



UNIVERSITI PUTRA MALAYSIA

***CHARACTERISTICS OF NaCl-SALINIZED CUCURBITS AND IMPACT OF
SILICON IN ALLEVIATING SALT STRESS***

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SILICON IN ALLEVIATING SALT STRESS**

By

MUHAMMAD NAJIB BIN OTHMAN GHANI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Master of Science**

November 2017

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Abstracts of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

CHARACTERISTICS OF NaCl-SALINIZED CUCURBITS AND IMPACT OF SILICON IN ALLEVIATING SALT STRESS

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November 2017

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Members of *Cucurbitaceae* are salt-sensitive plants and continuous fertilizer application without sufficient leaching may create saline environment that consequently reducing their growth and yield. Among other methods, silicon fertilization was reported to have some ability to reduce negative salinity effects on many plant species. This study was carried out to evaluate the physiological responses of four selected *Cucurbitaceae* to salinity stress, and to investigate the possible role of silicon in alleviating salinity effects on the crops. Four types of *Cucurbitaceae* viz. cucumber (*Cucumis sativa*), pumpkin (*Cucurbita moschata*), bitter gourd (*Momordica charantia*) and bottle gourd (*Lagenaria siceraria*) were subjected to four levels of NaCl (0, 25, 50, 75 mM) and data at vegetative stages; growth, leaf relative water content (RWC), mineral content in leaf, stem, root, relative chlorophyll content and proline concentration were collected. In the second experiment, a study involving two species of *Cucurbitaceae* that have shown highest (bitter gourd) and lowest (cucumber) salinity tolerant were performed. The species were subjected to two levels of NaCl (0, 50 mM) and three silicon concentrations (0, 50, 100 mg/L sodium silicate) and data on growth, leaf RWC, leaf mineral content, net photosynthesis and stomatal conductance at vegetative stage were collected.

Among all species, bitter gourd was least salt-sensitive while cucumber was most salt-sensitive. In terms of growth, bitter gourd had the lowest reduction of leaf area, plant height and stem dry weight while cucumber had the highest reduction of leaf area. Total reduction of plant dry matter as salinity increased to 75 mM NaCl was the least in bitter gourd, 44.40% and the highest in cucumber, 67.84%. Significant reduction of chlorophyll content was recorded at 75 mM in cucumber and pumpkin while in other species shown no significant

reduction. At 75 mM NaCl, cucumber leaf water status was significantly impaired by salinity which marked by highest accumulation of proline (3.55 times compared to control) and strong negative correlation between proline and RWC ($r = -0.83$, $p \leq 0.01$), whereas in other species no significant association was recorded.

When treated with 100 mg/L silicon, growth of salt-stressed and non-salt stressed plants for both species was significantly improved, with more beneficial effects recorded on the salt-stressed plants and salt-sensitive species. In terms of Na^+ ion, plants treated with 50 mg/L silicon had 11.11% significantly lower Na^+ ion in leaf compared to 0 mg/L silicon, regardless of species and salinity condition. Moreover, treatment of 100 mg/L silicon significantly increased net photosynthesis and stomatal conductance by 12.13 and 30.14% respectively, compared to 0 mg/L silicon treatment. Besides that, plants supplied with silicon at 100 mg/L also had significantly higher RWC compared to non-silicon supplied plants.

In conclusion, based on reduction in plant dry matter, accumulation of Na^+ and Cl^- ions in leaf, degradation of chlorophyll content and impaired leaf water status, bitter gourd exhibited a better adaptation to salinity stress than other *Cucurbitaceae* tested and application of silicon can alleviate salinity stress in both on salt-sensitive species (cucumber) and on a less salt-sensitive species (bitter gourd) by reducing sodium toxicity, increasing photosynthetic activity and improving leaf water status.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

CIRI-CIRI CUCURBIT YANG MENGALAMI KEMASINAN NaCl DAN IMPAK SILIKON DALAM MENGURANGKAN TEGASAN KEMASINAN

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Tumbuhan dari kalangan *Cucurbitaceae* adalah peka kemasinan dan pembajaan berterusan tanpa proses larut lesap yang mencukupi akan menyebabkan persekitaran bergaram yang mengurangkan pertumbuhan dan hasil. Antara kaedah-kaedah lain, pembajaan silikon dilaporkan berupaya mengurangkan kesan negatif kemasinan terhadap pelbagai spesies tumbuhan. Kajian ini telah dijalankan untuk menilai gerak balas fisiologi empat *Cucurbitaceae* terpilih terhadap tegasan kemasinan, dan menyelidik peranan silikon dalam mengurangkan kesan kemasinan terhadap tumbuhan. Empat jenis *Cucurbitaceae* iaitu timun (*Cucumis sativa*), labu manis (*Cucurbita moschata*), peria katak (*Momordica charantia*) dan labu botol (*Lagenaria siceraria*) telah diberikan empat tahap kemasinan NaCl (0, 25, 50, 75 mM) dan data peringkat vegetatif; pertumbuhan, kandungan air daun relatif, kandungan mineral daun, batang, akar, kandungan klorofil relatif dan kepekatan prolin telah diambil. Pada eksperimen kedua, kajian yang melibatkan dua spesies *Cucurbitaceae* yang menunjukkan ketahanan kemasinan tertinggi (peria katak) dan terendah (timun) telah dijalankan. Spesies-spesies tersebut telah diberikan dua tahap kemasinan NaCl (0, 50 mM) dan tiga kepekatan silikon (0, 50, 100 mg/L sodium silika) dan data pertumbuhan, kandungan air daun relatif, kandungan mineral daun, fotosintesis dan aliran stomata telah diambil pada peringkat vegetatif.

Antara semua spesies, peria katak adalah paling kurang peka kemasinan manakala timun adalah paling peka kemasinan. Dari segi pertumbuhan, pengurangan keluasan daun, ketinggian pokok dan berat kering batang, peria katak adalah yang terendah manakala timun mempunyai pengurangan keluasan daun yang tertinggi. Jumlah pengurangan bahan kering pokok

apabila kadar kemasinan meningkat ke 75 mM NaCl adalah terendah pada peria katak, 44.40% dan tertinggi pada timun, 67.84%. Pengurangan kandungan klorofil yang signifikan direkodkan pada 75 mM untuk timun dan labu manis manakala untuk spesies lain, tiada pengurangan signifikan direkodkan. Pada 75 mM NaCl, status air daun pada timun terjejas secara signifikan oleh kemasinan yang dijelaskan oleh pengumpulan prolin yang tertinggi (3.55 kali lebih tinggi berbanding kawalan) dan korelasi negatif antara prolin dan kandungan air daun relatif ($r = -0.83$, $p \leq 0.01$), manakala tiada korelasi signifikan direkodkan pada spesies lain.

Apabila dirawat dengan 100 mg/L silikon, pertumbuhan pokok yang tegas kemasinan dan tidak tegas kemasinan untuk kedua-dua spesies bertambah baik secara signifikan, dengan kesan bermanfaat direkodkan pada pokok yang tegas kemasinan dari spesies yang peka kemasinan. Dari segi ion Na^+ , pokok yang dirawat dengan 50 mg/L silikon mempunyai 11.11% lebih rendah ion Na^+ secara signifikan berbanding 0 mg/L silikon, tidak kira spesies atau keadaan kemasinan. Tambahan lagi, rawatan 100 mg/L silikon juga meningkatkan fotosintesis dan aliran stomata masing-masing sebanyak 12.13 dan 30.14% berbanding 0 mg/L rawatan silikon. Selain itu, pokok yang diberikan silikon pada kadar 100 mg/L juga mempunyai kandungan air daun relatif yang secara signifikan lebih tinggi berbanding pokok tanpa penambahan silikon.

Sebagai rumusan, berdasarkan pengurangan bahan kering pokok, pengumpulan ion-ion Na^+ dan Cl^- pada daun, pengurangan kandungan klorofil dan status air daun, peria katak menunjukkan penyesuaian terhadap tegasan kemasinan yang lebih baik berbanding *Cucurbitaceae* lain di dalam kajian ini dan aplikasi silikon boleh mengurangkan tegasan kemasinan pada spesies yang peka kemasinan (timun) dan kurang peka kemasinan (peria katak) dengan mengurangkan ketoksikan sodium, meningkatkan aktiviti fotosintesis dan menambahbaik status air daun.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

mM	Milimolar
mmol/L	milimolar per litre
RWC	Relative water content
P _n	Net photosynthesis
g _s	Stomatal conductance
μmol	micromole
DW	Dry weight
FW	Fresh weight
TW	Turgid weight
EC	Electrical conductivity
MDA	Malondialdehyde
ROS	Reactive oxygen species
dS/m	Decisiemens per metre

CHAPTER 1

INTRODUCTION

1.1 Background information

In 2015, the world population had reached 7.3 billion and with approximately 83 million addition increased annually, the current population is projected to reach 8.5 billion by 2030 (UN, 2015). Agricultural productivity in contrast, is not increasing at a required rate to keep up with the food demand. Many factors contributed to this problem such as availability of agricultural land, fresh water resources, ever-increasing biotic and abiotic stresses, and low economic activity in agricultural sector (Athar and Ashraf, 2009). It is believed that abiotic stresses are considered to be the key source of yield reduction (Munns and Tester, 2008). Salinity is one of the major abiotic stresses that affect production in agriculture. According to FAO statistics, over 800 million hectares of land throughout the world is presently salt-affected, equivalent to more than 6% of the world's total land area. Continuing salinization of arable land is expected to result in a 30% loss of agricultural land over the next 25 years and up to 50% loss by 2050 (Vahdati and Leslie, 2013).

Salinity causes detrimental effects on plant's life. Growth reduction is a consequence of numerous physiological responses including modifications of ion balance, water status, mineral nutrition, stomatal behavior, photosynthetic efficiency, carbon allocation and utilization (Kusvuran, 2011). The degree to which growth is reduced by salinity differs greatly among species and to a lesser extent among varieties within a species (Ghoulam *et al.*, 2002).

In combating salinity problem, numerous approaches and studies have been done in recent years. Traditional breeding programmes have proven to be time-consuming and very challenging due to the complexity of the traits since salt tolerance is the result of complex genetics and physiological interactions (Flowers, 2004). Alternatively, grafting onto rootstocks that capable in ameliorating salt-induced damage to the shoot is feasible but limitations may occur in compatibility of the scions and rootstocks (Cohen *et al.*, 2007) while chemical amendments or leaching salts application is not environmentally friendly (Cuartero *et al.*, 2006).

Silicon (Si) has essential function in healing plants in response to environmental stresses (Sahebi *et al.*, 2015). Application of silicon is a promising method that has been proven in alleviating salt stress in various important crops namely rice (Gong *et al.* 2006), barley (Liang *et al.*, 2003),

wheat (Tuna *et al.*, 2008), sugarcane (Ashraf *et al.*, 2010), soybean (Lee *et al.*, 2010), tomato (Romero-Aranda *et al.*, 2006), and zucchini (Savvas *et al.*, 2009). Salinity tolerance in plants treated with silicon is associated with the ability of the silicon to reduce ion toxicity and decrease oxidative damage under salt stress.

Cucurbitaceae family primarily comprises species consumed as food worldwide which consists of about 130 genera and 800 species (Jeffrey, 2005). Genetic diversity within the family is tremendous and the ability of *Cucurbitaceae* to tolerate salinity stress differs considerably among species. Majority of the cucurbits are reported moderately sensitive to salt stress such as cucumber (*Cucumis sativus* L.) and musk melon (*Cucumis melo* L.) while bitter gourd (*Momordica charantia* L.) is reported to have salinity tolerance (Pessarakli, 2016).

While studies of *Cucurbitaceae* species in response to salinity stress has been done extensively in other regions, information on salinity responses in local Malaysia *Cucurbitaceae* species is still lacking. Due to the tremendous genetic diversity within *Cucurbitaceae* family, the effects of salinity on Malaysian *Cucurbitaceae* species might be different from studies reported in other regions. Information gained from studies on the effects of salinity on physiological and biochemical aspects of *Cucurbitaceae* species could lead to identification of salt tolerant species among *Cucurbitaceae* in Malaysia. Furthermore, the alleviating effects of silicon against salinity stress could be beneficial in improving production in salt-stressed *Cucurbitaceae* as well as giving new perspectives in *Cucurbitaceae* research in the future.

1.2 Problem statements

Traditional breeding, grafting onto salt-tolerant rootstock and application of chemical amendments have its own limitation in overcoming salinity problems. Alternatively, agronomic practice approach which can alleviate salt stress such as application of silicon is more efficient and promising. While studies on effects of salinity stress on Cucurbit has been done extensively in other regions, information on salinity responses in local Malaysia Cucurbit species is still lacking and no comparative studies has been done previously in comparing their salinity tolerance level. Besides that, the alleviate effects of silicon in reducing salt stress in both susceptible and less susceptible Cucurbits are still equivocal and need to be elucidated further.

1.3 Aim and objectives of the study

The aim of the study is to differentiate salinity tolerance level among selected *Cucurbitaceae* species and to enhance salinity tolerance using silicon at vegetative stage. In achieving that, the objectives of this study include:

- i. To characterize the growth and physiological responses of *Cucurbitaceae* species to different levels of NaCl-induced salinity.
- ii. To determine the susceptible and salt-tolerant species among the *Cucurbitaceae* species studied.
- iii. To investigate the effects of varying concentrations of NaCl salinity and silicon application on physiological and biochemical properties in both susceptible and tolerant species.

REFERENCES

- Abdalmajid, N.A.M. (2012). *Growth and development of tomato (Lycopersicon esculentum Mill.) cultivars in vitro under salinity condition*, PhD Thesis, Universiti Putra Malaysia.
- Ahmad, P., Jaleel, C.A., Salem, M.A., Nabi, G. and Sharma, S. (2010). Roles of enzymatic and nonenzymatic antioxidants in plants during abiotic stress. *Critical Reviews in Biotechnology* 30(3):161–175.
- Ahmed, B.A., Inoue, M. and Moritani, S. (2010). Effect of saline water irrigation and manure application on the available water content, soil salinity, and growth of wheat. *Agriculture Water Management* 97:165-170.
- Akram, N.A. and Ashraf, M. (2011). Improvement in growth, chlorophyll pigments and photosynthetic performance in salt-stressed plants of sunflower (*Helianthus annuus* L.) by foliar application of 5-aminolevulinic acid. *Agrochimica* 55:94-104.
- Al-aghabary, K., Zhu, Z.J. and Shi, Q.H. (2004). Influence of silicon supply on chlorophyll content, chlorophyll fluorescence, and antioxidative enzyme activities in tomato plants under salt stress. *Journal of Plant Nutrition* 27:2101–2115.
- Anjum, M.A. (2008). Effect of NaCl concentration in irrigation water on growth and polyamine metabolism in two citrus rootstocks with different levels of salinity tolerance. *Acta Physiologiae Plantarum* 30(1): 43-52.
- Apse, M.P., Aharon, G.S., Snedden, W.A. and Blumwald, E. (1999). Salt tolerance conferred by overexpression of a vacuolar Na⁺/H⁺ antiport in arabidopsis. *Science* 285 (5431):1256-1258.
- Apse M.P. and Blumwald, E. (2007). Na⁺ transport in plants. *FEBS Letters* 581(12):2247–2254.
- Arfan, M., Athar, H.R. and Ashraf, M. (2007). Does exogenous application of salicylic acid through the rooting medium modulate growth and photosynthetic capacity in differently adapted spring wheat cultivars under salt stress?. *Journal of Plant Physiology* 6:685-694.
- Arzani, A. (2008). Improving salinity tolerance in crop plants: A Biotechnological View. *In Vitro Cellular & Developmental Biology-Plant* 44 (5):373-383.
- Ashraf, M. and Bhatti, A. (2000). Effect of salinity on growth and chlorophyll content of rice. *Pakistan Journal of Scientific and Industrial Research* 43:130-131.

- Ashraf, M. and Sultana, R. (2000). Combination effect of NaCl salinity and N-form on mineral composition of sunflower plants. *Biology Plant* 43:615-619.
- Ashraf, M. and Foolad, M.R. (2007). Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environmental and Experimental Botany* 59(2):206–216.
- Ashraf, M., Ozturk, M. and Athar, H.R. (2008). *Salinity and water stress: improving crop efficiency*. Netherlands : Springer, 244p.
- Ashraf, M., Rahmatullah, Ahmad, R., Bhatti, A.S., Afzal, M., Sarwar, A., Maqsood, M.A. and Kanwal, S. (2010). Amelioration of salt stress in sugarcane (*Saccharum officinarum* L.) by supplying potassium and silicon in hydroponics. *Pedosphere* 20(2):153-162.
- Athar, H.R. and M. Ashraf. (2009). Strategies for crop improvement against salt and water stress: An overview. In Ashraf, M., Ozturk, M. and Athar, H.R. (Eds.) *Salinity and water stress: Improving crop efficiency*. Netherlands : Springer, 1-16pp.
- Awang, Y., Shaharom, A.S., Mohamad, R.B. and Selamat, A. (2009). Chemical and physical characteristics of cocopeat-based media mixtures and their effects on the growth and development of *Celosia cristata*. *American Journal of Agricultural and Biological Science* 4(1):63–71.
- Bates, L., Waldren, R. and Teare, I. (1973). Rapid Determination of Free Proline for Water-Stress Studies. *Plant and Soil* 39 (1):205-207.
- Blumwald, E. (2000). Sodium transport and salt tolerance in plants. *Current Opinion in Cell Biology* 12:431–434.
- Carillo, P., Mastrolonardo, G., Nacca, F., Parisi, D., Verlotta, A., and Fuggi, A. (2008). Nitrogen metabolism in durum wheat under salinity: accumulation of proline and glycine betaine. *Functional Plant Biology* 35(5):412-426.
- Carillo, P., Annunziata, M. G., Pontecorvo, G., Fuggi, A. and Woodrow, P. (2011). Salinity stress and salt tolerance, In Shanker, A. (Ed). *Abiotic Stress in Plants Mechanisms and Adaptations*. InTech. 21-38pp.
- Cavins, T., Whipker, B., Fonteno, W., Harden, B., McCALL, I., and Gibson, J. (2000). Monitoring and managing pH and EC using the PourThru extraction method. *Horticulture Information Leaflet* 590:1–17.
- Chan, Y.K. (1978). Soils under coconut in Peninsular Malaysia. In "Proceedings of the International Conference on Cocoa and Coconuts" : 583-594. Incorporated Society of Planters

- Cohen, R., Burger, Y., Horev, C., Koren, A., and Edelstein, M. (2007). Introducing Grafted Cucurbits to Modern Agriculture: The Israeli Experience. *Plant disease* 91(8):916-923.
- Cuartero, J., Bolarín, M.C., Asíns, M.J., Moreno, V. (2006). Increasing salt tolerance in the tomato. *Journal of Experimental Botany* 57:1045–1058.
- Dadkhah, A. (2011). Effect of Salinity on Growth and Leaf Photosynthesis of Two Sugar Beet (*Beta vulgaris* L.) Cultivars. *Journal of Agricultural Science and Technology* 13:1001–1012.
- Dajic, Z. (2006). Salt Stress. In Madhava, K.V., Raghavendra, A.S. and Janardhan Reddy, K. (Eds.) *Physiology and Molecular Biology of Stress Tolerance in Plants*. Germany: Springer, 41-82 pp.
- Davenport, R., James, R., Zakrisson-Plogander, A., Tester, M. and Munns, R. (2005). Control of Sodium Transport in Durum Wheat. *Plant Physiology* 137:807-818.
- Eckardt, N.A. (2009). A new chlorophyll degradation pathway. *Plant Cell* 21: 700.
- El-Shrai, M., Mostafa, M.A., Sanaa, A. and Shehata, S. (2011). Physiological aspect of NaCl-salt stress tolerant among cucurbitaceous cultivars. *Australian Journal of Basic and Applied Sciences* 5(11):62-71.
- Epstein, E., Bloom, A.J. (2005). *Mineral nutrition of plants: principles and perspectives* (2nd ed). Sunderland : Sinauer, 600 pp.
- Fahad, S., Hussain, S., Matloob, A., Khan, F.A., Khaliq, A., Saud, S. and Huang, J. (2015). Phytohormones and plant responses to salinity stress: a review. *Plant Growth Regulator* 75:391.
- Farhoudi, R., Saeedipour, S. and Mohammadreza, D. (2011). The effect of NaCl seed priming on salt tolerance, antioxidant enzyme activity, proline and carbohydrate accumulation of Muskmelon (*Cucumis melo* L.) under saline condition. *African Journal of Agricultural Research* 6(6):1363-1370.
- Farias, M.E., Lima, Y.J.M, Melo, D.F. and Dizengremel, P. (1997). NaCl-induced changes of NAD (P) malic enzyme activities in *Eucalyptus citriodora* leaves. *Trees* 12:66-72.
- Flagella, Z., Trono, D., Pompa, M., Di Fonzo, N. and Pastore, D. (2006). Seawater stress applied at germination affects mitochondrial function in durum wheat (*Triticum durum*) early seedlings. *Functional Plant Biology* 33(4):357-366.

- Flowers, T.J. (2004). Improving crop salt tolerance. *Journal of Experimental Botany* 55(396):307–319.
- Furtana, B.G. and Tipirdamaz, R. (2008). Physiological and antioxidant response of three cultivars of cucumber (*Cucumis sativus* L.) to salinity. *Turkish Journal of Biology* 34:287-296.
- Gama, P.B.S., Inanaga, S., Tanaka, K., Nakazawa, R. (2007). Physiological response of common bean (*Phaseolus vulgaris* L.) seedlings to salinity stress. *African Journal of Biotechnology* 6(2):79–88.
- Genc, Y., McDonald, G.K. and Tester, M. (2007). Reassessment of tissue Na⁺ concentration as a criterion for salinity tolerance in bread wheat. *Plant, Cell and Environment* 30:1486–1498.
- Ghoulam, C., Foursy, A., Fares, K. (2002). Effects of salt stress on growth, inorganic ions and proline accumulation in relation to osmotic adjustment in five sugar beet cultivars. *Environmental and Experimental Botany* 47:39-50.
- Gong, H.J., Randall, D.P. and Flowers, T.J. (2006). Silicon deposition in root reduces sodium uptake in rice (*Oryza sativa* L.) seedlings by reducing bypass flow. *Plant Cell Environment* 29:1970–1979.
- Grattan, S.R. and Grieve, C.M. (1994). Mineral nutrient acquisition and response by plants grown in saline environments. In: Pessarakli, M. (Ed.), *Handbook of Plant and Crop Stress*. Marcel Dekker, New York, pp. 203-226.
- Grisso, R., Alley, M., Wysor, W.J., Holshouser, D. and Thomason, W. (2009). Precision Farming Tools: Soil Electrical Conductivity. *Virginia Cooperative extension Publication* 442-508.
- Haque, S.A. (2006). Salinity problems and crop production in coastal regions of bangladesh. *Pakistan Journal of Botany* 38(5):1359-1365.
- Hasegawa, P.M., Bressan, R.A., Zhu, J.K. and Bohnert, H.J. (2000). Plant cellular and molecular responses to high salinity. *Annual Review of Plant Physiology and Plant Molecular Biology* 51:463-499.
- Hashemi, A., Abdolzadeh, A. and Sadeghipour, H.R. (2010) Beneficial effects of silicon nutrition in alleviating salinity stress in hydroponically grown canola, *Brassica napus* L., plants. *Soil Science and Plant Nutrition* 56(2):244-253.
- Hashim, G. M. (2003). Salt-Affected Soils of Malaysia in —Proceedings of the Workshop on Soil Science in Malaysia towards 2020”Malaysian Society of Soil Science.

- Hayat, S., Hayat, Q., Alyemeni, M.N., Wani, A.S., Pitchel, J. and Ahmad, A. (2012). Role of proline under changing environments: a review. *Plant Signaling and Behavior* 7(11):1456-1466.
- Henriet, C., Draye, X., Oppitz, I., Swennen, R. and Delvaux, B. (2006). Effects, distribution and uptake of silicon in banana (*Musa* spp.) under controlled conditions. *Plant Soil* 287:359–374.
- Iwasaki, K., Maier, P., Fecht, M. and Horst, W.J. (2002). Leaf apoplastic silicon enhances manganese tolerance of cowpea. *Plant Physiology* 159:167-173.
- Izzo, R., Incerti, A. and Bertolla, C. (2008). Seawater irrigation : Effect on growth and nutrient uptake of sunflower plants. In Abdelley, C., Ozturk, M., Ashraf, M. and Gringnon, C. (Eds.) *Biosaline Agriculture and High Salinity Tolerance*. Switzerland : Verlag, 61-69pp.
- James, R.A., Blake, C., Byrt, C.S. and Munns, R. (2011). Major genes for Na⁺ exclusion, Nax1 and Nax2 (wheat HKT1;4 and HKT1;5), decrease Na⁺ accumulation in bread wheat leaves under saline and waterlogged conditions. *Journal of Experimental Botany* 62:2939-2947.
- James, R.A., Munns, R., Caemmerer, S.V., Trejo, C., Miller, C., Condon, O. (2006). Photosynthetic capacity is related to the cellular and subcellular partitioning of Na⁺, K⁺ and Cl⁻ in salt affected barley and durum wheat. *Plant Cell Environment* 29:2185–2197.
- James, R.A., Caemmerer, S., Condon, A.G., Zwart, A.B. and Munns, R. (2008). Genetic variation in tolerance to the osmotic stress component of salinity stress in durum wheat. *Functional Plant Biology* 35(2):111-123.
- Jamil, M., Lee, C.C., Rehman, S.U., Lee, D.B., Ashraf, M., Rha, E.S. (2005). Salinity (NaCl) tolerance of Brassica species at germination and early seedling growth. *Electronic Journal of Environmental, Agriculture and Food Chemistry* 4(4):970- 976.
- Jeffrey, C. (2005). A new system of *Cucurbitaceae*. *Botanicheskii Zhurnal* 90:332–335.
- Karimi, S., Rahemi, M., Maftoun, M. and Tavallali, V. (2009). Effects of long-term salinity on growth and performance of two pistachio (*Pistacia* L.) rootstocks. *Australian Journal of Basic and Applied Sciences* 3(3):1630-1639.

- Kaufman, P.B., Takeoka, Y., Carlson, T.J., Bigelow, W.C., Jones, J.D., Moore, P.H. and Ghosheh, N.S. (1979). Studies on silica deposition in sugarcane (*Saccharum* spp.) using scanning electron microscopy, energy-dispersive X-ray analysis, neutron activation analysis and light microscopy. *Phytomorphology* 29:185-193.
- Khan, M. and Srivastava, H. (1998). Changes in growth and nitrogen assimilation in maize plants induced by NaCl and growth regulators. *Biologia Plantarum* 41:93-99.
- Khan, M., Al-Mas'oudi, R.S.M, Al-Said, F. and Khan, I. (2013). Salinity effects on growth, electrolyte leakage, chlorophyll content and lipid peroxidation in cucumber (*Cucumis sativa* L.). *International Proceedings of Chemical, Biological & Environmental* 55:28.
- Khan, W.U.D, Aziz, T., Maqsood, M.A., Sabir, M., Ahmad, H.R., Ramzani, P.M.A. and Nasim, M. (2016). Silicon: A beneficial nutrient under salt stress, its uptake mechanism and mode of action. In Hakeem, K.R. et al. (Eds.) *Soil science: Agricultural and environmental perspectives*. Switzerland: Springer Intl. Pub, 287-301pp.
- Kumar, D.S., Sharathnath, K.V., Yogeswaran, P., Harani, A., Sudhakar, K., Sudha, P. and Banji, D. (2010). A medicinal potency of *Momordica charantia*. *International Journal of Pharmaceutical Science Review and Research* 1(2):95.
- Kusvuran, S. (2012). Effects of drought and salt stresses on growth, stomatal conductance, leaf water and osmotic potentials of melon genotypes (*Cucumis melo* L.). *African Journal of Agricultural Research* 7(5):775-781.
- Lechno, S., Zamski, K. and Tel-or, E. (1997). Salt stress-induced responses in cucumber plants. *Journal of Plant Physiology* 150: 206-211.
- Lee, S.K., Sohn, E.Y., Hamayun, M., Yoon, J.Y. and Lee, I.J. (2010). Effect of silicon on growth and salinity stress of soybean plant grown under hydroponic system. *Agroforestry System* 80:333–340.
- Li, Q., Ma, C., Shang, Q. (2007). Effects of silicon on photosynthesis and antioxidant enzymes of maize under drought stress. *Yingyong Shengtai Xuebao* 18:531–536.
- Li, T., Zhang, Y., Liu, H. (2010). Stable expression of Arabidopsis vacuolar Na⁺/H⁺ antiporter gene AtNHX1 and salt tolerance in transgenic soybean for over six generations. *Chinese Science Bulletin* 55:1127-1134.

- Liang, X., Wang, H., Hu, Y., Mao, L., Sun, L., Dong, T., Nan, W. and Bi, Y. (2015). Silicon does not mitigate cell death in cultured tobacco BY-2 cells subjected to salinity without ethylene emission. *Plant Cell Reports* 34:331–343.
- Liang, Y., Sun, W., Zhu, Y.G. and Christie, P. (2007). Mechanisms of silicon-mediated alleviation of abiotic stresses in higher plants: a review. *Environmental Pollution* 147:422–428.
- Liang, Y.C. (1999). Effects of silicon on enzyme activity, and sodium, potassium and calcium concentration in barley under salt stress. *Plant Soil* 209:217–224.
- Liang, Y.C. and Ding, R.X. (2002). Influence of silicon on microdistribution of mineral ions in roots of salt-stressed barley as associated with salt tolerance in plants. *Science China Series C : Life Sciences* 45:298–308.
- Liang, Y.C., Chen, Q., Liu, Q., Zhang, W.H. and Ding, R.X. (2003). Exogenous silicon (Si) increases antioxidant enzyme activity and reduces lipid peroxidation in roots of salt-stressed barley (*Hordeum vulgare* L.). *Journal of Plant Physiology* 160:1157–1164.
- Lim, C.P. (1981). Notes on soil classification in Sarawak. In “*Proceedings of the Seminar on Soil Correlation in Malaysia* (Paramanathan, S., ed) :39-101. Malaysian Society of Soil Science.
- Liu, W.G., Wang, L.Q. and Bai, Y.H. (2003). Research progress in the beneficial elements—silicon for plants. *Acta Botanica Boreali Occidentalia Sinica* 23:2248–2253.
- Ma, J.F. and Yamaji, N. (2006). Silicon uptake and accumulation in higher plants. *Trends in Plant Science* 11:392–397.
- Ma, J.F. and Yamaji, N. (2008). Functions and transport of silicon in plants. *Cellular and Molecular Life Sciences* 65:3049–3057.
- Mahajan, S. and Tuteja, N. (2005). Cold, salinity and drought stress : an overview. *Archives of Biochemistry and Biophysics* 444(2):139-158.
- Mansour, M.M.F. (2000). Nitrogen containing compounds and adaptation of plants to salinity stress. *Biologia Plantarum* 43(4):491-500.
- Mathur, N., Singh, J., Bohra, S., Bohra, A., Vyas, A. (2006). Biomass production, productivity and physiological changes in moth bean genotypes at different salinity levels. *American Journal of Plant Physiology* 1(2):210–213.

- Mazumdar, J. (2011). Phytoliths of pteridophytes. *South Africa Journal of Botany* 77:10–19.
- Mitani, N. and Ma, J.F. (2005). Uptake system of silicon in different plant species. *Journal of Experimental Botany* 56:1255–1261.
- Mohsenzadeh, S., Shahrtash, M. and Mohabatkar, H. (2011). Interactive effects of salicylic acid and silicon on some physiological responses of cadmium-stressed maize seedlings. *Iranian Journal of Science and Technology* 35(1):57-60.
- Monirifar, H. and Barghi, M. (2009). Identification and selection for salt tolerance in alfalfa (*Medicago sativa* L.) ecotypes via physiological traits. *Notulae Scientia Biologicae* 1:63-66.
- Munns, R. (2002). Comparative physiology of salt and water stress. *Plant Cell Environment* 25:239–50.
- Munns, R., James, R.A. and Lauchli, A. (2006). Approaches to increasing the salt tolerance of wheat and other cereals. *Journal of Experimental Botany* 57:1025–43.
- Munns, R. and Tester, M. (2008). Mechanisms of salinity tolerance. *Annual Review of Plant Biology* 59:651-681.
- Najqfian, S., Rahemi, M. and Tavalli, V. (2008). Effects of Salinity on Tolerance of Two Bitter Almond Rootstocks. *American-Eurasian Journal of Agricultural and Environmental Sciences* 3(2):264-268.
- Nikolic, M., Nikolic, N., Liang, Y., Kirkby, E.A. and Romheld, V. (2007). Germanium-68 as an adequate tracer for silicon transport in plants. Characterization of silicon uptake in different crop species. *Plant Physiology* 143(1):495–503.
- Noreen, Z., Ashraf, M. and Akram, N.A. (2010). Salt-induced modulation in some key gas exchange characteristics and ionic relations in pea (*Pisum sativum* L.) and their use as selection criteria. *Crop Pasture Science* 61:369-378.
- Parida, A.K., Das, A.B. and Mittra, B. (2004). Effects of salt on growth, ion accumulation, photosynthesis and leaf anatomy of the mangrove, *Bruguiera parviflora*. *Trees Structure Function*, 18: 167-174.
- Parker, R., Flowers, T. Moore, A. and Harpham, N. (2006). An accurate and reproducible method for proteome profiling of the effects of salt stress in the rice leaf lamina. *Journal of Experimental Botany* 57:1109–1118.

- Parveen, N. and Ashraf, M. (2010). Role of silicon in mitigating the adverse effects of salt stress on growth and photosynthetic attributes of two maize (*Zea mays* L.) cultivars grown hydroponically. *Pakistan Journal of Botany* 42(3):1675-1684.
- Perveen, S., Shahbaz, M. and Ashraf, M. (2010). Regulation in gas exchange and quantum yield of photosystem II (PSII) in salt stressed and non-stressed wheat plants raised from seed treated with triacontanol. *Pakistan Journal of Botany* 42:3073-3081.
- Pessarakli, M. and Szabolcs, I. (2010). Soil salinity and sodicity as particular plant /crop stress factor. In Pessarakli, M. (Ed.) *Handbook of Plant and Crop Stress* (3rd edition). Florida: CRC Press, Taylor & Francis Publishing Group, 3-21pp.
- Pessarakli, M. (2016). *Handbook of Cucurbits, Growth, Cultural Practices, and Physiology*. Florida : CRC Press, Taylor & Francis Publishing Group, 574pp.
- Pinheiro, H.A., Silva, J.V. and Endres, L. (2008). Leaf gas exchange, chloroplastic pigments and dry matter accumulation in castor bean (*Ricinus communis* L.) seedlings subjected to salt stress conditions. *Indian Crop Production* 27:385-392.
- Prajapati, R.P., Kalariya, M., Parmar, S.K., and Sheth, N.R. (2010). Phytochemical and pharmacological review of *Lagenaria siceraria*. *Journal of Ayurveda and Integrative Medicine* 1(4):266–272.
- Qin, J., Dong, W. Y., He, K. N., Yu, Y., Tan, G. D., Han, L., Dong, M., Zhang, D., Li, A.Z., Wang, Z.L. and Zhang, Y.Y. (2010). NaCl salinity-induced changes in water status, ion contents and photosynthetic properties of *Shepherdia argentea* (Pursh) Nutt. seedlings. *Plant, Soil and Environment* 56(7):325–332.
- Rai, M., Pandey, S. and Kumar, S. (2008). Cucurbit research in India: A retrospect. In Pitrat, M. (Ed.) *Cucurbitaceae 2008, Proc.IXth EUCARPIA Mtg. on Genetics and Breeding of Cucurbitaceae*, 21–24 May. INRA, Avignon, France, 285-293pp.
- Rajendran, K., Tester, M. and Roy, S.J. (2009). Quantifying the three main components of salinity tolerance in cereals. *Plant, Cell & Environment* 32(3):237-249.
- Raul, L., Andres, O., Armado, L., Bernardo, M. and Enrique, T. (2003). Response to salinity of three grain legumes for potential cultivation in arid areas. *Soil Science and Plant Nutrition* 49(3):329–336.

- Raza, S.H., Athar, H.R. and Ashraf, M. (2006). Influence of exogenously applied glycinebetaine on the photosynthetic capacity of two differently adapted wheat cultivars under salt stress. *Pakistan Journal of Botany* 38:341-351.
- Richards, L.A. (1954). Diagnosis and improvement of saline and alkali soils. *Soil Science* 78(2):154.
- Rizwan, M., Ali, S. and Ibrahim, M. (2015). Mechanisms of silicon-mediated alleviation of drought and salt stress in plants: a review. *Environmental Science and Pollution Research* 22(20):15416–15431.
- Romero-Aranda, R., Soria, T. and Cuartero, S. (2001). Tomato plant-water uptake and plant-water relationships under saline growth conditions. *Plant Sci.*, 160: 265-272.
- Romero-Aranda, R., Juado, O. and Cuartero, S. (2006). Silicon alleviates the deleterious salt effect on tomato plant growth by improving plant water status. *Journal of Plant Physiology* 163:847-855.
- Rouphael, Y., Cardarelli, M., Rea, E. and Colla, G. (2012). Improving melon and cucumber photosynthetic activity, mineral composition, and growth performance under salinity stress by grafting onto Cucurbita hybrid rootstocks. *Photosynthetica* 50(2):180-188.
- Sabir, P., Ashraf, M., Hussain, M. and Jamil, A. (2009). Relationship of photosynthetic pigments and water relations with salt tolerance of proso millet (*Panicum miliaceum* L.) accessions. *Pakistan Journal of Botany* 41:2957-2964.
- Sahebi, M., Hanafi, M.M., Siti Nor Akmar, A., Rafii, M.Y., Azizi, P., Tengoua, F., Nurul Mayzaitul Azwa, J., Shabanimofrad, M. (2015). Importance of silicon and mechanisms of biosilica formation in plants. *BioMed Research International* 2015:16.
- Sairam, R.K., Rao, V.K. and Srivastava, G.C. (2002). Differential response of wheat genotype to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration. *Plant Science* 163:1037-1046.
- Santos, C., Azevedo, H. and Caldeira, G. (2001). In situ and in vitro senescence induced by KCl stress: nutritional imbalance, lipid peroxidation and antioxidant metabolism. *Journal of Experimental Botany* 52:351-360.

- Savvas, D., Giotis, D., Chatzieustratiou, E., Bakea, M. and Patakioutas, G. (2009). Silicon supply in soilless cultivations of zucchini alleviates stress induced by salinity and powdery mildew infections. *Environmental and Experimental Botany* 65:11–17.
- Shabala, S., Shabala, S., Cuin, T.A., Pang, J., Percey, W., Chen, Z., Conn, S., Eing, C. and Wegner, L.H. (2010). Xylem ionic relations and salinity tolerance in barley. *The Plant Journal* 61(5):839–853.
- Shafieizargar, A., Awang, Y., Ajamgard, F., Juraimi, A.S., Othman, R., and Ahmadi, A.K. (2015). Assessing five citrus rootstocks for NaCl salinity tolerance using mineral concentrations, proline and relative water contents as indicators. *Asian Journal of Plant Sciences* 14(1):20–26.
- Shagufta, S., Sobia, N., Ashraf, M. and Nudrat, A. (2013). Salt stress affects water relations, photosynthesis, and oxidative defense mechanisms in *Solanum melongena* L. *Journal of Plant Interactions* (8)1:85-96.
- Shannon, M.C. and Grieve, C.M. (1999). Tolerance of vegetable crops to salinity. *Scientia Horticulturae* 78:5-38.
- Sharpley, A., Meisinger, N., Power, J.F. and Suarez, D.L. (1992). Root extraction of nutrients associated with long-term soil management. In: Stewart, B. (Ed.) *Advances in Soil Science*, vol. 19. Netherlands : Springer, 151 pp.
- Shi, Y., Wang, Y.C., Flowers, T.J. and Gong, H.J. (2013). Silicon decreases chloride transport in rice (*Oryza sativa* L.) in saline conditions. *Journal of Plant Physiology* 170:847–853.
- Shu, S., Guo, S., Sun, J. and Yuan, L.Y. (2012). Effects of salt stress on the structure and function of the photosynthetic apparatus in *Cucumis sativus* and its protection by exogenous putrescine. *Physiologia Plantarum* 146:285–296.
- Sin Seng Huat Seeds (2017). Retrieved from <http://gallery.greeneagle.com.my/>
- Slabu, C., Zorb, C., Steffens, D. and Schubert, S. (2009). Is salt stress of faba bean (*Vicia faba*) caused by Na⁺ or Cl⁻ toxicity? *Journal of Plant Nutrition and Soil Science* 172:644–650.
- Sommer, M., Kaczorek, D., Kuzyakov, Y. and Breuer, J. (2006). Silicon pool and fluxes in soils and landscapes—a review. *Journal of Plant Nutrition and Soil Science* 169:310–329.
- Stepian, P. and Klobus, G. (2006). Water relations and photosynthesis in *Cucumis sativus* L. leaves under salt stress. *Biologia Plantarum* 50(4):610-616.

- Szabados, L. and Savoure, A. (2010). Proline: a multifunctional amino acid. *Trends in Plant Science* 15(2):89-97.
- Tayebimeigooni, A., Awang, Y., Mahmood, M., Selamat, A. and Wahab, Z. (2012). Leaf water status, proline content, lipid peroxidation and accumulation of hydrogen peroxide in salinized Chinese kale (*Brassica alboglabra*). *Journal of Food, Agriculture and Environment* 10(2):371–374.
- Taylor, L. (2002). *Technical Data Report for Bitter melon (Momordica charantia) Herbal Secrets of the Rainforest* (2nd edition). Austin: Sage Press.
- Teakle, N.L. and Tyerman, S.D. (2010). Mechanisms of Cl^- transport contributing to salt tolerance. *Plant, Cell and Environment* 33:566–589.
- Travakkoli, E., Rengasamy, P. and McDonald, K. (2010). High concentrations of Na^+ and Cl^- ions in soil solution have simultaneous detrimental effects on growth of faba bean under salinity stress. *Journal of Experimental Botany* 61(15):4449-4459.
- Troncoso, A., Matte, C., Cantos, M., and Lavee, S. (1999). Evaluation Oalt Tolerance of in Vitro-Grown Grapevine Rootstock Varieties. *Vitis* 38(2):55-60.
- Tuna, A.L., Kaya, C., Higgs, D., Murillo-Amador, B., Aydemir, S., Girgin, A.R. (2008). Silicon improves salinity tolerance in wheat plants. *Environmental and Experimental Botany* 62:10–16.
- United Nations, Department of Economic and Social Affairs, Population Division (2015). *World Population Prospects: The 2015 Revision, Key Findings and Advance Tables*. Working Paper No. ESA/PWP.241.
- Vahdati, K. and Leslie, C. (Eds.).(2013). *Abiotic Stress - Plant Responses and Applications in Agriculture*. Retrieved from <https://www.intechopen.com/books/abiotic-stress-plant-responses-and-applications-in-agriculture>
- Vaughan, J.G. and Geissler, C.A. (2009). *The new oxford book of food plants 2nd edition*. Oxford : Oxford University Press, 249pp.
- Vijayvargiya, S. and Kumar, A. (2011). *Influence of salinity stress on plant growth and productivity: salinity stress influences on plant growth*. Germany: Lap Lambert Academic Publishers, 180 pp.

- Wanjogu, S.N., Muya, E.M., Gicheru, P.T. and Waruru, B.K. (2001). *Soil degradation: Management and rehabilitation in Kenya*. Proceedings of the FAO/ISCW expert consultation on Management of Degraded Soil in Southern and Eastern Africa (MADS-SEA) 2nd Networking meeting, Pretoria, South Africa, PR102-113.
- White, P.J. and Broadley, M.R. (2001). Chloride in soils and its uptake and movement within the plant: a review. *Annals of Botany* 88:967–988.
- Wong, N.C., Lee, B.S., Yuen, P.M., Wan Abdullah, W.Y and Mohd Ridzuan Saad. (2002). Effects of continuous mono-cropping of chrysanthemum flowers under rain-shelter on soil quality. In Shamshuddin, J., Hamdan, J. and Samsuri, A.W. (Eds.) *Sustainable Land Mangement*. Malaysian Society of Soil Science, 236-252 pp.
- Yadav, S., Irfan, M., Ahmad, A. and Hayat, S. (2011). Causes of Salinity and Plant manifestations to Salt Stress; a Review. *Journal of Environmental Biology/Academy of Environmental Biology, India* 32(5):667.
- Yang, Y., Wang, L., Tian, J., Li, J., Sun, J. and He, L. (2012). Proteomic study participating the enhancement of growth and salt tolerance of bottle gourd rootstock-grafted watermelon seedlings. *Plant Physiology and Biochemistry* 58:54–65.
- Yetisir, H. and Uygur, V. (2009). Plant growth and mineral element content of different gourd species and watermelon under salinity stress. *Turkish Journal of Agricultural Forestry* 33:65-77.
- Yin, L., Wang, S., Tanaka, K., Fujihara, S., Itai, A. and Deng, S. (2015). Silicon-mediated changes in polyamines participate in silicon-induced salt tolerance in *Sorghum bicolor* L. *Plant Cell Environment* 39(2):245-58.
- Yin, L.N., Wang, S.W., Li, J.Y., Tanaka, K. and Oka, M. (2013). Application of silicon improves salt tolerance through ameliorating osmotic and ionic stresses in the seedling of *Sorghum bicolor*. *Acta Physiologiae Plantarum* 35:3099-3107.
- Zhu, J., Bie, Z. and Li, Y. (2008). Physiological and growth responses of two different salt-sensitive cucumber cultivars to NaCl stress, *Soil Science and Plant Nutrition* 54(3):400-407.
- Zhu, J.K. (2001). Plant salt tolerance. *Trends in Plant Science* 6(2):66–71.
- Zhu, Y. and Gong, H. (2014). Beneficial effects of silicon on salt and drought tolerance in plants. *Agronomy for Sustainable Development* 34: 455-472.

Zhu, Z., Wei, G., Li, J., Qian, Q. and Yu, J. (2004). Silicon alleviates salt stress and increases antioxidant enzymes activity in leaves of salt-stressed cucumber (*Cucumis sativus* L.). *Plant Science* 167:527–533.



LIST OF PUBLICATIONS

- Najib, O.G., Yahya, A. and Firdaus, I. (2016). Assessing four *Cucurbitaceae* species for salt-tolerant rootstock using plant growth parameters as indicators. *National Seed Symposium, 23-24 August 2016, Sepang, Poster Presentation.*
- Najib, O.G., Yahya, A. and Firdaus, I. (2017). Assessing Salinity Tolerance Level of Four *Cucurbitaceae* Species Using Growth and Mineral Ion Content (Na^+ and Cl^-) as Indicators. *Malaysian Society of Plant Physiology Conference, 21-23 August 2017, Johor Bahru, Poster Presentation.*
- Najib, O.G., Yahya, A. and Firdaus, I. (2017). Impact of Silicon in Alleviating Salinity Stress in *Cucurbitaceae* Species. *MARDI Science and Technology Exhibition MSTE, 18-19 Sept 2017, MAEPS Serdang, Poster Presentation.*
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