



UNIVERSITI PUTRA MALAYSIA

***EVALUATION OF *Andrographis paniculata* Burm. F. EXTRACTS
AGAINST *Bemisia tabaci* GENNADIUS***

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**EVALUATION OF *Andrographis paniculata* Burm. F. EXTRACTS
AGAINST *Bemisia tabaci* GENNADIUS**

By

REZA TANHA NAJAFABADI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

January 2015

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To those that devoted their lives to science, nothing can give more happiness than making discoveries, but these cups of joy are full only when the results of their studies find practical application.

Louis Paster

DEDICATIONS

I Lovingly Dedicate This Thesis to My Parents for Their Immeasurable Supports in All Steps of My Life. Specially my wife and my father, who taught me that the best kind of knowledge to have is that which is learned for its own sake. It is also dedicated to my mother, who taught me that even the largest task can be accomplished if it is done one step at a time.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

EVALUATION OF *Andrographis paniculata* Burm. F. EXTRACTS AGAINST *Bemisia tabaci* GENNADIUS

By

REZA TANHA NAJAFABADI

January 2015

Chair: Professor Dzolkhifli Bin Omar, PhD

Faculty: Agriculture

Bemisia tabaci (Gennadius) (Hemiptera: Aleyrodidae) is one of the most destructive pests of vegetables and ornamental crops in the world. The development of resistance to most synthetic insecticides by this insect pest has necessitated a search for alternative methods of controlling it. Some botanical pesticides have been shown to provide satisfactory alternatives to chemical pesticides. *Andrographis paniculata*, an annual herb of the family Acanthaceae is primarily used for its medicinal properties, although insecticidal applications of this plant against insect pests of agricultural and medical importance have also been reported. This study aimed to evaluate the efficiency of various solvents in extracting the active compounds of *A. paniculata*, to determine the quality and quantity of the compounds and active ingredients in *A. paniculata* extract and to evaluate the toxicity of *A. paniculata* leaf extract against *B. tabaci*. Leaves of *A. paniculata* collected from Melaka, Malaysia were extracted with methanol, chloroform, and ethanol using Soxhlet extractor. Methanol was found to be the best solvent for the extraction of active compounds, especially andrographolide, from *A. paniculata* leaves. Chloroform gave the lowest yield of active compounds. Phytochemical screening of bioactive compounds in *A. paniculata* leaf extracts showed methanol extracts contained alkaloids, saponins, flavonoids, tannins, terpenoids and steroids. The levels of these bioactive compounds were generally low in chloroform and ethanol leaf extracts. The quantity andrographolide in methanolic *A. paniculata* leaf extract showed the highest amount (92.32 ppm) followed by ethanol (41.17 ppm) and chloroform (14.85 ppm). With regard to the toxicity of three solvent extracts (methanol, ethanol, and chloroform) of *A. paniculata* leaves against the adult and nymphal stages of *B. tabaci*, the most important finding was that *A. paniculata* leaf extract can be used as a bioinsecticide to control second instar nymphs and adults of *B. tabaci*. The LC₅₀ values for methanol, chloroform and ethanol extracts of *A. paniculata* against second instar

nymphs of *B. tabaci* were 39.48, 120.05, and 54.17 ppm, respectively. Among the three tested solvents, the crude methanol extract was found to have the most effective toxicity against second instar nymphs of *B. tabaci*. The toxicity of the three solvent extracts of *A. paniculata* was also tested against adults of *B. tabaci* after 96 hours of exposure. At this time point, the observed LC₅₀ ranged from 43.68 ppm for the methanol extract to 172.7 ppm for the chloroform extract. The LC₅₀ values for methanol, chloroform and ethanol extracts of *A. paniculata* against adult of *B. tabaci* were 43.68, 172.70, and 70.07 ppm, respectively. The larvicidal and adulticidal activity of *A. paniculata* against *B. tabaci* has not been previously studied in any detail, and this is the first report of its larvicidal and adulticidal activity against this pest.



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yang diuji, ekstrak metanol didapati mempunyai ketoksikan paling efektif terhadap instar kedua nimfa, *B. tabaci*. Ketoksikan ketiga-tiga pelarut untuk ekstrak *A. paniculata* diuji terhadap *B. tabaci* dewasa selepas pendedahan 96 jam menunjukkan nilai LC50 43.68, 172.70, dan 70.07 ppm untuk metanol, kloroform dan etanol ekstrak masing-masing. Aktiviti kebolehan membunuh larva dan *A. paniculata* terhadap *B. tabaci* belum diuji secara terperinci sebelum ini, dan ini adalah laporan pertama bagi aktiviti membunuh larva dan dewasa terhadap serangga ini.



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I certify that a Thesis Examination Committee has met on 23 January 2015 to conduct the final examination of Reza Tanha Najafabadi on her thesis entitled "Evaluation of *Andrographis paniculata* Burm.F. Extracts against *Bemisia tabaci* Gennadius " in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLEDGEMENTS		v
APPROVAL		vi
DECLARATION		viii
LIST OF TABLES		xiii
LIST OF FIGURES		xiv
LIST OF ABBREVIATIONS		xvi
CHAPTER		
1	INTRODUCTION	1
2	LITERATURE REVIEW	3
	2.1 Botanicals	3
	2.1.1 Definition and History	3
	2.1.2 Most Common Botanical Insecticides and their Mode of Action	4
	2.1.3 Advantages and Disadvantages of Botanicals	5
	2.1.4 Prospects of Botanical Insecticides	6
	2.1.5 Botanical Pesticides: for Richer and Poorer Countries	7
	2.2 <i>Andrographis paniculata</i>	8
	2.2.1 Botany	8
	2.2.2 History, Origin and Distribution	10
	2.2.3 Cultivation and Harvesting	11
	2.2.4 Extraction and Analytical Techniques of <i>A. paniculata</i>	11
	2.2.5 Main Content of <i>A. paniculata</i>	12
	2.2.6 Application	13
	2.2.6.1 Pharmaceutical	13
	2.2.6.2 Pesticidal	13
	2.2.6.3 Agriculturally Important Pests	14
	2.2.6.4 Medically Important Pests	15
	2.2.6.5 Examples of Botanicals against Whitefly	16
	2.3 <i>Bemisia tabaci</i>	17
	2.3.1 Ecology and Biology	17
	2.3.2 Taxonomic Position	19
	2.3.3 <i>Bemisia tabaci</i> Types of Damage	20
	2.3.3.1 Symptoms of Whitefly Infestation	21

2.3.4	Control of Whiteflies	22
2.3.4.1	Cultural Control	22
2.3.4.2	Biological Control	22
2.3.4.3	Chemical Control	23
2.3.5	Impact and Significance	24
3	MATERIALS AND METHODS	25
3.1	Extraction and Qualitative and Quantitative Analysis of Bioactive Compounds of <i>Andrographis paniculata</i>	25
3.2	The Efficiency of Different Solvents on the Extraction of Active Ingredient of <i>A. paniculata</i>	26
3.2.1	Collection of Plant Material	26
3.2.2	Preparation of Plant Extracts	26
3.3	Qualitative Screening of Bioactive Compounds in <i>A. paniculata</i> Leaves	26
3.3.1	Alkaloids Test	27
3.3.2	Saponin Test	27
3.3.3	Flavanoids Test	27
3.3.4	Tannins Test	27
3.3.5	Steroids and Terpenoids Tests	27
3.4	Quantitative Analysis of Andrographolide from Extracts of Different Solvents by HPLC	28
3.4.1	HPLC Method Development: Preparation of Sample and Standard Solution	28
3.4.2	Chromatographic Conditions	28
3.5	Toxicity of <i>Andrographis paniculata</i> Extracts against Adult and Nymphal Stage of <i>Bemisia Tabaci</i>	29
3.5.1	Host Plant	29
3.5.2	Insect Culture	30
3.5.3	Insecticides	30
3.5.4	Bioassay	31
3.5.4.1	Nymph Bioassay	31
3.5.4.2	Adult Bioassay	33
3.6	Data Analysis	35
4	RESULTS AND DISCUSSION	36
4.1	Extraction of <i>Andrographis paniculata</i> with Soxhlet Extractor	36
4.2	Qualitative Screening for Bioactive Compounds in <i>A. paniculata</i> Leaves	37
4.3	Quantitative Analysis of Andrographolide in <i>A. paniculata</i> Leaf Extracts by HPLC	39
4.4	Adult and Nymphal Mortality	44
4.4.1	Nymph Mortality	44
4.4.2	Adult Mortality	46
4.5	Larvicidal and Adulticidal Activities of <i>A. paniculata</i> Methanol Extract Extract against <i>B. tabaci</i>	50

4.6	Larvicidal and Adulticidal Activities of <i>A. paniculata</i> Chloroform Extracts against <i>B. tabaci</i>	51
4.7	Larvicidal and Adulticidal Activities of <i>A. paniculata</i> Ethanol Extracts against <i>B. tabaci</i>	53
4.8	Optimum Point	54
5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	64
	REFERENCES	66
	APPENDICES	93
	BIODATA OF STUDENT	106
	PUBLICATION	107

LIST OF TABLES

Table		Page
2.1	Properties of some Important Botanical Insecticides	4
2.2	Summary of Advantages and Disadvantages of Botanicals	5
4.1	Effect of Solvent Polarity on Extraction Yield	36
4.2	Qualitative Analysis of Bioactive Compounds in <i>A. paniculata</i> Leaf Extracts Prepared in Different Solvents	38
4.3	HPLC Quantification of Andrographolide in <i>A. paniculata</i> Leaf Extract	41
4.4	Larvicidal Activity of Different Solvent Extracts of <i>A. paniculata</i> against Second Instar Nymphs of <i>B. tabaci</i>	45
4.5	Adulticidal Activity of Different Solvent Extracts of <i>A. paniculata</i> against Adult of <i>B. tabaci</i> at 48 Hours	47
4.6	Adulticidal Activity of Different Solvent Extracts of <i>A. paniculata</i> against Adult of <i>B. tabaci</i> at 96 Hours	49
4.7	Mean squares ANOVA for methanol extract application at different stage	50
4.8	Mean comparison among different application on methanol extract at different stage	51
4.9	Mean Square for Chloroform Extract Application	52
4.10	Mean Comparison Rate of Chloroform Extract Application	52
4.11	Mean Square for Ethanol Extract Application	53
4.12	Mean Comparison Rate of Ethanol Extract Application	54

LIST OF FIGURES

Figure	Page
2.1 Taxonomic Tree of <i>A. paniculata</i>	8
2.2 Botanical Features of <i>A. paniculata</i>	10
2.3 Schematic Life Cycle of <i>B. tabaci</i>	18
2.4 Nymphs of <i>B. tabaci</i>	18
2.5 Sweet Potato Whitefly Adults, Nymphs, and Pupal Cases.	18
2.6 Taxonomic Tree of <i>B. tabaci</i>	19
2.7 Honeydew Produced by <i>B. tabaci</i> Can Result in the Growth of Black Sooty Mold on Tomato Cherry Fruits and Leaves.	21
2.8 Suspected Viral Disease Symptoms Due to Vectoring of Virus Diseases by <i>B. tabaci</i> . Left: Healthy Cherry Tomato Plants Right: Virus- Infected Cherry Tomato Plants with Significant Yield Reduction.	21
3.1 Potted EggPlant which has been trimmed until only Three Leaf Squares Remain	32
3.2 Eggplant Leaf which has been trimmed to an Approximate 5x7 cm Square	32
3.3 Adult Bioassay Steps	34
3.4 Log Dose-Probit Lines for Homogeneous and Heterogeneous Populations. 1 Homogenous Susceptible, 2, 3, and 4 Heterogeneous Resistant, and 5 Homogenous Resistant	35
4.1 HPLC Chromatogram of Standard Andrographolide	39
4.2 HPLC Chromatograms of Andrographolide (AP) from <i>A. paniculata</i> Leaves Extracted with Methanol Extract	40
4.3 HPLC Chromatograms of Andrographolide (AP) from <i>A. paniculata</i> Leaves Extracted with Chloroform Extract	40

4.4	HPLC Chromatograms of Andrographolide (AP) from <i>A. paniculata</i> Leaves Extracted with Ethanol Extract	41
4.5	HPLC Calibration Curves for the Determination of Andrographolide	42
4.6	Larvicidal Activity of Different Solvent Extracts of <i>A. paniculata</i> against Second Instar Nymphs of <i>B. tabaci</i>	55
4.7	Adulticidal Activity of Different Solvent Extracts of <i>A. paniculata</i> against Adult of <i>B. tabaci</i> at 48 Hours	56
4.8	Adulticidal Activity of Different Solvent Extracts of <i>A. paniculata</i> against Adult of <i>B. tabaci</i> At 96 Hours	57
4.9	Larvicidal Activities of <i>A. paniculata</i> Extract from Different Solvents	58
4.10	Adulticidal Activities of <i>A. paniculata</i> Extract from Different Solvents at 48 Hours	58
4.11	Adulticidal Activities of <i>A. paniculata</i> Extract from Different Solvents at 96 Hours	59

LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
CRD	Completely Randomized Design
DBM	Diamond back Moth
EPA	The U.S. Environmental Protection Agency
FOA	Food and Agriculture Organization
GC-MS	Gas Chromatography and Mass Spectroscopy
HIV	Human Immunodeficiency Virus
HPLC	High Performance Liquid Chromatography
HPTLC	High Performance Thin Layer Chromatography
IRAC	Insecticide Resistance Action Committee
IRM	Insecticide Resistance Management
IPM	Integrated Pest Management
LCL	Less-Than-Container Load
LC-MS	Liquid Chromatography and mass spectroscopy
LC ₅₀	Median Lethal Concentration
LC ₉₀	Lethal Concentration 90
LD ₅₀	Median Lethal Dose
LD ₉₀	Lethal Dose 90
LC	Liquid Chromatography
MECC	Micellar Electrokinetic Capillary Chromatography
MEKC	Microemulsion Electrokinetic Chromatographic
MHA	Muller Hinton Agar
nAChR	Nicotinic Acetylcholine Receptor
NSKE	Neem Seed Kernel Extract
PH	Parker-Hannifin
PPM	Parts Per Million
RH	Relative Humidity
RF	resistance factors
RP	Reversed-Phase

SAS	Statistical Analysis System
TLC	Thin Layer Chromatography
UCL	Ultra Chromatography Liquid
UPLC	Ultra Performance Liquid Chromatography
UV	Ultraviolet Visible
w/w	Weight for weight



CHAPTER 1

INTRODUCTION

Bemisia tabaci is one of the most important sap-sucking pests, affecting a diverse range of crops that includes fruits, vegetables, and fibre and ornamental crops. It causes damage through direct feeding, promoting sooty mold growth, and, most importantly, by vectoring some of the key plant viruses. The species has long confounded whitefly systematists because many whiteflies were described and at the same time synonymised due to considerable overlap in their morphological characteristics (Perring, 2001). Consequently, consensus was reached that *B. tabaci* is a cryptic species complex (Brown et al., 1995; De Barro et al., 2011a) representing at least some 28 species (Dinsdale et al., 2010; Hu et al., 2011). The species of this complex vary in terms of some features, including rate of development, expression of resistance, breadth and efficacy of virus transmission, attraction by natural enemies, host utilisation, damage to the physiological host, and endosymbionts (Kirk et al., 2000; Oliveira et al., 2001; Horowitz et al., 2007). Among these species, a few, namely the B and Q biotypes, are regarded as invasive and harbour far more potential to cause huge crop losses than endemic biotypes. For instance, following the introduction of the invasive B biotype to the United States in the mid-1980s, *B. tabaci* has cost more than \$2 billion in crop loss, crop damage, and pest control (Toscano et al., 1998). Non-chemical control of this pest is difficult to achieve, especially in cases in which the species vectors plant viruses. Thus, chemical control remains the primary method of control of *B. tabaci* (Medina-Ortega, 2011).

While most botanical insecticides fail to adequately compete against the newest generation of synthetic insecticides, their application still has merit in some scenarios. Such scenarios include application in industrialised countries in cases where human and animal health is paramount, in niche markets such as certified organic production, and most importantly, in developing countries, especially in tropical and subtropical zones (Isman, 2008). *Andrographis paniculata*, an annual herb of the family Acanthaceae (Kumar et al., 2012), is primarily used for its medicinal properties; however, use of this herb as an insecticide against insect pests of agricultural and medical importance has also been reported.

Andrographis paniculata was introduced to Malaysia (Valdiani et al., 2012) and is now being cultivated in some states for its medicinal properties. In fact, it is one of the most commonly used medicinal herbs in Malaysia (Hanapi et al., 2010). While the species has been researched for its medicinal applications in Malaysia (Mustaffa et al., 2011; Arifullah et al., 2013), studies regarding the insecticidal activity of this herb in Malaysia are non-existent. Its insecticidal, antifertility,

antifeedant and repellent activity against important agricultural pests of the orders Lepidoptera, Hemiptera, and Coleoptera orders have been tested (Qader et al., 2011). To the best of our knowledge, there have been no studies regarding its effect on whiteflies, including the sweet potato whitefly or *B. tabaci*. Recently, an invasive biotype of *B. tabaci*, the Q biotype, was reported in the Cameron Highlands and Kundasang in Malaysia (Shadmany et al., 2013). This biotype has a well-established reputation for enormous capacity to resist multiple classes of insecticides. Interestingly, a Q-type population from Cameron Highlands that was bioassayed against various insecticides showed very high levels of resistance to some of the tested materials (Shadmany et al., 2013). Because chemical control is still used as the primary method of control of this pest and because of the high potential of *B. tabaci* for the development of resistance to chemical agents, it is imperative to develop more environment-friendly chemical control agents, preferably with different modes of action, to better manage the pest and delay the onset of resistance. As mentioned above, the search for effective botanicals is justifiable in developing countries like Malaysia as it most likely carries a lower poisoning risk compared with conventional insecticides (Isman, 2008). Consequently, this study aims to evaluate the efficiency of various solvents in extracting the active compounds of *A. paniculata*, to determine the quality and quantity of the compounds and active ingredients in *A. paniculata* extract and to evaluate the toxicity of *A. paniculata* leaf extract against *B. tabaci*.

REFERENCES

- Abbassi, K., Atay-Kadiri, Z., & Ghaout, S. (2003). Biological Effects of Alkaloids Extracted from Three Plants of Moroccan Arid Areas on the Desert Locust. *Physiological Entomology*, 28(3), 232-236.
- Abbott, W. S. (1925). A Method of Computing the Effectiveness of an Insecticide. *Journal of Economic Entomology*, 18(2), 265-267.
- Abd-Rabou, S., & Simmons, A. M. (2010). Survey of Reproductive Host Plants of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in Egypt, Including New Host Records. *Entomological News*, 121(1), 456-465.
- Abu-Ghefreh, A. A., Canatan, H., & Ezeamuzie, C. I. (2009). In Vitro and In Vivo Anti-Inflammatory Effects of Andrographolide. *International Immunopharmacology*, 9(1), 313-318.
- A glossary of science, chemistry, environmental and laboratory terms. (2013). Retrieved 18th Sep 2013 from <http://www.caslab.com/Laboratory-Terms-B/>.
- Ahmad, I., Aqil, F., & Owais, M. (2006). Modern Phytomedicine: Turning Medicinal Plants into Drugs. In W. M. Bandaranayake (Ed.), *Quality Control, Screening, Toxicity, and Regulation of Herbal Drugs* (pp. 25-53). Germany: WILEY-VCH Verlag GmbH & Co.
- Akbar, S. (2011). *Andrographis paniculata*: a Review of Pharmacological Activities and Clinical Effects. *Alternative Medicine Review*, 16(1), 66-77.
- Akowuah, G. A., Ismail, Z., Norhayati, I., & Sadikun, A. (2005). The Effects of Different Extraction Solvents of Varying Polarities on Polyphenols of *Orthosiphon stamineus* and Evaluation of the Free Radical-Scavenging Activity. *Food Chemistry*, 9(3), 311-317.
- Akowuah, G. A., Zhari, I., Norhayati, I., & Mariam, A. (2006). HPLC and HPTLC Densitometric Determination of Andrographolides and Antioxidant Potential of *Andrographis Paniculata*. *Journal of Food Composition and Analysis*, 19(1), 118-126.
- Al-mazra'aw, M. S., & Ateyyat, M. (2009). Insecticidal And Repellent Activities of Medicinal Plant Extracts against the Sweet Potato Whitefly, *Bemisia tabaci* (Hom. Aleyrodidae) and Its Parasitoid *Eretmocerus mundus* (Hym. Aphelinidae). *Journal of Pesticide Science*, 8(2), 149-154.

- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic Local Alignment Search Tool. *Molecular Ecology*, 2(15), 403-410.
- Andrographis. (n.d.). Retrieved 15th Sep 2013, from http://www.oisat.org/control_methods/plants_in_pest_control/andrographis.html.
- Arifullah, M., Namsa, N. D., Mandal, M., Chiruvella, K. K., Vikrama, P., & Gopal, G. R. (2013). Evaluation of Anti-Bacterial and Anti-Oxidant Potential of Andrographolide and Echioidinin Isolated from Callus Culture of *Andrographis paniculata* Nees. *Asian Pacific journal of tropical biomedicine*, 3(8), 604-610.
- Arora, J. R., Swarup, R., & Gupta, S. V. (2000). The BIOME News. India: Department of Biotechnology, Ministry of Science and Technology, Government of India.
- Aslan, I., Ozbek, H., Calmasur, O., & SahIn, F. (2004). Toxicity of Essential Oil Vapours to Two Greenhouse Pests, *Tetranychus Urticae* Koch and *Bemisia Tabaci* Genn. *Industrial Crops and Products*, 19(4), 167-173.
- Ateyyat, M. A., Al-Mazra'awi, M., Abu-Rjai, T., & Shatnawi, M. A. (2009). Aqueous Extracts of Some Medicinal Plants are as toxic as Imidacloprid to the Sweet Potato Whitefly, *Bemisia tabaci*. *Journal of Insect Science*, 9(2), 1-6.
- Authority, V. (2010). A publication of the agri-food & veterinary authority of Singapore December 2010, 0-1.
- Avanigadda, S., & Vangalapati, M. (2010). Experimental and Modelling Studies of Andrographolide Extraction from *Andrographis paniculata*. *International Journal of Chemical, Environmental and Pharmaceutical Research*, 1(1), 32-36.
- Bagde, S., Khare, M., Patidar, R. K., & Singh, V. (2013). Antimicrobial Properties and Characterization of Phytoconstituents of the Leaf Extracts of Some Medicinal Plants. *Journal of Pharmacognosy and Phytochemistry*. 1(6), 121-142.
- Bajwa, A. A., & Ahmad, A. (2012). Potential Applications of Neem Based Products as Biopesticides. *The Health*, 3(4): 116-120.
- Banks, G. K., Colvin, J., Reddy, R. V. C., Maruthi, M. N., Muniyappa, V., Venkatesh, H. M., Kumar, M. K., Padmaja, A. S., Beitia, F. J., & Seal, S. E. (2001). First Report of the *Bemisia tabaci* B Biotype in India and an Associated Tomato Leaf Curl Virus Disease Epidemic. *Plant Disease*, 8(5), 231-231.

- Barati, R., Golmohammadi, G., Ghajarie, H., Zarabi, M., & Mansouri, R. (2013). Efficiency of Some Herbal Pesticides on Reproductive Parameters of Silverleaf Whitefly, *Bemisia tabaci* (Gennadius) (Hemiptera: Aleyrodidae). *Archives of Phytopathology and Plant Protection*, 6(4), 1-10.
- Barney, J. N., Hay, A. G., & Weston, L. A. (2005). Isolation And Characterization of Allelopathic Volatiles from Mugwort (*Artemisia vulgaris*). *Journal of Chemical Ecology*, 3(1), 247-265.
- Barwick, V. C. (1997). Strategies for solvent selection a literature review. *Trends in Analytical Chemistry*, 16(1), 293-309.
- Basile, A., Sorbo, S., Giordano, S., Ricciardi, L., Ferrara, S., & Montesano, D. (2000). Antimicrobial and Allelopathic Activity of Extract from *Castanea sativa* Leaves. *Fitoterapia*, 71(5), 110-116.
- Betarbet, R., Sherer, T. B., MacKenzie, G., Garcia-Osuna, M., Panov, A. V., & Greenamyre, J. T. (2000). Chronic Systemic Pesticide Exposure Reproduces Features of Parkinson's disease. *Nature Neuroscience*, 3(1), 1301-1306.
- Bobbarala, V., Koteswara Rao, P., Srinivasa Rao, G., & Aryamithra, D. (2009). Bioactivity of *Andrographis paniculata* against Selected Phytopathogens. *Journal of Pharmacy Research*, 2(3), 480-482.
- Bostanian, N. J., Wise, J. C., & Isaacs, R. (2012). Pesticides for Arthropod Control in Vineyards. In N. J. Bostanian., C. Vincent & R. Isaacs (Eds.), *Arthropod Management in Vineyards* (pp. 53-90). New York, NY: Springer.
- Botanical Insecticides. (n.d.). Retrieved 25th Oct 2013, from <http://Landscapeipm.tamu.edu/types-of-pest-control/chemical-control/organic/botanical/>.
- Bright, A. A., Babu, A., Ignacimuth, S., & Dorn, S. (2001). Efficacy of Crude Extracts of *Andrographis Paniculata* Nees. on *Callosobruchus Chinensis* L. During Post Harvest Storage of Cowpea. *Indian Journal Experience Biology*, 3(9), 715-718.
- Brown, A. E. (2005). *Mode of action of insecticides and related pest control chemicals for production agriculture, ornamentals and turf*. Pesticide Info Leaflet Nr 43:1-13. <http://pesticide.umd.edu>.
- Brown, J. K. (2007). The *Bemisia tabaci* complex: Genetic and Phenotypic Variability Drives Begomovirus Spread and Virus Diversification. *Plant Disease*, 1(1), 25-56.

- Brown, J. K., Frohlich, D. R., & Rosell, R. C. (1995). The Sweet Potato Or Silverleaf Whiteflies: Biotypes of *Bemisia tabaci* or Species Complex? *Annual Review of Entomology*, 40(1), 511-534.
- Calmasur, O., Aslan, I., & Sahin, F. (2006). Insecticidal and Acaricidal Effect of Three Lamiaceae Plant Essential Oils Against *Tetranychus Urticae* Koch and *Bemisia Tabaci* Genn. *Industrial Crops and Products*, 2(3), 140-146.
- Casida, J. E. (1973). *Pyrethrum the natural insecticide*. New York, NY: Academic Press.
- Cawthray, G. R. (2003). An Improved Reversed-Phase Liquid Chromatographic Method for the Analysis of Low-Molecular Mass Organic Acids in Plant Root Exudates. *Journal of Chromatography*, 1(1), 1011, 1233.
- Chaieb, K., Hajlaoui, H., Zmantar, T., Kahla-Nakbi, A. B., Rouabhia, M., Mahdouani, K., & Bakhrouf, A. (2007). The Chemical Composition and Biological Activity of Clove Essential Oil, *Eugenia caryophyllata* (*Syzigium aromaticum* L. Myrtaceae): a Short Review. *Phytotherapy research*, 21(6), 501-506.
- Chakraborty, S., Biswas, S., Sarkar Manna, J., Das, S., & Dey, R. (2011). Sol-Gel Derived Silica-Gel as a Controlled Delivery System of *Andrographis paniculata* Extract and Its Anti-Microbial Efficacy. *Springer*, 64(1), 198-193.
- Chao, W. W., Kuo, Y. H., Hsieh, S. L., & Lin, B. F. (2011). Inhibitory Effects of Ethyl Acetate Extract of *Andrographis paniculata* on NF-B Trans-Activation Activity and LPS-induced Acute Inflammation in Mice. *Evidence-Based Complementary and Alternative Medicine*, 1(1), 1-9.
- Chapman, R. F. (1998). *The Insects: Structure and Function* (4thEd). Cambridge: Cambridge University Press.
- Chenniappan, K., Chellamuthu, V., Dhandapani, A., & Kadarkarai, M. (2011). Synergistic Activity of *Andrographis paniculata* Nees with *Bacillus thuringiensis* Var *Israelensis* against Malarial Vector, *Anopheles Stephensi* Liston (Diptera: Culicidae). *Journal of the Entomological Research Society*, 1(3), 71-86.
- Chenniappan, K., & Kadarkarai, M. (2008). Oviposition Deterrent, Ovicidal and Gravid Mortality Effects of Ethanolic Extract of *Andrographis paniculata* Nees against the Malarial Vector *Anopheles stephensi* Liston (Diptera: Culicidae). *Entomological Research*, 3(8), 119-125.

- Chenniappan, K., & Kadarkarai, M. (2008). Synergistic Activity of *Andrographis Paniculata* Nees Extracts against the Larvae of the Malarial Vector *Anopheles stephensi* Liston (Diptera: Culicidae). *Journal of the Entomological Research Society*, 10(1), 112-124.
- Choi, D. W., Kim, J. H., Cho, S. Y., Kim, D. H., & Chang, S.Y. (2002). Regulation and quality control of herbal drugs in Korea. *Toxicology*, 181(182), 581-586.
- Chong, G. H. (2002). *Appropriate Drying and Storage Conditions of Hempedu Bumi (Andrographis Paniculata Nees)*. (Unpublished Doctoral Dissertation). Universiti Putra Malaysia.
- Chopra, R. N., Nayar, S. L., & Chopra, I. C. (1956). *Glossary of Indian Medicinal Plants*. New Delhi: CSIR - Council for Scientific and Industrial Research.
- Chua, K. H. (2007). *Studies on Andrographis Paniculata (Burm. F.) Nees (HDM 15) a Medicinal Native Plant of Brunei Darussalam*. (Unpublished Doctoral dissertation). Bradford University, United Kingdom.
- Chu, D., Zhang, Y. J., Brown, J. K., Cong, B., Xu, B. Y., Wu, Q. J., & Zhu, G. R. (2006). The Introduction of the Exotic Q Biotype of *Bemisia tabaci* from the Mediterranean Region into China on Ornamental Crops. *Florida Entomologist*, 8(9), 168-174.
- Cohen, A. C., & Byrne, D. N. (1992). *Geocoris Punctipes* as a Predator of *Bemisia tabaci*: a Laboratory Evaluation. *Entomologia Experimentalis ET Applicata*, 64(2), 195-202.
- Costa, H. S., Brown, J. K., & Byrne, D. N. (1991). Life History Traits of the Whitefly, *Bemisia tabaci* (Homoptera: Aleyrodidae) on Six Virus-Infected or Healthy Plant Species. *Environmental Entomology*, 20(1), 1102-1107.
- Crop Protection Compendium. 2012. Retrieved 5th Nov 2013, from <http://www.cabi.org/cpc/default.aspx?site=161&page=4063>.
- Cruz-Estrada, A., Gamboa-Angulo, M., Borges-Argaez, R., & Ruiz-Sanchez, E. (2013). Insecticidal Effects of Plant Extracts on Immature Whitefly *Bemisia tabaci* Genn. (Hemiptera: Aleyroideae). *Electronic Journal of Biotechnology*, 1(6), 1-9.
- Cui, Y., Wang, Y., Ouyang, X., Han, Y., Zhu, H., & Chen, Q. (2009). Fingerprint Profile of Active Components for *Andrographis paniculata* Nees by HPLC-DAD. *Sensing and Instrumentation for Food Quality and Safety*, 3(3), 165-179.

- Dalton, R. (2006). Whitefly infestations: The Christmas Invasion. *Nature*, 44(3), 898-900.
- De Barro, P. J., Liu, S. S., Boykin, L. M., & Dinsdale, A. B. (2011). *Bemisia tabaci*: A Statement of Species Status. *Annual Review of Entomology*, 5(6), 1-19.
- Delatte, H., Duyck, P. F., Triboire, A., David, P., Becker, N., Bonato, O., & Reynaud, B. (2009). Differential Invasion Success Among Biotypes: Case of *Bemisia tabaci*. *Biological Invasions*, 11(3), 1059-1070.
- Dennehy, T. J., Degain, B. A., Harpold, V. S., Zaborac, M., Morin, S., Fabrick, J. A., Nichols, R. L., Brown, J. K., Byrne, F. J., & Li, X. (2010). Extraordinary Resistance to Insecticides Reveals Exotic Q Biotype of *Bemisia tabaci* in the New World. *Journal of Economic Entomology*, 10(3), 2174-2186.
- Dev, S., & Koul, O. (1997). *Insecticides of Natural Origin*. Amsterdam: Harwood Academic.
- Dimetry, N. Z. (2012). Prospects of Botanical Pesticides for the Future in Integrated Pest Management Programme (IPM) With Special Reference to Neem Uses in Egypt. *Archives of Phytopathology and Plant Protection*, 45(10), 1138-1161.
- Dinsdale, A., Cook, L., Riginos, C., Buckley, Y. M., & De Barro, P. (2010). Refined Global Analysis of *Bemisia tabaci* (Hemiptera: Sternorrhyncha: Aleyrodoidea: Aleyrodidae) Mitochondrial Cytochrome Oxidase 1 to Identify Species Level Genetic Boundaries. *Annals of the Entomological Society of America*, 10(3), 196-208.
- Dubey, N. K., Shukla, R., Kumar, A., Singh, P., & Prakash, B. (2010). Prospects of Botanical Pesticides in Sustainable Agriculture. *Current Science*, 9(8), 479-480.
- Duke, S. O. (1990). Natural Pesticides from Plants. In J. Janick., & J. E. Simon (Eds.), *Advances in New Crops* (pp. 511-517). Portland: Timber Press, OR.
- Dutta, A., & Sukul, N. (1982). Filaricidal Properties of a Wild Herb *Andrographis paniculata*. *Journal of Helminthology*, 5(6), 81-84.
- Dodia, D.A., Patel, I.S., Pathak, A.R. (1998). Antifeedant Properties of Some Indigenous Plant Extracts against Larvae of *Helicoverpa armigera*. *Pestology*, 1(9), 21-22.

- Elango, G., Abdul Rahuman, A., Bagavan, A., Kamaraj, C., Abdus Zahir, A., Rajakumar, G., Marimuthu, S., & Santhoshkumar, T. (2010). Studies on Effects of Indigenous Plant Extracts on Malarial Vector, *Anopheles subpictus* Grassi (Diptera: Culicidae). *Trop Biomed*, 2(7), 143-154.
- Elango, G., Abdul Rahuman, A., Kamaraj, C., Bagavan, A., Abdus Zahir, A., Santhoshkumar, T., & Rajakumar, G. (2012). Efficacy of Medicinal Plant Extracts against Formosan Subterranean Termite, *Coptotermes formosanus*. *Industrial Crops and Products*, 36(1), 524-530.
- Elango, G., & Rahuman, A. A. (2011). Evaluation of Medicinal Plant Extracts Against Ticks and Fluke. *Parasitology Research*, 10(8), 513-519.
- Elango, G., Zahir, A. A., Bagavan, A., Kamaraj, C., Rajakumar, G., Santhoshkumar, T., Marimuthu, S., & Rahuman, A. A. (2011). Efficacy of Indigenous Plant Extracts on the Malaria Vector *Anopheles subpictus* Grassi (Diptera: Culicidae). *Indian Journal of Medicine Research*, 13(4), 375-383.
- El Kady, H., & Devine, G. J. (2003). Insecticide Resistance in Egyptian Populations of the Cotton Whitefly, *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Pest Management Science*, 59(8), 865-871.
- El-Wakeil, N. E. (2013). Botanical Pesticides and Their Mode of Action. *Gesunde Pflanzen*, 65(4), 125-149.
- Espinosa, P. J., Bielza, P., Contreras, J., & Lacasa, A. (2002). Insecticide Resistance in Field Populations of *Frankliniella occidentalis* (Pergande) in Murcia (South-east Spain). *Pest Management Science*, 5(8), 967-971.
- FAO. (2004). FAO statistic database. Available at <http://www.faostat.fao.org>
- Forget, G., Goodman, T., & Villiers, A. (1993). *Impact of pesticide use on health in developing countries*. Ottawa: International Development Research Centre.
- Frohlich, D. R., & Brown, J. K. (1994). Mitochondrial 16S Ribosomal Subunit as a Molecular Marker in *Bemisia tabaci* and Implications for Population Variability. *Bemisia Newsletter*, 8(1), 3-12.
- Frohlich, D. R., Torres-Jerez, I. I., Bedford, I. D., Markham, P. G., & Brown, J. K. (1999). A Phylogeographical Analysis of the *Bemisia tabaci* Species Complex Based on Mitochondrial DNA Markers. *Molecular Ecology*, 8(1), 1683-1691.
- Gennadius, P. (1889). Disease of Tobacco Plantation in the Trikonía: The Aleyrodid of Tobacco. *Ellenike Georgia*, 5(1), 1-13.

- Gerling, D., Alomar, O., & Arno, J. (2001). Biological Control of *Bemisia tabaci* Using Predators and Parasitoids. In S. E. Naranjo & P. C. Ellsworth (Eds.), *Special Issue: Challenges and Opportunities for Pest Management of Bemisia tabaci in the New Century* (pp. 779-799). New York, NY: Crop Protection.
- Gerling, D., & Mayer, R. T. (1996). *Bemisia: Taxonomy, Biology, Damage, Control and Management*. Andover, UK: Intercept Limited.
- Ghosh, K. B., Datta, K. A., Mandal, A., Dubey, K. P., & Halder, S. (2012). An Overview on *Andrographis Paniculata* (Burm. F.) Nees. *International Journal of Research in Ayurveda & Pharmacy*, 3(6), 752-760.
- Gilbertson, R. L. (2008). *UC IPM Pest Management Guidelines: Tomato*. California: UC ANR Publication Diseases.
- Gill, R. J. (1990). The Morphology of Whiteflies. In D. Gerling (Ed.), *Whiteflies: Their Bionomics. Pest Status and Management* (pp. 13-46). Andover, UK: Intercept Ltd.
- Goswami, B. K. & Vijayalakshmi, K. (1985). In Vitro and Pot Culture Studies on the Effect of Dry Products of *Andrographis paniculata*, *Calendula officinalis*, *Ephydra fluctunus* and *Solanum khasianum* on *Meloidogyne incognita*. *Indian Journal of Nematology*, 15(2), 264-266.
- Govindarajan, M. (2011). Evaluation of *Andrographis paniculata* Burm.f. (Family: Acanthaceae) Extracts against *Culex quinquefasciatus* (Say.) and *Aedes aegypti* (Linn.) (Diptera: Culicidae). *Asian Pacific Journal of Tropical Medicine*, 4(1), 176-181.
- Govindarajan, M., & Sivakumar, R. (2012). Adulticidal and Repellent Properties of Indigenous Plant Extracts against *Culex quinquefasciatus* and *Aedes aegypti* (Diptera: Culicidae). *Parasitology Research*, 1(10), 1607-1620.
- Govindarajan, M., Sivakumar, R. (2011). Mosquito Adulticidal and Repellent Activities of Botanical Extracts against Malarial Vector, *Anopheles stephensi* Liston (Diptera: Culicidae). *Asian Pacific Journal of Tropical Medicine*, 4(1), 941-947.
- Grainge, M., & Ahmed, S. (1988). *Handbook of Plants with Pest-Control Properties*. New York, NY: Wiley.
- Guleria, S., & Tiku, A. K. (2009). Botanicals in Pest Management: Current Status and Future Perspectives. In R. Peshin & A. K. Dhawan (Eds.), *Integrated Pest Management: Innovation-Development Process* (pp. 317-329). Netherlands: Springer.

- Gupta, S., Choudhry, M. A., Yadava, J. N. S., Srivastava, V., & Tandon, J. S. (1990). Antidiarrhoeal Activity of diterpenes of *Andrographis paniculata* (Kal-Megh) Against *Escherichia coli* Enterotoxin in InVivo Models. *Pharmaceutical Biology*, 28(4), 273–283.
- Gupta, V., & Srivastava, V. K. (1995). Evaluation Studies on Kalmegh (*Andrographis paniculata* Nees). *Indian Journal of Human Genetics Resource*, 8(1), 141-143.
- Hamburger, M., Baumann, D., & Adler, S. (2004). Supercritical Carbon Dioxide Extraction of Selected Medicinal Plants, Effects of High Pressure and Added Ethanol on Yield of Extracted Substances. *Phytochemical analysis*, 15(1), 46-54.
- Hammad, E. A. F., & McAuslane, H. J. (2006). Effect of *Melia azedarach* L. Extract on *Bemisia argentifolii* (Hemiptera: Aleyrodidae) and Its Biocontrol Agent *Eretmocerus rui* (Hymenoptera: Aphelinidae). *Environmental Entomology*, 3(5), 740-745.
- Hammad, E. A. F., Zournajian, H., & Talhouk, S. (2001). Efficacy of Extracts of *Melia Azedarach* L. callus, leaves and fruits Against Adults of the Sweetpotato Whitefly *Bemisia tabaci* (Hom: Aleyrodidae). *Journal of Applied Entomology*, 12(5), 483-488.
- Hanapi, N. A., Azizi, J., Ismail, S., & Mansor, S. M. (2010). Evaluation of Selected Malaysian Medicinal Plants on Phase I Drug Metabolizing Enzymes, CYP2C9, CYP2D6 and CYP3A4 Activities in vitro. *International journal of Pharmacology*, 6(4), 1-6.
- Hancke, J., Burgos, R., Caceres, D., & Wikman, G. (1995). A Double Blind Study with a New Monodrug Kan Jang: Decrease of Symptoms and Improvement in the Recovery from Common Colds. *Phytotherapy Research*, 9(8), 559–562.
- Handa, S. S., Khanuja, S. P. S., Longo, G., & Rakesh, D. D. (2008). Extraction Technologies for Medicinal and Aromatic Plants. In S. S. Handa (Ed.), *An Overview of Extraction Techniques for Medicinal and Aromatic Plants* (pp. 21-37). Trieste, Italy: International Centre for Science and High Technology.
- Haverty, M. L., & Robertson, J. L. (1982). Laboratory Bioassays for Selecting Candidate Insecticides and Application Rates for Field Tests on the Western Spruce Budworm. *Journal of Economic Entomology*, 75(2), 179-182.
- Harborne, J. B. (1977). *Introduction to ecological biochemistry*. London: Academic Press.

- Hedin, P. A., Hollingworth, R. M., Masler, E. P., Miyamoto, J., & Thompson, D. G. (1997). *Phytochemicals for Pest Control*. Washington, DC: American Chemical Society.
- Hermawan, W., Nakajima, S., Tsukuda, R., Fujisaki, K., & Nakasuji, F. (1997). Antifeedant Activity of Active Fractions from a Tropical Plant, *Andrographis paniculata* Nees against The Diamondback Moth. *Entomology Cornell*, 4(2), 134-138.
- Hermawan, W., Nakajima, S., Tsukuda, R., Fujisaki, K., & Nakasuji, F. (1997). Isolation of an Antifeedant Compound from *Andrographis paniculata* (Acanthaceae) against the Diamondback Moth, *Plutella xylostella* (Lepidoptera: Yponomeutidae). *Applied Entomology and Zoology*, 3(2), 551-559.
- Hilje, L. (2000). Agricultural Practices for Managing *Bemisia tabaci*: Integrated Pest Management. *Costa Rica*, 5(6), 22-30.
- Hilje, L., & Hanson, P. (1998). La Biodiversidad Tropicaly El Manejo Integrado De Plagas. *Manejo Integrado Plagas*, 48(1), 1-10.
- Hilje, L., & Hanson, P. (1998). Tropical biodiversity and integrated pest management: Integrated Pest Management. *Costa Rica*, 4(8), 1-10.
- Hilje, L., & Mora, G. A. (2006). Promissory Botanical Repellents/Deterrents for Managing two Key Tropical Insect Pests, The Whitefly *Bemisia tabaci* and the Mahogany Shootborer *Hypsipyla grandella*. In M. Rai & M. C. Carpinella (Eds.), *Naturally occurring bioactive compounds* (pp. 379-403). Amsterdam: Elsevier.
- Hilje, L., Costa, H. S., & Stansly, P. A. (2001). Cultural Practices for Managing *Bemisia tabaci* and Associated Viral Diseases. *Crop Protection*, 20(9), 801-812.
- Horowitz, A. R., & Ishaaya, I. (1996). Chemical Control of *Bemisia tabaci*: Management and Application. In: D. Gerling & R. T. Mayer (Eds.), *Bemisia: 1995 Taxonomy, Biology, Damage, Control and Management* (pp. 537-556). Andover, Hants, UK: Intercept Ltd.
- Horowitz, R., Denholm, I., & Morin, S. (2007). *Resistance to Insecticides in the TYLCV Vector, Bemisia tabaci: Tomato Yellow Leaf Curl Virus Disease*. Netherlands: Springer.
- Houndete, T. A., Ketoh, G. K., Hema, O. S., Brevault, T., Glietho, I. A., & Martin, T. (2010). Insecticide Resistance in Field Populations of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in West Africa. *Pest Management Science*, 66(11), 1181-1185.

- Hu, J., De Barro, P., Zhao, H., Wang, J., Nardi, F., & Liu, S. S. (2011a). An Extensive Field Survey Combined With a Phylogenetic Analysis Reveals Rapid and Widespread Invasion of Two Alien Whiteflies in China. *Plos One*, 6(1), 16-61.
- Hu, X., Dennehy, T. J., Ni, X., Zhao, H., Nichols, R. L., & Li, X. (2011b). Potential Adaptation of A Q Biotype Whitefly Population from Poinsettia to Field Crops. *Insect Science*, 1(8), 719-728.
- Hu, Y. M., & Wu, Z. F. (1995). The Chemical Component of Volatile Oil and Plantation of Several Species of Xin-yi. *Forest Science and Technology*, 13(2), 26-28.
- Index Kewensis, Royal Botanic Garden, Oxford University Press, Oxford. (Vol. 1-18)(1977-87). Vol. 1-18).
- Insang, A. (2001). Effect of Some Medicinal Plant Extracts on *Tetranychus truncatus* (Ehara). *King Mongkut's Agricultural Journal*, 1(9), 15-22.
- IRAC Susceptibility Test Methods Series. (2009). Retrieved July 31st, 2012, from http://www.iraconline.org/content/uploads/2009/09/Method_015_v3June09.pdf and http://www.iraconline.org/content/uploads/2009/09/Method_016_v3June09.pdf.
- Islam, M. T., Castle, S. J., & Ren, S. (2009). Compatibility of the Insect Pathogenic Fungus *Beauveria Bassiana* with Neem against Sweetpotato Whitefly, *Bemisia tabaci*, on Eggplant. *Entomologia Experimentalis et Applicata*, 13(4), 28-34.
- Isman, M. B. (2008). Botanical Insecticides. In J. L. Capinera (Ed.), *Encyclopedia of Entomology* (pp. 549-552). Vancouver, BC: Springer.
- Isman, M. B. (2006). Botanical Insecticides, Deterrents, and Repellents in Modern Agriculture and an Increasingly Regulated World. *Annual Review Entomology*, 5(1), 45-66.
- Isman, M. B. (2010). Botanical Insecticides, Deterrents, Repellents and Oils. In B. P. Singh (Ed.), *Industrial Crops and Uses* (pp. 433-445). New York, NY: © CAB International Uses.
- Jain, D. C., Gupta, M. M., Saxena, S., & Kumar, S. (2000). LC Analysis of Hepatoprotective Diterpenoids from *Andrographis paniculata*. *Journal of Pharmaceutical and Biomedical Analysis*, 2(2), 705-709.
- Jbilou, R., Ennabili, A., & Sayah, F. (2006). Insecticidal Activity of Four Medicinal Plant Extracts against *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *African Journal of Biotechnology*, 5(10), 936-940.

- Jiu, M., Zhou, X. P., Tong, L., Xu, J., Yang, X., Wan, F. H., & Liu, S. S. (2007). Vector-Virus Mutualism Accelerates Population Increase of an Invasive Whitefly. *PLOS ONE*, 2(1), 1-8. doi:10.1371/journal.pone.0000182.
- Joyce, A. L., Bellows, T. S., & Headrick, D. H. (1999). Reproductive Biology and Search Behavior of *Amitus bennetti* (Hymenoptera: Platygasteridae), a Parasitoid of *Bemisia argentifolii* (Homoptera: Aleyrodidae). *Environmental Entomology*, 28(2), 282-289.
- Kanokwan, J., & Nobuo, N. (2008). Pharmacological Aspects of *Andrographis paniculata* on Health and Its Major Diterpenoid Constitute Andrographolide. *Journal of Health Science*, 54(4), 370-381.
- Katakya, A., & Handique, P. J. (2010). A Brief Overview on *Andrographis paniculata* (Burm. F) Nees., a High Valued Medicinal Plant: Boon over Synthetic Drugs. *Asian Journal of Science and Technology*, 6(1), 113-118.
- Kathuria, V., & Kaushik, N. (2005). Feeding Inhibition of *Helicoverpa armigera* (Hübner) by *Eucalyptus camaldulensis* and *Tylophora indica* Extracts. *Insect Science*, 12(4), 249-254.
- Katz, J., Prescott, K., & Woolf, A. D. (1996) Strychnine Poisoning From a Cambodian Traditional Remedy. *The American journal of emergency medicine*, 14(5), 475-477.
- Kaul, V., Shankar, U., & Khushu, M. K. (2009). *Bio-Intensive Integrated Pest Management in Fruit Crop Ecosystem: Integrated Pest Management: Innovation-Development Process*. Netherlands: Springer.
- Khandelwal, K. R. (2005). Practical Pharmacognosy. In N. Prakashan (13th Ed.), *Technique and experiments* (pp. 157-160). India: Nirali Prakashan.
- Khare, C. P. (2007). *Andrographis paniculata*. In C. P. Khare (Ed.). *Indian Medicinal Plants An Illustrated Dictionary* (pp. 49-50). New Delhi: Springer.
- Kirar, K., Kaurav, D., Chourasiya, J., & Shukla, R. N. (2012). Extraction And Identification of Diterpenoid Lactone from *Andrographis paniculata*. *International Journal of Pharmaceutical Sciences Review*, 4(10), 53-56.
- Kirk, A. A., Lacey, L. A., Brown, J. K., Ciomperlik, M. A., Goolsby, J. A., Vacek, D. C., & Napompeth, B. (2000). Variation in the *Bemisia tabaci* s. 1. Species Complex (Homoptera: Aleyrodidae) and its Natural Enemies Leading to Successful Biological Control of Bemisia Biotype B in the USA. *Bulletin of Entomological Research*, 90(4), 317-327.

- Kordali, S., Cakir, A., Ozer, H., Cakmakci, R., Kesdek, M., & Mete, E. (2008). Antifungal, Phytotoxic and Insecticidal Properties of Essential Oil Isolated from Turkish *Origanum acutidens* and Its Three Components, Carvacrol, Thymol and P-Cymene. *Bioresource Technology*, 99(18), 8788-8795.
- Koteswara Rao, Y., Vimalamma, G., Venkata Rao, C., & Tzeng, Y. M. (2004). Flavonoids and Andrographolides from *Andrographis paniculata*. *Phytochemistry*, 6(5), 2317-2321.
- Koul, O., & Dhaliwal, G. S. (2001). *Phytochemical Biopesticides*. Amsterdam: Harwood Academy.
- Kulyal, P., Tiwari, U. K., Shukla, A., & Gaur, A.K. (2010). Chemical Constituents Isolated from *Andrographis paniculata*. *Indian Journal of Chemistry*, 4(9), 356-359.
- Kumar, A., Dora, J., Singh, A., & Tripathi, R. (2012). A Review on King of Bitter (Kalmegh). *International Journal of Research in Pharmacy and Chemistry*, 2(1), 116-124.
- Kumar, P., Poehling, H. M., & Borgemeister, C. (2005). Effects of Different Application Methods of Azadirachtin against Sweetpotato Whitefly *Bemisia tabaci* Gennadius (Hom. Aleyrodidae) on Tomato Plants. *Journal of Applied Entomology*, 12(9), 489-497.
- Kumar, S., Geetha, K. A., Jat, R. S., & Varghese, T. S. (2012-2013). *Annual Report Directorate of Medicinal and Aromatic Plants Research Cujarat, India*.
- Kumbhar, P. P., & Dewang, P. M. (2001). Monoterpenoids: Botanical Managers of Pests. In M. J. Khan (Ed.), *Agriculture Today: The National Agriculture Magazine* (pp. 47-49). Lajpat Nagar, New Delhi: Centre for Agriculture and Rural Development.
- Kumoro, A. C., & Hasan, M. (2006). Extraction of Diterpenoid Lactones of *Andrographis paniculata* using Liquid Solvents: Effect of Solvent's Hildebrand Solubility Parameter on Extraction Efficiency. *National Technical Postgraduate Symposium*, 4(1), 13-16.
- Kumoro, A. C., Hasan, M., & Singh, H. (2009). Effects of Solvent Properties on the Soxhlet Extraction of Diterpenoid Lactones from *Andrographis paniculata* Leaves. *Science Asia*, 35(1), 306-309.
- Kumoroa, A. C., Hasana, M., & Singha, H. (2010). Experimental and Modelling Studies of Andrographolide Extraction from *Andrographis Paniculata*. *International Journal of Chemical, Environmental and Pharmaceutical Research*, 1(1), 32-36.

- Kuppusamy, C., & Murugan, K. (2008). Oviposition Deterrent, Ovicidal and Gravid Mortality Effects of Ethanolic Extract of *Andrographis paniculata* Nees against the Malarial Vector *Anopheles stephensi* Liston (Diptera: Culicidae). *Entomological Research*, 3(8), 119–125.
- Lacey, L. A., Fransen, J. J., & Carruthers, R. (1996). Global distribution of naturally occurring fungi of Bemisia, their biologies and use as biological control agents. In D. Gerling & R. T. Mayer (Eds.), *Bemisia 1995: Taxonomy, Biology, Damage, Control and Management*, Intercept (pp. 133-401). Andover, UK: Intercept Limited.
- Latiff, A. (1994). Conservation of medicinal plant and aromatic plants resources through in situ and ex situ methods. Report of the 2nd Regional Meeting of Asian Region Countries on G15 Gene Bank for Medicinal and Aromatic Plants Project, p. 31– 43. Kuala Lumpur: Ministry of Agriculture.
- Lattoo, S. K., Khan, S., Dhar, A. K., Choudhary, D. K., Gupta, K. K., & Sharma, P. R. (2006). Genetics and Mechanism of Induced Male Sterility in *Andrographis paniculata* (Burm. f.) Nees and Its Significance. *Current Science*, 9(1), 515-519.
- Lee, Y. H., Charles, A. L., Kung, H. F., Ho, C. T., & Huang, T. C. (2010). Extraction of Nobiletin and Tangeretin from *Citrus depressa* (Hayata) by Supercritical Carbon Dioxide with Ethanol as Modifier. *Industrial crops and products*, 31(1), 59-64.
- Legg, J., Gerling, D., & Neuenschwander, P. (2003). *Biological Control of Whiteflies in Sub-Saharan Africa: Biological Control in IPM System in Africa*. Wallingford, UK: CAB International.
- Legg, J. P., Owor, B., Sseruwagi, P., & Ndunguru, J. (2006). Cassava Mosaic Virus Disease in East and Central Africa: Epidemiology and Management of a Regional Pandemic. *Advances in Virus Research*, 6(7), 355-418.
- Leonard, S., Capote, R., Germonprez, N., Van Puyvelde, L., Kimpe, N. D., & Vermeersch, H. (2003). Liquid Chromatographic Method for Analysis of Saponins in *Maesa Balansae* Extract Active against Leishmaniasis. *Journal of Chromatography*, 1(1)1012, 1039.
- Leshkowitz, D., Gazit, S., Reuveni, E., Ghanim, M., Czosnek, H., McKenzie, C., Shatters, R. L., & Brown, J. K. (2006). Whitefly (*Bemisia tabaci*) Genome Project: Analysis of Sequenced Clones from Egg, Instar, And Adult (viruliferous and non-viruliferous) cDNA libraries. *BMC genomics*, 7(1), 79-89.

- Li, M., Hu, J., Xu, F. C., & Liu, S. S. (2010). Transmission of Tomato Yellow Leaf Curl Virus by Two Invasive Biotypes and a Chinese Indigenous Biotype of the Whitefly *Bemisia tabaci*. *International Journal of Pest Management*, 5(6), 275-280.
- Li, S. J., Xue, X., Ahmed, M. Z., Ren, S. X., Du, Y. Z., Wu, J. H., Cuthbertson, A. G. S., & Qiu, B. L. (2011). Host Plants and Natural Enemies of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in China. *Insect Science*, 1(8), 101-120.
- Li, X., Degain, B. A., Harpold, V. S., Marçon, P. G., Nichols, R. L., Fournier, A. J., & Ellsworth, P. C. (2012). Baseline susceptibilities of B- and Q-biotype *Bemisia tabaci* to anthranilic diamides in Arizona. *Pest management science*, 68(1), 83-91.
- Li, Z. H., Zhou, X. P., Zhang, X., & Xie, Y. (2004). Molecular Characterization of Tomato-Infecting Begomoviruses in Yunnan, China. *Archives of Virology*, 14(9), 1721-1732.
- Lingampally, V., Solanki, V. R., Kaur, A., & Raja, S. S. (2013). Effect of Andrographolide on the Protein Content of *Tribolium confusum* (Duval). *International Journal of Pure and Applied Zoology*, 1(1), 70-79.
- Lingampally, V., Solanki, V. R., & Raja, S. S. (2012). Andrographolide: An effective Anti-Fertility Agent for the Control of *Tribolium confusum*. *Asian Journal of Plant Science and Research*, 2(1), 313-317.
- Liu, J., Li, M., Li, J. M., Huang, C. j., Zhou, X. P., Xu, F. C., Liu, S. S. (2010). Viral Infection of Tobacco Plants Improves Performance of *Bemisia tabaci* but More so for an Invasive than for an Indigenous Biotype of the Whitefly. *Journal of Zhejiang University - Science*, 1(1), 30-40.
- Liu, S. S., Colvin, J., & De Barro, P. J. (2012). Species Concepts as Applied to the Whitefly *Bemisia tabaci* Systematics: How Many Species Are There? *Journal of Integrative Agriculture*, 11(1), 176-186.
- Liu, S. S., De Barro, P. J., Xu, J., Luan, J. B., Zang, L. S., Ruan, Y. M., & Wan, F. H. (2007). Asymmetric Mating Interactions Drive Widespread Invasion and Displacement in a Whitefly. *Science*, 31(8), 1769-1772.
- Liu, T. X., & Stansly, P. A. (1995). Toxicity and Repellency of some Biorational Insecticides to *Bemisia argentifolii* on Tomato Plants. *Entomologia Experimentalis et Applicata*, 7(4), 137-143.
- Lowe, S., Browne, M., Boudjelas, S., & De Poorter, M. (2004). 100 of the World's Worst Invasive Alien Species a Selection from the Global Invasive Species Database. New Zealand.

- Maiti, K., Mukherjee, K., Murugan, V., Saha, B. P., & Mukherjee, P. K. (2010). Enhancing Bioavailability and Hepatoprotective Activity of Andrographolide from *Andrographis paniculata*, a Well-Known Medicinal Food, Through Its Herbosome. *Journal of Science Food Agriculture*, 9(1), 43-51.
- Majee, C., Gupta, B. K., Mazumder, R., & Chakraborty, G. S. (2011). HPLC Method Development and Characterization of Bio-Active Molecule Isolated from *Andrographis paniculata*. *International Journal of PharmTech Research*, 3(3), 1586-1592.
- Mamatha, A. (2011). Quantitative HPTLC Analysis of Andrographolide in *Andrographis Paniculata* Obtained from Different Geographical Sources (India). *International Journal of Pharmacy & Pharmaceutical Sciences*, 3(2), 42-44.
- Malahubban, M., Alimon, A. R., Sazili, A. Q., Fakurazi, S., & Zakry, F. A. (2013). Phytochemical analysis of *Andrographis paniculata* and *Orthosiphon stamineus* leaf extracts for their antibacterial and antioxidant potential. *Tropical biomedicine*, 30(3), 467-480.
- Marco, G. J., Hollingworth, R. M., & Durham, W. (1987). *Silent Spring Revisited*. Washington, DC: American Chemical Society.
- Martin, J. H. (2004). Whiteflies of Belize (Hemiptera: Aleyrodidae). Part 1: Introduction and Account of the Subfamily *Aleurodicinae* *Quaintance* and Baker. *Zootaxa*, 68(1), 3-119.
- Martin, N. A. (1999). Whitefly Biology, Identification and Life Cycle. *Crop and Food Research*, 9(1), 1-8.
- McAuslane, H. J. (2012). *Sweetpotato Whitefly B Biotype of Silverleaf Whitefly, Bemisia Tabaci (Gennadius) or Bemisia Argentifolii Bellows and Perring (Insect: Hemiptera: Aleyrodidae)*. (Unpublished Doctoral Dissertation). University of Florida.
- McKenzie, C. L., Anderson, P. K., & Villarreal, N. (2004). An Extensive Survey of *Bemisia tabaci* (Homoptera: Aleyrodidae) in Agricultural Ecosystems in Florida. *Florida Entomologist*, 8(7), 403-407.
- McLaughlin, G. A. (1973). *History of pyrethrum*. New York, NY: Academic.
- Medina-Ortega, K. J. (2011). *Poinsettia (Euphorbia pulcherrima Willd. ex Klotzsch: Euphorbiaceae) Resistance Mechanisms against the Silverleaf Whitefly, Bemisia tabaci (Gennadius) (Hemiptera: Aleyrodidae) Biotype B*. (Unpublished Doctoral dissertation). The Ohio State University.

- Meenatchisundaram, S., Parameswari, G., & Michael, A. (2009). Studies On Antivenom Activity of *Andrographis Paniculata* and *Aristolochia Indica* Plant Extracts Against *Daboia Russelli* Venom By In Vivo And In Vitro Methods. *Indian Journal of Science and Technology*, 2(4), 76–79.
- Misra, P., Pal, N. L., Guru, P. Y., Katiyar, J. C., Srivastava, V., & Tandon, J. S. (1992). Antimalarial Activity of *Adrographis paniculata* (Kalmegh) against *P. berghei* NK65 in *Mastomys natalensis*. *International Journal of Pharmacognosy and Phytochemic*, 3(1), 263-274.
- Misra, S. S. (1993). Potato Pests and Their Management. *Journal of Applied Zoological Research*, 4(1), 27-32.
- Mohan, M. C., Reddy, N. P., Devi, U. K., Kongara, R., & Sharma, H. C. (2007). Growth and Insect Assays of *Beauveria bassiana* with Neem to test Their Compatibility and Synergism. *Biocontrol Science and Technology*, 1(7), 1059-1069.
- Mordue, A. J., & Blackwell, A. (1993). Azadirachtin: an Update. *Journal of Insect Physiology*, 39(1), 903-924.
- Mound, L. A., & Halsey, S. H. (1978). *Whiteflies of the World: A Systematic Catalogue of the Aleyrodidae (Homoptera) With Host Plants and Natural Enemy Data*. Chichester: British Museum of (Natural History) and John Willey and Sons.
- Mukherjee, P. K. (2002). *Analytical Profiles of Selected Medicinal Plants: Quality Control of Herbal Drugs, an Approach to Evaluation of Botanicals*. New Delhi, India: Business Horizons.
- Muniramappa, R. P., & Farooqi, A. A., Gowda, H. G. R., & Maricapu, S. (1997). Influence of Macronutrients on Yield and Active Principle Content in Kalmegh. *Journal of Medicinal and Aromatic Plant Sciences*, 19(2), 1039-1042.
- Mustaffa, F., Indurkar, J., Ali, N. I. M., Hanapi, A., Shah, M., Ismail, S., & Mansor, S. M. (2011). A Review of Malaysian Medicinal Plants with Potential Antidiabetic Activity. *Journal of Pharmacy Research*, 4(11), 4217-4224.
- Nandi, R. P. (1992). Increase in Productivity Regime of Some Well-Known Medicinal and Aromatic Plants Used in Ayurvedic System of Medicines. *Advances in Plant Sciences*, 5(1), 274-282.
- Naranjo, S. E., & Legg, J. P. (2010). Biology and Ecology of *Bemisia tabaci*. In P. A. Stansly & S. E. Naranjo (Eds.), *Bemisia: Bionomics and Management of a Global Pest* (pp. 105-107). California: Springer.

- Nauen, R., Elbert, A., McCaffery, A., Slater, R., & Sparks, T. C. (2012). IRAC: Insecticide Resistance, And Mode of Action Classification of Insecticides. *Modern Crop Protection Compounds*, 1(3), 935-955.
- Nigg, H. N., Nordby, H. E., Beier, R. C., Dillman, A., Macias, C., & Hansen, R. C. (1993). Phototoxic Coumadin's in Limes. *Food Chemistry Toxicology*, 31(1), 331-335.
- Niranjan, A., Tewari, S. K., & Lehri, A. (2010). Biological Activities of kalmegh (*Andrographis paniculata* Nees). *Indian Journal of Natural Products and Resources*, 1(2), 125-135.
- Oliveira, M. R. V., Henneberry, T. J., & Anderson, P. (2001). History, Current Status, and Collaborative Research Projects for *Bemisia tabaci*. *Crop Protection*, 20(9), 709-723.
- Organic Farming: Pest and Disease Control. (n.d.). Retrieved 15th Sep 2013, from http://www.agritech.tnau.ac.in/org_farm/orgfarm_pestanddisease.html
- Osborne, L. S., & Landa, Z. (1992). Biological Control of Whiteflies with Entomopathogenic Fungi. *Entomologica Fennica*, 7(5), 456-471.
- Paiva, S. R., Figueiredo, M. R., Aragao, T. V., & Kaplan, M. A. C. (2003). Antimicrobial Activity in Vitro of Plumbagin Isolated from Plumbago Species. *Memories do Institute Oswaldo Cruz*, 9(8): 959-961.
- Palumbo, J. C., Horowitz, A. R., & Prabhaker, N. (2001). Insecticidal control and resistance management for *Bemisia tabaci*. *Crop Protection*, 20(1), 739-765.
- Panda, N., & Khush, G. S. (1995). *Host plant resistance to insects*. Wallingford, UK: CAB International-IRRI.
- Pandey, A., & Tripathi, S. (2014). Concept of Standardization, Extraction and Pre Phytochemical Screening Strategies for Herbal Drug. *Journal of Pharmacognosy and Phytochemistry*, 2(5), 115-119.
- Panneerselvam, C., & Murugan, K. (2013). Adulticidal, Repellent, and Ovicidal Properties of Indigenous Plant Extracts against the Malarial Vector, *Anopheles stephensi* (Diptera: Culicidae). *Parasitol Research*, 11(2), 679-692.
- Pant, N., Jain, D. C., Bhakuni, R. S., Prajapati, V., Tripathi, A. K., & Kumar, S. (2001). Zederone: a Sesquiterpenic Keto-dioxide from Curcuma Aromatica. *Indian Journal Chemistry*, 40(1), 87-88.

- Patnaik, S., Rout, K., Pal, S., Panda, P. K., Mukherjee, P. S., & Sahoo, S. (2011). Essential Oils of Aromatic and Medicinal Plants as Botanical Biocide for Management of Coconut Eriophyid Mite (*Aceria guerreronis* Keifer). *Psyche*, 20(11), 1-5.
- Pavel, D., & Jitka, M. (2004). Recent Advances in Analysis of Chinese Medicinal Plants and Traditional Medicines. *Journal of Chromatography*, 8(12), 3-21.
- Perring, T. M. (2001). The *Bemisia tabaci* Species Complex. *Crop Protection*, 20(1), 725-737.
- Perring, T. M., Cooper, A. D., Rodriguez, R. J., Farrar, C. A., & Bellows, T. S. (1993). Identification of a Whitefly Species by Genomic and Behavioral Studies. *Science*, 25(9), 74-77.
- Perry, A. S., Yamamoto, I., Ishaaya, I., & Perry, R. Y. (1998). *Insecticides in Agriculture and Environment: Retrospect and Prospects*. Berlin: Springer-Verlag.
- Pholphana, N., Rangkadilok, N., Thongnest, S., Ruchirawat, S., Ruchirawat, M., & Satayavivad, J. (2004). Determination and Variation of Three Active Diterpenoids in *Andrographis Paniculata* (Burm.F.) Nees. *Phytochemical Analysis*, 15(6), 365-371.
- Pickett, J. A., Wadhams, L. J., & Woodcock, C. M. (1997). Developing Sustainable Pest Control from Chemical Ecology. *Agriculture, Ecosystems & Environment*, 6(4), 149-156.
- Pinheiro, P. V., Quintela, E. D., Oliveira, J. P. D., & Seraphin, J. C. (2009). Toxicity of Neem Oil to *Bemisia tabaci* Biotype B Nymphs Reared on Dry Bean. *Pesquisa Agropecuaria Brasileira*, 44(4), 354-360.
- Polston, J. E., & Anderson, P. K. (1997). The Emergence of Whitefly-Transmitted Geminiviruses in Tomato in the Western Hemisphere. *Plant Disease*, 8(1), 1358-1369.
- Poornima, K., & Vadivelu, S. (1993). Comparative Efficacy of Nematicides, Oil Cakes and Plant Extracts in the Management of *Meloidogyne incognita*, *Pratylenchus delattrei* and *Rotilenchulus reniformis* on brinjal. *Indian Journal of Nematology*, 2(3), 170-173.
- Prakash, A., & Rao, J. (1996). *Section A: Botanical Pesticides against Insects: Botanical Pesticides in Agriculture*. Florida: CRC Press.
- Prakash, A., & Rao, J. (1997). *Botanical Pesticides in Agriculture*. Boca Raton: CRC Press.

- Prathanturarug, S., Soonthornchareonnon, N., Chuakul, W., & Saralamp, P. (2007). Variation in Growth and Diterpene Lactones Among Field-Cultivated *Andrographis paniculata*. *Journal of Natural Medicines*, 6(1), 159-163.
- Qader, S. W., Abdulla, M. A., Chua, L. S., Najim, N., Zain, M. M., & Hamdan, S. (2011). Antioxidant, Total Phenolic Content and Cytotoxicity Evaluation of Selected Malaysian Plants. *Molecules*, 16(4), 3433-3443.
- Quaintance, A. L., & Baker, A. C. (1913). Classification of the Aleyrodidae Part I. *Technical Series Bureau of Entomology U.S.*, 2(7), 75-77.
- Raaman, N. (2006). *Phytochemical Techniques*. Delhi: New India Publishing Agency.
- Rai, M., & Carpinella, M. C. (2006). *Naturally Occurring Bioactive Compounds* (Vol. 3). Chicago: Elsevier.
- Raina, A. P., Gupta, V., Sivaraj, N., & Dutta, N. (2013). *Andrographis paniculata* (Burm. f.) Wall. ex Nees (kalmegh), a Traditional Hepatoprotective Drug from India. *Genetic Resources and Crop Evolution*, 60(1), 1181-1189.
- Rajakumar, G., & Abdul Rahuman, A. (2011). Larvicidal Activity of Synthesized Silver Nanoparticles Using *Eclipta Prostrate* Leaf Extract against Filariasis and Malaria Vectors. *Acta tropica*, 118(3), 196-203.
- Rajagopal, S., Kumar, R. A., Deevi, D. S., Satyanarayana, C., & Rajagopalan, R. (2003). Andrographolide, a Potential Cancer Therapeutic Agent Isolated from *Andrographis paniculata*. *Journal of Experimental Therapeutics and Oncology*, 3(1), 147-158.
- Raja, N., Elumalai, K., Jayakumar, M., Jeyasankar, A., & Ignacimuthu, S. (2002). Antifeedant Activity of Solvent Extracts of 50 Plants on *Spodoptera litura* Fab. (Lepidoptera: Noctuidae) and *Helicoverpa armigera* Hubner (Lepidoptera: Noctuidae). *Malaysian Applied Biology Journal*, 3(1), 19-28.
- Ramlogan, R. (2004). *The Developing World and the Environment: Making the Case for Effective Protection of the Global Environment*. United State: University Press of America.
- Ramya, S., Gopinath, K., Karthikeyan, M., Sundarpandian, S. M., Periyathambi, N., Sundarajan, G., & Jayakumararaj, R. (2011). Effect of Crude Methanol Leaf Extracts of *Andrographis paniculata* (Burm.f) Nees on Larvae of *Helicoverpa armigera* (Hübner). *Environment & We an International Journal of Science & Technology*, 6(2), 21-28.

- Ramya, S., Rajasekaran, C., Sundararajan, G., Alaguchamy, N., & Jayakumararaj, R. (2008). Antifeedant Activity of Leaf Aqueous Extracts of Selected Medicinal Plants on VI instar larva of *Helicoverpa armigera* (Hübner). *Ethnobotanical Leaflets*, 1(2), 938-943.
- Rasdi, Z. M., Fauziah, I., Fairuz, K., Saiful, M. S., Jamaludin, B. M., Salmah, M. R., & Jusoff, K. (2009). Population Ecology of Whitefly, *Bemisia tabaci*, (Homoptera: Aleyrodidae) on Brinjal. *Journal of Agricultural Science*, 1(1), 1916-9752.
- Reddy, P. J., Krishna, D., Suryanarayana, M. U., & Jamil, K. (1992). A Microcomputer FORTRAN Program for Rapid Determination of Lethal Concentrations of Biocides in Mosquito Control. *Computer Applications in the Biosciences*, 8 (3), 209-213.
- Regnault-Roger, C., & Philogène, B. J. R. (2008). Past and Current Prospects for the Use of Botanicals and Plant Allelochemicals in Integrated Pest Management. *Pharmaceutical Biology*, 46(1), 41–52.
- Regnault-Roger, C., Philogène, B. J. R., & Vincent, C. (2005). Biopesticides of Plant Origin. *European Journal of Entomology*, 106(2), 224-313.
- Remia, K. M. (2012). Larvicidal and Pupicidal Effect of *Spilanthes acmella* and *Andrographis paniculata* on the Mosquito *Aedes aegypti*. *International Journal of Institutional Pharmacy and Life Sciences*, 2(1), 71-76.
- Renugadevi, G., Ramanathan, T., Shanmuga, P. R., & Thirunavukkarasu, P. (2013). Studies on Effects of *Andrographis paniculata* (Burm.f.) and *Andrographis lineata* nees (Family: Acanthaceae) extracts against two Mosquitoes *Culex quinquefasciatus* (Say.) and *Aedes aegypti* (Linn.). *Asian Pacific Journal of Tropical Medicine*, 6(1), 176-179.
- Ribeiro, S. G., Ambrozevicus, L. P., Avila, A. C., Bezerra, I. C., Calegario, R. F., Fernandes, J. J., Lima, M. F., de Mello, R. N., Rocha, H., & Zerbini, F. M. (2003). Distribution and Genetic Diversity of Tomato-Infecting Begomoviruses in Brazil. *Archives of Virology*, 14(8), 281-295.
- Robertson, J. L., Smith, K. C., Savin, N. E., & Lavigne, R. J. (1984). Effect of Dose Selection and Sample Size on the Precision of Lethal Dose Estimates in Dose Mortality Regression. *Journal of Economic Entomology*, 77(4), 833-837.
- Roditakis, E., Grispos, M., Morou, E., Kristoffersen, J. B., Roditakis, N., Nauen, R., & Tsagkarakou, A. (2009). Current Status of Insecticides Resistance in Q Biotype *Bemisia tabaci* Population from Crete. *Pest Management Science*, 6(5), 313-322.

- Roditakis, E., Roditakis, N. E., & Tsagkarakou, A. (2005). Insecticide Resistance in *Bemisia tabaci* (Homoptera: Aleyrodidae) Populations from Crete. *Pest Management Science*, 61(6), 577-582.
- Roy, S., Rao, K., Bhuvaneshwari, C., Giri, A., & Mangamoori, L. N. (2010). Phytochemical Analysis of *Andrographis paniculata* Extract and Its Antimicrobial Activity. *World Journal of Microbiology and Biotechnology*, 26(1), 85-91.
- Sanyal, U., Bhattacharyy, S., Patra, A., & Hazra, B. (2003). Liquid Chromatographic Separation of Derivatives of Diospyrin, a Bioactive Bisnaphthoquinonoid Plant-Product, and Analogous Naphthyl Compounds. *Phytochemical Analysis*, 10(17), 225-243.
- Saroya, A. S. (2010). *Herbalism, Phytochemistry and Ethnopharmacology*. New York, NY: Science Publishers, P.O.
- Sathyaseelan, V., & Bhaskaran, V. (2010). Efficacy of Some Native Botanical Extracts on the Repellency Property against the Pink Mealy Bug, *Maconellicoccus hirsutus* (Green) in Mulberry Crop. *Recent Research in Science and Technology*, 2(10), 35-38.
- Saunders, L. A. (2014). *Easy Organic Gardening*. Munich, Germany: OTB Ebook publishing.
- Saxena, R. C., Singh, R., Kumar, P., Yadav, S. C., Negi, M. P., Saxena, V. S., Joshua, A. J., Vijayabalaji, V., Goudar, K. S., Venkateshwarlu, K., & Amit, A. (2010). A Randomized Double Blind Placebo Controlled Clinical Evaluation of Extract of *Andrographis paniculata* (KalmCold) in Patients with Uncomplicated upper Respiratory Tract Infection. *Phytomedicine*, 1(7), 178-185.
- Schmutterer, H. (1969). *Pests of Crops in Northeast and Central Africa with Particular Reference to the Sudan*. Stuttgart: Gustav Fischer Verlag.
- Schmutterer, H., Ascher, K. R. S., & Rembold, H. (1982). *Natural Pesticides from the Neem Tree (Azadirachta indica A. Juss)*. Eschorn, Germany: GTZ.
- Scott, I. M., Jensen, H., Nicol, R., Lesage, L., Bradbury, R., Sanchez-Vindas, P., & Philogene, B. J. R. (2004). Efficacy of Piper (Piperaceae) Extracts for Control of Common Home and Garden Insect Pests. *Journal of economic entomology*, 97(4), 1390-1403.
- Shalan, E. A. S., Canyonb, D., Younesc, M. W. F., Abdel-Wahaba, H., & Mansoura, A. H. (2005). A Review of Botanical Phytochemicals with Mosquitocidal Potential. *Environment International*, 3(1), 1149-1166.

- Shadmany, M., Dzolkhifli, O., Rita, M. (2013). First Report of *Bemisia tabaci* (Hemiptera: Aleyrodidae) Biotype Q in Malaysia. *Florida Entomologist*, 9(6), 280-282.
- Shahi, M., Hanafi-Bojd, A. A., Iranshahi, M., Vatandoost, H., & Hanafi-Bojd, M. Y. (2010). Larvicidal efficacy of latex and extract of *Calotropis procera* (Gentianales: Asclepiadaceae) against *Culex quinquefasciatus* and *Anopheles stephensi* (Diptera: Culicidae). *Journal of Vector Borne Disease*, 47(1), 185-188.
- Shanker, C., & Uthamasamy, S. (2010). Evaluation of Some Medicinal Plants and Their Mixtures for Their Bio-Efficacy against Crop and Stored Product Pests. *Archives of Phytopathology and Plant Protection*, 4(3), 140-148.
- Sharafel, D. N. (1986). *Investigation into the Biology, Ecology and Control of the Cotton Aphid in the Sudan. Crop Pest Management in the Sudan; Proceeding of a Symposium Held in Khartoum.* (Unpublished Doctoral Dissertation). University of Khartoum.
- Sharaf, N. (1986). Chemical Control of *Bemisia tabaci*. *Agriculture, Ecosystems & Environment*, 1(7), 111-127.
- Sharma, M., & Sharma, R. G. (2013). Identification, Purification and Quantification of Andrographolide from *Andrographis Paniculata* (Burm. F.) Nees by HPTLC at Different Stages of Life Cycle of Crop. *Journal of Chemical and Pharmaceutical Research*, 3(1), 23-32.
- Sharma, M., Sharma, A., & Tyagi, S. (2012). Quantitative HPLC Analysis of Andrographolide in *Andrographis Paniculata* at Two Different Stages of Life Cycle of Plant. *Acta Chimica & Pharmaceutica Indica*, 2(1), 1-7.
- Sharma, S. N., Sinha, R. K., Sharma, D. K., & Jha, Z. (2009). Assessment of Intra-Specific Variability at Morphological, Molecular and Biochemical Level of *Andrographis Paniculata* (Kalmegh). *Current Science*, 96(3), 402-408.
- Sheeja, B. D., Sindhu, D., Ebanasar, J., & Jeeva, S. (2012). Larvicidal Activity of *Andrographis paniculata* (Burm.f) Nees against *Culex quinquefasciatus* Say (Insecta: Diptera-Culicidae), a Filarial Vector. *Asian Pacific Journal of Tropical Disease*, 2(2), 574-578.
- Sheeja, K., & Kuttan, G. (2007). Activation of Cytotoxic T Lymphocyte Responses and Attenuation of Tumor Growth in Vivo by *Andrographis paniculata* Extract and Andrographolide. *Immunopharmacology and Immunotoxicology*, 2(9), 81-93.

- Simmons, A. M., Harrison, H. F., & Ling, K. S. (2008). Forty-nine New Host Plant Species for *Bemisia tabaci* (Hemiptera: Aleyrodidae). *Entomological Science*, 1(1), 385-390.
- Singh, J., Tripathi, A., Gandha, R., Singh, S. C., Jain, S. P., Sharma, A., & Khanuja, S. P. S. (2007). Traditional Uses of *Andrographis Paniculata* (Kalmegh). *Journal of Medicinal and Aromatic Plant Sciences*, 2(9), 228–233.
- Sivakumar, M. & Marimuthu, T. (1986). Efficacy of Different Organic Amendments against Phytonematodes Associated with Betel Vine. *Indian Journal of Nematology*, 16(2), 278-287.
- Song, Y. X., Liu, S. P., Jin, Z., Qin, J. F., & Jiang, Z. Y. (2013). Qualitative and Quantitative Analysis of *Andrographis paniculata* by Rapid Resolution Liquid Chromatography/Time-of-Flight Mass Spectrometry. *Molecules*, 1(8), 12192-12207.
- Soto, F., Hilje, L., Mora, G. A., & Carballo, M. (2011). Phagodeterrence by *Quassia amara* (Simaroubaceae) Wood Extract Fractions on *Hypsipyla grandella* (Lepidoptera: Pyralidae) Larvae. *Revista de Biología Tropical*, 59(1), 487-499.
- Stoll, G. (2000). *Natural Crop Protection in the Tropics* (2th Eds.). Weikersheim, Germany: Margraf Verlag.
- Subramanian, R., Asmawi, M. Z., & Sadikun, A. (2012). A Bitter Plant with a Sweet Future? A Comprehensive Review Of An Oriental Medicinal Plant: *Andrographis paniculata*. *Phytochemistry Reviews*, 11(1), 39-75.
- Syed, A. R., Sivapragasam, A., Loke, W. H., & Mohamad Roff, M. N. (2000). *Whitefly infesting vegetables in Malaysia organized: the plant resource management seminar* (pp. 38-44). Sarawak, Malaysia: MAPPS, DOA Sarawak and SIAS.
- Tanwer, B. S., & Vijayvergia, R. (2010). Phytochemical Evaluation and Molluscicidal Activity of *Andrographis paniculata*. *Herba Polonica*, 5(6), 71-77.
- Thacker, J. M. R. (2002). *An introduction to arthropod pest control*. (Unpublished Doctoral Dissertation). Cambridge University Press, Cambridge.
- Tipakorn, N. (2002). *Effects of Andrographis Paniculata (Burm.F.) Nees on Performance, Mortality and Coccidiosis in Broiler Chickens*. (Unpublished Doctoral Dissertation). Georg University, Gottingen.

- Toscano, N. C., Castle, S. J., Henneberry, T. J., & Castle, N. P. (1998). Persistent Silverleaf Whitefly Exploits Desert Crop Systems. *California agriculture*, 5(2), 29-33.
- Trindade, M. S. A., Sousa, A. H., Maracaja, P. B., Junior, R. S., & Andrade, W. G. (2007). Aqueous Extracts and Oil of Neem Combined with Neonicotinoid Insecticides against *Bemisia tabaci* Biotype B in Melon. *Ciencia Rural*, 3(7), 1798-1800.
- Tripathi, A. K., Prajapati, V., Jain, D. C., & Saxena, S. (1999). Antifeedant, Oviposition-Deterrent and Growth-Inhibitory Activity of *Andrographis paniculata* against *Spilarctia obliqua*. *International Journal of Tropical Insect Science*, 1(9), 211-216.
- Trumble, J. T. (2002) Caveat Emptor: Safety Considerations for Natural Products Used in Arthropod Control. *American Entomologist-Lanham*, 48(1), 7-13.
- Tsai, T. R., Tseng, C. F., Chen, C. F., & Tsai, T. H. (2002). Identification and Determination of Geniposide Contained in Gardenia Jasminosides and in Two Preparations of Mixed Traditional Chinese Medicines. *Journal of Chromatography*, 9(6), 83-98.
- Valdiani, A., Kadir, M., Tan, S., Talei, D., Abdullah, M., & Nikzad, S. (2012). Nain-e Havandi *Andrographis paniculata* Present Yesterday, Absent Today: A Plenary Review on Underutilized Herb of Iran's Pharmaceutical Plants. *Molecular Biology Reports*, 3(9), 5409-5424.
- Varma, A., & Malathi, V. G. (2003). Emerging geminivirus problems: A Serious Threat to Crop Production. *Annals of Applied Biology*, 14(2), 145-164.
- Vijaykumar, K., Murthy, P. B. S., Kannababu, S., Syamasundar, B., & Subbaraju, G. V. (2007). Estimation of Adrographolide in *Andrographis paniculata* Herb, Extracts and Dosage forms. *International Journal of Applied Science and Engineering*, 5(1), 27-39.
- Viscarret, M. M., Botto, E. N., & Polaszek, A. (2000). Whiteflies (Hemiptera: Aleyrodidae) of Economic Importance and Their Natural Enemies (Hymenoptera: Aphelinidae, Signiphoridae) in Argentina. *Ecological Entomology*, 2(6), 5-11.
- Von Dohlen, C. D., & Moran, N. A. (1995). Molecular Phylogeny of the Homoptera a Paraphyletic Taxon. *Journal of Molecular Evolution*, 4(1), 211-223.
- Walker, G. P., & Gordh, G. (1989). The Occurrence of Apical Labial Sensilla in the Aleyrodidae and Evidence for a Contact Chemosensory Function. *Entomologia experimentalis et applicata*, 51(3), 215-224.

- Walker, G. P., Perring, T. M., & Freeman, T. P. (2010). Life History, Functional Anatomy, Feeding and Mating Behavior. In P. A. Stansly & S. E. Naranjo (Eds.), *Bemisia: Bionomics and Management of a Global Pest* (pp. 109-160). Netherlands: Springer.
- Walter, J. F. (1999). Commercial Experience with Neem Products. In F. R. Hall, & J. J. Menn (Eds.), *Biopesticides: Use and Delivery* (pp. 155-170). New Jersey: Humana Press.
- Wang, P., Crowder, D. W., & Liu, S. S. (2012). Roles of Mating Behavioral Interactions and Life History Traits in the Competition between Alien and Indigenous Whiteflies. *Bulletin of Entomological Research*, 10(2), 1-11.
- Ware, G. W. (1983). *Pesticides: Theory and application*. San Francisco: WH Freeman.
- Widiarta, I., Hermawan, W., Oya, S., Nakajima, S., & Akasuji, F. N. (1997). Antifeedant Activity of Constituents of *Andrographis paniculata* (Acanthaceae) against the Green Rice Leafhopper, *Nephotettix cincticeps* (Hemiptera: Cicadellidae). *Applied Entomology and Zoology*, 3(2), 561-566.
- Wilson, E. O. (1988). *Biodiversity*. Washington, D.C: National Academy Press.
- Wongkittipong, R., Prat, L., Damronglerd, S., & Gourdon, C. (2004). Solid-Liquid Extraction of Andrographolide from Plants, Experimental Study, Kinetic Reaction and Model. *Purification Technology*, 40(1), 147-154.
- Wraight, S. P., & Carruthers, R. L. (1999). Production, Delivery, and Use of Mycoinsecticides for Control of Insect Pests of Field Crops. In F. R. Hall & J. J. Menn (Eds.), *Methods in Biotechnology, Biopesticides: Use and Delivery* (pp. 233-269). Totowa, NJ: Humana Press.
- Xie, Y., & Zhou, X. P. (2003). Molecular Characterization of Squash Leaf Curl Yunnan Virus, a New Begomovirus and Evidence for Recombination. *Archives of Virology*, 14(8), 2047-2054.
- XueYun, Y., Boguang, Z., & QiHua, W. (2000). New Natural Insecticides: Plant Photoxins. *Journal of Nanjing Forestry*, 24(1), 77-80.
- Xu, J., Lin, K. K., & Liu, S. S. (2011). Performance on Different Host Plants of an Alien and an Indigenous *Bemisia Tabaci* from China. *Journal of Applied Entomology*, 13(5), 771-779.
- Yadav, J. S., & Singh, T. P. (2012). Phytochemical Analysis and Antifungal Activity of *Andrographis Paniculata*. *International journal of pharmaceutical research and Bio-science*, 1(4), 240-263.

- Yang, N. W., Li, A. L., Wan, F. H., Liu, W. X., & Johnson, D. (2010). Effects of Plant Essential Oils on Immature and Adult Sweetpotato Whitefly, *Bemisia tabaci* Biotype B. *Crop Protection*, 2(9), 1200-1207.
- Yatagi, M., Makihara, H., & Oba, K. (2002). Volatile Components of Japanese Cedar Cultivars as Repellents Related To Resistance to Cryptomeria Bark Borer. *Journal of Wood Science*, 48(1), 51-55.
- Yin, Q., Yang, H., Gong, Q., Wang, H., Liu, Y., Hong, Y., & Tien, P. (2001). Tomato Yellow Leaf Curl China Virus: Monopartite Genome Organization and Agroinfection of Plants. *Virus Research*, 8(1), 69-76.
- Yoopan, N., Thisoda, P., Rangkadilok, N., Sahasitiwat, S., Pholphana, N., Ruchirawat, S., & Satayavivad, J. (2007). Cardiovascular Effects of 14-deoxy-11,12-didehydroandrograph-olide and *Andrographis paniculata* Extracts. *Planta Medica*, 7(3), 503-511.
- Yu, H., Wan, F., & Guo, J. (2012). Different Thermal Tolerance and Hsp Gene Expression in Invasive and Indigenous Sibling Species of *Bemisia tabaci*. *Biological Invasions*, 1(4), 1587-1595.
- Yu, S. J. (2008). *The Classification of Insecticide: The Toxicology and Biochemistry of Insecticides*. Florida: Florida Entomologist.
- Zang, L. S., Chen, W. Q., & Liu, S. S. (2006). Comparison of Performance on Different Host Plants between the B Biotype and a Non-B Biotype of *Bemisia tabaci* from Zhejiang, China. *Entomologia Experimentalis Application*, 12(1), 221-227.
- Zang, L. S., Liu, S. S. (2007). A Comparative Study on Mating Behaviour between the B Biotype and a non-B Biotype of *Bemisia tabaci* (Hemiptera: Aleyrodidae) from Zhejiang, China. *Journal of Insect Behavior*, 2(1), 157-171.
- Zargar, M., Azizah, A. H., Roheeyati, A. M., Fatimah, A. B., Jahanshiri, F., & Pak-Dek, M. S. (2011). Bioactive Compounds and Antioxidant Activity of Different Extracts from *Vitex Negundo* Leaf. *Journal of Medicinal Plants Research*, 5(1), 2525-2532.
- Zaridah, M. Z., Idid, S. Z., Omar, A. W., & Khozirah, S. (2001). In Vitro Antifilarial Effects of Three Plant Species against Adult Worms of Subperiodic *Brugia Malayi*. *Journal of Ethnopharmacology*, 7(8), 79-84.
- Zhou, Z. (1987). Cultivation of *Andrographis paniculata*. *Chinese Health Science Journals*, 12(1), 15-8.