

UNIVERSITI PUTRA MALAYSIA

INVOLVEMENT OF TLR2, MAPKinase and NFκβ PATHWAYS IN REGULATION OF HUMAN BETA DEFENSIN 9 IN HUMAN CORNEAL EPITHELIAL CELLS STIMULATED WITH Pam3CSK4

NURUL HANA ZAINAL BAHARIN

FPSK(m) 2018 4



INVOLVEMENT OF *TLR2*, *MAPKinase* AND *NFκβ* PATHWAYS IN REGULATION OF HUMAN BETA DEFENSIN 9 IN HUMAN CORNEAL EPITHELIAL CELLS STIMULATED WITH *Pam*3CSK4



NURUL HANA BINTI HAJI ZAINAL BAHARIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

October 2017

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

INVOLVEMENT OF TLR2, MAPKinase AND NFκβ PATHWAYS IN REGULATION OF HUMAN BETA DEFENSIN 9 IN HUMAN CORNEAL EPITHELIAL CELLS STIMULATED WITH Pam3CSK4

By

NURUL HANA BINTI HAJI ZAINAL BAHARIN

October 2017

Chairman: Associate Professor Nazri Omar, PhDFaculty: Medicine and Health Sciences

Corneal epithelium was shown to provide immune surveillance against invading pathogens through a variety of pathogen recognition receptors (PRR) such as toll like receptors (TLRs) and nucleotide oligomerisation receptors (NLRs) which are capable of recognizing pathogen associated molecular patterns (PAMPs) such as lipopolysaccharide (LPS) and synthetic triacylated lipoprotein (Pam3CSK4), derived from various infection-causing microbes. The elimination of pathogenic microorganisms, including Gram-positive and negative bacteria, fungi and viruses, involves antimicrobial peptides (AMPs). A number of AMPs such as defensins have multiple functions in host defence. Defensins, produced by cells in the course of innate host defence, serve as signals which initiate, mobilise, and amplify adaptive host immune defences. Defensins use multiple cellular receptors, which endow them with the capacity to marshall adaptive host defences against microbial invaders. Human βdefensins (HBDs), one type of defensins family, are an important part of the innate host immune defense at the ocular surface. Unlike other defensins, expression of HBD9, also known as DEFB109, at the ocular surface is reduced during microbial infection, but the stimulation of HCECs by Pam3CSK4 was shown to upregulate HBD9 at the initial responds followed by a significant downregulation. The mechanism of infection or inflammation has been linked with alterations in several important cellular signaling pathways such as *TLR2*, *MAPKinase* and *NF\kappa\beta* pathways. These pathways are of interest from a therapeutic perspective, because targeting them may help to reverse, delay, or prevent inflammation. The main objectives in this study was to determine the involvement of *TLR2*, *MAPKinase* and *NF\kappa\beta* signaling pathways in the expression of DEFB109 in Pam3CSK4-stimulated HCECs. The techniques included cell culture stimulated with Pam3CSK4, the exposure of the cells to specific transcription factor inhibitors, qPCR and dot blot analysis. In this study, the evidence to indicate that TLR2 induces HBD9 mRNA and protein expression in a time- and



dose-dependent manner was presented. *TLR2* siRNA was observed interfering TLR2mediated induction of *DEFB109* expression. The involvement of *MAPKinase* pathways and *NF\kappa\beta* pathways in the expression of *DEFB109* gene also were observed. These pathway-specific molecules can be exploited to modulate the response of HBD9 during microbial infection. The variable expression of different AMPs to specific pathogens would suggest similar but subtly different pathways invoked by the pathogens, probably related to their PAMPs and different TLRs and other receptors they bind to.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENGLIBATAN LALUAN ISYARAT TLR2, MAPKinase DAN NFκβ KE ATAS EKSPRESI DEFB109 DI DALAM SEL EPITELIUM KORNEA TERANSANG OLEH Pam3CSK4

Oleh

NURUL HANA BINTI HAJI ZAINAL BAHARIN

Oktober 2017

Pengerusi : Profesor Madya Nazri Omar, PhD Fakulti : Perubatan dan Sains Kesihatan

Epitelium kornea didapati memberikan pengawasan imun terhadap serangan patogen melalui pelbagai reseptor pengeceman patogen (PRR) seperti reseptor tol (TLR) dan reseptor oligomerisasi nukleotida (NLRs) yang mampu mengenal pasti pola molekul berkaitan patogen (PAMP) seperti lipopolisakarida (LPS) dan lipoprotein triasilat sintetik (*Pam3CSK4*), yang diperoleh daripada pelbagai mikrob penyebab jangkitan. Pemusnaha mikroorganisma patogen termasuk bakteria Gram-positif dan negatif, kulat dan virus, melibatkan peptida antimikrob (AMP). Sejumlah AMP, seperti defensin mempunyai pelbagai fungsi dalam sistem pertahanan. Defensin yang dihasilkan oleh sel berfungsi sebagai isyarat yang mencetuskan, meningkatkan, dan mengukuhkan sistem pertahanan imun perumah. Defensin menggunakan pelbagai reseptor sel, yang membantu dalam sistem pertahanan perumah terhadap jangkitan mikrob. β-defensin manusia (HBDs) adalah salah satu ahli kumpulan defensin yang merupakan komponen penting dalam pertahanan inat di permukaan okular. Tidak seperti defensin lain, ekspresi HBD9 yang juga dekenali sebagai DEFB109 di permukaan okular berkurang semasa jangkitan mikrob, tetapi rangsangan Pam3CSK4 terhadap HCEC menunjukkan peningkatan ekspresi *DEFB109* pada gerak balas awal diikuti dengan penurunan ekspresi yang signifikan. Mekanisme jangkitan atau keradangan yang berlaku dikaitkan dengan perubahan dalam beberapa laluan isyarat sel penting seperti laluan TLR2, MAPKinase dan $NF\kappa\beta$. Laluan-laluan ini penting dari perspektif terapeutik, kerana menyasarkan laluan-laluan yang mungkin membantu merencatkan, memulihkan atau mencegah keradangan. Objektif utama dalam kajian ini adalah untuk menentukan penglibatan laluan isyarat TLR2, MAPKinase dan NFκβ dalam ekspresi DEFB109 di dalam HCEC yang dirangsang oleh Pam3CSK4. Teknikteknik yang terlibat dalam kajian ini termasuk, rangsangan sel kultur oleh Pam3CSK4, pendedahan sel-sel terhadap perencat faktor transkripsi khusus, qPCR dan analisis dot blot. Dalam kajian ini, keterangan menunjukkan bahawa TLR2 menginduksi mRNA



HBD9 dan ungkapan protein dalam cara yang berkaitan dengan masa dan dos yang diberikan. *TLR2* siRNA dilihat mengganggu induksi *TLR2*-pengantara *DEFB109*. Penglibatan jalur MAPKinase dan laluan NF $\kappa\beta$ dalam ekspresi gen *DEFB109* juga diperhatikan. Molekul khusus jalur ini boleh dieksploitasi untuk memodulasi tindak balas HBD9 semasa jangkitan mikrob. Ekspresi bervariasi dari AMP yang berbeza kepada patogen spesifik akan mencadangkan laluan yang serupa tetapi halus yang berbeza dipanggil oleh patogen, mungkin berkaitan dengan PAMP mereka dan TLR yang berlainan dan reseptor lain yang mereka sambungkan.

~

ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious, the Most Merciful All gratification are referred to Allah

All praise be to Allah, the Almighty for His consent for giving me the courage and strength in completing my Master study and research.

First and foremost, I would like to convey my deepest gratitude to my supervisor, Assoc. Prof. Dr. Nazri b. Omar, who greatly enriched my knowledge with his guidance and assistance. Devoid of his constant support, the master research would have not been accomplished.

I would like to thank the rest of my supervisors, Dr. Rafidah Md Saleh and Dr. Nor Shariza Nordin, for their guidance, encouragement, insightful comments and ideas for making this thesis more meaningful.

A sincere gratitude and appreciation also go to Faculty of Medicine and Health Sciences, Universiti Putra Malaysia, the place that has granted me the opportunity and amenities to collect the essential practical skills and the keen in fulfilling the research. Special note of thanks goes to all medical laboratory technologists and staff of Pathology's department, Surgery's department and Cell Signaling Laboratory at this faculty for their constructive assistance while grappling the handiness laboratory tasks.

Last but not least, a heartiest thank goes to my family and friends for their tireless love, support and motivation throughout my study. Thank you and may peace and blessing be upon those who read.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Nazri Omar, MD, PhD

Associate Professor Faculty Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Rafidah Md Saleh, MD

Senior Lecturer Faculty Medicine and Health Sciences Universiti Putra Malaysia (Member)

Nur Shariza Nordin, PhD Senior Lecturer Faculty Medicine and Health Sciences Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:

Date:

Name and Matric No.: Nurul Hana Binti Haji Zainal Baharin, GS42812

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: Name of Chairman of Supervisory Committee:	Associate Professor Dr. Nazri Omar
Signature: Name of Member of Supervisory Committee:	Dr. Rafidah Md Saleh
Signature: Name of Member of Supervisory Committee:	Dr. Nur Shariza Nordin,

TABLE OF CONTENTS

]	Page
ABST	RACT	,	i
ABST	RAK		iii
ACK	NOWL	EDGEMENTS	v
APPR	OVAL		vi
DECL		TION	V111
LISI LIST	OF IA OF FI	CIDEC	XII
LIST	OF FN	BREVIATIONS	xiv
			211
CHAI	PTER		
1	INTR	ODUCTION	1
	1.1	Background	1
	1.2	Problem Statement	3
	1.3	Research Objectives	4
		1.3.2 Specific Objectives	4 4
	1.4	Hypotheses	4
2	LITE	RATURE REVIEW	5
	2.1	Anatomy of the eye	5
	2.2	Cornea	6
		2.2.1 Epithelium layer	6 7
		2.2.2 Endothenum	7
		2.2.5 Submar's membrane	7
		2.2.5 Descemet's membrane	7
	2.3	Bacteria is one of The Factor of Inflammation or Infection of the	
		Eyes	8
	2.4	Antimicrobial Peptides (AMPs)	8
		2.4.1 History of AMPs	8
	2.5	2.4.2 Properties of AMPs	10
	2.5	Detensin	10
	26	2.5.1 Structure and function of defensins	10
	2.0	2.6.1 MAPKingse Pathways	14
		$2.6.2 NF\kappa\beta$ Pathways	14
-	_		_
3	MET	HODOLOGY	16
	$\frac{3.1}{2.2}$	Experimental design	16 17
	3.2 3.2	Nethodology	1/ 1Q
	3.3	3 3 1 Cell culture	10 19
		5.5.1 Concurate	10

		3.3.2	The optimum concentration of <i>Pam3CSK4</i> in for cell viability in stimulating the upregulation of <i>DEFB109</i>	19
		3.3.3	The cytotoxicity and growth kinetic test of drugs inhibitors of <i>MAPKinase</i> and $NF\kappa\beta$ pathways in	
		3.3.4	<i>Pam3CSK4</i> -stimulated HCECs Signaling pathways determination in the expression of	23
	3.4	Statistic	<i>DEFB109</i> al analysis	24 25
4	RESU	ULTS AN	D DISCUSSIONS	26
	4.1	The opti	mum concentration of <i>Pam3CSK4</i> in stimulating the	
		upregula	ation of DEFB109	26
		4.1.1	Pam3CSK4 Cytotoxicity Testing	26
		412	Pam3CSK4 modulates DEFB109 gene expression in	-
			HCECs	27
		413	A validation step to prove the expression protein of	_ /
		1.1.5	HBD9 in the <i>Pam3CSK4</i> stimulated HCECs by dot	
			hlot analysis	27
	12	The cyto	provide and growth kinetic test of drug inhibitors of	21
	7.2	MADKin	ase and NEkB pathways in <i>Pam3CSK4</i> stimulated	
			ase and WIKD pathways in TamsCSK4-stimulated	20
		1 2 1	The optimum concentration for MAPKingse and NEVR	50
		4.2.1	nethyper drugs inhibitor in Day 2CEKA stimulated	
			LCEC-	20
		122		30
		4.2.2	The effect of the compound on the growth kinetic of	21
		T 1 ·	immortalized HCECs	31
	4.3	The sign	naling pathways in the expression of <i>DEFB109</i>	34
		4.3.1	<i>TLR2</i> plays a key role in <i>DEFB109</i> gene expression in HCECs	34
		432	A validation step to prove the expression of HBD9	51
		4.5.2	protein in the HCEC treated with TLP2 siPNA by dot	
			blot analysis	35
		122	MAPKs and $NErch$ are involved in TLP2 induced	55
		4.5.5	DEEP100 gono	27
			DEF DIO9 gene	57
5	SUM	MARY A	ND CONCLUSION, LIMITATION OF THE STUDY	
	AND	RECOM	MENDATIONS FOR FUTURE RESEARCH	40
	5.1	Summar	y and conclusion	40
	5.2	Limitati	on of the study	40
	5.3	Recomn	nendations for future research	41
REF	ERENC	CES		42
APP	ENDIC	ES		55
BIOI	DATA (OF STUD	ENT	60
				50

xi

LIST OF TABLES

Table		Page
3.2	Experimental design of HCECs treated with different concentration of <i>Pam3CSK4</i>	20
3.3	Experimental design of HCECs treated with compound inhibitors	21

LIST OF FIGURES

Figur	e	Page
2.1	The cross axial sectional view of the anatomy part of human eye	5
2.2	Cross section of human cornea	6
2.6	Inflammatory pathways in the expression of human defensin	13
3.1	Experimental design	17
4.1	Cytotoxicity test of <i>Pam3CSK4</i> in HCECs	26
4.2	<i>Pam3CSK4</i> modulates <i>DEFB109</i> gene expression in HCECs	27
4.3	Dot blot analysis of HBD9 in the <i>Pam3CSK4</i> stimulated HCECs	28
4.4	HBD9 protein expression in HCECs incubated with <i>Pam3CSK4</i>	28
4.5	The cytotoxicity test for drugs inhibitors	31
4.6	The effect of drug inhibitors compound on the growth kinetic of HCECs	33
4.7	<i>DEFB109</i> mRNA in HCECs incubated with <i>TLR2</i> siRNA	35
4.8	The expression of HBD9 protein in HCECs incubated with <i>Pam3CSK4</i> after treated with <i>TLR</i> 2 siRNA	36
4.9	<i>DEFB109</i> protein in HCECs incubated with <i>Pam3CSK4</i> after treated with <i>TLR2</i> siRNA	37

LIST OF ABBREVIATIONS

	AA	Antibiotic-Antimycotic
	AMPs	Antimicrobial peptides
	ATF2	Activating transcription factor 2
	BAFF	B cell activating factor
	BPE	Bovine Pituitary extract
	BSA	Bovine serum albumin
	CAP18	Cationic antimicrobial protein of 18 kDA
	CAP35	Cationic antimicrobial protein of 35 kDA
	CCL20	Chemokine ligand 20
	CCR-6	Chemokine receptor 6
	cDNA	Complementary DNA
	DEFB109	Human beta defensin 9 gene
	EGF	Epidermal growth factor
	ERK	Extracellular regulated kinase
	gDNA	Genomic DNA
	HBD9	Human beta defensin 9
	HCECs	Human corneal epithelial cell
	НЕК293	Human embryonic kidney 293
	HNP	Herniated nucleus pulposus
	ΙΚΚα	Inhibitory κB kinase-alpha
	ΙΚΚβ	Inhibitory κB kinase-beta
	ΙΚΚγ	Inhibitory KB kinase-gamma

xiv

	IL-1	Interleukin-1
	IL-1β	Interleukin-1 Beta
	JIPS	JNK-interacting proteins
	JNK	c-Jun N-terminal kinase
	LL37	Leucine-leucine 37
	LPS	Lipopolysaccharide
	LTA	Lipoteicoid acid
	MAP2K	Mitogen activated protein kinase kinase
	МАРЗК	Mitogen activated protein kinase kinase kinase
	МАРК	Mitogen activated kinase
	MP1	Mouse protamine-1
	NADH	Nicotinamide adenine dinucleotide – hydrogen
	NAPDH	Nicotinamide adenine dinucleotide phosphate hydrogen
	NFĸB	Nuclear factor kappa beta
	Ng	Nitroglycerin
	NLRs	NOD like receptors
	p38	p38 mitogen activated kinase
	Pam3CSK4	Palmitoyloxy) ₃ -cysteinyl-serine-(lysine) ₄
	PAMPs	Pathogen associated molecular patterns
	PA01	Pseudomonas aeruginosa 01
	PGN	Peptidoglycan
	PRR	Pathogen recognition receptors

RANKL	Receptor Activator of Nuclear Factor Kappa B Ligand
siRNA	Short interference ribonucleic acid
TAK-1	TGFbeta activated kinase 1
TLR2	Toll-like receptor 2
TLRs	Toll-like receptors
TNFSF3	TNF superfamily 3
TNF-α	Tumor necrosis factor – alpha
TX	Texas
WB	Wipeout buffer

CHAPTER 1

INTRODUCTION

1.1 Background

Corneal epithelium is constantly exposed to numerous pathogens and to physical insult from the environment. It is responsible as a first line of defense, providing a barrier against external assaults (Chalovich & Eisenberg, 2005). Frequently, the damage of this highly specialized structure could lead to vision loss (Bashir et al., 2017). The inflammation of the eyes has become among the most severe issues in this world, proven by prevalence that has been stated in the population study in the United States, which dry eye, the symptom of keratitis, becoming among the most frequent eye disorders, ranges from 5% to less than 35% at various ages of the adult population (Kenneth, 2017).

One of the factors that can caused damage to the corneal epithelium is eye infections or inflammation, which it can be occurred when harmful microorganism such as bacteria, fungi and viruses invade any part of the eyeball or surrounding area (Fini & Stramer, 2005). Among the microorganisms, bacteria such as *Staphylococci*, *Haemophillus, Streptococci* and *Psedomonas* are the most frequently responsible for the inflammation of the eye (Akpek & Gottsch, 2003).

To fight against infections, corneal epithelium has several defense mechanisms such as immunological defense mechanism with features characteristic of the immune system. In the inflammatory response, the innate immune system employs Toll-like receptors (TLRs) to acknowledge and tie-up the pathogen associated molecular patterns (PAMPs). TLRs are the recognition system that utilizes PAMPs through epithelial cells in respond to the exposure of microbial infection (Akira et al., 2001). Like other mucosal epithelial cells, corneal epithelial cells will conscript inflammatory cells in response to pathogenic bacteria and their products (Solomon et al., 2004), by the recognition of the pathogen engage in the host response. This reaction will lead to the expression and the secretion of proinflammatory cytokines (Kumar et al., 2005).

The inflammatory response is characterized by coordinated activation of various signaling pathways that regulate expression of both pro- and anti-inflammatory mediators in tissues. The frequent pathways that were believed to be involved in the inflammatory response were TLRs, *MAPKinase* and NF-κB pathways. Among the TLRs family, *TLR2* was shown to accept a broad spectrum of PAMPs, including lipoteichoic acids (LTA), peptidoglycan (PGN) and bacterial lipoproteins from Grampositive bacteria cell wall (Takeuchi et al., 2000) which, successively, initiate MAPK- or NF-κB-dependent cascades that peak in a pro-inflammatory response. This reaction requires the secretion of chemokines, cytokines and defensins (Takeuchi et al., 2000).

In inflammatory response, cytokines and chemokines activate the humoral and cellular adaptive immune system. In contrast to the effects of cytokines and chemokines, the action of defensins is double-speared (Esche et al., 2005). Defensins have direct lytic effect on the infective agent and served as the rapid, first line of host defense. Thus, defensin was shown to play an important role in the innate immune systems (Roby & Nardo, 2013). They protect the host against invading organisms such as bacteria (Bals, 2000; Koczulla & Bals, 2007), fungi (Krishnakumari et al., 2009) and viruses (Mateua et al., 2003). In addition to the antimicrobial effect, defensins have also been proven as chemo-attractant (Niyonsaba et al., 2002), anti-cancer (Droin et al., 2009; Okumura et al., 2004) and promote wound healing (Koczulla & Bals, 2007; Steinstraesser et al., 2008)

Defensins are small cationic antimicrobial peptides between 20-50 amino acids with six evolutionary conserved cysteine residues (Ganz & Lehrer, 1995) They can be classified into α , β and θ -defensins. Among of these types of defensins, only α and β defensin have been isolated in human. To date, more than 30 genes belonging to the β -defensin family have been identified in human (Schutte et al., 2002), especially in ocular surface, the localization of human β -defensin (HBD)-1 to HBD3 was reported in superficial layers of normal and inflamed corneal (Terai et al., 2004). Besides, newer member of β -defensin family, *DEFB109*, also known as HBD9 was first reported on the ocular surface by Dua and co-workers in 2008 (Abedin et al., 2008). They demonstrated that *DEFB109* gene was expressed constitutively but was down regulated in the presence of ocular surface infection and inflammation. This was in agreement with earlier findings of Premratanachai and co-workers who demonstrated down regulation of the *DEFB109* gene in gingival candidiasis (Premratanachai et al., 2004). With this unique characteristic, the expression of HBD9 were found interesting to be investigated.

HBD9 is a relatively newly-found defensin in ocular surface and the pathways involve in the expression of this type of defensin was investigated. The existing data interestingly points towards possibly different or additional roles it plays in the host defence (Dua et al., 2014). This study extended the perimeter of our study regarding HBD9 as a potential source of broad spectrum efficacious therapeutic agent in the future. Although the more established defensins such as the HBD1-3 were shown to act via the *MAPKinase* pathway in tissue such as the lung and urinary tract, the pathway may varied for HBD9 in the ocular surface environment. In this project, the pathways that were believed to be involved inflammation were unravelled by blocking the mediator molecules in HCECs after treated with *Pam3CSK4* which is believed to express *DEFB109*, while observing the resultant relative mRNA expression. The importance of the involvement of nuclear factor kappa B (NF- κ B) and mitogen-activated protein kinase (MAPK) families molecules such as p38, c-Jun N-terminal kinase (JNK) and extracellular signal-regulated kinase (ERK), in infection and inflammation, have made all of these pathways interestingly to be studied (Mohammed et al., 2011). The results improved the current knowledge on the inflammation pathways involves in the expression of HBD9 in immortalized HCECs.

1.2 Problem Statement

Corneal epithelium was proven to provide immune defense, against attacking pathogens through a variety of pathogen recognition receptors (PRR) which are capable of recognizing PAMPs, derived from various infection-causing microbes (Mohammed et al., 2010). It will conscript inflammatory cells in response to pathogenic bacteria and their products (Solomon et al., 2004), by the recognition of the pathogen engage in the host response. This reaction will lead to the expression defensins.

Pam3CSK4 is one of the synthetic PAMPs that is believed to be associated in the HBD9 expression. Stimulation of HCECs with *Pam3CSK4* was shown to up regulated the expression of *DEFB109*, which is also known as HBD9 (Mohammed et al., 2010). *DEFB109* was first reported on the ocular surface by Dua and co-workers in 2008 (Abedin et al., 2008). In this study, they demonstrated that *DEFB109* gene was expressed constitutively but was down regulated in the presence of ocular surface infection and inflammation. With this special characteristic, HBD9 has become one of the defensins that draw attention of many researchers nowadays.

Although there were previous researches that studied the effect of *Pam3CSK4* on the expression of HBD9, the pathway associated in the expression of *DEFB109* have been not fully studied yet. Therefore in this research, various steps of the potential pathways in inflammation and infection of the eye such as *TLR2*, *MAPKinase* and *NF\kappa\beta* were tested and the expression of *DEFB109* was analyzed. The new information can be used as a reference in the future investigation. This analysis might be benefit by others scientist or researchers to increase understanding and find a new treatment method for corneal eye diseases. This will drive towards discovery of a broad spectrum, safe but effective resistant-free antibiotic in the future.

 \bigcirc

1.3 Research Objectives

1.3.1 General Objectives

To determine the signaling pathways involved in the expression of HBD9 at RNA and protein level in *Pam3CSK4*-stimulated culture HCECs.

1.3.2 Specific Objectives

- 1. To determine the optimal concentration and incubation time of *Pam3CSK4* in stimulating the upregulation of HBD9 in HCECs.
- 2. To assess the cytotoxicity of *TLR2* siRNA and inhibitors for *MAPKinase* and *NF\kappa\beta* pathways in stimulated HCECs.
- 3. To determine the effects on the expression of HBD9 in *Pam3CSK4*-stimulated HCECs after in treated by *TLR2* siRNA and inhibitors for *MAPKinase* and *NF\kappa\beta* pathways.

1.4 Hypotheses

- Ho 1: There is no significant difference between the expressions of *DEFB109* gene in HCECs before and after stimulated with *Pam3CSK4*
- H₁ 1: There is a significant difference between the expressions of *DEFB109* gene in HCECs before and after stimulated with *Pam3CSK4*
- Ho 2: There is no significant difference between the expressions of *DEFB109* gene in HCECs following the transfection of *TLR2* siRNA and inhibition of *MAPKinase* and *NF*- $\kappa\beta$ signaling pathways.
- H₁ 2: There is a significant difference between the expressions of *DEFB109* gene in HCECs following the transfection of *TLR2* siRNA and inhibition of *MAPKinase* and *NF*-κβ signaling pathways.

REFERENCES

- Abedin, A., Mohammed, I., Hopkinson, A., & Dua, H. S. (2008). A novel antimicrobial peptide on the ocular surface shows decreased expression in inflammation and infection. *Investigative Ophthalmology and Visual Science*, 49(1), 28–33.
- Abraham, S., Lawrence, T. & Kleiman, A. (2006). Antiinflammatory effects of dexamethasone are partly dependent on induction of dual specificity phosphatase 1. *The Journal of Experimental Medicine*, 203(8), 1883-1889.
- Agrawal, N., Dasaradhi, P. V., Mohmmed, A., Malhotra, P., Bhatnagar, R. K., & Mukherjee, S. K. (2003). RNA interference: *Biology, Mechanism & Applications*, 67(4), 657–685.
- Akira, S., Takeda, K., & Kaisho, T. (2001). Toll-like receptors: critical proteins linking inante and acquired immunity. *Nature Immunology*, 2(8), 675–680.
- Akpek, E. K. & Gottsch, J. D. (2003). Immune defense at the ocular surface. *Eye* (*Lond*), 17(8), 949-949.
- Bahar, A. & Ren, D. (2013). Antimicrobial peptides. *Pharmaceuticals*, 6(12), 1543–1575.
- Bals, R. (2000). Epithelial antimicrobial peptides in host defense against infection. *Respiratory Research*, 1(3), 141–150.
- Bals, R., Wang, X., Meegalla, R. L., et. al. (1999). Mouse β -defensin 3 is an inducible antimicrobial peptide expressed in the epithelia of multiple organs. *Infection and Immunity*, 67(7), 3542–3547.
- Bashir, H., Seykora, J. T., & Lee, V. (2017). Invisible Shield: Review of the Corneal Epithelium as a Barrier to UV Radiation, Pathogens, and Other Environmental Stimuli. *Journal of Ophthalmic and Vision Research*, *12*(3), 305–311.
- Bastian, A. & Schäfer, H. (2013). Human alpha-defensin 1 (HNP-1) inhibits adenoviral infection in vitro. *Regulatory Pept*ides, 101(1-3), 157-161.
- Beam, M. E., & Toshach, J. C. (2000). Letters to the Editors, A review, 28(3), 217–218.
- Becker, M. N., Diamond, G., Verghese, M. W. & Randell, S. H. (2000). CD14dependent lipopolysaccharide-induced beta-defensin-2 expression in human tracheobronchial epithelium. *Journal of Biology Chemistry*, 275(1), 29731– 29736.

- Bensch, K. W., Raida, M., Mägert, H. J., Schulz-Knappe, P. & Forssmann, W.G. (1995). hBD-1: A novel β-defensin from human plasma. *FEBS Letters*, *368(1)*, 331–335.
- Birchler, T., Seibl, R., Bchner, K., et. al. (2001). Human Toll-like receptor 2 mediates induction of the antimicrobial peptide human beta-defensin 2 in response to bacterial lipoprotein. *European Journal of Immunology*, 31(11), 3131–3137.
- Bonizzi, G. & Karin, M. (2004). The two NF-κB activation pathways and their role in innate and adaptive immunity. *Trends of Immunology*, 25(1), 280–288.
- Bonizzi, G., Bebien, M., Otero, D. C., et. al. (2004). Activation of IKKα target genes depends on recognition of specific κB binding sites by RelB:p52 dimers. *The Embo Journal*, 23(1), 4202–4210.
- Bourcier, T., Thomas, F., Borderie, V., Chaumeil, C. & Laroche, L. (2003). Bacterial keratitis: predisposing factors, clinical and microbiological review of 300 cases. *The British Journal of Ophthalmology*, 87(7), 834-838.
- Brackett, D. J., Lerner, M. R., Lacquement, M. A., He, R. & Pereira, H. A. (1997). A synthetic lipopolysaccharide-binding peptide based on the neutrophil-derived protein CAP37 prevents endotoxin-induced responses in conscious rats. *Infection and Immunity*, 65(7), 2803–11.
- Brown, K. L., & Hancock, R. E. (2006). Cationic host defense (antimicrobial) peptides. *Current Opinion in Immunology*, 18(1), 24–30.
- Cart, M. Y., Issue, C., Section, A., Board, E., About, M., & Issue, O. A. N. (2000). European Cytokine Network, 1(1), 8–11.
- Chalovic, M & Eisenberg, L. (2005). Ocular drug delivery. *The AAPSS Journal, 12*(3), 348-360.
- Chen, Y., Xie, D., Yin Li, W., et. al. (2010). RNAi targeting EZH2 inhibits tumor growth and liver metastasis of pancreatic cancer in vivo. *Cancer Letters*, 297(1), 109–116.
- Chi, H., Barry, R., Roth, R. J. & Wu, J. J. (2006). Dynamic regulation of pro and antiinflammatory cytokines by MAPK phosphatase 1 (MKP-1) in innate immune responses. *Proceedings of the National Academy of Sciences of the United States of America*, 103(7), 2274-2279.
- Cho, Y. S., Challa, S., Moquin, D., et. al. (2009). Phosphorylation-driven assembly on the RIP1-RIP3 complex regulates programmed necrosis and virus-induced inflammation. *Cell*, 137(6), 1112-1123.
- Conlon, J. M., & Sonnevend, A. (2010). Antimicrobial Peptides. A review, 618(2), 3–14.

- Coskun, P. E., Wyrembak, J., Derbereva, O., et. al. (2010). Systemic mitochondrial dysfunction and the etiology of Alzheimer's diseases and down syndrome dementia. *Journal of Alzheimers*, 20(2), 293-310.
- Cubrey, J. A., Steelman, L. S., Abrams, S. L., et. al. (2006). Roles of the RAF/MEK/ERK and PI3K/PTEN/AKT pathways in malignant transformation and drug resistance. *Advances in Enzyme Regulation*, 46(1), 249-279.
- Cuello, O. H., Caorlin, M. J. & Reviglio, V. E. (2002). Rhodococcus globerulus keratitis after laser in situ keratomileusis. *Journal of Cataract and Refractive Surgery*, 28(1), 2235-2237.
- Danielle, M. R., Li, L., Stephen, F., et. al. (2005). Characterization of growth and differentiation in a tolemerase-immortalized human corneal epithelial cell line. *Investigative Ophthalmology and Visual Science*, 2(46), 470-478.
- Dejardin, E., Droin, N. M., Delhase, M., et. al. (2002). The lymphotoxin-β receptor induces different patterns of gene expression via two NF-κB pathways. *Immunity*, 17(1), 525–535.
- Dermott, M., Rich, D., Cullor, J., et. al. (2006). The in vitro activity of selected defensins against an isolate of Pseudomonas in the presence of human tears. *The British Journal of Ophthalmology*, 90(5), 609–611.
- Dhillon, A. S., Hagan, S., Rath, O. & Kolch, W. (2007). MAP kinase signalling pathways in cancer. Oncogene, A review, 26(1), 3279-3290.
- Diamond, G., Beckloff, N., Weinberg, A. & Kisich, K. O. (2009). The roles of antimicrobial peptides in innate host defense. *Current Pharmaceutical Design*, 15(21), 2377–2392.
- Droin, N., Hendra, J.B., Ducoroy, P., et. al. (2009). Human defensins as cancer biomarkers and antitumour molecules. *Journal of Proteomics*, 72(1), 918-927.
- Dua, H., Otri, A. & Hopkison, A. (2014). In vitro studies on the antimicrobial peptide human beta-defensin 9 (HBD9): signaling pathways and pathogen-related response (an American Ophthalmological Society thesis). *Transactions of the American Ophthalmological Society*, 112(1), 50-73.
- Dubos, R. J. (1939). Studies on a bactericidal agent extracted from a soil bacillus: Preparation of the agent and its activity in vitro. *The Journal of Experimental Medicine*, 70(1), 1–10.
- Esche, C., Stellato, C. & Beck, L. (2005). Chemokines: Key players in innate and adaptive immunity. *Journal of Investigative Dermatology*, 125(4), 615-628.
- Eun, K. & Eui. J. C. (2010). Pathological roles of MAPK signaling pathways in human diseases. *Molecular Basis of Diseases*, 1802(1), 396-405.

- Fini, M. E., & Stramer, B. M. (2005). How the cornea heals: cornea-specific repair mechanisms affecting surgical outcomes. *Cornea*, 24(1), S2–S11.
- Fleiszig, S. M. & Evans, D. J. (2003). Contact lens infections: can they ever be eradicated? *Eye Contact Lens*, 29(1), S67-S61.
- Ganz, T. (2003). The role of antimicrobial peptides in innate immunity. *Integrative* and Comparative Biology, 43(2), 300–304.
- Ganz, T., & Lehrer, R. I. (1995). Defensins. Pharmacology & Therapeutics, 66(2), 191–205.
- Ghosh, S. & Karin, M. (2002). Missing pieces in the NF-κB puzzle. *Cell*, 109(1), S81–S96.
- Gordon, Y. J, Romanowski, E. G. & McDermott, A. M. (2005). A review of antimicrobial peptides and their therapeutic potential as anti-infective drugs *Current Eye Research*, 30(7), 505-515.
- Graham, J. E, Moore, J. E. & Jiru, X. (2007). Ocular pathogen or commensal: a PCRbased study of surface bacterial flora in normal and dry eyes. *Investigative Ophthalmology and Visual Science*, 48(1), 5616–5623.
- Groenink, J., Walgreen, W. E., Hof, W., Veerman, E. C. & Nieuw, A. V. (1999). Cationic amphipathic peptides, derived from bovine and human lactoferrins, with antimicrobial activity against oral pathogens. *FEMS Microbiology Letters*, 179(2), 217–22.
- Gunshefski, L., Mannis, M. J., Cullor., et. al. (1994). In vitro antimicrobial activity of Shiva-11 against ocular pathogens. *Cornea*, 13(3), 237–242.
- Hancock, R. E., & Diamond, G. (2000). The role of cationic antimicrobial peptides in innate host defences. *Trends in Microbiology*, 8(9), 402–410.
- Hancock, R. E., & Scott, M. G. (2000). The role of antimicrobial peptides in animal defenses. *Proceedings of the National Academy of Sciences of the United States of America*, 97(16), 8856–8861.
- Harder, J., Siebert, R., Zhang, Y., et. al. (1997). Mapping of the gene encoding human beta-defensin-2 (DEFB2) to chromosome region 8p22-p23.1. *Genomics*, 46(3), 472–475.
- Harris, F., Dennison, S. R. & Phoenix, D. A. (2009). Anionic Antimicrobial Peptides from Eukaryotic Organism. *Current Protein and Peptide Sciences*, 6(10), 585-606.

- Haynes, R. J., Tighe, P. J., & Dua, H. S. (1999). Antimicrobial defensin peptides of the human ocular surface. *The British Journal of Ophthalmology*, 83(1), 737– 741.
- Hertz, L., Robinson, S., Griffin, J. & Magistreet, P. (2003). Energy of neurotransmitter. *Science*, 5428(285), 639-639.
- Hill, C. P., Yee, J., Selsted, M. E., & Eisenberg, D. (1991). Crystal structure of defensin HNP-3, an amphiphilic dimer: mechanisms of membrane permeabilization. *Science*, 251(1), 1481-1485.
- Huang, L. C., Petkova, T. D., Reins, R. Y., Proske, R. J., & McDermott, A. M. (2006).
 Multifunctional roles of human cathelicidin (LL-37) at the ocular surface.
 Investigative Ophthalmology and Visual Science, 47(6), 2369–2380.
- Izadpanah, A. & Gallo, R. L. (2005). Antimicrobial peptides. *Journal of the American Academy of Dermatology*, *52*(3), 381–390
- Jester, J. V., Moller, P. T., Hung, J., et. al. (1999). The cellular basis of corneal transperancy: evidence for 'corneal crystallin. *Journal of Cell Science*, 112(1), 613-622.
- Jones, D. E. & Bevins, C. L. (1992). Paneth cells of the human small intestine express an antimicrobial peptide gene. *The Journal of Biological Chemistry*, 267(32), 23216-23225.
- Joyce, N. C. (2003). Proliferative capacity of the corneal endothelium. Progress in Retinal and Eye Research, 22(3), 359-89.
- Kagan, B. L., Ganz, T. & Lehrer, R. I. (1994). Defensins: A family of antimicrobial and cytotoxic peptides. *Toxicology*, 87(1-3), 131–149.
- Kahn, C. R., Young, E., Lee, I. H. & Rhim, S. (1993). Human corneal epithelial primary cultures and cell lines with extended life span: in vitro model for ocular studies. *Investigative Ophthalmology and Visual Science*, 34(1), 3429–3441.
- Kenneth, A. B., Jodi, L., Cynthia, M., et. al. (2017). Current Opinions and Modern Approaches in the Diagnosis and Treatment of Dry Eye Diseases. *New York Eye and Ear Infirmary of Mount Sinai and MedEdicus LLC, 1*(1), 1-3.

Khanna, M., Srivastava, L. M. & Kumar, P. (2003). Deffective interleukin-2 receptor expression in pulmonary tuberculosis. *Journal of Communication Diseases*, 35(2), 65-70.

- Kim, J., Campbell, B., Mahoney, N., Chan, K., Molyneux, R. & May, G. (2008). Chemosensitization prevents tolerance at Aspergilus fumigatus to antimycotic drugs. *Biochemical and Biophysical Research Communications*. 372(1): 266-271.
- Kindrachuk, J., Jenssen, H., Elliott, M., et. al. (2013). Manipulation of innate immunity by a bacterial secreted peptide: lantibiotic nisin Z is selectively immunomodulatory. *Innate Immunity*, 19(3), 315–27.
- Koczulla, R. & Bals, R. (2007). Cathelicidin Antimicrobial Peptides Modulate Angiogenesis. *Therapeutic Neovascularization-Quo Vadis?*, 12(21), 191-196.
- Krishnakumari, V., Ramgaraj, N. & Nagaraj, R. (2009). Antifungal activities of human beta defensin HBD-1 to HBD-3 and their C-terminal analog Phd1 to Phd. Antimicrobiology Journal, 53(1), 256-260.
- Kumar, A., Zhang, J., & Yu. F., S. (2005). Innate immune response of corneal epithelial cells to Staphylococcus aureus infection: Role of peptidoglycan in stimulating proinflammatory cytokine secretion. *Investigative Ophthalmology* and Visual Science, 45(10), 3513-3522.
- Kunt, H. (2016). Investigating science student teachers ideas about function and anatomical form of two human sensory organs, the eye and the ear. *International Journal of Environment and Science Education*, 11(5), 535-542.
- Lan, W., Petznick, A., Heryati, S., Rifada, M. & Tong, L. (2012). Nuclear factor-κβ: Central regulator in ocular surface inflammation and diseases. *Ocular Surface*, *10*(3), 137–148.
- Larrick, J. W., Hirata, M., Balint, R. F., Lee, J., Zhong, J., & Wright, S. C. (1995). Human CAP18: A novel antimicrobial lipopolysaccharide-binding protein. *Infection and Immunity*, 63(4), 1291–1297.
- Lawrence, T. (2009). The nuclear factor NF-кВ pathway in inflammation. *Cold Spring Harbor Perspectives in Biology*, *1*(6), a001651-a001651.
- Legarda, D., Klein-Patel, M. E., Yim, S., Yuk, M. H., & Diamond, G. (2005). Suppression of NF-κB-mediated β-defensin gene expression in the mammalian airway by the *Bordetella* type III secretion system. *Cellular Microbiology*, 7(4), 489–497.
- Leippe, M. (1999). Antimicrobial and cytolytic polypeptides of amaeboid protozoaeffector molecules of primitive phagocytes. *Developmental and Comparative Immunology*, 23(4-5), 267–279.
- Liu, A. Y., Destoumieux, D., Wong, A. V, et. al. (2002). Human beta-defensin-2 production in keratinocytes is regulated by interleukin-1, bacteria, and the state of differentiation. *The Journal of Investigative Dermatology*, *118*(2), 275–281.

- Liu, L., Wang, L., Jia, H. P., et. al. (1998). Structure and mapping of the human betadefensin HBD-2 gene and its expression at sites of inflammation. *Gene*, 222(2), 237–244.
- Livak K. J. & Schimittgen T. D. (2008). Analyzing real-time PCR data by the comprative C(t) method. Nature Protocol, 3(6), 1101-1108.
- Loppnow, H., Libby, P., Freudenberg, M., Krauss, J. H., Weckesser, J., & Mayer, H. (1990). Cytokine induction by lipopolysaccharide (LPS) corresponds to lethal toxicity and is inhibited by nontoxic Rhodobacter capsulatus LPS. *Infection and Immunity*, 58(11), 3743–3750.
- Ma, Y., Liu, C., Liu, X., et. al. (2010). Peptidomics and genomics analysis of novel antimicrobial peptides from the frog, Rana nigrovittata. *Genomics*, 95(1), 66–71.
- Massari, P., McClure, R., & Massari, P. (2015). TLR-dependent human mucosal epithelial cell responses to microbial pathogens. *A review*, *1*(1), 1-1
- Mateua, Michael, M. L., Zhimin, F., et al. (2003). Human epithelial Beta-defensins 2 and 3 inhibit HIV-1 replication. *AIDS*. 17(1), F39–F48.
- Matsushima, A., Kaisho, T., Rennert, P. D., et. al. (2001). Essential role of nuclear factor NF- κ B-inducing kinase and inhibitor of κ B (I κ B) kinase α in NF- κ B activation through lymphotoxin β receptor, but not through tumor necrosis factor receptor I. *Journal of Experimental Medicine*, 193(1), 631–663.
- McIntosh, R. S., Cade, J. E., Abed, M., et. al. (2005). The spectrum of antimicrobial peptide expression at the ocular surface. *Investigative Ophthalmology and Visual Science*. 46(4), 1379–138.
- Mohammed, I., Suleman, H., Otri, A. M., et. al. (2010). Localization and gene expression of human beta-defensin 9 at the human ocular surface epithelium. *Investigative Ophthalmology and Visual Science*, *51*(9), 4677–4682.
- Mohammed, I., Yeung, A., Abedin, A., Hopkinson, A., & Dua, H. S. (2011). Signalling pathways involved in ribonuclease-7 expression. *Cellular and Molecular Life Sciences*, 68(11), 1941–1952.
- Morrison, D. K. & Davis, R. J. (2003). Regulation of MAP Kinase signaling modules by scaffold proteins in mammals. *Annual Review of Cell and Developmental Biology*, 19(1), 91-118.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival. Application to Proliferation and Cytotoxicity Assay. *Journal of Immunological Methods*, 65(1), 56-63.

- Muller, L. J., Pels, E. & Vrensen, G. F (1995). Novel aspects of the ultrastructural organisation of human corneal keratocytes. *Investigative Ophthalmology and Visual Science*, *36*(13), 2557–2567.
- Narayanan, S., Miller, W. L., & McDermott, A. M. (2003). Expression of human betadefensins in conjunctival epithelium: relevance to dry eye disease. *Investigative Ophthalmology and Visual Science*, 44(9), 3795–801.
- Nijnik, A., Pistolic, J., Filewod, N. C. & Hancock, R. E. (2012). Signaling pathways mediating chemokine induction in keratinocytes by cathelicidin LL-37 and flagellin. *Journal of Innate Immunity*, *4*(4), 377–386.
- Ninomiya, T. J., Kishimoto, K., Hiyama, A., Inoue, J., Cao, Z. & Matsumoto, K. (1999). The kinase TAK-1 can activate the NIK-1 kappaB as well as the MAP kinase cascade in the IL-1 signaling pathways. *Nature*, *1*(398), 252-256.
- Niyonsaba, F., Iwabuchi, K., Matsuda, H., Ogawa, H., & Nagaoka, I. (2002). Epithelial cell-derived human beta-defensin-2 acts as a chemotaxin for mast cells through a pertussis toxin-sensitive and phospholipase C-dependent pathway. *International mmunology*, 14(4), 421–426.
- Nosjean (1999). Regulation of dendritic cell trafficking: A process that involves the participations of selective chemokines. *Journal of Leukocyte Biology*, 66(1), 252-262.
- Novack, D. V., Yin, L., Hagen-Stapleton, A., et. al. (2003). The IkB function of NFkB2 p100 controls stimulated osteoclastogenesis. *Journal of Experimental Medicine*, 198(1), 771–781.
- Ohgami, K., Ilieva, I. B., Shiratori, K., et. al. (2003). Effect of human cationic antimicrobial protein 18 Peptide on endotoxin-induced uveitis in rats. *Investigative Ophthalmology* Ohtani, S., Okada, T., Kagamiyama, H., Yoshizumi, H. & No, M. (1977). Complete a lethal primary protein structures for brewer's of two yeast subunits from of purothionin flour, *Journal of Biochemistry*, 82(3), 753-767.
- Okumura, K., Itoh, A., Isogai, E., et al. (2004). C-terminal domain of human CAP18 antimicrobial peptide induces apoptosis in oral squamous cell carcinoma SAS-H1 cells. *Cancer Letters*, 212(1), 185–194.
- Oppenheim, J. J., Biragyn, A., Kwak, L. W., & Yang, D. (2003). Roles of antimicrobial peptides such as defensins in innate and adaptive. *Immunity Annrheumdis and Rheumatology Disease*, 62(1), ii17–ii21.
- Patel, A. A., Cai, Y., Sang, Y., Blecha, F., & Zhang, G. (2005). Cross-species analysis of the mammalian-defensin gene family : presence of syntenic gene clusters and preferential expression in the male reproductive tract. *A review*, *1*(1), 5–17.

- Peng, W. C., Lau, W., Madoori, P. K., Forneris., F., Granneman, J. C., Clevers, H. (2013). Structures of Wnt-Antagonist ZNRF3 and its complex with R-spondin 1 and implication for signaling. *Plos One*, 8(12), e83110-e83110.
- Peters, B. M., Shirtliff, M. E. & Jabra-Rizk, M. A. (2010). Antimicrobial peptides: Primeval molecules or future drugs? *Plos Pathogens*, 6(10), e1001067e1001067.
- Pol, C. V. (2003). Basic anatomy and physiology of the human. *Basic Anatomy and Physiology of the Human Muscle, Ciliary*, 6(1), 238–246.
- Premratanachai, P., Joly, S., Johnson, G. K., et al. (2004). Expression and regulation of novel human beta-defensins in gingival keratinocytes. *Oral Microbiology and Immunology*, 19(2), 111–117.
- Radek, K. & Gallo, R. (2007). Antimicrobial peptides: natural effectors of the innate immune system. *Seminars in Immunopathology*, 29(1), 27–43.
- Raj, P. A. & Dentino, A. R. (2002). Current status of defensins and their role in innate and adaptive immunity. *Science*, 206, 9–18.
- Ramesh, S., Ramakrishnan, R., Bharati, M., Javahar A. M. & Viswanathan, S. (2010). Prevalence of bacterial pathogens causing ocular infections in South India. *Indian Journal of Pathology and Microbiology*, 2(53), 281-286.
- Rammelkamp, C. H. & Weinstein, L. (1941). Trycodine. A review, 1(1), 1-1
- Riedel, S., Zweytick, D. & Lohner, K. (2011). Membrane-active host defense peptides
 Challenges and perspectives for the development of novel anticancer drugs. *Chemistry* and Physics of Lipids, 164(8), 766–781.
- Robert, P. Y., Traccard, I. & Adenis, J. P. (2002). Multiplex detection of herpes viruses in tear fluid using the "stair primers" PCR method: prospective study of 93 patients. *Journal of Medical Virology*, 66(1), 506-11.
- Roby, K. D. & Nardo, A. D. (2013). Innate immunity and the role of the antimicrobial peptide cathelicidin in inflammatory skin disease. *Drug Discovery Today*. *Disease Mechanisms*, 10(3-4), e79-e82.
- Rosillo, M. A., Sanchez, H. M., Cardeno, A., et. al. (2012). *Pharmacology Research*, 66(3), 235-242.
- Salojin, K., Owusu, I. & Millerchip, K. (2006). Essential role of MAPK phosphatase-1 in the negative control of innate immune responses. *Journal of Immunology*, *176*(3), 1899-1907.

- Sato, T., Esaki, M., Fernandez, J. M., & Endo, T. (2005). Comparison of the proteinunfolding pathways between mitochondrial protein import and atomic-force microscopy measurements. *Proceedings of National Academic of Science*, 102(50), 17999-8004.
- Schaeffer, H. J. & Weber, M. J. (1999). Mitogen-activated protein kinases: specific messages from ubiquitous messengers. *Molecular and Cell Biology*, 19(1), 2435–2444.
- Schauber, J., Gallo, R. L., Udall, D., Kanada, K., Yamasaki, K. & Alexandrescu, D. (2009). NIH Public Access. *Journal of Allergy*, 122(4), 829–831.
- Schibli, D. J., Hunter, H. N., Aseyev, V., et. al. (2002). The solution structures of the human defensins lead to a better understanding of the potent bactericidal activity of HBD3 against *Staphylococcus aureus*, *Journal of Biological Chemistry*, 277(10), 8279–8289.
- Schuman, S., Terao, M., Garattini, E., et. al. (2009). Site directed mutagenesis of amino acid residues at the active site of mouse aldehyde oxidase AOX1. *Plos One*, 4(4), 5348-5348.
- Schutte, B. C., Mitros, J. P., Bartlett, J. A., et al. (2002). Discoveries of five conserved beta-defensin gene clusters using a computational search strategy. *Proceeding* of the National Academy of Sciences of the United States of America, 99(1), 2129-2133.
- Scott, M. G., Rosenberger, C. M., Gold, M. R., Finlay, B. B., & Hancock, R. E. (2000). An -helical cationic antimicrobial peptide selectively modulates macrophage responses to lipopolysaccharide and directly alters macrophage gene expression. *The Journal of Immunology*, 165(6), 3358–3365.
- Secker, G. & Daniels, J. (2016). Limbal epithelial stem cells of the cornea., *Stembook*, *I*(1), 1–18.
- Sen, C. K., Khanna, S., Roy, S. & Packer, L. (2000). Molecular basis of vitamin E action. Tocotrienol potently inhibits glutamate-induced pp60(c-Src) kinase activation and death of HT4 neuronal cells. *Journal of Biological Chemistry*, 275(1), 13049–13055.
- Senftleben, U., Cao, Y., Xiao, G., et. al. (2001). Activation by IKKα of a second, evolutionary conserved, NF-κ B signaling pathway. *Science*, 293(1), 1495– 1495.
- Sharma, S. (2000). Diagnosis of external ocular infections: microbiological processing and interpretation. *British Journal of Ophthalmology*, 1(1), 84-229.
- Solomon, A. W., Peeling, R. W., Foster, A., & Mabey, D. C. (2004). Diagnosis and Assessment of Trachoma. *Clinical Microbiology Reviews*, 17(4), 982–1011.

- Steinstraesser, L., Vranckx, J. J., Mohammadi, T. A., et. al. (2006). A novel titanum wound chamber for the study of wound infections in pigs. *Comprative Medicine*, 56(4), 279-285.
- Stokes, R. (1941). The production of bactericidal substances by aerobic sporulating Bacilli from cultures of an aerobic sporulating bacillus isolated from soil (strain B.G). *Journal of Bacteria*. 1(1), 629–640.
- Sumikawa, Y., Asada, H., Hoshino, K., et al. (2006). Induction of beta-defensin 3 in keratinocytes stimulated by bacterial lipopeptides through toll-like receptor 2.
 Microbes Infection, 8(6), 1513-1521.
- Szyk, A., Wu, Z., Tucker, K., Yang, D. E., Lu, W., & Lubkowski, J. (2006). Crystal structures of human α-defensins HNP4, HD5 and HD6, *Protein Science*, 1(1), 2749–2760.
- Ta, C. N., Chang, R. T. & Singh, K. (2003). Antibiotic resistance patterns of ocular bacterial flora: a prospective study of patients undergoing anterior segment surgery. *Ophthalmology*, 110(1), 1946–51.
- Takeuchi, O., Hoshino, K. & Akira, S. (2000). Cutting edge: TLR2-deficient and MyD88-deficient mice are highly susceptible to *Staphylococcus aureus* infection. *Journal of Immunology*, *165*(10), 5392–5396.
- Tang, Y. Q., Yuan, J., Osapay, G., et. al. (1999). A cyclic antimicrobial peptide produced in primate leukocytes by the ligation of two truncated alphadefensins. *Science*, 286(5439), 498–502.
- Terai, K., Sano, Y. & Kawasaki S. (2004). Effects of dexamethasone and cyclosporin on human beta-defensin in corneal epithelial cells. *Experimental Eye Research*. 79(1), 175–180
- Thi, T. H., Jean, M. P., Youri, G., Françoise, V. B. & Paul, M. T. (2011). *In vitro* selection of resistance to temocillin in Enterobacter aerogenes. *8th International symposium on Antibiotic and Resistance, Seoul, Korea*, 1(1), 6-8.
- Torii, S., Yamamoto, T., Tsuchiya, Y. & Nishida, E. (2006). ERK MAP kinase in G1 cell cycle progression and cancer. *Cancer Science*, *97*(8), 697-702.
- Tsutsumi, Y. & Nagaoka, I. (2002). NFkB mediated transcriptional regulation of human β -defensin-2 gene following lipopolysaccharide stimulation. *Journal of Leukocyte Biology*, 71(1), 154-162.
- Valore, E. V., Park, C. H., Quayle, J., Wiles, K. R., McCray, P. B. & Ganz, T. (1998). Human beta-defensin-1: an antimicrobial peptide of urogenital tissues. *The Journal of Clinical Investigation*, 101(8), 1633–1642.

- VanEpps, H. L. (2006). René Dubos: unearthing antibiotics. *The Journal of Experimental Medicine*, 203(2), 259-259.
- Vora, P., Youdim, L., Thomas, L. S. & Fukata, M. (2004). Beta-defensin-2 expression is regulated by TLR signaling in intestinal epithelial cells. *Journal of Immunology*, 173(9), 5398-5405.
- Wang, M.C., Bohmann, D. & Jasper, H. (2003). JNK signaling confers tolerance to oxidative stress and extends lifespan in Drosophila. *Developmental Cell*, 5(5), 811-816.
- Wehkamp, K., Schwichtenberg, J. & Schroder, J. M. (2006). Pseudomonas aeruginosa and IL- beta-mediated induction of human beta-defensin-2 in keratinocyte is control by NFkB and AP-1. *The Journal of Investigative Dermatology*, 126(1), 121-127.
- Weinberg, D. E., Nakanishi, K., Patel, D. J., & Bartel, D. P. (2011). The Inside-Out Mechanism of Dicers from Budding Yeasts. *Cell*, 146(2), 262–276.
- Welling, M. M., Hiemstra, P. S., Barselaar, M. T., et al. (1998). Antibacterial activity of human neutrophil defensins in experimental infections in mice is accompanied by increased leukocyte accumulation. *Journal of Clinical Investigation*, 102(8), 1583-1590.
- White, S. H., Wimley, W. C. & Selsted, M. E. (1995). Structure, function, and membrane integration of defensins. *Current Opinion in Structural Biology*, 5(4), 521–527.
- Whitmarsh, A. J. (2006). The JIP family of MAPK scaffold proteins. *Biochemical Society Transactions*, 34(5), 828-832.
- Willoughby, C. E., Ponzin, D., Ferrari, S., Lobo, A., Landau, K. & Omidi, Y. (2010). Review Anatomy and physiology of the human eye: effects of mucopolysaccharidoses disease on structure and function, Clinical & Experimental of Ophthalmology, 38(S1), 2–11.
- Wu, Y., Gao, B. & Zhu, S. (2017). New fungal defensin-like peptides provide evidence for fold change of proteins in evolution. *Bioscience Reports*, 37(1), BSR20160438.
- Yamagata, M., Rook, S. L., Sassa, Y., et. al. (2006). Bactericidal/permeabilityincreasing protein's signaling pathways and its retinal trophic and antiangiogenic effects. *Faseb Journal : Official Publication of the Federation of American Societies for Experimental Biology*, 20(12), 2058–67.
- Yang, D. (1999). Defensins: Linking innate and adaptive immunity through dendritic and T cell CCR6. *Science*, 286(5439), 525–528

- Yau, S. W., Azar, W. J., Sabin, M. A., Werther, G. A. & Russo, V. C. (2015). IGFBP-2 taking the lead in growth, metabolism and cancer. *Journal of Cell Communication and Signaling*, 9(2), 125–142.
- Zandi, E., Rothward, D. M., Delhase, M., Hayakawa, M. & Karin, M. (1997). The IkB kinase complex (IKK) contains two kinase subunits, IKKα and IKKβ, necessary for IkB phosphorylation and NF-kB activation. *Cell*, 91(1), 243-252.
- Zasloff, M. (2002). Organisms. Current Opinion in Immunology, 6870(415), 389–395.
- Zelezetsky, I. & Tossi, A. (2006). Alpha-helical antimicrobial peptides—Using a sequence template to guide structure–activity relationship studies. *Biochimica et Biophysica Acta (BBA) Biomembranes*, 1758(9), 1436–1449.
- Zeya, H. I. & Spitznagel, J. K. (1963). Antibacterial and enzymic basic proteins from leukocyte lysosomes: Separation and identification. *Science*, 142(3595), 1085–1087.
- Zhang, G. H., Mann, D. M. & Tsai, C. M. (1999). Neutralization of endotoxin in vitro and in vivo by a human lactoferrin-derived peptide. *Infection and Immunity*, 67(3), 1353–1358
- Zhao, Q., Wang, X., Nellin, L. D., et. al. (2005). MAPKinase phosphatase 1 controls innates immune responses and suppresses endotoxic shock. *Journal of Experimental Medicine*, 203(1), 1794-1794.
- Zhao, X., Wu, H., Lu, H., Li, G. & Huang, Q. (2013). LAMP: A database linking antimicrobial peptides. *Plos One*, 8(6), 1–6.
- Zhou, L., Huang, L. Q., Beuerman, R. W., et. al. (2004). Proteomic analysis of human tears: Defensin expression after ocular surface surgery. *Journal of Proteome Research*, 3(3), 410–416.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION :

TITLE OF THESIS / PROJECT REPORT :

INVOLVEMENT OF *TLR2*, *MAPKinase* AND *NFκβ* PATHWAYS IN REGULATION OF HUMAN BETA DEFENSIN 9 IN HUMAN CORNEAL EPITHELIAL CELLS STIMULATED WITH *Pam3CSK4*

NAME OF STUDENT: NURUL HANA BINTI HAJI ZAINAL BAHARIN

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (V)



CONFIDENTIAL

RESTRICTED

OPEN ACCESS

(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from		until	
-	(date)		(date)

Approved by:

(Signature of Student) New IC No/ Passport No.: (Signature of Chairman of Supervisory Committee) Name:

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]