

# **UNIVERSITI PUTRA MALAYSIA**

APPLICATION OF PHYLLOSPHERE BACTERIAL ANTAGONIST AGAINST RICE SHEATH BLIGHT

**SHAMIMA AKTER** 

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# APPLICATION OF PHYLLOSPHERE BACTERIAL ANTAGONIST AGAINST RICE SHEATH BLIGHT

By

SHAMIMA AKTER

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

January 2015

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# **ДЕДЈЕАЈЈО**Л

Ia

My affectionate parents with gratitude

and

My beloved husband for his sacrifices and understanding

Abstract of thesis presented to the Senate of Universiti Putra Malaysia, in fulfillment of the requirements for the degree of Doctor of Philosophy

# APPLICATION OF PHYLLOSPHERE BACTERIAL ANTAGONIST AGAINST RICE SHEATH BLIGHT

By

## SHAMIMA AKTER

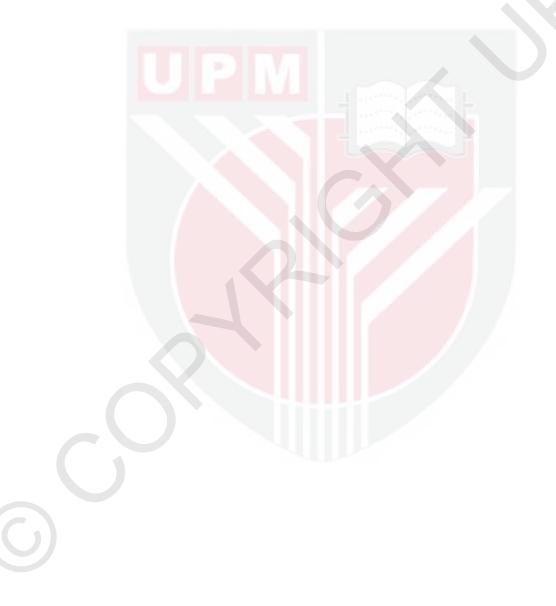
### January 2015

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Antagonistic bacteria originating from the rhizosphere are being used in sheath blight management. However, little is known about the potential of bacteria inhabiting the phyllosphere. Hence, a study was initiated with the aim of assessing the effective bacterial antagonists against the disease. A total of 325 bacterial isolates obtained from 100 rice plant samples collected from different locations of Malaysia and Bangladesh were preliminarily screened. Out of the 325 bacterial isolates, 14 were selected based on their ability to inhibit the growth of R. solani. In dual culture tests isolates KMB25, TMB33, PMB38, UMB20 and BMB42 showed 68.44, 60.89, 60.22, 50.00 and 48.22% inhibitions, respectively. In extracellular metabolites tests most of these isolates showed comparatively higher percentages of growth inhibition of the fungus than in dual culture tests. Selected isolates were negative to indole, methyl red, Voges Proskaeur, and starch hydrolysis but positive to catalase, urease, and nitrate reduction tests. Isolates KMB25, TMB33 and PMB38 were positive to gelatin liquefaction, while isolates UMB20 and BMB42 were negative to the test. Biolog identified P. fluorescens (UMB20), P. aeruginosa (KMB25, TMB33 and PMB38) and P. asplenii (BMB42) with the similarity index ranging from 0.52 to 0.70. The identities of the selected three bacterial isolates UMB20, KMB25 and BMB42 were further confirmed through 16S rDNA gene sequencing. According to the GenBank database of NCBL, UMB20 and BMB42 were identified as P. fluorescens and KMB25 as P. aeruginosa. Fungal growth inhibition ranging from 86.85 to 93.15% was obtained by these strains in volatile and 100% was in diffusible metabolites test. Among the 3 strains, UMB20 and BMB42 produced indole 3-acetic acid and chitinase, but not protease. All of them produced cellulase, siderophore, HCN, and ammonia and were able to solubilize phosphate. Strains UMB20 and BMB42 were preserved in peat and talc as single strains or in mixtures. The peat formulation was found to be more suitable than talc to retain longer shelf life of individuals and strain mixtures with sufficient viable cells. At 4°C of storage condition peat formulations were better than room temperature  $(28\pm2^{\circ}C)$  condition. In bioefficacy tests of peat based bacterial formulations under glass house conditions, the strain mixture and UMB20 alone significantly reduced the disease severity in terms of area under disease progress curve (AUDPC) compared to the untreated control. Percent reduction of AUDPC was 32.79, 32.58 and 21.19 for strain mixture (UMB20+BMB42), UMB20 and BMB42, respectively. Significantly lowest disease progression rate (0.01unit/day) was found in the strain mixture applied plants. In addition to disease suppression, the strain mixture enhanced the plant height, percentage of effective tillers per hill, and percentage of fertile spikelets per panicle. Effects of all the treatments on flag leaf area, total number of tillers and number of effective tillers were insignificant. Significantly highest weight of 100-grain (1.65 g) was obtained from the strain mixture applied plants. The *Pseudomonas* bacteria isolated from rice plants had the potential to inhibit the fungal growth *in vitro* and *in vivo* and possessed most of the plant growth promoting characteristics. They have the potential to be utilized as biocontrol agents for management of sheath blight in rice.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

# APLIKASI BAKTERIA ANTAGONIS FILOSFERA TERHADAP PENYAKIT HAWAR SELUDANG PADI

Oleh

## SHAMIMA AKTER

#### Januari 2015

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Bakteria antagonistik berasal dari rizosfera telah digunakan dalam pengurusan hawar seludang. Walau bagaimanapun, sedikit yang diketahui mengenai potensi bakteria yang mendiami filosfera. Oleh itu, satu kajian telah dimulakan dengan tujuan untuk menilai bakteria antagonis yang berkesan terhadap penyakit itu. Sejumlah 325 pencilan bakteria yang diperolehi daripada 100 sampel tanaman padi yang diambil dari lokasi yang berlainan di Malaysia dan Bangladesh yang pada awalnya telah disaring. Daripada 325 pencilan bakteria, 14 telah dipilih berdasarkan keupayaan mereka untuk menghalang pertumbuhan R. solani. Dalam ujian dua pencilan kultur, KMB25, TMB33, PMB38, UMB20 dan BMB42 telah menunjukkan 68,44, 60,89, 60,22, 50.00 dan 48.22% perencatan, masing-masing. Dalam ujian metabolik ekstrasel, kebanyakan pencilan ini menunjukkan peratusan yang agak tinggi di dalam perencatan pertumbuhan kulat daripada dalam ujian dua kultur. Pencilan terpilih adalah negatif untuk indol, metil merah, voges proskaeur, dan hidrolisis kanji tetapi positif untuk katalase, urease, dan ujian penurunan nitrat. Pencilan KMB25, TMB33 dan PMB38 pula positif kepada pencairan gelatin, manakala pencilan UMB20 dan BMB42 adalah negatif untuk ujian ini. Sistem biolog telah mengenal pasti P. fluorescens (UMB20), P. aeruginosa (KMB25, TMB33 dan PMB38) dan P. asplenii (BMB42) dengan indeks persamaan antara 0.52 – 0.70. Identiti tiga pencilan bakteria yang dipilih UMB20, KMB25 dan BMB42 disahkan lagi melalui penjujukan gen 16S rDNA. Menurut pangkalan data gen bank UMB20 dan BMB42 telah dikenal pasti sebagai P. fluorescens dan KMB25 sebagai P. aeruginosa. Perencatan pertumbuhan kulat yang terdiri daripada 86.85 – 93.15% telah diperolehi oleh strain ini dalam bentuk meruap dan 100% adalah dalam ujian peresapan metabolit. Di antara 3 jenis strain, UMB20 dan BMB42 menghasilkan indole 3-asetik asid dan kitinase, tetapi tidak protease. Kesemua mereka menghasilkan selulase, siderophore, HCN, dan ammonia dan mampu mencairkan fosfat. Strain UMB20 dan BMB42 adalah dipelihara dalam tanah gambut dan talkum sebagai jenis tunggal atau campuran. Formulasi gambut pula didapati lebih sesuai daripada talkum untuk mengekalkan jangka masa hidup bagi strain individu dan campuran dengan sel-sel hidup yang mencukupi. Penyimpanan gambut di dalam suhu 4°C keadaan sejuk adalah lebih baik daripada suhu bilik (28±2°C). Di dalam ujian bioefikasi gambut

iii

berasaskan formulasi bakteria di bawah keadaan rumah kaca, campuran strain dan UMB20 sahaja keterukan penyakit ini berkurangan dengan signifikan dari segi kawasan di bawah lengkung kemajuan penyakit (AUDPC) berbanding dengan kawalan yang tidak dirawat. Peratus pengurangan AUDPC adalah 32.79, 32.58 dan 21.19 masing-masing untuk campuran strain (UMB20 + BMB42), UMB20 dan BMB42. Kadar perkembangan penyakit terendah (0.01unit / hari) sangat signifikan untuk campuran strain kepad tanaman yang digunakan. Selain kawalan penyakit, campuran strain telah meningkatkan ketinggian tumbuhan, peratusan anak padi yang efektif bagi setiap bukit, dan peratus spikelet yang subur bagi setiap penikel. Kesan semua rawatan kepada kawasan keluasan daun, jumlah anak padi dan bilangan anak padi juga tidak ketara. Berat tertinggi yang signifikan bagi 100-bijian (1.65 g) telah diperolehi daripada campuran strain kepada tanaman yang digunakan. Bakteria Pseudomonas yang dipencilkan daripada tanaman padi mempunyai potensi untuk menghalang pertumbuhan kulat in vitro dan in vivo dan mempunyai kebanyakan ciriciri penggalak pertumbuhan tanaman. Ia juga mempunyai potensi untuk digunakan sebagai agen kawalan biologi untuk pengurusan hawar seludang untuk padi.

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I certify that a Thesis Examination Committee has met on 16 January 2015 to conduct the final examination of Shamima Akter on her thesis entitled "Application of Phyllosphere Bacterial Antagonist against Rice Sheath Blight" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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# TABLE OF CONTENTS

				Page
AI	BSTRA	АСТ		I
AI	BSTRA	4K		iii
AC	CKNO	WLED	GEMENTS	v
	PPRO			viii
		RATIO		xiv
		F TABI		xvi
		F FIGU		xviii
LI	ST OI	F ABBF	REVIATIONS	
CI	HAPT	ER		
1	INT	'RODU	CTION	1
2	т тт		JRE REVIEW	3
2	2.1	Rice		
	2.1		iseases	3 3
	2.2		blight of rice	4
	2.3		i blight pathogen <i>Rhizoctonia solani</i>	6
	2.5		gement of sheath blight	7
			Management with host resistance	7
			Cultural management	8
		2.5.3		8
		2.5.4	Management using botanicals	9
			Biocontrol	9
			2.5.5.1 Mechanisms of action deployed by bacterial antagonists	11
			2.5.5.2 Formulation	14
			2.5.5.3 Application of bacterial inoculants	14
			2.5.5.5 Application of bacterial moculants	15
3			N, CHARACTERIZATION AND IDENTIFICATION	18
			ERIA ANTAGONISTIC AGAINST SHEATH BLIGHT	
	3.1	Introd		18
	3.2		als and Methods	19
		3.2.1	Samples collection	19
		3.2.2	Isolation of microorganisms	19
		3.2.3	Determination of anastomosis group	20
		3.2.4	Pathogenicity test of R. solani on rice	20
		3.2.5	Screening of bacteria for antagonism against R. solani	20
		3.2.6	Hypersensitivity reaction test	21
		3.2.7	Morphological and cultural characterization	21
		3.2.8	Physiological characterization of bacterial isolates	21
		3.2.9	Biochemical characterization	21
		3.2.10	Intrinsic resistance profile of bacterial isolates against	23
		0.0.11	heavy metals and antibiotics	<u> </u>
		3.2.11	Selection of bacterial isolates	23
		3.2.12	Identification of bacteria using Biolog system	24

		3.2.13	Characterization of bacterial isolates using the 16S rDNA	25
			gene sequencing	
		3.2.14	Experimental design and statistical analysis	26
	3.3	Results		26
		3.3.1	Isolation, identification and anastomosis grouping of <i>R</i> . <i>solani</i>	26
		3.3.2	Pathogenicity of R. solani	27
		3.3.3	Isolation and screening of bacterial antagonists and	28
			hypersensitivity reaction	
		3.3.4	Morphological, cultural and physiological characteristics of bacterial isolates	29
		3.3.5	Biochemical characteristics	33
		3.3.6	Tolerance of bacterial isolates to different heavy metals and antibiotics	33
		3.3.7	Dual culture test	36
		3.3.8	Extracellular metabolites test	36
		3.3.9	Identification of bacteria using the Biolog system	38
		3.3.10	Molecular characterization of the isolates	38
	3.4			40
	3.5	Conclus		43
4	BIC	CONTR	EVALUATION OF MECHANISMS INVOLVED IN COL AND PLANT GROWTH PROMOTING S OF SELECTED PSEUDOMONAS STRAINS	44
	4.1	Introdu		44
	4.1		ls and Methods	44 45
	4.2	4.2.1	Microorganisms and culture conditions	45 45
		4.2.2	Broad spectrum antifungal activity of bacterial strains	45
			against common fungal pathogens of rice	
		4.2.3	Production of diffusible metabolites	45
		4.2.4	Production of volatile metabolites	46
		4.2.5	Detection of chitinase production	46
		4.2.6	Production of protease	47
		4.2.7	Production of $\beta$ -1,3-glucanase	47
		4.2.8	Production of other beneficial enzymes	48
		4.2.9	Indole 3-acetic acid (IAA) production	48
		4.2.10	Phosphate solubilization assay	48
		4.3.11	Detection of ammonia	48
		4.3.12	Hydrogen cyanide (HCN) production	49
		4.3.13	Siderophore detection	49
		4.3.14	Assessment of biofilm formation	49
	1.2	4.3.15	Experimental design and statistical analysis	50
	4.3	Results		50
		4.3.1	Antifungal activity of bacterial strains against common fungal pathogens of rice	50
		4.3.2	Production of volatile and diffusible metabolites	51
		4.3.3	Screening for lytic enzyme activities	51
		4.3.4	Screening for plant growth promoting traits	53
		4.3.5	Formation of biofilm by bacterial strains	55
	4.4	Discuss	•	56

xi

4	.5 Conclu	sions	59
S	HEATH B	OF BIOFORMULATED UMB20 AND BMB42 ON LIGHT AND YIELD ATTRIBUTES OF RICE UNDER USE CONDITION	60
	.1 Introdu		60
		ls and Methods	60
5	.2 Materia 5.2.1	Bacterial inoculants and pathogen	60
	5.2.2	Plant and carrier materials	60
	5.2.3	Experimental locations	61
	5.2.4	Preparation of bacterial cell suspension	61
	5.2.5	Compatibility test of the bacterial strains	61
	5.2.6	Preparation of formulations	61
	5.2.7	Enumeration of viable cells and calculation of viability	62
	0.1217	losses of bacterial strains in formulations	
	5.2.8	Evaluation of antagonistic ability of peat formulated	62
		stored bacterial strains against R. solani	
	5.2.9	Effect on germination of sclerotia	63
	5.2.10	Effect on seedling growth	63
	5.2.11	Raising and transplanting seedling	63
	5.2.12	Management practices	64
	5.2.13	Application of bacterial inoculants	64
	5.2.14	Pathogen inoculation	64
	5.2.15	Determination of rhizosphere colonization of peat formulated bacterial strains	64
	5.2.16	Assessment of disease	65
	5.2.1 <mark>7</mark>	Assessment of agronomic traits and yield contributing components	65
	5.2.18	Experimental design and statistical analysis	66
5	.3 Results		66
	5.3.1	Compatibility of bacterial strains	66
	5.3.2	Effect of storage period on the viability of bacterial strains in carrier materials	66
	5.3.3	Effect of storage period on viability losses of formulated bacterial inoculants	69
	5.3.4	Antagonistic activities of bacterial strains in formulations	69
	5.3.5	Effect on sclerotial germination	71
	5.3.6	Effect on seedling growth	72
	5.3.7	Rhizosphere colonization of bacterial strains	73
	5.3.8	Efficacy of formulated bacterial strains on disease	73
	5.3.9	suppression Effect of bacterial consortium on plant growth and	75
	<b>FO</b> 10	tillering ability	
-	5.3.10	Effect of bacterial inoculants on yield components	76
	.4 Discuss		77
5	.5 Conclus	Sions	80

6	SUMMARY, GENERAL CONCLUSION AND	82
	<b>RECOMMENDATIONS FOR FUTURE RESEARCH</b>	
	6.1 Summary	82
	6.2 Conclusions	82
	6.3 Recommendations	83
	FERENCES	84
	PENDICES	113
BIO	DATA OF STUDENT	131
LIS	T OF PUBLICATIONS	132



# LIST OF TABLES

Table		Page
2.1	Host range and diseases caused by Rhizoctonia solani AG-1 IA	7
2.2	Major diseases of rice with causal organisms and bacterial antagonists	11
3.1	Anastomosis grouping of different isolates of <i>Rhizoctonia solani</i> with the tester of <i>R. solani</i> AG-1 IA	27
3.2	Specification of bacterial isolate	29
3.3	Cultural and morphological characteristics of bacterial isolates	31
3.4	Growth of bacteria at different range of temperature, salinity and pH	32
3.5	Biochemical characteristics of bacterial isolates done by conventional method	33
3.6	Tolerance ability of bacterial isolates to different heavy metals after 24 hours of incubation	35
3.7	Resistance of bacterial isolates to different antibiotics after 48 hours of incubation	36
3.8	Effect of bacterial isolates on mycelial growth of <i>Rhizoctonia solani</i> after 3 days of incubation	37
3.9	Identification of <i>Pseudomonas</i> spp. by means of Biolog Microstation system based on carbon source utilization	38
4.1	Antagonistic activity of bacterial strains against common fungal pathogens of rice	50
4.2	Effect of volatile and diffusible metabolites on growth of <i>Rhizoctonia</i> solani after 5 days of incubation	51
4.3	Production of hydrolytic enzymes by bacterial strains	52
4.4	Production of growth related hormone and metabolites by bacterial strains	53
5.1	Viability of bacterial inoculants in peat formulation at $4^{\circ}$ C storage condition	68
5.2	Viability of bacterial inoculants in peat formulation at room temperature ( $28\pm2^{\circ}C$ ) storage condition	68
5.3	Viability of bacterial inoculants in talc formulation at 4°C storage	68

condition

5.4	Viability of bacterial inoculants in talc formulation at room temperature $(28\pm2^{\circ}C)$ storage condition	69
5.5	Effect of storage period on antagonistic activity of individuals and strain mixture in peat and talc formulations against the growth of <i>R</i> . <i>solani</i> at 4°C and room temperature ( $28\pm2^{\circ}C$ ) condition (RT)	71
5.6	Effect of peat formulated bacterial strains on seedling growth of rice	72
5.7	Effect of soil application of peat base formulation of bacterial strains on rhizosphere colonization of <i>Pseudomonas</i> in the rice roots	73
5.8	Effect of bacterial strains on sheath blight affected area, disease progression rate and percent tiller infection	74
5.9	Effect of bacterial strains on plant growth and tillering ability	76
5.10	Effect of bacterial strains on yield attributes in rice	77

# LIST OF FIGURES

Figure		Page
2.1	Sheath blight disease symptoms and culture of the pathogen <i>Rhizoctonia solani</i>	5
2.2	Disease cycle of sheath blight disease in rice	б
3.1	Mean values of percent sheath blight lesion length on rice cultivar IR50 with respect to different <i>R. solani</i> isolates	28
3.2	Growth inhibition of <i>Rhizoctonia solani</i> caused by bacterial isolates in dual culture test on potato dextrose agar after 72 hours of incubation	38
3.3	Amplification of 16S rDNA of bacterial isolates in 1% TAE buffer fragmented at 1500 base pairs	39
3.4	Phylogenetic tree showing the position of strains UMB20, KMB25 and BMB42	40
4.1	Effect of volatile metabolites produced by bacterial strains on the growth of <i>Rhizoctonia solani</i> after 5 days of incubation at 28±2°C	51
4.2	Effect of diffusible metabolites produced by bacterial strains on the growth of <i>Rhizoctonia solani</i> after 5 days of incubation at 28±2°C	51
4.3	Chitinase activities of bacterial strains on colloidal chitin amended basal medium after 2 days of incubation at 28±2°C	52
4.4	Proteolytic activities of bacterial strains on skim milk agar after 2 days of incubation at 28±2°C	53
4.5	Colorimetric quantification of indole acetic acid production by bacterial strains in TSB medium supplemented with L-Tryptophan @ 1 mg mL <sup>-1</sup> after 5 days of incubation at $30^{\circ}$ C	54
4.6	Phosphate solubilizing efficiency of bacterial strains on National Botanical Research Institute`s Phosphate medium after 7 days of incubation at 28±2°C	54
4.7	Hydrogen cyanide production by bacterial strains on Whatman No. 1 filter paper soaked in picric acid solution after incubation for 7 days at $30^{\circ}$ C	55
4.8	Siderophore production by bacterial strains on CAS-agar medium after 2 days of incubation at $28\pm2^{\circ}C$	55
4.9	Abilities of bacterial strains to form biofilm in 1% crystal violet after 24 hours of incubation in Luria-Bertany broth	56

6

5.1	Compatibility of bacterial strains UMB20 and BMB42	66
5.2	Percent viability losses of formulated bacterial strains as individuals and strain mixture in peat and talc at $4^{\circ}$ C and room temperature (28±2°C) storage conditions	70
5.3	Effect of formulated individual and strain mixture on germination of sclerotia of <i>Rhizoctonia solani</i> on potato dextrose agar after different soaking hours	72
5.4	Progression of sheath blight (as measured by % disease severity) over time after application of treatments	75
5.5	Effect of bacterial strains on average of 100-grain weight after adjusted at 14% moisture content	77

# LIST OF ABBREVIATIONS

%	Percent
PDI	Percent Disease Incidence
%RLH	Percent Relative Lesion Height
μg	Microgram
μL	Microliter
2, 4- DAPG	2, 4-Diacetylphloroglucinol
AG	Anastomosis Group
ANOVA	Analysis of Variance
AUDPC	Area Under Disease Progress Curve
bp	Base pairs
ĊAS	Chromo Azurol S
cfu	Colony Forming Units
CMC	Carboxymethyl cellulose
CRD	Completely randomized design
СТАВ	Cetyl Trimethyl-ammonium bromide
DMAB	<i>p</i> -dimethylaminobenzaldehyde
DNA	Deoxyribonucleic acid
DNS	Dinitrosalicylic acid
dNTP	Deoxyribonucleotide triphosphate
EDTA	Ethylenediaminetetraacetic acid
et al.	and others
GL	Gelatin Liquefaction
GN	Gram-Negative
ha	Hectare
HCN	Hydrogen Cyanide
HDTMA	Hexadecyletrimethyllammonium
HPLC	High Performance Liquid Chromatography
HR	Hypersensitivity reaction
IAA	Indole acetic acid
IAR	Intrinsic Antibiotic Resistance
ISR	Induced Systemic Resistance
KBA	Kings B Agar
KBB	Kings B Broth
kg	Kilogram
LBB	Luria-Bertani Broth
LSD	Least Significant Difference
LSD L-Trp	L-Tryptophan
M	Molar
	Milligram
mg mL	Milliliter
	Millimeter
mm mM	Millimolar
	Muriate of Potash
MP MP	
MR	Methyl red
N NA	Normal
NA	Nutrient Agar
NB	Nutrient Broth
NBRIP	National Botanical Research Institute's Phosphate

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nm	Nanometer
NPK	Nitrogen Phosphorus Penta oxide Potassium oxide
NR	Nitrate Reduction
°C	Degree Celsius
р.	Page
PCR	Polymerase Chain Reaction
PDA	Potato Dextrose Agar
PGPR	Plant Growth Promoting Rhizobacteria
PIRG	Percent Inhibition of Radial Growth
PO	Peroxidase
pp.	Pages
ppm	Parts per million
psi	Pound per square inch
Rf	Retention Factor
RH	Relative Humidity
rpm	Rotation per minute
SA	Salicylic acid
SAS	Statistical Analysis System
SDS	Sodium dodecyl sulphate
ShB	Sheath blight
SM	Succinic Medium
SMA	Skim Milk Agar
sp.	species (singular)
spp.	species (plural)
TAE	Tris - Acetate - EDTA
TSB	Trypticase Soya Broth
TSP	Triple Super Phosphate
UV	Ultra Violet
v/v	Volume per volume
VP	Voges Proskeaur
w/v	Weight per volume
w/w	Weight per weight

 $(\mathbf{G})$ 

### **CHAPTER 1**

#### **INTRODUCTION**

Rice (*Oryza sativa* L.) is the most widely cultivated food crop in the world and is the staple food for more than half of the world's population (Zeigler and Barclay, 2008). The major rice producer countries are China, India, Indonesia, Bangladesh, Vietnam, Thailand, Myanmar, Philippines, Brazil, and Japan (IRRI, 2008; FAO, 2013). In Malaysia, rice is the third most important crop next to rubber and oil palm. Selangor, Kedah, Penang and Perak are the main rice growing states in Peninsular Malaysia. Currently, rice self-sufficiency level is around 73% with an average yield of 3.7 t ha<sup>-1</sup> which is below the potential level (Akinbile *et al.*, 2011).

Yield potential of rice under farmer's field conditions has never been achieved due to be encountered by different biotic and abiotic stresses. Among the biotic stresses, diseases play important roles in reducing the yield. Rice is confronted with more than 70 diseases caused by fungi, bacteria, viruses or nematodes (Manidipa *et al.*, 2013). Out of these, sheath blight caused by *Rhizoctonia solani* AG-1 IA is the most important disease in tropical and subtropical rice growing countries of the world with the incidence of more than 90% (Mathivavan and Shanmugaiah, 2011) incurring yield losses up to 50% annually (Zheng *et al.*, 2013).

To combat the disease, different management practices including host resistance, cultural practices, and chemical and biological control measures are being used. The disease cannot be satisfactorily managed through host resistance alone because of the low level of inherent resistance to this pathogen. Unfortunately, no resistant rice cultivar against the pathogen is available elsewhere (Ou, 1985; Suudi et al., 2013). However, cultural practices have contributed to the management of the disease to some extent. Currently, management of this disease is depended on chemical fungicides. Widespread and indiscriminate use of chemical fungicides has been a serious concern of public health and scrutiny due to hazardous effects on the environment, non-target organisms and possible carcinogenicity of some chemicals (Heydari, 2007; Heydari et al., 2007). Other problems include, development of new resistant races of pathogens (Houssein et al., 2010), plant phytotoxicity (Foster and Hausbeck, 2010) and high cost of chemicals. Increasing awareness of these fungicide-related problems has emphasized the need for non-chemical and ecofriendly disease control methods. In this context, biological control of sheath blight is viewed as a viable and cost effective method with sustainable yield potential.

Biological control using bio-agents is an excellent alternative to chemicals and has been proven successful for controlling plant diseases in many countries. Various fungi from the genus *Aspergillus, Gliocladium, Paecilomyces, and Trichoderma* and bacteria from the genus *Bacillus, Pseudomonas, Serratia, Erwinia, Rhizobium* and *Paenibacillus* are good examples. Among them, bacteria under the genus *Pseudomonas* especially fluorescent pseudomonads are important candidates for biological control and have been successfully used in sheath blight management across the world. The beneficial effects of *P. fluorescens* have been attributed to the production of antibiotics, metabolites, phytohormones, siderophores, lytic enzymes, and hydrogen cyanide (HCN); and to phosphate solubilization, and induction of systemic resistance against different pathogens (Podile and Kishore, 2006)

*Pseudomonas* is known as an aggressive colonizer in both rhizosphere and phyllosphere. A great majority of antagonistic bacteria have been isolated from the rhizosphere, identified and used as biocontrol agents (Manjula *et al.*, 2002; Sessitsch *et al.*, 2004), and a few from the phyllosphere. The cells of microrganism inhabiting the phyllosphere are often exposed to various physical and chemical stresses due to fluctuation of environmental conditions. Thus, they possess the additional advantages in terms of survivability and virulence under stress conditions compared to the rhizosphere. Hence, there is a possibility to explore for more potential bacteria from this habitat. Until now, no indigenous phylloplane bacteria against *Rhizoctonia solani* on rice have been documented in Malaysia. Therefore, this study was designed to isolate antagonistic bacteria from rice plant samples collected from different rice growing regions of Peninsular Malaysia. It was hypothesized that these bacterial antagonists will be the effective biocontrol agents against *R. solani* causing sheath blight in rice.

To accomplish this, laboratory and glasshouse experiments were conducted with the following major objectives-

1) To characterize and identify the antagonistic bacteria isolated from rice plants

2) To determine the modes of action deployed in biocontrol of sheath blight and plant growth promoting activities by selected bacterial strains

3) To determine the efficacy of selected bioformulated bacterial strains against sheath blight under glasshouse conditions

#### REFERENCES

- Abbas, S.J., Ahmad, B. and Karlovsky, P. 2014. Real-time PCR (QPCR) assay for *Rhizoctonia solani* anastomoses group AG2-2 IIIB. *Pakistan Journal of Botany* 46(1): 353-356.
- Abdel-Kader, M.M., El-Mougy, N.S., Aly, M.D.E. and Lashin, S.M. 2012. Long Activity of Stored Formulated Bio-Agents Against Some Soil-Borne Plant Pathogenic Fungi Causing Root Rot of Some Vegetables. *Journal of Applied Sciences Research* 8(4): 1882-1892.
- Abo-Elyousr, K.A.M., Hashem, M. and Ali, E.H. 2009. Integrated control of cotton root rot of disease by mixing fungal biocontrol agents and resistance inducer. *Crop Protection* 28(4): 295-301.
- Adhipathi, P., Singh, V. and Meena, S.C. 2013. Virulence diversity of *Rhizoctonia* solani causing sheath blight disease in rice and its host pathogen interaction. *The Bioscan* 8(3): 949-952.
- Afsharmanesh, H., Ahmadzadeh, M., Javan-Nikkhah, M. and Behboudi, K. 2010. Characterization of the antagonistic activity of a new indigenous strain of *Pseudomonas fluorescens* isolated from onion rhizosphere. *Journal of Plant Pathology* 92(1): 187-194.
- Agrios, G.N. 1997. Plant Pathology (4<sup>th</sup> Edn.). London: Academic Press.
- Ahemad, M. and Khan, M.S. 2012. Effects of pesticides on plant growth promoting traits of *Mesorhizobium* strain MRC4. *Journal of the Saudi Society of Agricultural Sciences* 11(1): 63-71.
- Ahmad, S.G., Garg, V.K., Pandit, A.K., Anwar, A. and Aijaz, A. 2011. Disease Incidence of Paddy Seedlings in Relation to Environmental Factors under Temperate Agroclimatic Conditions of Kashmir Valley. *Journal of Research* and Development 11: 29-38.
- Ajit, N.S., Verma, R. and Shanmugan, V. 2006. Extracellular chitinase of fluorescent pseudomonads antifungal to *Fusarium oxysporum* f.sp. *dianthi* causing carnation wilt. *Current Microbiology* 52(4): 310-316.
- Akinbile, C.O., Abd El-Latif, K.M., Abdullah, R. and Yusoff, M.S. 2011. Rice production and water use efficiency for self-sufficiency in Malaysia: A review. *Trend in Applied Sciences Research* 6(10): 1127-1140.
- Akter, S., Rahman, M.M., Mia, M.A.T., Sharma, N.R. and Kohinoor, H. 2003. Interaction among three species of *Rhizoctonia* causing sheath blight disease complex of rice. *Bangladesh Journal of Plant Pathology* 19(1-2): 1-5.
- Alam, S., Khalil, S., Ayub, N. and Rashid, M. 2002. *In vitro* solubilization of inorganic phosphate by phosphate solubilizing microorganisms (PSM) from

maize rhizosphere. *International Journal of Agriculture and Biology* 4(4): 454-458.

- Al-Hinai, A.H., Al-Sadi, A.M., Al-Bahry, S.N., Mothershaw, A.S., Al-Said, F.A., Al-Harthi S.A. and Deadman, M.L. 2010. Isolation and characterization of *Pseudomonas aeruginosa* with antagonistic activity against *Pythium aphanidermatum. Journal of Plant Pathology* 92(3): 653-660.
- Ali, N.I., Siddiqui, L.A., Shaukat, S.S. and Zaki, M.J. 2001. Survival of *Pseudomonas aeruginosa* in various carriers for the inhibition of root-rot knot disease complex of mung bean. *Phytopathologia Mediterranea* 40(1): 108-112.
- Aliferis, K.A. and Jabaji, S. 2010. Metabolite composition and bioactivity of *Rhizoctonia solani* sclerotial exudates. *Journal of Agricultural and Food Chemistry* 58(13): 7604-7615.
- Anderson, N.A. 1982. The genetics and pathology of *Rhizoctonia solani*. Annual *Review of Phytopathology* 20: 329-347.
- Anitha, G. and Kumudini, B.S. 2014. Isolation and characterization of fluorescent pseudomonads and their effect on plant growth promotion. *Journal of Environmental Biology* 35(4): 627-634.
- Annonymous. 1993. Country report on crop loss due to sheath blight. In *Rice sheath blight management workshop*. China National Rice Research Institute, Hangzhou, China, 11-15 October 1993. Published by IRRI, the Philippines.
- Ardakani, S.S., Heydari, A., Khorasani, N. and Arjmandi, R. 2010a. Development of new bioformulations of *Pseudomonas fluorescens* and evaluation of these products against damping-off of cotton seedlings. *Journal of Plant Pathology* 92(1): 83-88.
- Ardakani, S.S., Heydari, A., Tayebi, L. and Mohammedi, M. 2010b. Promotion of cotton seedling growth characteristics by development and use of new bioformulations. *International Journal of Botany* 6(2): 95-100.
- Arora, N.K., Kim, M.J., Kang, S.C. and Maheswari, D.K. 2007. Role of chitinase and b-1,3-glucanase activities produced by a fluorescent pseudomonad and *in vitro* inhibition of *Phytophthora capsici* and *Rhizoctonia solani*. *Canadian Journal of Microbiology* 53(2): 207-212.
- Arumugam, K., Ramalingam, P. and Appu, M. 2013. Isolation of *Trichoderma* viride and *Pseudomonas fluorescens* organism from soil and their treatment against rice pathogens. *Journal of Microbiology and Biotechnology Research* 3(6): 77-81.
- Ashrafuzzaman, M., Hossen, F.A., Ismail, M.R., Hoque, M.A., Islam, M., Shahidullah, S.M. and Meon, S. 2009. Efficiency of plant growth promoting

Rhizobacteria (PGPR) for the enhancement of rice growth. *African Journal of Biotechnology* 8 (Suppl 7): 1247-1252.

- Bashan, Y. 1998. Inoculants of plant growth promoting rhizobacteria for use in agriculture. *Biotechnology Advances*. 16(4): 729-770.
- Bashan, Y. and de-Bashan, L.E. 2005. Bacteria. In *Encyclopedia of Soils in the Environment*, ed. D. Hillel, pp. 103-115. Oxford, UK: Elsevier.
- Bashan, Y. and de-Bashan, L.E. 2010. Chapter two-How the plant growth-promoting bacterium *Azospirillum* promotes plant growth-a critical assessment. *Advances in Agronomy* 108: 77-136.
- Bashar, M.A., Hossain, M.A., Rahman, M.M., Uddin, M.N. and Begum, M.N. 2010.
  Biological Control of Sheath Blight Disease of Rice by using Antagonistic Bacteria. Bangladesh Journal of Science and Industrial Research 45(3): 225-232.
- Belimov, A., Hontzeas, A., Safronova, N., Demchinskaya, V.I., Piluzza, S.V., Bulitta, G. and Glick, B.R. 2005. Cadmium-tolerant plant growth-promoting bacteria associated with the roots of Indian mustard (*Brassica juncea* L. Czern). Soil Biology and Biochemistry 37(2): 241-250.
- Bertin, J., Hermardinquer, J., Keul, M. and Randles, W.G.L 1971. Atlas of food crops. France: Paris, Mouton.
- Bhattacharyya, P.N. and Jha, D.K. 2012. Plant growth-promoting rhizobacteria (PGPR): Emergence in Agriculture. *World Journal of Microbiology and Biotechnology* 28(4): 1327-1350.
- Bhuvaneswari, V. and Raju, S.K. 2012. Efficacy of New Combination Fungicide against Rice Sheath Blight Caused by *Rhizoctonia solani* (Kuhn). *Journal of Rice Research* 5(1-2): 57-61.
- Biswas, A. 2001. Evaluation of rice germplasm for sheath blight (ShB) disease resistance in West Bengal, India. *Environment and Ecology* 19(1): 247-248.
- Biswas, A. 2006. Efficacy of biotos: a new botanical against sheath blight disease of rice. *Environment and Ecology* 24(Special 2): 484-485.
- Biswas, A. 2007. Evaluation of neem formulations against sheath blight disease of rice. *Indian Journal of Plant Protection* 35(2): 296-298.
- Bogino, P.C., Mercede Oliva, M.D.L., Sorroche, F.G. and Giordano, W. 2013. The Role of Bacterial Biofilms and Surface Components in Plant-Bacteria Associations. *International Journal of Molecular Sciences* 14(8): 15838-15859.

- Bora, T., Ozaktan, H., Gore, E. and Aslan, E. 2004. Biological control of *Fusarium* oxysporum f. sp. melonis by wettable powder formulations of the two strains of *Pseudomonas putida*. Journal of Phytopathology 152(8-9): 471-475.
- Boukaew, S. and Prasertsan, P. 2014. Suppression of rice sheath blight disease using a heat stable culture filtrate from *Streptomyces philanthi* RM-1-138. *Crop Protection* 61: 1-10.
- Bucio, J.L., Cuevas, C.C., Calderon, E.H., Becerra, C.V., Rodriguez, R.F., Rodriguez, L.I.M. and Cantero, E.V. 2007. *Bacillus megaterium* rhizobacteria promote growth and alter root system architecture through an auxin and ethylene independent signaling mechanism in *Arabidopsis thaliana*. *Molecular Plant-Microbe Interactions* 20(2): 207-217.
- Caesar, A.J. and Burr, T.J. 1991. Effect of conditioning, bataine, and sucrose on survival of rhizobacteria in powder formulations. *Applied and Environmental Microbiology* 57: 168-172.
- Cappuccino, J.G. and Sherman, N. 2008. Microbiology of soil. In *Microbiology: A Laboratory Manual*, p. 344. California, USA: The Benjamin/Cummings Publishing Company.
- Carling D.E. 1996. Grouping in *Rhizoctonia solani* by hyphal anastomosis reaction. In *Rhizoctonia species: taxonomy, molecular biology, ecology, pathology and disease control,* ed. B. Sneh, S. Jabaji-Hare, S. Neate, G. Dijst, pp. 35-47. Dordrecht, The Netherlands: Kluwer Academic Publishers.
- Carling, D.E., Baird, R.E., Gitaitis, R.D., Brainard, K.A. and Kuninaga, S. 2002. Characterization of AG-13, a newly reported anastomosis group of *Rhizoctonia solani*. *Phytopathology* 92(8): 893-899.
- Castric, K.F. and Castric, P. 1983. Method for rapid detection of cyanogenic bacteria. *Applied and Environmental Microbiology* 45(2): 701-702.
- Cattelan, A.J., Hartel, P.G. and Fuhrmann, J.J. 1999. Screening for plant growthpromoting rhizobacteria to promote early soybean growth. *Soil Science Society of American Journal* 63(6): 1670-1680.
- Cervantes, C., Chavez, J., Cardova, N.A., De Na Mora, P. and Velasco, J.A. 1986. Resistance to Metal by *Pseudomonas aeruginosa* Clinical Isolates. *Microbiology* 48(196-197): 159-163.
- Chaiharn, M. and Lumyong, S. 2009. Phosphate solubilization potential and stress tolerance of rhizobacteria from rice soil in Northern Thailand. *World Journal of Microbiology and Biotechnology* 25(2): 305-314.
- Chakravarty, G. and Kalita, M.C. 2011. Comparative evaluation of organic formulations of *Pseudomonas fluorescens* based biopesticides and their application in the management of bacterial wilt of brinjal (*Solanum melongena* L.). *African Journal of Biotechnology* 10(37): 7174-7182.

- Chandra, A., Saxena, R., Dubey A. and Saxena, P. 2007. Change in phenylalanine ammonialyase activity and isozyme patterns of polyphenol oxidase and peroxidase by salicylic acid leading to enhanced resistance in cowpea against *Rhizoctonia solani*. *Acta Physiologiae Plantarum* 29(3): 361-367.
- Chang, T.T. 1985. Crop history and genetic conservation in rice-a case study. *Iowa State Journal of Research* 59: 405-55.
- Chen, Y., Zhang, A.F., Wang, W.X., Zhang, Y. and Gao, T.C. 2012. Baseline sensitivity and efficacy of thifluzamide in *Rhizoctonia solani*. Annals of Applied Biology 161: 247-254.
- Chien, C.C., Lin, B.C. and Wu, C.H. 2013. Biofilm formation and heavy metal resistance by an environmental *Pseudomonas* sp. *Biochemical Engineering Journal* 78: 132-137.
- Chincholkar, S.B. and Mukerji, K.G. 2007. Biological Control of Plant Diseases. New York. London: The Haworth Press, Inc.
- Chowdhury, A.K. and Sarkar, S.C. 2006. Performance of different fungicides on the management of sheath blight of rice. *Pesticide Research Journal* 18(2): 157-158.
- Cigdem, K. and Merih, K. 2005. Effect of formulation on the viability of biocontrol agent, *Trichoderma harzianum* conidia. *African Journal of Biotechnology* 85: 483-486.
- Coelho, L., Chellemi, D.O. and Mitchell, D.T. 1999. Effect of solarization and cabbage amendments for the control of *Phytophthora* spp. in North Florida. *Plant Disease* 83: 293-299.
- Compant, S., Duffy, B., Nowak, J., Clément, C. and Barka, E.A. 2005. Use of plant growth-promoting bacteria for biocontrol of plant diseases: principles, mechanisms of action, and future prospects. *Applied and Environmental Microbiology* 71: 4951-4959.
- Cook, R.J. and Baker, F.B. 1983. The Nature and Practice of Biological Control of Plant Pathogens. St. Paul, Minnesota: American Phytopathological Society.
- Christen, C.L. and Marshall, R.T. 1984. Selected properties of lipase and protease of *Pseudomonas fluorescens* 27 produced in four medium. *Journal of Dairy Science* 67: 1680-1687.
- Crowley, D.E. and Kraemer, S.M. 2007. Function of siderophores in the plant rhizosphere. In *The Rhizosphere, Biochemistry and Organic Substances at the Soil-Plant Interface*, ed. R. *Pinton, Z. Varaniniand, and P. Nannipieri*, pp. 73-109. Florida, US: CRC Press (Tylor & Francis Group).

- Da Silva, P. and Nahas, E. 2002. Bacterial diversity in soil in response to different plans, phosphate fertilizers and liming. *Brazilian Journal of Microbiology* 33: 304-310.
- Dalla Serra, M., Menestrina, G., Carpaneto, A., Gambale, F., Fogliano, V. and Ballio, A. 2003. Molecular mechanisms of action of syringopeptins, antifungal peptides from *Pseudomonas syringae* pv. syringae. In *Pore-Forming Peptides and Proteins Toxins*, ed. G. Menestrina, M. Dalla Serra, and P. Lazarovici, pp. 272-295. London, UK: Taylor & Francis.
- Datnoff, L.E. and Rodrigues, F.A. 2005. The Role of Silicon in Suppressing Rice Diseases. *APSnet Features*. doi 10.1094/APSnetFeature-2005-0205.
- Datta, D. and Surajit, K. 1933. Principles and Practices of Rice Production, pp. 152-163. USA: A Wiley - Inter Science Publication.
- De Costa, D.M., Samarasinghe, S.S.T., Dias, H.R.D. and Dissanayake, D.M.N. 2008. Control of rice sheath blight by phyllosphere epiphytic microbial antagonists. *Phytoparasitica* 36(1): 52-65.
- De Lima e Silva, A.A., Ribeiro de Carvalho, M.A., L de Souza, S.A., Teixeira Dias, P.M., da Silva Filho, R.G., de Meirelles Saramago, C.S., de Melo Bento, C.A. and Ernesto Hofer. 2012. Heavy metal tolerance (Cr, Ag and Hg) in bacteria isolated from sewage. *Brazilian Journal of Microbiology* 43(4): 1620-1631.
- Dey, R., Pal, K.K., Bhatt, D.M. and Chauhan, S.M. 2004. Growth promotion and yield enhancement of peanut (*Arachis hypogaea* L.) by application of plant growth-promoting rhizobacteria. *Microbiological Research* 159: 371-394.
- Dezfuli, P.M., Sharif-Zadeh, F. and Janmohammadi, M. 2008. Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays* L.). *ARPN Journal of Agricultural and Biological Science* 3(3): 22-25.
- Diby, P., Anandaraj, M., Kumar, A. and Sarma, Y.R. 2005. Antagonistic mechanisms of fluorescent *Pseudomonas* against *Phytophthora capsici* Leonian in black pepper (*Piper nigrum* L.). *Journal of Spices and Aromatic Crops* 14(2): 94-101.
- DOA. 2011. Department of Agriculture. Keluasan Bertanam dan Pengeluaran Padi Malaysia Mengikut Negeri, 2006-2011.
- Duffy, B.K. and Weller, D.M. 1995. Use of *Gaeumannomyces graminis* var. *graminis* alone and in combination with fluorescent *Pseudomonas* spp. to suppress take-all of wheat. *Plant Disease* 79: 907-911.
- Duffy, B.K., Simon, A. and Weller, D.M. 1996. Combination of *Trichoderma koningii* with fluorescent pseudomonads for control of take-all on wheat. *Phytopathology* 86: 188-194.

- Duijff, B.J., Miejer, J.W., Bakker, P.A.H.M. and Schippers, B. 1993. Siderophore mediated competition for iron and induced resistance in the suppression of Fusarium wilt of carnation by *Pseudomonas* sp. *Netherland Journal of Plant Pathology* 99: 277-289.
- Dunne, C., Moenne-Loccoz, Y., McCarthy, J., Higgins, P., Powell, J., Dowling, D.N. and O'Gara, F. 1998. Combining proteolytic and phloroglucinol producing bacteria for improved biocontrol of *Pythium*-mediated damping-off of sugar beet. *Plant Pathology* 47: 299-307.
- Egambardieva, D. 2010. Growth response of wheat cultivars to bacterial inoculation in calcareous soil. *Plant, Soil and Environment* 56(12): 570-573.
- Ehteshami, S.M.R., Amin-Deldar, Z. and Khavazi, K. 2013. Foliar spraying of different strains of *Pseudomonas fluorescens* on quantitative characteristics, yield and yield components of two rice cultivars under greenhouse conditions. *Journal of Science Technology and Greenhouse Culture* 4(14): 99-107.
- Elangovan, C. and Gnanamanickam, S.S. 1992. Incidence of *Pseudomonas* fluorescens in the rhizosphere of rice and their antagonism towards *Sclerotium oryzae. Indian Phytopathology* 45: 358-361.
- El-Hassan, S.A. and Gown, S.R. 2006. Formulation and delivery of the bacterial antagonist *Bacillus subtilis* for management of lentil vascular wilt caused by *Fusarium oxysporum* f. sp. *lentis. Journal of Phytopathology* 154(3): 148-155.
- Esitken, A., Pirlak L, Turan, M. and Sahin, F. 2006. Effects of floral and foliar application of plant growth promoting rhizobacteria (PGPR) on yield, growth and nutrition of sweet cherry. *Horticultural Science* 110: 324-327.
- Estrada de los Santos, P., Bustillos-Cristales, M.R. and Caballero-Mellado, J. 2001. *Burkholderia*, a genus rich in plant-associated nitrogen fixers with wide environmental and geographic distribution. *Applied and Environmental Microbiology* 67: 2790-2798.
- FAO. 2011. Food and Agriculture Organization of the United Nations. Agricultural Statistics. Downloaded from <u>http://www.faostat.org.</u>
- FAO. 2013. Food and Agriculture Organization of the United Nations. Agricultural Statistics. Downloaded from <u>http://www.faostat.org.</u>
- Ferdous, J., Hanafi, M.M., Rafii, M.Y. and Muhammad, K. 2012. A quick DNA extraction protocol: Without liquid nitrogen in ambient temperature. *African Journal of Biotechnology* 11: 6956-6964.
- Ferdous, M.J. 2012. Investigation of phylloplane bacteria for bio-control of sheath blight of rice. MS Thesis, Bangladesh Agricultural University, Mymensingh, Bangladesh.

- Foster, J.M. and Hausbeck, M.K. 2010. Managing Phytophthora Crown and Root Rot in Bell Pepper Using Fungicides and Host Resistance. *Plant Disease* 94(6): 697-702.
- Gangopadhyay, S. and Chakrabarti, N.K. 1982. Sheath blight of rice. *Review of Plant Pathology* 61: 451-460.
- Garcia, O.A. and Sarmiento, M. 2000. A note on the viability of Azospirillum brasilense in turf used as carrier in inoculated grass seeds. Cuban Journal of Agricultural Science 34: 343-345.
- Gerhardt, P., Murray, R.G.E., Costilow, R.N., Nester, E.W., Wood, W.A., Krieg, N.R. and Phillips, G.B. 1981. Manual of Methods for General Bacteriology. p. 22. Washington, DC: American Society for Microbiology.
- Getha, K. and Vikineswary, S. 2002. Antagonistic effects of *Streptomyces violaceusniger* strain G10 on *Fusarium oxysporum* f.sp. *cubense* race 4: indirect evidence for the role of antibiosis in the antagonistic process. *Journal Industrial Microbiology and Biotechnology* 28: 303-310.
- Glick, B.R., Todorovic, B., Czarny, J., Cheng, Z., Duan, J. and McConkey, B. 2007. Promotion of plant growth by bacterial ACC deaminase. *Critical Reviews in Plant Sciences* 26: 227-242.
- Glick, B.R. 2012. Plant Growth-Promoting Bacteria: Mechanisms and Applications. *Scientifica* 2012, article ID 963401, http://dx.doi.org/10.6064/2012/963401.
- Gochfeld, M. 2003. Cases of mercury exposure, bioavailability, and absorption. *Ecotoxicology and Environmental Safety* 56(1): 174-179.
- Goldstein, A.H. 1986. Bacterial solubilization of mineral phosphates: Historical perspective and future prospects. *American Journal of Alternative Agriculture* 1: 51-57.
- Gonzalez, V.G., Onco, P.M.A. and Susan, V.R. 2006. Review Biology and Systematic of the form genus *Rhizoctonia*. Spanish Journal of Agricultural Research 4(1): 55-79.
- Goswami, B.K., Bhuiyan, K.A. and Mian, I.H. 2010. Morphogenic and pathogenic variations in the isolates of *Rhizoctonia solani* in Bangladesh. *Bangladesh Journal of Agricultural Research* 35(3): 375-380.
- Grist, D.H. 1986. Rice. 6<sup>th</sup> edition. Tropical Agriculture Series, pp. 69-98. Longman, London and New York.

- Groth, D.E. and Bollich, P.K. 2000. The effect of nitrogen rate and fungicide application on disease development and yield of rice. Pages 81–82 in: 28th Proceedings of the Rice Technical Working Group, Mississippi State Univ., Biloxi, MS, 27 Feb.- 1 Mar. 2000. Louisiana State Univ. Agric. Cent. Rice Res. Stn, Crowley, LS.
- Groth, D.E. and Bond, J.A. 2007. Effects of cultivars and fungicides on rice sheath blight, yield and quality. *Plant Disease* 91(12): 1647-1650.
- Guleria, S., Agarwal, R., Thind, T.S. and Sharma, T.R. 2007. Morphological and pathological variability in rice isolates of *Rhizoctonia solani* and molecular analysis of their genetic variability. *Journal of Phytopathology* 155: 645-661.
- Gupta, C.D., Dubey, R.C., Kang, S.C. and Maheshwari, D.K. 2001. Antibiotic mediated necrotrophic effect of *Pseudomonas* GRC2 against two fungal plant pathogens. *Current Science* 81: 91-94.
- Gupta, C.P., Kumar, B., Dubey, R.C. and Maheshwari, D.K. 2006. Chitinase mediated destructive antagonistic potential of *Pseudomonas aeruginosa* GRC1 against *Sclerotinia sclerotiorum* causing charcoal rot of peanut. *BioControl* 51: 821-35.
- Haferburg, G. and Kothe, E. 2010. Metallomics: lessons for metalliferous soil remediation. *Applied Microbiology and Biotechnology* 87: 1271-1280.
- Harvey, J., Keenan, K.P. and Gilmour, A. 2007. Assessing biofilm formation by *Listeria monocytogenes* strains. *Food Microbiology* 24: 380-392.
- Hashiba, T. 1984. Estimating method of severity and yield loss by rice sheath blight disease. *Bulletin of the Hokuriku National Agricultural Experiment Station* 26: 115-164.
- Hayat, R., Ali, S., Amara, U., Khalid, R. and Ahmed, I. 2010. Soil beneficial bacteria and their role in plant growth promotion: a review. *Annals of Microbiology* 60(4): 579-598.doi:10.1007/s13213-010-0117-1.
- Herman, M.A.B., Nault, B.A. and Smart, C.D. 2008. Effects of plant growth promoting rhizobacteria on bell pepper production and green peach aphid infestations in New York. *Crop Protection* 27: 996-1002.
- Heydari, A. 2007. Biological Control of Turfgrass Fungal Diseases. In *Hand Book of Turf grass Management and Physiology*, ed. M. Pessaraki. Florida, USA: CRC Press.
- Heydari, A., Misaghi, I.J. and Balestra, G.M. 2007. Pre-emergence herbicides influence the efficacy of fungicides in controlling cotton seedling damping-off in the field. *International Journal of Agricultural Research* 2: 1049-1053.
- Hien, N.T. 2007. The product BioGro and improvements in its performance. In *Efficient nutrient use in rice production in Vietnam achieved using inoculants*

*biofertilizers*, ed. I.R. Kennedy, A.T.M.A. Choudury, M.L. Kecskes, M.T. Rore, pp: 15-23. Proceedings of a project (SMCN/2002/073). Workshop Held in Hanoi, Vietnam 12-13 October 2007.

- Holt, J.G., Krieg, N.R., Sneath, P.H.A., Staley, J.T. and Williams, S.T. 1984. Bergeys Manual of Determinative Bacteriology. Baltimore, London: USA: Williams and Wilkins.
- Houssien, A.A., Ahmed, S.M. and Ismail, A.A. 2010. Activation of tomato plant defense response against *Fusarium* wilt disease using *Trichoderma harzianum* and salicylic acid under greenhouse conditions. Research Journal of Agriculture and Biological Sciences 6(3): 328-338.
- Hu, X.F., Ying, F.X., He, Y.B., Gao, Y.Y., Chen, H.M. and Chen, J.S. 2008. Characterization of *Pectobacterium carotovorum* sub sp. *carotovorum* causing soft-rot disease on *Pinellia ternata* in China. *European Journal of Plant Pathology* 120: 305-310.
- Huang, J.H., Yang, M. and Zhou, E.X. 2008. Cross pathogenicity of *Rhizoctonia* spp. isolated from thirteen plants on rice, sweet corn, cucumber and cabbage. *Journal of Huazhong Agricultural University* 27(2): 198-203.
- Hughes, M.N. and Poole, R.K. 1989. The functions of metals in microorganisms. In *Metals and Microorganisms*, pp. 22-27. London: Chapman and Hall.
- Hung, P.Q. and Annapurna, K. 2004. Isolation and Characterization of Endophytic Bacteria in Soybean (*Glycine* sp.) *Omonrice* 12: 92-101.
- Husen, E., Wahyudi, A.T., Suwanto, A. and Giyanto. 2011. Soybean Response to 1 Aminocyclopropane-1-Carboxylate Deaminase-Producing *Pseudomonas* under Field Soil Conditions. *American Journal of Agricultural and Biological Sciences* 6(2): 273-278.
- Innerebner, G., Knief, C. and Vorholt, J.A. 2011. Protection of *Arabidopsis thaliana* against leaf-pathogenic *Pseudomonas syringae* by *Sphingomonas* strains in a controlled model system. *Applied and Environmental Microbiology* 77(10): 3202-3210.
- Intana, W., Yenjit, P., Suwanno, T., Sattasakulchai, S., Suwanno, M. and Chamswarng, C. 2008. Efficacy of antifungal metabolites of *Bacillus* spp. for controlling tomato damping-off caused by *Pythium aphanidermatum*. *Walailak Journal of Science and Technology* 5(1): 29-38.
- IRRI. 2008. International Rice Research Institute: IRRI World Rice Statistics (WRS), Facts and Figures: 1960-2010.
- Islam, Z.R., Pamplona, A.D., Atkinson and Azucena, E.J. 2003. Biological control of rice disease. IRRI Rice Knowledge Bank. p. 24.

- ISTA. 1999. International Rules for Seed Testing Association. Seed Science and *Technology* 27: Supplement Rules: 27-31.
- Jamali, F., Sharifi-Tehrani, A, Lutz, M.I. and Maurhofer, M. 2009. Influence of host plant genotype, presence of a pathogen, and co inoculation with *Pseudomonas fluorescens* strains on the rhizosphere expression of hydrogen cyanide and 2, 4-diacetylphloroglucinol biosynthetic genes in *P. fluorescens* biocontrol strain CHA0. *Microbial Ecology* 57: 267-275.
- Jambhulkar, P.P. and Sharma, P. 2013. Promotion of rice seedling growth characteristics by development and use of bioformulation of *Pseudomonas fluorescens. Indian Journal of Agricultural Sciences* 83(2): 136-42.
- Jayaprakashvel, M. and Mathivanan, N. 2012. Morphological and pathological variations of rice sheath blight inciting south Indian *Rhizoctonia solani* isolates. *Archives of Phytopathology and Plant Protection* 45(4): 455-467.
- Jayaraj, J., Parthasarathi, T. and Radhakrishnan, N.V. 2007. Characterization of a *Pseudomonas fluorescens* strain from tomato rhizosphere and its use for integrated management of tomato damping-off. *Biocontrol* 52: 683-702.
- Jayaraj, J., Radhakrishnana, N.V., Kannan, R., Saktivel, K., Suaganaya, D., Venkatesan, S. and Velazhahan, R. 2005. Development of new formulations of *Bacillus* for management of tomato damping-off caused by *Pythium aphanidermatum*. *Biocontrol Science and Technology* 15: 55-65.
- Jensen, P.R. and Fenical, W. 1994. Strategies for the discovery of secondary metabolites from maraine bacteria. *Annual Review of Microbiology* 48: 559-684.
- Joshi, N., Brar, K.S., Pannu, P.P.S. and Singh, P. 2007. Field efficacy of fungal and bacterial antagonists against brown spot of rice. *Journal of Biological Control* 21(1): 159-162.
- Kalita, M.K., Pathak, K. and Saikia, M.K. 2000. Reaction of some rice cultivars against leaf and sheath blight of rice (*Rhizoctonia solani*) in southern Assam. *Annals of Agri Bio Research* 5(2): 163-164.
- Kamala, T. and Indra Devi, S. 2012. Biocontrol properties of indigenous *Trichoderma* isolates from North-East India against *Fusarium oxysoprum* and *Rhizoctonia solani*. *African Journal of Biotechnology* 11(34): 8491-8499.
- Kamaluddeen and Simon, S. 2013. Effect of soil solarization, bio-agents and organic composts on blast of paddy. *International Journal of Botany and Research* 4(3): 21-28.
- Kamensky, M., Ovadis, M., Chet, I. and Chernin, L. 2003. Soil-borne strain IC14 of Serratia plymuthica with multiple mechanisms of antifungal activity provides biocontrol of Botrytis cinerea and Sclerotinia sclerotiorum diseases. Soil Biology and Biochemistry 35: 323-331.

- Kandhari, J., Gupta, R.L. and Kandhari, J. 2003. Efficacy of fungicides and resistance inducing chemicals against sheath blight of rice. *Journal of Mycological Research* 41: 67-69.
- Kannapiran, E. and Sri Ramkumar, V. 2011. Isolation of phosphate solubilizing bacteria from sediments of Thondi coast, Palk Strait, Southeast coast of India. *Annals of Biological Research* 25: 157-163.
- Kararah, M.A., Barakat, F.M., Mikhail, M.S. and Fouly, H.M. 1985. Pathophysiology in garlic cloves inoculated with *Bacillus subtilis*, *Bacillus pumilus* and *Erwinia carotovora*. *Egyptian Journal of Phytopathology* 17(2): 131-140.
- Karnwal, A. 2009. Production of indole acetic acid by fluorescent *Pseudomonas* in the presence of L-Tryptophan and rice root exudates. *Journal of Plant Pathology* 91 (Suppl 1): 61-63.
- Karpagam, T. and Nagalakshmi, P.K. 2014. Isolation and Characterization of Phosphate Solubilizing Microbes from Agricultural soil. *International Journal of Current Microbiology and Applied Sciences* 3(3): 601-614.
- Karthikeyan, A. and Chandrasekaran, B. 2007. Bio efficacy of plant extracts in the management of sheath blight disease of rice. *Journal of Ecobiology* 21(3): 279-282.
- Karthikeyan, M., Radhika, K., Bhaskaran, R., Mathiyazhagan, S., Sandosskumar, R., Velazhahan, R. and Alice, D. 2008. Biological control of onion leaf blight disease by bulb and foliar application of powder formulation of antagonist mixture. Archives of Phytopathology and Plant Protection 41(6): 407-417.
- Karthikeyan, M., Radhika, K., Mathiyazhagan, S., Bhaskaran, R., Samiyappan, R. and Velazhahan, R. 2006. Induction of phenolics and defense-related enzymes in coconut (*Cocos nucifera* L.) roots treated with biological agents. *Brazilian Journal of Plant Physiology* 18(3): 367-377.
- Kavitha, K., Nakkeeran, S., Chadrasekar, G., Fernanado, W.G.D., Mathiyazhagan, S., Renukadevi, P. and Krishnamoorthy, A.S. 2003. Role of antifungal antibiotics, siderophores and IAA production in biocontrol of *Pythium aphanidermatum* inciting damping off in tomato by *Pseudomonas chlororaphis* and *Bacillus subtilis*. In *Proceedings of the 6<sup>th</sup> International Workshop on PGPR*, pp. 493-497. Organized by IISR, Calicut 5-10 October, 2003.
- Kazempour, M.N. 2004. Biological control of *Rhizoctonia solani* the causal agent of sheath blight of rice by antagonistic bacteria in greenhouse and field condition. *Plant Pathology Journal* 3(2): 88-96.
- Khabbaz, S.E. and Abbasi, P.A. 2014. Isolation, characterization, and formulation of antagonistic bacteria for the management of seedlings damping-off and root

rot disease of cucumber. *Canadian Journal of Microbiology* 60(1): 25-33 (doi: 10.1139/cjm-2013-0675).

- Khan, A.A. and Sinha, A.P. 2006. Influence of formulations, doses and time of application of fungal bio-agents on rice sheath blight. *Annals of Plant Protection Sciences* 14(1): 157-161.
- Khan, A.A. and Sinha, A.P. 2007. Biocontrol potential of *Trichoderma* species against sheath blight of rice. *Indian Phytopathology* 60(2): 208-213.
- Khan, M.S., Zaidi, A. and Wani, P.A. 2006. Role of phosphate solubilizing microorganisms in sustainable agriculture- a review. Agronomy for *Sustainable Development* 27: 29-43.
- Khan, M.S., Zaidi, A., Wani, P.A. and Oves, M. 2009. Role of plant growth promoting rhizobacteria in the remediation of metal contaminated soils. *Environmental Chemistry Letters* 7(1): 1-19.
- Khorshidi, Y.R., Ardakani, M.R., Ramezanpour, M.R., Khavazi, K. and Zargari, K. 2011. Response of yield and yield components of rice (*Oryza sativa* L.) to *Pseudomonas flourescens* and *Azospirillum lipoferum* under different nitrogen levels. *American-Eurasian Journal of Agricultural and Environmental Science* 10(3): 387-395.
- Khush, G.S. 1997. Origin, dispersal, cultivation and variation of rice. *Plant Molecular Biology* 35(1&2): 25-34.
- Kidarsa, T.A., Goebel, N.C., Zabriskie, T.M. and Loper, J.E. 2011. Phloroglucinol mediates cross-talk between the pyoluteorin and 2, 4-diacetylphloroglucinol biosynthetic pathways in *Pseudomonas fluorescens* Pf-5. *Molecular Microbiology* 81(2): 395-414.
- Kidoglu, F., Gul, A., Ozaktan, H. and Tuzel, Y. 2007. Effect of rhizobacteria on plant growth of different vegetables. ISHS Acta Horticulturae 801: International Symposium on High Technology for Greenhouse System Management.
- Kim, J. and Rees, D.C. 1994. Nitrogenase and biological nitrogen fixation. *Biochemistry* 33(2): 389-397.
- Kishore, G.K. Pande, S. and Podile, A.R. 2005. Phylloplane bacteria increase seedling emergence, growth and yield of field-grown groundnut (*Arachis hypogaea* L.). *Letters in Applied Microbiology* 40: 260-268.
- Kloepper, J.W. and Schroth, M.N. 1981. Development of powder formulation of rhizobacteria for inoculation of potato seed pieces. *Phytopathology* 71(6): 590-592.

- Krishnamurthy, K. and Gnanamanickam, S.S. 1998. Biological control by *Pseudomonas fluorescens* strain pf7-14: Evaluation of a marker gene and formulations. *Biological Control* 13(3): 158-165.
- Kumar, A.A., Kumar, S., Devi, S., Patil, C., Payal, C. and Sushila Negi, S. 2012c. Isolation, screening and characterization of bacteria from rhzospheric soils for different plant growth promotion (PGP) activities: an *in vitro* study. *Recent Research in Science and Technology* 4(1): 1-5.
- Kumar, C.S., Harikrishnan, H., Charulatha, R. and Shanmugaiah, V. 2013. Evaluation of *Pseudomonas* sp. VSMKU-4046 for Suppression of Sheath Blight (ShB of Rice in Detached Leaf Assay. In *Prospects in Bioscience: Addressing the issues*, ed. A. Sabu and Augustine, Springer India. DOI10.10077978-81-322-0810-5\_33 2013.
- Kumar, M., Singh, V., Singh, K.N. and Vikram, P. 2008. Morphological and virulence characterization of *Rhizoctonia solani* causing sheath blight of rice. *Environment and Ecology* 26(3): 1158-1166.
- Kumar, M.K.P., Gowda, D.K.S., Gowda K.T.P. and Vishwanath, K. 2012a. A New Carboxynilide Group Fungicide against Paddy Sheath Blight. *Research Journal of Agricultural Sciences* 3(2): 500-505.
- Kumar, K.V.K., Yellareddygari, S.K.R., Reddy, M.S., Kloepper, J.W. Lawrence, K.S. Zhou, X.G., Sudini, H., Groth, D.E., Raju, S.K. and Miller, M.E. 2012b. Efficacy of *Bacillus subtilis* MBI 600 against sheath blight caused by *Rhizoctonia solani* and on growth and yield of rice. *Rice Science* 19(1): 55-63.
- Lakshmipriya, V.P. and Sivakumaar, P.K. 2013. Optimization of certain growth parameters for the production of exopolysaccharides from *Azotobacter* species isolated from mangrove ecosystem. *Research Journal of Biological Sciences* 5(1): 27-33.
- Latha, P, Anand, T., Ragupathi, N., Prakasam, V. and Samiyappan, R. 2009. Antimicrbial activity of plant extracts and induction of systemic resistance in tomato plants by mixtures of PGPR strains and zimmu leaf extract against *Alternaria solani. Biological Control* 50: 85-93.
- Latif, M.A., Rafii Yusop, M., Motiur Rahman, M. and Bashar Talukdar, M. 2011. Microsatellite and minisatellite markers based DNA fingerprinting and genetic diversity of blast and ufra resistant genotypes. *Comptes Rendus Biology* 334(4): 282-289.
- Lee, F.N. and Rush, M.C. 1983. Rice Sheath blight: A major rice disease. *Plant Disease* 67: 829-832.
- Lelliott, R.A. and Stead, D.E. 1987. Methods for the Diagnosis of Bacterial Diseases of Plants. 1<sup>st</sup> Edn. p. 216. London: Blackwell Scientific Publications.

- Leong, J. 1986. Siderophores; their biochemistry and possible role in the biocontrol of plant pathogens. *Annual Review of Phytopathology* 24: 187-209.
- Li, J.G., Jiang, Z.Q., Xu, L.P., Sun, F.F. and Guo, J.H. 2008. Characterization of chitinase secreted by *Bacillus cereus* strain CH2 and evaluation of its efficacy against Verticillium wilt of eggplant. *Biocontrol* 53: 931-944.
- Lindow, S.E. and Brandl, M.T. 2003. Microbiology of the phyllosphere. *Applied and Environmental Microbiology* 69: 1875-1883.
- Liu, T., Shao, D., Kovi, M.R. and Xing, Y. 2010. Mapping and validation of QTL for spikelets per panicle and 1000-grain weight in rice (*Oryza sativa* L.). *Theoritical and Applied Genetics* 120: 933-942.
- Luo JinYan., Xie GuanLin., Li Bin., Luo Yuan Chan., Zhao LiHan., Wang Xiao., Liu Bo., and Li Wen. 2005. Gram-positive bacteria associated with rice in China and their antagonists against the pathogens of sheath blight and bakanae disease in rice. *Rice Science* 12(3): 213-218.
- Luong, M.C. and Heong, K.L. 2005. Effects of organic fertilizers on insect pest and diseases of rice. *Omonrice* 13: 26-33.
- Manidipa, R., Dutta, S.G. and Venkata, R.C. 2013. Pseudomonads: Potential Biocontrol agents of Rice Diseases. *Research Journal of Agriculture and Forestry Sciences* 1(9): 19-25.
- Manjula, K., Singh S.D. and Kishore, G.K. 2002. Role of endophytic bacteria in biological control of plant diseases. *Annual Review of Plant Pathology* 1: 231-252.
- Mathivanan, N. and Shanmugaiah, V. 2011. Management of sheath blight disease in rice by *Pseudomonas aeruginosa* MML2212. Proceedings of the 2nd Asian PGPR Conference, August 21-24, 2011, Beijing, China.
- Mathivanan, N., Prabavathy, V.R. and Vijayanandaraj, V.R. 2005. Application of talc formulations of *Pseudomonas fluorescens* Migula and *Trichoderma viride* Pers ex S.F. gray decrease the sheath blight and enhance the plant growth and yield in rice. *Journal of Phytopathology* 153(11-12): 697-701.
- Maurhofer, M., Keel, C., Haas, D. and Defago, G. 1995. Influence of plant species on disease suppression by *Pseudomonas fluorescens* strain CHA0 with enhanced antibiotic production. *Plant Pathology* 44(1): 40-50.
- Meena, B. 2011. Effect of *Pseudomonas fluorescens* Pfl formulation application on rhizosphere and phyllosphere population in groundnut. *International Journal of Plant Protection* 4 (1): 92-94.
- Meera, T. and Balabaskar, P. 2012 Isolation and characterization of *Pseudomonas* fluorescens from rice fields. International Journal of Food, Agriculture and Veterinary Sciences 2(1): 113-120.

- Mian, M.S., Akter, S., Ali, M.A. and Mia, M.A.T. 2004. Evaluation of some chemicals against sheath blight of rice. *Bangladesh Journal of Plant Pathology* 20(1-2): 59-61.
- Miller and Marvin, J. 2008. Siderophores (microbial iron chelators) and siderophoredrug conjugates (new methods for microbially selective drug delivery). University of NotreDame. Dame, 4/21/2008. <u>http://www.nd.edu/~mmiller1/page2.html</u>
- Miller, J.M. and Rhoden, D.L. 1991. Preliminary evaluation of Biolog, a carbon source utilization method for bacterial identification. *Journal of Clinical Microbiology* 29(6): 1143-1147.
- Min, C. and Hui, H.X. 2006. The research exploration to the effect of controlling rice sheath blight with *Bacillus* spp. Drt-11. *Southwest China Journal of Agricultural Sciences* 19(1): 53-57.
- Minorsky, P.V. 2008. On the inside. Plant Physiology 146: 323-324.
- Mirza, M.S., Mehnaz, S., Normand, P., Prigent-Combaret, C., Moenne-Loccoz, Y., Bally, R. and Malik, K.A. 2006. Molecular characterization and PCR detection of a nitrogen fixing *Pseudomonas* strain promoting rice growth. *Biology and Fertility of Soils* 43(2): 163-170.
- Mishra, D.S., Gupta, A.K., Prajapati, C.R and Singh, U.S. 2011. Combination of fungal and bacterial antagonists for management of root and stem rot disease of soybean. *Pakistan Journal of Botany* 43(5): 2569-2574.
- Mishra, M., Kumar, U., Mishra, P.K. and Prakash, V. 2010. Efficiency of plant growth promoting rhizobacteria for the enhancement of *Cicer arietinum* L growth and germination under salinity. *Advances in Biological Research* 4(2): 92-96.
- Molina, M.A., Godoy, P., Ramos-Gonzalez, M.I., Munoz, N., Ramos, J.L. and Espinosa Ergel, M. 2005. Role of iron and the TonB system in colonization of corn seeds and roots by *Pseudomonas putida* KT2440. *Environmental Microbiology* 7(3): 443-449.
- Montealegre, J., Reyes, R., Perez, L.M., Herrera, R. and Silva, P. 2003. Selection of bioantagonistic bacteria to be used in biological control of *Rhizoctonia solani* in tomato. *Electronic Journal of Biotechnology* 6(2): 115-127.
- Morath, S.U., Hung, R. and Bennett, J.W. 2012. Fungal volatile organic compounds: a review with emphasis on their biotechnological potential. *Fungal Biology Review* 26(2-3): 73-83.
- Motsara, M., Bhattacharyya, P. and Stava, B. 1995. Biofertilizer technology, marketing and usage-a source bookcum-glossary fertilizer development and consultation organization, New Delhi, India.

- Mubarik, N.R., Mahagiani, I. Anindyaputri, A., Santoso, S. and Rusmana, I. 2010. Chitinolytic bacteria isolated from chili rhizosphere: Chitinase characterization and its application as biocontrol for white fly (*Bemisia tabaci* Genn.). *American Journal of Agricutural and Biological Sciences* 5: 430-435.
- Mugilan, I., Gayathri, P., Elumalai, E.K. and Elango, R. 2011. Studies on improve Survivability and Shelf Life of Carrier Using Liquid Inoculation of *Pseudomonas striata. International Journal of Pharmaceutical & Biological Archives* 2(4): 1271-1275.
- Muthukumar, A., Eswaran, A., Nakkeeran, S. and Sangeetha, G. 2010. Efficacy of plant extracts and biocontrol agents against *Pythium aphanidermatum* inciting chilli damping-off. *Crop Protection* 29: 1483-1488.
- Nakkeeran, S., Fernando Dilantha, W.G. and Siddiqui, A.Z. 2005. Plant growth promoting rhizobacteria formulations and its scope in commercialization for the management of pests and diseases. In *PGPR: Biocontrol and Biofertilization* ed. Z.A. Siddiqui, pp. 257-296. Dordrecht, The Netherlands: Springer.
- Nakkeeran, S., Kavitha, K., Mathiyazhagan, S., Fernando, W.G.D., Chandrasekar, G. and Renukadevi, P. 2004. Induced systemic resistance and plant growth promotion by *Pseudomonas chlororaphis* strain PA-23 and *Bacillus subtilis* strain CBE4 against rhizome rot of turmeric (*Curcuma longa L.*). *Canadian Journal of Plant Pathology* 26: 417-418.
- Nandakumar, R., Babu, S., Viswanathan, R., Raguchander, T. and Samiyappan, R. 2001a. Induction of systemic resistance in rice against sheath blight disease by *Pseudomonas fluorescens*. Soil Biology and Biochemistry 33(4-5): 603-612.
- Nandakumar, R., Babu, S., Viswanathan, R., Sheela, J., Raguchander, T. and Samiyappan, R. 2001b. A new bio-formulation containing plant growth promoting rhizobacterial mixture for the management of sheath blight and enhanced grain yield in rice. *Biocontrol* 46(4): 493-510.
- Narayana Swamy, H., Sannaula, S. and Kumar, M.D. 2009. Screening of new fungicides against rice sheath blight disease. *Karnataka Journal of Agricultural Science* 22(2): 448-449.
- Nathan, P., Rathinam, X., Kasi, M., Abdul Rahman, Z. and Subramanian, S. 2011. A pilot study on the isolation and biochemical characterization of *Pseudomonas* from chemical intensive rice ecosystem. *African Journal of Biotechnology* 10(59): 12653-12656.
- Nautiyal, S. 1999. An efficient microbiological growth medium for screening phosphate solubilizing microorganisms. *FEMS Microbiology Letters* 170(1): 265-270.

- Neilands, J.B. 1995. Siderophores: Structure and Function of Microbial Iron Transport Compounds. *The Journal of Biological Chemistry* 270 (Suppl 45): 26723-26726.
- Neuhaus, J.M. 1999. Plant chitinases. In: *Pathogenesis related Proteins in Plants*, ed. S. Muthukrishnan, S.K. Datta, pp. 77-104. Washington, DC, USA: CRC Press.
- Noori, M.S.S. and Saud, H.M. 2012. Potential Plant Growth-Promoting Activity of *Pseudomonas* sp. Isolated from Paddy Soil in Malaysia as Biocontrol Agent. *Journal of Plant Pathology and Microbiology* 3:120. doi:10.4172/2157-7471.1000120.
- O'Brien, M. and Colwell, R. 1987. A Rapid Test for Chitinase Activity that uses 4-Methylumbelliferyl-N-Acetyl-β-D-Glucosaminide. *Applied and Environmental Microbiology* 53(7): 1718-1720.
- O'Sullivan, D.J. and O'Gara, F. 1992. Traits of fluorescent *Pseudomonas* spp. involved in suppression of plant root pathogens. *Microbiological Reviews* 56: 662-676.
- Oh, B.T., Hur, H., Lee, K.J., Shanthi, K., Byoung-Yul Soh, B.Y., Lee, W.J., Myung, H. and Kannan, S.K. 2011. Suppression of Phytophthora blight on pepper (*Capsicum annuum* L) by bacilli isolated from brackish environment. *Biocontrol Science and Technology* 21(11): 1297-1311.
- Ou, S.H. 1985. Rice Diseases. 2nd ed., pp. 272-286. Kew, Surrey, England: Commonwealth Mycological Institute.
- Ownley, B.H., Duffy. B.K. and Weller, D.M. 2003. Identification and manipulation of soil properties to improve the biological control performance of phenazineproducing *Pseudomonas fluorescens*. *Applied and Environmental Microbiology* 69: 3333-3343.
- Pan, S.Q., Ye, X.S. and Kuc, J. 1991. Association of  $\beta$ -1,3-glucanase Activity and Isoform Pattern with Systemic Resistance to Blue Mold in Tobacco Induced by Stem Injection with *Peronospora tabacina* or Leaf Inoculation with Tobacco Mosaic Virus. *Physiological and Molecular Plant Pathology* 39(1): 25-39.
- Pandey, A., Sharma, E. and Palni, L.M.S. 1998. Influence of bacterial inoculation on maize in upland farming systems of the Sikkim Himalaya. *Soil Biology and Biochemistry* 30(3): 379-384.
- Pandey, S., Gauchan, D., Malabayabas, M., Bool-Emrick, M. and Hardy, B. 2012. Patterns of adoption of improved rice varities and farm level impacts in stress-prone rainfed areas in South Asia. International Rice Research Institute.

- Panhwar, Q.A., Othman, R., Rahman, A.A., Meon, S. and Ismail, M.R. 2012. Isolation and characterization of phosphate-solubilizing bacteria from aerobic rice. *African Journal of Biotechnology* 11(11): 2711-2719.
- Panhwar, Q.A., Radziah, O., Zaharah Rahman, A., Sariah, M., Mohd Razi, I. and Naher, U.A. 2011. Contribution of phosphate-solubilizing bacteria in phosphorus bioavailability and growth enhancement of aerobic rice. *Spanish Journal of Agricultural Research* 9(3): 810-820.
- Paramageetham, C. and Prasada Babu, G. 2012. Antagonistic Activity of Fluorescent Pseudomonads against a Polyphagous Soil Born Plant Pathogen-*Sclerotium rolfsii*. 1: 436. doi:10.4172/scientificreports.436.
- Pareja, L., Fernandez-Alba, A.R., Cesio, V. and Heinzen, H. 2011. Analytical methods for pesticide residues in rice. *Trends in Analytical Chemistry* 30(2): 270-291.
- Parmeter, J.R.Jr., Sherwood, R.T. and Platt, W.D. 1969. Anastomosis grouping among isolates of *Thanatephorus cucumeris*. *Phytopathology* 59: 1270-1278.
- Parsek, M.R. and Fuqua, C. 2003. Biofilms, emerging themes and challenges in studies of surface-associated microbial life. *Journal of Bacteriology* 186: 4427-4440.
- Pathma, J., Kennedy, R.K. and Sakthivel, N. 2011. Mechanisms of fluorescent pseudomonads that mediate biological control of phytopathogens and plant growth promotion of crop plants. In *Bacteria in agrobiology: plant growth responses*, ed. D.K. Maheshwari, pp. 77-105. Berlin: Springer. doi: 10.1007/978-3-642-20332-9-4.
- Patricio, F.R.A., Kimati, H., Neto, J.T., Petenatti, A. and Barros, B.C. 2007. Soil solarization under greenhouse and field conditions to the control of *Rhizoctonia solani* AG-4. *Summa Phytopathologica* 33(3): 245-251.
- Patten, C.L. and Glick, B.R. 1996. Bacterial biosynthesis of indole 3-acetic acid. *Canadian Journal of Microbiology* 42(3): 207-220.
- Perez-Garcia, A., Romero, D. and Vicente, A. 2011. Plant Protection and Growth Stimulation by Microorganisms: Biotechnological Applications of Bacilli in Agriculture. *Current Opinion in Biotechnology* 22(2): 187-193.
- Pengnoo, A., Kusongwiriyawong, C., Nilratana, L. and Kanjanamaneesathian, M. 2000. Greenhouse and field trials of the bacterial antagonists in pellet formulations to suppress sheath blight of rice caused by *Rhizoctonia solani*. *BioControl* 45(2): 245-256.
- Phillips, K., Zaidan, F., Elizondo, O.R. and Lowe, K.L. 2012. Phenotypic characterization and 16S rDNA identification of culturable non-obligate halophilic bacterial communities from a hypersaline lake, La Sal del Rey, in extreme South Texas (USA). *Aquative Biosystems* 8.10.1186/2046-9063-8-5.

- Pimentel, J.P., Olivares, F., Pitard, R.M., Urquiaga, S., Akiba, F. and D'obereiner, J. 1991. Dinitrogen fixation and infection of grass leaves by *Pseudomonas rubrisubalbicans* and *Herbaspirillum seropedicae*. *Plant and Soil* 137 (1): 61-65.
- Pleban, S., Chernin, L. and Chet, I. 1997. Chitinolytic activity of an endophytic strain of *Bacillus cereus*. *Letters in Applied Microbiology* 25(4): 284-288.
- Podile, A.R. and Kishore, G.K. 2006. Plant growth promoting rhizobacteria. In *Plant Associated Bacteria*, ed. S.S. Gnanamanickam, pp. 195-230. Netherland: Springer.
- Ponmurugan, P. and Gopi, C. 2006. Distribution pattern and screening of phosphate solubilizing bacteria isolated from different food and forage crops. *Journal of Agronomy* 5(4): 600-604.
- Postma, J., Scheper, R.W.A. and Schilder, M.T. 2010. Effect of successive cauliflower plantings and *Rhizoctonia solani* AG 2-1 inoculations on disease suppressiveness of a suppressive and a conducive soil. *Soil Biology and Biochemistry* 42(5): 804-812.
- Pragash, G., Badri Narayanan, M.K., Ravindra Naik, P. and Sakthivel, N. 2009. Characterization of *Chryseobacterium aquaticum* strain PUPC1 producing a novel antifungal protease from rice rhizosphere soil. *Journal of Microbiology Biotechnology* 19(1): 99-107.
- Prasad, B. and Eizenga, G.C. 2008. Rice sheath blight disease resistance identified in *Oryza* spp. accessions. *Plant Disease* 92(11): 1503-1509.
- Prasad, P.S., Naik, M.K., Thimmegowda, P.R. and Nagaraju, P. 2010. Evaluation of rice genotypes against *Rhizoctonia solani*. *Journal of Plant Disease Sciences* 5(1): 61-64.
- Priya, C.S. and Kalaichelvan, P.T. 2013. Strategies for Antagonistic Activity of Local Actinomycete Isolates against Rice Fungal Pathogens. *Asian Journal of Experimental Biological Sciences* 2(4): 648-653.
- Rabindran, R. and Vidhyasekaran, P. 1996. Development of a formulation of *Pseudomonas fluorescens* PfALR2 for management of rice sheath blight. *Crop Protection* 15: 715-721.
- Radhajeyalakshmi, R., Meena, B., Thangavelu, R. Deborah, S.D., Vidhyasekaran, P. and Velazhahan, R. 2000. A 45 kDa chitinase purified from pearl millet (*Pennisetum glaucum* L.) (R. Br.) shows antifungal activity. *Journal of Plant Diseases and Protection* 107: 605-616.
- Rahimi, S., Wright, D.J. and Perry, R.N. 1998. Identification and localization of chitinase induced in the roots of potato plants infected with the potato cyst nematode *Globodera pallida*. *Fundamental and Applied Nematology* 21: 705-713.

- Rajkumar, M., Ae, N., Prasad, M.N.V. and Freitas, H. 2010. Potential of siderophore-producing bacteria for improving heavy metal phytoextraction. *Trends in Biotechnology* 28(3): 142-149.
- Ramezanpur, M.R., Popov, Y., Khavazi, K. and Rahmani, H.A. 2010. Genetic Diversity and Efficiency of Indole Acetic Acid Production by the Isolates of Fluorescent *Pseudomonas* from Rhizosphere of Rice (*Oryza Sativa* L). *American-Eurasian Journal of Agricultural and Environmental Sciences* 7(1): 103-109.
- Ranjan Nath., Laha, S.K., Bhattacharya, P.M. and Dutta, S. 2005. Evaluation of new fungicidal formulation for controlling the rice sheath blight disease. *Journal* of Mycopathological Research 43(1): 113-115.
- Rangarajan, S., Saleena, L.M. and Nair, S. 2002. Diversity of *Pseudomonas* spp. isolated form rice rhizosphere populations grown along a salinity gradient. *Microbial Ecology* 43(3): 280-289.
- Rangarajan, S., Saleena, L.M., Vasudevan, P. and Nair, S. 2003. Biological suppression of rice diseases by *Pseudomonas* spp. under saline soil conditions. *Plant and Soil* 251(1): 73-82.
- Rasouli, M.H., Khavazi, K., Rahimian, H., Malakouti, M.J. and Asadi-Rahmani, H. 2005. An evaluation of the potential of indigenous Fluorescent Pseudomonds of wheat rhizosphere for producing siderophore. *Iran Journal of Soil and Water Science* 20(1): 133-143.
- Reddy, B.P. Rani, J., Reddy, M.S. and Kumar, K.V.K. 2010a. Isolation of siderophore- producing strains of rhizobacterial fluorescent *pseudomonads* and their biocontrol against rice fungal pathogens. *International Journal of Applied Biology and Pharmaceutical Technology* 1(1): 133-137.
- Reddy, B.P., Rani, J., Reddy, M.S. and Kumar, K.V.K. 2010b. *In-vitro* antagonistic potential of *Pseudomonas fluorescens* isolates and their metabolites against rice sheath blight pathogen *Rhizoctonia solani*. *International Journal of Applied Biology and Pharmaceutical Technology* 1(2): 676-679.
- Redly, G.A. and Poole, K. 2003. Pyoverdine-mediated regulation of FpvA synthesis in *Pseudomonas aeruginosa*: involvement of a probable extracytoplasmicfunction sigma factor. *Journal of Bacteriology* 185(4):1261-1265.
- Reissig, W.H., Heinrichs, E.A., Litssinger, J.A., Moody, K., Fiedler, L., Mew, T.W. and Barrion, A.T. 1986. Illustrated guide to integrated pest management in rice. IRRI, Philippines.
- Reynolds, J. 2004. Lab Procedures Manual: Biochemical tests. Richland College. http://www.rlc. dcccd.edu/mathsci/Reynolds /micro/lab\_manual/TOC.html.

- Ramey, B.E., Koutsoudis, M., von Bodman, B.S. and Fuqua, C. 2004. Biofilm formation in plant-microbe associations. *Current Opinion in Microbiology* 7(6): 602-609.
- Rosales, A.M., Vantomme, R., Swings, J., De ley, J. and Mew, T.W. 1993. Identification of some bacteria from paddy antagonistic to several rice fungal pathogens. *Journal of Phytopathology* 138(3): 189-208.
- Roussel-Delif, L., Tarnawski, S., Hamelin, J., Pjilippot, L., Aragno, M. and Fromin, N. 2005. Frequency and diversity of nitrate reductase genes among nitratedissimilating *Pseudomonas* in the rhizosphere of perennial grasses grown in field conditions. *Microbial Ecology* 49(1): 63-72.
- Sahaf, B.Z., Moharramipour, S. and Meshkatalsadat, M.H. 2007. Chemical constituents and fumigant toxicity of essential oil from *Carum copticum* against two stored product beetles. *Insect Science* 14(3): 213-218.
- Saikia, R., Singh, B., Kumar, R. and Aroram, D.K. 2005. Detection of pathogenesisrelated proteins-chitinase and  $\beta$ -1,3-glucanse in induced chickpea. *Current Science* 89: 659-663.
- Salaheddin, K., Valluvaparidasan, V., Ladhalakshmi, D. and Velazhahan, R. 2010. Management of bacterial blight of cotton using a mixture of *Pseudomonas fluorescens* and *Bacillus subtilis*. Journal Plant Protection Sciences 46: 41-50.
- Sallam, N.A., Riad, S.N., Mohamed, M.S. and El-Eslam, A.S. 2013. Formulations of Bacillus spp. and Pseudomonas fluorescens for biocontrol of cantaloupe root rot caused by Fusarium solani. Journal of Plant Protection Research 53(3): 295-300.
- Santoyo, G., Orozco-Mosqueda, M.C. and Govindappa, M. 2012. Mechanisms of biocontrol and plant growth-promoting activity in soil bacterial species of *Bacillus* and *Pseudomonas*: a review. *Biocontrol Science and Technology* 22(8): 855-872.
- Saraf, M., Thakkar, A., Pandya, U., Joshi, M. and Parikh, J. 2013. Potential of plant growth promoting microorganisms as biofertilizers and biopesticides and it's exploitation in sustainable agriculture. *Journal of Microbiology and Biotechnology Research* 3(5): 54-62.
- Saranya Devi, K. and Sowndaram, S. 2014. Biological control of rhizobacteria against rice diseases caused by *Rhizoctonia solani* (Sheath blight) and *Sarocladium oryzae* (Sheath rot). *International Journal of Advanced Research* 2(5): 818-823.
- Sarkar, A., Islam, T., Biswsh, G.C, Alam, S., Hossain, M. and Talukdar, N.M. 2012. Screening for phosphate solubilizing bacteria inhabiting the rhizoplane of rice grown in acidic soil in Bangladesh. Acta Microbiologica et Emmunologica Hungarica 59(2): 199-213.

- Sarker, A., Talukder, N.M. and Islam, T. 2014. Phosphate solubilizing bacteria promote growth and enhance nutrient uptake by wheat. *Plant Science Today* 1(2): 86-93.
- SAS. 1987. Statistical Analysis System. Version 9.2. PC SAS User's Guide. SAS Institute Inc., Cary, NC, USA.
- Savary, S., Castilla, N.P., Elazegui, F.A., McLaren, C.G., Ynalvez, M.A. and Teng, P.S. 1995. Direct and indirect effects of nitrogen supply and disease source structure on rice sheath blight spread. *Phytopathology* 85: 959-965.
- Schaad, N.W. 1980. Laboratory guide for the identification of plant pathogenic bacteria. pp. 28-45. St. Paul. Minnesota: American Phytopathological Society.
- Schmidt, C.S., Lorenz, D., Wolf, G.A. and Jager, J. 2001. Biological control of grape vine dieback fungus. Eutypa latall: influence of formulation additives and transpajon mutagenesis on the antagonistic activity of *Bacillus subtilis* and *Erwinia herbicola. Journal of Phytopathology* 149(1): 437-445.
- Sehajpal, A., Arora, S. and Kaur, P. 2009. Evaluation of plant extracts against *Rhizoctonia solani* causing sheath blight of rice. *The Journal of Plant Protection Sciences* 1(1): 25-30.
- Senapoty, D. 2010. Efficacy of soil amendments for the management of rice sheath blight. *Indian Phytopathology* 63(1): 94-95.
- Seneviratne, G., Thilakaratne, R.M.M.S., Jayasekaran, A.P.D.A., Seneviratne, K.A.C.N., Padmathilake, K.R.E., De Silva, M.S.D.L. 2009. Developing beneficial microbial biofilms on roots of non-legumes: a novel biofertilizing technique. In *Microbial strategy for crop improvement*, ed. M.S. Khan, A. Zaidi and J. Musarrat, pp. 51-61. Berlin, Heidelberg: Springer.
- Sen, S., Biswas, G., Basu, S.K. and Acharya, K. 2012. Management of leaf spot disease of *Stevia rebaudiana* Bertoni with antagonistic bacteria. *Australian Journal of Crop Sciences* 62(2): 350-356.
- Sen, S., Rai, M., Acharya, R., Dasgupta, S., Saha, A. and Acharya, K. 2009. Biological control of pathogens causing the cymbidium pseudobulb rot complex using fluorescent *Pseudomonas* strain BRL-1. *Journal of Plant Pathology* 91: 617-621.
- SES. 2013. Standard Evaluation System for Rice. 5<sup>th</sup> ed. p. 25. INGER. Genetic Resource Center, International Rice Research Institute, Manila, Philippines.
- Sessitsch, A., Reiter, B. and Berg, G. 2004. Endophytic bacterial communities of field-grown potato plants and their plant-growth promoting and antagonistic abilities. *Canadian Journal of Microbiology* 50: 239-249.

- Shabayev, V.P. 2010. Effect of the introduction of the nitrogen-fixing bacteria *Pseudomonas putida* 23 on the nitrogen balance in soil. *Eurasian Soil Science* 43(4): 436-441.
- Shahjahan, A.K.M., Ahmed, H.U., Sharma, N.R. and Miah, S.A. 1990. Epidemiological studies of sheath blight of rice of rice caused by *Rhizoctonia solani* Kuhn. *Bangladesh Journal of Botany* 19(2): 125-133.
- Shahjahan, A.K.M., Sharma, N.R., Ahmed, H.U. and Miah, S.A. 1986. Yield loss in modern rice varieties due to sheath blight in Bangladesh. *Journal of Agricultural Research* 11(2): 82-90.
- Shali, A., Ghasemi, S., Ahmadian, G., Ranjbar, G., Dehestani, A., Khalesi, N., Motallebi, E. and Vahed, M. 2010. *Bacillus pumilus* SG2 chitinases induced and regulated by chitin, show inhibitory activity against *Fusarium* graminearum and Bipolaris sorokiniana. Phytoparasitica 38: 141-147.
- Shahaya Mary, R. and Dhanaseeli, M. 2012. Biocontrol Potential of Selected Actinomycete and its Metabolites against *Rhizoctonia solani*. *Indian Journal* of Natural Sciences 2(12): 965-972.
- Shyamala, L. and Sivakumaar, P.K. 2012. Antifungal Activity of Rhizobacteria Isolated from Rice Rhizosphere Soil against Rice Blast Fungus Pyricularia oryzae. International Journal of Pharmaceutical and Biological Archives 3(3): 692-696.
- Silva, J.C., Torres, D.B., Lustosa, D.C., Filippi, M.C.C. and Silva, G.B. 2012. Rice sheath blight biocontrol and growth promotion by *Trichoderma* isolates from the Amazon. *Amazonian Journal of Agricultural and Environmental Science* 54(4): 243-250.
- Singh, A. and Shahi, J.P. 2012. Banded leaf and sheath blight: an emerging disease of maize (*Zea mays* L.). *Maydica* 57: 215-219.
- Singh, N., Kumar, S., Bajpai, V.K., Dubey, R.C., Maheshwari, D.K. and Kang, S.C. 2010a. Biological control of *Macrophomina phaseolina* by chemotactic fluorescent *Pseudomonas aeruginosa* PN1 and its plant growth promotory activity in chir-pine. *Crop Protection* 29(10): 1142-1147.
- Singh, P.P., Shin, Y.C., Park, C.S. and Chung, Y.R. 1999. Biological control of Fusarium wilt of cucumber by chitinolytic bacteria. *Phytopathology* 89: 92-99.

- Singh, R. and Sinha, A.P. 2005. Influence of application methods of *Pseudomonas fluorescens* for managing rice sheath blight. *Indian Phytopathology* 58(4): 474-476.
- Singh, R. and Sinha, A.P. 2009. Biological control of sheath blight of rice with *Pseudomonas fluorescens. Indian Phytopathology* 62(3): 381-383.
- Singh, R., Singh, L.S., Prasad, D. Kureel, R.S., Sengar, R. and Singh, A. 2010b. Relationship of susceptibility and growth stages of plant for development of epidemic of sheath blight in rice. *Journal of Applied and Natural Science* 2(2): 230-233.
- Sivakamasundari, R. and Usharani, G. 2012. Studies on the influence of *Pseudomonas fluorescens* and chemicals on the biocontrol of sheath blight incidence in rice. *International Journal of Pharmaceutical and Biochemical Archieves* 3(4): 973-977.
- Smibert, R.M. and Krieg, N.R. 1994. Phenotypic characterization. In *Methods for General and Molecular Biology*, ed. P.R.G. Gerhardt, E., Murray, W.A. Wood and N.R. Krieg, pp. 607-654. Washington, DC: American Society for Microbiology.
- Sneh, B., Burpee, L. and Ogoshi, A. 1991. Cytomorphological key to *Rhizoctonia* spp. Identification of *Rhizoctonia* species, pp. 39-42. St. Paul, Minnesota, USA: The American Phytopathological Society Press.
- Sneh, B., Jabaji-Hare, S., Neate, S. and Dijst, G. 1996. Rhizoctonia species: Taxonomy, Molecular Biology, Ecology, Pathology, and Control. p. 578. Dordrecht, The Netherlands: Kluwer Academic Publisher.
- Soltani, A.A., Khavazi, K., Rahmani, H.A., Alikhani, H.A., Omidvari, M. and Dahaji, P.A. 2012. Evaluation of Biological Control Traits in Some Isolates of Fluorescent Pseudomonads and *flavobacterium*. *Journal of Agricultural Science* 4(1): 164-170.
- Somasegaran, P. and Hoben, H.J. 1994. Hand book for rhizobia: methods in legume *Rhizobium* technology. New York: Springer-Verlag.
- Someya, N., Nakajima, M., Watanabe, K., Hibi, T. and Akutsu, K. 2005. Potential of *Serratia marcescens* strain B2 for biological control of rice sheath blight. *Biocontrol Science and Technology* 15(1):105-109.
- Son, T.T.N., Diep, C.P. and Giang, T.T.M. 2006. Effect of Bradyrhizobia and phosphate solubilizing bacteria application on soybean in rotational system in the Mekong delta. *Omonrice* 14: 48-57.
- Spaepen, S. and Vanderleyden, J. 2011. Auxin and plant-microbe interactions. *Cold Spring Harbors Perspectives in Biology* <u>http://dx.doi.org/</u>10.1101/cshperspect.a 001438.

- Srinivasachary, Willocquet, L. and Savary, S. 2011. Resistance to rice sheath blight (*Rhizoctonia solani* Kuhn) [(telomorph: *Thanatephorus cucumeris* (A.B. Frank) Donk.] disease: current status and perspectives. *Euphytica* 178: 1-22.
- Srivastava, R. and Shalini. 2009. Antifungal Activity of Pseudomonas fluorescens Against Different Plant Pathogenic Fungi. The Internet Journal of Microbiology 7(2) DOI: 10.5580/1e69.
- Sudha, M., Shyamala Gowri, R., Prabhavathi, P., Astapriya, P., Yamuna Devi, S. and Saranya, A. 2012. Production and optimization of indole acetic acid by indigenous micro flora using agro waste as substrate. *Pakistan Journal of Biological Sciences* 15(1): 39-43.
- Sundaramoorthy, S., Karthiba, L., Raguchander, T. and Samiyappan, R. 2013. Ecofriendly approaches of potential microbial bioagents in management of sheath rot disease in rice caused by *Sarocladium oryzae* (Sawada). *Plant Pathology Journal* 12(2): 98-103.
- Sundaramoorthy, S., Raguchander, T., Ragupathi, N. and Samiyappan, R. 2012. Combinatorial effect of endophytic and plant growth promoting rhizobacteria against wilt disease of *Capsicum annum* L. caused by *Fusarium solani*. *Biological Control* 60: 59-67.
- Suparman, M. 2004. Antagonistic bacteria for controlling *Fusarium* f. sp. *lycopersici*. MS Thesis, Universiti Putra Malaysia, Malaysia.
- Surendran, M., Kannan, G.S., Kamala, N. and Leenakumary, S. 2011. Consortium of Fluorescent Pseudomonads for the Management of Rice Sheath Blight Disease. *Journal of Biological Control* 25(2): 156-159.
- Suryadi, Y., Susilowati, D., Akhdiya, A., Kadir, T.S. and Wibowo, B. 2013a. Efficacy of consortium bacteria for control rice diseases under System of Rice Intensification (SRI) in West Java-Indonesia. *Albanian Journal of Agricultural Sciences* 12(1): 143-147.
- Suryadi, Y., Susilowati, D.N., Riana, E. and Mubarik, N.R. 2013b. Management of rice blast disease (*Pyricularia oryzae*) using formulated bacterial consortium. *Emirates Journal of Food and Agriculture* 25(5): 349-357.
- Suudi, M., Park, J.M., Kang, W.R., Hwang, D.J., Kim, S. and Ahn, P. 2013. Quantification of Rice Sheath Blight Progression Caused by *Rhizoctonia solani*. *Journal of Microbiology* 51(3): 580-588.
- Swadling, I.R. and Jeffries, P. 1998. Antagonistic properties of two bacterial biocontrol agents of grey mould disease. *Biocontrol Science and Technology* 8: 439-448.
- Szkop, M. and Bielawski, W. 2013. A simple method for simultaneous RP-HPLC determination of indolic compounds related to bacterial biosynthesis of indole-3-acetic acid. *Antonie van Leeuwenhoek* 103: 683-691.

- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. *Molecular Biology* and Evolution 30: 2725-2729.
- Todar, K. 2004. *Pseudomonas* and related bacteria. Todar's online text book of bacteriology.http://textbookofbacteriology.net/Pseudomonas.etc.html accessed on 6 April 2006.
- Trivedi, P., Pandey, A. and Palni, L.M.S. 2008. *In vitro* evaluation of antagonistic properties of *Pseudomonas corrugata*. *Microbiological Research* 163: 329-336. DOI: 10.1016/j.micres.2006.06.007.
- Umashankari, J. and Sekar, C. 2011. Comparative evaluation of different bioformulations of PGPR cells on the enhancement of induced systemic resistance (ISR) in Rice *P. oryzae* pathosystem under upland condition. *Current Botany* 2(3): 12-17.
- Upma, D. and Kalha, C.S. 2011. *In vitro* evaluation of fungicides, botanicals and bioagents against *Rhizoctonia solani* causing sheath blight of rice and their integration for effective management of the disease under field conditions. *Plant Disease Research* 26(1): 14-19.
- Vansuyt, G., Robin, A., Briat, J.F., Curie, C. and Lemanceau, P. 2007. Iron acquisition from Fe-pyoverdine by *Arabidopsis thaliana*. *Molecular Plant-Microbe Interactions* 20: 441-447.
- Varma, A., Bakshi, M., Lou, B.C., Hartmann, A. and Oelmueller, R. 2012. *Piriformospora indica*: a novel plant growth-promoting mycorrhizal fungus. *Agricultural Research* 1: 117-131.
- Vasantha Devi, T., Malarvizhi, R., Sakthivel, N. and Gnanamanickam, S.S. 1989. Biological control of sheath blight of rice in India with antagonistic bacteria. *Plant and Soil* 119: 325-330.
- Velusamy, P., Ko, H.S. and Kim, K.Y. 2011. Determination of antifungal activity of *Pseudomonas* sp. A3 against *Fusarium oxysporum* by high performance liquid chromatography (HPLC). *Agriculture, Food and Analytical Bacteriology* 1: 15-23.
- Velusamy, P., Immanuel, J.E. and Gnanamanickam, S.S. 2013. Rhizosphere Bacteria for Biocontrol of Bacterial Blight and Growth Promotion of Rice. *Rice Science* 20(5): 356-362.
- Vidaver, A.K. 1976. Prospects for control of phytopathogenic bacteria by bacteriophages and bacteriocins. *Annual Review of Phytopathology* 14: 465-541.
- Vidhyasekaran, P. and Muthamilan, M. 1995. Development of formulations of *Pseudomonas fluorescens* for control of chickpea wilt. *Plant Disease* 79(8): 782-786.

- Virmal, V., Mercy, R.B. and Kannabiran, K. 2009. Antimicrobial activity of maraine actinomycete, *Norcadiopsis* spp. VITSVK5 (FJ973467). Asian Journal of Medical Science 1: 57-63.
- Vojtkova, H., Janulkova, R. and Svanova, P. 2012. Phenotypic characterization of *Pseudomonas* bacteria isolated from polluted sites of Ostrava, Czech Republic. *GeoScience Engineering* 58(3): 52-57.
- Wahyudi, A.T., Astuti, R.I. and Giyanto. 2011. Screening of *Pseudomonas* sp. Isolated from Rhizosphere of Soybean Plant as Plant Growth Promoter and Biocontrol Agent. *American Journal of Agricultural and Biological Sciences* 6(1): 134-141.
- Wang, Y.L., Liu, S.Y., Mao, X.Q., Zhang, Z., Jiang, H., Chai, R.Y., Qiu, H.P., Wang, J.Y, Du, X.F., Li, B. and Sun, G.C. 2013. Identification and characterization of rhizosphere fungal strain MF-91 antagonistic to rice blast and sheath blight pathogens. *Journal of Applied Microbiology* 114(5): 1480-1490.
- Wei, G., Kloepper, J.W. and Tuzan, S. 1996. Induced systemic resistance to cucumber diseases and increased plant growth by plant growth promoting rhizobacteria under field conditions. *Phytopathology* 86: 221-224.
- Wissing, F. 1974. Cyanide formation from oxidation of glycine by a *Pseudomonas* species. *Journal of Bacteriology* 117: 1289-1294.
- Xiao, Y., Liu, M.W., Li, G., Zhou, E.X., Wang, L.X., Tang, J., Tan, F.R., Zheng, G.P. and Li, P. 2008. Genetic diversity and pathogenicity variation of different *Rhizoctonia solani* isolates in rice from Sichuan Province, China. *Chinese Journal of Rice Science* 22(1): 87-92.
- Xu, J.M., Cheng, H.H., Koskinen, W.C., Molina, J.A.E. 1997. Characterization of potentially bioactive soil organic carbon and nitrogen by acid hydrolysis. *Nutrient Cycling in Agroecosystems* 49: 267-280.
- Yang, C.J., Zhang, Z.G., Shi, G.Y., Zhao, H.Y., Tao, L.C.H. and Hou, T.P. 2011. Isolation and identification of endophytic bacterium W4 against tomato *Botrytis cinerea* and antagonistic activity stability. *African Journal of Microbiological Research* 5(2): 131-136.
- Yang, D., Wang, B., Wang, J., Chen, Y. and Zhou, M. 2009. Activity and efficacy of *Bacillus subtilis* strain NJ-18 against rice sheath blight and Sclerotinia stem rot of rape. *Biological Control* 51: 61-65.
- Yi, C.Z, Gang, X.Z., Dong, G.T., Kun, S.N., Fu, Y.D., Fan, L. and Feng, L.Y. 2000. Evaluation and utilization of antagonistic bacteria against rice sheath blight. *Chinese Journal of Rice Science* 14(2): 98-102.

- Yu, X., Liu, X., Zhu, T.H., Liu, G.H. and Mao, C. 2011. Isolation and characterization of phosphate solubilizing bacteria from walnut and their effect on growth and phosphorus mobilization. *Biology and Fertility of Soils* 47(4): 437-446.
- Zachow, C., Grosch, R. and Berg, G. 2011. Impact of biotic and a-biotic parameters on structure and function of microbial communities living on sclerotia of the soil-borne pathogenic fungus *Rhizoctonia solani*. *Applied Soil Ecology* 48(2): 193-200.
- Zaidi, A., Khan, M.S., Ahemad, M. and Oves, M. 2009. Plant growth promotion by phosphate solubilizing bacteria. *Acta Microbiologica et Immunologica Hungarica* 56 (Suppl 3): 263-284.
- Zarandi, E.M., Shahidi Bonjar, G.H. and Padasht Dehkaei, F. 2009. Biological control of rice blast (*Magnaporthe oryzae*) by used of *Streptomyces sindeneusis* isolate 263 in green house. *American Journal of Applied Sciences* 6(1): 194-199.
- Zeigler, R. and Barclay, A. 2008. The relevance of rice. Rice 1: 3-10.
- Zhang, C.Q., Liu, Y.H., Ma, X.Y., Feng, Z. and Ma, Z.H. 2009. Characterization of sensitivity of *Rhizoctonia solani*, causing rice sheath blight to mepronil and boscalid. *Crop Protection* 28(5): 381-386.
- Zheng, A., Lin, R., Zhang, D., Qin, P., Xu, L., Ai, P., Ding, L., Wang, Y., Chen, Y., Liu, Y., Sun, Z., Feng, H., Liang, X., Fu, R., Tang, C., Li, Q., Zhang, J., Xie, Z., Deng, Q., Li, S., Wang, S., Zhu, J., Wang, L., Liu, H. and Li, P. 2013. The evolution and pathogenic mechanisms of the rice sheath blight pathogen. *Nature Communications* 4, Article number: 1424. pp. 1-10. Doi: 10.1038/ncomms 2427.
- Zhong, T.W., Wei, Z., Qi, O.Z., Wen, L.C., Jun, Z.J., Kun, W.Z. and Li., W. 2007. Analyses of temporal development and yield losses due to sheath blight of rice (*Rhizoctonia solani* AG-11A). *Agricultural Sciences in China* 6(9): 1074-1081.