



**UNIVERSITI PUTRA MALAYSIA**

***BIO-BASED CROTONIC ACID PRODUCTION ROUTE VIA DIRECT  
PYROLYSIS OF BACTERIAL POLYHYDROXYBUTYRATE INCLUSIONS***

**MOHD RAHIMI ZAKARIA @ MAMAT**

**FBSB 2015 16**



**BIO-BASED CROTONIC ACID PRODUCTION ROUTE VIA DIRECT  
PYROLYSIS OF BACTERIAL POLYHYDROXYBUTYRATE INCLUSIONS**

By

**MOHD RAHIMI ZAKARIA @ MAMAT**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirement for the Degree of Master of Science**

**March 2015**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purpose from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

## **BIO-BASED CROTONIC ACID PRODUCTION ROUTE VIA DIRECT PYROLYSIS OF BACTERIAL POLYHYDROXYBUTYRATE INCLUSIONS**

By

**MOHD RAHIMI ZAKARIA @ MAMAT**

**March 2015**

**Chairman : Hidayah Ariffin, PhD**

**Faculty : Biotechnology and Biomolecular Sciences**

Crotonic acid is a short-chain unsaturated carboxylic acid which can be utilized in a wide variety of applications such as resins, polymers, drug-ligand agents, and insecticides. Its current industrial production is from non-renewable petrochemical resource, using hydrocarbon as starting material which is converted into ethylene and several chemical intermediates before obtaining crotonic acid as a product of oxidation of crotonaldehyde. The utilization of hydrocarbon has become a concern due to various environmental and economical issues such as pollution, limited availability and increasing price of the hydrocarbon. Therefore, alternative methods have to be developed to reduce dependence on hydrocarbon and ensure sustainable economy and environment. This study was aimed to develop a simple production method for bio-based crotonic acid via pyrolysis of bacterial polyhydroxybutyrate (PHB) inclusions. Bacterial biomass containing PHB was obtained via fermentation of sugar by PHB producing bacteria, *Cupriavidus necator* NCIMB 11599 at limited phosphate concentration. PHB biomass was subjected to dynamic and isothermal pyrolysis to evaluate thermal degradation behavior and to collect the pyrolyzate produced. The pyrolyzate was analyzed by gas chromatography - mass spectrometry (GC-MS) and proton nuclear magnetic resonance ( $^1\text{H-NMR}$ ) to determine its composition. It was found that PHB inclusions with  $66\pm 3$  % PHB content had different thermo gravimetric (TG) profile compared to pure PHB. PHB inclusions exhibited multi-step degradation which was contributed by presence of PHB, water and bacterial cell components. PHB in the inclusions had temperature degradation range of 270 – 340 °C. Later, when PHB inclusion were pyrolyzed in glass tube oven at its maximum degradation temperature, it was found that the pyrolyzate consisted of 56.8 % monomer, 30.6 % dimer, 9.4 % trimer and 3.2 % impurities. Recovery yield of biocrotonic acid obtained in this study was ~63 %, which is approximately 30 % higher than the conventional petrochemical-based CA. Furthermore, the proposed method also has other advantages such as renewable raw materials with less and simple processing steps. Economic analysis also revealed that crotonic acid price from both proposed bio-based and petrochemical-based was comparable (USD 7.80 - 11.05 and USD 6.75 - 13.50 respectively).

In addition, effects of pyrolysis parameters (temperature and retention time), pretreatment and catalyst were also studied in order to increase biocrotonic acid yield

and to reduce the amount of impurities in the pyrolyzate. Results showed that no significant changes of biocrotonic acid yield for temperature range of 300 – 340 °C. Therefore, the temperature of maximum degradation rate of PHB was used for subsequent experiment. Meanwhile, retention time of 20 min showed highest biocrotonic acid recovery yield (66.25 %). Besides that, when the bacterial cells were pretreated by homogenization and combined homogenization with ethanol washing, biocrotonic acid recovery was further improved by 6 % and 8 %, respectively, indicating that improving PHB purity could reduce impurities content and increase the recovery yield. Meanwhile, addition of MgO and Mg(OH)<sub>2</sub> as catalysts resulted in significant selective formation of biocrotonic acid. Catalyzed biocrotonic acid production using MgO and Mg(OH)<sub>2</sub> yielded 64.07 % and 70.60 % biocrotonic acid with 96.10 % and 95.75% purity respectively. Overall, results obtained from this research showed that biocrotonic acid production from pyrolysis of bacterial inclusions could be a promising method for alternative industrial production of crotonic acid as it does not only provide renewable chemical, but also simpler in production method and yielded higher crotonic acid recovery.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

## **PENGHASILAN ASID KROTONIK BERASASKAN BIO MELALUI PIROLISIS SECARA TERUS BUTIRAN POLIHIDROKSIBUTIRAT DALAM BAKTERIA**

Oleh

**MOHD RAHIMI ZAKARIA @ MAMAT**

**Mac 2015**

**Pengerusi : Hidayah Ariffin, PhD**

**Fakulti : Bioteknologi dan Sains Biomolekul**

Asid krotonik adalah asid karboksilik tidak tepu berantai pendek yang boleh digunakan dalam pelbagai aplikasi seperti resin, polimer, ejen ligan dadah dan racun serangga. Penghasilan semasa asid krotonik di peringkat industri adalah dari proses petrokimia dengan menggunakan hidrokarbon sebagai bahan asas yang akan ditukar menjadi etilena dan beberapa bahan kimia perantaraan sebelum menghasilkan asid krotonik sebagai produk daripada pengoksidaan krotonaldehid. Penggunaan hidrokarbon telah menjadi satu kebimbangan disebabkan pelbagai isu-isu alam sekitar dan ekonomi seperti pencemaran, pengeluaran yang terhad dan harga hidrokarbon yang meningkat. Oleh itu, kaedah alternatif perlu dibangunkan untuk mengurangkan kebergantungan kepada hidrokarbon dan memastikan ekonomi dan alam sekitar yang mampan. Kajian ini bertujuan untuk membangunkan satu kaedah pengeluaran asid krotonik berasaskan bio yang mudah melalui pirolisis polihidroksibutirat (PHB) dalam bakteria. Biomassa bakteria yang mengandungi PHB diperolehi melalui fermentasi gula oleh bakteria yang menghasilkan PHB, *Cupriavidus necator* NCIMB 11599 dengan menghadkan kepekatan fosfat. PHB biomassa diuji melalui kaedah pirolisis dinamik dan isoterma untuk menilai tingkahlaku degradasi haba dan untuk mengumpul pirolizat yang dihasilkan. Pirolizat dianalisis dengan gas kromatografi - spektrometri jisim (GC-MS) dan proton nuklear resonans magnet (<sup>1</sup>H-NMR) untuk menentukan komposisinya. Telah didapati bahawa butiran PHB dengan kandungan PHB sebanyak 66±3 % mempunyai profil termo gravimetrik (TG) yang berbeza berbanding dengan PHB. Butiran PHB mempamerkan degradasi multi-langkah yang telah disumbangkan oleh kehadiran PHB, air dan komponen sel bakteria. PHB dalam butir granul mempunyai suhu degradasi di antara 270 - 340 °C. Kemudian, apabila butir PHB telah di pirolisis di dalam ketuhar tiub kaca pada suhu degradasi maksimum, didapati bahawa pirolizat terdiri daripada 56.8 % monomer, 30.6 % dimer, 9.4 % trimer dan 3.2 % bendasing. Hasil asid krotonik adalah kira-kira 63 %, yang hampir 30 % lebih tinggi daripada pengeluaran konvensional petrokimia. Tambahan pula, kaedah yang dicadangkan juga mempunyai kelebihan lain seperti bahan mentah yang boleh diperbaharui, langkah pemrosesan yang sedikit dan mudah. Analisis ekonomi juga mendedahkan bahawa harga asid krotonik dari kedua-dua kaedah berasaskan bio dan petrokimia adalah setanding (USD 7.80 – 11.05 dan USD 6.75-13.50 masing-masing).

Di samping itu, kesan parameter pirolisis, rawatan awal dan pemangkin juga dikaji untuk meningkatkan hasil asid krotonik-bio dan mengurangkan jumlah bendasing dalam pirolizat itu. Keputusan menunjukkan bahawa tiada perubahan yang besar kepada hasil asid krotonik-bio untuk lingkungan suhu 300 – 340 °C. Oleh itu, suhu untuk kadar degradasi maksima bagi PHB digunakan untuk eksperimen yang seterusnya. Sementara itu, masa tahanan selama 20 min menunjukkan nilai penghasilan asid krotonik yang paling tinggi (66.25 %). Selain itu, apabila sel-sel bakteria telah dirawat awal oleh penghomogenan dan gabungan penghomogenan dengan basuhan etanol, peningkatan hasil asid krotonik (kira-kira 11 % dan 16 %, masing-masing) diperhatikan, menunjukkan bahawa peningkatan ketulenan PHB boleh mengurangkan kotoran dan meningkatkan penghasilan. Sementara itu, penambahan MgO dan Mg(OH)<sub>2</sub> sebagai pemangkin menyebabkan pembentukan terpilih asid krotonik. Pengeluaran asid krotonik-bio menggunakan MgO dan Mg (OH)<sub>2</sub> menghasilkan 64.07 % dan 70.60 % asid krotonik-bio dengan 96.10 % dan 95.75 % ketulenan. Secara keseluruhan, keputusan yang diperolehi daripada kajian ini menunjukkan bahawa pengeluaran asid krotonik-bio daripada pirolisis butiran PHB dalam bakteria boleh menjadi kaedah yang berpotensi untuk pengeluaran alternatif asid krotonik-bio kerana ia bukan sahaja memberi kimia yang boleh diperbaharui, tetapi juga lebih mudah dan memberikan hasil asid krotonik-bio yang lebih tinggi.

## ACKNOWLEDGEMENTS

First and foremost I would like to say Alhamdulillah and thank my Lord, Allah S.W.T. for His mercy and guidance for which I have completed my research. Many people have contributed in their own way in the progress of my research study.

Firstly, I would like to express my utmost gratitude to my supervisor, Dr Hidayah Ariffin, for her kind support, patience, motivation, enthusiasm, enlightening lectures, and for many valuable discussions. I also would like to express my deepest gratitude to my co-supervisor Prof. Dr. Mohd Ali Hassan for detailed and constructive comments, guidance, suggestion and unconditional support that greatly eased my progress throughout this study.

My condensed thanks go to Mohd Nor Faiz and Che Mohd Hakimian for their kind help and supports during my study.

I also wish to extend my sincere gratitude to all friends, Ida Amalina, Elmy Nahida Othman, Nurhajirah Biran, Farisha, Falia Shazana, Huzairi, Farhan and others have who helped and gave me support, guidance, helpful suggestions as well as proof-read my paper and thesis.

I also would like to express my greatest gratitude to my father, Zakaria @ Mamat Bin Yunus, my mother, Mahazon Binti Mohd Ali and other family members for their love, prayer, and moral support. They prevented me from feeling lost and give up during my study and helped me by giving me unconditional supports.

This list is far from exhaustive, I pray for forgiveness from those I did not mention by name and include them in my heart-felt gratitude.



I certify that a Thesis Examination Committee has met on 23 March 2015 to conduct the final examination of Mohd Rahimi Zakaria @ Mamat on his thesis entitled “Bio-based crotonic acid production route via direct pyrolysis of bacterial polyhydroxybutyrate inclusions” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Umi Kalsom binti Md Shah, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Mohd. Yunus bin Abd. Shukor, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Nor Azowa binti Ibrahim, PhD**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Mohamad Suffian bin Mohamad Annuar, PhD**

Associate Professor  
University of Malaya  
Malaysia  
(External Examiner)



---

**ZULKARNAIN ZAINAL, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 17 June 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members for the Supervisory Committee were as follows:

**Hidayah Ariffin, PhD**

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

**Mohd Ali Hassan, PhD**

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

---

**BUJANG BIN KIM HUAT, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed, or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscript, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2013(Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: MOHD RAHIMI ZAKARIA @ MAMAT (GS31003)

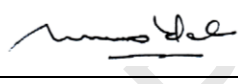
## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  \_\_\_\_\_

Name of  
Chairman of  
Supervisory  
Committee: Hidayah Ariffin, PhD

Signature: \_\_\_\_\_  \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee: Mohd Ali Hassan, PhD



© COPYRIGHT

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xvi
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	
1.1 Research overview	1
1.2 Problem statement	2
1.3 Objectives	2
<b>2 LITERATURE REVIEW</b>	
2.0 Introduction	3
2.1 Polyhydroxyalkanoates (PHA)	3
2.1.1 Polyhydroxybutyrate (PHB)	6
2.1.2 Production of PHA/PHB	6
2.1.3 Physical, mechanical and thermal properties of PHB	8
2.2 Recovery of PHA	9
2.2.1 Solvent extraction	11
2.2.2 Digestion method	12
2.2.2.1 Chemical	12
2.2.2.2 Biological	13
2.2.3 Mechanical cell disruption	14
2.2.3.1 Bead mill	14
2.2.3.2 High pressure homogenization	14
2.3 Degradation of PHA	16
2.3.1 Enzymatic degradation	16
2.3.2 Chemical degradation	17
2.3.3 Thermal degradation	19
2.4 Crotonic acid (CA)	19
2.4.1 Physical and chemical properties	20
2.4.2 Industrial Production of crotonic acid	20
2.4.3 Other method for crotonic acid production	22
2.4.3.1 Chemical-based production	22
2.4.3.2 Biological-based production	22
2.5 Thermal degradation of PHA for CA formation	23
2.5.1 Thermal depolymerization mechanisms of PHB	24

2.5.2	Factors affecting the composition of pyrolysis products	25
2.5.2.1	Temperature	25
2.5.2.2	Catalyst	26
2.6	Current and potential applications of CA	27
<b>3</b>	<b>GENERAL MATERIALS AND METHODS</b>	
3.1	Materials	30
3.2	Microorganism and maintenance	30
3.3	Production of bacterial biomass-containing PHB	30
3.3.1	Growth and production medium for <i>C. necator</i> NCIMB 11599	30
3.3.2	Production of bacterial biomass-containing PHB in 20L bioreactor	30
3.4	Preparation of PHB biomass for pyrolysis	31
3.5	Pyrolysis of PHB	31
3.5.1	Dynamic pyrolysis using thermo gravimetry analyzer (TGA)	31
3.5.2	Isothermal pyrolysis in glass tube oven (GTO)	31
3.6	Analytical procedures	32
3.6.1	PHB content determination	32
3.6.2	Gas chromatograph and mass spectrometry (GC-MS)	32
3.6.3	Proton-Nuclear Magnetic Resonance (NMR)	32
3.7	Overall research methodology	33
<b>4</b>	<b>PRODUCTION OF BIOCROTONIC ACID VIA PYROLYSIS OF BACTERIAL POLYHYDROXYBUTYRATE INCLUSION</b>	
4.1	Introduction	35
4.2	Materials and Methods	36
4.3	Result and Discussion	37
4.3.1	PHB production using glucose as carbon source	37
4.3.2	Thermal degradation temperature of PHB samples	38
4.3.3	Biocrotonic acid recovery	40
4.3.4	Comparison between bio-based and petrochemical-based CA	43
4.3.4.1	Availability of raw materials for CA production	43
4.3.4.2	Process steps	47
4.3.4.3	Biocrotonic acid recovery yield	47
4.3.4.4	Estimation of selling price and market potential of biocrotonic acid	48
4.4	Conclusion	51
<b>5.0</b>	<b>EFFECT OF PYROLYSIS PARAMETERS, PRETREATMENTS AND CATALYST ON RECOVERY YIELD OF BIOCROTONIC ACID</b>	
5.1	Introduction	52
5.2	Materials and Methods	54
5.3	Results and Discussion	56
5.3.1	Effect of temperature	56
5.3.2	Effect of reaction time	58
5.3.3	Effect of mechanical and chemical pretreatment	60

5.3.4 Effect of catalyst	65
5.4 Conclusion	68
<b>6.0 SUMMARY, CONCLUSIONS AND RECOMMENDATION FOR FUTURE WORK</b>	
6.1 Summary and conclusions	69
6.2 Impact of the findings of this research	69
6.3 Suggestion for future work	70
<b>REFERENCES</b>	71
<b>APPENDICES</b>	81
<b>BIODATA OF STUDENT</b>	86
<b>LIST OF PUBLICATIONS</b>	87



## LIST OF TABLES

Table		Page
2.1	Example of types of monomer and homopolymers	4
2.2	Classification of microbial PHA depending on several criteria	6
2.3	Physical properties of PHB and polypropylene	8
2.4	Solvent extraction methods which have been studied	11
2.5	Comparisons of different PHA recovery method	15
2.6	PHA degradation by chemical	18
2.7	Physical properties of <i>trans</i> and <i>cis</i> -CA	20
2.8	Pyrolysis product generated from different pyrolysis temperature	26
2.9	Catalysts used in PHB thermal degradation	27
2.10	Platform chemicals which can be produced from CA and their market opportunities	29
3.1	Composition of growth medium for <i>C. necator</i> NCIMB 11599	30
3.2	MSM composition for <i>C. necator</i> NCIMB 11599	31
4.1	Comparison of pyrolyzate analysis using GC-MS and <sup>1</sup> H-NMR. Isothermal pyrolysis was conducted for 30 min	39
4.2	Comparison between current production steps of crotonic acid and proposed alternative production of crotonic acid	45
4.3	Bacterial strain and carbon sources for PHB production	46
4.4	Comparison between petrochemical-based and bio-based production of CA	48
4.5	Summary of the production cost of crotonic acid from pyrolysis of PHB biomass	48
5.1	Relationship between pyrolysis temperature and products produced	53
5.2	Pyrolyzate composition with different temperature treatment for constant reaction time (30 min)	58
5.3	Pyrolyzate composition with different reaction time at 320°C	60
5.4	Pyrolyzate composition with different pretreatment (reaction time = 20 min)	65
5.5	Pyrolyzate composition with different catalysts (reaction time= 20 min)	68



## LIST OF FIGURES

Figure		Page
2.1	General structure of PHA	3
2.2	Picture of PHA-producing bacteria under phase contrast and light microscope	5
2.3	General chemical structure of PHB. n is the number of repeating units in the polymer	6
2.4	Methods for PHA recovery	10
2.5	Figure 2.5: TEM image of (a) <i>Comamonas sp.</i> EB172 cells before recovery and (b) PHA granules after recovery by NaOH	13
2.6	Degradation of PHA film on soil surface. The film degraded completely in less than 50 days.	16
2.7	Carbon cycle of PHA	17
2.8	Industrial production of CA	21
2.9	Biological pathway of CA in genetically modified <i>E.coli</i>	23
2.10	E1cB degradation mechanism of PHB	24
2.11	Overall thermal degradation pathways of PHB	25
3.1	Pyrolysis apparatus for CA production; (a) sample vial, (b) heating chamber, (c) collector, (d) cold trap, and (e) vacuum pump.	32
3.2	Overall experimental design for the bio-based production of CA	34
4.1	Growth and PHB profile of <i>C.necator</i> NCIMB 11599 in 20L bioreactor	38
4.2	Degradation profile of PHB and PHB inclusion	39
4.3	Total ion current (TIC) chromatogram of pyrolyzates of (a) pure PHB (pyrolysis at 290°C), (b) PHB inclusion (pyrolysis at 310°C). Isothermal pyrolysis was maintained for 30 min. Arrow shows new peak observed which has been identified as crotonamide	41
4.4	<sup>1</sup> H-NMR spectra of pyrolyzates of (a) pure PHB, (b) PHB inclusion. Both samples were pyrolyzed at their respective maximum temperature	42
4.5	Mass balance of the (a) PHB, (b) PHB inclusion	43
4.6	Schematic diagram of bio-based CA production via pyrolysis of PHB biomass	50
5.1	Composition of pyrolyzates for different pyrolysis temperatures.	57
5.2	Pyrolyzates' composition at different pyrolysis duration at 320 °C	59
5.3	TGA profile of pure PHB, untreated and treated PHB biomass	61
5.4	Transmission electron microscope image of a) untreated PHB biomass, b) pretreatment by homogenization, c)	62

5.5	pretreatment by homogenization and ethanol Composition of pyrolyzate of untreated and treated samples.	64
5.6	Thermal degradation profile for i) PHB biomass (ii) PHB biomass with MgO and (iii) PHB biomass with Mg(OH) <sub>2</sub>	65
5.7	Effect of catalysts' addition on the composition of pyrolyzate	67



## LIST OF ABBREVIATIONS

3HB	3-hydroxybutyrate
4HB	4-hydroxybutyrate
3HHx	3-hydroxyhexanoate
3HV	3-hydroxyvalerate
5HV	5-hydroxyvalerate
AMPS	Acrylamido-2-methyl-1-propanesulfonic acid
CA	Crotonic acid
CoA	Coenzyme A
CDCl <sub>3</sub>	Chloroform <i>-d</i>
CO <sub>2</sub>	Carbon dioxide
CuCl <sub>2</sub>	Copper chloride
DSC	Differential scanning calorimetry
EDTA	Ethylenediaminetetraacetic acid
FAB-MS	Fast atom bombardment mass spectrometry
FID	Flame ionization detector
GC-MS	Gas chromatography mass spectrometry
GTO	Glass tube oven
<sup>1</sup> H-NMR	Proton nuclear magnetic resonance
H <sub>2</sub> O	Water
HV	Hydroxyvalerate
LDL	Low density lipoprotein
MgO	Magnesium oxide
Mg(OH) <sub>2</sub>	Magnesium hydroxide
MSM	Mineral salt medium
MTBE	Methyl <i>tert</i> -butyl ether

NMR	Nuclear magnetic resonance
NPCM	Non-PHA cellular mass
OPFJ	Oil palm frond juice
PdCl <sub>2</sub>	Palladium chloride
PHA	Polyhydroxyalkanoate
PHB	Polyhydroxy 3-butyrate
PHBHHx	Polyhydroxybutyrate- <i>co</i> -hydroxyhexanoate
PHBV	Polyhydroxybutyrate- <i>co</i> -hydroxyvalerate
PP	Polypropylene
Py-GC	Pyrolysis-gas chromatography
Py-MS	Pyrolysis-mass spectrometry
SATVA	Subambient thermal volatilization analysis
SDS	Sodium dodecyl sulphate
T <sub>d</sub>	Degradation temperature
TEM	Transmission electron microscopy
TG	Thermo gravimetry
TGA	Thermo gravimetry analyzer
TIC	Total ion current
UV	Ultraviolet

# CHAPTER 1

## INTRODUCTION

### 1.1 Research overview

Fossil feedstocks are primarily used as energy source which cover about 75 % of the world energy demand. Besides that, they are also being utilized to produce variety of products such as polymers and chemicals (Dapsens et al., 2012). However, their usage has been always associated with harmful pollutants which consequently causing global warming and climate changes. In addition, petroleum has faced a drastic increase in price over the last decade due to limited availability and increased demand of the hydrocarbon (Jong et al., 2012). Therefore, efforts have been taken in finding sustainable resources and methods to reduce reliance on the non-renewable hydrocarbons for fuel, energy and chemicals (Armaroli and Balzani, 2007; Pacala and Socolow, 2004). Biomass becomes an interesting alternative for fossil hydrocarbons due to its abundance and ability to be transformed into different products (Dapsens et al., 2012). Furthermore, utilization of biomass can also stimulate development of rural agricultural regions (Jong et al., 2012).

One of the chemicals produced industrially using fossil hydrocarbon is crotonic acid (CA) (Schulz et al., 2003; Arpe, 2010). CA has found its importance in a wide variety of applications. For instance, it can be copolymerized with vinyl acetate, known under different commercial names such as Cevian, Gelva, and Vinac, (Schulz et al., 2003) and be used as a component in hair care and cosmetic products (Campain, 2010). It can also be used in other industries such as coating, absorbent, textile, paint, ceramic, agrochemical and polymer industries (Jasicka-Misiak et al., 2005; Van Walsem et al., 2011). Furthermore, it has great potential as a chemical building block for important chemicals with high market demand such as acrylic acid, butanol, propylene and maleic anhydride (Somleva et al., 2013). However, despite of all its importance, current production method of crotonic acid is from non-renewable petroleum through multiple complicated chemical conversions including steam cracking of heavy hydrocarbon, oxidation of ethylene, aldol condensation of acetaldehyde, dehydration of acetaldol and finally, oxidation of crotonaldehyde into crotonic acid (Arpe, 2010). With growing concern on the impact of hazardous pollutants to the environment, together with limited availability and increasing price of the hydrocarbon, people has started to search for alternative resources and bio-based processes which are safer to both planet and its inhabitant. Recently, there were reports on the use of modified bacteria to produce CA (Koch and Meurer, 2012; Mauch and Schmid, 2008). They manipulated specific bacterial metabolism pathways (2-oxoglutarate and 2-ketoglutarate pathways) which enable the bacterial cells to produce CA which will be secreted into the fermentation broth. However, they did not include the quantitative details and information on recovery methods.

There are many publications on CA as one of the degradation products of polyhydroxybutyrate (PHB) (Morikawa and Marchessault, 1981; Kopinke et al., 1995; Tokiwa and Calabia, 2004; Gonzalez et al., 2005; Ariffin et al., 2008). It has been

reported that PHB can be thermally degraded via a process called pyrolysis into its corresponding dehydrated monomers (*cis* and *trans*-CA), dimers, trimers and secondary products such as carbon dioxide (CO<sub>2</sub>) and propylene. Degradation products can be influenced by pyrolysis temperature as temperatures below 400 °C produce mainly dehydrated monomer and oligomer of PHB while above 400 °C, secondary products will be observed which resulted from further degradation of primary products (Grassie et al., 1984a; Kopinke et al., 1996; Gonzalez et al., 2005). PHB is thermally degraded via combination of degradation mechanisms including random β-elimination, α-deprotonation, trans-esterification and unzipping β-elimination (Ariffin et al., 2008).

In general, PHB is a bioplastic produced through bacterial fermentation. Nevertheless, due to brittle nature and narrow processing window, PHB can only be used in limited applications. Therefore in this study, PHB biomass produced from bacterial fermentation was used as feedstock for bio-based CA production. Feasibility and sustainability of the process were also evaluated.

## 1.2 Problem statement

The problem of current industrial production of CA lies with the use of hydrocarbons as raw material. Chemical synthesis of CA uses heavy hydrocarbons such as naphtha to obtain ethylene which can be subsequently converted into CA through multiple and complicated steps. Fossil hydrocarbon, which has high demand in energy and chemical industries, is facing several issues such as non-renewable, decreasing reservoir, elevated price, and more seriously, causing environmental pollutions. Low recovery yield of CA will require another step which is purification step which might influence the production cost and the number of byproducts which needs to be treated.

The second issue that led to this research is an unfavorable narrow processing temperature characteristic of PHB that limits its application as consumer plastics. PHB, in spite of having advantageous in terms of biodegradability and biocompatibility, has limited feasible applications due to brittleness and narrow processing temperatures. PHB is known to melt at approximately 180 °C and degrade at about 250 °C. This has caused difficulties during the molding processing.

Therefore, this research was aimed at finding alternative method for the production of CA, which is renewable and simple through thermal degradation of PHB. The use of PHB for production of CA can broaden its application.

## 1.3 Objectives of the study

The objectives of this study are:

- 1) To develop bio-based production of crotonic acid and to compare the proposed method with the chemically-synthesized crotonic acid.
- 2) To study the effects of selected pyrolysis parameters, pre-treatments and addition of catalyst on the recovery of bio-crotonic acid.

## REFERENCES

- Aoyagi, Y., Yamashita, K., Doi, Y., 2002. Thermal degradation of poly[(R)-3-hydroxybutyrate], poly[ $\epsilon$ -caprolactone], and poly[(S)-lactide]. *Polym. Degrad. Stab.* 76, 53–59.
- Ariffin, H., Nishida, H., 2009. Anhydride production as an additional mechanism of poly (3-hydroxybutyrate) pyrolysis. *J. Appl. Polym. Sci.* 11, 323–328. doi:10.1002/app
- Ariffin, H., Nishida, H., Shirai, Y., Hassan, M.A., 2008. Determination of multiple thermal degradation mechanisms of poly(3-hydroxybutyrate). *Polym. Degrad. Stab.* 93, 1433–1439. doi:10.1016/j.polymdegradstab.2008.05.020
- Ariffin, H., Nishida, H., Shirai, Y., Hassan, M.A., 2010. Highly selective transformation of poly[(R)-3-hydroxybutyric acid] into trans-crotonic acid by catalytic thermal degradation. *Polym. Degrad. Stab.* 95, 1375–1381. doi:10.1016/j.polymdegradstab.2010.01.018
- Armaroli, N., Balzani, V., 2007. The future of energy supply: Challenges and opportunities. *Angew. Chem. Int. Ed. Engl.* 46, 52–66. doi:10.1002/anie.200602373
- Arpe, H.J., 2010. *Industrial Organic Chemistry*, Fifth. ed. Wiley-VCH Verlag GmbH Co. KGaA, Weinheim, Germany.
- Ballistreri, A., Garozzo, D., Mario, G., Impallomeni, G., Montaudo, G., 1989. Analytical degradation: An approach to the structural analysis of microbial polyesters by different methods. *J. Anal. Appl. Pyrolysis* 16, 239–253.
- Belgacem, M., Gandini, A., 2008. *Monomers, polymers and composites from renewable resources*. Amsterdam: Elsevier.
- Bourque, D., Pomerleau, Y., Groleau, D., 1995. High-cell-density production of poly-b-hydroxybutyrate (PHB) from methanol by *Methylobacterium extorquens*: Production of high-molecular-mass PHB. *Appl. Microbiol. Biotechnol.* 44, 367–376.
- Braunegg, G., Sonnleitner, B., Lafferty, R., 1978. A rapid gas chromatographic method for the determination of poly- $\beta$ -hydroxybutyric acid in microbial biomass. *Eur. J. Appl. Microbiol.* 6, 29–37.
- Bridgewater, A., 2004. Biomass fast pyrolysis. *Therm. Sci.* 8, 21–49.
- Bridgewater, A.V., Toft, A.J., Brammer, J.G., 2002. A techno-economic comparison of power production by biomass fast pyrolysis with gasification and combustion. *Renew. Sustain. Energy Rev.* 6, 181–246. doi:10.1016/S1364-0321(01)00010-7

- Bury, D., Jelen, P., Kalab, M., 2001. Disruption of *Lactobacillus delbrueckii* ssp. *Bulgaricus* 11842 cells for lactose hydrolysis in dairy products: a comparison of sonication, high-pressure homogenization and bead milling. *Innov. Food Sci. Emerg. Technol.* 2, 23–29.
- Campaign, C., 2010. Semipermanent hair shaping method. US Pat. 7,744,859 2.
- Cavalheiro, J.M.B.T., de Almeida, M.C.M.D., Grandfils, C., da Fonseca, M.M.R., 2009. Poly(3-hydroxybutyrate) production by *Cupriavidus necator* using waste glycerol. *Process Biochem.* 44, 509–515. doi:10.1016/j.procbio.2009.01.008
- Chee, J.-Y., Tan, Y., Samian, M.-R., Sudesh, K., 2010. Isolation and characterization of a *Burkholderia* sp. USM (JCM15050) capable of producing Polyhydroxyalkanoate (PHA) from triglycerides, fatty acids and glycerols. *J. Polym. Environ.* 18, 584–592. doi:10.1007/s10924-010-0204-1
- Chee, J.Y., Yoga, S.S., Lau, N., Lau, N.S., Ling, S.C., Abed, R.M.M., Sudesh, K., 2010. Bacterially produced Polyhydroxyalkanoate (PHA): converting renewable resources into bioplastics. *Curr. Res. Technol. Educ. Top. Appl. Microbiol. Microb. Biotechnol.* 1395–1404.
- Chen, Y., Yang, H., Zhou, Q., Chen, J., Gu, G., 2001. Cleaner recovery of poly (3-hydroxybutyric acid) synthesized in *Alcaligenes eutrophus*. *Process Biochem.* 36, 501–506.
- Chisti, Y., Moo-Young, M., 1986. Disruption of microbial cells for intracellular products. *Enzyme Microb. Technol.* 8, 194–204. doi:10.1016/0141-0229(86)90087-6
- Dapsens, P., Mondelli, C., Pérez-Ramírez, J., 2012. Biobased chemicals from conception toward industrial reality: lessons learned and to be learned. *ACS Catal.* 2, 1487–1499.
- Diels, A.M., Michiels, C.W., 2006. High-pressure homogenization as a non-thermal technique for the inactivation of microorganisms. *Crit Rev Microbiol* 32, 201–216.
- Discher, D.E., Janmey, P., Wang, Y.-L., 2005. Tissue cells feel and respond to the stiffness of their substrate. *Science* (80-. ). 310, 1139–1143. doi:10.1126/science.1116995
- Doi, Y., 1990. Structure and properties of poly(3-hydroxybutyrate), in: *Microbial Polyesters*. VCH Publishers Incorporation, New York, USA.
- Doucha, J., Lívanský, K., 2008. Influence of processing parameters on disintegration of *Chlorella* cells in various types of homogenizers. *Appl. Microbiol. Biotechnol.* 81, 431–40. doi:10.1007/s00253-008-1660-6
- Eisenmann, J.L., Mcrae, W.A., 1962. Synthesis of unsaturated carboxylic acids. US3024275 A. doi:10.1074/JBC.274.42.30033.(51)



- Elbahloul, Y., Steinbüchel, A., 2009. Large-scale production of poly(3-hydroxyoctanoic acid) by *Pseudomonas putida* GP01 and a simplified downstream process. *Appl. Environ. Microbiol.* 75, 643–51. doi:10.1128/AEM.01869-08
- Fiorese, M. L., Freitas, F., Pais, J., Ramos, A.M., de Aragão, G.M.F., Reis, M. a. M., 2009. Recovery of polyhydroxybutyrate (PHB) from *Cupriavidus necator* biomass by solvent extraction with 1,2-propylene carbonate. *Eng. Life Sci.* 9, 454–461. doi:10.1002/elsc.200900034
- Goh, C.S., Tan, K.T., Lee, K.T., Bhatia, S., 2010. Bio-ethanol from lignocellulose: Status, perspectives and challenges in Malaysia. *Bioresour. Technol.* 101, 4834–41. doi:10.1016/j.biortech.2009.08.080
- Gonzalez, A., Irusta, L., Fernández-Berridi, M.J., Iriarte, M., Iruin, J.J., 2005. Application of pyrolysis/gas chromatography/Fourier transform infrared spectroscopy and TGA techniques in the study of thermal degradation of poly(3-hydroxybutyrate). *Polym. Degrad. Stab.* 87, 347–354. doi:10.1016/j.polymdegradstab.2004.09.005
- Grassie, N., Murray, E., Holmes, P., 1984a. The thermal degradation of poly (-D)- $\beta$ -hydroxybutyric acid): Part 1—Identification and quantitative analysis of products. *Polym. Degrad. Stab.* 6, 47–61.
- Grassie, N., Murray, E., Holmes, P., 1984b. The thermal degradation of poly (-D)- $\beta$ -hydroxybutyric acid): part 2—changes in molecular weight. *Polym. Degrad. Stab.* 6, 95–103.
- Haas, R., Jin, B., Zepf, F.T., 2008. Production of Poly(3-hydroxybutyrate) from waste potato starch. *Biosci. Biotechnol. Biochem.* 72, 253–256. doi:10.1271/bbb.70503
- Harrison, S., 1991. Bacterial cell disruption: a key unit operation in the recovery of intracellular products. *Biotechnol. Adv.* 9, 217–240.
- Hassan, M.A., 1997. Treatment of agro-industrial effluents for the production of bacterial polyhydroxyalkanoates in Malaysia. Okayama University, Japan.
- Hayashi, S., Ishii, T., Kawaguchi, S., Kimata, T., Misu, N., Yamanaka, S., 1991. Crotonic acid amide derivatives and insecticides containing the same. doi:US5066657
- Holland, S., Jolly, A., Yasin, M., Tighe, B., 1987. Polymers for biodegradable medical devices: II. Hydroxybutyrate-hydroxyvalerate copolymers: hydrolytic degradation studies. *Biomaterials* 8, 289–295.
- Holmes, P., Lim, G., 1990. Separation Process. US Patent 4910145.
- Irie, Y., Kajikawa, K., Takahashi, H., Fujiwara, T., 1993. Method for production of salt-resistant absorbent resin. US Pat. 5,264,495. doi:US5264495

- Iwata, T., Doi, Y., Kokubu, F., Teramachi, S., 1999. Alkaline Hydrolysis of Solution-Grown Poly[(R)-3-hydroxybutyrate] Single Crystals. *Macromolecules* 32, 8325–8330. doi:10.1021/ma991248f
- Jacquel, N., Lo, C.-W., Wei, Y.-H., Wu, H.-S., Wang, S.S., 2008. Isolation and purification of bacterial poly(3-hydroxyalkanoates). *Biochem. Eng. J.* 39, 15–27. doi:10.1016/j.bej.2007.11.029
- Jacquet, B., Mahieu, C., 1981. Hair lacquer and hair lotion compositions containing a copolymer having units of a vinyl allyl or methally ester of an  $\alpha$ -or  $\beta$ -cyclic carboxylic acid. US Pat. 4282203.
- Jasicka-Misiak, I., Wieczorek, P.P., Kafarski, P., 2005. Crotonic acid as a bioactive factor in carrot seeds (*Daucus carota L.*). *Phytochemistry* 66, 1485–91. doi:10.1016/j.phytochem.2005.04.005
- Jong, E.D., Higson, A., Walsh, P., Wellisch, M., 2012. Bio-based Chemicals Value Added Products from Biorefineries.
- Jung, W., 1993. Branched acrylate copolymer with polymerizable double bonds and methods for the production of the acrylate copolymer. US Pat. 5,227,432.
- Kapritchkoff, F.M., Viotti, A.P., Alli, R.C.P., Zuccolo, M., Pradella, J.G.C., Maiorano, A.E., Miranda, E.A., Bonomi, A., 2006. Enzymatic recovery and purification of polyhydroxybutyrate produced by *Ralstonia eutropha*. *J. Biotechnol.* 122, 453–62. doi:10.1016/j.jbiotec.2005.09.009
- Kawalec, M., Adamus, G., Kurcok, P., Kowalczyk, M., Foltran, I., Focarete, M.L., Scandola, M., 2007. Carboxylate-induced degradation of poly(3-hydroxybutyrate)s. *Biomacromolecules* 8, 1053–1058. doi:10.1021/bm061155n
- Kelly, W.J., Muske, K.R., 2004. Optimal operation of high-pressure homogenization for intracellular product recovery. *Bioprocess Biosyst. Eng.* 27, 25–37. doi:10.1007/s00449-004-0378-9
- Khanna, S., Srivastava, A.K., 2005. Recent advances in microbial polyhydroxyalkanoates. *Process Biochem.* 40, 607–619. doi:10.1016/j.procbio.2004.01.053
- Kim, K.J., Doi, Y., Abe, H., 2006. Effects of residual metal compounds and chain-end structure on thermal degradation of poly(3-hydroxybutyric acid). *Polym. Degrad. Stab.* 91, 769–777. doi:10.1016/j.polymdegradstab.2005.06.004
- Kim, K.J., Doi, Y., Abe, H., 2008. Effect of metal compounds on thermal degradation behavior of aliphatic poly(hydroxyalkanoic acid)s. *Polym. Degrad. Stab.* 93, 776–785. doi:10.1016/j.polymdegradstab.2008.01.026
- Kita, T., Narumiya, S., Narisada, M., Watanabe, F., Matsumoto, S., Doteuchi, M., Mizui, T., 1993. Phenolic thioethers, and their production and use. EP Pat. 0 348 203 B1.

- Koch, D., Meurer, G., 2012. Means and methods for producing crotonic acid. EP Pat. 2,511,377.
- Kopinke, F., Mackenzie, K., 1997. Mechanistic aspects of the thermal degradation of poly (lactic acid) and poly ( $\beta$ -hydroxybutyric acid). J. Anal. Appl. Pyrolysis 40, 43–53.
- Kopinke, F., Remmler, M., Mackenzie, K., 1996. Thermal decomposition of biodegradable polyesters -1 : Poly ( p -hydroxybutyric acid ). Polym. Degrad. Stab. 52, 25–38.
- Kunasundari, B., Sudesh, K., 2011. Isolation and recovery of microbial polyhydroxyalkanoates. Express Polym. Lett. 5, 620–634. doi:10.3144/expresspolymlett.2011.60
- Kuppens, T., Cornelissen, T., Carleer, R., Yperman, J., Schreurs, S., Jans, M., Thewys, T., 2010. Economic assessment of flash co-pyrolysis of short rotation coppice and biopolymer waste streams. J. Environ. Manage. 91, 2736–47. doi:10.1016/j.jenvman.2010.07.022
- Lafferty, R., Heinzle, E., 1979. Use of cyclic carbonic acid esters as solvents for poly-( $\beta$ -hydroxybutyric acid). US Patent 4140741.
- Lakshman, K., Shamala, T.R., 2006. Extraction of polyhydroxyalkanoate from *Sinorhizobium meliloti* cells using *Microbispora* sp. culture and its enzymes. Enzyme Microb. Technol. 39, 1471–1475. doi:10.1016/j.enzmictec.2006.03.037
- Lee, S.Y., Park, S.J., Park, J.P., Lee, Y., Lee, S.H., 2005. Economic aspect of biopolymer production, in: Steinbüchel, A., Doi, Y. (Eds.), Biotechnology of Biopolymers: From Synthesis to Patents. Wiley-VCH Verlag GmbH, pp. 307–315. doi:10.1086/431280
- Lehrle, R., Williams, R., 1994. Thermal degradation of bacterial poly (hydroxybutyric acid): mechanisms from the dependence of pyrolysis yields on sample thickness. Macromolecules 3782–3789.
- Letch, R.A., Linstead, R.P., 1932. The chemistry of the three-carbon system. Part XXVIII. Nitriles of butenoic and hexenoic acids. A note on the analysis of unsaturated nitriles by halogen addition. J. Chem. Soc. 443–456. doi:10.1039/jr9320000443
- Li, S., He, J., Yu, P., Cheung, M., 2003. Thermal degradation of poly (3-hydroxybutyrate) and poly (3-hydroxybutyrate-co-3-hydroxyvalerate) as studied by TG, TG–FTIR, and Py–GC/MS. J. Appl. Polym. Sci. 89, 1530–1536.
- Loo, C., Sudesh, K., 2007. Polyhydroxyalkanoates: bio-based microbial plastics and their properties. Malaysian Polym. J. 2, 31–57.
- López-Cortés, A., Rodríguez-Fernández, O., Latisnere-Barragán, H., Mejía-Ruíz, H.C., González-Gutiérrez, G., Lomelí-Ortega, C., 2009. Characterization of

polyhydroxyalkanoate and the phaC gene of *Paracoccus seriniphilus* E71 strain isolated from a polluted marine microbial mat. World J. Microbiol. Biotechnol. 26, 109–118. doi:10.1007/s11274-009-0149-5

Madison, L.L., Huisman, G.W., 1999. Metabolic engineering of poly(3-hydroxyalkanoates): from DNA to plastic. Microbiol. Mol. Biol. Rev. 63, 21–53.

Mamat, M.R.Z., Ariffin, H., Hassan, M.A., Mohd Zahari, M.A.K., 2014. Bio-based production of crotonic acid by pyrolysis of poly(3-hydroxybutyrate) inclusions. J. Clean. Prod. 83, 463–472. doi:10.1016/j.jclepro.2014.07.064

Marchessault, R., Monasterios, C., Jesudason, J., 1994. Chemical, enzymatic and microbial degradation of bacterial and synthetic poly- $\beta$ -hydroxyalkanoates. Polym. Degrad. Stab. 45, 187–196.

Mauch, K., Schmid, J., 2008. Biotechnological production of crotonic acid. WO Pat. 2009/ 046828 A1.

McChalicher, C., Srienc, F., Rouse, D., 2010. Solubility and degradation of polyhydroxyalkanoate biopolymers in propylene carbonate. AIChE J. 56, 1616–1625. doi:10.1002/aic

Melchior, M., Keul, H., Höcker, H., 1996. Depolymerization of poly to cyclic oligomers and polymerization of the cyclic trimer: an example of thermodynamic recycling. Macromolecules 29, 6442–6451.

Middelberg, A., 1995. Process-scale disruption of microorganisms. Biotechnol. Adv. 13, 491–551.

Mohammadi, M., 2011. Alternative recovery methods of intracellular polyhydroxyalkanoates from local isolate *Comamonas* sp. EB172. Universiti Putra Malaysia.

Mohammadi, M., Hassan, M.A., Shirai, Y., Man, H.C., Ariffin, H., Yee, L.-N., Mumtaz, T., Chong, M.-L., Phang, L.-Y., 2012. Separation and Purification of Polyhydroxyalkanoates from Newly Isolated *Comamonas* sp. EB172 by Simple Digestion with Sodium Hydroxide. Sep. Sci. Technol. 47, 534–541. doi:10.1080/01496395.2011.615788

Morikawa, H., Marchessault, R.H., 1981. Pyrolysis of bacterial polyalkanoates. Can. J. Chem. 59, 2306–2313. doi:10.1139/v81-334

Muhamad, I.I., Joon, L.K., Azemi, M., Noor, M., 2006. Comparing the Degradation of Poly- $\beta$ - ( hydroxybutyrate ), Poly- $\beta$  -( hydroxybutyrate-co-valerate )( PHBV ) and PHBV / Cellulose Triacetate Blend 1, 39–46.

Naoto, Y., 1994. Production method of crotonic acid and crotonic acid derivative. JP 06 087784.

- Narasimhan, K., Cearley, A., Gibson, M., Welling, S., 2008. Process for the solvent-based extraction of polyhydroxyalkanoates from biomass. US Patent 7378266 B2.
- Nguyen, S., Yu, G., Marchessault, R.H., 2002. Thermal degradation of poly(3-hydroxyalkanoates): preparation of well-defined oligomers. *Biomacromolecules* 3, 219–24.
- Nishida, H., Tokiwa, Y., 1995. Confirmation of colonization of degrading bacterium strain SC-17 on poly(3-hydroxybutyrate) cast film. *J. Environ. Polym. Degrad.* 3, 187–197. doi:10.1007/BF02068673
- Nonato, R., Mantelatto, P., Rossell, C., 2001. Integrated production of biodegradable plastic, sugar and ethanol. *Appl. Microbiol. Biotechnol.* 57, 1–5. doi:10.1007/s002530100732
- Omar, S., Rayes, A., Eqaab, A., Voß, I., Steinbüchel, A., 2001. Optimization of cell growth and poly ( 3-hydroxybutyrate ) accumulation on date syrup by a *Bacillus megaterium* strain. *Biotechnol. Lett.* 23, 1119–1123.
- Pacala, S., Socolow, R., 2004. Stabilization wedges: solving the climate problem for the next 50 years with current technologies. *Science* 305, 968–72. doi:10.1126/science.1100103
- Papantoniou, C., Grognet, J., 1978. Terpolymer of (a) crotonic acid (b) vinyl acetate and (c) allyl or methallyl esters. US Pat. 4,070,533.
- Park, C.-H., Damodaran, V.K., 1994. Biosynthesis of poly(3-hydroxybutyrate-co-3-hydroxyvalerate) from ethanol and pentanol by *Alcaligenes eutrophus*. *Biotechnol. Prog.* 10, 615–620.
- Peters, M.S., Timmerhaus, K.D., West, R.E., 2003. *Plant Design and Economics for Chemical Engineers*, Fifth. ed. McGraw-Hill, New York.
- Philip, S., Keshavarz, T., Roy, I., 2007. Polyhydroxyalkanoates: biodegradable polymers with a range of applications. *J. Chem. Technol. Biotechnol.* 247, 233–247. doi:10.1002/jctb
- Potter, T., Grubbs, H., 1992. Blocked polyisocyanates for the production of powder coatings with flat finishes. US Pat. 5,112,931. doi:US 5112931 A
- Ramsay, J., Berger, E., Ramsay, B., Chavarie, C., 1990. Recovery of poly-3-hydroxyalkanoic acid granules by a surfactant-hypochlorite treatment. *Biotechnol. Tech.* 4, 221–226.
- Ramsay, J., Berger, E., Voyer, R., Chavarie, C., Ramsay, B., 1994. Extraction of poly-3-hydroxybutyrate using chlorinated solvents. *Biotechnol. Tech.* 8, 589–594.

- Rittner, S., Gortz, H., Riedel, K., 1990. Process for the preparation of pure crotonic acids. US Pat. 4,918,225.
- Roberts, K.G., Gloy, B. a, Joseph, S., Scott, N.R., Lehmann, J., 2010. Life cycle assessment of biochar systems: estimating the energetic, economic, and climate change potential. *Environ. Sci. Technol.* 44, 827–33. doi:10.1021/es902266r
- Ryu, H.W., Hahn, S.K., Chang, Y.K., Chang, H.N., 1997. Production of poly(3-hydroxybutyrate) by high cell density fed-batch culture of *Alcaligenes eutrophus* with phosphate limitation. *Biotechnol. Bioeng.* 55, 28–32. doi:10.1002/(SICI)1097-0290(19970705)55:1<28::AID-BIT4>3.0.CO;2-Z
- Schulz, R.P., Blumentein, J., Kohlpaintner, C., 2003. Crotonaldehyde and Crotonic Acid, in: *Ullmann's Encyclopedia of Industrial Chemistry*. Wiley-VCH, Weinheim.
- Somleva, M.N., Peoples, O.P., Snell, K.D., 2013. PHA bioplastics, biochemicals, and energy from crops. *Plant Biotechnol. J.* 11, 233–52. doi:10.1111/pbi.12039
- Soykan, C., Coskun, R., Kirbag, S., 2007. Poly(crotonic acid-co-2-acrylamido-2-methyl-1-propanesulfonic acid)–metal complexes with copper(II), cobalt(II), and nickel(II): Synthesis, characterization and antimicrobial activity. *Eur. Polym. J.* 43, 4028–4036. doi:10.1016/j.eurpolymj.2007.06.033
- Špitalský, Z., Lacík, I., Lathová, E., Janigová, I., Chodák, I., 2006. Controlled degradation of polyhydroxybutyrate via alcoholysis with ethylene glycol or glycerol. *Polym. Degrad. Stab.* 91, 856–861. doi:10.1016/j.polymdegradstab.2005.06.019
- Sudesh, K., Abe, H., Doi, Y., 2000. Synthesis, structure and properties of polyhydroxyalkanoates: biological polyesters. *Prog. Polym. Sci.* 25, 1503–1555. doi:10.1016/S0079-6700(00)00035-6
- Sudesh, K., Iwata, T., 2008. Review Sustainability of Biobased and Biodegradable Plastics 1 Introduction : The Need for Biobased and Production of Biobased and Biodegradable. *CLEAN-Soil, Air, Water* 36, 433–442. doi:10.1002/clen.200700183
- Tamer, I.M., Moo-young, M., 1998. Disruption of *Alcaligenes latus* for Recovery of Poly (B-hydroxybutyric acid ): Comparison of High-Pressure Homogenization , Bead Milling , and Chemically Induced Lysis. *Ind. Eng. Chem. Res.* 37, 1807–1814.
- Tokiwa, Y., Calabia, B.P., 2004. Degradation of microbial polyesters. *Biotechnol. Lett.* 26, 1181–9. doi:10.1023/B:BILE.0000036599.15302.e5
- Ute, K., Tarao, T., Nakao, S., Kitayama, T., 2003. Preparation and properties of disyndiotactic poly(alkyl crotonate)s. *Polymer (Guildf).* 44, 7869–7874. doi:10.1016/j.polymer.2003.10.005

- Valappil, S.P., Misra, S.K., Boccaccini, a R., Keshavarz, T., Bucke, C., Roy, I., 2007. Large-scale production and efficient recovery of PHB with desirable material properties, from the newly characterised *Bacillus cereus* SPV. *J. Biotechnol.* 132, 251–8. doi:10.1016/j.jbiotec.2007.03.013
- Van Walsem, J., Anderson, E., Licata, J., Sparks, K.A., Mirley, C., Sivasubramanian, M., S., 2011. Process for producing a monomer component from a genetically modified polyhydroxyalkanoate biomass. WIPO patent, WO2011/100608A1. doi:WO2011100608A1
- Viout, A., Pasero, R., 1978. Polymers comprising vinyl esters-crotonic acid. US Pat. 4,129,711.
- Volova, T., 2004. Polyhydroxyalkanoate-plastic materials of the 21st century: Production, properties, and application. Nova Publisher, .
- Voon, P.T., 2005. Environmental friendly alternative methods for the recovery of intracellular polyhydroxyalkanoates (PHA). Universiti Putra Malaysia.
- Wampfler, B., Ramsauer, T., Rezzonico, S., Hischer, R., Köhling, R., Thöny-Meyer, L., Zinn, M., 2010. Isolation and purification of medium chain length poly(3-hydroxyalkanoates) (mcl-PHA) for medical applications using nonchlorinated solvents. *Biomacromolecules* 11, 2716–23. doi:10.1021/bm1007663
- Watt, B., Morgan, S., Fox, A., 1991. 2-Butenoic acid, a chemical marker for poly- $\beta$ -hydroxybutyrate identified by pyrolysis—gas chromatography/mass spectrometry in analyses of whole microbial cells. *J. Anal. Appl. Pyrolysis* 19, 237–249.
- Wolf, O., Crank, M., Patel, M., Marscheider-Weidemann, F., Schleich, J., Husing, B., Angerer, G., 2005. Techno-economic feasibility of large-scale production of bio-based polymers in Europe, European Communities. Spain.
- Yanagihara, T., Honda, T., 1994. Gas permeable and waterproof nonwoven fabric and process for its production. doi:US 4983450
- Yasothea, K., Aroua, M.K., Ramachandran, K.B., Tan, I.K.P., 2006. Recovery of medium-chain-length polyhydroxyalkanoates (PHAs) through enzymatic digestion treatments and ultrafiltration. *Biochem. Eng. J.* 30, 260–268. doi:10.1016/j.bej.2006.05.008
- Yiamsawas, D., Kangwansupamonkon, W., Chailapakul, O., Kiatkamjornwong, S., 2007. Synthesis and swelling properties of poly[acrylamide-co-(crotonic acid)] superabsorbents. *React. Funct. Polym.* 67, 865–882. doi:10.1016/j.reactfunctpolym.2007.05.011
- Yu, J., 2009. Recovery and purification of polyhydroxyalkanoates. US Pat. 7,514,525.

- Yu, J., Chen, L.X.L., 2006. Cost-effective recovery and purification of polyhydroxyalkanoates by selective dissolution of cell mass. *Biotechnol. Prog.* 22, 547–553. doi:10.1021/bp050362g
- Yu, J., Plackett, D., Chen, L.X.L., 2005. Kinetics and mechanism of the monomeric products from abiotic hydrolysis of poly[(R)-3-hydroxybutyrate] under acidic and alkaline conditions. *Polym. Degrad. Stab.* 89, 289–299. doi:10.1016/j.polymdegradstab.2004.12.026
- Yu, J., Stahl, H., 2008. Microbial utilization and biopolyester synthesis of bagasse hydrolysates. *Bioresour. Technol.* 99, 8042–8048. doi:10.1016/j.biortech.2008.03.071
- Yu, P.H., Chua, H., Huang, a L., Ho, K.P., 1999. Conversion of industrial food wastes by *Alcaligenes latus* into polyhydroxyalkanoates. *Appl. Biochem. Biotechnol.* 77-79, 445–54.
- Zahari, M.A.K.M., 2013a. Oil palm frond juice as a novel and renewable substrate for the production of poly(3-hydroxybutyrate) bioplastic. University Putra Malaysia.
- Zahari, M.A.K.M., 2013b. Scaling up of poly (3-hydroxybutyrate) production from oil palm frond juice by *Cupriavidus necator* (CCUG52238T). *Res. J. Chem. Environ.* 17, 18–24.
- Zahari, M.A.K.M., Ariffin, H., Mokhtar, M.N., Salihon, J., Shirai, Y., Hassan, M.A., 2012a. Factors affecting poly(3-hydroxybutyrate) production from oil palm frond juice by *Cupriavidus necator* (CCUG52238(T)). *J. Biomed. Biotechnol.* 2012, 1–8. doi:10.1155/2012/125865
- Zahari, M.A.K.M., Zakaria, M.R., Ariffin, H., Mokhtar, M.N., Salihon, J., Shirai, Y., Hassan, M.A., 2012b. Renewable sugars from oil palm frond juice as an alternative novel fermentation feedstock for value-added products. *Bioresour. Technol.* 110, 566–71. doi:10.1016/j.biortech.2012.01.119
- Zinn, M., Weilenmann, H.-U., Hany, R., Schmid, M., Egli, T., 2003. Tailored Synthesis of Poly([R]-3-hydroxybutyrate-co-3-hydroxyvalerate) (PHB/HV) in *Ralstonia eutropha* DSM 428. *Acta Biotechnol.* 23, 309–316. doi:10.1002/abio.200390039