



UNIVERSITI PUTRA MALAYSIA

***MOLECULAR MECHANISMS UNDERLYING ANTIPROLIFERATIVE
EFFECT OF TRICYCLOHEXYLPHOSPHINE GOLD (I)
MERCAPTOBENZOATE DERIVATIVES ON HUMAN BREAST AND
OVARIAN CARCINOMA CELL LINES***

ANG KOK PIAN

FPSK(p) 2018 5



**MOLECULAR MECHANISMS UNDERLYING ANTIPROLIFERATIVE
EFFECT OF TRICYCLOHEXYLPHOSPHINE GOLD (I)
MERCAPTOBENZOATE DERIVATIVES ON HUMAN BREAST AND
OVARIAN CARCINOMA CELL LINES**

By

ANG KOK PIAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

October 2017

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

MOLECULAR MECHANISMS UNDERLYING ANTIPROLIFERATIVE EFFECT OF TRICYCLOHEXYLPHOSPHINE GOLD (I) MERCAPTOBENZOATE DERIVATIVES ON HUMAN BREAST AND OVARIAN CARCINOMA CELL LINES

By

ANG KOK PIAN

October 2017

Chairman : Assoc. Prof. Roslida Abdul Hamid @ Abdul Razak, PhD
Faculty : Medicine and Health Sciences

Breast and ovarian cancers are in the list of top three female cancers. Hormonal problem and family genetic background play important role in these two types of cancer. In cancer research, p53 deems to be the key regulator in the apoptosis events (intrinsic and extrinsic). Many studies have been focusing on the therapeutic effects of the metal complexes including gold complexes due to its potential in exhibiting various medical therapeutic benefits. Thus, a series of gold complexes has been synthesized from the precursor tricyclohexylphosphine gold (I) and mercaptobenzoic acid ligands. The Tricyclohexylphosphinegold (I) R-mercaptobenzoate derivatives are yielded at different ligand position of ortho (2), meta (3) and para (4), labelled as CAU2, CAU3 and CAU4, respectively. The antiproliferative effect and their underlying mechanism(s) were investigated on both breast (MCF-7R) and ovarian (A2780) cancer cell lines via various *in vitro* assays. Molecular mechanisms of p53 of both cells upon treatments were analysed via Human p53 signaling pathway RT² profiler PCR array with all other supportive parameters such as cell proliferation, DNA fragmentation, cell invasion, cell cycle analysis, Annexin V/FITC and caspases (3, 8, 9 and 10). All compounds (CAU2, CAU3 and CAU4) exhibited strong cytotoxicity against MCF-7R and A2780 cell lines with IC₅₀ of 8.14 μ M, 7.26 μ M and 9.03 μ M against the former cell lines and 1.19 μ M, 2.28 μ M and 0.785 μ M against the latter cell lines, respectively. Apoptotic cell death was confirmed by DNA fragmentation and Annexin V assay, respectively. The treated compounds also induced caspases (caspase-3/7, -8, -9, -10) expressions at both cancer cells which led to apoptosis. Both types of cancer cells were arrested at S-phase checkpoint upon treatment with the treated compounds. All treated compounds were shown to induce both intrinsic and extrinsic apoptotic pathways, supported and confirmed by the data obtained from

Human p53 signaling pathway RT² profiler PCR array and caspases activities assay, respectively. The compounds were also able to significantly modulate several important gene expressions such as *p53*, *p73* and *Bax* via upregulation whilst simultaneously downregulated key anti-apoptotic gene *bcl-2*. Downregulation of *MDM2* gene has also been observed, as it served as the destructive factor for *p53* gene. The compounds also inhibited the NF- κ B signaling pathway via activation of Lys48-linked polyubiquitination thus led to NF- κ B degradation. Furthermore, the accumulation of reactive oxygen species (ROS) was observed upon the compounds' treatment, indicating the ROS generation thus led to the increment of mitochondrial membrane potential (MMP). Consequently, this led to the increase of cytochrome c releases from mitochondria, manifested by the results obtained by flow cytometric analysis. In conclusion, CAU2, CAU3 and CAU 4 exhibited significant anticancer effects against both breast and ovarian cancer by inducing intrinsic and extrinsic apoptotic cell death, respectively. These findings shed a light for furthering the research in the new discovery of novel chemotherapeutic agents.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doctor Falsafah

KESAN PROLIFERATIF DAN MEKANISME KOMPLEKS EMAS TERHADAP SEL MCF-7 DAN A2780

Oleh

ANG KOK PIAN

Oktober 2017

Pengerusi : Prof. Madya Roslida Abdul Hamid @ Abdul Razak, PhD
Fakulti : Perubatan dan Sains Kesihatan

Kanser payudara and ovary merupakan kanser utama di kalangan wanita. Masalah hormon dan latar belakang genetik keluarga memainkan peranan penting dalam kedua-dua jenis kanser. Dalam penyelidikan kanser, gen p53 dianggap sebagai pengatur utama dalam laluan apoptosis (intrinsik dan ekstrinsik). Banyak kajian telah memberi tumpuan kepada kesan terapeutik kompleks logam termasuk kompleks emas kerana potensinya mempamerkan pelbagai manfaat terapeutik perubatan. Oleh itu, satu siri kompleks emas telah disintesis daripada prekursor emas (I) trisikloheksilfosfina dan ligan asid merkaptobenzoik. Terbitan emas (1) trisikloheksilfosfina R-merkaptobenzoat dihasilkan pada kedudukan ligan yang berbeza iaitu pada kedudukan ortho (2), meta (3) dan para (4), dilabelkan sebagai CAU2, CAU3 dan CAU4. Kesan anti-proliferasi dan mekanisme sebatian-sebatian tersebut telah dikaji ke atas sel kanser payudara (MCF-7R) dan ovari (A2780) melalui pelbagai asai *in vitro*. Mekanisme molekul gen p53 pada kedua-dua sel kanser selepas rawatan dengan terbitan kompleks emas dianalisis melalui pemprofilan RT² PCR laluan isyarat p53 manusia dan kesemua parameter sokongan lain seperti kesan sitotoksik sel, pemecahan DNA, perebakan sel, analisis kitaran sel, Annexin V / FITC dan caspases (3, 8, 9 dan 10). Semua sebatian (CAU2, CAU3 dan CAU4) telah menunjukkan kesan sitotoksik terhadap sel-sel MCF-7R dan A2780 dengan nilai IC₅₀ masing-masing pada 8.14, 7.26 dan 9.03 µM terhadap sel MCF-7R dan 1.19, 2.28 dan 0.785 µM ke atas sel A2780. Kematian sel apoptotik disahkan oleh pemecahan DNA dan asai Annexin V. Semua sebatian yang dirawat menunjukkan keupayaan mengaruh laluan intrinsik dan ekstrinsik apoptosis, dan ini disokong dan disahkan melalui data yang diperolehi daripada pelbagai pemprofilan RT² PCR laluan isyarat p53 manusia dan asai aktiviti caspase. Terbitan kompleks emas juga telah mengaruh ekspresi caspases (caspase-3/7, -8, -9, -10) pada kedua-dua jenis sel kanser dan seterusnya menyebabkan apoptosis. Kedua-dua jenis sel kanser telah menunjukkan

kematian sel pada fasa-S selepas rawatan dengan terbitan kompleks emas. Sebatian-sebatian tersebut juga berupaya memodulasi secara ketara beberapa ekspresi gen yang penting seperti *p53*, *p73* dan *Bax* melalui peningkatan regulasi dan pada masa yang sama menurunkan regulasi gen utama anti-apoptosis *bcl2*. Penurunan regulasi gen *MDM2* juga dipamerkan kerana ia berfungsi sebagai faktor pemusnah gen *p53*. Sebatian-sebatian ini juga merencat laluan isyarat NF- κ B melalui pengaktifan poliubikuitinasi berkaitan Lys48 seterusnya membawa kepada penguraian NF- κ B. Tambahan pula, rawatan menggunakan sebatian-sebatian tersebut juga menyebabkan pengumpulan spesis oksigen reaktif (ROS) yang ditunjukkan melalui penjanaan ROS dan seterusnya menyebabkan peningkatan keupayaan membrane mitokondria (MMP). Akibatnya, ini membawa kepada peningkatan pengeluaran sitokrom c dari mitokondria, yang ditunjukkan oleh hasil yang diperolehi oleh analisis aliran sitometrik. Secara keseluruhannya, CAU2, CAU3 dan CAU4 mempamerkan kesan antikanser yang ketara terhadap kedua sel payudara dan ovari dengan mengaruh kematian sel apoptosis intrinsik dan ekstrinsik. Hasil daripada kajian ini akan memberi harapan untuk meneruskan penyelidikan dalam penemuan baru agen kemoterapi yang ulung.

ACKNOWLEDGEMENTS

First and foremost, it is a genuine pleasure to deliver my deep sense of thanks and gratitude to my supervisor Associate Professor Dr. Roslida, her dedication and keen interest as well as her overwhelming attitude to help her students had contributed for the completion of my works and thesis. Her timely advice, scholarly advice and scientific approach have helped me to a very great extend in the accomplishment of this task.

Besides, I would like to express my sincere thanks to the panel members of my supervisory comittee, Associate Professor Dr. Abdah, Associate Professor Dr. Cheah Yoke Kqueen and Distinguished Professor Dr. Edward Tiekink. Not forgetting their kind hands delivered to me when I was facing tough time at the beginning of my PhD candidature. Without the precious compounds and financial support of the research materials from Professor Dr. Edward Tiekink, it is impossible for me to complete my bench work and deliver my thesis today.

I would like to deliver my deepest thanks to Dr. Samuel Ooi and Dr. Ally Yeo for their assistance and advise during my experimental stage and write up. Without their kind help, it is not possible for me to come to this stage smoothly. Special thanks to Dr. Mohd Islahuddin for his moral support and time too during the difficult time.

My appreciation also extends to my friends and fellow colleagues, Jassy Lim and Dr. Arulselvan who provided me plenty of support, flexibility and advise for the completion of the thesis as well as publication.

Lastly, the greatest appreciation of thanks to be delivered to my family members, especially my dearest mother, a person who bless me all the time and never fail in providing her immeasurable support, encouragement, help and her care to me during the tough time of my candidature.

I certify that a Thesis Examination Committee has met on 26 October 2017 to conduct the final examination of Ang Kok Pian on his thesis entitled "Molecular Mechanisms Underlying Antiproliferative Effect of Tricyclohexylphosphine Gold (I) Mercaptobenzoate Derivatives on Human Breast and Ovarian Carcinoma Cell Lines" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Mohd Nasir bin Mohd Desa, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Huzwah binti Khazaai, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Patimah binti Ismail, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Ingo Ott, PhD

Professor
Braunschweig University of Technology
Germany
(External Examiner)



NOR AINI AB. SHUKOR, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 27 February 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Roslida Abdul Hamid @ Abdul Razak

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Cheah Yoke Kqueen, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Abdah Md Akim, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Edward Richard Tom Tiekink, PhD

Distinguished Professor
Faculty of Science
Sunway University
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Ang Kok Pian, GS30290

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Associate Professor
Dr. Roslida Abd Hamid @ Abd Razak

Signature: _____

Name of Member
of Supervisory
Committee:

Associate Professor
Dr. Abdah Md Akim

Signature: _____

Name of Member
of Supervisory
Committee:

Associate Professor
Dr. Cheah Yoke Kqueen

Signature: _____

Name of Member
of Supervisory
Committee:

Distinguished Professor
Edward Richard Tom Tiekink

TABLE OF CONTENTS

		Page
	ABSTRACT	i
	ABSTRAK	iii
	ACKNOWLEDGEMENTS	v
	APPROVAL	vi
	DECLARATION	viii
	LIST OF TABLES	xiv
	LIST OF FIGURES	xv
	LIST OF ABBREVIATIONS	xix
	CHAPTER	
1	INTRODUCTION	1
	1.1 Research Background	1
	1.2 Problem Statement	2
	1.2.1 Cells selection	3
	1.3 Objectives	4
	1.3.1 General objective	4
	1.3.2 Specific objectives	4
	1.4 Hypothesis	4
2	LITERATURE REVIEW	5
	2.1 Introduction of Cancer	5
	2.1.1 Overview of breast cancer	5
	2.1.2 Pathogenesis of breast cancer	6
	2.1.3 Overview of Ovarian Cancer	7
	2.1.4 Pathogenesis of ovarian cancer	8
	2.2 Apoptosis (Programmed Cell Death)	9
	2.2.1 Introduction of apoptosis	9
	2.2.2 The intrinsic pathway	11
	2.2.3 The extrinsic pathway	11
	2.2.4 Executional phase of apoptosis	12
	2.3 Signaling Pathways in Breast Cancer and Ovarian Cancer	13
	2.3.1 Cyclin dependent kinase	13
	2.3.2 Notch signaling pathway	13
	2.3.3 Wnt signaling pathway	14
	2.3.4 BRK signaling pathway	14
	2.3.5 Human epidermal growth factor receptor (HER) signaling	15
	2.3.6 Ataxiatelangiectasia mutated/Ataxia-telangiectasia mutated and Rad3 related (ATM/ATR) pathway	16
	2.3.7 The tumour suppressor gene - <i>p53</i>	16
	2.3.8 <i>p73</i> gene	17
	2.3.9 Bcl-2 proteins family	17
	2.3.10 The Nuclear Factor-kappaB (NF- kB)	18

2.4	Chemotherapy	19
2.4.1	History of chemotherapy	19
2.4.2	Chemotherapeutic agents classified by mechanism of action	21
2.4.2.1	DNA alkylation and DNA adduct formation	21
2.4.2.2	Nitrosoureas	21
2.4.2.3	Platinum-based agents	21
2.4.2.4	Antimetabolites	21
2.4.2.5	Topoisomerase inhibitor/DNA replication inhibitor	22
2.4.3	Chemotherapy regimen on breast cancer	22
2.4.4	Chemotherapy regime on ovarian cancer	22
2.4.5	Principle of combined regimen in chemotherapy	23
2.5	Medicinal Application of Metal-Based Complexes	23
2.6	Gold-Based Complexes	24
2.6.1	Introduction	24
2.6.2	Chrysotherapy	25
2.6.3	Gold complexes as potential anti-cancer treatment	25
2.6.4	Gold (I) phosphine complexes	26
2.6.5	Tricyclohexylphosphinegold (I) mercaptobenzoate	27
3	METHODOLOGY	30
3.1	Compounds	30
3.2	Cell Lines	30
3.3	Chemical and Reagents	30
3.4	Equipment	30
3.5	Cell Culture	31
3.5.1	Cell thawing	31
3.5.2	Cell maintenance	31
3.5.3	Cell sub-culturing	31
3.5.4	Cell harvesting and cell counting	32
3.5.5	Cell preservation and storage	33
3.5.6	Cell seeding	33
3.6	Drugs preparation	34
3.7	Cell Cytotoxicity Assay	34
3.7.1	Experimental design	34
3.7.2	Cell proliferation assay	35
3.8	Cell invasion assay	36
3.9	DNA Fragmentation	36
3.9.1	DNA extraction	37
3.9.2	Measurement of DNA fragmentation	37
3.10	Intracellular Reactive Oxygen Species (ROS) Measurement	37
3.11	Human Caspases Assay (Caspase-3, Caspase-8, Caspase-9,	37
3.11.1	Experimental design	37
3.11.2	Fluorometric quantitative Caspases ELISA test (Caspase-3, Caspase-8 and Caspase-9)	38
3.11.3	Fluorimetric quatitative ELISA Caspase-10 ELISA test	38

	3.11.4	Qualitative fluorescence morphological assessment of human Caspase-3, 8, and 9	39
	3.12	Measurement of Mitochondrial Cytochrome C	39
	3.13	Cell Apoptosis Analysis	39
	3.14	Cell Cycle Analysis	40
	3.15	Human p53 Signaling Pathway RT ² Profiler PCR Array	40
	3.15.1	Apoptosis induction	40
	3.15.2	RNA extraction	41
	3.15.3	RNA concentration determination	41
	3.15.4	Reverse transcription and cDNA synthesis	42
	3.15.5	Real-time PCR (qPCR)	42
	3.16	Measurement of Intracellular Levels of Reactive Oxygen Species (ROS)	43
	3.17	Measurement of Thioredoxin Reductase (TrxR) Inhibition	43
	3.18	<i>In vitro</i> Lys48 and Lys63 Ubiquitination Assay	43
	3.19	Statistical Analysis	44
4		RESULTS	45
	4.1	Inhibitory Studies	45
	4.1.1	Cytotoxicity analysis of MCF-7R and A2780	45
	4.1.1.1	Cell cytotoxicity assay on MCF-7R cells	45
	4.1.1.2	Cell cytotoxicity assay on A2780 cells	47
	4.1.2	Cell invasion	49
	4.1.3	Thioredoxin reductase (TrxR) activity	54
	4.2	Apoptosis	56
	4.2.1	DNA fragmentation	56
	4.2.2	Cell apoptosis analysis	58
	4.2.3	Cell cycle analysis	62
	4.2.4	Measurement of reactive oxygen species (ROS)	65
	4.2.5	Analysis of Caspases (Caspase-3/7, Caspase-8, Caspase-9 and Caspase-10)	67
	4.2.5.1	Qualitative and quantitative analysis of Caspase-3/7 expression	67
	4.2.5.2	Qualitative and quantitative analysis of Caspase-8 expression	73
	4.2.5.3	Qualitative and quantitative analysis of Caspase-9 expressionz Activation of caspase-9 for the treated cells (MCF-7R and A2780) with test compounds (CAU 2, CAU 3 and CAU4) was analyzed by qualitative and quantitative method.	79
	4.2.5.4	Analysis of Caspase-10 expression	85
	4.2.6	Measurement of mitochondrial cytochrome C	87
	4.2.7	Human p53 signaling pathway RT ² profiler PCR array	90
	4.3	Flow Cytometry Analysis on Ubiquitin Activities (Lys48 and Lys63)	96
	4.3.1	Ubiquitin activities (Lys48 and Lys63) of MCF-7R cells	97

4.3.2	Ubiquitin activities (Lys48 and Lys63) of A2780 cells	100
5	DISCUSSION	103
5.1	Inhibitory Effect of Gold Complexes	103
5.2	Gold Complexes Induces Apoptosis	108
5.2.1	Apoptosis analysis in breast cancer (MCF-7R)	108
5.2.2	Apoptosis analysis in ovarian cancer cells (A2780)	113
5.3	Cell Cycle Analysis	117
5.4	Polyubiquitin Analysis and NF- κ B Activities	118
6	CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	122
6.1	Recommendation for Future Research	122
	REFERENCES	124
	APPENDICES	150
	BIODATA OF STUDENT	166

LIST OF TABLES

Table		Page
2.1	The molecular weight of the studied compounds	29
4.1	IC ₅₀ value of CAU2, CAU3 and CAU4 towards MCF-7R cells in 24 h	45
4.2	IC ₅₀ value of CAU2, CAU3 and CAU4 towards A2780 cells in 24 h	47
4.3	p53-related gene expression of MCF-7R cells upon treated with compound CAU2, CAU3 and CAU4	90
4.4	p53-related gene expression of A2780 cells upon treated with compound CAU2, CAU3 and CAU4	94

LIST OF FIGURES

Figure		Page
2.1	Schematic diagram of breast cancer development	7
2.2	The formation of Type I and Type II serous ovarian carcinoma tumors	9
2.3	Cellular pathways of apoptotic cell death	10
2.4	Summary of the extrinsic and intrinsic pathways of apoptosis event	12
2.5	The chronology development of some anti-cancer drugs from the early days until today	20
2.6	Gold complexes used for rheumatoid arthritis	24
2.7	The chemical structure of $[\text{Au}(\text{dppe})_2]\text{Cl}$. R=Phosphine (Ph)	26
2.8	The precursor of gold complexes, tricyclohexylphosphinegold (I), $\text{Cy}_3\text{Pau}(\text{R-mba})$, R=2, 3, 4	27
2.9	The chemical structure of synthesized tricyclohexylphosphinegold (I) R-mercaptobenzoate, R= 2, 3, 4	28
3.1	The gridded square on the haemocytometer chamber	32
3.2	96-wells plate was used for the cell cytotoxicity assay	35
4.1	Concentration-dependent growth inhibition of CAU2 on MCF-7R cells after 24 h incubation	46
4.2	Concentration-dependent growth inhibition of CAU3 on MCF-7R cells after incubation for 24 h	46
4.3	Concentration-dependent growth inhibition of CAU4 on MCF-7R cells after incubation of 24 h	47
4.4	Concentration-dependent growth inhibition of CAU2 on A2780 cells after incubation for 24 h	48
4.5	Concentration-dependent growth inhibition of CAU3 on A2780 cells after 24 hb incubation	48

4.6	Concentration-dependent growth inhibition of CAU4 on A2780 cells after 24 h incubation	49
4.7	Cell invasion assay conducted using (a) MCF-7R and (b) A2780 cell lines treated with CAU2, CAU3 and CAU4	51
4.8	The Cell Invasion Assay of MCF-7R cells with Matrigel™ Invasion Chamber	52
4.9	The Cell Invasion Assay of A2780 cells with Matrigel™ Invasion Chamber	53
4.10	Thioredoxin reductase assay conducted in cultured (a) MCF-7R and (b) A2780 cells treated with CAU2, CAU3 and CAU4 for 24 hours	55
4.11	DNA fragmentation analysis of MCF-7R cells treated with compound CAU2, CAU3 and CAU4 at the concentration based of the IC50 value respectively	57
4.12	DNA fragmentation analysis of A2780 cells treated with compound CAU2, CAU3 and CAU4 at the concentration based of the IC50 value respectively	58
4.13	The quadrant graphs of cell apoptosis analysis of MCF-7R cells conducted by flow cytometry	60
4.14	The quadrant graphs of cell apoptosis analysis of A2780 cells conducted by flow cytometry	61
4.15	The quadrant graphs of cell cycle analysis of MCF-7R cells conducted by flow cytometry	63
4.16	The quadrant graphs of cell cycle analysis of A2780 cells conducted by flow cytometry	64
4.17	Production of ROS after treatment of MCF-7R (a) and A2780 (b) cells	66
4.18	Fluorescent microscopic analysis of in-situ expression of Caspase3/7 in MCF-7R cells	69
4.19	Fluorescent microscopy analysis of in-situ expression of Caspase-3/7 in A2780 cells	71
4.20	Quantitative analysis of Caspase-3/7 expression in (a) MCF-7R and (b) A2780 cells treated with CAU2, CAU3 and CAU4	72

4.21	Fluorescent microscopy analysis of in-situ expression of Caspase-8 expression in MCF-7R cells	75
4.22	Fluorescent microscopy analysis of in-situ expression of Caspase-8 in A2780 cells	77
4.23	Quantitative analysis of Caspase-8 expression on (a) MCF-7R and (b) A2780 cells treated with CAU2, CAU3 and CAU4	78
4.24	Fluorescent microscopy analysis of in-situ expression of Caspase-9 in MCF-7R cells	81
4.25	Fluorescent microscopy analysis of in-situ expression of Caspase-9 expression in A2780 cells	83
4.26	Quantitative analysis of Caspase-9 expression on (a) MCF-7R and (b) A2780 cells treated with CAU2, CAU3 and CAU4	84
4.27	Quantitative analysis of Caspase-10 expression on (a) MCF-7R and (b) A2780 cells treated with CAU2, CAU3 and CAU4	86
4.28	Cytochrome c measurement in MCF-7R cells	88
4.29	Cytochrome c measurement in A2780 cells	89
4.30	Flow cytometry analysis on the activities of ubiquitin Lys48 in MCF-7R cells	98
4.31	Flow cytometry analysis on the activities of ubiquitin Lys63 in MCF-7R cells	99
4.32	Flow cytometry analysis on the activities of ubiquitin Lys48 in A2780 cells	101
4.33	Flow cytometry analysis on the activities of ubiquitin Lys63 in A2780 cells	102
4.34	Involvement of thioredoxin reductase in the inhibition of cancer cells invasion and metastasis	108
4.35	Summary flow of apoptosis pathway of treated compounds (CAU2, CAU3 and CAU4) in MCF-7R cells	113
4.36	Summary flow of apoptosis pathway of treated compounds (CAU2, CAU3 and CAU4) in A2780 cells	116

4.37	Summary pathway of tested compounds (CAU2, 3 and 4) on MCF-7R cancer cell line	120
4.38	Summary pathway of tested compounds (CAU2, 3 and 4) on A2780 cancer cell line	121



LIST OF ABBREVIATIONS

APAF-1	Apoptosis protease activating factor-1
BRCA1	Tumor suppressor gene BRCA 1
BRCA2	Tumor suppressor gene BRCA 2
BRK	Breast tumor kinase
ATM/ATR	Ataxiatelangiectasia mutated/Ataxia-telangiectasia mutated and Rad related
CAD	Caspase activated DNase
CDK	Cyclin-dependent kinase
CDKI	Cyclin-dependent kinase inhibitors
DISC	Death-induced signaling complex
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
ER	Estrogen receptor
ERK	Extracellular regulated kinase
EtBr	Ethidium bromide
FADD	Fas-associated death domain protein
FasLG	Fas-ligand
FasR	Fas receptor
FBS	Fetal bovine serum
FLICA	Flurochrome inhibitors of caspase
HER2	Human epidermal growth receptor-2
ICAD	Inhibitor caspase activated DNase
ICAM-1	Intracellular adhesion molecule-1
IL	Interleukin
JNK	jun Amino-terminal Kinases
Lys	Lysine
MAPK	Mitogen activated protein kinase
MCP-1	Monocyte chemoattractant protein-1
MDM2	Mouse double minute 2 homolog
mg	Miligram

MIP-1	Macrophage inflammatory protein-1
mM	Milimolar
MMP	Matrix metalloproteinase
mRNA	Messenger ribonucleic acid
mTOR	Mammalian target of rapamycin
MTT	Dimethyl thiazolyl dephenyl tetrazolium
NF- κ B	Nuclear factor kappa-B
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
PDGF B	Platelet derived growth factor subunit B
RIP	Ribosome-inactivating protein
RNA	Ribonucleic acid
ROS	Reactive oxygen species
SD	Standard deviation
TAE	Tris-acetate-EDTA
TAK1	Mitotic Activated Protein Kinase Kinase Kinase
TNF	Tumor necrosis factor
TNFR	Tumor necrosis factor receptor
TRADD	TNF receptor-associated death domain protein
TrxR	Thioredoxin reductase
μ M	Micromolar
uPA	Urokinase-type plasminogen activator
VCAM-1	Vascular cell adhesion molecule-1
VEGF	Vascular endothelial growth factor

CHAPTER 1

INTRODUCTION

1.1 Research Background

The world of 21st century changes the diet lifestyle and living habits of people, which actually causes several health problems. Among the health problems, cancer is considered one of the most major public health issues. Up to date, there are total of 14.1 million of the new cancer cases has been reported and the mortality rate of cancer is 9.1 million at the year of 2012 (Globocan, 2017).

According to the figures obtained from global cancer statistic of the year 2012, three of the top leading cancers in females is breast, ovarian and cervical cancer. Environmental exposure, hormonal and family genetic backgrounds are closely correlated to an increased risk for breast cancer and ovarian.

According to the latest Health Facts (2013) released by the Ministry of Health Malaysia, cancer is one of the top five causes of death in both government and private hospitals. Besides, cancer has overtaken coronary heart disease as the 1st killer in 2014. In Malaysia, the rate of new cancer incidence has been increased from 32,000 in year 2008 to approximately 37,000 cases in year 2012. The mortality rate of cancer in Malaysia also increased from 20,100 death cases in year 2008 to 21,700 death cases in 2012.

Breast cancer is a major global health issue and also the killer among women of all ethnic backgrounds. In Malaysia, breast cancer affected 1 in every 19 Malaysian women whom are diagnosed with breast cancer by the age of 85. Approximately, 4000 women are diagnosed each year and it commonly affects the women aged between 35 to 60 and 40% of the incident rate age d below 50 years old. The risk factors of breast cancer including early menarche or late menopause, late stage at first full term pregnancy, high body mass index after menopause and exposure to ionizing radiation (Almutlaq *et al.*, 2017).

Ovarian cancer is the 4th most common cancer amongst Malaysian women. The statistic shows about 500 women were diagnosed with the disease each and every year. The known risk factors of ovarian cancer including nulliparity, late menopause, early menarche, use of infertility drugs and personal or family history of breast or ovarian cancer (Keng *et al.*, 2015).

Genetic cancer syndromes including mutations of BRCA1 or BRCA2 genes have been associated with an increased risk in both ovarian and breast cancer. Women with BRCA1 and BRCA2 mutations possess an estimated risk of 85% for breast cancer, 20% - 40% of the chances of developing ovarian cancer (Petruccioli *et al.*, 2010).

Several treatments for cancer are usually adopted by most of the medical institutions including radiotherapy, surgical removal and chemotherapy. Surgical resection of the tumour is very risky as it may cause severe pain and may also lead to organs dysfunction, post-operative infection and even death. Radiotherapy kills cancer cells by using the principle of radiation; at the same time, it harms the normal cells due to the accuracy and persistency to target the tumour spot for the delivery of radiation (Sharma *et al.*, 2001). Whereas, the most common therapy is chemotherapy that usually adopts anti-neoplastic drugs in killing rapid dividing cancer cells and further suppresses their proliferation.

In cancer research, the apoptosis and p53 are closely related as most of the time apoptosis is mediated by p53 (Volgerstein *et al.*, 2000). Besides, p53 capable of intervene apoptosis mechanisms of cancer at both intrinsic and extrinsic apoptosis pathways (Haupt *et al.*, 2003). Tumour suppressor gene – p53 also critically involved in cell cycle checkpoint and also activation of caspases that lead to apoptosis (Schuler *et al.*, 2000). Hence it is important for us to study the molecular mechanisms of the treated compounds in related to p53 as well as apoptosis.

Therefore, each of the elements involved in both intrinsic and extrinsic apoptosis pathways are to be studied in this research project. Cytochrome c, *Bcl-2*, *ATM*, *ATR*, *Apaf1*, *Bax*, caspase-3 and -9 are to be involved in intrinsic pathway of apoptosis. Whereby, caspase-3, -8 and -10, Bid, TNF/TNF-R and FAS/FADD are involved in extrinsic pathway of apoptosis (Ooi *et al.*, 2017).

1.2 Problem Statement

Most of the antineoplastic-based chemotherapy drugs are the metal-based drugs. Most commonly used metal-based antineoplastic drugs are made of platinum complexes. Platinum complexes are a family of metal-based drugs showing anti-cancer properties. Cisplatin, the conventional antineoplastic drug of this family is one of the most widely used and most effective cytotoxic agent in the treatment of solid tumours such as breast and ovarian cancers with the cure rate of 70% - 80% (Brezdan *et al.*, 2000; Donzelli *et al.*, 2004; Taguchi *et al.*, 2005). However, there is a limitation in the application of cisplatin to the cancer patients due to the development of resistance by tumour cells (Boulikas and Vougiouka, 2003).

Auronafin, [triethylphosphinegold (I) tetraacetathioglucose] is the gold based anti-inflammatory compound serendipitously discovered for its potential inhibitory activity on malignancies. The discovery of this gold compound inspired the researchers to determine and discover more anti-cancer potential from gold complexes. A hypothesis was made from the presence of phosphine ligand on gold complexes, which demonstrated significant cytotoxicity on cancer cells (Tiekink, 2002).

Tricyclohexylphosphine gold (I) R-mercaptobenzoate (R= 2, 3, 4) complexes were synthesized with the inclusion of cyclophosphine ligand and mercaptobenzoic acid ligand at different location on the structure. Based on the aforementioned justification, tricyclohexylphosphine gold (I) R-mercaptobenzoate (R= 2, 3, 4) complexes were successfully synthesized to investigate their potential in anticancer by determining their apoptosis pathway profile against human breast adenocarcinoma cell line (MCF-7R) and human ovarian carcinoma cell lines (A2780) in this current study. R is referred to the ligand position at the ortho (2), meta (3) and para (4), respectively.

1.2.1 Cells selection

Human ovarian carcinoma cell line (A2780) and human breast adenocarcinoma cell line - drug resistance (MCF-7R) were selected to be used as cell culture model in this experiment.

In this study, cisplatin was used as the drug control and A2780 cells are sensitive towards the drug control. Besides, A2780 is a very common cell line used in most of the ovarian cancer research yet can be compared the efficacy and sensitivity of the cancer cells with treated compounds (Hunakova *et al.*, 2005).

Apart from this, MCF-7R was selected to be the cell line used for breast cancer study, where this cell line is multi-drug resistant. This cell line is selected to compare the sensitivity of the cells with drug control (cisplatin) over the treated compounds (Bichat *et al.*, 1997). Besides, the chemoresistance mechanism could be elucidated too from the study.

1.3 Objectives

1.3.1 General objective

To investigate the anticancer potential of tricyclohexylphosphine gold (I) R-mercaptobenzoate (R= 2, 3, 4) complexes against human breast adenocarcinoma cell lines (MCF7-R and human ovarian carcinoma cell lines (A2780) and its underlying molecular mechanism(s).

1.3.2 Specific objectives

1. To determine and compare IC₅₀ values of tricyclohexylphosphine gold (I) R-mercaptobenzoate (R=2,3,4) compounds against MCF-7R and A2780 cell lines, respectively.
2. To evaluate the cell cycle analysis and mode of cells death on MCF-7R and A2780 cell lines when treated with tricyclohexylphosphine gold (I) R-mercaptobenzoate (R=2,3,4).
3. To assess the influence of caspases (caspase-3, caspase-8, caspase-9, caspases-10), cytochrome c and inducer of apoptosis in the pathway of apoptosis
4. To investigate the activity of Nuclear Factor-kappa B (NF- κB) in apoptosis through ubiquitin-detection as well as its tyrosine kinase activities
5. To manifest the cross talk signaling events amongst the biomarkers assessments with gene expression analysis of p-53 pathway via Human p-53 RT² Profiler PCR Array.

1.4 Hypothesis

Based on the principles and knowledge established on aurofin and platinum complexes, thus it is anticipated that tricyclohexylphosphine gold (I) R-mercaptobenzoate (R= 2, 3, 4) series may possess anti-proliferative activities on human breast adenocarcinoma cell lines and human ovarian carcinoma cell lines by inducing both intrinsic and extrinsic apoptosis pathway via modulation of multiple gene expressions

REFERENCES

- Aabo, K., Adams, M., Adnitt, P. (1998). Chemotherapy in advanced ovarian cancer: four systematic meta-analyses of individual patient data from 37 randomized trials. Advanced Ovarian Cancer Trialists' Group. *British Journal of Cancer*. 78: 1479–87
- Adhikary, A., Mohanty, S., Lahiry, L., Hossain, D. M. S., Chakraborty, S., Das, T. (2010). Theaflavins retard human breast cancer cell migration by inhibiting NF- κ B via p53-ROS cross-talk. *FEBS Letters*. 584: 7 – 14.
- Almutlaq, B. A., Almuazzi, R. F., Almuhayfir, A. A., Alfouzan, A. M., Alshammari, B. T., AlAnzi, H. F., Ahmed, H. G. (2017). Breast cancer in Saudi Arabia and its possible risk factors. *Journal of Cancer Policy*. 12: 83-89.
- Al-Saran, N., Subash-Babu, P., Al-Nouri, D. M., Alfawaz, H. A., and Alshatwi, A. A. (2016). Zinc enhances CDKN2A, pRb1 expression and regulates functional apoptosis via upregulation of p53 and p21 expression in human breast cancer MCF-7R cell. *Environmental Toxicology and Pharmacology*. 47: 19-27.
- Altieri, D. C. (2013). Targeting surviving in cancer. *Cancer Letters*. 332: 225-228.
- Anand, P., Kunnumakkara, A. B., Sundaram, C., Harikumar, K. B., Tharakan, S. T., Lais O. S. (2008). Cancer is a preventable disease that requires major lifestyle changes. *Pharmaceutical Research*. 25:2097–116.
- Anestal, K., Prast-Nielsen, S., Cenas, N., and Arner, E. S. (2008). Cell death by SecTRAPs: thioredoxin reductase as a prooxidant killer of cells. *PLoS One* 3: e1846.
- Arora, S., and Tandon, S. (2015). DNA fragmentation and cell cycle arrest: a hallmark of apoptosis induced by *Ruta graveolens* in human colon cancer cells. *Homeopathy*. 104: 36-47.
- Aubele, M., Auer, G., Walch, A.K., Munro, A., Atkinson, M. J., Braselmann, H., Fornander, T., Barlett, J.M. (2007)., PTK (protein tyrosine kinase)-6 and HER2 and 4, but not HER1 and 3 predict long-term survival in breast carcinomas. *British Journal of Cancer*. 96: 801–807.
- Baguley, B. C. (2002). A brief history of cancer chemotherapy. *Academic Press*.
- Barar, J., and Omid, Y. (2013). Dysregulated pH in tumour microenvironment checkmates cancer therapy. *Bioimpacts*. 3: 149-162.
- Bargou, R. C., Wagener, C., Bommert, K., Mapara, M. Y., Daniel, P. T., Arnold, W., Dietel, M., Guski, H., Feller, A., Royer, H. D., Dorken, B. (1996).

Overexpression of the death-promoting gene bax-a which is downregulated in breast cancer restores sensitivity to different apoptotic stimuli and reduces tumour growth in SCID mice. *Journal of Clinical Investigation*. 97: 2651 – 2659.

- Barker, K.T., Jackson, L.E., Crompton M. R. (1997). BRK tyrosine kinase expression in a high proportion of human breast carcinomas. *Oncogene*. 15: 799–805.
- Bast, R.C., Hennesy, B., Mills, G.B. (2009). The biology of ovarian cancer: new opportunities for translation. *Nature Reviews Cancer*. 9: 415-428.
- Becker, K., Gromer, S., Schirmer, R. H., Muller, S. (2000). Thioredoxin reductase as a pathophysiological factor and drug target. *European Journal of Biochemistry*. 20: 6118 – 6125.
- Becker, S., Dossus, L., & Kaaks, R. (2009). Obesity related hyperinsulinaemia and hyperglycaemia and cancer development. *Archives of Physiology and Biochemistry*, 115(2), 86–96.
- Benedek, T. G. (2004). The history of gold therapy in tuberculosis. *Journal of the History of Medicines and Allied Sciences*. 59: 50-89.
- Bhatia, M., T.C. Karlenius, T.C., G. Di Trapani, G., K.F. Tonissen, K.F. (2013). The Interaction Between Redox and Hypoxic Signalling Pathways in the Dynamic Oxygen Environment of Cancer Cells (Ed). *Carcinogenesis* (pp. 125-150). Croatia, Rijeka: InTech.
- Bhatia, M., McGrath, K. L., Di Trapani, G., Charoentong, P., Shah, F., King, M. M., Clarke, F. M., and Tonissen, K. F. (2016). The thioredoxin system in breast cancer cell invasion and migration. *Redox Biology*. 8: 68-78.
- Bichat, F., Mouawad, R., Solis-Recendez, G., Khayat, D., Bastian, G. (1997). Cytoskeleton alteration in MCF7R cells, a multidrug resistant human breast cancer cell line. *Anticancer Research*. 17; 3393-3401.
- Black, D. (1994). Familial breast cancer. BRCA1 down, BRCA2 to go. *Current Biology*. 4:1023 – 1024.
- Blanco, E., Ferrari, M. (2004). Emerging nanotherapeutic strategies in breast cancer. *Breast (Edinburgh, Scotland)* 23: 10–18.
- Bohgaki, M., Tsukiyama, T., Nakajima, A., Maruyama, S., Watanabe, M., Koike, T., Hatakeyama, S. (2008). Involvement of Ymer in suppression of NF- κ B activation by regulated interaction with lysine-63-linked polyubiquitin chain. *Biochimica et Biophysica Acta*. 1783: 826-837.
- Bonnefoy-Berard, N., Aouacheria, A., Verschelde, C., Quemeneur, L., Marcais,

- A., Marvel, J. (2004). Control of proliferation by Bcl-2 family members, *Biochimica et Biophysica Acta (BBA)* 1644: 159–168.
- Boulikas, T., Vougiouka, M. (2003). Cisplatin and platinum drugs at the molecular level (Review). *Oncology Reports*. 10: 1663–1682.
- Bowtell, D.D. (2010). The genesis and evolution of high-grade serous ovarian cancer. *Nature Reviews Cancer*. 10: 803–808.
- Brachova, P., Thiel, K. W. and Leslie, K. K. (2013). The consequence of oncomorphic TP53 mutations in ovarian cancer. *International Journal of Molecular Science*. 14: 19257-19275.
- Brezdan, C. B., Phillips, K. A., Abdolle, M., Bunston, T., Tannock, I. F. (2000). Cognitive function in breast cancer patients receiving adjuvant chemotherapy. *Journal of Clinical Oncology*. 18: 2695-2701.
- Brunelle, J. K., & Letai, A. (2009). Control of mitochondrial apoptosis by the Bcl-2 family. *Journal of Cell Science*, 122(4), 437–441.
- Bufalo, D. D., Biroccio, A., Trisciuglio, D., Bruno, T., Floridi, A., Aquino, A., Zupi, G. (2002). Bcl-2 has differing effects on the sensitivity of breast cancer cells depending on the antineoplastic drug used. *European Journal of Cancer*. 38: 2455 – 2463.
- Cadenas, C., Franckenstein, D, Schmidt, M., Gehrmann, M., Hermes, M., Geppert, B., Schormann, W., Maccour, L. J., Schug, M., Schumann, A., Wilhelm, C., Freis, E., Ickstadt, K., Rahnenfuler, J.m Baumbach, J., Sickmann, A., Hengstler, J. G. (2010). Role of thioredoxin reductase 1 and thioredoxin interacting protein in prognosis of breast cancer. *Breast Cancer Research*. 12: R44.
- Calabro, V., Mansueto, G., Parisi, T., Vivo, M., Calogero, R. A., La Mantia, G. (2001). The human MDM2 Oncoprotein increases the Transcriptional activity and the protein level of the p53 Homolog p63. *Journal of Biological Chemistry*, 277(4), 2674–2681.
- Campbell, T. L., Quadriatero, J. (2016). Data on skeletal muscle apoptosis, autophagy, and morphology in mice treated with doxorubicin. *Data in Brief*. 7: 786-793.
- Caruso, F., Villa, R., Rossi, M., Pettinari, C., Paduano, F., Pennati, M., Daidone, M. G., Zaffaroni, N. (2007). Mitochondria are primary targets in apoptosis induced by the mixed phosphine gold species chlorotriphenylphosphine-1,3-bis(diphenylphosphino) propanegold(I) in melanoma cell lines. *Biochemical Pharmacology*. 73: 773–781.

- Cattaruzza, L., Fegona, D., Mongiat, and Ronconi, L. (2010). Antitumour activity of gold(III)-dithiocarbamate derivatives on prostate cancer cells and xenografts. *International Journal of Cancer*. 128: 206-215.
- Chatterjee, A., Chang, X., Sen, T., Ravi, R., Bedi, A., and Sidransky, D. (2010). Regulation of p53 family member Isoform Np63 by the nuclear factor- κ B targeting Kinase I B Kinase. *Cancer Research*, 70(4), 1419–1429.
- Chen, E.Y., Mehra, K., Mehrad, M., Ning, G., Miron, A., Mutter, G.L., Monte, N., Quade, B., McKeon, F.D., Yassin, Y., Crum, C.P. (2010). Secretory cell outgrowth, PAX2 and serous carcinogenesis in the fallopian tube. *J. Pathol.* 222: 110–116
- Chen, T. C., Yu, M. C., Chien, C. C., Wu, M. S., Lee, Y. C., and Chen, Y. C. (2016). Nilotinib reduced the viability of human ovarian cancer cells via mitochondria-dependent apoptosis, independent of JNK activation. *Toxicology in vitro*. 31: 1-11.
- Chen, X., Shi, X., Zhao, C., Li, X., Lan, X., Liu, S., Huang H. (2014). Anti-rheumatic agent auranofin induced apoptosis in chronic myeloid leukemia cells resistant to imatinib through both Bcr/ABI-dependent and -independent mechanisms. *Oncotarget*. 5: 9118-9132.
- Chen, X., Zhou, H. J., Huang, Q., Lu, L., Min, W. (2014). Novel action and mechanism of auranofin in inhibition of vascular endothelial growth factor receptor-3-dependent lymphangiogenesis. *Anti-cancer Agents in Medicinal Chemistry*. 14: 946-954.
- Chene, G., Dauplat, J., Robin, N. R., Cayre, A., Llorca, F. P. (2013). Tu-be or not tu-be: That is the question. . . About serous ovarian carcinogenesis. *Critical Reviews in Oncology/Haematology*. 88: 134 – 143.
- Chiang, C. T., Chu, W. K., Chow, S. E., and Chen, J. K. (2009). Overexpression of delta Np63 in a human nasopharyngeal carcinoma cell line downregulates CKIs and enhances cell proliferation. *Journal of Cellular Physiology*. 219: 117-122.
- Choi, H.-N., Jin, H.-O., Kim, J.-H., Hong, S.-E., Kim, H.-A., Kim, E.-K., Noh, W. C. (2013). Inhibition of S6K1 enhances glucose deprivation-induced cell death via downregulation of anti-apoptotic proteins in MCF-7R breast cancer cells. *Biochemical and Biophysical Research Communications*. 432(1): 123–128.
- Circu, M. L., & Aw, T. Y. (2010). Reactive oxygen species, cellular redox systems, and apoptosis. *Free Radical Biology and Medicine*, 48(6), 749–762.

- Cleator S, Ahamed E, Coombes R, Palmieri CA. Update on the treatment of patients with hormone receptor-positive breast cancer. *Clinical Breast Cancer* 2009;2009(Suppl. 1): S6–S17.
- Clemons, M., Goss, P. E. (2000). Estrogen and the risk of breast cancer. cohorts of twins from Sweden, Denmark, and Finland. *New England Journal of Medicine*. 343:78–85.
- Cortes, J., O'Shaughnessy, J., Loesch, D. (2011). Eribulin monotherapy versus treatment of physician's choice in patients with metastatic breast cancer (embrace): a phase 3 open-label randomised study. *Lancet*. 377: 914–923.
- Cory, S., Adams, J. M. (2002). The Bcl2 family: regulators of the cellular life-or-death switch. *Nature Reviews Cancer*. 2: 647–656.
- Cruet, H. S., Glynn, M. T., Murillo, L. S., Coyne, S., Carty, M. P. (2008). Enhanced DNA-PK-mediated RPA2 hyperphosphorylation in DNA polymerase β -deficient human cells treated with cisplatin and oxaliplatin. *DNA Repair*. 7: 582 – 596.
- Cui, Y., Lu, P., Song, G., Liu, Q., Zhu, D., & Liu, X. (2016). Involvement of PI3K/Akt, ERK and p38 signaling pathways in emodin-mediated extrinsic and intrinsic human hepatoblastoma cell apoptosis. *Food and Chemical Toxicology*, 92, 26–37.
- Day, T. W., Huang, S., & Safa, A. R. (2008). C-fLIP knockdown induces ligand-independent DR5-, FADD-, caspase-8-, and caspase-9-dependent apoptosis in breast cancer cells. *Biochemical Pharmacology*, 76(12), 1694–1704.
- De Graeff, P., Criins, A. P., de Jong, S., Boezen, M., Post, W. J., de Vries, E. G., van der Zee, A. G., de Bock, G. H. (2009). Modest effect of p53, EGFR and HER-2/neu on prognosis in epithelial ovarian cancer: a meta-analysis. *British Journal of Cancer* 101: 149–159.
- Denamur, S., Boland, L., Beyaert, M., Verstraeten, S. L., Fillet, M., Tulkens, P.M., Bontemps, F., and Mingeot-Leclercq, M. P. (2016). Subcellular mechanisms involved in apoptosis induced by aminoglycoside antibiotics: Insights on p53, proteasome and endoplasmic reticulum. *Toxicology and Applied Pharmacology*. 30: 24-36.
- Derry, J. J., Prins, G. S., Ray, V., Tyner, A. L. (2003). Altered Localization and Activity of the Intracellular Tyrosine Kinase BRK/Sik in Prostate Tumour Cells. *Oncogene*. 22: 4212–4220.
- Deryugina, E. I., Quigley, J.P. (2006). Matrix metalloproteinases and tumour metastasis. *Cancer Metastasis Reviews*. 25: 9-34.

- DeVita, V. T., and Chu, E. (2008). A history of cancer chemotherapy. *Cancer Research*. 68: 8643 – 8653.
- Donzelli, E., Carfi, M., Miloso, M., Strada, A., Galbiati, A., Bayssas, M., Griffon-Etienne, G., Caveletti, G. (2004). Neurotoxicity of platinum compounds: comparison of the effects of cisplatin and oxaliplatin on the human neuroblastoma cell line SH-SY5Y. *Journal of neuro-oncology*. 67: 65-73.
- Du, G. J., Wang, C. Z., Qi, L. W., Zhang, Z. Y., Calway, T., He, T. C., Du, W., and Yuan, C. S. (2012). Caspase-mediated pro-apoptotic interaction of panaxadiol and irinotecan in human colorectal cancer cells. *Journal of Pharmacy and Pharmacology*. 64: 727-734.
- Dutta, J., Fan, Y., Gupta, N., Fan, G., Gélinas, C. (2006). Current insights into the regulation of programmed cell death by NF-kappaB. *Oncogene*. 25: 6800–6816.
- Easty, D. J., Mitchell, P.J., Patel, K., Flørenes, V. A., Spritz, R. A. (1997). Loss of expression of receptor tyrosine kinase family genes PTK7 and SEK in metastatic melanoma. *International Journal of Cancer*. 71:1061–1065.
- Elmore, S. (2007). Apoptosis: A review of programmed cell death. *Toxicologic Pathology*. 35: 495-516.
- Eng, K. H., Hanlon, B. M., Bradley, W. H., and Szender, J. B. (2015). Prognostic factors modifying the treatment-free interval in recurrent ovarian cancer. *Gynecologic Oncology*. 139: 228-235.
- Fader, A. N., Arriba, L. N., Frasure, H. E., & von Gruenigen, V. E. (2009). Endometrial cancer and obesity: Epidemiology, biomarkers, prevention and survivorship. *Gynecologic Oncology*, 114(1), 121–127.
- Fan, Y., Shi, Y., Liu, S., Mao, R., An, L., Zhao, Y., Zhang, H., Zhang, F., Xu, G., Qin, J., and Yang, J. (2012). Lys⁴⁸-linked TAK1 polyubiquitination at lysine-72 downregulates TNF- α induced NF- κ B activation via mediating TAK1 degradation. *Cellular Signaling*. 24: 1381-1389.
- Farina, A. R., Cappabianca, L., Desantis, G., Ianni, N. D., Ruggieri, P. D., Ragone, M., *et al.* (2011). Thioredoxin stimulates MMP-9 expression, deregulates the MMP-9/TIMP-1 equilibrium and promotes MMP-9 dependent invasion in human MDA-MB-231 breast cancer cells. *FEBS Letter*.
- Farina, A. R., Tacconelli, A., Cappabianca, L., DeSantis, G., Gulino, A., and Mackay, A. R. (2003). Thioredoxin inhibits microvascular endothelial capillary tubule formation. *Experimental Cell Research*. 291: 474-483.

- Farina, A. R., Tacconelli, A., Cappabianca, L., Masciulli, M. P., Holmgren, A., Beckett, G. J., *et al.* (2001). Thioredoxin alters the matrix metalloproteinase/tissue inhibitors of metalloproteinase balance and stimulates human SK-N-SH neuroblastoma cell invasion. *European Journal of Biochemistry*. 268: 405-413.
- Farrell, N. (2003). Metal complexes as drugs and chemotherapeutic agents. *Comprehensive Coordination Chemistry II*. 9: 809-840.
- Fatah, T. M. A. A., Arora, A., Mosely, P., Coveney, C., Perry, C., Johnson, K., Kent, C., Ball, G., Chan, S., Madhusudan, S. (2014). ATM, ATR and DNA-PKcs expressions correlate to adverse clinical outcomes in epithelial ovarian cancers. *BBA Clinical*. 2: 10 – 17.
- Ferlay, J., Soerjomataram, I., Ervik, M., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D. M., Forman, D., Bray, F. (2014). GLOBOCAN 2012 v1.1, Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. Lyon, France: International Agency for Research on Cancer.
- Ferlay, J., Steliarova-Foucher, E., Lortet-Tieulent, J., Rosso, S., Coebergh, J. W., Comber, H., Forman, D., Bray, F. (2013). Cancer incidence and mortality patterns in Europe: estimates for 40 countries in 2012. *European Journal of Cancer*. 49: 1374–1403.
- Filkelstein, A. E., Burrone, O. R., Walt, D. T., Misher, A. (1977). Effect of auranofin on DNA and protein synthesis in human lymphocytes. *Journal of Rheumatology*. 4: 245 – 251.
- Fillat, M. F., Gimeno, M. C., Laguna, A., Latorre, E., Ortego, L., and Villacampa, M. D. (2011). Synthesis, structure and bactericide activity of (Aminophosphone) gold(I) thiolate complexes. *European Journal of Inorganic Chemistry*. 1487-1495.
- Fiskus, W., Saba, N., Shen, M., Ghias, M., Liu, J., Gupta, S. D., Bhalla, K. N. (2014). Auranofin induces lethal Oxidative and Endoplasmic Reticulum stress and exerts potent preclinical activity against chronic lymphocytic leukemia. *Cancer Research*, 74(9), 2520–2532.
- Freed-Pastor, W. A., Prives, C. (2012). Mutant p53: one name, many proteins. *Genes & Development*. 26:1268 – 86.
- Fries, J. F., Bloch, D., Spitz, P., Mitchell, D. M. (1985). Cancer in rheumatoid arthritis: a prospective long-term study of mortality. *American Journal of Medicine*. 78: 56 - 69.

- Gadducci, A., Guerrieri, M. E., and Genazzani, A. R. (2012). New insights on the pathogenesis of ovarian carcinoma: molecular basis and clinical implications. *Gynecology and Endocrinology*. 28: 582-586.
- Gallahan, D., Callahan, R. (1987). Mammary tumourigenesis in feral mice: identification of a new int locus in mouse mammary tumour virus (Czech II)-induced mammary tumours. *Journal of Virology*. 61:66–74
- Gallegos, J. R., Litersky, J., Lee, H., Sun, Y., Nakayama, K., and Lu, H. (2007). SCF TrCP1 Activates and Ubiquitylates tAp63. *Journal of Biological Chemistry*. 1: 66–75.
- Gan DD, Macaluso M, Cinti C, Khalili K, Giordano A. How does a normal human cell become a cancer cell? *J Exp Clin Cancer Res* 2003; 22:509–16
- Gemignani, M. L., Armstrong, D. K. (2014). Breast cancer. *Gynaecologic Oncology*. 132: 264 – 267.
- Gandin, V., Fernandes, A. P., Rigobello, M. P., Dani, B., Sorrentino, F., Tisato, F., Bjornstedt, M., Bindoli, A., Sturaro, A., Rella, R., Marzano, C. (2010). Cancer cell death induced by phosphine gold(I) compounds targeting thioredoxin reductase. *Biochemical Pharmacology*. 79: 90-101.
- Giannakeas, V., Sopik, V., Narod, S. A. (2016). A model for ovarian cancer progression based on inherent resistance. *Gynecologic Oncology*. 142: 484-489.
- Globocan. (2012). Estimate cancer incidence, mortality & prevalence worldwide in 2012: Retrieved from http://globocan.iarc.fr/Pages/fact_sheets_cancer.aspx
- Gloss, B.S., Samimi, G. (2014). Epigenetic biomarkers in epithelial ovarian cancer. *Cancer Letters*. 342: 257 – 263.
- Grady, W. M., Pritchard, C. C. (2014). Molecular alterations and biomarkers in colorectal cancer. *Toxicologic Pathology*. 42: 124-139.
- Graziano, V., De Laurenzi, V. (2011). Role of p63 in cancer development. *Biochemica et Biophysica Acta*. 1816: 57-66.
- Schuler, M., Bossy-Wetzel, E., Goldstein, J. C., Fitzgerald, P., Green, D. R. (2000). *Journal of Biological Chemistry*. 275: 7337-7345.
- Gromer, S., Arscott, L. D., Williams, C. H. Jr., Schirmer, R. H., Becker, K. (1998). Human placenta thioredoxin reductase. Isolation of the selenoenzyme, steady state kinetics, and inhibition by therapeutic gold compounds. *Journal of Biological Chemistry*. 32: 20096 -20101.

- Gross, A., McDonnell, J. M., Korsmeyer, S. J. (1999). Bcl-2 family members and the mitochondria in apoptosis. *Genes & Development*. 13: 1899–1911
- Gupta, S., Kass, G.E., Szegezdi, E., Joseph, B. (2009). The mitochondrial death pathway: a promising therapeutic target in Diseases. *Journal of Cellular and Molecular Medicine*. 13: 1004–1033.
- Haber, G., Ahmed, N. U., Pekovic, V. (2012). Family history of cancer and its Association with breast cancer risk perception and repeat mammography. *American Journal of Public Health*. 102: 2322-2329.
- Hajra, K. M., & Liu, J. R. (2004). Apoptosome dysfunction in human cancer. *Apoptosis*, 9(6), 691–704.
- Halliwell, B. (2012). Free radicals and antioxidants: updating a personal view. *Nutrition Reviews*. 70: 257-265.
- Han, J., Zhang, L., Guo, H., Wysham, W. Z., Roque, D. R., Willson, A. K., Bae-Jump, V. L. (2015). Glucose promotes cell proliferation, glucose uptake and invasion in endometrial cancer cells via AMPK/mTOR/S6 and MAPK signaling. *Gynecologic Oncology*, 138(3), 668–675.
- Hansen, T. M., Rossi, M., Roperch, J. P., Ansell, K., Simpson, K., Taylor, D., Melino, G. (2007). Itch inhibition regulates chemosensitivity in vitro. *Biochemical and Biophysical Research Communications*. 1: 33–36.
- Harms, P. W., Hocker, T. L., Zhao, L., Chan, M. P., Andea, A. A., Wang, M., Harms, K. L., Wang, M. L., Carskadon, S., Palanisamy, N., Fullen D. R. (2016). Loss of p16 expression and copy number changes of CDKN2A in a spectrum of spitzoid melanocytic lesions. *Human Pathology*. 58: 152-160.
- Harvey, A. J., Pennington, C. J., Porter, S., Burmi, R. S., Edwards, D. R., Court W., Eccles, S. A., Crompton, M. R. (2009). Brk protects breast cancer cells from autophagic cell death induced by loss of anchorage. *American Journal of Pathology*. 175: 1226–1234.
- Haupt, S., Berger, M., Goldberg, C., Haupt, Y. (2003). Apoptosis – the p53 network. *Journal of Cell Science*. 116: 4077-4085.
- Hellfritsch, J., Kirsch, J., Schneider, M., Fluege, T., Wortmann, M., and Frijhoff, J. (2015). Knockout of mitochondrial thioredoxin reductase stabilizes prolyl hydroxylase 2 and inhibits tumour growth and tumour-derived angiogenesis. *Antioxidant and Redox Signaling*. 22: 938-950.
- Hennesy, B. T. and Markman, M. (2009). Development of novel agents for ovarian cancer. *Updates on Cancer Therapeutics*. 3: 119 – 132.

- Hernández-Reséndiz, I., Román-Rosales, A., García-Villa, E., López-Macay, A., Pineda, E., Saavedra, E., Rodríguez-Enríquez, S. (2015). Dual regulation of energy metabolism by p53 in human cervix and breast cancer cells. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*. 12: 3266–3278.
- Hoke, G. D., Rush, G. F., Mirabelli, C. K. (1989). The mechanism of acute cytotoxicity of triethylphosphine gold(I) complexes: III. Chlorotriethylphosphine gold(I)-induced alterations in isolated rat liver mitochondrial function. *Toxicology and Applied Pharmacology*. 99: 50 – 60.
- Hsu, P. P., & Sabatini, D. M. (2008). Cancer cell metabolism: Warburg and beyond. *Cell*. 5: 703–707.
- Huang, C., K. Jacobson, K., M.D. Schaller, M. D. (2004). A role for JNK-paxillin signaling in cell migration. *ABBV Cell Cycle*. 3: 4–6.
- Hunakova, L., Gronesova, P., Horvathova, E., Chalupa, I., Cholujova, D., Duraj, J., Sedlak, J. (2014). Modulation of cisplatin sensitivity in human ovarian carcinoma A2780 and SKOV3 cell lines by sulforaphane. *Toxicology Letters*. 230: 479-486.
- Hunters, A. M., LaCasse, E. C., and Korneluk, R. G. (2007). The inhibitors of apoptosis (IAPs) as cancer targets. *Apoptosis*. 129: 1543-1568.
- Idogawa, M., Ohashi, T., Sasaki, Y., Maruyama, R., Kashima, L., Suzuki, H. (2014). Identification and analysis of large intergenic non-coding RNAs regulated by p53 family members through a genome-wide analysis of p53-binding sites. *Human Molecular Genetics*. 23: 2847-2857.
- Indran, I. R., Tufo, G., Pervaiz, S., Brenner, C. (2011). Recent advances in apoptosis, mitochondria and drug resistance in cancer cells. *Biochimica et Biophysica Acta*. 1807: 735 – 745.
- Iorfida, M., Bagnardi, V., Rotmensz, N., Munzone, E., Bonanni, B., Viale, G., Pruneri, G., Mazza, M., Cardillo, A., Veronesi, P., Luini, A., Galimberti, V., Goldhirsch, A., Colleoni, M. (2014). Outcome of male breast cancer: a matched single-institution series. *Clinical Breast Cancer*. 5: 371-377.
- Isono, T., Chano, T., Kitamura, A., Yuasa, T. (2014). Glucose deprivation induces G2/M transition-arrest and cell death in N-GlcNAc2-Modified protein-producing renal carcinoma cells. *PLoS ONE*, 9(5), e96168.
- Isono, T., Chano, T., Okabe, H., Suzaki, M. (2013). Study of global Transcriptional changes of N-GlcNAc2 proteins-producing T24 bladder carcinoma cells under glucose deprivation. *PLoS ONE*. 8(4), e60397.

- Jaramillo, S., Muriana, F. J. G., Guillen, R., Jimenez-Araujo, A., Rodriguez-Arcos, R., Lopez, S. (2016). Saponins from edible spears of wild asparagus inhibit AKT, p70S6K, and ERK signalling, and induce apoptosis through G0/G1 cell cycle arrest in human colon cancer HCT-116 cells. *Journal of Functional Foods*. 26: 1-10.
- Jemal, A., Bray, F., Melissa, M., Ferlay, J., Ward, E., Forman, D. (2011). Global Cancer Statistics. *American Cancer Society*. 61: 69-90.
- Jiang, B. P., Le, L., Xu, L. J., Xiao, P. G. (2014). Minocycline inhibits ICAD degradation and the NF- κ B activation induced by 6-OHDA in PC12 cells. *Brain Research*. 1586: 1-11.
- Johnson, I. S., Armstrong, J. G., Gorman, M. (1963). The *Vinca* alkaloids: A new class of oncolytic agents. *Cancer Research*. 23: 1390–1427.
- Jung, E. B., Lee, S. C. (2014). Baicalein attenuates proteasome inhibition-induced apoptosis by suppressing the activation of the mitochondrial pathway and the caspase-8- and Bid-dependent pathways. *European Journal of Pharmacology*. 730: 116-124.
- Kaler, P., Godasi, B. N., Augenlicht, L., Klampfer, L. (2009). The NF- κ B/AKT-dependent Induction of Wnt Signaling in Colon Cancer Cells by Macrophages and IL-1 β . *Cancer Microenvironment*. 2: 69–80.
- Kamdje, A. H. N., Etet, P, F. N., Vecchio, L., Muller, J. M., Krampera, M., Lukong, K. E. (2014). Signaling pathway in breast cancer: Therapeutic targeting the microenvironment. *Cellular Signaling*. 26: 2843 – 2856.
- Kandoth, C., McLellan, M. D, Vandin, F., Ye, K., Niu, B., Lu, C. (2013) . Mutational landscape and significance across 12 major cancer types. *Nature*. 17(502): 333 – 9.
- Karakosta, A., Golias, C. H., Charalabopoulos, A., Peschos, D., Batistatou, A., Charalabopoulos, K. (2005). Genetic models of human cancer as a multistep process. Paradigm models of colorectal cancer, breast cancer, and chronic myelogenous and acute lymphoblastic leukaemia. *Journal of Experimental & Clinical Cancer Research*. 24:505–514.
- Karnofsky, D. A., Abelson, W. H., Craver, L. F., and Burchenal, J. H. (1948). The use of nitrogen mustards in the palliative treatment of carcinoma. *Cancer*. 1: 634 – 656.
- Keng, S. L., Abdul Wahab, S. B., Chiu, L. B., Yusuf, A. (2015). Awareness of ovarian cancer risk factor among Women in Malaysia: a preliminary study. *Asian Pacific Journal of Cancer Prevention*. 16: 537 – 540.
- Khalil, I., Brewer, M. A., Neyarapally, T., Runowickz, C. D. (2010). The potential

of biologic network models in understanding the etiopathogenesis of ovarian cancer. *Gynecologic Oncology*. 116: 282 - 285.

- Khamis, Z. I., Sahab, Z. J., Sang, Q. X. (2012). Active roles of tumour stroma in breast cancer metastasis. *International Journal of Breast Cancer*. 574025.
- Kim, N. H., Park, H. J., Oh, M. K., Kim, L. S. (2013). Antiproliferative effect of gold (I) compound auranofin through inhibition of STAT3 and telomerase activity in MDA-MB 231 human breast cancer cells. *BMB Reports*. 46: 59-64.
- Kim, S. J., Miyoshi, Y., Taguchi, T., Tamaki, Y., Nakamura, H., Yodoi, J., Kato, K., Noguchi, S. (2005). High thioredoxin expression is associated with resistance to docetaxel in primary breast cancer. *Clinical Cancer Research*. 11: 845-8430.
- Kindelberger, D.W., Lee, Y., Miron, A., Hirsch, M.S., Feltmate, C., Medeiros, F., Callahan, M.J., Garner, E.O., Gordon, R.W., Birch, C., Berkowitz, R.S., Muto, M.G., Crum, C.P. (2007) Intraepithelial carcinoma of the fimbria and pelvic serous carcinoma: evidence for a causal relationship. *American Journal of Surgical Pathology*. 31: 161–169.
- Kohn, K. W., Spears, C. L., and Doty, P. (1966). Inter-strand crosslinking of DNA by nitrogen mustard. *Journal of Molecular Biology*. 19: 266 – 288.
- Kuhn, E., Kurman, R.J., Vang, R., Sehdev, A.S., Han, G., Soslow, R., Want, T.L., Shih, L.M. (2012). TP53 mutations in serous tubal intraepithelial carcinoma and concurrent pelvic high-grade serous carcinoma - evidence supporting the clonal relationship of the two lesions, *Journal of Pathology*. 226: 421–426.
- Lamb, M.P., Ablett, K., Spence, G., Landberg, A.H., Sims, R. B., Clarke, R. B. (2013). Wnt pathway activity in breast cancer sub-types and stem-like cells. *PLoS One*. 7: e67811.
- LeBlanc, A.C. (2003). Natural cellular inhibitors of caspases. *Prog. Neuro-Psychopharmacol. Biological Psychiatry*. 27: 215–229.
- Lee, K. I., Soo, C. C., Yang, C. Y., Hung, D. Z., Lin, C. T., Lu, T. H., Liu, S. H., and Huang, C. F. (2016). Etoposide induces pancreatic β -cells cytotoxicity via the JNK/ERK/GSK-3 signaling-mediated mitochondria-dependent apoptosis pathway. *Toxicology in Vitro*. 36: 142-152.
- Lei, S., Shen, F., Chen, J., Feng, J., Cai, W., Shen, L., Hu, Z., and Xu, B. (2016). MiR-639 promoted cell proliferation and cell cycle in human thyroid cancer by suppressing CDKN1A expression. *Biomedicine and Pharmacotherapy*. 84: 1834 – 1840.

- Lengyel, E. (2009). Ovarian cancer development and metastasis. *American Journal of Pathology*. 177: 1053-1064.
- Li, B., Gao, Y., Rankin, G. O., Rojanasakul, Y., Cutler, S. J., Tu, Y., & Chen, Y. C. (2015). Chaetoglobosin K induces apoptosis and G2 cell cycle arrest through p53-dependent pathway in cisplatin-resistant ovarian cancer cells. *Cancer Letters*, 356(2), 418–433.
- Li, F. (2005). Role of surviving and its splice variants in tumorigenesis. *British Journal of Cancer*. 92: 212-216.
- Li, W., Khor, T. O., Xu, C., Shen, G., Jeong, W. S., Yu, S., Kong, A. N. (2008). Activation of Nrf-2 antioxidant signaling attenuates NF- κ B inflammatory response and elicits apoptosis. *Biochemical Pharmacology*. 76: 1485 – 1489.
- Li, Y. (2013). Palmitate induces H9c2 cell apoptosis by increasing reactive oxygen species generation and activation of the ERK1/2 signaling pathway. *Molecular Medicine Reports*. 3: 855 – 861.
- Liang, L., Fan, Y., Cheng, J., Cheng, D., Zhao, Y., Cao, B., Ma, L., An, L., Jia, W., Su, X., Yang, J., Zhang, H. (2013). TAK1 ubiquitination regulates doxorubicin-induced NF- κ B activation. *Cellular Signaling*. 25: 247-254.
- Lichtenstein, P., Holm, N. V., Verkasalo, P. K., Iliadou, A., Kaprio, J., Koskenvuo, M., Pukkala, E., Skytthe, A., Hemminki, K. (2000) Environmental and heritable factors in the causation of cancer-analyses of cohorts of twins from Sweden, Denmark & Finland. *New England Journal of Medicine*. 343: 78 – 85.
- Lipton, A., Köstler, W. J., Leitzel, K., Ali, S. M., Sperinde, J., Weidler, J., Paquet, A., Sherwood, T., Huang, W., Bates, M. (2010) Quantitative HER2 protein levels predict outcome in fluorescence *in situ* hybridization-positive patients with metastatic breast cancer treated with trastuzumab. *Cancer*. 116:5168–5178.
- Lincoln, D. T., Ali Emadi, E. M., Tonissen, K.F., Clarke, F. M. (2003). The thioredoxin–thioredoxin reductase system: over-expression in human cancer. *Anticancer Research*. 23: 2425–2433.
- Liu, J., Chang, F., Li, F., Fu, H., Wang, J., Zhang, S., Yin, D. (2015). Palmitate promotes autophagy and apoptosis through ROS-dependent JNK and p38 MAPK. *Biochemical and Biophysical Research Communications*, 463(3), 262–267.
- Liu, J.F., Konstantinopoulos, P. A. (2017). Homologous Recombination and BRCA Genes in Ovarian Cancer: Clinical Perspective of Novel

Therapeutics. *Translational Advances in Gynaecologic cancers*. C6: 111-128.

- Liu, T., Xiang, B., Guo, D., Sun, F., Wei, R., Zhang, G., Ai, H., Tian, X., Zhu, Z., Zheng, W., Wang, Y., Wang, W. (2016). Morronside promotes angiogenesis and further improves microvascular circulation after focal cerebral ischemia/reperfusion. *Brain Research Bulletin*. 127: 111-118.
- Liu, Y., Liu, G., Mei, J., Wang, J. (2016). The preventive effects of hyperoside on lung cancer in vitro by inducing apoptosis and inhibiting proliferation through Caspase-3 and P53 signaling pathway. *Biomedicine & Pharmacotherapy*, 83, 381–391.
- Llor, X., Serfas, M. S., Bie, W., Vasioukhin, V., Polonskaia, M., Derry, J., Abbott, C. M., Tyner, A. L. (1999). BRK/Sik expression in the gastrointestinal tract and in colon tumours. *Clinical Cancer Research*. 5: 1767–1777.
- Machado-Silva, A., Perrier, S., Bourdon, J. C. (2010). P53 family members in cancer diagnosis and treatment. *Seminars in Cancer Biology*, 20(1), 57–62.
- Makar, A. P., Baekelandt, M., Trope, C. G., Kristensen, G. B. (1995). The prognostic significance of residual disease, FIGO substage, tumour histology, and grade in patients with FIGO stage III ovarian cancer. *Gynaecologic Oncology*. 56: 175-180.
- Mamede, A. C., Tavares, S. D., Abrantes, A. M., Trindade, J., Maia, J. M., Botelho, M.F. (2011). The role of vitamins in cancer: a review. *Nutrition and Cancer*. 63: 479–94.
- Mamenta, E. L., Poma, E. E., Kaufmann, W. K., Delmastro, D. A., Grady, H. L., Chaney, S. G. (1994). Enhanced replicative bypass of platinum-DNA adducts in cisplatin-resistant human ovarian carcinoma cells lines. *Cancer Research*. 54: 3500 – 2505.
- Marino, G., Kroemer, G. (2013). Mechanisms of apoptotic phosphatidylserine exposure. *Cell Research*. 23: 1247-1248.
- Martin, A. M., Weber, B.L. (2000). Genetic and hormonal risk factors in breast cancer. *Journal of the National Cancer Institute*. 14: 1126-1135.
- Marzano, C., Gandin, V., Folda, A., Scurari, G., Bindoli, A., Rigobello, M. P. (2007). Inhibition of thioredoxin reductase by auranofin induces apoptosis in cisplatin-resistant human ovarian cancer cells. *Free Radical Biology and Medicine*. 42: 872 – 881.
- Masur, K., Vetter, C., Hinz, A., Tomas, N., Henrich, H., Niggemann, B., Zänker, K. S. (2010). Diabetogenic glucose and insulin concentrations

modulate transcriptom and protein levels involved in tumour cell migration, adhesion and proliferation. *British Journal of Cancer*. 2: 345–352.

- Mattioli, I., Sebald, A., Bucher, C., Charles, R.P., Nakano, H., Doi, T., Kracht, M., Schmitz, M.L., (2004). Transient and selective NF-kappa B p65 serine 536 phosphorylation induced by T cell costimulation is mediated by I kappa B kinase beta and controls the kinetics of p65 nuclear import. *Journal of Immunology*. 172: 6336.
- Mauro, C, Leow, S. C., Anso, E., Rocha, S., Thotakura, A.K., Tornatore, L. (2011). NF-kB controls energy homeostasis and metabolic adaptation by upregulating mitochondrial respiration. *Nature Cell Biology*. 13: 1272–9.
- Meek, D. W. (1998). Multisite Phosphorylation and the integration of stress signals at p53. *Cellular Signalling*, 10(3): 159–166.
- Melisi, D., Xia, Q., Paradiso, G., Ling, J., Moccia, J., Carbone, C., Budilon, A., Abbruzzese, J. L., and Chiao, P. J. (2011). Modulation of pancreatic cancer chemoresistance by inhibition of TAK1. *Journal of National Cancer Institution*. 103: 1190-1204.
- Miao, Y., Yan, Q., Li, S., Li, B., Feng, Y. (2016). Neutrophil to lymphocyte ratio and platelet to lymphocyte ratio are predictive of chemotherapeutic response and prognosis in epithelial ovarian cancer patients treated with platinum-based chemotherapy. *Cancer Biomarkers*. 17: 33 -40.
- Mirabelli, K., Hill, D.T., Faucette, L. F., McCabe, F. L., Girard, G. R., Bryan, D. B., Sutton, B. M., Bartus, J. O., Crooke, S. T., Johnson, R. K. (1987). Antitumour activity of bis-(diphenylphosphino) alkanes, their gold(I) coordination complexes, and related compounds, *Journal of Medicinal Chemistry*. 30: 2181.
- Mitchell, P. J., Barker, K. T., Martindale, J. E., Kamalati, T., P.N. Lowe, P. N., Page, M. J., Gusterson, B. A., Crompton, M. R. (1994). Cloning and characterisation of cDNAs encoding a novel non-receptor tyrosine kinase, brk, expressed in human breast tumours. *Oncogene*. 9: 2383–2390.
- Molhotra, V., Perry, M. C. (2003). Classical chemotherapy: mechanisms, toxicities and the therapeutic window. *Cancer Biology and Therapy*. 2: 2 – 4.
- Moll, U. M., S. Wolff, S., Speidel, D., Deppert, W. (2005). Transcription-independent pro apoptotic functions of p53, *Current Opinion of Cell Biology*. 17: 631–636.

- Muller, P. A., Vousden, K.H. (2014). Mutant p53 in cancer: new functions and therapeutic opportunities. *Cancer Cell*. 25:304 – 17.
- Mundt, H. M., Stremmel, W., Melino, G., Krammer, P. H., Schilling, T., Muller, M. (2010). Dominant negative (DeltaN) p63alpha induces drug resistance in hepatocellular carcinoma by interference with apoptosis signaling pathways. *Biochemical and Biophysical Research Communications*. 396: 335-341.
- Nardon, C., Schmitt, S. M., Yang, H., Zuo, J., Fregona, D., Dou, Q. P. (2014). Gold(III)-Dithiocarbamate Peptidomimetics in the Forefront of the Targeted Anticancer Therapy: Preclinical Studies against Human Breast Neoplasia. *PLoS ONE* 9(1): e84248.
- Nardon, C., Boscutti, G., Fregona, D. (2014). Beyond platinum: Gold complexes as anticancer agents. *Anticancer Research*. 34: 487-492.
- Nishinaka, Y., Nakamura, H., Masutani, H., Yodoi, J. (2001). Redox control of cellular function by thioredoxin: a new therapeutic direction in host defense. *Archivum Immunologiae et Therapia Experimentalis*. 49: 285-292.
- Nordberg, J., Arner, E. S. J. (2001). Reactive oxygen species, antioxidants and the mammalian thioredoxin system. *Free Radical Biology and Medicine*. 31: 1287-1312.
- Oberst, A., Malatesta, M., Aqeilan, R. I., Rossi, M., Salomoni, P., Murillas, R., Melino, G. (2007). The Nedd4-binding partner 1 (N4BP1) protein is an inhibitor of the E3 ligase itch. *Proceedings of the National Academy of Sciences*, 104(27), 11280–11285.
- Ooi, K. K., Yeo, C. I., Ang, K. P., Abdah, M. A., Cheah, Y. K., Seng, H. L., Tiekink, E. R. T. (2017). G2/M cell cycle arrest on HT 29 cancer cells and toxicity assessment of triphenylphosphane-gold(I) carbonimidodithioates Ph₃PAu[SC(OR)=NPh] R=Me Et and iPr during zebrafish development. *Journal of Inorganic Biochemistry*. 166: 173-181.
- Oommen, D., Dodd, N. J. F., Yiannakis, D., Moyeed, R., Jha, AN. (2016). Linking genotoxicity and cytotoxicity with membrane fluidity: A comparative study in ovarian cancer cell lines following exposure to auranofin. *Mutation Research*. 809: 43-49.
- Oommen, D., Yiannakis, D., and Jha, A. N. (2016). BRCA1 deficiency increases the sensitivity of ovarian cancer cells to auranofin. *Mutation Research*. 7784: 8-15
- Ortego, L., Cardoso, F., Martins, S., Fillat, M. F., Laguna, A., Meireles, M., Villacampa, M. D., and Gimeno, M. C. (2014). Strong inhibition of

- thioredoxin reductase by highly cytotoxic gold(I) complex. DNA binding studies. *Journal of Inorganic Biochemistry*. 130: 32-37.
- Ott, I. (2009). On the medicinal chemistry of gold complexes as anticancer drugs. *Coordination Chemistry Reviews*. 253: 1670 – 1681.
- Pang, Y., Qin, G., Wu, L., Wang, X., Chen, T. (2016). Artesunate induces ROS-dependent apoptosis via a Bax-mediated intrinsic pathway in Huh-7 and Hep3B cells. *Experimental Cell Research*. 347: 251-260.
- Parajuli, B., Lee, H. G., Kwon, S. H., Cha, S. D., Shin, S. J., Lee, G. H., Bae, I., Cho, C. H. (2013). Salinomycin inhibits Akt/NF- κ B and induces apoptosis in cisplatin resistant ovarian cancer cells. *Cancer Epidemiology*. 37: 512 – 517.
- Park, E. J., Park, S. W., Kim, H. J., Kwak, J.-H., Lee, D.-U., & Chang, K. C. (2014). Dehydrocostuslactone inhibits LPS-induced inflammation by p38MAPK-dependent induction of hemeoxygenase-1 *in vitro* and improves survival of mice in CLP-induced sepsis *in vivo*. *International Immunopharmacology*, 22(2), 332–340.
- Park, Y. J., Choi, C. I., Chung, K. H., Kim, K. H. (2016). Phorbilignan C induces apoptosis through a mitochondria-mediated intrinsic pathway in human breast cancer cells. *Bioorganic & Medicinal Chemistry Letters*. 26: 4645-4649.
- Parr, C., Watkins, G., Jiang, W. G. (2004). The possible correlation of Notch-1 and Notch-2 with clinical outcome and tumour clinicopathological parameters in human breast cancer. *International Journal of Molecular Medicine*. 14:779–86.
- Petro, B. J., Tan, R. C., Tyner, A. L., Lingen, M. W., Watanabe, K. (2004). Differential expression of the non-receptor tyrosine kinase BRK in oral squamous cell carcinoma and normal oral epithelium. *Oral Oncology*. 40: 1040–1047.
- Petrucci, N., Daly, M. B., Feldman, G. L. (2010). Hereditary breast and ovarian cancer due to mutations in BRCA1 and BRCA2. *Genetics in Medicine*. 12: 245 – 259.
- Piątkiewicz, P., Czech, A. (2011). Glucose metabolism disorders and the risk of cancer. *Archivum Immunologiae et Therapiae Experimentalis*, 3: 215–230.
- Piek, J.M., van Diest, P.J., Zweemer, R.P., J.W., Poort-Keesom, R.J., Menko, F.H., Gille, J.J., Jongsma, A.P., Pals, G., Kenemans, P., Verheijen, R.H. (2001). Dysplastic changes in prophylactically removed fallopian tubes of women predisposed to developing ovarian cancer, *Journal of*

Pathology. 195: 451–456.

- Portt, L., Norman, G., Clapp, C., Greenwood, M., Greenwood, M.T. (2011). Anti-apoptosis and cell survival: A review. *Biochimica et Biophysica Acta*. 1813: 238 – 259.
- Powis, G., Montfort, W. R. (2001). Properties and biological activities of Thioredoxins. *Annual Reviews in Pharmacology and Toxicology*. 46: 261-295.
- Rackham, O., Nichols, S. J., Leedman, P. J., Berners-Price, S. J., Filipovska, A. (2007). A gold(I) phosphine complex selectively induces apoptosis in breast cancer cells: implications for anticancer therapeutics targeted to mitochondria. *Biochemical Pharmacology*. 74: 992 – 1002.
- Rahman, N., Stratton M. R. (1998). The genetics of breast cancer susceptibility. *Annual Review in Genetics*. 32: 95-121.
- Redza-Dutordoir, M., Averill-Bates, D. A. (2016). Activation of apoptosis signaling pathways by reactive oxygen species. *Biochimica et Biophysica Acta*. 1863: 2977-2992.
- Reed, J. C. (1998). Bcl-2 family proteins. *Oncogene*. 17: 3225 – 3236.
- Reid, J. F., Gariboldi, M., Sokolava, V., Capabianco, P., Lampis, A., Perrone, F., Signoroni, S., Costa, A., Leo, E., Pilotti, S., Pierotti, M. A. (2009). Integrative approach for prioritizing cancer genes in sporadic colon cancer. *Genes Chromosomes Cancer*. 48: 953-962.
- Ribeiro-Silva, A., de Moura, H. B., do Vale, F. R., Zucoloto, S. (2005). The differential regulation of human telomerase reverse transcriptase and vascular endothelial growth factor may contribute to the clinically more aggressive behavior of p63-positive breast carcinomas. *International Journal of Biological Markers*. 20: 227-234.
- Ribeiro-Silva, A., Ramalho, L. N. Z., Garcia, S. B., Zucoloto, S. (2003). The relationship between p63 and p53 expression in normal and neoplastic breast tissue. *Archives of Pathology and Laboratory Medicine*. 127: 336-340.
- Rivlin, N., Koifman, G., Rotter, V. (2015). p53 orchestrates between normal differentiation and cancer. *Seminars of Cancer Biology*. 32: 10-17.
- Robaina, M. C. S., Faccion, R. S., Arruda, V. O., de Rezende, L. M. M., Vasconcelos, G. M., Apa, A. G., Bacchi, C. E., Klumb, C. E. (2015). Quantitative analysis of CDKN2A methylation, mRNA, and p16INK4a protein expression in children and adolescents with Burkitt lymphoma: Biological and clinical implications. *Leukemia Research*. 39: 248-256.

- Rohrs, S., Kutzner, N., Vlad, A., Grunwald, T., Ziegler, S., Muller, O. (2009). Chronological expression of Wnt target genes Ccnd1, Myc, Cdkn1a, Tfr3, Plf1 and Ramp3. *Cell Biology International*. 33: 501-508.
- Ronconi, L., Aldinucci, D., Dou, P., Fregona, D. (2010). Latest Insights into the Anticancer Activity of Gold(III)-Dithiocarbamate Complexes. *Anti-cancer Agents in Medicinal Chemistry*. 10: 283-292.
- Rosenberg, B., VanCamp, L., Trosko, J. E., Mansour, V. H. (1969). Platinum compounds: a new class of potent anti-tumour agents. *Nature*. 222: 385 – 386.
- Rowinsky, E. K., and Donehower, R. C. (1991). The clinical pharmacology and use of antimicrotubule agents in cancer chemotherapeutics. *Pharmacology and Therapeutics*. 52: 35–84.
- Rush, G. F., Smith, P. F., Hoke, G. D., Alberts, D. W., Snyder, R. M., Mirabelli, C. K. (1987). The mechanism of acute toxicity of triethylphosphine gold (I) complexes: II. Triethylphosphine gold chloride induced alterations in mitochondrial function. *Toxicology and Applied Pharmacology*. 90: 391 – 400.
- Ryan, B. M., O' Donovan, N., Duffy, M. J. (2009). Survivin: A new target for anti-cancer therapy. *Cancer Treatment Reviews*. 35: 553-562.
- Sabol, S. L., Li, R., Lee, T. Y., Abdul-Khalek, R. (1998). Inhibition of Apoptosis-Associated DNA Fragmentation Activity in Nonapoptotic Cells: The Role of DNA Fragmentation Factor-45 (DFF45/ICAD). *Biochemical and Biophysical Research Communications*. 253: 151-158.
- Sakurai, H. (2012). Targeting of TAK1 in inflammatory disorders and cancer. *Trends in Pharmacological Sciences*. 33: 522-530.
- Santini, S., Di Agostino, S., Coppari, E., Bizzarri, A. R., Blandino, G., Cannistraro, S. (2014). Interaction of mutant p53 with p73: A surface Plasmon resonance and atomic force spectroscopy study. *Biochimica et Biophysica Acta (BBA) - General Subjects*, 1840(6), 1958–1964.
- Santos, S., Silva, A. M., Matos, M., Monteiro, S. M., Alvaro, A. R. (2016). Copper induced apoptosis in Caco-2 and Hep-G2 cells: Expression of caspase 3, 8, and 9, AIF and p53. *Comparative Biochemistry and Physiology. Part C: Toxicology & Pharmacology*. 185-186: 138-146.
- Santos-Silva, M. C., Freitas, M. S., Assreuy, J. (2006). Involvement of NF-kappaB and glutathione in cytotoxic effects of nitric oxide and taxol on human leukaemia cells. *Leukaemia Research*. 30:145–52

- Sasada, T., Sono, H., Yodoi, J. (1996). Thioredoxin/adult t-cell leukemia-derived factor (ADF) and Redox regulation. *Journal of Toxicological Sciences*. 5: 285–287.
- Scaffidi, C., Fulda, S., Srinivasan, A., Friesen, C., Li, F., Tomaselli, K.J., Debatin, K.M., Krammer, P.H., Peter, M.E. (1998). Two CD95 (APO-1/Fas) signaling pathways. *EMBO Journal*. 17: 1675–1687.
- Schmandt, R. E., Bennett, M., Clifford, S., Thornton, A., Jiang, F., Broaduss, R. R., Sun, C. C., Lu, K. H., Sood, A. K., Gershenson, D. M. (2006). The BRK tyrosine kinase is expressed in high-grade serous carcinoma of the ovary *Cancer Biology and Therapy*. 5: 1136–1141.
- Sehouli, J., Stengel, D., Oskay-Oezcelik, G. (2008). Nonplatinum topotecan combinations versus topotecan alone for recurrent ovarian cancer: results of a phase III study of the North-Eastern German Society of Gynecological Oncology Ovarian Cancer Study Group. *Journal of Clinical Oncology*. 26: 3176–82.
- Seol, J. Y., Mihich, E. Berleth, E. S. (2015). TNF Apoptosis Protection Fraction (TAPF) prevents apoptosis induced by TNF, but not by Fas or TRAIL, via NF- κ B-induced increase in cFLIP. *Cytokine*. 75: 321-329.
- Sharma, H. K., Chhangte, L., Dolui, A. K. (2001). Traditional medicinal plants in Mizoram, India. *Fitoterapia*. 72: 146-161.
- Shaw, C. F. (1999). Gold-based therapeutic agents. *Chemical Reviews*. 99: 2589 – 2600.
- Shelar, S. B., Kaminska, K. K., Reddy, S. A., Kumar, D., Tan, C. T., Yu, V. C. (2015). Thioredoxin-dependent regulation of AIF-mediated DNA damage. *Free Radical Biology and Medicine*. 87: 125-136.
- Shi, J., Shen, H. M. (2008). Critical role of Bid and Bax in indirubin-3'-monoxime-induced apoptosis in human cancer cells. *Biochemical Pharmacology*. 75: 1729-1742.
- Shiloh, Y. (2003). ATM and related protein kinases: safeguarding genome integrity. *Nature Reviews Cancer*. 3:155 – 168.
- Si, L., Zheng, L., Xu, L., Yin, L., Han, X., Qi, H., Xu, Y., Wang, C., Peng, J. (2016). Dioscin suppresses human laryngeal cancer cells growth via induction of cell-cycle arrest and MAPK-mediated mitochondrial-derived apoptosis and inhibition of tumour invasion. *European Journal of Pharmacology*. 774: 105-117.
- Siegel, R., Ma, J. Zou, Z., Jemal, A. (2014) A cancer statistics. *CA: A Cancer Journal for Clinicians*. 64: 9-29.

- Simon, T. M., Kunishima, D. H., Vilbert, G. J., Lorber, A. (1981). Screening trail with the coordinated gold compound auronafin using mouse lymphocytic leukemia P388. *Cancer Research*. 41: 94 – 97.
- Singh, A., Sweeney, M. F., Yu, M., Burger, A., Greninger, P., Benes, C., Haber, D. A., Settleman, J. (2012). TAK1 inhibition promotes apoptosis in KRAS-dependent colon cancers. *Cell*. 148: 639-650.
- Slamon, D.J., Leyland-Jones, B., Shak, S. (2001). Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *New England Journal of Medicine*. 344: 783–792.
- Slee, E.A., Harte, M.T., Kluck, R.M., Wolf, B.B., Casiano, C.A., Newmeyer, D.D., Wang, H.G., Reed, J.C., Nicholson, D.W., Alnemri, E.S., Green, D.R., Martin, S.J. (1999). Ordering the cytochrome c-initiated caspase cascade: hierarchical activation of caspases-2, -3, -6, -7, -8, and -10 in a caspase-9-dependent manner. *Journal of Cell Biology*. 144: 281–292.
- Sohda, M., Misumi, Y., Oda, K. (2015). TNF α triggers release of extracellular vesicles containing TNFR1 and TRADD, which can modulate TNF α responses of the parental cells. *Archives of Biochemistry and Biophysics*. 587: 31-37.
- Sotillo, O., Renner, P., Dubus, J., Ruiz-Cabello, J., Martín-Caballero, M, Barbacid, A., Carnero, M., Malumbres, M. (2005). Cooperation between Cdk4 and p27kip1 in tumour development: a preclinical model to evaluate cell cycle inhibitors with therapeutic activity. *Cancer Research*. 65: 3846–3852.
- Stevens, C. M., Mylorie, A., Auerbach, C. (1950). Biological actions of mustard gas compound. *Nature*. 166: 1019 – 1021.
- Su, H. Y., Lai, H. C., Lin, Y. W., Liu, C. Y., Chen, C. K.,(2010). Epigenetic silencing of SFRP5 is related to malignant phenotype and chemoresistance of ovarian cancer through Wnt signaling pathway. *International Journal of Cancer*. 127: 555–56.
- Su, Y., Zhang, X., Sinko, P. J. (2007). Exploitation of drug-induced Bcl-2 overexpression for restoring normal apoptosis function: A promising new approach to the treatment of multidrug resistant cancer. *Cancer Letters*. 253: 115 – 123.
- Sullivan, K. D., Gallant-Behm, C. L., Henry, R. E., Fraikin, J. L., Espinosa, J. M. (2012). The p53 circuit board. *Biochimica et Biophysica Acta*. 1825: 229-244.

- Taguchi, T., Nazneen, A., Abid, M.R., Razzaque, M.S. (2005). Cisplatin associated nephrotoxicity and pathological events. *Contributions Nephrology*. 148: 107-121.
- Tannapfel, A., Engeland, K., Weinans, L., Katalinic, A., Hauss, J., Mössner, J., Wittekind, C. (1999). Expression of p73, a novel protein related to the p53 tumour suppressor p53, and apoptosis in cholangiocellular carcinoma of the liver. *British Journal of Cancer*. 7: 1069–1074.
- Thompson, D., Easton, D. (2004). The genetic epidemiology of breast cancer genes. *Journal of Mammary Gland Biology and Neoplasia*. 9:221–36.
- Tozawa, K., Kawai, N., Hayashi, Y., Sasaki, S., Kohri, K., Okamoto, T. (2003). Gold compounds inhibits adhesion of human cancer cells to vascular endothelial cells. *Cancer Letters*. 196: 93 – 100.
- Todd, A., Anderson, R. J., Pickles, G., Groundwater, P. W. (2010). Targeting the thioredoxin system in the treatment of certain cancers. *The Pharmaceutical Journal*. 284: 243-244.
- Tonissen, K.F., Di Trapani, G. (2009). Thioredoxin system inhibitors as mediators of apoptosis for cancer therapy. *Molecular Nutrition and Food Research*. 53: 87–103
- Trachootham, D., Alexandre, J., Huang, P. (2009). Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach? *Nature Reviews Drug Discovery*. 8: 579–591.
- Trembaly, C., Saw, J., Curtis, D. (2014). Cdkn1A (p21) is required for quiescence, therapeutic resistance and clonal evolution of pre-leukemic stem cells. *Experimental Hematology*. 42: S17.
- Tsujimoto, Y., Shimizu, S. (2006). Role of the mitochondrial membrane permeability transition in cell death. *Apoptosis*, 12(5), 835–840.
- Tsujimoto, Y., Shimizu, S. (2001). Bcl-2 family: life or death switch. *FEBS Letters*. 466: 6 – 10.
- Tu, D., Zhu, Z., Zhou, A. Y., Yun, C., Lee, K. Y., Yoms, A. V., Li, Y., Dunn, G. P., Chan, E., Thai, T., Yang, S., Ficarro, S. B., Marto, J. A., Jeon, H., Hahn, W. C., Barbic, D. A., Eck, M. J. (2013). Structure and ubiquitination-dependent activation of TANK-binding kinase 1. *Cell Reports*. 3: 747-758.
- Tutt, A., Ashworth, A. (2002). The relationship between the roles of BRCA genes in DNA repair and cancer predisposition. *Trends in Molecular Medicine*. 8: 571 – 576.

- Ukaji, T., Umezawa, K. (2014). Novel approaches to target NF- κ B and other
- Vicencio, J.M., Galluzzi, L., Tajeddine, N., Ortiz, C., Criollo, A., Tasmemir, E., Morselli, E., Ben Younes, A., Maiuri, M.C., Lavandro, S., Kroemer, G. (2008) Senescence, apoptosis or autophagy? When a damaged cell must decide its path—a mini-review. *Gerontology*. 54: 92 – 99.
- Ukaji, T., Lin, Y., Okada, S., Umezawa, K. (2017). Inhibition of MMP-2-mediated cellular invasion by NF- κ B inhibitor DHMEQ in 3D culture of breast carcinoma MDA-MB-231 cells: A model for early phase of metastasis. *Biochemical and Biophysical Research Communications*. 485: 76-81.
- Van Zijin, F., Krupitza, G., Mikulits, W. (2011). Initial steps of metastasis: Cell invasion and endothelial transmigration. *Mutation Research/Review in Mutation Research*. 728: 23-34.
- Veeck, J., C. Geisler, C., E. Noetzel, E., S. Alkaya, S., A. Hartmann, A., Knuchel, R., Dahl, E. (2008). Epigenetic inactivation of the secreted frizzled-related protein-5 (SFRP5) gene in human breast cancer is associated with unfavorable prognosis. *Carcinogenesis*. 29: 991–998.
- Videira, M., Reis, R. L., Brito, M. A. (2014). Deconstructing breast cancer cell biology and the mechanisms of multidrug resistance. *Biochimica et Biophysica Acta*. 1846: 312-325.
- Volgerstein B., Lane D., Levine, A. J. (2000). Surfing the p53 network. *Nature*. 408: 307-310.
- Vousden, K. H., Lu, X. (2002). Live or let die: the cell's response to p53. *Nature Reviews Cancer*. 2: 594 – 604.
- Wada, T., and Penninger, J. M. (2004). Mitogen-activated protein kinases in apoptosis regulation. *Oncogene*. 23: 2838-2849.
- Wang, F., Wang, L., Zhao, Y., Li, Y., Ping, G., Xiao, S., Wu, C. (2014). A novel small-molecule activator of procaspase-3 induces apoptosis in cancer cells and reduces tumour growth in human breast, liver and gallbladder cancer xenografts. *Molecular Oncology*. 8: 1640–1652.
- Wang, K., Ye, Y., Xu, Z., Zhang, X., Hou, Z., Cui, Y., Song, Y. (2010). Interaction between BRCA1/BRCA2 and ATM/ATR associate with breast cancer susceptibility in a Chinese Han population. *Cancer Genetics and Cytogenetics*. 200: 40 – 46.
- Wani, M., Taylor, H. L., Wall, M. E., Coggan, P., McPhail, A. T. (1971). Plant antitumour agents. VI. The isolation and structure of taxol a novel antileukemic and antitumour agent from *Taxus brevifolia*. *Journal of American Chemical Society*. 93: 2325–2327.

- Weber, M. L. (2013). Targeting apoptosis pathways in cancer by Chinese medicine. *Cancer Letters*. 332: 304 – 312.
- Weijzen, S., Rizzo, P., Braid, M., Vaishnav, R., Jonkheer, S. M., Zlobin, A. (2002). Activation of Notch-1 signaling maintains the neoplastic phenotype in human Ras-transformed cells. *Nature Medicine*. 8:979–86.
- Wei, L., Jin, X., Cao, Z., Li, W. (2016). Evodiamine induces extrinsic and intrinsic apoptosis of ovarian cancer cells via the mitogen-activated protein kinase/phosphatidylinositol-3-kinase/protein kinase B signaling pathways. *Journal of Traditional Chinese Medicine*. 36: 353-359.
- Welsh, S. J., Bellamy, W. T., Briehl, M. M., Powis, G. (2002). The redox protein thioredoxin 1 (Trx-1) increases hypoxia-inducible factor 1alpha protein expression: Trx-1 overexpression results in increased vascular endothelial growth factor production and enhanced tumour angiogenesis. *Cancer Research*. 62: 5089-5095.
- Whittington, R. M., Close, H. P. (1970). Clinical experience with mitomycin C (NSC-26980). *Cancer Chemotherapy Reports*. 54: 195–198.
- Wiese, C., Rudolph, J. H., Jakob, B., Fink, D., Tobias, F., Blantner, C., Taucher-Scholz, G. (2012). PCNA-dependent accumulation of CDKN1A into nuclear foci after ionizing irradiation. *DNA Repair*. 11: 511-521.
- Wu, G. S. (2004). The functional interactions between the p53 and MAPK signaling pathway. *Cancer Biology and Therapy*. 3: 151-161.
- Wu, Y., Zhou B.P., (2010). TNF-alpha/NF-kappaB/Snail pathway in cancer cell migration and invasion. *British Journal of Cancer*. 102: 639.
- Wu, Z., Wu, Y., Qin, Y. Li, X. (2014). Influences of sorting and cryopreservation on the mitochondrial membrane potential (MMP) and phosphatidylserine (PS) externalization in bovine sperm. *Livestock Science*. 168: 177-182.
- Yang, P.Y., Hu, D.N., Kao, Y.H., Lin, I.C., Chou, C.Y., Wu, Y.C. (2016). Norcantharidin induces apoptosis in human prostate cancer cells through both intrinsic and extrinsic pathways. *Pharmacological Reports*, 68(5), 874–880.
- Yang, Y., Yu, Y., Wang, J., Li, Y., Li, Y., Wei, J., Zheng, T., Jin, M., Sun, Z. (2017). Silica nanoparticles induced intrinsic apoptosis in neuroblastoma SH-SY5Y cells via CytC/Apaf-1 pathway. *Environmental Toxicology and Pharmacology*. 52: 161 – 169.

- Yasuhara, R., Irie, T., Suzuki, K., Sawada, T., Miwa, N., Sasaki, A., Tsunoda, Y., Nakamura, S., Mishima, K. (2015). The β -catenin signaling pathway induces aggressive potential in breast cancer by up-regulating the chemokine CCL5. *Experimental Cell Research*. 338: 22-31.
- Yip, C. H., Pathy, N. B., Teo, S. H. (2014). A review of breast cancer research in Malaysia. *The Medical Journal of Malaysia*. 69: 8-22.
- Yokomizo, A., Ono, M., Nanri, H., Makino, Y., Ohga, T. Wada, M. (1995). Cellular levels of thioredoxin associated with drug sensitivity to cisplatin, mitomycin C, doxorubicin and etoposide. *Cancer Research*. 55: 4293-4296.
- Yoo, M. H., Xu, X. M., Carlson, B. A., Gladyshev, V. N., Hatfield, D. L. (2006). Thioredoxin reductase 1 deficiency reverses tumour phenotype and tumourigenicity of lung carcinoma cells. *Journal of Biological Chemistry*. 281: 13005-13008.
- Yoo, M. H., Xu, X. M., Carlson, B. A., Patterson, A. D., Gladyshev, V. N., Hatfield, D.L. (2007). Targeting thioredoxin reductase 1 reduction in cancer cells inhibits self-sufficient growth and DNA replication. *PLoS One*, 2:e1112.
- Yoshikawa, Y., Murayama, A., Adachi, Y., Sakurai, H., Yasui, H. (2011). Challenge of studies on the development of new Zn complexes ($Zn(opt)_2$) to treat diabetes mellitus. *Metallomics*. 3: 686 – 692.
- Zaika, A. (2000). Oncogenes induce and activate endogenous p73 protein. *Journal of Biological Chemistry*. 14: 11310–11316.
- Zeren, T., Inan, S., Vatanserver, H. S., Sayhan. S. (2014). Significance of apoptosis related proteins on malignant transformation of ovarian tumours: A comparison between Bcl-2/Bax ratio and p53 immunoreactivity. *Acta Histochemica*. 116: 1251-1258.
- Zhang, H., Bai, M., Deng, T., Liu, R., Wang, X., Qu, Y., Duan, J., Zhang, L., Ning, T., Ge, S., Li, H., Zhou, L., Liu, Y., Huang, D., Ying, G., Ba. Y. (2016). Cell-derived microvesicles mediate the delivery of miR-29a/c to suppress angiogenesis in gastric carcinoma. *Cancer Letters*. 375: 331-339.
- Zhang, J., Li, Y., Duan, D., Yao, J., Gao, K., Fang, J. (2016). Inhibition of thioredoxin reductase by alantolactone prompts oxidative stress-mediated apoptosis of HeLa cells. *Biochemical Pharmacology*. 102: 34-44.
- Zhao, R., Choi, B. Y., Lee, M., Bode, A. M., Dong, Z. (2016). Implications of Genetic and Epigenetic Alterations of CDKN2A (p16INK4a) in Cancer. *EBioMedicine*. 8: 30-39.

Zhou, A. Y., Shen, R. R., Kim, E., Lock, Y. J., Xu, M., Chen, Z. J., Hahn, W. C. (2013). IKK ϵ -mediated tumorigenesis required k63-linked polyubiquitination by a cIAP1/cIAP2/TRAF2 E3 ubiquitin ligase complex. *Cell Reports*. 3: 724-733.

Zhou, Y.-Y., Li, Y., Jiang, W.-Q., Zhou, L.-F. (2015). MAPK/JNK signaling: A potential autophagy regulation pathway. *Bioscience Reports*. 35: e00199

Zorn, K.K., Bonome, T., Gangi, L., Chandramouli, G.V., Awtrey, C.S., Gardner, G.J., Barrett, J.C., Boyd, J., Birrer, M.J. (2005). Gene expression profiles of serous, endometrioid, and clear cell subtypes of ovarian and endometrial cancer. *Clinical Cancer Research*. 11: 6422–6430.

Zucchi, I., Astigiano, S., Bertalot, G., Sanzone, S., Cocola, C., and Pelucchi, P. (2008). Distinct populations of tumour-initiating cells derived from a tumour generated by rat mammary cancer stem cells. *Proceedings of the National Academy of Sciences*. 105: 16940-16945.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT :

MOLECULAR MECHANISMS UNDERLYING ANTIPROLIFERATIVE EFFECT OF TRICYCLOHEXYLPHOSPHINE GOLD (I) MERCAPTOBENZOATE DERIVATIVES ON HUMAN BREAST AND OVARIAN CARCINOMA CELL LINES

NAME OF STUDENT: ANG KOK PIAN

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]