

UNIVERSITI PUTRA MALAYSIA

CYTOTOXICITY OF SRJ23 AND ITS DERIVATIVES, EXPRESSION AND SRJ23 BINDING OF KRAS G12V ONCOPROTEIN

MICHELLE TAN SIYING

FPSK(M) 2018 13



CYTOTOXICITY OF SRJ23 AND ITS DERIVATIVES, EXPRESSION AND SRJ23 BINDING OF KRAS G12V ONCOPROTEIN

By

MICHELLE TAN SIYING

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

December 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

CYTOTOXICITY OF SRJ23 AND ITS DERIVATIVES, EXPRESSION AND SRJ23 BINDING OF KRAS G12V ONCOPROTEIN

By

MICHELLE TAN SIYING

December 2017

Chair: Professor Johnson Stanslas, PhD Faculty: Medicine and Health Sciences

Oncogenic KRas signaling is often associated with a poor prognosis of pancreatic cancer. Effort to target oncogenic KRas signaling persists for years but without much success due to its 'undruggable' property. In recent years, several small-molecule KRas inhibitors such as SRJ23 were developed to inhibit oncogenic KRas signaling at the guanine nucleotide exchange level. MTT cell viability assay was used to evaluate the cytotoxicity of SRJ23 and its derivatives towards breast, colon, prostate and pancreatic cancer cell lines. SRJ23 is a semi-synthetic derivative of andrographolide (AGP) which lacks a distinct selectivity towards specific cancer type by demonstrating an equally good cytotoxicity in breast, colon, prostate, and pancreatic cancer cell lines. A few novel derivatives of SRJ23 were synthesised to improve its target specificity. One of the derivatives (SRS 151) shows selective total growth inhibition on pancreatic cancer cells harbouring oncogenic KRas (MIA PaCa-2 and Capan-2). SRS157 (GI₅₀ = 2.4 μ M), although is as equally potent as SRJ23 (Gl₅₀ = 2.1 µM) in pancreatic cancer cell lines, was found selectively targeting pancreatic cancer. Previous study has revealed a direct binding of SRJ23 to KRas in silico. To validate a physical interaction between oncogenic KRas G12V and SRJ23 in vitro, KRas G12V was expressed by using Champion pET SUMO protein expression system and was used for X-ray crystallography to study the physical interaction between KRas G12V and SRJ23. The hanging drop vapour diffusion method in X-ray crystallography did not yield diffractable protein crystals, therefore saturation transfer difference-nuclear magnetic resonance (STD-NMR) was applied to study KRas G12V-SRJ23 interaction. The STD-NMR suggested a potential physical interaction between KRas G12V and SRJ23 that involves mainly the three-membered ring and the hydroxyl group on the lactone ring of SRJ23. In conclusion, the findings of this study showed that SRJ23 could be a promising anti-Ras drug.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

SITOTOKSIKSITI SRJ23 DAN DERIVATIF, EKSPRESI DAN INTERAKSI SRJ23 DENGAN KRAS G12V ONKOPROTEIN

Oleh

MICHELLE TAN SIYING

Disember 2017

Pengerusi: Profesor Johnson Stanslas, PhD Fakulti: Perubatan dan Sains Kesihatan

Isyarat sel berunsurkan KRas onkogenik sering dihubungkait dengan ramalan perkembangan kanser pancreas yang tidak menyenangkan hati. Usaha membanteras isyarat Ras onkogenik telah dijalankan bertahun-tahun tetapi tidak berjaya kerana sifat Ras yang sukar disasarkan oleh perencat. Sejak kebelakangan ini, beberapa molekul kecil seperti SRJ23 yang berpotensi menjadi perencat isyarat Ras onkogenik pada tahap 'guanine nucleotide exchange' telah ditemui. Ujian sitotoksik MTT digunakan untuk menentu aktiviti sitotoksik SRJ23 dan terbitannya terhadap payudara, kolon, prostat dan SRJ23 kompaun sintetik yang diterbit daripada pankreas. ialah andrographolide (AGP). Kompaun ini tidak mempunyai sasaran kanser yang nyata dengan menunjukkan aktiviti sitotoksik yang setara terhadap sel-sel kanser payudara, kolon, prostat dan pankreas. Beberapa terbitan SRJ23 telah disintesis agar terbitannya menyasarkan jenis kanser tertentu. Salah satu terbitannya (SRS151) didapati hanya bertindak ke atas pertumbuhan sel-sel kanser pankreas yang bergantung kepada ekspresi KRas onkogenik (MIA PaCa-2 and Capan-2). SRS157(GI₅₀ = 2.4 μ M) menunjukkan aktiviti sitotoksitik yang setanding SRJ23 (GI₅₀ = 2.1 μ M) ke atas sel-sel kanser pankreas, tetapi aktiviti sitotoksiknya adalah selektif ke atas kanser pankreas jika dibandingkan SRJ23. Kajian terdahulu membuktikan kewujudan ikatan langsung antara KRas dan SRJ23 in silico. Untuk mengesahkan interaksi fizikal antara protein onkogenik Kras G12V dan SRJ23 secara in vitro, KRas G12V dihasilkan melalui sistem ekspresi protein yang dikenali sebagai Champion pET SUMO dan digunakan dalam X-ray kristalografi untuk mengkaji kewujudan interaksi fizikal antara protein onkogenik KRas G12V dan SRJ23. Disebabkan oleh Xray kristalografi menggunakan teknik 'hanging drop vapour diffusion' tidak menghasilkan kristal protein yang sesuai untuk diffraksi, teknik ini telah diganti dengan 'saturation transfer difference-nuclear magnetic resonance' (STD-NMR) untuk mengkaji interaksi KRas G12V-SRJ23. Kajian STD-NMR



mencadangkan kewujudan interaksi antara KRas G12V dan SRJ23. Interaksi ini melibatkan bahagian-bahagian pada struktur SRJ23 iaitu 'three-membered ring' dan kumpulan hidroksil yang terikat pada 'lactone ring'. Pada kesimpulannya, hasil kajian ini menunjukkan SRJ23 berpotensi menjadi ubat anti-Ras.



ACKNOWLEDGEMENTS

Firstly, I would like to extend my heartfelt gratitude to my supervisor, Prof. Dr. Johnson Stanslas for providing me with this great opportunity to work on this meaningful research project. I would like to express my sincere appreciation to him for his valuable advices, suggestions and guidance which had made the completion of this thesis possible. Without his critical and encouraging support of my academic development over these years, I would not be able to complete the research project. I am grateful for his patience, dedication, intellectual support, trust and confidence in my work. I am truly thankful for his utmost understanding and caring when I was traumatised by the most painful life event that I have ever experienced.

I would also like to express my sincere thanks and appreciation to my cosupervisor, Dr. Ho Kok Lian, who had guided me through the work of protein expression and purification. I am grateful for his extensive help in taking time to read and comment on my thesis. I would like to thank Dr. Sreenivasa Rao Sagineedu for his guidance, support and valuable feedback during the investigation of this research work.

I am also thankful for the learning experience which I have gained from working with all the caring laboratory staff and lab mates especially Gracie Chia and Beh Poay Ling, and Dr. Azira Muhamad from Malaysia Genome Institute. I am grateful for the wonderful times we had in the laboratory and their kind assistance in guiding me to use some of the laboratory equipment.

Special thanks and appreciation to Teh Yuan Han, for supporting and inspiring me relentlessly with his genuine enthusiasm in research, while being a good friend of mine along the way to accomplish my Master's research.

Last but not least, I would like to thank my family members and my beloved late father, to whom I dedicate this thesis, for their stimulating encouragement, support and love which enabled me to complete the research.

I certify that a Thesis Examination Committee has met on 29 December 2017 to conduct the final examination of Michelle Tan Siying on her thesis entitled "CYTOTOXICITY OF SRJ23 AND ITS DERIVATIVES, EXPRESSION AND SRJ23 BINDING OF KRAS G12V ONCOPROTEIN" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science. Members of the Thesis Examination Committee were as follows:

Rajesh Ramasamy, PhD

Associate Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Chairman)

Subashini Chellappah Thambiah, PhD

Associate Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Internal Examiner)

Aman Shah bin Abdul Majid, PhD

Associate Professor Faculty of Medicine Quest International University Perak (Qiup) City Campus Malaysia (External Examiner)

> NOR AINI AB. SHUKOR, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 27 February 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Johnson Stanslas, PhD

Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Ho Kok Lian, PhD

Senior Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Sreenivasa Rao Sagineedu, PhD

Senior Lecturer Faculty of Pharmacy International Medical University (Member)



ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Michelle Tan Siying (GS35883)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013)are adhered to.

Signature	:
Name of Chairman of Supervisory Committee	: <u>Professor Dr. Johnson Stanslas</u>
Signature	:
Name of Member of Supervisory Committee	: <u>Dr. Ho Kok Lian</u>
Signature	
Name of Member of Supervisory Committee	: <u>Dr. Sreenivasa Rao Sagineedu</u>

TABLE OF CONTENTS

ABSTRACT ABSTRAK ACKNOWLI APPROVAL DECLARAT LIST OF TA LIST OF FIC LIST OF AB	EDGE ION BLES GURE	S S			Page i iv v vii xiii xiv xvii
CHAPTER					
1	INTE	RODUC	TION		
		Overvi			1
	1.2	Hypoth			4
	1.3	Object			4
	1.4	Specif	ic Objectiv	es	5
2					
2	2.1		RE REVIEV	(AGP) And Its Derivatives	6
	2.2		• •	Sarcoma Viral Oncogene	7
	2.2	Homo		Saroonia virai choogene	'
	2.3			TP Hydrolysis And Ras Signaling	9
	2.4			tions In Ras	11
	2.5		as Agents		14
	2.6			Expression System	16
	2.7			ein Purification	19
	2.8		stallization		20
	2.9			fer Difference-Nuclear Magnetic	22
		G12V	lance (STL	D-NMR) Of Compound-KRas	
		GIZV			
3	IN V		YTOTOXI	C ACTIVITIES OF SRJ23 AND	
		DERIVA			
	3.1	Introdu	uction		24
	3.2	Materi	als And Me	ethodology	25
		3.2.1	Materials		25
			3.2.1.1	Test Compounds	25
			3.2.1.2	Cell Lines	25
			3.2.1.3	Chemicals And Reagents	27
		3.2.2	Methodol		27
			3.2.2.1	Preparation Of Test	27
			3.2.2.2	Compounds Plating	27
			3.2.2.2	Compound Treatments	28
			3.2.2.3	MTT Cell Viability Assay	28
			3.2.2.5	Statistical Analysis	29
	3.3	Result			30

(C)

	3.3.1		rowth Inhibitory Effects Of SRJ23 erivatives Against Selected	30
3.4	Discus		ell Lines	39
3.5	Summ			40
IMM	UNODE	ETECTION	ION, PURIFICATION, , CHARACTERISATION AND F KRAS G12V PROTEIN	
4.1	Introdu			42
4.2	Materi	als And Me	ethodology	44
	4.2.1	Materials		44
		4.2.1.1	Recombinant DNA, Vector And Host	44
		4.2.1.2	Primers	44
		4.2.1.3	Chemicals And Molecular Biology Reagents	45
		4.2.1.4	Equipment And Instrumentation	46
	4.2.2	Methodol		47
		4.2.2.1	DNA Sequencing Of pEXV KRas G12V	47
		4.2.2.2	Gene Amplification	47
		4.2.2.3	Purification Of PCR Products	47
		4.2.2.4	Ligation Of <i>KRas G12V</i> Into pET SUMO	48
		4.2.2.5	Transformation And Plasmid Isolation For pET SUMO KRas G12V	48
		4.2.2.6	Plasmid Verification	48
		4.2.2.7	Pilot Expression And Large	49
		T.Z.Z.1	Scale Protein Expression For Purification	70
		4.2.2.8	Protein Purification Using	50
		4.2.2.0	Immobilized Metal-ion Affinity	00
		4.2.2.9	Chromatography (IMAC) Buffer Exchange And Cleavage Of N-terminal SUMO Fusion	50
		4.2.2.10	Protein By SUMO Protease Reverse IMAC To Remove The Cleaved N-Terminal SUMO Fusion Protein	50
		4.2.2.11	GDP Loading Of Purified Protein	51
		4.2.2.12	Native-PAGE Electrophoresis	51
		4.2.2.13	SDS-PAGE Electrophoresis	51
		4.2.2.14	Western Blot	52
		4.2.2.15	Identification Of Recombinant KRas Using Anti-human KRas Rabbit Polyclonal Antibody	53

		4.2.2.16	Quantification Of Purified KRas G12V Protein Concentration And Determination Of Their Molecular Weight	53
4.3	Result	s	3	54
	4.3.1	DNA And	PCR Verification Sequencing Of Ras G12V Plasmid	54
	4.3.2	Optimisa	tion And Pilot Expression In The n <i>Escherichia coli</i> And Cell Lysis	58
	4.3.3		on Of Recombinant KRas G12V	59
	4.3.4		rotease Digestion And Purification	61
	4.3.5	Verificatio	on Of Complete GDP Loading Of (Ras G12V	65
	4.3.6		Blot Analysis	67
	4.3.7		letection Of KRas G12V With	67
	1.0.1		an KRas Rabbit Polyclonal	01
	4.3.8		ation Of Purified KRas G12V	68
			oncentration	
4.4	Discus	ssion		68
4.5	Summ	nary		77
SAT MAC		ON TRANS	RYSTALLOGRAPHY & SFER DIFFERENCE-NUCLEAR NCE APPROACHES	70
5.1	Introd		a the shall a sur	78
5.2	5.2.1		ethodology	79 79
	5.2.1	5.2.1.1	Test Compounds	79
		5.2.1.1	Chemicals And Reagents	79
		5.2.1.2	Equipment And Instrumentation	80
		5.2.1.4	Crystallization Tools	80
		5.2.1.5	Software For NMR Analysis	81
5.3	Metho	dology	,	81
	5.3.1		creening Of GDP-bound KRas	81
		G12V	C C	
		5.3.1.1	Buffer Exchange Of Protein Into Crystallization Buffer	81
		5.3.1.2	Siliconizing The Coverslips For Hanging Drop Set-ups	81
		5.3.1.3	Hanging Drop Vapour Diffusion Method	81
	5.3.2	5.3.1.4 STD-NM	Crystallization Conditions	81 82
		5.3.2.1	Preparation Of Stock Concentration Of SRJ23, DCAI	82

C

			5.3.2.2	And Vismodegib Buffer Exchange Of Protein KRas G12V Into Deuterated	82
				NMR Buffer	
			5.3.2.3		82
			5.3.2.4	Bound KRas G12V STD-NMR Of DCAI-GDP Bound KRas G12V	83
			5.3.2.5	STD-NMR of Vismodegib-GDP Bound KRas G12V	83
	5.4	Result	S		84
		5.4.1		Screening Of Purified Protein With GDP	84
		5.4.2	STD-NIV	IR Of SRJ23 And KRas G12V	84
		5.4.3	STD-NIV	R Of DCAI And KRas G12V	89
		5.4.4	STD-NM G12V	IR Of Vismodegib And KRas	93
	5.5	Discus	ssion		97
	5.6	Summ	ary		104
6		OMME	NDATION	JSION, AND IS FOR FUTURE RESEARCH	
	6.1	Summ			105
	6.2				107
	6.3	Recon	nmendatio	ons For Future Research	108
REFERENCES APPENDICES BIODATA OF STUDENT LIST OF PUBLICATION					109 123 128 129

 \bigcirc

LIST OF TABLES

Table		Page
4.1	List of primers used in cloning of KRas G12V into pET SUMO	44
4.2	Molecular weights of peaks 1 and 2	65
5.1	Atoms from STD spectrum showing protons of SRJ23 which are in close contact with the receptor of KRas G12V protein	89



LIST OF FIGURES

Figure		Page
2.1	Members of Ras family have highly conserved G- domain but differ in their hypervariable C-terminal region	8
2.2	Schematic of the canonical Ras effector pathways (Raf-MEk-Erk and PI3K-Akt) and the mutations that activate these pathways in driving cancer	9
2.3	The three-dimensional structure of Ras consists of six β -sheets and five α -helices interconnected by a series of ten loops	10
2.4	In silico docking shows SRJ23 bind to switch I and II of KRas Q61H protein and inhibits the Ras-SOS interaction	11
2.5	Oncogenic mutation of G12 and G13 prevent formation of van der Waals bonds between Ras and GAP through steric hindrance and perturb the proper orientation of Q61 in Ras causing attenuation of GTP hydrolysis	13
2.6	Chemical structure of anti-Ras agents	16
2.7	Use of strong denaturants to obtain soluble protein	18
2.8	Champion pET SUMO expression system	18
2.9	SML-KRas G12C complex assume conformation similar to GDP-bound KRas G12C which cause SML-KRas G12C complex does not interact with downstream effectors	20
2.10	Chemical structure of compounds bound to mutant KRas	21
2.11	Crystal structure of the ligand DCAI in complex with full-length KRas-'GTP', where 'GTP' is a nonhydrolyzable analog of GTP	21
2.12	One-dimensional proton spectra showing STD- NMR results which identifies Ras-binding compounds	23
3.1	Scheme of synthesis of SRJ23 derivatives	26
3.2	GI_{50} values of SRJ23 and its derivatives in six cell lines of four cancer types (potency)	31
3.3	GI ₅₀ values of SRJ23 and its derivatives in six cell lines of four cancer types (selectivity)	32
3.4	TGI values of SRJ23 and its derivatives in six cell lines of four cancer types (potency)	34
3.5	TGI values of SRJ23 and its derivatives in six cell lines of four cancer types (selectivity)	35
3.6	LC ₅₀ values of SRJ23 and its derivatives in six cell lines of four cancer types (potency)	37
3.7	LC_{50} values of SRJ23 and its derivatives in six cell lines of four cancer types (selectivity)	38

4.1	Sequence validation of <i>KRas G12V</i> gene construct in a mammalian-expressing pEXV <i>KRas G12V</i> plasmid	56
4.2 4.3	Validation of in-frame cloning 12% SDS-PAGE analysis of over-expressed KRas G12V in crude lysates of BL21 (DE3) <i>E. Coli</i> using 1 mM IPTG	57 58
4.4	The expression and purification of recombinant full- length KRas G12V oncoprotein	60
4.5	SDS-PAGE analysis showing successful SUMO Protease cleavage product of 5 µg of KRas G12V protein for cleavage reaction carried out using 2.5U of SUMO Protease at 30°C for 3 hours in 10X SUMO Protease Buffer -salt	61
4.6	SDS-PAGE analysis of reverse immobilised metal ion affinity chromatography cleaved pET SUMO KRas G12V	62
4.7	SDS-PAGE analysis of Sephacryl S-200 purification	63
4.8	Size exclusion chromatography was carried out by using Hi Prep 16/60 Sephacryl S-200 column	64
4.9	The standard was plotted as the partition coefficient for each of the gel filtration standard components (K_{av}) against their respective molecular weights in a base-10 logarithmic scale	65
4.10	Protein band mobility shift of unloaded and GDP loaded native KRas G12V	66
4.11	Western blot analysis shows successful identification of 6xHis-SUMO Tag using anti-His mouse monoclonal antibody from the SUMO Protease cleavage reaction of N-terminal SUMO fusion KRas G12V at 16°C for 1 hour using 2.5U of SUMO Protease	67
4.12	Western blot analysis shows successful identification of purified KRas G12V using anti- human KRas rabbit polyclonal antibody	68
5.1	Microscopic image of KRas G12V-GDP protein crystals	84
5.2	Proton 1D spectrum of SRJ23 (1mM) in phosphate buffer containing deuterium oxide and 10% deuterated DMSO	86
5.3	Proton 1D spectrum of SRJ23 (0.4mM) incubated with KRas G12V protein (0.02mM) in phosphate buffer containing deuterium oxide and 10% deuterated DMSO	87
5.4	STD spectrum of SRJ23 (0.4mM) incubated with KRas G12V protein (0.02mM) in phosphate buffer containing deuterium oxide and 10% deuterated DMSO	88

C

5.5	Proton 1D spectrum of DCAI in Tris buffer containing deuterium oxide and 1% deuterated DMSO	90
5.6	Proton 1D spectrum of DCAI (0.25mM) incubated with KRas G12V protein (0.005mM) in Tris buffer containing deuterium oxide and 1% deuterated DMSO	91
5.7	STD spectrum of DCAI (0.25mM) incubated with KRas G12V (0.005mM) in Tris buffer containing deuterium oxide and 1% deuterated DMSO	92
5.8	Proton 1D spectrum of vismodegib in phosphate buffer containing deuterium oxide and 10% deuterated DMSO	94
5.9	Proton 1D spectrum of vismodegib (0.25mM) incubated with KRas G12V protein (0.005mM) in phosphate buffer containing deuterium oxide and	95
	10% deuterated DMSO	
5.10	STD spectrum of vismodegib (0.25mM) incubated with KRas G12V protein (0.005mM) in phosphate buffer containing deuterium oxide and 10% deuterated DMSO	96
5.11	Molecular docking simulations for ensemble docking of SRJ23 on KRas Q61H structure showed 5 representatives cluster centroids with the percentage of the total conformer they represented from the 75 unique KRas conformers used for docking.	99
5.12	Principles of STD-NMR experiment showing the protein in surface representation and the non-exchangeable protons of the compound as spheres	102
6.1	Ras signalling showing two best studied pathways; mitogen activated protein kinase (MAPK) through direct phosphorylation of Raf-MEK-ERK and phosphoinositide 3-kinase (PI3K) through indirect phosphorylation of AKT	106

 \bigcirc

LIST OF ABBREVIATIONS

PDAC	Pancreatic ductal adenocarcinoma
NSCLC	Non-small cell lung cancer
Ras-MAPK	Ras-mitogen activated protein kinase
NF-KB	Nuclear factor kappa-light-chain-enhancer of activated B cells
PI3K	Phosphoinositide 3-kinase
GEF	guanine nucleotide exchange factors
GDP	Guanine diphosphate
GTP	Guanine triphosphate
GAP	GTPase activating proteins
HVR	Hypervariable C-terminal region
Q61	Glutamine
AGP	Andrographolide
NCI	National Cancer Institute
MTT	Microculture tetrazolium
EGFR	Epidermal growth factor receptor
RaIGDS	Ral guanine nucleotide dissociation stimulator
PLC ₈	Phospholipase C_{ε}
FTIs	Farnesyltransferase inhibitors
Ub	Ubiquitin
<i>E. coli</i>	<i>Escherichia coli</i>
AP	Ammonium persulfate
PCR	Polymerase chain reaction
NMR	Nuclear magnetic resonance
STD-NMR	Saturation transfer difference-nuclear magnetic resonance
PDB	Protein Data Bank
FDA	Food and Drug Administration
HTS	High-throughput screening
NADH	Nicotinamide adenine dinucleotide
DMSO	Dimethyl sulfoxide
SD	Standard deviation
SAR	Structure-activity relationship
SI	Selective index
IMAC	Ion metal affinity chromatography
SOS	Son of sevenless
NOE	Nuclear overhauser effect
Ppm	Parts per million
MD	Molecular dynamic

xvii



CHAPTER 1

INTRODUCTION

1.1 Overview

Cancer is the leading cause of death worldwide caused by loss of normal regulatory mechanisms which inhibit cell cycle progression. Breast, prostate, and lung cancers are among the top cancers being diagnosed currently with lung cancer remains as the top cancer killer (Torre et al., 2015). Thyroid cancer is expected to replace colorectal cancer as the fourth leading cancer being diagnosed by year 2030, while melanoma and uterine cancer stand at the fifth and sixth position of the most common cancer, respectively (Rahib et al., 2014). Pancreatic cancer is the third leading cause of cancer-related death in the United States surpassing breast cancer and expected to surpass prostate and colorectal cancer by year 2030 to become the second leading cause of cancer-related death in the United States (Rahib et al., 2014). Pancreatic cancer is considered deadly as it has the highest mortality rate of all major cancers and surgical resection is a possible curative treatment for only a minority of pancreatic cancer patients (10 to 15%) in the early stages (Jemal et al., 2005). Pancreatic cancer patients have low survival rates with a mortality rate of 97 to 98% within five years of diagnosis (Ghaneh et al. 2007). Most pancreatic cancer patients often die from a cancer-related death due to the late diagnosis of neoplasm which often occurs at the later stages of the disease, after local spread and distant metastases (Li et al., 2004). In an exacerbating way, the symptoms mostly appeared to be unspecific in nature (Ghaneh et al. 2007). Approximately 10% of pancreatic cancer cases are familial. Pancreatic ductal adenocarcinoma (PDAC) which accounts for more than 80% of the malignant neoplasms of pancreas is the most common epithelial, exocrine pancreatic malignancy (Alexakis et al., 2004). PDAC harbours genetic abnormalities which affect the mechanisms controlling G1 to S cell cycle progression and cellular proliferation. Mutation in p16, p53, Smad4 tumour suppressors gene and Ras proto-oncogene are examples of genetic abnormalities often found in more than 80% of pancreatic cancer (Rozenblum et al., 1997, Naumann et al., 1996; Huang et al., 1996; Chen et al., 1996; Redston et al., 1994). The Ras gene was known as cancer-related genes with KRas being the most frequently altered gene in pancreatic cancer. Besides pancreatic cancer harbouring approximately 95% of KRas mutation, approximately 30% to 40% of colon cancers harbour KRas mutation (Arrington et al., 2012). KRas mutation is also found mainly in 16% to 40% of non-small cell lung cancer (NSCLC) (Nelson et al., 1996).

Despite current available diagnostic means, medical practitioners have met with challenges in early detection and differential diagnosis of pancreatic cancer. As a result, many academic and pharmaceutical researchers have paid greater attention to understanding molecular events which lead to the development of this cancer in order to provide a basis for development of effective strategies for prevention, diagnosis and treatment. Recently, the focus has switched to targeting the Ras signaling pathway as cancer is often associated with the deregulation of the Ras signaling pathway caused by mutation of *Ras* gene in more than 30% of all cancers (Bos, 1989; Prior et al., 2012). *Ras* has a central role in regulating growth and cell survival in a wide spectrum of human tumours, the Ras-mitogen activated protein kinase (Ras-MAPK) signaling pathway is a well-validated oncogenic cascade. As the earliest and most common genetic mutation, *Ras* mutation drives transformation and tumour progression in the pancreas as evidenced by sequence analysis (Lemoine et al., 1992).

Ras protein, an essential component in Ras signaling is a small molecular weight GTPase that functions as a molecular switch in regulating pathways involved in fundamental cellular processes such as proliferation, differentiation, motility, transcription and survival. This protein couples extracellular signal to intracellular effectors such as the MEK kinase pathway, nuclear factor kappa-light-chain-enhancer of activated B cells (NF-kB) andphosphoinositide 3-kinase (PI3K) pathway. The molecular switch cycles between "off" and "on" conformations and the cycling is controlled by two key regulators; guanine nucleotide exchange factors (GEF) which promote GDP dissociation and GTP binding, and GTPase activating proteins (GAP) which stimulate intrinsic GTPase activity in Ras. Ras bound to GDP is inactive and will transit to the active state when bound to GTP and this process is controlled by GEF. The active state of Ras bound to GTP will hydrolyse to its inactive state by GAP in which it turns off the Ras signaling.

The Ras protein is commonly referred to as p21^{ras} with a molecular weight of approximately 21kDa (Barbacid, 1987) and is encoded by three ubiquitously expressed genes known as NRas, HRasand KRas. The four different Ras proteins (HRas, NRas, and the KRas 4b and the less abundant KRas 4a splice variants) encoded by these three genes have a highly conserved primary amino acid sequencehowever differ in their hypervariable C-terminal region (HVR) as a result of lipid posttranslational modifications (Dekker and Hedberg, 2011). According to the COSMIC dataset, KRas is confirmed as the most frequently mutated isoform, being found in 22% of all tumours analysed, followed by 8% for NRas and 3% for HRas. Mutation in Ras genes render Ras oncoprotein to constitutively bound to GTP and results in activation of downstream effector pathways even in the absence of extracellular stimuli (Karnoub and Weinberg, 2008). Each Ras isoform has a distinctive codon mutation. Point mutations in codons 12, 13 and 61 of KRaswith codon 12 being the most common site of Ras activation in pancreatic adenocarcinoma results in expression of constitutively active mutated Ras proteins which transform cells into a malignant phenotype (Santos and Nebreda, 1989). This research focuses on targeting KRas especially on the mutation of codon 12 due to their



higher occurrence in 75% to 100% of pancreatic cancers (Mu et al., 2004; Cox and Der, 2010).

The cellular signaling networks of a living cell are quite complex. It is not the simple rule that molecular level outcome will be determined by environmental signal.Ras proteins have been coined "undruggable" as their activation. mechanism is poorly understood. Activating Ras mutation was found to be resistant to standard therapies (Lu et al., 2016). There is no clinically useful drug available as an anti-Ras therapy due to the "undruggable" properties of KRas in which the protein lacks pocket for small molecule to bind with high affinity. The resemblance of mutant protein and normal KRas protein from a structural point of view and similar GDP/GTP-binding domain with other members of Ras superfamily of small GTPases such as Ral (Nicely et al., 2004), results in specific therapies even more challenging (McCormick, 2016). Despite being "undruggable", there are a few interesting exceptions which were discovered over the past few years that lead to new insights on KRas structure and function. Structural biology and in silico drug design had discovered a few promising inhibitors using small molecules which reveal opportunities for interventions and developing KRas therapies. In order to understand the molecular details of Ras signaling cascades, there are five main approaches in targeting oncogenic Ras signaling using small molecules which are:

- 1) Inhibition of nucleotide exchange factor which abrogated the Ras-GTP formation
- 2) Interference with the Ras membrane association
- 3) Inhibition of Ras-effector interactions
- 4) Downregulation of Ras proteins
- 5) Enhancement of intrinsic Ras GTPase activity

Using ensemble docking and innovative cell-based assays, small-molecule ligands known as andrographolide (AGP) and its benzylidene derivatives such as SRJ23 are found to directly bind to transient pocket on KRas and inhibit the nucleotide exchange factor binding of both its wild type KRas and oncogenic mutant KRas G12V (Hocker et al., 2013). AGP has been shown to exhibit great *in vitro* anticancer activities in various cancer cells particularly breast and colon cancers by inducing G1 cell-cycle arrest and apoptosis. The benzylidene derivatives of AGP exhibit inhibitory potency at nanomolar to micromolar concentrations (Jada et al., 2008). One of the andrographolide derivatives known as SRJ23 showed enhanced cytotoxicity and selectivity than AGP in the National Cancer Institute (NCI) screenings (Jada et al., 2008). SRJ23 displayed growth inhibition at low micromolar concentration and selectivity towards PC-3 prostate cancer cell lines among the various prostate cancer cell lines tested Wong et al., 2009).

1.2 Hypothesis

Despite SRJ23 having good cytotoxicity in prostate cancer cell lines (Wong et al., 2014), this compound is yet to be tested in pancreatic cancer which commonly harbours KRas mutation. Therefore, this situation had led to testing the cytotoxicity of SRJ23 using microculture tetrazolium (MTT) cell proliferation assay in various pancreatic cancer cell lines such as MIA PaCa 2, Capan-2, and BxPC-3. The *in vitro* anticancer activity of SRJ23 on pancreatic cancer cell line was hypothesised to involve direct inhibition of SRJ23 to Ras protein. The derivatives of SRJ23 were synthesised in order to discover compound with better potency and selectivity than SRJ23. Derivatives of SRJ23 were hypothesised to have better cytotoxicity activity than SRJ23. Although one of the derivatives (SRS157) displayed greaterselectivity in Capan-2 pancreatic cancer cell line as compared to HCT116 colorectal cancer cell line but overall, SRJ23 still showed better potency as compared to its derivatives in various cancer cell lines such as MCF7 breast, BxPC-3 pancreatic, MIA PaCa 2 pancreatic, HCT116 colorectal cancer cell line and PC-3 prostate cancer line.

Furthermore, thein silico docking using 75 unique KRas conformers showed SRJ23 is able to bind to three distinct pocketsknown as p1, p2, and p3 of oncogenic mutant KRas Q61H which involved switch 1 and 2 (Hocker et al., 2013). Ras has reduced affinity for effectors and harbours open pockets in state 1 and it was hypothesised that small molecule inhibitor such as SRJ23 which can stabilise the state 1 conformation has the potential to inhibit Ras signalling through interference with either effector or nucleotide exchange factor. Using molecular dynamics simulations, KRas-SRJ23 complex was found to be stable with SRJ23 favoured binding at p1 and state 1-like conformers with an open switch 1.SRJ23 was hypothesised to form complex with KRas G12V while STD NMR signals were hypothesised to show protons of SRJ23 are in close contact with the surface of the binding pocket of KRas G12Vprotein, indicating interaction between SRJ23 and KRas G12V protein. With promising in vitro and in silico findings, this has prompted the research to focus on expression of KRas G12V using bacterial system and purification of protein to determine their potential interaction with SRJ23 through X-ray crystallography and saturation transfer difference-nuclear magnetic resonance (STD-NMR).

1.3 Objectives

The general objective of this study was to determine the potentiality of SRJ23 compound and its derivatives to be developed as an anti-Ras therapy.

1.4 Specific Objectives

The general objective could be achieved through the specific objectives as followed:

- To determine the *in vitro* cytotoxicity of SRJ23 and its derivatives among a panel of cancer cell linesand to discover compound with better potency and selectivity than SRJ23.
- ii) To express and purify KRas G12V.
- iii) To determine potential interaction between KRas G12V protein and SRJ23 compound through X-ray crystallography and saturation transfer difference-nuclear magnetic resonance.

REFERENCES

- Abraham, S.J., Muhamed, I., Nolet, R., Yeung, F., and Gaponenko, V. 2010. Expression, purification, and characterization of soluble K-Ras4B for structural analysis. *Protein Expr Purif*73(2): 125-131.
- Ahmadian, M.R. 2002. Prospects for anti-ras drugs. *Br. J. Haematol.*116(3): 511-518.
- Aiba, S., Tsunekawa, H., and Imanaka, T. 1982. "New approach to tryptophan production by *Escherichia coli*: genetic manipulation of composite plasmids *in vitro*". *Applied and Environmental Microbiol.* 43(2): 289-297.
- Alexakis, N., Halloran, C., Raraty, M., Ghaneh, P., Sutton, R., Neoptolemos, J.P. 2004. Current standards of surgery for pancreatic cancer. *Br. J. Surg*.91(11): 1410-1427.
- An, W.F. and Tolliday, N. 2010. Cell-based assays for high-throughput screening. *Mol. Biotechnol*.45(2): 180-186.
- Arrington, A.K., Heinrich, E.L., Lee, W., Duldulao, M., Patel, S., Sanchez, J., Garcia- Aguilar, J.,and Kim, J. 2012. Prognostic and predictive roles of KRAS mutation in colorectal cancer. *Int. J. Mol.Sci.* 13(10): 12153-12168.
- Badisa, R.B., Darling-Reed, S.F., Joseph, P., Cooperwood, J.S., Latinwo, L.M., Goodman, C.B. 2009. Selective cytotoxic activities of two novel synthetic drugs on human breastcarcinoma MCF-7 cells. *Anticancer Res.* 29(8): 2993-2996.
- Baines, A.T., Xu, D., and Der, C.J. 2011. Inhibition of Ras for cancer treatment: the search continues. *Future Med. Chem.* 3(14): 1787-1808.
- Baker, N.M. and Der, C.J. 2013. Cancer: Drug for an "undruggable" protein. *Nature.* 497(7451): 577-578.
- Banerjee, M., Chattopadhyay, S., Choudhuri, T., Bera, R., Kumar, S., Chakraborty, B., Mukherjee, S.K. 2016. Cytotoxicity and cell cycle arrest induced by andrographolide lead to programmed cell death of MDA-MB-231 breast cancer cell line. *J. Biomed. Sci.* 23-40.
- Banerjee, S., Deshpande, A.A., Mandi, N. & Padmanabhan, S. 2009. A novel cytokine derived fusion tag for over-expression of heterologous proteins in *E. coli. Int. J. of Biological Life Sciences* 5: 129-133.
- Barbacid, M. 1987. Ras genes. Annu. Rev. Biochem. 56: 779-827.
- Barker, J., Courtney, S., Hesterkamp, T., Ullmann, D., Whittaker, M. 2006. Fragment screening by biochemical assay. *Expert Opin. Drug Discov.* 1(3): 225-236.
- Berchtold, H., Reshetnikova, L., Reiser, C.O., Schirmer, N.K., Sprinzl, M., and Hilgenfeld, R. 1993. Crystal structure of active elongation factor Tu reveals major domain rearrangements. *Nature* 365(6442): 126-132.
- Bergfors, T.M. 1999. Protein crystallization: techniques, strategies, and tips: *a laboratory manual*, La Jolla, Calif: International University Line.
- Berman, H.M., Westbrook, J., Feng, Z., Gilliland, G., Bhat, T.N., Weissig, H., Shindyalov, I.N., and Bourne, P.E. 2000. The Protein Data Bank. *Nucleic Acids Res.* 28(1): 235-242.

- Bittner, M. and Vapnek, D. 1981. Versatile cloning vectors derived from the runaway-replication plasmid pKN402. *Gene* 15(4): 319-329.
- Blake, M.S., Johnston, K.H., Russell-Jones, G.J., and Gotschlich, E.C. 1984. A rapid, sensitive method for detection of alkaline phosphataseconjugated anti-antibody on Western blots. *Anal. Biochem.* 136(1): 175-179.
- Bollag G., Hirth P., Tsai J., Zhang J., Ibrahim P.N., Cho H., Spevak W., Zhang C., Zhang Y., Habets G., Burton E.A., Wong B., Tsang G., West B.L., Powell B., Shellooe R., Marimuthu A., Nguyen H., Zhang K.Y.J., Artis D.R., Schlessinger J., Su F., Higgins B., Iyer R., D'Andrea K., Koehler A., Stumm M., Lin P.S., Lee R.J., Grippo J., Puzanov I., Kim K.B., Ribas A., McArthur G.A., Sosman J.A., Chapman P.B., Flaherty K.T., Xu X., Nathanson K.L., Nolop K., 2010. Clinical efficacy of a RAF inhibitor needs broad target blockade in BRAF-mutant melanoma. *Nature*467: 596–599.
- Bornhorst, J.A. and Falke, J.J. 2000. Purification of proteins using polyhistidine affinity tags. *Methods Enzymol* 326: 245-254.
- Bos, J.L. 1989. Ras oncogenes in human cancer: a review. *Cancer Res.* 49: 4682-4689.
- Buhrman, G., Holzapfel, G., Fetics, S., Mattos, C. 2010. Allosteric modulation of Ras positions Q61 for a direct role in catalysis. *Proc. Natl. Acad. Sci. USA.* 107(11): 4931-4936.
- Burns, M.C., Sun, Q., Daniels, R.N., Camper, D., Kennedy, J.P., Phan, J., Olejniczak, E.T., Lee, T., Waterson, A.G., Rossanese, O.W., Fesik, S.W. 2014. Approach for targeting Ras with small molecules that activate SOS-mediated nucleotide exchange. *Proc. Natl. Acad. Sci. USA*. 111(9): 3401-3406.
- Cáceres, D.D., Hancke, J.L., Burgos, R.A., Sandberg, F., and Wikman, G.K. 1999. Use of visual analogue scale measurements (VAS) to assess the effectiveness of standardized Andrographis paniculata extract SHA-10 in reducing the symptoms of common cold. A randomized double blindplacebo study. *Phytomedicine* 6(4): 217-223.
- Cala, O., Guillière, F., Krimm, I. 2014. NMR-based analysis of protein-ligand interactions. *Anal. Bioanal. Chem.* 406(4): 943-956.
- Camps, M. 2010. Modulation of ColE1-like plasmid replication for recombinant gene expression. *Recent. Pat. DNA Gene Seq.* 4(1): 58-73.
- Capon, D.J., Seeburg, P.H., McGrath, J.P., Hayflick, J.S., Edman, U., Levinson, A.D., and Goeddel, D.V. 1983. Activation of Ki-ras 2 gene in human colon and lung carcinomas by two different point mutations. *Nature* 304(5926): 507-513.
- Catic, A., Misaghi, S., Korbel, G.A., and Ploegh, H.L. 2007. ElaD, a Deubiquitinating protease expressed by *E. coli. PLoS One* 2(4): e381.
- ChampionTM pET SUMO Expression System. N.d. Retrieved 31 July 2017 from https://www.thermofisher.com/order/catalog/product/K30001.
- ChampionTM pET SUMO Expression System. SUMO Protease. N.d. Retrieved 31 July 2017 from
 - https://tools.thermofisher.com/content/sfs/manuals/petsumo_man.pdf

- Chandrasekaran, C.V., Gupta, A., and Agarwal, A. 2010. Effect of an extract of Andrographis paniculata leaves on inflammatory and allergic mediators *in vitro. J. Ethnopharmacol.* 129(2): 203-207.
- Chataway, T. K. and Barritt, G. J. 1995. Purification of histidine-tagged ras and its use in the detection of ras binding proteins. *Mol and Cellular Biochem.* 144(2): 167-173.
- Chen, Z., Zhang, H., and Savarese, T. 1996. Gene deletion chemoselectivity: Codeletion of the genes for p16 (INK4), methylthioadenosine phosphorylase, and the alpha- and beta-interferons in human pancreatic cell carcinoma lines and its implications for chemotherapy. *Cancer Res.*56: 1083, 1996.
- Cooper, G.M. 1982. Cellular transforming genes. Science217(4562): 801-806.
- Cooper, G. M. 2000. The Cell: A Molecular Approach (2nd Edition),
 - Sunderland, MA: Sinauer Associates.
- Cox, A.D. and Der, C.J. 2010. Ras history: The saga continues. *Small GTPases* 1(1): 2-27.
- Cox, M.J. and Weber, P.C. 1988. An investigation of protein crystallization parameters using successive automated grid searches (SAGS). *J. Cryst. Growth* 90: 318– 324.
- Dalton, A.C. and Barton, W.A. 2014. Over-expression of secreted proteins from mammalian cell lines. *Protein Sci.* 23(5): 517-525.
- Das, U., Pati, H.N., Sakagami, H., Hashimoto, K., Kawase, M., Balzarini, J., De Clercq, E., Dimmock, J.R., 2011. 3, 5-Bis(benzylidene)-1-[3-(2hydroxyethylthio)propanoyl]piperidin-4-ones: a novel cluster of potent tumor-selective cytotoxins. J. Med. Chem. 54(9): 3445-3449.
- Davis, B. 2013. Screening protein-small molecule interactions by NMR. *Methods Mol. Biol.* 1008: 389-413.
- Dekker, F.J. and Hedberg, C. 2011. Small molecule inhibition of protein depalmitoylation as a new approach towards downregulation of oncogenic Ras signaling. *Bioorg. & Med. Chem.*19(4): 1376-1380.
- Dervan, P.B. 2001. Molecular recognition of DNA by small molecules. *Bioorg. Med.Chem.* 9(9): 2215-2235.
- Dessau, M.A. and Modis, Y. 2011. Protein Crystallization for X-ray Crystallography. J. Vis. Exp. 47: 2285.
- Drewry, D.H. and Macarron, R. 2010. Enhancements of screening collections to address areas of unmet medical need: an industry perspective. *Curr. Opin. Chem.* 14: 289-298.

Drews, J. 2000. Science 287: 1962.

- Ekalaksananan, T., Sookmai, W., Fangkham, S., Pientong, C., Aromdee, C., Seubsasana, S., and Kongyingyoes, B. 2015. Activity of Andrographolide and Its Derivatives on HPV16 Pseudovirus Infection and Viral Oncogene Expression in Cervical Carcinoma*Cells. Nutr. Cancer* 67(4): 687-696.
- EMBOSS Water, Pairwise Sequence Alignment (PROTEIN). N.d. Retrieved on 31 July 2017 from

http://www.ebi.ac.uk/Tools/psa/emboss_water/nucleotide.html

Eser, S., Schnieke, A., Schneider, G., and Saur, D. 2014. Oncogenic KRAS signaling in pancreatic cancer. *Br. J. Cancer* 111:817-822.

ExPASy Bioinformatics Resource Portal. N.d. Retrieved on 31 July 2017 from

http://web.expasy.org/translate/

- Fakruddin, M., Mohammad Mazumdar, R., Bin Mannan, K.S., Chowdhury, A., And Hossain, M.N., 2012. Critical Factors Affecting the Success of Cloning, Expression, and Mass Production of Enzymes by Recombinant *E. coli. ISRN Biotechnol*:590587.
- Fast, J.L., Cordes, A.A., Carpenter, J.F., and Randolph, T.W. 2009. Physical instability of a therapeutic Fc fusion protein: domain contributions to conformational and colloidal stability. *Biochemistry* 48(49): 11724-11736.
- Fernández-Medarde, A. and Santos, E. 2011. Ras in Cancer and Developmental Diseases. *Genes & Cancer* 2(3): 344-358.
- Fielding, L. 2007. NMR methods for the determination of protein–ligand dissociation constants. *Progress in Nuclear Magnetic Reson. Spectroscopy* 51: 219-242.
- Fielding, L., Rutherford, S., and Fletcher, D. 2005. Determination of proteinligand binding affinity by NMR: observations from serum albumin model systems. *Magn Reson Chem.* 43(6): 463-470.
- Fournier, D.B. and Gordon, G.B., 2000. COX-2 and colon cancer: potential targets for chemoprevention. *J. Cell Biochem. Suppl.* 34: 97-102.
- Frey, S. and Görlich, D. 2014. A new set of highly efficient, tag-cleaving proteases for purifying recombinant proteins. *J Chromatogr A* 1337: 95-105.
- German, L.R. and Eduardo, A.C. 2014. Recombinant protein expression in *Escherichia coli*: advances and challenges. *Front Microbiol* 5: 172.
- Gershenzon, J. and Dudareva, N. 2007. The function of terpene natural products in the natural world. *Nat. Chem. Biol.* 3(7): 408-414.
- Ghaneh, P. and Costello, E., and Neoptolemos, J.P. 2007. Biology and management of pancreatic cancer, *Gut* 56: 1134–1152.
- Gibbs, J.B., Sigal, I.S., Poe, M., and Scolnick, E.M. 1984. Intrinsic GTPase activity distinguishes normal and oncogenic ras p21 molecules. *Proc Natl Acad Sci USA* 81(18): 5704-5708.
- Glick, B.R. and Whitney, G.K., 1987. Factors affecting the expression of foreign proteins in *Escherichia coli*1(5). 277-282.
- Govindarajan, M. 2011. Evaluation of *Andrographis paniculata* Burm.f. (Family:Acanthaceae) extracts against *Culex quinquefasciatus* (Say.) and *Aedes aegypti* (Linn.) (Diptera:Culicidae). *Asian Pac. J. Trop. Med.* 4(3): 176-181.
- Gralla, J.D. 1990. Promoter Recognition and mRNA Initiation by *Escherichia coli* Es70. *Meth Enzymol* 185: 37-54.
- Graziano, M.P., Freissmuth, M., and Gilman, A.G. 1989. Expression of Gs alpha in *Escherichia coli*. Purification and properties of two forms of the protein. *J Biol Chem* 264(1): 409-418.
- Griffiths A.J.F., Gelbart W.M., Miller J.H. 1999. Modern Genetic Analysis. New York: W. H. Freeman. Retrieved on 31 July 2017 from https://www.ncbi.nlm.nih.gov/books/NBK21248/
- Grisshammer, R. and Tucker, J. 1997. Quantitative evaluation of neurotensin receptor purification by immobilised metal affinity chromatography. *Protein Expr. Purif.* 11(1): 53-60.

Gross, M., Sweet, R.W., Sathe, G., Yokoyama, S., Fasano, O., Goldfarb, M.,

Wigler, M., and Rosenberg, M. 1985. Purification and characterization of human H-ras proteins expressed in *Escherichia coli. Mol. Cell Biol.* 5(5): 1015-1024.

- Guan J.-Y., Keizers P.H.J., Liu W.-M., Löhr F., Skinner S.P., Heeneman E.A., Schwalbe H., Ubbink M., Siegal G., 2013. Small-molecule binding sites on proteins established by paramagnetic NMR spectroscopy. J. Am. Chem. Soc.135: 5859–5868.
- Güldenhaupt, J., Rudack, T., Bachler, P., Mann, D., Triola, G., Waldmann, H., Kötting, C., and Gerwert, K. 2012. N-Ras forms dimers at POPC membranes. *Biophys J.* 103(7): 1585-1593.
- Gysin, S., Salt, M., Young, A., and McCormick, F. 2011. Therapeutic strategies for targeting ras proteins. *Genes Cancer* 2(3): 359-372.
- Hall, A., Marshall, C.J., Spurr, N.K., and Weiss, R.A. 1983. Identification of transforming gene in two human sarcoma cell lines as a new member of the ras gene family located on chromosome 1. *Nature* 303(5916): 396-400.
- Hall, B.E., Bar-Sagi, D., Nassar, N. 2002. The structural basis for the transition from Ras-GTP to Ras-GDP. *Proc Natl Acad Sci USA* 99(19): 12138-12142.
- Hanahan, D. and Weinberg, R.A. 2011. Hallmarks of cancer: the next generation. *Cell* 144(5): 646-674.
- Hancock, J.F. and Parton, R.G. 2005. Ras plasma membrane signalling platforms. *Biochem J.* 389(1): 1-11.
- Haselhorst, T., Garcia, J.M., Islam, T. 2008. Avian influenza H5-containing virus-like particles (VLPs): host-cell receptor specificity by STD NMR spectroscopy. *Angew. Chem. Int. Ed. Engl.* 47: 1910-1912.
- Hassell, A.M., An, G., Bledsoe, R.K., Bynum, J.M., Carter, H.L. 3rd., Deng, S.J., Gampe, R.T., Grisard, T.E., Madauss, K.P., Nolte, R.T., Rocque, W.J., Wang, L., Weaver, K.L., Williams, S.P., Wisely, G.B., Xu, R., Shewchuk, L.M. 2007. Crystallization of protein-ligand complexes. *Acta. Crystallogr. D. Biol.Crystallogr.* 63(1): 72-79.\
- Henry, K.A., Zwieb, C., and Fried, H.M. 1997. Purification and biochemical characterization of the 19-kDa signal recognition particle RNA-binding protein expressed as a hexahistidine-tagged polypeptide in *Escherichia coli. Protein Expr. Purif.* 9(1): 15-26.
- Hocker, H.J., Cho, K.J., Chen, C.Y., Rambahal, N., Sagineedu, S.R., Shaari, K., Stanslas, J., Hancock, J.F., and Gorfe, A.A. 2013. Andrographolide Derivatives inhibit guanine nucleotide exchange and abrogate oncogenic Ras function. *Proc* Natl Acad Sci U.S.A 110(25): 10201-10206.
- Hopkins, A.L. and Groom, C.R. 2002. The druggable genome. *Nat Rev Drug Discovery* 1(9): 727-730.
- Huang, L., Goodrow, T., Zhang, S., Klein-Szanto, A., Chang, H., and Ruggeri,
 B. 1996. Deletion and mutation analyses of the p16/MTS-1 tumor suppressor gene in human ductal pancreatic cancer reveals a higher frequency of abnormalities in tumor derived cell lines than in primary ductal adenocarcinomas. *Cancer Res* 56: 1137, 1996.

Hughes, J.P., Rees, S., Kalindjian, S.B., Philpott, K.L. 2011. Principles of early Drug discovery. *Br J Pharmacol* 162(6): 1239-1249.

- Hunke, S. and Betton, J.M. 2003. Temperature effect on inclusion body formation and stress response in the periplasm of *Escherichia coli*. *Mol Microbiol* 50(5): 1579-1589.
- Hunter, J.C., Gurbani, D., Ficarro, S.B., Carrasco, M.A., Lim, S.M., Choi, H.G., Xie, T., Marto, J.A., Chen, Z., Gray, N.S., and Westover, K.D. 2014. In situ selectivity profiling and crystal structure of SML-8-73-1, an active site inhibitor of oncogenic K-Ras G12C. *Proc. Natl. AcadSci. U.S.A* 111(24): 8895-8900.
- Hunter, J.C., Manandhar, A., Carrasco, M.A., Gurbani, D., Gondi, S., Westover, K.D. 2015. Biochemical and Structural Analysis of Common Cancer-Associated KRAS Mutations. *Mol. Cancer Res.* 13(9): 1325-1335.
 Inclusion body purification & protein refolding. N.d. Retrieved 31 July 2017 from https://www.profacgen.com/inclusion-body-purificationprotein-refolding.htm
- Inouye, K., Mizutani, S., Koide, H., Kaziro, Y. 2000. Formation of the Ras dimer is essential for Raf-1 activation. *J. Biol. Chem.* 275(6): 3737-3740.
- Jada, S.R., Hamzah, A.S., Lajis, N.H., Saad, M.S., Stevens, M.F., Stanslas, J. 2006. Semisynthesis and cytotoxic activities of andrographolide analogues. *J. Enzyme Inhib. Med. Chem.* 21(2):145-155.
- Jada, S.R., Subur, G.S., Matthews, C., Hamzah, A.S., Lajis, N.H., Saad, M.S., Stevens, M.F., and Stanslas, J. 2007. Semisynthesis and in vitro anticancer activities of *Phytochemistry* 68(6): 904-912.
- Jada, S.R., Matthews, C., Saad, M.S., Hamzah, A.S., Lajis, N.H., Stevens, M.F.G., and Stanslas, J. 2008. Benzylidene derivatives of andrographolide inhibit growth of breast and colon cancer cells in vitro by inducing G1 arrest and apoptosis. *Br. J. Pharmacol*.155(5): 641-654.
- Janknecht, R., de Martynoff, G., Lou, J., Hipskind, R.A., Nordheim, A., and Stunnenberg, H.G. 1991. Rapid and efficient purification of native histidine-tagged protein expressed by recombinant vaccinia virus. *Proc. Natl. Acad. Sci. U.S.A* 88(20): 8972-8976.
- Jemal, A., Murray, T., Ward, E., Samuels, A., Tiwari, R.C., Ghafoor, A., Feuer, E.J., and Thun, M.J. 2005. *Cancer statistics, 2005. CA Cancer J. Clin. 55(1)*: 10-30.
- Karnoub, A.E. and Weinberg, R.A. 2008. Ras oncogenes: split personalities. *Nat. Rev. Mol. Cell Biol.*9: 517-531.
- Khan, F., Legler, P.M., Mease, R.M., Duncan, E.H., Bergmann-Leitner, E.S., Angov, E. 2012. Histidine affinity tags affect MSP1(42) structural stability and immunodominance in mice. *Biotechnol J* 7(1): 133-147.
- Khan, K.H. 2013. Gene expression in Mammalian cells and its applications. *Adv. Pharm. Bull.* 3(2): 257-263.
- Kleuss, C., Raw, A.S., Lee, E., Sprang, S.R., and Gilman, A.G. 1994. Mechanism of GTP hydrolysis by G-protein alpha subunits. *Proc. Natl. Acad. Sci*.USA 91(21): 9828-9831.
- Khow, O. and Suntrarachun, S. 2012. Strategies for production of active eukaryotic proteins in bacterial expression system. *Asian Pac J Trop Biomed* 2(2): 159-162.

Kortmann, M, Kuhl, V., Klaffl, S., and Bott, M. 2015. A chromosomally encoded

T7 RNA polymerase-dependent gene expression system for *Corynebacterium glutamicum*: construction and comparative evaluation at the single-cell level. *Microb. Biotechnol.* 8(2): 253-265.

- Kotar, A., Tomašič, T., Lenarčič Živković, M., Jug, G., Plavec, J., and Anderluh, M. 2016. STD NMR and molecular modelling insights into interaction of novel mannose-based ligands with DC-SIGN. Org. Biomol. Chem. 14(3): 862-875.
- Kumar, R.A., Sridevi, K., Kumar, N.V., Nanduri, S., and Rajagopal, S. 2004. Anticancer and immunostimulatory compounds from *Andrographis* paniculata. J Ethnopharmacol 92(2-3): 291-295.
- Kurien, B.T. and Scofield, R.H. 2003. Protein blotting: a review. *J Immunol Methods* 274(1-2): 1-15.
- Lacal, J.C., Santos, E., Notario, V, Barbacid, M., Yamazaki, S., Kung, H., Seamans, C., McAndrew, S., and Crowl, R. 1984. Expression of normal and transforming H-ras genes in *Escherichia coli* and purification of their encoded p21 proteins. *Proc Natl Acad Sci. USA* 81(17): 5305-5309.
- Lee, C.D., Sun, H.C., Hu, S.M., Chiu, C.F., Homhuan, A., Liang, S.M., Leng, C.H., and Wang, T.F. 2008. An improved SUMO fusion protein system for effective production of native proteins. *Protein Sci*.17(7): 1241-1248.
- Lee, M.J., Rao, Y.K., Chen, K., Lee, Y.C., Chung, Y.S., Tzeng, Y.M. 2010. Andrographolide and 14-deoxy-11, 12-didehydroandrographolide from *Andrographis paniculata* attenuate high glucose-induced fibrosis and apoptosis in murine renal mesangeal cell lines. *J. Ethnopharmacol.* 132(2): 497-505.
- Lemoine, N.R., Jain, S., and Hughes, C.M. 1992. Ki-ras oncogene activation in preinvasive pancreatic cancer. *Gastroenterology* 102: 230 236.
- Leshchiner, E.S., Parkhitko, A., Bird, G.H., Luccarelli, J., Bellairs, J.A., Escudero, S., Opoku-Nsiah, K., Godes, M., Perrimon, N., Walensky, L.D. 2015. Direct inhibition of oncogenic KRAS by hydrocarbonstapled SOS1 helices. *Proc. Natl. Acad. Sci. U.S.A* 112(6): 1761-1766.
- Li, G. and Zhang, X.C. 2004. GTP hydrolysis mechanism of Ras-like GTPases. *J. Mol. Biol*.340(5): 921-932.
- Li, D., Xie, K., Wolff, R., and Abbruzzese, J.L. 2004. Pancreatic cancer. *Lancet.* 363(9414): 1049-1057.
- Li, J., Huang, W., Zhang, H., farnesyltra, X., and Zhou, H. 2007. Synthesis of a ndrographolide derivatives and their TNF-alpha and IL-6 expression Inhibitory activities. *Bioorg. Med. Chem. Lett.* 17(24): 6891-6894.
- Lim, J.C., Jeyaraj, E.J., Sagineedu, S.R., Wong, W.S., and Stanslas, J. 2015. SRS06, a new semisynthetic andrographolide derivative with improved anticancer potency and selectivity, inhibits nuclear factor-kB nuclear binding in the A549 non-small cell lung cancer cell line. *Pharmacology* 95(1-2): 70-77.
- Lin, W.C., Iversen, L., Tu, H.L., Rhodes, C., Christensen, S.M., Iwig, J.S., Hansen, S.D., Huang, W.Y., and Groves, J.T. 2014. H-Ras forms dimmers on membrane surfaces via a protein-protein interface. *Proc. Natl. Acad. Sci. USA* 111(8): 2996-3001.

- Lodish, H., Berk, A., Zipursky, S.L., Matsudaira, P., Baltimore, D., Darnell, J. 2000. Section 20.4, receptor tyrosine kinases and Ras. Molecular cell biology (4th ed.). New York: W. H. Freeman.
- Lomenick, B., Olsen, R.W., and Huang, J. 2011. Identification of direct protein targets of small molecules. *ACS Chem. Biol.* 6(1): 34-46.
- Lowe, P.N., Page, M.J., Bradley, S., Rhodes, S., Sydenham, M., Paterson, H., and Skinner, R.H. 1991. Characterization of recombinant human Kirsten-ras (4B) p21 produced at high levels in *Escherichia coli* and insect baculovirus expression systems. *J. Biol. Chem.* 266(3): 1672-1678.
- Lu, J., Hunter, J., Manandhar, A., Gurbani, D., Westover, K.D. 2015. Structural dataset for the fast-exchanging KRAS G13D. *Data Brief* 17(5): 572-578.
- Lu, S., Jang, H., Zhang, J., Nussinov, R. 2016. Inhibitors of Ras-SOS Interactions. *Chem. Med. Chem.* 11(8): 814-821.
- Lu, S., Jang, H., Gu, S., Zhang, J., and Nussinov, R. 2016. Drugging Ras GTPase: a comprehensive mechanistic and signaling structural view. *Chem. Soc. Rev.* 45(18): 4929-4952.
- Lu, S., Jang, H., Nussinov, R., and Zhang, J. 2016. The Structural Basis of Oncogenic Mutations G12, G13 and Q61 in small GTPase K-Ras4B. *Sci. Rep.* 6: 21949.
- Luft, J.R., Wolfley, J.R., Said, M.I., Nagel, R.M., Lauricella, A.M., Smith, J.L., Thayer, M.H., Veatch, C.K., Snell, E.H., Malkowski, M.G., and Detitta, G.T. 2007. Efficient optimization of crystallization conditions by manipulation of drop volume ratio and temperature. *Protein Sci*.16(4): 715-722.
- Macha, M.A., Batra, S.K., Ganti, A.K. 2013. Profile of vismodegib and its potential in the treatment of advanced basal cell carcinoma. *Cancer Manag. Res.* 5: 197-203.
- Makrides, S.C. 1996. Strategies for achieving high-level expression of genes in *Escherichia coli. Microbiol. Rev.* 60(3): 512-538.
- Malakhov, M.P., Mattern, M.R., Malakhova, O.A., Drinker, M., Weeks, S.D., and Butt, T.R. 2004. SUMO fusions and SUMO-specific protease for efficient expression and purification of proteins. *J. Struct. Funct. Genomics* 5(1-2): 75-86.
- Malumbres, M. and Barbacid, M. 2003. RAS oncogenes: the first 30 years. *Nat. Rev. Cancer.* 3: 459-465.
- Marblestone, J.G., Edavettal, S.C., Lim, Y., Lim, P., Zuo, X., and Butt T.R. 2006. Comparison of SUMO fusion technology with traditional gene fusion systems: enhanced expression and solubility with SUMO. *Protein Sci.* 15(1): 182-189.
- Maurer, T., Garrenton, L.S., Oh, A., Pitts, K., Anderson, D.J., Skelton, N.J., Fauber, B.P., Pan, B., Malek, S., Stokoe, D., Ludlam, M.J., Bowman, K.K., Wu, J., Giannetti, A.M., Starovasnik, M.A., Mellman, I., Jackson, P.K., Rudolph, J., Wang, W., and Fang, G. 2012. Smallmolecule ligands bind to a distinct pocket in Ras and inhibit SOS-mediated nucleotide exchange activity. *Proc. Natl. Acad. Sci. U.S.A* 109(14): 5299-5304.

Mayer, M and Meyer, B. 1999. Characterization of ligand binding by saturation

transfer difference NMR spectroscopy. *Angew. Chem. Int. Ed.* 38: 1784-1788.

- Mayer, M. and Meyer, B. 2001. Group epitope mapping by saturation transfer difference NMR to identify segments of a ligand in direct contact with a protein receptor. *J. Am. Chem. Soc.* 123(25): 6108-6117.
- McCormick, F. 2016. K-Ras protein as a drug target. *J Mol Med (Berl)*.94(3): 253-258.McPherson A. 1999. Crystallization of biological macromolecules. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.
- Meena, R. and Harish, P. 2001. "Expression systems for production of heterologous proteins". *Current Science* 80(9): 1121-1128.
- Menon, V. and Bhat, S. 2010. Anticancer activity of andrographolide semisynthetic derivatives. *Nat. Prod. Commun.* 5(5): 717-720.
- Meyer, B. and Peters, T. 2003. NMR spectroscopy techniques for screening and identifying ligand binding to protein receptors. *Angew. Chem. Int. Ed. Engl.* 42(8): 864-890.
- Misra, P., Pal, N.L., Guru, P.Y., Kariya, J.C., Srivastava, V, Tandon, J.C. 1992. Antimalarial activity of *Andrographis paniculata* (Kamelgh) against Plasmodium berghei NK65 in Mastomys natalensis. *Int. J. Pharm.* 30: 263-274.
- Mosmann, T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J. Immunol. Methods* 65(1-2): 55-63.
- Mossessova, E. and Lima, C.D. 2000. Ulp1-SUMO crystal structure and genetic analysis reveal conserved interactions and a regulatory element essential for cell growth in yeast. *Mol. Cell* 5(5): 865-876.
- Mu, D.Q., Peng, Y.S., and Xu, Q.J.2004. Values of mutations of K-ras oncogene at codon 12 in detection of pancreatic cancer: 15-year experience. *World J. Gastroenterol.* 10(4): 471-475.
- Mulder, F.A., Hon, B., Muhandiram, D.R., Dahlquist, F.W., and Kay, L.E. 2000. Flexibility and ligand exchange in a buried cavity mutant of T4 Iysozyme studied by multinuclear NMR. *Biochemistry* 39(41): 12614-12622.
- Muratcioglu, S., Chavan, T.S., Freed, B.C., Jang, H., Khavrutskii, L., Freed, R.N., Dyba, M.A., Stefanisko, K., Tarasov, S.G., Gursoy, A., Keskin, O., Tarasova, N.I., Gaponenko, V., and Nussinov, R. 2015. GTP-Dependent K-Ras Dimerization. *Structure* 23(7): 1325-1335.
- Nan, X., Tamgüney, T.M., Collisson, E.A., Lin, L.J., Pitt, C., Galeas, J., Lewis, S., Gray, J.W., McCormick, F., and Chu, S. 2015. Ras-GTP dimers activate the Mitogen-Activated Protein Kinase (MAPK) pathway. *Proc. Natl. Acad. Sci. USA* 112(26): 7996-8001.
- Naumann, M., Savitskaia, N., Eilert, C., Schramm, A., Kalthoff, H., and Schmiegel, W. 1996. Frequent codeletion of p16/MTS1 and p15/MTS2 and genetic alterations in p16/MTS1 in pancreatic tumors. *Gastroenterology* 110: 1215.
- Nelson, M.A., Wymer, J., and Clements, N. Jr. 1996. Detection of K-ras gene mutations in non-neoplastic lung tissue and lung cancers. *Cancer Lett.* 103(1): 115-121.
- Neogy, S., Das, S., Mahapatra, S.K., Mandal, N., and Roy, S. 2008.

Amelioratory effect of Andrographis paniculata Nees on liver, kidney, heart, lung and spleen during nicotine induced oxidative stress. *Environ. Toxicol. Pharmacol.* 25(3): 321-328.

- Nettleship, J.E., Assenberg, R., Diprose, J.M., Rahman-Hug, N., and Owens, R.J. 2010. Recent advances in the production of proteins in insect and mammalian cells for structural biology. *J. Struct. Biol.* 172(1): 55-65.
- Nicely, N., Kosak, J., de Serrano, V., and Mattos, C. 2004. Crystal structures of Ral-GppNHp and Ral-GDP reveal two binding sites that are also present in Ras and Rap. *Structure*12(11): 2025-2036.
- Nikolov, D.B., Hu, S.H., Lin, J., Gasch, A., Hoffmann, A., Horikoshi, M., Chua, N.H., Roeder, R.G., and Burley, S.K. 1992. Crystal structure of TFIID TATA-box binding protein. *Nature* 360(6399): 40-46.
- Nilsson, J., Ståhl, S., Lundeberg, J., Uhlén, M., Nygren, P.A. 1997. Affinity fusion strategies for detection, purification, and immobilization of recombinant proteins. *Protein Expr. Purif.* 11(1): 1-16.
- O' Hagan, R.C. and Heyer, J. 2011. KRAS Mouse Models: Modeling Cancer Harbouring KRAS Mutations. *Genes Cancer* 2(3): 335-343.
- Om, P., Amit, K., Pawan, K. Ajeet 2013. Anticancer Potential of Plants and Natural Products: A Review. American J. of Pharmacol. Sci. 1(6): 104-115.
- Ooi, J.P., Kuroyanagi, M., Sulaiman, S.F., Muhammad, T.S., and Tan, M.L. 2011. Andrographolide and 14-deoxy-11,12didehydroandrographolide inhibit cytochrome P450s in HepG2 hepatoma cells. *Life Sci.* 88(9-10): 447-454.
- Pan, S.H. and Malcolm, B.A. 2000. Reduced background expression and Improved plasmid stability with pET vectors in BL21(DE3). *Biotechniques* 29(6): 1234-1238.
- Panavas, T., Sanders, C., and Butt, T. R. (2009). SUMO fusion technology for enhanced protein production in prokaryotic and eukaryotic expression systems. *Methods Mol. Biol.* 497: 303-317.
- Papageorge, A., Lowy, D., and Scolnick, E.M. 1982. Comparative biochemical properties of p21 ras molecules coded for by viral and cellular ras genes. *J. Virol.* 44(2): 509-519.
- Peciak, K., Tommasi, R., Choi, J. W., Brocchini, S., and Laurine, E. 2014.Expression of soluble and active interferon consensus in SUMO fusion expression system in *E. coli*.*Protein Expr. Purif.* 99: 18-26.
- Pellecchia, M., Sem, D.S., and Wüthrich K. 2002. NMR in drug discovery. *Nat. Rev. Drug. Discov.*1(3): 211-219.
- Peng, Y., Li, J., Sun, Y., Chan, J.Y, Sheng, D., Wang, K., Wei, P., Ouyang, P., Wang, D., Lee, S.M.Y., and Zhou, G. 2015. SAR studies of 3,14,19-Derivatives of Andrographolide on Anti-Proliferative Activity to Cancer Cells and Toxicity to Zebrafish: An *In Vitro* and *In Vivo* Study. *Royal Soc. Chem. Adv.* 5: 22510-22526.
- Peroutka lii, R.J., Orcutt, S.J., Strickler, J.E., Butt, T.R. 2011. SUMO fusion technology for enhanced protein expression and purification in prokaryotes and eukaryotes. *Methods Mol. Biol.* 705: 15-30.
- Pierpont, M.E., Magoulas, P.L., Adi, S., Kavamura, M.I., Neri, G., Noonan, J., Pierpont, E.I., Reinker, K., Roberts, A.E., Shankar, S., Sullivan, J., Wolford, M., Conger, B., Santa Cruz, M., Rauen, K.A. 2014. Cardio-

facio-cutaneous syndrome: clinical features, diagnosis, and management guidelines. *Pediatrics*134(4): 1149-1162.

- Pope, B. and Kent, H.M. 1996. High efficiency 5 minutes transformation of *Escherichia coli.Nucleic Acids Res.* 24(3): 536-537.
- Prior, I.A., Lewis, P.D., and Mattos, C. 2012. A comprehensive survey of Ras Mutations in cancer. *Cancer Res.* 72: 2457–2467.
- Psahoulia, F.H., Moumtzi, S., Roberts, M.L, Sasazuki, T., Shirasawa, S. and Pintzas, A. 2007. Quercetin mediates preferential degradation of oncogenic Ras and causes autophagy in HRAS-transformed human colon cells. *Carcinogenesis*28(5): 1021-1031.
- Pulciani, S., Santos, E., Lauver, A.V., Long, L.K., Aaronson, S.A., Barbacid, M. 1982. Oncogenes in solid human tumours. *Nature* 300: 539-542.
- Pusztaszeri, M., Soccal, P.M., Mach, N., Pache, J.L., and Kee, T.M. 2012. Cytopathological Diagnosis of Non Small Cell Lung Cancer: Recent Advances Including Rapid On-Site Evaluation, Novel Endoscopic Techniques and Molecular Tests. *J. Pulmonar. Respirat. Med.* 5:2.
- Puri, A., Saxena, R., Saxena, R.P., Saxena, K.C., Srivastava, V., and Tandon, J.S. 1993. Immunostimulant agents from *Andrographis paniculata*. J. Nat. Prod. 56(7): 995-999.
- Pierce, J. and Gutteridge, S. 1985. "Large-scale preparation of ribulose bisphosphate carboxylase from a recombinant system in *Escherichia coli* characterized by extreme plasmid instability." *Applied and Environ. Microbiol.* 49(5): 1094-1100.
- Pons, J., Evrard-Todeschi, N., Bertho, G., Gharbi-Benarous, J., Tanchou, V., Benarous, R., and Girault, J. P. 2008. Transfer-NMR and docking studies identify the binding of the peptide derived from activating transcription factor 4 to protein ubiquitin ligase beta-TrCP. Competition STD-NMR with beta-catenin. *Biochemistry* 47(1): 14-29.
- Prakash, A., Parsons, S.J., Kyle, S., McPherson, M.J. 2012. Recombinant production of self-assembling β-structured peptides using SUMO as a fusion partner. *Microb.Cell. Fact.* 11: 92.
- Quah, S.Y., Tan, M.S., Teh, Y.H., Stanslas, J. 2016. Pharmacological Modulation of Oncogenic Ras by Natural Products and Their Derivatives: Renewed Hope in the Discovery of Novel Anti-Ras Drugs. Pharmacol. Ther. 162: 35-57.
- Quan, Y., Liu, G., Yu, W., Nie, Z., Chen, J., Lv, Z., Zhang, Y. 2012. Expression, purification, and characterization of Ras protein (BmRas1) from Bombyx mori. *Comp. Funct. Genomics*747539.
- Rad-Malekshahi, M., Flement, M., Hennink, W.E., and Mastrobattista, E. 2014. Optimization of the recombinant production and purification of a selfassembling peptide in *Escherichia coli. Microb. Cell. Fact.* 13: 178.
- Rahib, L., Smith, B.D., Aizenberg, R., Rosenzweig, A.B., Fleshman, J.M., Matrisian, L.M. 2014. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. *Cancer Res.* 74(11): 2913-2921.
- Ranelletti, F.O., Maggiano, N., Serra, F.G., Ricci, R., Larocca, L.M., Lanza, P. 2000. Quercetin inhibits p21-RAS expression in human colon cancer cell lines and in primary colorectal tumors. *Int. J. of Cancer*85(3): 438-445.

- Razzaghi-Asl, N., Miri, R., Firuzi, O. 2016. Assessment of the Cytotoxic Effect of a Series of 1,4-Dihydropyridine Derivatives Against Human Cancer Cells. Iran J Pharm Res. 15(3):413-420.
- Redston, M. S., Caldas, C., Seymour, A. B., Hruban, R. H., da Costa, L., Yeo, C. J., and Kern, S. 1994. p53 mutations in pancreatic carcinoma and evidence of common involvement of homocopolymer tracts in DNA microdeletions. *Cancer Res.* 54: 3025.
- Rees D.C., Congreve M., Murray C.W., Carr R., 2004. Fragment-based lead discovery. *Nat. Rev. Drug. Discov*.3:660–672.
- Riss, T.L., Moravec, R.A., Niles, A.L., Duellman, S., Benink, H.A., Worzella, T.J., Minor, L. 2013. Cell Viability Assays. Assay Guidance Manual [Internet]. Retrieved 31 July 2017 from https://www.ncbi.nlm.nih.gov/pubmed/23805433/
- Rosano, G.L. and Ceccarelli, E.A. 2014. Recombinant protein expression in *Escherichia coli*: advances and challenges. *Front. Microbiol.* 5: 172.
- Rozenblum, E., Schutte, M., Goggins, M. 1997. Tumor-suppressive pathways in pancreatic carcinoma. *Cancer Res.* 57:1731 –1734.
- Russo Krauss, I., Merlino, A., Vergara, A., and Sica, F. 2013. An overview of Biological macromolecule crystallization. *Int. J. Mol. Sci.* 14(6): 11643-11691.
- Santos, E., Nebreda, A.R., Bryan, T., and Kempner, E.S. 1988. Oligomeric structure of p21 ras proteins as determined by radiation inactivation. *J. Biol. Chem.* 263(20): 9853 -9858.
- Santos, E. and Nebreda, A.R. 1989. Structural and functional properties of ras proteins. *FASEB J.* 3:2151-2163.
- Satakarni, M. and Curtis, R. 2011. Production of recombinant peptides as fusions with SUMO. *Protein. Expr. Purif.* 78(2): 113-119.
- Saturation transfer difference-nuclear magnetic resonance. N.d. Retrieved 31 July 2017 from http://glycopedia.eu/e-chapters/NMR-for-Structural-Glycoscience- 35/Saturation-Transfer-Difference
- Scheffzek, K., Ahmadian, M.R., Kabsch, W., Wiesmuller, L., Lautwein, A., Schmitz, F., Wittinghofer, A. 1997. The Ras-RasGAP complex: structural basis for GTPase activation and its loss in oncogenic Ras mutants. *Science*277(5324): 333-338.
- Schena, A., Griss, R., and Johnsson, K. (2015). Modulating protein activity using tethered ligands with mutually exclusive binding sites. *Nat. Commun.* 22(6): 7830.
- Schmitt, J., Hess, H., and Stunnenberg, H.G. 1993. Affinity purification of histidine-tagged proteins. *Mol. Biol. Rep.* 18(3):223-230.
- Sezonov, G., Joseleau-Petit, D., and D'Ari, R. 2007. Escherichia coli physiology in Luria-Bertani broth. *J. Bacteriol.* 189(23): 8746-8749.
- Sheeja, K., Guruvayoorappan, C., and Kuttan, G. 2007. Antiangiogenic activity Of Andrographis paniculata extract and andrographolide. *Int. Immunopharmacol.* 7(2): 211-221.
- Shih, T.Y., Weeks, M.O., Young, H.A., and Scholnick, E.M. 1979. Identification of a sarcoma virus-coded phosphoprotein in nonproducer cells transformed by Kirsten or Harvey murine sarcoma virus. *Virology* 96(1): 64-79.

- Shiloach, J. and Fass, R. 2005. Growing *E*. coli to high cell density: a historical perspective on method development. *Biotechnol. Adv.* 23(5): 345-357.
- Shima, F., Yoshikawa, Y., Ye, M., Araki, M., Matsumoto, S., Liao, J., Hu, L., Sugimoto, T., Ijiri, Y., Takeda, A., Nishiyama, Y., Sato, C., Muraoka, S., Tamura, A., Osoda, T., Tsuda, K., Miyakawa, T., Fukunishi, H., Shimada, J., Kumasaka, T., Yamamoto, M., and Kataoka, T. 2013. In silico discovery of small-molecule Ras inhibitors that display antitumor activity by blocking the Ras-effector interaction. *Proc. Natl. Acad. Sci. U.S.A* 110(20): 8182-8187.
- Singha, P.K., Roy, S., and Dev, S. 2003. Antimicrobial activity of *Andrographis paniculata. Fitoterapia* 74(7-8): 692-694.
- Singha, P.K., Roy, S., and Dev, S. 2007. Protective activity of andrographolide And arabinogalactan proteins from *Andrographis paniculara* Nees. against ethanol-induced toxicity in mice. *J. Ethnopharmacol.* 111(1): 13-21.
- Skinner, A.L. and Laurence, J.S. 2008. High-field solution NMR spectroscopy as a tool for assessing protein interactions with small molecule ligands. *J. Pharm. Sci.* 97(11): 4670-4695.
- Slabinski, L., Jaroszewski, L., Rodrigues, A.P., Rychlewski, L., Wilson, I.A., Lesley, lessons from structural genomics. *Protein Sci.* 16(11): 2472-2482.
- Smith, D.B. and Johnson, K.S. 1988. Modified glutathione S-transferase fusion Proteins for simplified analysis of protein - protein interactions. *Gene* 67: 31-40.
- Spiegel, J., Cromm, P.M., Zimmermann, G., Grossmann, T.N., Waldmann, H. 2014. Small-molecule modulation of Ras signaling. *Nat. Chem. Biol.* 10(8): 613-622.
- Sprangers, R., Velyvis, A., and Kay, L.E. 2007. Solution NMR of supramolecular complexes: providing new insights into function. *Nat. Methods.* 4(9): 697-703.
- Stehelin, D., Varmus, H.E., Bishop, J.M., Vogt, P.K. 1976. DNA related to the transforming gene(s) of avian sarcoma viruses is present in normal avian DNA. *Nature* 260(5547): 170-173.
- Sun, Q., Burke, J.P., Phan, J., Burns, M.C., Olejniczak, E.T., Waterson, A.G., Lee, T., Rossanese, O.W., Fesik, S.W. 2012. Discovery of small molecules that bind to K-Ras and inhibit Sos-mediated activation. *AngewChemInt Ed Engl* 51(25): 6140-6143.
- Tidyman, W.E. and Rauen, K.A. 2009. The RASopathies: developmental syndromes of Ras/MAPK pathway dysregulation. *Curr Opin Genet. Dev*.19: 230-236.
- Torre, L.A., Bray, F., Siegel, R.L., Ferlay, J., Lortet-Tieulent, J. and Jemal, A. 2015. Global cancer statistics, 2012. *CA: A Cancer J. for Clinicians* 65: 87-108.
- Traut, T.W. 1994. Physiological concentration of purines and pyrimidines. *Mol. Cell. Biochem.*140(1): 1-22.
- Trepe, K. 2006. Overview of bacterial expression systems for heterologous protein production: from molecular and bio- chemical fundamental to commercial systems. *Applied Microbiol. and Biotechnol.* 72(2): 211-222.

- Vector map of Champion pET SUMO. N.d. Retrieved 31 July 2017 from www.snapgene.com
- Vector map of pEXV KRas G12V. N.d. Retrieved 31 July 2017 from www.snapgene.com
- Veeresham, C. 2012. Natural products derived from plants as a source of drugs. J. Adv. Pharmaceutical Technology & Research 3(4): 200-201.
- Venkitakrishnan, R.P., Benard, O., Max, M., Markley, J.L., and Assadi-Porter, F.M. 2012. Use of NMR saturation transfer difference spectroscopy to study ligand binding to membrane proteins. *Methods. Mol. Biol.* 914: 47-63.
- Wang, Y., Kaiser, C.E., Frett, B., Li, H.Y. 2013. Targeting mutant KRAS for anticancer therapeutics: a review of novel small molecule modulators. *J. Med. Chem.* 56(13): 5219-5230.
- Wang, Y.S., Liu, D., and Wyss, D.F. 2004. Competition STD NMR for the detection of high-affinity ligands and NMR-based screening. *Magn. Reson. Chem.* 42(6): 485-489.
- Weisz, B., Giehl, K., Gana-Weisz, M., Egozi, Y., Ben-Baruch, G., and Marciano, D. 1999. A new functional Ras antagonist inhibits human pancreatic tumour growth in nude mice. *Oncogene*18(16): 2579-2588.
- Whyte, D. B., Kirschmeier, P., Hockenberry, T. N., Nunez-Oliva, I., James, L., Catino, J. J. 1997. K-Ras and N-Ras are geranylgeranylated in cells treated with farnesyl protein transferase inhibitors. *The Journal of Biol. Chem.* 272: 14459-14464.
- Williams, C. S., Goldman, A.P., Sheng, H., Morrow, J.D., and DuBois, R.N. 1999. Sulindac Sulfide, but not Sulindac Sulfone, Inhibits Colorectal Cancer Growth. *Neoplasia*1(2): 170-176.
- Wishart, D. 2005. NMR spectroscopy and protein structure determination: applications to drug discovery and development. *Curr. Pharm. Biotechnol.* 6(2): 105-120.
- Wong, H.C., Wong, C.C., Sagineedu, S.R., Loke, S.C., Lajis, N.H., Stanslas, J. 2014. SRJ23, a new semisynthetic andrographolide derivative: *in vitro* growth inhibition and mechanisms of cell cycle arrest and apoptosis in prostate cancer cells. *Cell. Biol. Toxicol*.30(5): 269-288.
- Wong, S., and Xie, M. 2009. GDC-0449-a potent inhibitor of the hedgehog pathway. *Bioorg. Med. Chem. Lett.* 19(19): 5576-5581.
- Zang, R., Ding, L., Tang, I-C., Wang, J., amd Yang, S-T. 2012. Cell-Based Assays in High-Throughput Screening for Drug Discovery. *Int. J. Biotechnol. for Wellness Industries*1: 31-51.
- Zhang, C. and Liu, Y. 2015. Targeting cancer with sesterterpenoids: the new Potential antitumor drugs. *J. Nat. Med.* 69(3): 255-266.
- Zhang, Z., Kuipers, G., Niemiec, Ł., Baumgarten, T., Slotboom, D.J., de Gier, J.W., and Hjelm, A. 2015. High-level production of membrane proteins in *E. coli* BL21(DE3) by omitting the inducer IPTG. *Microb. Cell. Fact.* 14: 142.
- Zheng, N, Perez Jde, J., Zhang, Z., Dominguez, E., Garcia, J.A., and Xie, Q. 2008. Specific and efficient cleavage of fusion proteins by recombinant plum pox virus NIa protease. *Protein. Expr. Purif.*57(2): 153-16