



UNIVERSITI PUTRA MALAYSIA

CELLULASE PRODUCTION BY THERMOPHILIC *Bacillus licheniformis* 2D55 AND ITS APPLICATION FOR RECOVERY OF FERMENTABLE SUGAR FROM RICE HUSK

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By

KAZEEM MUINAT OLANIKE

**Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in
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January 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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January 2018

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Faculty : Biotechnology and Biomolecular Sciences

Fermentable sugar production from various agro-waste biomass via enzymatic saccharification using mesophilic fungi cellulase has been the issue of interest for many researchers. However, research carried out using thermophilic bacterial cellulase for sugar production are limited. Cellulases, chemical pretreatment and enzymatic saccharification required for sugar production are very costly and often result in low sugar yield, thereby affecting the overall economics of the bioconversion process. Hence, the main aim of this study was to isolate, characterize and enhance the production of cellulolytic enzyme complex (CMCase, FPase and β -glucosidase) and Xylanase from agro-waste mixture (a mixture of untreated sugarcane bagasse and pretreated rice husk) by a locally isolated thermophilic *Bacillus licheniformis* 2D55 and utilize the crude thermostable cellulase enzyme for sugar production through enzymatic saccharification of rice husk subjected to high pressure steam pretreatment.

Cellulose degrading bacterium isolated from oil palm empty fruit bunch-chicken manure compost was identified by morphological, biochemical and 16S rRNA test and known as *B. licheniformis* 2D55. In basal medium with microcrystalline cellulose as carbon source, the bacterium produced cellulase at 50°C within 18-24 h. Among the various untreated and sodium hydroxide (NaOH) pretreated agro-waste biomass used, agro-waste mixture comprising of untreated sugarcane bagasse and pretreated rice husk results in overall improvement in the cellulolytic enzyme complex with CMCase at 0.37 U/mL, FPase at 0.29 U/mL, β -glucosidase at 0.006 U/mL and xylanase at 0.98 U/mL. The SEM image reveal deformity in bacteria cell grown on NaOH pretreated sugarcane bagasse only, which resulted in the low performance of the bacteria for cellulase production on the substrate.

Effect of nutritional and physicochemical factors were investigated for enhancing cellulase production. The CMCase, FPase, and β -glucosidase activities increased by 77.4-folds, 44.3-folds, and 10-folds, respectively. The crude enzyme was highly active and stable over broad temperature (50 to 80°C) and pH (3.5 to 10.0) ranges with optimum temperature at 65°C and 80°C for CMCase and FPase, respectively. While the optimum pH for CMCase and FPase was 7.5 and 6.0, respectively.

An operational condition was developed for high pressure steam pretreatment (HPSP) of rice husk for fermentable sugar recovery through enzymatic saccharification. The pretreatment at 200°C/1.85 MPa for 7 min was found to effectively enhance cellulose content of the rice husk. The scanning electron micrograph (SEM), fourier transform infrared (FTIR) and x-ray diffraction (XRD) analysis expressed effectiveness of the pretreatment. A two-step saccharification of pretreated rice husk at 60°C yielded reducing sugar at 0.581 g/g substrate that was equivalent to 73.5% saccharification.

Therefore, the strain *B. licheniformis* 2D55 has the potential of utilizing agro-waste mixture for higher cellulolytic enzyme production. The thermostable nature of the cellulase contributed to the efficient release of fermentable sugars at higher temperature. The application of high pressure steam pretreatment and two-step enzymatic saccharification provides greener technology and improve the fermentable sugar production. The results of this study could contribute to future research in thermophilic bioprocessing rice husk with the application of thermostable cellulase producing bacterium for improving biomass-sugar conversion.

Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGHASILAN SELULASE OLEH *Bacillus licheniformis* 2D55
TERMOFILIK DAN APLIKASINYA UNTUK PEMEROLEHAN GULA
FERMENTASI DARIPADA JERAMI PADI**

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Penghasilan gula fermentasi daripada pelbagai sisa biomass pertanian melalui pensakaridaan enzim menggunakan kulat selulase mesofilik menjadi isu yang diminati oleh ramai penyelidik. Walaubagaimanapun, kajian menggunakan bakteria selulase termofilik untuk penghasilan gula sangat terhad. Enzim selulase, pra-rawatan kimia dan pensakaridaan enzim yang diperlukan untuk penghasilan gula fermentasi memerlukan kos yang tinggi dan kerap menghasilkan gula yang rendah serta memberi kesan ekonomi keseluruhan kepada proses penukaran. Jerami padi merupakan substrat penghasilan gula yang kurang mendapat perhatian penyelidikan. Oleh itu, kajian ini bertujuan untuk memencil, mencari dan mengoptimum penghasilan kompleks enzim selulase (CMCase, FPase dan β -glucosidase) dan xylanase daripada campuran sisa pertanian (hampas tebu dan jerami padi terawat) oleh bakteria termofilik tempatan yang dipencilkan, *Bacillus licheniformis* 2D55 dan menggunakan enzim selulase termostabil mentah untuk penghasilan gula fermentasi melalui enzim pensakaridaan oleh jerami padi yang dirawat pada wap tekanan tinggi.

Bakteria pengurai selulosa yang dipencilkan daripada kompos tandan sawit kosong dan tinja ayam dikenalpasti melalui ujian morfologi, biokimia dan 16s rRNA dan dikenalpasti sebagai *Bacillus licheniformis* 2D55. Dalam media asas menggunakan mikrokristal selulosa sebagai sumber karbon, bakteria menghasilkan aktiviti CMCase dan FPase sebanyak 0.33 U/mL dan 0.09 U/mL, masing-masing pada suhu 50°C selama 18-24 jam. Di antara pelbagai sisa biomass pertanian tidak terawat dan menggunakan pra-rawatan natrium hidroksida, campuran sisa pertanian hampas tebu tidak terawat dan jerami padi terawat menunjukkan peningkatan keseluruhan penghasilan kompleks enzim selulase dengan CMCase sebanyak 0.37 U/mL, FPase (0.29 U/mL) β -glucosidase (0.006 U/mL) and xylanase (0.98 U/mL). Imej mikroskop

elektron menunjukkan pertumbuhan bakteri terencat di atas hampas tebu terawat oleh natrium hidroksida yang menyebabkan kurang prestasi bakteri untuk menghasilkan selulase daripada substrat.

Kesan faktor nutrisi dan fizikokimia dinilai untuk meningkatkan penghasilan enzim selulase. Penghasilan aktiviti CMCase, FPase, dan β -glukosidase meningkat 77.4-kali ganda, 44.3-kali ganda, and 10-kali ganda, masing-masing, menggunakan parameter optimum. Enzim mentah sangat aktif dan stabil pada julat suhu (50 ke 80°C) dan pH (3.5 ke 10) dengan suhu optima CMCase dan FPase ialah 65°C dan 80°C, masing-masing. Manakala, pH optima untuk CMCase dan FPase ialah 7.5 dan 6.0, masing-masing.

Kondisi pengoperasian pra-rawatan wap tekanan tinggi (PSTT) jerami padi dibuat untuk pemerolehan gula fermentasi melalui pensakaridaan enzim. Satu keadaan operasi telah dibangunkan untuk pra-rawatan wap tekanan tinggi terhadap sekam padi untuk pemerolehan gula fermentasi melalui sakarifikasi enzim. Pra-rawatan pada 200C/1.85MPa selama 7 min telah didapati berkesan menggalakkan kandungan selulosa sekam padi. Analisis scanning electron micrograph (SEM), fourier transform infrared (FTIR) dan x-ray diffraction (XRD) menunjukkan keberkesanan pra-rawatan tersebut. Dua langkah sakarifikasi pra-rawatan sekam padi pada 60°C menghasilkan gula penurun pada 0.581 g/g substrat yang bersamaan 73.5% sakarifikasi.

Oleh itu, *Bacillus licheniformis* 2D55 berpotensi menggunakan campuran biomas sisa pertanian untuk penghasilan enzim selulolitik yang lebih tinggi. Keadaan termostabil selulase membantu kecekapan penghasilan gula fermentasi pada suhu yang lebih tinggi. Penggunaan pra-rawatan wap tekanan tinggi dan dua langkah pensakaridaan enzim menawarkan teknologi hijau dan meningkatkan penghasilan gula fermentasi. Hasil dari kajian ini menyumbang kepada kajian akan datang dalam bioproses termofilik jerami padi oleh bakteria selulase termostabil untuk meningkatkan penukaran biomas kepada gula fermentasi.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

β	Beta
BLAST	Basic Local Alignment Search Tool
BLASTN	Nucleotide-nucleotide BLAST
BSA	Bovine serum albumin
CFU/ml	Colony –forming unit per millilitre
CMC	Carboxymethyl cellulose
CMCase	Carboxymethyl cellulase
dBa	Decibel
DNA	deoxyribonucleic acid
DNS	3,5-dinitrosalicylic acid
dNTPs	Deoxynucleotides
EDX	Electron diffraction x-ray
FPase	Filter paperase
FPU/ml	Filter paper unit per millilitre
FTIR	Fourier transform infrared
g	Gram
g/L	Gram per litre
g/ml	Gram per millilitre
HCL	Hydrochloric acid
HPLC	High performance liquid chromatography
KH ₂ PO ₄	Potassium dihydrogen phosphate
K ₂ HPO ₄	Potassium hydrogen phosphate
KI	Potassium iodide

kg	Kilogram
L	Litre
MgSO ₄ .7H ₂ O	Magnesium sulphate
min	Minutes
NaH ₂ C ₆ H ₅ O ₇	Sodium citrate
NaOH	Sodium hydroxide
NCBI	National Centre for Biotechnology Information
(NH ₄) ₂ SO ₄	Ammonium sulphate
NO ₃ C ₆ H ₅ O ₇ .2H ₂ O	Sodium acetate
°C	Degree centigrade
OD	Optical density
PEG	Polyethylene glycol
PCR	Polymerase chain reaction
%	Percentage
rpm	Rotations per minute
SA	Specific activity
SEM	Scanning electron microscope
U/mL	unit per millilitre
w/v	Weight per volume
XRD	X-ray diffraction

CHAPTER 1

INTRODUCTION

Fuel has been a significant factor that shapes the economic life style of human for generations. The global increase in world population results in huge energy demand. Increase in fuel prices, shortage of fossil fuel and major concern over environmental pollution has directed research attention towards fermentable sugar (substrate for biofuel) production from food sources such as cereals, starch, sugar beets and sugarcane. Unlike biofuel, burning fossil such as gas, coal and oil results in the generation of CO₂ which has been a major contribution to global warming (Eriksson and Gray, 2017; Stips *et al.*, 2016). However, the unsustainability of sugar production from food crops due to increase in food prices has guided research toward production of sugars from agro-waste biomass. Sugar production from agro-waste biomass is renewable, sustainable, and ensures food security. Likewise agro-waste biomass is cheap, highly abundant and is the only alternative major raw material for fuels and chemical production at present and in future (Simpson-Holley *et al.*, 2007).

Various agro-waste biomass such as sugarcane bagasse, date palm biomass, rice straw, corn cob and wheat straw have been applied in the production of sugars (Alrumman, 2016; Gupta and Parkhey, 2014). However, less attention have been focused on rice husk as substrate for sugar production

Rice is ranked as largest cultivated cereal crop and a staple food for more than half of the world's population (Johar *et al.*, 2012). Rice consumption in Malaysia is expected to increase from 2.30 million metric tonnes to 2.69 million metric tonnes by 2020 (Shafie *et al.*, 2014). About 1.2 million tonnes of rice husk was generated in 2015 and the Malaysian government has projected 7 million tonnes per year paddy residue by the year 2020 (Shafie *et al.*, 2014). In addition, the growth population was expected to reach 41.50 million by 2040 from 28.60 million in 2010 (DOSM, 2016). Consequently, it was then projected that in order to meet the demand of the population growth and the self-sufficiency level (SSL) of 90 % by the year 2060, an additional 1,320,000 tons per year of rice must be produced (Siwar *et al.*, 2014). Thus, resulting in higher generation of rice husk. However, rice husk disposal system through open burning and burying causes environmental pollution with great health risk. Therefore, rice husk will be suitable raw material for sugar production due to its abundance, low-cost, low commercial value.

Pretreatment followed by enzymatic saccharification are the majorly required for the bioconversion of agro-waste to sugars. For several decades fungi cellulase from *Trichoderma* and *Aspergillus* species have been the central focus of both academic and industrial researchers (Howard *et al.*, 2003; Narra *et al.*, 2014; Rana *et al.*, 2014). Therefore, saccharification reactors are conducted at low temperatures because fungi cellulase is limited by low thermostability, high enzyme usage and prolong hydrolysis

thereby resulting in low hydrolysis and high cost of sugar production. However, processing of agro-waste biomass at elevated temperatures offers several advantages such as improved hydrolysis, low risk of potential contamination, better substrate solubility thus improving the overall economics of the process (Rastogi *et al.*, 2010). Hence, developing thermostable cellulase enzymes would be the best strategy for rapid release of sugars. But the lack of robust cellulase that can efficiently function at elevated temperature and broad pH range remains a bottle neck in thermophilic digestion of agro-waste biomass. Thus, thermophilic cellulase producing bacteria are better reservoir for thermostable enzymes with great potential for developing viable conversion of agro-waste biomass to sugars.

Quite few thermophilic cellulase degrading bacteria such as *Geobacillus* sp., *Brevibacillus* sp., *Bacillus* sp. C1, *Bacillus valismortis* and *Bacillus stearothermophilus* have been isolated from gold mine, cow dung, garden soil and forest soil (Assareh *et al.*, 2012; Gaur and Tiwari, 2015; Rastogi *et al.*, 2009; Sadhu *et al.*, 2013). However, the harsh conditions such as active degradation, extreme temperature, pH, pressure, inhibitors and toxic metal experienced during composting makes it a potential source of robust bacteria since the environment is similar to those of industrial processes. Furthermore, thermophilic bacteria are also advantageous due to high growth rate, high stability, versatility, genetic amiability, resistance to relatively harsh condition and production of cellulosome (Multi-enzyme complex) (Maki *et al.*, 2009; Maki *et al.*, 2011b).

It is worth to note that sensitivity analysis and techno-economic modelling has suggested that carbon source is a key factor in cellulase production and could account for about 50 % of total enzyme cost (Humbird *et al.*, 2011; Ryu and Mandels, 1980). Therefore, selecting appropriate carbon source is very important. Carbon substrates such as sugars; (*e.g.*, lactose, sucrose cellobiose) and pure cellulose (*e.g.*, caboxymethyl cellulose-CMC, avicel, microcrystalline-MCC cellulose and solka floc) have been reported as good inducers of cellulase synthesis (Jyotsna *et al.*, 2015; Sethi *et al.*, 2013). The utilization of these substrates on industrial scale is uneconomical due to the high cost of substrate. Likewise single substrate agro-waste like rice straw, sugarcane bagasse, date palm leaves, wheat straw, rice bran have been reported for carbon substrate (Alrumman, 2016; Gaur and Tiwari, 2015; Yang *et al.*, 2014). However, using single substrate might not be sustainable in real biorefinary condition due to shortage in feedstock as a result of seasonal variation (Ebadian *et al.*, 2011; Rentizelas *et al.*, 2009). In addition different agro-waste carbon substrate induces cellulolytic enzymes at various levels. Therefore, combining agro-waste as cocktail carbon feed stock could help mitigate this effect and are uncommonly reported.

The recalcitrant nature of lignocellulose restrict hydrolysis. Rice husk biomass is majorly consist of cellulose (32 – 47 %), hemicellulose (19-27 %) and lignin (5-24 %) (Binod *et al.*, 2010). In the structure of rice husk biomass, cellulose act as a linear crystalline polymer of β -D-glucose unit that is rigid. While lignin act as a cement that binds cellulose fibres to hemicellulose, hemicellulose links cellulose fibre and lignin, giving the whole cellulose-hemicellulose-lignin complex a rigid structure that makes

it more difficult and resistant to degradation (Chandra *et al.*, 2007; Hendriks and Zeeman, 2009; Palonen, 2004). Hence, pretreatment becomes necessary to disrupt the structural matrix of the lignocellulose by disintegration of cellulose fibres from hemicelluloses and lignin. Over the years, pretreatment methods have mainly focused on acid, alkali, ionic liquids and organosolvent chemicals (Ang *et al.*, 2012; Dagnino *et al.*, 2013; Omidvar *et al.*, 2016; Zhang *et al.*, 2016). However, this method present great limitation to agro-waste conversion due its unsustainability, high cost of chemicals and environmental toxicity. Therefore, alternative green, eco-friendly, inexpensive, and sustainable method of high pressure steam pretreatment is suggested. High pressure steam allows exposure of biomass to steam at high temperature (160-240°C), and pressure (0.7 and 4.8 MPa) at low residence time (2-20 min). Unlike chemical pretreatment, high pressure steam does not require extreme washing, neutralization and catalyst. Therefore, high pressure steam pretreatment represent great significance to this study. The application of high pressure steam have been reported for pretreatment of oil palm mesocarp fibre (OPMF) (Yunos *et al.*, 2012) and oil palm biomass (Baharuddin *et al.*, 2013; Baharuddin *et al.*, 2012). However, no research have been aimed at applying high pressure steam for the pretreatment of rice husk for sugar production.

Improving process condition for efficient enzymatic saccharification is of upmost importance towards improving sugar yield (Alvira *et al.*, 2010). Similarly, efficient and rapid enzymatic saccharification remains one of the technical and economical bottlenecks in the overall bioconversion of lignocellulosic biomass to fermentable sugars (Berlin *et al.*, 2007; Walker and Wilson, 1991; Zhang *et al.*, 2009). A variety of method have been suggested to increase hydrolysis yield of sugars including; gradual substrate loading (Rosgaard *et al.*, 2007), addition of surfactant (Börjesson *et al.*, 2007), immobilization (Tsai and Meyer, 2014; Zhang *et al.*, 2016) and enzyme recycling (Ouyang *et al.*, 2013). However, conceptualizing the idea that enzymes bound to substrate after hydrolysis could be recycled by recycling the recovered solids in subsequent steps lead to exploitation of this method for improving sugar production.

Therefore, in this study the feasibility of isolating local thermophilic bacterium that can utilize insoluble agro-waste biomass for the production of cellulolytic enzyme complex (CMCase, FPase, and β -glucosidase and xylanolytic enzymes (xylanase) was investigated. The study also examined enhancement of the crude enzyme production, characterization and subsequent fermentable sugar production using the crude enzyme via enzymatic saccharification studies.

Thus the objectives of this study are:

1. To isolate and characterize thermophilic cellulose-degrading bacteria for utilization of agro-waste biomass for cellulase production.
2. To enhance cellulase production by the isolated bacterium (*B. licheniformis* 2D55) through formulation of nutritional and physiological conditions using one factor at a time and characterize the endoglucanase and exoglucanase enzyme.
3. To develop efficient pretreatment of rice husk under high pressure steam and to improve the production of fermentable sugars through efficient enzymatic saccharification using crude cellulase produced by thermophilic *B. licheniformis* 2D55.



REFERENCES

- Aanniz, T., Ouadghiri, M., Melloul, M., Swings, J., Elfahime, E., Ibijbjen, J., et al. (2015). Thermophilic bacteria in Moroccan hot springs, salt marshes and desert soils. *Brazilian Journal of Microbiology*, 46(2): 443-453.
- Abdel-Rahman, M. A., Nour El-Din, M., Refaat, B. M., Abdel-Shakour, E. H., Ewais, E. E.-D. and Alrefaey, H. M. A. (2016). Biotechnological application of thermotolerant cellulose-decomposing bacteria in composting of rice straw. *Annals of Agricultural Sciences*, 61(1): 135-143.
- Abdullah, R., Zafar, W., Nadeem, M., Iqtedar, M., Kaleem, A. and Naz, S. (2016). Partial purification and characterization of cellulases produced by *Bacillus* Strain. *Romanian Biotechnological Letters*, 21(1): 11103-11109.
- Abou-Taleb, K. A., Mashhoor, W., Nasr, S. A., Sharaf, M. and Abdel-Azeem, H. H. (2009). Nutritional and environmental factors affecting cellulase production by two strains of cellulolytic Bacilli. *Australian Journal of Basic and Applied Sciences*, 3(3): 2429-2436.
- Abraham, E., Deepa, B., Pothan, L. A., Jacob, M., Thomas, S., Cvelbar, U., et al. (2011). Extraction of nanocellulose fibrils from lignocellulosic fibres: a novel approach. *Carbohydrate polymers*, 86(4): 1468-1475.
- Absar, N. (2009). Purification and characterization of intracellular cellulase from *Aspergillus oryzae* ITCC-4857.01. *Mycobiology*, 37(2): 121-127.
- Acharya, S. and Chaudhary, A. (2011). Effect of nutritional and environmental factors on cellulases activity by thermophilic bacteria isolated from hot spring. *Journal of Scientific and Industrial Research*, 70:142-148.
- Acharya, S. and Chaudhary, A. (2012a). Bioprospecting thermophiles for cellulase production: a review. *Brazilian Journal of Microbiology*, 43(3): 844-856.
- Acharya, S. and Chaudhary, A. (2012b). Optimization of fermentation conditions for cellulases production by *Bacillus licheniformis* MVS1 and *Bacillus* sp. MVS3 isolated from Indian hot spring. *Brazilian Archives of Biology and Technology*, 55(4): 497-503.
- Adhyaru, D. N., Bhatt, N. S. and Modi, H. (2014). Enhanced production of cellulase-free, thermo-alkali-solvent-stable xylanase from *Bacillus altitudinis* DHN8, its characterization and application in sorghum straw saccharification. *Biocatalysis and Agricultural Biotechnology*, 3(2): 182-190.

- Agarwal, T., Saxena, M. K. and Chandrawat, M. (2014). Production and optimization of cellulase enzyme by *Pseudomonas aeruginosa* MTCC 4643 using sawdust as a substrate. *International Journal of Scientific and Research Publications*, 4(1): 1-3.
- Agbor, V. B., Cicek, N., Sparling, R., Berlin, A. and Levin, D. B. (2011). Biomass pretreatment: fundamentals toward application. *Biotechnology Advances*, 29(6), 675-685.
- Akhtar, M. S., Saleem, M. and Akhtar, M. W. (2001). Saccharification of lignocellulosic materials by the cellulases of *Bacillus subtilis*. *International Journal of Agriculture and Biology*, 3: 199-202.
- Alemдар, A. and Sain, M. (2008). Isolation and characterization of nanofibers from agricultural residues—wheat straw and soy hulls. *Bioresource Technology*, 99(6): 1664-1671.
- Alizadeh, H., Teymouri, F., Gilbert, T. I. and Dale, B. E. (2005). Pretreatment of switchgrass by ammonia fiber explosion (AFEX). *Applied Biochemistry and Biotechnology*, 124(1-3): 1133-1141.
- Alrumman, S. A. (2016). Enzymatic saccharification and fermentation of cellulosic date palm wastes to glucose and lactic acid. *Brazilian Journal of Microbiology*, 47(1): 110-119.
- Alvira, P., Tomás-Pejó, E., Ballesteros, M. and Negro, M. J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. *Bioresource Technology*, 101(13): 4851-4861.
- Ang, S. K. and Abd-Aziz, S. (2015). Potential Uses of Xylanase-Rich Lignocellulolytic Enzymes Cocktail for Oil Palm Trunk (OPT) Degradation and Lignocellulosic Ethanol Production. *Energy & Fuels*, 29(8): 5103-5116.
- Ang, T. N., Ngoh, G. C. and Chua, A. S. M. (2013). Comparative study of various pretreatment reagents on rice husk and structural changes assessment of the optimized pretreated rice husk. *Bioresource Technology*, 135: 116-119.
- Ang, T. N., Ngoh, G. C., Chua, A. S. M. and Lee, M. G. (2012). Elucidation of the effect of ionic liquid pretreatment on rice husk via structural analyses. *Biotechnology for Biofuels*, 5(67): 1-12.
- Annamalai, N., Rajeswari, M. V. and Balasubramanian, T. (2014). Enzymatic saccharification of pretreated rice straw by cellulase produced from *Bacillus carboniphilus* CAS 3 utilizing lignocellulosic wastes through statistical optimization. *Biomass and Bioenergy*, 68: 151-160.

- Annamalai, N., Rajeswari, M. V., Elayaraja, S. and Balasubramanian, T. (2013). Thermostable, haloalkaline cellulase from *Bacillus halodurans* CAS 1 by conversion of lignocellulosic wastes. *Carbohydrate polymers*, 94(1): 409-415.
- Annamalai, N., Rajeswari, M. V., Elayaraja, S., Thavasi, R., Vijayalakshmi, S. and Balasubramanian, T. (2012). Purification and characterization of thermostable alkaline cellulase from marine bacterium *Bacillus licheniformis* AU01 by utilizing cellulosic wastes. *Waste and Biomass Valorization*, 3(3): 305-310.
- Annamalai, N., Thavasi, R., Vijayalakshmi, S. and Balasubramanian, T. (2011). A novel thermostable and halostable carboxymethylcellulase from marine bacterium *Bacillus licheniformis* AU01. *World Journal of Microbiology and Biotechnology*, 27(9): 2111-2115.
- Anwar, Z., Gulfraz, M. and Irshad, M. (2014). Agro-industrial lignocellulosic biomass a key to unlock the future bio-energy: a brief review. *Journal of Radiation Research and Applied Sciences*, 7(2): 163-173.
- Arantes, V. and Saddler, J. N. (2010). Access to cellulose limits the efficiency of enzymatic hydrolysis: the role of amorphogenesis. *Biotechnology for Biofuels*, 3(4): 1-11.
- Ariffin, H., Abdullah, N., Umi Kalsom, M., Shirai, Y. and Hassan, M. (2006). Production and characterization of cellulase by *Bacillus pumilus* EB3. *International Journal of Engeneering and Technology*, 3(1): 47-53.
- Ariffin, H., Hassan, M. A., Shah, U. K. M., Abdullah, N., Ghazali, F. M. and Shirai, Y. (2008). Production of bacterial endoglucanase from pretreated oil palm empty fruit bunch by *Bacillus pumilus* EB3. *Journal of Bioscience and Bioengineering*, 106(3): 231-236.
- Asha, B. and Sakthivel, N. (2014). Production, purification and characterization of a new cellulase from *Bacillus subtilis* that exhibit halophilic, alkalophilic and solvent-tolerant properties. *Annals of Microbiology*, 64(4): 1839-1848.
- Askarieh, M. M., Chambers, A. V., Daniel, F. B. D., FitzGerald, P. L., Holtom, G. J., Pilkington, N. J., et al. (2000). The chemical and microbial degradation of cellulose in the near field of a repository for radioactive wastes. *Waste Management*, 20(1):93-106.
- Asmare, B. (2014). Biotechnological Advances for Animal Nutrition and Feed Improvement. *World Journal of Agricultural Research*, 2(3): 115-118.
- Assareh, R., Shahbani Zahiri, H., Akbari Noghabi, K., Aminzadeh, S. and Bakhshi khaniki, G. (2012). Characterization of the newly isolated *Geobacillus* sp. T1, the efficient cellulase-producer on untreated barley and wheat straws. *Bioresource Technology*, 120: 99-105.

- Atykyan, N., Revin, V. and Lyovina, E. (2016). Optimization of the production of cellulases by fungi *Lentinus tigrinus* VKM F-3616D and *Trichoderma viride* VKM F-1131 using submerged fermentation. *Journal of Biotechnology*, 231: S12-S13.
- Aygan, A. and Arikan, B. (2008). A new halo-alkaliphilic, thermostable endoglucanase from moderately halophilic *Bacillus* sp. C14 isolated from van soda lake. *International Journal of Agriculture and Biology*, 10(4): 369-374.
- Aygan, A., Karcioglu, L. and Arikan, B. (2011). Alkaline thermostable and halophilic endoglucanase from *Bacillus licheniformis* C108. *African Journal of Biotechnology*, 10(5): 789-796.
- Azadian, F., Badoei-dalfard, A., Namaki-Shoushtari, A. and Hassanshahian, M. (2016). Purification and biochemical properties of a thermostable, haloalkaline cellulase from *Bacillus licheniformis* AMF-07 and its application for hydrolysis of different cellulosic substrates to bioethanol production. *Molecular Biology Research Communications*, 5(3):143-155.
- Azadian, F., Badoei-dalfard, A., Namaki-Shoushtari, A., Karami, Z. and Hassanshahian, M. (2017). Production and characterization of an acidothermophilic, organic solvent stable cellulase from *Bacillus sonorensis* HSC7 by conversion of lignocellulosic wastes. *Journal of Genetic Engineering and Biotechnology*, 15(1): 187-196.
- Baek, S. Y., Lee, Y. J., Yun, H. J., Park, H. Y. and Yeo, S.-H. (2014). Characterization of alkaline cellulase from *Bacillus subtilis* 4-1 isolated from Korean traditional soybean paste. *Korean Journal of Food Preservation*, 21(3): 442-450.
- Bagudo, A., Argungu, A., Aliero, A., Suleiman, N. and Kalpana, S. (2014). *Bacillus subtilis* as an alternative source of beta-glucosidase. *International Journal of Cell and Molecular Biology*, 3(1), 1-9.
- Baharuddin, A. S., Sulaiman, A., Kim, D. H., Mokhtar, M. N., Hassan, M. A., Wakisaka, M., et al. (2013). Selective component degradation of oil palm empty fruit bunches (OPEFB) using high-pressure steam. *Biomass and Bioenergy*, 55: 268-275.
- Baharuddin, A. S., Yunus, N. S. H. M., Mahmud, N. A. N., Zakaria, R. and Yunus, K. F. M. (2012). Effect of high-pressure steam treatment on enzymatic saccharification of oil palm empty fruit bunches. *BioResources*, 7(3): 3525-3538.
- Bahrin, E. K., Baharuddin, A. S., Ibrahim, M. F., Razak, M. N. A., Sulaiman, A., Aziz, S. A., et al. (2012). Physicochemical property changes and enzymatic hydrolysis enhancement of oil palm empty fruit bunches treated with superheated steam. *BioResources*, 7(2): 1784-1801.

- Bajaj, B. K. and Singh, N. P. (2010). Production of xylanase from an alkalitolerant *Streptomyces* sp. 7b under solid-state fermentation, its purification, and characterization. *Applied Biochemistry and Biotechnology*, 162(6): 1804-1818.
- Bals, B., Wedding, C., Balan, V., Sendich, E. and Dale, B. (2011). Evaluating the impact of ammonia fiber expansion (AFEX) pretreatment conditions on the cost of ethanol production. *Bioresource Technology*, 102(2): 1277-1283.
- Banerjee, S., Sen, R., Morone, A., Chakrabarti, T., Pandey, R. and Mudliar, S. (2012). Improved wet air oxidation pretreatment for enhanced enzymatic hydrolysis of rice husk for bioethanol production. *Dynamic Biochemistry Process Biotechnology and Molecular Biology*, 6(2): 43-45.
- Banerjee, S., Sen, R., Mudliar, S., Pandey, R. A., Chakrabarti, T. and Satpute, D. (2011). Alkaline peroxide assisted wet air oxidation pretreatment approach to enhance enzymatic convertibility of rice husk. *Biotechnology Progress*, 27(3): 691-697.
- Banerjee, S., Sen, R., Pandey, R. A., Chakrabarti, T., Satpute, D., Giri, B. S., et al. (2009). Evaluation of wet air oxidation as a pretreatment strategy for bioethanol production from rice husk and process optimization. *Biomass and Bioenergy*, 33(12): 1680-1686.
- Bansal, N., Tewari, R., Soni, R. and Soni, S. K. (2012). Production of cellulases from *Aspergillus niger* NS-2 in solid state fermentation on agricultural and kitchen waste residues. *Waste Management*, 32(7): 1341-1346.
- Banu, A. and Ingale, S. (2012). Xylanase production by *Bacillus pumilus* AB-1 under solid state fermentation and its application. *Bulletin of Environmental and Scientific Research*, 1(1), 35-43.
- Baramée, S., Phitsuwan, P., Waeonukul, R., Pason, P., Tachaapaikoon, C., Kosugi, A., et al. (2015). Alkaline xylanolytic–cellulolytic multienzyme complex from the novel anaerobic alkalithermophilic bacterium *Cellulosibacter alkalithermophilus* and its hydrolysis of insoluble polysaccharides under neutral and alkaline conditions. *Process Biochemistry*, 50(4): 643-650.
- Barana, D., Salanti, A., Orlandi, M., Ali, D. S. and Zoia, L. (2016). Biorefinery process for the simultaneous recovery of lignin, hemicelluloses, cellulose nanocrystals and silica from rice husk and *Arundo donax*. *Industrial Crops and Products*, 86: 31-39.
- Barati, B. and Amiri, I. S. (2015). Introduction of cellulase and its application In: in silico engineering of disulphide bonds to produce stable cellulase: *SpringerBriefs in Applied Sciences and Technology*. Springer, Singapore. pp (1-5)

- Barcelos, C. A., Maeda, R. N., Betancur, G. J. V. and Pereira Jr, N. (2013). The essentialness of delignification on enzymatic hydrolysis of sugar cane bagasse cellulignin for second generation ethanol production. *Waste and Biomass Valorization*, 4(2): 341-346.
- Barnette, A. L., Lee, C., Bradley, L. C., Schreiner, E. P., Park, Y. B., Shin, H., et al. (2012). Quantification of crystalline cellulose in lignocellulosic biomass using sum frequency generation (SFG) vibration spectroscopy and comparison with other analytical methods. *Carbohydrate Polymers*, 89(3): 802-809.
- Barrington, S., Kim, J. S., Wang, L. and Kim, J.-W. (2009). Optimization of citric acid production by *Aspergillus niger* NRRL 567 grown in a column bioreactor. *Korean Journal of Chemical Engineering*, 26(2): 422-427.
- Begum, F., Absar, N. and Alam, M. S. (2009). Purification and characterization of extracellular cellulase from *A. oryzae* ITCC-4857.01. *Journal of Applied Sciences Research*, 26: 1646-1651.
- Behera, B. C., Mishra, R. R., Singh, S. K., Dutta, S. K. and Thatoi, H. (2016). Cellulase from *Bacillus licheniformis* and *Brucella* sp. isolated from mangrove soils of Mahanadi river delta, Odisha, India. *Biocatalysis and Biotransformation*, 34(1), 44-53.
- Behera, B. C., Sethi, B. K., Mishra, R. R., Dutta, S. K. and Thatoi, H. N. (2017). Microbial cellulases – diversity and biotechnology with reference to mangrove environment: A review. *Journal of Genetic Engineering and Biotechnology*, 3(1): 197-210
- Behera, S., Arora, R., Nandhagopal, N. and Kumar, S. (2014). Importance of chemical pretreatment for bioconversion of lignocellulosic biomass. *Renewable and Sustainable Energy Reviews*, 36: 91-106.
- Behera, S. S. and Ray, R. C. (2016). Solid state fermentation for production of microbial cellulases: Recent advances and improvement strategies. *International journal of Biological Macromolecules*, 86: 656-669.
- Berlin, A., Maximenko, V., Gilkes, N. and Saddler, J. (2007). Optimization of enzyme complexes for lignocellulose hydrolysis. *Biotechnology and Bioengineering*, 97(2): 287-296.
- Bertero, M., de la Puente, G. and Sedran, U. (2012). Fuels from bio-oils: Bio-oil production from different residual sources, characterization and thermal conditioning. *Fuel*, 95: 263-271.
- Beukes, N. and Pletschke, B. I. (2006). Effect of sulfur-containing compounds on *Bacillus* cellulosome-associated 'CMCase' and 'Avicelase' activities. *FEMS Microbiology Letters*, 264(2): 226-231.

- Bhalla, A., Bansal, N., Kumar, S., Bischoff, K. M. and Sani, R. K. (2013). Improved lignocellulose conversion to biofuels with thermophilic bacteria and thermostable enzymes. *Bioresource Technology*, 128: 751-759.
- Bhalla, A., Bischoff, K. M. and Sani, R. K. (2015). Highly thermostable xylanase production from a thermophilic *Geobacillus* sp. strain WSUCF1 utilizing lignocellulosic biomass. *Frontiers in Bioengineering and Biotechnology*, 3 (84):1-8.
- Bhat, M. K. (2000). Cellulases and related enzymes in biotechnology. *Biotechnology Advances*, 18(5): 355-383.
- Binod, P., Satyanagalakshmi, K., Sindhu, R., Janu, K. U., Sukumaran, R. K. and Pandey, A. (2012). Short duration microwave assisted pretreatment enhances the enzymatic saccharification and fermentable sugar yield from sugarcane bagasse. *Renewable Energy*, 37(1): 109-116.
- Binod, P., Sindhu, R., Singhanian, R. R., Vikram, S., Devi, L., Nagalakshmi, S., et al. (2010). Bioethanol production from rice straw: an overview. *Bioresource Technology*, 101(13): 4767-4774.
- Bischoff, K. M., Rooney, A. P., Li, X.-L., Liu, S. and Hughes, S. R. (2006). Purification and characterization of a family 5 endoglucanase from a moderately thermophilic strain of *Bacillus licheniformis* *Biotechnology Letters*, 28(21): 1761-1765.
- Bisswanger, H. (2014). Enzyme assays. *Perspectives in Science*, 1(6): 41-55.
- Börjesson, M. and Westman, G. (2015). Crystalline nanocellulose—preparation, modification, and properties In: P. Matheus (Ed.), *Cellulose-Fundamental Aspects and Current Trends*. (pp. 1-35). InTech publishers
- Börjesson, J., Engqvist, M., Sipos, B. and Tjerneld, F. (2007). Effect of poly(ethylene glycol) on enzymatic hydrolysis and adsorption of cellulase enzymes to pretreated lignocellulose. *Enzyme and Microbial Technology*, 41(1): 186-195.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1-2), 248-254.
- Brandt, A., Gräsvik, J., Hallett, J. P., & Welton, T. (2013). Deconstruction of lignocellulosic biomass with ionic liquids. *Green Chemistry*, 15(3): 550-583.
- Brett, C. T. (2000). Cellulose microfibrils in plants: biosynthesis, deposition, and integration into the cell wall. *International Review of Cytology*, 199: 161-199.

- Brijwani, K. and Vadlani, P. V. (2011). Cellulolytic enzymes production via solid-state fermentation: effect of pretreatment methods on physicochemical characteristics of substrate. *Enzyme Research*, ID 860134 2011: 1-10.
- Brodeur, G., Yau, E., Badal, K., Collier, J., Ramachandran, K. B. and Ramakrishnan, S. (2011). Chemical and physicochemical pretreatment of lignocellulosic biomass: a review. *Enzyme Research*, ID 787532, 2011: 1-17
- Brummer, V., Skryja, P., Jurena, T., Hlavacek, V. and Stehlik, P. (2014). Suitable technological conditions for enzymatic hydrolysis of waste paper by Novozymes® Enzymes NS50013 and NS50010. *Applied Biochemistry and Biotechnology*, 174(4): 1299-1308.
- Budihal, S. R., Agsar, D. and Patil, S. R. (2016). Enhanced production and application of acidothermophilic *Streptomyces* cellulase. *Bioresource Technology*, 200: 706-712.
- Calvo- Flores, F. G. and Dobado, J. A. (2010). Lignin as renewable raw material. *Chemistry and Sustainability Energy and Materials*, 3(11): 1227-1235.
- Cao, Y. and Tan, H. (2004). Structural characterization of cellulose with enzymatic treatment. *Journal of Molecular Structure*, 705(1): 189-193.
- Casas López, J., Sánchez Pérez, J., Fernández Sevilla, J., Acien Fernández, F., Molina Grima, E. and Chisti, Y. (2004). Fermentation optimization for the production of lovastatin by *Aspergillus terreus*: use of response surface methodology. *Journal of Chemical Technology and Biotechnology*, 79(10): 1119-1126.
- Chakdar, H., Kumar, M., Pandiyan, K., Singh, A., Nanjappan, K., Kashyap, P. L., et al. (2016). Bacterial xylanases: biology to biotechnology. *3 Biotech*, 6(2): 150.
- Chandra, R. P., Bura, R., Mabee, W., Berlin, d. A., Pan, X. and Saddler, J. (2007). Substrate pretreatment: The key to effective enzymatic hydrolysis of lignocellulosics In: Olsson L. (eds) *Biofuels*. Advances in biochemical engineering/biotechnology, Springer, Berlin, Heidelberg *Biofuels* 108:67-93.
- Chua, M. G. S. and Wayman, M. (1979). Characterization of autohydrolysis aspen (*P. tremuloides*) lignins. Part 1. Composition and molecular weight distribution of extracted autohydrolysis lignin. *Canadian Journal of Chemistry*, 57(10): 1141-1149.
- Chundawat, S. P. S., Beckham, G. T., Himmel, M. E. and Dale, B. E. (2011). Deconstruction of lignocellulosic biomass to fuels and chemicals. *Annual review of chemical and biomolecular engineering*, 2: 121-145.
- Coates, J. (2000). Interpretation of infrared spectra, a practical approach. *Encyclopedia of Analytical Chemistry*. John Wiley and Son Ltd.

- Collins, T., Gerday, C. and Feller, G. (2005). Xylanases, xylanase families and extremophilic xylanases. *FEMS Microbiology Reviews*, 29(1): 3-23.
- Cruys-Bagger, N., Tatsumi, H., Ren, G. R., Borch, K. and Westh, P. (2013). Transient kinetics and rate-limiting steps for the processive cellobiohydrolase Cel7A: effects of substrate structure and carbohydrate binding domain. *Biochemistry*, 52(49): 8938-8948.
- Da Vinha, F. N. M., Gravina-Oliveira, M. P., Franco, M. N., Macrae, A., da Silva Bon, E. P., Nascimento, R. P., et al. (2011). Cellulase production by *Streptomyces viridobrunneus* SCPE-09 using lignocellulosic biomass as inducer substrate. *Applied Biochemistry and Biotechnology*, 164(3): 256-267.
- Dagnino, E. P., Chamorro, E. R., Romano, S. D., Felissia, F. E. and Area, M. C. (2013). Optimization of the acid pretreatment of rice hulls to obtain fermentable sugars for bioethanol production. *Industrial Crops and Products*, 42: 363-368.
- Das, A., Bhattacharya, S. and Murali, L. (2010). Production of cellulase from a thermophilic *Bacillus* sp. isolated from cow dung. *American-Eurasian Journal of Agriculture and Environmental Science*, 8(6): 685-691.
- Datta, S., Holmes, B., Park, J. I., Chen, Z., Dibble, D. C., Hadi, M., et al. (2010). Ionic liquid tolerant hyperthermophilic cellulases for biomass pretreatment and hydrolysis. *Green Chemistry*, 12(2): 338-345.
- Dave, B. R., Sudhir, A. P. and Subramanian, R. B. (2015). Purification and properties of an endoglucanase from *Thermoascus aurantiacus*. *Biotechnology Reports*, 6: 85-90.
- de Souza Lima, M. M. and Borsali, R. (2004). Rodlike cellulose microcrystals: structure, properties, and applications. *Macromolecular Rapid Communications*, 25(7): 771-787.
- de Souza Moreira, L. R., Álvares, A. d. C. M., da Silva Jr, F. G., de Freitas, S. M. and Ferreira Filho, E. X. (2015). Xylan-degrading enzymes from *Aspergillus terreus*: Physicochemical features and functional studies on hydrolysis of cellulose pulp. *Carbohydrate polymers*, 134, 700-708.
- Deka, D., Bhargavi, P., Sharma, A., Goyal, D., Jawed, M. and Goyal, A. (2011). Enhancement of cellulase activity from a new strain of *Bacillus subtilis* by medium optimization and analysis with various cellulosic substrates. *Enzyme research*, ID 15656 2011: 1-8.
- Deka, D., Das, S. P., Sahoo, N., Das, D., Jawed, M., Goyal, D., et al. (2013). Enhanced cellulase production from *Bacillus subtilis* by optimizing physical parameters for bioethanol production. *ISRN Biotechnology*, ID 965310 2013: 1-11.

- Deka, D., Jawed, M. and Goyal, A. (2013). Purification and characterization of an alkaline cellulase produced by *Bacillus subtilis* (AS3). *Preparative Biochemistry and Biotechnology*, 43(3): 256-270.
- Demain, A. L., Newcomb, M. and Wu, J. H. D. (2005). Cellulase, clostridia, and ethanol. *Microbiology and Molecular Biology Reviews*, 69(1), 124-154.
- Deswal, D., Khasa, Y. P. and Kuhad, R. C. (2011). Optimization of cellulase production by a brown rot fungus *Fomitopsis* sp. RCK2010 under solid state fermentation. *Bioresource Technology*, 102(10): 6065-6072.
- Diaz, A., Le Toullec, J., Blandino, A., De Ory, I. and Caro, I. (2013). Pretreatment of rice hulls with alkaline peroxide to enhance enzyme hydrolysis for ethanol production. *Chemical Engineering Transactions*, 32: 949-954.
- Dissanayake, N. P. J. and Summerscales, J. (2013). Life cycle assessment for natural fibre composites. *Green Composites from Natural Resources*, 8: 157-186.
- DOE, U. S. (2008). US Department of Energy–Environmental justice strategy. Retrieved on 28 March 2018 from <https://www.energy.gov/nepa/downloads/environmental-justice-strategy-doe>–DOE Office of Science, Washington, DC.
- DOSM, (2016). Department of statistics Malaysia. Population projection (revised), Malaysia, 2010-2040. Retrieved on 28 March 2018 from https://www.dosm.gov.my/v1/index.php?r=column/cthemByCat&cat=118&bul_id=Y3kwU2tSNVFDOWp1YmtZYnhUeVBEdz09&menu_id=L0pheU43NWJwRWVSZklWdzQ4TlhUUT09
- Dover, R. S., Bitler, A., Shimoni, E., Trieu-Cuot, P. and Shai, Y. (2015). Multiparametric AFM reveals turgor-responsive net-like peptidoglycan architecture in live *Streptococci*. *Nature communications*, ID7193 6:1-10
- Dutta, S., De, S., Saha, B. and Alam, M. I. (2012). Advances in conversion of hemicellulosic biomass to furfural and upgrading to biofuels. *Catalysis Science and Technology*, 2(10), 2025-2036.
- Dutta, T., Sengupta, R., Sahoo, R., Sinha Ray, S., Bhattacharjee, A. and Ghosh, S. (2007). A novel cellulase free alkaliphilic xylanase from alkali tolerant *Penicillium citrinum*: production, purification and characterization. *Letters in Applied Microbiology*, 44(2): 206-211.
- Ebadian, M., Sowlati, T., Sokhansanj, S., Stumborg, M. and Townley-Smith, L. (2011). A new simulation model for multi-agricultural biomass logistics system in bioenergy production. *Biosystems Engineering*, 110(3): 280-290.
- Eggstein, M., Kreutz, F. H. and Bailey, J. L. (1967). Techniques in Protein Chemistry: Elsevier, (Bailey, J. L. (Ed.) Elsevier Science Publishers, Amsterdam, 340.

- El-Metwally, M. M., El-Sharkawy, A. M., El-Morsy, A. A. and El-Dohlob, S. M. (2014). Statistical optimization of rapid production of cellulases from *Aspergillus niger* MA1 and its application in bioethanol production from rice hulls. *African Journal of Biotechnology*, 13(36): 3702-3714.
- Erdei, B., Barta, Z., Sipos, B., Réczey, K., Galbe, M. and Zacchi, G. (2010). Ethanol production from mixtures of wheat straw and wheat meal. *Biotechnology for Biofuels*, 3(16): 1-9
- Eriksson, E. L. V. and Gray, E. M. (2017). Optimization and integration of hybrid renewable energy hydrogen fuel cell energy systems – A critical review. *Applied Energy*, 202: 348-364.
- FAO, (2015). The state of food insecurity in the world. Meeting the 2015 international hunger targets: taking stock of uneven progress. Retrieved on 28 March 2018 from <http://www.fao.org/policy-support/resources/resources-details/en/c/469455/>
- Farinas, C. S. (2015). Developments in solid-state fermentation for the production of biomass-degrading enzymes for the bioenergy sector. *Renewable and Sustainable Energy Reviews*, 52: 179-188.
- Fatokun, E. N., Nwodo, U. U. and Okoh, A. I. (2016). Classical optimization of cellulase and xylanase production by a marine *Streptomyces* species. *Applied Sciences*, 6(10): 286.
- Florencio, C., Couri, S. and Farinas, C. S. (2012). Correlation between agar plate screening and solid-state fermentation for the prediction of cellulase production by *Trichoderma* strains. *Enzyme Research*, 2012, 1-7.
- Gadde, B., Bonnet, S., Menke, C. and Garivait, S. (2009). Air pollutant emissions from rice straw open field burning in India, Thailand and the Philippines. *Environmental Pollution*, 157(5): 1554-1558.
- Galbe, M. and Zacchi, G. (2007). Pretreatment of lignocellulosic materials for efficient bioethanol production. In: Olsson L. (eds) *Biofuels*. Advances in biochemical engineering and biotechnology. Springer, Berlin, Heidelberg 108: 41-65
- Gao, M., Li, B., Yu, A., Liang, F., Yang, L. and Sun, Y. (2010). The effect of aeration rate on forced-aeration composting of chicken manure and sawdust. *Bioresource Technology*, 101(6): 1899-1903.
- Gao, W., Kim, Y. J., Chung, C. H., Li, J. and Lee, J. W. (2010). Optimization of mineral salts in medium for enhanced production of pullulan by *Aureobasidium pullulans* HP-2001 using an orthogonal array method. *Biotechnology and Bioprocess Engineering*, 15(5): 837-845.

- Gao, Z., Mori, T. and Kondo, R. (2012). The pretreatment of corn stover with *Gloeophyllum trabeum* KU-41 for enzymatic hydrolysis. *Biotechnology for Biofuels*, 5 (28): 1-11
- Garg, R., Srivastava, R., Brahma, V., Verma, L., Karthikeyan, S. and Sahni, G. (2016). Biochemical and structural characterization of a novel halotolerant cellulase from soil metagenome. *Scientific reports*, 6:1-15
- Gaur, R. and Tiwari, S. (2015). Isolation, production, purification and characterization of an organic-solvent-thermostable alkalophilic cellulase from *Bacillus vallismortis* RG-07. *BMC Biotechnology*, 15 (19) 1-12.
- Gincy, M. M., Sukumaran, R. K., Singhanian, R. R. and Pandey, A. (2008). Progress in research on fungal cellulases for lignocellulose degradation. *Journal of Scientific and Industrial Research* 67(11): 898-907.
- Goering, H. K. and Van Soest, P. J. (1970). Forage fiber analysis. Agricultural handbook no. 379. *US Department of Agriculture*, Washington, DC, 1-20.
- Gomaa, E. Z. (2013). Optimization and characterization of alkaline protease and carboxymethyl-cellulase produced by *Bacillus pumilus* grown on *Ficus nitida* wastes. *Brazilian Journal of Microbiology*, 44(2): 529-537.
- Gowdhaman, D. and Ponnusami, V. (2015). Production and optimization of xylooligosaccharides from corncob by *Bacillus aerophilus* KGJ2 xylanase and its antioxidant potential. *International journal of Biological Macromolecules*, 79: 595-600.
- Goyal, V., Mittal, A., Bhuwal, A. K., Singh, G., Yadav, A. and Aggarwal, N. K. (2014). Parametric optimization of cultural conditions for carboxymethyl cellulase production using pretreated rice straw by *Bacillus* sp. 313SI under stationary and shaking conditions. *Biotechnology Research International*, 2014: 1-7.
- Guo, G. L., Chen, W. H., Chen, W. H., Men, L.C. and Hwang, W. S. (2008). Characterization of dilute acid pretreatment of silvergrass for ethanol production. *Bioresource Technology*, 99(14); 6046-6053.
- Gupta, P. and Parkhey, P. (2014). A two-step process for efficient enzymatic saccharification of rice straw. *Bioresource Technology*, 173: 207-215.
- Gupta, R. and Ramnani, P. (2006). Microbial keratinases and their prospective applications: an overview. *Applied Microbiology and Biotechnology*, 70(1): 21-33
- Gupta, M., Sharma, M., Singh, S., Gupta, P. and Bajaj, B. K. (2015). Enhanced production of cellulase from *Bacillus licheniformis* K- 3 with potential for saccharification of rice straw. *Energy Technology*, 3(3), 216-224.

- Gupta, V. G. (2016). *Aspergillus* system properties and applications In: New and future developments in microbial biotechnology and bioengineering, Radarweg, Amsterdam, Netherlands Elsevier, B.V. pp 215-222
- Gusakov, A. V. (2011). Alternatives to *Trichoderma reesei* in biofuel production. *Trends in Biotechnology*, 29(9): 419-425.
- Haldar, D., Sen, D. and Gayen, K. (2016). A Review on the production of fermentable sugars from lignocellulosic biomass through conventional and enzymatic route—A comparison. *International Journal of Green Energy*, 13(12): 1232-1253.
- Hamzah, F., Idris, A. and Shuan, T. K. (2011). Preliminary study on enzymatic hydrolysis of treated oil palm (*Elaeis*) empty fruit bunches fibre (EFB) by using combination of cellulase and β 1-4 glucosidase. *Biomass and Bioenergy*, 35(3): 1055-1059.
- Han, W. and He, M. (2010). The application of exogenous cellulase to improve soil fertility and plant growth due to acceleration of straw decomposition. *Bioresource Technology*, 101(10): 3724-3731.
- Harmsen, P., Huijgen, W., Bermudez, L. and Bakker, R. (2010). Literature review of physical and chemical pretreatment processes for lignocellulosic biomass. *Biosynenergy*, 1-53. <http://library.wur.nl/WebQuery/wurpubs/fulltext/150289>. Accessed on 29/09/2017.
- Harris, A. D. and Ramalingam, C. (2010). Xylanases and its application in food industry: a review. *Journal of Experimental Sciences*, 1(7): 1-11
- Harun, N. A. F., Baharuddin, A. S., Zainudin, M. H. M., Bahrin, E. K., Naim, M. N. and Zakaria, R. (2012). Cellulase production from treated oil palm empty fruit bunch degradation by locally isolated *Thermobifida fusca*. *BioResources*, 8(1): 676-687.
- Harun, S., Balan, V., Takriff, M. S., Hassan, O., Jahim, J. and Dale, B. E. (2013). Performance of AFEX™ pretreated rice straw as source of fermentable sugars: the influence of particle size. *Biotechnology for Biofuels*, 6 (40): 1-17.
- Hasegawa, I., Khoo, T. H. and Mae, K. (2013). Direct saccharification of lignocellulosic biomass by hydrolysis with formic acid solution. *Green Processing and Synthesis*, 2(2): 143-149.
- He, Y., Chen, Z., Liu, X., Wang, C. and Lu, W. (2014). Influence of trace elements mixture on bacterial diversity and fermentation characteristics of liquid diet fermented with probiotics under air-tight condition. *PLOS ONE*, 9(12), e114218.

- Heck, J. X., de Barros Soares, L. H. and Ayub, M. A. Z. (2005). Optimization of xylanase and mannanase production by *Bacillus circulans* strain BL53 on solid-state cultivation. *Enzyme and Microbial Technology*, 37(4), 417-423.
- Hendriks, A. and Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*, 100(1): 10-18.
- Hilden, L. and Johansson, G. (2004). Recent developments on cellulases and carbohydrate-binding modules with cellulose affinity. *Biotechnology Letters*, 26(22): 1683-1693.
- Himmel, M. E., Ding, S.-Y., Johnson, D. K., Adney, W. S., Nimlos, M. R., Brady, J. W., et al. (2007). Biomass recalcitrance: engineering plants and enzymes for biofuels production. *Science*, 315(5813): 804-807.
- Hirasawa, K., Uchimura, K., Kashiwa, M., Grant, W. D., Ito, S., Kobayashi, T., et al. (2006). Salt-activated endoglucanase of a strain of alkaliphilic *Bacillus agaradhaerens*. *Antonie Van Leeuwenhoek*, 89(2): 211-219.
- Hongzhang, C. and Junying, Z. (2013). Clean production technology of integrated pretreatment for Lignocellulose. *African Journal of Agricultural Research*, 8(4): 339-348.
- Howard, R., Abotsi, E., Van Rensburg, E. J. and Howard, S. (2003). Lignocellulose biotechnology: issues of bioconversion and enzyme production. *African Journal of Biotechnology*, 2(12): 602-619.
- Hoyles, L., Honda, H., Logan, N. A., Halket, G., La Ragione, R. M. and McCartney, A. L. (2012). Recognition of greater diversity of *Bacillus* species and related bacteria in human faeces. *Research in Microbiology*, 163(1): 3-13.
- Hsieh, C.-w. C., Cannella, D., Jørgensen, H., Felby, C. and Thygesen, L. G. (2015). Cellobiohydrolase and endoglucanase respond differently to surfactants during the hydrolysis of cellulose. *Biotechnology for Biofuels*, 8(52):1-10.
- Hsieh, Y., Du, Y., Jin, F., Zhou, Z. and Enomoto, H. (2009). Alkaline pre-treatment of rice hulls for hydrothermal production of acetic acid. *Chemical Engineering Research and Design*, 87(1): 13-18.
- Hsu, T.-C., Guo, G.-L., Chen, W.-H. and Hwang, W.-S. (2010). Effect of dilute acid pretreatment of rice straw on structural properties and enzymatic hydrolysis. *Bioresource Technology*, 101(13): 4907-4913.
- Hu, D., Ju, X., Li, L., Hu, C., Yan, L., Wu, T., et al. (2016). Improved in situ saccharification of cellulose pretreated by dimethyl sulfoxide/ionic liquid using cellulase from a newly isolated *Paenibacillus* sp. LLZ1. *Bioresource Technology*, 201: 8-14.

- Huang, S., Deng, G., Yang, Y., Wu, Z. and Wu, L. (2015). Optimization of Endoglucanase Production from a Novel Bacterial Isolate, *Arthrobacter* sp. HPG166 and Characterization of Its Properties. *Brazilian Archives of Biology and Technology*, 58(5): 692-701.
- Huang, Y., Wei, X., Zhou, S., Liu, M., Tu, Y., Li, A., et al. (2015). Steam explosion distinctively enhances biomass enzymatic saccharification of cotton stalks by largely reducing cellulose polymerization degree in *G. barbadense* and *G. hirsutum*. *Bioresource Technology*, 181: 224-230.
- Humbird, D., Davis, R., Tao, L., Kinchin, C., Hsu, D., Aden, A., et al. (2011). Process design and economics for biochemical conversion of lignocellulosic biomass to ethanol: dilute-acid pretreatment and enzymatic hydrolysis of corn stover: National Renewable Energy Laboratory (NREL), Golden, CO. <https://www.osti.gov/scitech/biblio/1013269> Accessed on 29/09/2017.
- Ibrahim, M., Razak, M., Phang, L., Hassan, M. and Abd-Aziz, S. (2013). Crude cellulase from oil palm empty fruit bunch by *Trichoderma asperellum* UPM1 and *Aspergillus fumigatus* UPM2 for fermentable sugars production. *Applied Biochemistry and Biotechnology*, 170(6): 1320-1335.
- Iqbal, H. M. N., Kyazze, G. and Keshavarz, T. (2013). Advances in the valorization of lignocellulosic materials by biotechnology: an overview. *BioResources*, 8(2): 3157-3176.
- Irfan, M., Asghar, U., Nadeem, M., Nelofer, R. and Syed, Q. (2016). Optimization of process parameters for xylanase production by *Bacillus* sp. in submerged fermentation. *Journal of Radiation Research and Applied Sciences*, 9(2):139-147.
- Irfan, M., Mushtaq, Q., Tabssum, F., Shakir, H. A. and Qazi, J. I. (2017). Carboxymethyl cellulase production optimization from newly isolated thermophilic *Bacillus subtilis* K-18 for saccharification using response surface methodology. *AMB Express*, 7 (29): 1-9.
- Irfan, M., Tayyab, A., Hasan, F., Khan, S., Badshah, M. and Shah, A. A. (2017). Production and characterization of organic solvent-tolerant cellulase from *Bacillus amyloliquefaciens* AK9 isolated from hot spring. *Applied Biochemistry and Biotechnology*, 182 (4): 1390-1402.
- Jaiswal, A. and Ravindran, R. (2015). A Comprehensive review on pre-treatment strategy for lignocellulosic food Industry waste: challenges and opportunities. *Bioresource Technology*, 199: 92-102.
- Jensen, J., Morinelly, J., Aglan, A., Mix, A. and Shonnard, D. R. (2008). Kinetic characterization of biomass dilute sulfuric acid hydrolysis: Mixtures of hardwoods, softwood, and switchgrass. *AIChE Journal*, 54(6): 1637-1645.

- Jessen, J. E., Sveinsson, T., Scully, S. M. and Orlygsson, J. (2014). Ethanol production by a *Paenibacillus* species isolated from an Icelandic hot spring—Production yields from complex biomass. *Icelandic Agricultural Sciences* 28: 15-24.
- Jeya, M., Zhang, Y. W., Kim, I. W. and Lee, J.K. (2009). Enhanced saccharification of alkali-treated rice straw by cellulase from *Trametes hirsuta* and statistical optimization of hydrolysis conditions by RSM. *Bioresource Technology*, 100(21): 5155-5161.
- Ji, L., Yu, H., Liu, Z., Jiang, J. and Sun, D. (2015). Enhanced ethanol production with mixed lignocellulosic substrates from commercial furfural and cassava residues. *BioResources*, 10(1): 1162-1173.
- Jiang, W., Chang, S., Li, H., Oleskowicz-Popiel, P. and Xu, J. (2015). Liquid hot water pretreatment on different parts of cotton stalk to facilitate ethanol production. *Bioresource Technology*, 176: 175-180.
- Jiang, W., Peng, H., Li, H. and Xu, J. (2014). Effect of acetylation/deacetylation on enzymatic hydrolysis of corn stalk. *Biomass and Bioenergy*, 71: 294-298.
- Johar, N., Ahmad, I. and Dufresne, A. (2012). Extraction, preparation and characterization of cellulose fibres and nanocrystals from rice husk. *Industrial Crops and Products*, 37(1): 93-99.
- Jönsson, L. J. and Martín, C. (2016). Pretreatment of lignocellulose: formation of inhibitory by-products and strategies for minimizing their effects. *Bioresource Technology*, 199: 103-112.
- Jørgensen, H., Kristensen, J. B. and Felby, C. (2007). Enzymatic conversion of lignocellulose into fermentable sugars: challenges and opportunities. *Biofuels, Bioproducts and Biorefining*, 1(2): 119-134.
- Jurado, M. M., Suárez-Estrella, F., López, M. J., Vargas-García, M. C., López-González, J. A. and Moreno, J. (2015). Enhanced turnover of organic matter fractions by microbial stimulation during lignocellulosic waste composting. *Bioresource Technology*, 186: 15-24.
- Juturu, V. and Wu, J. C. (2012). Microbial xylanases: engineering, production and industrial applications. *Biotechnology Advances*, 30(6): 1219-1227.
- Juturu, V. and Wu, J. C. (2014). Microbial cellulases: Engineering, production and applications. *Renewable and Sustainable Energy Reviews*, 33: 188-203.
- Jyotsna, K. P., Rao, A. R. and Devaki, K. (2015). Effect of nutritional factors on cellulase production by *Lyccinibacillus sphaericus* from the gut of earthworm, *Eisenia foetida*. *Current Biotica*, 9(2): 145-152.

- Kabel, M. A., Bos, G., Zeevalking, J., Voragen, A. G. J. and Schols, H. A. (2007). Effect of pretreatment severity on xylan solubility and enzymatic breakdown of the remaining cellulose from wheat straw. *Bioresource Technology*, 98(10): 2034-2042.
- Kalim, B., Böhringer, N., Ali, N. and Schäberle, T. F. (2015). Xylanases—from microbial origin to Industrial application. *British Biotechnology Journal*, 7:1-20.
- Kamran, A., Ur Rehman, H., Ul Qader, S. A., Baloch, A. H. and Kamal, M. (2015). Purification and characterization of thiol dependent, oxidation-stable serine alkaline protease from thermophilic *Bacillus* sp. *Journal of Genetic Engineering and Biotechnology*, 13(1): 59-64.
- Karim, A., Nawaz, M. A., Aman, A. and Ul Qader, S. A. (2015). Hyper production of cellulose degrading endo (1,4) β -d-glucanase from *Bacillus licheniformis* KIBGE-IB2. *Journal of Radiation Research and Applied Sciences*, 8(2): 160-165.
- Karimi, K. and Taherzadeh, M. J. (2016). A critical review on analysis in pretreatment of lignocelluloses: Degree of polymerization, adsorption/desorption, and accessibility. *Bioresource Technology*, 203: 348-356.
- Karlsson, O., Tornaiainen, P., Dagbro, O., Granlund, K. and Morén, T. (2012). Presence of water-soluble compounds in thermally modified wood: carbohydrates and furfurals. *BioResources*, 7(3): 3679-3689.
- Kasana, R. C. and Gulati, A. (2011). Cellulases from psychrophilic microorganisms: a review. *Journal of Basic Microbiology*, 51(6): 572-579.
- Kasana, R. C., Salwan, R., Dhar, H., Dutt, S. and Gulati, A. (2008). A rapid and easy method for the detection of microbial cellulases on agar plates using Gram's iodine. *Current Microbiology*, 57(5): 503-507.
- Kashket, E. R. (1987). Bioenergetics of lactic acid bacteria: cytoplasmic pH and osmotolerance. *FEMS Microbiology Letters*, 46(3): 233-244.
- Kazeem, M. O., Shah, U. K. M. and Baharuddin, A. S. (2016). Enhanced Cellulase Production by a Novel Thermophilic *Bacillus licheniformis* 2D55: Characterization and Application in Lignocellulosic Saccharification. *BioResources*, 11(2): 5404-5423.
- Khatiwada, P., Ahmed, J., Sohag, M., Islam, K. and Azad, A. (2016). Isolation, screening and characterization of cellulase producing bacterial isolates from municipal solid wastes and rice straw wastes. *Journal of Bioprocessing & Biotechniques*, 6: 280-285.

- Khokhar, Z. U., Syed, Q., Nadeem, M., Irfan, M., Wu, J., Samra, Z. Q., et al. (2014). Enhanced production of cellulase by *Trichoderma reesei* using wheat straw as a carbon source. *World Applied Sciences Journal*, 30(9): 1095-1104.
- Kim, H.-J., Gao, W., Chung, C. H. and Lee, J. W. (2011). Statistical optimization for production of carboxymethylcellulase from rice hulls by a newly isolated marine microorganism *Bacillus licheniformis* LBH-52 using response surface method. *Journal of Life Science*, 21(8):1083-1093.
- Kim, B. K., Lee, B. H., Lee, Y. J., Jin, I. H., Chung, C. H. and Lee, J.W. (2009). Purification and characterization of carboxymethylcellulase isolated from a marine bacterium, *Bacillus subtilis* subsp. *subtilis* A-53. *Enzyme and Microbial Technology*, 44(6): 411-416.
- Kim, K. H., Tucker, M. and Nguyen, Q. (2005). Conversion of bark-rich biomass mixture into fermentable sugar by two-stage dilute acid-catalyzed hydrolysis. *Bioresource Technology*, 96(11): 1249-1255.
- Kim, T. H. and Lee, Y. (2006). Fractionation of corn stover by hot-water and aqueous ammonia treatment. *Bioresource Technology*, 97(2): 224-232.
- Kim, Y. K., Lee, S. C., Cho, Y. Y., Oh, H. J. and Ko, Y. H. (2012). Isolation of cellulolytic *Bacillus subtilis* strains from agricultural environments. *International Scholarly Research Network Microbiology*, ID 650563 2012: 1-9
- Kishishita, S., Fujii, T. and Ishikawa, K. (2015). Heterologous expression of hyperthermophilic cellulases of archaea *Pyrococcus* sp. by fungus *Talaromyces cellulolyticus*. *Journal of Industrial Microbiology & Biotechnology*, 42:137-141.
- Koo, B. W., Kim, H. Y., Park, N., Lee, S. M., Yeo, H. and Choi, I. G. (2011). Organosolv pretreatment of *Liriodendron tulipifera* and simultaneous saccharification and fermentation for bioethanol production. *Biomass and Bioenergy*, 35(5): 1833-1840.
- Korany, A. H., Ali, A. E., Essam, T. M. and Megahed, S. A. (2017). Optimization of cellulase production by *Halobacillus* sp. QLS 31 isolated from Lake Qarun, Egypt. *Applied Biochemistry and Biotechnology*, 183(1): 189-199.
- Korenaga, T., Liu, X. and Huang, Z. (2001). The influence of moisture content on polycyclic aromatic hydrocarbons emission during rice straw burning. *Chemosphere Global Change Science*, 3(1), 117-122.
- Kostylev, M. and Wilson, D. (2012). Synergistic interactions in cellulose hydrolysis. *Biofuels*, 3(1): 61-70.

- Kshirsagar, S. D., Waghmare, P. R., Chandrakant Loni, P., Patil, S. A. and Govindwar, S. P. (2015). Dilute acid pretreatment of rice straw, structural characterization and optimization of enzymatic hydrolysis conditions by response surface methodology. *RSC Advances*, 5(58): 46525-46533.
- Kshirsagar, S. D., Waghmare, P. R., Loni, P. C., Patil, S. A. and Govindwar, S. P. (2015). Dilute acid pretreatment of rice straw, structural characterization and optimization of enzymatic hydrolysis conditions by response surface methodology. *RSC Advances*, 5(58): 46525-46533.
- Kuhad, R. C., Gupta, R. and Singh, A. (2011). Microbial cellulases and their industrial applications. *Enzyme Research*, 2011, 1-10.
- Kumar, D. and Murthy, G. S. (2013). Stochastic molecular model of enzymatic hydrolysis of cellulose for ethanol production. *Biotechnology for Biofuels*, 6(63): 1-20.
- Kumar, L., Kumar, D., Nagar, S., Gupta, R., Garg, N., Kuhad, R. C., et al. (2014). Modulation of xylanase production from alkaliphilic *Bacillus pumilus* VLK-1 through process optimization and temperature shift operation. *3 Biotech*, 4(4): 345-356.
- Kumar, P., Barrett, D. M., Delwiche, M. J. and Stroeve, P. (2009a). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial & Engineering Chemistry Research*, 48(8): 3713-3729.
- Kumar, R., Mago, G., Balan, V. and Wyman, C. E. (2009). Physical and chemical characterizations of corn stover and poplar solids resulting from leading pretreatment technologies. *Bioresource Technology*, 100(17): 3948-3962.
- Kumar, R., Tabatabaei, M., Karimi, K. and Sárvári Horváth, I. (2016). Recent updates on lignocellulosic biomass derived ethanol-A review. *Biofuel Research Journal*, 3(1): 347-356.
- Kumar, S., Sharma, H. K. and Sarkar, B. C. (2011). Effect of substrate and fermentation conditions on pectinase and cellulase production by *Aspergillus niger* NCIM 548 in submerged (SmF) and solid state fermentation (SSF). *Food Science and Biotechnology*, 20(5): 1289-1298.
- Kumar, U., Kumar, B. and Pandey, K. M. (2014). SEM and EDAX study of pretreated rice Husk for the sorption of Cu (II) from waste water. *Applied Mechanics and Materials* 656: 17-22.
- Kumar, V., Marín-Navarro, J. and Shukla, P. (2016). Thermostable microbial xylanases for pulp and paper industries: trends, applications and further perspectives. *World Journal of Microbiology and Biotechnology*, 32(2): 1-10.

- Kupiainen, L. (2012). Dilute acid catalyzed hydrolysis of cellulose-extension to formic acid. *Desertation University of Ouluensis, C Technica*, 438.pp 1-68
- Ladeira, S. A., Cruz, E., Delatorre, A. B., Barbosa, J. B. and Martins, M. L. L. (2015). Cellulase production by thermophilic *Bacillus* sp. SMIA-2 and its detergent compatibility. *Electronic Journal of Biotechnology*, 18(2): 110-115.
- Lai, L. W. and Idris, A. (2016). Comparison of steam-alkali-chemical and microwave-alkali pretreatment for enhancing the enzymatic saccharification of oil palm trunk. *Renewable Energy*, 99: 738-746.
- Lairson, L. L., Henrissat, B., Davies, G. J. and Withers, S. G. (2008). Glycosyltransferases: structures, functions, and mechanisms. *Biochemistry*, 77: 521-555.
- Laurichesse, S. and Avérous, L. (2014). Chemical modification of lignins: Towards biobased polymers. *Progress in Polymer Science*, 39(7): 1266-1290.
- Lee, B. H., Kim, B. K., Lee, Y. J., Chung, C. H. and Lee, J.-W. (2010). Industrial scale of optimization for the production of carboxymethylcellulase from rice bran by a marine bacterium, *Bacillus subtilis* subsp. *subtilis* A-53. *Enzyme and Microbial Technology*, 46(1): 38-42.
- Lee, S. H., Doherty, T. V., Linhardt, R. J. and Dordick, J. S. (2009). Ionic liquid-mediated selective extraction of lignin from wood leading to enhanced enzymatic cellulose hydrolysis. *Biotechnology and Bioengineering*, 102(5): 1368-1376.
- Lee, Y. J., Kim, B. K., Lee, B. H., Jo, K. I., Lee, N. K., Chung, C.H., et al. (2008). Purification and characterization of cellulase produced by *Bacillus amyloliquefaciens* DL-3 utilizing rice hull. *Bioresource Technology*, 99(2): 378-386.
- Leite, P., Salgado, J. M., Venâncio, A., Domínguez, J. M. and Belo, I. (2016). Ultrasounds pretreatment of olive pomace to improve xylanase and cellulase production by solid-state fermentation. *Bioresource Technology*, 214: 737-746.
- Lenihan, P., Orozco, A., O'Neill, E., Ahmad, M. N. M., Rooney, D. W. and Walker, G. M. (2010). Dilute acid hydrolysis of lignocellulosic biomass. *Chemical Engineering Journal*, 156(2): 395-403.
- Li, X. and Yu, H. Y. (2012). Purification and characterization of an organic-solvent-tolerant cellulase from a halotolerant isolate, *Bacillus* sp. L1. *Journal of Industrial Microbiology and Biotechnology*, 39(8):1117-1124.

- Li, J., Henriksson, G. and Gellerstedt, G. (2007). Lignin depolymerization/repolymerization and its critical role for delignification of aspen wood by steam explosion. *Bioresource Technology*, 98(16): 3061-3068.
- Li, J., Zhang, X., Zhang, M., Xiu, H. and He, H. (2015). Ultrasonic enhance acid hydrolysis selectivity of cellulose with HCl-FeCl₃ as catalyst. *Carbohydrate polymers*, 117: 917-922.
- Li, S., Bashline, L., Lei, L. and Gu, Y. (2014). Cellulose synthesis and its regulation. *The Arabidopsis Book*, 14, 1-21
- Liang, Y.-L., Zhang, Z., Wu, M., Wu, Y. and Feng, J.X. (2014). Isolation, screening, and identification of cellulolytic bacteria from natural reserves in the subtropical region of China and optimization of cellulase production by *Paenibacillus terrae* ME27-1. *BioMed Research International*, 2014, 1-13
- Liang, Y., Feng, Z., Yesuf, J. and Blackburn, J. W. (2010). Optimization of growth medium and enzyme assay conditions for crude cellulases produced by a novel thermophilic and cellulolytic bacterium, *Anoxybacillus* sp. 527. *Applied Biochemistry and Biotechnology*, 160(6): 1841-1852.
- Liang, Y., Yesuf, J. and Feng, Z. (2010). Toward plant cell wall degradation under thermophilic condition: a unique microbial community developed originally from swine waste. *Applied Biochemistry and Biotechnology*, 161(18): 147-156.
- Liang, Y., Yesuf, J., Schmitt, S., Bender, K. and Bozzola, J. (2009). Study of cellulases from a newly isolated thermophilic and cellulolytic *Brevibacillus* sp. strain JXL. *Journal of Industrial Microbiology & Biotechnology*, 36(7): 961-970.
- Lim, J. S., Manan, Z. A., Alwi, S. R. W. and Hashim, H. (2012). A review on utilisation of biomass from rice industry as a source of renewable energy. *Renewable and Sustainable Energy Reviews*, 16(5): 3084-3094.
- Lim, W. S. and Lee, J. W. (2013). Effects of pretreatment factors on fermentable sugar production and enzymatic hydrolysis of mixed hardwood. *Bioresource Technology*, 130: 97-101.
- Liu, C. J., Miao, Y. C. and Zhang, K.W. (2011). Sequestration and transport of lignin monomeric precursors. *Molecules*, 16(1): 710-727.
- Liu, J. and Yang, J. (2007). Cellulase production by *Trichoderma koningii* AS3. 4262 in solid-state fermentation using lignocellulosic waste from the vinegar industry. *Food Technology and Biotechnology*, 45(4), 420-425
- Liu, C. F., Sun, R. C., Zhang, A. P. and Ren, J. L. (2007). Preparation of sugarcane bagasse cellulosic phthalate using an ionic liquid as reaction medium. *Carbohydrate Polymers*, 68(1): 17-25.

- Ludueña, L., Fasce, D., Alvarez, V. A. and Stefani, P. M. (2011). Nanocellulose from rice husk following alkaline treatment to remove silica. *BioResources*, 6(2): 1440-1453.
- Lugani, Y., Singla, R. and Sooch, B. S. (2015). Optimization of cellulase production from newly isolated *Bacillus* sp. Y3. *Journal of Bioprocessing & Biotechniques*, 5(11): 264
- Lusterio, D. D., Suizo, F. G., Labunos, N. M., Valledor, M. N., Ueda, S., Kawai, S., et al. (1992). Alkali-Resistant, Alkaline Endo-1, 4- β -glucanase Produced by *Bacillus* sp. PKM-5430. *Bioscience, Biotechnology, and Biochemistry*, 56(10): 1671-1672.
- Lynd, L. R., Weimer, P. J., van Zyl, W. H. and Pretorius, I. S. (2002). Microbial Cellulose Utilization: Fundamentals and Biotechnology. *Microbiology and Molecular Biology Reviews*, 66(3): 506-577.
- Maeda, R. N., Serpa, V. I., Rocha, V. A. L., Mesquita, R. A. A., Anna, L. M. M. S., de Castro, A. M., et al. (2011). Enzymatic hydrolysis of pretreated sugar cane bagasse using *Penicillium funiculosum* and *Trichoderma harzianum* cellulases. *Process Biochemistry*, 46(5): 1196-1201.
- Mahamud, M. R. and Gomes, D. J. (2012). Enzymatic saccharification of sugar cane bagasse by the crude enzyme from indigenous fungi. *Journal of Scientific Research*, 4(1): 227-238.
- Mahmud, N. A. N., Baharuddin, A. S., Bahrin, E. K., Sulaiman, A., Naim, M. N., Zakaria, R., et al. (2013). Enzymatic saccharification of oil palm mesocarp fiber (OPMF) treated with superheated steam. *BioResources*, 8(1): 1320-1331.
- Maitan-Alfenas, G. P., Visser, E. M. and Guimarães, V. M. (2015). Enzymatic hydrolysis of lignocellulosic biomass: converting food waste in valuable products. *Current Opinion in Food Science*, 1: 44-49.
- Maki, M., Leung, K. T. and Qin, W. (2009). The prospects of cellulase-producing bacteria for the bioconversion of lignocellulosic biomass. *International Journal of Biological Sciences*, 5(5): 500-516.
- Maki, M. L., Broere, M., Leung, K. T. and Qin, W. (2011a). Characterization of some efficient cellulase producing bacteria isolated from paper mill sludges and organic fertilizers. *International Journal Biochemistry and Molecular Biology*, 2(2):146-154.
- Maki, M. L., Broere, M., Leung, K. T. and Qin, W. (2011b). Characterization of some efficient cellulase producing bacteria isolated from paper mill sludges and organic fertilizers. *International Journal of Biochemistry and Molecular Biology*, 2(2): 146-154.

- Mandels, M. and Reese, E. (1999). Fungal cellulases and the microbial decomposition of cellulosic fabric. *Journal of Industrial Microbiology & Biotechnology*, 22(4): 225-240.
- Mandels, M. and Weber, J. (1969). The production of cellulases. *Advances in Chemistry* 95: 331-414
- Matkar, K., Chapla, D., Divecha, J., Nighojkar, A. and Madamwar, D. (2013). Production of cellulase by a newly isolated strain of *Aspergillus sydowii* and its optimization under submerged fermentation. *International Biodeterioration & Biodegradation*, 78: 24-33.
- Mawadza, C., Hatti-Kaul, R., Zvaunya, R. and Mattiasson, B. (2000). Purification and characterization of cellulases produced by two *Bacillus* strains. *Journal of Biotechnology*, 83(3): 177-187.
- Medie, F. M., Davies, G. J., Drancourt, M. and Henrissat, B. (2012). Genome analyses highlight the different biological roles of cellulases. *Nature Reviews Microbiology*, 10(3): 227-234.
- Meng, F., Ma, L., Ji, S., Yang, W. and Cao, B. (2014). Isolation and characterization of *Bacillus subtilis* strain BY-3, a thermophilic and efficient cellulase-producing bacterium on untreated plant biomass. *Letters in Applied Microbiology*, 59(3): 306-312.
- Meng, F., Xing, G., Li, Y., Song, J., Wang, Y., Meng, Q., et al. (2016). The optimization of *Marasmius androsaceus* submerged fermentation conditions in five-liter fermentor. *Saudi Journal of Biological Sciences*, 23(1): S99-S105.
- Meng, X. and Ragauskas, A. J. (2014). Recent advances in understanding the role of cellulose accessibility in enzymatic hydrolysis of lignocellulosic substrates. *Current Opinion in Biotechnology*, 27: 150-158.
- Meng, X., Wells, T., Sun, Q., Huang, F. and Ragauskas, A. (2015). Insights into the effect of dilute acid, hot water or alkaline pretreatment on the cellulose accessible surface area and the overall porosity of *Populus*. *Green Chemistry*, 17(8): 4239-4246.
- Menon, V. and Rao, M. (2012). Trends in bioconversion of lignocellulose: biofuels, platform chemicals and biorefinery concept. *Progress in Energy and Combustion Science*, 38(4), 522-550.
- Merino, S. T. and Cherry, J. (2007). Progress and challenges in enzyme development for biomass utilization In: Olsson L. (eds) *Biofuels*. Advances in biochemical engineering and biotechnology, Springer, Berlin, Heidelberg 108: 95-120

- Mesa, L., González, E., Cara, C., González, M., Castro, E. and Mussatto, S. I. (2011). The effect of organosolv pretreatment variables on enzymatic hydrolysis of sugarcane bagasse. *Chemical Engineering Journal*, 168(3): 1157-1162.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, 31(3): 426-428.
- Mirahmadi, K., Kabir, M. M., Jeihanipour, A., Karimi, K. and Taherzadeh, M. (2010). Alkaline pretreatment of spruce and birch to improve bioethanol and biogas production. *BioResources*, 5(2): 928-938.
- Mistry, T. B., Shaikh, N. M., Rana, H. N. and Patel, N. D. (2016). Isolation and screening of cellulase producing thermophilic bacteria from compost piles and optimization of cellulase production. *International Journal of Advanced Biotechnology and Research*, 7(1): 64-76.
- Mmango-Kaseke, Z., Okaiyeto, K., Nwodo, U. U., Mabinya, L. V. and Okoh, A. I. (2016). Optimization of cellulase and xylanase production by *micrococcus* species under submerged fermentation. *Sustainability*, 8(11): 1168.
- Mood, S. H., Golfeshan, A. H., Tabatabaei, M., Jouzani, G. S., Najafi, G. H., Gholami, M., et al. (2013). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renewable and Sustainable Energy Reviews*, 27: 77-93.
- Mooney, C. A., Mansfield, S. D., Touhy, M. G. and Saddler, J. N. (1998). The effect of initial pore volume and lignin content on the enzymatic hydrolysis of softwoods. *Bioresource Technology*, 64(2): 113-119.
- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y. Y., Holtzapple, M., et al. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology*, 96(6): 673-686.
- Mostafa, Y. S. and Alamri, S. A. (2014). Biosynthesis of thermostable-amylase by immobilized *Bacillus subtilis* in batch and repeat batch cultures using fortified date syrup medium. *African Journal of Microbiology Research*, 8(12): 1292-1301.
- Motta, F. L., Andrade, C. C. P. and Santana, M. H. A. (2013). A review of xylanase production by the fermentation of xylan: classification, characterization and applications In: Anuj, C. K., and Silvio, S. D., *Sustainable Degradation of Lignocellulosic Biomass-Techniques, Applications and Commercialization*: InTech. Open science pp.252-275
- Mussatto, S. I. and Roberto, I. C. (2004). Alternatives for detoxification of diluted-acid lignocellulosic hydrolyzates for use in fermentative processes: a review. *Bioresource Technology*, 93(1), 1-10.

- Nakagame, S., Chandra, R. and Saddler, J. (2011). The influence of lignin on the enzymatic hydrolysis of pretreated biomass substrates. In: Junyong, J. Y., Z, Xiao Z., Xuejun, J (eds.) Sustainable production of fuels, chemicals, and fibers from forest biomass, *American Chemical Society* (ACS) pp.145-167.
- Nakagame, S., Chandra, R. P. and Saddler, J. N. (2010). The effect of isolated lignins, obtained from a range of pretreated lignocellulosic substrates, on enzymatic hydrolysis. *Biotechnology and Bioengineering*, 105(5):871-879.
- Nascimento, P., Marim, R., Carvalho, G. and Mali, S. (2016). Nanocellulose produced from rice hulls and its effect on the properties of biodegradable starch films. *Materials Research*, 19 (1): 167-174
- nee'Nigam, P. S., Gupta, N. and Anthwal, A. (2009). Pretreatment of agro-industrial residues In: Singh nee' Nigam P., Pandey A. (eds) *Biotechnology for agro-industrial residues utilisation*. Springer, Dordrecht pp 13-33
- Nema, N., Alamir, L. and Mohammad, M. (2015). Production of cellulase from *Bacillus cereus* by submerged fermentation using corn husks as substrates. *International Food Research Journal*, 22(5):1831-.1836.
- Nierhaus, K. H. (2014). Mg²⁺, K⁺, and the ribosome. *Journal of Bacteriology*, 196(22): 3817-3819.
- Nilsson, D. and Hansson, P. A. (2001). Influence of various machinery combinations, fuel proportions and storage capacities on costs for co-handling of straw and reed canary grass to district heating plants. *Biomass and Bioenergy*, 20(4): 247-260.
- Noparat, P., Prasertsan, P., Sompong, O. and Pan, X. (2015). Dilute acid pretreatment of oil palm trunk biomass at high temperature for enzymatic hydrolysis. *Energy Procedia*, 79: 924-929.
- Nordin, N. I. A. A., Ariffin, H., Andou, Y., Hassan, M. A., Shirai, Y., Nishida, H., et al. (2013). Modification of oil palm mesocarp fiber characteristics using superheated steam treatment. *Molecules*, 18(8): 9132-9146.
- Oke, M. A., Annuar, M. S. M. and Simarani, K. (2016a). Enhanced endoglucanase production by *Bacillus aerius* on mixed lignocellulosic substrates. *BioResources*, 11(3): 5854-5869.
- Oke, M. A., Annuar, M. S. M. and Simarani, K. (2016b). Mixed lignocellulosic biomass degradation and utilization for bacterial cellulase production. *Waste and Biomass Valorization*, 8(3): 893-903

- Omidvar, M., Karimi, K. and Mohammadi, M. (2016). Enhanced ethanol and glucosamine production from rice husk by NaOH pretreatment and fermentation by fungus *Mucor hiemalis*. *Biofuel Research Journal*, 3(3): 475-481.
- Otero, D. M., Cadaval, C. L., Teixeira, L. M., Rosa, C. A., Sanzo, A. V. L. and Kalil, S. J. (2015). Screening of yeasts capable of producing cellulase-free xylanase. *African Journal of Biotechnology*, 14(23): 1961-1969.
- Ouyang, J., Dong, Z., Song, X., Lee, X., Chen, M. and Yong, Q. (2010). Improved enzymatic hydrolysis of microcrystalline cellulose (Avicel PH101) by polyethylene glycol addition. *Bioresource Technology*, 101(17): 6685-6691
- Ouyang, J., Liu, B., Zhang, M., Zheng, Z. and Yu, H. (2013). Enzymatic hydrolysis, adsorption, and recycling during hydrolysis of bagasse sulfite pulp. *Bioresource Technology*, 146: 288-293.
- Padilha, I., Carvalho, L., Dias, P., Grisi, T., Silva, F., Santos, S., et al. (2015). Production and characterization of thermophilic carboxymethyl cellulase synthesized by *Bacillus* sp. Growing on sugarcane bagasse in submerged fermentation. *Brazilian Journal of Chemical Engineering*, 32(1):35-42.
- Palmqvist, E. and Hahn-Hägerdal, B. (2000). Fermentation of lignocellulosic hydrolysates. I: inhibition and detoxification. *Bioresource Technology*, 74(1):17-24
- Palonen, H. (2004). Role of lignin in the enzymatic hydrolysis of lignocellulose. *VTT publication*, 520: 1-80
- Park, J. I., Steen, E. J., Burd, H., Evans, S. S., Redding-Johnson, A. M., Batth, T., et al. (2012). A thermophilic ionic liquid-tolerant cellulase cocktail for the production of cellulosic biofuels. *PLOS ONE*, 7(5), e37010.
- Patel, M. A., Ou, M. S., Ingram, L. O. and Shanmugam, K. T. (2005). Simultaneous saccharification and co- fermentation of crystalline cellulose and sugar cane bagasse hemicellulose hydrolysate to lactate by a thermotolerant acidophilic *Bacillus* sp. *Biotechnology Progress*, 21(5): 1453-1460.
- Pathan, A. K., Bond, J. and Gaskin, R. E. (2008). Sample preparation for scanning electron microscopy of plant surfaces—horses for courses. *Micron*, 39(8): 1049-1061.
- Paudel, Y. P. and Qin, W. (2015). Characterization of Novel Cellulase-producing bacteria isolated from rotting wood samples. *Applied Biochemistry and Biotechnology*, 177(5): 1186-1198.

- Peng, X., Qiao, W., Mi, S., Jia, X., Su, H. and Han, Y. (2015). Characterization of hemicellulase and cellulase from the extremely thermophilic bacterium *Caldicellulosiruptor owensensis* and their potential application for bioconversion of lignocellulosic biomass without pretreatment. *Biotechnology for Biofuels*, 8(131):1-14.
- Pérez, J., Munoz-Dorado, J., de la Rubia, T. and Martinez, J. (2002). Biodegradation and biological treatments of cellulose, hemicellulose and lignin: an overview. *International Microbiology*, 5(2): 53-63.
- Pillai, P., Mandge, S. and Archana, G. (2011). Statistical optimization of production and tannery applications of a keratinolytic serine protease from *Bacillus subtilis* P13. *Process Biochemistry*, 46(5): 1110-1117.
- Pillet, F., Formosa-Dague, C., Baaziz, H., Dague, E. and Rols, M.-P. (2016). Cell wall as a target for bacteria inactivation by pulsed electric fields. *Scientific Reports*, 6:19778
- Pode, R. (2016). Potential applications of rice husk ash waste from rice husk biomass power plant. *Renewable and Sustainable Energy Reviews*, 53: 1468-1485.
- Poornejad, N., Karimi, K. and Behzad, T. (2014). Ionic liquid pretreatment of rice straw to enhance saccharification and bioethanol production. *Journal of Biomass to Biofuel*, 1: 8-15.
- Potumarthi, R., Baadhe, R. R., Nayak, P. and Jetty, A. (2013). Simultaneous pretreatment and saccharification of rice husk by *Phanerochete chrysosporium* for improved production of reducing sugars. *Bioresource Technology*, 128: 113-117.
- Prasanna, H. N., Ramanjaneyulu, G. and Rajasekhar Reddy, B. (2016). Optimization of cellulase production by *Penicillium* sp. 3 *Biotech*, 6(162): 1-11.
- Premalatha, N., Gopal, N. O., Jose, P. A., Anandham, R. and Kwon, S. W. (2014). Optimization of cellulase production by *Enhydrobacter* sp. ACCA2 and its application in biomass saccharification. *Frontiers in Microbiology*, 6: 1046-1046.
- Priya, I., Dhar, M. K., Bajaj, B. K., Koul, S. and Vakhlu, J. (2016). Cellulolytic activity of thermophilic bacilli isolated from tattapani hot spring sediment in North West Himalayas. *Indian journal of microbiology*, 56(2): 228-231.
- Pu, Y., Hu, F., Huang, F., Davison, B. H. and Ragauskas, A. J. (2013). Assessing the molecular structure basis for biomass recalcitrance during dilute acid and hydrothermal pretreatments. *Biotechnology for Biofuels*, 6(15): 1-13.

- Qi, B., Chen, X., Su, Y. and Wan, Y. (2011). Enzyme adsorption and recycling during hydrolysis of wheat straw lignocellulose. *Bioresource Technology*, 102(3): 2881-2889.
- Quiroga, A. G., Silvera, A. B., Padilla, R. V., da Costa, A. C. and Maciel Filho, R. (2015). Continuous and semicontinuous reaction systems for high-solids enzymatic hydrolysis of lignocellulosics. *Brazilian Journal of Chemical Engineering*, 32(4): 805-819.
- Rahnama, N., Mamat, S., Shah, U. K. M., Ling, F. H., Rahman, N. A. A. and Ariff, A. B. (2013). Effect of alkali pretreatment of rice straw on cellulase and xylanase production by local *Trichoderma harzianum* SNRS3 under solid state fermentation. *BioResources*, 8(2): 2881-2896.
- Ram, L., Kaur, K. and Sharma, S. (2014). Screening isolation and characterization of cellulase producing microorganisms from soil. *International Journal of Pharmarceutical Science Invention* 3(3): 12-18.
- Ramírez-Núñez, J., Romero-Medrano, R., Nevárez-Moorillón, G. V. and Gutiérrez-Méndez, N. (2011). Effect of pH and salt gradient on the autolysis of *Lactococcus lactis* strains. *Brazilian Journal of Microbiology*, 42(4): 1495-1499.
- Rana, V., Eckard, A. D., Teller, P. and Ahring, B. K. (2014). On-site enzymes produced from *Trichoderma reesei* RUT-C30 and *Aspergillus saccharolyticus* for hydrolysis of wet exploded corn stover and loblolly pine. *Bioresource Technology*, 154: 282-289.
- Rastogi, G., Bhalla, A., Adhikari, A., Bischoff, K. M., Hughes, S. R., Christopher, L. P., et al. (2010). Characterization of thermostable cellulases produced by *Bacillus* and *Geobacillus* strains. *Bioresource Technology*, 101(22): 8798-8806.
- Rastogi, G., Muppidi, G. L., Gurram, R. N., Adhikari, A., Bischoff, K. M., Hughes, S. R., et al. (2009). Isolation and characterization of cellulose-degrading bacteria from the deep subsurface of the Homestake gold mine, Lead, South Dakota, USA. *Journal of Industrial Microbiology & Biotechnology*, 36(4): 585-598.
- Rasul, F., Afroz, A., Rashid, U., Mehmood, S., Sughra, K. and Zeeshan, N. (2015). Screening and characterization of cellulase producing bacteria from soil and waste (molasses) of sugar industry. *International Journal of Bioscience*, 6(3): 230-238.
- Rawat, R. and Tewari, L. (2012). Purification and characterization of an acidothermophilic cellulase enzyme produced by *Bacillus subtilis* strain LFS3. *Extremophiles*, 16(4), 637-644.

- Reddy, K. V., Lakshmi, T. V., Reddy, A. V. K., Bindu, V. H. and Narasu, M. L. (2016). Isolation, screening, identification and optimized production of extracellular cellulase from *Bacillus subtilis* using cellulosic waste as carbon source. *International Journal of Current Microbiology and Applied Sciences*, 5(4), 442-451.
- Reed, C. J., Lewis, H., Trejo, E., Winston, V. and Evilia, C. (2013). Protein adaptations in archaeal extremophiles. *Archaea*, 2013. ID373275 1-14
- Rentizelas, A., Tatsiopoulou, I. and Tolis, A. (2009). An optimization model for multi-biomass tri-generation energy supply. *Biomass and Bioenergy*, 33(2): 223-233.
- Rey, M. W., Ramaiya, P., Nelson, B. A., Brody-Karpin, S. D., Zaretsky, E. J., Tang, M., et al. (2004). Complete genome sequence of the industrial bacterium *Bacillus licheniformis* and comparisons with closely related *Bacillus* species. *Genome Biology*, 5(10): 1-12.
- Rojas- Rejón, Ó. A., Poggi- Varaldo, H. M., Ramos- Valdivia, A. C., Ponce- Noyola, T., Cristiani- Urbina, E., Martínez, A., et al. (2016). Enzymatic saccharification of sugar cane bagasse by continuous xylanase and cellulase production from *cellulomonas flavigena* PR- 22. *Biotechnology Progress*, 32(2): 321-326.
- Ryu, D. D. and Mandels, M. (1980). Cellulases: biosynthesis and applications. *Enzyme and Microbial Technology*, 2(2) :91-102.
- Sadhu, S., Ghosh, P. K., Aditya, G. and Maiti, T. K. (2014). Optimization and strain improvement by mutation for enhanced cellulase production by *Bacillus* sp. (MTCC10046) isolated from cow dung. *Journal of King Saud University - Science*, 26(4) :323-332.
- Sadhu, S., Ghosh, P. K., De, T. K. and Maiti, T. K. (2013). Optimization of cultural condition and synergistic effect of lactose with carboxymethyl cellulose on cellulase production by *Bacillus* sp. isolated from fecal matter of elephant (*Elephas maximus*). *Advances in Microbiology*, 3: 280-288
- Sadhu, S., Saha, P., Sen, S. K., Mayilraj, S. and Maiti, T. K. (2013). Production, purification and characterization of a novel thermotolerant endoglucanase (CMCase) from *Bacillus* strain isolated from cow dung. *SpringerPlus*, 2(1): 1-10.
- Saha, B. C. and Cotta, M. A. (2007). Enzymatic saccharification and fermentation of alkaline peroxide pretreated rice hulls to ethanol. *Enzyme and Microbial Technology*, 41(4): 528-532.
- Saini, J. K., Saini, R. and Tewari, L. (2015). Lignocellulosic agriculture wastes as biomass feedstocks for second-generation bioethanol production: concepts and recent developments. *3 Biotech*, 5(4): 337-353.

- Salwanee, S., Wan Aida, W. M., Mamot, S. and Maskat, M. Y. (2013). Effects of enzyme concentration, temperature, ph and time on the degree of hydrolysis of protein extract from viscera of tuna (*euthynnus affinis*) by using alcalase. *Sains Malaysiana*, 42(3): 279-287.
- Saranraj, P., Stella, D. and Reetha, D. (2012). Microbial cellulases and its applications: A review. *International Journal of Biochemistry Biotechnological Science*, 1:1-12.
- Saritha, M. and Arora, A. (2012). Biological pretreatment of lignocellulosic substrates for enhanced delignification and enzymatic digestibility. *Indian journal of microbiology*, 52(2): 122-130.
- Saritha, M., Arora, A. and Lata. (2012). Biological pretreatment of lignocellulosic substrates for enhanced delignification and enzymatic digestibility. *Indian journal of microbiology*, 52(2): 122-130.
- Sawle, L. and Ghosh, K. (2011). How do thermophilic proteins and proteomes withstand high temperature? *Biophysical Journal*, 101(1) :217-227.
- Scheller, H. V. and Ulvskov, P. (2010). Hemicelluloses. *Plant Biology*, 61(1) : 263.
- Schwarz, W. (2001). The cellulosome and cellulose degradation by anaerobic bacteria. *Applied Microbiology and Biotechnology*, 56 (5-6): 634-649.
- Scully, S. M. and Orlygsson, J. (2014). Recent advances in second generation ethanol production by thermophilic bacteria. *Energies*, 8(1): 1-30.
- Seo, J. K., Park, T. S., Kwon, I. H., Piao, M. Y., Lee, C. H. and Ha, J. K. (2013). Characterization of cellulolytic and xylanolytic enzymes of *Bacillus licheniformis* JK7 isolated from the rumen of a native Korean Goat. *Asian Australaiaian Journal of Animal Science*, 26(1): 50-58.
- Sethi, S., Datta, A., Gupta, B. L. and Gupta, S. (2013). Optimization of cellulase production from bacteria isolated from soil. *International Scholarly Research notices Biotechnology*, 2013.
- Shabeb, M., Younis, M., Hezayen, F. and Nour-Eldein, M. (2010). Production of cellulase in low-cost medium by *Bacillus subtilis* KO strain. *World Applied Sciences Journal*, 8(1) :35-42.
- Shafie, S. M., Masjuki, H. H. and Mahlia, T. M. I. (2014). Rice straw supply chain for electricity generation in Malaysia: Economical and environmental assessment. *Applied Energy*, 135: 299-308.

- Shahriarinnour, M., Abdul Wahab, M. N., Mohamad, R., Mustafa, S. and Ariff, A. (2011). Cyanobacterial biomass as n-supplement to oil palm empty fruit bunch (OPEFB) fibre for improvement of cellulase production by *Aspergillus terreus* in submerged fermentation. *BioResources*, 6(2) : 1696-1706
- Shajahan, S., Moorthy, I. G., Sivakumar, N. and Selvakumar, G. (2016). Statistical modeling and optimization of cellulase production by *Bacillus licheniformis* NCIM 5556 isolated from the hot spring, Maharashtra, India. *Journal of King Saud University-Science*, 29(3): 302-310.
- Shallom, D. and Shoham, Y. (2003). Microbial hemicellulases. *Current Opinion in Microbiology*, 6(3), 219-228.
- Shamsudin, S., Shah, U. K. M., Zainudin, H., Abd-Aziz, S., Kamal, S. M. M., Shirai, Y., et al. (2012). Effect of steam pretreatment on oil palm empty fruit bunch for the production of sugars, *Biomass and Bioenergy*, 36: 280-288.
- Shankar, T. and Isaaiarasu, L. (2011). Cellulase production by *Bacillus pumilus* EWBCM1 under varying cultural conditions. *Middle-East Journal of Scientific Research*, 8(1): 40-45.
- Sharma, M. and Kumar, A. (2013). Xylanases: an overview. *British Biotechnology Journal*, 3(1): 1-28
- Sharma, M. and Kumar Bajaj, B. (2017). Optimization of bioprocess variables for production of a thermostable and wide range pH stable carboxymethyl cellulase from *Bacillus subtilis* MS 54 under solid state fermentation. *Environmental Progress and Sustainable Energy*, 36(4): 1123-1130
- Sharma, S., Sharma, V. and Kuila, A. (2016). Cellulase production using natural medium and its application on enzymatic hydrolysis of thermo chemically pretreated biomass. *3 Biotech*, 6:139. 1-11
- Silva, G. G. D., Guilbert, S. and Rouau, X. (2011). Successive centrifugal grinding and sieving of wheat straw. *Powder technology*, 208(2):266-270.
- Silva, J. C. R., Guimarães, L. H. S., Salgado, J. C. S., Furriel, R. P. M., Polizeli, M. L. T. M., Rosa, J. C., et al. (2013). Purification and biochemical characterization of glucose–cellobiose-tolerant cellulases from *Scytalidium thermophilum*. *Folia Microbiologica*, 58(6): 561-568.
- Simpson-Holley, M., Higson, A. and Evans, G. (2007). Bring on the biorefinery. *Chemical engineering*, 795: 46-48.
- Sindhu, R., Binod, P., Janu, K. U., Sukumaran, R. K. and Pandey, A. (2012). Organosolvent pretreatment and enzymatic hydrolysis of rice straw for the production of bioethanol. *World Journal of Microbiology and Biotechnology*, 28(2), 473-483.

- Singh, J. and Kaur, P. (2012). Optimization of process parameters for cellulase production from *Bacillus* sp. JS14 in solid substrate fermentation using response surface methodology. *Brazilian Archives of Biology and Technology*, 55(4), 505-512.
- Singh, K. and Singh, R. K. (2015). Role of enzymes in fruit juices clarification during processing: a review. *International Journal of Biological Technology*, 6: 114-124.
- Singh, R., Shukla, A., Tiwari, S. and Srivastava, M. (2014). A review on delignification of lignocellulosic biomass for enhancement of ethanol production potential. *Renewable and Sustainable Energy Reviews*, 32: 713-728.
- Singh, S., Dikshit, P. K., Moholkar, V. S. and Goyal, A. (2015). Purification and characterization of acidic cellulase from *Bacillus amyloliquefaciens* SS35 for hydrolyzing *Parthenium hysterophorus* biomass. *Environmental Progress and Sustainable Energy*, 34(3): 810-818.
- Singh, S., Moholkar, V. and Goyal, A. (2014). Optimization of carboxymethylcellulase production from *Bacillus amyloliquefaciens* SS35. *Biotech*, 4(4): 411-424.
- Singhania, R. R., Patel, A. K., Sukumaran, R. K., Larroche, C. and Pandey, A. (2013). Role and significance of beta-glucosidases in the hydrolysis of cellulose for bioethanol production. *Bioresource Technology*, 127: 500-507.
- Singhania, R. R., Sukumaran, R. K., Patel, A. K., Larroche, C. and Pandey, A. (2010). Advancement and comparative profiles in the production technologies using solid-state and submerged fermentation for microbial cellulases. *Enzyme and Microbial Technology*, 46(7): 541-549.
- Singla, A., Paroda, S., Dhamija, S. S., Goyal, S., Shekhawat, K., Amachi, S., et al. (2012). Bioethanol production from xylose: problems and possibilities. *Journal of Biofuels*, 3(1): 1-17.
- Siwar, C., Diana, N., Yasar, M. and Morshed, G. (2014). Issues and challenges facing rice production and food security in the granary areas in the East Coast Economic Region (ECER), Malaysia. *Research Journal of Applied Sciences, Engineering and Technology*, 7(4): 711-722.
- Smidt, E. and Schwanninger, M. (2005). Characterization of waste materials using FTIR spectroscopy: Process monitoring and quality assessment. *Spectroscopy Letters*, 38(3): 247-270.
- Sreedharan, S., Prakasan, P., Sasidharan, S. and Sailas, B. (2016). An overview on fungal cellulases with an industrial perspective. *Journal of Nutrition and Food Sciences*, 6 (1) 461-473

- Sreeja, S., Jeba, M., Sharmila, J., Steffi, T., Immanuel, G. and Palavesam, A. (2013). Optimization of cellulase production by *Bacillus altitudinis* APS MSU and *Bacillus licheniformis* APS2 MSU, gut isolates of fish *Etroplus suratensis*. *International Journal of Advance. Research Technology* 2(4): 401-406.
- Sriariyanun, M., Tantayotai, P., Yasurin, P., Pornwongthong, P. and Cheenkachorn, K. (2016). Production, purification and characterization of an ionic liquid tolerant cellulase from *Bacillus* sp. isolated from rice paddy field soil. *Electronic Journal of Biotechnology*, 19: 23-28.
- SriBala, G., Chennuru, R., Mahapatra, S. and Vinu, R. (2016). Effect of alkaline ultrasonic pretreatment on crystalline morphology and enzymatic hydrolysis of cellulose. *Cellulose*, 23(3): 1725-1740.
- Stephen, J. D., Mabee, W. E. and Saddler, J. N. (2012). Will second- generation ethanol be able to compete with first- generation ethanol? Opportunities for cost reduction. *Biofuels, Bioproducts and Biorefining*, 6(2): 159-176.
- Stips, A., Macias, D., Coughlan, C., Garcia-Gorriz, E. and San Liang, X. (2016). On the causal structure between CO₂ and global temperature. *Scientific Reports*, 6:1-9
- Subramaniyam, R. and Vimala, R. (2012). Solid state and submerged fermentation for the production of bioactive substances: a comparative study. *International Journal of Science Nature*, 3: 480-486.
- Sukumaran, R. K., Singhania, R. R. and Pandey, A. (2005). Microbial cellulases- production, applications and challenges. *Journal of Scientific and Industrial Research*, 64(11): 832.
- Sultana, A. and Kumar, A. (2011). Optimal configuration and combination of multiple lignocellulosic biomass feedstocks delivery to a biorefinery. *Bioresource Technology*, 102(21): 9947-9956.
- Sun, F. and Chen, H. (2008). Organosolv pretreatment by crude glycerol from oleochemicals industry for enzymatic hydrolysis of wheat straw. *Bioresource Technology*, 99(13): 5474-5479.
- Sun, S., Sun, S., Cao, X. and Sun, R. (2016). The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials. *Bioresource Technology*, 199: 49-58.
- Sun, Y., Lin, L., Deng, H., Li, J., He, B., Sun, R., et al. (2008). Structural changes of bamboo cellulose in formic acid. *BioResources*, 3(2): 297-315.

- Sureshkumar, C. and Prathaban, M. (2015). Optimization of process parameters for carboxymethyl cellulase production under submerged fermentation by *Streptomyces lividians*. *Journal of Academia and Industrial Research* 3(8): 371-379.
- Svetlitchnyi, V. A., Kensch, O., Falkenhan, D. A., Korseska, S. G., Lippert, N., Prinz, M., et al. (2013). Single-step ethanol production from lignocellulose using novel extremely thermophilic bacteria. *Biotechnology for Biofuels*, 6(31): 1-15.
- Tadesse, H., & Luque, R. (2011). Advances on biomass pretreatment using ionic liquids: an overview. *Energy and Environmental Science*, 4(10), 3913-3929.
- Taherzadeh, M. J. and Karimi, K. (2007). Acid-based hydrolysis processes for ethanol from lignocellulosic materials: a review. *BioResources*, 2(3): 472-499.
- Tandy, S., Healey, J. R., Nason, M. A., Williamson, J. C., Jones, D. L. and Thain, S. C. (2010). FT-IR as an alternative method for measuring chemical properties during composting. *Bioresource Technology*, 101(14): 5431-5436
- Tengerdy, R. P. and Szakacs, G. (2003). Bioconversion of lignocellulose in solid substrate fermentation. *Biochemical Engineering Journal*, 13(2): 169-179.
- Thomas, L., Larroche, C. and Pandey, A. (2013). Current developments in solid-state fermentation. *Biochemical Engineering Journal*, 81: 146-161.
- Thomsen, M. H. and Haugaard-Nielsen, H. (2008). Sustainable bioethanol production combining biorefinery principles using combined raw materials from wheat undersown with clover-grass. *Journal of Industrial Microbiology and Biotechnology*, 35(5): 303-311.
- Torigoe, K., Hasegawa, S., Numata, O., Yazaki, S., Matsunaga, M., Boku, N., et al. (2000). Influence of emission from rice straw burning on bronchial asthma in children. *Pediatrics International*, 42(2): 143-150.
- Trivedi, N., Gupta, V., Kumar, M., Kumari, P., Reddy, C. R. K. and Jha, B. (2011a). An alkali-halotolerant cellulase from *Bacillus flexus* isolated from green seaweed *Ulva lactuca*. *Carbohydrate polymers*, 83(2): 891-897.
- Tsai, C. T. and Meyer, A. S. (2014). Enzymatic cellulose hydrolysis: enzyme reusability and visualization of β -glucosidase immobilized in calcium alginate. *Molecules*, 19(12): 19390-19406.
- Turner, P., Mamo, G. and Karlsson, E. N. (2007). Potential and utilization of thermophiles and thermostable enzymes in biorefining. *Microbial Cell Factories*, 6(9):1-23.

- USDA (2015). United States Department of Agriculture, Foreign agricultural service
USDA commodity intelligence report database. Retrieved on 15 September,
2017 from
https://www.pecad.fas.usda.gov/highlights/2015/06/Southeast_Asia/Index.htm
- Vaid, S. and Bajaj, B. K. (2016). Production of ionic liquid tolerant cellulase from *Bacillus subtilis* G2 using agroindustrial residues with application potential for saccharification of biomass under one pot consolidated bioprocess. *Waste and Biomass Valorization*, 8(3): 949-964
- van Dyk, J. S., Sakka, M., Sakka, K. and Pletschke, B. I. (2009). The cellulolytic and hemi-cellulolytic system of *Bacillus licheniformis* SVD1 and the evidence for production of a large multi-enzyme complex. *Enzyme and Microbial Technology*, 45(5): 372-378.
- van Dyk, J. S., Sakka, M., Sakka, K. and Pletschke, B. I. (2010). Identification of endoglucanases, xylanases, pectinases and mannanases in the multi-enzyme complex of *Bacillus licheniformis* SVD1. *Enzyme and Microbial Technology*, 47(3): 112-118.
- Van Nguyen, N. and Ferrero, A. (2006). Meeting the challenges of global rice production. *Paddy and Water Environment*, 4(1): 1-9.
- Vanzin, G. F., Madson, M., Carpita, N. C., Raikhel, N. V., Keegstra, K. and Reiter, W. - D. (2002). The mur2 mutant of *Arabidopsis thaliana* lacks fucosylated xyloglucan because of a lesion in fucosyltransferase AtFUT1. *Proceedings of the National Academy of Sciences*, 99(5): 3340-3345.
- Vardon, D. R., Franden, M. A., Johnson, C. W., Karp, E. M., Guarnieri, M. T., Linger, J. G., et al. (2015). Adipic acid production from lignin. *Energy & Environmental Science*, 8(2): 617-628.
- Varga, E., Schmidt, A. S., Réczey, K. and Thomsen, A. B. (2003). Pretreatment of corn stover using wet oxidation to enhance enzymatic digestibility. *Applied Biochemistry and Biotechnology*, 104(1): 37-50.
- Vasudeo, Z. and Lew, C. (2011). Optimization of culture conditions for production of cellulase by a thermophilic *Bacillus* strain. *Journal of Chemistry and Chemical Engineering*, 5(6): 521-527.
- Veith, B., Herzberg, C., Steckel, S., Feesche, J., Ouml, r., Maurer, K. H., et al. (2004). The complete genome sequence of *Bacillus licheniformis* DSM13, an organism with great industrial potential. *Journal of Molecular Microbiology and Biotechnology*, 7(4): 204-211.

- Vyas, A. K., Putatunda, C., Singh, J. and Vyas, D. (2016). Cellulase Production by *Bacillus subtilis* M1 using pretreated groundnut shell based liquid state fermentation. *Biotropia-The Southeast Asian Journal of Tropical Biology*, 23(1): 28-34.
- Walker, L. P. and Wilson, D. B. (1991). Enzymatic hydrolysis of cellulose: an overview. *Bioresource Technology*, 36(1): 3-14.
- Wang, G., Zhang, X., Wang, L., Wang, K., Peng, F. and Wang, L. (2012). The activity and kinetic properties of cellulases in substrates containing metal ions and acid radicals. *Advances in Biological Chemistry*, 2:290-395
- Wang, K., Jiang, J. X., Xu, F. and Sun, R. C. (2009). Influence of steaming pressure on steam explosion pretreatment of Lespedeza stalks (*Lespedeza crytobotrya*): Part 1. Characteristics of degraded cellulose. *Polymer Degradation and Stability*, 94(9): 1379-1388.
- Wang, S. Y., Hu, W., Lin, X. Y., Wu, Z. H. and Li, Y. Z. (2012). A novel cold-active xylanase from the cellulolytic Myxobacterium *Sorangium cellulosum* So9733-1: gene cloning, expression, and enzymatic characterization. *Applied Microbiology and Biotechnology*, 93(4): 1503-1512.
- Wei, G. Y., Gao, W., Jin, I. I. H., Yoo, S. Y., Lee, J. H., Chung, C. H., et al. (2009). Pretreatment and saccharification of rice hulls for the production of fermentable sugars. *Biotechnology and Bioprocess Engineering*, 14(6): 828-834.
- Wenzel, M., Schöning, I., Berchtold, M., Kämpfer, P. and König, H. (2002). Aerobic and facultatively anaerobic cellulolytic bacteria from the gut of the termite *Zootermopsis angusticollis*. *Journal of Applied Microbiology*, 92(1): 32-40.
- Willrodt, C., Hoschek, A., Bühler, B., Schmid, A. and Julsing, M. K. (2015). Decoupling production from growth by magnesium sulfate limitation boosts De Novo limonene production. *Biotechnology and Bioengineering* 113(6): 1036-1314
- Wilson, D. B. (2009). Cellulases and biofuels. *Current Opinion in Biotechnology*, 20(3): 295-299.
- Wood, I. P., Cao, H.-G., Tran, L., Cook, N., Ryden, P., Wilson, D. R., et al. (2016). Comparison of saccharification and fermentation of steam exploded rice straw and rice husk. *Biotechnology for Biofuels*, 9(1) : 193.
- Wood, T. M. and Bhat, K. M. (1988). Methods for measuring cellulase activities. *Methods in Enzymology*, 160: 87-112.

- Wyman, C. E., Dale, B. E., Elander, R. T., Holtzapple, M., Ladisch, M. R. and Lee, Y. Y. (2005). Coordinated development of leading biomass pretreatment technologies. *Bioresource Technology*, 96(18): 1959-1966.
- Xu, J., Gao, Z., Wu, B. and He, B. (2017). Lactose-induced production of a complete lignocellulolytic enzyme system by a novel bacterium *Bacillus* sp. BS-5 and its application for saccharification of alkali-pretreated corn cob. *Cellulose*, 24(5): 2059-2070.
- Xu, Z., Wang, Q., Jiang, Z., Yang, X. and Ji, Y. (2007). Enzymatic hydrolysis of pretreated soybean straw. *Biomass and Bioenergy*, 31(2):162-16
- Yahya, A., Sye, C. P., Ishola, T. A. and Suryanto, H. (2010). Effect of adding palm oil mill decanter cake slurry with regular turning operation on the composting process and quality of compost from oil palm empty fruit bunches. *Bioresource Technology*, 101(22): 8736-8741.
- Yang, B., Dai, Z., Ding, S. Y. and Wyman, C. E. (2011). Enzymatic hydrolysis of cellulosic biomass. *Biofuels*, 2(4):421-449.
- Yang, C.-Y. and Fang, T. J. (2015). Kinetics for enzymatic hydrolysis of rice hulls by the ultrasonic pretreatment with a bio-based basic ionic liquid. *Biochemical Engineering Journal*, 100: 23-29.
- Yang, F., Jin, E.-S., Zhu, Y., Wu, S., Zhu, W., Jin, Y., et al. (2015). A mini-review on the applications of cellulose-binding domains in lignocellulosic material utilizations. *BioResources*, 10(3): 6081-6094.
- Yang, H.-H., Tsai, C.-H., Chao, M.-R., Su, Y.-L. and Chien, S.-M. (2006). Source identification and size distribution of atmospheric polycyclic aromatic hydrocarbons during rice straw burning period. *Atmospheric Environment*, 40(7): 1266-1274.
- Yang, J., Zhang, X., Yong, Q. and Yu, S. (2011). Three-stage enzymatic hydrolysis of steam-exploded corn stover at high substrate concentration. *Bioresource Technology*, 102(7): 4905-4908.
- Yang, W., Meng, F., Peng, J., Han, P., Fang, F., Ma, L., et al. (2014). Isolation and identification of a cellulolytic bacterium from the Tibetan pig's intestine and investigation of its cellulase production. *Electronic Journal of Biotechnology*, 17(6) : 262-267.
- Yin, L. J., Lin, H. H. and Xiao, Z. R. (2010). Purification and characterization of a cellulase from *Bacillus subtilis* YJ1. *Journal of Marine Science and Technology*, 18: 466-471.

- Yu, C. T., Chen, W. H., Men, L. C. and Hwang, W. S. (2009). Microscopic structure features changes of rice straw treated by boiled acid solution. *Industrial Crops and Products*, 29(2): 308-315.
- Yu, G., Yano, S., Inoue, H., Inoue, S., Endo, T. and Sawayama, S. (2010). Pretreatment of rice straw by a hot-compressed water process for enzymatic hydrolysis. *Applied Biochemistry and Biotechnology*, 160(2): 539-551.
- Yu, H.Y. and Li, X. (2015). Alkali-stable cellulase from a halophilic isolate, *Gracilibacillus* sp. SK1 and its application in lignocellulosic saccharification for ethanol production. *Biomass and Bioenergy*, 81(0): 19-25.
- Yunos, N. S. H. M., Baharuddin, A. S., Yunos, K. F. M., Naim, M. N. and Nishida, H. (2012). Physicochemical property changes of oil palm mesocarp fibers treated with high-pressure steam. *BioResources*, 7(4) : 5983-5994.
- Zahoor, S., Javed, M. and Aftab, M. (2011). Cloning and expression of β -glucosidase gene from *Bacillus licheniformis* into *E. coli* BL 21 (DE3). *Biologia*, 66(2) : 213-220.
- Zainudin, M. H. M., Hassan, M. A., Tokura, M. and Shirai, Y. (2013). Indigenous cellulolytic and hemicellulolytic bacteria enhanced rapid co-composting of lignocellulose oil palm empty fruit bunch with palm oil mill effluent anaerobic sludge. *Bioresource Technology*, 147: 632-635.
- Zakaria, M. R., Hirata, S. and Hassan, M. A. (2014). Combined pretreatment using alkaline hydrothermal and ball milling to enhance enzymatic hydrolysis of oil palm mesocarp fiber. *Bioresource Technology*, 169: 236-243.
- Zakaria, M. R., Hirata, S. and Hassan, M. A. (2015). Hydrothermal pretreatment enhanced enzymatic hydrolysis and glucose production from oil palm biomass. *Bioresource Technology*, 176 :142-148.
- Zakaria, M. R., Norraahim, M. N. F., Hirata, S. and Hassan, M. A. (2015). Hydrothermal and wet disk milling pretreatment for high conversion of biosugars from oil palm mesocarp fiber. *Bioresource Technology*, 181: 263-269.
- Zakzeski, J., Bruijninx, P. C. A., Jongerius, A. L. and Weckhuysen, B. M. (2010). The catalytic valorization of lignin for the production of renewable chemicals. *Chemical Reviews*, 110(6): 3552-3599.
- Zambare, V. P., Bhalla, A., Muthukumarappan, K., Sani, R. K. and Christopher, L. P. (2011). Bioprocessing of agricultural residues to ethanol utilizing a cellulolytic extremophile. *Extremophiles*, 15(5): 611-618.

- Zarafeta, D., Kissas, D., Sayer, C., Gudbergsdottir, S. R., Ladoukakis, E., Isupov, M. N., et al. (2016). Discovery and characterization of a thermostable and highly halotolerant GH5 cellulase from an icelandic hot spring isolate. *PLOS ONE*, 11:1-18
- Zeng, R., Hu, Q., Yin, X.-Y., Huang, H., Yan, J.-B., Gong, Z.-W., et al. (2016). Cloning a novel endo-1, 4- β -d-glucanase gene from *Trichoderma virens* and heterologous expression in *E. coli*. *AMB Express*, 6(1): 108.
- Zeng, R., Xiong, P. and Wen, J. (2006). Characterization and gene cloning of a cold-active cellulase from a deep-sea psychrotrophic bacterium *Pseudoalteromonas* sp. DY3. *Extremophiles*, 10(1): 79-82.
- Zhan, F. X., Wang, Q. H., Jiang, S. J., Zhou, Y. L., Zhang, G. M. and Ma, Y. H. (2014). Developing a xylanase XYNZG from *Plectosphaerella cucumerina* for baking by heterologously expressed in *Kluyveromyces lactis*. *BMC Biotechnology*, 14(1): 107.
- Zhang, D., Hegab, H. E., Lvov, Y., Dale Snow, L. and Palmer, J. (2016). Immobilization of cellulase on a silica gel substrate modified using a 3-APTES self-assembled monolayer. *SpringerPlus*, 5(1): 48.
- Zhang, H., Wang, Y. J., Lü, F., Chai, L. N., Shao, L. M. and He, P. J. (2015). Effects of dilute acid pretreatment on physicochemical characteristics and consolidated bioprocessing of rice straw. *Waste and Biomass Valorization*, 6(2) : 217-223.
- Zhang, J., Shi, H., Xu, L., Zhu, X. and Li, X. (2015). Site-directed mutagenesis of a hyperthermophilic endoglucanase cel12b from *Thermotoga maritima* based on rational design. *PLOS ONE*, 10: 1-14
- Zhang, K., Agrawal, M., Harper, J., Chen, R. and Koros, W. J. (2011). Removal of the fermentation inhibitor, furfural, using activated carbon in cellulosic-ethanol production. *Industrial & Engineering Chemistry Research*, 50(24):14055-14060.
- Zhang, K., Pei, Z. and Wang, D. (2016). Organic solvent pretreatment of lignocellulosic biomass for biofuels and biochemicals: a review. *Bioresource Technology*, 199: 21-33.
- Zhang, L. and Sun, X. (2015). Effects of earthworm casts and zeolite on the two-stage composting of green waste. *Waste Management*, 39: 119-129.
- Zhang, M., Ouyang, J., Liu, B., Yu, H., Jiang, T. and Cai, C. (2013). Comparison of hydrolysis efficiency and enzyme adsorption of three different cellulosic materials in the presence of poly(ethylene glycol). *BioEnergy Research*, 6(4): 1252-1259

- Zhang, X., Qin, W., Paice, M. G. and Saddler, J. N. (2009). High consistency enzymatic hydrolysis of hardwood substrates. *Bioresource Technology*, 100(23): 5890-5897.
- Zhang, Y. H. P., Berson, E., Sarkanen, S. and Dale, B. E. (2009). Sessions 3 and 8: pretreatment and biomass recalcitrance: fundamentals and progress. *Applied Biochemistry and Biotechnology*, 153(3): 80-83
- Zhang, Y. H. P., Himmel, M. E. and Mielenz, J. R. (2006). Outlook for cellulase improvement: screening and selection strategies. *Biotechnology Advances*, 24(5): 452-481.
- Zhang, Y. H. P. and Lynd, L. R. (2004). Toward an aggregated understanding of enzymatic hydrolysis of cellulose: noncomplexed cellulase systems. *Biotechnology and Bioengineering*, 88(7):797-824.
- Zhao, C., Lu, X., Deng, Y., Huang, Y. and Liu, B. (2015). Purification and characterization of thermostable cellulase from consortium xm70 in terrestrial hot spring with sugarcane bagasse. *Tropical Journal of Pharmaceutical Research*, 14(4):591-598.
- Zhao, H., Jones, C. L., Baker, G. A., Xia, S., Olubajo, O. and Person, V. N. (2009). Regenerating cellulose from ionic liquids for an accelerated enzymatic hydrolysis. *Journal of Biotechnology*, 139(1): 47-54.
- Zheng, Y., Zhao, J., Xu, F. and Li, Y. (2014). Pretreatment of lignocellulosic biomass for enhanced biogas production. *Progress in Energy and Combustion Science*, 42: 35-53.