



**UNIVERSITI PUTRA MALAYSIA**

***ANTIOXIDANT-MEDIATED DEFENSE RESPONSE OF *Carica papaya* L.  
VAR. EKSOTIKA AGAINST A COMPATIBLE *Erwinia mallotivora*  
STRAIN BT-MARDI***

**SUHAINA SUPIAN**

**FBSB 2015 15**



**ANTIOXIDANT-MEDIATED DEFENSE RESPONSE OF  
*Carica papaya* L. VAR. EKSOIKA AGAINST A COMPATIBLE  
*Erwinia mallotivora* STRAIN BT-MARDI**

By

**SUHAINA SUPIAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfilment of the Requirements for the Degree of Master of  
Science**

**February 2015**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**ANTIOXIDANT-MEDIATED DEFENSE RESPONSE OF *Carica papaya* L. VAR. EKSOTIKA AGAINST A COMPATIBLE *Erwinia mallotivora* STRAIN BT-MARDI**

By

**SUHAINA SUPIAN**

**February 2015**

**Chairman : Associate Professor Mohd Puad Abdullah, PhD**

**Faculty : Biotechnology and Biomolecular Sciences**

Papaya dieback disease, caused by the bacterium *Erwinia mallotivora*, is the most devastating papaya disease in Malaysia. Most papaya cultivars in Malaysia are susceptible to this disease which suggests that the molecular basis of the interaction between these cultivars and the pathogen is based on a compatible interaction, leading to disease development. However, in certain pathosystems, such interaction could also develop moderate defense response mediated by antioxidant system. Hence, it was hypothesized that early defense response in compatible interaction of papaya and *E. mallotivora* is mediated by antioxidant system. Therefore, this study was aimed to determine the expression of antioxidant-related genes and proteins and activity of antioxidant-related enzymes in the early response of Eksotika papaya to *E. mallotivora* infection. In addition, this study also optimized the inoculation procedures for artificial infection of Eksotika papaya with this pathogen. Two important parameters in the artificial infection of *E. mallotivora*, inoculum concentration and leaf position, affected significantly the disease development in papaya. The probability of the infection depends on the number of bacteria inoculated where greater number of bacteria is required to defeat host defenses as well as the position of leaf where lower or older leaf is less metabolically active thus, lead to higher susceptibility to this infection. Therefore, for efficient artificial infection of papaya with this pathogen, the use of second leaf from the top and inoculum concentration of  $10^7$  CFU/inoculation site was recommended. Using these optimized parameters, the inoculation of papaya with this pathogen was carried out and the inoculated leaf samples were collected at 0, 2, 4, 8, 12 and 24 h of post-infection (hpi). Following the bacterial challenge, through semi-quantitative reverse-transcription-PCR (RT-PCR) method, the expression of two selected antioxidant marker genes [superoxide dismutase (*SOD*) and peroxidase] was analyzed. *SOD* gene was found to be transiently up-regulated at 2 hpi whereas, peroxidase gene was up-regulated throughout the experimental

period. Through enzyme assays, enzymatic activity of SOD and peroxidase also showed a significant progressive increase occurred after 8 hpi (p-value<0.05). These observations were further confirmed through protein profiling of the infected leaf proteome at 4 and 8 hpi using two dimensional-polyacrylamide gel electrophoresis (2D-PAGE) and mass spectrometry analysis where a significant up-regulation of SOD protein at 4 hpi was observed (p-value<0.10). Overall, the findings in this study proved that antioxidant system is involved in early defense response of papaya to *E. mallotivora* infection which may suggest that this defense pathway is also activated in this compatible interaction. This study provides new insights into molecular mechanisms underlying this interaction.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains (*Bioteknologi Tumbuhan*)

**TINDAKBALAS PERTAHANAN AWAL *Carica papaya* L. VAR. EKSOTIKA MELAWAN STRAIN BT-MARDI *Erwinia mallotivora* YANG SERASI YANG DIPERANTARAI OLEH SISTEM ANTIOKSIDAN**

Oleh

**SUHAINA SUPIAN**

**Februari 2015**

**Pengerusi : Profesor Madya Mohd Puad Abdullah, PhD**

**Fakulti : Bioteknologi dan Sains Biomolekul**

Penyakit mati rosot betik, yang disebabkan oleh bakteria *Erwinia mallotivora*, adalah penyakit betik yang paling dahsyat di Malaysia. Kebanyakan kultivar betik yang ditanam di Malaysia adalah rentan kepada penyakit ini di mana ini menunjukkan bahawa asas molekul interaksi antara kultivar dan patogen ini adalah berasaskan interaksi yang serasi, yang membawa kepada pembangunan penyakit. Walau bagaimanapun, dalam sistem-sistem tumbuhan-patogen yang tertentu, interaksi seperti itu boleh juga membangunkan tindak balas pertahanan yang sederhana yang diperantarai oleh sistem antioksidan. Oleh itu, dihipotesiskan bahawa tindak balas pertahanan awal dalam interaksi serasi antara betik dan *E. mallotivora* diperantarai oleh sistem antioksidan. Oleh itu, kajian ini bertujuan untuk menentukan ekspresi gen dan protin yang berkaitan antioksidan dan aktiviti enzim yang berkaitan antioksidan dalam tindak balas awal betik Eksotika terhadap jangkitan *E. mallotivora*. Selain itu, kajian ini juga mengoptimumkan prosedur inokulasi untuk jangkitan buatan betik Eksotika dengan patogen ini. Dua parameter yang penting dalam jangkitan buatan *E. mallotivora*, kepekatan inokulum dan kedudukan daun, mempengaruhi pembangunan penyakit pada betik secara signifikan. Kebarangkalian jangkitan bergantung kepada jumlah bakteria yang diinokulasi di mana bilangan bakteria yang lebih besar diperlukan untuk mengalahkan pertahanan perumah dan kedudukan daun di mana daun yang lebih rendah atau lebih tua adalah kurang aktif secara metabolisma yang membawa kepada kerentanan yang lebih tinggi terhadap jangkitan ini. Oleh yang demikian, untuk jangkitan buatan betik dengan patogen ini dengan efisien, penggunaan daun kedua dari atas dan kepekatan inokulum pada  $10^7$  CFU/tapak inokulasi adalah diusulkan. Menggunakan parameter-parameter yang dioptimumkan ini, inokulasi betik dengan patogen ini telah dijalankan dan sampel daun yang diinokulasi telah dikumpulkan pada

0, 2, 4, 8, 12 dan 24 jam selepas jangkitan (hpi). Berikutan cabaran bacteria tersebut, melalui kaedah kuantitatif separa transkripsi terbalik-PCR (RT-PCR), pengekspresan dua gen penanda antioksidan yang dipilih [superoxide dismutase (SOD) dan peroxidase] telah dianalisa. Didapati gen SOD telah diregulasi tinggi secara sementara pada 2 hpi manakala, gen peroxidase telah diregulasi tinggi sepanjang tempoh eksperimen. Melalui asai-asai enzim, aktiviti enzim SOD dan peroxidase juga menunjukkan peningkatan progresif yang ketara berlaku selepas 8 hpi (nilai- $p < 0.05$ ). Pemerhatian ini telah disahkan lagi melalui profil protin pada proteom daun yang dijangkiti pada 4 dan 8 hpi menggunakan dua dimensi- elektroforesis gel poliakrilamida (2D-PAGE) dan analisis spektrometri jisim di mana regulasi tinggi SOD protin yang ketara pada 4 hpi telah diperhatikan (nilai- $p < 0.10$ ). Secara keseluruhannya, penemuan-penemuan dalam kajian ini membuktikan bahawa sistem antioksidan terlibat dalam tindak balas pertahanan awal betik terhadap jangkitan *E. mallotivora* yang boleh mencadangkan bahawa laluan pertahanan ini juga diaktifkan dalam interaksi serasi ini. Kajian ini memberi pemahaman baru ke dalam mekanisma molekul yang mendasari interaksi ini.

## ACKNOWLEDGEMENTS

I would like to express my deepest gratitude to my main supervisor, Assoc. Prof. Dr. Mohd Puad Abdullah for continuous encouragement, guidance and constructive comments throughout the execution of my study. Heartfelt appreciation for his patience and efforts in helping me to complete this study. My sincere appreciation also goes to my committee members, Dr. Noor Baiti Saidi and Dr. Wee Chien Yeong, who have been so helpful in providing me valuable advice and beneficial suggestions for this study. I also thank them for their precious time and countless effort in encouraging me to complete this thesis.

My gratitude also goes to my colleagues, Lina Rozano, Nazrul Hisham, Muhamad Zulhimi and Ayu Nazreena for their helps, advices, supports and importantly, for being such a great team partners. A special thanks to Pn. Noriha Mat Amin for providing me the *Erwinia mallotivora* isolate and for her helpful advices.

Lastly, my deepest appreciation to my husband, Muhamad Rozaini, and my one and only son, Mirza Rifqi for their wonderful supports, sacrifices, loves and understanding. I also thank my parents, En. Supian and Pn. Halijah, for their encouragement and motivation throughout my study.



I certify that a Thesis Examination Committee has met on 25 February 2014 to conduct the final examination of Suhaina binti Supian on her thesis entitled "Antioxidant-Mediated Defense Response of *Carica papaya* L. Var. Eksotika Against a Compatible *Erwinia mallotivora* Strain BT-MARDI" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Muhajir bin Hamid, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Mohd Arif b Syed, PhD**

Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Janna Ong binti Abdullah, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Sreeramanan Subramaniam, PhD**

Associate Professor  
School of Biological Sciences  
Universiti Sains Malaysia  
(External Examiner)



---

**ZULKARNAIN ZAINAL, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 15 April 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science (Plant Biotechnology). The members of the Supervisory Committee were as follows:

**Mohd Puad bin Abdullah, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Noor Baity Binti Saidi, PhD**

Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Member)

**Wee Chien Yeong, PhD**

Biotechnology Research Centre  
Malaysian Agricultural Research and Development Institute  
(Member)

---

**(Bujang bin Kim Huat, PhD)**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in theUniversiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: **Suhaina Supian (GS32808)**

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: \_\_\_\_\_

## TABLE OF CONTENTS

		Page
	<b>ABSTRACT</b>	i
	<b>ABSTRAK</b>	iii
	<b>ACKNOWLEDGEMENTS</b>	v
	<b>APPROVAL</b>	vi
	<b>DECLARATION</b>	viii
	<b>LIST OF TABLES</b>	xiv
	<b>LIST OF FIGURES</b>	xv
	<b>LIST OF ABBREVIATIONS</b>	xvii
<b>CHAPTER</b>		
<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
 <b>2</b>	<b>LITERATURE REVIEW</b>	<b>3</b>
	2.1 Plant and pathogen interaction	3
	2.1.1 Compatible and incompatible interaction	3
	2.1.2 Early event and components of compatible interaction	4
	2.1.3 Significance of compatible interaction study at molecular level	10
	2.2 Papaya dieback and its causal agent	11
	2.2.1 <i>Carica papaya</i> L.	11
	2.2.2 <i>Erwinia mallotivora</i>	13
	2.2.3 Status of papaya dieback disease	14
	2.3 Methods to study host-pathogen interaction at gene, protein and enzyme level	15
	2.3.1 Semi-quantitative gene expression analysis	15
	2.3.2 Protein expression analysis	16
	2.3.3 Enzyme assays	17
 <b>3</b>	<b>OPTIMIZATION OF INOCULATION PROCEDURE FOR ARTIFICIAL INFECTION OF <i>Carica papaya</i> L. VAR. EKSOTIKA WITH <i>Erwinia mallotivora</i></b>	 <b>19</b>
	3.1 Introduction	19
	3.2 Material and methods	19
	3.2.1 Papaya material	19
	3.2.2 Bacterial isolate	19

3.2.3	Germination and planting of papaya	19
3.2.4	Determination of colony-forming units of <i>E. mallotivora</i>	20
3.2.5	Pathogenicity test	20
3.2.6	Optimization of concentration of <i>E. mallotivora</i> inoculum	20
3.2.7	Optimization of leaf position	21
3.2.8	Statistical analysis	21
3.3	Results and discussion	21
3.3.1	Pathogenicity test	21
3.3.2	Effect of concentration of <i>E. mallotivora</i> inoculum on disease development	22
3.3.3	Effect of leaf position on disease development	26
3.4	Conclusion	30
<b>4</b>	<b>DETERMINATION OF THE EXPRESSION PATTERNS AND ACTIVITIES OF ANTIOXIDANT-RELATED GENES AND ENZYMES IN EARLY RESPONSE OF <i>Carica papaya</i> L. VAR. EKSOTIKA TO <i>Erwinia mallotivora</i> INFECTION</b>	<b>31</b>
4.1	Introduction	31
4.2	Materials and methods	31
4.2.1	Materials	31
4.2.2	Chemical reagents	31
4.2.3	Plant inoculation and sample collection	32
4.2.4	Gene expression analysis	32
4.2.5	Enzyme assays	34
4.2.6	Correlation analysis of gene expression and enzyme activity	37
4.2.7	Statistical analysis	37
4.3	Results and discussion	37
4.3.1	Yield and quality of RNA	37
4.3.2	Optimum parameters of RT-PCR	38
4.3.3	Expression of internal reference gene	42
4.3.4	Crude extract	42
4.3.5	Changes of SOD in papaya in response to <i>E. mallotivora</i> infection	43
4.3.6	Changes of peroxidase in papaya in response to <i>E. mallotivora</i> infection	47
4.4	Conclusion	52

<b>5</b>	<b>PROFILING AND IDENTIFICATION OF ANTIOXIDANT-RELATED PROTEIN IN EARLY RESPONSE OF <i>Carica papaya</i>L. VAR. EKSOTIKA TO <i>Erwinia mallotivora</i> INFECTION</b>	<b>53</b>
5.1	Introduction	53
5.2	Materials and methods	53
5.2.1	Materials	53
5.2.2	Chemical reagents	53
5.2.3	Protein extraction	53
5.2.4	Qualitative and quantitative quantification of total protein	54
5.2.5	2D-PAGE	54
5.2.6	Image analysis	55
5.2.7	Protein identification through MALDI-TOF/TOF and bioinformatics analysis	56
5.2.8	Correlation analysis of gene expression, protein expression and enzyme activity	56
5.3	Results and discussion	56
5.3.1	Protein extract	56
5.3.2	Protein separation through 2D-PAGE	57
5.3.3	Analysis of differentially expressed proteins in papaya in response to <i>E. mallotivora</i> infection	58
5.3.4	Identification of differentially expressed proteins through MALDI-TOF/TOF and their functional classification	61
5.3.5	Involvement of antioxidant-related protein in early response of papaya to <i>E. mallotivora</i> infection	65
5.3.6	Correlation of antioxidant-related gene, protein and enzyme in early response of papaya to <i>E. mallotivora</i> infection	66
5.4	Conclusion	67
<b>6</b>	<b>CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>68</b>

REFERENCES	70
APPENDICES	87
BIODATA OF STUDENT	113
LIST OF PUBLICATIONS	114





## LIST OF TABLES

Table		Page
4.1	Primer sequences for the targeted genes	33
4.2	RNA yield and purity of infected and control samples	38
5.1	Differentially expressed proteins in papaya leaves following <i>E. mallotivora</i> infection identified by MALDI-TOF/TOF analysis	62

## LIST OF FIGURES

Figure		Page
2.1	Enzymatic reaction of SOD	7
2.2	Enzymatic reaction of peroxidase	8
2.3	Dieback disease symptoms on papaya	11
3.1	Pathogenicity test of <i>E. mallotivora</i> isolate	22
3.2	Disease development on leaves inoculated with <i>E. mallotivora</i> inoculum at concentration of $10^3$ , $10^4$ and $10^5$ CFU/inoculation site	24
3.3	Disease development on leaves inoculated with <i>E. mallotivora</i> inoculum at concentration of $10^6$ and $10^7$ CFU/inoculation site	25
3.4	Effect of <i>E. mallotivora</i> inoculum concentration on development of papaya dieback disease	26
3.5	Effect of leaf position on development of papaya dieback disease	27
3.6	Disease development on first, second and third leaves from the top inoculated with <i>E. mallotivora</i>	28
3.7	Disease development on fourth and fifth leaves from the top inoculated with <i>E. mallotivora</i>	29
4.1	Oxidation of guaiacol by peroxidase in the presence of hydrogen peroxide to form tetraguaiacol and water	36
4.2	Total RNA (1 µg) of infected and control papaya leaves	38
4.3	Determination of optimum annealing temperature for RT-PCR reaction	39
4.4	Determination of optimum number of amplification cycle for RT-PCR reaction	41

4.5	A constant expression of <i>EIF</i> gene in infected and control samples	42
4.6	SDS-PAGE of crude extracts (20 µg) from infected and control papaya leaves	43
4.7	Expression analysis of <i>SOD</i> gene in infected and control papaya plants through RT-PCR	44
4.8	<i>SOD</i> enzyme activity in infected and control papaya plants at different time points	45
4.9	Correlation of <i>SOD</i> gene expression and its corresponding enzyme activity in papaya plants following <i>E. mallotivora</i> infection	47
4.10	Expression analysis of peroxidase gene in infected and control papaya plants through RT-PCR	48
4.11	Peroxidase enzyme activity in infected and control papaya plants at different time points	50
4.12	Correlation of peroxidase gene expression and its corresponding enzyme activity in papaya plants after <i>E. mallotivora</i> infection	51
5.1	SDS-PAGE of total proteins (20 µg) from infected and control papaya leaves	57
5.2	Separation of total proteins from papaya leaves through 2D-PAGE	59
5.3	Representative 2D-PAGE gel images of the infected and control leaf proteomes at different time points	60
5.4	Correlation of changes in <i>SOD</i> gene expression, protein expression and enzyme activity in papaya in response to <i>E. mallotivora</i> infection	67

## LIST OF ABBREVIATIONS

2D-PAGE	Two Dimensional Polyacrylamide Gel Electrophoresis
BLAST	Basic Local Alignment Search Tool
bp	base pair
BSA	Bovine serum albumin
cDNA	complementary DNA
CFU	Colony-forming unit
CHAPS	3-[(3-cholamidopropyl)dimethylammonio]-1-propanesulfonate
DNA	Deoxyribonucleic acid
DTT	Dithiothreitol
EDTA	Ethylenediamine tetraacetic acid
EIF	Eukaryotic initiation factor 4A
g	gram
ha	hectar
HCl	Hydrogen chloride
HR	Hypersensitive response
IEF	Isoelectric focusing
IPG	immobilized pH gradient
LB	Luria-Bertani
MALDI-TOF/TOF	Matrix-Assisted Laser Desorption/Ionization Time-of-Flight
MARDI	Malaysian Agricultural and Research Development Institute
min	minute
mRNA	messenger RNA

NBT	Nitroblue tetrazolium
OD	Optical density
OEE1	Oxygen evolving enhancer protein 1
PAMPs	Pathogen-associated molecular patterns
PCR	Polymerase Chain Reaction
pI	Isoelectric point
PR protein	Pathogenesis-related protein
rDNA	ribosomal DNA
RNA	Ribonucleic acid
ROS	Reactive oxygen species
rpm	revolutions per minute
rRNA	ribosomal RNA
RT-PCR	Reverse Transcription-Polymerase Chain Reaction
SDS	Sodium dodecyl sulfate
SDS-PAGE	Sodium Dodecyl Sulphate-Polyacrylamide Gel Electrophoresis
sec	second
SOD	Superoxide dismutase
TCA	Trichloroacetic acid
µg	microgram
v/v	volume per volume
w/v	weight per volume

## CHAPTER 1

### INTRODUCTION

Papaya dieback, caused by *Erwinia mallotivora* (*E. mallotivora*) bacterium, is one of the most destructive diseases to *Carica papaya* L. (*C. papaya*) in Malaysia. This disease has severely affected most papaya fields in Malaysia. First incidence of papaya dieback disease was reported in Batu Pahat, Johor in late 2003. Since then, this disease was spread aggressively throughout Malaysia including Sabah and Sarawak. In 2006, this disease had affected approximately 800 ha of papaya fields and caused the destruction of 1 million trees with total yield losses of 200,000 metric tons, which equivalent to US\$ 58 million (Maktar *et al.*, 2008). Due to major losses of papaya yield, total national export volume in Peninsular Malaysia plunged from 57,113 tons in 2003 to 10,323 tons in 2008 (Eng, 2011). This disease is difficult to control due to ineffective chemical sprays and none resistant variety to this disease identified through breeding and genetic engineering. In addition, there is limited information underlying the interaction of papaya and *E. mallotivora* at molecular level.

To date, there is no report yet on the molecular basis of papaya early response towards *E. mallotivora* infection. Basic information on molecular mechanisms involved during the compatible interaction of papaya and *E. mallotivora* at the early stage of interaction is still unknown. Lack of the information underlying this pathosystem limits the options to find an effective solution to overcome and control this disease. Thus, investigating the interaction at the molecular level would be helpful to identify key enzymes or genes that play defensive roles in papaya challenged with the pathogen. This will eventually provide better understanding of the defense mechanisms involved in this pathosystem.

To understand the defense mechanisms in plants upon pathogen infection, research should be carried out at the very early stage of the infection process. This is because plant defense system is triggered as soon as the pathogen is in contact with the host plant. During the early stage of plant and pathogen interaction, plants activate their first line of defenses including oxidative burst. Oxidative burst constitutes plant's early defense against pathogen infection by producing a rapid, transient and high amount of reactive oxygen species (ROS) in the cell. Over accumulation of ROS is, however, capable of causing oxidative damage to plant cell (Sharma *et al.*, 2012; Mendoza, 2011).

To prevent cell damage caused by oxidation, antioxidant system is induced to control and equilibrate ROS to acceptable levels (Mendoza, 2011). SOD and peroxidase are two of the major enzymes involved in this system. The involvement of these two enzymes in plant defense mechanism has been reported in many plant-pathogen systems (Viljevac *et al.*, 2009; Mandal *et al.*, 2008; Venisse *et al.*, 2001). These enzymes have been proven to be involved

in defense mechanism during early interaction of plant and pathogen even in the compatible interaction (Sgherri *et al.*, 2013; Mandal *et al.*, 2011). However, in the case of papaya and *E. mallotivora* interaction, the involvement and role of these enzymes are unknown.

Therefore, in the present study, it was hypothesized that the antioxidant system, particularly SOD and peroxidase, are involved in the early defense response of papaya to *E. mallotivora* infection. To prove this, changes of gene expression and enzymatic activity of both SOD and peroxidase in papaya plants in response to this infection were analyzed through semi-quantitative RT-PCR and enzyme assays, respectively. To further confirm the involvement of SOD and peroxidase proteins during the early interaction of papaya and this pathogen, protein expression analysis and identification were carried out through 2D-PAGE and mass spectrometry analysis. Hence, the present study was carried out with the following objectives:

- 1) to optimize the inoculation procedure for artificial infection of *Carica papaya* L. var. Eksotika with *Erwinia mallotivora*,
- 2) to determine the expression patterns and activities of the antioxidant genes and enzymes in early response of *Carica papaya* L. var. Eksotika to *Erwinia mallotivora* infection, and
- 3) to profile and identify antioxidant-related proteins in early response of *Carica papaya* L. var. Eksotika to *Erwinia mallotivora* infection.

## REFERENCES

- Able, A.J., Guest, D.I., & Sutherland, M.W. (1998). Use of a new tetrazolium-based assay to study the production of superoxide radicals by tobacco cell cultures challenged with avirulent zoospores of *Phytophthora parasitica* var *nicotianae*. *Plant Physiology*, 117, 491-499.
- Afroz, A., Ali, G.M., Mir, A., & Komatsu, S. (2011). Application of proteomics to investigate stress-induced proteins for improvement in crop protection. *Plant Cell Reports*, 30, 745-763.
- Aimé, S., Cordier, C., Alabouvette, C., & Olivain, C. (2008). Comparative analysis of pr gene expression in tomato inoculated with virulent *Fusarium oxysporum* f. sp. *lycopersici* and the biocontrol strain *F. oxysporum* fo47. *Physiological and Molecular Plant Pathology*, 73, 9-15.
- Al-Whaibi, M.H. (2011). Plant heat-shock proteins: A mini review. *Journal of King Saud University – Science*, 23, 139-150.
- Ali, A., & Alqurainy, F. (2006). Activities of antioxidants in plants under environmental stress In N. Motohashi (Ed.), *The lutein - prevention and treatment for age-related diseases* (pp. 187-256).
- Almagro, L., Ros, L.V.G., Belchi-Navarro, S., Bru, R., Barcelo, A.R., & Pedreno, M.A. (2009). Class III peroxidases in plant defence reactions. *Journal of Experimental Botany*, 60(2), 377-390.
- Alscher, R.G., Erturk, N., & Heath, L.S. (2002). Role of superoxide dismutases (SODs) in controlling oxidative stress in plants. *Journal of Experimental Biology*, 53(372), 1331-1341.
- Amey, R.C., Schleicher, T., Slinn, J., Lewis, M., Macdonald, H., Neill, S.J., & Spencer-Phillips, P.T.N. (2008). Proteomic analysis of a compatible interaction between *Pisum sativum* (pea) and the downy mildew pathogen *peronospora viciae*. *European Journal of Plant Pathology*, 122, 41-55.
- Amir, R. (2008). Towards improving methionine content in plants for enhanced nutritional quality. *Functional Plant Science and Biotechnology*, 2(1), 36-46.
- Aravind, G., Bhowmik, D., Duraivel, S., & Harish, G. (2013). Traditional and medicinal uses of *Carica papaya*. *Journal of Medicinal Plants Studies*, 1(1), 7-15.
- Asada, K., Takahashi, M., & Nagate, M. (1974). Assay and inhibitors of spinach superoxide dismutase. *Agricultural and Biological Chemistry*, 38 (2), 471-473.



- Balbi-Peña, M.I., Schwan-Estrada, K.R.F., & Stangarlin, J.R. (2012). Differential occurrence of the oxidative burst and the activity of defence-related enzymes in compatible and incompatible tomato-*Oidium neolycopersici* interactions. *Australasian Plant Pathology*, *41*, 573-586.
- Ballmoos, C.V., Wiedenmann, A., & Dimroth, P. (2009). Essentials for ATP synthesis by F<sub>1</sub>F<sub>0</sub> ATP synthases. *Annual Review of Biochemistry*, *78*, 649-672.
- Barras, F., Gijsegem, F., & Chatterjee, A.K. (1994). Extracellular enzymes and pathogenesis of soft-rot *Erwinia*. *Annual Review of Phytopathology*, *32*, 201-234.
- Beauchamp, C., & Fridovich, I. (1971). Superoxide dismutase: Improved assays and an assay applicable to acrylamide gels. *Analytical Biochemistry*, *44*, 276-287.
- Bézier, A., Lambert, B., & Baillieux, F. (2002). Study of defense-related gene expression in grapevine leaves and berries infected with *Botrytis cinerea*. *European Journal of Plant Pathology*, *108*, 111-120.
- Blokhina, O., & Fagerstedt, K.V. (2010). Reactive oxygen species and nitric oxide in plant mitochondria: Origin and redundant regulatory systems. *Physiologia Plantarum*, *138*, 447-462.
- Bohler, S., Sergeant, K., Lefèvre, I., Jolivet, Y., Hoffmann, L., Renaut, J., Dizengremel, P. & Hausman, J. (2010). Differential impact of chronic ozone exposure on expanding and fully expanded poplar leaves. *Tree Physiology*, *30*, 1415-1432.
- Boguszewska, D., & Zagdańska, B. (2012). ROS as signaling molecules and enzymes of plant response to unfavorable environmental conditions. In V. Lushchak & H. M. Semchyshyn (Eds.), *Oxidative Stress – Molecular Mechanisms and Biological Effects* (pp. 341-362).
- Bolton, M.D. (2009). Primary metabolism and plant defense—fuel for the fire. *Molecular Plant-Microbe Interactions*, *22*(5), 487-497.
- Boyd, L.A., Ridout, C., O'Sullivan, D.M., Leach, J.E., & Leung, H. (2013). Plant-pathogen interactions: disease resistance in modern agriculture. *Trends in Genetics*, *29*(4), 233-240.
- Bringans, S., Eriksen, S., Kendrick, T., Kaur, R., Gopalkrishnakone, P., & Lipscombe, R. (2008). Proteomic analysis of the venom of *Heterometrus longimanus* (asian black scorpion). *Proteomics*, *8*(5), 1081-1096.
- Brosnan, J.T. & Brosnan, M.E. (2006). The sulfur-containing amino acids: an overview. *The Journal of Nutrition*, *136*(6), 1636S-1640S.

- Buonaurio, R. (2008). Infection and plant defense responses during plant-bacterial interaction *Plant-microbe interactions* (pp. 169-197). Kerala, India: Research Signpost.
- Bürstenbinder, K., Rzewuski, G., Wirtz, M., Hell, R. & Sauter, M. (2007). The role of methionine recycling for ethylene synthesis in *Arabidopsis*. *The Plant Journal*, *49*, 238-249.
- Chan, Y.K. (1987). Backcross method in improvement of papaya (*Carica papaya* L.). *Malaysian Applied Biology*, *16*, 95-100.
- Chan, Y.K., & Baharuddin, A.G. (2010). Rejuvenating the flagging papaya industry in Malaysia: The role of MAFC. *Acta Horticulturae*, *1*, 37-40.
- Chan, Y.K., & Ong, C.A. (2003). Field performance of papaya lines selected for tolerance to ringspot virus disease. *Journal of Tropical Agriculture and Food Science*, *31*(2), 129-137.
- Chandra, S., Martin, G.B., & Low, P.S. (1996). The pto kinase mediates a signaling pathway leading to the oxidative burst in tomato. *Proceedings of the National Academy of Sciences*, *93*, 13393-13397.
- Chang, S.W., & Hwang, B.K. (2003). Effects of plant age, leaf position, inoculum density and wetness period on *Bipolaris coicis* infection in adlays of differing resistance. *Plant Disease*, *87*(7), 821-826.
- Chaudhary, B. (2013). Plant domestication and resistance to herbivory. *International Journal of Plant Genomics*, *2013*, 1-14. doi: 10.1155/2013/572784
- Cheng, Z., McConkey, B.J., & Glick, B.R. (2010). Proteomic studies of plant-bacterial interactions. *Soil Biology & Biochemistry*, *42*, 1673-1684.
- Choi, H.W., Kim, Y.J., Lee, S.C., Hong, J.K., & Hwang, B.K. (2007). Hydrogen peroxide generation by the pepper extracellular peroxidase *CaPO2* activates local and systemic cell death and defense response to bacterial pathogens. *Plant Physiology*, *145*, 890-904.
- Choquer, M., Boccara, M., & Vidal-Cros, A. (2003). A semi-quantitative RT-PCR method to readily compare expression levels within *Botrytis cinerea* multigenic families in vitro and in planta. *Current Genetics*, *43*, 303-309.
- Clarke, S.F., Guya, P.L., Burritta, D.J., & Jameson, P.E. (2002). Changes in the activities of antioxidant enzymes in response to virus infection and hormone treatment. *Physiologia Plantarum*, *114*, 157-164.
- Condemine, G., Castillo, A., Passeri, F., & Enard, C. (1999). The pectin repressor coregulates synthesis of exopolysaccharides and virulence factors in *Erwinia chrysanthemi*. *Molecular Plant-Microbe Interactions*, *12*(1), 45-52.

- Cordell, G.A. (2009). Sustainable drugs and global health care. *Química Nova*, 32(5), 1356-1364.
- Córdova-Campos, O., Adame-Álvarez, R.M., Acosta-Gallegos, J.A., & Heil, M. (2012). Domestication affected the basal and induced disease resistance in common bean (*Phaseolus vulgaris*). *European Journal of Plant Pathology*, 134, 367-379.
- Cosio, C., Vuillemin, L., Meyer, M.D., Kevers, C., Pene, C., & Dunand, C. (2009). An anionic class III peroxidase from Zucchini may regulate hypocotyl elongation through its auxin oxidase activity. *Planta*, 229, 823-836.
- Crosse, J.E., Goodman, R.N., & Shaffer, W.H. (1972). Leaf damage as a predisposing factor in the infection of apple shoots by *Erwinia amylovora*. *Phytopathology*, 62, 176-182.
- Dahal, D., Pich, A., Braun, H.P., & Wydra, K. (2010). Analysis of cell wall proteins regulated in stem of susceptible and resistant tomato species after inoculation with *Ralstonia solanacearum*: A proteomic approach. *Plant Molecular Biology*, 73(6), 643-658.
- Daudi, A., Cheng, Z., O'Brien, J.A., Mammarella, N., Khan, S., Ausubel, F.M., & Bolwel, G.P. (2012). The apoplastic oxidative burst peroxidase in *Arabidopsis* is a major component of pattern-triggered immunity. *The Plant Cell*, 24, 275-287.
- Efferth, T., & Greten, H.J. (2014). Traditional medicine with plants – present and past. *Medicinal & Aromatic Plants*, 3(3), 1-3.
- Elavarthi, S., & Martin, B. (2010). Spectrophotometric assays for antioxidant enzymes in plants. In R. Sunkar (Ed.), *Methods in molecular biology* (Vol. 639, pp. 273-281).
- Eng, L. (2011, 3 July 2011). Bacterial dieback of papaya trees. *New Sunday Tribune*.
- Eyles, S.J., & Gierasch, L.M. (2010). Nature's molecular sponges: Small heat shock proteins grow into their chaperone roles. *PNAS*, 107(7), 2727-2728.
- Faize, M., Burgos, L., Faize, L., Petri, C., Barba-Espin, G., P.Díaz-Vivancos, Clemente-Moreno, M. J., Alburquerque, N. & Hernandez, J.A. (2012). Modulation of tobacco bacterial disease resistance using cytosolic ascorbate peroxidase and Cu,Zn-superoxide dismutase. *Plant Pathology*, 61, 858-866.
- FAO. (2000). The state of food insecurity in the world (SOFI). Rome, Italy: FAO, UN.
- FAOSTAT. (2012). Available from <http://faostat.fao.org/>

- Fatibello-Filho, O., & Vieira, I.D.C. (2002). Analytical use of vegetal tissue and crude extract as enzymatic source. *Química Nova*, 25(3), 455-464.
- Fiori, M., Viridis, S., & Schiaffino, A. (2005). Phenotypic and genetic characterization of *Erwinia carotovora* ssp. *carotovora* (Jones) Bergey et al. isolates from grafted tomato in Sardinia, Italy. *Phytopathologia Mediterranea*, 44, 50-57.
- Flint-Garcia, S.A. (2013). Genetics and consequences of crop domestication. *Journal of Agricultural and Food Chemistry*, 61, 8267-8276.
- Flood, J. (2010). The importance of plant health to food security. *Food Security*, 2, 215-231.
- Flor, H.H. (1971). Current status of the gene-for-gene concept. *Annual Review of Phytopathology*, 9, 275-296.
- Foyer, C.H., Lelandais, M., & Kunert, K.J. (1994). Photooxidative stress in plants. *Physiologia Plantarum*, 92(4), 696-717.
- Foyer, C.H., & Noctor, G. (2005). Redox homeostasis and antioxidant signaling: A metabolic interface between stress perception and physiological responses. *The Plant Cell*, 17, 1866-1875.
- Freeman, W.M., Walker, S.J., & Vrana, K.E. (1999). Quantitative RT-PCR: Pitfalls and potential. *BioTechniques*, 26(1), 112-125.
- Freitas-Astúa, J., Bastianel, M., Locali-Fabris, E.C., Novelli, V.M., Silva-Pinhati, A.C., Basilio-Palmieri, A.C., Targon, M.L.P.N. & Machado, M.A. (2007). Differentially expressed stress-related genes in the compatible citrus-*Citrus leprosis* virus interaction. *Genetics and Molecular Biology*, 30(3), 980-990.
- Garavaglia, B.S., Thomas, L., Gottig, N., Zimaro, T., Garofalo, C.G., Gehring, C., & Ottado, J. (2010). Shedding light on the role of photosynthesis in pathogen colonization and host defense. *Communicative & Integrative Biology*, 3(4), 382-384.
- García-Limones, C., Hervás, A., Navas-Cortés, J.A., Jiménez-Díaz, R.M., & Tena, M. (2002). Induction of an antioxidant enzyme system and other oxidative stress markers associated with compatible and incompatible interactions between chickpea (*Cicer arietinum* L.) and *Fusarium oxysporum* f. sp. *ciceris*. *Physiological and Molecular Plant Pathology*, 61(6), 325-337.
- Garrod, A.E. (1909). Inborn errors of metabolism. London, UK: Frowde, Hodder & Stoughton.
- Giannopolitis, C.N., & Ries, S.K. (1977). Superoxide dismutases. I. Occurrence in higher plants. *Plant Physiology*, 59, 309-314.

- Gibb, K.S. (1996). Phytoplasmas associated with papaya diseases in Australia. *Plant Disease*, 80(2), 174-178.
- Gill, S.S., & Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry*, 48, 909-930.
- Glanemann, C., Loos, A., Gorret, N., Willis, L.B., O'Brien, X.M., Lessard, P.A., & Sinskey, A.J. (2003). Disparity between changes in mRNA abundance and enzyme activity in *Corynebacterium glutamicum*: Implications for DNA microarray analysis. *Applied Microbiology and Biotechnology*, 61, 61-68.
- Glawe, D.A. (1992). Thomas J. Burrill, pioneer in plant pathology. *Annual Review of Phytopathology*, 30, 17-24.
- Goto, M. (1976). *Erwinia mallotivora* sp. nov., the causal organism of bacterial leaf spot of *Mallotus japonicus* Muell. Arg. *International Journal Of Systematic Bacteriology*, 26(4), 467-473.
- Gottwald, E., Müller, O., & Polten, A. (2001). Semiquantitative reverse transcription-polymerase chain reaction with the Agilent 2100 Bioanalyzer. *Electrophoresis*, 22, 4016-4022.
- Grant, J.J., & Loake, G.J. (2000). Role of reactive oxygen intermediates and cognate redox signaling in disease resistance. *Plant Physiology*, 124, 21-29.
- Guo, R., Yu, F., Gao, Z., An, H., Cao, X., & Guo, X. (2011). *GhWRKY3*, a novel cotton (*Gossypium hirsutum* L.) *WRKY* gene, is involved in diverse stress responses. *Molecular Biology Reports*, 38, 49-58.
- Guthrie, J.N., Walsh, K.B., P.T.Scott, & Rasmussen, T.S. (2001). The phytopathology of Australian papaya dieback: A proposed role for the phytoplasma. *Physiological and Molecular Plant Pathology*, 58, 23-30.
- Hakmaoui, A., Pérez-Bueno, M.L., García-Fontana, B., Camejo, D., Jiménez, A., Sevilla, F., & Barón, M. (2012). Analysis of the antioxidant response of *Nicotiana benthamiana* to infection with two strains of pepper mild mottle virus. *Journal of Experimental Biology*, 63(15), 5487-5496.
- Hanifei, M., Dehghani, H., & Choukan, R. (2013). The role of antioxidant enzymes and phenolic compounds in disease resistance to *Fusarium oxysporum* f. sp. *melonis* race 1.2. *International Journal of Agronomy and Plant Production*, 4(8), 1985-1996.
- Hell, R., & Kruse, C. (2007). Sulfur in biotic interactions of plants. In M. J. Hawkesford & L. J. D. Kok (Eds.), *Sulfur in plants an ecological perspective* (Vol. 6, pp. 197-224).



- Herrera-Estrella, A., & Chet, I. (1999). Chitinases in biological control. In P. Jolles & R. A. A. Muzzarelli (Eds.), *Chitin and chitinases* (Vol. 87, pp. 171-184).
- Hesse, H., Kreft, O., Maimann, S., Zeh, M., & Hoefgen, R. (2004). Current understanding of the regulation of methionine biosynthesis in plants. *Journal of Experimental Botany*, 55(404), 1799-1808.
- Heyno, E., Mary, V., Schopfer, P., & Krieger-Liszky, A. (2011). Oxygen activation at the plasma membrane: Relation between superoxide and hydroxyl radical production by isolated membranes. *Planta*, 234(1), 35-45.
- Hilaire, E., Young, S.A., Willard, L.H., McGee, J.D., Sweat, T., Chittoor, J.M., Guikema, J.A. & Leach, J.E. (2001). Vascular defense responses in rice: Peroxidase accumulation in xylem parenchyma cells and xylem wall thickening. *Molecular Plant-Microbe Interaction*, 14(12), 1411-1419.
- Hiraga, S., Sasaki, K., Ito, H., Ohashi, Y., & Matsui, H. (2001). A large family of class III plant peroxidases. *Plant and Cell Physiology*, 42(5), 462-468.
- Hossain, M.M., Shibata, S., Aizawa, S.-I., & Tsuyumu, S. (2005). Motility is an important determinant for pathogenesis of *Erwinia carotovora* subsp. *carotovora*. *Physiological and Molecular Plant Pathology*, 66, 134-143.
- <http://primer3.sourceforge.net>
- <http://www.fao.org>
- <http://www.mardi.gov.my>
- <http://www.moa.gov.my>
- Hyun, M.W., Yun, Y.H., Kim, J.Y., & Kim, S.H. (2011). Fungal and plant phenylalanine ammonia-lyase. *Mycobiology*, 39(4), 257-265.
- Imbeaud, S., Gaudens, E., Boulanger, V., Barlet, X., Zaborski, P., Eveno, E., Odilo Mueller, O., Schroeder, A. & Auffray, C. (2005). Towards standardization of RNA quality assessment using user-independent classifiers of microcapillary electrophoresis traces. *Nucleic Acids Research*, 33(6), 12.
- Jiang, M., Miao, L., & He, C. (2012). Overexpression of an oil radish superoxide dismutase gene in broccoli confers resistance to downy mildew. *Plant Molecular Biology Reporter*, 30, 966-972.
- Jones, J.D.G., & Dangl, J.L. (2006). The plant immune system. *Nature*, 444, 323-329.

- Kaur, P., Jost, R., Sivasithamparam, K., & Barbetti, M.J. (2011). Proteome analysis of the *Albugo candida*–*Brassica juncea* pathosystem reveals that the timing of the expression of defence-related genes is a crucial determinant of pathogenesis. *Journal of Experimental Botany*, 62(3), 1285-1298.
- Kim, S.G., Wang, Y., Lee, K.H., Park, Z.-Y., Park, J., Wu, J., Kwon, S.J., Lee, Y.H., Agrawal, G.K., Rakwal, R., Kim, S.T. & Kang, K.Y. (2013). In-depth insight into in vivo apoplastic secretome of rice-*Magnaporthe oryzae* interaction. *Journal of Proteomics*, 78, 58-71.
- Knox, J.P. (1995). Developmentally regulated proteoglycans and glycoproteins of the plant cell surface. *The FASEB Journal* 9, 1004-1012.
- Krah, A., Wessel, R., & Pleissner, K.P. (2004). Assessment of protein spot components applying correspondence analysis for peptide mass fingerprint data. *Proteomics*, 10, 2982-2986.
- Krishnan, H.B., & Natarajan, S.S. (2009). A rapid method for depletion of rubisco from soybean (*Glycine max*) leaf for proteomic analysis of lower abundance proteins. *Phytochemistry*, 70(17-18), 1958-1964.
- Kundu, S., Chakraborty, D., Kundu, A., & Pal, A. (2013). Proteomics approach combined with biochemical attributes to elucidate compatible and incompatible plant-virus interactions between *Vigna mungo* and mungbean yellow mosaic india virus. *Proteome Science*, 11(15), 1-14.
- Kuvalekar, A.A., & Gandhe, K.R. (2010). Hydrogen peroxide generation and lignification by peroxidases from *Acacia eburnea* infected with *Ravenelia esculenta*. *Plant, Soil and Environment*, 56(9), 419-428.
- Laemmli, U.K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227, 680 - 685.
- Lamb, C., & Dixon, R.A. (1997). The oxidative burst in plant disease resistance. *Annual Review of Plant Physiology and Plant Molecular Biology*, 48, 251–275.
- Lambrechts, L. (2010). Dissecting the genetic architecture of host–pathogen specificity. *PLoS Pathogens*, 6(8).
- Langenkämper, G., Manac'h, N., Broin, M., Cuiñé, S., Becuwe, N., Kuntz, M., & Rey, P. (2001). Accumulation of plastid lipid-associated proteins (fibrillin/CDSP34) upon oxidative stress, ageing and biotic stress in Solanaceae and in response to drought in other species. *Journal of Experimental Botany*, 52(360), 1545-1554.
- Lapin, D., & Ackerveken, G.V.D. (2013). Susceptibility to plant disease: More than a failure of host immunity. *Trends in Plant Science*, 18(10), 546-554.

- Larbi, N.B., & Jefferies, C. (2009). 2D-DIGE: Comparative proteomics of cellular signalling pathways. In C. E. McCoy & L. A. J. O'Neill (Eds.), *Methods in molecular biology, toll-like receptors* (Vol. 517, pp. 105-132).
- Lin, C. C., & Kao, C. H. (2001). Abscisic acid induced changes in cell wall peroxidase activity and hydrogen peroxide level in roots of rice seedlings. *Plant Science*, 160, 323-329.
- Lin, M., Kuo, T., & Lin, C. (1998). Molecular cloning of a cDNA encoding copper/zinc superoxide dismutase from papaya fruit and overexpression in *Escherichia coli*. *Journal of Agricultural and Food Chemistry*, 46, 344-348.
- Liu, B., White, D.T., Walsh, K.B., & Scott, P.T. (1996). Detection of phytoplasmas in dieback, yellow crinkle, and mosaic diseases of papaya using polymerase chain reaction techniques. *Australian Journal of Agricultural Research*, 47(3), 387-394
- Lodish, H., Berk, A., & Zipursky, S.L. (2000). Section 10.1 bacterial gene control: The Jacob-Monod model. In W. H. Freeman (Ed.), *Molecular cell biology. 4th edition*. New York.
- Mahmood, T., Jan, A., Kakishima, M., & Komatsu, S. (2006). Proteomic analysis of bacterial-blight defense-responsive proteins in rice leaf blades. *Proteomics*, 6(22), 6053-6065.
- Maimann, S., Hoefgen, R., & Hesse, H. (2001). Enhanced cystathionine  $\beta$ -lyase activity in transgenic potato plants does not force metabolite flow towards methionine. *Planta*, 214(2), 163-170.
- Maimann, S., Wagner, C., Kreft, O., Zeh, M., Willmitzer, L., Höfgen, R., & Hesse, H. (2000). Transgenic potato plants reveal the indispensable role of cystathionine beta-lyase in plant growth and development. *The Plant Journal*, 23(6), 747-758.
- Maimbo, M., Ohnishi, K., Hikichi, Y., Yoshioka, H., & A. Kiba. (2007). Induction of a small heat shock protein and its functional roles in nicotiana plants in the defense response against *Ralstonia solanacearum*. *Plant Physiology*, 145(4), 1588-1599.
- Maktar, N.H., Kamis, S., Yusof, F.Z.M., & Hussain, N.H. (2008). *Erwinia papayae* causing papaya dieback in malaysia. *Plant Pathology*, 57(4), 774.
- Maldonado, A.M., Echevarría-Zomeño, S., Jean-Baptiste, S., Hernández, M., & Jorrín-Novo, J.V. (2008). Evaluation of three different protocols of protein extraction for *Arabidopsis thaliana* leaf proteome analysis by two-dimensional electrophoresis. *Journal of Proteomics*, 71(4), 461-472.



- Mandal, S., Das, R.K., & Mishra, S. (2011). Differential occurrence of oxidative burst and antioxidative mechanism in compatible and incompatible interactions of *Solanum lycopersicum* and *Ralstonia solanacearum*. *Plant Physiology and Biochemistry*, 49, 117-123.
- Mandal, S., Mitra, A., & Mallick, N. (2008). Biochemical characterization of oxidative burst during interaction between *Solanum lycopersicum* and *Fusarium oxysporum* f. sp. *lycopersici*. *Physiological and Molecular Plant Pathology*, 72, 56-61.
- Mann, M., & Jensen, O.N. (2003). Proteomic analysis of post-translational modifications. *Nature Biotechnology*, 21, 255 - 261.
- Marone, M., Mozzett, S., Ritis, D.D., Pierelli, L., & Scambia, G. (2001). Semiquantitative RT-PCR analysis to assess the expression levels of multiple transcripts from the same sample. *Biological Procedures Online*, 3(1), 19-25.
- Martinelli, F., Uratsu, S.L., Albrecht, U., Reagan, R.L., Phu, M.L., Britton, M., Buffalo, V., Fass, J., Leicht, E., Zhao, W., Lin, D., Souza, R.D., Davis, C.E., Bowman, K.D. & Dandekar, A.D. (2012). Transcriptome profiling of citrus fruit response to Huanglongbing disease. *PLoS ONE*, 7(5).
- Mat Amin N, Bunawan H, R., A.R., & Jaganath IB. (2011). *Erwinia mallotivora* sp., a new pathogen of papaya (*Carica papaya*) in peninsular Malaysia. *International Journal of Molecular Sciences*, 12, 39-45.
- Matsumura, H., Reich, S., Ito, A., Saitoh, H., Kamoun, S., Winter, P., Kahl, G., Reuter, M., Kruger, D.H. & Terauchi, R. (2003). Gene expression analysis of plant host-pathogen interactions by superSAGE. *PNAS*, 100(26), 15718-15723
- Matuschke, I. (2009). Rapid Urbanization and Food Security: Using Food Density Maps to Identify Future Food Security Hotspots. In *International Association of Agricultural Economists Conference*, Beijing, China.
- McCord, J. M., & Fridovich, I. (1969). Superoxide dismutase, an enzymic function for erythrocyte hemoglobin. *The Journal of Biological Chemistry*, 244(22), 6049-6055.
- McKenzie, C.L., Shatters, R.G., Doostdar, H., Lee, S.D., Inbar, M., & Mayer, R.T. (2002). Effect of *Gemini* virus infection and *Emisia* infestation on accumulation of pathogenesis-related proteins in tomato. *Archives of Insect Biochemistry and Physiology*, 49, 203-214.
- Mendoza, M. (2011). Oxidative burst in plant-pathogen interaction. *Biocología Vegetal*, 11(2), 67-65.
- Milli, A., Cecconi, D., Bortesi, L., Persi, A., Rinalducci, S., Zamboni, A., Gianni Zoccatelli, G., Lovato, A., Zolla, L. & Polverari, A. (2012). Proteomic

analysis of the compatible interaction between *Vitis vinifera* and *Plasmopara viticola*. *Journal Of Proteomics*, 75, 1284-1302.

- Minibayeva, F., Kolesnikov, O., Chasov, A., Beckett, R.P., Lühje, S., Vylegzhaniina, N., Buck, F. & Böttger, A. (2009). Wound-induced apoplastic peroxidase activities: Their roles in the production and detoxification of reactive oxygen species. *Plant, Cell & Environment*, 32(5), 497-508.
- Mohr, H., & Schopfer, P. (1995). *Plant Physiology*. Springer Science & Business Media.
- Mordecai, E.A. (2011). Pathogen impacts on plant communities: Unifying theory, concepts, and empirical work. *Ecological Monographs*, 81(3), 429–441.
- Moussaoui, A.E., Nijs, M., Paul, C., Wintjens, R., Vincentelli, J., Azarkan, M., & Looze, Y. (2001). Revisiting the enzymes stored in the laticifers of *Carica papaya* in the context of their possible participation in the plant defence mechanism. *Cellular and Molecular Life Sciences*, 58, 556-570.
- Mukherjee, A.K., Lev, S., Gepstein, S., & Horwitz, B.A. (2009). A compatible interaction of *Alternaria brassicicola* with *Arabidopsis thaliana* ecotype DiG: Evidence for a specific transcriptional signature. *BMC Plant Biology*, 9(31), 1-11.
- Muthamilarasan, M., & Prasad, M. (2013). Plant innate immunity: An updated insight into defense mechanism. *Journal of Biosciences*, 38(2), 433–449.
- Nakasone, H.Y., & Paull, R.E. (1998). *Tropical fruits*: CAB International Wallingford, UK.
- Nanda, A.K., Andrio, E., Marino, D., Pauly, N., & Dunand, C. (2010). Reactive oxygen species during plant-microorganism early interactions. *Journal of Integrative Plant Biology*, 52(2), 195-204.
- Oh, C.-S., & Beer, S.V. (2005). Molecular genetics of *Erwinia amylovora* involved in the development of fire blight. *FEMS Microbiology Letters*, 253, 185-192.
- Oliveira, J.T.A., Andrade, N.C., Martins-Miranda, A.S., Soares, A.A., Gondim, D.M.F., Araújo-Filho, J.H., Freire-Filho, F.R. & Vasconcelos, I.M. (2012). Differential expression of antioxidant enzymes and PR-proteins in compatible and incompatible interactions of cowpea (*Vigna unguiculata*) and the root-knot nematode *Meloidogyne incognita*. *Plant Physiology and Biochemistry*, 51, 145-152.
- Ong, S.-E., & Mann, M. (2005). Mass spectrometry-based proteomics turns quantitative. *Nature Chemical Biology*, 1, 252 - 262.

- Padliya, N.D., & Cooper, B. (2006). Mass spectrometry-based proteomics for the detection of plant pathogens. *Proteomics*, 6, 4069-4075.
- Padliya, N.D., Garrett, W.M., Campbell, K.B., Tabb, D.L., & Cooper, B. (2007). Tandem mass spectrometry for the detection of plant pathogenic fungi and the effects of database composition on protein inferences. *Proteomics*, 7(21), 3932-3942.
- Park, J.M., & Paek, K.H. (2007). Recognition and response in plant-pathogen interactions. *Journal of Plant Biology*, 50(2), 132-138.
- Passardi, F., Penel, C., & Dunand, C. (2004). Performing the paradoxical: How plant peroxidases modify the cell wall. *Trends in Plant Science*, 9(11), 534-540.
- Pérombelon, M.C.M. (2002). Potato diseases caused by soft rot *Erwinias*: An overview of pathogenesis. *Plant Pathology*, 51, 1-12.
- Posch, A., Franz, T., Hartwig, S., Knebel, B., Al-Hasani, H., Passlack, W., Kunz, N., Hinze, Y., Li, X., Kotzka, J. & Lehr, S. (2013). 2D-ToGo workflow: Increasing feasibility and reproducibility of 2-dimensional gel electrophoresis. *Archives of Physiology and Biochemistry*, 119(3), 108-113.
- Premanandh, J. (2011). Factors affecting food security and contribution of modern technologies in food sustainability. *Journal of the Science of Food and Agriculture*, 91(15), 2707-2714.
- Que, Y., Xu, L., Lin, J., Ruan, M., Zhang, M., & Chen, R. (2011). Differential protein expression in sugarcane during sugarcane-*Sporisorium scitamineum* interaction revealed by 2-DE and MALDI-TOF/TOF-MS. *Comparative and Functional Genomics*, 10.
- Quiroga, M.N., Guerrero, C., Botella, M.A., Barcelo, A., Amaya, I., Medina, M. I., Alonso, F.J., Forchetti, S.M.D., Tigier, H. & Valpuesta, V. (2000). A tomato peroxidase involved in the synthesis of lignin and suberin. *Plant Physiology*, 122, 1119-1127.
- Rabilloud, T. (1998). Use of thiourea to increase the solubility of membrane proteins in two-dimensional electrophoresis. *Electrophoresis*, 19(5), 758-760.
- Rabilloud, T.h., Adessi, C., Giraudel, A., & Lunard, J. (2009). Improvement of the solubilization of proteins in two-dimensional electrophoresis with immobilized pH gradients. *Electrophoresis*, 18(3-4), 307-316.
- Ralph, J., Bunzel, M., Marita, J.M., Hatfield, R.D., Lu, F., Kim, H., Schatz, P.F., Grabber, J.H. & Steinhardt, H. (2004). Peroxidase-dependent cross-linking reactions of p-hydroxycinnamates in plant cell walls. *Phytochemistry Reviews*, 3, 79-96.

- Ramakrishna, H., Murthy, S.S., Divya, R., MamathaRani, D.R., & Murthy, G.P. (2012). Hydroxy radical and DPPH scavenging activity of crude protein extract of *Leucas linifolia*: A folk medicinal plant *Asian Journal of Plant Science and Research*, 2(1), 30-35.
- Ramírez, V., Agorio, A., Coego, A., á-Andrade, J.G., Hernández, M.J., Balaguer, B., Ouwerkerk, P.B.F., Zarra, I. & Vera, P. (2011). MYB46 modulates disease susceptibility to *Botrytis cinerea* in *Arabidopsis*. *Plant Physiology*, 155, 1920-935.
- Rao, G.P., Chaturvedi, Y., Priya, M., & Mall, S. (2011). Association of a 16SRII group phytoplasma with dieback disease of papaya in India. *Bulletin of Insectology*, 64, 105-106.
- Redzuan, R.A., Bakar, N.A., Rozano, L., Badrun, R., Amin, N.M., & Raih, M.F.M. (2014). Draft genome sequence of *Erwinia mallotivora* BT-mardi, causative agent of papaya dieback disease. *Genome Announcements*, 2(3).
- Reimers, P.J., Guo, A., & Leach, J.E. (1992). Increased activity of a cationic peroxidase associated with an incompatible interaction between *Xanthomonas oryzae* pv *oryzae* and rice (*Oryza sativa*). *Plant Physiology*, 99, 1044-1050.
- Restrepo, S., Myers, K.L., Pozo, O.D., Martin, G.B., Hart, A.L., Buell, C.R., Fry, W.E. & Smart, C.D. (2005). Gene profiling of a compatible interaction between *Phytophthora infestans* and *Solanum tuberosum* suggests a role for carbonic anhydrase. *Molecular Plant-Microbe Interactions*, 18(9), 913-922.
- Reuveni, M., Tuzun, S., Cole, J.S., Siegel, M.R., & Kuc, J. (1986). The effects of plant age and leaf position on the susceptibility of tobacco to blue mold caused by *Peronospora tabacina*. *Phytopathology*, 76, 455-458.
- Reymond, J., Fluxa, V.S., & Maillard, N.I. (2009). Enzyme assays. *Chemical Communications*, 34-46.
- Río, L.A.D., Sandalio, L.M., Corpas, F.J., Palma, J.M., & Barroso, J.B. (2006). Reactive oxygen species and reactive nitrogen species in peroxisomes. Production, scavenging, and role in cell signaling. *Plant Physiology*, 141, 330-335.
- Riaz, M., Bockus, W.W., & Davis, M.A. (1991). Effects of wheat genotype, time after inoculation, and leaf age on conidia production by *Drechslera tritici-repentis*. *Phytopathology*, 81(10), 1298-1302.
- Rodrigues, S.P., Ventura, J.A., Aguilar, C., Nakayasu, E.S., Almeida, I.C., Fernandes, P.M.B., & Zingali, R.B. (2011). Proteomic analysis of papaya (*Carica papaya* L.) displaying typical sticky disease symptoms. *Proteomics*, 11, 2592-2602.

- Rodrigues, S.P., Ventura, J.A., Zingalia, R.B., & Fernandesb, P.M.B. (2009). Evaluation of sample preparation methods for the analysis of papaya leaf proteins through two-dimensional gel electrophoresis. *Phytochemical Analysis*, 20, 456–464.
- Rodriguez, M.A.D., Brommonschenkel, S.H., Matsuoka, K., & Mizubuti, E.S.G. (2006). Components of resistance to early blight in four potato cultivars: Effect of leaf position. *Journal of Phytopathology*, 154, 230-235.
- Roje, S. (2006). S-Adenosyl-L-methionine: Beyond the universal methyl group donor. *Phytochemistry*, 67, 1686-1698.
- Roldan-Arjona, T., & Ariza, R.R. (2009). Repair and tolerance of oxidative DNA damage in plants. *Mutation Research*, 681, 169-179.
- Rosenthal, J.P., & Dirzo, R. (1997). Effects of life history, domestication and agronomic selection on plant defence against insects: Evidence from maizes and wild relatives. *Evolutionary Ecology*, 11, 337-355.
- Ruibal, C., Castro, A., Carballo, V., Szabados, L., & Vidal, S. (2013). Recovery from heat, salt and osmotic stress in *Physcomitrella patens* requires a functional small heat shock protein PpSHSP16.4. *BMC Plant Biology*, 13(174), 18.
- Ruz, L., Moragrega, C., & Montesinos, E. (2008). Evaluation of four whole-plant inoculation methods to analyze the pathogenicity of *Erwinia amylovora* under quarantine conditions. *International Microbiology*, 11, 111-119.
- Sarowar, S., Zhao, Y., Soria-Guerra, R.E., Ali, S., Zheng, D., Wang, D., & Korban, S.S. (2011). Expression profiles of differentially regulated genes during the early stages of apple flower infection with *Erwinia amylovora*. *Journal of Experimental Botany*, 62 (14), 4851-61.
- Sgherri, C., Ranieri, A., & Quartacci, M.F. (2013). Antioxidative responses in *Vitis vinifera* infected by grapevine fanleaf virus. *Journal of Plant Physiology*, 170, 121– 128.
- Sharma, P., Jha, A.B., Dubey, R.S., & Pessarakli, M. (2012). Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *Journal of Botany*, 2012, 26.
- Sharma, P., Jha, A. B., Dubey, R. S., & Pessarakli, M. (2014). Reactive oxygen species generation, hazards, and defense mechanisms in plants under environmental (abiotic and biotic) stress conditions. In M. Pessarakli (Ed.), *Handbook of Plant and Crop Physiology, Third Edition* (pp. 509–548).
- Shi, J., Pagliaccia, D., Morgan, R.N., Qiao, Y., Pan, S., Vidalakis, G., & Ma, W. (2014). Novel diagnosis for citrus stubborn disease by detection of a *Spiroplasma citri*-secreted protein. *Phytopathology*, 104(2), 188-195.



- Singh, D.K., Maximova, S.N., Jensen, P.J., Lehman, B.L., Ngugi, H.K., & McNellis, T.W. (2010). Fibrillin4 is required for plastoglobule development and stress resistance in apple and arabidopsis. *Plant Physiology*, 154(3), 1281-1293.
- Singh, N.K., Kumar, K.R.R., Kumar, D., Shukla, P., & Kirti, P.B. (2013). Characterization of a pathogen induced thaumatin-like protein gene *AdTLP* from *Arachis diogeni*, a wild peanut. *PLoS ONE*, 8(12), 1-18.
- Song, J., & Bent, A.F. (2014). Microbial pathogens trigger host DNA double-strand breaks whose abundance is reduced by plant defense responses. *PLoS Pathogens*, 10(4), 1-11.
- Sreedevi, S., Remani, K.N., & Benjamin, S. (2013). Biotic stress induced biochemical and isozyme variations in ginger and tomato by *Ralstonia solanacearum*. *American Journal of Plant Sciences*, 4, 1601-1610.
- Strahler, J.R., & Hanash, S.M. (1991). Immobilized pH gradients: Analytical and preparative use. *METHODS: A Companion to Methods in Enzymology*, 3(2), 109-114.
- Subramanian, B., Bansal, V.K., & Kav, A.V. (2005). Proteome-level investigation of *Brassica carinata*-derived resistance to *Leptosphaeria maculans*. *Journal of Agricultural and Food Chemistry*, 53, 313-324
- Sudhakar, N., & RM., T.V. (2014). Potential medicinal properties of *Carica papaya* linn. - a mini review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(2), 1-4.
- Swarupa, V., Ravishankar, K.V., & Rekha, A. (2014). Plant defense response against *Fusarium oxysporum* and strategies to develop tolerant genotypes in banana. *Planta*, 239, 735-751.
- Thakker, J.N., Patel, S., & Dhandhukia, P.C. (2013). Induction of defense-related enzymes in banana plants: Effect of live and dead pathogenic strain of *Fusarium oxysporum* f. sp. *cubense*. *ISRN Biotechnology*, 6.
- Ukeda, H., Kawana, D., Maeda, S., & Sawamura, M. (1999). Spectrophotometric assay for superoxide dismutase based on the reduction of highly water-soluble tetrazolium salts by xanthine-xanthine oxidase. *Bioscience, Biotechnology and Biochemistry*, 63(3), 485-488.
- Venisse, J.S., Gullner, G., & Brisset, M.N. (2001). Evidence for the involvement of an oxidative stress in the initiation of infection of pear by *Erwinia amylovora*. *Plant Physiology*, 125, 2164-2172.
- Verdonck, L., Mergaert, J., Rijckaert, C., Swings, J., Kersters, K., & Ley, J.D. (1987). Genus *Erwinia*: Numerical analysis of phenotypic features. *International Journal Of Systematic Bacteriology*, 37(1), 4-18.

- Viljevac, M., Dugalic, K., Stolfa, I., Dermic, E., Cvjetkovic, B., Sudar, R., Kovacevic, J., Cesar, V., Lepedus, H. & Jurkovic, Z. (2009). Biochemical basis of apple leaf resistance to *Erwinia amylovora* infection. *Food Technology Biotechnology*, 47(3), 281–287.
- Vogel, C., & Marcotte, E.M. (2013). Insights into the regulation of protein abundance from proteomic and transcriptomic analyses. *Nature Reviews Genetics*, 13(4), 227-232.
- Voloudakis, A.E., Marmey, P., Delannoy, E., Jalloul, A., Martinez, C., & Nicole, M. (2006). Molecular cloning and characterization of *Gossypium hirsutum* superoxide dismutase genes during cotton–*Xanthomonas campestris* pv. *malvacearum* interaction. *Physiological and Molecular Plant Pathology*, 68, 119-127.
- Vrancken, K., Holtappels, M., Schoofs, H., Deckers, T., & Valcke, R. (2013). Pathogenicity and infection strategies of the fire blight pathogen *Erwinia amylovora* in Rosaceae: State of the art. *Microbiology*, 159, 823-832.
- Walker, S.J., Worst, T.J., & Vrana, K.E. (2003). Semiquantitative real-time PCR for analysis of mRNA levels. In J. Q. Wang (Ed.), *Methods in molecular medicine* (Vol. 79, pp. 211-227).
- Walsh, K.B., Guthrie, J.N., & White, D.T. (2006). Control of phytoplasma diseases of papaya in Australia using netting. *Australasian Plant Pathology*, 35, 49-54.
- Wang, J., Liu, K., Li, D., Zhang, Y., Zhao, Q., He, Y., & Gong, Z. (2013). A novel peroxidase *CanPOD* gene of pepper is involved in defense responses to *Phytophthora capsici* infection as well as abiotic stress tolerance *International Journal of Molecular Sciences*, 14, 3158-3177.
- Wang, X., Liu, W., Chen, X., Tang, C., Dong, Y., Ma, J., Huang, X., Wei, G., Han, Q., Huang, L. & Kang, Z. (2010). Differential gene expression in incompatible interaction between wheat and stripe rust fungus revealed by cDNA-AFLP and comparison to compatible interaction. *BMC Plant Biology*, 10(9), 1-15.
- Wee, C. Y., Hanam, H.M., Adly, M.Z.M.W., & Khairun, H.N. (2014). Expression of defense-related genes in papaya seedling infected with *Erwinia mallotivora* using real-time PCR. *Journal of Tropical Agriculture and Food Science*, 42 (1), 73-82.
- Welinder, K.G., Justesen, A.F., Kjærsgård, I.V.H., Jensen, R.B., Rasmussen, S.K., Jespersen, H.M., & Duroux, L. (2002). Structural diversity and transcription of class III peroxidases from *Arabidopsis thaliana*. *European Journal of Biochemistry*, 269(24), 6063-6081.
- Wit, P.J.G.M.d. (2007). How plants recognize pathogens and defend themselves. *Cellular and Molecular Life Sciences*, 64, 2726 – 2732.

- Wojtaszek P. (1997). Oxidative burst: An early plant response to pathogen infection. *Biochemical Journal*, 322, 681-692.
- Wurms, K.V., Chee, A.A., Reglinski, T., & Taylor, J.T. (2007). Suitability of phenylalanine ammonia lyase and chitinase activities as biochemical markers of soft rot resistance in *Actinidia chinensis* kiwifruit. *New Zealand Plant Protection*, 60, 228-234.
- Xi, J., Wang, X., Li, S., Zhou, X., Yue, L., Fan, J., & Hao, D. (2006). Polyethylene glycol fractionation improved detection of low-abundant proteins by two-dimensional electrophoresis analysis of plant proteome. *Phytochemistry*, 67(21), 2341-2348.
- Xiang, X., Ning, S., Jiang, X., Gong, X., Zhu, R., & Wei, D. (2010). Protein extraction methods for two-dimensional electrophoresis from *Baphicacanthus cusia* (nees) bremek leaves - a medicinal plant with high contents of interfering compounds. *Agricultural Sciences in China*, 9(10), 1530-1537.
- Yang, F., Jensen, J.D., Svensson, B., Jørgensen, H.J.L., Collinge, D.B., & Finnie, C. (2010). Analysis of early events in the interaction between *Fusarium graminearum* and the susceptible barley (*Hordeum vulgare*) cultivar scarlett. *Proteomics*, 10, 3748-3375.
- Yi, X., McChargue, M., Laborde, S., Frankel, L.K., & Bricker, T.M. (2005). The manganese-stabilizing protein is required for photosystem II assembly/stability and photoautotrophy in higher plants. *The Journal of Biological Chemistry*, 280(16), 16170 -16174.
- Zhu, X., Li, X., Chen, W., Chen, J., Lu, W., Chen, L., & Fu, D. (2012). Evaluation of new reference genes in papaya for accurate transcript normalization under different experimental conditions. *PLoS ONE*, 7(8), 14.
- Zipfel, C. (2008). Pattern-recognition receptors in plant innate immunity. *Current Opinion in Immunology*, 20, 10-16.



## LIST OF PUBLICATIONS

### Poster and Proceeding

Suhaina Supian, Noor Baiti Saidi, Wee Chien Yeong & Mohd Puad Abdullah (2014). Potential antioxidant enzymes to combat papaya dieback disease. Proceeding of the BioJohor Biotechnology Conference, 25-27 August 2014, Puteri Pacific Hotel, Johor Bahru, Johor.

Suhaina Supian, Noor Baiti Saidi, Wee Chien Yeong & Mohd Puad Abdullah (2014). Deciphering the defense mechanism in papaya for combating dieback disease. Pameran Sains dan Teknologi MARDI Edisi ke-9, 4-5 November 2014. Silver medal.

Suhaina Supian, Noor Baiti Saidi, Wee Chien Yeong & Mohd Puad Abdullah (2014). Deciphering the defense mechanism in papaya for combating dieback disease. Malaysian Agricultural Innovation Challenge 2014. 6-8 November 2014.

### Biotech Colloquium

Suhaina Supian, Noor Baiti Saidi, Wee Chien Yeong & Mohd Puad Abdullah (2012). Identification and characterization of proteins related to papaya dieback disease. 18<sup>th</sup> Biotech Colloquium, 7 & 21 December 2012, Universiti Putra Malaysia. Pp 32.

Suhaina Supian, Noor Baiti Saidi, Wee Chien Yeong & Mohd Puad Abdullah (2013). Early molecular responses of papaya plants to *Erwinia mallotivora* infection. 20<sup>th</sup> Biotech Colloquium, 6-7 November 2013, Universiti Putra Malaysia. Pp 3.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : \_\_\_\_\_

TITLE OF THESIS / PROJECT REPORT :

Antioxidant-mediated defense response of *Carica papaya* L. var Eksotika against a compatible *Erwinia mallotivora* strain BT-MARDI

NAME OF STUDENT : Suhaina Supian

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

\*Please tick (✓)

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from \_\_\_\_\_ until \_\_\_\_\_  
(date) (date)

Approved by:

\_\_\_\_\_  
(Signature of Student)  
New IC No/ Passport No.: 851105-14-5376

\_\_\_\_\_  
(Signature of Chairman of Supervisory Committee)  
Name:

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted. ]