

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF COLONIZATION OF AN ENDOPHYTIC FUNGUS, Hendersonia toruloidea ON THIAMINE BIOSYNTHESIS IN OIL PALM SEEDLINGS (Elaeis guineensis Jacq.)

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FBSB 2018 6



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Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECTS OF COLONIZATION OF AN ENDOPHYTIC FUNGUS, Hendersonia toruloidea ON THIAMINE BIOSYNTHESIS IN OIL PALM SEEDLINGS (Elaeis guineensis Jacq.)

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October 2017

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Thiamine, or vitamin B1 plays an indispensable role in many metabolic reactions. Besides that, thiamine is also associated with the induction of systemic acquired resistance (SAR) in plants and having a role in boosting plant's immunity and defense system. In Malaysia, oil palm productivity is hampered by basal stem rot disease caused by a pathogenic fungus, Ganoderma boninense and proper disease management have yet to be discovered. Application of endophytes as biocontrol agent is a promising measure to prevent the disease. Hendersonia toruloidea is an endophytic fungus originally isolated from oil palm roots which have been shown to have excellent biocontrol activity in oil palm seedlings. Previous studies showed that this endophyte is able to suppress G. boninense infection in oil palm seedlings. This work aimed to investigate the responses in oil palm seedlings, specifically on the expressions of thiamine biosynthesis genes upon application of *H. toruloidea*. Seven months old oil palm seedlings were inoculated with *H. toruloidea* and microscopy analyses were carried out to visualize the colonization of the fungus. Total RNA was extracted from oil palm leaves at day 1, 7, 15 and 30 post inoculation. Quantitative real-time PCR (qPCR) was performed to measure the level of expression of four key thiamine biosynthesis genes, namely THI4, THIC, TH1 and TPK. The results showed of up to 12-fold of increase in the expression of all gene transcripts at day 1 post inoculation. At subsequent days of day 7, day 15 and 30 post inoculation, the relative expression of these genes were shown to be downregulated. Thiamine accumulation was observed via HPLC analysis at day 7 post inoculation and subsequently attenuated until day 30. This work provides first evidence of enhancement of thiamine biosynthesis by endophytic colonization in oil palm and suggesting the role of thiamine in stress protection in oil palm seedlings.

KESAN KOLONISASI KULAT ENDOFIT Hendersonia toruloidea TERHADAP BIOSINTESIS VITAMIN B1 DALAM KELAPA SAWIT (Elaeis guineensis Jacq.)

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Vitamin B1 memainkan peranan yang penting dalam metabolisma dalam semua kehidupan. Selain itu, vitamin B1 juga terlibat dalam induksi rintangan sistemik dalam tumbuhan dan memainkan peranan penting dalam immuniti dan pertahanan tumbuhan. Di Malaysia, produktiviti kelapa sawit adalah terjejas disebabkan oleh penyakit pereputan pangkal akar yang disebabkan oleh kulat Ganoderma boninense. Namun, pengawalan terbaik dan efektif terhadap kulat Ganoderma boninense belum dikenalpasti. Pengawalan penyakit BSR telah dilakukan dengan menggunakan agen kawalan biologi iaitu melalui penggunaan endofit terhadap anak pokok kelapa sawit. Kulat Hendersonia toruloidea merupakan kulat endofit yang telah dijumpai dalam akar kelapa sawit dan telah menunjukkan keberkesanan yang memberangsangkan terhadap kawalan penyakit BSR. Oleh itu, kajian ini bertujuan untuk menyelidiki kesan kolonisasi kulat endofit *H. toruloidea* terhadap biosintesis vitamin B1 di dalam anak pokok kelapa sawit. Kulat endofit H. toruloidea telah diinokulasi ke atas anak benih kelapa sawit berusia 7 bulan di nurseri. Analisa mikroskopi telah dijalankan untuk mengkaji kolonisasi kulat endofit di dalam akar kelapa sawit. RNA telah diekstrak daripada daun kelapa sawit pada hari ke 1, 7, 15 dan 30 hari usai inokulasi. Tindak balas rangkaian polimerase secara secara kuantitatif masa sebenar (qPCR) telah dijalankan untuk melihat pengkspresan empat gen biosintesis B1 yang utama iaitu THI4, THIC, TH1 dan TPK. Hasil penemuan menunjukkan pengekspresan gen biosintesis vitamin B1 meningkat sehingga 12 kali ganda. Selepas 15 dan 30 hari usai inokulasi, pengekspresan gen vitamin B1 telah menurun. Analisa HPLC menunjukkan kolonisasi kulat endofit telah menyebabkan pengumpulan metabolit vitamin B1 di dalam daun kelapa sawit. Hasil kajian ini berjaya menunjukkan kolonisasi H. toruloidea meningkatkan biosintesis vitamin B1 di dalam pokok dan mencadangkan peranan vitamin B1 dalam perlindungan kelapa sawit terhadap tekanan.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

°C Degree celsius

% Percentage

A_{260nm} Optical density at wavelength 260 nanometer

A_{280nm} Optical density at wavelength 280 nanometer

μL Microliter

μm Micrometre

μmoles Micromoles

mg Milligram

ATP Adenosine triphosphate

bp Base pair

cDNA Complementary deoxyribonucleic acid

CTAB Cetyl trimethyammonium bromide

DNA Deoxyribonucleic acid

EDTA Ethylenediaminetetraacetic acid

g Gram

HCl Hydrochloric acid

kb Kilobase

L Litres

LB Luria Bertani

M Moles

mRNA Messenger RNA

nm Nanomoles

NaOH Sodium hydroxide

PDA Potato dextrose agar

PCR Polymerase chain reaction

qPCR Quantitative real-time PCR

RNA Ribonucleic acid

RT Reverse trancriptase



CHAPTER 1

INTRODUCTION

Plants are sessile organisms that are inevitably exposed to unfavourable biotic and abiotic stresses, namely salinity, drought, and also microbial pathogens. Climate changes and attack by pathogenic diseases can severely hamper the productivity of important crop plants, including oil palm. Oil palm is one of Malaysia's important commodity. However, the productivity of oil palm is threatened by basal stem rot disease caused by a fungus, *Ganoderma boninense* which resulted in major economic losses (Paterson, 2007; Rees *et al.*, 2009). Current research in oil palm is accelerating towards finding ways to control the disease and is focusing on the detailed molecular mechanism in the plant-pathogenic interaction (Ho and Tan, 2014). Recently, studies of plant and microorganism interactions are significantly attracting interest because it has been demonstrated that microorganisms such as endophytes play a role in alleviating stresses in their host plants (Boivin *et al.*, 2016).

Endophytes are microorganisms that colonize the insides of plant tissues without causing any disease (Wilson, 1995). It is widely documented that endophytes formed a beneficial mutualistic relationship with plants (Hernández-Montiel *et al.*, 2013; Seerangan and Thangavelu, 2014). It is suggested that plant-endophyte mutualism is formed through direct and indirect mechanisms. Direct mechanism include antibiosis, and indirect is through production of biochemical compounds that are associated to alleviating stresses through induced systemic resistance (ISR), thereby enhancing plant's immune system and preventing pathogenic attack (Alquéres *et al.*, 2010; Gao *et al.*, 2010).

Relatively, no studies have been done on the role of thiamine in stress protection in oil palm. Thiamine or vitamin B1, an enzymatic cofactor in metabolic reactions, is involved in plant adaptation and alleviation of biotic and abiotic stresses in plants (Tunc-Ozdemir *et al.*, 2009; Rapala-Kozik *et al.*, 2008; Goyer, 2010). It was observed that there was an accumulation of thiamine when the plants were subjected to salinity stress, oxidative stress and pathogenic attack (Rapala-Kozik *et al.*, 2008; Tunc-Ozdemir *et al.*, 2009; Zhou *et al.*, 2013). It is now understood that thiamine formed an indirect role in enhancing anti oxidative capacity in the plants, which is important in defense responses (Zhou *et al.*, 2013). Yet, the exact mechanism of biosynthesis of thiamine in response to stresses is still poorly understood.

Thiamine is involved in adaptation to biotic and abiotic stresses and application of endophytes enhance the synthesis of defence metabolites that is associated with ISR/SAR (Zheng *et al.*2015). This led to a hypothesis that thiamine biosynthesis in oil palm will be upregulated by colonisation by endophytes. In this study, the endophytic fungus *Hendersonia toruloidea* was chosen as the strain of choice due to its excellent

colonization ability and also its ability to suppress *Ganoderma boninense* disease infection in oil palm (Idris *et al.*, 2013).

1.1 Objectives

The objectives of this study are:

- 1. To determine the localization and colonisation of endophytic fungus in oil palm by scanning electron microscopy and transmission electron microscopy
- 2. To determine the level of expression of thiamine biosynthesis genes upon colonisation of endophytic fungus by quantitative PCR
- 3. To quantify the total thiamine accumulation in oil palm upon colonisation by endophytic fungus by HPLC

The first objective was to perform microscopy analysis to study the colonisation and morphological pattern of the endophytic fungus. It was expected that the endophytic *H. toruloidea* will actively colonizing oil palm root tissues. Scanning electron microscopy (SEM) revealed the morphology of the *H. toruloidea* while transmission electron microscopy (TEM) was carried out to examine the structure and localisation of *H. toruloidea* in the oil palm root.

The second objective was to determine the expressions of thiamine biosynthesis genes in oil palm upon successive colonisation of *H. toruloidea*. The successful colonization of endophytic fungus was hypothesized to cause an upregulation of the thiamine biosynthesis genes in oil palm. The gene expression study was performed using quantitative real time-PCR (qRT-PCR), a simple, high throughput technology that enable us to measure gene expression in real time. The expression of thiamine biosynthesis genes was examined over a time course of 1, 7, 15 and 30 days post inoculation and the result will reflect the changes in transcript abundances of thiamine biosynthesis genes upon colonization of *H. toruloidea*.

The third objective of this study was to measure total thiamine content in oil palm upon colonization of the endophytic fungus. The upregulation of thiamine biosynthesis genes was expected to cause the increase in total thiamine and its intermediates accumulation overall. It was performed using High Performance Liquid Chromatography (HPLC). Since gene expressions are not necessarily translated into functional protein, the measurement of total thiamine and its intermediate content will verify that thiamine biosynthesis genes are expressed to synthesise total thiamine or the synthesis of its intermediates might be involved in other mechanisms in the overall metabolic pathways.

REFERENCES

- Abidin, A. A. Z., Wong, S. Y., Abdul Rahman, N. S., Che Idris, and Balia Yusof, Z. N. (2016). Osmotive, oxidative, and salinity stresses upregulate the expressions of thiamine (vitamin B1) biosynthesis genes (*THIC & THI1/THI4*) in oil palm (*Elaeis guineensis*). *Journal of Oil Palm Research*, 28(3): 308–319.
- Ahn, I. P., Kim, S., and Lee, Y. H. (2005). Vitamin B1 functions as an activator of plant disease resistance. *Plant Physiology*, 138(3): 1505–15.
- Ahn, I. P., Kim, S., Lee, Y. H., and Suh, S. C. (2007). Vitamin B1-induced priming is dependent on hydrogen peroxide and the NPR1 gene in *Arabidopsis*. *Plant Physiology*, 143(2): 838–48.
- Alquéres, S. M. C., Oliveira, J. H. M., Nogueira, E. M., Guedes, H. V, Oliveira, P. L., Câmara, F., and Martins, O. B. (2010). Antioxidant pathways are up-regulated during biological nitrogen fixation to prevent ROS-induced nitrogenase inhibition in *Gluconacetobacter diazotrophicus*. Archives of Microbiology, 192(10): 835–41.
- Ariffin, D., Idris, A. S., and Marzuki, A. (Eds.). (1996). Proceedings of the 1996 PORIM International Palm Oil Congress (Agriculture). PORIM, Malaysia. p. 317-329.
- Asano, Y., Katsumoto, H., Inokuma, C., Kaneko, S., Ito, Y., and Fujiie, A. (1996). Cytokinin and thiamine requirements and stimulative effects of riboflavin and α-ketoglutaric acid on embryogenic callus induction from the seeds of *Zoysia japonica steud. Journal of Plant Physiology*, 149(3–4): 413–417.
- Bahuguna, R. N., Joshi, R., Shukla, A., Pandey, M., and Kumar, J. (2012). Thiamine primed defense provides reliable alternative to systemic fungicide carbendazim against sheath blight disease in rice (*Oryza sativa L.*). *Plant Physiology and Biochemistry*, 57: 159–67.
- Balia Yusof, Z. N., Borhan, F. P., Mohamad, F. A., and Rusli, M. H. (2015). The effect of *Ganoderma boninense* infection on the expresions of thiamine (Vitamin B1) biosynthesis genes in oil palm. *Journal of Oil Palm Research*, 27(1): 12–18.
- Barcelos, E., Rios, S. de A., Cunha, R. N. V., Lopes, R., Motoike, S. Y., Babiychuk, E., and Kushnir, S. (2015). Oil palm natural diversity and the potential for yield improvement. *Frontiers in Plant Science*, 6: 190.
- Bustin, S. A., Benes, V., Garson, J. A., Hellemans, J., Huggett, J., Kubista, M., et al. (2009). The MIQE guidelines: Minimum Information for publication of quantitative real-time PCR experiments. *Clinical Chemistry*, 55, 611–622.
- Begley, T. P., Downs, D. M., Ealick, S. E., McLafferty, F. W., Van Loon, A. P. G. M., Taylor, S., and Xi, J. (1999). Thiamin biosynthesis in prokaryotes. *Archives of*

- Microbiology, 171(5): 293-300
- Boivin, S., Fonouni-Farde, C., and Frugier, F. (2016). How Auxin and Cytokinin Phytohormones Modulate Root Microbe Interactions. *Frontiers in Plant Science*, 7: 1240.
- Boubakri, H., Wahab, M. A., Chong, J., Bertsch, C., Mliki, A., and Soustre-Gacougnolle, I. (2012). Thiamine induced resistance to *Plasmopara viticola* in grapevine and elicited host-defense responses, including HR like-cell death. *Plant Physiology and Biochemistry*. 57: 120–33.
- Cabezas, L., Calderon, C., Medina, L. M., Bahamon, I., Cardenas, M., Bernal, A. J., and Restrepo, S. (2012). Characterization of cellulases of fungal endophytes isolated from *Espeletia* spp. *Journal of Microbiology* 50(6): 1009–13.
- Chan, P. L., Rose, R. J., Abdul Murad, A. M., Zainal, Z., Leslie Low, E. T., Ooi, L. C. L., et al. (2014). Evaluation of reference genes for quantitative real-time PCR in oil palm elite planting materials propagated by tissue culture. *PLoS One*, 9(10): e110079
- Choudhary, D. K., Prakash, A., and Johri, B. N. (2007). Induced systemic resistance (ISR) in plants: mechanism of action. *Indian Journal of Microbiology*, 47(4): 289–97.
- Colinas, M., and Fitzpatrick, T. B. (2015). Natures balancing act: examining biosynthesis de novo, recycling and processing damaged vitamin B metabolites. *Current Opinion in Plant Biology*, 25: 98–106.
- Dhillon, R. S., Hooda, M. S., Pundeer, J. S., Ahlawat, K. S., and Chopra, I. (2011). Effects of auxins and thiamine on the efficacy of techniques of clonal propagation in *Jatropha curcas L. Biomass and Bioenergy*, 35(4): 1502–1510.
- Djonovic, S., Vargas, W. A, Kolomiets, M. V, Horndeski, M., Wiest, A., and Kenerley, C. M. (2007). A proteinaceous elicitor Sm1 from the beneficial fungus *Trichoderma virens* is required for induced systemic resistance in maize. *Plant Physiology*, 145: 875–889.
- Dong, W., Stockwell, V. O., and Goyer, A. (2015). Enhancement of thiamin content in *Arabidopsis thaliana* by metabolic engineering. *Plant and Cell Physiology*, 56(12): 2285–2296.
- Dong, W., Thomas, N., Ronald, P. C., and Goyer, A. (2016). Overexpression of thiamin biosynthesis genes in rice Increases leaf and unpolished grain thiamin content but not resistance to *Xanthomonas oryzae* pv. *oryzae*. *Frontiers in Plant Science*, 7: 616.
- Du, Q., Wang, H., and Xie, J. (2011). Thiamin (vitamin B1) biosynthesis and regulation: A rich source of antimicrobial drug targets? *International Journal of Biological Sciences*. 7(1): 41-52.

- Ferdous, A. S., Islam, T., Alam, S. S., and Khan, H. (2015). Identification of stable reference genes for quantitative PCR in jute under different experimental conditions: An essential assessment for gene expression analysis. *Australian Journal of Crop* Science, 9: 646–655.
- Ganley, R. J., Sniezko, R. A., and Newcombe, G. (2008). Endophyte-mediated resistance against white pine blister rust in *Pinus monticola*. Forest Ecology and Management, 255(7): 2751–2760.
- Gao, F. K., Dai, C. C., and Liu, X. Z. (2010). Mechanisms of fungal endophytes in plant protection against pathogens. *African Journal of Microbiology Research*, 4(13): 1346–1351.
- Gindro, K., Pezet, R., and Viret, O. (2003). Histological study of the responses of two *Vitis vinifera* cultivars (resistant and susceptible) to *Plasmopara viticola* infections. *Plant Physiology and Biochemistry*, 41(9): 846–853.
- Goyer, A. (2010). Thiamine in plants: Aspects of its metabolism and functions. *Phytochemistry*, 71(14–15): 1615–1624.
- Goyer, A., and Haynes, K. G. (2011). Vitamin B1 Content in Potato: Effect of Genotype, Tuber Enlargement, and Storage, and Estimation of Stability and Broad-Sense Heritability. *American Journal of Potato Research*, 88(4): 374–385.
- Gond, S. K., Verma, V. C., Mishra, A., Kumar, A., and Kharwar, R. N. (2010). Role of Fungal Endophytes in Plant Protection. *Management of Fungal Plant Pathogens*, 183–197.
- Guan, J., Hasnain, G., Garrett, T. J., Chase, C. D., Gregory, J., Hanson, A. D., and Mccarty, D. R. (2014). Divisions of labor in the thiamin biosynthetic pathway among organs of maize. *Frontiers in Plant Science*, 5(370): 1–11.
- Hamada, A. M., and Jonsson, L. M. V. (2013). Thiamine treatments alleviate aphid infestations in barley and pea. *Phytochemistry*, 94: 135–41.
- Hardoim, P. R., van Overbeek, L. S., Berg, G., Pirttilä, A. M., Compant, S., Campisano, A., et al. (2015). The hidden world within plants: Ecological and Evolutionary considerations for defining functioning of microbial endophytes. *Microbiology Molecular Biology Reviews*, 79: 293–320.
- Helliwell, K. E., Scaife, M. A., Sasso, S., Araujo, A. P. U., Purton, S., and Smith, A. G. (2014). Unraveling Vitamin B12-Responsive Gene Regulation in Algae. *Plant Physiology*, *165*(1): 388–397.
- Hernández-Montiel, L. G., Rueda-Puente, E. O., Cordoba-Matson, M. V., Holguín-Peña, J. R., and Zulueta-Rodríguez, R. (2013). Mutualistic interaction of rhizobacteria with arbuscular mycorrhizal fungi and its antagonistic effect on *Fusarium oxysporum* in Carica papaya seedlings. *Crop Protection*, 47: 61–66.

- Ho, C.L., and Tan, Y.C. (2014). Molecular defense response of oil palm to *Ganoderma* infection. *Phytochemistry*, 114: 168-177.
- Hoppenau, C. E., Tran, V. T., Kusch, H., Aßhauer, K. P., Landesfeind, M., Meinicke, P., and Braus, G. H. (2014). *Verticillium dahliae* VdTHI4, involved in thiazole biosynthesis, stress response and DNA repair functions, is required for vascular disease induction in tomato. *Environmental and Experimental Botany*, 108: 14–22.
- Hushiarian, R., Yusof, N. A., and Dutse, S. W. (2013). Detection and control of *Ganoderma boninense*: strategies and perspectives. *SpringerPlus*, 2: 555.
- Idris, A.S., Noor Haida, S and Nur Rashyeda, R. (2010). GanoEF1- A fungal biocontrol agent for *Ganoderma* in oil palm. Bangi: MPOB Information Series: 501.
- Idris, A. S., Ahmad Kushari, D., Nur Rashyeda, R., Madihah, A. Z., Haida, N., and Sebran. (2013). Compositions for controlling Ganoderma disease in plants and method thereof by using endophytic fungus, *Hendersonia* GanoEF1. US Patent No. WO2015170961.
- Izzati. N.A. and Abdullah. F. (2008). Disease suppression in *Ganoderma* infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*, 44: 101-107.
- Jeun, Y., Park, K., Kim, C., Fowler, W., and Kloepper, J. (2004). Cytological observations of cucumber plants during induced resistance elicited by rhizobacteria. *Biological Control*, 29(1): 34–42.
- Johansson, T., Le Quéré, A., Ahren, D., Söderström, B., Erlandsson, R., Lundeberg, J., et al. (2004). Transcriptional responses of *Paxillus involutus* and *Betula pendula* during formation of ectomycorrhizal root tissue. *Molecular Plant and Microbe Interaction*, 17: 202–15.
- Kamarudin, A. N., Song, L. K., Idris A.S., Lamasudin, D. U and Balia Yusof, Z. N. (2017). Enhancement of thiamine biosynthesis in oil palm seedlings by colonization of endophytic fungus *Hendersonia toruloidea*. *Frontiers in Plant Science*, 8: 1799.
- Kloepper, J. W., McInroy, J. A., Liu, K., and Hu, C. H. (2013). Symptoms of fern distortion syndrome resulting from inoculation with opportunistic endophytic fluorescent *Pseudomonas* spp. *Plos One*, 8(3): e58531
- Koskimäki, J. J., Hokkanen, J., Jaakola, L., Suorsa, M., Tolonen, A., Mattila, S., and Hohtola, A. (2009). Flavonoid biosynthesis and degradation play a role in early defence responses of bilberry (*Vaccinium myrtillus*) against biotic stress. *European Journal of Plant Pathology*, 125(4): 629–640.
- Khan, A., Bassett, S., Voisey, C., Gaborit, C., Johnson, L., Christensen, M., et al. (2010). Gene expression profiling of the endophytic fungus Neotyphodium

- lolii in association with its host plant perennial ryegrass. *Australasian Plant Patholology*, 39: 467.
- Kozyrovska, N. O. (2013). Crosstalk between endophytes and a plant host within information processing networks. *Biopolymers and Cell*, 29(3): 234–243.
- Kumara, P. M., Shweta, S., Vasanthakumari, M. M., Sachin, N., Manjunatha, B. L., Jadhav, S. S., and Ganeshaiah, K. N. (2014). Endophytes and Plant Secondary Metabolites Synthesis: Molecular and Evolutionary Perspective. In *Advances in Endophytic Research*, (pp.177–190). Springer India.
- Larriba, E., Jaime, M. D. L. A., Nislow, C., Martín-Nieto, J., and Lopez-Llorca, L. V. (2015). Endophytic colonization of barley (*Hordeum vulgare*) roots by the nematophagous fungus *Pochonia chlamydosporia* reveals plant growth promotion and a general defense and stress transcriptomic response. *Journal of Plant Research*, 128: 665–678.
- Lee, S., Whitaker, V. M., and Hutton, S. F. (2016). Mini Review: Potential Applications of Non-host Resistance for Crop Improvement, *Frontiers in Plant Science*, 7: 997.
- Li, M., Petteys, B. J., McClure, J. M., Valsakumar, V., Bekiranov, S., Frank, E. L., and Smith, J. S. (2010). Thiamine biosynthesis in Saccharomyces cerevisiae is regulated by the NAD+-dependent histone deacetylase Hst1. *Molecular and Cellular Biology*, 30(13): 3329–41.
- Liddicoat, C., Hucker, B., Liang, H., and Vriesekoop, F. (2015). Thiamin analysis in red wine by fluorescence reverse phase-HPLC. *Food Chemistry*, 177: 325–329.
- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and. *Methods*, 25: 402–408.
- Lopez-Llorca, L. V., Bordallo, J. J., Salinas, J., Monfort, E., and López-Serna, M. L. (2002). Use of light and scanning electron microscopy to examine colonisation of barley rhizosphere by the nematophagous fungus *Verticillium chlamydosporium*. *Micron*, *33*(1): 61–67.
- Machado, C. R., Praekelt, U. M., de Oliveira, R. C., Barbosa, a C., Byrne, K. L., Meacock, P. A., and Menck, C. F. (1997). Dual role for the yeast THI4 gene in thiamine biosynthesis and DNA damage tolerance. *Journal of Molecular Biology*, 273(1): 114–21.
- Malaysian Palm Oil Board, Economic & Industry Development Division (MPOB). (2015, February). Overview of the Malaysian Oil Palm Industry. Retrieved from
 - http://bepi.mpob.gov.my/images/overview/Overview_of_Industry_2015.pdf.

- Malaysian National News Agency. (2014, May 24). Oil palm industry to contribute RM200bil to Malaysia's export value. *The Star*. Retrieved from http://www.thestar.com.my/business/business-news/2014/05/21/oil-palm-musa/
- Mohammed, C. L., Rimbawanto, A., and Page, D. E. (2014). Management of basidiomycete root- and stem-rot diseases in oil palm, rubber and tropical hardwood plantation crops. *Forest Pathology*, 44(6): 428-446
- Mimura, M., Zallot, R., Niehaus, T. D., Hasnain, G., Gidda, S. K., and Nguyen, T. N. (2016). *Arabidopsis* TH2 encodes the orphan enzyme thiamin monophosphate phosphatase'. *The Plant Cell*, 28(10): 2683-2696
- Monaim, M. F. A. (2011). Role of riboflavin and thiamine in induced resistance against charcoal rot disease of soybean. *African Journal of Biotechnology*, 10(53): 10842–10855.
- Nongbri, P. L., Vahabi, K., Mrozinska, A., Seebald, E., Sun, C., Sherameti, I., et al. (2013). Balancing defense and growth—Analyses of the beneficial symbiosis between *Piriformospora indica* and *Arabidopsis thaliana*. *Symbiosis*, 58: 17–28.
- Nurrashyeda, R., Idris, A.S., and Ramle, M. (2011) Viability test of alginate granular formulation of *Hendersonia* GanoEF1 against *Ganoderma boninense in vitro*. In: Proceedings of the Third International Seminar Integrated Oil Palm Pests and Management. MPOB, Malaysia. p. 111-115.
- Nurrashyeda, R and Idris, A.S. (2013). GanoEF1 Biofertiliser Colonization of *Hendersonia* GanoEF1 in oil palm roots. In: Proceedings of the Fifth International Seminar: Sustainable Management of Pests and Diseases in Oil Palm- The Way Forward. MPOB, Malaysia.
- Paerl, R. W., Bertrand, E. M., Allen, A. E., Palenik, B., and Azam, F. (2015). Vitamin B1 ecophysiology of marine picoeukaryotic algae: Strain-specific differences and a new role for bacteria in vitamin cycling. *Limnology and Oceanography*, 60(1): 215–228.
- Palacios, O. A., Bashan, Y., and de-Bashan, L. E. (2014). Proven and potential involvement of vitamins in interactions of plants with plant growth-promoting bacteria—an overview. *Biology and Fertility of Soils*, 50(3): 415–432.
- Palanisamy, K., Ansari, S. A., Kumar, P., and Gupta, B. N. (1998). Adventitious rooting in shoot cuttings of *Azadirachta indica* and *Pongamia pinnata*. *New Forests*, 16(1): 81–88.
- Paterson, R. (2007). *Ganoderma* disease of oil palm -a white rot perspective necessary for integrated control. *Crop Protection*, 26: 1369–1376.

- Pavlo, A., Leonid, O., Iryna, Z., Natalia, K., and Anna, P. (2011). Endophytic bacteria enhancing growth and disease resistance of potato (*Solanum tuberosum L.*). *Biological Control*, 56(1): 43–49.
- Pinto, E., Pedersén, M., Snoeijs, P., Van Nieuwerburgh, L., and Colepicolo, P. (2002). Simultaneous detection of thiamine and its phosphate esters from microalgae by HPLC. *Biochemical and Biophysical Research Communications*, 291(2): 344–348.
- Podolich, O., Ardanov, P., Zaets, I., Pirttilä, A. M., and Kozyrovska, N. (2014). Reviving of the endophytic bacterial community as a putative mechanism of plant resistance. *Plant and Soil*, 388(1): 367-377
- Pourcel, L., Moulin, M., and Fitzpatrick, T. B. (2013). Examining strategies to facilitate vitamin B1 biofortification of plants by genetic engineering. *Frontiers in Plant Science*, 4: 160.
- Rapala-Kozik, M., Kowalska, E., and Ostrowska, K. (2008). Modulation of thiamine metabolism in *Zea mays* seedlings under conditions of abiotic stress. *Journal of Experimental Botany*, 59(15): 4133–4143
- Rapala-Kozik, M., Wolak, N., Kujda, M., and Banas, A. K. (2012). The upregulation of thiamine (vitamin B1) biosynthesis in *Arabidopsis thaliana* seedlings under salt and osmotic stress conditions is mediated by abscisic acid at the early stages of this stress response. *BMC Plant Biology*, 12: 2.
- Rees, R. W., Flood, J., Hasan, Y., Potter, U., and Cooper, R. M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*); mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology*, *58*: 982–989.
- Saikkonen, K., Wäli, P., Helander, M., and Faeth, S. H. (2004). Evolution of endophyte-plant symbioses. *Trends in Plant Science*, 9: 275–280.
- Sapak, Z., Meon, S., and Ahmad, Z. A. M. (2008). Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agriculture and Biology*, 10(2): 127–132.
- Salter, M. G., and Conlon, H. E. (2007). Extraction of plant RNA. *Methods in Molecular Biology*, 362: 309–314.
- Seerangan, K., and Thangavelu, M. (2014). Arbuscular Mycorrhizal and Dark Septate Endophyte Fungal Associations in South Indian Aquatic and Wetland Macrophytes. *Journal of Botany*, 2014: 1–14.
- Shoresh, M., Yedidia, I., and Chet, I. (2005). Involvement of Jasmonic Acid/Ethylene Signaling Pathway in the Systemic Resistance Induced in Cucumber by *Trichoderma asperellum* T203. *Phytopathology*, 95(1): 76–84.
- Spiering, M. J., Greer, D. H., and Schmid, J. (2006). Effects of the fungal endophyte, Neotyphodium lolii, on net photosynthesis and growth rates of perennial

- ryegrass (*Lolium perenne*) are independent of in planta endophyte concentration. *Annals of Botany*, 98: 379–387.
- Su, Z.Z., Mao, L.J., Li, N., Feng, X-X., Yuan, Z.L., Wang, L.W., and Zhang, C.L. (2013). Evidence for biotrophic lifestyle and biocontrol potential of dark septate endophyte *Harpophora oryzae* to rice blast disease. *PloS One*, 8(4): e61332
- Sundram, S. (2013). First report: Isolation of endophytic *Trichoderma* from oil palm (*Elaeis guineensis Jacq*.) and their *in vitro* antagonistic assessment on *Ganoderma boninense*. *Journal of Oil Palm Research*, 25(3): 368–372.
- Tarkka, M. T., Lehr, N., Hampp, R., and Schrey, S. D. (2008). Plant behavior upon contact with Streptomycetes. *Plant signaling & behavior*, *3*(11): 917–919.
- Torres, M. S., White, J. F., Zhang, X., Hinton, D. M., and Bacon, C. W. (2012). Endophyte-mediated adjustments in host morphology and physiology and effects on host fitness traits in grasses. *Fungal Ecology*, 5(3): 322–330.
- Tunc-Ozdemir, M., Miller, G., Song, L., Kim, J., Sodek, A., Koussevitzky, S., and Shintani, D. (2009). Thiamin confers enhanced tolerance to oxidative stress in *Arabidopsis. Plant Physiology*, 151(1): 421–32.
- Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., and Speleman, F. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology*, *3*(7): research0034.1–0034.11
- Waller, F., Achatz, B., Baltruschat, H., Fodor, J., Becker, K., Fischer, M., and Kogel, K.-H. (2005). The endophytic fungus *Piriformospora indica* reprograms barley to salt-stress tolerance, disease resistance, and higher yield. *Proceedings of the National Academy of Sciences of the United States of America*, 102(38): 13386–13391.
- Wang, G., Ding, X., Yuan, M., Qiu, D., Li, X., Xu, C., and Wang, S. (2006). Dual function of rice OsDR8 gene in disease resistance and thiamine accumulation. *Plant Molecular Biology*, 60(3): 437–449.
- Wang, X., Anzhi, R., and Yubao, G. (2014). Effect of endophyte infection on fungal disease resistance of *Leymus chinensis*. *Acta Ecol. Sin.* 34, 6789–6796. doi:10.5846/stxb201303080377.
- Wang, L., Ye, Liu, H., Liu, X., Wei, C., Huang, Y., and Tu, J. (2016). Both overexpression and suppression of an *Oryza sativa* NB-LRR-like gene OsLSR result in autoactivation of immune response and thiamine accumulation. *Scientific Reports*, 6(866): 24079.
- Waqas, M., Khan, A. L., Hamayun, M., Shahzad, R., Kim, Y.-H., Choi, K.-S., and Lee, I.-J. (2015). Endophytic infection alleviates biotic stress in sunflower through

- regulation of defence hormones, antioxidants and functional amino acids. *European Journal of Plant Pathology*, 141(4), 803–824.
- Wilson, D. (1995). Endophyte: The Evolution of a Term, and Clarification of Its Use and Definition. *Oikos*, 73(2), 274–276.
- Wong, L., Bong, C.H., and Idris, A. S. (2012). *Ganoderma* Species Associated with Basal Stem Rot Disease of Oil Palm. *American Journal of Applied Sciences*, 9(6), 879–885.
- Wong, S. Y., Abdul Aziz, S. D., and Balia Yusof, Z. N. (2016). Osmotic stress upregulates the transcription of thiamine (vitamin B1) biosynthesis genes (THIC and THI4) in oil palm (*Elaies guineensis*). *African Journal of Biotechnology*, 15(29): 1566–1574.
- Woodward, J. B., Abeydeera, N. D., Paul, D., Phillips, K., Rapala-Kozik, M., Freeling, M., and Scanlon, M. J. (2010). A maize thiamine auxotroph is defective in shoot meristem maintenance. *The Plant Cell*, 22(10): 3305–3317.
- Zeng, Y., and Yang, T. (2002). RNA isolation from highly viscous samples rich in polyphenols and polysaccharides. *Plant Molecular Biology Reporter*, 20(4): 417.
- Zheng, Y. K., Qiao, X.-G., Miao, C.P., Liu, K., Chen, Y.W., Xu, L. H., and Zhao, L.-X. (2015). Diversity, distribution and biotechnological potential of endophytic fungi. *Annals of Microbiology*, 56: 529–542.
- Zhou, J., Sun, A., and Xing, D. (2013). Modulation of cellular redox status by thiamine-activated NADPH oxidase confers *Arabidopsis* resistance to *Sclerotinia sclerotiorum*. *Journal of Experimental Botany*, 64(11): 3261–3272.