



**UNIVERSITI PUTRA MALAYSIA**

***TISSUE CULTURE, ITS MORPHOLOGICAL AND BIOCHEMICAL  
INTEGRITY AND ANTIOXIDANT PROFILING ON  
Cucumis sativus L. cv. MTi2***

**TG AZIA FARAHIN KU HASAN**

**FS 2015 51**



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INTEGRITY AND ANTIOXIDANT PROFILING ON  
*Cucumis sativus* L. cv. MTi2**

**By**

**TG AZIA FARAHIN KU HASAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfillment of the Requirement for the Degree of Master of  
Science**

**November 2015**

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## DEDICATION

Special dedication goes to my loving parents, families, my supervisor, my co-supervisor, and fellow friends for all their continuous support, guidance, motivation and love.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

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**November 2015**

**Chairman : Rosimah Nulit, PhD**  
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Cucumber is one of the high-nutritional vegetable and is one of the most important crops worldwide. It is rich in vitamin B, vitamin C, proteins, minerals, amino acids and many other constituents. It is beneficial for people to consume cucumber which offer health benefits and aid in disease prevention such as cancer. Cucumber variety MTi2 is one of the famous cucumber variety in Malaysia that consumed by local people. Therefore it is important to maintain the good supply of this favorable variety and to conserve the species to ensure its continuance in the future. Also, the outbreak of viral disease prompts us to look alternative method to safe keep viable stock plant. Besides, the conventional means of propagation commonly yield non uniform plants that may reduce the crop quality. This might be a huge problem for large scale multiplication industries and for conservation purpose of the species. As an alternative, *in vitro* regeneration was carry out to optimize the protocol in obtaining uniform plantlet through culturing shoot apical meristem (SAM) of cucumber. Nevertheless, *in vitro* regeneration is also associated with somaclonal variation that may result of producing non uniform plantlet. Therefore the SAM culture was optimized in this study to generate identical plantlet. The effect of plant growth regulators (PGR) in *in vitro* regeneration was also studied through application of 10 treatments of full strength MS media supplemented with different concentration and combination of PGR, with MS basal media served as control treatment. The result showed that meristem tissue of cucumber can be regenerated into whole plant through *in vitro* micropropagation. The combination treatment of 0.01 mgL<sup>-1</sup> IAA and 0.1 mgL<sup>-1</sup> KIN produced highest percentage survival of plantlets which was 88%. Comparative study was done in term of morphology, anatomy, biochemical and antioxidant profiling between cucumber plant and cucumber plantlet. The finding showed that the plantlet possess similar characteristics as the cucumber plant in term of morphology, anatomy, total protein content, total phenolic content and total flavonoid content. In conclusion, identical plantlet can be produced through *in vitro* regeneration of SAM of cucumber, thus this technique can be

implemented for conservation and for large-scale multiplication purpose. This study had also identify and compared the antioxidant activity between seed, young fruit and matured fruit of cucumber. Antioxidant play vital roles in neutralize the effects of free radicals where free radicals are molecules that can damage the cellular components in human body. Phenolic and flavonoid are examples of antioxidant properties measured in this study. The finding showed that the seed of cucumber possess considerate amount of antioxidant properties which is 0.2 mg/ml flavonoid content and 0.02 mg/ml phenolic content. Therefore, the seed of cucumber can be utilized as a dietary source of natural antioxidant.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**Kultur Tisu, Integriti Morfologi dan biokimia serta Profil Antioksida ke  
atas *Cucumis sativus* L. cv. MTi2**

Oleh

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Timun merupakan salah satu sayur-sayuran yang tinggi nutrisi dan merupakan salah satu tanaman yang paling penting di seluruh dunia. Ia kaya dengan vitamin B, vitamin C, protein, mineral, asid amino dan lain-lain unsur. Pengambilan timun adalah bermanfaat kepada manusia yang memberi manfaat kesihatan dan membantu mencegah penyakit seperti penyakit kanser. Timun MTi2 adalah salah satu variasi timun yang terkenal di Malaysia yang dimakan oleh masyarakat tempatan. Oleh itu, adalah penting untuk mengekalkan bekalan yang baik dari variasi timun ini dan untuk memulihara spesies untuk memastikan kelangsungannya pada masa akan datang. Juga, wabak virus penyakit memerlukan kami untuk mencari kaedah alternatif untuk menyimpan tumbuhan yang berdaya maju dengan selamat. Selain itu, cara penghasilan secara konvensional biasanya menghasilkan tumbuhan yang tidak seragam yang boleh mengurangkan kualiti tanaman. Ini mungkin menjadi satu masalah yang besar kepada industri penghasilan tanaman skala besar dan kepada pemuliharaan spesies ini. Sebagai alternatif, pertumbuhan semula *in vitro* dilakukan untuk mengoptimumkan protokol dalam menghasilkan anak pokok yang seragam dengan mengkulturkan meristem apikal pucuk timun. Walau bagaimanapun, pertumbuhan semula *in vitro* adalah berkait dengan perubahan somaklonal yang boleh menyebabkan penghasilan anak pokok yang tidak seragam. Oleh itu, pengkulturan SAM telah dioptimumkan dalam kajian ini untuk menghasilkan anak pokok yang sama. Kesan daripada pengawal selia pertumbuhan tumbuhan (PGR) dalam pertumbuhan semula *in vitro* juga telah dikaji melalui penggunaan 10 rawatan MS media dengan kekuatan penuh ditambah dengan kepekatan dan kombinasi PGR yang berbeza, dengan media basal MS bertindak sebagai rawatan kawalan. Keputusan menunjukkan tisu meristem timun boleh ditumbuhkan semula menjadi keseluruhan pokok melalui mikropropagasi *in vitro*. Rawatan kombinasi  $0.01 \text{ mgL}^{-1}$  IAA dan  $0.1 \text{ mgL}^{-1}$  KIN menghasilkan peratusan kelangsungan hidup anak pokok yang tertinggi iaitu 88%. Kajian perbandingan telah dilakukan dari segi morfologi, anatomi, biokimia dan profil antioksida ke atas pokok

timun dan klon timun. Hasil kajian menunjukkan bahawa klon-klon menunjukkan ciri-ciri yang sama seperti pokok timun dari segi morfologi, anatomi, jumlah kandungan protein, jumlah kandungan fenolik dan jumlah kandungan flavonoid. Sebagai kesimpulan, klon yang serupa boleh dihasilkan melalui pertumbuhan semula *in vitro* menggunakan meristem apikal pucuk timun, oleh itu teknik ini boleh diaplikasikan untuk tujuan pemuliharaan dan penghasilan tanaman berskala besar. Kajian ini juga telah mengenalpasti dan membanding profil antioksidan pada biji, buah muda dan buah matang timun. Antioksidan memainkan peranan penting dalam meneutralkan kesan radikal bebas yang mana radikal bebas adalah molekul yang boleh memusnahkan komponen selular dalam badan manusia. Fenolik dan flavonoid adalah contoh-contoh antioksida yang diukur dalam kajian ini. Hasil kajian menunjukkan bahawa biji timun mengandungi jumlah antioksidan yang boleh dipertimbangkan iaitu 0.2 mg/ml kandungan flavonoid dan 0.02 mg/ml kandungan fenolik. Oleh itu, bahagian biji timun boleh diambil sebagai sumber antioksidan semula jadi.



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I certify that a Thesis Examination Committee has met on (date of viva voce) to conduct the final examination of Tg Azia Farahin Ku Hasan on her thesis entitled

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## LIST OF ABBREVIATIONS

SAM	shoot apical meristem
°C	degree Celsius
%	percentage
g	gram
mg	milligram
PGR	plant growth regulator
MS	Murashige and Skoog
DNA	Deoxyribonucleic acid
IAA	Indole-3-Acetic Acid
KIN	Kinetin
ml	milliliter
ppm	parts per million
N	normal
NaOH	sodium hypochlorite
pH	negative logarithm of hydrogen ion concentration
Mg <sup>L</sup> <sup>-1</sup>	milligram per liter
mm	millimeter
psi	pounds per square inch
v/v	volume per volume
LAF	laminar air flow
ANOVA	Analysis of Variance
g <sup>L</sup> <sup>-1</sup>	gram per liter
CRD	completely randomized design
cm	centimeter

min	minute
cm <sup>3</sup>	cubic centimeters
DPX	diputal petroleum xylene
nm	nanometer
μl	micrometer
H <sub>2</sub> SO <sub>4</sub>	sulfuric acid
mg/ml	milligram per milliliter
rpm	revolutions per minute
mM	millimolar
SE	standard error
DPPH	2, 2-diphenyl-1-picrylhydrazyl
μl/ml	microliter per milliliter
μm	micrometer
mgCl <sub>2</sub>	magnesium chloride
DTT	dithiothreitol
EDTA	ethylenediaminetetraaceticacid
EGTA	ethyleneglycoltetraaceticacid
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
KOH	potassium hydroxide

## CHAPTER 1

### INTRODUCTION

#### 1.1 General Introduction

Nowadays, with rapid development of fast food industries that offer delicious but disease-carried food, it is important for people to consume natural nutritious foods in order to keep the body healthy. Vegetables are one of the natural food sources that benefit people as they are the cheapest sources of proteins, vitamins, minerals and amino acids (Liu, 2004).

One of the popular vegetables crop is cucumber. Cucumber (*Cucumis sativus*) belongs to Cucurbitaceae family and is popularly used as salads. Cucumber is rich in potassium, oxalic acid and phosphorus (Ahmad and Anis, 2005). It contains phenolic content that offer health benefit to people (Liu, 2004) and could prevent cancer from its antioxidant action (Cai *et al.*, 2004). It is good to eat raw cucumber as there are more nutrients in raw vegetables compared to cooked ones (Sivakumar *et al.*, 2010). Traditionally, the seeds of cucumber are used for headache, burning and insomnia, while the leaves are used to treat dyspepsia (Gogte, 2000; Khandelwal, 2000; Shah *et al.*, 2013). The leaf of cucumber has also been used for jaundice and bleeding disorder (Shah *et al.*, 2013).

People love to eat raw cucumber as realizing its nutritional value besides enjoying its uniquely cool, refreshing taste and great crunchiness. In Malaysia, cucumber has great demand from local consumers. Cucumber variety MTi2 are the local variety of cucumber in Malaysia that famously consumed by Malaysian and widely grown throughout the country. Besides, cucumber seed that imported from outside of the country are expensive. It is important to maintain or increase the self sufficiency of this cucumber variety in order to maintain good plant stock.

Nevertheless, the conventional means of propagation may produce non uniform plants that may diminish the quality of the yield (Yadav *et al.*, 2012). Besides, the conventional propagation of cucumber always has high exposure to disease and pests, thus reducing the yield and quality. Growing cucumber in farms is highly associated with variety of diseases such as cucumber mosaic virus, angular leaf spot, bacteria wilt disease, downy mildew disease and many more. Cucumber farming is also easily affected by viruses and bacterial that may lead to insufficient cucumber supply to the world market. For example in 2011, there was an outbreak of cucumber issue

in Europe that affects hundreds of people and caused the death of 14 people (Torry, 2011). The cucumber was infected with *E. coli* from contaminated water used to water the crops (Poullter, 2011). This issue lead to the diminished of cucumber crops of many areas around the world especially Spain, and some Europe countries were reported to ban the sale of cucumbers (“E. coli Cucumber Scare”, 2011).

Therefore, it was important to conserve the cucumber especially of favorable variety to ensure the continuance of cucumber stock if the world was affected by cucumber crisis. Besides, commercially-important variety of cucumber must be conserved to produce uniform products. The finding of this study may lead to production of plantlet with similar characteristics as the mother plant. Recently, *in vitro* propagation was used as alternative method to overcome the problems faced during conventional propagation. *In vitro* propagation possesses many advantages over conventional propagation. This include production of genetically stable plants and virus free plants (Moghaleb *et al.*, 1999), for conservation of plant species (Yadav *et al.*, 2012), multiple plant of favorable traits, introduce new traits into selected plants and growing cultivars in shorten time (Taji *et al.*, 2002).

## **1.2 Problem Statements, Justifications and Objectives of Study**

However, tissue culture is always subjected to somaclonal variation, which results to production of plants that vary from the parent plant. In order to produce plant with the same favorable characteristics as the parent plant, tissue culture techniques must be optimized. This is important as no variation wanted in the production of plant variety that was high demanded from local people and has high commercial value. Here, it is necessary to obtain protocol to produce uniform plantlets that may be useful for industrial large scale multiplication and for conservation of the species.

In this study, shoot apical meristem (SAM) was used as explants to be culture *in vitro*. Meristem culture might produce genetically stable plantlet that may result of producing plantlet with similar characteristics as parent plant, besides provide explant that is free from viruses, bacterial and fungal pathogen for the culture (Grout, 1999). Apical meristem is highly organized structure. Meristematic cells of apical meristem can direct the pattern formation of organs in plant, thus help to reduce somaclonal variation, which is the variation seen in regenerated plants from tissue culture. This study had evaluated the best treatment for culturing cucumber variety MTi2 to obtain uniform plantlet. The protocol for SAM culture would be optimized in this study by using auxin and cytokinin as plant growth regulator. Comparison study was carried out in term of morphology, anatomy, biochemical and antioxidant profiling to detect whether the plantlet produced expressed somaclonal variation. The finding was useful for the species conservation, for



large scale multiplication and for future research. To date, there is no study found on the *in vitro* propagation of cucumber by using SAM.

This study was also carried out to compare the antioxidant activity between different parts of cucumber plant. People did realize the high nutritional content in cucumber fruit. However, they didn't realize which part of the edible parts of cucumber possesses higher nutritional value. Recent researches showed that there was higher nutritional content in seed rather than in edible fruit part of many plant species. As cucumber are always eaten together with the seed, this study was done to find out whether the seed of cucumber possess higher nutrient content compare to fruit parts, in term of antioxidant profiling. Different part of the plant may contain different antioxidant activity. There is also no report found on the comparative study of antioxidant profiling between edible parts of cucumber plant. Antioxidant nutrients are important for human. A few roles possess by antioxidant are in preventing cardiovascular disease, macular degeneration, pathogenic process related to cancer, asthma, besides improve immune function (McDermott, 2000).

Hence, the objectives of this study are:

- (1) To regenerate *C. sativus* cv. MTi2 plants *in vitro* by using shoot apical meristem.
- (2) To compare the morphology, anatomy, biochemical and antioxidant profiling between *C. sativus* plant and *C. sativus* plantlet.
- (3) To compare the antioxidant profiling in different edible parts of *C. sativus* plant.



## REFERENCES

- Abou-Jawdah, Y., Sobh, H., Fayad, A., Lecoq, H., Delecolle, B., Trad-Ferre, J. (2000). Cucurbit yellow stunting disorder virus - a new threat to cucurbits in Lebanon. *Journal of Plant Pathology*.82(1): 55-60.
- Agarwal, M., Kumar, A., Gupta, R., Upadhyaya, S. (2012). Extraction of polyphenol, flavonoid from *emblica officinalis citrus limon, cucumis sativus* and evaluation of their antioxidant activity. *Oriental Journal of Chemistry*. 28(2): 993-998.
- Ahmad, A., Zhong, H., Wang, W., Sticklen, M. B. (2002). Shoot apical meristem: *In vitro* regeneration and morphogenesis in wheat (*Triticuma estivum* L.). *In vitro cell. Dev. Biol. Plant*. 38: 163-167.
- Ahmad, N., Anis, M. (2005). In vitro mass propagation of *Cucumis sativus* L. from nodal segments. *Turkish Journal of Botany*. 29: 237-240.
- Ahmad, N., Fazal, H., Abbasi, B. H., Rashid, M., Mahmood, T., Fatima, N. (2010). Efficient regeneration and antioxidant potential in regenerated tissues of *Piper nigrum* L. *Plant Cell, Tissue and Organ Culture*. 102:129–134.
- Alam, M. F., Banu, M. L. A., Swaraz, A. M., Parvez, S., Hossain, M., Khalekuzzaman, M., Ahsan, N. (2004). Production of virus free seeds using meristem culture in tomato plant under tropical conditions. *J Plant Biotechnol*. 6:221-227.
- Alrahman, N. M. A., Shibli, R. A., Ereifej, K., Hindiyeh, M. Y. (2005). Influence of salinity on growth and physiology of *in Vitro* grown cucumber (*Cucumis Sativus* L. ). *Jordan Journal of Agricultural Sciences*. 1(1): 93-105.
- Assimopoulou, A. N., Boskou, D., Papageoriou, V. P. (2004). Antioxidant activity of alkannin, shikonin and *Alkanna tinctoria* root extracts in oil substrates. *Food Chemistry*. 87: 433-438.
- Aversano, R., Savarese, S., De Nova, J. M., Frusciante, L., Punzo, M., Carputo, D. (2009). Genetic stability at nuclear and plastid DNA level in regenerated plants of *Solanum* species and hybrids. *Euphytica*. 165:353-361.
- Azima, A. M. and Ismail, O. (2009) Challenges on Idle agriculture land management: an institutional perspective in Malaysia. *European Journal of Social Sciences*. 9(1): 39-47.
- Baidez, A. G., Gomez, P., Del Rio, J. A., Ortuno, A. (2007). Dysfunctionality of the xylem in *Olea europaea* L. plants associated with the infection

process by *Verticillium dahlia* Kleb. Role of phenolic compounds in plant defense mechanism. *J. Agric. Food Chem.* 55: 3373-3377.

Bairu, M. W., Aremu, A. O., Staden, J. V. (2011). Somaclonal variation in plants: causes and detection methods. *Plant Growth Regul.* 63: 147-173.

Bandurski, R. S., Cohen, J. D., Slovin, J. P., Reinecke, D. M. (1995). Auxin biosynthesis and metabolism. In: Davies, P.J. (Ed.), *Plant Hormones*. Dordrecht: Kluwer Academic Publishers. (pp. 39-65).

Bar-Nun, N., Mayer, A. M. (1990). Cucurbitacins protect cucumber tissue against infection by *Botrytis cinerea*. *Phytochemistry*. 29(3): 787-791.

Benkova, E., Michniewicz, M., Sauer, M., Teichmann, T., Seifertova, D., Jurgens, G., Friml, J. (2003). Efflux-dependent auxin gradients as a common module for plant organ formation. *Cell*. 115: 591-602.

Benzion, G., Phillips, R. L. (1988). Cytogenetic stability of maize tissue cultures: a cell line pedigree analysis<sup>1</sup>. *Genome*. 30:318-325.

Berg, L. R. (2008). *Introductory Botany: Plants, People and the Environment*, 2<sup>nd</sup> edition. Thomson Brooks/Cole. (pp. 116-159).

Bhatt, A., Naidoo, Y., Nicholas, A. (2010). The foliar trichomes of *Hypoestes aristata* (Vahl) Sol. ex Roem. & Schult var *aristata* (Acanthaceae) a widespread medicinal plant special in tropical sub-Saharan Africa: with comments on its possible phylogenetic significance. *Biological Research*. 43: 403-409.

Bhojwani, S. S. and Rajdhan, M. K. (1996). Plant tissue culture: theory and practice. Revised: *Elsevier*, Amstredam. (pp 483-586).

Bidabadi, S.S., Meon, S. And Wahab, Z. (2010). Study of Genetic and Phenotypic Variability among Somaclones Induced by BAP and TDZ in Micropropagated Shoot Tips of Banana (*Musa* spp.) using RAPD marker. *Journal of Agriculture Science*. 2(3): 49-60.

Biswas, M. K., Hossain, M., Islam, R. (2007). Virus free plantlets production of strawberry through meristem culture. *World Journal of Agricultural Sciences*. 3(6): 757-763.

Blachburn, G. A. (1998). Quantifying chlorophyll and carotenoids at leaf and canopy scales. *Remote Sensing of Environment*. 66: 273-285.

Bondet, V., Brand-Williams, W., Berset, C. (1996). Kinetics and mechanisms of antioxidant activity using the DPPH Free Radical method. *Lebensm. Wiss. U. Technol.* 30: 609-615.

- Bordallo, P. N., Silva, D. H., Maria, J., Cruz, C. D., Fontes, E. P. (2004). Somaclonal variation on *in vitro* callus culture potato cultivars. *Horticultura Brasileira*.22(2): 300-304.
- Bornman, C. H. (1993). Micropropagation and somatic embryogenesis. In: Hayward, M. D., Bosemark, N. O., Romagosa, T. (eds.), *Plant Breeding; Principles and prospects*. Netherland: Springer Netherland Publisher. (pp. 246-260).
- Bradford, M. (1976). A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye-binding. *Analytical Biochemistry*. 72: 248-254.
- Brand-Williams, W., Cuvelier, M. E., Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. *Journal of Lebensmittel-Wissenschaft u-Technology*. 28: 25-30.
- Brandt, K. & Molgaard, J. P. (2001). Featured Article Organic agriculture: does it enhance or reduce the nutritional value of plant foods?. *Journal of the Science of Food and agriculture*. 81: 924-931.
- Cai, Y., Luo, Q., Sun, M., Corke, H. (2004). Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Sciences*. 74: 2157–2184.
- Candan, N. (2003). Changes in chlorophyll-carotenoid contents, antioxidant enzyme activities and lipid peroxidation levels in Zn-stressed *Mentha pulegium*. *Turkish Journal of Chemistry*. 27: 21-30.
- Cao, X & Hammerschlag, F. A. (2000). Improved shoot organogenesis from leaf explants of high bush blueberry. *Hortscience*. 35: 945-947.
- Carles, C. C. & Fletcher, J. C. (2003). Shoot apical meristem maintenance: the art of a dynamic balance. *Trends in Plant Science*. 8(8): 394-401.
- Chandra, S., Bandopadhyay, R., Kumar, V., Chandra, R. (2010). Acclimatization of tissue culture plantlets: from laboratory to land. *Biotechnol Lett*. 32: 1199-1250
- Chang, w. D., Huang, W. W., Cheng, C. C., Chang, Y. S., Tsay, H. S. (1994). The production of secondary metabolites from Chinese medicinal herbs by suspension cell and tissue culture. In. Proc. 7th Int. Congr of SABRAO and WASS, Taipei, Taiwan: *Academia Sinica*, 16-19 Nov. 535-540.
- Chatenet, M., Delage, C., Ripolles, M., Irey, M., Lockhart, B. L. E., Rott, P. (2001). Detection of sugar-cane yellow leaf virus in quarantine and production of virus-free sugarcane by apical meristem culture. *Plant Disease*. 85(11): 1177-1180.

- Chaudhry, Z., Afroz, A., Rashid, H. (2007). Effect of variety and plant growth regulators on callus proliferation and regeneration response of three tomato cultivars (*Lycopersicon esculentum*). *Pakistan Journal of Botany*. 39(3): 857-869.
- Chehregani, A., Mohsenzadeh, F., Ghanad, M. (2011). Male and female gametophyte development in *Cichorium intybus*. *International Journal of Agriculture and Biology*. 13: 1560-8530.
- Chen, C., Ertl, J. R., Leisner, S. M., Chang, C. (1985). Localization of cytokinin biosynthetic sites in pea plants and carrot roots. *Plant Physiol*. 78: 510-513.
- Chu, Y. F., Sun, J., Wu, X., Liu, R. H. (2002). Antioxidant and antiproliferative activities of common vegetables. *J Agric Food Chem*. 50:6910-7000.
- Cushnie, T. P. T. and Lamb, A. J. (2005). Antimicrobial activity of flavonoids. *International Journal of Antimicrobial Agents*. 26: 343-356.
- Daub, M. E. (1986). Tissue culture and the selection of resistance to pathogens. *Annu Rev Phytopathol*. 24: 159–186.
- Davies, P. J. (1995). *Plant Hormones: Physiology, Biochemistry and Molecular Biology*. Dordrecht: Kluwer Publisher. (pp. 319).
- Department of Statistics, Malaysia. (2013). Available online: <http://www.statistics.gov.my>.
- DelBano, M. J., Lorente, J., Castillo, J., Benavente-Garcia, O., DelRio, J. A., Ortuno, A., Quirin, K., Gerard, D. (2003). Phenolic diterpenes, flavones, and rosmarinic acid distribution during the development of leaves, flowers, stems and roots of *Rosmarinus officinalis*. *J. Agric. Food Chem*. 51: 4247-4253.
- Dewilde, W. J. J. O., Duyfjes, B. E. E. (2010). *Cucumis sativus* L. forma *hardwickii* (Royle). *Thai Forest Bulletin*. 38: 98-107.
- Dickison, W. C. (2000). *Integrative Plant Anatomy*. USA: Elsevier.
- Dixon, R. A., Gonzales, R. A. (eds.), (1985). *Plant cell culture: a practical approach*. Oxford University Press, New York Published by British Library Cataloguing in Publication Data.
- Duncan, R. R. (1997). Tissue culture-induced variation and crop improvement. *Advances in Agronomy*. 58:201-240.
- E. Baylis (ed.), *Plant biotechnologies for developing countries*. UK: The Trinity Press. (pp. 241–243).

- E. coli* cucumber scare: Spain angry at German claims. (2011). *BBC News*. Retrieved October 3, 2012, from <http://www.bbc.co.uk/news/world-europe-13605910>.
- El-Hela (2013). Phenolic content, antioxidant potential and *ADES AEGYPTII* ecological friend larvicidal activity of some selected Egyptian plants. *J. Egyptian Soc. Parasitology*. 43(1): 215-234.
- Elmastas, M., Isildak, O., Turkekul, I., Temur, N. (2007). Determination of antioxidant activity and antioxidant compounds in wild edible mushrooms. *Journal of Food Composition and Analysis*. 20: 337-345.
- Evans, D.A. (1988). Applications of somaclonal variation, In: Mizrahi. (ed.), *Biotechnology in agriculture*. New York: Allan R. Liss Publisher. (pp. 203-223).
- Evans, D. A., Sharp, W. R., Medina-Filho, H. P. (1984). Somaclonal and gametoclonal variation. *American Journal of Botany*. 71:759-774.
- Fahn. A. (1990). *Plant Anatomy*. 4<sup>th</sup> edition. Pergamon Press, incorporated. (pp. 185-270).
- Ferreira, S. A. & Boley, R. A. (1992). *Cucumber mosaic virus*. Department of Plant Pathology, University of Hawaii, Manoa. Retrieved from <http://www.extento.hawaii.edu/kbase/Crop/Type/cucvir.htm>.
- Food and Agricultural Organization of United Nations. (2014). FAOSTAT. Available online: <http://faostat.fao.org>.
- Franco, P., Vittorio, S., Robert, A. (2002). *Plants in cosmetics*. Press-Council of Europe, Europe.
- Friml, J. (2003). Auxin transport-shaping the plant. *Curr. Opin. Plant Biol.* 6: 7-12.
- Fujita, H & Mochizuki, A. (2006). The Origin of the Diversity of Leaf Venation Pattern. *Developmental Dynamics*. 235:2710-2721.
- Galperin, M., Patlis, L., Ovadia, A., Wolf, D., Zelcer, A., Kenigsbuch, D. (2003). A melon genotype with superior competence for regeneration and transformation. *Plant Breed.* 122: 66-69.
- Gambley, R. L. & Dodd, W. A. (1990). An *in vitro* technique for the production of *de novo* multiple shoots in cotyledon explants of cucumber (*Cucumis sativus* L.). *Plant Cell Tissue Organ Culture*. 20: 177-183.
- Garcia-Gonzales, R., Quiroz, K., Carraso, B., Caligari, P. (2010). Plant tissues culture: current status, opportunities and challenges. *Ciencia Investigation Agraria*. 37(3): 5-30.



- Gaspar, T., Kevers, C., Penel, C., Greppin, H., Reid, D. M., Thorpe, T. A. (1996). Plant hormones and plant growth regulators in plant tissue culture. *In Vitro Cell Dev. Biol.* 32: 272-289.
- Ghebretinsae, A. G., Thulin, M., Barber, J. C. (2007). Relationships of cucumbers and melons unraveled: molecular phylogenetics of cucumis and related genera (benincaseae, cucurbitaceae). *American Journal of Botany*. 94(7): 1256–1266.
- Gogte, V. M. (2000). *Ayurvedic Pharmacology and Therapeutic Uses of Medicinal Plants*. Mumbai: Chaukhamba Publisher. (pp. 663).
- Grout, B. W. W. (1999). Meristem-tip culture for propagation and virus elimination. In: R. D. Hall (Ed.), *Methods in molecular biology, plant cell culture protocols*. New Jersey: Human Press. (pp. 115-125).
- Guo, C., Yang, J., Wei, J., Li, Y., Xu, J., Jiang, Y. (2003). Antioxidant activities of peel, pulp and seed fractions of common fruits as determined by FRAP assay. *Nutrition Research*. 23(12): 1719-1726.
- Hammerschlag, F. A. (1992). Somaclonal variation. In: F. A. Hammerschlag & R. E. Litz. (eds.), *Biotechnology of perennial fruits*. Wallingford: CAB International. (pp. 35–55).
- Han, X. Z., Shen, T., Lou, H. X. (2007). Dietary polyphenols and their biological significance. *Int. J. Mol. Sci.* 8: 950-988.
- Handley, L. W. & Chambliss, O. L. (1979). *In vitro* propagation of *Cucumis sativus* L. *HortScience*. 14: 22-23.
- Haque, M. S., Wada, T., Hattori, K. (2003). Shoot regeneration and formation from shoot and root meristem of Garlic cv Bangladesh Local. *Asian J. Plant Sci.* 2: 23-27.
- Harborne, J. B. (1989). General procedures and measurements of total phenolics. In Dey, P. M. and Harborne, J. B. (eds.). *Methods in plant biochemistry. Vol1*. Academic Press Limited, London. (pp. 1-28).
- Hartmann, H. T., Kester, D. E., Davies, F. T., Geneve, R. (2010). *Hartmann & Kester's Plant Propagation: Principles and Practices, 8<sup>th</sup> edition*. (pp. 1-57). New Jersey: Prentice Hall.
- Heim, T. & Bobilya, A. (2002). Flavanoids antioxidants: chemistry, metabolism and structure activity relationships. *J Nutr Biochem.* 13: 572-584.
- Hong, S. H., Choi, S. A., Yoon, H., Cho, K. (2011). Screening of *Cucumis sativus* as a new arsenic-accumulating plant and its arsenic accumulation in hydroponic culture. *Environmental Geochemistry and Health*. 33:143–149.

- Horejsi, T. and Staub, J. E. (1999). Genetic variation in cucumber (*Cucumis sativus* L.) as assessed by random amplified polymorphic DNA. *Genetic Resources and Crop Evolution*. 46: 337-350.
- Houghton, P. J. (2002). Chromatography of the chromone and flavonoid alkaloids. *Journal of Chromatography*. 967: 75-84.
- Huang, W. Y. and Cai, Y. Z. (2010). Natural phenolic compounds from medicinal herbs and dietary plant: potential use for cancer prevention. *Nutrition and Cancer*. 62(1): 1-20.
- Huetteman, C. A., Preece, J. E. (1993). Thidiazuron: a potent cytokinin for woody plant tissue culture. *Plant Cell Tissue Organ Culture*. 33: 105-119.
- Hwang, S. and Tang, C. Y. (2000). Improved resistance to Fusarium wilt through somaclonal variation in Cavendish bananas. In: Proc. Conf. on Challenges for Banana Production and Utilization in 21st Century, Trichy, India: *Association for Improvement in Production and Utilization of Banana (AIPUB)*, 24-25 Sept 1996. (pp. 195–208).
- Ishag, S., Osman, M. G., Khalafalla, M. M. (2009). Effect of growth regulators, explant and genotype on shoot regeneration in tomato (*Lycopersicon esculentum* c.v. Omdurman). *International Journal of Sustainable Crop Production*. 4(6): 7-13.
- Jacquemond, M. (2012). Cucumber Mosaic Virus. In: Loebenstein, G. and Lecoq, H. (Eds.), *Advances in Virus research*. (pp. 439-487). London, UK: Elsevier.
- James, A. D. (1997). *The Green Pharmacy*. (pp. 394-459). Rodale Press, Pennsylvania.
- Javanmardi, J., Stushnoff, C., Locke, E., Vivanco, J. M. (2003). Antioxidant activity and total phenolic content of Iranian *Ocimum* accessions. *Food Chemistry*. 83: 547-550.
- Jayaprakasha, G. K., Girenavar, B., Patil, B. S. (2008). Radical scavenging activities of Rio Red grapefruits and Sour orange fruit extracts in different in vitro model systems. *Bioresource Technology*. 99: 4484–4494.
- Johnson, G. N., Scholes, J. D., Horton, P., Young, A. J. (1993). Relationships between carotenoid composition and growth habit in British plant species. *Plant Cell Environment*. 16: 681-686.
- Jose, J., Nimisha, K., Anu, M. A., Nambisan, P. (2012). Evaluation of somaclonal variation in callus cultures of *Jatropha curcas* maintained on different hormonal combinations using RAPD markers. *World Journal of Agricultural Sciences*. 8(6): 616-623.

- Kahkonen, M. P., Hopia, A. I., Vuorela, H. J., Rauha, J., Pihlaja, K., Kujala, T. S., Heinonen, M. (1999). Antioxidant Activity of Plant Extracts Containing Phenolic Compounds. *J. Agric. Food Chem.* 47: 3954-3962.
- Kapoor, L. D. (1990). *CRC handbook of Ayurvedic medicinal plants*. CRC Press LLC, Florida.
- Karp, A. (1989). Can genetic instability be controlled in plant tissue cultures?. *Plant Tissue Culture*. 58: 2–11.
- Kathal, R., Bhatnagar, S. P., Bhojwani, S. S. (1988). Regeneration of plants from leaf explants of *Cucumis melo* cv. Pusa sharbati. *Plant Cell reports*. 7: 449-222.
- Kaushik, A., Jijta, C., Kaushik, J. J., Zeray, R., Ambesajir, A., Beyene, L. (2012). FRAP (Ferric reducing ability of plasma) assay and effect of *Diplazium esculentum* (Retz) Sw. (a green vegetable of North India) on central nervous system. *Indian Journal of natural Products and Resources*. 3(2): 228-231.
- Khandelwal, R. (2000). *Practical Pharmacognosy Techniques and Experiments*. 9th edition. Pune: Nirali Prakashan. (pp. 75).
- Kim, S., Chang, J., Cha, H. C., Lee, K. (1988). Callus growth and plant regeneration in diverse cultivars of cucumber (*Cucumis sativus* L.). *Plant Cell, Tissue and Organ Culture*. 12:67-74.
- Knuttel, A. J. (1989). Tissue culture troubles. *Nurseryman*. 170(11):43-49.
- Kochert, G. (1978). Carbohydrate determination by the phenol sulfuric acid method. In Helebust, J. A. and Craig, J. S. (ed.). *Hand book of phycologia method*. Cambridge University Press, Cambridge. (pp. 56-97).
- Kozai, T., Kubota, C., Jeong, B. R. (1997). Environmental control for the large-scale production of plants through *in vitro* techniques. *Plant Cell, Tissue and Organ Culture* 51: 49-56
- Kumar, D., Kumar, S., Singh, J., Narender, Rashmi, Vashistha, B. D., Singh, N. (2010). Free radical scavenging and analgesic activities of *Cucumis sativus* L. fruit extract. *Journal of Young Pharmacists*. 2(4): 365-368.
- Kumar, H. G. A., Murthy, H. N., Paek, K. Y. (2003). Embryogenesis and plant regeneration from anther cultures of *Cucumis sativus* L. *Science Hort*. 98: 213-222.
- Kumar, V., Parvatam, G., Ravishankar, G. A. (2009). AgNO<sub>3</sub> – a potential regulator of ethylene activity and plant growth modulator. *Electronic Journal of Biotechnology*. 12(2): 1-15.



- Larkin, P. J., Scowcroft, W. R. (1981). Somaclonal variation - a novel source of variability from cell cultures for plant improvement. *Theoretical and Applied Genetics*. 60:197-214.
- Lavie, D. and Glotter, E. (1971). The cucurbitanes; a group of tetracyclic triterpenes. Progress in the Chemistry of Organic Natural Products, New York, USA. (pp. 352-354).
- Letham, D. and Gollnow, B. (1985). Regulators of cell division in plant tissues; cytokinin metabolism in relation to radish cotyledon expansion and senescence. *J. Plant Growth Regul.* 4: 129-145.
- Li, H. P., Huang, T., Wang, C. X., Liao, Y. C. (2009). An efficient regeneration system of berley cultivars from leaf base segments. *Biol. Plant.* 53: 733-736.
- Lichtenthaler, H. K. and Wellburn, A. R. (1985). Determination of total carotenoids and chlorophylls A and B of leaf in different solvents. *Biochemical Society Transactions*. 11: 591-592.
- Liu, R. H. (2003). Health benefits of fruits and vegetables are from additive and synergistic combination of phytochemicals. *American Journal of Clinical Nutrition*. 78:517-520.
- Liu, R. H. (2004). Potential synergy of phytochemicals in cancer prevention: mechanism of action1. *The journal of nutrition*. 134: 3479S-3485S.
- Liza, L. N., Nasar, A. N. M., Zinnah, K. M. N., Chowdhury, M. A. N., Ashrafuzzaman, M. (2013). *In vitro* growth media effect for regeneration of tomato (*Lycopersicon esculentum*) and evaluation of the salt tolerance activity of callus. *Journal of Agricultural and Sustainability*. 3(2): 132-143.
- Lough, T. J. and Lucas, W. J. (2006). Integrative plant biology: role of phloem long-distance macromolecular trafficking. *Annu. Rev. Plant. Biol.* 57: 203-232.
- Malepszy, S., Niemirowicz-Szczytt, K. (1991). Sex determination in cucumber (*Cucumis sativus*) as a model system for molecular biology. *Plant Science*. 80: 39-47.
- Maisuthisakul, P., Pongsawatmanit, R., Gordon, M. H. (2007). Characterization of the phytochemicals and antioxidant properties of extracts from Teaw (*Cratoxylum formosum* Dyer). *Food Chemistry*. 100(4): 1620-1629.
- Malek, M. A., Bari Miah, M. A., Al-Amin, M., Khanam, D., Khatun, M. (2007). *In vitro* regeneration in pointed goured. *Bangladesh J Agri Res*. 32(3): 461-471.

- Mandal, A., Maiti, A., Chowdhury, B., Elanchezhian, R. (2001). Isoenzyme markers in varietal identification of banana. *In Vitro Cell Dev. Biol. Plant.* 37: 599–604.
- Mantle, D., Eddeb, F., Pickering, A. T. (2000). Comparison of relative antioxidant activities of British medicinal plant species *in vitro*. *Journal of Ethnopharmacology.* 72: 47-51.
- Martinez, O., Reyes, L. M., Beltran, M. (1998). Chemovariability in the genus *Musa*: similarities and differences. *Infomusa.* 7(2):16-20.
- Matsumoto, T., Sakai, A., Yamada, A. (1994). Cryopreservation of *in vitro*-grown apical meristems of wasabi (*Wasabia japonica*) by vitrification and subsequent high plant regeneration. *Plant Cell Reports.* 13:442-446.
- Mauseth, J. D. (1991). *Botany: An Introduction to Plant Biology*. Philadelphia: Saunders publisher. (pp. 348-415).
- Mauseth, J. D. (2009). *Botany: An Introduction to Plant Biology*, 4<sup>th</sup> edition. Jones and Bartlett Publishers, Incorporated. (pp. 92-181).
- McDermott, J. H. (2000). Antioxidant nutrients: current dietary recommendations and research update. *Journal of the American Pharmaceutical Association.* 40(6): 785-799.
- Melo, E. D. A., Lima, V. L. A. G., Maciel, M. I. S., Caetano, A. C. S., Leal, F. L. L. (2006). Polyphenol, ascorbic acid and total carotenoid contents in common fruits and vegetables. *Braz J Food Technol.* 9:89-94.
- Michalak, A. (2006). Phenolic compounds and their antioxidant activity in plants growing under heavy metal stress. *Polish Journal of Environmental Study.* 15(4): 523-530.
- Michel, T., Destandau, E., Le Floch, G., Lucchesi, M. E., Elfakir, C. (2012). Antimicrobial, antioxidant and phytochemical investigations of sea buckthorn (*Hippophaë rhamnoides* L.) leaf, stem, root and seed. *Food Chemistry.* 131(3): 754-760.
- Minano, H. S., Benito, M. E. G., Martin, C. (2009). Molecular characterization and analysis of somaclonal variation in chrysanthemum cultivars using RAPD markers. *Scientia Horticulture.* 122: 238-243.
- Mineo, L. (1990). *Plant Tissue Culture Techniques*. In. C. Goldman, C. A. (Ed.), *Tested Studies for Laboratory Teaching*. (pp. 151-174).
- Mishra, A. K & Bhatnagar, S. P. (1995). Direct shoot regeneration from the leaf explant of *Cucumis sativus* L. *Phytomorphol.* 45:47-55.

- Moghaleb, R. E. A., Fujita, K. (1999). Plant regeneration from hypocotyl and cotyledon explants of tomato (*Lycopersicon esculentum*). *Soil Science and Plant Nutrition* 45: 639-646.
- Mukherjee, P. K., Nema, N. K., Maity, N., Sarkar, B., K. (2013). Phytochemical and therapeutic potential of cucumber. *Fitoterapia*. 84: 227–236.
- Murch, S. J., Krishna, R. S., Saxena, P. K. (2000). Phyto-pharmaceuticals: massproduction, standardization, and conservation. *Herbal Medicine*. 4(2): 39-43.
- Nadolska-Orczyk, A. & Malepszy, S. (1989). *In vitro* culture of *Cucumis sativus* L. *Theor Appl Genet*. 78: 836-840.
- Naliwajski, M. R., Sklodowska, M. (2014). The oxidative stress and antioxidant systems in cucumber cells during acclimation to salinity. *Biologia Plantarum*. 58(1): 47-54.
- Nayar, N. M., More, T. A. (1998) *Cucurbits*. Enfield. USA: Science Publishers.
- Nema, N. K., Maity, N., Sarkar, B., Mukherjee, P. K. (2011). *Cucumis sativus* fruitpotential antioxidant, anti-hyaluronidase, and anti-elastase agent. *Arch Dermatol Res*. 11(303): 47-52.
- Nik Mustapha, N. H., Hashim, N. M. F., Abu Hassan, F. (2013). Potential of idle land for mixed vegetable and fruit farming using linear programming. *Asian Journal of Empirical Research*. 3(4): 388-400.
- Nobre, J., Davey, M. R., Lazzeri, P. A., Cannel, M. E. (2000). Transformation of barley scutellum protoplast: regeneration of fertile transgenic plants. *Plant Cell Reports*. 9:1000-1005.
- Nurul Izzah, A., Aminah, A., Md Pauzi, A., Lee, Y. H., Wan Rozita, W. M. Siti Fatimah, D. (2012). Patterns of fruits and vegetable consumption among adults of different ethnics in Selangor, Malaysia. *International Food Research Journal*. 19(3): 1095-1107.
- Obi, R. K. (2003). Antiviral potential of vegetables: can they be cost-effective agents for human disease?.In. R. R. Watson *et al.* (eds.), *Nutrients, Dietary Supplements, and Nutraceuticals: Cost Analysis Versus Clinical Benefits*. London: Springer Science+Business Media. (pp. 259-276).
- Oropeza, M., Guevara, P., Garcia. E., Ramirez, J. L. (1995). Identification of sugarcane (*Saccharum* spp) somaclonal variants resistant to sugarcane mosaic virus via RAPD markers. *Plant Mol. Biol. Rep*. 13: 182-191.

- Orton, T. J. (1984). Somaclonal variation: Theoretical and practical considerations. In: J. P. Gustafson (Ed.), *Gene manipulation in plant improvement*. New York: Plenum Press. (pp. 427-468).
- Pandey, B. P. (2000). *Economic Botany*. New Delhi: S. Chand & Co. Ltd. (pp. 76-77).
- Peschke, V. M., Phillips, R. L. (1992). Genetic implications of somaclonal variation in plants. *Advances in Genetics*. 30:41–75.
- Peter, K. V., Abraham, Z. (2007). *Biodiversity in horticultural crops*. New Delhi: Daya publishing house.
- Phillips, R. L., Kaepplert, S. M., Olhoft, P. (1994) Genetic instability of plant tissue cultures: breakdown of normal controls. In: Proc Natl Acad Sci USA: 5222–5226.
- Pierik, R. L. M. (1987). *In vitro culture of higher plants*. Dordrecht: Kluwer Academic Publishers.
- Plader, W., Malepszy, S., Burza, S., Rusinowski, Z. (1998). The relationship between the regeneration system and genetic variability in the cucumber (*Cucumis sativus* L.). *Euphytica*. 103(1): 9-15.
- Plant tissue culture: environmental condition, methods, types and application. (2013). *Biology Discussion*. Retrieved from <http://www.biologydiscussion.com/>
- Poulson, H. E., Prieme, H., Loft, S. (1998). Role of oxidative DNA damage in cancer initiation and promotion. *Eur. J. Cancer Prev*. 71: 9-16.
- Polston, J. E., Hladky, L., Akad, F., Wintermantel, W. M. (2008). First report on cucurbit yellow stunting disorder virus in cucurbits in Florida. *Plant Disease*. 92(8): 1251.
- Pramanik, M. H. R., Nagai, M., Asao, T., Matsui, Y. (2000). Effects of temperature and photoperiod on phytotoxic root exudates of cucumber (*cucumis sativus*) in hydroponic culture. *Journal of Chemical Ecology*. 26(8): 1953-1967.
- Prasad, K. N., Chew, L. Y., Khoo, H. E., Kong, K. W., Azlan, A., Ismail, A. (2010). Antioxidant Capacities of Peel, Pulp, and Seed Fractions of *Canarium odontophyllum* Miq. Fruit. *Journal of Biomedicine and Biotechnology*. 2010: 871379.
- Pyrzynska, K. & Pekal, A. (2013). Application of free radical diphenylpicrylhydrazyl (DPPH) to estimate the antioxidant capacity of food samples. *Analytical Methods*. 17(5): 4288-4295.

- Radford, A. E., Dickison, W. C., Massey, J. R., Bell, C. R. (1974). *Vascular Plant Systematics*. New York: Harper and Row Publishers. (pp. 6-135).
- Rahman, A. H. M. M. (2004). Taxonomic studies of cucurbits grown in the Northern Parts of Bangladesh. Master of Philosophy Thesis, University of Rajshahi.
- Rahman, A. H. M. M., Anisuzzaman, M., Ferdous, A. A. K. M., Rafiul, I., Naderuzzaman, A. T. M. (2008). Study of nutritive value and medicinal uses of cultivated cucurbits. *Journal of Applied Sciences Research*. 4(5): 555-558.
- Rajasekaran, K., Mullins, M. G., Nair, Y. (1983). Flower formation *In vitro* by hypocotyl explants of cucumber (*Cucumis sativus* L.). *Annals of Botany*. 52: 417-420.
- Rani, V., Singh, K. P., Shiran, B., Nandy, S., Goel, S., Devarumath, R. M., Sreenath, H. L., Raina, S. N. (2000). Evidence for new nuclear and mitochondria genome organizations among high-frequency somatic embryogenesis derived plants of allotetraploid *Coffee Arabica* L. *Plant Cell Reports*. 19(10): 1013-1020.
- Raupach, G. S., Liu, L., Murphy, J. F., Tuzun, S., Kloepper, J. W. (1996). Induced systemic resistance in cucumber and tomato against cucumber mosaic virus using plant growth-promoting rhizobacteria (PGPR). *Plant Disease*. 80(8): 891-894.
- Raven, P. H. (1992). *Biology of Plants*. New York: Worth Publisher. (pp. 545-572).
- Re, R., Pellegrini, N., Pannala, A., Yang, M., Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical and Biological Medicine*. 26: 1231-1237.
- Reed, B. M. (1999). Design a micropropagation system: Workshop presentation from the 1998 SIVB Congr. On *in vitro* Biology. *In Vitro Cellular and Developmental Biology- Plant* 35: 275-284.
- Ren, Y., Zhang, Z., Liu, J., Staub, J. E., Han, Y., Cheng, Z., Li, X., Lu, J., Miao, H., Kang, H., Xie, B., Gu, X., Wang, X., Du, Y., Jin, W., Huang, S. (2009). An integrated genetic and cytogenetic map of the cucumber genome. *PLoS ONE*. 4(6): e5795.
- Rice-Evans, C. A., Miller, N. J., Paganga, G. (1997). Antioxidant properties of phenolic compounds. *Trends in plant science*. 2(4): 152-159.
- Rohman, A., Riyanto, S., Yuniarti, N., Saputra, W. R., Utami, R., Mulatsih, W. (2010). Antioxidant activity, total phenolic, and total flavonoid of



- extracts and fractions of red fruit (*Pandanus conoideus* Lam). *International Food Research Journal*. 17: 97-106.
- Rzepka-Plevnes, D., Kulpa, D., Wajda, A. (2009). Initiation of *in vitro* cultures of *Lycopersicon peruvianum* var. *humifusum*. *Journal of Food Agricultural & Environment*. 7:576-580.
- Saad, A. I. M & Elshahed, A. M. (2012). Plant Tissue Culture Media. *Intech Open Science*. 2:29-40.
- Sablowski, R. (2004). Plant and animal stem cells: conceptually similar, molecularly distinct?. *Trends in Cell Biology*. 14(11): 605-611.
- Sahijram, L., Soneji, J. R., Bollamma, K. T. (2003). Analyzing somaclonal variation in micropropagated bananas (*Musa* spp.). *In vitro Cellular & Developmental Biology*. 39:551-556.
- Saikia, M. & Handique, P. J. (2013). Antioxidant and antibacterial activity of leaf, bark, pulp and seed extracts of seabuckthorn (*Hippophae salicifolia* D. Don) of Sikkim Himalayas. *Journal of Medicinal Plants Research*. 7(19): 1330-1338.
- Sakakibara, H., Honda, Y., Nakagawa, S., Ashida, H., Kanazawa, K. (2003). Simultaneous determination of all polyphenols in vegetables, fruits, and teas. *Journal of Agricultural and Food Chemistry*. 51(3): 571–581.
- Salisbury, F. D. and Ross, C. W. (1992). *Plant Physiology*. Belmont: Wadsworth Publisher. (pp. 357-407).
- Saritha, K. V., Prakash, E., Ramamurthy, N., Naidu, C. V. (2002). Micropropagation of *Spilanthes acmella* Murr. *Biologia Plantarum*. 45(4): 581-584.
- Scheres, B. (2005). Cells: a plant biology perspectives. *Cell*. 122: 499-504.
- Sebastian, P., Schaefer, H., Telford, I. R. H., Renner, S. S. (2010). Cucumber (*Cucumis sativus*) and melon (*C. melo*) have numerous wild relatives in Asia and Australia, and the sister species of melon is from Australia. *PNAS*. 107(32): 14269-14273.
- Sekioka, T. T. & Tanka, J. S. (1981). Differentiation in callus cultures of Cucumber (*Cucumis sativus* L.). *HortScience*. 16: 45.
- Selvaraj, N., Vasudevan, A., Manickavasagam, M., Ganapathi, A. (2006). *In vitro* organogenesis and plant formation in cucumber. *Biologia Plantarum*. 50(1): 123-126.
- Shah, P., Joshi, Y., Dongare, P., Dhande, S., Kadam, V. (2013). Free radical scavenging activity of leaves of *Cucumis sativus*. *International Journal of Phytopharmacy*. 3(3): 72-75.

- Sharma, U. K., Sharma, K., Sharma, N., Sharma, A., Singh, H. P., Sinha, A. K. (2008). Microwave-assisted efficient extraction of different parts of *Hippophae rhamnoides* for the comparative evaluation of antioxidant activity and quantification of its phenolic constituents by reverse-phase high-performance liquid chromatography (RP-HPLC). *Journal of Agricultural and Food Chemistry*. 56(2): 374–379.
- Siddhuraju, P., Mohan, P. S., Becker, K. (2002). Studies on the antioxidant activity of Indian Laburnum (*Cassia fistula* L.): a preliminary assessment of crude extracts from stem bark, leaves, flowers and fruit pulp. *Food Chemistry*. 79(1): 61-67.
- Silvarajan, L. (2013). *In vitro* regeneration of *Oryza sativa* (MR219) and *Zea mays* (Thai Super Sweet) by SAM and RAM. Master Thesis, University Putra Malaysia.
- Silvarajan, L., Nulit, R., Qamaruz Zaman, F. (2012). Effects of plant growth regulators on *in vitro* regeneration of Malaysia Indica Rice (*Oryza sativa* L.) cv. MR219 by shoot apical meristem. *Asian Journal of Agricultural Research*. 6(4): 180-187.
- Simon, S. and Petrasek, J. (2011). Why plants need more than one type of auxin. *Plant Science*. 180: 454-460.
- Singh. M. N., Mishra, A. K., Bhatnagar, S. P. (1996). *In vitro* production of plants from cotyledon explant of *Cucumis melo* L. and their successful transfer to field. *Phytomorphology*. 46: 395-402.
- Sivakumar, T. T., Nair, B. J., Panicker, A. Phytochemicals - The natural fighters against oral cancer. *Trivandrum Dental Journal*. 1(1): 33-41.
- Skirvin, R. M., McPheeters, K. D., Norton, M. (1994). Sources and frequency of somaclonal variation. *Hortscience*. 29(11): 1232-1237.
- Slater, A., Scott, N., Fowler, M. (2003). *Plant biotechnology-the genetic manipulation of plants*. (pp. 35-53). Oxford University Press.
- Song, J. Y., Mattson, N. S., Jeong, B. R. (2011). Efficiency of shoot regeneration from leaf, stem, petiole and petal explants of six cultivars of *Chrysanthemum morifolium*. *Plant Cell Tissue Organ Culture*. 107: 295-304.
- Soong, Y. & Barlow, P. J. (2004). Antioxidant activity and phenolic content of selected fruit seeds. *Food Chemistry*. 88: 411-417.
- Sotiroidis, G., Melliou, E., Sotiroidis, T. G., Chinou, I. (2010). Chemical analysis, antioxidant and antimicrobial activity of three greek cucumber (*cucumis sativus*) cultivars. *Journal of Food Biochemistry*. 34: 61–78.

- Srivastava, A. K., Diengdoh, L. C., Rai, R., Bag, T. K. (2012). Tissue Culture - Technology Harnessed for Potato Seed Production. *Keanean Journal of Science*. 1: 80-86.
- Stipp, L. C. L., Mendes, B. M. J., Piedade, S. M. D. S., Rodriguez, A. P. M. (2001). *In vitro* morphogenesis of *Cucumis melo* var. inodorus. *Plant cell Tissue and Organ Culture*. 65: 81-89.
- Surh, Y. J. (2003). Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer*. 3: 768-780.
- Taiz, L. and Zeiger, E. (1998). *Auxins*. In *Plant Physiology*. Sunderland: Sinauer Associates Publishers. ( pp. 543–589).
- Taji, A., Kumar, P. P., Lakshmanan, P. (2002). *In vitro* Plant breeding. New York: Food Products Press. (pp. 167).
- Tan, C., Chang, W. C., Drew, R. (2005). Somaclonal variation: a tool for the improvement of cavendish banana cultivars. In. Proc. 2nd int. Symposium on Biotech of Tropical & Subtropical Species Taipei, Taiwan: *Acta Hort* 692. 61-65.
- Tanurdzic, M. and Banks, J. A. (2004). Sex determining mechanisms in land plants. *Plant Cell*. 16: 61-71.
- Tanwer, B. S., Faheem, M., Singhl, S., Khan, M., Shahzad, A. (2011). *In vitro* regeneration of multiplication shoots in *Catharanthus roseus*- An important medicinal plant. *Pelagia Research Library*. 0976-8610.
- Teisson, C., De Langhe, E. (1989). Biotechnologies for banana and plantain. In: Baylis, E. (ed.), *Plant biotechnologies for developing countries*. UK: The Trinity Press. (pp. 241–243).
- Thorpe, T. (2007). History of plant tissue culture. *Molecular Biotechnology*. 37: 169-180.
- Torry, H. (2011,). Europe in a pickle over deadly cucumber *E-coli* Infection. *The Wall Street Journal*. Retrieved October 3, 2013, from <http://blogs.wsj.com/source/2011/05/31/europe-in-a-pickle-over-deadly-cucumber-e-coli-infection>.
- Poulter, S. (2011). Major food alert as two britons are diagnosed with fatal food poisoning bug traced to organic cucumbers. *Mail Online*. Retrieved October 3, 2013, from <http://www.dailymail.co.uk/health/article-1391526/Cucumber-E-coli-2people-Britain-diagnosed-5-killed-Germany.html>.
- Trigiano, R. N. and Gray, D. J. (2000). *Plant Tissue Culture Concepts and Laboratory Exercises*. Second edition. Humana Press. (pp. 75-86).



- Tringali, C. (2001). Bioactive compounds from natural sources (isolation, characterization and biological properties). London: Taylor & Francis Group.
- Trulson, A. J. & Shahin, E. A. (1986). *In vitro* plant regeneration in the genus *Cucumis*. *Plant Science*. 47: 35-43.
- Ugandhar, T., Venkateshwarlu, M., Begum, G., Srilatha, T., Jaganmohanreddy, K. (2011). *In vitro* plant regeneration of cucumber (*Cucumis sativus* L.) from cotyledon and hypocotyl explants. *Science Research Reporter*. 1(3): 164-169.
- Valizadeh, M., Kazemi Tabar, S, K. Nematzadeh, G. A. (2007). Effect of plant growth regulators on callus induction and regeneration of Cumin (*Cuminum cyminum*). *Asian Journal of Agricultural Research*. 1: 17-22.
- Vasil, I. K. & Thorpe, T. A. (1994). *Plant Cell and Tissue Culture*. Dordrecht: Kluwer Academic Publishers.
- Veeriah, S., Kautenburger, T., Habermann, N., Sauer, J., Dietrich, H. (2006). Apple flavonoids inhibit growth of HT29 human colon cancer cells and modulate expression of genes involved in the biotransformation of xenobiotics. *Mol. Carcinogen*. 45: 164-174.
- Velioglu, Y. S., Mazza, G., Gao, L., Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *J. Agric. Food Chem*. 46: 4113-4117.
- Venkateshwarlu, M. (2012). Direct multiple shoots proliferation of muskmelon (*cucumis melo* (L.) from shoot tip explants. *International Journal of Pharma and Bio Sciences*. 3(2): 645-652.
- Verdeil, J., Alemanno, L., Niemenak, N., Tranbarger, T. J. (2007). Pluripotent versus totipotent plant stem cells: dependence versus autonomy?. *Trends in Plant Science*. 12(6): 245-252.
- Verzelloni, E., Tagliazucchi, D., Conte, A. (2007). Relationship between antioxidant properties and the phenolic and flavonoid content in traditional balsamic vinegar. *Food Chemistry*. 105(2): 564-571.
- Vidal, M. D. C. and DeGarcia, E. (2000). Analysis of *Musa* spp.: Somaclonal variant resistant to yellow sigatoka. *Plant Mol. Biol*. 18: 23-31.
- Viehmanna, I., Bortlova, Z., Vitamvas, J., Cepkova, P. H., Eliasova, K., Svobodova, E., Travnickova, M. (2014). Assessment of somaclonal variation in somatic embryo-derived plants of yacon [*Smallanthus sonchifolius* (Poepp. and Endl.) H. Robinson] using inter simple sequence repeat analysis and flow cytometry. *Electronic Journal of Biotechnology*. 17: 102–106.

- Vuylsteke, D. R., Ortiz, R. (1996). Field performance of conventional vs. *in vitro* propagules of plantain (*Musa* spp., AAB Group). *HortScience*. 31(5):862–865.
- Wang, H., Gan, D., Zhang, X., Pan, Y. (2010). Antioxidant capacity of the extracts from pulp of *Osmanthus fragrans* and its components. *Food Sci. Technol.* 43: 319–325.
- Wang, G. L., Qin, Z. W., Zhou, X. Y. (2007). Genetic analysis and SSR markers of tuberculate trait in *Cucumis sativus*. *Chin Bull Bot.* 24: 168-172.
- Wang, Q. & Wang, L. (2012). An evolutionary view of plant tissue culture: somaclonal variation and selection. *Plant Cell Rep.* 31:1535–1547.
- Wang, X. H., Lazzeri, P. A., Lorz, H. (1992). Chromosomal variation in dividing protoplasts derived from cell suspensions of barley (*Hordeum vulgare* L.). *Theor Appl Genet.* 85:181-185.
- Wehner, T. C. & Locy, R. D. (1981). *In vitro* adventitious shoot and root formation of cultivars and lines of *Cucumis sativus* L. *HortScience*. 16: 759-760.
- Weigel, D. and Jurgens, G. (2002). Stem cells that make stems. *Nature*. 415: 751-754.
- Xin, W., Liu, Z., Song, Y., Hou, T., Xiang, F. (2012). Direct shoot regeneration from *Arabidopsis thaliana* shoot apical meristem. *Biologia Plantarum*. 56(4): 601-606.
- Yadav, K., Singh, N., Verma, S. (2012). Plant tissue culture: a biotechnological tools for solving the problem of propagation of multipurpose endangered medicinal plants in India. *Journal of Agricultural Technology*. 8(1): 305-318.
- Yannerelli, G. G., Gallego, S. M., Tamaro, M. L. (2006). Effect of UV-B radiation on the activity and isoforms of enzymes with peroxidase activity in sunflower cotyledons. *Environmental and Experimental Botany*. 56: 174-181.
- Yildiz, M. (2012). The prerequisite of the success in plant tissue culture: high frequency shoot regeneration. *Intech Open Science*. 4: 63-90
- Zaiton, S., Sariah, M., Zainal Abidin, M. A. (2006). Isolation and characterization of microbial endophytes from oil palm roots: Implication as biocontrol agents against Ganoderma. *The Planter*. 82: 587-597.
- Zhang, W., He, H., Guan, Y., Du, H., Yuan, L., Li, Z., Yao, D., Pan, J., Cai, R. (2010). Identification and mapping of molecular markers linked to the

tuberculate fruit gene in the cucumber (*Cucumis sativus* L.). *Theoretical and Applied Genetics*. 120: 645-654.

Zhishen, J., Mengcheng, T., Jianming, W. (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *J. Food Chem.* 64: 555-559.

Zitter, T. A. (1984). Virus disease of leafy vegetables and celery. *Vegetable Crops*. (pp. 737).

Ziv, M., Gadasi, G. (1986). Enhanced embryogenesis and plant regeneration from cucumber (*Cucumis sativus* L.) callus by activated charcoal in solid/liquid double-layer cultures. *Plant Science*. 47: 155-122.

