

UNIVERSITI PUTRA MALAYSIA

INDUCTION OF APOPTOSIS BY 2',3'-EPOXYISOCAPNOLACTONE AND 8-HYDROXYISOCAPNOLACTONE-2',3'- DIOL ISOLATED FROM MICROMELUM MINUTUM IN HUMAN T-LYMPHOCYTE LEUKEMIA CEM-SS CELLS

TAN BOON KEAT.

IB 2006 5



INDUCTION OF APOPTOSIS BY 2',3'-EPOXY ISOCAPNOLACTONE AND 8-HYDROXYISOCAPNOLACTONE-2',3'-DIOL ISOLATED FROM *MICROMELUM MINUTUM* IN HUMAN T-LYMPHOCYTE LEUKEMIA CEM-SS CELLS

TAN BOON KEAT

MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA

2006



INDUCTION OF APOPTOSIS BY 2',3'-EPOXY ISOCAPNOLACTONE AND 8-HYDROXYISOCAPNOLACTONE-2',3'-DIOL ISOLATED FROM *MICROMELUM MINUTUM* IN HUMAN T-LYMPHOCYTE LEUKEMIA CEM-SS CELLS

By

TAN BOON KEAT

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

March 2006



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

INDUCTION OF APOPTOSIS BY 2',3'EPOXYISOCAPNOLACTONE AND 8HYDROXYISOCAPNOLACTONE-2',3'-DIOL ISOLATED FROM MIRCOMELUM MINUTUM IN HUMAN T-LYMPHOCYTE LEUKEMIA CEM-SS CELLS

By

TAN BOON KEAT

March 2006

Chairman: Professor Abdul Manaf Ali, PhD

Faculty: Institute of Bioscience

2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol are two bioactive compounds isolated from the leaves of Micromelum minutum. The cytotoxic effect of the compounds was tested on a variety of human cell lines respectively using MTT assay. They were found to be most sensitive against human T-lymphoblastic leukemia cells (CEM-SS). The inhibition effect of 2',3'-epoxyisocapnolactone hydroxyisocapnolactone-2',3'-diol at 50% of cell population (IC₅₀) was found to be 4.6 $\mu g/ml$ (13.5 μM) and 3 $\mu g/ml$ (7.8 μM) on CEM-SS cells, respectively. Besides that, the inhibitor effect of the compounds on other human cells were found to be 13.4 µg/ml (39.2 μ M) and 9.0 μ g/ml (23.9 μ M) on cervical carcinoma cells (HeLa), 14.2 μ g/ml (41.5 μ M) and 7.7 μg/ml (20.5 μM) on colon adenocarcinoma cells (HT29), 7.4 μg/ml (21.6 μM) and 5.9 µg/ml (15.7 µM) on hepatocarcinoma cells (HepG2), 6.5 µg/ml (19.0 µM) and 7.1 μ g/ml (18.9 μ M) on transform liver cells (Chang). For comparative purposes, the IC₅₀



of several clinical cytotoxic drugs against CEM-SS cells were determined. The inhibitor effect of the compounds were more significant compared with methotrexate $[IC_{50} = >30]$ $\mu g/ml$ (66.1 μM)], cytosine arabinoside [IC₅₀ = >30 $\mu g/ml$ (123.5 μM)] and colchicines [IC₅₀ = 8 μ g/ml (20.1 μ M)]. The compounds also shown near similar IC₅₀ concentration as compare with cis-diamine dichloroplatinum [IC₅₀ = 3 μ g/ml (10.1 μ M)], vinorelbine [IC₅₀ = 3 μ g/ml (3.9 μ M)] and doxorubicin [IC₅₀ = 2.4 μ g/ml (4.1 μ M)]. Furthermore, from proliferation assay study, the compounds were significantly inhibiting the proliferation of cells at IC₅₀ value. From the morphological observation and agarose gel electrophoresis, apoptosis of the compounds on CEM-SS cells was determined. By using phase contrast, fluorescence and electron microcopies, observation on morphological alterations indicating apoptosis was evaluated. From DNA fragmentation, Acridine orange and Propidium iodide staining and DNA content analyses, the compounds were confirmed to have ability in promoting apoptosis. However, the percentage of apoptosis induced is low and the event is time-dependent. At high concentration of 10 µg/m, 2',3'and 8-hydroxyisocapnolactone-2',3'-diol induced necrosis. epoxyisocapnolactone Furthermore, 8-hydroxyisocapnolactone-2',3'-diol also exhibited better cytotoxicity compared to 2',3'-epoxyisocapnolactone. The induction time for apoptosis by 8hydroxyisocapnolactone-2',3'-diol in CEM-SS is earlier than 2',3'-epoxyisocapnolactone, which is 4 hours and 12 hours after treatment. Based on the results obtained, 2',3'epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol are able to induced apoptosis.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KESAN INDUKSI APOPTOSIS OLEH 2',3'EPOXYISOCAPNOLACTONE DAN 8HYDROXYISOCAPNOLACTONE-2',3'-DIOL YANG DIASINGKAN DARI MICROMELUM MINUTUM KE ATAS JUJUKAN SEL CEM-SS T-LYMPHOCYTE LEUKEMIK MANUSIA

Oleh

TAN BOON KEAT

Mac 2006

Pengerusi : Profesor Abdul Manaf Ali, PhD

Fakulti : Institusi Biosains

2',3'-epoxyisocapnolactone dan 8-hydroxyisocapnolactone-2',3'-diol merupakan dua jenis sebatian yang diasingkan dari daun Micromelum minutum. Kesan sitotoksik oleh kedua-dua sebatian ke atas pertumbuhan perbagai jenis jujukan kanser sel manusia telah diuji dengan teknik MTT. Mereka didapati lebih sensitive ke atas jujukan sel T-(CEM-SS). Kesan perencatan lymphoblastik leukemik manusia epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol pada 50% dari sel populasi (IC₅₀) didapati sebanyak 4.6 μg/ml (13.5 μM) dan 3 μg/ml (7.8 μM) masingmasing ke atas sel CEM-SS. Selain itu, kesan perencatan oleh kedua-dua sebatian aktif di atas jujukan sel manusia yang lain juga didapati sebanyak 13.4 μg/ml (39.2 μM) dan 9.0 μ g/ml (23.9 μ M) ke atas sel servikal karsinoma (HeLa), 14.2 μ g/ml (41.5 μ M) dan 7.7 μg/ml (20.5 μM) ke atas sel adenokarsinoma usus (HT29), 7.4 μg/ml (21.6 μM) dan 5.9 μg/ml (15.7 μM) ke atas sel hepatokarsinoma (HepG2), 6.5 μg/ml (19.0 μM) dan 7.1



ug/ml (18.9 μM) ke atas sel hati yang normal (Chang). Sebagai bandingan, IC₅₀ dari beberapa jenis ubat pasaran ke atas sel CEM-SS juga diujikan. Kesan perencatan oleh kedua-dua sebatian semulajadi adalah lebih berkesan berbanding dengan ubat-ubatan pasaran seperti methotrexate [IC50 = >30 $\mu g/ml$ (66.1 μM)], cytosine arabinoside [IC50 = >30 $\mu g/ml$ (123.5 μM)] dan colchicines [IC₅₀ = 8 $\mu g/ml$ (20.1 μM)]. Kedua-dua sebatian juga menunjukan aktiviti IC50 yang agak sama berbanding dengan cis-diamine dichloroplatinum [IC50 = 3 μ g/ml (10.1 μ M)], vinorelbine [IC50 = 3 μ g/ml (3.9 μ M)] dan doxorubicin [IC $_{50}$ = 2.4 $\mu g/ml$ (4.1 μM)]. Selain itu, dari pengajian atas teknik pertumbuhan, kedua-dua sebatian adalah ketara dalam perencatan pertumbuhan sel pada IC50. Kesan apoptosis bagi kedua-dua sebatian aktif ke atas sel CEM-SS telah diperolehi daripada pemerhatian morfologi dan elektrophoresis gel agaros. Dengan mengunakan teknik mikroskop fasa perbezaan, fluoresent dan elektron, pemerhatian ke atas perbezaan morfologi yang berkaitan dengan apoptosis telah dijalankan. Dari keputusan DNA fragmentasi, pewarnaan AO/PI dan pengajian kandungan DNA, kedua-dua sebatian aktif telah menunjukan kebolehannya dalam menrangsangkan induksi apoptosis. Akan tetapi, peratus bagi induksi apoptosis adalah rendah and kemunculan kesan apoptosis adalah rawatan. Bila 2',3'-epoxyisocapnolactone bergantung pada masa hydroxyisocapnolactone-2',3'-diol dirawat dengan dos yang tinggi (10 μg/ml), ia akan menrangsangkan nekrosis. Selebih daripada itu, 8-hydroxyisocapnolactone-2',3'-diol juga menunjukkan aktiviti sitotoksik yang lebih baik berbanding dengan 2',3'epoxyisocapnolactone. Kemunculan kesan apoptosis di dalam 8-hydroxyisocapnolactone-2',3'-diol adalah lebih awal berbanding dengan 2',3'-epoxyisocapnolactone, iaitu 4 jam and 12 jam selepas rawatan. Berdasarkan keputusan yang diperolehi, kedua-dua sebatian



semulajadi (2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol) boleh disimpulkan sebagai agen sitotoksik potensi yang boleh merangsangkan apoptosis.



ACKNOWLEDGEMENTS

First of all, I would like to express my most sincere thanks and appreciation to my supervisor, Professor Dr. Abdul Manaf bin Ali for his valuable guidance, suggestions, encouragement and advice throughout my project and preparation of the thesis.

I would also like to extend my gratefulness to my co-supervisors; Professor Dr. Mawardi Rahmani and Dr. Noorjahan Alitheen for their support and valuable discussion in making this research a success. Thank you for your kindness to help when I need it most.

Not to be forgotten to express my sincere thanks to Associate Professor Dr. Raha Rahim and Associate Professor Dr. Salmaan Inayat Hussain for their generosity to help and to allow me to use the equipment in their laboratories. Special thanks to Dr. Anthony Ho, his encouragement and patient enlightend me during my difficulties.

I am also very grateful to all my friends in UPM especially to Yih Yih, Ching Ling, Maddie, Yang Ping, Chyan Leong, Dr. Lim, Dr. Khor, Dr Yiap, Varma, Dr. Shuhaimi, Dr. Siti and Dr. Majid. Thank you for willing to share with me your knowledge. Learning from you all is one of the most beautiful things in my life.

Last but not least, my deepest appreciation to my parents who have made me who I am today and my sister, Boon Li and Boon Hooi who have always been there for me.



TABLE OF CONTENTS

ABSTRACT ABSTRAK ACKNOWLEDGEMENTS APPROVAL DECLARATION LIST OF TABLES LIST OF FIGURES LIST OF ABBREVIATION			ii iv vii vii x xii xiv	
CHA	APTER			
1	INTR	ODUCTION OF THE PROPERTY OF TH	ON	1
2	LITE	RATURE	REVIEW	5
	2.1	Cancer		5
			Biology of Tumor	6
			Classification of Cancer	6
			Leukemia	7
	2.2	Chemot		9
			Herbal Medicine	10
			Micromelum minutum	11
	2.3		ology of Cell Death	13
	2.4			13
	2.5	Apopto		14
			Morphology of Apoptosis	16
			Biochemical Regulation of Apoptosis	20
			2.5.2.1 Endonuclease Activation	20
			2.5.2.2 Phagocytosis	23
			2.5.2.3 Intracellular Signaling	24
			2.5.2.4 Caspase	27
			Molecular Regulation of Apoptosis	33
			2.5.3.1 Bcl-2 Family	33
			2.5.3.2 Tumor Suppressor Genes	36
3	MAT		AND METHODS	40
	3.1		compounds	40
	3.2	Cell line		40
	3.3	Cells re		41
	3.4		ne Maintenance	41
	3.5		cicity Assay	42
			MTT Assay	44
	3.6		ation Assay	45
	3.7	_	ology Assessment	45
			Phase Contrast Microscopy	45
		3.7.2	Fluorescent Microscopy	46



		[Acridine Orange/ Propidium Iodide (AO/PI) Staining]	
		3.7.3 Electron Microscopy	47
		[Scanning Electron Microscopy (SEM) – (JEOL-JSM 6400)]	
	3.8	DNA Fragmentation Assay	48
	3.9	Flow Cytometric Analyses	49
	3.10	Statistical Analyses	50
4	RESULTS AND DISCUSSION		51
	4.1	MTT Assay	51
	4.2	Proliferation Assay	61
	4.3	DNA Fragmentation Assay	
	4.4	Morphological Assessment of Cell Death	69
		4.4.1 Phase Contrast Microscopy	69
		4.4.2 Fluorescence Microsopy	74
		4.4.3 Electron Microscopy	84
	4.5	Cell Cycle Analyses	88
5	CON	CLUSION	94
REF	ERENC	ES	97
BIO	DATA (OF THE AUTHOR	107



LIST OF TABLES

Table		Page	
1	Differential features and significance of necrosis and apoptosis		
2	The Caspase Family	32	
3	Serial dilution gradient	43	
4	The inhibition concentration of 50% (IC50) of both natural compounds isolated from <i>Mircomelum minutum</i> (2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol) against various cell lines determined by using MTT assay	59	
5	The inhibition concentration of 50% (IC ₅₀) of various standard cytotoxic compounds against CEM-SS cells determined by using MTT assay	59	
6	The percentage of CEM-SS cells distribution in different cell cycle phase for untreated cells, cells treated with 2',3'-epoxyisocapnolactone in IC50 and cells treated with 8-hydroxyisocapnolactone-2',3'-diol in IC50 by using flow cytometry analysis	93	



LIST OF FIGURES

Figure		Page
1	The chemical structure of 2',3'-epoxyisocapnolactone isolated from the leaves of <i>Micromelum minutum</i>	12
2	The chemical structure of 8-hydroxyisocapnolactone-2',3'-diol isolated from the leaves of <i>Micromelum minutum</i>	12
3	Illustration of the morphological features of necrosis and apoptosis	18
4	Distinct pathways to apoptosis converge on activation of caspase	31
5	The intrinsic and extrinsic pathways leading to apoptosis	35
6	Molecular structure of MTT and formazan	53
7	Cytotoxic activity of 2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol against CEM-SS cells after 72 hours of incubation. Viability was determined with MTT Assay and data represents mean of triplicate \pm SD	54
8	Cytotoxic activity of 2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol against HT-29 cells after 72 hours of incubation. Viability was determined with MTT Assay and data represents mean of triplicate \pm SD	55
9	Cytotoxic activity of 2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol against HeLa cells after 72 hours of incubation. Viability was determined with MTT Assay and data represents mean of triplicate \pm SD	56
10	Cytotoxic activity of 2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol against HepG2 cells after 72 hours of incubation. Viability was determined with MTT Assay and data represents mean of triplicate \pm SD	57
11	Cytotoxic activity of 2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol against Chang cells after 72 hours of incubation. Viability was determined with MTT Assay and data represents mean of triplicate \pm SD	58
12	The cytotoxic effect of 2',3'-epoxyisocapnolactone on CEM-SS cell concentration compared with untreated cell. Data represents mean of triplicate ± SD	65



Figure		Page
13	The cytotoxic effect of 8-hydroxyisocapnolactone-2',3'-diol on CEM-SS cell concentration compared with untreated cell. Data represents mean of triplicate \pm SD	66
14	Effect of 2',3'-epoxyisocapnolactone and hydroxyisocapnolactone-2',3'-diol on DNA fragmentation	68
15	Phase contrast microscopy examination of untreated CEM-SS cells	71
16	Phase contrast microscopy examination of CEM-SS cells treated with 2',3'-epoxyisocapnolactone	72
17	Phase contrast microscopy examination of CEM-SS cells treated with 8-hydroxyisocapnolactone-2',3'-diol	73
18	Fluorescence microscopy examination of untreated CEM-SS cells	78
19	Fluorescence microscopy examination of CEM-SS cells treated with 2',3'-epoxyisocapnolactone	79
20	Fluorescence microscopy examination of CEM-SS cells treated with 8-hydroxyisocapnolactone-2',3'-diol	80
21	The percentage of viable, apoptotic and necrotic CEM-SS cell in the population after treated with 2',3'-epoxyisocapnolactone at various time courses	81
22	The percentage of viable, apoptotic and necrotic CEM-SS cell in the population after treated with 8-hydroxyisocapnolactone-2',3'- diol at various time courses	82
23	The percentage of viable, apoptotic and necrotic CEM-SS cell in the untreated population and population after treated with 0.1% DMSO at various time courses	83
24	Scanning electron microscopy examination of untreated CEM-SS cells	85
25	Scanning electron microscopy examination of CEM-SS cells treated with 2',3'-epoxyisocapnolactone	86
26	Scanning electron microscopy examination of CEM-SS cells treated with 8-hydroxyisocapnolactone-2',3'-diol	87



Page 27 DNA fluorescence histograms of the effect of 2',3'epoxyisocapnolactone on the progression through the cell cycle 28 DNA fluorescence histograms of the effect of 8hydroxyisocapnolactone-2',3'-diol on the progression through the cell cycle



LIST OF ABBREVIATIONS

% Percentage

ALL Acute Lymphocytic Leukemia

ANLL Acute Non-Lymphocytic Leukemia

AO Acridine Orange

ATCC American Type Culture Collection

ATP Adenosine triphosphate

bp Base pair

CEM-SS T-Lymphoblastic Leukemia

CGM Complete Growth Medium

Chang Normal Liver Cell

CO₂ Carbon dioxide

Da Dalton

DMSO Dimethylsulfoxide

DNA Deoxyribonucleic acid

EDTA Ethylenediaminetetraacetic acid Di-Sodium Salt

ELISA Enzyme linked immunosorbent assay

FBS Fetal Bovine Serum

HeLa Cervical Carcinoma

HepG2 Hepato Carcinoma

HT29 Colon Carcinoma

IC₅₀ Inhibition Concentration 50%

mg Milligram

ml Milliliter

MTT 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide

nm Nanometer

O.D. Optical Density

PBS Phosphate Buffered Saline

PI Propidium Iodide

RNA Ribonucleic acid

RPM Rotation per Minute

SDS Sodium dodecyl sulfate

SEM Scanning Electron Microscope

UV Ultra violet

μg Microgram

μl Microliter



CHAPTER 1

INTRODUCTION

Cancer chemotherapy, the treatment or control of cancer using anticancer drugs which are highly toxic medications that destroy cancer cells by interfering with their growth or preventing their reproduction (Altman and Sarg, 1992). It has played a role in cancer treatment for almost half a century. Years of testing and research have proved chemotherapy to be an effective cancer treatment. It may be the only treatment, or it may be used in combination with other treatments, including surgery and radiation therapy. Chemotherapy works by killing rapidly dividing cells. These cells include cancer cells, which continuously divide to produce more cells, and healthy cells that divide quickly, such as those in bone marrow, gastrointestinal tract, reproductive system and hair follicles. Healthy cells usually recover shortly after chemotherapy is complete (Mayo Foundation for Medical Education and Research, 2003). It differs from surgery or radiation in that it is always used as a systemic treatment. In chemotherapy the medicines travel throughout the whole body or system rather than being confined or localized to one area such as the breast, lung, or colon. Thus chemotherapy can reach cancer cells that may have spread to other parts of the body.

More than 100 drugs are currently used for chemotherapy, either alone or in combination.

Many more are expected to become available. These chemotherapy medicines are vary widely in their chemical composition, how they are taken, their usefulness in treating



specific forms of cancer, and their side effects. New medications are first developed through laboratory research in test tubes and animals. Then, their safety and effectiveness are tested for clinical trials in humans (American Cancer Society, 2001).

Chemotherapy drugs are divided into several categories based on how they affect specific chemical substances within cancer cells, which cellular activities or processes the drug interferes with, and which specific phases of the cell cycle the drug affects. Most chemotherapy is given as a combination of drugs that work together to kill cancer cells. Some of the types of chemotherapy medications commonly used to treat cancer include: 1) Alkylating agents. These medications interfere with the growth of cancer cells by blocking the replication of DNA. Examples of alkylating agents include busulfan, cisplatin, carboplatin, chlorambucil, cyclophosphamide, ifosfamide, dacarbazine (DTIC), mechlorethamine (nitrogen mustard), and melphalan. 2) Antimetabolites. These drugs block the enzymes needed by cancer cells to live and grow. Examples of antimetabolites include 5-fluorouracil, capecitabine, methotrexate, gemcitabine, cytarabine (ara-C), and fludarabine. 3) Antitumor antibiotics. These antibiotics are different from those used to treat bacterial infections. It interferes with DNA, blocking certain enzymes and cell division, and changing cell membranes. Examples of antitumor antibiotics include dactinomycin, daunorubicin, doxorubicin (Adriamycin), idarubicin, and mitoxantrone. 4) Mitotic inhibitors. These drugs inhibit cell division or hinder certain enzymes necessary in the cell reproduction process. Examples of mitotic inhibitors include paclitaxel, docetaxel, etoposide (VP-16), vinblastine, vincristine, and vinorelbine. 5) Nitrosoureas.



These medications impede the enzymes that help repair DNA. Examples of nitrosoureas include carmustine (BCNU) and lomustine (CCNU) (American Cancer Society, 2001).

Previous studies have demonstrated that a wide range of anticancer agents, including chemotherapeutic agents, hormones, and various biologicals, induce apoptosis in malignant cells *in vitro* (Mesner *et al*, 1997; Kaufmann and Earnshaw, 2000). It is important to emphasize that this treatment-induced apoptosis is not merely a tissue culture phenomenon. Serial examination of peripheral blood mononuclear cells from acute leukemia patients undergoing induction therapy has demonstrated that various agents, including cytarabine, mitoxantrone, etoposide, paclitaxel, and topotecan, cause a marked increase in the number of apoptotic blasts (Li *et al*, 1994). Characteristic apoptotic changes have also been described in solid tumors after treatment of mice with various cytotoxic drugs, including cytarabine, 5-fluorouracil (5FU), fludarabine, doxorubicin, cyclophosphamide, cisplatin, etoposide, dactinomycin, and camptothecin (Kaufmann and Earnshaw, 2000).

For chemotherapy, natural products have been important sources of medicines for many traditional communities around the world. Natural products or their structural relatives comprise about 50% of the drugs that are used in cancer chemotherapy (Mann, 2002). In Malaysia, out of 12,000 species of higher plants which are found in this country, that are more than 1,000 species are said to have therapeutic properties and currently being used in the local traditional medicine system (Said, 1995). According to another report by Burkill in 1966, there are about 6,000-7,000 species of higher plants that have been



reported to have therapeutic or medicinal properties in Peninsular Malaysia and its surrounding islands (Burkill, 1966). Since the used of natural products in cancer chemotherapy have growth tremendously, the study of mechanism and mode of action of plant extracts become more and more important. In many research, plant related derivatives can be synthesized by knowing their biochemical reaction against cancer, and these derivatives can even have greater effect over the original compounds.

In this study, two natural compounds, 2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol, which were isolated from the leaves of *Micromelum* minutum, were tested for their cytotoxic activity against human T-lymphoblastic leukemia cells (CEMSS)

The objectives of this study were:

- To evaluate the cytotoxic and antiproliferative activities of 2',3'-epoxyisocapnolactone and 8-hydroxyisocapnolactone-2',3'-diol against human T-lymphoblastic leukemic cells (CEM-SS) in terms of proliferation, morphological changes and the mode of cell death.
- To investigate the induction of apoptosis by 2',3'-epoxyisocapnolactone and 8hydroxyisocapnolactone-2',3'-diol in treated CEM-SS cells.



CHAPTER 2

LITERATURE REVIEW

2.1 Cancer

Cancer, a general term for more than 100 diseases which characterized by an uncontrolled, abnormal growth of cells appear in different parts of the body that can spread to other parts of the body (Altman and Sarg, 1992). It is a potentially fatal disease caused mainly by environmental factors that mutate genes encoding critical cell-regulation proteins. The resultant aberrant cell behavior leads to expensive masses of abnormal cells that destroy surrounding normal tissue and can spread to vital organs resulting in disseminated, commonly a harbinger of imminent patient death (Alison, 2002). Cancer cells contain many genetic alterations that accumulate as tumors develop. Over the last 20 years, considerable information has been gathered on regulation of cell growth and proliferation leading to the identification of the involvement of specific genes at the molecular level (Macdonald and Ford, 1997).

The incident of cancer is rising with doubling in new cancer cases and cancers related deaths expected over the next 20 years (Alison, 2002). Part of the reason for this rise is that life expectancy is steadily rising and most cancers are more common in an ageing population. More significantly, a globalization of unhealthy lifestyles, particularly

