

UNIVERSITI PUTRA MALAYSIA

DEVELOPMENT OF RICE STARCH-FISH GELATIN EDIBLE FILMS

ROZHIN AHANGARI

FSTM 2015 40



DEVELOPMENT OF RICE STARCH-FISH GELATIN EDIBLE FILMS



By

ROZHIN AHANGARI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

March 2015

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright ©Universiti Putra Malaysia



DEDICATION

Specially dedicated to,

My beloved mother, Shahnaz Ghari Rasi

For her invaluable love, precious support and encouragement from the beginning of my study



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

DEVELOPMENT OF RICE STARCH-FISH GELATIN EDIBLE FILMS

By

ROZHIN AHANGARI

March 2015

Chairman: Professor Jamilah Bt. Bakar, PhD

Faculty: Food Science and Technology

Edible films of protein or carbohydrate have overall suitable mechanical properties but generally exhibit poor barrier properties. The use of a biopolymer like starch can be an interesting because this polymer is relatively cheap, abundant, biodegradable, and edible. The aim of this study was to incorporate fish gelatin and tannic acid (TA) into rice starch based edible films and investigate barrier, physicochemical and antimicrobial inhibition properties towards Escherichia coli, Listeria monocytogenes, Salmonella typhimurium, and Bacillus cereus by disc diffusion test in 25 and 4 °C. Film forming solutions of different concentrations of rice starch to fish gelatin (0 to 4 %) were prepared. The best film (fish gelatin: 1 %) containing sorbitol was incorporated with different concentration of TA (0.15, 0.3, 0.45, and 0.6 %) in acidic and alkaline pH. Film containing 0.45 % TA was used to wrap sausage in order to extend shelf life in chilled storage (4 °C) in compare with industrial package (LDPE) as a control. The addition of fish gelatin has a significant effect (p<0.05), resulting in lower tensile strength (TS), higher elongation at the break (EAB), water vapor permeability (WVP), oxygen permeability (PO₂), and film solubility (FS), but made films slightly darker than rice starch edible film (control). The barrier properties of incorporated TA films were found to improve significantly (p<0.05) as compared to the non-treated. The addition of TA increased TS (10.83-62.68 MPa), decreased the permeability values, solubility and EAB (4.44 to 1.02 g.mm.m⁻².day⁻¹.KPa⁻¹), (0.136 to 0.0025 %), and (27.97 to 16.82 %), respectively. The possible interactions between the two main components were evaluated by X-ray diffractometer (XRD) and Fourier-transform infrared spectroscopy (FTIR). The amide I and II band the most useful peak of the secondary structure of fish gelatin molecule shifted from 1534.54 to 1647.3 cm⁻¹ in the edible films. In the TA incorporated films the characteristic peak of starch at 1536 cm⁻¹ shifted to 1427 cm⁻¹ and the amide peak of fish gelatin at $1642-1646 \text{ cm}^{-1}$ indicating interaction between the amide groups and hydroxyl groups (-OH) of starch, fish gelatin, and TA. An amorphous state was observed in the film instead of the crystalline structure of starch granule, indicating molecular miscibility. The microstructure of the film showed smoother surfaces with decreasing fish gelatin, while increasing the TA content made the film more compact due to the networking introduced by tannic acid as observed in SEM. The comparison of inhibition zone was E. coli>L. monocytogenes>S. typhimurium> B. cereus with the increase of TA concentration. E. coli was more sensitive to the TA than L. monocytogenes, S. typhimurium, and B. cereus. The inhibition zoon in L. monocytogenes, S. typhimurium, and B. cereus in lower concentration of TA and control was not significant. However, in higher concentration of TA, there is no significant difference between inhibition zone of S. typhimurium and B. cereus either in room or in chilled storage. All sausages became drier than commercial wrapped sausages (MC: 69.97-27.55 %) and in relation to water activity; the average initial values were 0.950 and decreased to 0.850 and 0.920 during 4-week storage in sample and control, respectively. The sausages became more reddish (a value: 17.79-23.66) during chilled storage. As water activity decreased significantly (p<0.05), the total count decreased during storage (from 4.82 to 4.15 CFU/g in sample). The thiobarbituric acid reactive substances increased (1.90-4.06 µmol MA equivalent/per µmol PL) as well as control (1.90-5.24 µmol MA equivalent/per µmol PL), however; as compared to control there is lower TBARS after week 2. These results indicate that antimicrobial packaging with the inclusion of tannic acid as a natural antimicrobial could

be considered as an effective technique to extend shelf life to 30 days in compare with control \leq 2 weeks in 4 °C.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah.

PEMBANGUNAN FILEM BOLEH MAKAN DARIPADA KANJI BERAS DAN GELATIN IKAN

Oleh

ROZHIN AHANGARI

Mac 2015

Pengerusi: Professor Jamilah Bt. Bakar, PhD

Fakulti: Sains dan Teknologi Makanan

Filem boleh dimakan yang berasaskan protein dan karbohidrat mempunyai ciri-ciri mekanikal yang baik, namun kurang berkesan sebagai penghadang. Penggunaan biopolimer seperti kanji menjadi perhatian kerana polimer ini lebih murah, sumbernya yang banyak, biodegradasi dan selamat dimakan. Kajian ini bertujuan untuk menghasilkan filem berasaskan kanji beras yang mengabungkan gelatin ikan dan asid tannik (TA). Ciriciri penghadang, fizikal kimia dan kesan filem tersebut terhadap bakteria Escherichia coli, Listeria monocytogenes, Salmonella typhimurium, and Bacillus cereus diuji melalui ujian resapan cakera pada suhu 25 ^oC dan 4 ^oC. Campuran larutan kanji beras dan gelatin ikan pada kadar kepekatan yang berbeza (0-4 %) telah disediakan untuk membuat filem. Formulasi filem terbaik (1% gelatin ikan) yang mengandungi sorbitol kemudiannya dicampurkan dengan TA pada kepekatan yang berbeza (0.15, 0.30, 0.45 dan 0.60 %) pada pH yang berasid dan beralkali. Filem yang mengandungi 0.45 % TA telah digunakan untuk membalut sosej dengan tujuan meningkatkan tempoh penyimpanan di suhu sejuk (4 °C) manakala pembungkus industri LDPE digunakan pada sosej sebagai rujukan kawalan. Penambahan gelatin ikan memberi kesan yang ketara (p < 0.05) dalam merendahkan kekuatan regangan (TS), meningkatkan pemanjangan takat putus (EAB), kebolehtelapan wap air (WVP), kebolehtelapan oksigen (PO₂) dan kebolehlarutan filem tetapi menghasilkan warna yang lebih gelap daripada filem bolehmakan kanji beras (kawalan). Namun, penambahan gelatin ikan menyebabkan filem yang terhasil berwarna lebih gelap. Filem yang mengandungi TA didapati mempunyai ciri-ciri penghadang yang lebih baik (p<0.05) berbanding dengan filem yang tidak mengandungi TA. Penambahan TA meningkatkan TS (daripada 10.83 hingga 62.68 MPa), menurunkan WVP (daripada 4.44 ke 1.02 g.mm.m⁻ ².hari⁻¹.kPa⁻¹), kebolehlarutan (daripada 0.136 ke 0.0025 %) dan EAB (daripada 27.97 ke 16.82 %). Sinar-X diffractometer (XRD) and spektrometer inframerah jelmaan Fourier (FT-IR) telah digunakan untuk mengkaji hubungan intraksi antara dua komponen tersebut. Jalur amida I and amida II yang merupakan struktur sekunder penting bagi molekul gelatin ikan telah berganjak daripada 1534.54 ke 1647.30 sm⁻¹ di dalam filem eksperimen. Sementara filem yang mempunyai bahan TA menunjukkan ganjakan puncak kanji daripada 1536 sm⁻¹ ke 1427 sm⁻¹, dengan puncak amida gelatin ikan pada 1642-1646 sm⁻¹. Keadaan ini menunjukkan wujudnya interaksi antara kumpulan amida dengan kumpulan hidroksil kanji, gelatin ikan dan TA. Bentuk amorfus dapat dilihat di dalam struktur filem berbanding struktur kristal yang menunjukkan kebolehcampuran molekul-molekul. Analisis SEM menunjukkan permukaan mikrostruktur filem semakin licin dengan penurunan kepekatan gelatin ikan. Sementara itu, peningkatan TA menjadikan mikrostruktur filem lebih padat disebabkan oleh rangkaian struktur yang diperkenalkan oleh TA. Kajian antimikrob mendapati peningkatan kepekatan TA menunjukkan zon perencatan yang paling tinggi pada E. coli, diikuti dengan L. monocytogenes, S. typhimurium dan B. cereus. Bakteria E. coli didapati lebih sensitif pada TA berbanding L. monocytogenes, S. typhimurium dan B. cereus. Zon perencatan bakteria L. monocytogenes, S. typhimurium dan B. cereus didapati kurang ketara pada sosej kawalan dan filem yang mengandungi kepekatan TA yang lebih rendah. Tiada perbezaan ketara didapati pada zon perencatan bakteria S. typhimurium dan B. cereus yang disimpan pada suhu bilik atau suhu penyimpanan dingin, walaupun kepekatan TA digunakan pada kadar yang tinggi. Selepas 30 hari tempoh penyimpanan, semua sosej didapati menjadi kering dengan kandungan air menurun daripada 69.97 % ke 27.55

%. Kajian penyimpanan mendapati aktiviti air filem telah menurun daripada 0.950 kepada 0.850 (sampel filem) dan 0.920 (sosej kawalan), masing-masing. Sosej menjadi merah sepanjang penyimpanan pada suhu sejuk dangan peningkatan nilai 'a' daripada 17.79 sehingga 23.66. Jumlah kiraan bakteria menurun daripada 4.82 CFU/g kepada 4.15 CFU/g (sampel filem) masing-masing semasa penyimpanan, memandangkan aktiviti air menurun dengan ketara (p<0.05). Kedua-dua filem (1.90-4.06 µmol MA kesetaraan/per µmolPL) dan sosej kawalan (1.90-5.24 µmol MA equivalent/per µmolPL) menunjukkan peningkatan nilai TBARS. Walaubagaimanapun, filem eksperimen menunjukkan nilai TBARS yang lebih rendah pada minggu kedua berbanding sosej kawalan. Keputusan ini menunjukkan bahawa potensi TA (bahan antibakteria semulajadi) dalam penghasilan pembungkusan antibakteria merupakan kaedah yang berkesan untuk memanjangkan jangka hayat sosej sehingga 30 hari, berbanding sosej kawalan yang hanya mempunya jangka hayat \leq dua minggu pada 4 °C.



ACKNOWLEDGEMENTS

At the outset, I would like to thank God who is the Most Merciful, for all His blessings, without which I wouldn't be able to achieve this feat.

I would like to take this opportunity to express my profound appreciation to my supervisor, Prof. Dr. Jamilah Bt. Bakar from the Department of Food Technology, Faculty of Food Science and Technology, Universiti Putra Malaysia. Her remarkable level of knowledge and support has led me to work confidently and her critical comments and suggestions instructed the study in the right direction. I could not have finished this dissertation without her generous guidance, and patience.

Special thank goes to my co-supervisors Prof. Dr. Russly Abdul Rahman and Assoc. Prof. Dr. Roselina Karim for their supportive supervisions and invaluable advice throughout the entire progress of this project.

I would like to thank my family and friends. My beloved mother (Shahnaz Ghari Rasi) who gave me life and love in the first place; her great trust, dedication and generosity have backed me in my entire life. With special regards to my dear brothers, Mohammad Amin Ahangari and Ehsan Ahangari, who support and encourage me to proceed to completion.

There are many more who deserve to be thanked whose names I may have forgotten to mention, but their priceless help, friendship and advice will always been appreciated.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy.

The members of the Supervisory Committee were as follows:

Jamilah Bt. Bakar, PhD Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Chairman)

Russly Abdul Rahman, PhD Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

Roselina Karim, PhD

Associate Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:	
Name and Matric No.:		

TABLE OF CONTENTS

			Page
ABS	STRAC	CT	i
ABS	STRAF	X	iii
AC	KNOW	/LEDGEMENTS	v
API	PROVA	AL	vi
DEC	CLAR	ATION	viii
LIS	TOF	TABLES	XV
LIS	T OF I	FIGURES	xvii
LIS	T OF A	ABBREVIATIONS R	xx 1
1	INT	RODUCTION	1
	1.1	Food packaging technology	1
	1.2	Developments in food packaging	1
	1.3	New food packaging technologies	1
	1.4	Scope of study	3
	1.5	Objectives of study	3
2	LIT	ERATURE REVIEW	4
	2.1	Edible and biodegradable films	4
		2.1.1 Film-forming materials	5
		2.1.1.1 Plasticizers	7
		2.1.1.2 Additives	7
		2.1.2 Film-forming mechanisms	8
	2.2	Active packaging	9
		2.2.1 Oxygen scavenging	10
		2.2.2 Absorbing and controlling of moisture	10
		2.2.3 CO ₂ -scavengers and emitters	11
		2.2.4 Ethanol generating systems	11
		2.2.5 Antimicrobial active packaging	12

3 EFFECT OF FISH GELATIN ON PROPERTIES OF RICE STARCH EDIBLE FILMS PLASTICIZED WITH GLYCEROL AND SORBITOL 14

3.1	Introduction		1.	4

3.2	Materia	ls and methods	15
	3.2.1	Film formation	15
	3.2.2	Determination of thickness and light absorption	15
	3.2.3	Determination of film solubility	15
	3.2.4	Determination of tensile strength and elongation tests	15
	3.2.5	Determination of water vapor permeability (WVP)	16
	3.2.6	Determination of oxygen permeability	16
	3.2.7	Determination of color	16
	3.2.8	Differential scanning calorimetry (DSC) test	16
	3.2.9	Fourier transform infrared (FTIR) test	17
	3.2.10	X-ray diffraction test	17
	3.2.11	Scanning electron microscopy (SEM) test	17
	3.2.12	Statistical analysis	17
3.3	Results	and discussion	17
	3.3.1	Film observation	17
	3.3.2	Thickness and light absorption	18
	3.3.3	Film solubility	19
	3.3.4	Tensile strength and elongation tests	21
	3.3.5	Water vapor permeability	22
	3.3.6	Oxygen permeability	23
	3.3.7	Color	25
	3.3.8	Differential scanning calorimetry (DSC)	25
	3.3.9	Fourier transform infrared (FTIR)	31
	3.3.10	X-ray diffraction	34
	3.3.11	Scanning electron microscopy (SEM)	36
3.3.12	2 0	Conclusion	39

4PHYSICOCHEMICAL AND ANTIMICROBIAL PROPERTIES OF RICE STARCH-
FISH GELATIN FILM INCORPORATED WITH TANNIC ACID PREPARED IN
ACIDIC/ALKALINE MEDIUM404.1Introduction40

4.2 Materials and methods			41
	4.2.1	Film formation	41
	4.2.2	Determination of thickness and light absorption	41
	4.2.3	Determination of film solubility	41

	4.2.4	Determination of tensile strength and elongation tests	41
	4.2.5	Determination of water vapor permeability (WVP)	41
	4.2.6	Determination of oxygen permeability	42
	4.2.7	Determination of color	42
	4.2.8	Differential scanning calorimetry (DSC) test	42
	4.2.9	Fourier transform infrared (FTIR) test	42
	4.2.10	X-ray diffraction test	42
	4.2.11	Scanning electron microscopy (SEM) test	42
	4.2.12	Determination of biodegradation	42
	4.2.1 <mark>3</mark>	Antimicrobial test	42
	4.2. <mark>1</mark> 4	Statistical analysis	43
4.3	Result	s and discussion	43
	4.3.1	Film observation	43
	4.3.2	Thickness and light absorption	43
	4.3.3	Film solubility	45
	4.3.4	Tensile strength and elongation tests	46
	4.3.5	Water vapor permeability	47
	4.3.6	Oxygen permeability	48
	4.3.7	Color	49
	4.3.8	Differential scanning calorimetry (DSC) analysis	52
	4.3.9	Fourier transform infrared (FTIR)	55
	4.3.10	X-ray diffraction	58
	4.3.11	Scanning electron microscopy (SEM)	59
	4.3.12	Biodegradation	63
	4.3.13	Antimicrobial activity	64
	4.3.14	Conclusion	72
PAC	CKAGIN	G OF SAUSAGE USING RICE STARCH-FISH GELATIN	FILM
INC	ORPOR	ATED WITH TANNIC ACID FOR EXTENDED SHELF LIFE	73
5.1	Introdu	ction	73
5.2	Materia	ls and methods	74
	5.2.1	Film formation	74
	5.2.2	Storage study	74

5.2.3 Visual observation of stored sausage5.2.4 Moisture content

5

74 74

		5.2.5	Water activity	74
		5.2.6	pH	75
		5.2.7	Color	75
		5.2.8	Rancidity test	75
		5.2.9	Head space analysis of packed sausage	75
		5.2.10	Total microbial count test	75
		5.2.11	Statistical analysis	75
	5.3	Results	and discussion	87
		5.3.1	Visual observation	76
		5.3.2	Moisture content and water activity	77
		5.3.3	pH of packed sausage	78
		5.3.4	Color of packed sausage	79
		5.3.5	Rancidity of packed sausage	79
		5.3.6	Headspace analysis of package	80
		5.3.7	Total microbial count	81
		5.3.8	Conclusion	82
6	CO	NCLUSI	ONS AND RECOMMENDATIONS	83
	6.1	Conclus	ions	83
	6.2	Recomm	nendations	84
REFI	EREN	ICES		85
APPE	ENDI	CES		117
BIOD	BIODATA OF STUDENT 128			
LIST	OF I	PUBLIC	ATIONS	129

G

LIST OF TABLES

Table Page
2.1. Materials used for edible films and coatings 6
3.1. Transparency and solubility of rice starch-fish gelatin films containing glycerol and sorbitol as plasticizers 20
3.2. Mechanical properties of rice starch-fish gelatin films plasticized with glycerol or sorbitol 21
3.3. Water vapor permeability (WVP) and Oxygen permeability (PO ₂) of rice starch- fish gelatin films containing glycerol and sorbitol as plasticizers 23
3.4. Effect of fish gelatin and plasticizers on brightness (L), greenness (a), yellowness (b), and the difference of color (ΔE^*) of rice starch-fish gelatin films plasticized glycerol or sorbitol 24
3.5. DSC thermal properties of rice starch-fish gelatin films plasticized with glycerol and sorbitol 25
3.6. X-ray diffraction data of rice starch-fish gelatin films plasticized with glycerol and sorbitol 36
4.1. Transparency and thickness of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker 45
4.2. Solubility of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker 46
4.3. The TS (MPa) and EAB (%) properties of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker 46
4.4. Water vapor permeability (WVP) and oxygen permeability (PO ₂) of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker 48
4.5. Effect of tannic acid as a cross-linker on brightness (L), greenness (a), yellowness (b), and the difference of color (ΔE^*) of rice starch-fish gelatin films plasticized with sorbitol 51
4.6. DSC thermal properties of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker 53
4.7. X-ray diffraction data of plasticized rice starch-fish gelatin edible films containing tannic acid 58
4.8. Effect of antimicrobial film on <i>E. coli</i> in room and chilled temperature (Agar diffusion method photograph) 68
 4.9. Effect of antimicrobial film on <i>L. monocytogenes</i> in room and chilled temperature (Agar diffusion method photograph) 69

- 4.10. Effect of antimicrobial film on *S. typhimurium* in room and chilled temperature (Agar diffusion method photograph) 70
- 4.11. Effect of antimicrobial film on *B. cereus* in room and chilled temperature (Agar diffusion method photograph) 71
- 5.1. Changes in physicochemical attributes of sausage wrapped with rice starch-fish gelatin edible films incorporated with tannic acid during 30-day chilled storage78
- 5.2. Effect of rice starch-fish gelatin edible films incorporated with tannic acid on color of sausage 79



LIST OF FIGURES

Figure	Page	
2.1. Effect of surface tension of film solution on films' adhesion during a film-fo	orming process 5	
2.2. Various ways for modifying the characteristics of edible films and coatings	9	
3.1. Film formation, (A) low plasticizer (containing either < 20 % Glycerol or high plasticizer (containing either >20 % Glycerol or >40% Sorbitol) and edible film (containing either 20 % Glycerol or 40% Sorbitol)	r <40% Sorbitol), (B) (C) present developed 18	
3.2. Thickness of rice starch-fish gelatin films containing glycerol and sorbitol a	s plasticizers 19	
3.3. Transparency of rice starch-fish gelatin films containing glycerol and sorbit	ol as plasticizers 19	
3.4. Effects of fish gelatin concentration on film solubility (FS) of the rice star films containing glycerol and sorbitol as plasticizers	rch-fish gelatin edible 20	
3.5. Water vapor permeability (WVP) of rice starch-fish gelatin films containing as plasticizers	g glycerol and sorbitol 22	
3.6. Oxygen permeability (PO ₂) of rice starch-fish gelatin films containing gly plasticizers	ycerol and sorbitol as 24	
3.7. DSC thermograph for (A) glycerol plasticized rice starch-fish gelatin edible plasticized rice starch-fish gelatin edible films	films and (B) sorbitol 26	
3.8. DSC thermograph for Tg calculation of rice starch-fish gelatin films pla with the fish gelatin of 1 % (A) and 4 % (B)	sticized with glycerol 27	
3.9. DSC thermograph for Tg calculation of rice starch-fish gelatin films plastici the fish gelatin of 1 % (A) and 4 % (B)	zed with sorbitol with 28	
3.10.DSC thermograph for Δ Hm calculation of rice starch-fish gelatin films pla with the fish gelatin of 1 % (A) and 4 % (B)	asticized with glycerol 29	
3.11. DSC thermograph for Δ Hm calculation of rice starch-fish gelatin films pl with the fish gelatin of 1 % (A) and 4 % (B)	asticized with sorbitol 30	
3.12. FTIR spectra of rice starch-fish gelatin films plasticized with glycerol with % (A), 2 % (B), and 4 % (C)	th the fish gelatin of 0 32	
3.13. FTIR spectra of rice starch-fish gelatin films plasticized with sorbitol with % (A), 2 % (B), and 4 % (C)	the fish gelatin of 0 33	
3.14. X-ray diffraction patterns for (A) glycerol plasticized rice starch-fish ge (B) sorbitol plasticized rice starch-fish gelatin edible films	latin edible films and 35	

3.15. Scanning electron microscopy images (500X) of rice starch-fish gelatin films plasticized with glycerol with the fish gelatin of 0 % (A), 1 % (B), 2 % (C), 3 % (D), and 4 % (E)37
3.16. Scanning electron microscopy images (500X) of rice starch-fish gelatin films plasticized with sorbitol with the fish gelatin of 0 % (A), 1 % (B), 2 % (C), 3 % (D), and 4 % (E)39
4.1. Sorbitol plasticized rice starch-fish gelatin film without tannic acid (A) and incorporated with tannic acid (B)43
 4.2. Thickness of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker
4.3. Transparency of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker 44
4.4. Effects of tannic acid ratios on film solubility (FS) of the rice starch-fish gelatin edible films containing sorbitol as plasticizers 45
4.5. Water vapor permeability (WVP) of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker
4.6. Oxygen permeability (PO ₂) of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid as a cross-linker
4.7. DSC thermograph for (A) acidic and (B) alkaline rice starch-fish gelatin incorporated with tannic acid edible films 54
4.8. FTIR spectra of cross-linked rice starch-fish gelatin films plasticized with sorbitol with tannic acid of A (0), B (0.15 %) and C (0.45 %) in acidic type 56
4.9. FTIR spectra of cross-linked rice starch-fish gelatin films plasticized with sorbitol with tannic acid of A (0), B (0.15 %) and C (0.45 %) in alkaline type 57
4.10. X-ray diffraction patterns for (A) acidic and (B) alkaline plasticized rice starch fish gelatin edible films containing tannic acid 59
 4.11. Scanning electron microscopy images (500X) of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid A (Control), B (0.15 %), C (0.30 %), D (0.45 %), and E (0.60 %) in acidic type
4.12. Scanning electron microscopy images (500X) of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid A (Control), B (0.15 %), C (0.30 %), D (0.45 %), and E (0.60 %) in alkaline type 62
4.13. Carbon dioxide evolutions of rice starch-fish gelatin films plasticized with sorbitol containing tannic acid in (A) acidic type and (B) alkaline type
4.14. Agar diffusion methods photograph, inhibition zone of <i>Escherichia coli</i> 66

- 4.15. Agar diffusion methods photograph, inhibition zone of *Listeria monocytogenes*4.16. Agar diffusion methods photograph, inhibition zone of *Salmonella typhimurium*4.17. Agar diffusion methods photograph, inhibition zone of *Bacillus cereus*67
 5.1. Effect of rice starch-fish gelatin edible films incorporated with tannic acid on (a) appearance,
- 5.2. O₂ concentration in the headspace of rice starch-fish gelatin edible films incorporated with tannic

77

81

(b) color, (c) odor, and (d) overall acceptability of sausage

acid

5.3. CO₂ concentration in the headspace of rice starch-fish gelatin edible films incorporated with tannic acid 81



LIST OF ABBREVIATIONS

A&I	Active and intelligent
ANOVA	Analysis of variance
ASTM	American society for testing and materials
ATCC	American type culture collection
a_w	Water activity
B. cereus	Bacillus cereus
СМС	Carboxy methylcellulose
CFU	Colony forming unit
CO ₂	Carbon dioxide
C ₂ H ₄	Ethylene
ΔH_m	Melting enthalpy
DSC	Differential scanning calorimetry
EAB	Elongation at the break
EAS	Electronic article surveillance
E. coli	Escherichia coli
EFSA	European food safety authority
ERH	Equilibrium relative humidity
FS	Film solubility
FTIR	Fourier-transform infrared spectroscopy
GA	Glutaraldehyde
GLY	Glycerol
GRAS	Generally recognized as safe
НРС	Hydroxypropyl cellulose
НРМС	Hydroxypropyl methylcellulose
IMF	Intermediate moisture foods

L. monocytogenes	Listeria monocytogenes
LDPE	Low density polyethylene
MA	Malonaldehyde
MAP	Modified atmosphere packaging
MC	Methylcellulose
O ₂	Oxygen
OTR	Oxygen transmission rate
PA	Proanthocyanidin
PO ₂	Oxygen permeability
PL	Phospholipid
RFID	Radio frequency identification
RH	Relative humidity
S. typhimurium	Salmonella typhimurium
SEM	Scanning electron microscopy
SOR	Sorbitol
SD	Standard deviation
SO ₂	Sulfur dioxide
ТА	Tannic acid
ТВА	Thiobarbituric acid
TBARS	Thiobarbituric acid reactive substances
T _m	Melting temperature
T _g	Transition temperature
TS	Tensile strength
TEP	1, 1, 3, 3-tetraethoxypropane
TSA	Tripton soy agar
TSB	Tripton soy broth
TTI	Time-temperature integrator

UV	Ultra violet
WVP	Water vapor permeability
WVTR	Water vapor transmission rate
XRD	X-ray diffractometer



CHAPTER 1

INTRODUCTION

1.1 Food packaging technology

One of the essential processes to keep quality of food products for storage and transportation is packaging (Kelsey, 1985). It delays quality deterioration and helps distribution and advertising. Also, the basic purposes of packaging are protection, containment, information and convenience (Kelsey, 1985). Good packaging considerably contributes to a business profit in addition to preserve food quality. In the process of delivery, the quality of a food product declines biologically and chemically, beside the more obvious physical and noticeable visual damage. The attribute of the packaged food is directly related to the food and packaging material qualities. Most food products' quality decline because of mass transfer, such as the migration of packaging constituents into the food, absorption of moisture, oxygen permeability, flavor loss, and absorption of unpleasant odor (Debeaufort, Quezada-Gallo, & Voilley, 1998; Kester & Fennema, 1986).

1.2 Developments in food packaging

In recent time, about 150 million tons of plastics are being manufactured annually, all over the world. As a result, it had led to critical environmental pollution because of wasted and non-degradable polymers. The recycling of wasted plastics is limited and plastics cannot be reused forever, that is, wasted plastics are finally set to be burnt or buried in ground (Parra, Lugao, & Ponce, 2004). The continuous growing of consumer demand in convenience and food quality has supported more research in edible films and biopolymers coatings (Rodriguez, Oses, Ziani, & Mate, 2006). Edible films and coatings are useful materials produced these days from edible biopolymers and food-grade additives. Most biopolymers are naturally occurring polymers, either proteins or carbohydrates (Gennadios, Hanna, & Kurth, 1997). Plasticizers and other additives are incorporated with the film-forming biopolymers to improve film physical characteristics or to form extra functionalities (Kester & Fennema, 1986).

The application of edible films and coatings is an easy technique to develop the physical property of the food products (Cuq, Gontard, & Guilbert, 1995). Food products can be protected from oxidation, moisture absorption/desorption, microbial growth, and other chemical reactions (Kester & Fennema, 1986). The most common purposes of edible films and coatings are that they are barriers against oils, gas or vapors, and also they are carriers of active materials such as antioxidants, antimicrobials, colors and flavors (Guilbert & Gontard, 1995; Krochta & De Mulder-Johnston, 1997).

Preservation, convenience and the other basic purposes of packaging are definitely significant, but its disposal should be treated as an important characteristic of packaging improvement. This problem may challenge package development in the future. The food manufacturing uses a large amount of packaging materials. Therefore, any small decrease in the amount of materials used for each package would cause a substantial cost drop, and may decrease solid waste problems. Packaging technology has done an effort to decrease the volume and weight of materials in order to reduce resources and costs (Testin & Vergano, 1990). Another important subject in food packaging is that it should be natural and environmentally friendly. To plan environmentally friendly packaging systems that are more natural, involves partial replacement of synthetic packaging materials with biodegradable or edible materials. Furthermore, a subsequent decline in the application of entire amount of materials and a growth in the amount of biodegradable and recyclable materials will occur.

1.3 New food packaging technologies

Beyond the barrier function and recyclability, there has been more research and progress about the introduction of new functions to food packaging systems. Among these, important new functional packaging systems include active packaging, modified atmosphere packaging (MAP) and edible films/coatings. The increase of new active packaging functionalities has been promising due to technological progresses in food processing and packaging technology, leading to higher standards of rule, improve sanitation, and better health and safety.

Various new determinations have been presented in active packaging technologies, as well as oxygenscavenging and atmosphere control, antimicrobial activity, intelligent functions, edibility, biodegradability etc. Food packaging has achieved further success than its conventional barrier characteristic. Active packaging systems prolong the shelf life of food products by preserving their quality longer and enhance their safety by securing foods from pathogens. The decision to consider active packaging for a food or beverage is usually based on factors normally elaborated in any package selection. These considerations consist of economic improvement, process engineering limitations, convenience in use, environmentally friendly effects, and subordinate effects resulting from some other modification in the processing or packaging.

Antimicrobial packaging, a promising form of active packaging is directly related to food microbial safety, as well as shelf-life extension by preventing the growth of spoilage and/or pathogenic microorganisms. The growth of spoilage microorganisms reduces the food shelf life, while the growth of pathogenic microorganisms endangers public health. Antimicrobial active packaging systems consist of packaging materials, in-package atmospheres and packaged foods, and are able to kill or inhibit microorganisms that cause food-borne illnesses (Han, 2000; Han, 2003).

Quality may consist of different characteristics such as sensory quality, toxicological quality, and microbial quality, based on deterioration factors and determination procedures. Microbial food quality relates to all three sets of factors, since the growth of bacteria produces undesirable odors and serious toxins, changes the color, taste and texture of food, and also decreases the shelf life of products. Due to the most food packaging systems consist of the food products, the headspace atmosphere and the packaging materials. Any of these three elements of food packaging systems could have an antimicrobial component to enhance antimicrobial efficiency. Antimicrobial active packaging can destroy or prevent target microorganisms and it can be made by applying antimicrobial packaging materials and/or antimicrobial agents inside the package space or inside foods (Han, 2000; Han, 2003).

For the consumers' tendency for foods without chemical preservative, food producers are now applying natural antimicrobials to sterilize and/or prolong shelf life of foods. Herbal extracts have multiple natural compounds, and are recognized to have an extensive antimicrobial range against different microorganisms. Various natural antimicrobial agents such as organic acid, enzymes, and plant extracted essential oils incorporated into packaging materials. Organic acids are usually applied as natural antimicrobial agents because their effectiveness is commonly well understood and cost effective. The precise selection of organic acids is necessary to have effective antimicrobial agents.

Edible films and coatings which carry active elements have been introduced as an active food packaging application (Cuq et al., 1995; Han, 2000, and 2001). Actually, edible films and coatings which are used for packaging of food without active constituents is also regarded as an active food packaging, as these films are edible and biodegradable not be present in conventional packaging constituents (Cuq et al., 1995; Han, 2002).

One of the important factors is the solubility and reactivity of the antimicrobial agents and polymers to the solvents. The reactivity relates to the activity loss of the reactive antimicrobial agents, even though the solubility relates to the homogeneous distribution of the agents in the polymeric materials. Thus, the compatibility of antimicrobial agent and packaging material is a critical factor. The pH of the system is also important. Most antimicrobial chemicals modify their activity with different pH. The pH of the packaging system is generally influenced by the pH of the packed foods, and thus consideration of the food composition with the naturally chemical antimicrobial agent is essential, along with consideration of the packaging substantial reaction with the chemical nature of the agents (Han, 2003). Storage and distribution conditions are further important factors, as well as the storage temperature and time. The microbial growth profile, chemical reaction kinetics, and the distribution profile of antimicrobial agents in the food were affected by the time-temperature combination. In order to inhibit microbial growth, storage at the favorable temperature range for microbial growth should be prevented or decreased for the entire period of storage and distribution.

The incorporation of antimicrobial agents can affect the physical and mechanical properties of packaging materials. A considerable amount of the agent may permeate into the packaging material without any decline of its physical and mechanical properties, if the antimicrobial agent is compatible with the packaging components (Han & Floros, 1997). However, further antimicrobial agent that is not capable of being combined with

packaging components will decline physical strength and mechanical properties (Cooksey, Gremmer, & Grower, 2000).

1.4 Scope of study

Plastic polymers are not biodegradable and recycling is energy intensive and leads to critical environmental pollution. The plastic materials can also impose health hazard due to monomer leaching and migration into foods. The food manufacturing uses a large amount of variety of packaging materials (cost) and single edible film has shortcomings such as poor mechanical properties and fewer barriers to moisture. So composite tannic acid incorporated rice starch-fish gelatin edible film is a good modification to overcome above problems. There is no research on this combination for food active packaging application.

1.5 Objectives of study

- I. Using rice starch and fish gelatin to produce edible and biodegradable films with glycerol and sorbitol as plasticizers and evaluate the functional properties
- II. Evaluate the cross-linking capacity of tannic acid by evaluating both the structural improvement and its influence on physicochemical properties
- III. Determine effectiveness of tannic acid incorporated rice starch-fish gelatin films on quality and microbiological characteristics of sausage during refrigerated storage

This thesis consists of three working chapters, rice starch-fish gelatin edible film plasticized with glycerol and sorbitol produced, characterized and its physicochemical properties investigated in first working chapter. In second working chapter, tannic acid were incorporated into the best rice starch-fish gelatin edible film plasticized with sorbitol and then physicochemical properties and inhibition properties against *Escherichia coli* 0157:H7, *Listeria monocytogenes, Salmonella typhimurium*, and *Bacillus cereus* of incorporated tannic acid film were investigated. Best rice starch-fish gelatin edible film plasticized with sorbitol incorporated with tannic acid were applied to wrap sausage and stored in chilled temperature. Quality and microbiological characteristics of sausage during refrigerated storage determined and compared with commercial wrapped sausage in the last working chapter.

REFERENCES

Adams, M. R., & Moss, M. O. (2002). Food microbiology. London: Royal Society Chemistry.

- Aelenei, N., Popa, M. I., Novac, O., Lisa, G., & Balaita, L. (2009). Tannic acid incorporation in chitosan-based microparticles and in vitro controlled release. *Journal of Materials Science: Materials Medicine*, 20, 1095-1102.
- Ahn, J., Grun, I. U., & Mustapha, A. (2004). Antimicrobial and antioxidant activities of natural extracts in vitro and in ground beef. *Journal of Food Protection*, 67, 148-155.
- Akiyama, H., Fujii, K., Yamasaki, O., Oono, T., & Iwatsuki, K. (2001). Antibacterial action of several tannins against Staphylococcus aureus. Journal of Antimicrobial Chemotherapy, 48, 487-491.
- Al-Hassan, A. A., & Norziah, M. H. (2012). Starch-gelatin edible films: Water vapor permeability and mechanical properties as affected by plasticizers. *Food Hydrocolloids*, 26 108-117.
- Al-Hassan, A. A., & Norziah, M. H. (2012). Starch-gelatin edible films: Water vapor permeability and mechanical properties as affected by plasticizers. *Journal of Food Hydrocolloids*, 26 108-117.
- Al-Saidi, G. S., Al-Alawi, A., Rahman, M. S., & Guizani, N. (2012). Fourier transform infrared (FTIR) spectroscopic study of extracted gelatin from shaari (Lithrinus microdon) skin: effects of extraction conditions. *International Food Research Journal*, 19 (3), 1167-1173.
- Ali-Shtayeh, M. S., Yaghmour, M. R., Faidi, Y. R., Salem, K., & Al-Nury, M. A. (1998). Antimicrobial activity of 20 plants used in folkloric medicine in the Palestinian area. *Journal of Ethnopharmacology*, 60, 265-270.
- An, D., Hwang, Y., Cho, S., & Lee, D. (1998). Packaging of fresh curled lettuce and cucumber by using low density polyethylene films impregnated with antimicrobial agents. *Journal of the Korean Society of Food Science and Nutrition*, 274, 675-681.
- AOAC. (2000). Official methods of analysis. Maryland: Association of Official Analytical Chemists International.
- AOAC. (2005). FDA Bacteriological Analytical Manual. Washington, DC: Association of Official Analytical Chemists International.
- Appendini, P., & Hotchkiss, J. H. (2002). Review of antimicrobial food packaging. *Innovative Food Science & Emerging Technologies*, *3*, 113-126.
- Arvanitoyannis, I. (2002). Formation and properties of collagen and gelatinfilms and coatings: In A. Gennadios, Protein-based films and coatings. Boca Raton: CRC Press LCC.
- Arvanitoyannis, I., Psomiadou, E., & Nakayama, A. (1996). Edible films made from sodium caseinate, starches, sugars or glycerol: part 1. *Carbohydrate Polymers*, *31*, 179-192.
- ASTM. (1990). Standard Test Methods for Oxygen Transmission Rate through Dry Packages using a Coulometric Sensor. Designation D3985-81. Annual book of ASTM standards (pp. 1177-1182). Philadelphia: American Society for Testing and Materials.
- ASTM. (1992). Annual Book of ASTM Standards. Philadelphia: American Society for Testing and Materials.
- ASTM. (1995). Standard test method for tensile properties of thin plastic sheeting. Designation 882-95. Annual book of ASTM standards Philadelphia: American Society for Testing and Materials.

- Auras, R., Arroyo, B., & Selke, S. (2009). Production and properties of spin-coated cassava-starch-glycerolbeeswax films. *Starch/Stärke*, 61, 463-471.
- Avena-Bustillos, R. J., Olsen, C. W., Olson, D. A., Chiou, B., Yee, E., & Bechtel, P. J., et al. (2006). Water vapor permeability of mammalian and fish gelatin films. *Journal of Food Science*, *71*(4), 202-207.
- Baldwin, E. A., Nispero-Carriedo, M. O., & Baker, R. A. (1995). Edible coatings for lightly processed fruits and vegetables. *Horticulture Science*, *30*(1), 35-38.
- Baldwin, E. A., Nispero-Carriedo, M. O., Hagenmaier, R. D., & Baker, R. A. (1997). Use of lipids in coatings for food products. *Food Technology*, 51(6), 56-64.
- Banejee, R., Chen, H., & Wu, J. (1996). Milk protein-based edible film mechanical strength changes due to ultrasound process. *Journal of Food Science*, *61*(4), 824-828.
- Bigi, A., Borghi, M., Cojazzi, G., Fichera, A. M., Panzavolta, S., & Roveri, N. (2000). Structural and mechanical properties of cross-linked drawn gelatin films. *Journal of Thermal Anal Calorimetry*, 61, 451-459.
- Bigi, A., Cojazzi, G., Panzavolta, S., Roveri, N., & Rubini, K. (2002). Stabilization of gelatin films by crosslinking with genipin. *Biomaterials*, 23, 4827-4832.
- Bigi, A., Cojazzi, G., Panzavolta, S., Rubini, K., & Roveri, N. (2001). Mechanical and thermal properties of gelatin films at different degrees of glutaraldehyde crosslinking. *Biomaterials*, 22, 763-768.
- Bigi, A., Panzavolta, S., & Rubini, K. (2004). Relationship between triple-helix content and mechanical properties of gelatin films. *Biomaterials*, 25, 5675-5680.
- Bogracheva, T. Y., Cairns, P., & Noel, T. R. (1999). The effect of mutant genes at the r, rb, rug3, rug4, rug5 and lam loci on the granular structure and physico-chemical properties of pea seed starch. *Carbohydrate Polymer, 39*, 303-314.
- Bogracheva, T. Y., Morris, V. J., Ring, S. G., & Hedley, C. L. (1998). The granular structure of C-type pea starch and its role in gelatinization. *Biopolymers*, 45, 323-332.
- Bogracheva, T. Y., Wang, Y. L., Wang, T. L., & Hedley, C. L. (2002). Structural studies of starches with different water contents. *Biopolymers*, 64, 268-281.
- Bourtoom, T., & Chinnan, M. S. (2008a). Preparation and properties of rice starch-chitosan blend biodegradable film. *Food Science and Technology*, 41, 1633-1641.
- Bourtoom, T., & Chinnan, M. S. (2008b). Preparation and properties of rice starch-chitosan blend biodegradable film. *Journal of Food Science and Technology*, 41, 1633-1641.
- Bourtoom, T., Chinnan, M. S., Jantawat, P., & Sanguandeekul, R. (2006). Effect of plasticizer type and concentration on the properties of edible film from water-soluble fish proteins in surimi wash-water. *Food Science and Technology International*, *12*(2), 119-126.
- Bozic, M., Gorgieva, S., & Kokol, V. (2012). Laccase-mediated functionalization of chitosan by caffeic and gallic acids for modulating antioxidant and antimicrobial properties. *Carbohydrate Polymers*, 87(4), 2388-2398.

Briston, J. H. (1988). Plastic films: New York: John Wiley & Sons.

Brody, A., Strupinsky, E., & Kline, L. (2001). Active packaging for food applications. Boca Rotan: CRC press.

Brody, A. L. (2002). Active and intelligent packaging: the saga continues. Food Technology, 56(12), 65-66.

- Cachaldora, A., García, G., Lorenzo, J. M., & García-Fontán, M. C. (2013). Effect of modified atmosphere and vacuum packaging on some quality characteristics and the shelf-life of "morcilla", a typical cooked blood sausage. *Meat Science*, 93 220-225.
- Cao, N., Fu, Y., & He, J. (2006). Mechanical properties of gelatin films cross-linked respectively by ferulic acid and tannin acid. *Food Hydrocolloids*, 21, 575-584.
- Cao, N., Fu, Y., & He, J. (2007). Mechanical properties of gelatin films cross-linked, respectively, by ferulic acid and tannin acid. *Food Hydrocolloids*, 21, 575-584.
- Carvalho, R. A., & Grosso, C. R. F. (2004). Characterization of gelatin-based films modified with transglutaminase, glyoxal and formaldehyde. *Food Hydrocolloids*, 18, 717-726.
- Carvalho, R. A., & Grosso, C. R. F. (2006). Properties of chemically modified gelatin films. *Brazilian Journal* of Chemical Engineering, 23(1), 45-53.
- Cerqueiraa, M. A., Bartolomeu, W. S. S., Teixeira, J. A., & Vicente, A. A. (2012a). Effect of glycerol and corn oil on physicochemical properties of polysaccharidefilms - A comparative study. *Food Hydrocolloids*, 27, 175-184.
- Cerqueiraa, M. A., Bartolomeu, W. S. S., Teixeira, J. A., & Vicente, A. A. (2012b). Effect of glycerol and corn oil on physicochemical properties of polysaccharidefilms - A comparative study. *Journal of Food Hydrocolloids*, 27, 175-184.
- Chambi, H., & Grosso, C. (2006). Edible films produced with gelatin and casein cross-linked with transglutaminase. *Food Research International*, *39*, 458-466.
- Chan, E. W. C., Lim, Y. Y., & Chew, Y. L. (2007). Antioxidant activity of Camellia sinensis leaves and tea from a low land plantation in Malaysia. *Food Chemistry*, 102, 1214-1222.
- Chillo, S., Flores, S., Mastromatteo, M., Conte, A., Gerschenson, L., & Del Nobile, M. A. (2008). Influence of glycerol and chitosan on tapioca starch-based edible film properties. *Journal of Food Engineering*, 88 159–168.
- Chiou, B., Avena-Bustillos, R., Bechtel, P. J., Jafri, H., Narayan, R., Imam, S. H., Glenn, G.M., & Orts, W. J. (2008). Cold water fish gelatin films: Effects of cross-linking on thermal, mechanical, barrier, and biodegradation properties. *European Polymer Journal*, 44, 3748-3753.
- Chung, K. T., Lu, Z., & Chou, M. W. (1998). Mechanism of inhibition of tannic acid and related compounds on the growth of intestinal bacteria. *Food and Chemical Toxicology*, 36, 1053-1060.
- Chung, K. T., Stevens Jr., S. E., Lin, W. F., & Wei, C. I. (1993). Growth inhibition of selected food-borne bacteria by tannic acid, propyl gallate and related compounds. *Letters in Applied Microbiology*, 17, 29-32.
- Chung, K. T., Wong, T. Y., Wei, C. I., Huang, Y. W., & Lin, Y. (1998). Tannins and human health: a review. *Critical Reviews in Food Science and Nutrition, 38*, 421-464.
- Chung, S. K., Cho, S. H., & Lee, D. S. (1998). Modified atmosphere packaging of fresh strawberries by antimicrobial plastic films. *Korean Journal of Food Science and Biotechnology*, 30, 1140-1145.

- Cisneros-Zevallos, L., Saltveit, M. E., & Krochta, J. M. (1997). Hygroscopic coatings control surface white discoloration of peeled (minimally processes) carrots during storage. *Journal of Food Science*, 62(2), 363-366,398.
- Cooksey, D. K., Gremmer, A., & Grower, J. (2000). Characteristics of nisin-containing corn zein pouches for reduction of microbial growth in refrigerated shredded Cheddar cheese. In Book of Abstracts, 2000 Annual Meeting (p. 188). Chicago: Institute of Food Technologists.
- Cuq, B., Gontard, N., Cuq, J. L., & Guilbert, S. (1996). Functional properties of myofibrillar protein based biopackaging as affected by film thickness. *Journal of Food Science*, 61(3), 580-584.
- Cuq, B., Gontard, N., & Guilbert, S. (1995). Edible films and coatings as active layers. In. M. Rooney. Active Food Packaging (pp. 111-142). Glasgow: Blackie.
- Cuq, B., Gontard, N., Cuq, J. L., Guilbert, S. (1997). Selected functional properties of fish myofibrillar proteinbased films as affected by hydrophilic plasticizers. *Journal of Agricultural and Food Chemistry*, 45, 622-626.
- Cuvelier, M. E., Richard, H., & Berset, C. (1992). Comparison of the antioxidative activity of some acidphenols: structure-activity relationship. *Bioscience, Biotechnology, and Biochemistry*, 56, 324-325.
- Daniels, R. (1973). Edible Coatings and Soluble Packaging. Park Ridge: Noyes Data Corporation.
- Dawson, P. L., Acton, J. C., & Ogale, A. A. (2002). *Biopolymer films and potential applications to meat and poultry products*. Paper presented at the Proceedings of the 55th Reciprocal Meat Conference.
- Debeaufort, F., Quezada-Gallo, J. A., & Voilley, A. (1998). Edible films and coatings: tomorrow's packaging: a review. *Critical Review Food Science Nutrition*, 38(2), 299-313.
- DeCarvalho, R. A., & Grosso, C. R. F. (2004). Characterization of gelatin based films modified with transglutaminase, glyoxal and formaldehyde. *Food Hydrocolloids*, 18(5), 717-726.
- Díaz, P., Nieto, G., Garrido, M. D., & Bañón, S. (2008). Microbial, physic-chemical and sensory spoilage during the refrigerated storage of cooked pork loin processed by the sous vide method. *Meat Science*, 80, 287-292.
- Dobias, J., Voldrich, M., Marek, M., & Derovsky, M. (1998). Active packaging-immobilization of preservatives on/in packaging materials. *Lebensmittelchemie*, *52*, 33-36.
- Dong Sun, L. (2005). *Packaging containing natural antimicrobial or antioxidant agants*. In J.H., Han, Innovation in Food Packaging: Elsevier Science & Technology Books.
- Donhowe, I. G., & Fennema, O. (1993). The effects of plasticizers on crystallinity, permeability, and mechanical properties of methylcellulose films. *Journal of Food Processing and Preservation*, 17, 247-257.
- Ellis, R. P., Cochrane, M. P., & Dale, M. F. B. (1998). Starch production and industrial use. *Journal of the Science of Food and Agriculture*, 77, 289-311.
- Emmambuxa, M. N., Stading, M., & Taylor, J. R. N. (2004). Sorghum kafirin film property modification with hydrolysable and condensed tannins. *Journal of Cereal Science*, 40 127-135.
- Escamilla-García, M., Calderón-Domínguez, G., Chanona-Pérez, J. J., Farrera-Rebollo, R. R., Andraca-Adame, J. A., Arzate-Vázquez, I., . . . Moreno-Ruiz, L. A. (2013). Physical and structural characterisation of

zein and chitosan edible films using nanotechnology tools. International Journal of Biological Macromolecules, 61 196-203.

- Eswaranandam, S., Hettiarachchy, N. S., & Johnson, M. G. (2004). Antimicrobial activity of citric, lactic malic, or tartaric acids and nisin-incorporated soy protein film against *Listeria monocytogenes*, *Escherichia coli* O157:H7, and *Salmonella gaminara*. Journal of Food Science, 69, FMS79-FMS84.
- Fakhoury, F. M., Martelli, S. M., Bertan, L. C., Yamashita, F., Mei, L. H. I., & Queiroz, F. P. C. (2012). Edible films made from blends of manioc starch and gelatin eInfluence of different types of plasticizer and different levels of macromolecules on their properties. *Food Science and Technology*, 49 149-154.
- Farhoosh, R., Golmovahhed, G. A., & Khodaparast, M. H. H. (2007). Antioxidant activity of various extracts of old tea leaves and black tea wastes (Camellia sinensis leaves). *Food Chemistry*, *100*, 231-236.
- Floros, J. D., Dock, L. L., & Han, J. H. (1997). Active packaging technologies and applications. *Food Cosmetics and Drug Packaging*, 20, 10-17.
- Fonkwe, L. G., Narsimhan, G., & Cha, A. S. (2003). Characterization of gelation time and texture of gelatin and gelatin-polysaccharide mixed gels. *Food Hydrocolloids*, 17(6),18, 871-883,717-726.
- Galimpin-Johan, S. M. C., Abdul Rahman, R., Bakar, J., Che Man, Y. B., & Rusul, G. (2007). Pasteurization, development and storage of sous vide rendang (spicy beef stew). *Journal of Foodservice*, 18, 251-263.
- Garcia, M. A., Martino, M. N., & Zaritzky, N. E. (2000). Microstructural characterization of plasticized starchbased films. *Starch/Stärke*, 52(4), 118-124.
- Gennadios, A., Ghorpade, V. M., Weller, C. L., & Hanna, M. A. (1996). Heat curing of soy protein films. Biological Systems Engineering, 39(2), 575-579.
- Gennadios, A., Hanna, M. A., & Kurth, L. B. (1997). Application of edible coatings on meats, poultry and seafoods: a review. *Food Science and Technology*, *30*(4), 337-350.
- Gennadios, A., Rhim, J. W., Handa, A., Weller, C. L., & Hanna, M. A. (1998). Ultraviolet radiation affects physical and molecular properties of soy protein films. *Journal of Food Science*, 63(2), 225-228.
- Gennadios, A., & Weller, C. L. (1990). Edible films and coatings from wheat and corn proteins. Food *Technology*, 44(10), 63-69.
- Gomez-Guillen, M. C., Ihl, M., Bifani, V., Silva, A., & Montero, P. (2007). Edible films made from tuna-fish gelatin with antioxidant extracts of two different murta ecotypes leaves (Ugni molinae Turcz). Food Hydrocolloids, 21(7), 1133-1143.
- González, A., Strumia, M. C., & Alvarez Igarzabal, C. I. (2011). Cross-linked soy protein as material for biodegradable films: Synthesis, characterization and biodegradation. *Journal of Food Engineering*, 106 331-338.
- Guan, Y. L., Liu, X. F., Zhang, Y. P., & Yao, K. D. (1998). Study of phase behavior of chitosan/viscose rayon blend film. *Journal of Applied Polymer Science*, 67, 1965-1972.
- Guerrero, P., Stefani, P. M., Ruseckaite, R. A., & de la Caba, K. (2011). Functional properties of films based on soy protein isolate and gelatin processed by compression molding. *Journal of Food Engineering*, 105 65-72.
- Guilbert, S. (2002). *Edible and biodegradable coating/film systems*. In J.H., Han. Active Food Packaging (pp. 4-10). Winnipeg: SCI Publication and Communication Services.

- Guilbert, S., Cuq, B., & Gontard, N. (1997). Recent innovations in edible and/or biodegradable packaging materials. *Food Additives and Contaminants*, 14(6-7), 741-751.
- Guilbert, S., & Gontard, N. (1995). Edible and biodegradable food packaging. In P., Ackermann, M., Jagerstad, & T., Ohlsson. Foods and Packaging Materials-Chemical Interactions (pp. 159-168). Cambridge: The Royal Society of Chemistry.
- Guilbert, S., Gontard, N., & Gorris, L. G. M. (1996). Prolongation of the shelf life of perishable food products using biodegradable films and coatings. *Food Science and Technology*, 29, 10-17.
- Gullen, J. S., & Vaylen, N. E. (1994). 5322701, US patent.
- Han, J., Castell-Perez, M. E., & Moreira, R. G. (2007a). The influence of electron beam irradiation of antimicrobial-coated LDPE/polyamide films on antimicrobial activity and film properties. *Food Science and Technology*, 40, 1545-1554.
- Han, J., Castell-Perez, M. E., & Moreira, R. G. (2007b). The influence of electron beam irradiation of antimicrobial-coated LDPE/polyamide films on antimicrobial activity and film properties. *Journal of LWT*, 40, 1545-1554.
- Han, J., & Floros, J. (1998). Potassium sorbate diffusivity in American processed and mozzarella cheeses. *Journal of Food Science*, 63, 435-437.
- Han, J., & Floros, J. (1998). Simulating diffusion model and determining diffusivity of potassium sorbate through plastics to develop antimicrobial packaging film. *Journal of Food Processing and Preservation*, 222 107-122.
- Han, J. H. (2000). Antimicrobial food packaging. Food Technology, 54(3), 56-65.
- Han, J. H. (2001). *Design of edible and biodegradable films/coatings containing active ingredients*. Paper presented at the Active Biopolymer Films and Coatings for Food and Biotechnological Uses, Seoul, Korea,10th IUFoST World Congress Organization.
- Han, J. H. (2002). *Protein-based edible films and coatings carrying antimicrobial agents*. In A., Gennadios. Protein-based Films and Coatings (pp. 485-499). Boca Raton: CRC Press.
- Han, J. H. (2003). *Antimicrobial food packaging*. In R., Ahvenainen. Novel Food Packaging Techniques (pp. 50-70). Cambridge: Woodhead Publishing Ltd.
- Han, J. H. (2003). Design of antimicrobial packaging systems. International Review of Food Science and Technology, 11, 106-109.
- Han, J. H., & Floros, J. D. (1997). Casting antimicrobial packaging films and measuring their physical properties and antimicrobial activity. *Journal of Plastic Film Sheeting*, 13, 287-229.
- Han, J. H., & Gennadios, A. (2005). *Innovations in food packaging*. In J.H., Han. Edible films and coatings: a review: Elsevier Academic Press.
- Han, J. H., & Krochta, J. M. (1999). Wetting properties and water vapor permeation of whey-protein-coatedpaper. American Society of Agricultural and Biological Engineers, 42(5), 1375-1382.
- Han, J. H., & Krochta, J. M. (2001). Physical properties and oil absorption of whey protein coated paper. *Journal of Food Science*, 66(2), 294-299.

- Henning, S., Metz, R., & Hammes, W. (1986). New aspects for the application of nisin to food products based on its mode of action. *International Journal of Food Microbiology*, 33 135-141.
- Hoffman, K. L., Han, I. Y., & Dawson, P. L. (2001). Antimicrobial effects of corn zein films impregnated with nisin, lauric acid, and EDTA. *Journal of Food Protection*, 64, 885-889.
- Hong, S., Park, J., & Kim, D. (2000). Antimicrobial and physical properties of food packaging films incorporated with some natural compounds. *Food Science and Biotechnology*, 91, 38-42.
- Hotchkiss, J. (1997). Food packaging interactions influencing quality and safety. Food Additives and Contaminants, 14(6-7), 601-607.
- Igene, J. O., & Pearson, A. M. (1979). Role of phospholipids and triglycerides in warmed-over flavor development in meat model systems. *Journal of Food Science*, 44, 1285-1290.
- Igene, J. O., Pearson, A. M., Dugan, L. R., & Price, J. F. (1980). Role of triglycerides and phosphohpids on development of rancidity in model meat systems during frozen storage. *Food Chemistry*, 5, 263-276.
- Jagannath, J. H., Nanjappa, C., Das Gupta, D. K., & Bawa, A. S. (2003). Mechanical and barrier properties of edible starch-protein-based films. *Journal of Applied Polymer Science*, 88, 64-71.
- Jamshidian, M., Arab Tehrany, E., Cleymand, F., Leconte, S., Falher, T., & Desobrya, S. (2012). Effects of synthetic phenolic antioxidants on physical, structural, mechanical and barrier properties of poly lactic acid film. *Journal of Carbohydrate Polymers*, 87 1763-1773.
- Jongjareonrak, A., Benjakul, S., Visessanguan, W., & Tanaka, M. (2006). Effects of plasticizers on the properties of edible films from skin gelatin of big eye snapper and brown stripe red snapper. *European Food Research and Technology*, 222, 229-235.
- Jung, D., Bodyfelt, F., & Daeschel, M. (1992). Influence of fat emulsifiers on the efficacy of nisin in inhibiting Listeria monocytogenes in fluid milk. Journal of Dairy Science, 752 387-393.
- Kelsey, R. J. (1985). Packaging in Today's Society. Lancaster: Technomic Pub Hshing Co.
- Kerry, J. F., O'Grady, M. N., & Hogan, S. A. (2006). Past, current and potential utilisation of active and intelligent packaging systems for meat and muscle-based products: A review. *Meat Science*, 74 113-130.
- Kester, J. J., & Fennema, O. R. (1986). Edible films and coatings: a review. Food Technology, 48(12), 47-59.
- Kim, S., Nimni, M. E., Yang, Z., & Han, B. (2005). Chitosan/gelatin-based films cross-linked by proanthocyanidin. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 75B(2), 442-450.
- Kim, S., Ruengwilysup, C., & Fung, D. Y. C. (2004). Antibacterial effect of water-soluble tea extracts on foodborne pathogens in laboratory medium and ina food model. *Journal of Food Protection*, 67, 2608-2612.
- Kim, S. J., & Ustunol, Z. (2001). Solubility and moisture sorption isotherms of whey-protein-based edible films as influenced by lipid and plasticizer incorporation. *Journal of Agricultural and Food Chemistry*, 49(9), 438-4391.
- Kong, J., & Yu, S. (2007). Fourier transform infrared spectroscopic analysis of protein secondary structures. Acta Biochimica et Biophysica Sinica, 39 (8), 549-559.

- Krochta, J. M. (2002). Proteins as raw materials for films and coatings: definitions, current status, and opportunities. In A., Gennadios. Protein-based Films and Coatings (pp. 1-41). Boca Raton: CRC Press.
- Krochta, J. M., & De Mulder-Johnston, C. (1997). Edible and biodegradable polymer films: challenges and opportunities. *Food Technology*, *51*(2), 61-74.
- Ku, K. J., Hong, Y. H., & Song, K. B. (2008). Mechanical Properties of a Gelidium corneum Edible Film Containing Catechin and Its Application in Sausages. *Food Chemistry*, 73, 217-221.
- Kubota, N., Kikuchi, Y., Mizuhara, Y., Ishihara, T., & Takita, Y. (1993). Solid-phase modification of chitosan hydrogel membranes and permeability properties of modified chitosan membranes. *Journal of Applied Polymer Science*, *50*, 1665-1670.
- Kumosinski, T. F., & Farrell, H. M. (1993). Determination of the global secondary structure of proteins by Fourier transform infrared (FTIR) spectroscopy. *Trends in Food Science & Technology*, *4*, 169.
- Lacroix, M., & Ouattara, B. (2000). Combined industrial processes with irradiation to assure innocuity and preservation of food products a review. *Food Research International*, 33(2), 719-724.
- Laohakunjit, N., & Noomhorm, A. (2004a). Effect of Plasticizers on Mechanical and Barrier Properties of Rice Starch Film. *Starch/Stärke*, *56*, 348-356.
- Laohakunjit, N., & Noomhorm, A. (2004b). Effect of Plasticizers on Mechanical and Barrier Properties of Rice Starch Film. *Journal of Starch/Stärke*, *56*, 348–356.
- Lee, D., Hwang, Y., & Cho, S. (1998). Developing antimicrobial packaging film for curled lettuce and soybean sprouts. *Food Science Biotechnology*, 7, 117-121.
- Lee, K. T., Choi, W. S., & Yoon, C. S. (2003). Effects of micro-perforated film on the quality and shelf life improvements of pork loins during chilled storage. *Meat Science*, *66*, 77-82.
- Lim, L. T., Mine, Y., & Tung, M. A. (1999). Barrier and tensile properties of transglutaminase cross-linked gelatin films as affected by relative humidity, temperature, and glycerol content. *Journal of Food Science*, 64(4), 616-622.
- Liu, J., Kerry, J. F., & Kerry, J. P. (2007). Application and assessment of extruded edible casings manufactured from pectin and gelatin/sodium alginate blends for use with breakfast pork sausage. *Meat Science*, 75 196-202.
- Liu, L., Liu, C. K., Fishman, M. L., & Hicks, K. B. (2007). Composite films from pectin and fish skin gelatin or soybean flour protein. *Journal of A gricultural and Food Chemistry*, 55(6), 2349-2355.
- Liu, Z. (2002). *Starch: from granules to biomaterials*. In S.G., Pandalai. Recent Research Developments in Applied Polymer Science (pp. 189-219). Trivandrum: Research Signpost.
- Mali, S., & Grossmann, M. V. E. (2003). Effects of yam starch films on storability and quality of fresh strawberries. *Journal of Agricultural and Food Chemistry*, 7005-7011.
- Mali, S., Grossmann, M. V. E., Garcia, M. A., Martino, M. N., & Zartizky, N. E. (2006). Effects of controlled storage on thermal, mechanical and barrier properties of plasticized biofilms from different starch sources. *Journal of Food Engineering*, 75, 453-460.

- Mali, S., Grossmann, M. V. E., Garcia, M. A., Martino, M. N., & Zartizky, N. E. (2006). Effects of controlled storage on thermal, mechanical and barrier properties of plasticized biofilms from different starch sources. *Journal of Food Engineering*, 75, 453-460.
- Mali, S., Sakanaka, L. S., Yamashita, F., & Grossmann, M. V. E. (2005). Water sorption and mechanical properties of cassava starch films and their relation to plasticizing effect. *Carbohydrate Polymers*, 60 (3), 283-289.
- Maqsood, S., Benjakul, S., & Khansaheb Balange, A. (2012). Effect of tannic acid and kiam wood extract on lipid oxidation and textural properties of fish emulsion sausages during refrigerated storage. *Food Chemistry*, 130 408-416.
- Mariniello, L., Di Pierro, P., Esposito, C., Sorrentino, A., Masi, P., & Porta, R. (2003). Preparation and mechanical properties of edible pectin-soy flour films obtained in the absence or presence of transglutaminase. *Journal of Biotechnology*, 102, 191-198.
- Mark, A. M., Roth, W. B., Mehltretter, C. L., & Rist, C. E. (1966). Oxygen permeability of amylomaize starch films. *Food Technology*, 20, 75-77.
- Martucci, J. F., & Ruseckaite, R. A. (2009). Biodegradation of three-layer laminate films based on gelatin under indoor soil conditions. *Polymer Degradation and Stability*, 94 (8), 1307-1313.
- Mathew, S., & Abraham, T. E. (2008). Characterisation of ferulic acid incorporated starch-chitosan blend films. *Food Hydrocolloids*, 22, 826-835.
- Mau, J. L., Chen, C. P., & Hsieh, P. C. (2001). Antimicrobial effect of extracts from Chinese chive, cinnamon, and corini fructus. *Journal of Agricultural and Food Chemistry*, 49, 183-188.
- McHugh, T. H., Avena-Bustillos, R., & Krochta, J. M. (1993). Hydrophilic edible films: modified procedure for water vapor permeability and explanation of thickness effects. *Journal of Food Science*, 58(4), 899-903.
- McHugh, T. H., & Krochta, J. M. (1994). Sorbitol -vs glycerol- plasticized whey protein edible films: integrated oxigen permeability and tensile property evaluation. *Journal of Agricultural and Food Chemistry*, 42(4), 841-845.
- Melton, S. L. (1983). Methodology for following lipid oxidation in muscle foods. *Food Technology*, 37, 105-111.
- Micard, X., Belamri, R., Morel, M. H., & Guilbert, S. (2000). Properties of chemically and physically treated wheat gluten films. *Journal of Agricultural and Food Chemistry*, 48, 2948-2953.
- Miller, K. S., Chiang, M. T., & Krochta, J. M. (1997). Heat curing of whey protein films. *Journal of Food Science*, 62(6), 1189-1193.
- Miller, K. S., Upadhyaya, S. K., & Krochta, J. M. (1998). Permeability of d-limonene in whey protein films. *Journal of Food Science*, 63(2), 244-247.
- Mohammadi Nafchi, A., Lai Hoong, C., & Alias, A. K. (2011). Effects of plasticizers on thermal properties and heat sealability of sago starch films. *Food Hydrocolloids*, 25 56-60.
- Muyonga, J. H., Cole, C. G. B., & Duodu, K. G. (2004). Fourier transform infrared (FTIR) spectroscopic study of acid soluble collagen and gelatin from skins and bones of young and adult Nile perch (Lates niloticus). *Food Chemistry*, 83(3), 325-332.

- Nielsen, P. V., & Rios, R. (2000). Inhibition of fungal growth on bread by volatile components from spices and herbs, and the possible application in active packaging, with special emphasis on mustard essential oil. *International Journal of Food Microbiology*, 60, 219-229.
- Ochs, D. (2000). Antimicrobials. In H., Zwefel. Plastics Additives Handbook (pp. 647-680). Munich: Hanser.
- Oellingrath, I. M., & Slinde, E. (1985). Color, pigment and iron content of meat loaves with blood, blood emulsion, or mechanically deboned meat added. *Journal of Food Science*, 50, 1551-1555.
- Ou, S., Wang, Y., Tang, S., Huang, C., & Jackson, M. G. (2005). Role of ferulic acid in preparing edible films from soy protein isolate. *Journal of Food Engineering*, 70(2), 205-210.
- Ouattara, B., Simard, R., Piette, G., Begin, A., & Holley, R. (2000). Diffusion of acetic and propionic acids from chitosan-based antimicrobial packaging films. *Journal of Food Science*, 65, 768-772.
- Ouattara, B., Simard, R. E., Piette, G., Be'gin, A., & Holley, R. A. (2000). Inhibition of surface spoilage bacteria in processed meats by application of antimicrobial films prepared with chitosan. *International Journal of Food Microbiology*, 62, 139-148.
- Ozdemir, M., & Floros, J. D. (2004). Active food packaging technologies. *Critical Reviews in Food Science* and Nutrition, 44(3), 185-193.
- Padgett, T., Han, I. Y., & Dawson, P. L. (1998). Incorporation of food-grade antimicrobial compounds into biodegradable packaging films. *Journal of Food Protection*, 61, 1330-1335.
- Park, H. J. (1991). Edible coating for fruits and vegetables: Determination of gas diffusivities, prediction of internal gas composition and effects of the coating on shelf life. Ph.D. dissertation, University of Georgia, Athens.
- Park, H. J., Weller, C. L., Vergano, P. J., & Testin, R. F. (1993). Permeability and mechanical properties of cellulose-based edible films. *Journal of Food Science*, 58(6), 1361-1364.
- Park, S. K., Rhee, C. O., Bae, D. H., & Hettiarachchy, N. S. (2001). Mechanical properties and water-vapor permeability of soy-protein films affected by calcium salts and glucono-d-lactone. *Journal of Agricultural and Food Chemistry*, 49(5), 2308-2312.
- Parra, D. F., Lugao, A. B., & Ponce, P. (2004). Mechanical properties and water vapor transmission in some blend of Cassava starch films. *Carbohydrate Polymer*, 37, 545-550.
- Pascat, B. (1986). *Study of some factors affecting permeability*. In M., Mathlouthi. Food packaging and preservation: theory and practice (pp. 2-24). New York: Elsevier.
- Pawlak, A., & Mucha, A. (2003). Thermogravimetric and FTIR studies of chitosan blends. *Thermochimica Acta*, 396(1-2), 153-166.
- Perez-Gago, M. B., & Krochta, J. M. (2002). Formation and properties of whey protein films and coatings. In A., Gennadios. Protein-based Films and Coatings (pp. 159-180). Boca Raton: CRC Press.
- Petersen, K., Nielsen, P. V., Bertelsen, G., Lawther, M., Olsen, M. B., Nilsson, N. H., & Mortensen, G. (1999). Potential of biobased materials for food packaging. *Trends in Food Science & Technology*, 10, 52-68.
- Pfeiffer, T., & Menner, M. (1999). Modified atmosphere packaging for self service fresh meat-change of gas atmosphere during storage. *Fleischwirtschaft*, 79 (12), 79-84.

- Pikul, J., & Kummerow, F. A. (1991). Thiobarbituric acid reactive substance formation as affected by distribution of polyenoic fatty acids in individual phospholipids. *Journal of Agricultural and Food Chemistry*, 39, 451-457.
- Pinotti, A., García, M. A., Martino, M., & Zaritzky, N. (2007). Study on microstructure and physical properties of composite films based on chitosan and methylcellulose. *Food Hydrocolloids*, 21, 66-72.
- Polnaya, F. J., Talahatu, J., Haryadi, & Marseno, D. W. (2012). Properties of biodegradable films from hydroxypropyl sago starches. *Asian Journal of Food and Agro-Industry*, 5 (3), 183-192.
- Pranoto, Y., Lee, C. M., & Park, H. J. (2007). Characterizations of fish gelatin films added with gellan and k-carrageenan. *Food Science and Technology*, 40(5), 766-774.
- Pranoto, Y., Lee, C. M., Park, H. J. (2007). Characterizations of fish gelatin films added with gellan and k-carrageenan. *Journal of Food Science and Technology*, 40(5), 766-774.
- Prodpran, T., Benjakul, S., & Phatcharat, S. (2012). Effect of phenolic compounds on protein cross-linking and properties of film from fish myofibrillar protein. *International Journal of Biological Macromolecules*, 51, 774-782.
- Pyla, R., Kim, T. J., Silva, J. L., & Jung, Y. S. (2010). Enhanced antimicrobial activity of starch-basedfilm impregnated with thermally processed tannic acid, a strong antioxidant. *International Journal of Food Microbiology*, 137 154-160.
- Quintavalla, S., & Vicini, L. (2002). Antimicrobial food packaging in meat industry. *Meat Science*, 62, 373-380.
- Rhim, J. M. (1998). Modification of soy protein film by formaldehyde. Korean Journal of Food Science and Technology, 30(2), 372-378.
- Rico-Pena, D., & Torres, J. (1991). Sorbic acid and potassium sorbate permeability of an edible methylcellulose-palmitic acid film: water activity and pH effects. *Journal of Food Science*, 562, 497-499.
- Rivero, S., García, M. A., & Pinotti, A. (2010). Crosslinking capacity of tannic acid in plasticized chitosan films. *Carbohydrate Polymers*, 82, 270-276.
- Robles-Martinez, C., Cervantes, E., & Ke, P. J. (1982). Recommended Method For Testing the Objective Rancidity Development in Fish Based on TBARS Formation: Canadian Technical Report of Fisheries and Aquatic Sciences.
- Rodri'guez, M., Ose's, J., Ziani, KH., Mate, J.I. (2006). Combined effect of plasticizers and surfactants on the physical properties of starch based edible films. *Journal of Food Research International, 39*, 840-846.
- Rodriguez, M., Oses, J., Ziani, K., & Mate, J. I. (2006). Combined effect of plasticizers and surfactants on the physical properties of starch based edible films. *Food Research International*, *39*, 840-846.
- Rooney, M. L. (1995). Active packaging in polymer films. Glasgow: Blackie Academic & Professional.
- Roth, W. B., & Mehltretter, C. L. (1970). Films from mixture of viscose and alkali high-amylose corn starch. *Journal of Applied Polymer Science*, 14, 1387-1389.
- Sabato, S. F., Ouattara, B., Yu, H., D'Aprano, G., Le Tien, C., & Mateescu, M. A. (2000). Mechanical and barrier properties of cross linked soy and whey protein based films. *Journal of Agricultural and Food Chemistry*, 49, 1397-1403.

- Sanchez-Moreno, C., Larrauri, A. J., & Saura-Calixto, F. (1999). Free radical scavenging capacity and inhibition of lipid oxidation of wines, grape juices and related polyphenolic constituents-history, production and role in disease prevention. *Food Research International*, *32*, 407-412.
- Santiago-Silva, P., Soares, N. F. F., Nóbrega, J. E., Júnior, M. A. W., Barbosa, K. B. F., Volp, A. C. P., Zerdas, E.R.M.A., & Würlitzer, N. J. (2009). Antimicrobial efficiency of film incorporated with pediocin (ALTA®2351) on preservation of sliced ham. *Food Control*, 20, 85-89.
- Scalbert, A. (1991). Antimicrobial properties of tannin. Phytochemistry, 30, 3875-3883.
- Sebti, I., & Coma, V. (2002). Active edible polysaccharide coating and interactions between solution coating compounds. *Carbohydrate Polymers*, 49, 139-144.
- Seyfzadeh, M., Motalebi, A. A., Kakoolaki, S., & Gholipour, H. (2013). Chemical, microbiological and sensory evaluation of gutted kilka coated with whey protein based edible film incorporated with sodium alginate during frozen storage. *Iranian Journal of Fisheries Sciences*, *12*(1), 140-153.
- Siragusa, G. R., & Dickinson, J. S. (1993). Inhibition of Listeria monocytogenes, Salmonella Typhimurium and Escherichia coli O157:H7 on beef muscle tissue by lactic or acetic acid contained in calcium alginate gels. Journal of Food Safety, 13(2), 147-158.
- Siripatrawan, U., & Noipha, S. (2012). Active film from chitosan incorporating green tea extract for shelf life extension of pork sausages. *Food Hydrocolloids*, 27 102-108.
- Skurtys, O., Acevedo, C., Pedreschi, F., Enrione, J., Osorio, F., & Aguilera, J. M. (2010). Food Hydrocolloid Edible Films and Coatings: Food Science and Technology: Nova Science Publishers.
- Slavutsky, A. M., & Bertuzzi, M. A. (2014). Water barrier properties of starch films reinforced with cellulose nanocrystals obtained from sugarcane bagasse. *Carbohydrate Polymers*, *110* 53-61.
- Smith, J. P., Hoshino, J., & Abe, Y. (1995). *Interactive packaging involving sachet technology*. In M.L., Rooney. Active Food Packaging (pp. 143-173). London: Blackie Academic and Professional.
- Smith, J. P., Ramaswamy, H. S., & Simpson, B. K. (1990). Developments in food packaging technology. Part II. Storage aspects *Trends in Food Science & Technology*, 1, 111-118.
- Smitha, B., Sridhar, S., & Khan, A. A. (2005). Chitosan-sodium alginate polyion complexes as fuel cell membranes. *European Polymer Journal*, 41, 1859-1866.
- Sobral, P. J. A. (2000). Influe[^]ncia da espessura sobre certas propriedades de biofilmes a[^] base de protei[^]nas miofibrilares. *Pesquisa Agropecua[^]ria Brasileira*, 35(6), 1251-1259.
- Sobral, P. J. A., & Habitante, A. M. Q. B. (2001). Phase transitions of pigskin gelatin. *Food Hydrocolloids*, 15(4-6), 377-382.
- Sobral, P. J. A., Menegalli, F. C., Hubinger, M. D., & Roques, M. A. (2001). Mechanical, water vapor barrier and thermal properties of gelatin based edible films. *Food Hydrocolloids*, *15*, 423-432.
- Sobral, P. J. A., Monterrey-Quintero, E. S., & Habitante, A. M. Q. B. (2002). Glass transition of Nile tilapia myofibrillar protein films plasticized by glycerin and water. *Journal of Thermal Analysis and Calorimetry*, 67(2), 499-504.
- Sorrentino, A., Gorrasi, G., & Vittoria, V. (2007). Potential perspectives of bio-nanocomposites for food packaging applications. *Trends in Food Science & Technology, 18*, 84-95.

- Sothornvit, R., & Krochta, J. M. (2000). Plasticizer effect on oxygen permeability of beta-lactoglobulin films. *Journal of Agricultural and Food Chemistry*, 48, 6298-6302.
- Sothornvit, R., & Krochta, J. M. (2000). Water vapor permeability and solubility of films from hydrolyzed whey protein. *Journal of Food Science*, 65 (4), 700-703.
- Sothornvit, R., & Krochta, J. M. (2001). Plasticizer effect on mechanical properties of β-lactoglobulin films. Journal of Food Engineering, 50, 149-155.
- Stevens, M. (2012). Permeation and its impact on Packaging. MOCON. Minneapolis, MN USA.
- Stuchell, Y. M., & Krochta, J. M. (1994). Enzymatic treatments and thermal effects on edible soy protein films. *Journal of Food Science*, 59, 1322-1337.
- Su, J. F., Huang, Z., Yuan, X. Y., Wang, X. Y., & Li, M. (2010). Structure and properties of carboxymethyl cellulose/soy protein isolate blend edible films crosslinked by Maillard reactions. *Carbohydrate Polymers*, 79(1), 145-153.
- Suppakul, P., Miltz, J., Sonneveld, K., & Bigger, S. W. (2003). Active packaging technologies with an emphasis on antimicrobial packaging and its applications. *Journal of Food Science*, 68, 408-420.
- Taguri, T., Tanaka, T., & Kouno, I. (2004). Antimicrobial activity of 10 different plant polyphenols against bacteria causing food-borne disease. *Biological & Pharmaceutical Bulletin*, 27, 1965-1969.
- Testin, R. F., & Vergano, P. J. (1990). Food Packaging, Food Protection and the Environment. A Workshop Report. *IFT Food Packaging Division*: The Institute of Food Technologists, Chicago.
- Theivendran, S., Hettiarachchy, N. S., & Johnson, M. G. (2006). Inhibition of *Listeria monocytogenes* by nisin combined with grape seed extract or green tea extract in soy protein film coated on turkey frankfurters. *Journal of Food Science*, *71*, M39-M44.
- Thomas, D. J., & Atwell, W. A. (1997). Starches. St Paul: Eagan Press.
- Thomazine, M., Carvalho, R. A., & Sobral, P. I. A. (2005). Physical properties of gelatin films plasticized by blends of glycerol and sorbitol. *Journal of Food Science*, 70(3), 172-176.
- Trezza, T. A., & Krochta, J. M. (2000). Color stability of edible coatings during prolonged storage. *Journal of Food Science*, 65 (1), 1166-1169.
- Vanin, F. M., Sobral, P. J. A., Menegalli, F. C., Carvalho, R. A., & Habitante, A. M. Q. B. (2005). Effects of plasticizers and their concentrations on thermal and functional properties of gelatin based films. *Food Hydrocolloids*, 19(5), 899-907.
- Vanin, F. M., Sobral, P. J. A., Menegalli, F. C., Carvalho, R. A., Habitante, A. M. Q. B. (2005). Effects of plasticizers and their concentrations on thermal and functional properties of gelatin-based films. *Journal of Food Hydrocolloids*, 19(5), 899-907.
- Veiga-Santos, P., Oliveira, L. M., Cereda, M. P., & Scamparini, A. R. P. (2007). Sucrose and inverted sugar as plasticizer. Effect on cassava starch-gelatin film mechanical properties, hydrophilicity and water activity. *Food Chemistry*, 103 255-262.
- Vermeiren, L., Devlieghere, F., Van Beest, M., de Kruijf, N., & Debevere, J. (1999). Developments in the active packaging of foods. *Trends in Food Science & Technology*, 10, 77-86.

- Vicentini, N. M., Dupuy, N., Leitzelman, M., Cereda, M. P., & Sobral, P. J. A. (2005). Prediction of cassava starch edible film properties by chemometric analysis of infrared spectra. *Spectroscopy Letters: An International Journal for Rapid Communication*, 38, 749.
- Viga-Santos, P., Oliveria, L. M., Cereda, M. P., & Scamparini, A. R. P. (2007). Sucrose and inverted sugars as plasticizer. Effect on cassava starch-gelatin film mechanical properties, hydrophilicity and water activity. *Food Chemistry*, 103, 255-262.
- Vojdani, F., & Torres, A. (1990). Potassium sorbate permeability of methylcellulose and hydroxypropyl methylcellulose coatings. Effects of fatty acids. *Journal of Food Science*, 553, 841-846.
- Vojdani, F., & Torres, J. A. (1989). Potassium sorbate permeability of methylcellulose and hydroxypropyl methylcellulose multilayer films. *Journal of Food Processing and Preservation*, 13, 417-430.
- Were, L., Hettiarachcky, N. S., & Coleman, M. (1999). Properties of cysteine-added soy protein-wheat gluten films. *Journal of Food Science*, 64(3), 514-518.
- Wu, H.-C. H., & Sarko, A. (1978). The double-helical molecular structure of crystalline B-amylose. *Journal of Carbohydrate Research*, 61, 7-25.
- Wu, H. C. H., & Sarko, A. (1978). The double-helical molecular structure of crystalline B-amylose. *Carbohydrate Research*, 61, 2740.
- Wu, Y., Rhim, J. W., Weller, C. L., Lamouz, F., Cuppet, S., & Schnepf, M. (2000). Moisture loss and lipid oxidation for precooked beef patties stored in edible coatings and films. *Journal of Food Science*, 65 300-304.
- Wu, Y., Weller, C. L., Hamouz, F., Cuppett, S. L., & Schnepf, M. (2002). Development and application of multicomponent edible coatings and films: a review. Advance Food and Nutrition Research, 44, 347-394.
- Xie, L., Hettiarachchy, N. S., Jane, M. E., & Johnson, M. G. (2003). Antimicrobial activity of ginkgo biloba leaf extract on *Listeria monocytogenes. Journal of Food Science*, 68, 268-270.
- Xu, P., Uyama, H., Whitten, J. E., Kobayashi, S., & Kaplan, D. L. (2005). Peroxidase-catalyzed in situ polymerization of surface oriented caffeic acid. *Journal of the American Chemical Society*, 127(33), 11745-11753.
- Xu, Y. X., Kim, K. M., Hanna, M. A., & Nag, D. (2005). Chitosan-starch composite film: preparation and characterization. *Journal of Industrial Crops and Products*, 21 185-192.
- Yildirim, M., & Hettiarachchy, N. S. (1997). Biopolymers produced by cross-linking soybean 11s globulin with whey proteins using transglutaminase. *Journal of Food Science*, 62(2), 270-275.
- Yilmaz, Y. (2006). Novel uses of catechins in foods. Trends in Food Science & Technology, 17, 64-71.
- Yin, Y. J., Yao, K. D., Cheng, G. X., & Ma, J. B. (1999). Properties of polyelectrolyte complex films of the chitosan and gelatin. *Polymer International*, 48, 429-433.

PUBLICATIONS

- Bakar, J., Ahangari, R., Abdul Rahman, R., & Karim, R., (2014). Edible packaging film. Universiti Putra Malaysia Patent. (PI2014702538)
- Ahangari, R., Bakar, J., Abdul Rahman, R., & Karim, R., (2014). Properties of rice starch edible films incorporated with fish gelatin, Food Hydrocolloid (submitted)

UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION: _

TITLE OF THESIS / PROJECT REPORT:

DEVELOPMENT OF RICE STARCH-FISH GELATIN EDIBLE FILMS

NAME OF STUDENT:

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.

- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as:

Date:

Date: