



UNIVERSITI PUTRA MALAYSIA

***ANALYTICAL TECHNIQUES FOR DETECTION OF ADULTERATION IN
CRUDE PALM OIL***

ANAND KUMAR INTHIRAM

FBSB 2015 18



UPM
UNIVERSITI PUTRA MALAYSIA
BERILMU BERBAKTI

**ANALYTICAL TECHNIQUES FOR DETECTION OF ADULTERATION IN
CRUDE PALM OIL**

By

ANAND KUMAR INTHIRAM

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

March 2015

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

ANALYTICAL TECHNIQUES FOR DETECTION OF ADULTERATION IN CRUDE PALM OIL

By

ANAND KUMAR INTHIRAM
March 2015

Chair : Professor Dr. Lai Oi Ming, PhD
Faculty : Biotechnology and Biomolecular Sciences

In recent years, the demand and consumption for Crude Palm Oil (CPO) has gradually increased in world market. However, production to fulfill the rising demand is rather slow. The gap between demand and production thus lead to potential frauds or adulteration of the CPO. Therefore, we are investigating the possibility of using analytical techniques such as gas chromatograph (GC), high performance liquid chromatograph (HPLC), differential scanning calorimetric (DSC) and Fourier transform infrared (FTIR) spectroscopy to detect CPO adulteration with sludge oil (SO) and used vegetable oil (UVO/UVOA). Investigations conducted with individual fatty acids and triacylglycerol by GC and HPLC analysis, respectively showed that it was very difficult to distinguish between genuine CPO from adulterated CPO. However, combination of fatty acids and triacylglycerols with multivariate statistical analysis produced more promising results. With these techniques, contaminated CPO with adulterants between 2-5% was easily detected by principle component analysis (PCA) and cluster analysis (Dendrogram). DSC thermograms showed that detection of adulteration was difficult to be determined by using heating thermogram due to the polymorphism of oils and fats. However, the profile of DSC cooling thermograms showed changes in peak intensity as the amount of adulterant concentrations varied in CPO. This provided a positive indication that detection of adulterants in CPO could be determined from the changes in peak intensity. In parallel, application of multivariate and regression analysis were able to identify adulterant contents as low as 2% accurately in CPO. FTIR spectroscopy was the last technique applied to detect SO and UVO/UVOA adulteration. FTIR techniques seems to be the least promising technique to detect adulteration of CPO. However adulteration with used vegetable oil containing animal fat (UVOA) was clearly highlighted and was able to be discriminated by cluster analysis Dendrogram at level of 2 % using FTIR spectra. Nevertheless, further study is required to improve the level of detection as well as accuracy for discriminating genuine CPO from the adulterated CPO.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

TEKNIK ANALISA UNTUK MENGESAN PENGADUKAN DALAM MINYAK SAWIT MENTAH

Oleh

ANAND KUMAR INTHIRAM
Mac 2015

Pengerusi : Profesor . Dr. Lai Oi Ming, PhD
Fakulti : Bioteknologi dan Sains Biomolekul

Semenjak kebelakangan ini, permintaan dan penggunaan untuk minyak sawit mentah (CPO) telah beransur-ansur meningkat di pasaran dunia . Walau bagaimanapun, pengeluaran CPO bagi memenuhi permintaan menjadi suatu cabaran . Jurang di antara permintaan dan pengeluaran CPO membawa kepada penipuan atau pencampuran minyak murah yang lain di dalam CPO. Oleh itu, potensi menggunakan teknik analisa seperti kromatografi gas (GC) , kromatografi cecair prestasi tinggi (HPLC) , permeteran kalori pengimbasan kebezaan (DSC) dan Spektroskopi infra-merah (FTIR) untuk mengenalpasti pencampuran CPO dengan minyak enapcemar (SO) dan minyak sayuran terguna (UVO/UVOA) disiasat dalam kajian ini. Kajian yang dijalankan dengan menentukan komposisi asid lemak dan kandungan trigliserida dengan analisis GC dan HPLC, masing-masing menunjukkan bahawa ia adalah amat sukar untuk membezakan CPO tulen daripada CPO yang diaduk. Walau bagaimanapun, gabungan penentuan asid lemak dan kandungan trigliserida dengan analisis statistik multivariat menghasilkan keputusan yang lebih merangsangkan . Dengan aplikasi ini , CPO diaduk dengan bahan aduk di antara 2 hingga 5% boleh dikesan melalui analisis komponen prinsipal (PCA) dan analisis kelompok (dendrogram). Termogram pencairan DSC menunjukkan bahawa pengesanan pencemaran adalah sukar untuk dikenal pasti disebabkan oleh sifat polimorfik minyak dan lemak. Walau bagaimanapun , profil termogram penyejukan DSC menunjukkan perubahan pada intensiti puncak apabila jumlah kepekatan minyak aduk seperti SO dan UVO/UVOA meningkat di dalam CPO. Ini memberikan satu petunjuk yang positif bahawa pengesanan bahan asing dalam CPO boleh ditentukan melalui perubahan intensiti puncak. Pada masa yang sama , penggunaan analisis multivariat dan regresi dapat mengenal pasti kandungan bahan aduk serendah 2 % dalam CPO. Spektroskopi FTIR adalah teknik terakhir yang digunakan untuk mengesan SO dan UVO / UVOA. Teknik FTIR yang digunakan tidak berkesan untuk mengenai pasti bahan adukan dalam CPO. Walau bagaimanapun pencemaran dengan minyak sayur-sayuran yang digunakan yang mengandungi lemak haiwan (UVOA) dapat diskriminasi melalui analisis kelompok , dendrogram di peringkat 2%. Walau bagaimanapun , kajian lanjut diperlukan untuk meningkatkan tahap pengesanan dan ketepatan pengesan CPO tulen daripada yang diaduk.

ACKNOWLEDGEMENTS

I would like to express my deepest appreciation to Genting Plantation (GenP) for giving me a chance to carry out this trial and the facilities and studentship support. In addition, I owe particular thanks to my supervisors Prof. Dr. Lai Oi Ming, and co-supervisors Prof Dr. Tan Chin Ping, Dr. Seyed Hamed Mirhosseini and Dr. Rosfariza Mohamad for your valuable guidance, assistance and encouragement in bringing this project to a successful end. I would also thank my supervisory committee members for spent their valuable time in checking my thesis. It is also my duty to record my thankfulness to Dr. Cheah Suan Choo, Dr. Lee Chong Hee and Mr Chua Kia Ling research management staffs in ACGT/GGT for their kind assistance and valuable opinion during the running of this project. A special note of thanks to my labmates at IBS, UPM for helping me during the instrusement run. Last but not least, I would like to express my heartfelt appreciation to my family members for giving me endless support, encouragement and understanding.

APPROVAL SHEETS (Replace with printed copy from UPM)

I certify that a Thesis Examination Committee has met on 13th March 2015 to conduct the final examination of ANAND KUMAR INTHIRAM on his thesis entitled “DETECTION OF CRUDE PALM OIL ADULTERATION BY CHEMICAL PROPERTIES USING CHROMATOGRAPHIC, THERMAL AND SPECTRAL TECHNIQUES.” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Umi Kalsom binti Md Shah, PhD
Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

H Ng Paik San, PhD
Associate Professor
Faculty of Forestry
Universiti Putra Malaysia
(Internal Examiner)

H Ng Paik San, PhD
Associate Professor
Faculty of Forestry
Universiti Putra Malaysia
(Internal Examiner)

Jumat Salimon, PhD
Professor
Faculty of Science and Technology
University Kebangsaan Malaysia
(External Examiner)

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Lai Oi Ming, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Tan Chin Ping, PhD

Professor
Faculty of Food Science and Technology,
Universiti Putra Malaysia
(Member)

Syed Hamed Mirhosseini, PhD

Associate Professor
Faculty of Food Science and Technology,
Universiti Putra Malaysia
(Member)

Rosfarizan Mohamad, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 15 April 2015

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: _____

Name and Matric No.: Anand Kumar Inthiram, GS31023

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Signature:

Name of Chairman of
Supervisory Committee:
Prof. Dr. Lai Oi Ming

Name of Member of Supervisory
Committee:
Prof. Dr. Tan Chin Ping

Signature:

Signature:

Name of Member of Supervisory
Committee:
Dr. Seyed Hamed Mirhosseini

Name of Member of Supervisory
Committee:
Dr. Rosfarizan Mohamad

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	ii
ACKNOWLEDGEMENTS	iii
APPROVAL	iv
DECLARATION	Vi
LIST OF TABLES	x
LIST OF FIGURES	xi
LIST OF ABBREVIATIONS	xii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	5
2.1 Chromatographic Techniques	6
2.1.1 Gas Chromatographic (GC)	6
2.1.2 High Performance Liquid Chromatographic	8
2.2 Spectrum Techniques	9
2.2.1 Nuclear Magnetic Resonance (NMR)	9
2.2.2 Infrared (IR)	10
2.2.3 Fourier Transform Infrared (FTIR)	11
2.3 Thermal Techniques	12
2.3.1 Differential Scanning Calorimetry (DSC)	13
2.4 Other Techniques	13
2.4.1 DNA Based Technology	13
2.4.2 Electronic Nose	14
2.5 Summary : Techniques and applications used for the determination of oil and fats authenticity	16
3 MATERIALS AND METHODS	21
3.1 Materials	21
3.1.1 Crude Palm Oil (CPO)	21
3.1.2 Sludge Oil (SO)	21
3.1.3 Vegetable Oil	21
3.1.3.1 Preparation of used vegetable oil without animal fat (UVO)	21
3.1.3.2 Preparation of used vegetable oil with animal fat (UVOA)	21
3.1.4 Admixture of Samples	22
3.1.5 Storage of Samples	22
3.2 Laboratory Analysis	23
3.2.1 Fatty Acid Composition Analysis	23
3.2.1.1 Gas Chromatograph Methodology	23
3.2.2 Triacylglycerol (TAG) Composition Analysis	23
3.2.2.1 High Performance Liquid Chromatograph (HPLC)	24

	methodology	
3.2.3	Thermal Analysis	24
3.2.3.1	Differential Scanning Calorimetry (DSC) Methodology	24
3.2.4	Spectrum Analysis	25
3.2.4.1	Fourier Transform Infrared (FTIR) Methodology	
3.3	Quality Quantification	25
3.3.1	Free Fatty Acid (FFA)	25
3.3.2	Deterioration of Bleachability Index (DOBI)	25
3.4	Statistical Analysis	26
4	RESULTS AND DISCUSSION	27
4.1	Crude Palm Oil and admixtures quality quantification analysis	27
4.2	Detection of presense of adulterant in Crude Palm Oil by chromatographic techniques	29
4.2.1	Fatty acid and triacylglycerol composition of Crude Palm Oil and admixtures	29
4.2.2	Multivariate Analysis : Principle Component Analysis (PCA)	33
4.2.3	Multivariate Analysis : Dendrogram	35
4.3	Detection of present of adulterant in Crude Palm Oil by Differential Scanning Calorimetry (DSC) techniques	36
4.3.1	Monitoring of CPO adulteration by DSC cooling thermogram	37
4.3.2	Monitoring of CPO adulteration by DSC heating thermogram	45
4.3.3	Quantification of adulterant in CPO by cooling thermogram	47
4.3.4	Multivariate Analysis : Principle Component Analysis (PCA) based on DSC thermogram of TAG	49
4.3.5	Multivariate Analysis : Dendrogram based on DSC thermogram of TAG	51
4.4	Detection of present of adulterant in Crude Palm Oil by Fourier Transform Infrared Spectroscopy (FTIR) techniques	52
4.4.1	Interpretation of FTIR spectrum	52
4.4.2	Multivariate Analysis : Principle Component Analysis (PCA) based on FTIR Spectrum	58
4.4.3	Multivariate Analysis : Dendrogram based on FTIR spectrum	59
5	CONCLUSIONS AND RECOMMENDATION FOR FUTURE WORK	60
5.1	Conclusion	60
5.2	Limitations and Recommendations for future work	61
	REFERENCES	62
	APPENDIX	73
	LIST OF PUBLICATIONS	81

LIST OF TABLES

Table	Title	Page
2.1	Techniques and applications used for the determination of oil and fats authenticity	20
3.1	The temperature settings of the oven in gas chromatography system.	30
3.2	The evaporative light scattering detector's setting.	31
3.3	DSC Thermal Setting	32
4.1	FFA percentage (%) and DOBI value of genuine Crude Palm Oil (CPO) and admixtures with Sludge Oil (SO)	37
4.2	FFA percentage (%) and DOBI value of genuine Crude Palm Oil (CPO) and admixtures with Used vegetable oil containing animal fats (UVOA)	37
4.3	FFA percentage (%) and DOBI value of genuine Crude Palm Oil (CPO) and admixtures with Used vegetable oil (UVO)	38
4.4	Fatty Acid Composition (%) and Triacylglycerol EN (%) of Crude Palm Oil (CPO) and admixture with Sludge Oil (SO)	40
4.5	Fatty Acid Composition (%) and Triacylglycerol EN (%) of Crude Palm Oil (CPO) and admixture with Used vegetable oil containing animal fats (UVOA)	41
4.6	Fatty Acid Composition (%) and Triacylglycerol EN (%) of Crude Palm Oil (CPO) and admixture with Used vegetable oil (UVO)	42
4.7	Eigen analysis of the correlation matrix, fatty acid composition and triacylglycerol profile	45
4.8	Triacylglycerol of Crude Palm Oil (CPO) and admixtures with Sludge Oil (SO)	50
4.9	Triacylglycerol of Crude Palm Oil (CPO) and admixtures with Used vegetable oil (UVO)	51
4.10	Triacylglycerol of Crude Palm Oil (CPO) and admixtures with Used vegetable oil containing animal fats (UVOA)	52
4.11	Peak Intensity (Mean \pm sd) of T2 cooling thermogram based on the concentration (%) of Crude Palm Oil (CPO)	59
4.12	Eigen analysis of the correlation matrix, triacylglycerol profile	63

LIST OF FIGURES

Figure	Title	Page
4.1	Principle Component Analysis (PCA) scores of crude palm oil and admixtures	44
4.2	PCA loading distribution of fatty acid composition and triacylglycerol	46
4.3	Cluster Dendrogram Crude Palm Oil and admixtures	48
4.4	DSC Cooling thermograms of Crude Palm Oil (CPO) and adulterant	53
4.5	DSC Cooling thermograms of Crude Palm Oil (CPO) and Sludge oil (SO) admixtures	54
4.6	DSC Cooling thermograms of Crude Palm Oil (CPO) and Used Vegetable Oil (UVO) admixtures.	55
4.7	DSC Cooling thermograms of Crude Palm Oil (CPO) and Used Vegetable Oil Containing Animal Fat (UVOA) admixtures.	56
4.8	DSC Heating thermogram of Crude Palm Oil (CPO) and adulterant	58
4.9	Regression graph of concentration of CPO and CPO admixtures with peak intensity of T2 cooling thermogram	60
4.10	Regression graph of concentration of CPO (Actual) vs concentration of CPO (Predicted)	61
4.11	Principle Component Analysis (PCA) scores of Crude Palm Oil (CPO) and adulterant admixtures	62
4.12	PCA Loading distribution of triacylglycerol	63
4.13	Cluster Dendrogram of Crude Palm Oil (CPO) and adulterant admixtures	65
4.14	FTIR spectra of CPO and adulterant (SO,UVO,UVOA)	68
4.15	FTIR spectra of CPO and adulterated with SO and admixtures at frequency of 4000-650cm ⁻¹	69
4.16	FTIR spectra of CPO and adulterated with UVO and admixtures at frequency of 4000-650cm ⁻¹	70
4.17	FTIR spectra of CPO and adulterated with UVOA and admixtures at frequency of 4000-650cm ⁻¹	71
4.18	Principle Component Analysis (PCA) scores of Crude Palm Oil (CPO) and adulterant admixtures	73
4.19	Cluster Dendrogram of Crude Palm Oil (CPO) and used vegetable oil containing animal fats (UVOA)	74

LIST OF ABBREVIATIONS

CPO	Crude Palm Oil
PCA	Principle Component Analysis
SIMCA	Standard in Multivariate Data Analysis
PLS	Partial Least Square
GC	Gas Chromatograph
HPLC	High Performance Liquid Chromatograph
FT-IR	Fourier Transform Infrared
NMR	Nuclear Magnetic Resonance
NIR	Near Infrared Spectroscopy
DNA	Deoxyribonucleic Acid
FAME	Fatty Acid Methyl Esters
DSC	Differential Scanning Calorimetry
PCR	Polymerase chain reaction
SNP	Single nucleotide polymorphism
UVO	Used Vegetable Oil
UVOA	Used Vegetable oil Containing Animal Fats
SO	Sludge Oil
CI	Confident Interval
Df	Degree of Freedom

CHAPTER 1

INTRODUCTION

In the recent years, a great interest has been devoted to the use of vegetable fats in human diet, especially regarding olive oil and other vegetable edible oil. Since these oils have a vast implication for human health due to their nutritional properties. Most olive oil and some vegetable oils are rich in monounsaturated fatty acids, antioxidants and phytosterols which may contribute to prevention of heart disease and possible for cancer prevention (Giugliano et al., 2005). The importance of vegetable oils to the global economy becomes clear when considering the amount of vegetable oils produced and consumed worldwide. With exports and imports booming, strict control that aims to combat illegal attempts to boost profits is necessary. The increased attention has stimulated the interest in food authenticity. It covers many aspects including adulteration, mislabeling, characterization and misleading origin (Che Man et al., 2005). As in the global economy, traceability can be defined as the ability to track any food, feed, food-producing animal or substance that will be used for consumption, along all steps of production, processing and distribution (Costa et al., 2012).

Various vegetable oils have been reported to be adulterated, as adulteration of oil and fats falls into two main ways; vegetable oils adulterated with cheaper vegetable oils or vegetable oils adulterated with animal fats. For example, olive oil and virgin olive oil which are the two most expensive oils compared to other edible oils (Al-Ismail, 2010) were commonly adulterated with hazelnut oil, sunflower oil and soybean oils (Christopoulou et al., 2002; Gamazo-Vazquez, 2003; Oussama, 2012). Although the adulteration of vegetable oils does not pose a threat to the consumer's health, in the specific cases of adulteration of olive oil with hazelnut may possess potential food allergenic which lead to life threatening risk.

Whilst, lard contamination was detected in vegetable oil namely palm oil, palm kernel oil, canola oil as well in refined, bleached, deodorized palm olein (Marikkar et al., 2005 & Che Man et al., 2005). Pork and lard in food are serious matters from a religious point of view such as Islam as their religion prohibits their followers from consuming any food containing pork or lard (Che Man et al., 2005). The high content of saturated fatty acids and low proportion of polyunsaturated fatty acids in animal fats has major health risk factor, resulting in coronary heart disease (Marikkar et al., 2003).

There are several ways in which food can be mislabeled or characterized by including the substitution of one ingredient by a similar but cheaper one and over-declaring a quantitative ingredient (Gimenez et al., 2010). Also, the quality and uniqueness of edible vegetable oils namely virgin olive oil is a result of the different cultivar, environment and cultural practices. Thus, quality of the product is closely linked to its geographical origin and therefore the subject of adulteration is based on the mixing of oils from one region with those of lower quality grades for the case of virgin olive oil (Cosio et al., 2006). This practice is a commercial exploitation of olive oil and causes

major loss in economic value, it has been estimated a loss of 4 million euros per year for countries in the European Union due to these activities (Zabaras et al., 2004).

Relatively, adulteration doesn't only focus on the highest commercial value product such as olive oil and virgin olive oil. Virgin coconut oil, a newcomer in oils and fats markets was also counterfeited with palm kernel oil (Manaf et al., 2006, Rohman et al., 2009 & 2010). Adulteration practices have also extended to other edible oils such as Cod Liver and Red Fruits (*Pandanus conoideus*) plant in the pandan family. Cod Liver oil has been a focus of a growing interest due to its nutritional advantages. It contains high levels of long chains n-3 fatty acids cis-5,8,11,14,17- eicosapentaenoic (EPA) and cis-4,7,10,13,16,19-docosahexaenoic (DHA) which are believed to play a preventive role in cardiovascular disease and in the alleviation of other health problems (Rohman and Che Man, 2009). Cod Liver oil was adulterated with canola, corn, soybean and walnut oil which have a similar color which therefore makes it difficult to detect using naked eyes (Rohman and Che Man, 2011). Similar to Cod Liver oil, Red fruits which provides several treatments for degenerative disease such as cancer was adulterated with common edible vegetable oils with similar fatty acid profiles such as canola and rice bran oil (Rohman et al., 2010)

As edible vegetable oils market expands, their authenticity has become an important subject from both commercial and health perspective. From the legislative point of view, quality standards have been established through the requirement of quality labels that specify the chemical composition of each product. In olive oil, the International Olive Oil Council (IOOC) has introduced methods and limits for the trade standard in olive oil. Among the established methods for determining the authenticity of olive oils were by determining fatty acids and triacylglycerol composition (Christopoulou et al., 2004; Mariani et al., 2006; Monfreda et al., 2012; Ruiz-Samblas et al., 2012). From the economical point of view, product authentication is essential to avoid unfair competition that can create a destabilized market and disrupt the regional economy and even the national economy (Cordella, 2002). This is done by selling adulterated products at below the cost of the pure production which can drive producers and packers to be out of business. Without direct evidence of adulteration, these impacts result in bad economic growth as well as reduce consumer's confidence towards the product (Fairchild et al., 2003).

Similar to the common edible oils such as olive oil, virgin olive oil and virgin coconut, in recent years there has been rising production (supply) and consumption (demand) of crude palm oil. The demand for domestic consumption has gradually increased with a rather slow growth in production (Ramli, 2011). This widening gap between demand and production thus leads to frauds or adulteration. In Nigeria, the adulteration is believed to be practiced by producers in order to increase the quantity of crude palm oil (CPO), for the sole purpose of profit maximization (Okogeri, 2013). The adulteration practice is normally done without considering its possible effect on the quality of palm oil and the health of consumers. Red dye commonly used in the coloring of leather, coloring of clothes, calabashes and as a body pigment is added into crude palm oil. This additive is to improve the colour and thus increase the market value without knowing the effect of the red dye on consumers health (Okonkwa et al., 2010; Okogeri, 2013).

At present in Malaysia, there is no published investigation of adulteration of crude palm oil reported to our best knowledge. However, possible adulterants which could be used include sludge oil and used cooking oil (from deep frying) which were added into crude palm oil due to their abundance supply and similarity to the genuine oil (author, personal communication). The oil palm is a monocotyledon belonging to the genus *Elaeis*. It is a perennial tree crop and the highest oil producing plant which yield averagely at 3.7 tonnes of oil per hectare per year in Malaysia (Sundram, 2001). The crop is unique in that it produces two types of oil. The fleshy mesocarp produces palm oil which is used mainly for its edible properties and the kernel produces palm kernel oil which has wide application in the oleochemical industry. Currently, Malaysia and Indonesia produce about 85% of world's crude palm oil.

Extraction of crude palm oil (CPO) from fresh fruit bunches require steam for sterilization and water for dilution, which finally contribute to substantial amounts of water being discharged as sludge oil or palm oil mill effluent (POME) (Wafiti et al., 2010). This generated about 60% of sludge oil for every tonne of fresh fruit bunches (FFB) processed in mills. As for recycled oil, countries in Asia such as China, Malaysia, Indonesia, Thailand, Hong Kong, and India produces about 40,000 tonnes per year of used cooking oil or waste cooking oil (Ismail, 2005). Improper waste management of used cooking oil leads to discharge to environment which leads to environment pollution or returned to human for consumption through food chains (Hanisah et al., 2013).

The increased demand for edible food oil caused entailed shortages of materials. Consequently, to fulfill the demand, common type of edible food oils were adulterated with cheaper materials as well as to improve the visual appearance of impure food oil. Despite the existence of food regulations which ensure that the standards are fulfilled, the constant evolution of adulteration practices have resulted in continuous amendment of the food regulation over time. As customers' concern about the quality and safety of the edible oil increases, significant progress in natural science and analytical science were established to ensure food quality. Advances in analytical chemistry provided knowledge about the composition and properties of food which could be used to differentiate between genuine and adulterated products. In detecting adulteration in olive oil, analytical chemistry instrument such as chromatograph, thermal and spectral analysis were used to characterize the chemical compositions of olive oil and its adulterants, this analytical method is adapted in these studies.

Chromatographic techniques consist of gas and liquid chromatography which analyses are based on the separation of components. Advantages of chromatographic technology are its high separation efficiency and most accurate quantify results (Zhang et al., 2011). Whereby, thermal analysis by differential scanning calorimetry (DSC) allows the physical change that occur upon heating and cooling were able to be determined. These technique possesses the advantages of being relatively quick and simply to carry out with minimal sample preparation (Marikkar et al., 2003). Similarly by spectroscopy techniques, samples are analyses rapidly and non-destructively. Information about the molecular bonds present and therefore give details of the types of molecules present in the samples. These techniques provides a relatively low financial cost of obtaining and

running the instrument (Reid et al., 2006).

Common edible oils such as olive oil and virgin coconut oil are the primary targets for adulteration in European countries. In the countries at tropical belt of Africa, Southeast Asia and parts of Brazil, crude palm oil is a main commodity to be adulterated. With an increase in demand for crude palm oil in these regions, it is not surprising that attempts by some devious suppliers to maximize revenues by counterfeiting and adulterating practices are conducted. Taken that, the main objectives of this project is to determine adulteration of crude palm oil by characterisation of the physiochemical properties of crude palm oil and its adulterant. To achieve the main objectives of the project, specific studies were designed as following :

- i. To investigate current quality quantification by Free Fatty Acid, FFA and Deterioration of Bleachability Index, DOBI is able to detect adulteration.
- ii. To characterize the crude palm oil, adulterant and admixtures of crude palm oil and adulterant via chromatographic techniques.
- iii. To characterize the crude palm oil, adulterant and admixtures of crude palm oil and adulterant via thermal behaviour based on the crystallization and melting profiles.
- iv. To characterize the molecular absorption and transmission of crude palm oil, adulterant and admixtures of crude palm oil and adulterant via fourier transform infrared (FTIR)
- v. To process obtain results by multivariate analysis and regression modelling to quantify concentration of adulterant in crude palm oil blends.

REFERENCES

- Ainie, K., Siew, W.L., Tan, Y.A., Noraini, I., Mohtar, Y. Tang, T.S., & Nuzul Amri, I. 2005. MPOB test method : A compendium of test on plam oil products, palm kernel product , fatty acids, food related products and other. *Malaysia Palm Oil Board*.
- Ainie, K., Siew, W.L., Tan, Y.A. & Ma, A.N. 1995. Characterization of a by product of palm oil milling. *Elaeis*, 7 (2) : 165-173.
- Agiomyrgianaki, A., Petrakis, P.V. & Dais, P. 2010. Detection of refined olive oil adulteration with refined hazelnut oil by employing NMR spectroscopy and multivariate statistical analysis.
- Agrimontic, C., Vietina, M., Pafundo, S. & Marmioli, N. 2011. The use of food genomics to ensure the traceability of olive oil. *Trends in Food Science & Technology*, 22: 237-244.
- Al-Ismail, K., Alsaed, A.K., Ahmad, R. & Al-Dabbas. 2010. Detection of olive oil adulteration with some plant oils by GLC analysis of sterols using polar column. *Food Chemistry*, 121: 1255-1259.
- Aparocop, R., Aparicio-Ruiz, R. 2000. Authentication of vegetable oils by chromatographic methods. *J. Chromatogr. A.*, 881 : 93 -104.
- Aktas, N. & Kaya, M. 2001. Detection of beef body fat and margarine in butterfat by differential scanning calorimetry. *Journal of Thermal Analysis and Calorimetry*, 66; 795-801.
- Al-Ismail, K.M., Alsaed, A.K., Ahmad, R., & Al-Dabbas, M. 2010. Detection of olive oil adulteration with some plant oils by GLC analysis of sterols using polar column. *Food Chemistry*, 121: 1255-1259.
- Angerosa, F., Servili, M., Selvaggini, R., Taticchi, A., Esposto, S. & Monteroda, G. 2004. Volatile Compounds in Virgin Olive Oils Occurrence and Their Relationship with the Quality. *J. Chromatogr. A*, 1054; 17-31.

- Ainie, K., Siew, W., L., Tan, Y., A., Noraini, I., Mohtar, Y., Tang, T., S. & Nurul Amri, I. 2005. MPOB Test Method : A Compendium of Test on Palm Oil Products, Palm Kernel Product, Fatty acids, Food Related Products and other. *Malaysia Palm Oil Board*.
- Bohacenko, I. & Kopicova, Z. 2001. Detection of Olive Oils Authenticity by Determination of Their Sterol content Using LC/GC. *Czech Journal of Food Science*, 19; 97-103.
- Cerretani, L., Maggio, R.M., Barnaba, C., Toschi, T.G & Chiavaro, E. 2011. Application of partial least square regression to differential scanning calorimetry for fatty acid quantitation in olive oil. *Food Chemistry*, 127: 1899-1904.
- Cercari, L., Rodriguez-Estrada, M.T. & Lercker, G. 2003. Solid-phase extraction thin layer chromatography gas chromatography method for the detection of hazelnut oil in olive oil by determination of esterified sterols. *Journal of Chromatography*, 985: 211-220.
- Che Man, Y.B., Haryati, T., Ghazali, H.M., & Asbi, B.A. 1999. Composition and thermal profile crude palm oil and its products. *JAOCs*, 76 (2) : 237- 242.
- Chen, H., Angiuli, M., Ferrari, C., Tombari, E., Salvetti, G. & Bramanti, E. 2011. Tocopherol speciation as first screening for the assessment of extra virgin olive oil quality by reversed-phase high-performance liquid chromatography/fluorescence detector.
- Chiavaro, E., Vittadini, E., Rodriguez-Estrada, M.T., Cerretani, L. & Bendini, A. 2008. Differential scanning calorimetry application to the detection of refined hazelnut oil in extra virgin olive oil. *Food Chemistry*, 110 ; 248-256.
- Christopoulou, E., Lazaraki, M., Komaitis, M. & Kaselimis, K. 2004. Effectiveness of determinations of fatty acids and triacylglycerol for the detection of adulteration of olive oils with vegetable oils. *Food Chemistry*, 84; 463-474.
- Christy, A.A., Kasemsumran, S. & Ozaki, Y. 2004. The detection and quantification of adulteration in olive oil by near-infrared spectroscopy and chemometrics. *Anal. Sci.* 20, 935-940.
- Cordella, C., Moussa, I., Martel, A., C., Sbirrazzuoli, N. & Lizzani-Cuvelier, L. 2002. Recent Developments in Food Characteriation and Adulteration Detection :

Techniques-Oriented Perspectives. *Journal of Agricultural and Food Chemistry*, 50; 1751-1764.

- Cosio, M.S., Ballabio, D., Benedetti, S. & Gigliottic, C. 2006. Geographical origin and authentication of extra virgin olive oils by an electronic nose in combination with artificial neural network. *Analytica Chimica Acta*, 567; 202-210.
- Costa, J., Mafra, I. & Oliveira, P.P. 2012. Advances in vegetable oils authentication by DNA-based markers. *Trends in Food Science & technology*, 26: 43-55.
- Cuncha, S., C. & Oliveira, M.B.P.P. 2006. Discrimination of vegetable oils by triacylglycerols evaluation of profile using HPLC/ELSD. *Food Chemistry*, 95; 518-524.
- Damirchi, S., A., Savage, P., G. & Dutta, P., C. 2005. Sterol Fractions in Hazelnut and Virgin Olive Oils and 4,4-Dimethylsterols as Possible Markers for Detection of Adulteration of Virgin Olive Oil. *JAOCS*, 82; 717- 725.
- Diaz, T., G., Meras, I., D., Casas, J., S. & Franco, M.F. 2005. Characterization of virgin olive oils according to its triacylglycerol and sterols composition by chemometric methods. *Food Control*, 16, 339-347.
- Dourtoglou, V., G., Dourtoglou, TH., Antonoploulos, A., Stefaou, E., Lalas, S. & Poulos, S. C. 2003. Detection of Olive Oil Adulteration Using Principal Component Analysis Applied on Total and Regio FA Content. *JAOCS*, 80; 203-207.
- Downey, G., McLytyre, P. & Davies, A.N. 2002. Detecting and quantifying sunflower oil adulteration in extra virgin olive oils from the eastern Mediterranean by visible and near infrared spectroscopy. *Journal of Agricultural and Food Chemistry*. 20; 5520-5525.
- Fairchild, G.F., Nichols, P.J., & Capps, O. 2003. Observations on economic adulteration of high value food products. *Journal of Food Distribution Research*, 34: 38-45.
- Fang, G., Goh, J.Y., Tay, M., Lau, H.F. & Li, S.F.Y. 2013. Characterization of oils and fats by ¹H NMR and GC/MS fingerprinting : Classification, prediction and detection of adulteration. *Food Chemistry*, 138: 1461-1469.

- Fauhl, C., Reniero, F. & Guillou, C. 2000. ^1H NMR as a tool for the analysis of the mixtures of virgin olive oils with oils of different botanical origin. *Magnetic Resonance in Chemistry*, 38; 436-443.
- Fadzilah, K. & Mashitah, M.D. 2010. Cellulases production in palm oil mill effluent : Effect of aeration and agitation. *Journal of Applied Science*, 10(24) : 3307-3312.
- Fasciotti, M., Annibal, D. & Netto, P. 2010. Optimization and application of methods of triacylglycerol evaluation for characterization of olive oil adulteration by soybean oil with HPLC-APCI-MS-MS. *Talanta*, 81: 1116-1125.
- Flores, G., Castillo, M., L., R., Herraiz, M. & Blanch, G., P. 2006. Study of the Adulteration of Olive Oil with Hazelnut oil by on-line coupled High Performance Liquid Chromatographic and Gas Chromatographic Analysis of Filbertone. *Food Chemistry*, 97; 742-749.
- Fragaki, G., Spyros, A., Siragakis, G., Salivaras, E. & Dais, P. 2005. Detection of extra virgin olive adulteration with lampante olive oil and refined olive oil using nuclear magnetic resonance spectroscopy and multivariate statistical analysis. *Journal of Agriculture and Food Chemistry*. 53; 2810-2816.
- Gamazo-Vazquez, J., Garcia-Falcon, M., S. & Simal-Gandara, J. 2003. Control of Contamination of Olive Oil by sunflower seed oil in bottling plants by GC-MS of fatty acid methyl esters. *Food Control*, 14; 463-467.
- Gan, H.L., Che Man, Y.B., Tan, C.P., Noraini, I, I. & Nazimah, S.A.H. 2005. Characterisation of vegetable oils by surface acoustic wave sensing electronic nose. *Food Chemistry*, 89: 507-518.
- Gimenez, M.J., Piston, F., Martin, A. & Atienza, S.G. 2010. Application of real time PCR on the development of molecular markers and to evaluate critical aspects for olive oil authentication. *Food Chemistry*, 118: 482-487.
- Gurdeniz, G. & Ozen, B. 2009. Detection of adulteration of extra virgin olive oil by chemometric analysis of mid-infrared spectral data. *Food Chemistry*, 116; 519-525.
- Giugliano, D. & Esposito, K. 2005. Mediterranean diet and cardiovascular health. *Annals of the New York Academy of Sciences*, 1056, 253-260.
- Hanisah, K., Kumar, S. & Tajul, A.Y. 2013. The management of waste cooking oil ; A Preliminary Survey. *Health and Environment Journal*, 4(1) :76-81.

- Hajimahmoodi, M., Vander Heyden, Y., Sadeghi, N., Jannat, B., Oveisi, M., R., & Shahbazian, S. 2005. Gas Chromatographic Fatty Acid Fingerprints and Partial Least Squares Modeling as a basis for the Simultaneous Determination of Edible Oil Mixtures. *Talanta*, 66; 1108-1116.
- Ismail, R. 2005. Palm oil and palm olein frying applications. *Asia Pac. J. Clin. Nutr.*, 14(4) : 414-419.
- James, D., Scott, S.M. O'Hare, W.T. Ali, Z. & Rowell, F.J. 2004. Clasification of fresh edible oils using a coated piezoelectric sensor array-based electronic nose with soft computing approach for pattern recognition. *Transactions of the institute of Measurement and Control*, 26:3-18.
- Jin, Q., Zhang, T., Shan, L., Liu, Y. & Wang, X. 2008. Melting and solidification of palm kernel oil, tallow and palm olein blends in the preparation of shortening. *J. Am. Oil Chem. Soc.*, 85 ; 23-28.
- Katz, M.H. 2003. Multivariable analysis : A primer for readers of medical research. *Ann Intern Med*, 138:644-650.
- Kasemsumra, S., Kang, N., Christy, A. & Ozaki, Y. 2005. Partial Least Square processing of near infrared spectra for discrimination and quantification of adulterated olive oil. *Spectrosc. Lett.* 38, 839-851.
- Khalid, M., Al-Ismail, Alsaed, A., K., Ahmad, R. & Al-Dabbas, M. 2010. Detection of Olive Oil Adulteration with some Plant Oils by GLC analysis of Sterols using polar column. *Food Chemistry*, 121; 1255-1259.
- Kumar, S., Kahlon, T. & Chaudhary, S. 2011. A rapid screening for adulterants in olive oil using DNA barcodes. *Food Chemistry*, 127: 1335-1341.
- Lerma-Garcia, M.J., Ramis-Ramos, G., Herrero-Martinex, J.M. & Simo-Alfonso, E.F. 2009. Authentication of extra virgin olive oils by fourier transform infrared spectroscopy. *Food Chemistry*, in press.

- Lee, D.S., Lee, K.S., Kim, H.J., Kim, S.O. & Kim, K. 2001. Reversed phase liquid chromatographic determination of triacylglycerol composition in sesame oils and the chemometric detection of adulteration. *Analytica Chimica Acta*, 429; 321-330.
- Mata, P., Dominguez-Vidal, D., Bosque-Sendra, J.M., Ruiz-Medina, A., Cuadros-Rodriguez, L. & Ayora-Canada, M.J. 2012. Olive oil assessment in edible oil blends by means of ATR-FTIR and chemometric. *Food Control*, 23; 449-455.
- Maggio, R.M., Kaufman, T.S., Carlo, M.D., Cerretani, L., Cichelli, A.A. & Compagnone, D. 2009. Monitoring of fatty acid composition in virgin olive oil by fourier transform infrared spectroscopy coupled with partial least squares. *Food Chemistry*, 114; 1549-1554.
- Maggio, R.M., Cerretani, L., Chiavaro, E., Kaufman, T.S., & Bendini, A. 2010. A novel chemometric strategy for the estimation of extra virgin olive oil adulteration with edible oils. *Food Control*, 21; 890-895.
- Margari, M.T. & Okogeri, O. 2001. Simultaneous determination of phenolic compounds and tocopherols in virgin olive oil using HPLC and UV detection. *Food Chemistry*, 74; 377-383.
- Marina, A.M., Che Man, Y.B. & Amin, I. Use of the SAW sensor electronic nose for detecting the adulteration of virgin coconut oil with RBD palm kernel olein. *J. Am Oil Chem. Soc.*, 87:263-270.
- Marikkar, J.M.N., Ghazali, H.M., Che Man, Y.B., & Lai, O.M. 2003. Differential scanning calorimetric analysis for determination for some animal fats as adulterants in palm olein. *Journal of Food Lipids*, 10; 63-79.
- Marikkar, J.M.N., Ghazali, H.M., Che Man, Y.B., & Lai, O.M. 2002. The use of cooling and heating thermograms for monitoring of tallow, lard and chicken fat adulteration in canola oil. *Food Research International*, 35 ; 1007-1014.
- Marikkar, J.M.N., Ghazali, H.M., Long, K. & Lai, O.M. 2003. Lard uptake and its detection in selected food products deep fried in lard. *Food Research International*, 36: 1047-1060.
- Marikkar, J.M.N., Ghazali, H.M., Che Man, Y.B., & Lai, O.M. 2002. Compositional and thermal analysis of RBD palm oil adulterated with lipase-catalyzed interesterified lard. *Food Chemistry*, 2002; 249-258.

Marikkar, J.M.N., Ghazali, H.M., Che Man, Y.B., & Lai, O.M. 2001. Detection of lard and randomized lard as adulterants in refined bleached-deodorized palm oil by differential scanning calorimetry. *Journal of the American Oil Chemists' Society*, 78; 1113-1119.

Marikkar, J.M.N., Ghazali, H.M., Che Man, Y.B., Peiris, T.S.G. & Lai, O.M. 2005. Distinguishing lard from other animal fats in admixtures of some vegetable oils using liquid chromatographic data coupled with multivariate data analysis. *Food Chemistry*, 91: 5-14.

Marikkar, J.M.N. 2004. Detection of lard and interesterified lard as adulterants in some vegetable oils and selected fried food product. Thesis

Mariani, C., Bellan, G., Lestini, E. & Ramon Aparicio. 2006. The Detection of the Presence of Hazelnut Oil in Olive Oil by Free and Esterified Sterols. *Eur. Food Res. Technol*, 223; 655-661.

Marina, A.B., Che Man, Y., Hamid, Z.S.A., Ismail, A. & Abidin, S.Z. 2006. Analysis of adulteration of virgin coconut oil by palm kernel olein fourier transform infrared spectroscopy. *Journal of Food Lipids*, 14; 111-121.

Marina, A.M, Che Man, Y.B, Nazimah, S.A.H. & Amin, I. 2008. Monitoring the adulteration of virgin coconut oil by selected vegetable oils using differential scanning calorimetry. *Journal of Food Lipid*, 16 ; 50-61.

Martin, Y.G., Oliveros, M.C.C., Pavon, J.L.P., Pinto, C.G. & Cordero, B.M. 2001. Electronic nose based on metal oxide semiconductor sensors and pattern recognition techniques: Characterisation of vegetable oils. *Analytica Chimica Acta*, 449: 69-80.

Manaf, M.A., Che Man, Y., Abdul Hamid, N., S. & Abidin, S., Z. Analysis of adulteration of virgin coconut oil by palm kernel olein using fourier transform infrared spectroscopy. *Journal of Food Lipid*, 14; 111-121.

Mannina, L. Dugo, G., Salvo, F., Cicero, L., Ansanelli, G. & Segre, A. Italian and Argentine olive oils: a NMR and gas chromatographic study. *Grasas Y Aceites*, 52: 380-388.

Mata-Espinosa, P., Bosque-Sendra, J.M., Bro, R. & Cuadros-Rodriguez, L. 2011. Olive Oil Quantification of Edible Vegetable Oil Blends using triacylglycerol Chromatographic fingerprint and chemometric tools. *Talanta*, 2011. 177-182. .

- Meza-Marguez, O., Gallardo-Velazquez, T. & Osorio-Revilla, G. 2010. Application of mid infrared spectroscopy with multivariate analysis and soft independent modeling of class analogies (SIMCA) for the detection of adulterants in minced beef. *Meat Science*, 86(2): 511-519
- Mildner-Szkudlarz, S. & Jelen, H., H. 2008. The potential of different techniques for volatile compound analysis coupled with PCA for the detection of the adulteration of olive oil with hazelnut oil. *Food Chemistry*, 110; 751-761.
- Morsy, N. & Sun, D.W. 2013. Robust linear and non linear model of NIR spectroscopy for detection and quantification of adulterants in fresh and frozen thawed minced beef. *Meat Science*, 93(2): 292-302.
- Monfreda, M., Gobbi, L. & Grippa, A. 2012. Blends of Olive Oil and Sunflower oil : Characterisation and Olive Oil quantification using fatty acids composition and chemometric tools. *Food Chemistry*, 134; 2283-2290.
- Ogrinc, N., Kosir, I.J., Spangenberg, J.E. & Kidric, J. 2003. The application of NMR and MS methods for detection of adulteration of wine, fruit juices and olive oil. A review. *Anal. Bioanal Chem*, 376; 424-430.
- Okogeri, O. 2013. Adulteration of Crude Palm Oil with Red Dye from Sheath of Sorghum Bicolor. *Food Science and Quality Management*, 17;1-7.
- Okonkwo, S.,I. & Ogbunike, R.,U. 2010. Assessment of Level of Adulteration in Palm Oil (*Elaeis Guineensis*) within Ihiala Local Government Area of Anambra State Nigeria. *Journal of Basic Physical Research*, 1; 13-16.
- Oliveros, M.C.C., Pavon, J.L.P., Pinto, C.G., Laespada, M.E.F., Cordero, B.M. & Forina, M. 2002. Electronic nose based on metal oxide semiconductor sensors as a fast alternative for the detection of adulteration of virgin olive oils. *Analytica Chimica Acta*, 459: 219-228.
- Oussama, A., Elabadi, F., Platikanov, S., Kzaiber, F. & Tauler, R. 2012. Detection of olive oil adulteration using FT-IR spectroscopy and PLS with variable important of projection (VIP) scores. *J. Am. Oil Chem. Soc.*,
- Ozdemir, D. & Ozturk, B. 2007. Near infrared spectroscopic determination of olive oil adulteration with sunflower and corn oil. *Journal of Food and Drug Analysis*, 15; 40-47.

- Ozen, B.F & Mauer, L.J. 2002. Detection of hazelnut oil adulteration using FTIR spectroscopy. *J Agric Food Chem* 50:3898–3901.
- Tan, Y.A. 2013. Palm oil quality standard for trading. PORAM Course on “Operational and Commercial Aspects of Palm Oil Trade.
- Timm, N.H. 2002. Applied Multivariate Analysis. Springer text in statistics.
- Tian, X., ang, J. & Cui, S. 2013. Analysis of pork adulteration in minced mutton using electronic nose of metal oxide sensors. *Journal of Food Engineering*, 119(4):744-749.
- Ramon Aparicio & Ramon Aparicio-Ruiz. 2000. Authentication of Vegetable Oils by Chromatographic. *Journal of Chromatograph A*, 881 ; 93-104.
- Rohman, A., Sisindari, Erwanto, Y. & Che Man, Y.B. 2011. Analysis of pork adulteration in beef meatball using fourier transform infrared (FTIR) spectroscopy. *Meat Science*, 88(1): 91-95.
- Rohman, A. & Che Man, Y.B. 2009. Analysis of Cod-Liver oil adutleation using fourier transform infrared (FTIR) spectroscopy. *J Am Oil Chem Soc*, 86:1149-1153.
- Rohman, A., Che Man, Y.B. & Riyanto, S. 2010. Authentication analysis fo red fruits(pandanus conoideus lam) oil using FTIR spectroscopy in combination with chemometrics. *Phytochemical Analysis*, 22:462-467.
- Rohman, A. & Che Man, Y.B. 2011. Application of fourier transform infrared (FTIR) spectroscopy combined with chemometrics for authentication of cod liver oil. *Vibrational Spectroscopy*, 55: 141-145
- Ruiz-Samblas, C., Marini, F., Cuadros-Rodriquez, L. & Goncales-Casado, A. 2012. Quantification of blending of olive oil and edible vegetable oil by triacylglycerol fingerprint gas chromatography and chemometric tools. *Journal of Chromatography*.
- Reid, L., M., O'Donnell, C., P. & Downey, G. 2006. Recent technological advances for the determination of food authenticity. *Trends in Food Sciene & Technology*, 17 : 344-353.

- Ramli Abdullah. 2011. World Palm Oil Supply, Demand, Price and Prospects: Focus on Malaysian and Indonesian Palm Oil Industries. *Oil Palm Industry Economic Journal*, 11, 13-25
- Sun, W.D. 2008. Modern techniques for food authentication. *Academic Press*.
- Sundram, K. 2001. Palm Oil : Chemistry and Nutrition Updates. *Malaysia Palm Oil Board (MPOB)*, 1-23.
- Smejkalova, D. & Piccola, A. 2010. High power gradient diffusion NMR spectroscopy for the rapid assessment of extra-virgin olive oil adulteration. *Food Chemistry*, 118;153-158.
- Syahriza, Z.A., Che Man, Y.B., Selamat, J. & Bakar, J. 2005. Detection of lard adulteration in cake formulation by fourier transform infrared (FTIR) spectroscopy. *Food Chemistry*, 95: 365-371.
- Tay, A., Singh, R.K., Krishnan, S.S & Gore, J.P. 2002. Authentication of olive oil adulterated with vegetable oils using fourier transform infrared spectroscopy. *LWT Food Sci Technol* 35:99-103
- Tasioula-Margari, M. & Okogeri, O. 2001. Simultaneous determination of phenolic compounds and tocopherols in virgin olive oil using HPLC and UV detection. *Food Chemistry*, 74; 377-383.
- Vigli, G., Philippidis, A., Spyros, A., & Dais, P. 2003. Classification of edible oils by employing P-31 and H-1 NMR spectroscopy in combination with multivariate statistical analysis. A proposal for the detection of seed oil adulteration in virgin olive oil. *Journal of Agriculture and Food Chemistry*, 51; 5715-5722.
- Vlachos, N., Skopelitis, Y., Psaroudaki, M., Konstantinidaou, V., Chatzilazarou, A., & Tegou, E. 2006. Application of Fourier transform infrared spectroscopy to edible oils. *Analtuca Chimica Acta*, 459-465.
- Wafti, N.S., Laum, H.L.N. & Choo, Y.M. 2010. Refining technology of sludge oil for industrial application. *MPOB Information Series*, ISSN 1511-7871.
- Wilson, A.D. & Baietto, M. 2009. Application and advances in electronic nose technologies. *Sensors*, 9: 5099-5148.

Wu, Y., Zhang, H., Han, J., Wang, B., Wang, Ju, X. & Chen, Y. 2011. PCR-CE-SSCP Applied to Detect Cheap Oil Blended in Olive Oil. *Eur Food Res Technol*, 233; 313-324.

Yang, H & Irudayaraj, J. 2001. Comparison of Near infrared, fourier transform infrared, and fourier transform raman methods for determining olive pomace oil adulteration in extra virgin olive oil. *JOACS*, 78 : 889-895.

Zhang,H., Wu,Y., Li,Y., Wang,B., Han,J., Ju,J. & Chen,Y. 2012. PCR-CE-SSCP used to authenticate edible oils. *Food Control*, 27: 322-329.

Zhang, J., Zhang, X., Dediu, L. & Victor, C. 2011. Review of the Current Application of Fingerprinting Allowing Detection of Food Adulteration and Fraud in China. *Food Control*, 22; 1126-1135.

Zabaras, D. & Gordon, M.H. 2004. Detection of pressed hazelnut oil in virgin olive oil by analysis of polar components : improvement and validation of the method. *Food Chemistry*, 84; 474-483

PUBLICATIONS

Published

Inthiram, A.K., Mirhosseini, H., Tan, C.H., Mohamad, R. & Lai, O.M. 20145. Application of Multivariate Analysis for Detection of Crude Palm Oil Adulteration through Fatty Acid Composition and Triacylglycerol Profile. *PERTANIKA*.

