



UNIVERSITI PUTRA MALAYSIA

***INVESTIGATION OF MELANOGENESIS USING NEONATAL NORMAL
HUMAN EPIDERMAL MELANOCYTE TREATED WITH MAWA YOUNG
COCONUT WATER***

SARA ANSARI

FPSK(m) 2015 65



**INVESTIGATION OF MELANOGENESIS USING NEONATAL NORMAL
HUMAN EPIDERMAL MELANOCYTE TREATED WITH MAWA
YOUNG COCONUT WATER**

By

SARA ANSARI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Master of Science**

June 2015

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

**INVESTIGATION OF MELANOGENESIS USING NEONATAL NORMAL
HUMAN EPIDERMAL MELANOCYTE TREATED WITH MAWA
YOUNG COCONUT WATER**

By

SARA ANSARI

June 2015

Chair : Professor Fauziah Othman, PhD
Faculty : Medicine and Health Sciences

The actual color of skin is determined by the type, size and amount of melanin synthesized by melanocytes, and also deposition pattern of melanin in the surrounding keratinocytes. Melanin is synthesized through a series of oxidative reactions and by the enzyme called tyrosinase. Besides tyrosinase, other melanogenic enzymes including tyrosinase related protein-1 (TRP-1) and tyrosinase related protein-2 (TRP-2) also known as dopachrome tautomerase (DCT) are involved in melanin synthesis pathway. Overproduction and aggregation of melanin in the human skin, can cause dark-skinned and also aesthetics problems which encourage researches to develop cosmetic agents with high efficacy and less side effects. Hence, inhibition of tyrosinase activity or melanogenic pathways to have skin lightening are challenging subjects challenge for many researchers. Today, many famous whitening agents such as kojic acid and hydroquinone have been used commercially in whitening creams and other products such as lotion. Due to some adverse effects of these whitening agents such as poor penetration and skin irritation, a natural tyrosinase inhibitor with less harmful side effects, and also low cost producer are always in demand. The objective of this study was to investigate the effect of young coconut water on melanogenesis using neonatal normal human epidermal melanocytes. In this study, cell viability assay was performed to investigate a safe concentration of young coconut water on neonatal normal human epidermal melanocyte. In addition, young coconut water evaluated for *in vitro* cellular tyrosinase activity and melanin content in neonatal normal human epidermal melanocyte. In the present study, the protein levels of tyrosinase and other tyrosinase enzymes including tyrosinase related protein-1 (TRP-1) and tyrosinase related protein-2 (TRP-2) also called dopachrome tautomerase (DCT) which are involved in melanogenesis pathway were determined using western blot method. In this study, skin melanocytes were treated with different concentration of young coconut water and compared with untreated cells. The result from MTT assay showed that young coconut water exhibited no cytotoxicity on melanocytes at 10 mg/ml and half-maximal cytotoxicity concentration (CC_{50}) was 13.12 mg/ml. This study indicated that young coconut water reduced the tyrosinase activity by inhibition of its activity with an IC_{50} (half-maximal inhibitory concentration) value of 10 mg/ml and also down regulated the protein level of tyrosinase. Results from western blot demonstrated that the protein level of tyrosinase related protein-2 significantly decreased at 8 and 10 mg/ml of young

coconut water by 0.351 and 0.280 fold, respectively. There was no significant reduction in protein level of tyrosinase related protein-1 (TRP-1). Although, young coconut water at low concentrations did not significantly reduce tyrosinase activity. In conclusion, young coconut water at 8 and 10 mg/ml obviously reduced the protein level of tyrosinase and tyrosinase related protein-2 (TRP-2) with more than 80% viability. This result indicated that young coconut water might be considered as a potential whitening agent in cosmetics.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**KAJIAN TENTANG MELANOGENESIS MENGGUNAKAN MELANOSIT
EPIDERMIS MANUSIA NORMAL NEONATAL DIRAWAT DENGAN
AIR KELAPA MUDA MALAYSIA**

Oleh

SARA ANSARI

Jun 2015

Pengerusi : Profesor Fauziah Othman, PhD
Fakulti : Perubatan dan Sains Kesihatan

Warna kulit asal ditentukan oleh jenis, saiz dan jumlah melanin disintesis oleh melanosit, dan juga pemendapan corak melanin dalam sekitar keratinosit. Melanin disintesis melalui siri tindak balas pengoksidaan dan oleh enzim dipanggil tirosinase. Selain tirosinase, enzim melanogenik lain termasuk protein tirosinase-1 (TRP-1) dan protein tirosinase-2 (TRP-2) juga dikenali sebagai dopachrome tautomerase (DCT) terlibat dalam laluan sintesis melanin. Pengeluaran berlebihan dan pengagregatan melanin dalam kulit manusia, boleh menyebabkan kulit hitam dan juga masalah estetik yang menggalakkan penyelidik untuk menghasilkan ejen kosmetik dengan keberkesanan yang tinggi dan kesan sampingan yang kurang. Oleh itu, perencatan aktiviti tirosinase atau laluan melanogenik untuk mempunyai kulit yang lebih cerah merupakan satu cabaran bagi ramai penyelidik. Kini, banyak agen pemutih terkenal seperti asid kojik dan hidrokuinon telah digunakan secara komersial dalam krim pemutih dan produk-produk lain seperti losyen. Berikutan beberapa kesan buruk agen pemutihan tesis seperti kurang penembusan dan kerengsaan kulit, perencat tirosinase semula jadi dengan kesan sampingan yang kurang berbahaya, dan juga pengeluar kos rendah adalah sentiasa dalam permintaan. Objektif kajian ini adalah untuk mengkaji kesan air kelapa muda terhadap melanogenesis menggunakan melanosit epidermis manusia normal neonatal. Dalam kajian ini, asai kebolehhidupan sel telah dijalankan untuk mengkaji kepekatan air kelapa muda yang selamat pada melanosit epidermis manusia normal neonatal. Selain itu, air kelapa muda dinilai untuk aktiviti sel tirosinase *in vitro* dan kandungan melanin dalam melanosit epidermis manusia normal neonatal. Dalam kajian ini, tahap protein tirosinase dan enzim tirosinase lain termasuk protein tirosinase-1 (TRP-1) dan protein tirosinase-2 (TRP-2) juga dipanggil dopachrome tautomerase (DCT) yang terlibat dalam laluan melanogenesis ditentukan dengan menggunakan kaedah pemendapan western. Dalam kajian ini, kulit melanosit telah dirawat dengan kepekatan air kelapa muda yang berbeza dan dibanding dengan sel-sel tidak dirawat. Hasil dari asai MTT menunjukkan bahawa air kelapa muda mempamerkan tiada kesitotoksikan pada melanosit di 10 mg / ml dan kepekatan kesitotoksikan setengah maksimal (CC₅₀) ialah 13.12 mg / ml. Kajian Kajian ini menunjukkan bahawa air kelapa muda mengurangkan aktiviti tirosinase oleh merencat aktiviti dengan IC₅₀ (kepekatan setengah maksimal rencatan) nilai 10 mg / ml dan juga turun pengaturan tahap protein tirosinase. Keputusan daripada pemendapan

western menunjukkan bahawa tahap protein tirosinase-2 menurun secara signifikan pada 8 dan 10 mg / ml air kelapa muda masing-masing dengan 0.351 dan 0.280 kali ganda. Tiada pengurangan yang signifikan dalam tahap protein tirosinase-1 (TRP-1). Selain itu, air kelapa muda pada kepekatan yang rendah tidak mengurangkan aktiviti tirosinase secara signifikan. Kesimpulannya, air kelapa muda pada 8 dan 10 mg / ml jelas mengurangkan tahap protein tirosinase dan protein tirosinase-2 (TRP-2) dengan lebih 80% kebolehidupan. Keputusan ini menunjukkan bahawa air kelapa muda mungkin dianggap sebagai agen pemutihan yang berpotensi dalam kosmetik.



ACKNOWLEDGEMENTS

I would like to express my sincere gratitude to my supervisor Prof. Dr. Fauziah Othman for her constant support, guidance and motivation. It would never have been possible for me to take this work to completion without her incredible support and encouragement.

I am heartily offered my regards and blessings to my supervisory member, Dr. Chee Hui Yee whose encouragement, guidance and support from the initial to the final level enabled me to develop an understanding of the subject. I am grateful to the members of Department of Human Anatomy, Department of Medical Microbiology and Parasitology, Department of Immunology, Faculty of Medicine and Health Sciences, University Putra Malaysia for their companionship. I would like to express special thanks to all my labmates and members of the Clinical and Molecular Virology Laboratory and Human Anatomy Laboratory for their kind support during the study.

My sincere appreciation is dedicated to my parents for their immeasurable love and care. They have always encouraged me to explore my potential and pursue my dreams. I need to express my gratitude towards my brothers who made it their priority to cheer me up and ease the pain of separation.

At last I wish to thank my friends especially; Mr. Mohamad Reza Etemadi, thank you for being always supportive and helpful and Mrs. Malahat Rezaei, for her usual help and also those whose names are not mentioned here but this does not mean that I have forgotten their help.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Fauziah Othman, PhD

Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

Chee Hui Yee, PhD

Senior Lecturer

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Sara Ansari GS31157

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of

Chairman of

Supervisory

Committee: Fauziah Othman, PhD

Signature: _____

Name of

Member of

Supervisory

Committee: Chee Hui Yee, PhD

TABLE OF CONTENTS

	ABSTRACT	Page
	<i>ABSTRACT</i>	i
	ACKNOWLEDGEMENTS	iii
	APPROVAL	v
	DECLARATION	vi
	LIST OF TABLES	viii
	LIST OF FIGURS	xii
	LIST OF ABBREVIATIONS	xiii
		xv
	CHAPTER	
1	INTRODUCTION	1
	1.1 Background of Study	1
	1.2 Problem Statement	2
	1.3 Objectives	3
2	LITERATURE REVIEW	4
	2.1 Pigmentation	4
	2.2 Anatomy of Skin	4
	2.2.1 Subcutaneous Tissue	4
	2.2.2 Dermis	4
	2.2.3 Epidermis	5
	2.3 Melanocyte	6
	2.3.1 Melanin	7
	2.3.2 Melanogenesis Regulatory proteins	9
	2.3.3 Melanogenesis Enzymes	10
	2.3.4 Regulation of Gene Expression of Tyrosinase Enzymes	12
	2.4 Melanogenesis Inhibitors	13
	2.4.1 Quinone-related products	13
	2.4.2 Natural and non-hydroquinone agents	13
	2.5 Coconut	14
	2.5.1 Taxonomy of Coconut Palm	14
	2.5.2 Coconut fruit	15
	2.5.3 Coconut Water or Coconut Juice	15
	2.5.4 Coconut as a cosmetics and beauty product	16
	2.5.5 Coconut Water as Medicine	17
3	METHODOLOGY	19
	3.1 Sample Preparation	19
	3.2 Primary Human Cell Culture	19
	3.2.1 Preparation of Medium	19
	3.2.2 Cultivation of Neonatal Normal Human Epidermal Melanocyte	19
	3.2.3 Subculture of Neonatal Normal Human Epidermal Melanocyte	19
	3.3 Cell Cytotoxicity Assay (MTT Assay)	20
	3.4 Assay of Human Tyrosinase Activity	21

3.4.1	Protein Measurement	21
3.4.2	Mushroom Tyrosinase Standard Curve	22
3.5	Enzymatic Assay of Mushroom Tyrosinase in a Cell-Free System	22
3.6	Melanin Content Measurement	22
3.6.1	Synthetic Melanin Standard Curve	23
3.7	SDS-page and Western Blot Analysis	23
3.7.1	Detection of Protein using Enhanced Chemiluminescence Substrate	24
3.8	Statistical Analysis	24
4	RESULTS	27
4.1	Cell Cytotoxicity of Young Coconut Water on Neonatal Normal Human Epidermal Melanocyte	27
4.2	Effect of Young Coconut Water on Human Tyrosinase Activity	30
4.2.1	Quantification of Cellular Tyrosinase Concentration in Treated and Untreated Cells using Mushroom Tyrosinase Standard Curve	32
4.3	Effect of Young Coconut Water on Mushroom Tyrosinase Inhibition Activity in a Cell-Free System	34
4.4	Effect of Kojic Acid on Mushroom Tyrosinase Activity	36
4.5	Effect of Young Coconut Water on Melanin Content in Neonatal Normal Human Epidermal Melanocyte	37
4.5.1	Quantification of Melanin Content in treated and untreated samples Using Synthetic Melanin Standard Curve	41
4.5.2	Comparison of MTT Assay, Tyrosinase Activity and Melanin Content	42
4.6	Determination of Protein Concentration	44
4.7	Western Blot Analysis	46
4.7.1	The Effect of Young Coconut Water on Tyrosinase Expression by Western Blotting	46
4.7.2	Western Blot Analysis of the Expression Level of Tyrosinase Related Protein-1 in Neonatal Normal Human Epidermal Melanocyte Treated with Young Coconut Water	49
4.8	The Inhibitory Effect of Young Coconut Water on Expression of Tyrosinase Related Protein-2	52
5	DISCUSSION	56
5.1	Discussion	56
6	CONCLUSION, LIMITATIONS & RECOMMENDATIONS	59
6.1	Conclusion	59
6.2	Limitations and Shortcoming	59
6.3	Recommendations for Future Studies	59
	REFERENCES	60
	BIODATA OF STUDENT	66

LIST OF TABLES

Table	Page
4.1. Cell cytotoxicity assay of neonatal normal human epidermal melanocyte treated with young coconut water	29
4.2. Cellular tyrosinase inhibition activity in neonatal normal human epidermal melanocyte treated with young coconut water	31
4.3. The OD of mushroom tyrosinase at varying unit/ml. All the values are presented as mean \pm SD of triplicate experiments	33
4.4. Mushroom tyrosinase inhibition activity treated with young coconut water in a cell-free system	34
4.5. Mushroom tyrosinase inhibition activity treated with kojic acid in a cell-free system	36
4.6. Comparison of melanin content in different concentrations of young coconut water	38
4.7. The number of cells in control cells and treated samples using hemocytometer	39
4.8. Cell viability of neonatal normal human epidermal melanocyte treated with young coconut water	40
4.9. Measurements of synthetic melanin at 475 nm using ELIZA reader	41
4.10. Comparison of tyrosinase activity and melanin content against cell viability	43
4.11. The OD of Pierce protein standards at 660 nm (n=2)	44
4.12. Quantification of protein concentration in treated and untreated samples	45
4.13. Quantification of tyrosinase band intensity at different concentrations of young coconut water	48
4.14. Quantification of tyrosinase related protein-1 band intensity at different concentrations of young coconut water	51
4.15. Quantification of tyrosinase related protein-2 band intensity at different concentrations of young coconut water	53

LIST OF FIGURES

Figure	Page
2.1. The photomicrograph of the different layers of human skin	5
2.2. Melanogenesis pathway	9
2.3. Tyrosinase molecule structure with functional units	11
2.4. Melanin Synthetic Pathway	12
3.1. Hemocytometer lam under inverted microscope	20
3.2. Flowchart of overall schematic methodology	26
4.1. Phase-contrast micrographs of morphological changes of neonatal normal human epidermal melanocytes treated with young coconut water	28
4.2. Cell cytotoxicity assay of young coconut water on neonatal normal human epidermal melanocyte	30
4.3. Inhibitory effect on cellular tyrosinase activity of young coconut water in neonatal normal human epidermal melanocyte	32
4.4. Mushroom tyrosinase standard curve	33
4.5. Mushroom Tyrosinase inhibition activity curve treated with young coconut water in a cell-free system	35
4.6. Effect of kojic acid on mushroom tyrosinase activity in a cell-free system	37
4.7. Effect of young coconut water on the melanin content of neonatal normal human epidermal melanocyte	38
4.8. Comparison of melanin content against cell viability after treating with young coconut water	40
4.9. Synthetic melanin standard curve within ranges of (0-50 µg/ml)	42
4.10. Comparison of inhibitory effect of young coconut water on cell viability, tyrosinase activity and melanin content	43
4.11. Pierce 660 nm protein standard curve	45
4.12. The effect of young coconut water on expression of tyrosinase in neonatal normal human epidermal melanocyte	47
4.13. Bar chart of the ratio of tyrosinase expression in treated samples/ control cells (Control value taken as one-fold in each sample)	49
4.14. Effect of young coconut water on expression of tyrosinase related protein-1 in neonatal normal human epidermal melanocyte	50
4.15. Bar chart of the ratio of tyrosinase related protein-1 expression in treated samples/ control cells (Control value taken as one-fold in each sample)	51

4.16.	Effect of young coconut on expression level of tyrosinase related protein-2	52
4.17.	Bar chart of the ratio of tyrosinase related protein-2 expression in treated samples/ control cells (Control value taken as one-fold in each sample)	54



LIST OF ABBREVIATIONS

ACTH	Adrenocorticotrophic hormone
APS	Ammonium per sulfate
AHA	α -hydroxyacid
B1	Thiamin
B2	Riboflavin
B3	Niacin or Niacinamide
B5	Pantothenic acid
B6	Pyridoxine
B7	Biotin
B9	Folic acid
BPE	Bovine pituitary extract
CaCl ₂	Calcium chloride
CC ₅₀	Half maximal cell cytotoxicity
DHICA	5, 6-dihydroxyindole-2-carboxylic acid
DMSO	Dimethyl sulfoxide
DTT	Dithiothreitol
DNA	Deoxyribonucleic acid
FBS	Fetal Bovine Serum
FGF	Fibroblast growth factor
rhFGF-B	rh fibroblast growth factor-B
GAs	Gibberellins
GA	Gentamicin sulfate & Amphotericin-B
HQ	Hydroquinone
HepG2	Human hepatoma cell line
IC ₅₀	Half maximal inhibitory concentration
L-DOPA	Dihydroxyphenylalanine
MAWA	Malaysian Red Dwarf” mother & “West African Tall
mT	Milli-Torr
α -MSH	Melanocyte-stimulating hormon
MITF	Microphthalmia-associated Transcription Factor
MAPK	Mitogen-activated protein kinase
MGM-4	Melanocyte growth medium
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NHEM-neo	Neonatal normal human epidermal melanocyte

OD	Optical density
PMA	Phorbol 12-myristate 13-acetate
PBS	Phosphate-buffered saline
PIH	Post Inflammatory Hyperpigmentation
PAR-2	Protease Activated Receptor-2
PAK	Phenylalanine hydroxylase
PKA	Protein kinase A
POMC	Proopiomelanocortin
rh-insulin	Recombinant human insulin
ROS	Reactive Oxygen Species
SDS	Sodium dodecyl sulfate
TYR	Tyrosinase
TRP-1	Tyrosinase related protein-1
TRP-2 (DCT)	Tyrosinase related protein-2 (Dopachrome tautomerase)
TNS	Trypsin neutralizing solution
TBS	Tris-Buffered Saline
TBS-T	Tris-Buffered Saline-Tween
TCW	Tender Coconut Water
TH1	Tyrosine hydroxylase isoenzyme 1
UV	Ultraviolet
UPM	Universiti Putra Malaysia
YCW	Young Coconut Water

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Today, a variety of cosmetics and clinical products are specially used in Asian countries to balance the colour of skin without making undesirable changes on the skin such as irritation or abnormal pigmentation. Since some of whitening agent are expensive and unsafe, the cosmetic factories are looking for safe, efficient and novel cosmetic agents. These products are tested on human skin to examine their effect on the melanogenesis. Hence, to investigate and evaluate the effect of whitening agents, the pigment producing cells such as melanocytes are being used (Costin & Raabe, 2013).

Melanocytes are dendritic cells that located in deepest layer of skin. The colour of the human skin is determined by type, size, amount, and deposition of melanin pigment in the melanocyte cells. The melanogenesis is enhanced by exposure of skin to UV radiation and also by activation of the melanogenesis enzyme, tyrosinase (Momtaz *et al.*, 2008; Gillbro & Olsson, 2011). Tyrosinase is a bifunctional copper- containing enzyme and involves two different catalytic reactions: first, tyrosinase hydroxylates the L-tyrosine to the dihydroxyphenylalanine (L-DOPA) and then converts L-DOPA to dopaquinone. In series of non-enzymatic reactions, dopaquinone is converted to the dopachrome (Baurin *et al.*, 2002). There are two types of melanin pigments: eumelanin (a dark brown-black pigment) and pheomelanin (a light red-yellow pigment). Besides, tyrosinase (Poole *et al.*, 2013), tyrosinase related protein 1 (TRP-1) and tyrosinase related protein 2 (TRP-2) also known as dopachrometaturase (DCT), are involved in melanogenesis. Dopachrome is spontaneously transformed to eumelanin or by TRP-2, enzymatically converted to 5, 6-dihydroxyindole-2-carboxylic acid (DHICA) to form eumelanin. The TRP-1 increases tyrosinase stability and the ratio of eumelanin to pheomelanin. The pheomelanin pathway which is determined by the presence of cysteine, is required to convert dopaquinone to cysteinyl-dopa in order to form pheomelanin. Tyrosinase inhibitors might suppress melanin production in the melanocyte (Gillbro & Olsson, 2011; Ebanks *et al.*, 2009).

Recently, several melanogenesis inhibitors are used in the cosmetic and pharmaceutical companies as skin-whitening agents (Hanamura *et al.*, 2008). Many of skin whitening agents decrease the total melanin production. Nowadays, some of these skin-lightening inhibitors such as kojic acid, arbutin (Maeda & Fukuda, 1991) and hydroquinone (Jimbow *et al.*, 1974) are used in cosmetics and beauty products (Gillbro & Olsson, 2011).

Coconut is the coconut palm fruit botanically known as *Cocos nucifera.*, and grows in tropical regions like Malaysia, Indonesia and India. The coconut water is widely consumed as refreshing beverage in the world especially in tropical areas because it is nutritious and provides health benefits for the body (Yong *et al.*, 2009). It was reported that coconut water has a lightening effect on hyperpigmentation such as melanoma. The study showed that coconut water can reduce melanin production on the mouse melanoma (S91 cell line) by reducing the enzymes involved in melanogenesis pathway such as TRP-2 enzyme (Mahalingam *et al.*, 2009).

In Malaysia “Mawa” is considered as high quality and unique coconut that is used for producing food and beverage product. Mawa coconut is cross-hybrid fruit, originated from “Malaysian Red Dwarf” mother and “West African Tall”. Among natural beverages in tropical regions, coconut water has huge potential commercialization industry (Pau & Chan, 1985). In this study, to examine cytotoxic effect of young coconut water (YCW) on skin cells (melanocytes), different concentrations of the extract were tested on neonatal normal human epidermal melanocytes (NHEM-neo).

1.2 Problem Statement

Since, many skin-whitening agents are used for treating and preventing hyperpigmentation disorders in cosmetics industries, it is important to consider the issue of safety of these lightening-agents on human skin (Kim *et al.*, 2013b). Some depigmenting agents such as hydroquinone (HQ), kojic acid and arbutin are widely used in cosmetic products. It was found that some of these lightening agents such as HQ produces a lot of side effects namely, skin irritation and destruction of melanocytes of the human skin. To avoid the risk of mutagenesis, HQ is prohibited in cosmetics production by European Union and United States. Other skin whitening agents such as kojic acid and arbutin have limited efficacy due to poor skin penetration and instability (Ubeid *et al.*, 2009).

Therefore, it is a main challenge for the cosmetics industries to apply whitening agents with less side effects. Thus, it is necessary to find natural lightening agents with more efficacy and less cytotoxicity effect on human skin. Whitening cosmetics are products with high consumption and daily usage, especially in tropical regions. Hence, the natural, inexpensive and achievable whitening resources are in demand. In this study, the effect of YCW on melanogenesis of human skin cells was undertaken to investigate its whitening effect.

1.3 Objectives

General Objective

- To investigate the effect of young coconut water on melanogenesis using neonatal normal human epidermal melanocyte

Specific Objectives

- To determine the cytotoxicity of young coconut water on neonatal normal human epidermal melanocyte using MTT assay.
- To determine the effect of young coconut water on cellular tyrosinase activity in neonatal normal human epidermal melanocyte.
- To determine melanin content of neonatal normal human epidermal melanocyte treated with young coconut water.
- To access changes in protein level of tyrosinase, tyrosinase related protein-1 and tyrosinase related protein-2 known as dopachrome tautomerase in neonatal normal human epidermal melanocyte treated with young coconut water using western blot technique.

REFERENCES

- Anurag, P. & T. Rajamohan. 2003. Cardioprotective effect of tender coconut water in experimental myocardial infection. *Plant Foods for Human Nutrition*. 58(3): 1-12.
- Aoki, Y., T. Tanigawa, H. Abe & Y. Fujiwara. 2007. Melanogenesis inhibition by an oolong tea extract in b16 mouse melanoma cells and UV-induced skin pigmentation in brownish guinea pigs. *Bioscience, Biotechnology and Biochemistry*. 71(8): 1879-1885.
- Barciszewski, J., F. Massino & B. F. Clark. 2007. Kinetin—a multiactive molecule. *International Journal of Biological Macromolecules*. 40(3): 182-192.
- Baurin, N., E. Arnoult, T. Scior, Q. T. Do. & P. Bernard. 2002. Preliminary screening of some tropical plants for anti-tyrosinase activity. *Journal of Ethnopharmacology*. 82: 155-158.
- Bouwstra, J. A., G. S. Gooris, J. A. van der Spek & W. Bras. 1991. Structural investigations of human stratum corneum by small-angle X-ray scattering. *Journal of Investigative Dermatology*. 97(6): 1005-1012.
- Bullough, W., & Laurence, E. 1960. The control of mitotic activity in mouse skin: dermis and hypodermis. *Experimental Cell Research*. 21(2): 394-405.
- Campbell-Falck, D., T. Thomas, T. M. Falck, N. Tutuo & K. Clem. 2000. The intravenous use of coconut water. *The American Journal of Emergency Medicine*. 18(1): 108-111.
- Cannon, B., & Nedergaard, J. (2008). Developmental biology: neither fat nor flesh. *Nature*. 454(7207): 947-948.
- Chen, J., Z. Sun, Y. Zhang, X. Zeng, C. Qing, J. Liu, *et al.* 2009. Synthesis of gibberellin derivatives with anti-tumor bioactivities. *Bioorganic & Medicinal Chemistry Letters*. 19(18): 5496-5499.
- Cheng, K-T., F-L. Hsu, S-H. Chen, P-K. Hsieh, H-S. Huang, C.-K. Lee, *et al.* 2007. New constituent from *Podocarpus macrophyllus* var. *macrophyllus* shows anti-tyrosinase effect and regulates tyrosinase-related proteins and mRNA in human epidermal melanocytes. *Chemical & Pharmaceutical Bulletin*. 55(5): 757-761.
- Choi, Y. K., Y. K. Rho, K. H. Yoo, Y. Y. Lim, K. Li, B. J. Kim, *et al.* 2010. Effects of vitamin C vs. multivitamin on melanogenesis: comparative study *in vitro* and *in vivo*. *International Journal of Dermatology*. 49(2): 218-226.
- Costin, G.-E. & V. J. Hearing. 2007. Human skin pigmentation: melanocytes modulate skin color in response to stress. *The FASEB Journal*. 21(4): 976-994.

- Costin, G.-E. & H. Raabe. 2013. Optimized *in vitro* pigmentation screening assay using a reconstructed three dimensional human skin model. *Romanian Journal of Biochemistry*. 50(1): 15-27.
- Crotty, B. A., A. Johnson & A. P. Znaiden. 1998. Vitamin C delivery system, Google Patents.
- DebMandal, M. & S. Mandal. 2011. Coconut (*Cocos nucifera* L.: Arecaceae): In health promotion and disease prevention. *Asian Pacific Journal of Tropical Medicine*. 4(3): 241-247.
- Del Marmol, V. & F. Beermann. 1996. Tyrosinase and related proteins in mammalian pigmentation. *FEBS Letters*. 381(3): 165-168.
- Ebanks, J. P., R. R. Wickett & R. E. Boissy. 2009. Mechanisms regulating skin pigmentation: the rise and fall of complexion coloration. *International Journal of Molecular Sciences*. 10(9): 4066-4087.
- Effiong, G., P. Ebong, E. Eyong, A. Uwah & U. Ekong. 2010). Amelioration of chloramphenicol induced toxicity in rats by coconut water. *J App Sci Res*. 6: 331-335.
- Gillbro, J. & M. Olsson. 2011. The melanogenesis and mechanisms of skin-lightening agents—existing and new approaches. *International Journal of Cosmetic Science*. 33(3): 210-221.
- Hakozaki, T., L. Minwalla, J. Zhuang, M. Chhoa, A. Matsubara, K. Miyamoto, *et al.* 2002. The effect of niacinamide on reducing cutaneous pigmentation and suppression of melanosome transfer. *British Journal of Dermatology*. 147(1): 20-31.
- Hanamura, T., E. Uchida & H. Aoki. 2008. Skin-lightening effect of a polyphenol extract from acerola (*Malpighia emarginata* DC.) fruit on UV-induced pigmentation. *Bioscience, Biotechnology, and Biochemistry*. 72(12): 3211-3218.
- Hirobe, T. 1995. Structure and function of melanocytes: microscopic morphology and cell biology of mouse melanocytes in the epidermis and hair follicle. *Histology and Histopathology*. 10(1): 223-237.
- Huang, H-C., S-H. Chiu & T-M. Chang. 2011. Inhibitory effect of [6]-gingerol on melanogenesis in B16F10 melanoma cells and a possible mechanism of action. *Bioscience, Biotechnology, and Biochemistry*. 75(6): 1067-1072.
- Itaya, A., F. Ma, Y. Qi, Y. Matsuda, Y. Zhu, G. Liang, *et al.* 2002. Plasmodesma-mediated selective protein traffic between “symplasmically isolated” cells probed by a viral movement protein. *The Plant Cell Online*. 14(9): 2071-2083.

- Jackson, J. C., A. Gordon, G. Wizzard, K. MacCook & R. Rolle. 2004. Changes in chemical composition of coconut (*Cocos nucifera*) water during maturation of the fruit. *Journal of the Science of Food and Agriculture*. 84(9): 1049-1052.
- Jeon, S., N-H. Kim, B.-S. Koo, J-Y. Kim & A-Y. Lee. 2009. Lotus (*Nelumbo nuficera*) flower essential oil increased melanogenesis in normal human melanocytes. *Experimental & Molecular Medicine*. 41(7): 517-524.
- Jiang, Z., S. Li, Y. Linu, P. Deng, J. Huang & G. He. 2011. Sesamin induces melanogenesis by microphthalmia-associated transcription factor and tyrosinase up-regulation via cAMP signaling pathway. *Acta Biochim Biophys Sin (Shanghai)*. 43(10): 763-770.
- Jimbow, K., H. Obata, M. A. Pathak & T. B. Fitzpatrick. 1974. Mechanism of depigmentation by hydroquinone. *Journal of Investigative Dermatology*. 62(4): 436-449.
- John, A. 2012. Epidermal Pigmentation, Nucleotide Excision Repair and Risk of Skin Cancer. *Journal of Carcinogenesis & Mutagenesis*.
- Kim, S. S., C.-G. Hyun, Y. H. Choi & N. H. Lee. 2013a. Tyrosinase inhibitory activities of the compounds isolated from *Neolitsea aciculata* (Blume) Koidz. *Journal of Enzyme Inhibition and Medicinal Chemistry*. 28(4): 685-689.
- Kim, S. S., M-J. Kim, Y. H. Choi, B. K. Kim, K. S. Kim, K. J. Park, *et al.* 2013b. Down-regulation of tyrosinase, TRP-1, TRP-2 and MITF expressions by citrus press-cakes in murine B16 F10 melanoma. *Asian Pacific Journal of Tropical Biomedicine*. 3(8): 617-622.
- Kim, Y-J. & H. Uyama. 2005. Tyrosinase inhibitors from natural and synthetic sources: structure, inhibition mechanism and perspective for the future. *Cellular and Molecular Life Sciences CMLS*. 62(15): 1707-1723.
- Kobayashi, T., K. Urabe, A. Winder, C. Jimenez-Cervantes, G. Imokawa, T. Brewington, *et al.* 1994. Tyrosinase related protein 1 (TRP1) functions as a DHICA oxidase in melanin biosynthesis. *The EMBO Journal*. 13(24): 5818.
- Kormos, B., N. Belso, A. Bebes, G. Szabad, S. Bacsá, M. Szell, *et al.* 2011. *In vitro* dedifferentiation of melanocytes from adult epidermis. *PloS One*. 6(2): e17197.
- Leyden, J. & W. Wallo. 2011. The mechanism of action and clinical benefits of soy for the treatment of hyperpigmentation. *International Journal of Dermatology*. 50(4): 470-477.
- Maeda, K. & M. Fukuda. 1991. *In vitro* effectiveness of several whitening cosmetic components in human melanocytes. *J Soc Cosmet Chem*. 42(2): 261-268.

- Makpol, S., N. N. M. Arifin, Z. Ismail, K. Chua, Y. A. M. Yusof & W. Z. W. Ngah. 2009. Modulation of melanin synthesis and its gene expression in skin melanocytes by palm tocotrienol rich fraction. *African Journal of Biochemistry Research*. 3(12): 385-392.
- Masamoto, Y., H. Ando, Y. Murata, Y. Shimoishi, M. Tada & K. Taka and K. Takahata. 2003. Mushroom tyrosinase inhibitory activity of esculetin isolated from seed of *Euphorbia lathyris* L. *Bioscience, Biotechnology, and Biochemistry*. 67(3): 631-634.
- Meredith, P. & T. Sarana. 2006. The physical and chemical properties of eumelanin. *Pigment Cell Research*. 19(6): 572-594.
- Miyamura, Y., S. G. Coelho, R. Wolber, S. A. Miller, K. Wakamatsu, B. Z. Zmudzka, *et al.* 2007. Regulation of human skin pigmentation and responses to ultraviolet radiation. *Pigment Cell Research*. 20(1): 2-13.
- Montaz, S., B. M. Mapunya, P. J. Houghton, C. Edgerly, A. Hussein, S. Naidoo, *et al.* 2008. Tyrosinase inhibition by extracts and constituents of *Sideroxylon inerme* L. stem bark, used in South Africa for skin lightening. *Journal of Ethnopharmacology*. 119(3): 507-512.
- Murphy, K. 2007. From http://commons.wikimedia.org/wiki/File:PPO_figure.jpeg.
- Montagna, W. 1974. The Structure and Function of Skin 3E: *Elsevier*.
- Murisier, F. & F. Beermann. 2006. Genetics of pigment cells, lessons from the tyrosinase gene family.
- Nevin, K. & T. Rajamohan. 2006. Virgin coconut oil supplemented diet increases the antioxidant status in rats. *Food Chemistry*. 99(2): 260-266.
- Norman, A. W. 1998. Sunlight, Season, skin pigmentation, vitamin D, and 25-hydroxyvitamin D: integral components of the vitamin D endocrine system. *The American Journal of Clinical Nutrition*. 67(6): 1108-1110.
- Park, S. H., D. S. Kim, W. G. Kim, I. J. Ryoo, D. H. Lee, C. H. Huh, *et al.* 2004. Terrein: a new melanogenesis inhibitor and its mechanism. *Cellular and Molecular Life Sciences CMLS*. 61(22): 2878-2885.
- Parvez, S., M. Kang, H. S. Chung & H. Bae. 2007. Naturally occurring tyrosinase inhibitors: mechanism and applications in skin health, cosmetics and agriculture industries. *Phytotherapy Research*. 21(9): 805-816.
- Parvez, S., M. Kang, H. S. Chung, C. Cho, M. C. Hong, M. K. Shim, *et al.* 2006. Survey and mechanism of skin depigmenting and lightening agents. *Phytotherapy Research*. 20(11): 921-934.

- Pau, T. Y. & E. Chan. 1985. Optimum density for Mawa hybrid coconuts (PB-121) Oleagineux 1985, 40. 189-195. *Museums_Hybrids, Coconut (EBBD, 190314938)*.
- Poole, D. P., S. Amadesi, N. A. Veldhuis, F. C. Abogadie, T. Lieu, W. Darby, *et al.* 2013. Protease-activated receptor 2 (PAR2) protein and transient receptor potential vanilloid 4 (TRPV4) protein coupling is required for sustained inflammatory signaling. *Journal of Biological Chemistry*. 288(8): 5790-5802.
- Prades, A., M. Dornier, N. Diop & J-P. pain. 2012. Coconut water uses, composition and properties: a review. *Fruits*. 67(2): 87-107.
- Radenahmad, N., F. Saleh, I. Sayoh, K. Sawangjaroen, P. Subhadhirasakul, P. Boonyoung, *et al.* 2012. Young coconut juice can accelerate the healing process of cutaneous wounds. *BMC Complementary and Alternative Medicine*. 12(1): 252.
- Raposo, G. & M. S. Marks. 2007. melanosomes—dark organelles enlighten endosomal membrane transport. *Nature Reviews Molecular Cell Biology*. 8(10): 786-797.
- Régnier, M., C. Tremblaye & R. Schmidt. 2005. Vitamin C affects melanocyte dendricity via keratinocytes. *Pigment Cell Research*. 18(5): 389.
- Riley, P. 1997. Melanin. *The International Journal of Biochemistry & Cell Biology*. 29(11): 1235-1239.
- Saat, M., R. Singh, R. G. Sirisinghe & M. Nawawi. 2002. Rehydration after exercise with fresh young coconut water, carbohydrate-electrolyte beverage and plain water. *Journal of Physiological Anthropology and Applied Human Science*. 21(2): 93-104.
- Seawan, N., S. Koysomboon & K. Chantrapromma. 2011. Anti-tyrosinase and anti-cancer activities of flavonoides from *Blumea blasmifera* DC. *J Med Plants Res*, 5(6): 1018-1025.
- Shibahara, S., K. Takeda, K-I. Yasumoto, T. Uono, K-I. Watanabe, H. Saito, *et al.* 2001. Microphthalmia-associated transcription factor (MITF): multiplicity in structure, function, and regulation. *Journal of Investigative dermatology Symposium Proceedings, Nature Publishing Group*.
- Srivastav, P. & S. Durgaprasad. 2008. Burn wound healing property of *Cocos nucifera*: An appraisal. *Indian Journal of Pharmacology*. 40(4): 144.
- Teulat, B., C. Aldam, R. Trehin, P. Lebrun, J. Barker, G. Arnold, *et al.* 2000. An analysis of genetic diversity in coconut (*Cocos nucifera*) populations from across the geographic range using sequence-tagged microsatellites (SSRs) and AFLPs. *Theoretical and Applied Genetics*. 100(5): 764-771.
- Tsatmali, M., J. Ancans & A. J. Thody. 2002. Melanocyte function and its control by melanocortin peptides. *Journal of Histochemistry & Cytochemistry*. 50(2): 125-133.

- Ubeid, A. A., L. Zhao, Y. Wang & B. M. Hantash. 2009. Short-sequence oligopeptides with inhibitory activity against mushroom and human tyrosinase. *Journal of Investigative Dermatology*. 129(9): 2242-2249.
- Vachtenheim, J. & J. Duchon. 1996. Melanogenic factors regulation of gene expression. *SB Lek*. 97: 41-47.
- Videira, I. F. D. S., D. F. L. Moura & S. Magina. 2013. Mechanisms regulating melanogenesis. *Anais Brasileiros de Dermatologia*. 88(1): 76-83.
- Wang, K.-H., R.-D. Lin, F.-L. Hsu, Y.-H. Huang, H.-C. Chang, C.-Y. Huang, *et al.* 2006. Cosmetic applications of selected traditional Chinese herbal medicines. *Journal of Ethnopharmacology*. 106(3): 353-359.
- Yong, J. W., L. Ge, Y. F. Ng & S. N. Tan. 2009. The chemical composition and biological properties of coconut (*Cocos nucifera* L.) water. *Molecules*. 14(12): 5144-5164.
- Zandi, K., B-T. Teoh, S-S. sam, P-F. Wong, M. R. Mustafa & S. AbuBakar. 2011. Antiviral activity of four types of bioflavonoid against dengue virus type-2. *Virology*. 8-560.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT :

INVESTIGATION OF MELANOGENESIS USING NEONATAL NORMAL HUMAN EPIDERMAL
MELANOCYTE TREATED WITH MAWA YOUNG COCONUT WATER

NAME OF STUDENT : SARA ANSARI

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

☐

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

☐

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

☐

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

☐

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]