

UNIVERSITI PUTRA MALAYSIA

SCREENING AND ISOLATION OF CYTOTOXIC COMPOUNDS FROM LOCAL MARINE AAPTOS SPECIES

KEE CHENG LING.

IB 2006 13



SCREENING AND ISOLATION OF CYTOTOXIC COMPOUNDS FROM LOCAL MARINE AAPTOS SPECIES

By

KEE CHENG LING

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

March 2006

.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

SCREENING AND ISOLATION OF CYTOTOXIC COMPOUNDS FROM LOCAL MARINE AAPTOS SPECIES

By

KEE CHENG LING

March 2006

Chairman: Professor Abdul Manaf Ali, PhD

Institute: Bioscience

Over the recent years, marine sponges have been the target of research as a source of new chemicals with therapeutic potential. They have been proven to have a high strike rate especially for cytotoxic compounds. Thirteen species of marine sponges collected from Pulau Kapas, Pulau Bidong and Pulau Redang were preliminarily identified as *Aaptos* sp. (1), *Xestospongia exigua* (2), unidentified sp. (3), *Xestospongia* sp. (4), *Xestospongia testudinaria* (5), *Callyspongia* sp. (6), *Theonella* sp. (7), *Theonella* sp. (8), *Sigmaducia ambolnenuris* (9), *Ircinia* sp. (10), *Dysedia* sp. (11), unidentified sp. (12) and *Ircinia Halisarca* (13). The crude methanol extracts of these samples were screened for cytotoxic activities against a panel of cell lines, namely HL-60 (promyelocytic leukemia), CEM-SS (T-lymphoblastic leukemia), MCF-7 (breast cancer), HeLa (cervical cancer), HT-29 (colon cancer) and L929 (murine fibrosarcoma from mouse) using a colorimetric tetrazolium (MTT) assay. Crude extracts from 1 and 10 were active against all six cell lines with CD₅₀ values ranging from 1.05 to 24.1 μ g/ml whereas extracts 2, 3 and 8 showed activity only against HL-60, CEM-SS and HT-29 with CD₅₀ values ranging from 12.95 to 29.5

ii

µg/ml. Aaptos sp. (1) was chosen for further investigations due to its abundance and strong cytotoxic activity. Bioassay guided isolation and purification of compounds afforded three cytotoxic alkaloids. H19 was identified as the previously isolated aaptamine [1] and the two orange compounds were established as the new aaptaminoid alkaloids, O1 and O2. All three compounds exhibited significant cytotoxic activity against CEM-SS cells with respective CD₅₀ values of 15.0, 5.3 and 6.7 µg/ml. When tested against 3T3 (normal mouse fibroblast), all three compounds displayed weak cytotoxicities. The CD₅₀ of compound O1 and O2 were determined as 21.2 and 21.0 µg/ml respectively. On the other hand, compound H19 did not achieve a CD₅₀ Phase contrast microscopic analysis showed that compound H19, O1 and O2 induced apoptosis in CEM-SS cells. The apoptotic features observed include cell shrinkage, condensation of chromatin material, membrane blebbing and the formation of apoptotic bodies. Due to insufficient quantity of O1 and O2, only H19 was subjected to subsequent evaluation using fluorescence microscopy. These results further supported that H19 induce apoptosis in CEM-SS as exemplified by the morphological changes observed.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENYARINGAN DAN PEMENCILAN SEBATIAN SITOTOKSIK DARIPADA SPESIS MARIN TEMPATAN AAPTOS SPESIES

Oleh

KEE CHENG LING

Mac 2006

Pengerusi: Profesor Abdul Manaf Ali, PhD

Institut: Biosains

Kebelakangan ini, sepan marin telah menjadi tumpuan perhatian para pennyelidik sebagai sumber kajian untuk penemuan sebatian yang berpotensi terapeutik. Sepan marin ini telah terbukti mempunyai kadar penemuan yang tinggi terutamanya sebatian sitotoksik. Tiga belas spesies sepan marin telah dikutip dari Pulau Kapas, Pulau Bidong serta Pulau Redang dan dikenalpasti sebagai *Aaptos* sp. (1), *Xestospongia exigua* (2), sp. tidak dikenalpasti (3), *Xestospongia sp.* (4), *Xestospongia testudinaria* (5), *Callyspongia* sp. (6), *Theonella* sp. (7), *Theonella* sp. (8), *Sigmaducia ambolnenuris* (9), *Ircinia* sp. (10), *Dysedia* sp. (11), sp. tidak dikenalpasti (12) dan *Ircinia halisarca* (13). Tiga belas ekstrak mentah metanol daripada sepan-sepan ini telah disaring untuk aktiviti sitotoksik melalui kaedah kolorimetri tetrazolium (MTT) ke atas satu panel sel kanser, antaranya HL-60 (leukemia promielositik), CEM-SS (leukemia T-limfoblastik), MCF-7 (kanser payu dara), HeLa (kanser serviks), HT-29 (kanser kolon) dan L929 (fibrosarkoma murin dari tikus). Ekstrak mentah 1 dan 10 telah menunjukkan aktiviti sitotoksik ke atas keenam-enam jenis sel dengan CD₅₀ daripada 1.05 hingga 24.1 µg/ml manakala



ekstrak mentah 2, 3 dan 8 menunjukkan aktiviti sitotoksik hanya ke atas HL-60, CEM-SS dan HT-29 dengan nilai CD₅₀ antara 12.95 dengan 29.5 µg/ml. Aaptos sp. telah dipilih untuk kajian seterusnya disebabkan aktiviti sitotoksiknya yang tinggi dan juga kuantitinya yang mencukupi. Teknik pengasingan biocerakinan berpandu telah menghasilkan tiga sebatian alkaloid. H19 telah dipencilkan sebagai aaptamina [1] yang pernah dipencilkan dahulu dan dua lagi sebatian jingga dikenalpasti sebagai alkaloid aaptaminoid yang baru iaitu O1 dan O2. Ketiga-tiga sebatian ini telah menunjukkan sifat ketoksikan yang kuat terhadap sel CEM-SS masing-masing dengan nilai CD₅₀ 15.0, 5.3 dan 6.7 µg/ml. Apabila dikaji pada sel 3T3 (fibroblast tikus biasa), ketiga-tiga sebatian ini mempamerkan aktiviti sitotoksik yang lemah. Sebatian O1 dan O2 telah memberikan nilai CD50 21.2 dan 21.0 µg/ml manakala sebatian H19 tidak mencapai satu nilai CD₅₀. Analisis daripada mikroskopi fasa kontras menunjukkan bahawa ketiga-tiga sebatian H19, O1 dan O2, memberi kesan aruhan apoptosis pada sel CEM-SS yang dikaji. Morfologi apoptosis yang diperhatikan termasuk pengecutan sel, kondensasi kromatin serta penghasilan tompok-tompok membran dan jasad-jasad apoptotik. Oleh sebab kekurangan sebatian O1 dan O2, hanya H19 telah dikaji seterusnya dengan menggunakan Pemerhatian ke atas penukaran morfologi sel telah mikrosop fluoresen. membuktikan lagi bahawa H19 memberi kesan aruhan apoptosis ke atas sel CEM-SS.



ACKNOWLEDGEMENTS

First and foremost, I would like to express my deepest gratitude to my supervisor, Professor Dr. Abdul Manaf Ali, and co-supervisors, Professor Dr. Md. Nordin Haji Lajis, Associate Professor Dr. Khozirah Shaari and Professor Dr. Faizah Shaharom for their invaluable guidance, advice, constructive comments and encouragement throughout the course of my study.

Special thanks goes to Encik Jasnizat and his colleagues from University College of Science and Technology Malaysia (KUSTEM) for their kind assistance and warm hospitality during my sampling trip at the beautiful islands of Terengganu. I had a wonderful time there.

To all my friends at the Laboratory of Phytomedicine (now Laboratory of Natural Products), especially Kak Rohaya, Faridah, Suryati, Kak Habibah and Shahrin, thank you for making me feel welcomed when I first joined the lab to do my chemistry work. Not to forget the ever-friendly science officer Din, thank you for helping me run my samples. My appreciation also extends to all my seniors, Tony, Yang Mooi, Boon Keat and Madihah for their guidance and encouragement. Also to all my friends back at the lab, Aya, Kak Asmah, Kak Izan and Aida to name a few, you have made life in the lab more colourful and lively.

Above all, I would like to express my utmost gratitude and appreciation to my beloved father, mother and brother, thank you for your everlasting love and support.

vi



TABLE OF CONTENTS

ABSTRACT	ii
ABSTRAK	iv
ACKNOWLEDGEMENTS	vi
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvii

CHAPTER

1	INT	RODUCTION	1
2	LIT	ERATURE REVIEW	4
	2.1	Natural Product	4
	2.2	Marine Natural Product	5
		2.2.1 Marine Environment – The Deep Sea	7
		2.2.2 Marine Environment as a Rich Source of	
		Biodiversity	7
		2.2.3 Bioactive Compounds from Marine Resources	11
		2.2.4 Cytotoxic and Antitumour Compounds	12
		2.2.5 Marine Natural Products in Clinical	
		Development	25
		2.2.6 Some Commercially Available Marine	35
		Bioproducts	
	2.3	Sponges	36
		2.3.1 What are sponges?	36
		2.3.2 History and Taxonomy of Marine Sponges	36
		2.3.3 Sponges – Target of Natural Product Scientists	
		2.3.4 The Genus <i>Aaptos</i> and The Aaptamine Family	37
	2.4	Cell Death	41
	2.5	Necrosis and Apoptosis	41
	2.6	Apoptosis – Target for Cancer Therapy	45
3	MAT	FERIALS AND METHODS	47
	3.1	Screening of Extracts	47
		3.1.1 Animal Material	47
		3.1.2 Sample Preparation and Extraction	47
		3.1.3 Cell Lines Maintenance	48
	3.2	Microtitration Cytotoxic Assay (MTT Assay)	50
	3.3	Morphology Assessment	51
		3.3.1 Phase Contrast Microscopy	51

3.3.2Fluorescent Microscopy (Acridine Orange (AO)
and Propidium Iodide (PI) staining)51

х

3.4 Is	olation of Cytotoxic Compounds from sponge,	
A	aptos sp.	52
3	4.1 General Instrumentation	52
3	4.2 Animal Material	53
3	4.3 Extraction and Fractionation of Crude Extract	
	Aaptos sp.	53
3	4.4 Isolation of Alkaloids from the Chloroform	
	Fraction	56
RESUL	IS AND DISCUSSION	59
4.1 S	creening of Cytotoxicity from Malaysian sponges	59
4.2 Is	olation and Structural Characterization of Alkaloids	
fi	om Aaptos sp.	72
4	2.1 Characterization of Compound H19 as	
	Aaptamine [1]	72
4	2.2 Characterization of Compound O1	88
4	2.3 Characterization of Compound O2	104
4.3 C	ytotoxic Activity of H18, H19, O1 and O2	118
4.4 N	Iorphological Assessment of Apoptosis	121
4	4.1 Phase Contrast Microscopy	121
4	4.2 Fluorescence Microscopy (Acidine Orange /	
	Propidium Iodide Staining Analysis)	128
CONCL	USION	135
REFER	ENCES	138
RIODA	TA OF THE AUTHOR	150

.

4

5





LIST OF TABLES

Table		Page
2.1	Numbers of Described Species of Living Organisms (Wilson, 1988)	8
2.2	Marine pharmacology in 1999: compounds with anthelmintic, antibacterial, anticoagulant, antifungal, antimalarial, antiplatelet, antituberculosis and antiviral activities (Mayer and Hanmann, 2002)	13
2.3	Marine pharmacology in 1999: compounds with anti-inflammatory, immunosuppressant and fibrinolytic effects and affecting the cardiovascular, nervous and sympathomimetic systems (Mayer and Hanmann, 2002).	15
2.4	2001–2 Antitumour pharmacology of marine natural products with established mechanisms of action (Mayer, 2004)	17
2.5	2001–2 Antitumour pharmacology of marine natural products with undetermined mechanism of action (Mayor, 2004)	21
2.6	Marine natural products and derivatives in clinical development (Haefner, 2003).	24
2.7	Examples of commercially available marine bioproducts (Pomponi, 1999)	35
2.8	Differential features and significance of necrosis and apoptosis (Wyllie, 1998)	44
3.1	2-fold dilution gradient (modified from Shier, 1991)	51
4.1	The CD_{50} (cytotoxic dose at 50%) of various crude methanol extracts against various cell lines determined using MTT assay.	66
4.2	Proton chemical shifts of H18 and H19 compared with those of aaptamine obtained from literature values.	77
4.3	¹ H (500 MHz) and ¹³ C (125 MHz) NMR data for H19, with short $({}^{1}J)$ and long $({}^{2}J \& {}^{3}J)$ connectivity obtained from gHSQC and gHMBC experiments.	78
4.4	¹ H (500 MHz) and ¹³ C (125 MHz) NMR data for O1, with short (¹ J) and long (² J, ³ J & ⁴ J) connectivity obtained from gHSQC and gHMBC experiments using gHX nanoprobe.	101



- 4.5 ¹H (500 MHz) and ¹³C (125 MHz) NMR data for O2, with short (¹J) 116 and long (²J, ³J & ⁴J) connectivity obtained from gHSQC and gHMBC experiments using gHX nanoprobe.
- 4.6 Suggested guidelines for distinguishing between non-specific 120 cytotoxicity and antineoplastic activity (Wilson, 1986).
- 4.7 Study of CEM-SS cell populations treated with H19 at different 133 concentrations and time course.



Figure		Page
2.1	Distribution of samples with significant cytotoxicity in the NCI's preclinical screen (Garson, 1994)	6
2.2	Cytotoxic ctivities by phylum (Garson, 1994)	38
2.3	Compounds from the aaptamine class.	40
2.4	Morphological manifestations of necrosis (Duvall and Wyllie, 1986).	43
2.5	Morphology of apoptosis (Duvall and Wyllie, 1986).	43
3.1	Extraction and fractionation of Aaptos sp.	55
3.2	Isolation scheme of alkaloids from the chloroform fraction.	57
4.1	Principle of the MTT assay.	59
4.2	Cytotoxicity of crude extract 1 <i>Aaptos</i> sp., 3 Unidentified, 8 <i>Theonella</i> sp. and 10 <i>Ircinia</i> sp. against HL60 cells.	61
4.3	Cytotoxicity of crude extract 1 <i>Aaptos</i> sp., 3 Unidentified and 10 <i>Ircinia</i> sp. against CEM-SS cells.	61
4.4	Cytotoxicity of crude extract 1 <i>Aaptos</i> sp. and 10 <i>Ircinia</i> sp. against HeLa.	62
4.5	Cytotoxicity of crude extract 1 Aaptos sp. 2 Xestospongia exigna, 8 Theonella sp. and 10 Ircinia sp. against HT-29.	62
4.6	Cytotoxicity of crude extract 1 <i>Aaptos</i> sp. and 10 <i>Ircinia</i> sp. against MCF-7.	63
4.7	Cytotoxicity of crude extract 1 Aaptos sp. 2 Xestospongia exigna and 10 Ircinia sp. against L929.	63
4.8	Sponge species with cytotoxic activity.	65
4.9	Morphology changes of various cell lines induced by crude extract 1 Aaptos sp. and crude extract 10 Ircinia sp.	71
4.10	The ESIMS spectrum of H19.	75
4.11	IR spectrum of H19.	76

LIST OF FIGURES

xiv

4.12	¹ H NMR Spectrum of H19 in CD ₃ OD.	79
4.13	COSY Spectrum of H19 in CD ₃ OD.	80
4.14	¹³ C NMR Spectrum of H19 in CD ₃ OD.	81
4.15	HSQC Spectrum of H19 in CD ₃ OD.	82
4.16a	HMBC Spectrum of H19 in CD ₃ OD (Expansion 1).	83
4.16b	HMBC Spectrum of H19 in CD ₃ OD (Expansion 2).	84
4.17	Molecular structure of compound H19 showing the numbering of hydrogens and the selected HMBC correlations.	85
4.18	¹ H NMR Spectrum of H18 in CDCl _{3.}	86
4.19	¹³ C NMR Spectrum of H18 in CDCl ₃	87
4.20a	The ESIMS full spectrum of O1.	91
4.20b	The ESIMS spectrum of O1.	92
4.20c	The ESIMS spectrum of O1.	93
4.21a	¹ H NMR Spectrum of O1 in CDCl ₃	94
4.21b	¹ H NMR Spectrum of O1 in CDCl ₃ (Expansion on region δ 7.0 - δ 7.5)	95
4.22a	HSQC Spectrum of O1 in CDCl ₃ (Expansion 1).	96
4.22b	HSQC Spectrum of O1 in CDCl ₃ (Expansion 2)	97
4.23a	HMBC Spectrum of O1 in CDCl ₃ (Expansion 1).	98
4.23b	HMBC Spectrum of O1 in CDCl ₃ (Expansion 2).	99
4.23c	HMBC Spectrum of O1 in CDCI ₃ (Expansion 3).	100
4.24	Partial structures A and B making up the complete molecular structure for compound O1.	102
4.25	COSY Spectrum of O1 in CDCl ₃ .	103
4.26	The ESIMS spectrum of O2	106
4.27	¹ H NMR Spectrum of O2 in CDCl ₃ .	107

4.27a	Expanded ¹ H NMR Spectrum of O2 in CDCl ₃ .	108
4.28a	COSY Spectrum of O2 in CDCl ₃ (Expansion 1).	109
4.28b	COSY Spectrum of O2 in CDCl ₃ (Expansion 2).	109
4.29a	HSQC Spectrum of O2 in CDCl ₃ (Expansion 1).	110
4.29b	HSQC Spectrum of O2 in CDCl ₃ (Expansion 2).	111
4.30a	Full HMBC Spectrum of O2 in CDC1 ₃ .	112
4.30b	HMBC Spectrum of O2 in CDCl ₃ (Expansion 1).	113
4.30c	HMBC Spectrum of O2 in CDCl ₃ (Expansion 2).	114
4.30d	HMBC Spectrum of O2 in CDCl ₃ (Expansion 3).	115
4.31	Molecular structure of compound O2 showing some selected HMBC correlations.	117
4.32	Cytotoxicity of H18, H19, O1 and O2 against CEM-SS. The respective CD_{50} were 10.9 µg/ml, 15.0 µg/ml, 5.3 µg/ml and 6.7 µg/ml.	119
4.33	Cytotoxicity of H18, H19, O1 and O2 against 3T3. CD_{50} of O1 and O2 were 21.2 µg/ml and 21.0 µg/ml respectively.	119
4.34	Sequence of morphological changes of CEM-SS treated with 30 μ g/ml of O1 (Phase contrast microscopic examination).	122
4.35	Sequence of morphological changes of CEM-SS treated with 30 μ g/ml of O2 (Phase contrast microscopic examination).	123
4.36	Sequence of morphological changes of CEM-SS treated with 30 μ g/ml of H19 (Phase contrast microscopic examination).	124
4.37	Sequence of morphological changes of CEM-SS treated with 15 μ g/ml of H19 (Phase contrast microscopic examination).	126
4.38	Sequence of morphological changes of CEM-SS treated with 30 µg/ml of DOX (Phase contrast microscopic examination)	127
4.39	Apoptotic and necrotic morphology of CEM-SS cells stained with AO/PI.	130
4.40	The percentage of viable, apoptotic and necrotic CEM-SS cells at different time course using AO/PI double staining.	131



LIST OF ABBREVIATIONS

AO	Acridine orange
ATCC	American type culture collection
atm	Atmospheres
CD ₅₀	Cytotoxic dose at 50%
CNS	Central nervous system
COSY	Correlation spectroscopy
DMSO	Dimethyl sulfoxide
DOX	Doxorubicin
EGFR	Epidermal growth factor receptor
EIMS	Electron impact mass spectrometry
ESIMS	Electron spray impact mass spectrometry
FBS	Fetal bovine serum
FTIR	Fourier transform infrared spectroscopy
GC-MS	Gas chromatography mass spectrometry
HMBC	Heteronuclear multiple bond correlation
HPLC	High performance liquid chromatography
HSQC	Heteronuclear single quantum coherence
HSV-1	Herpes simplex virus type 1
IR	Infrared
J	Coupling constant
LCMS	Liquid chromatography mass spectrometry
m/z	Mass-to-charge-ratio
MAPK	Mitogen activated protein kinase



mg	Miligram
ml	Mililiter
MTT	3-(4,5)-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NCI	National cancer institute
NF-ĸB	Nuclear factor kB
nm	Nanometer
NMR	Nuclear magnetic resonance
OD	Optical density
PBS	Phosphate buffered saline
PI	Propidium iodide
РКС	Protein kinase C
rpm	Round per minute
TLC	Thin layer chromatography
TNFa	Tumor necrosis factor
UV	Ultraviolet
VEGF	Vascular endothelial growth factor
%	Percentage
μg	Microgram
λ_{\max}	Wavelength at which maximum absorption occurs in UV
	spectroscopy
° C	Celsius

xviii



CHAPTER 1

INTRODUCTION

For decades, natural products have been the targets of researchers as an important source of new pharmaceutical compounds. "Natural products" are known as organic compounds of natural origin that are unique to one living organism including plants, animals or microorganisms. They are also known as secondary metabolites, which are synthesized by secondary metabolic pathways (Williams *et al.*, 1989). These natural products, be it plant, marine or microorganism origin, are sources that could lead to discovery of new and unique chemical entities with pharmaceutical potential.

The marine environment is a rich source of both biological and chemical diversity. This diversity has been the source of structurally unique chemical compounds for industrial development as pharmaceuticals, cosmetics, nutritional supplements, molecular probes, fine chemicals and agrochemicals (Kijjoa and Sawangwong, 2004). In the last 25 years, natural products derived from marine organisms have been the focus of many investigations. Since the initial discovery of Ara-C by Bergman in 1951, more than 12,000 compounds have been isolated from marine microorganisms, algae, sponges, soft corals, and marine invertebrates such as bryozoans, echinoderms, molluscs and ascidians (Faulkner, 2001).



These marine derived compounds are known to have various bioactivities such as antibacterial, antifungal, antiinflammatory, antiplatelet, antiprotozoa, as well as antiviral activities (Mayer, 2000). Nevertheless, they are also well known for their anticancer or cytotoxic properties (Mayer, 1999). To date, there are approximately 300 patents for marine metabolites and more than 10 of these are in various stages of clinical trials (Faulkner, 2001).

Over the years, it appears that pharmaceutical investigations of marine natural products have tended to focus on the anticancer therapeutics. These efforts have yielded a considerable number of drug candidates, which are in various stages of clinical trials including bryostatin-1, dolastatin-10, ecteinascidin-743, aplidine and kahalalide F. Clinical results from these investigations have proven the compounds to be promising anticancer agents. These results clearly anticipated the potential of the marine ecosystem in cancer therapy. During the last decade about 2500 new metabolites with antiproliferative activity have been reported (Jimeno *et al.*, 2004).

Macroorganisms such as sponges and tunicates are important sources of bioactive marine metabolites. These sessile, soft-bodied marine invertebrates that lack obvious physical defences are prime candidates to possess bioactive metabolites (Faulkner, 2000). A screening program done by the US National Cancer Institute (NCI) has shown that sponges contain the widest range of secondary metabolites and most of these natural products show biological activity, which are often applicable for medical use (Garson, 1994). Therefore, the present study was carried out to evaluate the cytotoxicity of crude extracts and isolated bioactive compounds from sponges,



collected from the east-coastal waters of Malaysia, mainly Pulau Bidong, Pulau Perhentian and Pulau Redang.

The objectives of this study are:

- To screen for cytotoxic compounds from marine sponges using a colorimetric tetrazolium (MTT) assay against various cancer cell lines.
- 2. To isolate the active compounds from selected sponge through bioassay guided fractionation technique.
- 3. To elucidate the structure of the isolated active compounds.
- To study the effect of the isolated compounds on CEM-SS cells in terms of morphological changes and mode of cell death induced by the compounds.



CHAPTER 2

LITERATURE REVIEW

2.1 Natural Product

The term 'natural product' is commonly known as organic compounds derived from natural sources including plants, animals as well as microorganisms. They are also known as secondary metabolites, previously regarded as waste products, which have no apparent function (Verpoorte, 1998) and appear to be non-essential to the organisms that produce them. These secondary metabolites are synthesized by secondary metabolic pathways (Williams *et al.*, 1989) and probably only activated during periods of stress caused by nutritional shortage or predator attack (Mann, 1986).

The role of natural products as a source for remedies has been recognized since ancient times. Natural products have provided mankind essential materials for shelter, furniture, food, clothing, writing and colouring materials, weapons, gifts, and most importantly, for the treatment of numerous diseases. This 'gift' from nature continues to contribute a great deal in the discovery and development of novel bioactive compounds, especially those with medicinal value. According to Martin and Bohlin (2004), natural products have been the most powerful source of drugs and it is also the single most successful strategy for the discovery of new medicines. Over 60% of drugs used today are of natural product origin (Newman, 2003) or are



based on natural product models and the world wide market in these plant-derived drugs alone is currently amounted to US\$22 billion per annum (Jaspars, 2001).

2.2 Marine Natural Product

Historically, plants have served as the major source of medicinally useful natural products, developed from a legacy of folk medicine based on herbal remedies (Carte, 1993; Cordell, 2000). A prominent example of plant-derived drug is paclitaxel (taxol), isolated from the plant, *Taxus brevifolia*. Taxol was first discovered in 1967 and it took 20 years to reach its first real clinical responses observed with ovarian cancer in 1987. Today, it is undoubtedly the best selling anticancer drug, famous for its anticancer activity and used for the effective treatment against refractory breast and ovarian cancer (Mann, 2002).

With regards to bioactive compounds of pharmaceutical potential, how does the marine environment fare in comparison to the more traditional areas such as terrestrial microorganisms and plants? In 1999, Munro and co-workers cited in their paper that, based on a cytotoxic screening done by the US National Cancer Institute (NCI), marine invertebrates were found to have a much higher incidence of significant cytotoxic activity. The data in Figure 2.1 shows that almost 2% of the 6,540 screened marine animals showed cytotoxic activity compared to less than 1% of 18,293 terrestrial plants and over 8000 microorganisms. This clearly indicated that marine organisms are a preferred source for study.





