



UNIVERSITI PUTRA MALAYSIA

***RAPID DETECTION OF Ganoderma-INFECTED OIL PALMS BY
MICROWAVE ERGOSTEROL EXTRACTION***

MUNIROH BINTI MD SAAD

FP 2015 81



**RAPID DETECTION OF *Ganoderma*-INFECTED OIL PALMS BY
MICROWAVE ERGOSTEROL EXTRACTION**

By

MUNIROH BINTI MD SAAD

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirement for the Degree of Master of Science**

January 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purpose from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

RAPID DETECTION OF *Ganoderma*-INFECTED OIL PALMS BY MICROWAVE ERGOSTEROL EXTRACTION

By

MUNIROH BINTI MD SAAD

January 2015

Chairman : Associate Professor Zainal Abidin Mior Ahmad, PhD
Faculty : Agriculture

Basal stem rot (BSR) is a devastating disease to oil palm in Malaysia. Detection of BSR was based on foliar symptoms and presence of *Ganoderma* fruiting body. Enzyme-Linked Immunosorbent Assays-Polyclonal Antibody (ELISA-PAB) and Polymerase Chain Reaction (PCR) has been proposed as an early detection method for the disease. However, these techniques are complex, time consuming and have accuracy limitations. In plantations, census needs to be carried out regularly at 6 months interval. Therefore, a robust, rapid and reliable early detection method is highly desirable for large scale BSR monitoring. An ergosterol method was developed, whereby it correlated well with the degree of infection in oil palm. This current study was designed to develop a simpler, rapid and efficient ergosterol detection method with utility in the field using microwave assisted extracted (MAE) method. Pure *Ganoderma* mycelium (isolated from Gua Musang Felda) was used for optimizing the extraction method with different microwave settings based on microwave power level, temperature and combination of microwave power level and temperature available on the microwave model (Sharp Jet Convectional Grill, model TTAG A437 with capacity 1.5 cu. ft) with three different exposure time (10, 20 and 30 seconds). The microwave setting with highest ergosterol concentration extracted which is the combination of 70°C and medium high (MH) power with 30 seconds exposure time was used for the subsequent experiment involving *Ganoderma* inoculated germinated oil palm seeds, seedlings and infected field palms for validation of the protocol and also development of semi-quantitative thin layer chromatography (TLC) method. The extraction procedure involves extracting a small amount of *Ganoderma* mycelium and *Ganoderma*-infected oil palm stem tissues (0.4g and 1.0g respectively) suspended in low volumes of solvent followed by irradiation in a conventional microwave oven with the best combination of 70°C and MH power for 30 seconds, resulting in simultaneous extraction and saponification. Ergosterol was detected in all infected samples and not detected in healthy samples by thin layer chromatography

and high performance liquid chromatography with diode array detection. Healthy samples were described as palms apparently normal and free from disease compared to infected palms that showed appearance of foliar symptoms and fruiting body at the base of palm trunk or on the trunk of the fallen palm and also at palm stump. The method was particularly effective in extracting high yields of ergosterol ($22.65 \mu\text{g g}^{-1}$) from infected oil palm and enables rapid analysis of field samples on site allowing infected oil palms to be treated or culled very rapidly. Positive relationship was observed between ergosterol content and inoculation period, where ergosterol concentration increased directly with the increase of inoculation period from day 3 to week 28 ($1.04 \mu\text{g g}^{-1}$ and $28.22 \mu\text{g g}^{-1}$ respectively) in the inoculated oil palm seedlings. In germinated seeds, ergosterol concentration also increased with the increase in the inoculation period from 6 hours to 168 hours after inoculation ($0.96 \mu\text{g g}^{-1}$ and $8.24 \mu\text{g g}^{-1}$ respectively) which means the *G. boninense* mycelial mass colonizing the roots of the palms also increased. Ergosterol was detected as early as day three after inoculation in diseased oil palm seedlings. These results indicated the early establishment of *G. boninense* in the root tissues. This present study did not wash or surface sterilize the roots before extraction of ergosterol, therefore ergosterol can be detected earlier on the root tissues once the fungus attached to the root tissue surface. The result also showed that external disease severity can be observed as early as week eight after inoculation. As disease progresses, palms show a pale appearance, retarded growth and finally death. Besides that, TLC analysis also showed a good correlation with the high performance liquid chromatography (HPLC) quantification where the intensity of the spot produced by TLC analysis showed high intensity of spot for higher ergosterol concentration quantified by HPLC and vice versa. Therefore, a semi-quantitative TLC analysis can be applied for handling large amount of samples during field survey. Based on the present work, it can be concluded that MAE is an efficient method to be used in ergosterol detection where ergosterol acts as biomarker for detection of BSR disease. Besides, TLC analysis can be used for detection of ergosterol in field palms, as it is easier and can be carried out on site and suitable for large field survey during the census.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KAEDAH PANTAS MENGESAN KELAPA SAWIT YANG DIJANGKITI
DENGAN *Ganoderma* MENGGUNAKAN KAEDAH EKTRAK GELOMBANG
KETUHAR**

Oleh

MUNIROH BINTI MD SAAD

Januari 2015

Pengerusi : Profesor Madya Zainal Abidin Mior Ahmad, PhD
Fakulti : Pertanian

Reput pangkal batang (BSR) adalah penyakit yang merbahaya dalam industri kelapa sawit di Malaysia. Pengesanan penyakit BSR kelapa sawit yang disebabkan oleh *Ganoderma* adalah berdasarkan simptom pada daun dan pengeluaran jasad berbuah. Enzyme-Linked Immunosorbent Assays-Polyclonal Antibody (ELISA-PAB) dan polymerase chain reaction (PCR) telah dicadangkan sebagai kaedah pengesanan awal penyakit ini. Walaubagaimanapun, teknik-teknik ini adalah rumit, memerlukan masa yang lama dan mempunyai had-had ketepatan. Di ladang, bancian perlu dijalankan secara berkala dalam tempoh setiap 6 bulan. Oleh itu, kaedah pengesanan awal yang mudah, pantas dan berkesan adalah sangat wajar diaplikasikan untuk pemantauan penyakit BSR pada skala besar. Satu kaedah pengesanan ergosterol telah dibangunkan yang berkait rapat dengan tahap jangkitan *Ganoderma* pada kelapa sawit. Kajian semasa ini telah direka bertujuan untuk membangunkan kaedah pengesanan ergosterol yang lebih ringkas, cepat, dan efisien dengan kemudahan di ladang menggunakan kaedah ekstrak ketuhar gelombang mikro (MAE). Miselium tulen *Ganoderma* telah digunakan untuk mengoptimumkan kaedah pengekstrakan dengan menggunakan tetapan ketuhar gelombang mikro yang berbeza berdasarkan kuasa dan suhu gelombang mikro dan juga kombinasi kuasa dan suhu ketuhar gelombang mikro yang terdapat pada ketuhar gelombang mikro model (Sharp Jet Convectional Grill, model TTAG A437 dengan saiz 1.5 cu. ft) . Tetapan ketuhar gelombang mikro yang menghasilkan kepekatan ergosterol tertinggi iaitu kombinasi 70°C dan kuasa sederhana rendah, dipanaskan selama 30 saat telah digunakan untuk eksperimen berikutnya yang melibatkan inokulasi *Ganoderma* pada benih bercambah, anak benih kelapa sawit, dan tisu kelapa sawit untuk mengesahkan protocol dan juga pembangunan kaedah separa- kuantitatif thin layer chromatography (TLC). Prosedur pengekstrakan ini melibatkan pengekstrakan miselium

Ganoderma dan tisu batang kelapa sawit yang dijangkiti *Ganoderma* dalam kuantiti yang sedikit (0.4g dan 1.0g masing-masing). Tisu tersebut direndam di dalam metanol dengan aruhan sinaran di dalam ketuhar gelombang mikro konvensional dengan kombinasi 70°C dan kuasa sederhana tinggi selama 30 saat, menghasilkan pengekstrakan dan saponifikasi berlaku secara serentak. Ergosterol dikesan dalam semua sampel yang dijangkiti dan tidak dikesan pada sampel yang tidak dijangkiti dengan menggunakan teknik kromatografi TLC dan high performance liquid chromatography (HPLC) diikuti pengesanan pelbagai diod. Sampel yang tidak dijangkiti penyakit adalah sampel yang kelihatan normal dan bebas dari penyakit, berbanding kelapa sawit yang dijangkiti penyakit yang menunjukkan gejala pada daun dan pembentukan jasad berbuah pada dasar batang kelapa sawit atau pada batang kelapa sawit yang tumbang dan juga pada tunggul kelapa sawit. Kaedah ini berkesan dalam mengekstrak ergosterol dengan kepekatan yang tinggi (22.65 $\mu\text{g g}^{-1}$) daripada tisu kelapa sawit yang dijangkiti dan ini membolehkan sampel ladang dianalisis lebih pantas dan membenarkan kelapa sawit yang dijangkiti penyakit dirawat dengan cepat. Hubungan yang positif dapat dilihat antara kandungan ergosterol dan masa inokulasi, di mana kepekatan ergosterol meningkat selari dengan peningkatan masa inokulasi daripada hari ke tiga hingga minggu ke lapan selepas inokulasi (masing-masing 1.04 $\mu\text{g g}^{-1}$ dan 28.22 $\mu\text{g g}^{-1}$) pada anak benih kelapa sawit. Pada anak benih bercambah kelapa sawit, kepekatan ergosterol juga meningkat selari dengan peningkatan masa inokulasi daripada enam jam hingga 168 jam selepas inokulasi (masing-masing 0.96 $\mu\text{g g}^{-1}$ dan 8.24 $\mu\text{g g}^{-1}$) bermaksud jumlah miselium *G. boninense* yang meliputi akar sawit juga meningkat. Ergosterol dapat dikesan seawal hari ke tiga selepas di inokulasi pada benih kelapa sawit. Keputusan ini menunjukkan kehadiran awal *G. boninense* pada tisu akar. Di dalam kajian ini, tisu akar tidak dicuci atau di steril sebelum di ekstrak, jadi ergosterol dapat dikesan lebih awal pada tisu akar setelah kulat melekat pada permukaan tisu akar. Hasil kajian ini juga menunjukkan skala tahap penyakit luaran dapat dilihat seawal lapan minggu selepas inokulasi. Semakin penyakit merebak, kelapa sawit menunjukkan gejala pucat, pertumbuhan terbantu, dan akhirnya mati. Selain itu, analisis TLC juga menunjukkan korelasi yang baik dengan pengiraan HPLC di mana keamatan tempok yang dihasilkan oleh analisis TLC menunjukkan keamatan paling tinggi bagi pengiraan HPLC yang paling tinggi dan sebaliknya. Oleh itu, analisis separa-kuantitatif TLC boleh digunakan untuk mengendalikan jumlah sampel yang besar dalam kajian lapangan. Berdasarkan kajian ini, kesimpulan yang boleh dibuat adalah, kaedah pengekstrakan menggunakan ketuhar gelombang mikro (MAE) adalah efisien dan boleh digunakan dalam pengesanan ergosterol di mana ergosterol berperanan sebagai penanda untuk pengesanan awal penyakit BSR. Selain itu, analisis TLC boleh digunakan untuk mengesan ergosterol di dalam tisu kelapa sawit di ladang kerana ia lebih mudah dan boleh dilaksanakan di-situ dan sesuai untuk kajian lapangan yang berskala besar.

ACKNOWLEDGEMENTS

‘In the nama of Allah, the most gracious, the most compassionate’

This project would not have been possible without the blessing from Allah the almighty and moral support from beloved people for the strength to complete this study.

I would like to express my sincere appreciation to my former advisor, Professor Sariah Meon, for her encouragement, guidance, support and critics during my study. Without her continued support and guidance, this study would not have been the same as presented here.

I am deeply grateful to my current advisor, Associate Professor Dr. Zainal Abidin Mior Ahmad and supervisory committee member, Dr. Nusaibah Syd Ali for their valuable advice, staff of Department of Plant Protection and Institute of Tropical Agriculture, Universiti Putra Malaysia for all their help in laboratory work and not to forget the staff of Institute of Bioscience (Microscopy Unit) for their kind assistance during SEM analysis. Special thanks to my lab mate Nurul Wahida Ramli who always helped and supported me in completion of this study.

Finally, I would like to extend my appreciation to my lovely parents, sister and brothers for their fully support and love. Moreover, the support, advice and help given by my dearest friends also highly appreciated.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master in Science. The members of the supervisory committee were as follows:

Zainal Abidin Mior Ahmad, Ph.D

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Nusaibah Syd Ali, Ph.D

Senior Lecturer,
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PH.D
Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirm that:

- This thesis is my original work;
- Quotation, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are full-owned by Universiti Putra Malaysia, as according to the U niversiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtain from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the university Putra Malaysia (Graduate studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: Date:

Name and Matric No.:

Declaration by Members of Supervisory Committee

This is confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to

Signature:

Signature:

Name of
Chairman of
Supervisory
Committee: Assoc. Prof. Dr Zainal Abidin
Mior Ahmad

Name of
member of
Supervisory
Committee: Dr. Nusaibah Syd Ali



TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	vii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xvii

CHAPTER

1	INTRODUCTION	1
2	LITERATURE REVIEW	
2.1	Basal Stem Rot (BSR) in oil palm	3
2.1.1	Causal Agent	4
2.1.2	Status of BSR disease in oil palm in Malaysia	4
2.2	Disease epidemiology	
2.2.1	Mycelial contact	4
2.2.2	Ganoderma Basidiospore	5
2.3	Disease Symptoms	
2.3.1	External Symptoms	6
2.3.2	Internal Symptoms	6
2.4	Current Management of BSR	
2.4.1	Cultural Control	7
2.4.2	Chemical Control	8
2.4.3	Biological Control	8
2.4.4	Resistance Variety	9
2.5	Detection of BSR	10
2.6	Ergosterol in Fungal Membrane	11
2.7	Ergosterol Biosynthesis Pathway	13
2.8	Ergosterol as Biomarker	15
2.9	Extraction, Detection and Quantification of Ergosterol	16
3	DEVELOPMENT OF A SIMPLE, RAPID, AND RELIABLE ERGOSTEROL EXTRACTION METHOD FROM <i>GANODERMA</i> AND <i>GANODERMA</i>-INFECTED TISSUE USING MICROWAVE-ASSISTED EXTRACTION (MEA)	
3.1	Introduction	18
3.2	Materials and Methods	

3.2.1	Mycelial Culture of <i>Ganoderma</i>	19
3.2.2	Ergosterol Extraction Using Microwave	22
3.2.3	Comparison Between Different Methods of Ergosterol Extraction	23
3.2.4	Detection and Quantification of Ergosterol	24
3.2.5	Statistical Analysis	25
3.3	Results	
3.3.1	Identification of Mycelial Culture of <i>Ganoderma</i>	26
3.3.2	Ergosterol Extraction using Microwave Assisted Extraction (MAE)	27
3.3.3	Comparison Between Different Methods of Ergosterol Extraction	34
3.4	Discussion	37
4	RELATIONSHIP BETWEEN <i>GANODERMA</i>, GROWTH, ERGOSTEROL CONCENTRATION, AND BSR DEVELOPMENT; AND VALIDATION OF MAE METHOD	
4.1	Introduction	39
4.2	Materials and Methods	
4.2.1	Germinated Seeds	40
4.2.2	Oil Palm Seedlings	44
4.2.3	Infected Oil Palm Field Tissue	49
4.2.4	Statistical Analysis	49
4.3	Results	
4.3.1	Germinated Seeds	52
4.3.2	Oil Palm Seedling	59
4.3.3	Infected Oil Palm Field Tissue	64
4.4	Discussion	66
5	SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	70
	REFERENCES	72
	APPENDICES	88`
	BIODATA OF STUDENT	101
	PUBLICATION	102

LIST OF TABLES

Table		Page
3.1	Comparison between different extraction methods of ergosterol from 0.4 g fresh weight of <i>Ganoderma</i> mycelium.	35
3.2	Comparisons of efficiency of ergosterol extraction based on MAE and NAE method	36
4.1	Description of external BSR symptoms based on disease category Level	50



LIST OF FIGURES

Figure		Page
2.1	Structure of an ergosterol molecule	12
2.2	Sterol Biosynthesis Pathway	14
3.1	Pure culture of 14 days old <i>Ganoderma boninense</i> maintained on malt extract agar, (a) Top view; (b) Bottom view	20
3.2	14 days old <i>Ganoderma</i> mycelial growth in malt extract broth	20
3.3	PCR amplification of 14 days old <i>Ganoderma</i> mycelial culture. M: Marker; L1-L4: amplified band of 14 days old <i>Ganoderma</i> mycelial culture.	26
3.4	(a) TLC analysis effect of microwave temperature on 0.5 g fresh weight of <i>Ganoderma</i> mycelium extracted using MAE method. Lanes 1-3, 40°C (10, 20 & 30 sec); lanes 4-6, 70°C (10, 20 & 30 sec), lane 7, Ergosterol standard. (b) Ergosterol concentration from 0.5 g fresh weight of <i>Ganoderma boninense</i> mycelium with various microwave temperature and exposure time. (40=40°C and 70=70°C). Bars \pm SE (standard error) of triplicate determinations.	28
3.5	(a) TLC analysis on effect of microwave power on 0.5 g fresh weight of <i>Ganoderma</i> mycelium extracted using MAE method. Lanes 1, Ergosterol Standard; 2-4, Medium high power (10, 20 & 30 sec); lanes 5-7, Medium power (10, 20, & 30 sec); lanes 8-10, High power (10, 20, & 30 sec). (b) Ergosterol concentration from 0.5 g fresh weight of <i>Ganoderma boninense</i> mycelium with various microwave power and exposure time. (M=Medium, MH= Medium high, H=High). Bars represent \pm SE (standard error) of triplicate determinations.	29
3.6	(a) TLC analysis on effect of combination temperature and microwave power on 0.5 g fresh weight of <i>Ganoderma</i> mycelium extracted using MAE method. Lanes 1, Ergosterol standard; lanes 2-4, 40°C & Medium high power (10, 20, & 30 sec); lanes 5-7, 70°C & medium high power (10, 20, & 30 sec); lanes 8-10, 40°C & medium power (10, 20, & 30 sec); 11-13, 70°C & medium power (10, 20, & 30 sec). (b) Ergosterol Concentration from 0.5 g fresh weight of <i>Ganoderma boninense</i> mycelium with various microwave temperatures, power levels and exposure times. (M40= Medium, 40°C, M70= Medium, 70°C, MH40= Medium high, 40°C, MH70= Medium high, 70°C). Bars represent \pm SE (standard error) of triplicate determinations	30

3.7	(a) TLC analysis of different weight of <i>Ganoderma</i> mycelium. Lanes: 1, Ergosterol standard; 2, 0.025g; 3, 0.05g; 4, 0.1g; 5, 0.2g; 6, 0.4g; 7, 0.6g; 8, 0.8g; 9, 1.0g. (b) Effect of sample weights of <i>Ganoderma boninense</i> mycelium on ergosterol concentration extracted using MAE method with microwave setting of medium high power and 70°C, for 30 seconds. Bars represent \pm SE (standard error) of triplicate determinations.	32
3.8	(a) TLC analysis of different weight of infected oil palm tissues. Lanes: 1, Ergosterol standard; 2, 0.25g; 3, 0.5g; 4, 1.0g; 5, 1.5g ; 6, 2.0g ; 7, 2.5g; 8, 3.0g; 9, 3.5g; 10, 4.0g. (b) Effect of infected oil palm tissue sample weight on ergosterol concentration extracted using MAE method with microwave setting of medium high power and 70°C, for 30 seconds. Bars represent \pm SE (standard error) of triplicate determinations.	33
3.9	TLC analysis of different methods of ergosterol extraction. Lane 1; Ergosterol standard. Lane 2-6; MAE method. Lane 7-11; NAE method. Lane 12-16; USE method. As can be determined an approximate concentration of ergosterol can be obtained from the size of the spots.	35
4.1	(a) Malt Extract Agar slant containing 14 day old <i>Ganoderma</i> culture; (b) Germinated seed inoculated on 14 day old <i>Ganoderma</i> culture	41
4.2	Formation of brown color zone after 7 days of inoculated roots plated on the GSM media as positive indication of the presence of <i>Ganoderma</i>	42
4.3	Rubber wood block in polypropylene bag fully colonized by <i>G. boninense</i> mycelium after four weeks of incubation.	45
4.4	Inoculation procedure of oil palm seedling with <i>G. boninense</i> colonized RWB. (a) Fully colonized RWB in the polybag. (b) Roots placed in close contact with RWB. (c) Roots contact with RWB. (d) RWB covered with 3 kg of soil mixture.	47
4.5	Visual assessment of external symptoms of healthy and BSR infected palms. (a) Healthy palms; (B) category A. Arrow shows presence of basidiocarp; (C) category B. Arrow shows presence of frond skirting; (D) Category C of infection shows fallen palm.	51
4.6	Percentage of infection on <i>Ganoderma</i> selective media after inoculation.	52

4.7	Ergosterol detection from uninoculated and inoculated germinated seeds by TLC. Ergosterol standard: lane 1, Uninoculated seedlings: lane 2-4, Inoculated seedlings (6, 12, 24, 48, 72, 96, 120, 144 and 168 hrs): lane 5-13.	53
4.8	Ergosterol concentration of the germinated seeds in un-inoculated and inoculated seeds. Bars represent SE (standard error) of triplicate determinations.	54
4.9	Relationship of inoculation period and ergosterol concentration of oil palm germinated seeds.	55
4.10	PCR amplification of inoculated and non-inoculated germinated seeds. L1, non-inoculated germinated seed; L2-L10, inoculated germinated seeds (6, 12, 24, 48, 72, 96, 120, 144, 168 hours); M: 100bp Marker	56
4.11	Comparison between inoculated and non-inoculated root (A) non-inoculated root, (B) 6 hrs after inoculation, (C) 24 hrs after inoculation, (D) 48 hrs after inoculation with <i>Ganoderma</i> culture.	58
4.12	Ergosterol detection from uninoculated and inoculated oil palm seedlings by TLC. Ergosterol standard; lane 1, Uninoculated seedlings; lane 3-6 and Inoculated seedlings (day 3, 7, 14, week 4, 12, 16, 20, 24, 28): lane 7-16.	59
4.13	Ergosterol concentration and disease severity percentage in inoculated seedlings. Bars represent SE (standard error) of triplicate determinations. (D= day; W= week; DS= disease severity).	60
4.14	Relationship of ergosterol concentration and internal disease severity of oil palm seedling from <i>G. boninense</i> .	61
4.15	Relationship of ergosterol concentration and external disease severity of oil palm seedling from <i>G. boninense</i> .	61
4.16	Comparison between healthy root and damage root: (A) Healthy root. (B) Arrow shows root with the presence of white mycelium 3 days after inoculation. (C) Arrow shows rotting root. (D) Arrow shows lesion of root 16 weeks after inoculation.	62
4.17	Comparison between healthy palm (left) and infected palm (right) 16 weeks after inoculation. Infected palms shows stunted growth.	63
4.18	Comparison between (A) healthy bole and (B) infected bole 20 weeks after inoculation.	63

- 4.19 TLC analysis of ergosterol from field palms based on different level of external BSR infection. Lane 1: Ergosterol standard, Lane2-4: Healthy palms, Lane 5-7: Category A palms, Lane 8-10: Category B palms, Lane 11-13: Category C palms. (Healthy= (Palms apparently normal and free from disease; Category A= free from foliar symptom with appearance of fruiting body at the base of trunk; Category B= appearance of foliar symptoms and presence of fruiting body at base of trunk; Category C= fruiting body appearance at the base of palm trunk or on the trunk of the fallen palm and also at palm stump). 64
- 4.20 Average ergosterol ($\mu\text{g g}^{-1}$) compared to degree of BSR external symptoms (Healthy= (Palms apparently normal and free from disease; Category A= free from foliar symptom with appearance of fruiting body at the base of trunk; Category B= appearance of foliar symptoms and presence of fruiting body at base of trunk; Category C= fruiting body appearance at the base of palm trunk or on the trunk of the fallen palm and also at palm stump). Bars represent \pm SE (standard error) of triplicate determination. 65

LIST OF ABBREVIATIONS

AE	Alkaline Extraction
bp	Base Pair
BSR	Basal Stem Rot
C	Carbon
CRD	Complete Randomize Design
DNA	Deoxyribonucleic acid
DS	Disease Severity
FW	Fresh Weight
GIS	Geographical Information System
GSM	<i>Ganoderma</i> Selective Media
Ha	Hectare
HPLC	High Performance Liquid Chromatography
hrs	Hours
IPP	Isopentenyl Diphosphate
ITS	Internal Transcribe Spacer
LSD	Least Significant Different
MAE	Microwave Assisted Extraction
MEA	Malt Extract Agar
MEB	Malt Extract Broth
MH	Medium High
Min	minute
NADPH	Nicotinamide Adenine Dinucleotide Phosphate
NAE	Non-alkaline Extraction
nm	Nanometer
PCR	Polymerase Chain Reaction
PIRG	Percentage of Radial Growth
RAPDs	Random Amplified Polymorphic DNA
Rf	Retention Value
RNA	Ribonucleic Acid
RWB	Rubber Wood Block
SE	Standard error
Sec	Second
SEM	Scanning Electron Microscopy
SFE	Supercritical Fluid Extraction
TLC	Thin Layer Chromatography
USE	Ultrasonicator Extraction Method
UV	Ultraviolet

CHAPTER 1

INTRODUCTION

Oil palm (*Elaeis guineensis*) is a monocotyledon in the family Arecaceae (formerly Palmae) within the subfamily Coccoideae (Corley and Tinker, 2003). It is a major crop that grows in the tropical areas, especially in Southeast Asia. Palm oil is used worldwide for the production of food, cosmetics, pharmaceuticals, biodiesel and oleo chemical industry, where it is being used for making soaps and detergents (Kalam and Masjuki, 2002; Corley and Tinker, 2003; Turner *et al.*, 2008). Oil palm industry contributes to the Malaysian economy and also in the development of country's rural areas (Chin, 2008). In Malaysia, cultivation of oil palm has increased year by year with 1.5 million hectares (ha) in the 1985 to 5.39 million ha in 2014 (MPOB, 2014).

Oil palm is subjected to numerous devastating diseases such as basal stem rot (BSR), vascular wilt, spear rot-bud rot, sudden wither and red ring (Corley and Tinker, 2003). However, BSR is the major disease encountered by Malaysian palms which is caused by *Ganoderma* species (Idris *et al.*, 2011). Several attempts have been made to control BSR using various control methods; however until now none of the methods gave good control of *G. boninense* infection in established oil palm plantation due to late detection of *Ganoderma*. In plantation, census needs to be done regularly, practically at 3-6 months interval. To date, a practicable early detection method of the disease is still lacking. Basal stem rot is detected based on foliar associated symptoms and production of basidiomata at the base of infected stem. However, by the time visible symptoms appear, the palms are already at a serious stage and usually half of the basal tissues have been killed by the fungus (Idris, 2009). Enzyme-linked immunosorbent assays-polyclonal antibody (Idris and Rafidah, 2008) as well as PCR based techniques involving specific *Ganoderma* primers (Bridges *et al.*, 2000; Utomo and Niepold, 2000; Yamoaka, *et al.*, 2000) has been proposed as early detection method of the disease. However these methods are complicated and time consuming for early detection of the disease in oil palm fields. Moreover, there are some limitations with PCR technique which require to be addressed before applying for detection of *Ganoderma* (Paterson, 2007a; Paterson *et al.* 2008; Paterson and Lima, 2009).

Ergosterol is a primary sterol in cell membranes of filamentous fungi and is either absent or present in a minor component in the majority of higher plants (Madonna *et al.*, 2001). First data published on the use of ergosterol analysis as a diagnostic method to detect BSR support the view that ergosterol has the utility for the detection of BSR in oil palm (Mohd Aswad *et al.*, 2011). Ergosterol is specific to fungi and indicates live fungal biomass. Therefore, Parkinson and Coleman (1991) reported that ergosterol assay is commonly considered to be the most promising tools for detection and quantification of

fungal biomass. Parsi and Gorecki, (2006) also reported that the detection of ergosterol as fungal biomarker could be considered to be the method of choice

Previous study used non-alkaline extraction (NAE), alkaline extraction (AE), and ultrasonicator extraction (USE) methods for extraction of ergosterol. The organic solvent based method (classical) typically required large samples, large reagent volume, labor intensive, time consuming and also reported that AE and USE give low ergosterol concentration when compared to NAE method (Mohd Aswad *et al.*, 2011). Therefore, more reliable extraction methods are required for the extraction of ergosterol. Young (1995) has developed microwave assisted extraction (MAE) method for extraction of ergosterol which required smaller samples and reagent volume, more economical in term of chemicals used, and using convectional equipment (Domestic Microwave). Microwave assisted extraction is therefore more convenient than other methods in terms of time for sample preparation, cost and sample size. In addition, more samples can be extracted at one time and a suitable, simple, rapid, and reliable extraction method for ergosterol detection in field palms during census.

Thus, present study is undertaken to apply ergosterol as bio indicator of *Ganoderma* infection grown in field palms with simple extraction method. The aims of this study are (I) to optimize a simple and rapid ergosterol extraction method from *Ganoderma* infected tissue using MAE, (ii) to establish the relationship between *Ganoderma*, ergosterol concentration, and BSR development; and to validate the efficiency of MAE method for extraction of ergosterol and test the sensitivity of TLC analysis for detection of ergosterol using artificially inoculated germinated seeds and seedlings; and infected field palm samples.

REFERENCES

- Abdul Razak, J., Ahmad, H., Ramdhan, K., Idris, A.S., Abdul Rahim, S., Aminul, R. and Fauzi, I. (2004). Mechanical Trunk Injection for Control of Ganoderma MPOB Information Series. MPOB TT No. 215.
- Abdullah, R., Yeun L.H., Rashdan, M.M., Leaw, C.L., Alizah, Z., Yap, S.P. And Wee, Y.H.(1999). Genetically modified oil palm for pest and disease control. In: Plant Protection in the Tropics: Tropical plant protection in the information age. (15-18th March, 1999, Kuala Lumpur, Malaysia). Malaysian Plant Protection Society. Sivapragasam, A.; Ismail, A.A.; Sidam, A.K.; Cheah, U.B.; Chung, G.F.; Chia, T.H.; Dzolkhifli, O.; Ho, T.H.; Hussan, A.K.; Lee, S.S., Lim, J.L.; Lum, K.Y.; Mohamed, S.; Nathan, G.; Ong, C.A.; Vijaysegaran, S. And Zainal Abidin, M.A. Eds., 1999, P. 47-50.
- Abdullah, S.A., (2003). Fragmented forest in tropical landscape: the case of the state of Selangor, Peninsular Malaysia. *Journal of Environment Science* 15, 267–270.
- Ariffin, D. and Idris, A.S. (1991). A selective medium for the isolation of *Ganoderma* from diseased tissues. In: Yusof, B. *et al.* (eds), Proceedings of the 1991 PORIM International Palm Oil Conference-Progress, Prospects and Challenges Towards the 21st Century-Module 1, Agriculture. Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia, pp 517-519.
- Ariffin, D., Idris A.S. and Singh, G. (2000). Status of *Ganoderma* in oil palm. In: Flood, J. *et al.*, (eds), *Ganoderma* Diseases of Perennial Crops. CAB International Publishing, Wallingford, UK, pp 249-666.
- Ariffin, D., Idris, A.S. and Abdul Halim, H. (1989). Significance of the black line within oil palm tissue decay by *Ganoderma boninense*. *Journal of Elaeis*, 1:11-16.
- Aust, S.D. and J.D. Stahl, (1998). Biodegradation of Dioxin and Dioxin-Like Compounds by White-Rot. Fungi. In Biodegradation of Dioxin and Furans. Rolf-Michael Wittich (Ed.). Springer-Verlag and R.G. Landes Company, pp: 61-73.
- Axelsson, B-O., Saraf, A. and Larsson, L. (1995). Determination of ergosterol in organic dust by gas chromatography–mass spectrometry. *Journal of Chromatography B*, 666:77–84.
- Azahar TM, Mustapha CJ, Mazliham S, Patrice B (2011). Temporal analysis of basal stem rot disease in oil palm plantations: An analysis on peat soil. *Int J Eng Tech*. 3:96-101

- Barajas-Aceves, M., Hassan, M., Tinoco, R., Vazquez-Duhalt, R., (2002). Effect of pollutants on the ergosterol content as indicator of fungal biomass. *Journal of Microbiological Methods* 50, 227– 236.
- Barrett-Bee, K. and Ryder, N. (1992). Biochemical aspects of ergosterol biosynthesis inhibition. In : Sutcliffe JA, Georgopadakou NH eds. *Emerging Targets in antibacterial and antifungal chemotherapy*. New York: Chapman and Hall. Pp 410-436.
- Benhamou, N., Chet, I. (1996). Parasitism of sclerotia of *Sclerotium rolfsii* by *Trichoderma harzianum*: ultrastructural and cytochemical aspects of the interaction. *Phytopathology* 86, 405–416.
- Bindler, G.N., Piadé, J.J. and Schulthess, D. (1988) Evaluation of selected steroids as chemical markers of past or presently occurring fungal infections on tobacco. *Beitrage Zur Tabakforschung International* 14, 127–134.
- Bjurman, J., (1999). Fungal and microbial activity in external wooden panels as determined by finish, exposure, and construction techniques. *Int. Biodeterior. Biodegrad.* 43, 1–5.
- Breton, F., Hasan, Y., Hariadi, Lubis Z. and de Franqueville H. (2005). Characterization of parameters for the development of an early screening test for Basal Stem Rot tolerance in oil palm progenies. In: *Proceedings of Agriculture, Biotechnology and Sustainability Conference. Technological Breakthroughs and Commercialization-The Way Forward, PIPOC 2005 MPOB International Palm Oil Congress, 25-29 September 2005, Kuala Lumpur, Malaysia.*
- Breton, F., Hasan, Y., Hariadi, S., Lubis, Z., de Franqueville, H. (2006). Characterization of parameters for the development of an early screening test for basal stem rot tolerance in oil palm progenies. *Journal of Oil Palm Research (Special issue, April 2006)*, 24–36.
- Bridge, P.D., Ogrady, E.B., Pilotti, C.A. and Sanderson, F.R. (2000). Development of Molecular Diagnostics for the Detection of *Ganoderma* Isolates Pathogenic to Oil Palm. In: Flood, J. *et al.* (eds), *Ganoderma Diseases of Perennial Crops*. CAB International Publishing, Wallingford, UK, pp 225-234.
- Castro Moretzsohn de, M., Bragagnolo, N. and Toledo Valentini, S. (2002). The relationship between fungi growth and aflatoxin production with ergosterol content of corn.
- Chander, K., Dyckmans, J., Joergensen, R.G., Meyer, B., Raubuch, M., (2001). Different sources of heavy metals and their longterm effects on soil microbial properties. *Biology and Fertility of Soils* 34, 241– 247.

- Chin, P.F.K. (2008). Malaysian efforts in developing responsible practises in the Palm Oil Industry, edited version of the keynote address by the Malaysian Minister of Plantation Industries and Commodities at the World Sustainable Palm Oil Conference, London, 15 September 2008. Published in the Global Oils & Fats Business Magazine .Vol. 5, Issue No. 4.
- Chong, K.P. (2010). *The role of phenolics in the interaction between oil palm and Ganoderma boninense the casual agent of basal stem rot*. PhD thesis, University of Nottingham.
- Chong, K.P., Lum, M.S., Foong, C.P., Wong, C.M.V.L. and Atong, M. (2011). First identification of *Ganoderma boninense* isolated from Sabah based on PCR and sequence homology. African Journal of Biotechnology 10: 14718-14723.
- Chung, G.F., (1991). Preliminary results on trunk injection of fungicides against *Ganoderma* basal stem rot in oil palm. In: Ariffin, D. and Sukaimi, J. (Eds.), Proceedings of *Ganoderma* workshop, Bangi, Selangor, Malaysia. Palm Oil Research Institute of Malaysia 81-97.
- Chung, G.F. (2005). Management of *Ganoderma* diseases in oil palm to minimise spreading in the fields. The Planter, 81 (957):765-773.
- Chung G.F. (2011). Management of ganoderma diseases in Oil palm plantations. Planter. 87(1022):325–339.
- Corley, R.H.V. and Tinker P.B. (2003). The Oil Palm. 4th edition. Oxford, UK: Blackwell Publishing.
- Darmono, T.W. (2000). *Ganoderma* in oil palm in Indonesia: current status and prospective use of antibodies for the detection of infection. In: Flood, J. et al., (eds), *Ganoderma* Diseases of Perennial Crops. CAB International Publishing, Wallingford, UK, pp 249-266.
- Daum G, Lees ND, Bard M, Dickson R. (1998). Cell biology and molecular biology of lipids of *Saccharomyces cerevisiae*. Yeast Biochemistry,14:1471–510.
- Dawson-Andoh, B.E. (2002). Ergosterol content as a measure of biomass of potential biological control in liquid cultures. Journal of Holz als Roh-und Werkstoff, 60: 115-117.
- Dharmaputra, O.S., H.S. Tjitrosomo and A.L. Abadi. (1989). Antagonistic effect of four fungal isolates to *Ganoderma boninense*, the causal agent of basal stem rot of oil palm. Journal of Biotropia 3: 41–49.
- Donald, W. and Mirocha, C. (1997) Chitin as a measure of fungal growth in stored corn and soybean seed. Cereal Chemical 54, 466–474.

- Doyle, J.J. and Doyle J.L., (1987). A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem. Bull.* 19:11-15.
- Durand-Gasselín T, Asmady H, Flori A, Jacquemard JC, Hayun Z, Breton F, De Franqueville H, (2005). Possible sources of genetic resistance in oil palm (*Elaeis guineensis* Jacq.) to basal stem rot caused by *Ganoderma boninense*—prospects for future breeding. *Mycopathologia* 159, 93–100.
- Farid C. and Giancarlo C. (2012). *Microwave-assisted Extraction for Bioactive Compounds: Theory and Practice*. New York: Springer Science & Business Media, 2012.
- Fischer, G. and Dott, W. (2002). Quality assurance and good laboratory practice in the mycological laboratory: compilation of basic techniques for the identification of fungi. *International Journal of Hygiene and Environment*. 205, 433–442.
- Flood, J, Hasan, Y, Turner, P.D. and O’Grady, E.B. (2000). The spread of *Ganoderma* from infective sources in the field and its implications for management of the disease in oil palm. In: Flood, J. *et al.* (eds), *Ganoderma Diseases of Perennial Crops*. CAB International Publishing, Wallingford, UK, pp 101– 112.
- Flood, J., Hasan, Y. and Foster, H. (2003). *Ganoderma* diseases of oil palm- an interpretation from Bah Lias Research Station. *The Planter* 78, 689–710.
- Frey, B., Buser, H-R. and Schüepp, H. (1992). Identification of ergosterol in vesicular-arbuscular mycorrhizae. *Biology Fertility Soils*, 13: 229-234.
- Frostegard, A., Baath, E. (1996). The use of phospholipid fatty acid analysis to estimate bacterial and fungal biomass in soil. *Biology and Fertility of soils*, 22, 59-65.
- Ganzler, K.; Salgo, A.; Valko, K. (1986). Microwave extraction. A novel sample preparation method for chromatography. *Journal of Chromatography* 371, 299-306.
- Garcia-Ayuso LE, Luque De, and Castro MD. (1999). A multivariate study of the performance of a microwave assisted Soxhlet extractor for olive seeds. *Analytica Chimica Acta*. 382.
- Garcia-Ayuso LE, Luque De and Castro MD (2001). Employing focused microwaves to counteract conventional soxhlet extraction drawbacks. *Trends in Analytical Chemistry*, 20: 28
- Gawrysiak-Witulska, M., Wawrzyniak, J., Ryniecki, A. and Perkowski, J. (2008). Relationship of ergosterol content and fungal contamination and assessment of technological quality of malting barley preserved in a metal silo using the near-ambient method. *Journal of Stored Production Research* 44:360–365.

- Gedye, R.; Smith, F.; Westaway, K.; Ali, H.; Baldisera, L.; Laberge, L.; Rousell, J. (1986). The use of microwave ovens for rapid organic synthesis. *Tetrahedron Letters* 27,279-282.
- George, S.T., Chung, G.F. and Zakaria, K. (1996). Updated Results (1990-1005) on trunk injection of fungicides of the control of Ganoderma basal stem rot. In: Ariffin, D, *et al.* (Eds.), Proceedings of the 1996 PORIM International Palm Oil Congress-Agriculture. Palm Oil Research Institute of Malaysia, Selangor, Malaysia 508-515.
- Gessner, M.O. (2005). Ergosterol as a measure of fungal biomass. In: Graça, M.A.S., Barlocher, F. and Gessner, M.O. (Eds.), *Methods to study litter decomposition: a practical guide*. 189- 196, Dordrecht, The Netherlands. *Springer*.
- Gessner M.O., Chauvet E. (1993). Ergosterol-to-biomass conversion factors for aquatic hyphomycetes. *Applied and Environmental Microbiology* 59:502–507.
- Gessner, M.O. and Newell, S.Y. (2002). Biomass, growth rate and production of filamentous fungi in plant litter. In: Hurst, C.J. *et al.*, (eds.), *Manual of Environmental Microbiology*, 2nd ed., ASM Press, Washington, DC, pp. 390-408.
- Gessner, M.O., Bauchrowitz, M.A., Escutier, M., (1991). Extraction and quantification of ergosterol as a measure of fungal biomass in leaf litter. *Microbial Ecology* 22, 285– 291.
- Gibbons, G.F., Mitropoulos, K.A. and Myant, N.B. (1982). *Biochemistry of cholesterol*. Elsevier Biomed Press, Amsterdam.
- Gong, P., Guan, X. and Witter, E. (2001). A rapid method to extract ergosterol from soil by physical disruption. *Applied Soil Ecology*, 17: 285-289.
- Grant, W.D. and West, A.W. (1986). Measurement of ergosterol, diaminopimelic acid and glucosamine in soil: evaluations indicators of microbial biomass. *Journal of Microbiological Methods*, 6: 47-53.
- Griffin, D.H. (1994). *Fungal physiology*. New York: Journal Wiley Publication.
- Griffiths, H.M., Jones, D.G., Akers, A., (1985). A bioassay for predicting the resistance of wheat leaves to *Septoria nodorum*. *Annals of Applied Biology* 107, 293– 300.
- Gutarowska, B., Zakowska, Z., (2002). Elaboration and application of mathematical model for estimation of mould contamination of some building materials based on ergosterol content determination. *International Biodeterioration and Biodegradation* 49, 299– 305.

- Hammel, K.E., Jensen, K.A., Jr. M.D. Mozuch, L.L. Landucci, M. Tien and E.A. Pease, (1993). Lignolysis by a Purified Lignin Peroxidase. *Journal of Biological Chemistry* 268(17): 12274-12281.
- Hasan, Y. and Turner, P.D. (1994). Research at BAH LIAS Research Station on basal stem rot of oil palm. In: Holderness, M. (Eds.), *Proceedings of the 1st International Workshop on Perennial Crop Diseases caused by Ganoderma*, 28 November- 2 December 1994. UPM, Serdang, Selangor, Malaysia 19-24.
- Hasan, Y. and Turner, P.D. (1998). The comparative importance of different oil palm tissues as infection sources for basal stem rot in replantings. *The Planter* 74, 119–35.
- Hasan, Y., Foster, H.L. and Flood, J. (2005). Investigations on the causes of upper stem rot (USR) on standing mature oil palms. *Mycopathologia* 159, 109–12.
- Henson, J.M. and French, R. (1993). The polymerase chain reaction and plant disease diagnosis. *Annual Review of Phytopathology* 31: 81-109.
- Ho, C.T. and Khairudin, H. (1997). Usefulness of soil moulding treatments in prolonging productivity of prime-aged *Ganoderma* infected palms. *The Planter*, Kuala Lumpur **73**: 239-244.
- Ho, Y.W. and Nawawi, A. (1986). *Ganoderma boninense* from basal stem rot of oil palm (*Elaeis guineensis*) in Peninsular Malaysia. *Pertanika Journal of Tropical Agricultural Science*, **8**: 425-428.
- Hoong, H.W. (2007). *Ganoderma* Diseases of Oil Palm In Sabah. *The Planter*, **83** (974): 299-313.
- Idris, A.S., Ariffin, D., Swinburne and Watt, T.A. (2000). The identity of *Ganoderma* species responsible for basal stem rot (BSR) disease of oil palm in Malaysia – Pathogenicity test. MPOB Information Series No. 103(77).
- Idris, A.S., Khushairi, A., Ismail, S. and Arifin, D. (2002). Selection for partial resistance in oil palm to *Ganoderma* basal stem rot. Seminar: elevating the national oil palm productivity and recent progress in the management of Peat and *Ganoderma*, Kuala Lumpur, Malaysia, 6–7 May, 2002, p. 11.
- Idris, A.S., Ismail, S., Ariffin, D., and Ahmad, H., (2003). Control of *Ganoderma*-Infected palm-development of pressure injection and field applications. MPOB Information Series No. 131.
- Idris, A.S., Kushairi, A., Ismail, S. and Ariffin, D. (2004). Selection for partial resistance in oil palm progenies to *Ganoderma* basal stem rot. *Journal of Oil Palm Research*, 16, 12-8.

- Idris, A.S., Kushairi, A., Ariffin, D. and Basri, M.W. (2006). Technique for inoculation of oil palm geminated seeds with *Ganoderma*. MPOB Information Series, MPOB TT No. 314, June 2006, Malaysian Palm Oil Board, Bangi, Selangor, Malaysia. 4pp.
- Idris, A.S. and Rafidah, A.R. (2008). Polyclonal antibody for detection of *Ganoderma*. MPOB Information Series 430, MPOB, Malaysia.
- Idris, A.S. (2009). Basal stem rot in Malaysia - Biology, Economic Importance, Epidemiology, Detection and Control. Paper presented at the International workshop on awareness, detection and control of oil palm devastating diseases, Kuala Lumpur, Malaysia. 6 November 2009.
- Idris, A.S., Mazliham, M.S. and Madihah, A.Z. (2009). Current technologies for detection of *Ganoderma* in oil palm. In: Non-edited Proceedings of the 2009 Agriculture, Biotechnology and Sustainability Conference (PIPOC), Malaysian Palm Oil Board (MPOB). Kuala Lumpur: 9-12 November 2009. pp. 81-99.
- Idris, A.S., Rajinder, S., Madihah, A.Z. and Wahid, M.B. (2010). Multiplex PCR-DNA kit for early detection and identification of *Ganoderma* species in oil palm. MPOB Information Series 531, MPOB, Malaysia.
- Idris, A.S., Mior, M.H.A.Z., Maizatul, S.M. and Kushairi, A. (2011). Survey on status of *Ganoderma* disease of oil palm. Proc. Of the PIPOC 2011 International Palm Oil Congress – Agriculture Conference. MPOB, Bangi, pp. 235-238.
- Izzati, M.Z. and Abdullah, F. (2008). Disease suppression in *Ganoderma*-infected oil palm seedling treated with *Trichoderma harnianum*. Plant Protection Science 44(3): 101-107.
- Johnson, B.N., McGill, W.B., (1990). Comparison of ergosterol and chitin as quantitative estimates of mycorrhizal infection and *Pinus contorta* seedling response to inoculation. Canadian Journal of Forest Research 20, 1125– 1131.
- Kalam, M.A. and Masjuki, H.H. (2002). Biodiesel from palm oil- an analysis of its properties and potential. Biomass Bioenergy. 23: 471-479.
- Karin Thevissen, Kathelijne K.A. Ferket, Isabelle E.J.A. François, Bruno P.A. Cammue (2003). Interactions of antifungal plant defensins with fungal membrane components . Science direct, 24 : 1705–1712.
- Kaspersson, A.(1986). The role of fungi in deterioration of stored feeds. PhD Thesis Report 31, Uppsala, Sweden: Department of Microbiology, Swedish University of Agricultural Sciences.

- Kaufmann, B., and Christen, P. (2002). Recent extraction techniques for natural products: microwave assisted extraction and pressurized solvent extraction. *Phytochemical Analysis*, 13(2), 105–113.
- Khairuddin, H. (1990). Basal stem rot of oil palm: incidence, etiology and control. Master of Agriculture Science thesis, Universiti Pertanian Malaysia, Selangor, Malaysia.
- Khairudin, H. (1991). Pathogenicity of three *Ganoderma* species on oil palm seedlings. *Journal of Perak Planters Association*. 43-49
- Larsen, T., Axelsen, J. and Ravn, H.W. (2004). *Journal of Chromatography A*, 1026, 301-304.
- Lee, C., Howarth, R.W., Howes, B.L., (1980). Sterols in decomposing *Spartina alterniflora* and the use of ergosterol in estimating the contribution of fungi to detrital nitrogen. *Limnology and Oceanography* 25, 290–303.
- Lim H., and Fong Y. (2005). Research on basal stem rot (BSR) of ornamental palms caused by basidiospores from *Ganoderma boninense*. *Mycopathologia*. 159(1):171–179.
- Lim, T.K., Chung, G.F. and Ko, W.H. (1992). Basal stem rot of oil palm caused by *Ganoderma boninense*. *Plant Pathology* 1: 147-152.
- Lim, K.H., Chuah, J.H. and Ho, C.H. (1993). Effect of soil heaping on *Ganoderma*-infected oil palm. In: Jalani et al., (eds) Proceeding of the 1993 PORIM International Palm Oil Congress 'Update and Vision' (Agriculture). 20-25 September, 1993. Palm oil Research Institute of Malaysia, Bangi, Selangor, Malaysia. Pp. 735-738.
- Liu, L., Kloepper, J.W. and Tuzun, S. (1995). Induction of systemic resistance in cucumber against bacterial angular leaf sport by plant growth promoting rhizobacteria. *Journal of Phytopathology*, 85: 843-847.
- Lopez-Avila, V., Young, R., Beckert, W.F. (1994). Microwave-assisted extraction of organic compounds from standard reference soils and sediments. *Anal. Chem.* 66 (7), 1097–1106.
- Madonna, A.J., Voorhees, K.J. and Hadfield, T.L. (2001). Rapid detection of taxonomically important fatty acid methyl ester and steroid biomarkers using in situ thermal hydrolysis/methylation mass spectrometry (THM-MS): implication for bioaerosol detection. *Journal of Analytical and Applied Pyrolysis* 61: 65-89.
- Malaysia Palm Oil Board, (2014). Oil Palm Planted Area By State As At December 2014. [Http://Bepi.Mpob.Gov.My/Index.Php/Statistics/Area/132-Area-2014/713-Oil-Palm-Planted-Area-Dec-2014.Html](http://Bepi.Mpob.Gov.My/Index.Php/Statistics/Area/132-Area-2014/713-Oil-Palm-Planted-Area-Dec-2014.Html)

- Mandal, V., Mohan, Y., Hemalatha, S. 2007. Microwave assisted extraction—an innovative and promising extraction tool for medicinal plant research. *Pharmacognosy Reviews* 1(1), 7-18.
- Mazliham, M.S. (2008). Design of a decision support system handling imperfect scattered data: application to early detection of *Ganoderma* fungus infection in oil palm trees. Ph.D thesis, Universite de La Rochelle, France, 185 pp.
- Miller, J.D., Laflamme, A.M., Sobol, Y., Lafontaine, P., Greenhalgh, R., (1988). Fungi and fungal products in some Canadian houses. *International Biodeterioration and Biodegradation* 24, 103–120.
- Mille-Lindblom, C., Von Wachenfeldt, E. and Tranvik, L.J. (2004). Ergosterol as a measure of living fungal biomass: persistence in environmental samples after fungal death. *Journal of Microbiological Methods*, 59: 253-262.
- Mille-Lindblom, C., Fischer, H. and Tranvik L.J. (2006). Antagonism between bacteria and fungi: substrate competition and a possible tradeoff between fungal growth and tolerance towards bacteria. 113: 233-242.
- Miller, J.D., Young, J.C., Trenholm, H.L., (1983). Fusarium toxins in field corn: I. Time course of fungal growth and production of deoxynivalenol and other mycotoxins. *Canadian Journal of Botany* 61,3080–3087.
- Miller, R.N.G., Holderness. M., Bridge, P.D., Chung, G.F. and Zakaria, M.H. (1999). Genetic diversity of *Ganoderma* in oil palm plantings. *Plant Pathology* 48, 595–603.
- Mingos, D.M.P., Baghurst, D.R. (1991). Applications of microwave dielectric heating effects to synthetic problems in chemistry. *Chemical Society Reviews* 20, 1-47.
- Mior, M.H, Idris, A.S., Wahid, O. and Kushairi, A.D. (2009). Spatial, Temporal and Hotspot Analysis of Basal Stem Rot Disease Caused by *Ganoderma*. In: Non-edited Proceedings of the 2009 Agriculture, Biotechnology and Sustainability Conference (PIPOC), Malaysian Palm Oil Board (MPOB). Kuala Lumpur: 9-12 November 2009. pp. 1371-1383.
- Mohd Aswad, A.W., Sariah, M., Paterson, R.R.M., Zainal Abidin, M.A., Lima, N. (2011). Ergosterol analysis of oil palm seedlings and plants infected with *Ganoderma* . *Crop protection* 30: 1438-1442.
- Moncalvo, J.M., Wang, H.H. and Hseu, R.S. (1995). Phylogenetic relationships in *Ganoderma* inferred from the internal transcribed spacers and 25S ribosomal DNA sequences. *Mycologia*, 87: 223-238.

- Montgomery, H.J., Montreal, C.M., Young, J.C. and Seifert, K.A. (2000). Determination of soil fungal biomass from soil ergosterol analyses. *Soil Biology and Biochemistry* 32:1207-1217.
- Morpurgo, G., Serlupi-Crescenzi, G., Tecce, G., Valente, F. and Venettacci, D. (1964). Influence of ergosterol on the physiology and the ultra structure of *Saccharomyces cerevisiae*. *Nature*, **201**: 897-899.
- Mottonen, M., J7rvinen, E., Hokkanen, T.J., Kuuluvainen, T., Ohtonen, R., (1999). Spatial distribution of soil ergosterol in the organic layer of a mature Scots pine (*Pinus sylvestris* L.) forest. *Soil Biology and Biochemistry* 31, 503–516.
- Mudge, S.M., Norris, C.E., (1997). Lipid biomarkers in the Conwy Estuary (North Wales, UK): a comparison between fatty alcohols and sterols. *Marine Chemistry* 57, 61–84.
- Naewbanij, M. Seib, P.A., Burroughs, R. (1984). Determination of Ergosterol using thin-layer chromatography and ultraviolet spectroscopy. *Cereal Chemistry* 61(5): 385-388.
- Navaratnam, S.J. and Chee, K.L. (1965). Root inoculation of oil palm seedlings with *Ganoderma sp.* *Plant Disease* 49, 1011–2.
- Newell S.Y. (1992). Estimating fungal biomass and productivity in decomposing litter. in *The fungal community*, eds Carroll G. C., Wicklow D. T. (Marcel Dekker, Inc. New York, N.Y), 2nd ed. pp 521–561.
- Newell S. Y. (1994). Total and free ergosterol in mycelia of saltmarsh ascomycetes with access to whole leaves or aqueous extracts of leaves. *Applied Environmental and Microbiology* 60:3479–3482.
- Newell, S.Y. (2001). Fungal biomass and productivity, in: *Methods in Microbiology*, 30, 357–372.
- Newell, S.Y., Fallon, R.D., Miller, J.D., (1986). Measuring fungal-biomass dynamics in standing-dead leaves of a salt-marsh vascular plant. In: Moss, S.T. (Ed.), *The Biology of Marine Fungi*. Cambridge University Press, Cambridge, pp. 19– 25.
- Newell, S.Y., Arsuffi, T.L., Fallon, R.D. (1988). Fundamental procedures for determining ergosterol content of decaying plant material by liquid chromatography. *Applied and Environmental Microbiology* 54, 1876–1879.
- Ng, H-E., Raj, S.S.A., Wong, S.H., Tey, D. and Tan, H.M. (2007). Estimation of fungal growth using the ergosterol assay: a rapid tool in assessing the microbiological status of grains and feeds. *Letters in Applied Microbiology*, 46: 113-118.

- Onuska, F.I., Terry, K.A. (1993). Extraction of pesticides from sediments using a microwave technique. *Chromatographia* 36, 191-194.
- Osswald, W.F., Hfl, W., Elstner, E.F., (1986). Ergosterol as a biochemical indicator of fungal infection in spruce and fir needles from different sources. *Z. Naturforsch.* 41, 542– 546.
- Pan, X., Niu, G., & Liu, H. (2002). Comparison of microwave-assisted extraction and conventional extraction techniques for extraction of tanshinones from *salvia miltiorrhiza bunge*. *Biochemical Engineering Journal*, 12(1), 71-77.
- Parkinson D. and Coleman D. C. (1991) . Microbial communities activity and biomass. *Agriculture, Ecosystems and environment*, 34, 3-33.
- Parsi, Z. and Górecki, T. (2006). Determination of ergosterol as an indicator of fungal biomass in various samples using non-discriminating flash pyrolysis. *Journal of Chromatography A*, **1130**: 145-150.
- Pasanen, A.-L., Yli-Pietila, K., Pasanen, P., Kalliokoski, P., Tarhanen, J.,(1999). Ergosterol content in various fungal species and biocontaminated building materials. *Applied Environmental and Microbiology* 65, 138– 142.
- Paterson, R.R.M. (2006). *Ganoderma*- a therapeutic fungal biofactory. *Phytochemistry* 67, 1985-2001.
- Paterson, R.R.M., (2007a). Internal amplification controls have not been employed in diagnostic fungal PCR hence potential false negative results. *Journal of Applied Microbiology* 102: 1369-1376.
- Paterson, R.R.M. (2007b). *Ganoderma* disease of oil palm - a white rot perspective necessary for integrated control. *Crop Protection* **26**,1369–1376.
- Paterson, R.R.M. and Lima, N. (2009). Mutagens manufactured in fungal culture may affect DNA/RNA of producing fungi. *Journal of Applied Microbiology*,106: 1070-1080.
- Paterson, R.R.M., Holderness, M., Kelley, J., Miller, R.N.G. and O’Grady, E. (2000). *In vitro* Biodegradation of Oil Palm Stem Using Macroscopic Fungi from South-East Asia: Preliminary Investigation. In: Flood, J. *et al.*, (eds), *Ganoderma* Diseases of Perennial Crops. CAB International Publishing, Wallingford, UK, pp. 129-139.
- Paterson, R.R.M., Sariah, M., Lima, N., Zainal Abidin, M.A. and Santos, C., (2008). Mutagenic and inhibitory compounds produced by fungi affect detrimentally diagnosis and phylogenetic analyses. *Current Bioactive Compounds*, 4: 245–257.
- Paterson, R.R.M., Sariah, M., Lima, N., (2013). How will climate change affect oil palm fungal disease. *Crop. Prot.* 46, 113–120.

- Pilotti, C.A. (2005). Stem rots of oil palm caused by *Ganoderma boninense*: pathogen biology and epidemiology. *Mycopathologia* 159, 129–137.
- Pilotti, C.A., Sanderson, F.R. and Aitken, E.A.B. (2002). Sexuality and interactions of monokaryotic and dikaryotic mycelia of *Ganoderma boninense*. *Mycological Research* 11, 1315–1322.
- Pilotti, C.A., Sanderson, F.R., Aitken, E.A.B. and Armstrong, W. (2004). Morphological variation and host range of two *Ganoderma* species from Papua New Guinea. *Mycopathologia* 158, 251–265.
- Proestos, C., and Komaitis, M. (2008). Application of microwave assisted extraction to the fast extraction of plant phenolic compounds. *LWT-Food Science and Technology*, 41: 652-659. *Pub. British Med. Bull.*, 49:479-724.
- Ravelet, C., Grosset, C., Alary, J., (2001a). Quantitation of ergosterol in river sediment by liquid chromatography. *Journal of Chromatography Sci.* 39, 239– 242.
- Ravelet, C., Grosset, C., Krivobok, S., Montuelle, B., Alary, J., (2001b). Pyrene degradation by two fungi in a freshwater sediment and evaluation of fungal biomass by ergosterol content. *Applied Microbiology and Biotechnology* 56, 803–808.
- Rees, R.W., Flood, J., Hasan, Y. and Cooper, R.M. (2007). Effects of inoculum potential, shading and soil temperature on root infection of oil palm seedlings by the basal stem rot pathogen *Ganoderma boninense*. *Plant Pathology* 56, 862–70.
- Rees, R.W., Flood, J., Hasan, Y., Potter, U. and Cooper, R.M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*); mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology*, 58: 982-989.
- Rees, R. W., Flood, J., Hasan, Y., Wills, M. A., & Cooper, R. M. (2011). *Ganoderma boninense* basidiospores in oil palm plantations: evaluation of their possible role in stem rots of *Elaeis guineensis*. *Plant Pathology*, **61(3)**, 1365-3059.
- Roberts C. W., McLeod, R., Rice, D. W., Ginger, M., Chance, M. L. and Goad, L. J. (2003). Fatty acid and sterol metabolism: potential antimicrobial targets in apicomplexan and trypanosomatid parasitic protozoa. *Molecular and Biochemical Parasitology*, vol. 126, no.2, pp. 129–142.
- Salmonawicz, B. and Nylund, J-E. (1988). High performance liquid chromatography determination of ergosterol as a measure of ectomycorrhiza infection in Scots pine. *European Journal of Forest Pathology*, 18: 291-298.
- Sanderson, F.R. (2005). An insight into spore dispersal of *Ganoderma boninense* on oil palm. *Mycopathologia* 159, 139–411.

- Sanderson, F.R., Pilotti, C.A. and Bridge, P.D. (2000). Basidiospores: Their influence on our thinking regarding a control strategy for Basal Stem Rot of oil palm. In: Flood, J. *et al.* (eds), *Ganoderma Diseases of Perennial Crops*. CAB International Publishing, Wallingford, UK, pp. 113-121.
- Saraf, A., Larsson, L., Burge, H., Milton, D., (1997). Quantification of ergosterol and 3-hydroxy fatty acids in settled house dust by gas chromatography-mass spectrometry: comparison with fungal culture and determination of endotoxin by a *Limulus* amoebocyte lysate assay. *Applied Environmental and Microbiology* 63, 2554–2559.
- Sariah, M., Hussin, M.Z., Miller, R.N.G. and Holderness, M. (1994). Pathogenicity of *Ganoderma boninense* tested by inoculation of oil palm seedlings. *Plant Pathology* 43, 507–10.
- Sariah, M., Choo, C.W., Zakaria, H. and Norihan, M.S. (2005). Quantification and characterisation of *Trichoderma* spp. from different ecosystems. *Mycopathologia* 159: 113-117.
- Schnürer J. (1993). Comparison of methods for estimating the biomass of three food-borne fungi with different growth patterns. *Applied Environmental Microbiology* 59:552–555.
- Schwadorf, K. and Muller, H.-M. (1989). Determination of ergosterol in cereals, mixed feed components and mixed feeds by liquid chromatography. *Journal of Analytical Chemistry*, 72: 457– 462.
- Seitz, L.M., Mohr, H.E., Burroughs, R. Sauer, D.B. 1977. Ergosterol as an indicator of fungal invasion in grains. *Cereal Chemistry* 54, 1207-1217.
- Seitz, L.M., Sauer, D.B., Burroughs, R., Mohr, H.E., Hubbard, J.D. 1979. Ergosterol as a measure of fungal growth. *Journal of Phytopathology* 69, 1202-1203.
- Shamala, S., D. Chris, O. Sioban and Idris, A.S.(2006). Preliminary studies on the development of monoclonal antibodies against mycelia of *Ganoderma boninense*, the causal pathogen of basal stem rot of oil palm. *Malaysian Journal of Microbiology* 2: 30-34.
- Singh, G., (1990). *Ganoderma- The scourge of oil palm in the coastal areas*. In: Ariffin, D. and Jalani, S., (Eds.), In: *Proceedings of the Ganoderma Workshop*, 11 September 1990. Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia 7-35.
- Singh, G. (1991). *Ganoderma- the scourge of oil palms in the coastal areas*. *Planter* 67: 421-444.

- Souza, W. and Rodrigues, J.L.F. (2009). Sterol Biosynthesis Pathway as Target for Anti-Drugs. *Interdisciplinary Perspectives on Infectious Diseases* Volume 2009, Article ID 642502, 19 pages.
- Stahl, P.D. and Parkin, T.B. (1996). Relationship of soil ergosterol concentration and fungal biomass. *Soil Biology Biochemistry*, 28: 847-855.
- Starmans DAJ and Nijhuis HH (1996) Extraction of secondary metabolites from plant material: a review. *Trends Food Science and Technology*, 7: 191-197.
- Steyaert, R.L. (1967). Les *Ganoderma* palmicoles. *Bulletin du Jardin Botanique Nationale Belgique* 37, 465-492.
- Sticher, O. (2008). Natural product isolation. *Natural Product Reports*, 25: 517-554.
- Suberkropp K., Gessner M.O., Chauver E. (1993). Comparison of ATP and ergosterol as indicators of fungal biomass associated with decomposing leaves in streams. *Applied Environmental Microbiology* 59:3367-3372.
- Susanto, A. (2002). kajian pengendalian hayati *Ganoderma boninense* Pat. Penyebab penyakit busuk pangkal batang kelapa sawit. Disertai IPB, Bogor.
- Susanto, A. and Sudharto. (2003). Status of *Ganoderma* disease on oil palm in Indonesia. *Third International Workshop on Ganoderma disease of Perennial Crops*, March 24-26, Medan, Indonesia.
- Susanto, A. Sudharto, P.S. and Purba, R.Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, **159**: 153-157.
- Teh, K.S. and Sariah, M., (1999). Improved inoculation technique for testing pathogenicity of *Ganoderma boninense* on oil palm seedlings. In: *Plant Protection in the Information Age*. Fourth MAAPS International Conference on Plant Protection in the Tropics 142-145.
- Teh, C.L., Tey, C.C., and Normahnani, M.N. (2010). Integrated Disease Management of *Ganoderma* in Sabah. Paper presented in EMPA Seminar on *Ganoderma* Disease of Oil Palm in East Malaysia at Sabah Hotel, Sandakan, on 5 April 2010. Management of *Ganoderma* diseases in oil palm plantations 7 pp.
- Thompson, A. (1931). Stem rot of the oil palm in Malaya. *Bulletin Department of agriculture, Straits Settlements and F.M.S.*, *Science Series* 6:23 pp.
- Toh Choon R.L., Sariah M., Siti Mariam M.N., (2012). Ergosterol from the soilborne fungus *Ganoderma boninense*. Short communication; *Journal of Basic Microbiology* 52: 608-612.

- Trojanowski, J., A. Hüttermann, K. Haider and J.G.H. Wessels, (1985). Degradation of Lignin and Lignin Related Compounds by Protoplasts Isolated from *Fomes annosus*. *Archives of Microbiology* 140(4): 326-330.
- Turner, P. D. (1981). Oil Palm disease and Disorders. *Oxford University Press*.
- Turner, E.C., Snaddon, J.L., Fayle, T.M., Foster, W.A. (2008). Oil Palm Research in Context: Identifying the Need for Biodiversity Assessment. *PLoS ONE* 3(2):e1572.
- Urbina, J.A. (1997). Lipid biosynthesis pathways as chemotherapeutic targets in kinetoplastid parasites. *Parasitology*, vol. 114, supplement 1, pp. S91–S99, 1997.
- Utomo, C. and Niepold, F. (2000). Development of diagnostic methods for detecting *Ganoderma* infected oil palms. *Journal of Phytopathology* 148: 507-514.
- Wang, L., Weller, C.L. 2006. Recent advances in extraction of nutraceuticals from plants. *Trends in Food Science and Technology* 17, 300-312.
- Wallander, H., Massicotte, H.B., Nylund, J.E. (1997). Seasonal variation in protein, ergosterol and chitin in five morphotypes of *Pinus sylvestris* L. ectomycorrhizae in a mature Swedish forest. *Soil Biology and Biochemistry* 29, 45–53.
- Weete, J.D. and Gandhi, S.R. (1997). *Lipids*, 32, 1309. Springer–Verlag Publication.
- Whipps, J. and Lewis, D. (1980). Methodology of a chitin assay. *Transactions of the British Mycological Society* 79, 178–179.
- Wijesekera, H.T.R., Wijesundera, R.L.C. and Rajapakse, C.N.K. (1996). Hyphal interactions between *Trichoderma viridae* and *Ganoderma boninense* Pat. cause of coconut root and bole rot. *Sri Lanka J. Nat. Sci.*, 24: 217–219.
- Xue, H. Q., Upchurch, R. G., and Kwanyuen, P. 2006. Ergosterol as a quantifiable biomass marker for *Diaporthe phaseolorum* and *Cercospora kikuchii*. *Plant Disease* 90:1395-1398.
- Yamoaka, M., Hayakawa, S., Tsukamoto, M., Kurane, R., Idris, A.S., Mohd Haniff, H. and Ariffin, D. (2000). Diagnosis of basal stem rot of oil palm by foliar analysis and PCR-based detection of *Ganoderma* in oil palm. In the Proceedings of the 23rd Malaysian Society for Microbiology Symposium, Langkawi, Kedah 19-20 November 2000. pp. 4.
- Young, J.C. 1995. Microwave-assisted extraction of the fungal metabolite ergosterol and total fatty acids. *Journal of Agricultural and Food Chemistry* 43 (11), 2904-2910.

Young, J.C., and Games, D.E. 1993. Supercritical fluid extraction and supercritical fluid chromatography of the fungal metabolite ergosterol. *Journal of Agricultural and Food Chemistry* 41, 577-581.

Yuan, J.P, Wang, J.H., Liu, X., Kuang, H.C. and Huang, X.N. (2006). Determination of ergosterol in ganoderma spore lipid from the germinating spores of *Ganoderma lucidium* by HPLC. *Journal Agriculture Food Chemistry*, 54: 6172-6176.

Yuan, J.P., Hai Wang, J., Liu, X., Cong Kuang, H. and Yan Zhao, S. (2007). Simultaneous determination of free ergosterol and ergosteryl esters in *Cordyceps sinensis* by HPLC. *Food Chemistry*, 105: 1755-1759.

Zaiton, S. (2006). Bacterial endophytes from oil palm (*Elaeis guineensis*) and their antagonistic activity against *Ganoderma boninense*. Master of Science, Universiti Putra Malaysia, Selangor, Malaysia.

Zill, G. Engelhardt, G. and Wallniifer, P.R. (1988). Determination of ergosterol as a measure of fungal growth using Si 60 HPLC. *Z Lebensm Unters Forsch (European Food Research Technology)*, 187: 246-249.