



UNIVERSITI PUTRA MALAYSIA

***PHENOTYPIC, MOLECULAR AND MATING TYPE ANALYSES OF
EXSEROHILUM ROSTRATUM ASSOCIATED WITH LEAF SPOTS OF
RICE***

NOR AZIZAH KUSAI

FS 2015 54



**PHENOTYPIC, MOLECULAR AND MATING TYPE ANALYSES OF
Exserohilum rostratum ASSOCIATED WITH
LEAF SPOTS OF RICE**

By

NOR AZIZAH BINTI KUSAI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of
Master of Science**

August 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

**PHENOTYPIC, MOLECULAR AND MATING TYPE ANALYSES OF
Exserohilum rostratum ASSOCIATED WITH
LEAF SPOTS OF RICE**

By

NOR AZIZAH BINTI KUSAI

August 2015

Chairman : Nur Ain Izzati Mohd Zainudin, PhD
Faculty : Science

Exserohilum rostratum is one of the plant pathogens causing leaf spot disease, which affects wide range of plant species, mainly grasses. There has been no report of *E. rostratum* causes the disease on rice plant particularly on leaves, therefore this study was conducted. During a series of sampling in March to August 2013, leaf spot is one of common symptoms found in rice plantations in Malaysia. The objectives of this study were to identify fungi isolated from leaf spot of rice based on phenotypic and molecular approaches, to determine whether or not *E. rostratum* isolates were pathogenic, to analyze the mating type of *E. rostratum* using Polymerase Chain Reaction (PCR) and to examine the sexual development of *E. rostratum* isolates. All isolated fungi were purified and identified based on their phenotypic features for species delimitation and confirmed by sequencing the internal transcribed spacers (ITS) region. Seventy-eight isolates were identified; *E. rostratum* (45 isolates), *Bipolaris sorokiniana* (10 isolates), *Curvularia hawaiiensis* (8 isolates), *C. geniculata* (6 isolates), *C. eragrostidis* (6 isolates), *C. aerea* (2 isolates) and *C. lunata* (1 isolate). A phylogenetic tree was constructed using neighbor-joining method showed the same species grouped in the same clade. Clade I consisted of *E. rostratum* while Clade II were *Bipolaris sorokiniana* and *Curvularia* species. Based on pathogenicity test, some isolates of *E. rostratum* were pathogenic and produced leaf spot symptoms towards rice. In this study, *E. rostratum* is minor or secondary pathogen to rice because it showed smaller lesion compared to infection by *B. oryzae*, the main pathogen of brown spot disease. Mating type (*MAT*) of all isolates of *E. rostratum* were amplified using two sets of primers (NM162MAT1_1F1Seto and NM163MAT1_1R1Seto for *MAT1-1*; NM164MAT1_2F1Seto and NM165MAT1_2R1Seto for *MAT1-2*). Three isolates were confirmed carried *MAT1-1* allele while 42 isolates are *MAT1-2* isolates. The imbalance distribution of both *MAT* may caused by ratio of modes of reproduction and selection pressure in the field. The opposite *MAT* isolates were crossed on Sach's medium for 3 weeks in quadruplicate. Nine isolates were indicated as fertile, which produced mature pseudothecia and ascospores, 12 isolates produced barren pseudothecia (empty asci) and the remaining 24 isolates were non-fertile without formation of pseudothecia. Three isolates of *E. rostratum* from this study; one *MAT1-1* isolate (UPM1109) and two isolates of *MAT1-2* (UPM1247 and UPM1152) are highly fertile and suggested to

be used as tester strains. The developed tester strains may facilitate other researchers to access the fertility on other related study on other *E. rostratum* isolates.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**FENOTIP, MOLEKULAR DAN ANALISIS JENIS PENGAWANAN
EXSEROHILUM ROSTRATUM BERASSOSIASI DENGAN BINTIK DAUN
PADA PADI**

Oleh

NOR AZIZAH BINTI KUSAI

Ogos 2015

Pengerusi : Nur Ain Izzati Mohd Zainudin, PhD
Fakulti : Sains

Exserohilum rostratum merupakan salah satu patogen tumbuhan yang menyebabkan bintik daun yang mana menjejaskan pelbagai jenis spesies tumbuhan terutamanya rumput. Tiada laporan berkenaan *E. rostratum* menyebabkan penyakit pada pokok padi terutamanya pada daun, oleh itu kajian ini dijalankan. Semasa siri pensampelan dijalankan antara Mac hingga Ogos 2013, bintik daun adalah salah satu simptom yang biasa ditemui di sawah padi di Malaysia. Objektif kajian ini adalah untuk memencilkan dan mengecam kulat daripada bintik daun padi berdasarkan pendekatan fenotip dan molekul, untuk mengkaji sama ada pencilan-pencilan *E. rostratum* adalah patogenik, untuk menganalisis jenis pengawanan *E. rostratum* menggunakan *Polymerase Chain Reaction (PCR)* dan untuk mengkaji perkembangan seksual pencilan *E. rostratum*. Kesemua pencilan kulat telah dipencilkan dan dicamkan berdasarkan ciri fenotip untuk pencecaman species dan disahkan dengan analisis jujukan kawasan *Internal Transcribed Spacer (ITS)*. Tujuh puluh lapan pencilan telah berjaya dicamkan; *E. rostratum* (45 pencilan), *Bipolaris sorokiniana* (10 pencilan), *Curvularia hawaiiensis* (8 pencilan), *C. geniculata* (6 pencilan), *C. eragrostidis* (6 pencilan), *C. aerea* (2 pencilan) and *C. lunata* (1 pencilan). Pohon filogeni telah dibina menggunakan kaedah *neighbor-joining*, menunjukkan bahawa spesies yang sama dikelaskan dalam clad yang sama. Clad 1 terdiri daripada *E. rostratum* manakala Clad II adalah *Bipolaris sorokiniana* dan spesies *Curvularia*. Berdasarkan ujian kepatogenan, beberapa pencilan *E. rostratum* adalah patogenik dan menghasilkan simptom bintik daun terhadap padi. Di dalam kajian ini, *E. rostratum* ialah patogen minor atau sekunder kepada padi kerana menunjukkan lesi bersaiz kecil berbanding jangkitan oleh *B. oryzae*, patogen utama penyakit bintik perang. Jenis pengawanan (*MAT*) untuk semua pencilan *E. rostratum* diamplikasikan menggunakan dua set primer (NM162MAT1_1F1Seto and NM163MAT1_1R1Seto untuk *MAT1-1*; NM164MAT1_2F1Seto and NM165MAT1_2R1Seto untuk *MAT1-2*). Tiga pencilan telah disahkan mempunyai *MAT1-1* manakala 42 pencilan adalah *MAT1-2*. Ketidakeimbangan pengagihan untuk kedua-dua *MAT* mungkin disebabkan nisbah cara pembiakan dan tekanan pemilihan di lapangan. Pencilan yang bertentangan *MAT* telah dikacukkan di atas media Sach selama tiga minggu dengan empat replikasi. Sembilan pencilan adalah subur, di mana menghasilkan pseudotesia dan ascospora

matang, 12 pencilan menghasilkan pseudotesia yang mandul (askus kosong) dan baki 24 pencilan adalah tidak subur tanpa pembentukan pseudotesia. Tiga pencilan *E. rostratum* daripada kajian ini, satu pencilan *MATI-1* (UPM1109) dan dua pencilan *MATI-2* (UPM1247 dan UPM1152) adalah paling subur dan dicadangkan untuk digunakan sebagai strain penguji. Strain yang ditemui dapat membantu penyelidik lain untuk memeriksa kesuburan strain *E. rostratum* yang lain di dalam kajian yang berkaitan.



ACKNOWLEDGEMENT

Bismillahirrahmanirrahim...

Alhamdulillah, all praise to Almighty ALLAH S.W.T. for the strength and His blessing in completing this thesis.

I would like to express my sincere and deepest gratitude to my supervisor, Dr. Nur Ain Izzati Mohd Zainudin for her patience, motivation, guidance and immense knowledge. Her invaluable help of constructive comments and advices throughout the experiment and thesis have contributed to the success of this research. Not forgotten to my co-supervisors, Dr. Mohd Termizi Yusof and Dr. Shahrizim Zulkifly for their support and contribution in this research. Besides, my thanks go to Director of MARDI Seberang Perai, Tuan Haji Azmi Abd Razak that helps me a lot in this work.

Sincere thanks to Kak Hidayah and Kak Nik for their full cooperation and facilities during my working for this research. My acknowledgement also goes to all staff of Department of Biology, Faculty of Science.

I want to express my deep thanks to my Mycology Lab members, Madiah, Maryam, Munirah, and seniors, Suhaida, Abby, and Nithiya who have given their opinion and time. Special thanks to my friends, Ain, Intan and Adibah who always give moral support every time I faced difficulties.

Finally, my deepest gratitude goes to my beloved parents, Kusai b. Mohd Taib and Aishah bt Mat Desa and also my siblings for their endless love, prayers and encouragement. To those who indirectly contributed in this research, your kindness means a lot to me. Thank you very much.

May Allah bless you all...

I certify that a Tesis Examination Committee has met on (10 August 2015) to conduct the final examination of Nor Azizah Kusai on her tesis entitled “Phenotypic, Molecular and Mating Type Analyses of *Exserohilum rostratum* Associated with Leaf Spot of Rice” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Master of Science.

Members of the Thesis Examination Committee were as follows:

Muskhazli bin Mustafa, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Wan Zuhainis binti Saad, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Science
Universiti Putra Malaysia
(Internal Examiner)

Hideyuki Nagao (Dhaakirullah bin Abdullah), PhD

Associate Professor
School of Biological Sciences
Universiti Sains Malaysia
Malaysia
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 5 November 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Nur Ain Izzati binti Mohd Zainudin, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Mohd Termizi bin Yusof, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Shahrizim bin Zulkifly, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No. : Nor Azizah binti Kusai, GS35553

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of
Chairman of
Supervisory
Committee: Nur Ain Izzati Mohd Zainudin, PhD

Signature: _____
Name of
Member of
Supervisory
Committee: Mohd Termizi Yusof, PhD

Signature: _____
Name of
Member of
Supervisory
Committee: Shahrizim Zulkifly, PhD

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xiv
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 Classification and Characteristics of <i>E. rostratum</i>	4
2.1.1 Current Taxonomy	4
2.1.2 Phenotypic Characteristics	4
2.1.3 Mating Types and Sexual Development of <i>E. rostratum</i>	5
2.2 Rice (<i>Oryza sativa</i> L.)	6
2.2.1 The Importance of Rice	6
2.2.2 Rice Plantation in Malaysia	7
2.2.3 Disease of Rice	7
2.2.4 Rice Planting Method	8
2.3 Disease of Rice Associated with Leaf Spot Symptoms	9
2.3.1 Disease Symptoms	9
2.3.2 Disease Development	10
2.3.3 Leaf Spot Disease of Rice in Malaysia	10
2.4 <i>Exserohilum rostratum</i> as Plant Pathogen	11
2.5 Molecular Identification of Fungi based on Internal Transcribed Spacer (ITS) Sequence Analysis	13
3 MATERIALS AND METHODS	
3.1 Culture Media and Incubation Condition	15
3.2 Fungal Isolation	15
3.3 Phenotypic Characterization	16
3.3.1 Macroscopic Characteristics	17
3.3.2 Microscopic Characteristics	17
3.4 Molecular Characterization	17
3.4.1 DNA Extraction	17
3.4.2 PCR Amplification of Internal Transcribed Spacer (ITS) Region	18
3.4.3 Gel Electrophoresis	18
3.4.4 Gel Purification	18
3.4.5 ITS Gene Sequencing and Phylogenetic Tree	19
3.5 Pathogenicity Test	19

3.5.1	Plant Inoculation	19
3.5.2	Disease Assessment and Data Analysis	19
3.6	Mating Type and Sexual Development of <i>E. rostratum</i>	19
3.6.1	Amplification of Mating Type (<i>MAT</i>) Gene	19
3.6.2	Crossing Procedure	20
4	RESULTS	
4.1	Phenotypic Characteristics	21
4.1.1	<i>Exserohilum rostratum</i>	23
4.1.2	<i>Bipolaris sorokiniana</i>	28
4.1.3	<i>Curvularia hawaiiensis</i>	30
4.1.4	<i>Curvularia eragrostidis</i>	32
4.1.5	<i>Curvularia geniculata</i>	34
4.1.6	<i>Curvularia lunata</i>	36
4.1.7	<i>Curvularia aerea</i>	37
4.2	Molecular Characterization	39
4.3	Pathogenicity Test	44
4.4	Mating Type Analyses of <i>E. rostratum</i>	45
4.5	Sexual Crosses of <i>E. rostratum</i>	47
5	DISCUSSION	
5.1	Isolation and Identification of Fungi Associated with Leaf Spots of Rice	50
5.2	Pathogenicity of <i>E. rostratum</i>	53
5.3	Mating Type and Sexual Development of <i>E. rostratum</i>	54
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	
6.1	Summary and Conclusion	57
6.2	Recommendations and Future Research	57
	REFERENCES	59
	APPENDICES	76
	BIODATA OF STUDENT	154
	LIST OF PUBLICATIONS	155

LIST OF TABLES

Table		Page
2.1	Characteristics of species in the genus <i>Exserohilum</i>	13
3.1	Sampling sites and growth stage of the rice plants showed leaf spot symptom	16
4.1	List of fungal isolates based on sampling locations	22
4.2	Morphological characteristics <i>E. rostratum</i> isolates	25
4.3	Morphological characteristics of <i>B. sorokiniana</i> isolates	29
4.4	Morphological characteristics of <i>C. hawaiiensis</i> isolates	31
4.5	Morphological characteristics of <i>C. eragrostidis</i> isolates	33
4.6	Morphological characteristics of <i>C. geniculata</i> isolates	35
4.7	Morphological characteristics of <i>C. aerea</i> isolates	38
4.8	GenBank accession number of isolated fungal isolates	41
4.9	Mating type and fertility status of <i>E. rostratum</i>	48

LIST OF FIGURES

Figure		Page
2.1	Rice growing areas in Peninsular Malaysia	11
2.2	Internal transcribed spacer region in ribosomal RNA	14
3.1	Rice leaves showing leaf spot symptoms	15
4.1	The percentage isolates isolated from infected rice leaves according to species	21
4.2	Morphological characteristics of <i>E. rostratum</i>	24
4.3	Morphological characteristics of <i>B. sorokiniana</i>	28
4.4	Morphological characteristics of <i>C. hawaiiensis</i>	30
4.5	Morphological characteristics of <i>C. eragrostidis</i>	32
4.6	Morphological characteristics of <i>C. geniculata</i>	34
4.7	Morphological characteristics of <i>C. lunata</i>	36
4.8	Morphological characteristics of <i>C. aeria</i>	37
4.9	Banding pattern of amplification of ITS region	39
4.10	Neighbour-joining tree showing the relationship between 78 isolates of <i>E. rostratum</i> , <i>Bipolaris</i> and <i>Curvularia</i> species generated from ITS sequences.	43
4.11	Lesion on leaves after nine days inoculated with <i>E. rostratum</i> isolates.	44
4.12	Lesion sizes of brown leaves spots on day nine after inoculation.	45
4.13	Banding pattern of mating type of <i>E. rostratum</i> isolates	46
4.14	Sexual stages of <i>E. rostratum</i>	49

LIST OF ABBREVIATIONS

%	Percentage
°C	Degree celcius
µl	Microliter
µm	Micrometer
µM	Micromolar
×g	Centrifugal force
bp	Base pair
Ca(NO ₃) ₂	Calcium nitrate
CaCO ₃	Calcium carbonate
cm	centimeter
CM	Complete medium
CMX	Complete medium xylose
DNA	Deoxyribonucleic acid
dNTP	Deoxynucleotide triphosphate
dpi	days post-inoculation
EtBr	Ethidium bromide
g	gram
h	hour
ha	hectares
IADA	Kawasan Pembangunan Pertanian Bersepadu
ITS	Internal transcribed spacer
K ₂ HPO ₄	Potassium hydrogen phosphate
KADA	Lembaga Kemajuan Pertanian Kemubu
kb	kilo base
KETARA	Kawasan Pembangunan Pertanian Bersepadu Terengganu Utara
kg	kilogram
L	liter
MADA	Lembaga Kemajuan Pertanian MUDA
MAT	Mating type
mg	miligram
MgSO ₄	Magnesium sulphate
min	minutes

ml	milliter
ng	nanogram
PCR	Polymerase chain reaction
rDNA	Ribosomal deoxyribonucleic acid
rpm	revolution per minute
s	second
TBE	Tris-Boric acid-EDTA
V	Volt
WA	Water agar



CHAPTER 1

INTRODUCTION

Kingdom of Fungi is composed of most diverse species on Earth consisting of pathogens, saprobes and endophytes. According to The Dictionary of Fungi by Kirk *et al.* (2008), there are about 99000 known species recorded. The ascomycetes fungus, *Exserohilum* is an important phytopathogens that cause high yield losses and pathogenic towards human and animals. *Exserohilum rostratum* is a plant pathogen that affects more than 30 plants species comprises of 28 genera in 11 families (Lin *et al.*, 2011) including the important crops such as sugarcane, tomato, sweet sorghum and wheat which leads severe yield losses (Ahmadpour *et al.*, 2013; Cardona and González, 2007; Luan *et al.*, 2004). *Exserohilum rostratum* has been reported to cause leaf spot and blight in many plant species particularly grasses (Sharma *et al.*, 2014). However, there have been no reports found on disease of rice plant caused by *E. rostratum*.

Rice (*Oryza sativa*); classified under family Poaceae is an important cereal plants cultivated across the world. Rice supplied with variety of nutrient and consumed as staple food for humans and animals in many parts of the world. Rice consumption was projected to increase by 2030 with 873 million tonnes compared to year 2000 (Kubo and Purevdorj, 2004). People in Southeast Asia; consume rice as more than 50% of their daily diet composition (Fitzgerald *et al.*, 2009). A high demand of rice gives impact on rice industry over the world and gets a lot of attention by researchers. Rice production has contributed in Malaysia's agricultural sector development. Up to 2013, Malaysia has 688207 hectares of rice cultivation, which is the third largest crop cultivated (Department of Agriculture, 2014a). In Malaysia, wetland rice is grown in Peninsular Malaysia as the main rice cultivation compared to upland rice, which can only be found in Borneo Island. Rice production plays important roles in Malaysia as national's food security, self-sufficiency and source of country's income (Fahmi *et al.*, 2013). With that, the studies on etiology and disease distribution of rice are important to help in disease management.

Abiotic and biotic pressure are the limiting factors in rice cultivation and always give significant problems to rice growing in field especially in large cultivation areas. Biotic pressures such as pests and pathogens caused severe damage and losses of rice yield (Oerke *et al.*, 2012). Fungi are major pathogens infecting rice during seedling up to maturing stage occurring on all parts of plants. Leaf spot, blast, blight and smut are the common diseases found in field caused by infestation of several fungal species. Leaf spots such as narrow brown spot, brown spot and blast disease frequently found in all rice growing areas. However, in Asia, especially Malaysia, limited number of references was available on the disease. Therefore, etiology of ascomycete fungi including *Exserohilum* species was studied.

Phenotypic characteristics are the primary step in identifying plant pathogens. Characterization based on phenotype of *E. rostratum* was identified through macroscopic and microscopic level. Both levels provide information to determine and limit the species identification. However, this step always leads to misidentification because of lack of standardized terminology and high subjectivity depends on environmental condition (Guarro *et al.*, 1999). Therefore, phytopathology researchers have extensively used molecular method to confirm the species identification. In recent years, characterization of DNA sequences have been used to determine fungal species based on phylogenetic species concept (Taylor *et al.*, 2000). Advances of molecular studies have enabled to distinguish small DNA sequences between species. Target genes such as rRNA, translation elongation factors 1-alpha (*tefl- α*), β -tubulin, glyceraldehyde-3-phosphate dehydrogenase (GPDH) are generally used to provide sufficient information to determine species (Manamgoda *et al.*, 2012; Geiser *et al.*, 2004). Internal transcribed spacer of rRNA was widely used for species differentiation between *Exserohilum* since it clearly defined interspecific variation (Sharma *et al.*, 2014; Wu and Turgeon, 2013; Lin *et al.*, 2011).

To make the study more comprehensible, biological species concept are also applied in characterization of *E. rostratum* based on sexual development. Development of fungus progeny sexually requires nuclear fusion and meiosis processes. This process leads to exchange of genetic materials between two organisms and form variation and recombination of progeny. The progeny produced through sexual stage is more vigor and have better chances to survive in harsh environment. Sexually produced generations will display variation in their traits such as virulence phenotype (Alters and Alters, 2006). Sexual reproduction can be applied in investigation of species invasion, conservation biology, pathogen evolution and control method of pathogen (Billiard *et al.*, 2012). Billiard *et al.* (2012) also state that sexual reproduction important which can affect the ecological process of the species through adaptation or colonization.

The study of mating type of organism has increased throughout the years since a century ago. Mate selection and mating process is significant event in the life of a fungus. Mating type (*MAT*) is used as a term in filamentous fungi and non-filamentous fungi to indicate genetic regulation of mating specificity (Kronstad and Staben, 1997). *MAT* consists of two functional alleles, *MAT1-1* and *MAT1-2* that occupy in single locus chromosome (Debuchy and Turgeon, 2006). Sexual reproduction process was initiated when there is interaction between opposite *MAT* alleles under favorable condition. The sexual development of fungus will form fruiting body known as ascocarps and ascospores as the progeny. Traditionally, *MAT allele* was determined using sexual crosses that required tester strains. However, this approach is time-consuming method. Mating type analyses using molecular technique is new insight and recently used in mating type studies because of rapid, effective, and accurate routine technique (Steenkamp *et al.*, 2000; Gafur *et al.*, 1997).

Exserohilum rostratum is heterothallic fungus and the teleomorph stage can only be obtained by crossing between opposite mating type. The teleomorph stage of *E. rostratum* known as *Setosphaeria rostrata* and the fruiting body formed is called pseudothecia with bitunicate ascus. Leonard (1976) firstly described teleomorph stage

of *E. rostratum* under laboratory condition using in-vivo culture. As *E. rostratum* becoming important plant pathogens and found in broad range host, the studies on their sexual development are crucial. The tester strains for *E. rostratum* are still undeveloped, to study the specific aspect of *E. rostratum* such as genetic, physiological and plant-pathogen interaction. Therefore, this study was conducted to develop the standard tester strain for *E. rostratum* studies.

The result will provide the information on diversity of *E. rostratum* associated with leaf spots of rice in Peninsular Malaysia, as references for related species and facilitate other researchers on *E. rostratum* studies.

Therefore, the objectives of this study were;

- 1) To isolate and identify *Exserohilum rostratum* and other associated fungi based on morphological and molecular characterization
- 2) To determine whether or not the *E. rostratum* isolates isolated from leaf spot of rice were pathogenic
- 3) To determine the mating type and to examine the sexual development of *E. rostratum* isolates

REFERENCES

- Agrios, G. N. (2005). *Plant Pathology*. UK: Elsevier Academic Press Publication.
- Ahmadpour, A., Karami, S., Heidarian, Z., & Javan-Nikkhah, M. (2013). *Exserohilum rostratum* causing sugarcane leaf spot in Iran. *Australasian Plant Disease Notes*, 8(1): 97-99.
- Alcorn, J. L. (1988). The taxonomy of " Helminthosporium" species. *Annual Review of Phytopathology*, 26(1): 37-56.
- Alters, S., & Alters, B. (2006). *Biology: Understanding Life* (Vol. 1). USA: John Wiley & Sons, Inc.
- Amatulli, M. T., Spadaro, D., Gullino, M. L., & Garibaldi, A. (2010). Molecular identification of *Fusarium* spp. associated with bakanae disease of rice in Italy and assessment of their pathogenicity. *Plant Pathology*, 59(5): 839-844.
- Amin, E. N. E., & Abdalla, M. H. (1980). Survival of *Curvularia lunata* var. *aeria* in soil. *Mycopathologia*, 71(3): 137-140.
- Arie, T., Christiansen, S. K., Yoder, O. C., & Turgeon, B. G. (1997). Efficient cloning of ascomycete mating type genes by PCR amplification of the conserved MAT HMG Box. *Fungal Genetics and Biology*, 21(1): 118-130.
- Ariyawansa, H. A., Thambugala, K. M., Manamgoda, D. S., Jayawardena, R., Camporesi, E., Boonmee, S., Wanasinghe, D. N., Phookamsak, R., Hongsanan, S., & Singtripop, C. (2015). Towards a natural classification and backbone tree for Pleosporaceae. *Fungal Diversity*, 71(1): 85-139.
- Azmi, M., & Abdullah, M. Z. (1998). A Manual For Identification and Control of Padi Angin (Weedy Rice) in Malaysia. Serdang: MARDI Publication.
- Azmi, A. R., Nur Ain Izzati, M. Z., Siti Nordahliawate, S., Nor Azliza, I., Nik Mohd Izham, N. M., & Baharuddin, S. (2009). Sheath brown rot disease of rice caused by *Pseudomonas fuscovaginae* in the Peninsular Malaysia. *Journal of Plant Protection Research*, 49(3): 244-249.
- Azzam, O., & Chancellor, T. C. B. (2002). The biology, epidemiology, and management of rice Tungro disease in Asia. *Plant Disease*, 86(2): 88-100.
- Bellemare, M. F. (2015). Rising food prices, food price volatility, and social unrest. *American Journal of Agricultural Economics*, 97(1): 1-21.
- Berbee, M., Pirseyedi, M., & Hubbard, S. (1999). *Cochliobolus* phylogenetics and the origin of known, highly virulent pathogens, inferred from ITS and glyceraldehyde-3-phosphate dehydrogenase gene sequences. *Mycologia*, 91: 964-977.

- Bhargava, T., & Hamer, J. E. (2001). Molecular Aspects of Host-Pathogen Interactions. In S. Sreenivasaprasad & R. Johnson (Ed.), *Major Fungal Diseases of Rice* (pp 61-86). Netherlands: Kluwer Academic Publishers.
- Bi, Q., Wu, D., Zhu, X., & Turgeon, B. G. (2013). *Cochliobolus heterostrophus* Llm1 – A Lae1-like methyltransferase regulates T-toxin production, virulence, and development. *Fungal Genetics and Biology*, 51: 21-33.
- Bihon, W., Wingfield, M. J., Slippers, B., Duong, T. A., & Wingfield, B. D. (2014). MAT gene idiomorphs suggest a heterothallic sexual cycle in a predominantly asexual and important pine pathogen. *Fungal Genetics and Biology*, 62: 55-61.
- Billiard, S., Lopez Villavicencio, M., Hood, M. E., & Giraud, T. (2012). Sex, outcrossing and mating types: unsolved questions in fungi and beyond. *Journal of Evolutionary Biology*, 25(6): 1020-1038.
- Blumenstein, A., Vienken, K., Tasler, R., Purschwitz, J., Veith, D., Frankenberg-Dinkel, N., & Fischer, R. (2005). The *Aspergillus nidulans* phytochrome FphA represses sexual development in red light. *Current Biology*, 15(20): 1833-1838.
- Braus, G. H., Krappmann, S., & Eckert, S. E. (2002). Sexual Development in Ascomycetes in Fruiting Body Formation of *Aspergillus nidulans*. In H. D. Osiewacz (Ed.), *Molecular Biology of Fungal Development* (pp. 200-226). Switzerland: Marcel Dekker Inc.
- Brewer, M. T., Cadle-Davidson, L., Cortesi, P., Spanu, P. D., & Milgroom, M. G. (2011). Identification and structure of the mating-type locus and development of PCR-based markers for mating type in powdery mildew fungi. *Fungal Genetics and Biology*, 48(7): 704-713.
- Brunelli, K. R., Dunkle, L. D., Sobrinho, C. A., Fazza, A. C., & Camargo, L. E. A. (2008). Molecular variability in the maize grey leaf spot pathogens in Brazil. *Genetics and Molecular Biology*, 31(4): 938-942.
- Brunings, A. M., Datnoff, L. E, Palmateer, A. J., Locke, J. C., & Krause, C. R. (2009). Exserohilum leaf spot on tiger grass. *Plant Health Progress*. doi: 10.1094/PHP-2009-1215-01-RS.
- Bunawan, H., Dusik, L., Bunawan, S. N., & Mat Amin, N. (2014). Rice Tungro disease: from identification to disease control. *World Applied Sciences Journal*, 31(6): 1221-1226.
- Callan, B. E., & Carris, L. M. (2004). Fungi on Living Plant Substrata, Including Fruits. In G. M. Mueller, G. F. Bills, M. S. Foster (Ed.), *Biodiversity of Fungi: Inventory and Monitoring Methods* (pp 105- 214). UK: Academic Press.

- Calvo, A. M., Wilson, R. A., Bok, J. W., & Keller, N. P. (2002). Relationship between secondary metabolism and fungal development. *Microbiology and Molecular Biology Reviews*, 66(3): 447-459.
- Cardona, R., & González, M. S. (2007). First report of *Exserohilum rostratum* associated with rice seed in Venezuela. *Plant Disease*, 91: 226.
- Chakrabarti, N. K. (2001). Epidemiology and Disease Management of Brown Spot Disease in India. In S. Sreenivasaprasad & R. Johnson (Ed.), *Major Fungal Diseases of Rice* (pp 293-306). Netherlands: Kluwer Academic Publisher.
- Chandra, N. S., Wulff, E. G., Udayashankar, A. C., Nandini, B. P., Niranjana, S. R., Mortensen, C. N., & Prakash, H. S. (2011). Prospects of molecular markers in *Fusarium* species diversity. *Applied Microbiology and Biotechnology*, 90(5): 1625-1639.
- Chang, T. T. (2003). Origin, Domestication, and Diversification. In C. W. Smith & R. H. Dilday (Ed.), *Rice: Origin, History, Technology and Production* (pp 3-26). Canada: John Wiley & Sons, Inc.
- Chaudhary, R.C., Nanda, J. S., & Tran, D. V. (2002). *Guidelines For Identification of Field Constraints To Rice Production*. Rome: Food and Agriculture Organization of United Nation
- Chen, C., & Dickman, M. B. (2005). cAMP blocks MAPK activation and sclerotial development via Rap-1 in a PKA-independent manner in *Sclerotinia sclerotiorum*. *Molecular Microbiology*, 55(1): 299-311.
- Cho, Y., Cramer Jr, R. A., Kim, K. H., Davis, J., Mitchell, T. K., Figuli, P., Pryor, B. M., Lemasters, E., & Lawrence, C. B. (2007). The Fus3/Kss1 MAP kinase homolog Amk1 regulates the expression of genes encoding hydrolytic enzymes in *Alternaria brassicicola*. *Fungal Genetics and Biology*, 44(6): 543-553.
- Choi, W., & Dean, R. A. (1997). The adenylate cyclase gene MAC1 of *Magnaporthe grisea* controls appressorium formation and other aspects of growth and development. *The Plant Cell*, 9(11): 1973-1983.
- Chu, Z., Fu, B., Yang, H., Xu, C., Li, Z., Sanchez, A., Park, Y. J., Bennetzen, J. L., Zhang, Q., & Wang, S. (2006). Targeting xa13, a recessive gene for bacterial blight resistance in rice. *Theoretical and Applied Genetics*, 112(3): 455-461.
- Condon, B. J., Wu, D., Kraševc, N., Horwitz, B. A., & Turgeon, B. G. (2014). Comparative Genomics of *Cochliobolus* Phytopathogens. In R.A Dean, Lichens-Park, A. & Kole C. (Ed.), *Genomics of Plant-Associated Fungi: Monocot Pathogens* (pp. 41-67). Berlin Heidelberg: Springer.
- Consolo, V. F., Cordo, C. A., & Salerno, G. L. (2005). Mating-type distribution and fertility status in *Magnaporthe grisea* populations from Argentina. *Mycopathologia*, 160(4): 285-290.

- Coppin, E., Debuchy, R., Arnaise, S., & Picard, M. (1997). Mating types and sexual development in filamentous ascomycetes. *Microbiology and Molecular Biology Reviews*, 61(4): 411-428.
- Cother, E. J., Noble, D. H., Van De Ven, R. J., Lanoiselet, V., Ash, G., Vuthy, N., Visarto, P., & Stodart, B. (2010). Bacterial pathogens of rice in the Kingdom of Cambodia and description of a new pathogen causing a serious sheath rot disease. *Plant Pathology*, 59(5): 944-953.
- Crous, P. W., Groenewald, J. Z., Groenewald, M., Caldwell, P., Braun, U., & Harrington, T. C. (2006). Species of *Cercospora* associated with grey leaf spot of maize. *Studies in Mycology*, 55: 189-197.
- Cruz, A., Marín, P., González-Jaén, M. T., Aguilar, K. G. I., & Cumagun, C. J. R. (2013). Phylogenetic analysis, fumonisin production and pathogenicity of *Fusarium fujikuroi* strains isolated from rice in the Philippines. *Journal of the Science of Food and Agriculture*, 93(12): 3032-3039.
- Cui, R. Q., & Sun, X. T. (2012). First report of *Curvularia lunata* causing leaf spot on lotus in China. *Plant Disease*, 96(7): 1068-1068.
- Da Cunha, K. C., Sutton, D. A., Fothergill, A. W., Cano, J., Gené, J., Madrid, H., De Hoog, S., Crous, P. W., & Guarro, J. (2012). Diversity of *Bipolaris* species in clinical samples in the United States and their antifungal susceptibility profiles. *Journal of Clinical Microbiology*, 50(12): 4061-4066.
- Dai, S., & Beachy, R. N. (2009). Genetic engineering of rice to resist rice tungro disease. *In Vitro Cellular & Developmental Biology - Plant*, 45: 517-524.
- Dallagnol, L. J., Rodrigues, F. Á., Mielli, M. V. B., Ma, J. F., & Datnoff, L. E. (2008). Defective active silicon uptake affects some components of rice resistance to brown spot. *Phytopathology*, 99(1): 116-121.
- Dayakar, B. V., Narayanan, N. N., & Gnanamanickam, S. S. (2000). Cross-compatibility and distribution of mating type alleles of the rice blast fungus *Magnaporthe grisea* in India. *Plant disease*, 84(6): 700-704.
- Debuchy, R., & Turgeon, B. G. (2006). Mating-type Structure, Evolution, and Function in Euscomycetes. In U. Kües, & R. Fischer (Ed.), *Growth, Differentiation and Sexuality* (pp. 293-323). Germany: Springer.
- Dehpour, A. A., Alavi, S. V., & Majad, A. (2007). Light and scanning electron microscopy studies on the penetration and infection processes of *Alternaria alternata*, causing brown spot on Minneola Tangelo in the West Mazandaran-Iran. *World Applied Science Journal*, 2: 68-72.
- Dela Paz, M., Goodwin, P. H., Raymundo, A. K., Ardales, E. Y., & Vera Cruz, C. M. (2006). Phylogenetic analysis based on ITS sequences and conditions affecting the type of conidial germination of *Bipolaris oryzae*. *Plant Pathology*, 55: 756-765.

- Department of Agriculture (2013). *Paddy Statistics of Malaysia*. Department of Agriculture Peninsular Malaysia.
- Department of Agriculture (2014a). *Paddy Production Survey Report Malaysia Main Season 2012/2013*. Department of Agriculture Peninsular Malaysia.
- Department of Agriculture (2014b). *Paddy Statistics of Malaysia*. Department of Agriculture Peninsular Malaysia.
- Devendra, C., & Sevilla, C. C. (2002). Availability and use of feed resources in crop–animal systems in Asia. *Agricultural Systems*, 71: 59-73.
- Devendra, P. Y., & Abdul Aziz, A. R. (1994). Credit, technology and paddy farm production: a case of Tanjong Karang and Beranang, Malaysia. *The Developing Economics*, 32: 67-83.
- Donofrio, N. M., Hu, J., Mitchell, T. K., & Wilson, R. A. (2014). Facilitating the Fungus: Insights from the Genome of the Rice Blast Fungus, Magnaporthe Oryzae. In R. A. Dean, A. Lichen-Park & C. Kole (Ed.), *Genomics of Plant-Associated Fungi: Monocot Pathogens* (pp. 141-160). Berlin Heidelberg: Springer.
- Druka, A., & Hull, R. (1998). Variation of rice Tungro viruses: further evidence of two rice Tungro bacilliform virus strains and possibly several rice Tungro spherical virus variants. *Journal of Phytopathology*, 146(4): 175-178.
- Duong, T. A., de Beer, Z. W., Wingfield, B. D., & Wingfield, M. J. (2013). Characterization of the mating-type genes in *Leptographium procerum* and *Leptographium profanum*. *Fungal Biology*, 117(6): 411-421.
- Duveiller, E. M., & Sharma, R. C. (2012). Wheat Resistance to Spot Blotch or Foliar Blight. In I. Sharma (Ed.), *Disease Resistance in Wheat* (pp 120-135). India: CABI.
- Dyer, P. S., & O'Gorman, C. M. (2012). Sexual development and cryptic sexuality in fungi: insights from *Aspergillus* species. *FEMS Microbiology Reviews*, 36(1): 165-192.
- Fahmi, Z., Samah, B. A., & Abdullah, H. (2013). Paddy industry and paddy farmers well-being: a success recipe for agriculture industry in Malaysia. *Asian Social Science*, 9(3): 177.
- Fajolu, O. L., Vu, A. L., Dee, M. M., Zale, J., Gwinn, K. D., & Ownley, B. H. (2012). First report of leaf spot and necrotic roots on switchgrass caused by *Curvularia lunata* var. *aeria* in the United States. *Plant Disease*, 96(9): 1372-1372.
- Fan, Y., Ma, J., Gui, X., An, X., Sun, S., & Dong, J. (2007). Distribution of mating types and genetic diversity induced by sexual recombination in *Setosphaeria turcica* in northern China. *Frontiers of Agriculture in China*, 1(4): 368-376.

- Food and Agricultural Organization of United Nation, N.d. Retrived 27 November 2014 from <http://www.fao.org/docrep/t0567e/t0567e0d.htm>.
- Food and Agricultural Organization of United Nation, N.d. Retrived 30 August 2015 from <http://www.fao.org/docrep/005/y4347e/y4347e14.htm>.
- Ferguson, L. M., & Carson, M. L. (2004). Spatial diversity of *Setosphaeria turcica* sampled from the Eastern United States. *Phytopathology*, 94(8): 892-900.
- Ferreira, A. V. B., An, Z., Metzberg, R. L., & Glass, N. L. (1998). Characterization of mat A-2, mat A-3 and Δ matA mating-type mutants of *Neurospora crassa*. *Genetics*, 148(3): 1069-1079.
- Fitzgerald, M. A., McCouch, S. R., & Hall, R. D. (2009). Not just a grain of rice: the quest for quality. *Trends in Plant Science*, 14(3): 133-139.
- Gafur, A., Tanaka, C., Ouchi, S., & Tsuda, M. (1997). A PCR-based method for mating type determination in *Cochliobolus heterostrophus*. *Mycoscience*, 38(4): 455-458.
- Geiser, D. M., del Mar Jiménez-Gasco, M., Kang, S., Makalowska, I., Veeraraghavan, N., Ward, T. J., Zhang, N., Kuldau, G. A., & O'Donnell, K. (2004). FUSARIUM-ID v. 1.0: A DNA Sequence Database for Identifying Fusarium. In G. Mule, J. A. Bailey, B. M. Cooke, & A. Logrieco (Ed.), *Molecular Diversity and PCR-detection of Toxigenic Fusarium Species and Ochratoxigenic Fungi* (pp. 473-479). Netherlands: Springer.
- Gnanamanickam, S. S. (2009). *Biological Control of Rice Diseases*. USA: Springer Science & Business Media.
- Goddard, M. R., Godfray, H. C. J., & Burt, A. (2005). Sex increases the efficacy of natural selection in experimental yeast populations. *Nature*, 434(7033): 636-640.
- Groenewald, M., Groenewald, J. Z., Harrington, T. C., Abeln, E. C. A., & Crous, P. W. (2006). Mating type gene analysis in apparently asexual *Cercospora* species is suggestive of cryptic sex. *Fungal Genetics and Biology*, 43(12): 813-825.
- Groth, D. , & Fleet, L. (2003). Rice Diseases. In C. W. Smith & R. H. Didlay (Ed.), *Rice: Origin, History, Technology, and Production* (pp. 413- 436). USA: John Wiley & Sons Inc.
- Gu, S., Li, P., Wu, Min, Hao, Z., Gong, X., Zhang, X., Wang, Y., Cao, Z., Fan, Y., Han, J., & Dong, J. (2014). *StSTE12* is required for the pathogenicity of *Setosphaeria turcica* by regulating appressorium development and penetration. *Microbiological Research*, 169(11): 817-823.
- Guarro, J., Gené, J., & Stchigel, A. M. (1999). Developments in fungal taxonomy. *Clinical Microbiology Reviews*, 12(3): 454-500.

- Hawksworth, D. L. (2001). The magnitude of fungal diversity: the 1.5 million species estimate revisited. *Mycological Research*, 105(12): 1422-1432.
- Hibbett, D. S., Ohman, A., Glotzer, D., Nuhn, M., Kirk, P., & Nilsson, R. H. (2011). Progress in molecular and morphological taxon discovery in fungi and options for formal classification of environmental sequences. *Fungal Biology Reviews*, 25(1): 38-47
- Honda, Y., Ueki, M., Okada, G., Onose, R., Usami, R., Horikoshi, K., & Osada, H. (2001). Isolation, and biological properties of a new cell cycle inhibitor, curvularol, isolated from *Curvularia* sp. RK97-F166. *The Journal of Antibiotics*, 54(1): 10-16.
- Horbach, R., Navarro-Quesada, A. R., Knogge, W., & Deising, H. B. (2011). When and how to kill a plant cell: Infection strategies of plant pathogenic fungi. *Journal of Plant Physiology*, 168(1): 51-62.
- Hosokawa, M., Tanaka, C., & Tsuda, M. (2003). Conidium morphology of *Curvularia geniculata* and allied species. *Mycoscience*, 44(3): 227-237.
- Hubballi, M., Sornakili, A., Nakkeeran, S., Anand, T., & Raguchander, T. (2011). Virulence of *Alternaria alternata* infecting noni associated with production of cell wall degrading enzymes. *Journal of Plant Protection Research*, 51(1): 87-92.
- Index Fungorum (2015). N.d. Retrieved <http://www.indexfungorum.org/>.
- Indira, S., Xu, X., Iamsupasit, N., Shetty, H. S., Vasanthi, N. S., Singh, S. D., & Bandyopadhyay, R. (2002). Diseases of Sorghum and Pearl Millet in Asia. In J. F. Leslie (Ed.), *Sorghum and Millet Diseases* (pp 393-402). Iowa, USA: Blackwell Publishing.
- Izumitsu, K., Yoshimi, A., Kubo, D., Morita, A., Saitoh, Y., & Tanaka, C. (2009). The MAPKK kinase ChSte11 regulates sexual/asexual development, melanization, pathogenicity, and adaptation to oxidative stress in *Cochliobolus heterostrophus*. *Current Genetics*, 55(4): 439-448.
- Jia, Y., McAdams, S. A., Bryan, G. T., Hershey, H. P., & Valent, B. (2000). Direct interaction of resistance gene and avirulence gene products confers rice blast resistance. *The EMBO Journal*, 19(15): 4004-4014.
- Jones, M. C., Gough, K., Dasgupta, I., Rao, B. L. Subba, Cliffe, J., Qu, R., Shen, P., Kaniewska, M., Blakebrough, M., Davies, J. W., Beachy, R. N. & Hull, R. (1991). Rice tungro disease is caused by an RNA and a DNA virus. *Journal of General Virology*, 72(4): 757-761.
- Kato, H. (2001). Rice blast disease. *Pesticide Outlook*, 12(1): 23-25.

- Khush, G. S. (1997). Origin, Dispersal, Cultivation and Variation of Rice. In S. Takuji & M. Graham (Ed.), *Oryza: From Molecule To Plant* (pp. 25-34). Netherlands: Springer.
- Kim, H.-S., Han, K.-Y., Kim, K.-J., Han, D.-M., Jahng, K.-Y., & Chae, K.-S. (2002). The veA gene activates sexual development in *Aspergillus nidulans*. *Fungal Genetics and Biology*, 37(1): 72-80.
- Kirk, P. M., Cannon, P. F., Minter, D. W., & Stalpers, J.A. (2008). *Dictionary of the Fungi*. Wallingford, UK: CABI.
- Kobayashi, H., Sano, A., Aragane, N., Fukuoka, M., Tanaka, M., Kawaura, F., Fukuno, Y., Matsuishi, E., & Hayashi, S. (2008). Disseminated infection by *Bipolaris spicifera* in an immunocompetent subject. *Medical Mycology*, 46(4): 361-365.
- Kocal, N., Sonnewald, U., & Sonnewald, S. (2008). Cell wall-bound invertase limits sucrose export and is involved in symptom development and inhibition of photosynthesis during compatible interaction between tomato and *Xanthomonas campestris* pv *vesicatoria*. *Plant Physiology*, 148(3): 1523-1536.
- Kornerup, A., & Wanscher, J. H. (1983). *Methuen Handbook of Colour* (Third ed.). London, UK: Methuen.
- Kronstad, J. W., & Staben, C. (1997). Mating type in filamentous fungi. *Annual Review of Genetics*, 31(1): 245-276.
- Kubo, M., & Purevdorj, M. (2004). The future of rice production and consumption. *Journal of Food Distribution Research*, 35(1): 128-142.
- Larena, I., Salazar, O., González, V., Julián, M. C., & Rubio, V. (1999). Design of a primer for ribosomal DNA internal transcribed spacer with enhanced specificity for ascomycetes. *Journal of Biotechnology*, 75: 187-194.
- Latterell, F. M., & Rossi, A. E. (1983). Gray leaf spot of corn: a disease on the move. *Plant Disease*, 67(8): 842-847.
- Leach, J., Lang, B. R., & Yoder, O. C. (1982). Methods for selection of mutants and in vitro culture of *Cochliobolus heterostrophus*. *Journal of General Microbiology*, 128(8): 1719-1729.
- Lee, B.-M., Park, Y.-J., Park, D.-S., Kang, H.-W., Kim, J.-G., Song, E.-S., Park, I.-C., Yoon, U.-H., Hahn, J.-H., & Koo, B.-S. (2005). The genome sequence of *Xanthomonas oryzae* pathovar *oryzae* KACC10331, the bacterial blight pathogen of rice. *Nucleic Acids Research*, 33(2): 577-586.
- Lee, N., D'Souza, C. A., & Kronstad, J. W. (2003). Of smuts, blasts, mildews, and blights: cAMP signaling in phytopathogenic fungi. *Annual Review of Phytopathology*, 41(1): 399-427.

- Lengeler, K. B., Davidson, R. C., D'Souza, C. A., Harashima, T., Shen, W.-C., Wang, P., Pan, X., Waugh, M., & Heitman, J. (2000). Signal transduction cascades regulating fungal development and virulence. *Microbiology and Molecular Biology Reviews*, 64(4): 746-785.
- Leonard, K. J. (1976). Synonymy of *Exserohilum halodes* with *E. rostratum*, and induction of the ascigerous state, *Setosphaeria rostrata*. *Mycologia*, 68: 402–411.
- Leonard, K. J., & Suggs, E. G. (1974). *Setosphaeria prolata*, the ascigerous state of *Exserohilum prolatum*. *Mycologia*, 66: 281-297.
- Leslie, J. F., & Klein, K. K. (1996). Female fertility and mating type effects on effective population size and evolution in filamentous fungi. *Genetics*, 144: 557–567.
- Leslie, J. F., & Summerell, B. A. (2006). *The Fusarium Laboratory Manual*. UK: Blackwell Publishing Ltd.
- Leslie, J. F., Summerell, B. A., Bullock, S., & Doe, F. J. (2005). Description of *Gibberella sacchari* and neotypification of its anamorph *Fusarium sacchari*. *Mycologia*, 97(3): 718-724.
- Leung, H., Borromeo, E. S., Bernardo, M. A., & Notteghem, J. L. (1988). Genetic analysis of virulence in the rice blast fungus *Magnaporthe grisea*. *Phytopathology*, 78(9): 1227-1233.
- Li, D. M., & de Hoog, G. S. (2009). Cerebral phaeohyphomycosis—a cure at what lengths? *The Lancet Infectious Diseases*, 9(6): 376-383.
- Lin, S.-H., Huang, S.-L., Li, Q.-Q., Hu, C.-J., Fu, G., Qin, L.-P., Ma, Y.-F., Xie, L., Cen, Z.-L., & Yan, W.-H. (2011). Characterization of *Exserohilum rostratum*, a new causal agent of banana leaf spot disease in China. *Australasian Plant Pathology*, 40(3): 246-259.
- Litvintseva, A. P., Marra, R. E., Nielsen, K., Heitman, J., Vilgalys, R., & Mitchell, T. G. (2003). Evidence of sexual recombination among *Cryptococcus neoformans* serotype A isolates in sub-Saharan Africa. *Eukaryotic Cell*, 2(6): 1162-1168.
- Loro, M., Valero-Jiménez, C. A., Nozawa, S., & Márquez, L. M. (2012). Diversity and composition of fungal endophytes in semiarid Northwest Venezuela. *Journal of Arid Environments*, 85: 46-55.
- Luan, F. G., Qiang, S., Ma, D.Y., & Riziwangguli. (2004). The primary studies on isolation and identification of wheat black embryo in Xinjiang. *Xinjiang Agriculture Science*, 41(5): 357-360.

- Maclean, J. , Hardy, B., & Hettel, G. (2013). *Rice Almanac, 4th edition: Source Book for One of the Most Important Economic Activities on Earth* (Fourth ed.). Phillippine: Global Rice Science Partnership.
- Maji, A., & Nath, R. (2015). Pathogenicity tset by using different inoculation methods on *Xanthomonas campestri* pv *campestris* caused of black rot of cabbage. *IMPACT: International Journal of Research in Applied, Natural and Social Sciences*, 3(2): 53-58.
- Manamgoda, D. S., Cai, L., Bahkali, A. H., Chukeatirote, E., & Hyde, K. D. (2012). A phylogenetic and taxonomic re-evaluation of the *Bipolaris* - *Cochliobolus* - *Curvularia* Complex. *Fungal Diversity*, 56: 131-144.
- Manamgoda, D. S., Cai, L., Bahkali, A. H., Chukeatirote, E., & Hyde, K. D. (2011). *Cochliobolus*: an overview and current status of species. *Fungal Diversity*, 51(1): 3-42.
- Manamgoda, D.S., Rossman, A. Y., Castlebury, L. A., Crous, P. W., Madrid, H., Chukeatirote, E., & Hyde, K. D. (2014). The genus *Bipolaris*. *Studies in Mycology*, 79:221-288.
- Martin, R. R., James, D., & Lévesque, C. A. (2000). Impacts of molecular diagnostic technologies on plant disease management. *Annual Review of Phytopathology*, 38(1): 207-239.
- Masratul Hawa, M. , Salleh, B., & Latiffah, Z. (2009). First report of *Curvularia lunata* on red-fleshed dragon fruit (*Hylocereus polyrhizus*) in Malaysia. *Plant Disease*, 93(9): 971-971.
- Mathre, D. E., Johnston, R. H., & Grey, W. E. (2003). Diagnosis of common root rot of wheat and barley. *Plant Health Progress* 10. doi: 10.1094/PHP-2003-0819-01-DG.
- McCartney, H. A., Foster, S. J., Fraaije, B. A., & Ward, E. (2003). Molecular diagnostics for fungal plant pathogens. *Pest Management Science*, 59(2): 129-142.
- Meldrum, S. I., Platz, D. G. J., & Ogle, H. J. (2004). Pathotypes of *Cochliobolus sativus* on barley in Australia. *Australasian Plant Pathology*, 33(1): 109-114.
- Mew, T. M., & Gonzales, P. (2002). *A Handbook of Rice Seedborne Fungi*. Los Bafios (Philippines): International Rice Research Institute.
- Michereff, S. J., Silveira, N. S. S., Reis, A., & Mariano, R. L. R. (1994). Epiphytic bacteria antagonistic to *Curvularia* leaf spot of yam. *Microbial Ecology*, 28(1): 101-110.
- Moore-Landecker, E. (1992). Physiology and biochemistry of ascocarp induction and development. *Mycological Research*, 96(9): 705-716.

- Morita, A., Saitoh, Y., Izumitsu, K., & Tanaka, C. (2012a). Teleomorph formation of *Setosphaeria monoceras*, a perfect state of *Exserohilum monoceras*, by Japanese isolates. *Mycoscience*, 53(2): 144-146.
- Morita, A., Saitoh, Y., Izumitsu, K., & Tanaka, C. (2012b). Molecular organization of the mating type (Mat) locus of *Exserohilum monoceras* (*Setosphaeria monoceras*), a bioherbicide agent for *Echinochloa* weeds. *Mycoscience*, 53(2): 92-101.
- Motlagh, M. R. S. (2011). Evaluation of *Curvularia lunata* as a biological control agent in major weeds of rice paddies. *Life Science Journal*, 8(2): 81-91.
- Motlagh, M. R. S., & Kavian, B. (2008). Characterization of new *Bipolaris* Spp.: the causal agent of rice Brown Spot Disease in the North of Iran. *International Journal of Agriculture and Biology*, 10: 638-642.
- Msikita, W., Yaninek, J. S., Ahounou, M., Baimey, H., & Fagbemissi, R. (1997). First report of *Curvularia lunata* associated with stem disease of cassava. *Plant Disease*, 81(1): 112-112.
- Mulè, G., González-Jaén, M. T., Hornok, L., Nicholson, P., & Waalwijk, C. (2005). Advances in molecular diagnosis of toxigenic *Fusarium* species: A review. *Food Additives & Contaminants*, 22(4): 316-323.
- Muto, M., Mulabagal, V., Huang, H. C., Takahashi, H., Tsay, H. S., & Huang, J. W. (2006). Toxicity of black nightshade (*Solanum nigrum*) extracts on *Alternaria brassicicola*, causal agent of black leaf spot of Chinese cabbage (*Brassica pekinensis*). *Journal of Phytopathology*, 154(1): 45-50.
- Narayanasamy, P. (2008). *Molecular Biology in Plant Pathogenesis and Disease Management: Disease Management* (Vol. 3). Netherlands: Springer Science & Business Media.
- Nilsson, R. H., Kristiansson, E., Ryberg, M., Hallenberg, N., & Larsson, K.-H. (2008). Intraspecific ITS variability in the kingdom Fungi as expressed in the international sequence databases and its implications for molecular species identification. *Evolutionary Bioinformatics Online*, 4: 193-201.
- Nithiyaa, P., Nur Ain Izzati, M. Z., Umi Kalsom, Y., & Salleh, B. (2012). Diversity and morphological characteristics of *Aspergillus* species and *Fusarium* species isolated from cornmeal in Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 35(1): 103-116.
- Nur Ain Izzati, M. Z., Azmi, A. R., & Salleh, B. (2008). Bakanae disease of rice in Malaysia and Indonesia: etiology of the causal agent based on morphological, physiological and pathogenicity characteristics. *Journal of Plant Protection Research*, 48(4): 475-485.

- Nur Ain Izzati, M. Z., Azmi, A. R., & Baharuddin, S. (2008). Secondary metabolite profiles and mating populations of *Fusarium* species in section *Lesiola* associated with bakanae disease of rice. *Malaysian Journal of Microbiology*, 4(1): 6-13.
- Nur Ain Izzati, M. Z., & Wan Hasmida, W. I. (2011). Isolation of microfungi from malay traditional vegetables and secondary metabolites produced by *Fusarium* species. *Sains Malaysiana*, 40:437-444.
- Nur Ain Izzati, M. Z., Wu, D., & Turgeon, B. G. (2013). *Deletion of CPS1 Reduces Virulence of Cochliobolus miyabeanus, Causal Agent of Brown Spot Disease in Rice*. Paper presented at the International Congress of the Malaysian Society for Microbiology 2013 (ICMSM2013), Langkawi Lagoon Resort, Malaysia.
- O'Brien, H. E., Parrent, J. L., Jackson, J. A., Moncalvo, J.-M., & Vilgalys, R. (2005). Fungal community analysis by large-scale sequencing of environmental samples. *Applied and Environmental Microbiology*, 71(9): 5544-5550.
- Oerke, E. C., Dehne, H. W., Schönbeck, F., & Weber, A. (2012). *Crop Production and Crop Protection: Estimated Losses in Major Food and Cash Crops*. Netherlands: Elsevier.
- Ohm, R. A., Feau, N., Henrissat, B., Schoch, C. L., Horwitz, B. A., Barry, K. W., Condon, B. J., Copeland, A. C., Dhillon, B., Glaser, F., Nesse, C. N., Kosti, I., LaButti, K., Lindquist, E. A., Lucas, S., Salamov, A. A., Bradshaw, R. E., Ciuffetti, L., Hamelin, R. C., Kema, G. H. J., Lawrence, C., Scott, J. A., Spatafora, J. W., Turgeon, B. G., de Wit, P.J.G.M., Zhong, S., Goodwin, S. B., & Grigoriev, I. V. (2012). Diverse lifestyles and strategies of plant pathogenesis encoded in the genomes of eighteen Dothideomycetes fungi. *PLoS Pathogens*, 8(12): e1003037.
- Oide, S., Liu, J., Yun, S.-H., Wu, Dongliang, Michev, A., Choi, M. Y., Horwitz, B. A., & Turgeon, B. G. (2010). Histidine kinase two-component response regulator proteins regulate reproductive development, virulence, and stress responses of the fungal cereal pathogens *Cochliobolus heterostrophus* and *Gibberella zeae*. *Eukaryotic cell*, 9(12): 1867-1880.
- Oliver, R. P., Lord, M., Rybak, K., Faris, J. D., & Solomon, P. S. (2008). Emergence of tan spot disease caused by toxigenic *Pyrenophora tritici-repentis* in Australia is not associated with increased deployment of toxin-sensitive cultivars. *Phytopathology*, 98(5): 488-491.
- Omar, O. (2008). *Rice Production and Potential For Hybrid Rice in Malaysia*. Paper presented at the International Plantation Industry Conference and Expo - IPiCEX 2008, Shah Alam.
- Paoletti, M., Seymour, F. A., Alcocer, M. J. C., Kaur, N., Calvo, A. M., Archer, D. B., & Dyer, P. S. (2007). Mating type and the genetic basis of self-fertility in the model fungus *Aspergillus nidulans*. *Current Biology*, 17(16): 1384-1389.

- Pöggeler, S., Nowrousian, M., & Kück, U. (2006). Fruiting-body Development in Ascomycetes. In U. Kries & R. Fischer (Ed.), *Growth, Differentiation and Sexuality* (pp. 325-355). Germany: Springer.
- Pratt, R. G. (2000). Diseases caused by dematiaceous fungal pathogens as potential limiting factors for production of bermudagrass on swine effluent application sites. *Agronomy Journal*, 92(3): 512-517.
- Pratt, R. G. (2003). An excised-leaf inoculation technique for evaluating host-pathogen interactions and quantitative resistance of bermudagrass genotypes to dematiaceous Hyphomycetes. *Phytopathology*, 93(12): 1565-1571.
- Pratt, R. G. (2005). Variation in occurrence of dematiaceous Hyphomycetes on forage bermudagrass over years, sampling times, and locations. *Phytopathology*, 95(10): 1183-1190.
- Pratt, R. G. (2006). Johnsongrass, yellow foxtail, and broadleaf signalgrass as new hosts for six species of *Bipolaris*, *Curvularia*, and *Exserohilum* pathogenic to bermudagrass. *Plant Disease*, 90(4): 528-528.
- Pratt, R. G., & Brink, G. E. (2007). Forage bermudagrass cultivar responses to inoculations with relationships to field persistence. *Crop Science*, 47(1): 239-244.
- Preeti, V., S., Kavitha, V Brindha, P., Babujee, L., & Gnanamanickam, S. S. (2002). Biological Control of Rice Diseases. In S. S. Samuel (Ed.), *Biological Control of Crop Diseases* (pp 11-32). USA: Marcel Dekker Inc.
- Raju, N. B., & Perkins, D. D. (1978). Barren perithecia in *Neurospora crassa*. *Canadian Journal of Genetics and Cytology*, 20(1): 41-59.
- Rau, D., Maier, F. J., Papa, R., Brown, A. H. D., Balmas, V., Saba, E., Schaefer, W., & Attene, G. (2005). Isolation and characterization of the mating-type locus of the barley pathogen *Pyrenophora teres* and frequencies of mating-type idiomorphs within and among fungal populations collected from barley landraces. *Genome*, 48(5): 855-869.
- Reddy, P. P. (2015). *Climate Resilient Agriculture for Ensuring Food Security*. India: Springer.
- Reyna, N. S., & Yang, Y. (2006). Molecular analysis of the rice MAP kinase gene family in relation to *Magnaporthe grisea* infection. *Molecular Plant-Microbe Interactions*, 19(5): 530-540.
- Rossmann, A. Y., & Palm-Hernández, M. E. (2008). Systematics of plant pathogenic fungi: why it matters. *Plant Disease*, 92(10): 1376-1386.
- Ruger-Herreros, C., Rodríguez-Romero, J., Fernández-Barranco, R., Olmedo, M., Fischer, R., Corrochano, L. M., & Canovas, D. (2011). Regulation of conidiation by light in *Aspergillus nidulans*. *Genetics*, 188(4): 809-822.

- Saha, A., Dasgupta, S., & Saha, D. (2001). Discovery of *Curvularia eragrostidis* on tea (*Camellia sinensis*(L.) O. Ktze) leaves from clonal-cutting nurseries in North Bengal. *Environment and Ecology*, 19(4): 846-848.
- Salleh B., & Sulaiman B. (1984). *Fusarium* associated with naturally diseases plants in Penang. *Journal of Plant Protection Tropics*, 1:47-53.
- Salleh, B., Safinat, A., Julia, L., & Teo, C. H. (1996). Brown spot caused by *Curvularia* spp., a new disease of asparagus. *BIOTROPIA-The Southeast Asian Journal of Tropical Biology*, 9: 26-37.
- Sayler, R. J., Cartwright, R. D., & Yang, Y. (2006). Genetic characterization and real-time PCR detection of *Burkholderia glumae*, a newly emerging bacterial pathogen of rice in the United States. *Plant Disease*, 90(5): 603-610.
- Schafer, M., & Kotanen, P. M. (2004). Impacts of naturally-occurring soil fungi on seeds of meadow plants. *Plant Ecology*, 175(1): 19-35.
- Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., Chen, W., Bolchacova, E., Voigt, K., & Crous, P. W. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences*, 109(16): 6241-6246.
- Schubler, A., Schwarzott, D., & Walker, C. (2001). A new fungal phylum, the Glomeromycota: phylogeny and evolution. *Mycological Research*, 105(12): 1413-1421.
- Selvaraj, K., & Fofana, B. (2012). An Overview of Plant Photosynthesis Modulation by Pathogen Attacks. In N. Mohammad Mahdi (Ed.), *Advances in Photosynthesis-Fundamental Aspects*. Croatia: INTECH Open Access Publisher.
- Sharma, K., Goss, E. M., Dickstein, E. R., Smith, M. E., Johnson, J. A., Southwick, F. S., & van Bruggen, A. H. C. (2014). *Exserohilum rostratum*: Characterization of a cross-kingdom pathogen of plants and humans. *PLoS one*, 9(10): e108691.
- Shen, S., Hao, Z., Gu, S., Wang, J., Cao, Z., Li, Z., Wang, Q., Li, P., Hao, J., & Dong, J. (2013). The catalytic subunit of cAMP-dependent protein kinase A StPKA-c contributes to conidiation and early invasion in the phytopathogenic fungus *Setosphaeria turcica*. *FEMS Microbiology Letters*, 343(2): 135-144.
- Sivanesan, A. (1987). *Graminicolous Species of Bipolaris, Curvularia, Drechslera, Exserohilum and Their Teleomorphs* (Vol. 159). Wallingford, UK: C.A.B. International.
- Staben, C. (1995). Sexual Reproduction in Higher Fungi. In N. A. R. Gow & G. M. Gadd (Ed.), *The Growing Fungus* (pp. 383-422). London: Chapman and Hall.

- Steenkamp, E. T., Wingfield, B. D., Coutinho, T. A., Zeller, K. A., Wingfield, M. J., Marasas, W. F.O., & Leslie, J. F. (2000). PCR-based identification of *MAT-1* and *MAT-2* in the *Gibberella fujikuroi* species complex. *Applied and Environmental Microbiology*, 66(10): 4378-4382.
- Takano, Y., Kikuchi, T., Kubo, Y., Hamer, J. E., Mise, K., & Furusawa, I. (2000). The *Colletotrichum lagenarium* MAP kinase gene CMK1 regulates diverse aspects of fungal pathogenesis. *Molecular Plant-Microbe Interactions*, 13(4): 374-383.
- Tan, R. X., Jensen, P. R., Williams, P. G., & Fenical, W. (2004). Isolation and structure assignments of rostratins A–D, cytotoxic disulfides produced by the marine-derived fungus *Exserohilum rostratum*. *Journal of Natural Products*, 67(8): 1374-1382.
- Tanaka, Y., Murata, N., & Kato, H. (1979). Behavior of nuclei and chromosomes during ascus development in the mating between either rice-strain or weeping lovegrass-strain and ragi-strain of *Pyricularia*. *Annual Phytopathology Society of Japan*, 45(2): 182-191.
- Taylor, J. W., Geiser, D. M., Burt, A., & Koufopanou, V. (1999). The evolutionary biology and population genetics underlying fungal strain typing. *Clinical Microbiology Reviews*, 12(1): 126-146.
- Taylor, J. W., Jacobson, D. J., Kroken, S., Kasuga, T., Geiser, D. M., Hibbett, D. S., & Fisher, M. C. (2000). Phylogenetic species recognition and species concepts in fungi. *Fungal Genetics and Biology*, 31: 21-32.
- TeBeest, D.O., Guerber C. & Ditmore M. (2007). Rice blast. *The Plant Health Instructor*. doi: 10.1094/PHI-I-2007-0313-07
- Timmer, C. P. (2014). Food security in Asia and the Pacific: The rapidly changing role of rice. *Asia & the Pacific Policy Studies*, 1(1): 73-90.
- Tosiah, S., Kadir, J., Sariah, M., Juraimi, A. S., Lo, N. P., & Soetikno, S. (2009). Survey and evaluation of native fungal pathogens for biocontrol of barnyard grass (*Echinochloa crus-galli* complex). *Journal of Tropical Agriculture and Food Science*, 37(1): 119-128.
- Tripati, D. P. (2006). *Introductory Mycology*. India: Kalyani Publishers.
- Truong, H. X., & Gergon, E. B. (2014). Current Reserach on Fungal Pathogens Associated with Rice. In J. K. Misra, P. T. Jalpa, K. D. Sunil & V. Csaba (Ed.), *Fungi From Different Substrates* (pp 260-308). U.S: CRC Press, Taylor & Francis Group.
- Turgeon, B. G. (1998). Application of mating type gene technology to problems in fungal biology. *Annual Review of Phytopathology*, 36: 115-137.

- Turgeon, B. G., & Yoder, O. C. (2000). Proposed nomenclature for mating type genes of filamentous ascomycetes. *Fungal Genetics and Biology*, 31(1): 1-5.
- Ueyama, A., & Tsuda, M. (1975). Culture conditions and the formation of perfect stage of *Helminthosporium oryzae*, *Cochliobolus miyabeanus*. *Annual Phytopathology Society of Japan*, 41: 447-452.
- Viji, G., & Uddin, W. (2002). Distribution of mating type alleles and fertility status of *Magnaporthe grisea* causing gray leaf spot of perennial ryegrass and St. Augustinegrass Turf. *Plant Disease*, 86(8): 827-832.
- Ward, E., Foster, S. J., Fraaije, B. A., & McCartney, H. A. (2004). Plant pathogen diagnostics: immunological and nucleic acid-based approaches. *Annals of Applied Biology*, 145(1): 1-16.
- Ward, J. M. J., Stromberg, E. L., Nowell, D. C., & Nutter, F. W. (1999). Gray leaf spot: a disease of global importance in maize production. *Plant Disease*, 83(10): 884-895.
- Wei, H., Requena, N., & Fischer, R. (2003). The MAPKK kinase SteC regulates conidiophore morphology and is essential for heterokaryon formation and sexual development in the homothallic fungus *Aspergillus nidulans*. *Molecular Microbiology*, 47(6): 1577-1588.
- Westcott, C. (2001). *Westcott's Plant Disease Handbook* (6th Ed.). United State of America: Kluwer Academic Publisher.
- White, T. J. , Bruns, T., Lee, S., & Taylor, J. (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In M. A. Innis, D. H. Gelfand, J. J. Sninsky & T. J. White (Eds.), *PCR Protocols : A guide to Methods and Applications* (pp. 315-322). San Diego, U: Academic Press.
- Wijayawardene, N.N., Crous, P.W., Kirk, P.M., Hawksworth, D.L., Boonmee, S., Braun, U., Chomnunti, P., Dai, D.Q., D'souza, M.J., Diederich, P., Dissanayake, A., Doilom, M., Hongsanan, S., Jones, E.B.G., Groenewald, J.Z., Jayawardena, R., Lawrey, J.D., Liu, J.K., Lücking, R., Madrid, H., Manamgoda, D.S., Muggia, L., Nelsen, M.P., Phookamsak, R., Suetrong, S., Tanaka, K., Thambugala, K.M., Wikee, S., Zhang, Y., Aptroot, A., Ariyawansa, H.A., Bahkali, A.H., Bhat, J.D., Gueidan, C., De Hoog, G.S., Knudsen, K., McKenzie, E.H.C., Miller, A.N., Mortimer, P.E., Wanasinghe, D.N., Phillips, A.J.L., Raja, H.A., Slippers, B., Shivas, R.S., Taylor, J.E., Wang, Y., Woudenberg, J.H.C., Piątek, M., Cai, L., Jaklitsch, W.M., & Hyde, K.D. (2014). Naming and outline of dothideomycetes–2014 including proposals for the protection or suppression of generic names. *Fungal Diversity*, 69(1):1–55.
- Wirsel, S., Horwitz, B., Yamaguchi, K., Yoder, O. C., & Turgeon, B. G. (1998). Single mating type-specific genes and their 3' UTRs control mating and fertility in *Cochliobolus heterostrophus*. *Molecular and General Genetics MGG*, 259(3): 272-281.

- Wu, D., & Turgeon, B. G. (2013). *Setosphaeria rostrata*: Insights from the sequenced genome of *Setosphaeria turcica*. *Fungal Genetics and Biology*, 61: 158-163.
- Wu, D., Oide, S., Zhang, N., Choi, M.Y., & Turgeon, B. G. (2012). ChLae1 and ChVel1 regulate T-toxin production, virulence, oxidative stress response, and development of the maize pathogen *Cochliobolus heterostrophus*. *PLoS Pathogens*, 8(2): e1002542.
- Wulff, E. G., Sorensen, J. L., Lübeck, M., Nielsen, K. F., Thrane, U., & Torp, J. (2010). *Fusarium* spp. associated with rice Bakanae: ecology, genetic diversity, pathogenicity and toxigenicity. *Environmental Microbiology*, 12(3): 649-657.
- Xu, J. (2006). Fundamentals of fungal molecular population genetic analyses. *Current Issues in Molecular Biology*, 8(2): 75.
- Zhang, S.-R., Hao, Z.-M., Wang, L.-H., Shen, S., Cao, Z.-Y., Xin, Y.-Y., Hou, M.-L., Gu, S.-Q., Han, J.-M., & Dong, J.-G. (2012). StRas2 regulates morphogenesis, conidiation and appressorium development in *Setosphaeria turcica*. *Microbiological Research*, 167(8): 478-486.
- Zhong, S., & Steffenson, B. J. (2001). Virulence and molecular diversity in *Cochliobolus sativus*. *Phytopathology*, 91(5): 469-476.
- Zhu, Y., & Qiang, S. (2004). Isolation, pathogenicity and safety of *Curvularia eragrostidis* isolate QZ-2000 as a bioherbicide agent for large crabgrass (*Digitaria sanguinalis*). *Biocontrol Science and Technology*, 14(8): 769-782.

LIST OF PUBLICATIONS

Journal:

Nor Azizah Kusai, Madihah Mior Zakuan Azmi, Shahrizim Zulkifly, Mohd Termizi Yusof, and Nur Ain Izzati Mohd Zainudin. (2015). Morphological and molecular characterization of *Curvularia* and related species associated with leaf spot disease of rice in Peninsular Malaysia. Published online in *Rendiconti Lincei*. DOI 10.1007/s12210-015-0458-6.

Nor Azizah Kusai, Madihah Mior Zakuan Azmi, Mohd Termizi Yusof, Azmi Abd Razak and Nur Ain Izzati Mohd Zainudin (2015). Morphological, molecular, virulence and development of *Setosphaeria rostrata* associated with leaf spot of rice. Accepted with major correction in *Mycologia*.

Proceedings:

Nor Azizah Kusai, Mohd Termizi Yusof, Shahrizim, Zulkifly, Azmi Abd Razak, and Nur Ain Izzati Mohd Zainudin (2014). *Morphological Characterization of Setosphaeria rostrata and Cochliobolus species Isolated from Diseased-Rice Leaves Samples in Selangor*. Paper presented at the Malaysia International Biological Symposium (i-SIMBIOMAS 2014), Palm Garden Hotel, Putrajaya.

Nor Azizah Kusai, Mohd Termizi Yusof and Nur Ain Izzati Mohd Zainudin (2014). *Mating Type Analyses of Setosphaeria rostrata Isolated From Brown Spot Disease of Rice in Peninsular Malaysia*. Paper presented at the Monash Science Symposium 2014, Monash University Campus Bandar Sunway.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT :

PHENOTYPIC, MOLECULAR AND MATING TYPE ANALYSES OF *Exserohilum rostratum*
ASSOCIATED WITH LEAF SPOTS OF RICE

NAME OF STUDENT : NOR AZIZAH BINTI KUSAI

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]