

UNIVERSITI PUTRA MALAYSIA

BIODEGRADATION OF OIL PALM FIBERS USING LOCALLY ISOLATED FUNGI (Pycnoporus sanguineus) THROUGH PLANT BIOMECHANICS APPROACH

FARAH NADIA BINTI OMAR

FK 2018 20



BIODEGRADATION OF OIL PALM FIBERS USING LOCALLY ISOLATED FUNGI (*Pycnoporus sanguineus*) THROUGH PLANT BIOMECHANICS APPROACH



By

FARAH NADIA BINTI OMAR

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in fulfilment of the requirement for the Degree of Doctor of Philosophy

November 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



UPM

(C)

Dedicated to my family For your endless love, support and encouragement Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

BIODEGRADATION OF OIL PALM FIBERS USING LOCALLY ISOLATED FUNGI (Pycnoporus sanguineus) THROUGH PLANT BIOMECHANICS APPROACH

By

FARAH NADIA BINTI OMAR

November 2017

Chairman Faculty Azhari Samsu Baharuddin, PhD Engineering

Utilization of lignocellulosic OPEFB fiber has tremendously seen in Malaysia due to the cellulose and hemicellulose content. Conversion of these biopolymers into valuable products remains a challenging task with the presence of the recalcitrant lignin and scattering silica bodies on the fiber surface. Therefore, this study investigates the mechanical behaviour of the complex lignocellulosic OPEFB fiber containing silica bodies and provide an in-depth understanding of the delignification of OPEFB by fungi for further bioconversion into wide range of biomaterial applications. The microstructure of silica bodies on OPEFB fiber surface was modelled using finite element method, based on the results obtained from scanning electron microscope (SEM) images, tensile tests and X-ray microtomography (micro-CT) images. Silica body geometry, possible anisotropy/ orthotropy, debonding between the interface of the silica body and fiber, fiber thickness and presence of vascular bundle in the OPEFB were investigated through 2D and 3D models and analysed by commercial finite element software, Abaqus.

In 2D model, silica bodies contribute on integrity or strength of the fiber, however, in the 3D model, the effect of silica bodies on the elasticity of the fiber was insignificant when the thickness of the fiber is larger than 0.2 mm. In the developed representative volume element (RVE) and micro-CT models, the simulation results show that the difference of the fiber model with and without silica bodies are larger under shear than compression and tension. However, in comparison to geometrical effect (silica bodies), lignin, cellulose, and hemicellulose components of the fiber are responsible for the complex mechanical and interface behavior of oil palm fibers.

Hence, screening and isolation of lignin degrading fungi for deconstruction of lignin polymer in OPEFB was carried out. About 47 isolated fungi collected from environmental samples with six fungi were able to decolorize selective agar media, indicating possible presence of lignin-degrading enzymes; laccase and peroxidases. The highest producer of ligninolytic enzymes was identified as *Pycnoporus sanguineus* which able to utilize raw OPEFB fiber through solid state fermentation (SSF) with an increment of 1.37 folds of ligninolytic enzymes production as compared to submerged fermentation (SmF). Optimization study of different substrate pre-treatments (sodium hydroxide, Soxhlet extraction), incubation temperatures (20-40°C), ABTS concentrations (0-4%) and substrate amounts (3-15 g) on ligninolytic enzymes production was carried out. Results showed that the optimum conditions for *P. sanguineus* to produce highest laccase (15.49 U/g) with Klason lignin removal at 7.11% were using extractive-free OPEFB fiber, incubation temperature at 30°C, supplemented with 4 mM of ABTS and with 10 g of substrate loading size. Effectiveness of *P. sanguineus* for OPEFB degradation was further evaluated with the different ratio of fiber, fungi and palm oil mill effluent (POME) sludge as inoculum.

The relationship between structural OPEFB fiber degradation and delignification process by *P. sanguineus* was studied through tensile testing data, enzymatic and lignin component data, and micro-CT images. The highest total lignin loss (35.81%) and total phenolic content produced (78.03%) was determined at a condition ratio of fiber to fungi (60:40), yielding of laccase and MnP of 0.18 and 0.02, respectively while production rate of laccase and MnP were 0.98 U/g/d and 0.11 U/g/d, respectively. Micro-CT results revealed that the delignification process damaged the fiber based on the volume reduction data where 14.11% of volume reduction was observed with treated fiber while 11.21% volume reduction was achieved with untreated fiber. It is suggested that *P. sanguineus* could be a potential lignin degrader of OPEFB fiber before being manipulated for other valuable products production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

PENGURAIAN FIBER KELAPA SAWIT MENGGUNAKAN KULAT YANG DIPENCILKAN KAWASAN SETEMPAT (Pycnoporus sanguineus) MELALUI PENDEKATAN BIOMEKANIK TUMBUHAN

Oleh

FARAH NADIA BINTI OMAR

November 2017

Penyelia Fakulti

:

Azhari Samsu Baharuddin, PhD Kejuruteraan

Penggunaan sisa lignoselulosa gentian tandan kelapa sawit kosong (OPEFB) sangat banyak di Malaysia kerana kandungan selulosa dan hemiselulosa. Penukaran biopolimer ini kepada produk bernilai telah terhalang oleh kehadiran lignin dan partikel silika di permukaan gentian. Oleh itu, kajian penyelidikan tingkah laku mekanikal gentian OPEFB kompleks yang mengandungi partikel silika dijalankan dan memberi kefahaman proses penguraian lignin dalam OPEFB oleh kulat untuk seterusnya melalui proses biopenukaran kepada pelbagai jenis aplikasi biomaterial. Mikrostruktur partikel silika di gentian OPEFB telah disimulasi dengan menggunakan kaedah 'finite element' berdasarkan keputusan dari gambar imbasan mikroskop elektron (IME), ujian tegangan dan mikro-tomografi (mikro-CT). Geometri partikel silika, kebarangkalian anisotrophy/ orthotropy, peleraian ikatan antara permukaan partikel silika dan gentian, ketebalan gentian, dan kehadiran bukaan "vascular bundle' telah dikaji menggunakan model 2D dan 3D didalam perisian komersil "finite element", Abaqus.

Dalam model 2D, partikel silika menyumbang kepada integriti gentian, manakala dalam model 3D kesan partikel silika kepada kekenyalan gentian tidak signifikan pada ketebalan gentian melebihi 0.2 mm. Dalam model 'representative volume element' (RVE) dan model mikro-CT, keputusan simulasi menunjukkan perbezaan model gentian dengan dan tanpa partikel silika adalah besar di bawah mod ricih berbanding mampatan dan tegangan. Walau bagaimanapun, jika dibanding dengan kesan geometri (partikel silika), komponen lignin, selulosa dan hemiselulosa bertanggungjawab kepada tingkahlaku mekanikal dan antara permukaan yang kompleks pada gentian kelapa sawit. Oleh itu, penyaringan dan pemencilan kulat pengurai lignin dijalankan untuk menguraikan polimer lignin dalam OPEFB. Sebanyak 47 kulat dikutip dari kawasan sekitar dan enam kulat mampu menyah-warna agar media saringan menunjukkan kehadiran enzim pengurai lignin; laccase dan peroksida. Kulat yang menghasilkan enzim tertinggi dipilih dan dikenali sebagai Pycnoporus sanguineus yang mampu menguraikan gentian OPEFB dan menghasilkan enzim yang tinggi melalui fermentasi fasa pepejal iaitu peningkatan sebanyak 1.37 kali ganda berbanding dengan fermentasi fasa terendam. Kajian pengoptimuman pelbagai pra-rawatan substrat (natrium menggunakan hidroksida, pengestrakan Soxhlet), suhu pengeraman (20-40°), kepekatan ABTS (0-4%) dan jumlah substrat (3-15 g) kepada penghasilan enzim ligninolitik dijalankan. Keputusan menunjukkan keadaan optimum penghasilan enzim *laccase* tertinggi (15.49 U/g) dengan penyingkiran Klason lignin sebanyak 7.11% oleh P. sanguineus adalah menggunakan gentian OPEFB bebas ekstraktif, suhu pengeraman 30°C, dibekalkan 4 mM ABTS dan dengan menggunakan 10 g substrat.

Kecekapan P.sanguineus untuk menguraikan gentian OPEFB dikaji lebih mendalam dengan menggunakan pelbagai nisbah gentian, kulat dan enap cemar dari sisa efluen kilang sawit (POME) sebagai inokulum. Hubungan antara penguraian struktur gentian OPEFB dan proses penguraian lignin oleh P. sanguineus dikaji melalui data ujian tegangan, enzim dan data komponen lignin, dan imej mikro-CT. Penyingkiran tertinggi lignin (35.81%) dan jumlah kandungan fenolik yang terhasil (78.03%) dikenalpasti pada keadaan nisbah gentian kepada kulat (60:40) menghasilkan laccase dan MnP masing-masing 0.18 dan 0.02, manakala kadar penghasilan laccase dan MnP masing-masing adalah 0.98 U/g/d dan 0.11 U/g/d. Keputusan mikro-CT menunjukkan proses penguraian lignin telah merosakkan gentian berdasarkan data pengurangan isipadu di mana 14.11% pengurangan isipadu telah diperolehi dari gentian terawat manakala 11.21% pengurangan isipadu diperolehi dari gentian yang tidak terawat. Ini membuktikan P. sanguineus berpotensi untuk menjadi kulat pengurai lignin gentian OPEFB sebelum gentian ini dimanipulasi untuk kegunaan pembuatan produk berharga.

ACKNOWLEDGEMENT

Alhamdulillah, thank you to Merciful God for giving me strength and courage to finish this project. My biggest appreciation goes to my main supervisor and supervisory committee, Dr Azhari Samsu Baharuddin, Dr Mohd Afandi P Mohammed, Dr Umi Kalsom Md Shah and Prof Kenji Sakai for their endless guidance, supports and valuable wisdoms. To all fellow lecturers in Faculty of Engineering that always inspire me and sharing precious know-how both directly and indirectly. I also would like to express my gratitude to Ministry of Science, Technology and Innovation of Malaysia (MoSTI) and Universiti Putra Malaysia for their financial aids throughout my studies.

I humbly thank you to all the technicians and laboratory staffs in Faculty of Engineering for their cooperation and kind help. A special appreciation also goes to my fellow Bioprocess Research Group: Halimatun, Suhaiza, Loo, Ida, Termizi and others for their joyful and memorable friendship moments. The appreciation is beyond words.

Up and foremost, my indebtedness goes to my parents and my husband as well as my family for their faith, understanding, and prayers. They have always been my rock and always will be. Last but not least, I would like to thank to everyone who involved directly and indirectly for completion of my studies. I am truly treasure your help and guidance for all this while. Only God could ever repay each one of you. Thank you. I certify that a Thesis Examination Committee has met on 20 November 2017 to conduct the final examination of Farah Nadia binti Omar on her thesis entitled "Biodegradation of Oil Palm Fibers using Locally Isolated Fungi (*Pycnoporus sanguineus*) through Plant Biomechanics Approach" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Mohd Noriznan bin Mokhtar, PhD Associate Professor Ing. Faculty of Engineering Universiti Putra Malaysia (Chairman)

Luqman Chuah Abdullah, PhD Professor Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Rosnah binti Shamsudin, PhD Associate Professor Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Minato Wakisaka, PhD Professor Kyushu Institute of Technology Japan (External Examiner)

NOR AINI AB. SHUKOR, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 29 January 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows;

Azhari Samsu Baharuddin, PhD

Senior Lecturer Faculty of Engineering Universiti Putra Malaysia (Chairman)

Mohd Afandi P Mohammed, PhD

Senior Lecturer, Faculty of Engineering, Universiti Putra Malaysia (Member)

Umi Kalsom Md Shah, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Kenji Sakai, Phd

Professor Faculty of Agriculture Kyushu University, Japan (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

___Date:

Name and Matric No: Farah Nadia Binti Omar (GS 35338)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Chairman of Supervisory Committee: Dr Azhari Samsu Baharuddin

Signature:

Member of Supervisory Committee: Dr Mohd Afandi P Mohammed

Signature:

Member of Supervisory Committee: Dr Umi Kalsom Md Shah

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENT	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xx
LIST OF SYMBOLS	xxi

IAPT	ER			
INT	RODUC	TION		
1.1	Lignin	degradat	ion of OPEFB	1
1.2	Oil pal	m fiber bi	omechanics	2
1.3	Proble	m stateme	ent	2
1.4	Scope	of researc	h and thesis structure	4
ттт	RATIE	RE REVIE	W	
	-			7
				ç
	-	0		10
2.0	-			11
	2.0.1	OPEFB	and incentation characteristics of	
		2.3.1.1	Morphology of OPEFB	11
		2.3.1.2	Mechanical Behavior of OPEFB	13
		2.3.1.3	Silica Bodies on OPEFB Fiber	14
	2.3.2	Chemica	al characteristics of OPEFB	17
	2.3.3	Microm	echanics modelling of natural fiber	19
			0	
2.4	Biodeg			21
	2.4.1			21
	2.4.2	Ligninol	ytic enzymes-producing	23
		0		
	2.4.3			24
		2.4.3.1	Laccase	24
		2.4.3.2	Lignin Peroxidase	25
		2.4.3.3	ē	26
		2.4.3.4		28
	2.4.4	Ferment	0	28
		2.4.4.1		28
		2.4.4.2	Solid State Fermentation	29
	INT] 1.1 1.2 1.3 1.4 LITH 2.1 2.2 2.3	 1.1 Lignin 1.2 Oil pal 1.3 Proble 1.4 Scope LITERATUE 2.1 Overvi 2.2 Oil pal 2.3 Oil pal 2.3.1 2.3.2 2.3.3 2.4 Biodeg 2.4.1 2.4.2 2.4.3 	INTRODUCTION1.1Lignin degradat1.2Oil palm fiber bi1.3Problem stateme1.4Scope of researchLITERATURE REVIE2.1Overview of lign2.2Oil palm lignoce2.3Oil palm empty2.3Oil palm empty2.3.1Physical0PEFB2.3.1.12.3.2Chemica2.3.3Micromeand oil p2.4Biodegradation2.4.1Pretreatt2.4.2Ligninol2.4.3Ligninol<	INTRODUCTION 1.1 Lignin degradation of OPEFB 1.2 Oil palm fiber biomechanics 1.3 Problem statement 1.4 Scope of research and thesis structure IITERATURE REVIEW 2.1 Overview of lignocellulosic biomass 2.2 Oil palm lignocellulosic biomass resources 2.3 Oil palm empty fruit bunch 2.3.1 Physical and mechanical characteristics of OPEFB 2.3.1.1 Morphology of OPEFB 2.3.1.2 Mechanical Behavior of OPEFB 2.3.3 Silica Bodies on OPEFB Fiber 2.3.2 Chemical characteristics of OPEFB 2.3.3 Micromechanics modelling of natural fiber and oil palm 2.4 Biodegradation of OPEFB 2.4.1 Pretreatment of OPEFB 2.4.2 Ligninolytic enzymes-producing microorganisms 2.4.3 Ligninolytic enzymes 2.4.3.1 Laccase 2.4.3.2 Lignin Peroxidase 2.4.3.3 Manganese Peroxidase 2.4.3.4 Lignin mediator system 2.4.4 Fermentation Strategies 2.4.4.1 Submerged Fermentatio

		2.4.4.3	Factors influencing SSF of lignin degradation	31
	2.4.5	Lignino	lytic enzymes production and biomass	33
	2.1.0	0	ition by white rot fungi	00
	2.4.6		ication and biodegradation effects on	34
		0	characteristics	-
2.5	Concl	uding rem		36
		Ũ		
			ALS AND METHODS	
3.1	Mater		1	37
	3.1.1		al reagents	37
	3.1.2		e preparation	37
			Oil palm empty fruit bunch	37
		3.1.2.2	Palm oil mill effluent sludge	38
3.2	Metho			20
	3.2.1		echanical modelling of silica bodies on	38
		OPEFB 1		20
		3.2.1.1	2D single particle model	38
		0.01.0	development	10
		3.2.1.2	2D multi particle model	40
		0.0.1.0	development	42
		3.2.1.3	-	43
		3.2.1.4		44
		3.2.1.5		46
	2.2.2	T	development	4 77
	3.2.2		biodegradation of OPEFB fiber	47
		3.2.2.1	Fungal screening and isolation	48
		3.2.2.2	Culture conditions for ligninolytic	48
		2222	enzymes production	10
		3.2.2.3	Development of fungal consortia	49
		3.2.2.4	Sampling procedures for ligninolytic enzymes production	49
3.3	Analy	tical meth	· · ·	49
0.0	-		nation of lignocellulose content	49
	3.3.2		nation of ligninolytic enzymes	51
	0.0.	activitie		01
		3.3.2.1	Laccase	51
		3.3.2.2	Lignin peroxidase	51
		3.3.2.3	Manganese peroxidase	52
	3.3.3		nation of total phenolic content	52
	3.3.4		ical testing of OPEFB fiber	52
	3.3.5		g electron microscope	53
	3.3.6		icrotomography	54
	3.3.7	2	transform infrared spectroscopy	54

4	FIBE		CHANIC	AL ANALISIS AND MODELING O	FOPEFB		
	4.1		uction		55		
	4.2	Physical analysis					
		4.2.1		ls and methods	56		
			4.2.1.1		56		
			4.2.1.2		56		
			4.2.1.3	0 1	57		
			4.2.1.4	5 0 1 5	57		
		4.2.2		and discussion	57		
			4.2.2.1		57		
				results			
	4.3	Micro	mechanics	s modelling of silica bodies on OPEFB	64		
		4.3.1		le particle models	64		
			4.3.1.1		64		
				fraction			
			4.3.1.2		65		
				circular silica bodies			
			4.3.1.3		66		
				fiber			
			4.3.1.4	Effect of interface between silica	67		
				bodies and fiber			
		4.3.2	2D mult	ti particle models	69		
			4. <mark>3.2.1</mark>	Effect of silica bodies arrangement	69		
				on OPEFB			
			4. <mark>3.2.2</mark>	Effect of interface debonding	70		
				between silica bodies and fiber			
		4.3.3	3D mod	lels of OPEFB fiber	72		
			4.3.3.1	Effect of fiber thickness	73		
			4.3.3.2	Effect of vascular bundle presence	74		
		4.3.4	Constitu	ative model	75		
		4.4.4	Compar	rison of micro-CT and RVE modelling	77		
	4.4	Conclu	uding ren	narks	84		
5				OCAL ISOLATED FUNGI FOR OPE	FB		
				N	05		
		Introd		.1 1	85		
	5.2		ials and m		86		
		5.2.1	Oil paln		86		
		5.2.2		al pretreatments of OPEFB fiber	86		
		5.2.3		ng and isolation of ligninolytic fungi	87		
		E 0 4		vironmental sample	07		
		5.2.4 5.2.5		conditions and sampling procedures	87		
		5.2.5	-	oment of compatible fungal consortia	87		
		5.2.6		e analyses	87		
			5.2.6.1	Laccase	87		
			5.2.6.2	Lignin peroxidase	88		

ANALVER AND MODELING OF OPEED 4 R т

		5.2.6.3 Manganese peroxidase	88
	5.2.7	Identification of ligninolytic fungi	88
	5.2.8	Determination of lignocellulose content	88
	5.2.9	Determination of total phenolic content	88
5.3	Results	s and discussion	89
	5.3.1	Characterization of substrate OPEFB	89
	5.3.2	Screening and isolation of ligninolytic fungi	89
	5.3.3	Identification of ligninolytic fungi	92
	5.3.4	Preliminary study of ligninolytic enzymes	92
		from Pycnoporus sanguineus	
		5.3.4.1 Development of compatible fungal consortia	92
		5.3.4.2 Effect of fermentation strategies on	97
		ligninolytic enzymes production	
	5.3.5	Effect of SSF parameters on laccase production	99
		5.3.5.1 Effect of substrate pretreatment	99
		5.3.5.2 Effect of incubation temperature	100
		5.3.5.3 Effect of ABTS concentration	101
		5.3.5.4 Effect of substrate amount	102
	5.3.6	Application of optimal conditions for laccase	103
		production	
5.4	Conclu	iding Remarks	105
6 OPE		DEGRADATION AND MICROSTRUCTURE S	TUDIES
6.1	Introd		106
6.2		als and methods	108
	6.2.1		108
	6.2.2		108
	6.2.3	POME sludge	108
	6.2.4	Microorganism and maintenance	108
	6.2.5	Solid state fermentation	108
	6.2.6	Tensile tests	109
	6.2.7	X-ray microtomography	109
	6.2.8	0 1	109
6.3	6.2.9 Rocult	Determination of total phenolic content s and discussion	109 110
0.5	6.3.1		110
	6.3.2	Characterization of OPEFB and POME sludge Raw versus pretreated fibers	110
	6.3.3	Biodegradation experiments results	110
	6.3.4	Solid state fermentation effectiveness	112
	6.3.5	X-ray microtomography analysis	115
	6.3.6	Relationship between physico-chemical and	120
	0.0.0	structural analyses of fibers degradation	120
6.4	Conclu	iding remarks	121
011		0	

7			ON AND RECOMMENDATIONS FOR ESEARCH	
	7.1	Concl	usions	122
		7.1.1	Micromechanics studies of OPEFB fiber	122
		7.1.2	Utilization of local isolated fungi for OPEFB degradation	122
		7.1.3	OPEFB degradation and microstructure studies	122
	7.2	Recon	nmendations for future works	123

REFERENCES APPENDICES BIODATA OF STUDENT LIST OF PUBLICATIONS

 \bigcirc

123 151 157

158

LIST OF TABLES

Table		Page
2.1	Categories of lignocellulosic biomass and their product	s 8
2.2	Diameter of oil palm fiber and other natural fibers	12
2.3	Fiber dimension value of OPEFB fiber	12
2.4	Mechanical properties of palm oil fiber and other natural fibers	13
2.5	Chemical properties and composition of OPEFB	17
2.6	Summary of pretreatment technologies on OPEFB for various purposes	23
2.7	Ligninolytic enzymes produced by microorganisms through SSF technique	30
2.8	Chemical inducers of laccase in white rot fungi	32
2.9	Comparison of delignification process	34
3.1	Materials parameters used in the 2D micromechanics model of OPEFB	43
3.2	Material parameters used to model oil palm fiber	45
3.3	Screening medium for ligninolytic enzyme producing fungi	48
5.1	Characterization of lignocellulosic OPEFB	89
5.2	The ability of local isolated fungi to produce clearing zone on RBBR agar plates after 3 days and 7 days of incubation	90
5.3	Ligninolytic enzymes production for different isolates using raw OPEFB and commercial Kraft lignin (KL)	91
5.4	Maximum ligninolytic enzymes production from monoculture and co-culture of local isolated of ligninolytic fungi	105
5.5	Laccase enzyme production from different species of	103
	rotting fungi through solid state fermentation	
6.1	Characterization of OPEFB and POME sludge	110
6.2	Loss of lignin and total phenolic compound from solid state fermentation of raw OPEFB and treated OPEFB	113
6.3	with respective ratio of POME sludge inoculum Kinetic data from solid state fermentation of OPEFB	116

G

LIST OF FIGURES

Figure		Page
1.1	Overview of the overall experimental design	6
2.1	Structure of lignocellulose consisting fractions of cellulose, hemicelluloses and lignin	7
2.2	Oil palm biomass wastes generated in Malaysia in 2009	10
2.3	Photographs of (a) an empty fruit bunch; (b) shredded OPEFB; (c) press-shredded OPEFB in the mill	11
2.4	(a) General uptake of silicone in a monocot plants (b) Silica bodies in the OPEFB fiber surfaces	16
2.5	Lignocellulose components in plant cell walls	18
2.6	Mechanism of laccase action for both phenolic and non- phenolic substrates	25
2.7	Catalytic reaction of lignin peroxidase	26
2.8	Catalytic cycle of manganese peroxidase	27
2.9	Fiber surface of OPEFB. (a) Untreated OPEFB, (b) fungal pretreated OPEFB, (c) fungal pretreated OPEFB strand covered by fungal mycelium, (d) cross section of fungal pretreated OPEFB	35
3.1	(a) The microstructure finite element model of OPEFB; and simulation images result of 10 spikes filler (b) and circular filler (c) models. The S22 colour scale indicates principal stress in y direction (unit in MPa).	38
3.2	2D micromechanics models of OPEFB surface: (a) random arrangement multi-particles model, (b) a single particle with 10 spikes, and (c) a simulation image of a multi-particles model where the scale is in MPa (von- Mises stress)	41
3.3	Traction versus separation of the cohesive zone model under Mode 1 (normal tension mode)	41
3.4	3D finite element model of silica body embedded halfway in the matrix fiber	43
3.5	Springs and dampers arrangement used for the Parallel Rheological Network (PRN) model	44
3.6	Geometry and modelling results of (a) RVE and (b) micro- CT elements models. The simulation results shown are the von-Mises stress (scales shown in MPa).	47
3.7	Tensile test configuration of an oil palm fiber consisting of "C-shaped" paper to hold the fiber.	53
4.1	Silica bodies embedded in OPEFB fiber	56
4.2	SEM images of OPEFB fiber and silica bodies. (a) Random arrangement of silica bodies on the fiber surface, (b) aligns arrangement of silica bodies, (c) close-up view of silica bodies, (d) cross-section of OPEFB (undeformed	58

G

	sample), (e) cross-section at fractured region, and (f) sample surface after fracture with some silica bodies absent (craters)	
4.3	Silica bodies diameter distribution from SEM images	59
4.4	Slices of micro-CT scan of raw OPEFB fiber across the fiber length.	60
4.5	Cellular area fraction at different sample's height (from micro-CT scan slices). The dotted line shows the average value of cellular area fraction.	61
4.6	FTIR spectra of OPEFB fibres to highlight the trace of Si from the fibre containing silica bodies.	61
4.7	Typical mechanical test results of untreated OPEFB fiber with (a) tensile test (b) cyclic test (c) stress relaxation	63
4.8	Effect of filler volume fraction under: a) uniaxial tension, b) uniaxial compression, and c) simple shear.	65
4.9	Effect of silica-body geometry under: (a) uniaxial tension, (b) uniaxial compression, and (c) simple shear. Note that all the results are shown at 15 % filler volume fraction	66
4.10	(a) Traction versus separation of the cohesive damage model under Mode I, Mode II or Mode III. Parametric study of CDM under uniaxial tension for: (b) critical stress, and (c) different cohesive energy. (d) shows close-up of results using different cohesive energy	68
4.11	Modeling results of 2D multi-particles where the curves "Random", "Aligned", and "Clustered" refer to the arrangement of silica bodies in the models.	70
4.12	Parametric study of 2D models with CZM: (a) results under different critical stress, (case 1), (b) results under different cohesive energy (case 2). The multi-particle model with random arrangement is used.	71
4.13	2D modelling results using anisotropy material models and CZM	72
4.14	Comparison of 3D models and the experimental results	73
4.15	3D finite element results for different matrix thicknesses, t. (a) Perfect silica body and fiber interface and (b) results with CZM activated using parameters in Table 3.1.	73
4.16	Modelling results with and without the inclusion of vascular bundle opening	74
4.17	Tensile tests results and material model fit under (a) cyclic, and (b) stress relaxation	76
4.18	Sensitivity study of the PRN model by excluding different functions of the model under (a)–(c) cyclic, and (d) stress relaxation	77

	4.19	Tensile modelling results from the RVE and micro-CT finite element models.	78
	4.20	Shear modelling results from the RVE and micro-CT finite element models	79
	4.21	Compression modelling results from the RVE and micro- CT finite element models	80
	4.22	Comparison between RVE and micro-CT models with and without silica bodies	81
	4.23	Different interaction between oil palm fiber	82
	4.24	Replot from Xiang et al. (2015) of elastic and plastic strains (from loading and unloading tensile tests) with cellulose content of alkali pre-treated oil palm fibers	83
	5.1	Schematic diagram of interactions between two different fungal strains, which were grown adjacently (4 cm apart) on MEA culture media (based on the observations of Porter (1924) and Stahl & Christensen (1992))	93
	5.2	Compatibility studies of lignin degraders on Malt Extract Agar (MEA) solid media. Mutual intermingling interactions on b and h; partial mutual intermingling interactions on a, e and i; replacement/ invasion interaction on f and g; deadlock interaction on c, d and j. All pictures were taken on 7 days incubation period	95
	5.3	The effect of fermentation cultures on ligninolytic enzyme production of <i>P.sanguineus</i> and total phenolic content (TPC).	97
	5.4	Ligninolytic enzymes profiles of laccase (•) and manganese peroxidase (•) of <i>Pycnoporus sanguineus</i> . Solid state fermentation was conducted at 30°C of incubation temperature for 10 days	98
	5.5	Effect of substrate pretreatment on laccase production and lignin component loss by <i>P.sanguineus</i> in solid state fermentation.	99
	5.6	Effect of incubation temperature on laccase production and lignin component loss by <i>P.sanguineus</i> in solid state fermentation.	100
	5.7	Effect of ABTS concentration on laccase production and lignin component loss by <i>P.sanguineus</i> in solid state fermentation	102
	5.8	Effect of substrate loading on laccase production and lignin component loss by <i>P.sanguineus</i> in solid state fermentation.	103
9	5.9	Production of laccase and lignin component loss of OPEFB by <i>P.sanguineus</i> under optimum condition (4 mM ABTS, 30°C, substrate amount 10g)	104

- 6.1 Stress strain curves of raw and alkaline treated OPEFB 111 fibers. Note that three duplicates of raw and pre-treated fibers were shown
- 6.2 OPEFB fiber surface before (left) and after (right) alkaline pretreatment. Silica bodies are clearly seen on the fiber and empty craters are resulted from the removal of silica bodies
- 6.3 Production of laccase, MnP and Klason lignin of local isolated Pycnoporus sanguineus during solid state fermentation of OPEFB fibers
- Images from micro-CT scan analyses showed raw fiber 6.4 before degradation at 500 and 10 000 um (a-b), after degradation at 500 and 10 000 um (c-d), treated fiber before degradation at 500 and 10 000 um (e-f) and treated fiber after degradation at 500 and 10 000 um (g-h).
- 6.5 3D images of OPEFB before (left) and after (right) 119 delignification process by *P.sanguineus*.
- Volume of treated and raw OPEFB fiber for both before 6.6 119 (Week 0) and after (Week 5) biodegradation by P.sanguineus with respective Klason lignin content. Results are mean of duplicate analyses.

112

118

114

LIST OF ABBREVIATIONS

ABTS	2,2`-azinobis-(3-ethyl-)benzothiazoline-6-sulphonate
MnP	Manganese peroxidase
LiP	Lignin peroxidase
OPEFB	Oil palm empty fruit bunch
POME	Palm oil mill effluent
SSF	Solid state fermentation
SmF	Submerged fermentation
FEA	Finite element analysis
TPC	Total phenolic compound
KL	Klason lignin
Micro-CT	Micro computed tomography
Si	Silica bodies
SiO ₂	Silica oxide
H ₂ SO ₄	Sulphuric acid
NaOH	Sodium hydroxide
RVE	Representative volume element

LIST OF SYMBOLS

°C	degree celcius
g/L	gram per litre
%	percentage
rev/min	revolutions per minute
β	beta
O ₂	oxygen
CO ₂	carbon dioxide
H ₂ O	water
h	hour
min	minutes
μm	micrometer
mm	milimeter
Pa	Pascal
U/g	unit per gram

CHAPTER 1

INTRODUCTION

1.1 Lignin degradation of OPEFB

Malaysia is blessed with valuable oil palm tree plantation where it has been as one of the major exporter of palm oil in the world where total oil palm planted area reported was 5.39 million hectares where it covered more than 73% of agricultural land that makes oil palm as a potential renewable biomass to be exploited for better use (Awaluddin et al., 2015). About 95.38 million tonnes of fresh fruit oil palm bunches were processed and the estimation of oil palm biomass generated from its process was 40.55 milliom tonnes (Loh, 2017). Oil palm empty fruit bunch (OPEFB) alone were contribute about 7.34 million tonnes where current practice of OPEFB manipulation involves incineration of OPEFB to produce bunch ash and further applied as soil conditioner and soil fertilizer and straight dumping on the field as soil mulching agent (Zainudin et al., 2012).

Some researchers have maximized the usage of OPEFB fibers as part of biocomposite materials in construction industries (Hassan et al., 2010) while some researchers use OPEFB as the main feedstock in bioconversion process into value added products such as fermentable sugars (Abu Bakar et al., 2012; Zainudin et al., 2012), biofuel (Sudiyani and Hermiati, 2010; Nieves et al., 2011), organic acids (Akhtar et al., 2014) and others. The cellulose and hemicelluloses is the most intriguing materials in the utilizing of the OPEFB as potential feedstock for the production of biofuel (Jeon et al., 2014; Kim and Kim, 2013), biochemicals (Reeb et al., 2014; Katinonkul et al., 2012). However, the utilization of cellulose and hemicellullose is hindered with the high content of lignin.

Lignin makes up of 15-40% of the dry matter of woody plant gives the rigidity and strength to cell walls and resilient towards degradation (Naseem et al., 2016). It is a highly stable biopolymer made of three cross-linked phenylpropane units and it present interlocking the cellulose and hemicelluloses polymers with strong ether bonds (C-O-C) and normal hydrogen bonds (C-C). Degradation of lignin in lignocellulosic biomass has been reported using various methods; 1) physical pretreatment such as by high pressure steam (Baharuddin et al., 2013); 2) chemical pretreatments by sodium hydroxide (Palamae et al., 2017; Zulkiple et al., 2016; Muryanto et al., 2015); and 3) biological pretreatment by fungal and ligninolytic enzyme. A number of fungi (white and brown rot) and some bacteria are effective as a lignin degrader due to their ability to produce lignolytic enzymes. Ligninolytic enzymes can be categorized as peroxidases (lignin peroxidase, manganese peroxidase, versatile peroxidase) and oxidative enzymes (laccase) could depolymerize the lignin polymers into smaller compounds through oxidative and electron transfer process (Bugg and Rahmanpour et al., 2015). Lignin can be precipitated as droplets on the surface of cellulose and hemicellulose, making them less accessible to enzymes attack. Hence, lignin removal is crucial in further utilization of cellulose and hemicellulose as it tends to adsorb the hydrolytic enzymes more easily and consequently reduce the effectiveness of the hydrolytic enzymes to access the cellulose and hemicellulose sites (Mishra et al., 2017; Li et al., 2009).

In addition, OPEFB fiber has some distinct features on its fiber surface, where random scattering protrusion silica bodies are found. These silica bodies are embedded half-way through the fiber surface, and it is made of silica oxide (SiO₂). It has been reported that silica bodies play a big role in providing mechanical support, strength and rigidity of the plant (Neethirajan et al., 2009; Ma and Yamaji, 2006). The presence of silica bodies in plant has been numerously studied especially on fermentable sugar production (Nurul Hazirah et al., 2016; Shamsudin et al., 2012). However, to this date, there are limited studies investigates the role of silica bodies in providing strength and rigidity towards plant particularly for oil palm tree. This issue, however, will be addressed and explained in this thesis focusing the presence of silica bodies on OPEFB fiber.

1.2 Oil palm fiber biomechanics

Micromechanics is a study of materials by understanding the interaction between constituent materials at microscopic level. Theoretically, it helps to compute and predict the behavior, properties and failure mechanisms of the materials. The main idea of micromechanics is to replace the original material with imaginary microscopic material so that the analysis of the original material could be understand and simplified (Yu, 2016). Micromechanics study have been used widely in building of materials as such each properties and behavior of the building material will be simulated and the overall performance of the material will be evaluated. A simple way to witness the micromechanics study is when natural fiber is used as reinforcement to other composite materials. The behavior of fibers will be simulated at various conditions and barriers and success and failure mechanisms of the overall materials will be evaluated. The micromechanics study of natural fibers like woody and plant cells have been well established (Hayot et al., 2012; Burgert and Dunlop, 2011). However, very limited studies are available in the literature that involves the study of the silica bodies on OPEFB fiber and their contributions to the mechanical behavior of OPEFB fiber. Only recently, there are studies on micromechanics of oil palm fiber performed by a research group in UPM. Hanipah et al. (2016) and Xiang et al. (2015) utilized numerical approach of micromechanics of oil palm fiber and revealed the viscoelastic properties as evident from stress relaxation curves. Likewise, in another study conducted by Wang et al. (2014), finite element analyses study of royal palm at tissue level was performed where cellular structure of the palm was reconstructed with polynomial area weighted tessellation model in order to simulate the vascular tissue behavior and area ratio and parameter ratio of adjacent cells were calculated and compared.

Deeper understanding and investigations of oil palm fiber cellular and tissue structure could be performed with both numerical and analytical micromechanics approaches. The behavior, properties, response and failure mechanisms could be understood and explored. This is essential especially if the utilization of oil palm fiber in composites or any other purposes are required if one aims to utilize it in its most possible way.

1.3 Problem statements

Micromechanics study of silica bodies on OPEFB fiber and its contribution on the fiber integrity has yet been studied, where this information could provide the fundamental background on its behavior, properties as well as other mechanisms. Up to date, there are no detailed models available that discuss the mechanics of the oil palm fiber specifically with silica bodies. Hence, development of model through numerical and analytical methods of micromechanics is essential to predict the behavior of the fiber by providing an in depth understanding of the effects of silica bodies physiologies and structures towards the fiber strength and therefore, may contribute to the decision of the degree of pretreatment for silica bodies removal needed especially in industries with natural fibers utilization like biocomposites and fiber bioconversion process. By knowing this information, it would minimize the energy, time and money spent on the silica bodies removal treatment process. This studies also provides deeper understanding of silica bodies and role of OPEFB fiber as a bioresources material, and the models could also be used for other natural fiber modelling as well.

Unnecessary compound and by-products formation from lignin degradation could be eliminated through biological treatments as it is substrate specific and involve no harsh chemicals. However, involvement of biological treatments usually lead to delayed response and achievements. Therefore, microbes with high production of lignin-degrading enzymes is preferred for lignin degradation to occur effectively. Removal of lignin is important as such it intensely being incorporated into emergent lignocellulose biorefineries. Additionally, the mechanism of structural degradation between fungi and OPEFB during lignin degradation process is an intriguing subject of research and up to this date, there are no comprehensive studies conducted and discussed in the literatures. The OPEFB biodegradation studies is important not only to solve the solid waste disposal in Malaysia, but also to prepare the fiber for holocellulose utilization which would later on would be greatly useful for numerous valuable products generations such as biosugars, carboxymethyl cellulose etc.

To fill in the gaps mentioned above, a study on micromechanics study on silica bodies on OPEFB fiber was conducted and the degradation of lignin by fungi was evaluated through structural and physico-chemical data analyses. The objectives of this study therefore are:

- 1. To determine the effect of silica bodies on OPEFB fiber integrity through solid mechanics approach.
- 2. To optimize the environmental conditions for laccase production from local isolated white rot fungi, *P. sanguineus*.
- 3. To study the relationship between structural and physico-chemical behavior of OPEFB after degradation process by *P.sanguineus*.

1.4 Scope of research and thesis structure

This study is principally concerned about the micromechanics study of OPEFB and its relationship with biodegradation of OPEFB fiber by local isolated white rot fungi. During this research, and in depth study has been performed in studying the feedstock, OPEFB fiber in terms of the micromechanics behavior and modeling of the silica bodies onto the surfaces of the fiber. A 2D model was adopted to explore the effect of silica bodies' arrangement and spiked geometry of silica bodies. 3D models were later developed in order to further investigate with the contribution of silica bodies towards fiber integrity. On the other hand, white rot fungi was isolated and the ability to perform the degradation on OPEFB was evaluated. The performance of the fungi was evaluated in both submerged and solid state fermentation. Based on the lignocellulosic content and phenol content, the fungi show some potential in depolymerizing the lignocellulosic content in OPEFB. Finally, the relationship between structural and physico-chemical behavior of degraded OPEFB fibers were reported and discussed in detail.

In this thesis, there are 5 chapters will be included in which each chapter will explain independent topics. In the Chapter 1, a brief introduction of the overall research was written together with objectives of the study and the scope of the research. In the Chapter 2, extensive literature review was written covering

current available knowledge on the micromechanics of natural fiber OPEFB, fermentation strategies and biodegradation of lignin process through fungal degradation. In Chapter 3, the first objective of the study was elaborated in which to study the micromechanics modeling of the silica bodies on the OPEFB fibers where constitutive material behavior (stress-strain), 2D and 3D modelling were performed in order to investigate the oil palm fiber behavior. In the Chapter 4, the second objective of the research was explained in which to explore the potential of the local isolate white rotting fungi for OPEFB biodegradation in solid state fermentation. In the Chapter 5, the third and final objective was well intricate in which to study the utilization of the micro computed tomography in the microstructure behavior of the degraded OPEFB fibers and the relationship between structural and physico-chemical behavior of degraded OPEFB was discussed. In the final chapter, chapter 6, final conclusions and some of recommendations were mentioned. Appendix and references used in this entire study was listed at the back of the thesis. The overview of the experimental design reported in the thesis is provided in Figure 1.1.

OPEFB FIBER PREPARATION

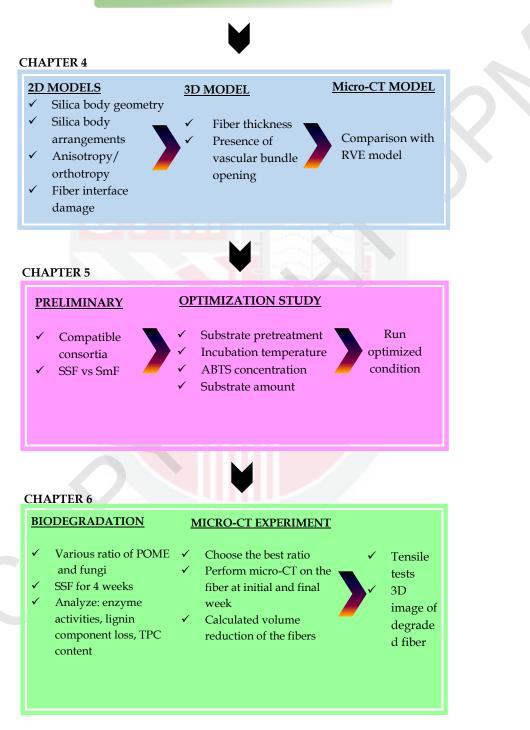


Figure 1.1: Overview of the overall experimental design

REFERENCES

- Abaqus (2015) User Manual version 6.9, Hibbitt Karlsson and Sorensen, Providence, RI.
- Abdel-Hamid, A. M., Solbiati, J. O., & Cann, I. K. O. (2013). Insights into lignin degradation and its potential industrial applications. *Advances in Applied Microbiology*, 82, 1–28.
- Abd-Elsalam, H. and El-Hanafy, A. (2009). Lignin biodegradation with ligninolytic bacterial strain and comparison of *Bacillus subtilis* and *Bacillus* sp. isolated from Egyptian soil. *American-Eurasian Journal of Agricultural and Environmental Sciences*, 5 (1), 39-44.
- Abdul Khalil, H. P. S., Siti Alwani, M., Ridzuan, R., Kamarudin, H., and Khairul, A. (2008). Chemical composition, morphological characteristics, and cell wall structure of malaysian oil palm fibers. *Polymer-Plastics Technology and Engineering*, 47 (3), 273–280.
- Abdullah, N., and Sulaiman, F. (2013). The oil palm wastes in Malaysia. *Biomass Now – Sustainable Growth and Use*, 75–100.
- Abu Bakar, N. K., Zanirun, Z., Abd-Aziz, S., Ghazali, F. M., Hassan, M. A. (2012). Production of fermentable sugars from oil palm empty fruit bunch using crude cellulase cocktails with *Trichoderma asperellum* UPM1 and *Aspergillus fumigatus* UPM2 for bioethanol production. *BioResources*, 7 (3), 3627–3639.
- Adejoye, O. D., & Fasidi, I. O. (2009). Biodegradation of agro-wastes by some Nigerian white-rot fungi. *Bioresources*, 4 (2), 816–824.
- Adler, D. C., and Buehler, M. J. (2013). Mesoscale mechanics of wood cell walls under axial strain. *Soft Matter*, 9 (29), 7138–7144.
- Akhtar, J., Idris, A., Teo, C. L., Lai, L. W., Hassan, N., & Khan, M. I. (2014). Comparison of delignification of oil palm empty fruit bunch (efb) by microwave assisted alkali/acid pretreatment and conventional pretreatment method. *International Journal of Advances in Chemical Engineering and Biological Sciences*, 1 (2), 155–157.
- Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W., & Lipman, D. J. (1997). Gapped BLAST and PSI-BLAST: A new generation of protein database search programs. *Nucleic Acids Research*, 25 (17), 3389– 3402.
- AOAC (1997). Official methods of analysis of AOAC International. Method 952.03, 16th ed., ed. Cunniff, P., Maryland: AOAC International.

- Arganda-Carreras, I., Kaynig, V., Rueden, C., Schindelin, J., Cardona, A., Seung, H. S. (2016). Trainable_Segmentation: release v3.1.2 [Data set]. Zenodo. doi:10.5281/zenodo.59290
- Arun, A., and Eyini, M. (2011). Comparative studies on lignin and polycyclic aromatic hydrocarbons degradation by basidiomycetes fungi. *Bioresource Technology*, 102 (17), 8063–8070.
- Asgher, M., Bashir, F., & Iqbal, H. M. N. (2014). A comprehensive ligninolytic pre-treatment approach from lignocellulose green biotechnology to produce bio-ethanol. *Chemical Engineering Research and Design*, 92 (8), 1571– 1578.
- Asgher, M., Nasir Iqbal, H. M., & Asad, M. J. (2012). Kinetic characterization of purified laccase produced from. *BioResources*, 7 (1), 1171–1188.
- Ashik, K. P. and Sharma, R. S. (2015). A review on mechanical properties of natural fiber reinforced hybrid polymer composites. *Journal of Minerals and Materials Characterization and Engineering*, *3*, 420-426.
- Askarinejad, S., and Rahbar, N., (2015) Toughening mechanisms in bioinspired multilayered materials. *Journal of Royal Society of Interface*, 12 (102), 20140855.
- Awalludin, M. F., Sulaiman, O., Hashim, R., & Nadhari, W. N. A. W. (2015). An overview of the oil palm industry in Malaysia and its waste utilization through thermochemical conversion, specifically via liquefaction. *Renewable and Sustainable Energy Reviews*, 50, 1469–1484.
- Aziz, M. M. A., Hamad, A. W., Maleka, A. M., & Jakarni, F. M. (2015). Effect of viscoelastic behavior of cellulose oil palm fiber (COPF) modified 60-70 asphalt binder for deterioration for roads and highways. *Jurnal Teknologi*, 75 (11), 17-23.
- Baharuddin, A. S., Hock, L. S., Yusof, M. Z. M., Abdul, N. A. R., Shah, U. K. M., Hassan, M. A., Wakisaka, M., Sakai, K., Shirai, Y. (2010). The effect of palm oil mill effluent (POME) anaerobic sludge from 500 m³ of closed anaerobic methane digested tank on pressed-shredded empty fruit bunch (EFB) composting process. *African Journal of Biotechnology*, 9 (16), 2427–2436.
- Baharuddin, A. S., Sulaiman, A., Kim, D. H., Mokhtar, M. N., Hassan, M. A., Wakisaka, M., Shirai, Y., Nishida, H. (2013). Selective component degradation of oil palm empty fruit bunches (OPEFB) using high-pressure steam. *Biomass and Bioenergy*, 55, 268–275.
- Baharuddin, A. S., Wakisaka, M., Shirai, Y., & Abd-Aziz, S. (2009). Cocomposting of empty fruit bunches and partially treated palm oil mill

efluents in pilot scale. International Journal of Agricultural Research, 4, 69-78.

- Begley, M. R., Philips, N. R., Compton, B. G., Wilbrink, D. V., Ritchie, R. O., Utz, M., (2012). Micromechanical models to guide the development of synthetic 'brick and mortar' composites. *Journal of Mechanical Physics and Solids*, 60 (8), 1545–1560.
- Benvenuto, M. L., Fernández Honaine, M., Osterrieth, M. L., & Morel, E. (2015). Differentiation of globular phytoliths in Arecaceae and other monocotyledons: Morphological description for paleobotanical application. *Turkish Journal of Botany*, 39 (2), 341–353.
- Bhargav, S., Panda, B. P., Ali, M., and Javed, S. (2008). Solid-state Fermentation : An Overview. *Chemical Biochemical Engineering*, 22 (1), 49–70.
- Bolhassan, M. H. (2012). Diversity of Polyporales and the application of *Ganoderma australe* (fr.) pat. in biopulping of empty fruit bunches of *Elaeis guineensis*. PhD Thesis, Universiti Malaya, Malaysia.
- Boonyuen, N., Manoch, L., Luangsa-ard, J. J., Piasai, O., Chamswarng, C., Chuaseeharonnachai, C., Ueapattanakit, J., Arnthong, J., Sri-indrasutdhi, V. (2014). Decomposition of sugarcane bagasse with lignocellulose-derived thermotolerant and thermoresistant Penicillia and Aspergilli. *International Biodeterioration and Biodegradation*, 92, 86–100.
- Bourbonnais, R. and Paice, M. G. (1990). Oxidation of non phenolic substrates: An expanded role of laccase in lignin biodegradation. *FEBS Letters*, 267(1), 99-102.
- Bourbonnais, R., Paice, M. G., Reid, I. D., Lanthier, P., & Yaguchi, M. (1995). Lignin oxidation by laccase isozymes from *Trametes versicolor* and role of the mediator in kraft lignin depolymerization. *Applied and Environmental Microbiology*, 61 (5), 1876–1880.
- Brandt, A., Grasvik, J., Hallet, J. P., Welton, T. (2013). Deconstruction of lignocellulosic biomass with ionic liquids. *Green Chemistry*, 15, 550-583.
- Brijwani, K., Rigdon, A., and Vadlani, P. V. (2010). Fungal laccases: production, function, and applications in food processing. *Enzyme Research*, 10, 149748, 1–10.
- Brodeur, G., Yau, E., Badal, K., Collier, J., Ramachandran, K. B., Ramakrishnan, S. (2011). Chemical and physicochemical pretreatment of lignocellulosic biomass: A review. *Enzyme Research*, e787532.
- Brunecky, R., Alahuta, M., Xu, Q., Donohoe, B. S., Crowley, M. F., Kataeva, I. A., Yang, S. J., Resch, M. G., Adams, M. W. W., Lunin, V. V., Himmel, M. E.,

Bomble, Y. J. (2013). Revealing nature's cellulase diversity: The digestion mechanism of *Caldicellulosiruptor bescii* CelA. *Science* 342, 1513–1516.

- Bugg, T. D. H., Ahmad, M., Hardiman, E. M., Rahmanpour, R. (2011). Pathways for degradation of lignin in bacteria and fungi. *Natural Product Reports*, 28 (12), 1883–96.
- Bugg, T. D., and Rahmanpour, R. (2015). Enzymatic conversion of lignin into renewable chemicals. *Current Opinion in Chemical Biology*, 29, 10–17.
- Burgert, I. (2006). Exploring the micromechanical design of plant cell walls. *American Journal of Botany*, 93 (10), 1391–1401.
- Burgert, I., and Dunlop, J. W. C. (2011). Micromechanics of cell walls. *Mechanical Integrity of Plant Cells and Plants*, 9,27-52.
- Burgert, I., and Fratzl, P., (2009). Actuation systems in plants as prototypes for bioinspired devices. *Philosophical Transactions of Royal Society A*, 367, 1541– 1557
- Burgert, I., and Keplinger, T. (2013). Plant micro- and nanomechanics: experimental techniques for plant cell-wall analysis. *Journal of Experimental Botany*, 64 (15), 4635-4649.
- Campilho, R. D. S. G., Moura, D. C., Gonçalves, D. J. S., Da Silva, J. F. M. G., Banea, M. D., Da Silva, L. F. M. (2013). Fracture toughness determination of adhesive and co-cured joints in natural fibre composites. *Composites Part B: Engineering*, 50, 120–126.
- Castilho, L. R., Polato, C. M. S., Baruque, E. A., Sant'Anna, G. L., & Freire, D. M. G. (2000). Economic analysis of lipase production by *Penicillium restrictum* in solid-state and submerged fermentations. *Biochemical Engineering Journal*, 4 (3), 239–247.
- Catherine, H., Penninckx, M., and Frédéric, D. (2016). Product formation from phenolic compounds removal by laccases: A review. *Environmental Technology and Innovation*, *5*, 250–266.
- Cave, I. D., and Hutt, L. (1968). The anisotropic elasticity of plant cell wall. *Wood Science Technology*, 2, 268–278.
- Chaturvedi, V., & Verma, P. (2013). An overview of key pretreatment processes employed for bioconversion of lignocellulosic biomass into biofuels and value added products. *3 Biotech*, *3* (5), 415–431.
- Chen, M., Zeng, G., Tan, Z., Jiang, M., Li, H., Liu, L., Zhu, Y., Yu, Z., Liu, Y., Xie, G. (2011). Understanding lignin-degrading reactions of ligninolytic

enzymes: Binding affinity and interactional profile. *PLoS ONE*, 6 (9), e25647.

- Chenthamarakshan, A., Parambayil, N., Miziriya, N., Soumya, P. S., Kiran Lakshi, M. S., Ramgopal, A., Dileep, A., Nambisan, P. (2017). Optimization of laccase production from *Marasmiellus palmivorus* LA1 by Taguchi method of design of experiments. *BMC Biotechnology*, 17(12), 1-10.
- Chhaya, U., and Gupte, A. (2013). Effect of different cultivation conditions and inducers on the production of laccase by the litter-dwelling fungal isolate *Fusarium incarnatum* LD-3 under solid substrate fermentation. *Annals of Microbiology*, 63 (1), 215–223.
- Chi, Y., Hatakka, A., and Maijala, P. (2007). Can co-culturing of two white-rot fungi increase lignin degradation and the production of lignin-degrading enzymes? *International Biodeterioration and Biodegradation*, 59 (1), 32–39.
- Chin, S. X., Chia, C. H., Zakaria, S., Fang, Z., Ahmad, S. (2015). Ball milling pretreatment and diluted acid hydrolysis of oil palm empty fruit bunch (EFB) fibres for the production of levulinic acid. *Journal of the Taiwan Institute of Chemical Engineers*, *52*, 85–92.
- Choi, W. I., Park, J. Y., Lee, J. P., Oh, Y. K., Park, Y. C., Kim, J. S., Park, J. M., Kim, C. H., Lee, J. S. (2013). Optimization of NaOH-catalyzed steam pretreatment of empty fruit bunch. *Biotechnology for Biofuels*, 29, 170-178.
- Chow, M. J., H'ng, P. S., Chin, K. L., Chai, E. W., Md Tahir, P., Lee, S. H., Lum, W. C., Chuah, L., Maminski, M. (2015). Empty Fruit Bunches in the Race for Energy, Biochemical and Material Industry in Agricultural Biomass Based Potential Materials, ed. Hakeem K. R., Springer International Publishing, Switzerland, pp375–389.
- Christopher, L. P., Yao, B., and Ji, Y. (2014) Lignin biodegradation with laccasemediator systems. *Fronties in Energy Research*, 2(12), 1-13.
- Chundawat, S. P., Beckham, G. T., Himmel, M. E., & Dale, B. E. (2011). Deconstruction of lignocellulosic biomass to fuels and chemicals. *Annual Review Chemical Biomolecular Engineering*, 2, 121–145.
- Coconi-Linares, N., Magana-Ortiz, D., Gusman-Ortiz, D. A., Fernandez, F., Loske, A. M., Gomez-Lim, M. A. (2014). High yield production of manganese peroxidase, lignin peroxidase and versatile peroxidase in *Phanerochaete chrysosporium. Applied Microbiology and Biotechnology*, 98, 9283-9294.
- Couto, S. R., Gundín, M., Lorenzo, M., Sanromán, M. A. (2002). Screening of supports and inducers for laccase production by *Trametes versicolor* in semi-

solid-state conditions. Process Biochemistry, 38 (2), 249-255.

- Couto, S. R. and Sanroman, M. A. (2005). Application of solid-state fermentation to ligninolytic enzyme production. *Biochemical Engineering*, 22, 211-219.
- Currie, H. A., and Perry, C. C. (2007). Silica in plants: Biological, biochemical and chemical studies. *Annals of Botany*, 100, 1383–1389.
- d'Almeida, J. R. M., Aquino, R. C. M. P., & Monteiro, S. N. (2006). Tensile mechanical properties, morphological aspects and chemical characterization of piassava (Attalea funifera) fibers. *Composites Part A: Applied Science and Manufacturing*, *37* (9), 1473–1479.
- da Costa Sousa, L., Chundawat, S. P., Balan, V., Dale, B. E. (2009). "Cradle-tograve" assessment of existing lignocellulose pretreatment technologies. *Current Opinion in Biotechnology*, 20 (3), 339–347.
- Dashtban, M. (2009). Fungal Bioconversion of Lignocellulosic Residues; Opportunities and Perspectives. *International Journal of Biological Sciences*, 5 (6), 578–595.
- Dekker, R. F. H., Barbosa, A. M., Giese, E. C., Godoy, S. D. S., Covizzi, L. G. (2007). Influence of nutrients on enhancing laccase production by *Botryosphaeria rhodina* MAMB-05. *International Microbiology*, 10, 177-185.
- Demir, A., Aytar, P., Gedikli, S., Çabuk, A., Arisoy, M. (2011). Laccase production with submerged and solid state fermentation : Benefit and cost analysis. *Journal of Biology and Chemistry*, 39 (3), 305–313.
- Dias, A. A., Freitas, G. S., Marques, G. S. M., Sampaio, A., Fraga, I. S., Rodrigues, M. A. M., Evtuguin, D. V., Bezerra, R. M. F. (2010). Enzymatic saccharification of biologically pre-treated wheat straw with white-rot fungi. *Bioresource Technology*, 101 (15), 6045–6050.
- Díaz, R., Alonso, S., Sánchez, C., Tomasini, A., Bibbins-Martínez, M., Díaz-Godínez, G. (2011). Characterization of the growth and laccase activity of strains of *Pleurotus ostreatus* in submerged fermentation. *BioResources*, 6 (1), 282–290.
- Dinis, M. J., Bezerra, R. M. F., Nunes, F., Dias, A. A., Guedes, C. V., Ferreira, L. M. M., Cone, J. W., Marques, G. S. M., Barros, A. R. N., Rodrigues, M. A. M. (2009). Modification of wheat straw lignin by solid state fermentation with white-rot fungi. *Bioresource Technology*, 100 (20), 4829–4835.
- Dong, X. Q., Yang, J. S., Zhu, N., Wang, E. T., & Yuan, H. L. (2013). Sugarcane bagasse degradation and characterization of three white-rot fungi. *Bioresource Technology*, 131, 443–451.

- Dong, Y. C., Wang, W., Hu, Z., Fu, M. L., Chen, Q. (2012). The synergistic effect on production of lignin-modifying enzymes through submerged cocultivation of *Phlebia radiata*, *Dichomitus squalens* and *Ceriporiopsis subvermispora* using agricultural residues. *Bioprocess and Biosystems Engineering*, 35 (5), 751–760.
- Dwivedi, P., Vivekanand, V., Pareek, N., Sharma, A., Singh, R. P. (2011). Cocultivation of mutant *Penicillium oxalicum* SAU E-3.510 and *Pleurotus ostreatus* for simultaneous biosynthesis of xylanase and laccase under solidstate fermentation. *New Biotechnology*, *28* (6), 616–626.
- Ehrman, T. (1996). Determination of acid soluble lignin in biomass. NREL CAT Task Laboratory Analytical Procedure #004.
- Elices, M., Guinea, G. V., Gomez, J., Planas, J. (2002). The cohesive zone model: Advantages, limitations and challenges. *Engineering Fracture Mechanics*, 69 (2), 137–163.
- Elisashvili, V., Penninckx, M., Kachlishvili, E., Tsiklauri, N., Metreveli, E., Kharziani, T., & Kvesitadze, G. (2008). *Lentinus edodes* and *Pleurotus species* lignocellulolytic enzymes activity in submerged and solid-state fermentation of lignocellulosic wastes of different composition. *Bioresource Technology*, 99 (3), 457–462.
- Ellaiah, P., Adinarayana, K., Bhavani, Y., Padmaja, P., Srinivasulu, B. (2002). Optimization of process parameters for glucoamylase production under solid state fermentation by a newly isolated *Aspergillus* species. *Process Biochemistry*, 38 (4), 615–620.
- Eshelby, J. D. (1957). The determination of the elastic field of and ellipsoidal inclusion and related problems. *Proceedings of The Royal Society A*, 241:376–396.
- Fang, J. Y., and Ma, X. L. (2006). In vitro simulation studies of silica deposition induced by lignin from rice. *Journal of Zhejiang University. Science. B*, 7 (4), 267-271.
- Ferhan, M., Leao, A. L., Itamar, S. de M., M, S. (2012). Ligninases production and partial purification of mnp from brazilian fungal isolate in submerged fermentation. *Fermentation Technology*, *1* (5), 1–7.
- Fisher, A. B., and Fong, S. S. (2014). Lignin biodegradation and industrial implications. *AIMS Bioengineering*, 1 (2), 92–112.
- Flores, E. I. S. and Friswell, M. I. (2012). Multi-scale finite element model for a new material inspired by the mechanics and structure of wood cell-walls.

Journal of Mechanical Physics and Solids, 60 (7), 943-951.

- Fonseca, M. I., Shimizu, E., Zapta, P. D., Villalba, L. L. (2010). Copper inducing effect on laccase production of white rot fungi native from Misiones (Argentina). *Enzyme and Microbial Technology*, 46, 534-539.
- Fujian, X., Hongzhang, C., & Zuohu, L. (2001). Solid-state production of lignin peroxidase (LiP) and manganese peroxidase (MnP) by *Phanerochaete chrysosporium* using steam-exploded straw as substrate. *Bioresource Technology*, 80 (2), 149–151.
- Fu, K., Fu, S., Zhan, H., Zhou, P., Liu, M., Liu, H. (2013). A newly isolated wood rot fungus for laccase production in submerged cultures. *BioResources*, 8 (1), 1385-1397.
- Gamauf, C., Metz, B., and Seiboth, B. (2007). Degradation of Plant Cell Wall Polymers by Fungi. *Environmental and Microbial Relationships - The Mycota IV*, 325–340.
- Gao, X., Zou, C., Wang, L., & Zhang, F. (2006). Silicon decreases transpiration rate and conductance from stomata of maize plants. *Journal of Plant Nutrition*, 29 (9), 1637–1647.
- Garcia-Nunez, J. A., Ramirez-Contreras, N. E., Rodriguez, D. T., Silva-Lora, E., Frear, C. S., Stockle, C., Garcia-Perez, M. (2016). Evolution of palm oil mills into bio-refineries: Literature review on current and potential uses of residual biomass and effluents. *Resources, Conservation and Recycling*, 110, 99–114.
- Gassara, F., Brar, S. K., Tyagi, R. D., Verma, M., Surampalli, R. Y. (2010). Screening of agro-industrial wastes to produce ligninolytic enzymes by *Phanerochaete chrysosporium. Biochemical Engineering Journal*, 49 (3), 388–394.
- Giardina, P., Faraco, V., Pezzella, C., Piscitelli, A., Vanhulle, S., & Sannia, G. (2010). Laccases: A never-ending story. *Cellular and Molecular Life Sciences*, 67 (3), 369–385.
- Gierlinger, N., Sapei, L., and Paris, O. (2008). Insights into the chemical composition of *Equisetun hyemale* by high resolution Raman imaging. *Planta*, 227, 969-980.
- Goh, C. S., Tan, K. T., Lee, K. T., & Bhatia, S. (2010). Bio-ethanol from lignocellulose: Status, perspectives and challenges in Malaysia. *Bioresource Technology*, *101* (13), 4834–4841.
- Gomaa, O. M., and Momtaz, O. A. (2015) Copper induction and differential expression of laccase in Aspergillus flavus. *Brazilian Journal of Microbiology*.

46 (1), 285-292.

- Gomes, E., Aguiar, A. P., Carvalho, C. C., Bonfá, M. R. B., Da Silva, R., & Boscolo, M. (2009). Ligninases production by basidiomycetes strains on lignocellulosic agricultural residues and their application in the decolorization of synthetic dyes. *Brazilian Journal of Microbiology*, 40 (1), 31– 39.
- Govumoni, S. P., Koti, S., Kothagouni, S. Y., Venkateshwar, S., & Linga, V. R. (2013). Evaluation of pretreatment methods for enzymatic saccharification of wheat straw for bioethanol production. *Carbohydrate Polymers*, 91 (2), 646–650.
- Guerriero, G., Hausman, J. F., Strauss, J., Ertan, H., & Siddiqui, K. S. (2016). Lignocellulosic biomass: Biosynthesis, degradation, and industrial utilization. *Engineering in Life Sciences*, *16*(1), 1–16.
- Gunawan, F. E., Homma, H., Brodjonegoro, S. S., Hudin, A. B., Zainuddin, A. (2009). Mechanical properties of oil palm empty fruit bunch fiber. *Journal* of Solid Mechanical and Material Engineering, 3(7), 943-951.
- Gupte, A., Gupte, S., and Patel, H. (2007). Ligninolytic enzyme production under solid-state fermentation by white rot fungi. *Journal of Scientific and Industrial Research*, 66 (8), 611–614.
- Haghighi Mood, S., Hossein Golfeshan, A., Tabatabaei, M., Salehi Jouzani, G., Najafi, G. H., Gholami, M., Ardjmand, M. (2013). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renewable and Sustainable Energy Reviews*, 27, 77–93.
- Hamisan, A. F., Abd-Aziz, S., Kamaruddin, K., Shah, U. K. M., Shahab, N., Hassan, M. A. (2009). Delignification of oil palm empty fruit bunch using chemical and microbial pretreatment methods. *International Journal of Agriculture Research*, 4, 250-256.
- Hamzah, F., Idris, A., and Shuan, T. K. (2011). Preliminary study on enzymatic hydrolysis of treated oil palm (*Elaeis*) empty fruit bunches fibre (EFB) by using combination of cellulase and β ,1-4 glucosidase. *Biomass and Bioenergy*, 35 (3), 1055–1059.
- Han, M., Kim, Y., Kim, Y., Chung, B., Choi, G. W. (2011). Bioethanol production from optimized pretreatment of cassava stem. *Korean Journal of Chemical Engineering*, 28 (1), 119–125.
- Hanipah, S. H., Mohammed, M. A. P., and Baharuddin, A. S. (2016). Non-linear mechanical behaviour and bio-composite modelling of oil palm mesocarp fibres. *Composite Interfaces*, 23 (1), 37–49.

- Hanipah, S. H., Yu Xiang, L., Mohammed, M. A. P., Samsu Baharuddin, A. (2017). Study of non-linear mechanical behavior of oil palm mesocarp fibers. *Journal of Natural Fibers*, 14 (2), 153–165.
- Hassan, A., Salema, A. A., Ani, F. N., & Bakar, A. A. (2010). A review on oil palm empty fruit bunch fiber-reinforced polymer composite materials. *Polymer Composites*, 31 (12), 2079–2101.
- Hatakka, A., and Hammel, K. (2010). Fungal biodegradation of lignocelluloses. *The Mycota*, 319–340.
- Hattaka, A. (1994). Lignin-modifying enzymes from selected white-rot fungi: Production and role from in lignin degradation. *FEMS Microbiology Reviews*, 13 (3), 125–135.
- Hayot, C. M., Farousech, E., Goel, A., Avramova, Z., Turner, J. A. (2012). Viscoelastic properties of cell walls of single living plant cells determined by dynamic nanoindentation. *Journal of Experimental Botany*, 53 (7), 2525-2540.
- He, C., Wang, L., Liu, J., Liu, X., Li, X., Ma, J., Lin, Y., Xu, F. (2015). Evidence for 'silicone' within the cell walls of suspension-cultured rice cells. *New Phytologist*, 200, 700-709.
- Hendriks, A. T. W. M., and Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*, 100 (1), 10–18.
- Hernandez, C., Da Silva, A.M., Ziarelli, F., Perraud-Gaime, I., Gutierrez-Rivera, B., Garcia-Perez, J. A., Alarcon, E. (2016). Laccase induction by synthetic dyes in *Pycnoporus sanguineus* and their possible use for sugar cane bagasse delignification. Applied Microbiology Biotechnology, 101(3), 1189-1201.
- Hill, R. (1952). The elastic behavior of crystalline aggregate: Proceedings of the Physical Society, London, Section A, *65* (5), 349-354.
- Hill, R. (1965). A self-consistent mechanics of composite materials. *Journal of Mechanical and Physics of Solids*, 13, 213-222.
- Hildén, K. S., Mäkelä, M. R., Hakala, T. K., Hatakka, A., & Lundell, T. (2006). Expression on wood, molecular cloning and characterization of three lignin peroxidase (LiP) encoding genes of the white rot fungus Phlebia radiata. *Current Genetics*, 49 (2), 97–105.
- Ho, H. L. (2014). Biodiversity, Bioprospecting and development effects of medium formulation and culture conditions on microbial xylanase production using agricultural extracts in submerged fermentation (SmF) and solid state fermentation (SSF): A review. *Journal of Biodiversity*,

Bioprospecting and Development, 1 (3), 1–13.

- Honda, Y., Watanabe, T., & Kuwahara, M. (2002). Pre-treatment of oil palm empty fruit bunch by white rot fungi for enzymatic saccharification. *Wood Research*, *89*, 19–20.
- Hurtado, J. A., Lapczyk, I., Govindarajan, S. M. (2013). Parallel rheological framework to model non-linear viscoelasticity, permanent set, and Mullins effect in elastomers. *Constitutive Models for Rubber VIII*, Edited by Asier Alonso CRC Press, pp 95–100.
- Iberahim, N. Z., Md-Jahim, J., Harun, S., Mohd-Nor, M. T., Hassan, O. (2013). Sodium hydroxide pretreatment and enzymatic hydrolysis of oil palm mesocarp fiber. *International Journal of Chemical Engineering Applications*, 4(3), 101–105.
- Ibrahim, S. M., Badri, K. H., and Hassan, O. (2012). A study on glycerolysis of oil palm empty fruit bunch fiber. *Sains Malaysiana*, 41(12), 1579–1585.
- Irshad, M. and Asgher, M. (2011). Purification and characterization of LiP produced by schyzophyllum commune IBL-06 using banana stalk in solid state cultures. *African Journal of Biotechnology*, 10 (79), 18234–18242.
- Isroi, Ishola, M. M., Millati, R., Syamsiah, S., Cahyanto, M. N., Niklasson, C., Taherzadeh, M. J. (2012). Structural changes of oil palm empty fruit bunch (OPEFB) after fungal and phosphoric acid pretreatment. *Molecules*, 17 (12), 14995–15012.
- Isroi, Millati, R., Syamsiah, S., Niklasson, C., Cahyanto, M. N., Lundquist, K., Taherzadeh, M. J. (2011). Biological pretreatment of lignocelluloses with white-rot fungi and its applications: A review. *BioResources*, 6 (4), 5224– 5259.
- Jeon, H., Kang, K. E., Jeong, J. S., Gong, G., Choi, J. W., Abimanyu, H., Ahn, B. S., Suh, D. J., Choi, G. W. (2014). Production of anhydrous ethanol using oil palm empty fruit bunch in a pilot plant. *Biomass and Bioenergy*, 67, 99–107.
- Jönsson, L. J. and Martín, C. (2016). Pretreatment of lignocellulose: Formation of inhibitory by-products and strategies for minimizing their effects. *Bioresource Technology*, 199, 103–112.
- Kachlishvili, E., Penninckx, M. J., Tsiklauri, N., Elisashvili, V. (2006). Effect of nitrogen source on lignocellulolytic enzyme production by white-rot basidiomycetes under solid-state cultivation. World Journal of Microbiology and Biotechnology, 22 (4), 391–397.

Kadam, A. A., Telke, A. A., Jagtap, S. S., Govindwar, S. P. (2011). Decolorization

of adsorbed textile dyes by developed consortium of *Pseudomonas* sp. SUK1 and *Aspergillus ochraceus* NCIM-1146 under solid state fermentation. *Journal of Hazardous Materials*, *189* (1–2), 486–494.

- Kalia, S., Kaith, B. S. and Kaur, I. (2007). A short review on rubber / clay nanocomposites with emphasis on mechanical properties. *Polymer Engineering and Sciences*, 49 (7), 12553–1272.
- Kamal Bahrin, E. (2012) Optimization and characterization of oil palm empty fruit bunch fermentation for cellulase producyion by *Botrysphaeria rhodina* UPM3, PhD Thesis, Universiti Putra Malaysia, Malaysia.
- Kamal Bahrin, E., Baharuddin, A. S., Ibrahim, M. F., Abdul Razak, M. N., Sulaiman, A., Abd-Aziz, S., Hassan, M. A., Shirai, Y., Nishida, H. (2012). Physicochemical property changes and enzymatic hydrolysis enhancement of oil palm empty fruit bunches treated with superheated steam. *BioResources*, 7 (2), 1784–1801.
- Kamcharoen, A., Champreda, V., Eurwilaichitr, L., and Boonsawang, P. (2014). Screening and optimization of parameters affecting fungal pretreatment of oil palm empty fruit bunch (EFB) by experimental design. *International Journal of Energy and Environmental Engineering*, 5 (4), 303–312.
- Kanmani, P., Karuppasamy, P., Pothiraj, C., Arul, V. (2009). Studies on lignocellulose biodegradation of coir waste in solid state fermentation using *Phanerocheate chrysosporium* and *Rhizopus stolonifer*. *African Journal of Biotechnology*, 8 (24), 6880–6887.
- Karp, S. G., Faraco, V., Amore, A., Birolo, L., Giangrande, C., Soccol, V. T., Pandey, A., Soccol, C. R. (2012). Characterization of laccase isoforms produced by *Pleurotus ostreatus* in solid state fermentation of sugarcane bagasse. *Bioresource Technology*, 114, 735–739.
- Katinonkul, W., Lee, J. S., Ha, S. H., & Park, J. Y. (2012). Enhancement of enzymatic digestibility of oil palm empty fruit bunch by ionic-liquid pretreatment. *Energy*, 47 (1), 11–16.
- Kausar, H., Sariah, M., Mohd Saud, H., Zahangir Alam, M., Razi Ismail, M. (2010). Development of compatible lignocellulolytic fungal consortium for rapid composting of rice straw. *International Biodeterioration and Biodegradation*, 64 (7), 594–600.
- Keiluweit, M., Nico, P., Harmon, M. E., Mao, J., Pett-Ridge, J., Kleber, M. (2015). Long-term litter decomposition controlled by manganese redox cycling. *Proceedings of the National Academy of Sciences*, 112 (38), E5253–E5260.

Kennedy, J. M. and Moeller, H. H. (1990). Thermal and Mechanical Behavior of

Metal Matrix and Ceramic Matrix Composites, *American Society for Testing and Materials*, Philadelphia, USA. pp 136-151.

- Koyani, R. D., and Rajput, K. S. (2015). Bioprocessing and biotechniques solid state fermentation: comprehensive tool for utilization of lignocellulosic through biotechnology, *Journal of Bioprocessing and Biotechniques*, 5 (10), 1-15.
- Kratky, L. and Jirout, T. (2011). Biomass size reduction machines for enhancing biogas production. *Chemical Engineering and Technology*, 34 (3), 391–399.
- Krishna, C. (2005). Solid-state fermentation systems An overview. *Criticals Reviews in Biotechnology*, 25, 1-30.
- Kristiani, A., Effendi, N., Aristiawan, Y., Aulia, F., Sudiyani, Y. (2015). Effect of combining chemical and irradiation pretreatment process to characteristic of oil palm's empty fruit bunches as raw material for second generation bioethanol. *Energy Procedia*, 68, 195–204.
- Kuhar, F., Castiglia, V., and Levin, L. (2015). Enhancement of laccase production and malachite green decolorization by co-culturing *Ganoderma lucidum* and *Trametes versicolor* in solid-state fermentation. *International Biodeterioration and Biodegradation*, 104, 238-243.
- Kumar, P., Barrett, D. M., Delwiche, M. J., Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial and Engineering Chemistry Research*, 48(8), 3713–3729.
- Kumar, V. V., Kirupha, S. D., Periyaraman, P. (2011). Screening and induction of laccase activity in fungal species and its application in dye decolorization. *African Journal of Miicrobiology Research*, 5 (11), 1261–1267.
- Kuwahara, M., Glenn, J. K., Morgan, M. A., Gold, M. H. (1984). Separation and characterization of two extracellular H₂O₂-dependent oxidases from ligninolytic cultures of *Phanerochaete chrysosporium*, 169 (2), 247–250.
- Lai, C., Zeng, G. M., Huang, D. L., Zhao, M. H., Huang, H. L., Huang, C., Wei, Z., Li, N. J., Xu, P., Zhang, C., Xie, G. X. (2013). Effect of ABTS on the adsorption of *Trametes versicolor* laccase on alkali lignin. *International Biodeterioration and Biodegradation*, 82, 180–186.
- Law, K.-N., Wan Daud, W. R., Ghazali, A. (2007). Morphological and chemical nature of fiber strands of oil palm empty-fruit-bunch (OPEFB). *BioResources*, 2 (3), 351–362.
- Levin, L., Herrmann, C., Papinutti, V. L. (2008). Optimization of lignocellulolytic enzyme production by the white-rot fungus *Trametes trogii* in solid-state

fermentation using response surface methodology. *Biochemical Engineering Journal*, 39 (1), 207–214.

- Li, X., Jia, R., Li, P., Ang, S. (2009). Response surface analysis for enzymatic decolorization of Congo red by manganese peroxidase. *Journal of Molecular Catalysis B: Enzymatic*, 56 (1), 1–6.
- Limbert, G., Bryan, R., Cotton, R., Young, P., Hall-Stoodley, L., Kathju, S., Stoodley, P. (2013). On the mechanics of bacterial biofilms on nondissolvable surgical sutures: A laser scanning confocal microscopy-based finite element study. *Acta Biomaterials*, 9 (5), 6641–6652.
- Liu, J., Sidhu, S. S., Wang, M. L., Tonnis, B., Habteselassie, M., Mao, J., & Huang, Q. (2015). Fungal pretreatment of switchgrass for improved saccharification and simultaneous enzyme production. *Journal of Cleaner Production*, 104, 480–488.
- Liu, K., and Jiang, L., (2011). Bio-inspired design of multiscale structures for function integration. *Nano Today* 6 (2), 155–175.
- Loh, S. K. (2017). The potential of the Malaysian oil palm biomass as a renewable energy source. *Energy Conversion and Management* 141, 285-298.
- Lomascolo, A., Uzan-Boukhris, E., Herpoël-Gimbert, I., Sigoillot, J. C., Lesage-Meessen, L. (2011). Peculiarities of *Pycnoporus* species for applications in biotechnology. *Applied Microbiology and Biotechnology*, 92 (6), 1129–1149.
- Lonsdale, R., Harvey, J. N., and Mulholland, A. J. (2012). A practical guide to modelling enzyme-catalysed reactions. *Chemical Society Review*, 41 (8), 3025-3038.
- Ma, J. F., and Yamaji, N. (2006). Silicon uptake and accumulation in higher plants. *Trends in Plant Science*, 11 (8), 392–397.
- Ma, J. F., Yamaji, N., and Mitani-Ueno, N. (2011). Transport of silicon from roots to panicles in plants. *Proceedings of the Japan Academy. Series B: Physical and biological sciences*, 87(7), 377-385.
- Ma, K., and Ruan, Z. (2015). Production of a lignocellulolytic enzyme system for simultaneous bio-delignification and saccharification of corn stover employing co-culture of fungi. *Bioresource Technology*, 175, 586–593.
- Maciel, M. J. M., Castro e Silva, A., & Ribeiro, H. C. T. (2010). Industrial and biotechnological applications of ligninolytic enzymes of the basidiomycota: A review. *Electronic Journal of Biotechnology*, *13*(6), 1–13.

Magnusson, M. S., and Östlund, S. (2013). Numerical evaluation of interfibre

joint strength measurements in terms of three-dimensional resultant forces and moments. *Cellulose*, 20 (4), 1691–1710.

- Mäkelä, M. R., Lundell, T., Hatakka, A., Hildén, K. (2013). Effect of copper, nutrient nitrogen, and wood-supplement on the production of ligninmodifying enzymes by the white-rot fungus *Phlebia radiata*. *Fungal Biology*, 117(1), 62–70.
- Marklund, E., Eitzenberger, J., and Varna, J. (2008). Nonlinear viscoelastic material model including stiffness degradation for hem/lignin composites. *Composites Sciences and Technology*, *68*, 2156-2162.
- Martín-Sampedro, R., Rodríguez, A., Ferrer, A., García-Fuentevilla, L. L., Eugenio, M. E. (2012). Biobleaching of pulp from oil palm empty fruit bunches with laccase and xylanase. *Bioresource Technology*, 110, 371–378.
- Martinez, A. T., Speranza, M., Ruiz-dueñas, F. J., Ferreira, P., Camarero, S., Guillén, F., Martinez, M. J., Gutierrez, A., del Río, J. C. (2005). Biodegradation of lignocellulosics: Microbial, chemical, and enzymatic aspects of the fungal attack of lignin. *International Microbiology*, *8*, 195–204.
- Masran, R., Zanirun, Z., Bahrin, E. K., Ibrahim, M. F., Lai Yee, P., Abd-Aziz, S. (2016). Harnessing the potential of ligninolytic enzymes for lignocellulosic biomass pretreatment. *Applied Microbiology and Biotechnology*, 1–16.
- Matsubara, M., Lynch, J. M., and De Leij, F. A. A. M. (2006). A simple screening procedure for selecting fungi with potential for use in the bioremediation of contaminated land, *Enzyme and Microbial Technology*, *39* (7), 1365-1372.
- Maza, M., Pajot, H. F., Amoroso, M. J., Yasem, M. G. (2014) Post-harvest sugarcane residue degradation by autochthonous fungi. *International Biodeterioration and Biodegradation*, 87, 18-25.
- Mazaheri, H., Lee, K. T., Bhatia, S., & Mohamed, A. R. (2010). Subcritical water liquefaction of oil palm fruit press fiber in the presence of sodium hydroxide: An optimisation study using response surface methodology. *Bioresource Technology*, 101 (23), 9335–9341.
- Mazumder, S., Basu, S. K., Mukherjee, M. (2009). Laccase production in solidstate and submerged fermentation by *Pleurotus ostreatus*. *Engineering in Life Sciences*, 9 (1), 45–52.
- Md Yunos, N. S. H., Baharuddin, A. S., Md Yunos, K. F., Hafid, H. S., Busu, Z., Mokhtar, M. N., Sulaiman, A., Md Som, A. (2015). The physicochemical characteristics of residual oil and fibers from oil palm empty fruit bunches. *BioResources*, 10 (1), 14-29.

- Mihai, L. A., Alayyash, K., and Goriely, A. (2015). Paws, pads and plants: the enhanced elasticity of cell-filled load-bearing structures. *Proceedings of the Royal Society A*, 471, 20150107.
- Mishnaevsky, L. (2007). *Computational mesomechanics of composites: Numerical analysis of the effect of microstructures of composites on their strength and damage resistance,* John Wiley and Sons, Chichester.
- Mishra, A., Kumar, S., Kumar Pandey, A. (2011). Laccase production and simultaneous decolorization of synthetic dyes in unique inexpensive medium by new isolates of white rot fungus. *International Biodeterioration and Biodegradation*, 65 (3), 487–493.
- Mishra V., Jana A.K., Jana M.M., Gupta A. (2017). Enhancement in multiple lignolytic enzymes production for optimized lignin degradation and selectivity in fungal pretreatment of sweet sorghum bagasse. *Bioresource Technology*, 236, 49-59.
- Mohammed, I. K., Charalambides, M. N., Williams, J. G., Rasburn, J. (2013). Modelling the deformation of a confectionery wafer as a non-uniform sandwich structure. *Journal of Material Sciences*, 48 (6), 2462–2478.
- Mohammed, L., Ansari, M. N. M., Pua, G., Jawaid, M., Islam, M. S. (2015). A review on natural fiber reinforced polymer composite and its applications. *International Journal of Polymer Sciences*, Article ID 243947, 15 pages.
- Moilanen, U., Kellock, M., Varnai, A., Andberg, M., Vikari, L. (2014). Mechanisms of laccase-mediator treatments improving the enzymatic hydrolysis of pre-treated spruce. *Biotechnology for Biofuels*, 7 (177), 1-13.
- Molla, A. H., Fakhru'l-Razi, A., Abd-Aziz, S., Hanafi, M. M., Alam, M. Z. (2001). In-vitro compatibility evaluation of fungal mixed culture for bioconversion of domestic wastewater sludge. *World Journal of Microbiology and Biotechnology*, 17 (9), 849–856.
- Mood, S. H., Golfeshan, A. H., Tabatabaei, M., Jouzani, G. S., Najafi, G. H., Gholami, M., Ardjmand, M. (2013). Lignocellulose biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renewable and Sustainable Energy Reviews*, 27, 77-93.
- Mori, T., and K. Tanaka (1973). Average stress in matrix and average elastic energy of materials with misfitting inclusions. *Acta Metallurgica*, 21:571-574.
- Morozova, O. V., Shumakovich, G. P., Shleev, S. V., Yaropolov, Y. I. (2007). Laccase-mediator systems and their applications: A review. *Applied Biochemistry and Microbiology*, 43 (5), 523–535.

- Moulia, B. (2013). Plant biomechanics and mechanobiology are convergent paths to flourishing interdisciplinary research. *Journal of Experimental Botany*, 64 (15), 4617-4633.
- Muryanto, T. E., Abimayu, H., Cahyono, A., Cahyono, E. T., Sudiyani, Y. (2015). Alkaline delignification of oil palm empty fruit bunch using black liquor from pretreatment. *Procedia Chemistry*, 16, 99–105.
- Mussatto, S., and Teixeira, J. (2010). Lignocellulose as raw material in fermentation processes. *Applied Microbiology an Microbial Biotechnology*, *2*, 897–907.
- Mwaikambo, L. Y., and Ansell, M. P. (2006). Mechanical properties of alkali treated plant fibers and their potential as reinformcement materials II. Sisal fibers. *Journal of Material Sciences*, 41, 2497-2508.
- Nafu, Y. R., Foba-tendo, J., Njeugna, E., Oliver, G., Cooke, K. O. (2015). Extraction and characterization of fibres from the stalk and spikelets of empty fruit bunch. *Journal of Applied Chemistry*, 2015, 1–10.
- Nascimento, D. C. O., Ferreira, A. S., Monteiro, S. N., Aquino, R. C. M. P., Kestur, S. G. (2012). Studies on the characterization of piassava fibers and their epoxy composites. *Composites Part A: Applied Science and Manufacturing*, 43 (3), 353–362.
- Naseem A., Tabasum S., Zia K.M., Zuber M., Ali M., Noreen A. (2016). Ligninderivatives based polymers, blends and composites: A review. *International Journal of Biological Macromolecules* 93, 296-313.
- Neethirajan, S., Gordon, R., Wang, L. (2009). Potential of silica bodies (phytoliths) for nanotechnology. *Trends in Biotechnology*, 27 (8), 461–467.
- Nerud, F., and Misurcova, Z. (1996). Distribution of Ligninolytic Enzymes in Selected White-Rot Fungi. *Folia Microbiology*, 41 (3), 1988–1990.
- Ngo, T. T., Kohl, J. G., Paradise, T., Khalily, A., Simonson, D. L., (2015) Improving mechanical properties of thermoset biocomposites by fiber coating or organic oil addition. *International Journal of Polymer Sciences*, 840823, 1–7.
- Nicholson, D. J., Leavitt, A. T., and Francis, R. C. (2014). A three-stage klason method for more accurate determinations of hardwood lignin content. *Cellulose Chemistry and Technology*, *48*, 53–59.
- Nieves, D. C., Karimi, K., and Horváth, I. S. (2011). Improvement of biogas production from oil palm empty fruit bunches (OPEFB). *Industrial Crops and Products*, 34 (1), 1097–1101.

Nishiyama, Y. (2009). Structure and properties of the cellulose microfibril.

Journal of Wood Science, 55 (4), 241-249.

- Nitta, Y., Goda, K., Noda, J., & Lee, W. (2011). Effect of Alkali-Treatment on Tensile Properties of Kenaf Long Fibres using Data-based Cross-sectional Area Approximation Method. 18th International Conference on Composite Materials, 21-26th August 2011, Jeju Island, Korea, (June), 24–28.
- Nordin, N. I. A. A., Ariffin, H., Andou, Y., Hassan, M. A., Shirai, Y., Nishida, H., Yunus, W. M. Z. W., Karuppuchamy, S., Ibrahim, N. A. (2013). Modification of oil palm mesocarp fiber characteristics using superheated steam treatment. *Molecules*, *18* (8), 9132–9146.
- Nurul Hazirah, C. H., Markom, M, Harun, S, Hassan, O. (2016). The effect of various pretreatment methods on empty fruit bunch for glucose production. *Malaysian Journal of Analytical Science*, 20 (6), 1474-1480.
- Omar, F. N., Hanipah, S. H., Xiang, L. Y., Mohammed, M. A. P., Baharuddin, A. S., Abdullah, J. (2016) Micromechanical modelling of oil palm empty fruit bunch fibers containing silica bodies. *Journal of Mechanical Behaviour of Biomedical Materials*, 62, 106–118
- Omar, F.N., Mohammed, M. A. P., and Baharuddin, A. S. (2014a) Microstructure modelling of silica bodies from oil palm empty fruit bunch (OPEFB) fibres. *BioResources*, 9 (1), 938-951.
- Omar, F.N., Mohammed, M. A. P., and Baharuddin, A. S. (2014b) Effect of silica bodies on the mechanical behaviour of oil palm empty fruit bunch fibres. *BioResources*, 9 (4), 7041-7058.
- Othman, J., and Jafari, Y. (2014). Selected research issues in the Malaysian agricultural sector. *Jurnal Ekonomi Malaysia*, 48 (2), 127–136.
- Palamae, S., Dechatiwongse, P., Choorit, W., Chisti, Y., Prasertsan, P. (2017). Cellulose and hemicellulose recovery from oil palm empty fruit bunch (EFB) fibers and production of sugars from the fibers. *Carbohydrate Polymers*, 155, 491–497.
- Pan, Y., and Pelegri, A. A. (2011). Progressive Damage Analysis of Random Chopped Fiber Composite Using Finite Elements. *Journal of Engineering Materials and Technology*, 133, 1-7.
- Patel, H., Gupte, S., Gahlout, M., Gupte, A. (2014). Purification and characterization of an extracellular laccase from solid-state culture of *Pleurotus ostreatus* HP-1. *3 Biotech*, *4*, 77-84.
- Pickering, K. L., Efendy, M. G. A., Le, T. M. (2016). A review of recent developments in natural fibre composites and their mechanical

performance. Composite: Part A, 83, 98-112.

- Piñeros-Castro, Y., and Velásquez-Lozano, M. (2014). Biodegradation kinetics of oil palm empty fruit bunches by white rot fungi. *International Biodeterioration and Biodegradation*, 91, 24–28.
- Pinto, P. A., Dias, A. A., Fraga, I., Marques, G., Rodrigues, M. A. M., Colaco, J., Sampaio, A., Bezerra, R. M. F. (2012). Influence of ligninolytic enzymes on straw saccharification during fungal pretreatment. *Bioresource Technology*, 111, 261-267.
- Piscitelli, A., Giardina, P., Lettera, V., Pezzella, C., Sannia, G., Faraco, V. (2011). Induction and transcriptional regulation of laccases in fungi. *Current Genomics*, 12 (2), 104–112.
- Placet, V., Cisse, O., and Boubakar, L. (2014). Nonlinear tensile behaviour of elementary hemp fibres. Part 1: Investigation of the possible origins using repeated progressive loading with in situ microscopic observations. *Composite Part A*, 56, 319–327.
- Plácido, J., and Capareda, S. (2015). Ligninolytic enzymes: a biotechnological alternative for bioethanol production. *Bioresources and Bioprocessing*, 2 (1), 23.
- Pointing, S. B., Jones, E. B. G., Vrijmoed, L. L. P. (2000). Optimization of laccase production by *Pycnoporus sanguineus* in submerged liquid culture. *Mycologia*, 92 (1), 139–144.
- Porter, C. L. (1924). Concerning the characters of certain fungi asexhibited by their growth in the presence of other fungi. *American Journal of Botany*, 11, 168–188.
- Preston, R. D. (1952). The molecular architecture of plant cell walls. John Wiley & Sons, New York pp 31-71.
- Prychid, C. J., Rudall, P. J., Gregory, M., Url, S., Rudall, P. J., Gregory, M. (2011). Systematics and biology of silica bodies in monocotyledons Published by : Springer on behalf of New York Botanical Garden Press 69 (4), 377–440.
- Qi-He, C., Krügener, S., Hirth, T., Rupp, S., Zibek, S. (2011). Co-cultured production of lignin-modifying enzymes with white-rot fungi. *Applied Biochemistry and Biotechnology*, 165 (2), 700–718.
- Qing, H., and Mishnaevsky, L. (2009). 3D hierarchical computational model of wood as a cellular material with fibril reinforced, heterogeneous multiple layers. *Mechanics of Materials*, 41 (9), 1034–1049.

- Qing, H., and Mishnaevsky, L. (2011). A 3D multilevel model of damage and strength of wood: Analysis of microstructural effects. *Mechanics of Materials*, 43 (9), 487–495.
- Rajan, A., Kurup, J. G., and Abraham, T. E. (2010). Solid state production of manganese peroxidases using arecanut husk as substrate. *Brazilian Archives* of *Biology and Technology*, 53 (3), 555–562.
- Ramirez-Cavazos, L. I., Junghanns, C., Ornelas-Soto, N., Cardenas-Chavez, D. L., Hernandez-Luna, C., Demarche, P., Enaud, E., Garcia-Morales, R., Agathos, S. N., Parra, R. (2014). Purification and characterization of two thermostable laccases from *Pycnoporus sanguineus* and potential role in degradation of endocrine disrupting chemicals. *Journal of Molecular Catalysis B: Enzymatic*, 108, 32-42.
- Rasband, W. S. (2012). ImageJ, U.S. National Institutes of Health, Bethesda, MD.
- Ratnasingam, J., Tek, T. C., and Farrokhpayam, S. R. (2008). Tool wear characteristics of oil palm empty fruit bunch particleboard. *Journal of Applied Sciences*, 8 (8), 1594–1596.
- Reeb, C. W., Hays, T., Venditti, R. A., Gonzalez, R., Kelley, S. (2014). Supply chain analysis, delivered cost, and life cycle assessment of oil palm empty fruit bunch biomass for green chemical production in Malaysia. *Bioresources*, 9 (3), 5385–5416.
- Renge, V. C., Khedkar, S. V, and Nandurkar, N. R. (2012). Enzyme Synthesis by Fermentation Method: A Review. *Scientific Reviews and Chemical Communications*, 2 (4), 585–590.
- Resch, M. G., Donohoe, B. S., Baker, J. O., Decker, S. R., Bayer, E. A., Beckham, G. T., Himmel, M. E. (2013). Fungal cellulases and complexed cellulosomal enzymes exhibit synergistic mechanisms in cellulose deconstruction. *Energy Environmental Sciences*, 6, 1858–1867.
- Reuss, A. (1929). Berechnung der Fließgrenze von Mischkristallen auf Grund der Plastizitätsbedingung für Einkristalle. Journal of Applied Mathematics and Mechanics, 9, 49–58.
- Risdianto, H., Sofianti, E., Suhardi, S. H., & Setiadi, T. (2012). Optimisation of laccase production using white rot fungi and agriculture wastes in solid state fermentation. *ITB Journal of Engineering Science*, 44 (1), 93–105.
- Rosli, N. S., Harun, S., Md Jahim, J., & Othaman, R. (2017). Chemical and physical characterization of oil palm empty fruit bunch. *Malaysian Journal* of Analytical Sciences, 21(1), 188–196.

- Ruqayyah, T. I. D., Jamal, P., Alam, M. Z., & Mirghani, M. E. S. (2013). Biodegradation potential and ligninolytic enzyme activity of two locally isolated *Panus tigrinus* strains on selected agro-industrial wastes. *Journal of Environmental Management*, 118, 115–121.
- Sabrina, D. T., Gandahi, A. W., Hanafi, M. M., Mahmud, T. M. M., & Nor Azwady, A. A. (2012). Oil palm empty-fruit bunch application effects on the earthworm population and phenol contents under field conditions. *African Journal of Biotechnology*, 11(19), 4396–4406.
- Saha, B. C., Qureshi, N., Kennedy, G. J., Cotta, M. A. (2016). Biological pretreatment of corn stover with white-rot fungus for improved enzymatic hydrolysis. *International Biodeterioration and Biodegradation*, 109, 29–35.
- Sakurai, T., and Kataoka, K. (2007). Structure and function of type I copper in multicopper oxidases. *Cellular and Molecular Life Sciences*, 64 (19), 2642– 2656.
- Salmén, L. (2004). Micromechanical understanding of the cell-wall structure. *Comptes Rendus - Biologies*, 327 (9), 873–880.
- Sánchez, C. (2009). Lignocellulosic residues: Biodegradation and bioconversion by fungi. *Biotechnology Advances*, 27(2), 185–194.
- Santhanam, N., Vivanco, J. M., Decker, S. R., Reardon, K. F. (2011). Expression of industrially relevant laccases: Prokaryotic style. *Trends in Biotechnology*, 29 (10), 480–489.
- Saraiva, J. A., Tavares, A. P. M., and Xavier, A. M. R. B. (2012). Effect of the inducers veratryl alcohol, xylidine, and ligninosulphonates on activity and thermal stability and inactivation kinetics of laccase from *Trametes versicolor*. *Applied Biochemistry and Biotechnology*, 167, 685-693.
- Sarnthima, R., Khammuang, S., and Svasti, J. (2009). Extracellular ligninolytic enzymes by *Lentinus polychrous* Lev. under solid-state fermentation of potential agro-industrial wastes and their effectiveness in decolorization of synthetic dyes. *Biotechnology and Bioprocess Engineering*, 14 (4), 513–522.
- Schroyen, M., Vervaeren, H., Van Hulle, S. W. H., Raes, K. (2014). Impact of enzymatic pretreatment on corn stover degradation and biogas production. *Bioresource Technology*, 173, 59–66.
- Senawi, R., Alauddin, S. M., Saleh, R. M., Shueb, M. I. (2013). Polylactic acid/empty fruit bunch fiber biocomposite: influence of alkaline and silane treatment on the mechanical properties. International Journal of Bioscience, Biochemistry and Bioinformatics, 3 (1), 59-61.

- Shamsudin, S., Md Shah, U. K., Zainudin, H., Abd-Aziz, S., Mustapa Kamal, S. M., Shirai, Y., Hassan, M. A. (2012). Effect of steam pretreatment on oil palm empty fruit bunch for the production of sugars. *Biomass and Bioenergy*, 36, 280–288.
- Sharma, R. K., and Arora, D. S. (2010). Production of lignocellulolytic enzymes and enhancement of in vitro digestibility during solid state fermentation of wheat straw by *Phlebia floridensis*. *Bioresource Technology*, 101 (23), 9248– 9253.
- Shinoj S., Visvanathan R., Panigrahi S., Kochubabua M. (2011). Oil palm fiber (OPF) and its composites: A review. *Industrial Crops and Products*, 33, 7–22.
- Shirkavand, E., Baroutian, S., Gapes, D. J., Young, B. R. (2016). Combination of fungal and physicochemical processes for lignocellulosic biomass pretreatment - A review. *Renewable and Sustainable Energy Reviews*, 54, 217– 234.
- Shuit, S. H., Tan, K. T., Lee, K. T., Kamaruddin, A. H. (2009). Oil palm biomass as a sustainable energy source: A Malaysian case study, *Energy*, *34*, 1225-1235.
- Simarani, K., Hassan, M. A., Abd-Aziz, S., Wakisaka, M., Shirai, Y. (2009). Effect of palm oil mill sterilization process on the physicochemical characteristics and enzymatic hydrolysis of empty fruit bunch. Asian Journal of Biotechnology, 1 (2), 57-66.
- Singanathan, J. (2010). Production of methane from palm oil mill effluent using membrane anaerobic system, Degree Thesis, Universiti Malaysia Pahang, Malaysia.
- Singh, P., Sulaiman, O., Hashim, R., Peng, L. C., Singh, R. P. (2012). Biodegradation study of pycnoporus sanguineus and its effects on structural and chemical features on oil palm biomass chips. *BioResources*, 1 (3), 210–227.
- Singhania, R. R., Patel, A. K., Soccol, C. R., Pandey, A. (2009). Recent advances in solid-state fermentation. *Biochemical Engineering Journal*, 44 (1), 13–18.
- Singleton, V. L., Orthofer, R., and Lamuela-Ravent, R. M. (1998). Analysis of total phenols and other oxidation substrates and antioxidants by means of folinciocalteu reagent. *Methods in Enzymology*, 299, 152–178.
- Sluiter, J. B., Ruiz, R. O., Scarlata, C. J., Sluiter, A. D., Templeton, D. W. (2010). Compositional analysis of lignocellulosic feedstocks: 1. Review and description of methods. *Journal of Agricultural and Food Chemistry*, 58, 9043-9053.

- Sreekala, M. S., Kumaran, M. G., Joseph, R. and Thomas, S. (2001). Stressrelaxation behaviour in composites based on short oil-palm fibres and phenol formaldehyde resin. *Composite Science and Technology*, 61, 1175-1188.
- Sreekala, M. S., Kumaran, M. G., and Thomas, S. (1997). Oil palm fibers: Morphology, chemical composition, surface modification, and mechanical properties. *Journal of Applied Polymer Science*, 66 (5), 821–835.
- Stahl, P. D. and Christensen, M. (1992). In vitro mycelial interactions among members of a soil microfungal community. *Soil Biology and Biochemistry*, 24, 309–316.
- Stanbury, PF. Whitaker, A. Hall, S. J. (2013). Principles of fermentation technology. *Journal of Chemical Information and Modeling*, 53 (9), 1689–1699.
- Subramaniyam, R., and Vimala, R. (2012). Solid state and submerged fermentation for the production of bioactive substances: A comparative study. *International Journal of Science and Nature*, 3 (2012), 480–486.
- Sun, F. H., Li, J., Yuan, Y. X., Yan, Z. Y., & Liu, X. F. (2011). Effect of biological pretreatment with *Trametes hirsuta* yj9 on enzymatic hydrolysis of corn stover. *International Biodeterioration and Biodegradation*, 65 (7), 931–938.
- Sun, S., Sun, S., Cao, X., Sun, R. (2016). The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials. *Bioresource Technology*, 199, 49–58.
- Suzuki, S., Ma, J. F., Yamamoto, N., Hattori, T., Sakamoto, M. and Umezawa, T. (2012). Silicon deficiency promotes lignin accumulation in rice. *Plant Biotechnology*, 29, 391–394.
- Tan, H., Huang, Y., Liu, C., Geubelle, P. H. (2005). The Mori-Tanaka method for composite materials with nonlinear interface debonding. *International Journal of Plasticity*, 21, 1890-1918.

TAPPI (2002) Acid insoluble lignin in wood and pulp. T222om-02 2002 (pp. 3-7).

- Thakur, S., Shrivastava, B., Ingale, S., Kuhad, R. C., Gupte, A. (2013). Degradation and selective ligninolysis of wheat straw and banana stem for an efficient bioethanol production using fungal and chemical pretreatment. *3 Biotech*, *3* (5), 365-372.
- Thomas, L., Larroche, C., & Pandey, A. (2013). Current developments in solidstate fermentation. *Biochemical Engineering Journal*, *81*, 146–161.

Tien, M., and Kirk, T. K. (1988). Lignin peroxidase of Phanerochaete chrysosporium.

Methods in Enzymology, 161, 238–249.

- Umikalsom, M. S., Ariff, A. B., Zulkifli, H. S., Tong, C. C., Hassan, M. A., Karim, M. I. A. (1997). The treatment of oil palm empty fruit bunch fibre for subsequent use as substrate for cellulase production by *Chaetomium globosum* kunze. *Bioresource Technology*, 62, 1–9.
- Umikalsom, M. S., and Sariah, M. (2006). Utilization of microbes for sustainable agriculture in Malaysia: current status. Bio prospecting and management of microorganisms. *National Conference on Agro Biodiversity Conservation and Sustainable Utilization*, pp 27–29.
- Underwood, E. E. (1970). Quantitative stereology, Addidon-Wesley, New York.
- United Plantations (2015) Annual Report for 2015, United Plantations Berhad.
- Usha, K. Y., Praveen, K., and Reddy, B. R. (2014). Enhanced production of ligninolytic enzymes by a mushroom *Stereum ostrea*. *Biotechnology Research International*, ID 815495.
- Uzan, E., Nousiainen, P., Balland, V., Sipila, J., Piumi, F., Navarro, D., Asther, M., Record, E., Lomascolo, A. (2010). High redox potential laccases from the ligninolytic fungi *Pycnoporus coccineus* and *Pycnoporus sanguineus* suitable for white biotechnology: From gene cloning to enzyme characterization and applications. *Journal of Applied Microbiology*, 108 (6), 2199–2213.
- Valasek, P., Ruggiero, A., and Muller, M. (2017). Experimental description of strength and tribological characteristic of EFB oil palm fibres/epoxy composites with technologically undemanding preparation. *Composites Part B: Engineering*, 122, 79-88.
- Van Bockhaven, J., De Vleesschauwer, D., and Hofte, M. (2013). Towards establishing broad-spectrum disease resistance in plants: Silicon leads the way. *Journal of Exprimental Botany*, 64 (5), 1281-1293.
- Van Heerden, A., Le Roux, N. J., Swart, J., Gardner-Lubbe, S., Botha, A. (2008). Assessment of wood degradation by *Pycnoporus sanguineus* when cocultured with selected fungi. *World Journal of Microbiology and Biotechnology*, 24 (11), 2489–2497.
- Vikineswary, S., Abdullah, N., Renuvathani, M., Sekaran, M., Pandey, A., Jones, E. B. G. (2006). Productivity of laccase in solid substrate fermentation of selected agro-residues by *Pycnoporus sanguineus*. *Bioresource Technology*, 97 (1), 171–177.

Voigt, W. (1887). Ueber das Doppler'sche Princip Nachr. Ges. Wiss. G"ottingen 8,

41–51. An English version can be found on Ernst, A. and Hsu, J. P. (2001). First proposal of the universal speed of light by Voigt in 1887, *Chinese Journal of Physics*, 39, 211–230.

- Wahab, A. G. (2015) GAIN Report, USDA Foreign Agricultural Service Global Agricultural Information.http://agriexchange.apeda.gov.in/marketreport/Reports/P alm%20Oil%20PSD%20Revisions_Kuala%20Lumpur_Malaysia_12-10-2015.pdf, Retrieved on 11 January 2016.
- Wan, C., and Li, Y. (2011). Effectiveness of microbial pretreatment by *Ceriporiopsis subvermispora* on different biomass feedstocks. *Bioresource Technology*, 102 (16), 7507–7512.
- Wan Razali, W. A., Baharuddin, A. S., Talib, A. T., Sulaiman, A., Naim, M. N., Hassan, M. A., Shirai, Y. (2012). Degradation of oil palm empty fruit bunches (OPEFB) fibre during composting process using in-vessel composter. *BioResources*, 7(4), 4786–4805.
- Wan Razali, W. A., Samsu Baharuddin, A., Zaini, L. A., Mokhtar, M. N., Taip, F. S., & Zakaria, R. (2014). Effect of seed sludge quality using oil palm empty fruit bunch (OPEFB) bio-char for composting. *BioResources*, 9 (2), 2739–2756.
- Webber, J. F., and Hedger, J. N. (1986). Comparison of interactions between *Ceratocystis ulmi* and Elm bark saprobes in vitro and in vivo. *Transactions of the British Mycological Society*, 86, 93-101.
- Wegst, U. G. K., Bai, H., Saiz, E., Tomsia, A. P., Ritchie, R. O. (2015). Bioinspired structural materials. *Natural Materials*, 14 (1), 23–36.
- White, T. J., Bruns, T., Lee, S., & Taylor, J. (1990). Amplification and direct sequencing of fungal ribosomal RNA Genes for Phylogenetics. PCR Protocols: A Guide to Methods and Applications, (January), 315–322.
- Xiang, L. Y., Hanipah, S. H., Mohammed, M. A. P., Baharuddin, A. S., Lazim, A. M. (2015). Microstructural, mechanical and physicochemical behaviours of alkali pre-treated oil palm stalk fibers. *BioResources*, 10(2), 2783-2796.
- Xiang, L. Y., Mohammed, M. A. P., Baharuddin, A. S. (2016). Characterisation of microcrystalline cellulose from oil palm fibres for food applications. *Carbohydrate Polymers*, *148*, 11-20.
- Xu, C., Ma, F., & Zhang, X. (2009). Lignocellulose degradation and enzyme production by Irpex lacteus CD2 during solid-state fermentation of corn stover. *Journal of Bioscience and Bioengineering*, 108 (5), 372–375.

Yamamoto, T., Nakamura, A., Iwai, H., Ishii, T., Ma, J.F., Yokoyama, R.,

Nishitani, K., Satoh, S., Furukawa, J. (2012). Effect of silicon deficiency on secondary cell wall synthesis in rice leaf. *Joural of Plant Research*, 125, 771–779.

- Yamanaka, S., Sato, K., Ito, F., Komatsubara, S., Ohata, H., Yoshino, K. (2012). Roles of silica and lignin in horsetail (*Equisetum hyemale*), with special reference to mechanical properties. Journal of Applied Physics, 111, 1-5.
- Ying Ying, T., Teong, L. K., Wan Abdullah, W. N., Peng, L. C. (2014). The effect of various pretreatment methods on oil palm empty fruit bunch (EFB) and kenaf core fibers for sugar production. *Procedia Environmental Sciences*, 20, 328–335.
- Yokoyama, S. (2008). The Asian Biomass Handbook. A guide for biomass production and utilization. The Japan Institute of Energy.
- Yu, J., Zhang, J., He, J., Liu, Z., & Yu, Z. (2009). Combinations of mild physical or chemical pretreatment with biological pretreatment for enzymatic hydrolysis of rice hull. *Bioresource Technology*, 100(2), 903-908.
- Yu, W. (2016) An introduction to micromechanics. Applied Mechanics and Materials, 828,3-24.
- Yusoff, M. Z. M., Salit, M. S., and Ismail, N. (2009). Tensile properties of single oil palm empty fruit bunch (OPEFB) fibre. *Sains Malaysiana*, 38 (4), 524–529.
- Zadrazil, F., Gonser, A., Lang, E., (1999). Influence of incubation temperature on the secretion of extracellular ligninolytic enzymes of *Pleurotus* sp. and *Dichomitus squalens* into oil. In: *Proceedings of Conference of Enzymes and Environment*, Granada, Spain, pp. 12–16.
- Zainudin, M. H. M., Nor' Aini, A. R., Abd-Aziz, S., Funaoka, M., Shinano, T., Shirai, Y., Wakisaka, M., Hassan, M. A. (2012) Utilization of glucose recovered by phase separation system from acid-hydrolysed oil palm empty fruit bunch for bioethanol production. *Pertanika Journal of Tropical Agricultural Sciences*, 35 (1), 117-126.
- Zanirun, Z. (2016). Lignin pretreatment of oil palm empty fruit bunch using ligninolytic enzyme mediator and cellulose hydrolysis dor fermentable sugar production. *PhD Thesis*, Universiti Putra Malaysia.
- Zanirun, Z., Bahrin, E. K., Lai-Yee, P., Hassan, M. A., Abd-Aziz, S. (2015). Enhancement of fermentable sugars production from oil palm empty fruit bunch by ligninolytic enzymes mediator system. *International Biodeterioration and Biodegradation*, 105, 13–20.

Zhang, J., Ren, X., Chen, W., Bao, J. (2012). Biological pretreatment of corn stover

by solid state fermentation of *Phanerochaete chrysosporium*. Frontiers of *Chemical Science and Engineering*, 6 (2), 146–151.

- Zheng, Y., Zhao, J., Xu, F., Li, Y. (2014). Pretreatment of lignocellulosic biomass for enhanced biogas production. *Progress in Energy and Combustion Science*, 42 (1), 35–53.
- Zhu, C., Bao, G., and Huang, S (2016). Optimization of laccase production in the white rot fungus *Pleurotus ostreatus* (ACCC 52857) induced through yeast extract and copper. *Biotechnology & Biotechnology Equipment*, 30 (2), 270-276.
- Zhuang, J., and Marchant, M. (2007). Economic analysis of cellulase production methods for bio-ethanol. *Applied Engineering in Agriculture*, 23 (5), 679–687.
- Zhuo, R., Ma, L., Fn, F., Gong, Y., Wan, X., Jiang, M., Zhang, X., Yang, Y. (2011). Decolorization of different dyes by a newly isolated white-rot fungi strain *Ganoderma* sp.En3 and cloning and functional analysis of its laccase gene. *Journal of Hazardous Materials*, 192, 855-873.
- Zimbardi, A. L. R. L., Camargo, P. F., Carli, S., Neto, S. A., Meleiro, L. P., Rosa, J. C., De Andrade, A. R., Jorge, J. A., Furriel, R. P. M. (2016) A high redox potential laccase from *Pycnoporus sanguineus* RP15: Potential application for dye decolorization. *International Journal of Molecular Science*, 17 (672), 1-24.
- Zulkiple, N., Maskat, M. Y. and Hassan, O. (2016). Pretreatment of oil palm empty fruit bunch (OPEFB) with aquaeous ammonia for high production of sudar. *Proceedia Chemistry* 18, 155-161.