



**UNIVERSITI PUTRA MALAYSIA**

***SOMATIC CELL COUNT, TEAT CHARACTERISTICS, MILK  
COMPOSITION AND SINGLE NUCLEOTIDE POLYMORPHISMS IN  
CANDIDATE GENES FOR MASTITIS IN FRIESIAN AND JERSEY COWS***

**UMAR MOHAMMED SANI**

**FP 2018 18**



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By

**UMAR MOHAMMED SANI**

**Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the degree of Master of Science**

**December 2017**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**SOMATIC CELL COUNT, TEAT CHARACTERISTICS, MILK COMPOSITION AND SINGLE NUCLEOTIDE POLYMORPHISMS IN CANDIDATE GENES FOR MASTITIS IN FRIESIAN AND JERSEY COWS**

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**December 2017**

**Chairman : Professor Jothi Malar Panandam, PhD**  
**Faculty : Agriculture**

Mastitis is a clinical condition of dairy animals characterized by inflammation of the mammary gland. It is classified as either clinical or subclinical, in which the former is associated with physical, chemical and bacteriological changes in milk as well as changes in udder morphological characteristics in dairy cows. Somatic cell count (SCC) is an important indicator of udder health and mastitis. Two types of immunity, natural or innate and acquired immunity, act together to protect the animals' mammary gland especially at early stage of infection. The bovine toll-like receptor 4 gene (*TLR4*), mannose-binding lectin 1 gene (*MBL1*) and peptidoglycan recognition protein 2 gene (*PGLYRP2*) have been identified to be responsible for these immune responses. There is limited information on the single nucleotide polymorphisms (SNPs) in these genes in the Malaysian dairy cattle populations, and reports on their association with mastitis resistance or susceptibility is lacking. This study was undertaken to investigate the udder mastitis status, milk somatic cell score (SCS), teat characteristics, some milk composition traits and SNPs in three candidate genes for bovine mastitis resistance in Friesian and Jersey cows, in order to identify potential genetic markers that may be used in marker assisted selection.

The Friesian and Jersey lactating cows in a commercial dairy farm were used in this research. Milk somatic cell count (SCC) was estimated using a somatic cell counter and converted to SCS, and mastitis status was established using California mastitis test (CMT). Teat lengths were measured, their functioning tested, and the orifices observed and scored. Milk composition traits were determined by infrared spectroscopy. DNA was extracted from the milk of 40 infected and 37 healthy cows, and the individual DNA samples were screened and genotyped for five SNP loci in

the exons and introns of *TLR4*, *MBL1* and *PGLYRP2* using the PCR-restriction fragment polymorphism (PCR-RFLP) technique.

There was no significant ( $P > 0.05$ ) differences in the numbers of infected (based on California mastitis test) and functional teats between Friesian and Jersey cows reared under common management. There was also no significant differences in SCS between the breeds nor between the lactations. However, the late lactation stage milk had the highest SCS ( $P < 0.05$ ). Udder quarters, lactation stage and lactation number had a significant ( $P < 0.05$ ) effect on the teat length. The mean rear quarter teat length was significantly ( $P < 0.05$ ) lower than the front quarter teats. Significant relationship between teat length and quarter mastitis status was observed. The shorter the teat length, the higher was the tendency for the udder quarter to be infected. Teat-end hyperkeratosis score showed a significant differences between breeds and udder quarters. Lactation number, lactation stage, and breed had a significant ( $P < 0.05$ ) effects on milk lactose, but not on solid non-fat. Lactation number and lactation stage had significant ( $P < 0.05$ ) effects on milk fat and protein. There was no significant influence of the genotypes at the investigated loci of the *TLR4*, *MBL 1* and *PGLYRP-2* genes on SCS, milk fat, protein and lactose. There was a significant ( $P < 0.05$ ) association between teat condition and mastitis status. Therefore, udder morphological traits should be included in selection and breeding for mastitis resistance. A larger herd size, evaluation of more SNP loci, and analysis of data from complete lactation cycles of each animal would provide a more valid information on the effects of various influencing factors and informative markers for SCS and milk composition traits for the Friesian and Jersey dairy cattle in Malaysian.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KIRAAN SE SOMATIC, CIRI TEAT, KOMPOSISI SUSU DAN  
POLIMORFISME NUKLEOTIDA TUNGGAL DALAM GEN CALON  
UNTUK MASTITIS DI LEMBU FRIESIAN DAN JERSEY**

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Mastitis adalah penyakit haiwan tenusu yang dicirikan oleh inflamasi kelenjar mamari. Ia diklasifikasikan sebagai klinikal atau subklinikal, di mana yang pertama dikaitkan dengan perubahan fizikal, kimia dan bakteriologi dalam susu serta perubahan dalam ciri morfologi udder dalam lembu tenusu. Bilangan sel somatik (SCC) adalah penunjuk penting dalam kesihatan udder dan mastitis. Dua jenis keimunan, semula jadi atau innat dan imuniti yang diperolehi, bertindak bersama untuk melindungi kelenjar mamari mamalia terutamanya pada peringkat awal jangkitan. Gen *bovine toll-like receptor 4 (TLR4)*, gen mannose-binding lectin 1 (*MBL1*) dan gen peptidoglycan recognition protein 2 (*PGLYRP2*) telah dikenal pasti bertanggungjawab terhadap tindak balas imun ini. Terdapat maklumat terhad mengenai *single nucleotide polymorphism (SNP)* dalam gen ini dalam populasi lembu tenusu di Malaysia, dan laporan mengenai perkaitan mereka dengan resistan terhadap mastitis atau rentan adalah kurang. Kajian ini dijalankan untuk menyelidik status mastitis udder, skor sel somatik susu (SCS), ciri puting, beberapa sifat komposisi susu dan SNPs dalam tiga gen calon untuk mastitis bovine pada lembu betina Friesian dan Jersey, supaya dapat mengenal pasti penanda genetic yang berpotensi yang dapat diguna dalam *marker assisted selection*.

Lembu Friesian dan Jersey yang berlaktasi di sebuah ladang tenusu komersil digunakan dalam kajian ini. Kiraan sel somatik susu (SCC) dianggarkan menggunakan pengira sel somatik dan ditukar kepada SCS, dan status mastitis ditentukan menggunakan ujian California mastitis (CMT). Panjang puting diukur, fungsinya diuji, dan pembukaan diperhatikan dan dicatat. Ciri komposisi susu ditentukan oleh spektroskopi inframerah. DNA diekstrak daripada susu 40 ekor lembu betina yang dijangkiti dan 37 ekor yang sihat, dan sampel DNA individu telah ditapis dan

digenotipkan untuk lima loki SNP dalam exon dan intron *TLR4*, *MBL1* dan *PGLYRP2* menggunakan teknik PCR-restriction fragment polymorphism (PCR-RFLP).

Tidak terdapat perbezaan signifikan ( $P < 0.05$ ) dalam jumlah puting yang dijangkiti (berdasarkan ujian mastitis California) dan berfungsi di antara lembu Friesian dan Jersey yang dipelihara di bawah pengurusan yang sama. Juga tidak terdapat perbezaan yang signifikan pada SCS antara baka atau antara laktasi. Walau bagaimanapun, susu peringkat laktasi lewat mempunyai SCS tertinggi ( $P < 0.05$ ). Sukuan udder, peringkat laktasi dan nombor laktasi mempunyai kesan yang signifikan ( $P < 0.05$ ) ke atas panjang puting. Min panjang puting bahagian belakang adalah lebih rendah secara signifikan ( $P < 0.05$ ) daripada sukuan udder depan. Perkaitan yang signifikan antara panjang puting dan status mastitis sukuan diperhatikan. Semakin panjang puting, semakin tinggi prevalensi mastitis. Skor hiperkeratosis hujung puting menunjukkan perbezaan yang signifikan antara baka dan kuar udang. Nombor laktasi, peringkat laktasi dan baka mempunyai kesan yang signifikan ( $P < 0.05$ ) terhadap laktosa susu, tetapi bukan terhadap lemak tidak padat. Nombor laktasi dan peringkat laktasi mempunyai kesan yang signifikan ( $P < 0.05$ ) terhadap lemak dan protein susu. Tidak terdapat perkaitan yang signifikan SCS, lemak, protein dan laktos susu dengan genotip di loki yang disiasat bagi gen *TLR4*, *MBL 1* dan *PGLYRP-2*, walaupun beberapa genotip telah mempunyai SCS yang rendah berbanding dengan yang lain. Terdapat perkaitan signifikan ( $P < 0.05$ ) antara keadaan puting dan status mastitis. Oleh itu, sifat morfologi udder perlu dimasukkan dalam pemilihan dan pembiakbakaan untuk resistan terhadap mastitis. Saiz gerompok yang lebih besar, penilaian lebih banyak loki SNP, dan analisis data dari kitaran laktasi yang lengkap setiap haiwan akan memberikan maklumat yang lebih sah mengenai kesan pelbagai faktor yang mempengaruhi dan penanda bermaklumat bagi SCS dan ciri komposisi susu untuk lembu tenusu Friesian dan Jersey di Malaysia.



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I certify that a Thesis Examination Committee has met on 5 December 2017 to conduct the final examination of Umar Mohammed Sani on his thesis entitled "Somatic Cell Count, Teat Characteristics, Milk Composition and Single Nucleotide Polymorphisms in Candidate Genes for Mastitis in Friesian and Jersey Cows" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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## LIST OF ABBREVIATIONS

CMT	California mastitis test
bp	base pair
DNA	deoxyribonucleic acid
<i>E. coli</i>	<i>Escherichia coli</i>
FAO	Food and Agricultural Organisation
DVS	Department of Veterinary Service
IDF	International Dairy Federation
IMI	Intramammary infection
kb	kilobase
MBL	mannose binding lectin
M	molar
NMC	National Mastitis Council
PGLYRP	peptidoglycan recognition proteins
PMN	polymorphonuclear neutrophils
SCC	somatic cell count
SCS	somatic cell score
TLR	toll like receptor
RE	restriction enzyme

# CHAPTER 1

## INTRODUCTION

The demand for milk and other dairy product is on the increase globally in which Malaysia is not an exception. Milk is a complex mixture secreted from the udder of mammals which comprises of lactose, proteins, fat, and vitamins among other components. The bulk of milk used worldwide is from cows although milk from buffalo, sheep, goat, as well as camel, are of commercial importance (FAO, 2007). Milk is an ideal food for the young mammals, human and animals (Vaclavik and Christian, 2014). Milk from normal healthy Holstein - Friesian dairy cows comprises 87% water, 3.8% fat, 3.4% protein, 4.5% lactose in addition to other components such as 1.3% minerals and a trace of epithelial cells (Ruegg, 2001).

Mastitis is defined as an inflammatory response of the mammary gland (Internal Dairy Federation, 1987). Contagious and environmental pathogens usually cause it. The pathogen, usually bacteria, enters the mammary gland through the teat canal. When they succeed in overcoming the innate defense mechanism of the animal, they localized in the udder and multiply. Here, the microbes produce toxins that destroy cell membranes and damage milk-producing tissues (Hillerton & Berry, 2005; Zhao & Lacasse, 2008). Damage to the mammary gland epithelial tissues is triggered not only by the bacteria but also by the host immune action. When this happens, the alveoli will become fragile, with resultant permeability to other extracellular fluid which mix with the milk and result in visible change in the milk and the udder (Zhao & Lacasse, 2008). Mastitis is classified as clinical mastitis, the form associated with inflammation of the mammary gland (visible swelling and reddening), wateriness of the milk, sore teat, presence of pus, flakes, and blood in the milk, and higher somatic cell count (SCC) (Zhao & Lacasse, 2008; Fedota *et al.*, 2015 ), or as sub-clinical mastitis, where the infected animals show no physical signs of the disease, the milk appear normal but milk yield is reduced and SCC elevated (DeGraves & Fetrow, 1993; Shitandi & Kihumbu, 2004).

Milk somatic cells are milk epithelial cells, usually shed from the inner lining of the mammary glands, and leukocytes which originated from the blood cells. Somatic cells composed of 75% leucocytes (which comprises neutrophils, macrophages, lymphocytes, erythrocytes) and 25% epithelial cells. (Sharma *et al.* 2011; Zhao & Lacasse, 2008). When there is a presence of infectious bacteria, injury in the mammary gland tissues and stress, there will be influx somatic cells notably polymorphonuclear neutrophils (PMN) to neutralize the effect of the invading pathogen. Therefore, SCC is an important indicator of udder health and mastitis (Pantoja *et al.*, 2009).

Selection for mastitis resistance/resilience or susceptibility mainly focuses on SCC or somatic cell score (SCS), the transformed values of SCC, as an indicator of udder health and mastitis. Accuracy of this selection may be improved if optimum udder

health index, udder and teat morphometric are added to SCC in breeding programs (Singh & Bansal, 2013; Dube *et al.*, 2008). However, mastitis is a polygenic trait. The greatest challenge faced by molecular geneticists is the identification of markers for genes that control the phenotypic variation in mastitis resistance/ resilience (Sender *et al.*, 2013; Ogorevc *et al.*, 2009).

A number of candidate genes for mastitis resistance have been reported, which have the potential of reducing mastitis susceptibility in dairy herds (Turner, 2003; Garred *et al.*, 2006; Tydell *et al.*, 2006; Dziarski & Gupta, 2010; Kumar & Yadav, 2012; Gupta *et al.*, 2015;). However, application of marker assisted selection strategy to control mastitis requires understanding of the genetic variations in the local herds, and identifying informative single nucleotide polymorphisms (SNPs) and genotypes associated with mastitis.

### **1.1 Research Problem**

There are limited information on the SNPs related to mastitis resistance or susceptibility in the Malaysian dairy cattle herds. In addition there is a lack of reports on udder morphological characteristics in local dairy cattle populations. There are also no published works investigating the association of SNPs in the candidate genes with SCS, udder or teat characteristics or milk composition traits.

### **1.2 Objectives**

The general objective of the study was to evaluate the udder mastitis status, milk SCS, teat characteristics, some milk composition traits and SNPs in candidate genes for bovine mastitis resistance in Friesian and Jersey cows in one dairy cattle farm in Malaysia, in order to identify potential genetic markers that may be used in marker assisted selection.

The specific objectives were:

1. To estimate the milk SCC and SCS of Friesian and Jersey cows at different lactations and lactation stages in a dairy cattle herd.
2. To evaluate the functioning of udders, teat characteristics and milk composition traits in the above dairy cows.
3. To identify single nucleotide polymorphisms in three candidate genes for bovine mastitis.
4. To determine the association of SCC, SCS, udder morphological characteristics and milk composition traits with the SNPs and genotypes at the three candidate genes for mastitis resistance in the Friesian and Jersey cows.



## REFERENCES

- Abiola, O. Joe M. Angel, Philip A., Alexander A. Bachmanov, John K. Belknap..... (2003). The nature and identification of quantitative trait loci: a community's view. *Nature Reviews Genetic* 4, 911–916.
- AHDB Dairy (2016). Technical note of the United Kingdom Agriculture and Horticulture Development Board (AHDB) Available online <https://dairy.ahdb.org.uk/about-ahdb-dairy/what-is-ahdb-dairy>
- AkhandPratap, D. K. V. P. K. and A. S. (2014). Effect of pregnancy, lactation stage, parity and age on yield and components of raw milk in Holstein Friesian cows in organized Dairy form in Allahabad/n. *IOSR Journal of Agriculture and Veterinary Science*, 7(2), 112–115.
- Alhussien, M., Kaur M., Manjari P., Kimothi S.P., Mohanty A.K., & Dang A.K. (2015) A comparative study on the blood and milk cell counts of healthy, subclinical and clinical mastitis Karan Fries cows, *Veterinary World*,8(5), 685-689.
- Ali, A.K.A., Shook GE (1980): An optimum transformation for somatic cell concentration in milk. *Journal of Dairy Science*, 63,487-490
- Armant, M. A., & Fenton, M. J. (2002). Toll-like receptors: a family of pattern-recognition receptors in mammals. *Genome Biology*, 3(8), 3011.1-3011.6.
- Arora, R. and Bhojak, N. (2013). Physiochemical and environmental factors responsible for change in milk composition of milking Animal. *International Journal of Engineering and Science*, 2(1), 275-277
- Ashwell, M.S., Heyen, D.W., Da, Y., Sonstegard, T.S., Tassel, C.P. & Lewin, H.A., (2002). Detection of QTL affecting milk production in 6 Dairy Bull DANN Repository grandsire families. Proceeding of the XXVIII International Conference on Animal Genetics, August 11-15, 2002. Göttingen, Germany.
- Langoni, H., Penachio DD, Nóbrega DB, de Freitas F.G., & Simone Lucheis S.B, ( 2012). Somaticell<sup>®</sup> as a screening method for somatic cell count from bovine milk. *Ciência Rural*, 42, 1095–1101.
- Bannerman, D.D., Paape M.J., Goff J.P., Kumari K., Lippolis J.D. & Hope J.C. (2004) Innate immune response to intramammary infection with *Serratia marcescens* and *Streptococcus uberis*. *Journal of Veterinary Resource*, 35(6), 681- 700
- Bannerman, D. D., Kauf, a C. W., Paape, M. J., Springer, H. R., & Goff, J. P. (2008). Comparison of Holstein and Jersey innate immune responses to *Escherichia coli* intramammary infection. *Journal of Dairy Science*, 91(6), 2225–2235.



- Barnouin, J., Bord, S., Bazin, S., & Chassagne, M. (2005). Dairy Management Practices Associated with Incidence Rate of Clinical Mastitis in Low Somatic Cell Score Herds in France. *Journal of Dairy Science*, 88(10), 3700–3709
- Barnouin, J., Bord, S., Bazin, S., & Chassagne, M. (2005). Dairy management practices associated with incidence rate of clinical mastitis in low somatic cell score herds in France. *Journal of Dairy Science*, 88(10), 3700–3709.
- Beecher, C., Daly, M., Childs, S., Berry, D. P., Magee, D. A., McCarthy, T. V., & Giblin, L. (2010). Polymorphisms in bovine immune genes and their associations with somatic cell count and milk production in dairy cattle. *BMC Genetics*, 11, 99.
- Berry, D. P., J. M. Lee, K. A. Macdonald, K. Stafford, L. Matthews, and J. R. Roche. 2007. Associations among body condition score, body weight, somatic cell count, and clinical mastitis in seasonally calving dairy cattle. *Journal of Dairy Science*, 90, 637–648.
- Besier, J., Lind, O., & Bruckmaier, R. M. (2015). Dynamics of teat-end vacuum during machine milking: types, causes and impacts on teat condition and udder health – a literature review. *Journal of Applied Animal Research*, 2119, 1–10.
- Bharti, P., Bhakat, C., Pankaj, P. K., Bhat, S. A., Prakash, M. A., & Thul, M. R. (2015). Relationship of udder and teat conformation with intra-mammary infection in crossbred cows under hot-humid climate. *Veterinary World*, 8(7), 898-901
- Bhoite UY and Padekar RN (2002). Factors affecting milk yield and composition. *Indian Journal of Animal Research*, 36, 67-69
- Bhutto, A. L., Murray, R. D., & Woldehiwet, Z. (2010). Udder shape and teat-end lesions as potential risk factors for high somatic cell counts and intra-mammary infections in dairy cows. *The Veterinary Journal*, 183(1), 63–67.
- Bhutto, A. L., Murray, R. D., & Woldehiwet, Z. (2012). California mastitis test scores as indicators of subclinical intra-mammary infections at the end of lactation in dairy cows. *Research in Veterinary Science*, 92(1), 13–7.
- Boneca, I. G. (2009). Mammalian PGRPs in the spotlight. *Cell Host & Microbe*, 5(2), 109–111.
- Boniface, B. and Gyau, A. and Stringer, R. (2009): Relationship quality as the predictor of long term relationship in the Malaysian dairy industry. Proceedings of the 4th International Conference of the Asian Academy of Applied Business (AAAB 2009) (17. December 2009): pp. 766-776
- Boniface, B., Silip J. J. and Ahmad, A. (2007): Dairy cattle management: survey on dairy cattle lactation trend in Sabah. Munich Personal RePEc Archive Paper No. 23781, <https://mpra.ub.uni-muenchen.de/id/eprint/23781>

- Bradley, A. J. (2002). Bovine mastitis: An evolving disease. *The Veterinary Journal*, 164(2), 116–128.
- Bretschneider, G., Arias, D., & Cuatrin, A. (2015). Comparative evaluation of udder and body conformation traits of first lactation 3/4 Holstein x 1/4 Jersey versus Holstein cows. *Archivos de Medicina Veterinaria*, 47(1), 85–89.
- Buckley, F. (2012). Evaluation of Jersey, Jersey × Holstein- Friesian and Holstein-Friesian cows under Irish grass-based spring milk production systems, <http://www.teagasc.ie/publications/> 1–4. [Accessed date, 26/09/2016]
- Buitkamp, J., & Götz, K.-U. (2004). Use of milk samples from a milk evaluation program for the genotyping of cows. *Archiv Tierzucht Dummerstorf*, 47(1), 15–26.
- Chesnokov Yu.V., and Artemyeva A.M. (2015). Evaluation of the measure of polymorphism information of genetic diversity. *Agricultural Biology*, 5 (50), 571-578
- Cargill, E. J. and J. E. Womack (2007). Detection of polymorphisms in bovine toll-like receptors 3, 7, 8, and 9. *Genomics*, 89(6), 745-755.
- Carolina, B. M., dos Reis, Barreiro J.R., Mestieri L., de Felício Porcionato M.A., & dos Santos M.V. (2013). Effect of somatic cell count and mastitis pathogens on milk composition in Gyr cows. *BMC Veterinary Research*, 9, 67.
- Carlen, E., Strandberg, E. & Roth, A., (2004). Genetic parameters for clinical mastitis, somatic cell score and production in the first three lactations of Swedish Holstein cows. *Journal of Dairy Science*, 87, 3062–3070.
- Carvajal, M., Huircan, P., & Lepori, a. (2013). Single nucleotide polymorphisms in immunity-related genes and their association with mastitis in Chilean dairy cattle. *Genetics and Molecular Research*, 12(3), 2702–2711.
- Castillo-Juarez, H., Oltenacu, P. A., & Cienfuegos-Rivas, E. G. (2002). Genetic and phenotypic relationships among milk production and composition traits in primiparous Holstein cows in two different herd environments. *Livestock Production Science*, 78(3), 223–231.
- Cengiz, M., Kaynar, O., Cannazik, O., Ileriturk, M., Cengiz, S., & Hayirli, A. (2015). Sampling factors causing variability in milk constituents in early lactation cows. *Veterinari Medicina*, 60(1), 6–15.
- Cerón-Muñoz, M., Tonhati, H., Duarte, J., Oliveira, J., Muñoz-Berrocal, M., & Jurado-Gómez, H. (2002). Factors affecting somatic cell counts and their relations with milk and milk constituent yield in Buffaloes. *Journal of Dairy Science*, 85(11), 2885–2889.

- Chrystal, M.A., Seykora A.J & Hansen L.B. (1999). Heritabilities of teat end shape and teat diameter and their relationships with somatic cell score. *Journal of Dairy Science*, 82, 2017–2022.
- Chachare, R.A. & Walkunde, T.R. (2011). Effect of stage of lactation on pre and post milking udder, teat and milk vein characteristics in gaolao cattle, *Research. Journal of Animal Husbandary and Dairy Science*, 2 (1&2), 27-29.
- Chaiyotwittayakun, A., Aiumlamai, S., Chanlun, A., & Srisupa, S. (2008). Alternative method for determination of milk somatic cell count in dairy cows, Proceedings of the 15th Congress of the Federation of Asian Veterinary Associations - World Organisation for Animal Health Joint Symposium on Emerging Diseases, 27-30 October, 2008, Bangkok, Thailand.
- Chrystal, M.A., Seykora, A.J., Hansen, L.B., Freeman, A.E., Kelley, D.H., & Healey, M.H., 2001. Heritability of teat-end shape and the relationship of teat-end shape with somatic cell score for an experimental herd of cows. *Journal of Dairy Science*, 84, 2549–2554.
- Chye, F. Y., Abdullah, A., & Ayob, M. K. (2004). Bacteriological quality and safety of raw milk in Malaysia. *Food Microbiology*, 21(5), 535–541.
- Cinar, M., Serbester, U., Ceyhan, A., & Gorgulu, M. (2015). Effect of somatic cell count on milk yield and composition of first and second lactation dairy Cows. *Italian Journal of Animal Science*, 14(1), 3646.
- Czerniewicz, M., Kielczewska, K., & Kruk, A. (2006). Comparison of some physicochemical properties of milk from Holstein-Friesian and Jersey cows. *Polish Journal of Food and Nutrition Science*, 15(1), 61–64.
- Czerniewicz, M., Kielczewska, K., & Kruk, A. (2006). Comparison of some physicochemical properties of milk from Holstein-Friesian and Jersey cows. *Polish Journal of Food and Nutrition Science*, 15(1), 61–64.
- d'Angelo, F., Santillo, A., Sevi, A., & Albenzio, M. (2007). Technical note: A simple salting-out method for DNA extraction from milk somatic cells: investigation into the goat CSN1S1 gene. *Journal of Dairy Science*, 90(7), 3550–2.
- de Pinho Manzi, M., Nóbrega, D. B., Faccioli, P. Y., Troncarelli, M. Z., Menozzi, B. D., & Langoni, H. (2012). Relationship between teat-end condition, udder cleanliness and bovine subclinical mastitis. *Research in Veterinary Science*, 93(1), 430–434.
- DeGraves, F. J., & Fetrow, J. (1993). Economics of mastitis and mastitis control. veterinary clinics of North America: *Food Animal Practice*, 9(3), 421–434.
- Dhakal, I. P. (2006). Normal somatic cell count and subclinical mastitis in Murrah buffaloes. *Journal of Veterinary Medicine Series B: Infectious Diseases and Veterinary Public Health*, 53(2), 81–86.

- Dohoo, I. R., & Meek, A. H. (1982). Somatic cell counts in bovine milk. *Canadian Veterinary Journal* 23, 119–125.
- Dommett, R. M., Klein, N., & Turner, M. W. (2006). Mannose-binding lectin in innate immunity: past, present and future. *Tissue Antigens*, 68(3), 193–209.
- Dong, F. (2006). The outlook for Asian dairy markets: The role of demographics, income, and prices. *Food Policy*, 31(3), 260–271.
- Dube, B., Dzama, K., & Banga, C. B. (2008). Genetic analysis of somatic cell score and udder type traits in South African Holstein cows. *South African Journal of Animal Sciences*, 38(1), 1–11.
- Dziarski, R., & Gupta, D. (2010). Review: Mammalian peptidoglycan recognition proteins (PGRPs) in innate immunity. *Innate Immunity*, 16(3), 168–174.
- Erdem, H., Atasever S., & Kul E. (2007) Some Environmental Factors Affecting Somatic Cell Count of Holstein Cows, *J.Appl. Anim. Res.* 32:(2), 173-176
- Erdem, H., S. Atasever & E. Kul, (2010). A study on somatic cell count of Jersey cows. *Asian Journal of Animal and Veterinary Advances*, 5, 253-259.
- FAO, (2010) Global dairy sector: Status and trends. In: Hemme, T. & J. Otte (eds), Status and Prospects for Smallholder Milk Production - A Global Perspective [Online], Food and Agriculture Organization of the United Nations, [www.fao.org/docrep/012/i1522e/i1522e.pdf](http://www.fao.org/docrep/012/i1522e/i1522e.pdf).
- FAO, (2007) State of the World's Farm Animal Genetic Resources [Online], Food and Agriculture Organization of the United Nations, [www.fao.org/Ag/AGAInfo/programmemes/en/genetics/documents/AH473e01.pdf](http://www.fao.org/Ag/AGAInfo/programmemes/en/genetics/documents/AH473e01.pdf).
- FAO, (2016). Food Outlook Trade and Markets Division of FAO under Global Information and Early Warning System (GIEWS), Food and Agriculture Organization of the United Nations, <http://www.fao.org/worldfoodsituation>.
- Farrell, H. M., Jimenez-Flores, R., Bleck, G. T., Brown, E. M., Butler, J. E., Creamer, L. K., ... Swaisgood, H. E. (2004). Nomenclature of the proteins of cows' milk - sixth revision. *Journal of Dairy Science*, 87(6), 1641–74.
- Fedota, O. M., Yu, R. S., Bolotin, V. I., Klochko, I. O., & Kharkov, V. N. K. (2015). Genetics of resistance to clinical mastitis in cows - A review. *Journal for Veterinary Medicine, Biotechnology and Biosafety*, 1(4), 2411-0388
- Ferris, C. P., Heins, B. J., & Buckley, F. (2014). Crossbreeding in dairy cattle : Pros and Cons. *Advances in Dairy Technology*, 26, 223–243.
- Fox, P. F., & Mcsweeney, P. L. H. (1998). Dairy Chemistry and Biochemistry. London, Blackie Academic & Professional, 478 pp.



- Garred, P., Larsen, F., Seyfarth, J., Fujita, R., & Madsen, H. O. (2006). Mannose-binding lectin and its genetic variants. *Genes and Immunity*, 7(2), 85–94.
- Gellrich, K., Meyer, H. H. D., & Wiedemann, S. (2014). Composition of major proteins in cow milk differing in mean protein concentration during the first 155 days of lactation and the influence of season as well as shortterm restricted feeding in early and mid-lactation. *Czech Journal of Animal Science*, 59(3), 97–106.
- Gellrich, K., Meyer, H. H. D., & Wiedemann, S. (2014). Composition of major proteins in cow milk differing in mean protein concentration during the first 155 days of lactation and the influence of season as well as shortterm restricted feeding in early and mid-lactation. *Czech Journal of Animal Science*, 59(3), 97–106.
- Georgi Radoslavov, P. H. (2015). A review of methods for genotyping milk proteins in cattle. *Advances in Dairy Research*, 3, 144, 2329-888.
- Gleeson, D.E., Meaney, W.J., O’Callaghan, E.J., & Rath, M.V., (2004). Effect of teat hyperkeratosis on somatic cell counts of dairy cows. *International Journal of Applied Research in Veterinary Medicine*, 2, 115–122.
- Gomberg, M., Paneth, F., Hafeditz, W., Mayo, F. R., Lewis, F. M., Menapace, L. W., ... Blaney, B. (2001). Relationship between teat-end callosity or hyperkeratosis and mastitis. Proceedings of the 34<sup>th</sup> Annual Convention of the American Association of Bovine Practitioners, Vancouver, British Columbia, Canada, 13 - 15 September, 2001.
- Gruet, P., Maincent, P., Berthelot, X., & Kaltsatos, V. (2001). Bovine mastitis and intramammary drug delivery: review and perspectives. *Advanced Drug Delivery Reviews*, 50(3), 245–259.
- Gupta, P. H., Patel, N. A., Rank, D. N., Joshi, C. G., & Husbandry, A., (2015). Genetic polymorphism of toll-like receptors 4 gene by polymerase chain reaction-restriction fragment length polymorphisms , polymerase chain reaction-single-strand conformational polymorphism to correlate with mastitic cows, *Veterinary World* 8, 6–11.
- Gurmessa, J., & Melaku, A. (2012). Effect of lactation stage , pregnancy , parity and age on yield and major components of raw milk in bred cross Holstein Friesian cows. *World Journal of Dairy & Food Sciences*, 7(2), 146–149.
- Haghkhah, M., Ahmadi, M. R., Gheisari, H. R., & Kadivar, A. (2011). Preliminary bacterial study on subclinical mastitis and teat. *Turkish Journal of Veterinary and Animal Science*. 35(6), 387-394
- Haile-Mariam, M., Goddard, M. E., & Bowman, P. J. (2001). Estimates of genetic parameters for daily somatic cell count of Australian Dairy Cattle. *Journal of Dairy Science*, 84(5), 1255–1264.

- Halasa, T., Huijps, K., Østerås, O., & Hogeveen, H. (2007). Economic effects of bovine mastitis and mastitis management: A review. *Veterinary Quarterly*, 29(1), 18–31.
- Higgins, S., Moore, R. K., & Kennedy, B. W. (1980). Heritabilities of teat conformation traits and their relationships with somatic cell counts in Holsteins. *Canadian Journal of Animal Science*, 60, 231-239.
- Hillerton, J. E., & Berry, E. A. (2005). Treating mastitis in the cow - A tradition or an archaism. *Journal of Applied Microbiology*, 98(6), 1250–1255.
- Hopkins, P., & Sriskandan, S. (2005). Mammalian Toll-like receptors: to immunity and beyond. *Clinical and experimental immunology*, 140(3), 395-407.
- Hortet, P., & Seegers, H. (1998). Review article Calculated milk production losses associated with elevated somatic cell counts in dairy cows : review and critical discussion. *Veterinary Research*, 29, 497–510.
- Hristov, P., & Radoslavov G (2015) A review of methods for genotyping milk proteins in cattle. *Journal of Advance Dairy Research*, 3, 144, 2329-888.
- IDF, (2016). The World Dairy Situation 2016. Bulletin of the International Dairy Federation 485/2016. Retrieved from [www.fil-idfa.org](http://www.fil-idfa.org).
- IDF, (1987) Bovine Mastitis: Definition and guidelines for diagnosis. International Dairy Federation, Brussels, 24 pp.
- Jánosi, S., & Baltay, Z. (2004). Correlations among the somatic cell count of individual bulk milk, result of the California Mastitis Test and bacteriological status of the udder in dairy cows. *Acta Veterinaria Hungarica*, 52(2), 173–83.
- Karp A., Kresovich S., Bhat K.V., Ayada W.G., & Hodgkin T. (1998). Molecular tools in plant genetic resources conservation: a guide to the technologies. International Plant Genetic Resources Institute (IPGRI) Technical Bulletin No. 2.
- Kashyap, D. R., Wang, M., Liu, L.-H., Boons, G.-J., Gupta, D., & Dziarski, R. (2011). Peptidoglycan recognition proteins kill bacteria by activating protein-sensing two-component systems. *Nature Medicine*, 17(6), 676–83.
- Kayastha, R.B., Zaman G. & Goswami, R.N. (2008) Factors affecting the milk constituents of native cattle of Assam. *Indian Journal of Animal Resource*, 42 (4), 270-272.
- KerroDego, O., van Dijk, J.E., & Nederbragt, H. (2002). Factors involved in the early pathogenesis of bovine *Staphylococcus aureus* mastitis with emphasis on bacterial adhesion and invasion: A review. *Veterinary Quarterly*, 24 (4): 181-198.

- Khatkar, M. S., Thomson, P. C., Tammen, I., & Raadsma, H. W. (2004). Quantitative trait loci mapping in dairy cattle: review and meta-analysis. *Genetics, Selection, Evolution*, 36(2), 163–190.
- Kittivachra, R., Sanguandeeikul, R., Sakulbumrungsil, R., Phongphanphanee, P., & Srisomboon, J. (2006). Determination of essential nutrients in raw milk. *Songklanakarin Journal of Science and Technology*, 28(suppl. 1), 115–120.
- Korhonen, H. & L. Kaartinen, (1995). Changes in the composition of milk induced by mastitis. In: Jyva. (ed.), *The bovine udder and mastitis*, Gum. Finland, pp: 76-82
- Kuczyńska, B., Puppel, K., Golebiewski, M., Kordyasz, M., Grodzki, H., & Brzozowski, P. (2012). Comparison of fat and protein fractions of milk constituents in Montbeliarde and Polish Holstein-Friesian cows from one farm in Poland. *Acta Veterinaria Brno*, 81(2), 139–144.
- Kumar, R., & Yadav, B. R. (2012). Molecular analysis of TLR4 gene and its association with intra-mammary infections in Sahiwal cattle and Murrah buffaloes. *Indian Journal of Biotechnology*, 11, 267-273.
- Kumaresan, G. (2013). Somatic cell pattern and composition of milk of Holstein Friesian cross bred Cattle, *International Journal of Science, Environment and Technology*, 2 (6), 1421–1425.
- Lacy-hulbert, B. Y. S. J., & Hillerton, J. E. (1995). Physical characteristics of the bovine teat canal and their influence on susceptibility to streptococcal infection, *Journal of Dairy Research*, 62, 395-404.
- Langoni, H, Penachio D.D, Nóbrega DB, de Freitas F.G. & Lucheis S.B.(2012). Somaticell<sup>®</sup> as a screening method for somatic cell count from bovine milk. *Ciência Rural*, 4,1095–1101.
- Lakic, B., Wredle, E., Svennersten-Sjaunja, K., & Östensson, K. (2009). Is there a special mechanism behind the changes in somatic cell and polymorphonuclear leukocyte counts, and composition of milk after a single prolonged milking interval in cows? *Acta Veterinaria Scandinavica*, 51(1), 1751-0147.
- Lal, D. & Narayanan, K.M. (1984) Effect of lactation number of the animal on the fat and solids-not-fat contents of milk. *Indian Journal of Animal Science*, 54 (9), 835-839.
- LeBlanc, S. J., Lissemore, K. D., Kelton, D. F., Duffield, T. F., & Leslie, K. E. (2006). Major advances in disease prevention in dairy cattle. *Journal of Dairy Science*, 89(4), 1267–79.
- Levene, H (1949) On a matching problem arising in genetics. *The Annals of Mathematical Statistics*, 20 (1), 91-94.



- Lindmark Månsson, H. (2008). Fatty acids in bovine milk fat. *Food & Nutrition Research*, DOI: 10.3402/fnr.v52i0.1821.
- Liu, M., Ju Z., Li Q., Wang H., Huang J., Li J., Zhong J.& Wang C. (2010) Polymorphisms of MBL1 gene intron 1 and exon 2 and its association with mastitis and milk quality in Chinese Holstein cattle [C] [abstract]. *Scientia Agricultura Sinica*, 43(11), 2363–2371.
- Liu, J., Ju, Z., Li, Q., Huang, J., Li, R., Li, J., ... Wang, C. (2011). Mannose-binding lectin 1 haplotypes influence serum MBL-A concentration, complement activity, and milk production traits in Chinese Holstein cattle. *Immunogenetics*, 63(11), 727–742.
- Li Yang, F., & Shan Li, X. (2013). Somatic Cell Counts Positive Effects on the DNA yield extracted directly from Murrah Buffalo milk. *Journal of Current Research in Science*, 1(5), 392–395.
- Lucy, M. C. (2001). Reproductive loss in high-producing dairy cattle: where will it end? *Journal of Dairy Science*, 84(6), 1277–93.
- Mackle, T. R., Bryant, A. M., Petch, S. F., Hooper, R. J., & Auldist, M. J. (1999). Variation in the composition of milk protein from pasture-fed dairy cows in late lactation and the effect of grain and silage supplementation. *New Zealand Journal of Agricultural Research*, 42(2), 147–154.
- Malek dos Reis, C. B., Barreiro, J. R., Mestieri, L., Porcionato, M. A. de F., & dos Santos, M. V. (2013). Effect of somatic cell count and mastitis pathogens on milk composition in Gyr cows. *BMC Veterinary Research*, 9(1), 1746-6148
- Martínez, M. E., Calderón, C., De la Barra, R., De La Fuente, F., & Gonzalo, C. (2011). Udder morphological traits and milk yield of Chilota and suffolk down sheep breeds. *Chilean Journal of Agricultural Research*, 71(March), 90–95.
- Mather, I. H. (2000). Nomenclature for Major Proteins of the Milk-Fat Globule Membrane A Review. *Journal of Dairy Science*, 83(2), 203–247.
- Mehrzaad, J., Duchateau L., Pyorala S., & Burvenich C. (2002). Blood and milk neutrophil chemiluminescence and viability in primiparous and pluriparous dairy cows during late pregnancy, around parturition and early lactation. *Journal of Dairy Science*, 85:3268–3276.
- Meilina, H., Kuroki, S., Jinendra, B. M., Ikuta, K., & Tsenkova, R. (2009). Double threshold method for mastitis diagnosis based on NIR spectra of raw milk and chemometrics. *Biosystems Engineering*, 104(2), 243–249.
- Mein, G.A., Neijenhuis F, Morgan WF, Reinemann D.J., Hillerton J.E..... (2001): Evaluation of bovine teat condition in commercial dairy herd: Non-infectious factors. Proceedings of the 2<sup>nd</sup> International Symposium on Mastitis and Milk Quality, pp. 347–356.

- Mein, G.A, Reinemann D, Schuring N,& Ohnstad I. (2004) Milking machines and mastitis risk: a storm in a teacup. Proceedings of the 43rd Annual Meeting of the National Mastitis Council, Charlotte, Carolina, US, pp. 176-188.
- Menzies, M., & Ingham, A. (2006). Identification and expression of Toll-like receptors 1-10 in selected bovine and ovine tissues. *Veterinary Immunology and Immunopathology*, 109(1–2), 23–30.
- Mesquita, A.Q., Rezende, C.S.M., Mesquita, A.J., Garcia, E.A., Jardim, D.V & Kipnis, A.P.J. (2012) Association of TLR4 Polymorphisms with subclinical mastitis in Brazilian Holsteins. *Brazian Journal Microbiology*, 43(2), 692-697.
- Modh, R. H., Islam, M. M., Patel, Y. G., Modi, R. J., & Wadhvani, K. N. (2017). Effect of parity on udder and teat biometry and its association with milk yield in Gir cows, *International Journal of Science, Environment and Technology*, 6(3), 2068–2073.
- Mollenhorst, H., van der Tol, P. P. J., & Hogeveen, H. (2010). Somatic cell count assessment at the quarter or cow milking level. *Journal of Dairy Science*, 93(7), 3358–64.
- Mononecor, I., Wrlron, J. W., Arren, O. & Beno Sonc, H. (1983). Stage of lactation effects, repeatabilities and influences on weaning weights of yield and composition of milk in beef cattle. *Canadian Journal of Animal Science*, 63, 751-761.
- Moon, J. S., Koo, H. C., Joo, Y. S., Jeon, S. H., Hur, D. S., Chung, C. I., ... Park, Y. H. (2007). Application of a new portable microscopic somatic cell counter with disposable plastic chip for milk analysis. *Journal of Dairy Science*, 90(5), 2253-2259
- Muhammad, & S.G. (2008). Somatic cell count as an indicator of udder health status under modern dairy production: a review. *Pakistan Veterinary journal*, 28(4), 194–200.
- Muhasin Asaf, V. N., Kumar, A., Rahim, A., Sebastian, R., Mohan, V., Dewangan, P., & Panigrahi, M. (2014). An overview on single nucleotide polymorphism studies in mastitis research. *Veterinary World*, 7(6), 416–421.
- Murphy, M. A., Shariou, M. R. & Moran, C. (1995) DNA from milk. Proceedings of the Australian Association of Animal Breeding and Genetics 11, 696.
- Murphy, M. A., Shariflou, M. R., & Moran, C. (2006). High quality genomic DNA extraction from large milk samples. *Journal of Dairy Research*, (2002), 645–649.
- Nobrega, D. B., & Langoni, H. (2011). Breed and season influence on milk quality parameters and in mastitis occurrence. *Pesquisa Veterinaria Brasileira*, 31(12), 1045–1052.

- Nakov, D., Hristov, S., Andonov, S., & Trajchev, M. (2014). Udder-related risk factors for clinical mastitis in dairy cows. *Veterinarski Arhiv*, 84(2), 111–127.
- Neijenhuis, F., Klungel, G. H., & Hogeveen, H. (2001b). Recovery of cow teats after milking as determined by ultrasonographic scanning. *Journal of Dairy Science*, 84(12), 2599-2606.
- Neijenhuis, F., Mein G.A., Britt J.S., Reinemann D.J., Hillerton J.E.,... (2001a) Relationship Between Teat-End Callosity or Hyperkeratosis and Mastitis. Proceedings of the AABP-NMC International Symposium on Mastitis and Milk Quality, Vancouver, BC, Canada. 13-15, September 2001
- Nobrega, D. B., & Langoni, H. (2011). Breed and season influence on milk quality parameters and in mastitis occurrence. *Pesquisa Veterinaria Brasileira*, 31(12), 1045–1052.
- Nyamushamba, G. B., Halimani, T. E., Imbayarwo-Chikosi, V. E., & Tavirimirwa, B. (2014). Comparative evaluation of non-genetic factors affecting milk yield and composition of Red Dane and Jersey cattle in Zimbabwe. *SpringerPlus*, 3(1), 88.
- Ogorevc, J., Kunej, T., Razpet, A., & Dovc, P. (2009). Database of cattle candidate genes and genetic markers for milk production and mastitis. *Animal Genetics*, 40(6), 832–851.
- Ohnstad, I. (2012). Teat condition scoring as a management tool. *Livestock, National Animal disease information service (NADIS) UK*, 17(7), 34–40. 23 Retrieved 26th March 2017, <http://doi.org/10.1111/j.2044-3870.2012.00165.x>
- Osman A. (1984). Dairy production. In: Vijchulata, P., Bongsu T.A. & Dahan M.M. (eds.). *Animal Industry in Malaysia*. Faculty of Veterinary Medicine & Animal Sciences, Universiti Pertanian Malaysia, Serdang. pp. 41-49.
- Oviedo-Boyso, J., Valdez-Alarcón, J. J., Cajero-Juárez, M., Ochoa-Zarzosa, A., López-Meza, J. E., Bravo-Patiño, A., & Baizabal-Aguirre, V. M. (2007). Innate immune response of bovine mammary gland to pathogenic bacteria responsible for mastitis. *The Journal of Infection*, 54(4), 399–409.
- Ozrenk, E., & Inci, S. S. (2008). The effect of seasonal variation on the composition of cow milk in Van Province. *Pakistan Journal of Nutrition*, 7(1), 161–164.
- Paape, M. J., Wiggans, G. R., Bannerman, D. D., Thomas, D. L., Sanders, A. H., Contreras, A., ... Miller, R. H. (2007). Monitoring goat and sheep milk somatic cell counts. *Small Ruminant Research*, 68(1–2), 114–125.
- Palladino, R. A., Buckley, F., Prendiville, R., Murphy, J. J., Callan, J., & Kenny, D. A. (2018). A comparison between Holstein-Friesian and Jersey dairy cows and their F1 hybrid on milk fatty acid composition under grazing conditions. *Journal of Dairy Science*, 93(5), 2176–2184.

- Panandam, J.M. and A.K. Raymond (2010). Development of the Mafriwal dairy Cattle of Malaysia. Available online: <http://agtr.ilri.cgiar.org/casestudy/jothi/Jothi.htm>.
- Pantoja, J. C. F., Hulland, C., & Ruegg, P. L. (2009). Dynamics of somatic cell counts and intramammary infections across the dry period. *Preventive Veterinary Medicine*, 90(1–2), 43–54.
- Patel, Y. G., Trivedi, M. M., Rajpura, R. M., Savaliya, F. P., & Parmar, M.(2016) Udder And Teat Measurements And Their Relation With Milk Production In Crossbred Cows. *International Journal of Science, Environment and Technology*, 5 (5), 3048 – 3054
- Pavol, M., Makovický, P., Nagy, M., Rimárová, K., & Diabelková, J. (2014). Genetic parameters for somatic cell count, LOGSCC and somatic cell score among breeds: Improved Valachian, Tsigai, Lacaune and their crosses. *Acta Veterinaria-Beograd*, 64, 386–396.
- Peeler, E. J., Green M.J., Fitzpatrick J.L., Morgan K.L & Green L.E. (2000): Risk factors associated with clinical mastitis in low somatic cell count British dairy herds. *Journal of Dairy Science*. 83, 2464-2472
- Petlane, M., Noor, R. R., & Maheswari, R. R. A. (2013). Relationship between somatic cell counts, mastitis and milk quality in Ettawah grade and PESA goats. *Walailak Journal of Science and Technology*, 10(6), 607–613.
- Phatsara, C., Jennen, D. G. J., Ponsuksili, S., Murani, E., Tesfaye, D., Schellander, K., & Wimmers, K. (2007). Molecular genetic analysis of porcine mannose-binding lectin genes, MBL1 and MBL2, and their association with complement activity. *International Journal of Immunogenetics*, 34(1), 55–63.
- Pintana, P. (2010). Division of Livestock sciences genetic analysis of milk fatty acid compositions based on infrared data; \* Animal Breeding and Genomics Centre, Wageningen University, PO Box 338, 6700AH Wageningen, the Netherlands.
- Pisestyani, H., & Septiani, Y. N. (2016). Teat length and lactation period as a predisposition factor of subclinical mastitis in dairy cattle in Bandung, Indonesia, *Journal of Life Sciences* 10, 1–6.
- Pokorska, J., Kułaj, D., Dusza, M., Żychlińska-Buczek, J., & Makulska, J. (2016). New rapid method of DNA isolation from milk somatic cells. *Animal Biotechnology*, 27(2), 113–117.
- Prendiville, R., Pierce, K. M., & Buckley, F. (2009). An evaluation of production efficiencies among lactating Holstein-Friesian, Jersey, and Jersey×Holstein-Friesian cows at pasture. *Journal of Dairy Science*, 92(12), 6176–6185.
- Prendiville, R., Pierce, K. M., & Buckley, F. (2010). A comparison between Holstein-Friesian and Jersey dairy cows and their F1 cross with regard to milk yield,



somatic cell score, mastitis, and milking characteristics under grazing conditions. *Journal of Dairy Science*, 93, 2741–50.

Rajčević, M., Potočnik, K., & Levstek, J. (2003). Correlations between somatic cells count and milk composition with regard to the season. *Agriculturae Conspectus Scientificus*, 68(3), 221–226.

Raymond, A.K. & Ratnakumar D. (1997). Genetic improvement of cattle, buffalo and small ruminants in Malaysia. In: Report on breeding programmes for ruminants in Asia. Expert discussion held in Kandalama (Matale), Sri Lanka, 31 March – 4 April 1997. MLD & RI, Sri Lanka and GTZ, Germany. pp 77-91

Roos, Y. H. (2009). Solid and liquid states of lactose. In: *Advanced Dairy Chemistry. Volume 3: Lactose, water, salts and minor constituents*, McSweeney P.L.H. and P.F. Fox (eds.), Springer, DOI 10.1007/978-0-387-84865-5\_2,

Ruegg P.L. & Pantoja J.C.F (2013): Understanding and using somatic cell counts to improve milk quality. *Irish Journal of Agricultural and Food Research* 52, 101–117.

Ruegg, P. L. (2001). Milk secretion and quality standards. *Milking and Milk Quality* No. 404, The Babcock Institute, University of Wisconsin, USA. [http://babcock.cals.wisc.edu/downloads/du/du\\_404.en.pdf](http://babcock.cals.wisc.edu/downloads/du/du_404.en.pdf).

Rupp, R., & Boichard, D. (1999). Genetic parameters for clinical mastitis, somatic cell score, production, udder type traits, and milking ease in first lactation Holsteins. *Journal of Dairy Science*, 82(10), 2198–204.

Sandrucci, A., Bava, L., Zucali, M., & Tamburini, A. (2014). Management factors and cow traits influencing milk somatic cell counts and teat hyperkeratosis during different seasons. *Revista Brasileira de Zootecnia*, 43(9), 505–511.

SNP FAQ Archive [Internet]. General information about single nucleotide polymorphisms. Bethesda (MD): National Center for Biotechnology Information (US); 2005-. Retrieved from: <https://www.ncbi.nlm.nih.gov/books/NBK44382/>

Sarikaya, H., Schlamberger, G., Meyer, H. H. D., & Bruckmaier, R. M. (2006). Leukocyte populations and mRNA expression of inflammatory factors in quarter milk fractions at different somatic cell score levels in dairy cows. *Journal of Dairy Science*, 89(7), 2479–86.

Sarkar, U., Gupta, A.K., Sarkar, V., Mohanty, T.K., Raina, V.S. & Prasad, S. (2006). Factors affecting test day milk yield and milk composition in dairy animals. *Journal of Dairying, Foods & H.S.* 25 (2), 129-132.

SAS Institute Inc. 2013. What's New in SAS® 9.4. Cary, NC: SAS Institute Inc.

- Schepers, A. J., Lam, T. J. G. M., Schukken, Y. H., Wilmink, J. B. M., & Hanekamp, W. J. A. (1997). Estimation of variance components for somatic cell counts to determine thresholds for uninfected quarters. *Journal of Dairy Science*, 80(8), 1833–1840.
- Schrick, F. N., Hockett, M. E., Saxton, A. M., Lewis, M. J., Dowlen, H. H., & Oliver, S. P. (2001). Influence of subclinical mastitis during early lactation on reproductive parameters. *Journal of Dairy Science*, 84(6), 1407–1412.
- Schutz, M. M., Hansen, L. B., Steuernagel, G. R., & Kuck, A. L. (1990). Variation of milk, fat, protein, and somatic cells for dairy cattle. *Journal of Dairy Science*, 73(2), 484–493.
- Sender, G., Korwin-Kossakowska, A., Pawlik, A., Hameed, K. G. A., & Oprządek, J. (2013). Genetic basis of mastitis resistance in dairy cattle – A review / Podstawy genetyczne odporności krów mlecznych na zapalenie wymienia – artykuł przeglądowy. *Annals of Animal Science*, 13(4), 663–673.
- Sharma, D. K.; Jallewar, P. K.; & Sharma, K. K. (2010). Antibioqram of bacteria isolated from bovine subclinical mastitis. *Indian Veterinary Journal*, 87, 407-407
- Sharma, N., Singh, N. K., & Bhadwal, M. S. (2011). Relationship of somatic cell count and mastitis: An overview. *Asian-Australasian Journal of Animal Sciences*, 24(3), 429–438.
- Shitandi, A., & Kihumbu, G. (2004). Assessment of the California mastitis test usage in smallholder dairy herds and risk of violative antimicrobial residues. *Journal of Veterinary Science*, 5, 5–9.
- Singh, V. (2014). Association between teat condition score and udder health status in dairy bovine. *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*, 7(5), 09–11.
- Singh, R. S., & Bansal, B. K. (2013). Udder health in relation to udder and teat morphometry in Holstein Friesian × Sahiwal crossbred dairy cows. *Tropical Animal Health Production*, 46(1):93-8
- Sladek, Z., Rysanek, D., Ryznarova, H., & Faldyna, M. (2006). The role of neutrophil apoptosis during experimentally induced *Streptococcus uberis* mastitis. *Veterinarni Medicina*, 51(9), 437–447.
- Smirnova, I., Poltorak, a, Chan, E. K., McBride, C., & Beutler, B. (2000). Phylogenetic variation and polymorphism at the toll-like receptor 4 locus (TLR4). *Genome Biology*, 1(1), research002.1–002.10.
- Sodeland M., Kent M.P., Olsen H.G., Opsal M.A., Svendsen M., Sehested E., Hayes B.J., & Lien S. (2011). Quantitative trait loci for clinical mastitis on chromosomes 2, 6, 14 and 20 in Norwegian Red cattle. *Animal Genetic*, 42, 457–

- Sterrett, A. E., Wood, C. L., McQuerry, K. J., & Bewley, J. M. (2013). Changes in teat-end hyperkeratosis after installation of an individual quarter pulsation milking system. *Journal of Dairy Science*, 96(6), 4041–4046.
- Sudhakar, K., Panneerselvam S., Thiruvenkadan A.K, Abraham, J. & Vinodkumar G. (2013). Factors effecting milk composition of crossbred dairy cattle in southern India. *International Journal of Food, Agriculture and Veterinary Sciences*, 3 (1), 229-233.
- Tančin, V., Ipema, B., Hogewerf, P., & Mačuhová, J. (2006). Sources of variation in milk flow characteristics at udder and quarter Levels. *Journal of Dairy Science*, 89(3), 978–988.
- Taponen, S., Simojoki, H., Haveri, M., Larsen, H. D., & Pyörälä, S. (2006). Clinical characteristics and persistence of bovine mastitis caused by different species of coagulase-negative staphylococci identified with API or AFLP. *Veterinary Microbiology*, 115(1–3), 199–207.
- Turner, M. . (2003). The role of mannose-binding lectin in health and disease. *Molecular Immunology*, 40(7), 423–429.
- Tydell, C. C., Yuan, J., Tran, P., & Michael, E. (2006). Bovine peptidoglycan recognition protein-S: Antimicrobial Activity, localization, secretion, and binding properties. *Journal of immunology*. 176 (2) 1154-1162
- Usman, T., Yu, Y., Liu, C., Fan, Z., & Wang, Y. (2014). Comparison of methods for high quantity and quality genomic DNA extraction from raw cow milk. *Genetic and Molecular Research*, 13(2), 3319-3328.
- Vaclavik V.A., Christian E.W. (2014) Milk and milk products. In: Essentials of Food Science. Springer, New York, NY. pp 201-229.
- Van den Bornea, B.H.P., Vernooija J.C.M., Lupindub A.M., van Schaikc G., Frankenab F., Lamc T.J.G.M. & Nielen M. (2011). Relationship between somatic cell count status and subsequent clinical mastitis in Dutch dairy cows. *Preventive Veterinary Medicine*, 102, 265– 273.
- Vesterinen, H. M., Corfe, I. J., Sinkkonen, V., Iivanainen, A., Jernvall, J., & Laakkonen, J. (2015). Teat morphology characterization with 3D imaging. *The Anatomical Record*, 298(7), 1359–1366.
- Waage, S., Sviland, S., & Ødegaard, S. A. (1998). Identification of Risk Factors for Clinical Mastitis in Dairy Heifers. *Journal of Dairy Science*, 81(5), 1275–1284.
- Walsh, S., Buckley, F., Berry, D.P., Rath, M., Pierce, K., Byrne, N. & Dillon, P. (2007). Effects of breed, feeding system, and parity on udder health and milking characteristics. *Journal of Dairy Science*, 90, 5767–5779.



- Wang, C., Liu, M., Li, Q., Ju, Z., Huang, J., Li, J., ... Zhong, J. (2011). Three novel single-nucleotide polymorphisms of MBL1 gene in Chinese native cattle and their associations with milk performance traits. *Veterinary Immunology and Immunopathology*, 139(2–4), 229–36.
- Wang, H. L., Li, Z. X., Chen, L., Yang, J., Wang, L. J., He, H., ... Liu, X. L. (2013). Polymorphism in PGLYRP-2 gene by PCR-RFLP and its association with somatic cell score and percentage of fat in Chinese Holstein. *Genetics and Molecular Research*, 12(4), 6743–6751.
- Wang, H. L., Li, Z. X., Wang, L. J., He, H., Yang, J., Chen, L., ... Liu, X. L. (2013). Polymorphism in PGLYRP-1 gene by PCR-RFLP and its association with somatic cell score in Chinese Holstein. *Research in Veterinary Science*, 95(2), 508–14.
- Wang, X., Ju, Z., Huang, J., Hou, M., Zhou, L., Qi, C., & Wang, C. (2012). The relationship between the variants of the bovine MBL2 gene and milk production traits, mastitis, serum MBL-C levels and complement activity. *Veterinary Immunology and Immunopathology*, 148(3–4), 311–9.
- Wang, X., Xu, S., Gao, X., Ren, H., & Chen, J. (2007). Genetic polymorphism of TLR4 gene and correlation with mastitis in cattle. *Journal of Genetics and Genomics*. 34(5), 406–12.
- Washburn, S. P., S. L. White, J. T. Green Jr., and G. A. Benson. 2002. Reproduction, mastitis and body condition of seasonally calved Holstein and Jersey cows in confinement or pasture systems. *Journal of Dairy Science*, 85:105–111.
- Weiss, D., M. Weinfurtner, & R. M. Bruckmaier. 2004. Teat anatomy and its relationship with quarter and udder milk flow characteristics in dairy cows. *Journal of Dairy Science*. 87:3280–3289
- Yang, L., Yang, Q., Yi, M., Pang, Z. H., & Xiong, B. H. (2013). Effects of seasonal change and parity on raw milk composition and related indices in Chinese Holstein cows in northern China. *Journal of Dairy Science*, 96(11), 6863–6869.
- Yeh, F., Yang R. and Boyle T. (2000). A Software for Population Genetics Data Analysis, POPGENE Version 1.32. Department of Renewable Resources University of Alberta, Edmonton, Canada.
- Yuan, Z., Li, J., Li, J., Gao, X., & Xu, S. (2013). SNPs identification and its correlation analysis with milk somatic cell score in bovine MBL1 gene. *Molecular Biology Reports*, 40(1), 7–12.
- Zadoks, R. N., Allore, H. G., Barkema, H. W., Sampimon, O. C., Wellenberg, G. J., Gröhn, Y. T., & Schukken, Y. H. (2001). Cow- and Quarter-Level Risk Factors for *Streptococcus uberis* and *Staphylococcus aureus* Mastitis. *Journal of Dairy Science*, 84(12), 2649–2663.

Zhao, X., & Lacasse, P. (2008). Mammary tissue damage during bovine mastitis: causes and control. *Journal of Animal Science*, 86(13 Suppl), 57–65.

Zwertvaegher, I., Van Weyenberg, S., Piepers, S., Baert, J., & De Vliegher, S. (2012). Variance components of teat dimensions in dairy cows and associated factors. *Journal of Dairy Science*, 95(9), 4978–4988.



## LIST OF PUBLICATIONS

- Sani, U.M., J.M. Panandam and H. Yaakub (2016). Udder quarters status in Friesian and Jersey cows affected with subclinical mastitis. Proceeding of the 37th Annual Conference of the Malaysian Society of Animal Production, Malacca, Malaysia, 1-3 June 2016, pp.84–85.
- Sani, U.M., J.M. Panandam, H. Yaakub, S. Sumita and Z.A. Rahman (2017). Association analysis of teat morphology and udder quarter mastitis status in Friesian and Jersey cows. Proceeding of the 4th ASEAN Regional Conference on Animal Production, Senai, Johor, Malaysia.
- Sani, U.M., J.M. Panandam, H. Yaakub, S. Sumita and Z.A. Rahman (2017). Association analysis of single nucleotide polymorphism in bovine PGLYRP-2 gene with milk somatic cell score and some milk composition traits in Friesian and Jersey cows. Proceeding of the 12th Malaysia International Genetic Congress, 25 - 27 September, 2017, Bangi-Putrajaya Malaysia.



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