composition of rat groups and food intake	80
6.2	Names, accession number and primer sequences used in the study	83
6.3	Biochemical parameters	85
7.1	Food composition and animal groups	97
7.2	Names, accession number, and primer sequences used in the study	99
7.3	Final body weights, food intake and biochemical parameters	101
8.1	Groupings, food composition and food intake	114
8.2	Names, accession number and primer sequences used in the study	117
8.3	Lipid profile and insulin resistance index	121
9.1	Animal groups, food composition, food intake and body weight	135
9.2	Names, accession number and primer sequences used in the study	138
9.3	Biochemical parameters	139

LIST OF FIGURES

Figure		Page
2.1	The pathogenesis of cardiovascular disease	6
3.1	ABTS results showing the antioxidant potentials of edible bird's nest before and after simulated gastrointestinal digestion	33
3.2	Oxygen radical absorbance capacity of edible bird's nest before and after simulated gastrointestinal digestion	34
3.3	Cytotoxic effects of water extract and digested samples of edible bird's nest on HepG2 cells	35
3.4	Cytoprotective effects of edible bird's nest against H ₂ O ₂ in HepG2 cells	36
3.5	Effect of edible bird's nest on intracellular ROS determination using DCFH-DA	37
3.6	AOPI staining of HepG2 cells after treatment with edible bird's nest and $\rm H_2O_2$	38
4.1	Representative electropherogram following gene analysis on GenomeLab GeXP genetic analysis system expression	48
4.2	Effects of Edible Bird's nest on body weight changes in high fat diet- fed rats over 12 weeks	50
4.3	Effects of edible bird's nest on oral glucose tolerance test in fed high fat diet- fed rats	53
4.4	Effects of edible bird's nest on serum adiponectin, serum leptin, and serum F2-isoprostane in high fat diet- fed rats	54
4.5	Effects of edible bird's nest on hepatic and adipose tissue mRNA levels of insulin receptor, insulin receptor substrate 2 and Phosphoinositide-3-kinase in high fat diet- fed rats	55
4.6	Effects of edible bird's nest on hepatic and adipose tissue mRNA levels of mammalian target of rapamycin, protein kinase C zeta, inhibitor of kappa light polypeptide gene enhancer in B-cells, kinase beta, and mitogen-activated protein kinase 1 in high fat diet- fed rats	56

4.7	Effects of edible bird's nest on hepatic and adipose tissue mRNA levels of Glucokinase, potassium inwardly rectifying channel, subfamily J, member 11, and pyruvate kinase-liver isoform in high fat diet- fed rats	57
4.8	Proposed schematic showing targets of edible bird's nest action in the insulin signaling pathway.	58
5.1	Effect of edible bird's nest on serum antioxidant status and liver thiobarbituric acid reactive species after 12 weeks intervention	70
5.2	Effect of edible bird's nest on serum markers of inflammation after 12 weeks intervention	71
5.3	Effect of edible bird's nest on hepatic mRNA levels of antioxidant genes after 12 weeks intervention	72
5.4	Effect of edible bird's nest on hepatic mRNA levels of inflammation-related genes after 12 weeks intervention	73
6.1	Effects of EBN on APTT, PT, BT in HFD-fed rats.	86
6.2	Effects of EBN on platelet count, platelet aggregation, and RBC count in HFD-fed rats	87
6.3	Effects of EBN on serum OxLDL, serum adiponectin, and serum leptin in HFD-fed rats	88
6.4	Effects of EBN on serum thromboxane, serum vWF, and serum prostacyclin in HFD-fed rats	89
6.5	Effects of EBN on hepatic tissue mRNA levels of leptin, adipoq, and NOS 3 in HFD-fed rats	90
6.6	Effects of EBN on hepatic tissue mRNA levels of vWF, and PAI -1 in HFD-fed rats	91
7.1	Effects of SA on body weight changes in HFD-fed rats over 12 weeks	102
7.2	Effects of SA on oral glucose tolerance test in HFD-fed rats	103
7.3	Effects of SA on hepatic mRNA levels	104
7.4	Effects of SA on adipose tissue mRNA levels	105
7.5	Proposed schematic showing targets of SA transcriptional regulation of insulin signaling pathway	107

8.1	Body weight changes in high fat diet-fed rats over 12 weeks of intervention	119
8.2	Oral glucose tolerance test in high fat diet-fed rats after 12 weeks of intervention	122
8.3	Serum alanine transaminase, aspartate transaminase, alkaline phosphatase, urea, creatinine and uric acid in high fat diet-fed rats after 12 weeks of intervention	123
8.4	Serum markers of inflammation (C-reactive protein; interleukin 6; tumor necrosis factor alpha) in high fat diet-fed rats after 12 weeks of intervention	124
8.5	Oxidative stress markers (serum TAS; liver TBARS) in high fat diet-fed rats after 12 weeks of intervention	125
8.6	Hepatic mRNA levels of glutathione peroxidase, glutathione reductase, superoxide dismutase 1 and 2 in high fat diet-fed rats after 12 weeks of intervention	126
8.7	Hepatic mRNA levels of C-reactive protein and nuclear factor kappa beta in high fat diet-fed rats after 12 weeks of intervention	127
9.1	Effects of sialic acid on activated partial thromboplastin time, prothrombin time, bleeding time in high fat diet-fed rats	140
9.2	Effects of sialic acid on platelet count, platelet aggregation, red blood cell count in high fat diet-fed rats	141
9.3	Effects of sialic acid on serum OxLDL, serum leptin, and serum adiponectin in high fat diet-fed rats	142
9.4	Effects of sialic acid on serum vWF, serum thromboxane, and serum prostacyclin, in high fat diet-fed rats	143
9.5	Effects of sialic acid on the mRNA levels of hepatic leptin and adiponectin genes in high fat diet-fed rats	144
9.6	Effects of sialic acid on the mRNA levels of hepatic von Willebrand factor and plasminogen activator inhibitor-1 genes in high fat diet-fed rats	145

LIST OF ABBREVIATIONS

AAPH 2,2'-azobis (2-amidinopropane) dihydrochloride

ABTS 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulphonic acid)

Adipoq Adiponectin

ALP Alkaline phosphatase
ALT Alanine transaminase

AO-PI Acridine orange-propidium iodide

ApoB Apolipoprotein B

APTT Activated partial thromboplastin time

AST Aspartate transaminase

BT Bleeding time

CAT Catalase

Ccl2 Pyruvate kinase

CHD Coronary heart disease

CRP C-reactive protein

CVD Cardiovascular disease

CYP7A1 Cholesterol 7 alpha-hydroxylase

DCFH-DA Dichloro-dihydro-fluorescein diacetate

EBN Edible bird's nest

EBNH High dose edible bird's nest
EBNL Low dose edible bird's nest
EGF Epidermal growth factor
FGF Fibroblast growth factor

Gck Glucokinase

Gpx Glutathione peroxidase
Gsr Glutathione reductase
H2O2 Hydrogen peroxide

HDL High-density lipoprotein

HFD High fat diet

HMGCR HMG-CoA reductase

HOMA-IR Homeostatic model assessment of insulin resistance

IDL Intermediate-density lipoprotein

Ikbkb Inhibitor of kappa light polypeptide gene enhancer in B-cells,

kinase beta

IKK-β Inhibitor of nuclear factor kappa-B kinase subunit beta

IL-1 Interleukin-1IL-6 Interleukin-6Insr Insulin receptor

IRS Insulin receptor substrate $K_2S_2O_8$ Potassium persulphate

KCNJ11 Potassium inwardly rectifying channel, subfamily J, member

11

LDL lactate dehydrogenase

LDL Low-density lipoprotein

LDLR Low-density lipoprotein receptor

LXR Liver X receptor

MAPK Mitogen-activated protein kinase
MCP1 Monocyte chemotactic protein 1

M-CSF Monocyte-macrophage colony-stimulating factor

MDA Malondialdehyde

mTOR Mammalian target of rapamycin

MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl-tetrazolium

bromide)

NaHCO₃ Sodium bicarbonate

Neu5Ac N-acetylneuraminic acid
Neu5Gc N-glycolylneuraminic acid

NF-κB Nuclear transcription factor kappa-B

NOS3 Nitric oxide synthase 3

OGTT Oral glucose tolerance test

ORAC Oxygen radical absorbance capacity
OxLDL Oxidized low density lipoprotein
PAI-1 Plasminogen activator inhibitor-1

PC Platelet count

PCSK9 Proprotein convertase subtilisin/kexin type 9

PDGF Platelet derived growth factor

PEP Phosphoenolpyruvate

PG Prostaglandins

PI3K Phosphoinositide-3-kinase

Pk Pyruvate kinase

Prkcz Protein kinase C, zeta
PT Prothrombin time

RBC Red blood cell

ROS Reactive oxygen species

SA Sialic acid

SAH High dose sialic acid SAL Low dose sialic acid

SIM Simvastatin

SOD1 Superoxide dismutase 1

SOD2 Superoxide dismutase 2

SREBP Sterol regulatory element-binding proteins

TBARS Thiobarbituric acid reactive substances

TC Total cholesterol

TG Triglyceride

TGFβ Transforming growth factor β
 TNF-α Tumor necrosis factor-alpha
 VLDL Very low-density lipoprotein

vWF Von Willebrand factor

WHO World Health Organization

CHAPTER 1

INTRODUCTION

Morbidity and mortality associated with cardiovascular diseases (CVD) is severe and significant. CVDs cause nearly 30% of all death worldwide. It accounts for more than 17 million deaths globally every year. By 2030, the figure is estimated to grow to 23.6 million. Around 80% of CVDs burden can be found in the low and middle income countries [WHO, 2015]. The fact that these countries already have a huge burden of communicable diseases, the growing burden of CVDs and other non-communicable diseases may be too much to bear.

Atherosclerosis has been reported to be central to most CVD morbidity and mortality, and is promoted by hypercholesterolemia, insulin resistance, coagulation, inflammation and oxidative stress [Libby et al., 2002; Van Gaal et al., 2006; Pawlak et al., 2003; Chuang et al., 2007]. There are many therapeutic options available for managing CVDs, and most are targeted at the factors that promote CVD. Limited successes have been recorded despite wide availability and use of these drugs, and in fact the burden of CVDs is expected to grow in the near future if no urgent action is taken. Insights into the pathogenesis of CVDs and other chronic diseases have indicated that diets play more significant role than was previously anticipated [Estruch et al., 2013; O'Keefe and Cordain, 2004; Joint WHO/FAO Expert Consultation, 2003]. Based on the contribution of diet to the development of CVDs and side effects associated with currently used drugs, there are increasing efforts to also look at diets that can prevent and manage CVDs [Shikany and White Jr, 2000; Amine et al., 2002; Mccullough et al., 2000].

Studies have indicated that the dietary intervention can improve CVD through diverse mechanisms [Erdman, 2000; Stephens et al., 2010; Imam et al., 2014] including anti-inflammation, antioxidation, and amelioration of hyperlipidemia and other metabolic problems. Whole foods have been shown to mediate some of these effects [Imam et al., 2014; He et al., 1995; Lithander and Mahmud, 2015], although in some cases, individual bioactives are better at producing such effects [Ryan et al., 2007; Hu et al., 2015]. Irrespective of the form (whole foods, extracts or bioactive fractions), however, the outcomes are promising. In this regard, oats have been used to manage CVD through lowering cholesterol levels [He et al., 1995], while lupin and germinated brown rice have been shown to regulate oxidative stress, lipid metabolism and insulin sensitivity [Lithander and Mahmud, 2015; Belski et al., 2011; Imam et al., 2014]. There is increasing awareness of the synergistic effects of foods to address the burden of diseases like CVD, which have multiple underlying perturbations, which has prompted the search for foods that have multiple bioactive compounds that are able to regulate several perturbations at once.

Edible bird's nest is used traditionally by the Chinese for its health promoting properties. Its many years of use have not recorded any major safety concerns, prompting its continuous and wide spread use. Scientific evidence supporting the use of EBN for medicinal purposes is very scarce, and information on the bioactives

responsible for its functional effects is sadly lacking. Additionally, there are limited in vitro tests that have indicated that EBN may have antioxidant and anti-inflammatory properties [Aswir and Wan Nazaimoon, 2011; Kim et al., 2012]. Thus, based on the antioxidant and anti-inflammatory effects of EBN, it was hypothesized that it may be beneficial in managing CVD, since both effects could independently improve CVD. Hence, this study was aimed to investigate the functional effects of EBN on selected parameters of CVD. In this study, the antioxidant potentials of EBN in vitro, and its effects on lipid metabolism, insulin resistance, inflammation, oxidative stress and anticoagulation were determined including underlying transcriptional regulation, which have all been associated with CVD, in a continuing study using an animal model of hyperlipidemia. The regulation of these different parameters by EBN indicated that it is a good candidate for use in the management of CVD among Malaysians, where EBN is produced extensively. Additionally, Malaysia is one of the largest exporters of EBN and as such this benefit can be extended to the international community. In view of the increasing drive to use safer alternatives for the management of chronic diseases like CVD, the results from this study could have profound implications on the health of majority of individuals since EBN is consumed by majority of people in Asia, where CVD is the leading cause of death. By extension, the use of EBN by people around the world could also impact positively on global mortality due to CVD, which remains the highest cause of morbidity and mortality all over the world. Similarly, in view of the possible contributory effects of the major constituent of EBN, sialic acid, to the functional effects of EBN, it was hypothesized that sialic acid, if it is able to produce similar effects to EBN, could also be used in isolation as a nutraceutical for individuals that prefer to use supplements instead of foods. Overall, the results could impact on the health of majority of people, because the metabolic disturbances targeted in the study underlie the major causes of death globally, especially CVD.

General objective:

To study the cardioprotective effects of edible bird's nest and the nutrigenomic basis for such effects, *in vitro* and *in vivo*.

Specific objectives:

- 1. To determine the antioxidant activities of EBN extracts and its cytotoxicity *in vitro* using HepG2 cell line.
- 2. To determine the effects of EBN and sialic acid on lipid metabolism and insulin resistance in high fat diet-induced hypercholesterolemic Sprague Dawley rat model, including the underlying nutrigenomic basis for understanding mechanistic action.
- 3. To determine the antioxidant and anti-inflammatory effects of EBN and sialic acid in high fat diet-induced hypercholesterolemic Sprague Dawley rat model, including the underlying nutrigenomic basis for understanding mechanistic action
- 4. To determine the anti-coagulation effects of EBN and sialic acid in high fat diet-induced hypercholesterolemic rat model, including the underlying nutrigenomic basis for understanding mechanistic action.

Hypotheses of the study were:

- 1. EBN has high antioxidant potential and is non-toxic to human cell.
- 2. EBN and sialic acid have lipid-regulating properties.
- 3. EBN and sialic acid can improve CVD risk factors such as insulin resistance, hypercoagulation, oxidative stress and inflammation.



REFERENCES

- [1] Abidin, F.Z., Hui, C.K., Luan, N.S., Ramli, E.S.M., Hun, L.T., and Ghafar, N.A. (2011). Effects of edible bird's nest (EBN) on cultured rabbit corneal keratocytes. *BMC Complementary and Alternative Medicine*. 11(1): 94.
- [2] Afman, L., and Müller, M. (2006). Nutrigenomics: from molecular nutrition to prevention of disease. *Journal of the American Dietetic Association*. 106(4): 569-576.
- [3] Aikawa, M., Rabkin, E., Sugiyama, S., Voglic, S.J., Fukumoto, Y., Furukawa, Y., Shiomi, M., Schoen, F.J., and Libby, P. (2001). An HMG-CoA reductase inhibitor, cerivastatin, suppresses growth of macrophages expressing matrix metalloproteinases and tissue factor *in vivo* and *in vitro*. *Circulation*. 103(2): 276-283.
- [4] Aksamitiene, E., Kiyatkin, A.B., and Kholodenko, B.N. (2012). Cross-talk between mitogenic Ras/MAPK and survival PI3K/Akt pathways: a fine balance. *Biochemical Society Transactions*. 40(1):139-46.
- [5] Al-Naqeep, G., Al-Zubairi, A. S., Ismail, M., Amom, Z. H., and Esa, N. M. (2011). Antiatherogenic potential of Nigella sativa seeds and oil in diet-induced hypercholesterolemia in rabbits. *Evidence-Based Complementary and Alternative Medicine*. 2011:213628.
- [6] Al-Naqeep, G., and Ismail, M. (2009). Regulation of apolipoprotein A-1 and apolipoprotein B100 genes by thymoquinone rich fraction and thymoquinone in HepG2 cells. *Journal of Food Lipids*. 16(2): 245-258.
- [7] Al-Naqeep, G., Ismail, M., and Allaudin, Z. (2009). Regulation of low-density lipoprotein receptor and 3-hydroxy-3-methylglutaryl coenzyme A reductase gene expression by thymoquinone-rich fraction and thymoquinone in HepG2 cells. *Journal of Nutrigenetics and Nutrigenomics*. 2(4-5): 163-172.
- [8] Al-Naqeeb, G., Ismail, M., and Al-Zubairi, A. S. (2009). Fatty acid profile, α-tocopherol content and total antioxidant activity of oil extracted from Nigella sativa seeds. *International Journal of Pharmacology*, 5(4): 244-250.
- [9] Al-Naqeep, G., Ismail, M., and Yazan, L.S. (2009). Effects of thymoquinone rich fraction and thymoquinone on plasma lipoprotein levels and hepatic low density lipoprotein receptor and 3-hydroxy-3-methylglutaryl coenzyme A reductase genes expression. *Journal of Functional Foods*. 1(3): 298-303.
- [10] Amarowicz, R., Pegg, R.B., Rahimi-Moghaddam, P., Barl, B., and Weil, J.A. (2004). Free-radical scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies. *Food Chemistry*. 84(4): 551-562.
- [11] Amine, E., Baba, N., Belhadj, M., Deurenbery-Yap, M., Djazayery, A., Forrester, T., Galuska, D., Herman, S., James, W., M'Buyamba, J., Katan, M., Key, T., Kumanyika, S., Mann, J., Moynihan, P., Musaiger, A., Prentice, A., Reddy, K., Schatzkin, A., Seidell, J., Simpopoulos, A., Srianujata, S., Steyn, N., Swinburn, Boyd, Uauy, R., Wahlqvist, M., Zhao-su, W. and Yoshiike, N. (2002). Diet, nutrition and the prevention of chronic diseases: report of a Joint WHO/FAO Expert Consultation. *World Health Organization*.
- [12] Andallu, B., and Radhika, B. (2000). Hypoglycemic, diuretic and hypocholesterolemic effect of winter cherry (Withania somnifera, Dunal) root. *Indian Journal of Experimental Biology*. 38(6): 607-609.
- [13] Anderson, K.M., Odell, P.M., Wilson, P.W., and Kannel, W.B. (1991). Cardiovascular disease risk profiles. *American Heart Journal*. 121(1): 293-298.

- [14] Arbabi, S., and Maier, R.V. (2002). Mitogen-activated protein kinases. *Critical Care Medicine*. 30(1): S74-S79.
- [15] Arici, M., and Walls, J. (2001). End-stage renal disease, atherosclerosis, and cardiovascular mortality: is C-reactive protein the missing link?. *Kidney International*. 59(2): 407-414.
- [16] Arkan, M.C., Hevener, A.L., Greten, F.R., Maeda, S., Li, Z.W., Long, J.M., Wynshaw-Boris, A., Poli, G., Olefsky, J., and Karin, M. (2005). IKK-β links inflammation to obesity-induced insulin resistance. *Nature Medicine*. 11(2): 191-198.
- [17] Aswir, A.R., and Wan Nazaimoon, W.M. (2011). Effect of edible bird's nest on cell proliferation and tumor necrosis factor-alpha (TNF-α) release *in vitro*. *International Food Research Journal*, 18(3), 1073-1077.
- [18] Au, A.L.S., Kwan, Y.W., Kwok, C.C., Zhang, R.Z., and He, G.W. (2003). Mechanisms responsible for the *in vitro* relaxation of ligustrazine on porcine left anterior descending coronary artery. *European Journal of Pharmacology*. 468(3): 199-207.
- [19] Babji, A.S., Yusop, S.M., and Ghassem, M. (2011). A review of research developments of Malaysian's biodiversified resources. *In Proceedings of 12th ASEAN Food Conference*. 115-122.
- [20] Barter, P.J., Ballantyne, C.M., Carmena, R., Cabezas, M.C., Chapman, M., Couture, P., Graaf, J.D., Durrington, P.N., Faergeman, O., Frohlich, J., Furberg, C.D., Gagne, C., Haffner, S.M., Humphrise, S.E., Jungner, I., Krauss, R.M., Kwiterovich, P., Marcovina, S., Packard, C.J., Pearson, T.A., Srinath Reddy, K., Rosenson, R., Sarrafzadegan, N., Sniderman, A.D., Stalenhoef, A.F., Stein, E., Talmud, P.J., Tonkin, A.M., Walldius, G., and Williams, K.M.S. (2006). Apo B versus cholesterol in estimating cardiovascular risk and in guiding therapy: report of the thirty-person/ten-country panel. *Journal of Internal Medicine*. 259(3): 247-258.
- [21] Belski, R., Mori, T.A., Puddey, I.B., Sipsas, S., Woodman, R.J., Ackland, T.R., Beilin, L.J., Dove, E.R., Carlyon, N.B., Jayaseena, V., and Hodgson, J.M. (2011). Effects of lupin-enriched foods on body composition and cardiovascular disease risk factors: a 12-month randomized controlled weight loss trial. *International Journal of Obesity*. 35(6): 810-819.
- [22] Brachmann, S.M., Ueki, K., Engelman, J.A., Kahn, R.C., and Cantley, L.C. (2005). Phosphoinositide 3-kinase catalytic subunit deletion and regulatory subunit deletion have opposite effects on insulin sensitivity in mice. *Molecular and Cellular Biology*. 25(5): 1596-1607.
- [23] Bravata, D.M., Gienger, A.L., McDonald, K.M., Sundaram, V., Perez, M.V., Varghese, R., Kapoor, J.R., Ardehali, R., Owens, D.K., and Hlatky, M.A. (2007). Systematic review: the comparative effectiveness of percutaneous coronary interventions and coronary artery bypass graft surgery. *Annals of Internal Medicine*. 147(10): 703-716.
- [24] Breda, E., Cavaghan, M.K., Toffolo, G., Polonsky, K.S., and Cobelli, C. (2001). Oral glucose tolerance test minimal model indexes of β-cell function and insulin sensitivity. *Diabetes*. 50(1): 150-158.
- [25] Broadhurst, C.L., Polansky, M.M., and Anderson, R.A. (2000). Insulin-like biological activity of culinary and medicinal plant aqueous extracts *in vitro*. *Journal of Agricultural and Food Chemistry*. 48(3): 849-852.
- [26] Brunton L, Parker K, Blumenthal D, Buxton I. (2008). Goodman & Gilman's the pharmacological basis of therapeutics. 1st Edition. McGrawHill, New York. pp 528-543 and 603-615.

- [27] Cao, H., Polansky, M.M., and Anderson, R.A. (2007). Cinnamon extract and polyphenols affect the expression of tristetraprolin, insulin receptor, and glucose transporter 4 in mouse 3T3-L1 adipocytes. *Archives of Biochemistry and Biophysics*. 459(2): 214-222.
- [28] Cao, Y., Xu, J., Wang, J.F., You, Y.Y., and Xue, C.H. (2012). Studies on immunomodulation function of Indonesia white edible bird's nest on hypoimmune mice. *Acta Nutrimenta Sinica*. 34(2): 168-171.
- [29] Carreira, H., Pereira, M., Alves, L., Lunet, N., and Azevedo, A. (2012). Dyslipidaemia, and mean blood cholesterol and triglycerides levels in the Portuguese population: a systematic review. *Arquivos de Medicina*. 26(3): 112-123.
- [30] Chan, P.S., and Cervoni, P. (1986). Prostaglandins, prostacyclin, and thromboxane in cardiovascular diseases. *Drug Development Research*. 7(4): 341-359.
- [31] Chua, K.H., Lee, T.H., Nagandran, K., Yahaya, N.H.M., Lee, C.T., Tjih, E.T.T., and Aziz, R.A. (2013). Edible Bird's nest extract as a chondro-protective agent for human chondrocytes isolated from osteoarthritic knee: *in vitro* study. *BMC Complementary and Alternative Medicine*. 13(1): 19.
- [32] Chuang, K.J., Chan, C.C., Su, T.C., Lee, C.T., and Tang, C.S. (2007). The effect of urban air pollution on inflammation, oxidative stress, coagulation, and autonomic dysfunction in young adults. *American Journal of Respiratory and Critical Care Medicine*. 176(4): 370-376.
- [33] Cromwell, W.C., Otvos, J.D., Keyes, M.J., Pencina, M.J., Sullivan, L., Vasan, R.S., Vilson, P.W.F., and D'Agostino, R.B. (2007). LDL particle number and risk of future cardiovascular disease in the Framingham Offspring Study—implications for LDL management. *Journal of Clinical Lipidology*. 1(6): 583-592.
- [34] Crook, M.A., Earle, K., Morocutti, A., Yip, J., Viberti, G., and Pickup, J.C. (1994). Serum sialic acid, a risk factor for cardiovascular disease, is increased in IDDM patients with microalbuminuria and clinical proteinuria. *Diabetes Care*. 17(4): 305-310.
- [35] Cui, J., Panse, S., and Falkner, B. (2011). The role of adiponectin in metabolic and vascular disease: a review. *Clinical Nephrology*. 75(1): 26-33.
- [36] Dávalos, A., Gómez-Cordovés, C., and Bartolomé, B. (2004). Extending applicability of the oxygen radical absorbance capacity (ORAC-fluorescein) assay. *Journal of Agricultural and Food Chemistry*. 52(1): 48-54.
- [37] Davie, E.W., Fujikawa, K., and Kisiel, W. (1991). The coagulation cascade: initiation, maintenance, and regulation. *Biochemistry*. 30(43): 10363-10370.
- [38] De Taeye, B., Smith, L.H., and Vaughan, D.E. (2005). Plasminogen activator inhibitor-1: a common denominator in obesity, diabetes and cardiovascular disease. *Current Opinion in Pharmacology*. 5(2): 149-154.
- [39] DeFronzo, R.A., and Ferrannini, E. (1991). Insulin resistance: a multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes Care*. 14(3): 173-194.
- [40] Dentali, F., Romualdi, E., and Ageno, W. (2007). The metabolic syndrome and the risk of thrombosis. *Haematologica*. 92(3): 297-299.
- [41] Deraman, N. (2012). Antioxidant studies of cave edible bird's nest. Bachelor of science thesis. Faculty of applied sciences, Universiti teknologi Mara.
- [42] Doran, A.C., Meller, N., and McNamara, C.A. (2008). Role of smooth muscle cells in the initiation and early progression of atherosclerosis. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 28(5): 812-819.

- [43] Erdman, J.W. (2000). Soy protein and cardiovascular disease a statement for healthcare professionals from the Nutrition Committee of the AHA. *Circulation*. 102(20): 2555-2559.
- [44] Estruch, R., Ros, E., Salas-Salvadó, J., Covas, M.I., Corella, D., Arós, F., and Martínez-González, M.A. (2013). Primary prevention of cardiovascular disease with a Mediterranean diet. *New England Journal of Medicine*. 368(14): 1279-1290.
- [45] Fahy, E., Subramaniam, S., Murphy, R.C., Nishijima, M., Raetz, C.R., Shimizu, T., Spener, F., Meer, G.V., Wakelam, M.J.O and Dennis, E.A. (2009). Update of the LIPID MAPS comprehensive classification system for lipids. *Journal of Lipid Research*. 50(Supplement): S9-S14.
- [46] Farquhar, J., Wood, P., Breitrose, H., Haskell, W., Meyer, A., Maccoby, N., Alexander, J., Brown J.R.B, Mcalister, A, Nash, J and Stern, M. (1977). Community education for cardiovascular health. *The Lancet*. 309(8023): 1192-1195.
- [47] Farràs, M., Valls, R.M., Fernández-Castillejo, S., Giralt, M., Solà, R., Subirana, I., Motilva, M.J., Konstantinidou, V., Covas, M.I., and Fitó, M. (2013). Olive oil polyphenols enhance the expression of cholesterol efflux related genes *in vivo* in humans. A randomized controlled trial. *The Journal of Nutritional Biochemistry*. 24(7): 1334-1339.
- [48] Feng, L., Ke, N., Cheng, F., Guo, Y., Li, S., Li, Q., and Li, Y. (2011). The protective mechanism of ligustrazine against renal ischemia/reperfusion injury. *Journal of Surgical Research*. 166(2): 298-305.
- [49] Ferguson, J.F., Phillips, C.M., McMonagle, J., Pérez-Martínez, P., Shaw, D.I., Lovegrove, J.A., Helal, O., Defoort, C., Gjelstad, I.M.F., Drevon, C.A., Blaak, E.E., Saris, W.H.M., Leszczyńska-Gołabek, I., Kiec-Wilk, B., Risérus, U., Karlström, B., Lopez-Miranda, J., and Roche, H.M. (2010). NOS3 gene polymorphisms are associated with risk markers of cardiovascular disease, and interact with omega-3 polyunsaturated fatty acids. *Atherosclerosis*. 211(2): 539-544.
- [50] Frostegård, J. (2002). Autoimmunity, oxidized LDL and cardiovascular disease. *Autoimmunity Reviews*. 1(4): 233-237.
- [51] Fujioka, H.B., (1998). The pharmacological effects of edible bird's nest extracts: Inotropic effect. International Journal of Traditional Chinese Medicine. 20(3): 58.
- [52] Fukai, T., Folz, R.J., Landmesser, U., and Harrison, D.G. (2002). Extracellular superoxide dismutase and cardiovascular disease. *Cardiovascular Research*. 55(2): 239-249.
- [53] Furie, B., and Furie, B.C. (1988). The molecular basis of blood coagulation. *Cell*. 53(4): 505-518.
- [54] Gable, D.R., Hurel, S.J., and Humphries, S.E. (2006). Adiponectin and its gene variants as risk factors for insulin resistance, the metabolic syndrome and cardiovascular disease. *Atherosclerosis*. 188(2): 231-244.
- [55] Gillingham, L. (2011). Efficacy of high-oleic canola and flaxseed oils for cardiovascular disease risk reduction. University of Manitoba Thesis, Canada.
- [56] Go, A.S., Mozaffarian, D., Roger, V.L., Benjamin, E.J., Berry, J.D., Blaha, M.J., and Stroke, S.S. (2014). Heart disease and stroke statistics--2014 update: a report from the American Heart Association. *Circulation*. 129(3): e28.
- [57] Gogg, S., Smith, U., and Jansson, P.A. (2009). Increased MAPK activation and impaired insulin signaling in subcutaneous microvascular endothelial cells in type 2 diabetes: the role of endothelin-1. *Diabetes*. 58(10): 2238-2245.

- [58] Gong, B., Yu, J., Li, H., Li, W., and Tong, X. (2012). The effect of KCNJ11 polymorphism on the risk of type 2 diabetes: a global meta-analysis based on 49 case-control studies. *DNA and Cell Biology*. 31(5): 801-810.
- [59] Gopaul, K.P., and Crook, M.A. (2006). Sialic acid: a novel marker of cardiovascular disease?. Clinical Biochemistry. 39(7): 667-681.
- [60] Gordon, D.J., Probstfield, J.L., Garrison, R.J., Neaton, J.D., Castelli, W.P., Knoke, J.D., Jacobs Jr, D.R., Bangdiwala, S., and Tyroler, H.A. (1989). High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. *Circulation*. 79(1): 8-15.
- [61] Gordon, T., Castelli, W.P., Hjortland, M.C., Kannel, W.B., and Dawber, T.R. (1977). High density lipoprotein as a protective factor against coronary heart disease: the Framingham Study. *The American Journal of Medicine*. 62(5): 707-714.
- [62] Guo, C.T., Takahashi, T., Bukawa, W., Takahashi, N., Yagi, H., Kato, K., Hidari, K.I.P.J., Miyamoto, D., Suzuki, T., and Suzuki, Y. (2006). Edible bird's nest extract inhibits influenza virus infection. *Antiviral Research*. 70(3): 140-146.
- [63] Hamanishi, T., Furuta, H., Kato, H., Doi, A., Tamai, M., Shimomura, H., Sakagashira, H., Nishi, M., Sasaki, H., Sanke, T., and Nanjo, K. (2004). Functional variants in the glutathione peroxidase-1 (GPx-1) gene are associated with increased intima-media thickness of carotid arteries and risk of macrovascular diseases in Japanese type 2 diabetic patients. *Diabetes*. 53(9): 2455-2460.
- [64] Han, J.Y., Fan, J.Y., Horie, Y., Miura, S., Cui, D.H., Ishii, H., Hibi, T., Tsuneki, H., and Kimura, I. (2008). Ameliorating effects of compounds derived from Salvia miltiorrhiza root extract on microcirculatory disturbance and target organ injury by ischemia and reperfusion. *Pharmacology and Therapeutics*. 117(2): 280-295.
- [65] Hanukoglu, I. (1992). Steroidogenic enzymes: structure, function, and role in regulation of steroid hormone biosynthesis. *The Journal of Steroid Biochemistry and Molecular Biology*. 43(8): 779-804.
- [66] He, J., Klag, M.J., Whelton, P.K., Mo, J.P., Chen, J.Y., Qian, M.C., Mo, P.S., and He, G.Q. (1995). Oats and buckwheat intakes and cardiovascular disease risk factors in an ethnic minority of China. *The American Journal of Clinical Nutrition*. 61(2): 366-372.
- [67] Heinrich, J., Schulte, H., Schönfeld, R., Köhler, E., and Assmann, G. (1995). Association of variables of coagulation, fibrinolysis and acute-phase with atherosclerosis in coronary and peripheral arteries and those arteries supplying the brain. *Thrombosis and Haemostasis*. 73(3): 374-379.
- [68] Hsieh, C.L., Lin, Y.C., Yen, G.C., and Chen, H.Y. (2007). Preventive effects of guava (Psidium guajava L.) leaves and its active compounds against α-dicarbonyl compounds-induced blood coagulation. *Food Chemistry*. 103(2): 528-535.
- [69] Hu, W.S., Ting, W.J., Chiang, W.D., Pai, P., Yeh, Y.L., Chang, C.H., Lin, W.T., and Huang, C.Y. (2015). The Heart Protection Effect of Alcalase Potato Protein Hydrolysate Is through IGF1R-PI3K-Akt Compensatory Reactivation in Aging Rats on High Fat Diets. *International Journal of Molecular Sciences*. 16(5): 10158-10172.
- [70] Imam, M.U., and Ismail, M. (2013). Nutrigenomic effects of germinated brown rice and its bioactives on hepatic gluconeogenic genes in type 2 diabetic rats and HepG2 cells. *Molecular Nutrition and Food Research*. 57(3): 401-411.

- [71] Imam, M.U., Ishaka, A., Ooi, D.J., Zamri, N.D. M., Sarega, N., Ismail, M., and Esa, N.M. (2014). Germinated brown rice regulates hepatic cholesterol metabolism and cardiovascular disease risk in hypercholesterolaemic rats. *Journal of Functional Foods*. 8: 193-203.
- [72] Imam, M.U., Ismail, M., Omar, A.R., and Ithnin, H. (2013). Research Article The Hypocholesterolemic Effect of Germinated Brown Rice Involves the Upregulation of the Apolipoprotein A1 and Low-Density Lipoprotein Receptor Genes. *Experimental Diabetes Research*. 2013: 134694
- [73] Imam, M.U., Musa, S.N.A., Azmi, N.H., and Ismail, M. (2012). Effects of white rice, brown rice and germinated brown rice on antioxidant status of type 2 diabetic rats. *International Journal of Molecular Sciences*. 13(10): 12952-12969.
- [74] Ip, J.H., Fuster, V., Badimon, L., Badimon, J., Taubman, M.B., and Chesebro, J.H. (1990). Syndromes of accelerated atherosclerosis: role of vascular injury and smooth muscle cell proliferation. *Journal of the American College of Cardiology*. 15(7): 1667-1687.
- [75] Ismail, M., Al-Naqeep, G., and Chan, K.W. (2010). Nigella sativa thymoquinone-rich fraction greatly improves plasma antioxidant capacity and expression of antioxidant genes in hypercholesterolemic rats. *Free Radical Biology and Medicine*. 48(5): 664-672.
- [76] Ismail, M., Al-Naqeeb, G., Mamat, W.A., and Ahmad, Z. (2010). Gamma-oryzanol rich fraction regulates the expression of antioxidant and oxidative stress related genes in stressed rat's liver. *Nutrition and Metabolism*. 7(23): 1-13.
- [77] Jacobs, D.R., Gross, M.D., and Tapsell, L.C. (2009). Food synergy: an operational concept for understanding nutrition. *The American Journal of Clinical Nutrition*. 89(5): 1543S-1548S.
- [78] Jenkins, D.J., Kendall, C.W., Vidgen, E., Augustin, L.S., van Erk, M., Geelen, A., Parker, T., Faulkner, D., Vladimir, V., Josse, R.G., Leiter, L.A., and Connelly, P.W. (2001). High-protein diets in hyperlipidemia: effect of wheat gluten on serum lipids, uric acid, and renal function. *The American Journal of Clinical Nutrition*. 74(1): 57-63.
- [79] Jia, J.H., Chen, K.P., Chen, S.X., Liu, K.Z., Fan, T.L., and Chen, Y.C. (2008). Breviscapine, a traditional Chinese medicine, alleviates myocardial ischaemia reperfusion injury in diabetic rats. *Acta Cardiologica*. 63(6): 757-762.
- [80] Jiang, F., Qian, J., Chen, S., Zhang, W., and Liu, C. (2011). Ligustrazine improves atherosclerosis in rat via attenuation of oxidative stress. *Pharmaceutical Biology*. 49(8): 856-863.
- [81] Kamisoyama, H., Honda, K., Tominaga, Y., Yokota, S., and Hasegawa, S. (2008). Investigation of the anti-obesity action of licorice flavonoid oil in diet-induced obese rats. *Bioscience, Biotechnology, and Biochemistry*. 72(12): 3225-3231
- [82] Kanda, T., and Takahashi, T. (2004). Interleukin-6 and cardiovascular diseases. *Japanese Heart Journal*. 45(2): 183-193.
- [83] Kang, Y.S., Cha, J.J., Hyun, Y.Y., and Cha, D.R. (2011). Novel CC chemokine receptor 2 antagonists in metabolic disease: a review of recent developments. *Expert Opinion on Investigational Drugs*. 20(6): 745-756.
- [84] Katcher, H.I., Legro, R.S., Kunselman, A.R., Gillies, P.J., Demers, L.M., Bagshaw, D.M., and Kris-Etherton, P.M. (2008). The effects of a whole grain–enriched hypocaloric diet on cardiovascular disease risk factors in men and women with metabolic syndrome. *The American Journal of Clinical Nutrition*. 87(1): 79-90.

- [85] Kathan, R.H., and Weeks, D.I. (1969). Structure studies of collocalia mucoid: I. Carbohydrate and amino acid composition. *Archives of Biochemistry and Biophysics*. 134(2): 572-576.
- [86] Khamzina, L., Veilleux, A., Bergeron, S., and Marette, A. (2005). Increased activation of the mammalian target of rapamycin pathway in liver and skeletal muscle of obese rats: possible involvement in obesity-linked insulin resistance. *Endocrinology*. 146(3): 1473-1481.
- [87] Khera, A.V., Cuchel, M., de la Llera-Moya, M., Rodrigues, A., Burke, M.F., Jafri, K., French, B.C., Phillips, J.A., Mucksavage, M.L., Wilensky, R.L., Mohler, E.R., Rothblat, G.H and Rader, D.J. (2011). Cholesterol efflux capacity, high-density lipoprotein function, and atherosclerosis. *New England Journal of Medicine*. 364(2): 127-135.
- [88] Khymenets, O., Fitó, M., Covas, M.I., Farré, M., Pujadas, M.A., Muñoz, D., Konstantinidou, V., and Torre, R.D.L. (2009). Mononuclear cell transcriptome response after sustained virgin olive oil consumption in humans: an exploratory nutrigenomics study. *OMICS A Journal of Integrative Biology*. 13(1): 7-19.
- [89] Kianbakht, S., and Hajiaghaee, R. (2011). Anti-hyperglycemic effects of saffron and its active constituents, crocin and safranal, in alloxaninduced diabetic rats. *Journal of Medicinal Plant.* 10(39): 82-89.
- [90] Kim, K.C., Kang, K.A., Lim, C.M., Park, J.H., Jung, K.S., and Hyun, J.W. (2012). Water extract of edible bird's nest attenuated the oxidative stress-induced matrix metalloproteinase-1 by regulating the mitogen-activated protein kinase and activator protein-1 pathway in human keratinocytes. *Journal of the Korean Society for Applied Biological Chemistry*. 55(3): 347-354.
- [91] Kohl 3rd, H.W. (2001). Physical activity and cardiovascular disease: evidence for a dose response. *Medicine and Science in Sports and Exercise*. 33(6 Suppl): S472-83.
- [92] Konstantinidou, V., Covas, M.I., Muñoz-Aguayo, D., Khymenets, O., de la Torre, R., Saez, G., Tormos, M.D.C., Toledo, E., Marti, A., Ruiz-Gutiérrez, V., Mendez, M.V.R., and Fito, M. (2010). *In vivo* nutrigenomic effects of virgin olive oil polyphenols within the frame of the Mediterranean diet: a randomized controlled trial. *The FASEB Journal*. 24(7): 2546-2557.
- [93] Koon, L.C. (2000). Features Bird's nest soup Market demand for this expensive gastronomic delicacy threatens the aptly named edible-nest SwiXets with extinction in the east. *Wildlife Conservation*. 103(1): 30–35.
- [94] Kou, J., Sun, Y., Lin, Y., Cheng, Z., Zheng, W., Yu, B., and Xu, Q. (2005). Anti-inflammatory activities of aqueous extract from Radix Ophiopogon japonicus and its two constituents. *Biological and Pharmaceutical Bulletin*. 28(7): 1234-1238.
- [95] Lamming, D.W., Ye, L., Katajisto, P., Goncalves, M.D., Saitoh, M., Stevens, D.M., Davis, J.M., Salmon, A.B., Sabatini, D.M., and Baur, J.A. (2012). Rapamycin-induced insulin resistance is mediated by mTORC2 loss and uncoupled from longevity. *Science*. 335(6076): 1638-1643.
- [96] Li, C.T., Wang, H.B., and Xu, B.J. (2013). A comparative study on anticoagulant activities of three Chinese herbal medicines from the genus Panax and anticoagulant activities of ginsenosides Rg1 and Rg2. *Pharmaceutical Biology*. 51(8): 1077-1080.
- [97] Li Xiong, S., Li, A., Huang, N., Lu, F., and Hou, D. (2011). Antioxidant and immunoregulatory activity of different polysaccharide fractions from tuber of Ophiopogon japonicus. *Carbohydrate Polymers*. 86(3): 1273-1280.

- [98] Libby, P. (2006). Inflammation and cardiovascular disease mechanisms. The *American Journal of Clinical Nutrition*. 83(2): 456S-460S.
- [99] Libby, P., Ridker, P.M., and Maseri, A. (2002). Inflammation and atherosclerosis. *Circulation*. 105(9): 1135-1143.
- [100] Liimatta, M., Towle, H.C., Clarke, S., and Jump, D.B. (1994). Dietary polyunsaturated fatty acids interfere with the insulin/glucose activation of L-type pyruvate kinase gene transcription. *Molecular Endocrinology*. 8(9): 1147-1153.
- [101] Lim, S.S., Vos, T., Flaxman, A.D., Danaei, G., Shibuya, K., Adair-Rohani, H., and Davis, A. (2013). A comparative risk assessment of burden of disease and injury attributable to 67 risk factors and risk factor clusters in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *The Lancet*. 380(9859): 2224-2260.
- [102] Lin, M.C., Hsu, P.C., and Yin, M.C. (2013). Protective effects of Houttuynia cordata aqueous extract in mice consuming a high saturated fat diet. *Food and Function*. 4(2): 322-327.
- [103] Lindberg, G., Eklund, G. A., Gullberg, B., and Råstam, L. (1991). Serum sialic acid concentration and cardiovascular mortality. *Bmj.* 302(6769): 143-146.
- [104] Lindbohm, N., Gylling, H., Miettinen, T.E., and Miettinen, T.A. (1999). Sialic acid content of LDL and lipoprotein metabolism in combined hyperlipidemia and primary moderate hypercholesterolemia. *Clinica Chimica Acta*. 285(1): 69-84.
- [105] Lithander, F.E., and Mahmud, A. (2015). The effect of red yeast rice on serum and haemodynamic risk factors for cardiovascular disease. *Proceedings of the Nutrition Society*, 74(OCE1): E77.
- [106] Liu, Z.B., Wu, L.J.T., Wang, H.S., Ding, Y.X., and Su, Y.F., (2009). Study on anti-inflammatory effects of kusnezoff monkshood leaf extracts. *Tianjin Journal of Traditional Chinese Medicine*, 26(1): 75-77.
- [107] Lozano, R., Naghavi, M., Foreman, K., Lim, S., Shibuya, K., Aboyans, V., and Cross, M. (2013). Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. The Lancet. 380(9859): 2095-2128.
- [108] Luepker, R.V., Murray, D.M., Jacobs Jr, D.R., Mittelmark, M.B., Bracht, N., Carlaw, R., Crow, R, Elmer, P, Finnegan, J and Folsom, A.R. (1994). Community education for cardiovascular disease prevention: risk factor changes in the Minnesota Heart Health Program. *American Journal of Public Health*. 84(9): 1383-1393.
- [109] Ma, F., and Liu, D. (2012). Sketch of the edible bird's nest and its important bioactivities. *Food Research International*. 48(2): 559-567.
- [110] Madamanchi, N.R., Vendrov, A., and Runge, M.S. (2005). Oxidative stress and vascular disease. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 25(1): 29-38
- [111] Malik, S., Wong, N.D., Franklin, S., Pio, J., Fairchild, C., and Chen, R. (2005). Cardiovascular disease in US patients with metabolic syndrome, diabetes, and elevated C-reactive protein. *Diabetes Care*. 28(3): 690-693.
- [112] Marwick, T.H. (2008). Diabetic heart disease. *Postgraduate Medical Journal*. 84(990): 188-192.
- [113] Matsukawa, N., Matsumoto, M., Bukawa, W., Chiji, H., Nakayama, K., Hara, H., and Tsukahara, T. (2011). Improvement of bone strength and dermal thickness due to dietary edible bird's nest extract in ovariectomized rats. *Bioscience, Biotechnology, and Biochemistry*. 75(3): 590-592.

- [114] Matthews, D.R., Hosker, J.P., Rudenski, A.S., Naylor, B.A., Treacher, D.F., and Turner, R.C. (1985). Homeostasis model assessment: insulin resistance and β-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*. 28(7): 412-419.
- [115] Maxwell, K.N., Soccio, R.E., Duncan, E.M., Sehayek, E., and Breslow, J.L. (2003). Novel putative SREBP and LXR target genes identified by microarray analysis in liver of cholesterol-fed mice. *Journal of Lipid Research*. 44(11): 2109-2119.
- [116] McCullough, M. L., Feskanich, D., Rimm, E. B., Giovannucci, E. L., Ascherio, A., Variyam, J. N., ... & Willett, W. C. (2000). Adherence to the Dietary Guidelines for Americans and risk of major chronic disease in men. The *American Journal of Clinical Nutrition*, 72(5), 1223-1231.
- [117] Mendis, S., Puska, P., and Norrving, B. (2011). Global atlas on cardiovascular disease prevention and control. *World Health Organization*.
- [118] Mohamad, S. (2013). Effect of Nigella sativa seeds and honey mixture on cardiovascular disease risk factors in hypercholesterolemic adults. Doctoral dissertation, Terengganu: Universiti Malaysia Terengganu.
- [119] Mohamed, A.K., Bierhaus, A., Schiekofer, S., Tritschler, H., Ziegler, R., and Nawroth, P.P. (1999). The role of oxidative stress and NF κB activation in late diabetic complications. *Biofactors*. 10(2-3): 157-167.
- [120] Moncada, S., and Vane, J.R. (1979). The role of prostacyclin in vascular tissue. *In Federation Proceedings*. 38(1): 66-71.
- [121] Morrow, J.D. (2005). Quantification of isoprostanes as indices of oxidant stress and the risk of atherosclerosis in humans. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 25(2): 279-286.
- [122] Müller, M., and Kersten, S. (2003). Nutrigenomics: goals and strategies. *Nature Reviews Genetics*. 4(4): 315-322.
- [123] Nagandran, K., Hui, C.K., Tjih, E.T.T., and Hun, L.T. (2009). Edible bird's nest extract rejuvenate skin by increased fibroblast proliferation and skin matrixes synthesis. *In: International Conference on Biotechnology for the Wellness Industry (ICBWI)*, 2009, PWTC Kuala Lumpur.
- [124] Nagandran, K., Hui, C.K., Yahaya, M., Hamdan, N., Tin, L.C., and Hun, L.T. (2009). Edible bird's nest extract promoted cartilage matrix expression and suppressed the catabolic genes expression in osteoarthritic chondrocytes. *In: Proceedings of the 2nd ICBWI*, PWTC Kuala Lumpur.
- [125] Narayanan, S. (1994). Sialic acid as a tumor marker. *Annals of Clinical and Laboratory Science*. 24(4): 376-384.
- [126] Nigam, P.K., Narain, V.S., and Kumar, A. (2006). Sialic acid in cardiovascular diseases. *Indian Journal of Clinical Biochemistry*. 21(1): 54-61.
- [127] Nowak, D., Kałucka, S., Białasiewicz, P., and Król, M. (2001). Exhalation of H 2 O 2 and thiobarbituric acid reactive substances (TBARs) by healthy subjects. *Free Radical Biology and Medicine*. 30(2): 178-186.
- [128] O'Keefe, J. H., & Cordain, L. (2004). Cardiovascular disease resulting from a diet and lifestyle at odds with our Paleolithic genome: how to become a 21st-century hunter-gatherer. *In Mayo Clinic Proceedings* 79(1),101-108.
- [129] Olefsky, J.M. (1976). The insulin receptor: its role in insulin resistance of obesity and diabetes. *Diabetes*. 25(12): 1154-1161.
- [130] Owens, A.P., Byrnes, J.R., and Mackman, N. (2014). Hyperlipidemia, tissue factor, coagulation, and simvastatin. *Trends in Cardiovascular Medicine*. 24(3): 95-98.

- [131] Ozanne, S.E., Olsen, G.S., Hansen, L.L., Tingey, K.J., Nave, B.T., Wang, C.L., Hartil, K., Petry, C.J., Buckley, A.J., and Mosthaf-Seedorf, L. (2003). Early growth restriction leads to down regulation of protein kinase C zeta and insulin resistance in skeletal muscle. *Journal of Endocrinology*. 177(2): 235-241.
- [132] Oztürk, L.K., Yarat, A., and Emekli, N. (2006). Effect of fenofibrate on serum and tissue sialic acid levels in short-term experimental hypercholesterolemia. *Arzneimittel-Forschung*. 57(12): 770-776.
- [133] Papathanassoglou, E., El-Haschimi, K., Li, X.C., Matarese, G., Strom, T., and Mantzoros, C. (2006). Leptin receptor expression and signaling in lymphocytes: kinetics during lymphocyte activation, role in lymphocyte survival, and response to high fat diet in mice. *The Journal of Immunology*. 176(12): 7745-7752.
- [134] Pari, L., and Latha, M. (2006). Antihyperlipidemic effect of Scoparia dulcis (sweet broomweed) in streptozotocin diabetic rats. *Journal of Medicinal Food*. 9(1): 102-107.
- [135] Patel, S.B., Reams, G.P., Spear, R.M., Freeman, R.H., and Villarreal, D. (2008). Leptin: linking obesity, the metabolic syndrome, and cardiovascular disease. *Current Hypertension Reports*. 10(2): 131-137.
- [136] Pawlak, K., Naumnik, B., Brzósko, S., Pawlak, D., and Myśliwiec, M. (2003). Oxidative stress-a link between endothelial injury, coagulation activation, and atherosclerosis in haemodialysis patients. *American Journal of Nephrology*. 24(1): 154-161.
- [137] Poirier, S., Mayer, G., Benjannet, S., Bergeron, E., Marcinkiewicz, J., Nassoury, N., Mayer, H., Nimpf, J., Prat, A., and Seidah, N.G. (2008). The proprotein convertase PCSK9 induces the degradation of low density lipoprotein receptor (LDLR) and its closest family members VLDLR and ApoER2. *Journal of Biological Chemistry*, 283(4), 2363-2372.
- [138] Popa, C., Netea, M.G., Van Riel, P.L., van der Meer, J.W., and Stalenhoef, A.F. (2007). The role of TNF-α in chronic inflammatory conditions, intermediary metabolism, and cardiovascular risk. *Journal of Lipid Research*. 48(4): 751-762.
- [139] Purkayastha, S., Zhang, G., and Cai, D. (2011). Uncoupling the mechanisms of obesity and hypertension by targeting hypothalamic IKK-[beta] and NF-[kappa] B. *Nature Medicine*. 17(7): 883-887.
- [140] Qin, L., Peng, X., Zhang, S.H., Wang, L., and Liu, F. (2000). Influence of monkshood root-peony root combination on inflamation-induced agents and free radicals. *China Journal of Chinese Materia Medica*. 25(6): 370.
- [141] Raines, E.W., and Ross, R. (1993). Smooth muscle cells and the pathogenesis of the lesions of atherosclerosis. *British Heart Journal*. 69(1 Suppl): S30.
- [142] Rajappa, M., Ikkruthi, S., Nandeesha, H., Satheesh, S., Sundar, I., Ananthanarayanan, P.H., and Harichandrakumar, K.T. (2013). Relationship of raised serum total and protein bound sialic acid levels with hyperinsulinemia and indices of insulin sensitivity and insulin resistance in non-diabetic normotensive obese subjects. *Diabetes and Metabolic Syndrome: Clinical Research and Reviews*. 7(1): 17-19.
- [143] Rashed, A.A., and Nazaimoon, W.W. (2010). Effect of Edible Bird's Nest on Caco-2 Cell Proliferation. *Journal of Food Technology*. 8(3): 126-130.
- [144] Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., and Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology and Medicine*. 26(9): 1231-1237.

- [145] Ridker, P.M., Buring, J.E., Cook, N.R., and Rifai, N. (2003). C-reactive protein, the metabolic syndrome, and risk of incident cardiovascular events an 8-year follow-up of 14 719 initially healthy American women. *Circulation*. 107(3): 391-397.
- [146] Ritchie, M.E. (1998). Nuclear factor-κB is selectively and markedly activated in humans with unstable angina pectoris. *Circulation*. 98(17): 1707-1713.
- [147] Roberts, L.J., and Morrow, J.D. (2000). Measurement of F 2-isoprostanes as an index of oxidative stress *in vivo*. *Free Radical Biology and Medicine*. 28(4): 505-513.
- [148] Rocha-Pereira, P., Santos-Silva, A., Rebelo, I., Figueiredo, A., Quintanilha, A., and Teixeira, F. (2001). Dislipidemia and oxidative stress in mild and in severe psoriasis as a risk for cardiovascular disease. *Clinica Chimica Acta*. 303(1): 33-39.
- [149] Roh, K.B., Lee, J., Kim, Y.S., Park, J., Kim, J.H., Lee, J., and Park, D. (2011). Mechanisms of edible bird's nest extract-induced proliferation of human adipose-derived stem cells. *Evidence-Based Complementary and Alternative Medicine*. 2012: 797520.
- [150] Rosenkranz, A.R., Schmaldienst, S., Stuhlmeier, K.M., Chen, W., Knapp, W., and Zlabinger, G.J. (1992). A microplate assay for the detection of oxidative products using 2', 7'-dichlorofluorescin-diacetate. *Journal of Immunological Methods*. 156(1): 39-45.
- [151] Ross, R. (1993). The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature*. 362: 801 809.
- [152] Rudijanto, A. (2007). The role of vascular smooth muscle cells on the pathogenesis of atherosclerosis. *Acta Medica Indonesiana*. 39(2): 86-93.
- [153] Ryan, D., Kendall, M., and Robards, K. (2007). Bioactivity of oats as it relates to cardiovascular disease. *Nutrition Research Reviews*. 20(02): 147-162.
- [154] Rydén, L., Grant, P.J., Anker, S.D., Berne, C., Cosentino, F., Danchin, N., Deaton, C., Escaned, J., Hammes, H.P., Huikuri, H., Marre, M., Marx, N., Mellbin, L., Ostergren, J., Patrono, C., Seferovic, P., Uva, M.S., Taskinen, M.R., Tendera, M., Tuomilehto, J., Valensi, P., and Zamorano, J.L. (2014). ESC Guidelines on diabetes, pre-diabetes, and cardiovascular diseases developed in collaboration with the EASD. European Heart Journal. 35(27): 1824.
- [155] Sadikario, S., Donev, D., and Kragelj, L.Z. (2007). Cardiovascular diseases prevention and control. *Health Promotion and Disease Prevention*. 2007: 531.
- [156] Saengkrajang, W., and Matan, N. (2011). Antimicrobial activities of the edible bird's nest extracts against food-borne pathogens. *Thai Journal of Agricultural Science*. 44(5): 326-330.
- [157] Sampathkumar, S.G., Li, A., and Yarema, K.J. (2006). Sialic acid and the central nervous system: perspectives on biological functions, detection, imaging methods and manipulation. *CNS and Neurological Disorders-Drug Targets*. 5(4): 425-440.
- [158] Saraswat, M., Suryanarayana, P., Reddy, P.Y., Patil, M.A., Balakrishna, N., and Reddy, G.B. (2010). Antiglycating potential of Zingiber officinalis and delay of diabetic cataract in rats. *Molecular Vision*. 16: 1525.
- [159] Schmid, J.A., and Birbach, A. (2008). IκB kinase β (IKKβ/IKK2/IKBKB)—A key molecule in signaling to the transcription factor NF-κB. *Cytokine and Growth Factor Reviews*. 19(2): 157-165.

- [160] Schmitz-Peiffer, C. (2002). Protein kinase C and lipid induced insulin resistance in skeletal muscle. *Annals of the New York Academy of Sciences*. 967(1): 146-157.
- [161] Shah, A., Mehta, N., and Reilly, M.P. (2008). Adipose inflammation, insulin resistance, and cardiovascular disease. *Journal of Parenteral and Enteral Nutrition*. 32(6): 638-644.
- [162] Shen, Z.Q., Chen, P., Duan, L., Dong, Z.J., Chen, Z.H., and Liu, J.K. (2004). Effects of fraction from Phyllanthus urinaria on thrombosis and coagulation system in animals. *Journal of Chinese Integrative Medicine*. 2(2): 106-110.
- [163] Shikany, J. M., & White Jr, G. L. (2000). Dietary guidelines for chronic disease prevention. *Southern Medical Journal*, 93(12), 1138-1151.
- [164] Shukri, R., Mohamed, S., and Mustapha, N.M. (2010). Cloves protect the heart, liver and lens of diabetic rats. *Food Chemistry*. 122(4): 1116-1121.
- [165] Sprenger, N., Julita, M., Donnicola, D., and Jann, A. (2009). Sialic acid feeding aged rats rejuvenates stimulated salivation and colon enteric neuron chemotypes. *Glycobiology*. 19(12): 1492-1502.
- [166] St-Onge, M.P., Aban, I., Bosarge, A., Gower, B., Hecker, K.D., and Allison, D.B. (2007). Snack chips fried in corn oil alleviate cardiovascular disease risk factors when substituted for low-fat or high-fat snacks. *The American Journal of Clinical Nutrition*. 85(6): 1503-1510.
- [167] Staprans, I., Pan, X.M., Rapp, J.H., Grunfeld, C., and Feingold, K.R. (2000). Oxidized cholesterol in the diet accelerates the development of atherosclerosis in LDL receptor—and apolipoprotein E—deficient mice. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 20(3): 708-714.
- [168] Stephens, A.M., Dean, L.L., Davis, J.P., Osborne, J.A., and Sanders, T.H. (2010). Peanuts, peanut oil, and fat free peanut flour reduced cardiovascular disease risk factors and the development of atherosclerosis in Syrian golden hamsters. *Journal of Food Science*. 75(4): H116-H122.
- [169] Stephens, J.M., Lee, J., and Pilch, P.F. (1997). Tumor necrosis factor-α-induced insulin resistance in 3T3-L1 adipocytes is accompanied by a loss of insulin receptor substrate-1 and GLUT4 expression without a loss of insulin receptor-mediated signal transduction. *Journal of Biological Chemistry*. 272(2): 971-976.
- [170] Su, Q., Tsai, J., Xu, E., Qiu, W., Bereczki, E., Santha, M., and Adeli, K. (2009). Apolipoprotein B100 acts as a molecular link between lipid induced endoplasmic reticulum stress and hepatic insulin resistance. *Hepatology*. 50(1): 77-84.
- [171] Subramaniam, S., Fahy, E., Gupta, S., Sud, M., Byrnes, R.W., Cotter, D., Dinasarapu, A.R and Maurya, M.R. (2011). Bioinformatics and systems biology of the lipidome. *Chemical Reviews*. 111(10): 6452-6490.
- [172] Sultan, M.T., Butt, M.S., and Anjum, F.M. (2009). Safety assessment of black cumin fixed and essential oil in normal Sprague Dawley rats: Serological and hematological indices. *Food and Chemical Toxicology*. 47(11): 2768-2775.
- [173] Surdo, P.L., Bottomley, M.J., Calzetta, A., Settembre, E.C., Cirillo, A., Pandit, S., Ni, Y.G., Hubbard, B., Sitlani, A and Carfí, A. (2011). Mechanistic implications for LDL receptor degradation from the PCSK9/LDLR structure at neutral pH. *EMBO Reports*. 12(12): 1300-1305.

- [174] Tachibana, N., Matsumoto, I., Fukui, K., Arai, S., Kato, H., Abe, K., and Takamatsu, K. (2005). Intake of soy protein isolate alters hepatic gene expression in rats. *Journal of Agricultural and Food Chemistry*. 53(10): 4253-4257.
- [175] Tanasescu, M., Leitzmann, M.F., Rimm, E.B., Willett, W.C., Stampfer, M.J., and Hu, F.B. (2002). Exercise type and intensity in relation to coronary heart disease in men. *Jama*. 288(16): 1994-2000.
- [176] Taylor, R.S., Brown, A., Ebrahim, S., Jolliffe, J., Noorani, H., Rees, K., Skidmore, B., Stone, J.A., thompson, D.R., and Oldridge, N. (2004). Exercise-based rehabilitation for patients with coronary heart disease: systematic review and meta-analysis of randomized controlled trials. *The American Journal of Medicine*. 116(10): 682-692.
- [177] Thu, V.T., Kim, H.K., Ha, S.H., Yoo, J.Y., Park, W.S., Kim, N., Oh, G.T., and Han, J. (2010). Glutathione peroxidase 1 protects mitochondria against hypoxia/reoxygenation damage in mouse hearts. *Pflügers Archiv-European Journal of Physiology*. 460(1): 55-68.
- [178] Traving, C., and Schauer, R. (1998). Structure, function and metabolism of sialic acids. *Cellular and Molecular Life Sciences*. 54(12): 1330-1349.
- [179] Trujillo, E., Davis, C., and Milner, J. (2006). Nutrigenomics, proteomics, metabolomics, and the practice of dietetics. *Journal of the American Dietetic Association*. 106(3): 403-413.
- [180] Ulmius, M., Johansson-Persson, A., Krogh, M., Olsson, P., and Önning, G. (2011). An oat bran meal influences blood insulin levels and related gene sets in peripheral blood mononuclear cells of healthy subjects. *Genes and Nutrition*. 6(4): 429-439.
- [181] Vahanian, A., Alfieri, O., Andreotti, F., Antunes, M.J., Barón-Esquivias, G., Baumgartner, H., Borger, M.A., Carrel, T.P., Bonis, M.D., and Habib, G. (2012). Guidelines on the management of valvular heart disease (version 2012). *European Heart Journal*. 33(19): 2451-2496.
- [182] Van Gaal, L.F., Mertens, I.L., and Christophe, E. (2006). Mechanisms linking obesity with cardiovascular disease. *Nature*. 444(7121): 875-880.
- [183] Van Schie, M.C., Van Loon, J.E., De Maat, M.P.M., and Leebeek, F.W.G. (2011). Genetic determinants of von Willebrand factor levels and activity in relation to the risk of cardiovascular disease: a review. *Journal of Thrombosis and Haemostasis*. 9(5): 899-908.
- [184] Varki, A. (2008). Sialic acids in human health and disease. *Trends in Molecular Medicine*. *14*(8): 351-360.
- [185] Vimala, B., Hussain, H., and Nazaimoon, W.W. (2012). Effects of edible bird's nest on tumour necrosis factor-alpha secretion, nitric oxide production and cell viability of lipopolysaccharide-stimulated RAW 264.7 macrophages. *Food and Agricultural Immunology*. 23(4): 303-314.
- [186] Vischer, U.M. (2006). von Willebrand factor, endothelial dysfunction, and cardiovascular disease. *Journal of Thrombosis and Haemostasis*. 4(6): 1186-1193.
- [187] Wang, B. (2009). Sialic acid is an essential nutrient for brain development and cognition. *Annual Review of Nutrition*. 29: 177-222.
- [188] Weintraub, W.S., Daniels, S.R., Burke, L.E., Franklin, B.A., Goff, D.C., Hayman, L.L., Lloyd-Jones, D., Pandey, D.K., Sanchez, E.J., Schram, A.P and Whitsel, L.P. (2011). Value of primordial and primary prevention for cardiovascular disease a policy statement from the American Heart Association. *Circulation*. 124(8): 967-990.

- [189] Wendel, A. (1980). Glutathione peroxidase. *Enzymatic Basis of Detoxication*. 1: 333-353.
- [190] Joint WHO/FAO Expert Consultation on Diet, Nutrition and the Prevention of Chronic Diseases (2003). World Health Organization technical report series. 916(i-viii), Geneva, Switzerland.
- [191] Wilson, P.W., D'Agostino, R.B., Levy, D., Belanger, A.M., Silbershatz, H., and Kannel, W.B. (1998). Prediction of coronary heart disease using risk factor categories. *Circulation*. 97(18): 1837-1847.
- [192] Woodhill, J.M., Palmer, A.J., Leelarthaepin, B., McGilchrist, C., and Blacket, R.B. (1978). Low fat, low cholesterol diet in secondary prevention of coronary heart disease. *In Drugs, Lipid Metabolism, and Atherosclerosis.* 109: 317-330.
- [193] World Health Organization. (2015). Cardiovascular diseases. Accessed 29 October, 2015 (http://www.who.int/mediacentre/factsheets/fs317/en/).
- [194] Wu, Y.J., Hong, C.Y., Lin, S.J., Wu, P., and Shiao, M.S. (1998). Increase of vitamin E content in LDL and reduction of atherosclerosis in cholesterol-fed rabbits by a water-soluble antioxidant-rich fraction of Salvia miltiorrhiza. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 18(3): 481-486.
- [195] Xu, J., Wang, Y., Xu, D.S., Ruan, K.F., Feng, Y., and Wang, S. (2011). Hypoglycemic effects of MDG-1, a polysaccharide derived from Ophiopogon japonicas, in the ob/ob mouse model of type 2 diabetes mellitus. *International Journal of Biological Macromolecules*. 49(4): 657-662.
- [196] Yap, T.A., Garrett, M.D., Walton, M.I., Raynaud, F., de Bono, J.S., and Workman, P. (2008). Targeting the PI3K–AKT–mTOR pathway: progress, pitfalls, and promises. *Current Opinion in Pharmacology*. 8(4): 393-412.
- [197] Yew, M.Y., Koh, R.Y., Chye, S.M., Othman, I., and Ng, K.Y. (2014). Edible bird's nest ameliorates oxidative stress-induced apoptosis in SH-SY5Y human neuroblastoma cells. *BMC Complementary and Alternative Medicine*. 14(1): 391.
- [198] Young, M.E., McNulty, P., and Taegtmeyer, H. (2002). Adaptation and maladaptation of the heart in diabetes: part II potential mechanisms. *Circulation*. 105(15): 1861-1870.
- [199] Zhao, G.R., Xiang, Z.J., Ye, T.X., Yuan, Y.J., and Guo, Z.X. (2006). Antioxidant activities of Salvia miltiorrhiza and Panax notoginseng. *Food Chemistry*. 99(4): 767-774.
- [200] Zhou, Q.S., Zhao, Y.M., Bai, X., Li, P.X., and Ruan, C.G. (1992). Effect of new-breviscapine on fibrinolysis and anticoagulation of human vascular endothelial cells. *Acta Pharmacologica Sinica*. 13(3): 239-242.