

UNIVERSITI PUTRA MALAYSIA

PHYTOTOXICITY OF OIL PALM RESIDUES USED AS MULCH FOR LETTUCE, TOMATO, CUCUMBER AND AMARANTH SEEDLINGS

MOHD ZAKWAN BIN ZAMRI

FP 2016 88



PHYTOTOXICITY OF OIL PALM RESIDUES USED AS MULCH FOR LETTUCE, TOMATO, CUCUMBER AND AMARANTH SEEDLINGS

By

MOHD ZAKWAN BIN ZAMRI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

April 2016

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PHYTOTOXICITY OF OIL PALM RESIDUES USED AS MULCH FOR LETTUCE, TOMATO, CUCUMBER AND AMARANTH SEEDLINGS

By

MOHD ZAKWAN BIN ZAMRI

April 2016

Chair: Associate Professor Siti Hajar Ahmad, PhD Faculty: Agriculture

Wastes of the Malaysian oil palm industry included empty fruit bunch (EFB), palm oil mill effluent (POME), oil palm frond (OPF), chopped trunk (OPT), palm pressed mesocarp fibre (PPMF) and palm kernel shell (PKS), amounting to173.02 million tonnes in 2014. Utilization of these residues is important to maximize uses of resources and overcome environmental pollution. Raw EFB and PPMF have been widely used in oil palm plantation, vegetable farming and landscape industry, as mulching and organic fertilizer. OPT and OPF are produced from old palm and left to rot in the field. Fresh plant wastes could release phytotoxic compounds, affecting growth of other plants. Therefore, the objective of the study were (1) to determine the phytotoxic effect of oil palm wastes, EFB, PPMF, OPF and OPT, using seedling bioassay and seed germination test on four vegetable: lettuce (Lactuca sativa), tomato (Solanum lycopersicum), cucumber (*Cucumis sativus*) and green amaranth (*Amaranthus viridis*), and (2) to identify phenolic acid phytotoxic compounds in extracts of selected raw oil palm wastes using thin layer chromatography (TLC). In Experiment 1, oil palm wastes were water extracted to produce aqueous extracts of OPT, EFB, PPMF and OPF. The application OPF aqueous extracts caused greatest radicle length reduction of lettuce, tomato, cucumber and green amaranth seedlings by61.9%, 65%, 53% and 52.6%, respectively, compared with control. PPMF and EFB extracts reduced radicle length significantly compared with OPT extracts, but degree of inhibition on radicle length treated with PPMF was higher than EFB aqueous extracts, with more than 20% radicle reduction for all seedlings, except cucumber seedlings, compared with control. However, OPT aqueous extracts showed no inhibition in radicle growth, hypocotyl growth and fresh and dry weights of all seedlings and total seed germination percentage and mean germination time. Thus, OPT extracts did not release any phytotoxic compounds and it can be concluded that OPF released phytotoxic compounds and degree of inhibition was higher compared with other wastes. PPMF inhibited seedling growth, with greater inhibition compared with EFB and OPT extracts. In Experiment 2, OPF and PPMF were further extracted using four solvents, hexane, diethyl ether, chloroform and ethyl acetate, to determine presence of phytotoxic compounds using seedling bioassay and seed germination test of lettuce,

tomato, cucumber and green amaranth. OPF diethyl ether extract exhibited highest inhibition on radicle length of lettuce (48,3%) and tomato (62,6%), but lower for cucumber (27.9%) and green amaranth (26.4%) seedlings. The OPF diethyl ether extract also reduced hypocotyl length of lettuce and green amaranth seedlings significantly compared to control. PPMF diethyl ether extract also reduced radicle length of lettuce, cucumber and green amaranth and hypocotyl length of green amaranth seedlings compared to control. Both solvent extracts showed high inhibition of radicle and hypocotyl length of seedlings compared with other treatments. Phenolic acid compounds present in OPF diethyl ether and PPMF diethyl ether extracts were identified using TLC and compared with 10 phenolic acid standards OPF extracts contained 4-hydroxybenzoic acid and syringic acid, while PPMF diethyl ether extracts contain 4-hydroxybenzaldehyde and syringic acid. OPF and PPMF contained phytotoxic compounds affecting plant growth and their utilization as mulching materials should be avoided to ensure that plants grow without inhibition. However, availability of OPT for mulching is limited since it is only available during oil palm replanting season. Thus, utilization of other readily available and safe oil palm wastes is essential to ensure that seedlings growth is not affected with mulching materials applied. EFB is one of the waste products of oil palm that would be safe to use and easily obtained since it has lower toxicity on seedling growth compared with OPF and PPMF wastes.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

KEFITOTOKSIKAN DALAM SISA KELAPA SAWIT SEBAGAI SUNGKUPAN BAGI ANAK POKOK SALAD, TOMATO, TIMUN DAN BENIH AMARANTH

Oleh

MOHD ZAKWAN BIN ZAMRI

April 2016

Pengerusi: Profesor Madya Siti Hajar Ahmad, PhD Fakulti: Pertanian

Sisa daripada industry kelapa sawit Malaysia termasuk tandan kosong (EFB), efluen kilang minyak sawit (POME), pelepah kelapa sawit (OPF), batang cincang (OPT), gentian mesokarpa kelapa yang ditekan (PPMF) dan kulit isi rong sawit (PKS) berjumlah 173.02 juta tan metric pada tahun 2014. Penggunaan sisa-sisa ini adalah penting untuk memaksimumkan penggunaan sumber dan mengatasi pencemaran alam sekitar. EFB dan PPMF mentah telah digunakan secara meluas di ladang kelapa sawit, sayur-sayuran dan industry landskap, sebagai sungkupan dan baja organik. OPT dan OPF dihasilkan daripada kelapa tua dan dibiarkan mereput di lapangan. Sisa tumbuhan segar boleh melepaskan sebatian fitotoksik sehingga menjejaskan pertumbuhan tumbuhan lain. Oleh itu, objektif kajian ini adalah (1) untuk menentukan kesan fitotoksik sisa kelapa sawit, EFB, PPMF, OPF dan OPT, menggunakan bioesei anak benih dan ujian percambahan benih kepada empat sayur-sayuran: salad (Lactuca sativa), tomato (Solanum lycopersicum), timun (Cucumis sativus) dan amaranth hijau (Amaranthus viridis), dan (2) untuk mengenal pasti sebatian fitotoksik asid fenolik dalam ekstrak sisa kelapa sawit mentah terpilih menggunakan kromatografi lapisan nipis (KLN). Dalam Eksperimen 1, sisa kelapa sawit adalah ekstrak akueus OPT, EFB, PPMF dan OPF. Aplikasi ekstrak akueus OPF menyebabkan pengurangan panjang radikel tertinggi bagi benih salad, tomato, timun dan amaranth hijau sebanyak 61.9%, 65%, 53% dan 52.6% masing-masing, berbanding dengan kawalan. PPMF dan ekstrak EFB mengurangkan panjang radikal dengan ketara berbanding dengan ekstrak OPT, tetapi tahap perencatan panjang radikal yang dirawat dengan PPMF adalah lebih tinggi daripada ekstrak akueus EFB, lebih daripada 20% pengurangan radikel untuk semua benih, kecuali anak benih timun, berbanding dengan kawalan. Walaubagaimanapun, ekstrak akueus OPT tidak menunjukkan perencatan pertumbuhan radikel, pertumbuhan hipokotil dan berat segar dan kering untuk semua benih dan jumlah peratusan percambahan benih dan purata masa percambahan. Oleh itu, ekstrak OPT tidak mengeluarkan sebarang sebatian fitotoksik, dan ia boleh membuat kesimpulan bahawa OPF mengeluarkan sebatian fitotoksik dan tahap perencatan adalah lebih tinggi berbanding dengan sisa lain. PPMF menghalang pertumbuhan anak benih, dengan

perencatan yang lebih besar berbanding dengan ekstrak EFB dan OPT. Dalam Eksperimen 2. OPF dan PPMF telah diekstrak menggunakan empat pelarut, heksana, dietileter, kloroform dan etil asetat, untuk menentukan kehadiran sebatian fitotoksik menggunakan bioesei anak benih dan benih ujian percambahan salad, tomato, timun dan amaranth hijau. Ekstrak dietileter OPF menunjukkan perencatan tertinggi bagi panjang radikal anak benih salad (48.3%) dan tomato (62.6%), tetapi lebih rendah untuk timun (27.9%) dan amaranth hijau (26.4%). Ekstrak dietileter OPF juga mengurangkan panjang hipokotil anak benih salad dan amaranth hijau dengan ketara berbanding dengan kawalan. Ekstrak dietileter PPMF juga mengurangkan panjang radikal anak benih salad, timun dan amaranth hijau dan panjang hipokotil anak benih amaranth hijau berbanding kawalan. Kedua-dua ekstrak pelarut menunjukkan perencatan radikel dan panjang hipokotil anak benih yang tinggi berbanding dengan rawatan lain. Sebatian asid fenolik di dalam ekstrak dietileter OPF dan PPMF telah dikenalpasti menggunakan KLN dan dibandingkan dengan 10 standard asid fenolik. Ekstrak OPF mengandungi asid 4-hidroksibenzoik dan asid siringik, manakala ekstrak dietileter PPMF mengandungi 4-hidroksibenzaldehid dan asid siringik. OPF dan PPMF mengandungi sebatian fitotoksik yang menjejaskan pertumbuhan tumbuhan dan penggunaannya sebagai bahan sungkupan perlu dielakkan untuk memastikan bahawa tumbuhan dapat tumbuh tanpa perencatan. Walaubagaimanapun, Ketersediaan OPT sebagai sungkupan adalah terhad kerana jahanya boleh didapati ketika musim penanaman semula kelapa sawit. Oleh itu, penggunaan bahan buangan kelapa sawit lain yang sedia ada dan selamat adalah penting untuk memastikan bahawa pertumbuhan anak pokok tidak terjejas dengan bahan-bahan sungkupan yang digunakan. EFB adalah salah satu daripada bahan buangan kelapa sawit yang selamat untuk digunakan dan mudah diperolehi kerana ia mempunyai ketoksikan yang lebih rendah pada pertumbuhan anak benih berbanding dengan sisa OPF dan PPMF.

ACKNOWLEDGEMENTS

In the name of Almighty ALLAH, Who provided me with the strength, wisdom and will to complete my master study. May His name be glorified and praised.

First and foremost, I would like to offer my heartfelt appreciation and utmost gratitude to my supervisor Associate Professor Dr. Siti Hajar Ahmad for her continuous support and invaluable guidance for my Master study, for her patience, motivation and enthusiasm. During my master study, she provided sound advice, good teaching and friendly company, and shared a lot of her expertise, research insight and best ideas. I simply could not imagine having a better advisor and friendlier mentor for Master study. I believe that one of the main gains of my master study was working with Associate Professor Dr. Siti Hajar Ahmad.

With a great deal of luck, I got an excellent Supervisory Committee. I owe an immense debt to Professor Datin Dr. Rosenani Abu Bakar for her encouragement, insightful comments and critical review. This thesis could not have been done without her strong supervision.

I am deeply indebted to my dear family (Mr. Zamri bin Zamli and Mrs Sharifah Marzita bt Syed Abdul Hamid), who deserve special mention for their inseparable support. To my sisters (Siti Nabihah bt Zamri, Nur Hamizah bt Zamri, Nur Zawanah bt Zamri and Siti Nur Zarifah bt Zamri) and brother (Muhamad Hazim bin Zamri), thank you for being supportive and caring siblings.

I would like to thank UPM for providing Graduate Research Fellowship (GRF) and research facilities to conduct my master study. I consider it an honor to work with all the administrative and technical staffs of the Faculty Agriculture, Mr. Azhar Othman. My sincere appreciation to Mrs Nor Elliza Tajidin, Mrs Bunga Raya Ketaren, Mrs Munirah, Miss Azimah Hamidon, Mrs Nur Indah Abdul Shukor, Miss Esther Yap Shiau Ping, Miss Tan Xue Yi, Miss Surisa Phornvillay who helped me with my work and writing.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Siti Hajar Ahmad, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Rosenani Binti Abu Bakar, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

BUJANG KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

Date:

Name and Matric No.: Mohd Zakwan Bin Zamri (GS34054)

Declaration by Members of Supervisory Committee

This is to confirm that:

C

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of	
Committee:	Siti Hajar Ahmad
Signature: Name of Member of Supervisory	
Committee	Rosenani Binti Abu Bakar

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF APPENDICES	xviii
LIST OF ABBREVIATION	XX

CHAPTER

1	GENE	RAL INTRO	DUCTION	1		
2	LITEI	RATURE RE	EVIEW	3		
	2.1 Oil Palm Waste			3		
		2.1.1	Empty Fruit Bunch	3		
		2.1.2	Oil Palm Frond	4		
		2.1.3	Oil Palm Trunk	5		
		2.1.4	Palm Oil Mill Effluent	5		
		2.1.5	Palm Pressed Mesocarp Fiber	6		
		2.1.6	Palm Kernel Shell	7		
	2.2	Mulching		7		
	2.2	2.2.1	Definition of Mulching	, 7		
		2.2.2	Types of Mulching	8		
		2.2.3	Advantages of Using Mulching	8		
		2.2.4	Disadvantages of Using Mulching	8		
	2.3	Phytotoxicit	y	9		
		2.3.1	Factors of Phytotoxicity	9		
		2.3.2	Methods of Releasing Phytotoxic Compounds from			
			Plants			
		2.3.3	Compounds Involve in Phytotoxicity	10		
		2.3.4	Effect of Phytotoxic Compound	11		
	2.4 Phenolic Acids		ids	12		
		2.4.1	Effect of Phenolic Acids on Plant Growth	12		
		2.4.2	Factors Influencing the Phytotoxicity of Phenolic	12		
			Acids Compound			
	2.5	Seed and Se	edling Bioassay	13		
		2.5.1	Leaf Lettuce (Lactuca sativa)	14		
		2.5.2	Tomato (Solanum lycopersicum)	14		
		2.5.3	Green Amaranth (Amaranthus viridis)	14		
		2.5.4	Cucumber (Cucumis sativus)	15		

3	DETE	RMINING P	RESENCE OF PHYTOTOXIC EFFECT IN OIL	16
	PALM	I WASTES	USING SEEDLING BIOASSAY AND SEED	
	GERN	IINATION		
	3.1	Introduction		16
	3.2	Materials and	d Methods	17
		3.2.1	Sample Collection	17
		3.2.2	Bioassay Test	18
		3.2.3	Seedling Bioassay	18
		3.2.4	Seed Germination	19
		3.2.5	Mean Germination Time	19
		3.2.6	Experimental Design and Statistical Analysis	20
	3.3	Results and I	Discussion	20
		3.3.1	Radicle Length	20
		332	Hypocotyl Length	22
		3 3 3	Radicle Diameter	24
		334	Seedling Fresh Weight	26
		335	Seedling Dry Weight	28
		336	Total Carmination Rate	20
		3.3.0	Mean Germination Time	30
	3 /	Conclusion	Wear Germination Time	30
	5.4	Conclusion		52
1	FYTR	ACTION A	ND IDENTIFICATION OF PHENOLIC ACID	33
-	COM	DOUND IN	DI DALM EDOND AND DALM DESSED	55
	MESC	CADD FIDE	D USING SEED AND SEEDLING DIOASSAV	
		CARI FIDI	CHDOMATOCDADUV	
		Introduction	CHROMATOGRAFHT	22
	4.1	Matorials on	d mathada	33
	4.2		Extraction of Water Soluble Divitatoria Compound	34
		4.2.1	Dertification of Aqueous Extract using Different	34
		4.2.2	Faithfolding of Aqueous Extract using Different	54
		122	Disassay of the Solvent Extraction Erections	25
		4.2.3	Bioassay of the Solvent Extraction Fractions	55 25
		4.2.4	Seeding Bloassay	33 25
		4.2.5	Seed Germination	33 25
		4.2.6	Identification of Phenolic Acid Phytotoxic	35
		107	Compounds using Thin Layer Chromatography	27
	1 2	4.2.7	Experimental Design and Statistical Analysis	36
	4.3	Results and I	Discussion	30
		4.3.1	Radicle Length	36
		4.3.2	Hypocotyl Length	39
		4.1.1	Radicle Diameter	42
		1010		45
		4.3.4	Seedling Fresh Weight	
		4.3.4 4.3.5	Seedling Dry Weight	46
		4.3.4 4.3.5 4.3.6	Seedling Dry Weight Total Germination Rate	46 48
		4.3.4 4.3.5 4.3.6 4.3.7	Seedling Dry Weight Total Germination Rate Mean germination time	46 48 49
		4.3.4 4.3.5 4.3.6 4.3.7 4.3.8	Seedling Fresh Weight Seedling Dry Weight Total Germination Rate Mean germination time Qualitative Analysis on Identification of Phenolic	46 48 49 51
		4.3.4 4.3.5 4.3.6 4.3.7 4.3.8	Seedling Fresh Weight Seedling Dry Weight Total Germination Rate Mean germination time Qualitative Analysis on Identification of Phenolic Acid Phytotoxic Compounds By Thin Layer	46 48 49 51
		4.3.4 4.3.5 4.3.6 4.3.7 4.3.8	Seedling Fresh Weight Seedling Dry Weight Total Germination Rate Mean germination time Qualitative Analysis on Identification of Phenolic Acid Phytotoxic Compounds By Thin Layer Chromatography	46 48 49 51

xi

5 SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR 54 FUTURE RESEARCH

REFERENCES	56
APPENDICES	71
BIODATA OF STUDENT	77
LIST OF PULICATIONS	78



LIST OF TABLES

Table3.1	Dry weights of <i>Lactuca sativa</i> and <i>Solanum lycopersicum</i> seedlings treated with waste aqueous extracts of oil palm trunk, empty fruit bunch, palm pressed mesocarp fiber and oil palm frond
3.2	Total seed germination rate of lettuce (<i>Lactuca sativa</i>), <i>tomato</i> (<i>Solanum lycopersicum</i>), <i>cucumber (Cucumis sativus</i>) and <i>green amaranth (Amarantus viridis)</i> treated with waste aqueous extracts of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond OPF.
3.3	Mean germination time of green amaranth (<i>Amarantus viridis</i>) to germinate when treated with waste aqueous extracts of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond OPF.
4.1	Hypocotyl length of cucumber <i>(cucumis sativus)</i> seedlings treated with control (distilled water), remaining aqueous and waste solvent (hexane, diethyl ether, chloroform, ethyl acetate) extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF).
4.2	Radicle diameter of cucumber (<i>Cucumis sativus</i>) seedlings treated with control (distilled water), remaining aqueous and waste solvent (hexane, diethyl ether, chloroform, ethyl acetate) extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF).
4.3	Fresh weight of lettuce (<i>Lactuca sativa</i>), cucumber(<i>Cucumis sativus</i>) and green amaranth (<i>Amaranthus viridis</i>) seedlings treated with distilled water (control), remaining aqueous and waste solvent (hexane, diethyl ether, chloroform, ethyl acetate) extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF).
4.4	Dry weight of tomato (<i>Solanum lycopersicum</i>) and, green amaranth (<i>Amaranthus viridis</i>) seedlings treated with distilled water (control), remaining aqueous and waste solvent (hexane, diethyl ether, chloroform, ethyl acetate) extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF).
4.5	Total germination rate of lettuce (<i>Lactuca sativa</i>), tomato(<i>Solanum lycopersicum</i>), cucumber (<i>Cucumis sativus</i>)

C

Page 28

31

30

39

45

42

48

and green amaranth (*Amaranthus viridis*) seedlings treated with distilled water (control), remaining aqueous and waste solvent (hexane, diethyl ether, chloroform, ethyl acetate) extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF).

- 4.6 Mean germination time of lettuce (*Lactuca sativa*), cucumber (*Cucumis sativus*) and, green amaranth (*Amaranthus viridis*) seedlings treated with distilled water (control), remaining aqueous and waste solvent (hexane, diethyl ether, chloroform, ethyl acetate) extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF).
- 4.7 Identified phenolic acids in oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF) diethyl ether extracts by thin layer chromatography eluted with ethyl acetate:chloroform (4:6)

52

LIST OF FIGURES

Figure

3.1

3.2

3.3

3.4

Measuring of seedling hypocotyl and length	Page 18
Radicle length of a) lettuce (<i>Lactuca sativa</i>), b) tomato (Solanum lycopersicum), c) cucumber (<i>Cucumis sativus</i>) and d) green amaranth (<i>Amaranthus viridis</i>) treated with waste aqueous extract of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond (OPF). Means, with the same letter above individual vertical bars, are not significantly different by using LSD test, P	21
= 0.05. n = 4.	
Hypocotyl lengths of a) lettuce (<i>Lactuca sativa</i>), b) tomato (<i>Solanum lycopersicum</i>), c) cucumber (<i>Cucumis sativus</i>) and d) green amaranth (<i>Amaranthus viridis</i>) treated with waste aqueous extract of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond (OPF). Means, with the same letter above individual vertical bars, are not significantly different by using LSD test, $P = 0.05$. $n = 10$.	23
Radicle diameter of a) lettuce (<i>Lactuca sativa</i>), b) tomato (<i>Solanum lycopersicum</i> , c) cucumber(<i>Cucumis sativus</i>) and d) green amaranth (<i>Amaranthus viridis</i>) treated with waste	25

(Solant green aqueous extract of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond (OPF). Means, with the same letter above individual vertical bars, are not significantly different by using LSD test, P=0.05. n = 4.

3.5 Fresh weight of a) lettuce (Lactuca sativa), b) tomato (Solanum 27 lycopersicum), c) cucumber(Cucumis sativus) and d) green amaranth (Amaranthus viridis) seedlings treated with waste aqueous extract of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond (OPF). Means, with the same letter above individual vertical bars, are not significantly different by using LSD test, P=0.05. n = 4.

29 3.6 Dry weight of a) cucumber (Cucumis sativus) and b) green amaranth (Amaranthus viridis) seedlings treated with waste aqueous extracts of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond (OPF). Bars with different lower-case letters are significantly different (P = 0.05) by LSD test. n = 4.

3.7 Mean germination time of lettuce (Lactuca sativa, treated with 31

XV

waste aqueous extracts of oil palm trunk (OPT), empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), and oil palm frond (OPF). Bars with different lower-case letters are significantly different ($P \le 0.05$) by LSD test. n = 4.

- 4.1 Radicle length of lettuce (*Lactuca sativa*), tomato (*Solanum lycopersicum*), cucumber (*Cucumis sativus*) and green amaranth (*Amaranthus viridis*) seedlings treated with distilled water (control), waste solvent hexane (HEX), diethyl ether (DE), chloroform (CHL), ethyl acetate (EA) and remaining aqueous (RA) extracts of oil palm frond (OPF) and pressed mesocarp fiber (PPMF). Values of the vertical bars are means of n = 4. Bars with different lower-case letters are significantly different (P ≤ 0.05), by Duncan multiple range test
- 4.2 Hypocotyl length of a) lettuce (Lactuca sativa), b) tomato (Solanum lycopersicum) and c) green amaranth (Amaranthus viridis) seedlings treated with distilled water (control), waste solvent hexane (HEX), diethyl ether (DE), chloroform (CHL), ethyl acetate (EA) and remaining aqueous (RA) extracts of oil palm frond (OPF) and pressed mesocarp fiber (PPMF). Values of the vertical bars are means of n = 4. Bars with different lower-case letters are significantly different (P ≤ 0.05), by Duncan multiple range test.
- 4.3 Radicle diameter of a) lettuce (*Lactuca sativa*), b) tomato (*Solanum lycopersicum*) and c) green amaranth (*Amaranthus viridis*) seedlings treated with distilled water (control), waste solvent hexane (HEX), diethyl ether (DE), chloroform (CHL), ethyl acetate (EA) and remaining aqueous (RA) extracts of oil palm frond (OPF) and pressed mesocarp fiber (PPMF). Values of the vertical bars are means of n = 4. Bars with different lower-case letters are significantly different ($P \le 0.05$), by Duncan multiple range test
- 4.4 Fresh weight of a) tomato (*Solanum lycopersicum*) seedlings treated with distilled water (control), waste solvent hexane (HEX), diethyl ether (DE), chloroform (CHL), ethyl acetate (EA) and remaining aqueous (RA) extracts of oil palm frond (OPF) and pressed mesocarp fiber (PPMF). Values of the vertical bars are means of n = 4. Bars with different lower-case letters are significantly different ($P \le 0.05$), by Duncan multiple range test.
- 4.5 Dry weight of a) lettuce (*Lactuca sativa*) and b) cucumber (*Cucumis sativus*) seedlings treated with distilled water (control), waste solvent hexane (HEX), diethyl ether (DE), chloroform (CHL), ethyl acetate (EA) and remaining aqueous (RA) extracts of oil palm frond (OPF) and palm pressed

38

41

44

46

mesocarp fiber (PPMF). Values of the vertical bars are means of n = 4. Bars with different lower-case letters are significantly different (P \leq 0.05), by Duncan multiple range test.

- 4.6 Mean germination time of a) tomato (*Solanum lycopersicu*) seedlings treated with distilled water (control), waste solvent hexane (HEX), diethyl ether (DE), chloroform (CHL), ethyl acetate (EA) and remaining aqueous (RA) extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF). Values of the vertical bars are means of n = 4. Bars with different lower-case letters are significantly different ($P \le 0.05$), by Duncan multiple range test.
- 4.7 Thin layer chromatography profile of diethyl ether extracts of oil palm frond (OPF) and palm pressed mesocarp fiber (PPMF) eluted with ethyl acetate:chloroform (4:6).

52

LIST OF APPENDICES

Appendix 1	ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of lettuce seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	Page 71
2	ANOVA table showing dry weight, total seed germination and mean germination time of lettuce seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	71
3	ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of tomato seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	71
4	ANOVA table showing dry weight, total seed germination and mean germination time of tomato seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	72
5	ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of cucumber seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	72
6	ANOVA table showing dry weight, total seed germination and mean germination time of cucumber seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	72
7	ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of green amaranth seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	73
8	ANOVA table showing dry weight, total seed germination and mean germination time of green amaranth seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	73
9	ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of lettuce seedling treated with PPMF and OPF aqueous extract.	73
10	ANOVA table showing dry weight, total seed germination and mean germination time of lettuce seedling treated with EFB, OPT, PPMF and OPF aqueous extract.	74
11	ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of tomato	74

seedling treated with PPMF and OPF solvent extract.

- 12 ANOVA table showing dry weight, total seed germination and mean germination time of tomato seedling treated with PPMF and OPF solvent extract.
- 13 ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of cucumber seedling treated with PPMF and OPF solvent extract.
- 14 ANOVA table showing the dry weight, total seed germination and mean germination time of cucumber seedling treated with PPMF and OPF solvent extract.
- 15 ANOVA table showing the radical length, hypocotyls length, radical diameter, and fresh weight of green amaranth seedling treated with PPMF and OPF solvent extract.
- 16 ANOVA table showing the dry weight, total seed germination and mean germination time of green amaranth seedling treated with PPMF and OPF solvent extract.

76

75

75

LIST OF ABBREVIATIONS

EFB	Empty fruit bunch
OPF	Oil palm fiber
PPMF	Palm press mesocarp fiber
OPT	Oil palm trunk
TLC	Thin layer chromatography
ANOVA	Analysis of variance
g	Gram
mm	Millimeter
cm	Centimeter
mg	Milligram
DMRT	Duncan multiple range test
LSD	Least significant difference
POME	Palm oil mill effluent
PKS	Palm kernel shell
FFB	Fresh fruit bunch
K	Potassium
Ν	Nitrogen
Mg	Magnesium
Р	Phosphorous
С	Carbon
m	Meter
C	Degree Celsius
%	Percentage
g/L	Gram per liter
Mm	Millimeter
EC	Electrical conductivity
$mS cm^{-1}$	Millisiemens per centimeter
SAS	Statistical Analysis System
UPM	Universiti Putra Malaysia
USEPA	United States Environmental Protection Agency
USFDA	United States Food and Drug Administration
OECD	Organization for Economic Coorperation and Development
kg	Kilogram
m/v	Mass per volume
rpm	Revolutions per minute
DNA	Deoxyribonucleic acid
NH ₃	Ammonia

C

CHAPTER 1

GENERAL INTRODUCTION

Oil palm (*Elaeis guineensis Jacq.*), in the family of Arecaceae is considered as the largest plantation industry in Malaysia and one of the major contributions towards the economic growth in some developing countries. It is cultivated in about 43 countries with over 12 million hectare in 2009. In Malaysia, the area under cultivation was increased by 5 times from 4 decades ago (Teoh, 2010). This is due to the suitable environment with high temperature between 22 to 33° C, high rainfall (>2000 mm) and sufficient sunlight (>7 hours) for these crops to grow (Tiquia *et al*, 1996). According to the Malaysia and 52.1% of total world population consumed palm oil (Anon., 2014). The oil palm industry produces broad ranges of product from crude oil to cosmetics product (Mohammad *et al.*, 2012). Besides some of oil palm wastes have been burnt in boiler to produce steam for electricity generation used in the palm oil mill industry (Sheil *et al.*, 2009).

However, the increasing oil palm production has released abundant agricultural residues in Malaysia. There are several type of residues produced from oil palm industry which are empty fruit bunches (EFB), palm oil mill effluent (POME), oil palm fronds (OPF), chopped trunks (OPT), palm pressed mesocarp fibre (PPMF) and palm kernel shell (PKS) (Basiron and Weng, 2004). Thus, it is advisable that waste products of these industry be investigated with the objective of using the raw material for other industry to get desired product (Singh *et al.*, 2011). Therefore, the utilization of these residues is important to maximize the uses of resources and to overcome environmental pollution (Ahmad, 2001). Many researches were carried out to support the possibilities of converting oil palm residues into many value added products such as EFB and POME which have been used as organic fertilizer and mulch in oil palm areas, while pressed fibre and shell are used as a fuel to generate steam and energy used for the operation of the mill. Other than that, palm kernel is effectively used to produce palm kernel cake for residual livestock food (Ahmad, 2001).

Besides that, raw EFB and PPMF have been widely used in vegetable farming and landscape industry as mulching and organic fertilizer (Yusof, 2006). However, fresh plant wastes used for organic fertilizer and mulching need to decompose to avoid the harmful accumulation of lipids and volatile compounds that can inhibit plants growth. Besides, decomposition is essential to redu00ce phytotoxin compound (Zucconi *et al*, 1981).

Phytotoxicity is the negative effects of substances arising from decomposing crop residues on growth and yield of the crops. The phytotoxicity of plant residue has been associated with the presence of various organic compounds including water soluble phenolic compounds that widely distributed all over the part of various plants species (Rice, 1984). Application of POME to seedling is proven to inhibit the growth of seedling significantly. The inhibitory effect of raw POME on plant growth is similar to

the phytotoxicity exhibited by other types of crop residues (Radziah *et al*,1997). However the degree of phytotoxicity depends on the types of crop residues, decomposition period and the sensitivity of plant root system toward phytotoxic compound. There are several type of compounds was classified as phytotoxic such as phenolic acid, heavy metal, ammonia, excessive accumulation of salt, lipid and organic acid (Radziah *et al*, 1997). However, phenolic acid is commonly identified as phytotoxin and inhibit growth of seedling and reduce crop yield (Einhellig, 1985)

In Malaysia, there are a lot of vegetable farms present in high or lowland. One of the most important planting materials that are highly demanded in vegetable production is mulching material, compost and organic fertilizer. Oil palm wastes such as EFB and OPMF, can be used as mulching material in vegetable production (Yusof, 2006) while OPF commonly been put under oil palm tree between the row to act as mulching and conserve soil structure (Mohammad *et* al., 2012). A study from Kim and Rahman, (2002) proved that mulching with EFB improved the soil exchangeable potassium, calcium and magnesium and pH. Mulching material gave many advantages to the plant growth such as increases in soil organic matter content, improves soil structure, improves soil water retention and increases cation exchange capacity (Kim and Rahman, 2002). Abdul Mutalib *et al.*, (2009) showed that EFB wastes in combination with inorganic fertilizer could support shoot growth of cabbage. However, the information of phytotoxity effect on different type of oil palm wastes is still not available except for POME.

A series of seed germination and seedling bioassay tests were undertaken on 4 selected vegetable seedlings; *Amaranthus viridis* (green amaranth), *Lactuca sativa* (lettuce), *Solanum lycopersicum* (tomato) and *Cucumis sativus* (cucumber);

- To determine the phytotoxic effect of four different oil palm wastes, empty fruit bunch (EFB), palm pressed mesocarp fiber (PPMF), oil palm frond (OPF) and oil palm trunk (OPT).
- To identify the phenolic acid phytotoxic compounds that present in extracts of selected raw oil palm waste by using thin layer chromatography (TLC).

REFERENCES

- Abu-Awwad, A. (1999). Irrigation water management for efficient water use in mulched onion. *Journal of Agronomy and Crop Science*, 183(1), 1-7.
- Ahmad, F. (2001, September). Sustainable agriculture system in Malaysia. In Regional Workshop on Integrated Plant Nutrition System (IPNS), Development in Rural Poverty Alleviation, United Nations Conference Complex, Bangkok, Thailand (pp. 18-20).
- Ahmad, A. L., Ismail, S., and Bhatia, S. (2003). Water recycling from palm oil mill effluent (POME) using membrane technology. *Journal Desalination*, 157(5), 87-95.
- Al-Amri, S. M. (2013). Improved growth, productivity and quality of tomato (Solanumlycopersicum L.) plants through application of shikimic acid. Saudi Journal of Biological Sciences, 20(4), 339-345.
- Alancar, S. R., Silva, M. A. P., Matos, M. F., Santos, M. A. F., Generino, M. E. M., Torquato, I. H. S., and Crispim, M. K. M. (2015). Biological Activity of *Bambusa vulgaris* Schrad. Ex JC Wendl.(Poaceae). *Journal of Agricultural Science*, 7(6), 150.
- Ali, M. F., and Abbas, S. (2006). A review of methods for the demetallization of residual fuel oils. *Fuel Processing Technology*, 87(7), 573-584.
- Alimon, A. (2004). The nutritive value of palm kernel cake for animal feed. *Palm Oil Dev*, 40(1), 12-14.
- Anonymus, (2001). Proper Mulching Techniques .http://www.treesaregood.com/tree care/resources/propermulching.pdf
- Anonymus,(2015),http://bepi.mpob.gov.my/index.php/statistics/production/125production-2014/659 production-of-crude-oil-palm-2014.html.
- Ariffin, A., and Tan, Y. A. (1990). The effects of handling of FFB on the formation of FFA and the subsequent quality of crude palm oil. Paper presented at the Proceedings of the International Palm Oil Development Conference Agriculture, Kuala Lumpur.
- Balke, N. E., Davis, M. P., and Lee, C. C. (1987). Conjugation of allelochemicals by plants. In G. R. Waller (Ed.), *Allelochemicals: Role in Agriculture and Forestry* (pp. 214-227). Washington, D.C.
- Barche, S., and Nair, R. (2014). Mulching-An Effective Conservation Technique in Olericulture. *Popular Kheti*, 2(2), 49-55.
- Barkosky, R. R., Einhellig, F. A., and Butler, J. L. (2000). Caffeic acid-induced changes in plant– water relationships and photosynthesis in leafy spurge *Euphorbia esula*. *Journal of Chemical Ecology*, *26*(9), 2095-2109.

- Basiron, Y., and Weng, C. K. (2004). The oil palm and its sustainability. *Journal of Oil Palm Research*, *16*(1), 231-244.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109(4), 289-295.
- Batish, D. R., Setia, N., Singh, H., and Kohli, R. (2004). Phytotoxicity of lemonscented eucalypt oil and its potential use as a bioherbicide. *Crop Protection*, 23(12), 1209-1214.
- Batish, D. R., Singh, H. P., Setia, N., Kaur, S., and Kohli, R. K. (2006). Chemical composition and phytotoxicity of volatile essential oil from intact and fallen leaves of *Eucalyptus citriodora*. *Zeitschrift für Naturforschung C*, 61(7-8), 465-471.
- Batish, D. R., Kaur, M., Singh, H. P., and Kohli, R. K. (2007). Phytotoxicity of a medicinal plant, *Anisomeles indica*, against Phalaris minor and its potential use as natural herbicide in wheat fields. *Crop Protection*, 26(7), 948-952.
- Baziramakenga, R., Simard, R. R., & Leroux, G. D. (1994). Effects of benzoic and cinnamic acids on growth, mineral composition, and chlorophyll content of soybean. *Journal of Chemical Ecology*, 20(11), 2821-2833.
- Baziramakenga, R., Leroux, G. D., & Simard, R. R. (1995). Effects of benzoic and cinnamic acids on membrane permeability of soybean roots. *Journal of Chemical Ecology*, 21(9), 1271-1285.
- Bhowmik, P. C., and Doll, J. D. (1984). Corn and soybean response to allelopathic effects of weed and crop residues. *Agronomy Journal*, 74(4), 601-606.
- Bhowmik, P. C. (2003). Challenges and opportunities in implementing allelopathy for natural weed management. *Crop Protection*, 22(4), 661-671.
- Blum, U., and Dalton, B. R. (1985). Effects of ferulic acid, an allelopathic compound, on leaf expansion of cucumber seedlings grown in nutrient culture. *Journal of chemical ecology*, *11*(3), 279-300.
- Blum, U. (1996). Allelopathic interactions involving phenolic acids. *Journal of Nematology*, 28(3), 259.
- Blum, U., Shafer, S. R., & Lehman, M. E. (1999). Evidence for inhibitory allelopathic interactions involving phenolic acids in field soils: concepts vs. an experimental model. *Critical Reviews in Plant Sciences*, *18*(5), 673-693.
- Bogatek, R., Gniazdowska, A., Zakrzewska, W., Oracz, K., and Gawronski, S. W. (2006).Allelopathic effects of sunflower extracts on mustard seed germination and seedling growth. *Biologia Plantarum*, 50(1), 156-158.
- Bradow, J. M. (1991). Relationships between chemical structure and inhibitory activity of C6 through C9 volatiles emitted by plant residues. *Journal of chemical ecology*, *17*(11), 2193-2212.

- Braids, O. C., and Miller, R. H. (1975). Fat, waxes and resins in soil. In J. E. Gieseking (Ed.), Soil Components. Vol 1. Organic Components (pp. 343-367). New York: Springer-Verlag.
- Bravo, J., Monente, C., Juániz, I., De Peña, M. P., & Cid, C. (2013). Influence of extraction process on antioxidant capacity of spent coffee. *Food Research International*, 50(2), 610-616.
- Buerkert, A., Bationo, A., and Dossa, K. (2000). Mechanisms of residue mulch-induced cereal growth increases in West Africa. Soil Science society of Amerca Journal, 64(1), 346-358.
- Burgos, N., Talbert, R., Kim, K., and Kuk, Y. (2004). Growth inhibition and root ultrastructure of cucumber seedlings exposed to allelochemicals from rye (*Secale cereale*). Journal of chemical ecology, 30(3), 671-689.
- Camargo, C. E. D. O., Ferreira Filho, A. W. P., & Salomon, M. V. (2004). Temperature and pH of the nutrient solution on wheat primary root growth.*Scientia Agricola*, *61*(3), 313-318.
- Campiglia, E., Mancinelli, R., Radicetti, E., & Caporali, F. (2010). Effect of cover crops and mulches on weed control and nitrogen fertilization in tomato (Lycopersicon esculentum Mill.). *Crop Protection*, 29(4), 354-363.
- Castro, A. D., and Helena, A. (2006). *Atmospheric* NH₃ Deposition, S and N metabolism in curly kale. (Ph.D. thesis), Suniversity of Groningen.
- Chan, K. W., Watson, I., & Lim, K. C. (1980). Use of oil palm waste material for increased production. *Planter*, *57*(658), 14-37.
- Chan, K. (1999). Biomass production in the palm oil industry. In G. Singh (Ed.), Oil palm and the environment: A Malaysian perspective (pp. 41-53). Kuala Lumpur: Malaysian Oil Palm Growers Council.
- Chaves, N., Sosa, T., and Escudero, J. C. (2001). Plant growth inhibiting flavonoids in exudate of *Cistus ladanifer* and in associated soils. *Journal of chemical ecology*, 27(3), 623-631.
- Cheung, Y. H., Wong, M. H., and Tam, N. F. Y. (1989). Root and shoot elongation as an assessment of heavy metal toxicity and 'Zn Equivalent Value' of edible crops.In *Environmental Bioassay Techniques and their Application* (pp. 377-383). Springer Netherlands.
- Chick, T. A., and Kielbaso, J. J. (1998). Allelopathy as an inhibition factor in ornamental tree growth: implications from the literature. *Journal of Arboriculture*, 24(5), 274-279.
- Chon, S. U., Choi, S. K., Jung, S., Jang, H. G., Pyo, B. S., and Kim, S. M. (2002). Effects of alfalfa leaf extracts and phenolic allelochemicals on early seedling growth and root morphology of alfalfa and barnyard grass. *Crop protection*, 21(10), 1077-1082.

- Chon, S. U., and Kim, D. K. (2005). Allelopathic potential of *Xanthium occidentale* extracts and residues. *Korean Journal of Weed Science*, 30(2), 349-358.
- Chung, I. M., and Miller, D. A. (1995). Natural herbicide potential of alfalfa residue on selected weed species. *Agronomy Journal*, 87(5), 920-925.
- Cooperband, L., Stone, A., Fryda, M., and Ravet, J. (2003). Relating compost measures of stability and maturity to plant growth. *Compost science and utilization*, 11(2), 113-124.
- Czabator, F. J. (1962). Germination value: an index combining speed and completeness of pine seed germination. *Forest Science*, 8(4), 386-396.
- Danks, M. L., Fletcher, J. S., and Rice, E. L. (1975). Influence of Ferulic Acid on Mineral Depletion and Uptake of ⁸⁶Rb by Paul's Scarlet Rose Cell-Suspension Cultures. *American Journal of Botany*, 749-755.
- Deraman, M. (1993). Palm Oil Research Inst Malaysia. Bulletin, 26, 1-5.
- Desai, B. B. (2004). Seeds handbook: Processing and storage: CRC Press.
- Dilipkumar, M., Mazira, C., and Chuah, T. (2015). Phytotoxicity of different organic mulches on emergence and seedling growth of goosegrass (*Eleusine indica*). *Journal Tropical Agriculture and Food Science*, 43(2), 145-153.
- DOA. (2013). Agro Food Statistics 2012. Kuala Lumpur: Strategic Planning and InternationalDivision. Malaysia.
- Einhellig, F. A., Leather, G. R., and Hobbs, L. L. (1985). Use of Lemna minor L. as a bioassay in allelopathy. *Journal of chemical Ecology*, *11*(1), 65-72.
- Ellnain-Wojtaszek, M., Kruczynski, Z., and Kasprzak, J. (2001). Analysis of the content of flavonoids, phenolic acids as well as free radicals from *Ginkgo biloba* L. leaves during the vegetative cycle. *Acta poloniae pharmaceutica*, 58(3), 205-210.
- Ernest, R., and Buffington, L. (1981). Crop residue. In T. A. McClure and E. S. Lipsinky (Eds.), *Handbook of bisolar research materials*. Boca Raton: FL7 CRC Press.
- Fageria, N. K. (2007). Yield physiology of rice. *Journal of Plant Nutrition*, 30(6), 843-879.
- Fan, D., Hodges, D. M., Zhang, J., Kirby, C. W., Ji, X., Locke, S. J., ... & Prithiviraj, B. (2011). Commercial extract of the brown seaweed Ascophyllum nodosum enhances phenolic antioxidant content of spinach (Spinacia oleracea L.) which protects Caenorhabditis elegans against oxidative and thermal stress. *Food Chemistry*, 124(1), 195-202.
- Gagliardo, R. W., and Chilton, W. S. (1992). Soil transformation of 2 (3H)benzoxazolone of rye into phytotoxic 2-amino-3H-phenoxazin-3-one. *Journal of Chemical Ecology*, 18(10), 1683-1691.

- Garau, G., Mele, E., Castaldi, P., Lauro, G. P., and Deiana, S. (2015). Role of polygalacturonic acid and the cooperative effect of caffeic and malic acids on the toxicity of Cu (II) towards triticale plants (× *Triticosecale Wittm*). *Biology* and Fertility of Soils, 51(5), 535-544.
- Gao, X., Li, M. E. I., Gao, Z., Li, C., and Sun, Z. (2009). Allelopathic effects of *Hemistepta lyrata*on the germination and growth of wheat, sorghum, cucumber, rape, and radish seeds. *Weed Biology and Management*, 9(3), 243-249.
- Gerig, T. M., and Blum, U. (1991). Effects of mixtures of four phenolic acids on leaf area expansion of cucumber seedlings grown in Portsmouth B1 soil materials. *Journal of Chemical Ecology*, 17(1), 29-40.
- Gilman, E. F., Beeson, R. C., and Meador, D. (2012). Impact of Mulch on Water Loss from a Container Substrate and Native Soil. *Arboriculture and Urban Forestry*, 38(1), 18.
- Gross, E. M., Erhard, D., & Iványi, E. (2003). Allelopathic activity of Ceratophyllum demersum L. and Najas marina ssp. intermedia (Wolfgang) Casper. *Hydrobiologia*, 506(1-3), 583-589.
- Guenzi, W. D., McCalla, T. M., and Norstadt, F. A. (1967). Presence and Persistence of Phytotoxic Substances in Wheat, Oat, Corn and Sorghum Residues. *Agronomy Journal*, 59, 163-165.
- Han, C. M., Pan, K. W., Wu, N., Wang, J. C., and Li, W. (2008). Allelopathic effect of ginger on seed germination and seedling growth of soybean and chive. *Scientia horticulturae*, 116(3), 330-336.
- Hassan, M., and Shirai, Y. (2003). Palm biomass utilization in Malaysia for the production of bioplastic. Paper presented at the Biomass-Asia-workshop, <u>www.</u> biomassasiaworkshop. jp/presentation files/21 Alihassan. pdf.
- Hassen, A., Belguith, K., Jedidi, N., Cherif, A., Cherif, M., & Boudabous, A. (2001). Microbial characterization during composting of municipal solid waste. *Bioresource technology*, 80(3), 217-225.
- Haugland, E., and Brandsaeter, L. O. (1996). Experiments on bioassay sensitivity in the study of allelopathy. *Journal of Chemical Ecology*, 22(10), 1845-1859.
- Henson, I. E., Betitis, T., Tomda, Y., and Chase, L. D. (2012). The estimation of frond base biomass (FBB) of oil palm. *Journal of Oil Palm Research*, 24(12), 1473-1479.
- Holappa, L. D., and Blum, U. (1991). Effects of exogenously applied ferulic acid, a potential allelopathic compound, on leaf growth, water utilization, and endogenous abscisic acid levels of tomato, cucumber and bean. *Journal Chemistry Ecology*, 17, 865-886.

- Huang, Y., Zhang, W., Zheng, X., Li, J., and Yu, Y. (2004). Modeling methane emission from rice paddies with various agricultural practices. *Journal of Geophysical Research: Atmospheres* (1984–2012),109(D8).
- Hundal, I. S., Sandhu, K. S., Singh, D., & Sandha, M. S. (2000). Effect of different types of mulching and herbicidal treatments on nutrient uptake in tomato (Lycopersicon esculentum). *Haryana Journal of Horticultural Sciences*, 29(3/4), 242-244.
- Husain, Z., Zainac, Z., and Abdullah, Z. (2002). Briquetting of palm fibre and shell from the processing of palm nuts to palm oil. *Biomass and Bioenergy*, 22(6), 505-509.
- Iannucci, A., Fragasso, M., Platani, C., Narducci, A., Miullo, V., and Papa, R. (2013). Dynamics of release of allelochemical compounds from roots of wild oat (*Avena fatua L.*). Agrochimica, 56(2), 90-103.
- Igwe, J., and Onyegbado, C. (2007). A review of palm oil mill effluent (POME) water treatment. *Global Journal of Environmental Research*, 1(2), 54-62.
- Ilori, O. J., and Ilori, O. O. (2012). Allelochemicals: types, activities and usage in Pest control. J Sci Sci Ed, Ondo, 3(1), 106-110.
- Inderjit, K., and Olosfsdotter, M. (1998). Using and improving laboratory bioassays in rice allelopathy research. Allelopathy in Rice, International Rice Research Institute, Manila, 45-45.
- Islam, A. K. M., and Kato-Noguchi, H. (2014). Phytotoxic Activity of *Ocimum tenuiflorum*Extracts on Germination and Seedling Growth of Different Plant Species. *The Scientific World Journal*, 2014.
- Jordan, D., Kremer, R., Bergfield, W., Kim, K., and Cacnio, V. (1995). Evaluation of microbial methods as potential indicators of soil quality in historical agricultural fields. *Biology and Fertility of Soils*, 19(4), 297-302.
- Jorgensen, H. (1985). Treatment of empty fruit bunches for recovery of residual oil and additional steam production. *Journal of the American Oil Chemists' Society*, 62(2), 282-286.
- Jouyban, Z. (2012). The Effects of Salt stress on plant growth. Technical Journal of Engineering and Applied Sciences, 2(1), 7-10.
- Kamarudin, H., mohamad, H., Ariffin, D., and Johri, S. (1997). An estimated availability of oil palm biomass in Malaysia *PORIM Occ Paper* (Vol. 37): Palm Oil Research Institute of Malaysia.
- Kapanen, A., and Itävaara, M. (2001). Ecotoxicity tests for compost applications. *Ecotoxicology* and environmental safety, 49(1), 1-16.

- Kapustka, L. A. (1997). Selection of phytotoxicity tests for use in ecological risk assessments. In: Wang, W.; Gorsuch, J.W.; Hughes, D. Plants for environmental studies. New York: CRC Press, 1997. pp.516-548.
- Kaur, S., Singh, H. P., Batish, D. R., and Kohli, R. K. (2011). Chemical characterization and allelopathic potential of volatile oil of *Eucalyptus* tereticornis against Amaranthus viridis. Journal of Plant Interactions, 6(4), 297-302.
- Kaya, Y., Aksakal, O., Sunar, S., Erturk, F. A., Bozari, S., Agar, G., and Battal, P. (2015). Phytotoxical effect of *Lepidium draba* L. extracts on the germination and growth of monocot (*Zea mays* L.) and dicot (*Amaranthus retroflexus* L.) seeds. *Toxicology and Industrial Health*, 31(3), 247-254.
- Kerdsuwan, S., and Laohalidanond, K. (2011).*Renewable energy from palm oil empty fruit bunch*. INTECH Open Access Publisher.
- Khalil, H. S. A., Alwani, M. S., and Omar, A. K. M. (2007). Chemical composition, anatomy, lignin distribution, and cell wall structure of Malaysian plant waste fibers. *BioResources*, 1(2), 220-232.
- Kimber, R. W. L. (1973). Phytotoxicity from plant residues. II. The effect of time of rotting of straw from some grasses and legumes on the growth of wheat seedlings. *Plant Soil*, (38), 235-491.
- Kim. C. L., & Rahman, Z. A. (2002). The effects oil palm empty fruit bunches on oil palm nutrition and yield, and soil chemical properties. *Journal of Oil Palm Research*, 14(2), 1-9.
- Kozlowska, M., & Krzywanski, Z. (1993, September). The possible role of phenolic compounds in red raspberry resistance to Didymella applanata (Niessl) Sacc. In *International Symposium on Natural Phenols in Plant Resistance 381* (pp. 671-674).
- Komilis, D. P., and Tziouvaras, I. S. (2009). A statistical analysis to assess the maturity and stability of six composts. *Waste Management*, 29(5), 1504-1513.
- Koller, D., & Hadas, A. (1982). Water relations in the germination of seeds. In*Physiological Plant Ecology II* (pp. 401-431). Springer Berlin Heidelberg.
- Kumar, S. D., & Bhardwaj, R. L. (2012). Effect of mulching on crop production under rainfed condition: A Review. *Int. J. Res. Chem. Environ*,2(2), 8-20.
- Lee, J. G., Lee, B. Y., and Lee, H. J. (2006). Accumulation of phytotoxic organic acids in reused nutrient solution during hydroponic cultivation of lettuce (Lactuca sativa L.). Scientia horticulturae, 110(2), 119-128.
- Leh, C. P., Rosli, W. W., Zainuddin, Z., and Tanaka, R. (2008). Optimisation of oxygen delignification in production of totally chlorine-free cellulose pulps from oil palm empty fruit bunch fibre. *Industrial Crops and Products*, 28(3), 260-267.

- Lerch, R., Barbarick, K., Sommers, L., and Westfall, D. (1992). Sewage sludge proteins as labile carbon and nitrogen sources. *Soil Science Society of America Journal*, 56(5), 1470-1476.
- Li, H.-M., L. Altschmied and J. Chory, 1993 Arabidopsis mutants nabe et al., 1996 Action spectra for phytochrome A- and Bdefine downstream branches in the phototransduction pathway. specific photoinduction of seed germination in Arabidopsis thaliGenes Devel. 8: 339–349
- Lim, K. H. (1986). *Optimising land application of digested palm oil mill effluent.* (Ph.D. thesis), State University of Ghent.
- Loh, S. K., Lai, M. E., Ngatiman, M., Lim, W. S., Choo, Y. M., Zhang, Z. and Salimon, J. (2013). Zero discharge treatment technology of palm oil mill effluent. *Journal of Oil Palm Research*, 25(3), 273-281.
- Lorestani, A. (2006). Biological treatment og palm oilmill effluent (POME) using an up-flow anaerobic sludge fixed film (UASFF) bioreactor. (Ph.D), Universiti Sains Malaysia.
- Ma, A. N. (1995). A novel treatment for palm oil mill effluent. Palm Oil Research Institute Malaysia (PORIM), 29, 201-212.
- Mahmoodzadeh, H., and Mahmoodzadeh, M. (2013). Allelopathic potential of soybean (*Glycine max* L.) on the germination and root growth of weed species. *Life Science Journal*, *10*, 63-69.
- McQualter, R. B., Chong, B. F., Meyer, K., Van Dyk, D. E., O'Shea, M. G., Walton, N. J., ... & Brumbley, S. M. (2005). Initial evaluation of sugarcane as a production platform for p-hydroxybenzoic acid. *Plant biotechnology journal*,3(1), 29-41.
- Mengel, K., & Kirkby, E. A. (1978). Principles of plant nutrition. *Principles of plant nutrition*.
- Mitelut, A. C., and Popa, M. E. (2011). Seed germination bioassay for toxicity evaluation of different composting biodegradable materials. *Romanian Biotechnological Letters*, *16*(1), 121-129.
- Mohammad, N., Alam, M. Z., Kabbashi, N. A., and Ahsan, A. (2012). Effective composting of oil palm industrial waste by filamentous fungi: A review.*Resources, Conservation and Recycling*,58, 69-78.
- Moradi, A., Teh, C. B. S., Goh, K. J., Husni, M. H. A., & Ishak, C. F. (2014). Decomposition and nutrient release temporal pattern of oil palm residues.*Annals* of applied biology, 164(2), 208-219.
- Moucawi, J., Eliane, F., andJambu, P. (1981). Decomposition of lipids in soils: Free and esterified fatty acids, alcohols and ketones. *Soil Biology & Biochemistry*, 13, 461-468.

- MPOB. (2002). Journal of Oil Palm Research. 21st edition, MPOB, Bangi. 131 pp. http://www.etawau.com/OilPalm/OilPalmFiber.htm
- MPOC, 2010. Palm Oil: A Success Story in Green Technology 471 Innovations. 472 http://www.akademisains.gov.my/download/asmic/asmic2010/Plenary12.pdf.47 3Accessed October 10, 2011
- Mulumba, L. N., and Lal, R. (2008). Mulching effects on selected soil physical properties. *Soil and Tillage Research*, *98*(1), 106-111.
- Narwal, S. S. (1999). Allelopathy update. Volume 2: Basic and applied aspects. Science Publishers, Inc..
- Nimbal, C. I., Pedersen, J. F., Yerkes, C. N., Weston, L. A., & Weller, S. C. (1996). Phytotoxicity and distribution of sorgoleone in grain sorghum germplasm. *Journal of Agricultural and Food Chemistry*, 44(5), 1343-1347.
- Oerke, E. C. (2006). Crop losses to pests. *The Journal of Agricultural Science*, 144(01), 31-43.
- Olofsdotter, M., Rebulanan, M., Madrid, A., Dali, W., Navarez, D., & Olk, D. C. (2002). Why phenolic acids are unlikely primary allelochemicals in rice. *Journal of Chemical Ecology*, 28(1), 229-242.
- Omar, A. H., Ishida, M., I., M. S., and Z., A. T. (1996). Oil-palm fronds as a roughage feed source for ruminants in Malaysia. ASPAC, *Food and Fertilizer Technology Center*, 1-8.
- Öncel, I., Keleş, Y., and Üstün, A. S. (2000). Interactive effects of temperature and heavy metal stress on the growth and some biochemical compounds in wheat seedlings. *Environmental Pollution*, *107*(3), 315-320.
- Othman, R., Hashim, A., and Abdul Rahman, Z. (1997). The phytotoxic effects of palm oil dry solids on plant growth. *Pertanika Journal of Tropical Agricultural Science*, 20(2), 91-99.
- Padmaja, K., Prasad, D. D. K., and Prasad, A. R. K. (1990). Inhibition of chlorophyll synthesis in *Phaseolus vulgaris* L. seedlings by cadmium acetate.*Photosynthetica*, 24(3), 399-405.
- Paepatung, N., Kullavanijaya, P., Loapitinan, O., Songkasiri, W., Noppharatana, A. C.
 P., and Chaiprasert, P. (2006). Assessment of biomass potential for biogas production in Thailand. *Final report submitted to The Joint Graduate School of Energy and Environment, Bangkok, Thailand (in Thai).*
- Pardo, J. M. (2010). Biotechnology of water and salinity stress tolerance. Current Opinion in Biotechnology, 21(2), 185-196.
- Parker, J. B., James, L., Jarvis, J., and Parks, S. (2010). *Commercial Greenhouse Cucumber Production*. Industry and Investment. Australia.

- Peng, S. L., Wen, J., and Guo, Q. F. (2004). Mechanism and active variety of allelochemicals. *Acta Botanica Sinica*, 46(7), 757-766.
- Perez, J., Dela Rubia, T., Moreno, J., and Martinez, J. (1992). Phenolic content and antibacterial activity of olive oil waste waters. *Environmental Toxicology and Chemistry*, 11(4), 489-495.
- Ping, L. Y., Sung, C. T. B., Joo, G. K., and Moraidi, A. (2012). Effects of Four Soil ConservationMethods on Soil Aggregate Stability. *Malaysian Journal of Soil Science*, 16, 43-56.
- Pramanik, M. H. R., Minesaki, Y., Yamamoto, T., Matsui, Y., and Nakano, H. (2001). Growth inhibitors in rice-straw extracts and their effects on Chinese milk vetch (Astragalus sinicus) seedlings. Weed Biology and Management, 1(2), 133-136.
- Prasertsan, S., and Prasertsan, P. (1996). Biomass residues from palm oil mills in Thailand: An overview on quantity and potential usage. *Biomass and Bioenergy*, 11(5), 387-395.
- Premuzic, Z., Palmucci, H. E., Tamborenea, J., and Nakama, M. (2007). Chlorine disinfection: effects on hydroponics lettuce.*The Society for Advancement of Horticulture*, 9(1), 62-65.
- Putnam, A. R., and Tang, C. S. (1986). *The science of allelopathy*: John Wiley and Sons Inc. New York, USA.
- Radziah, O., Rahmani, M., and Hashim, A. (1997). Phytotoxicity of Phenolic Acids Extracted from Palm Oil Dry Solids. *Pertanika Journal of Tropical Agricultural Science*,20(2/3), 147-155.
- Radziah O. (2001) Alleviation of phytotoxicity of raw POME by microorganism: Retrieved Sept. 2005, from www.agri.upm.edu.my/agrosearch/v3n2/irpa3.htm.
- Rice, E. L. (1984) Allelopathy. New York: Academic Press
- Robbins, R. J. (2003). Phenolic acids in foods: an overview of analytical methodology. *Journal of agricultural and food chemistry*, 51(10), 2866-2887.
- Romeo, J. T., and Weidenhamer, J. D. (1998). Bioassays for allelopathy in terrestrial plants *Methods in Chemical Ecology Volume 2* (pp. 179-211): Springer.
- Rosnah, M. Y., Ghazali, A., Wan Rosli, W. D., and Dermawan, Y. M. (2010). Influence of alkaline peroxide treatment duration on the pulpability of oil palm empty fruit bunch. *World Applied Sciences Journal*, 8(2), 185-192.
- Sabrina, B., Reda, D. M., & Houria, E. B. (2012). Induction of antioxidant enzyme system by a nitrogen fertilizer Npk in wheat Triticum durum. *Advances in Environmental Biology*, 85-89.

- Salvador V. H, Lima R. B, dos Santos W. D, Soares A. R, Böhm P. A. F, and Marchiosi R, (2013). Cinamic Acid Increases Lignin Production and Inhibits Soybean Root Growth. *Plos one*, 8(7).
- Santos, D. W. D., Ferrarese, M. L. L., Nakamura, C. V., Mourão, K. S. M., Mangolin, C. A., & Ferrarese-Filho, O. (2008). Soybean (Glycine max) root lignification induced by ferulic acid. The possible mode of action. *Journal of chemical ecology*, 34(9), 1230-1241.
- Saroa, G., and Lal, R. (2003). Soil restorative effects of mulching on aggregation and carbon sequestration in a Miamian soil in central Ohio. *Land Degradation and Development*, 14(5), 481-493.
- Selim, S. M., Zayed, M. S., Atta, H. M. (2012). Evaluation of phytotoxicity of compost during composting process. *Nature and Science*, 10(2), 69-77.
- Shanker, A. K., Cervantes, C., Loza-Tavera, H., & Avudainayagam, S. (2005). Chromium toxicity in plants. *Environment international*, *31*(5), 739-753.
- Sheil, D., Casson, A., Meijaard, E., Van Noordwjik, M., Gaskell, J., Sunderland-Groves, J., ... & Kanninen, M. (2009). The impacts and opportunities of oil palm in Southeast Asia: What do we know and what do we need to know? (No. CIFOR Occasional Paper no. 51, p. 67p). Center for International Forestry Research (CIFOR), Bogor, Indonesia.
- Shinoj, S., Visvanathan, R., Panigrahi, S., and Kochubabu, M. (2011). Oil palm fiber (OPF) and its composites: A review. *Industrial Crops and Products*, 33(1), 7-22.
- Sikorska, M., Matlawska, I., Glowniak, K., and Zgorka, G. (2000). Qualitative and quantitative analysis of phenolic acids in *Asclepias syriaca*. L. *Acta Poloniae Pharmaceutica*, 57, 69-72.
- Silva, F. V. D., Santos, R. L. D. A. D., Fujiki, T. L., Leite, M. S., & Fileti, A. M. F. (2010). Design of automatic control system for the precipitation of bromelain from the extract of pineapple wastes. *Food Science and Technology* (*Campinas*), 30(4), 1033-1040.
- Singh, R. P., & Agrawal, M. (2009). Use of sewage sludge as fertiliser supplement for Abelmoschus esculentus plants: Physiological, biochemical and growth responses. *International Journal of Environment and Waste Management*, 3(1-2), 91-106.
- Singh, G., KimHuan, L., Leng, T., and Kow, D. L. (1999). *Oil palm and the environment: a Malaysian perspective*: Malaysian Oil Palm Growers' Council.
- Singh, H. P., Batish, D. R., and Kohli, R. K. (2001). Allelopathy in agroecosystems: an overview. *Journal of Crop Production*, 4(2), 1-41.

- Singh, H. P., Batish, D. R., Pandher, J. K., and Kohli, R. K. (2005). Phytotoxic effects of *Parthenium hysterophorus* residues on three Brassica species. *Weed Biology* and Management, 5(3), 105-109.
- Singh, R., Embrandiri, A., Ibrahim, M., and Esa, N. (2011). Management of biomass residues generated from palm oil mill: Vermicomposting a sustainable option. *Resources, Conservation and Recycling*, 55(4), 423-434.
- Siqueira, J. O., Nair, M. G., Hammerschmidt, R., and Safir, G. R. (1991). Significance of phenolic compounds in plant-soil-microbial systems. *Critical Review Plant Science*, 10, 63-121.
- Somporn P., Shabbir H. Gheewala., Savitri G., (2004), Environmental evaluation of biodiesel production from palm oil in a life cycle perspective. Thai National News Bureau Public Relation Department Thailand.
- Somrat, K. and Krongkaew, L. 2011. *Renewable Energy from Palm Oil Empty Fruit Bunch The Waste*. Incineration Research Center, Department of Mechanical and Aerospace Engineering, King Mongkut's University of Technology North Bangkok Thailand
- Sosulski, F. (1979). Organoleptic and nutritional effects of phenolic compounds on oilseed protein products: a review. *Journal of the American Oil Chemists'* Society, 56(8), 711-715.
- Sreekala, M., Kumaran, M., and Thomas, S. (1997). Oil palm fibers: Morphology, chemical composition, surface modification, and mechanical properties. *Journal* of Applied Polymer Science, 66(5), 821-835.
- Stalikas, C. D. (2007). Extraction, separation, and detection methods for phenolic acids and flavonoids. *Journal of separation science*, *30*(18), 3268-3295.
- Stichnothe, H., and Schuchardt, F. (2011). Life cycle assessment of two palm oil production systems. *Biomass and Bioenergy*, *35*(9), 3976-3984.
- Suhaimi, M., and Ong, H. K. (2001). Composting empty fruit bunches of oil palm. *Extension Bulletin-Food & Fertilizer Technology Center*, (505), 1-8.
- Sulaiman, F., Abdullah, N., Gerhauser, H., and Shariff, A. (2010). A perspective of oil palm and its wastes. *Journal of Physical Science*, *21*(1), 67-77.
- Sulaiman, O., Salim, N., Nordin, N. A., Hashim, R., Ibrahim, M., and Masatoshi, S. (2012). The Potential of Oil Palm Trunk Biomass as An Alternative Source for Compressed Wood. *Bio Resource*.7(2), 2688-2706.
- Sumathi, S., Chai, S. P., and Mohamed, A. R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, 2404-2421.

- Sung, C. T. B., Joo, G. K., Chien, L. C., and Seng, S. T. (2010). Short-term changes in the soilphysical and chemical properties due to different soil and water conservation practices ina sloping land oil palm estate. *Pertanika Journal of Tropical Agriculture Science*, 34(1), 41-62.
- Suzuki, M., Tsukamoto, T., Inoue, H., Watanabe, S., Matsuhashi, S., Takahashi, M., and Nishizawa, N. K. (2008). Deoxymugineic acid increases Zn translocation in Zn-deficient rice plants. *Plant Molecular Biology*,66(6), 609-617.
- Tam, N., and Tiquia, S. (1994). Assessing toxicity of spent pig litter using a seed germination technique. *Resources, Conservation and Recycling*, 11(1), 261-274.
- Teoh, C. H. (2010). *Key sustainability issues in the palm oil sector*. A discussion paper for multi-stakeholders consultations (Commissioned by the World Bank Group).
- Tiquia, S., Tam, N., and Hodgkiss, I. (1996). Microbial activities during composting of spent pig-manure sawdust litter at different moisture contents. *Bioresource Technology*, 55(3), 201-206.
- Trautman, T. D. (2001). Risk communication—the perceptions and realities. Food Additives and Contaminants, 18(12), 1130-1134.
- Trifonova, R., Postma, J., Verstappen, F. W., Bouwmeester, H. J., Ketelaars, J. J., and van Elsas, J.D. (2008). Removal of phytotoxic compounds from torrefied grass fibres by plant beneficial microorganisms. *FEMS Microbiology Ecology*, 66(1), 158-166
- Tuck, C. O., Pérez, E., Horváth, I. T., Sheldon, R. A., and Poliakoff, M. (2012). Valorization of biomass: deriving more value from waste. *Science*, 337(6095), 695-699.
- Turk, M. A., and Tawaha, A. M. (2002). Allelopathic effect of black mustard (*Brassica nigra* L.) on germination and growth of wild oat (*Avena fatua* L.).Crop Protection, 22(4), 673-677.
- Vairappan, C. S., and Yen, A. M. (2008). Palm oil mill effluent (POME) cultured marine microalgae as supplementary diet for rotifer culture. *Journal of Applied Phycology*, 20(5), 603-608.
- Vaughan, D., and Ord, B. (1990). Influence of phenolic acids on morphological changes in roots of *Pisum sativum*. Journal of the Science of Food and Agriculture, 52(3), 289-299.
- Wagner, H., & Bladt, S. (1996). *Plant drug analysis: a thin layer chromatography atlas*. Springer Science & Business Media.
- Waller, G. R., Feng M-C, Fujii Y (1999) Biochemical analysis of allelopathic compounds: plants, microorganisms, and soil secondary metabolites. In: Inderjit, Daskshini KMM, Foy CL (eds)

- Wang, W. (1985). The use of plant seeds in toxicity tests of phenolic compounds. *Environment International*, 11(1), 49-55.
- Wang, X., Dong, Y., Han, S., and Wang, L. (2000). Structure-phytotoxicity relationship: comparative inhibition of selected nitrogen-containing aromatics to root elongation of *Cucumis sativus.Bulletin of Environmental Contamination* and Toxicology, 65(4), 435-442.
- Wang, H., Wang, Y., and Yang, Y. (2011). Effects of exogenous phenolic acids on roots of poplar hydroponic cuttings. In*Remote Sensing, Environment and Transportation Engineering (RSETE), 2011 International Conference on*(pp. 8245-8249).
- Waters, E. R., and Blum, U. (1987). Effects of single and multiple exposures of ferulic acid on the vegetative and reproductive growth of *Phaseolus vulgaris* BBL-290. Am. *Journal Botany*, 74, 1635-1645.
- Weidner, S., Amarowicz, R., Karamać, M., & Dąbrowski, G. (1999). Phenolic acids in caryopses of two cultivars of wheat, rye and triticale that display different resistance to pre-harvest sprouting. *European Food Research and Technology*, 210(2), 109-113.
- Weigel, H. J., and Jäger, H. J. (1980). Subcellular distribution and chemical form of cadmium in bean plants. *Plant Physiology*, 65(3), 480-482.
- Wong, M. (1985). Phytotoxicity of refuse compost during the process of maturation. Environmental Pollution Series A, Ecological and Biological, 37(2), 159-174.
- Wong, M. H., & Chu, L. M. (1985). The responses of edible crops treated with extracts of refuse compost of different ages. *Agricultural Wastes*, 14(1), 63-74.
- World bank. (2013). Agricultural land (% of land area). Retrieve on Sept 2015 <u>http://data.worldbank.org/indicator/AG.LND.AGRI.ZS</u>.
- Xuan, T. D., Shinkichi, T., Khanh, T. D., & Chung, I. M. (2004). Biological control of weeds and plant pathogens in paddy rice by exploiting plant allelopathy: an overview. *Crop Protection*, 24(3), 197-206.
- Yacob, S., Hassan, M. A., Yoshihito Shirai a, Wakisaka, M., and Subash, S. (2005). Baseline study of methane emission from open digesting tanks of palm oil mill effluent treatment. *Chemosphere*, 59, 1575–1581.
- Yamada, H., Tanaka, R., Sulaiman, O., Hashim, R., Hamid, Z., Yahya, M. and Nirasawa, S. (2010). Old oil palm trunk: a promising source of sugars for bioethanol production. *Biomass and Bioenergy*, 34(11), 1608-1613.
- Yruela, I. (2005). Copper in plants. *Brazilian Journal of Plant Physiology*,17(1), 145-156.
- Yusoff, S. (2006). Renewable energy from palm oil–innovation on effective utilization of waste. *Journal of cleaner production*, *14*(1), 87-93.

- Yusoff, M., Zuhri, M., Salit, M. S., Ismail, N., and Wirawan, R. (2010). Mechanical properties of short random oil palm fibre reinforced epoxy composites. *Sains Malaysiana*, 39(1), 87-92.
- Zain AZ, Koh Mok Poh, Mohd. Yusoff MN, Khoo Kean Choon and Mohd. Nasir N. (1994). Commercial Utilization of Oil Palm Empty Fruit Bunches. Processing of the Third National Seminar on Utilisation of Oil Palm Tree and Other Palms, Forest Research Institute Malaysia, Kuala Lumpur.
- Zheng, W., & Wang, S. Y. (2001). Antioxidant activity and phenolic compounds in selected herbs. *Journal of Agricultural and Food chemistry*,49(11), 5165-5170.
- Zhang, B. J., Yang, W. P., & Wang, Y. Y. (2001). Studies on the dynamic change of soil water of dibbling wheat in film-mulched field. *Journal of Northwest SCI-TECH University of Agriculture and Forestry*, 29, 70-73.
- Zhang, S., Li, P., Yang, X., Wang, Z., and Chen, X. (2011). Effects of tillage and plastic mulch on soil water, growth and yield of spring-sown maize. *Soil and Tillage Research*, 112(1), 92-97.
- Zucconi, F., Forte, M., Monaco, A. and De Bertoldi, M. 1981. Biological evaluation of compost maturity. *BioCycle*, 22(4):27-29
- Zucconi, F., Monaco, A. and Forte, M. (1985) Phytotoxins during the stabilisation of organicmatter. In Composting of Agricultural and Other Wastes, ed. J. K. R. Gasser, Elsevier, New York, pp. 73-86.
- Zucconi, F., and Bertoldi, M. D. (1987). Organic Waste Stabilization Throughout Composting and Its Compatibility with Agricultural Uses. *Global Bioconversions*, 109-137.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : ____

TITLE OF THESIS / PROJECT REPORT :

PHYTOTOXICITY OF OIL PALM RESIDUES USED AS MULCH FOR LETTUCE, TOMATO, CUCUMBER AND AMARANTH SEEDLINGS

NAME OF STUDENT: MOHD ZAKWAN BIN ZAMRI

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (v)



(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from_		_ until _	
	(date)		(date)

Approved by:

(Signature of Student) New IC No/ Passport No.: (Signature of Chairman of Supervisory Committee) Name:

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]