

UNIVERSITI PUTRA MALAYSIA

PREPARATION, CHARACTERISATION AND RELEASE OF CARVACROL ENCAPSULATED IN GELLAN HYDROGEL AND CHITOSAN NANOPARTICLES FOR ANTIBACTERIAL APPLICATION

NORAFIDA HASNU

FS 2018 15



PREPARATION, CHARACTERISATION AND RELEASE OF CARVACROL ENCAPSULATED IN GELLAN HYDROGEL AND CHITOSAN NANOPARTICLES FOR ANTIBACTERIAL APPLICATION



By

NORAFIDA BINTI HASNU

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

November 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PREPARATION, CHARACTERISATION AND RELEASE OF CARVACROL ENCAPSULATED IN GELLAN HYDROGEL AND CHITOSAN NANOPARTICLES FOR ANTIBACTERIAL APPLICATION

By

NORAFIDA BINTI HASNU

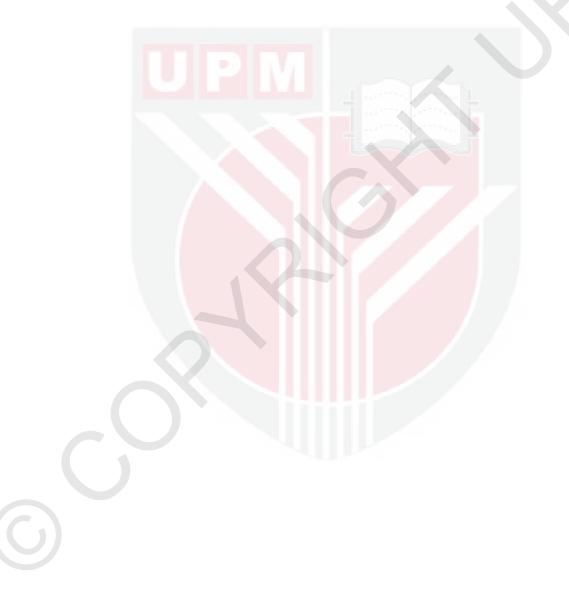
November 2017

Chair : Adila Binti Mohamad Jaafar, PhD Faculty : Science

Studies of plant materials as natural compound such as carvacrol (Carv) for antibacterial agents have gained much attention in the scientific research. It has been proven to be the potential agent in the treatment of infections and is safe for human and animal consumption. However, this free standing bioactive compound is unstable in the harsh environment conditions which easily evaporates and prone to degradation due to volatilisation and chemical reaction. In this study, the encapsulation technology helps to provide protection in order to enhance the effectiveness and the release manner thereby improving cost effectiveness of the product. Thus, Carv was encapsulated in two types of host materials which are gellan gum hydrogel thin film (GG-Carv TF) and chitosan nanoparticles (CNP-Carv). Besides that, the release properties are studied for further antibacterial application against Gram-negative bacteria, E. coli. Based on the result, GG-Carv TF showed combination of both functional groups from GG and Carv in FTIR spectra. The CHN analysis further confirmed the encapsulation as evidence of the changes in the element percentage. The swelling and degradation percentage increased with time and the decreasing patterns can be observed as the concentration of Carv increased in the range of 680.94-424.20 % and 26.83-2.67 %, respectively. Highest accumulated release of Carv from GG-Carv TF was recorded with 97.6 %. From the kinetic fitting model, pseudo-second order was observed to fit the GG-Carv TF release profile with $r^2 > 0.9$. GG-Carv TF exhibited the significant antibacterial activity against E. coli with clear inhibition zone of 20 mm while the detection of the bacterial growth by optical density also displayed the continued decrease in sustained and controlled manner. Meanwhile, the encapsulation with the other host, CNP-Carv, the results showed the increment in the size distribution average to 139 nm prior to the blank



size of CNP with only 56 nm. This result was complementary with the size of nanoparticles on surface morphology observed using FESEM. The FTIR spectrum also revealed the combination of both functional groups from the CNP and Carv, proving that Carv was successfully encapsulated. Highest accumulated release of Carv from GG-Carv TF was recorded with 28.0 %. From the kinetic fitting model, pseudo-second order was observed to fit the GG-Carv TF release profile with $r^2 > 0.9$. CNP-Carv exhibited significant antibacterial activity against *E. coli* with the notable inhibition zone of 20 mm and the detection of bacterial growth by optical density also showed a continued decline of bacterial growth. Hence, the gellan gum hydrogel and chitosan nanoparticles are proven to be effective carrier of carvacrol for further antibacterial application.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia bagi memenuhi keperluan untuk ijazah Master Sains

PENYEDIAAN, PENCIRIAN DAN PELEPASAN CARVACROL TERKANDUNG DALAM GELLAN HIDROGEL DAN KITOSAN NANOPARTIKEL UNTUK APLIKASI ANTIBAKTERIA

Oleh

NORAFIDA BINTI HASNU

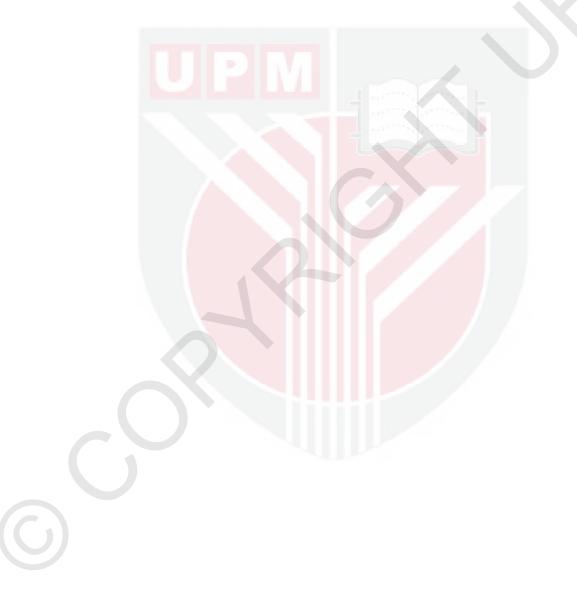
November 2017

Pengerusi : Adila Binti Mohamad Jaafar, PhD Fakulti : Sains

Penyelidikan ke atas bahan-bahan tumbuhan sebagai sebatian semula jadi seperti Carvacrol (Carv) telah mendapat banyak perhatian dalam kajian saintifik. Carv telah terbukti sebagai agen yang berpotensi untuk merawat jangkitan dan selamat untuk penggunaan manusia dan haiwan. Walau bagaimanapun, sebatian bioaktif berdiri bebas ini adalah tidak stabil dalam keadaan persekitaran yang sukar di mana ianya mudah tersejat dan mudah terdedah kepada kerosakan akibat tindak balas kimia. Dalam kajian ini, teknologi pengkapsulan membantu memberi perlindungan untuk meningkatkan keberkesanan dan tatacara pembebasan serta meningkatkan penjimatan kos produk. Oleh itu, Carv telah dikapsulkan ke dalam dua jenis hos iaitu gellan gam hidrogel (GG-Carv TF) dan kitosan nanopartikel (CNP-Carv). Selain itu, kajian turut dijalankan terhadap cara pembebasan untuk tujuan aplikasi antibakteria terhadap gram-negatif bakteria, E. coli. Berdasarkan keputusan kajian, GG-Carv TF menunjukkan gabungan kedua-dua kumpulan berfungsi daripada GG dan Carv di dalam spektra FTIR. Analisis CHN seterusnya mengesahkan pengkapsulan dengan bukti perubahan dalam peratusan unsur. Peratusan pembesaran dan degradasi juga meningkat selaras dengan masa dan corak penurunan juga dapat dilihat dengan meningkatnya kepekatan dalam anggaran 680.94-424.20 % dan 26.83-2.67 % masing-masing. Perlepasan tertinggi Carv daripada GG-Carv TF didapati sebanyak 97.6 %. Model kinetic-pseudo kedua didapati sesuai dengan profil perlepasan dengan $r^2 > 0.9$. GG-Carv TF turut mempamerkan aktiviti antibakteria yang siknifikan terhadap E. coli dengan saiz 20 mm zon perencatan yang jelas serta pengesanan pembiakan bakteria melalui cara ketumpatan optik juga menunjukkan penurunan yang berterusan dan berkala. Selain itu, pengkapsulan menggunakan hos CNP-Carv pula menunjukkan peningkatan saiz purata 139 nm selepas pengkapsulan berbanding



hanya 56 nm sebelumnya. Keputusan ini juga selaras dengan saiz yang dilihat melalui morfologi permukaan melalui FESEM. FTIR spektra turut mendedahkan gabungan kedua-dua kumpulan berfungsi daripada CNP dan Carv dan secara tidak langsung membuktikan kejayaan pengkapsulan. Perlepasan tertinggi Carv daripada CNP-Carv sebanyak 28.0 %. Daripada model kinetic-pseudo kedua didapati sesuai dengan profil perlepasan dengan $r^2 > 0.9$. CNP-Carv mempamerkan aktiviti antibakteria yang ketara terhadap *E. coli* dengan saiz 20 mm zon perencatan serta pengesanan pembiakan bakteria melalui cara ketumpatan optik juga menunjukkan penurunan. Oleh itu, gellan gam hidrogel dan kitosan nanopartikel telah terbukti untuk menjadi hos pembawa yang berkesan untuk aplikasi antibakteria selanjutnya.



ACKNOWLEDGEMENTS

Alhamdulillah, I thank God for everything. My praises to Allah for giving me the strength, confidence and courage to complete this thesis despite difficulties and problems encountered throughout this study.

I would to express my sincere appreciation to my supervisors, Dr. Adila Mohamad Jaafar, Dr Mas Jaffri Masarudin and Prof. Dr. Zulkarnain Zainal for for their patience, motivation, enthusiasm, and immense knowledge. Their guidance helped me in all the time of research and writing of this thesis.

I would like to thank the sponsorship of SGRA scheme from UPM. I am grateful that the entire laboratory officers at Chemistry Department of Faculty of Science, Faculty of Biotechnology and Biomolecular, Institute of Advanced Technology and Institute of Bioscience for assisting me throughout the study.

I thank my fellow labmates, Asma Najaj Anuar and Nurul Jannah Jamaluddin, also my junior in the research group, Nur Aisyah Nasuha Mohd Azam, for their help during conducting the lab work and friendship throughout the years of my study.

My deepest gratitude goes to my family for their infinite love and support throughout my life. Thanks to the most important person in my life; my parents, Hasnu Bin Ismail and Noor Aini Binti Muhammat for their sincere love that are not paid off. This thesis is simply impossible without them. Thank you for everything. To them I dedicate this thesis. I certify that a Thesis Examination Committee has met on 9 November 2017 to conduct the final examination of Norafida bt Hasnu on her thesis entitled "Preparation, Characterization and Release of Carvacrol Encapsulated in Gellan Hydrogel and Chitosan Nanoparticles for Antibacterial Application" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Norhazlin binti Zainuddin, PhD Senior Lecturer Faculty of Science Universiti Putra Malaysia (Chairman)

Jaafar bin Abdullah, PhD Senior Lecturer Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Azlan bin Kamari, PhD Associate Professor Universiti Pendidikan Sultan Idris Malaysia (External Examiner)

NOR AINI AB. SHUKOR, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 29 January 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Adila Mohamad Jaafar, PhD

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Chairman)

Mas Jaffri Masarudin, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Science Universiti Putra Malaysia (Member)

Zulkarnain Zainal, PhD

Professor Faculty of Science Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No.:	

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	
Name of Chairman of Supervisory Committee:	Dr. Adila Mohamad Jaafar
Signature:	
Name of Member of Supervisory Committee:	Dr. Mas Jaffri Masarudin
Signature: Name of Member of Supervisory Committee:	Prof. Dr. Zulkarnain Zainal

TABLE OF CONTENT

			Page
ABSTR	RACT		i
ABSTR	AK		iii
ACKN	OWLEDG	EMENTS	v
APPRO	DVAL		vi
DECLA	ARATION		viii
LIST O	OF TABLES	8	xii
LIST O)F FIGURI	ES	xiii
LIST O	OF ABBRE	VIATIONS	XV
CHAP	TER		
1	INT	RODUCTION	
		Nanotechnology	1
	1.2	05	2
	1.3		3
	14	Delivery Problem Statement	4
	1.5	Objectives of the Study	5
2	LIT	ERATURE REVIEW	
	2.1	Host Material	6
		2.1.1 Gellan Gum Hydrogel	7
		2.1.2 Chitosan Nanoparticles	11
	2.2	Guest Anion	13
	2.2	2.2.1 Plant-Derived Sources of Antimicrobials	13
		2.2.2 Carvacrol	14
		TEDIAL SAND METHODS	
3		TERIALS AND METHODS	10
	3.1 3.2	Materials Apparatus	19 19
	3.3	Preparation of GG-Carv TF	19
	3.4	Preparation of CNP-Carv	20
	3.5	Preparation of Pseudo Extra Cellular Fluid (PECF)	22
		Buffer Solution	
	3.6	Characterisations	22
		3.6.1 Fourier Transform-Infrared (FTIR)	22
		3.6.2 Carbon, Hydrogen and Nitrogen Analysis (CHN)	22

Х

		3.6.3 3.6.4 3.6.5	Scanning Electron Microscopy (SEM) Swelling Test Degradation Test	23 23 23
		3.6.6	Dynamic Light Scattering (DLS)	24
	3.7		e Study Controlled Balance study of Corry from	24
		3.7.1	Controlled Release study of Carv from GG-Carv TF and CNP-Carv	24
		3.7.2	Kinetic Study of GG-Carv TF and CNP-Carv	25
	3.8	Antiba	acterial Activity	25
		3.8.1	Disc Diffusion Test of GG-Carv TF and CNP-Carv	25
		3.8.2	Detection of Bacterial Growth by Optical Density (OD) of GG-Carv TF and CNP-Carv	26
	_			
4			AND DISCUSSION	• •
	4.1	Gellan 4.1.1	Gum Hydrogel	28 28
			Fourier Transform-Infrared (FTIR) Elemental Analysis	28 29
		4.1.3		30
		4.1.4		32
		4.1.5	Degradation Percentage	34
		4.1.6	Release Study of GG-Carv TF	35
		4.1.7	Antibacterial Activity of GG-Carv TF	39
	4.2	Chitos	an Nanoparticles	45
		4.2.1		45
		4.2.2	Elemental Analysis	46
			Size Distribution Analysis	47
			Morphological Analysis	47
		4.2.5		48
		4.2.6	Antibacterial Activity of CNP-Carv	53
5			ION AND RECOMMENDATIONS FOR RESEARCH	
	5.1		Conclusion	60
	5.2		Comparison Between Gellan Gum Hydrogel and Chitosan Nanoparticles in Controlled Released Study	61
	5.3		Recommendations for Future Research	62
REFERENCI	ES			63
			81	
BIODATA OF STUDENT 8			83	
LIST OF PUI	BLIC	ATION	NS	84

 \bigcirc

xi

LIST OF TABLES

Table		Page
2.1	Type of various nanocomposites and hydrogels with their antibacterial application in drug delivery system	7
2.2	Properties of Carvacrol	15
3.1	Trial formulation of volume and concentration of working solution of chitosan and TPP	21
4.1	Weight percentage of carbon, C and hydrogen, H for pure GG TF and encapsulated GG-Carv TF with various concentration of Carv	30
4.2	Rate constant and half life obtained from data fitting using kinetic models on the release of Carv from GG in PECF buffer solution	38
4.3	Disc diffusion test result of pure GG TF, GG-Carv 08, positive control (Kanamycin) and negative control (Ampicillin) against tested bacteria (<i>E. coli</i>) as indicated by the zone of inhibition	40
4.4	Weight percentage of carbon, C and hydrogen, H for blank CNP and encapsulated CNP-Carv	46
4.5	Rate constant and half life obtained from pseudo-second order fitting on the release of Carv from CNP in PECF buffer solution	52
4.6	Disc diffusion test result of blank CNP, CNP-Carv 08, positive control (Kanamycin) and negative control (Ampicillin) against tested bacteria (<i>E. coli</i>) as indicated by the zone of inhibition	54

LIST OF FIGURES

Figure		Page	
2.1	Two Forms of Gellan Gum	9	
2.2	Molecular structure of chitin and chitosan	11	
2.3	Schematic structure of polymer nanoparticles	12	
2.4	Molecular structure of Carvacrol	15	
2.5	Locations and mechanisms in the bacterial cell thought to be sites of action for carvacrol	17	
3.1	Scheme of interaction of chitosan with sodium tripolyphosphate upon ionic gelatin routes	21	
3.2	Measuring zones of inhibition	26	
4.1	FTIR spectra of Carv, pure GG TF and GG-Carv TF	29	
4.2	VPSEM surface micrograph of pure GG TF and GG-Carv TF at 1000x magnification	31	
4.3	VPSEM cross sectional of pure GG TF and GG-Carv TF at 1000 times magnification	32	
4.4	The Swelling Percentage of GG-Carv TF	33	
4.5	The Degradation Percentage of GG-Carv TF	34	
4.6	Release profile of carvacrol from gellan gum hydrogel in PECF buffer solution with pH 5.5	36	
4.7	Data fitting using kinetic models of Carv release from GG	37	
4.8	The appearance of inhibition zones after 18 hours of incubation of pure GG TF, GG-Carv 08 and Carv 08	41	

 \bigcirc

xiii

4.9	The antibacterial effects of GG-Carv TF and free standing Carv on cell viability	42
4.10	The antibacterial effects of GG-Carv TF and free standing Carv on cell growth	44
4.11	FTIR spectra of Carv, blank CNP and CNP-Carv	46
4.12	The size distribution of CNP-Carv	47
4.13	The surface morphologyanalysis of CNP and CNP-Carv at 100 000x of magnification	48
4.14	Release profile of carvacrol from chitosan nanoparticles in PECF buffer solution with pH 5.5	49
4.15	Data fitting using kinetic models of Carv release from CNP	51
4.16	Disc diffusion test of blank CNP, positive and negative control	55
4.17	Disc diffusion test of CNP-Carv and free standing Carv	56
4.18	The antibacterial effects of CNP-Carv and free standing Carv on cell viability	58
4.19	The antibacterial effects of CNP-Carv and free standing Carv on cell growth	59

C

 \bigcirc

LIST OF ABBREVIATIONS

Carvacrol
Gellan Gum Hydrogels Thin Films
Gellan Gum Hydrogels-Carvacrol Thin Films
Chitosan Nanoparticles
Chitosan Nanoparticles-Carvacrol
Pseudo Extra Cellular Fluid
Fourier Transform-Infrared
Carbon, Hydrogen and Nitrogen
Variable Pressure Scanning Electron Microscopy
Field Emission Scanning Electron Microscopy
Dynamic Light Scattering
Ultraviolet-visible Spectrophotometer
Optical Density

CHAPTER 1

INTRODUCTION

1.1 Nanotechnology

Nanotechnology is the evolutionary technology of science. It is the study and application of extremely small things and can be used across all of other science fields, such as chemistry, biology, physics, materials science, engineering and etc. It also involves the manipulation of materials at the molecular level. The development of these nano materials is a result of fundamental research done by scientists and industries which have showed improvements to the existing products. It is commonly employed in several critical fields such as drug delivery, environmental science, catalysis, engineering and many other areas of society for better and advanced application.

Richard Feynman, a Nobel Prize winner and a physicist has become well known with his famous lecture entitled 'There's Plenty of Room at the Bottom' in 1959 (Feynman, 1965). He has played an important role in catalysing nanotechnology research area by inspiring others to indulge in this promising research area. Feynman describes nanotechnology as science of building things from the bottom up with atomic precision. Feynman's ideas have influenced Drexler in writing a book entitled 'Engines Creation, The Coming Era of Nanotechnology', which he forecasted that nanotechnology will sweep the world within ten to fifty years and explained that nanotechnology will definitely give an impact to human life in politics, information science, defences, human relations and etc (Drexler, 1986).

Nanotechnology has been leading the current research trend and become very popular in the development of the latest scientific technology which encompasses many disciplines. It is a field of applied science and technology covering a broad range of topics. Currently, nanotechnology considered as highly multidisciplinary field compromising knowledge from other fields. The main aim is to control matter between 1-100 nanometres in size. It covers the study of functional system in a much deeper and extensive research at the molecular scale. Commonly nanotechnology means the ability to construct different material starting from the very bottom, using the latest techniques and tools as to invent high performance and completely distinguished products.

In human daily life, research and technological development has a primary impact in boosting the efficiency of a product. Technology has enable people to create new things and make many scientific endeavours become possible in assisting human in

1

life. Technology has such immense benefits in every phase of human lives, that it has not only made life easier but also raised the standard of living for every individual and society. The major advancement in technology is because of the increased of scientific research these days. Research in science has been the pillar of technology and therefore it is considered to be the most looked upon subject in the modern world due to its technological success.

1.2 Current Trend of Nanotechnology Research

The advantage of high surface-to-volume ratio in nano material has been manipulated by researchers worldwide to attain novel and useful application. Often in various cases, smaller nano materials are preferred, and it is generally found that specific applications require particle diameters of 30 nm or less to provide significant improvements over the use of 'conventional' nano or micron scale particles. Changes from bigger size material to nano structures involve the manipulation of structural, thermodynamic, electronic, spectroscopic, electromagnetic, dynamic and chemical features which could create potentials to various applications.

Coordination in multidisciplinary research of nanotechnology has enhanced various critical research fields such as drug delivery, tissue engineering, catalysis, filtration, efficiency of energy production and etc. Currently, the use of tailored nanoparticles has opened the avenue to optimize the capacity and transport beneficial material. Host material such as gellan gum hydrogel (GG) and chitosan nanoparticles (CNPs) have been modified to uptake the beneficial material, transport and finally release it at the targeted place. The development of nanotechnology has given the opportunities to synthesis, characterise, manipulate and organise the host material for various promising application.

Over the past decades, great attention has been focused on biopolymer-based hydrogels such as gellan gum hydrogel (GG) as the potential carriers in controlled drug delivery. Hydrogels are three-dimensional, hydrophilic, polymeric networks capable of imbibing large amount of water or biological fluids, resembling biological tissues. Because of these properties, great interest was devoted to these systems for biomedical applications. Indeed, the physicochemical properties of the hydrogels can be tuned by varying the crosslinking degree (physical and/or chemical), thus making these networks suitable devices for a modulated drug delivery.

 \bigcirc

In conjunction to the increasing demand of this delivery system tremendously, CNPs polymers also concomitantly attract the attention of the researchers. They are capable of carrying and delivering a continuous supply of biologically active molecules into a specific environment. These systems are able to reduce the amount of active agent

required for treatment by maintaining an effective concentration in the system applied over a certain period of time. Currently, most of the recently developed delivery systems consist of natural and synthetic polymers, polymer blends and composites of organic and inorganic materials that form membranes, capsules or micelles, depending on the application required. However, natural polymers such as CNPs are the most preferred.

The natural active molecule, carvacrol, which is the main constituents of oregano oil, has proven very usable in various applications, such as anti-microbial agents, antifungal, antiviral, antiparasitic, antioxidant, analgesic, anti-inflammatory and anti-cancer properties. It has a broad commercial interest because of the significant antimicrobial properties and able to inhibit or control the growth of pathogenic and/or spoiling bacteria. The previous reports have indicated that essential oils containing carvacrol (phenolic compounds) is one of the compounds showing the highest antibacterial performances (Meng *et al.*, 1998 and Dorman *et al.*, 2000). This finding directly highlighted the ability of carvacrol to inhibit bacterial growth and becomes the most promising idea for the development of natural antibacterial agent as the replacement of current used synthetic antibacterial agent.

Besides that, the existing hosts that aid to incorporate the active substance is tend to slowly degrade and lose their activity or become hazardous, by propagating a chain of oxidation reactions (Poshadri, 2010). Thus, there is a great need for the safe delivery systems. These two hosts, GG and CNPs, both are equally important for the encapsulation and delivery of natural active molecules such as carvacrol in diverse technological applications encompassing multidisciplinary areas. Their applications in human daily life bring the revival and new hopes in our technological system. Based on the foregoing, we can see that the combination of these natural polymers and natural active agents are widely used in various fields depending on the intended and specific application.

1.3 Nanotechnology Tools for Efficient Antibacterial Delivery

Malaysia is one of the countries that have high cases of bacterial outbreak of new and re-emerging infections due to the suitable temperature and condition for the growth of most bacteria. In recent years, an increasing number of bacterial outbreaks have been recorded and probably there should be more cases that were not detected or reported. There are many types of bacteria does exist, some of which cause illness through the infection and disease that has been associated with microorganisms like bacteria, fungi, viruses and parasites.



The infectious diseases are easily spread in various ways both direct and indirect contact. Most commonly, the outbreaks take place due to the ingestion of pathogenic bacteria like *Salmonella Typhi, Escherichia coli, Staphylococcus aureus, Vibrio cholera, Campylobacter jejuni,* and *Listeria monocytogenes* (Abdul-Mutalib, 2015). This has caused concerns for the wellbeing of Malaysians living locally and travelling abroad. The effort for prevention of diseases through the importance of hygiene has brought the matter to the forefront of public awareness.

Despite the discovery of new antibiotics, treatment of infections often fails to eradicate the pathogens completely. One major reason is that many antibacterial are difficult to be transported to the targeted area and the delivery time is short before the growth of bacteria can be completely inhibited due to the disadvantages of free standing antibacterial agent to the temperature, pressure, heat and environment.

Indeed, the challenge is to design the means of carrying this antibacterial agent direct to the targeted area such as human skin in order to prevent the infection and inhibit the bacterial growth. The crucial factor that should be taken into account is to provide the protection to this antibacterial agent by encapsulation technology with the intention to prolong and sustain the delivery time until the growth of the bacteria can be completely inhibited. This encapsulation technology is not merely enhances the efficiency of the release manner, but improving the cost effectiveness and the usage of the compound could also be maximised.

1.4 Problem Statement

Carvacrol is a natural compound and the main constituents of the oregano oil. It has been proven very usable and notorious for wide application including a useful source of antimicrobial compounds. Carvacrol has a significant antibacterial property to inhibit or control the growth of bacteria and has been recognised for its potential as antibacterial agent. The most compelling finding, carvacrol could be an alternative approach to control the spread of pathogenic organisms as well as the development of resistance to the conventional chemical antibiotics (Nostro *et al.*, 2012).

C

However, carvacrol is unstable in the harsh environment conditions. It is volatile, easily evaporates and prone to degradation during the processing owing to direct exposure of heat, pressure, light or oxygen (Charlier *et al.*, 2007). To elucidate this matter, carvacrol is encapsulated in gellan gum hydrogel and chitosan nanoparticles to provide stability and protection. This technology is hoped to extend the shelf life and sustain the release manner thereby the usage of the compound can be directly maximised. Therefore, in this study, the research was carried out to prepare carvacrol encapsulated in gellan gum hydrogel thin film (GG-Carv TF) and chitosan nanoparticles (CNP-Carv) and further characterised for antibacterial application.

1.5 Objectives of the Study

Carvacrol has been identified by previous researcher to have significant antibacterial properties and has been recognised for its potential as antibacterial agent. However, due to the disadvantages of this free standing carvacrol to the environment condition, the usage of this active compound is not fully maximised as it easily evaporated and degraded when directly exposed to the heat, oxygen, pressure or light (Charlier *et al.*, 2007). Thus, this study is attempt to bridge the gap between the issues of disadvantages of carvacrol and providing the protection from the host of gellan gum hydrogel and chitosan nanoparticles to ensure the function of carvacrol can be directly maximised.

Due to this concern, the major aim of this study is to encapsulate carvacrol in gellan gum hydrogel thin film and chitosan nanoparticles in order to enhance the effectiveness and release manner of carvacrol. The antibacterial properties were also studied for further application. Therefore, the specific objectives for this study are as follows :

- 1. To prepare and characterise the host of gellan gum hydrogel thin film (GG TF) and chitosan nanoparticles (CNP).
- 2. To prepare and characterise gellan gum hydrogel-Carv thin film (GG-Carv TF) and chitosan nanoparticles-Carv (CNP-Carv).
- 3. To study the release manner of both GG-Carv TF and CNP-Carv.
- 4. To study the antibacterial application of GG-Carv TF and CNP-Carv.

REFERENCES

- Abdul-Mutalib, N.A., Syafinaz, A.N., Sakai, K. and Shirai, Y. 2015. An overview of foodborne illness food safety in Malaysia. *International Food Research Journal*. 22(3): 896-901.
- Achinewhu, S.C., Ogbonna, C.C. and Hart, A.D. 1995. Chemical composition of indigenous wild herbs, spices, nuts and leafy vegetables used as food. *Plant Foods Hum. Nutr.* 32: 31-36.
- Acosta, E. 2009. Bioavailability of nanoparticles in nutrient and nutraceutical delivery. *Curr. Opin. Colloid Interface Sci.* 14: 3–15.
- Agnihotri, S.A., Nadagouda, N., Mallikarjuna, T., Aminabhavi, M. 2004. Recent Advances on Chitosan-Based Micro- and Nanoparticles in Drug Delivery. J. *Control Release*.100:5-28.
- Akgül, A. and Kivanç, M. 1988. Inhibitory effects of selected Turkish spices and oregano components on some foodborne fungi. *International Journal of Food Microbiology*. 6: 263-268.
- Akgül, A., Kivanç, M. and Sert. S. 1991. Effect of carvacrol on growth and toxin production by Aspergillus flavus and Aspergillus parasiticus. Sciences des Aliments 11:361-370.
- Aligiannis, N., Kalpoutzakis, E., Mitaki, S., Chinou, I.B. 2001. Composition and antimicrobial activity of the essential oil of two *Origanum* species. *J Agric Food Chem.* 49: 4168-4170.
- Anirudhan, T.S., Nima, J. and Divya, P.L. 2015. Synthesis, characterization and in vitro cytotoxicity analysis of a novel cellulose based drug carrier for the controlled delivery of 5-Fluorouracil, an anticancer drug. *Applied Surface Science*. 355: 64-73.
- Arfa, B.A., Combes, S., Preziosi-Belloy, L., Gontard, N., and Chalier, P. 2006. Antimicrobial activity of carvacrol related to its chemical structure. *Lett. in Appl. Microbiol.* 43: 149-154.

- Armum, P.V. 2000. Drug delivery market poised for five years of strong growth. *Chemical Market Reporter*. 258: 16-23.
- Arrebola, M.L., Navarro, M.C., Jimenez, J. and Ocana, F.A. 1994. Yield and composition of the essential oil of *Thymus serpylloides subsp. serpylloides*. *Phytochemistry*. 36:67-72.
- Arregui, L.M., Veramendi, J., Mingo-Castel, A. M. 2003. Effect of gelling agents on in vitro tuberization of six potato cultivars. *Am J Potato Res.* 20:1–4.
- Atta, S., Khaliq, S., Islam, A., Javeria, I., Jamil, T. 2015. Injectable biopolymer based hydrogels for drug delivery applications. *International Journal of Biological Macromolecules*. 80: 240-245.
- Au Natural Herbals. Chitosan history. Retrieved 22 November 2001 from http://www.chitosan-weightloss.net/history.html
- Aydin, S., Basaran, A.A. and Basaran, N. 2005. The effects of thyme volatiles on the induction of DNA damage by the heterocyclic amine IQ and mitomycin C. *Mutat Res.* 581:43-53.
- Azzouz, M. A., and Bullerman. L. B. 1982. Comparative antimycotic effects of selected herbs, spices, plant components and commercial antifungal agents. *Journal of Food Protection*. 45: 1298-1301.
- Bagamboula, C.F., Uyttendaele, M. and Debevere. 2004. Inhibitory effect of thyme and basil essential oils, carvacrol, thymol, estragol, linalool and p-cymene towards *Shigella sonnei* and *S. flexneri. Food Microbiol*. 21: 33-42.
- Bishop, C. D. 1995. Antiviral activity of the essential oil of *Melaleuca alternifolia* (Maiden and Betche) Cheel (tea tree) against tobacco mosaic virus. *Journal* of Essential Oil Research. 7: 641-644.
- Boyle, W. 1955. Spices and essential oils as preservatives. *The American Perfumer and Essential Oil Review*. 66: 25-28.
- Burt, S. 2004. Essential oils: their antibacterial properties and potential applications in foods –a review. *Int. J. Food Microbiol.* 94: 223-253.

- Burt, S. 2004. Essential oils: their antibacterial properties and potential applications in foods-a review. *Int. J. Food Microbiol.* 94: 223-253.
- Carson, C. F., Cookson, B. D. Farrelly. H. D. and Riley, T. V. 1995. Susceptibility of methicillin-resistant *Staphylococcus aureus* to the essential oil of *Melaleuca alternifolia*. Journal of Antimicrobial Chemotherapy. 35: 421-424.
- Chalier, P., Arfa, A.B., Belloy L.P. Gontard, N. 2007 Carvacrol losses from soy protein coated papers as a function of drying conditions. *Journal of Applied Polymer Science*. 106: 611-620.
- Chami, F., Chami, N., Bennis, S., Bouchikhi, T. and Remmal, A. 2005. Oregano and clove essential oils induce surface alteration of *Saccharomyces cerevisiae*. *Phytother. Res.* 19:405-408.
- Chen, L. and Subirade, M. 2005. Chitosan/beta-lactoglobulin core-shell nanoparticles as nutraceutical carriers. *Biomaterials*. 26: 6041–6053.
- Chen, L., Remondetto, G. E. and Subirade, M. 2006. Food protein-stabilized nanoemulsions as potential delivery systems for poorly water-soluble drugs: preparation, in vitro characterization and pharmacokinetics in rats. *Trends Food Sci. Technol.* 17: 272–283.
- Colombo, P. and Sciutto, A.M. 1996. Nutritional aspects of chitosan employment in hypocaloric diet. *Acta Toxicol Ther*. 17: 278–302.
- Costa, P. and Lobo, J.M.S. 2001. Modelling and comparison of dissolution profiles. *European Journal of Pharmaceutical Sciences*. 13(2):123-133.
- Cowan, M.M. 1999. Plant products as antimicrobial agents. *Clin. Microbiol. Rev.* 12: 564 582.
- De Vincenzi, M., Stammati, A., De Vincenzi, A. and Silano, M. 2004. Constituents of aromatic plants: carvacrol. *Fitoterapia*. 75:801–804.
- Deans, S. G., and G. Ritchie. 1987. Antibacterial properties of plant essential oils. International Journal of Food Microbiology. 5:165-180.

- Demitri, C., De Benedicts, V. M., Madaghiele, M., Corcione, C.E. and Mafezzoli, A. 2016. Nanostructured active chitosan-based films for food packaging applications: Effect of graphene on mechanical properties. *Measurements*. 90: 418-423.
- Di Pasqua, R., Betts, G., Hoskins, N., Edwards, M., Ercolini, D., and Mauriello, G. 2007. Membrane toxicity of antimicrobial compounds from essential oils. *J. Agric. Food Chem.* 55:4863-4870.
- Dong, W. and Bodmeier, R. 2006. Encapsulation of lipophilic drugs within enteric microparticles by a novel coacervation method. *Int. J. Pharm.* 326: 128–138.
- Dorman, H.J.D. and Deans, S.G. 2000. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *Journal of Applied Microbiology*. 88:308–316A.
- Dorman, H.J.D. and Deans, S.G. 2000. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. J. Appl. Microbiol. 88: 303-316.
- Drexler, K.E. 1986. Engines of Creation, *The Coming Era of Nanotechnology*. pp. 3-291. New York. Anchor Books.
- Duclairoir, C, Orecchioni, A.M., Depraetere, P., Osterstock, F. and Nakache, E. 2003. Evaluation of gliadins nanoparticles as drug delivery systems: a study of three different drugs. *Int. J. Pharm.* 253: 133–144.
- Duffy, C.V. David, L. and Crouzier, T. 2015. Covalently-crosslinked mucin biopolymer hydrogels for sustained drug delivery. *Acta Biomater*. 20: 51-59.
- Feynman, R.P. 1965. The Feynman lectures on Physics. Amer J. Physic. 33(9):750-752.
- Ferris, C.J., Gilmore, K.J., and Wallace, G.G. 2013. Modified gellan gum hydrogels for tissue engineering applications. *Soft Matter*. 9(14):3705-3711.
- Force, M., Sparks, W.S. and Ronzio, R.A. 2000. Inhibition of enteric parasites by emulsified oil of oregano *in vivo*. *Phytother Res.* 14: 213-4.

- Friedman, M., Henika, P.R. and Mandrell, R.E. 2002. Bactericidal activities of plant essential oils and some of their isolated constituents against Campylobacter jejuni, Escherichia coli, Listeria monocytogenes, and Salmonella enterica. J. Food Prot. 52:6042-6048.
- Friedman, M., Henika, P.R., and Mandrell, R.E. 2002. Bactericidal activities of plant essential oils and some of their isolated constituents against Campylobacter jejuni, Escherichia coli, Listeria monocytogenes, and Salmonella enterica. J. Food Prot. 65:1545-1560.
- Friedman, M., Henika, P.R., Levin, C. E., and Mandrell, R.E. 2004. Antibacterial activities of plant essential oils and their components against *Escherichia coli* O157:H7 and *Salmonella enterica* in apple juice. *J. Agric. Food Chem.* 52: 6042-6048.
- García, G.C.A., Argemí, A., Sousa, A.R.S.D., Duarte, C.M.M. and Saurina, J. 2010. Encapsulation efficiency of solid lipid hybrid particles prepared using the PGSS[®] technique and loaded with different polarity active agents. *Journal of Supercritical Fluids*. 54: 342-347.
- García, G.C.A, Alnaief, M. and Smirnova, I. 2011. Polysaccharide-based aerogels-Promising biodegradable carriers for drug delivery systems. *Carbohydrate Polymers*. 86: 1425-1438.
- Ghayempour, S., Montazer, M., Mahmoudi, Rad, M. 2016. Tragacanth gum biopolymer as reducing and stabilizing agent in biosonosynthesis of urchinlike ZnO nanorod arrays: A low cytotoxic photocatalyst with antibacterial and antifungal properties. *Carbohydr Polym.* 136: 232-241.
- Gaysinsky, S., Davidson, P.M., Bruce, B.D., and Weiss, J. 2005. Growth inhibition of *Escherichia coli* O157:H7 and *Listeria monocytogenes* by carvacrol and eugenol encapsulated in surfactant micelles. *J. Food Prot.* 68: 2259-2566.
- Gill, A.O. and Holley, R.A. 2006. Disruption of *Escherichia coli*, *Listeria monocytogenes* and *Lactobacillus sakei* cellular membranes by plant oil aromatics. *Int. J. Food Microbiol.* 108: 1-9.
- Guterres, S.S., Alves, M.P. and Pohlmann, A.R. 2007. Polymeric nanoparticles, nanospheres and nanocapsules, for cutaneous applications. *Drug Target Insights*. 2: 147–157.

- Hajhashemi, V., Ghannadi, A. and Pezeshkian, S.K. 2002. Antinociceptive and antiinflammatory effects of *Satureja hortensis L.* extracts and essential oil. *J Ethnopharmacol.* 82: 83-7.
- Harris, J.E. 1985. Gelrite as an agar substitute for cultivation of mesophilic Methanobacterium and Methanobrevibacter species. *Appl Environ Microbiol*. 50:1107 1109.
- Helander, I.M., Alakomi, H., Latva-Kala, K., Mattila-Sandholm, T., Pol, I., Smid, E.J., Gorris, L.G.M. and von Wright, A. 1998. Characterization of the action of selected essential oil components on Gram-negative bacteria. J. Agr. Food Chem. 46:3590–3595.
- Ichi, T., Koda, T., Asai, I. and Hatanaka, A. 1986. Effects of gelling agents on *in vitro* culture of plant tissues. *Agric Biol Chem.* 50 (9): 2397–2399.
- Illum, L. 1998. Chitosan and its use as a pharmaceutical excipient. *Pharm. Res.*15: 1326-1331.
- Jagur, G.J. 2010. Polymeric gels and hydrogels for biomedical and pharmaceutical application. *Polymers for Advanced Technologies*. 21: 27-47.
- Jang, K. I. and Lee, H.G. 2008. Stability of chitosan nanoparticles for L-ascorbic acid during heat treatment in aqueous solution. J. Agric. Food Chem. 56: 1936–1941.
- Jansson, P.E., Lindberg, B. and Sandford, P.A. 1983. Structural studies of gellan gum, an extracellular polysaccharide elaborated by *Pseudomonas elodea*. *Carbohydrate Res.* 124-135.
- Jayashree, T. and Subramanyam, C. 1999. Antiaflatoxigenic activity of eugenol is due to inhibition of lipid peroxidation. *Letters in Applied Microbiology*. 28: 179-183.
- Jia, Z., Shen, D. and Xu, W. 2001. Synthesis and antibacterial activities of quaternary ammonium salt of chitosan. *Carbohydr. Res.* 333: 1–6.

- Jing, S., Li, L., Ji, D., Takiguchi, Y. and Yamaguchi, T. 1997. Effect of chitosan on renal function in patients with chronic renal failure. *J Pharm Pharmacol*. 49: 721–3.
- Juglal, S., Govinden, R. and Odhav, B. 2002. Spice oils for the control of cooccurring mycotoxin-producing fungi. *Journal of Food Protection*. 65: 683-687.
- Juneja, V.K. and Friedman, M. 2007. Carvacrol, Cinnamaldehyde, Oregano oil, and thymol inhibit *Clostridium perfringens* spore germination and outgrowth in ground turkey during chilling. *J. Food Prot.* 70:218-222.
- Juneja, V.K. and Friedman, M. 2008. Carvacrol and Cinnamaldehyde facilitate thermal destruction of *Escherichia coli* O157:H7 in raw ground beef. *J. Food Prot.* 71:1604-1611.
- Juven, B. J., Kanner, J., Schved, F. and Weisslowicz, H. 1994. Factors that interact with the antibacterial action of thyme essential oil and its active constituents. *Journal of Applied Bacteriology*. 76:626-631.
- Kang, D., Zhang, H. B., Nitta, Y., Fang, Y. P. and Nishinari, K. 2015. Gellan. Polysaccharides: Bioactivity and Biotechnology. 1627-1682.
- Karatzas, A.K., Kets, E.P.W., Smid, E.J. and Bennik, M.H.J. 2001. The combined action of carvacrol and high hydrostatic pressure on *Listeria monocytogenes* Scott A. J. Appl Microbiol. 90: 463-469.
- Karkabounas, S., Kostoula, O.K., Daskalou, T., Veltsistas, P., Karamousis, M. and Zelovitis, I. 2006. Anticarcinogenic and antiplatelet effects of carvacrol. *Exp Oncol.* 28: 121-5.
- Karpouhtsis, I., Pardali, E. Feggou, E. Kokkini, S. Scouras, Z. G. and Mavragani-Tsipidou, P. 1998. Insecticidal and genotoxic activities of oregano essential oils. *Journal of Agricultural and Food Chemistry*. 46: 1111-1115.
- Karpouhtsis, I., Pardali, E., Feggou, E., Kokkini, S., Scouras, Z.G., Mavragani-Tsipidou, P. 1998. Insecticidal and genotoxic activities of oregano essential oils. J Agric Food Chem. 46: 1111-1115.

- Kaushik, V and Roos, Y.H. 2007. Limonene encapsulation in freeze-drying of gum Arabic–sucrose–gelatin systems. *LWT Food Sci. Technol.* 40: 1381–1391.
- Kendra,, D. F. and Hadwiger, L. A. 1984. Characterization of the smallest chitosan oligomer that is maximally antifungal to *Fusarium solani* and elicits pisatin formation by *Pisum sativum*. *Exp. Mycol.* 8: 276–281.
- Keong, L.C. Halim, A.S. 2009. *In vitro* models in biocompatibility assessment for biomedical-grade chitosan derivatives in wound management. *Int. J. Mol. Sci.* 10: 1300-1313.
- Kim, D.G., Jeong, Y. I., Choi, C., Roh, S.H., Kang, S.K., Jang, M.K. and Naha, J.W. 2006. Retinol-encapsulated low molecular water-soluble chitosan nanoparticles. *Int. J. Pharm.* 319: 130–138.
- Kisko, G. and Roller, S. 2005. Carvacrol and p-cymene inactivate *Escherichia coli* O157:H7 in apple juice. *BioMed Central Microbiol*. 1-9.
- Knobloch, K., Pauli, A. Iberl, B. Weigand, H. and Weis, N. 1989. Antibacterial and antifungal properties of essential oil components. *Journal of Essential Oil Research.* 1:119-128.
- Knowles, J.R., Roller, S., Murray, D.B., and Naidu, S. 2005. Antimicrobial action of carvacrol at different stages of dual-species biofilm development by *Staphylococcus aureus* and *Salmonella enterica* serovar Typhimurium. *Appl Environ. Microb.* 71:797-803.
- Koda, T., Ichi, T., Yamagishi, H. and Yoshikawa, H. 1988. Effects of phytohormones and gelling agents on plant regeneration from protoplasts of red cabbage. *Agric Biol Chem.* 52(9): 2337–2340.
- Kodama, T., Harada, Y., Ueda, M., Shimizu, K.I., Shuto, K. and Komarneni, S. 2001. Selective exchange and fixation of strontium ions with ultrafine Na-4mica. Langmuir. 17(16):4881-4886.
- Konstantopoulou, I., Vassilopoulou, L. Mavragani-Tsipidou, P. and Scouras. Z. G. 1992. Insecticidal effects of essential oils. A study of the effects of essential oils extracted from eleven Greek aromatic plants on *Drosophila auraria*. *Experientia*. 48: 616-619.

- Kumeria, T., Mon, H., Aw, M.S., Gulati, K. and Santos, A. 2015. Advanced biopolymer-coated drug-releasing titania nanotubes (TNTs) implants with simultaneously enhanced osteoblast adhesion and antibacterial properties. *Colloids and Surfaces B: Biointerfaces*. 130: 255-263.
- Kwon, J.A., Yu, C.B. and Park, H.D. 2003. Bactericidal effects and inhibition of cell separation of cinnamic aldehyde on *Bacillus cereus. Lett. Appl. Microbiol.* 37: 61-65.
- Lacroix, M., Borsa, J., Chiasson, F. and Ouattara, B. 2004. The influence of atmosphere conditions on *Escherichia coli* and *Salmonella typhi* radiosensitization in irradiated ground beef containing carvacroland tetrasodioum pyrophosphate. *Radiat. Phys. Chem.* 71:59-62.
- Lagouri, V.G., Blekas, M., Tsimidou, S., Kokkini, S., and Boskou, D. 1993. Composition and antioxidant activity of essential oils from oregano plants grown wild in Greece. Z. Lebensm Unters. For-schung. 197: 20-23.
- Lalita, K. and Rangrong, Y. 2011. Preparation, characterization and *in vitro* release study of carvacrol-loaded chitosan nanoparticles. J. Colloids Surf B Biointerfaces. 84:163-171.
- Lambert, R.J.W. Skandamis, P.N. Coote, P.J. and Nychas, G.J.E. 2001. A study of the minimum inhibitory concentration and node of action of oregano essential oil, Tymol and Carvacrol. J. Appl. Microbiol. 91: 453- 462.
- Lambert. R.J.W., Skandamis, P.N., Coote, P.J., and Nychas, G.-J.E. 2001. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol. *J. Appl. Microbiol.* 91: 453-462.
- Lee, C.M., Lim, S., Kim, G.Y. Kim, D.W., Rhee, J.H. and Lee, K.Y. 2005. Rosin nanoparticles as a drug delivery carrier for the controlled release of hydrocortisone. *Biotechnol. Lett.* 27: 1487–1490.
- LeHoux, J. G., and F. Grondin. 1993. Some effects of chitosan on liver function in the rat. *Endocrinology*. 132: 1078-1084.

- Liechty, W.B., Caldorera, M.M, Phillips, M.A., Schoener, C., Peppas, N.A. 2011. Advanced molecular design of biopolymers for transmucosal and intracellular delivery of chemotherapeutic agents and biological therapeutics. *Journal of Controlled Release*. 155: 119-127.
- Lin, C.C. and Cassida, L.E. 1984. Gelrite as a gelling agent for the growth of thermophilic microorganisms. *Appl Environ Microbiol.* 47:427–429.
- Ling, L., He, J., Wei, M., Evans, D.G. and Duan, X. 2006. Uptake of chloride ion from aqueous solution by calcined layered double hydroxides: Equilibrium and kinetics studies. *Water Research*. 40: 735-743.
- Lu, Y. and Wu, C. 2010. Reduction of *Salmonella enterica* contamination on grape tomatoes by washing with thyme oil, thymol, and carvacrol as compared with chlorine treatment. *J. Food Prot.* 73:2270–2275.
- Mahmoud, S. S., and Croteau, R. B. 2002. Strategies for transgenic manipulation of monoterpene biosynthesis in plants. *Trends in Plant Science*. 7: 366-373.
- Majeti, N.V. and Kumar, R. 2000. A review of chitin and chitosan applications. *React. Funct. Polym.* 46: 1–27.
- Manjunath, K. and Venkateswarlu, V. 2005. Pharmacokinetics, tissue distribution and bioavailability of clozapine solid lipid nanoparticles after intravenous and intraduodenal administration. J. Control. Release. 107: 215–228.
- Mari, M., Bertolini, P. and Pratella. G. C. 2003. Non-conventional methods for the control of post-harvest pear diseases. *Journal of Applied Microbiology*. 94: 761-766.
- Meng, J.H., Zhao, S.H., Doyle, M.P., Joseph, S.W. 1998. Antibiotic resistance of *E. coli* O157: H7 and O157: NM isolated from animals, food and humans. *J Food Prot.* 61:1511–1514.
- Morris, E.R., Nishinari, K. and Rinaudo. M. 2012. Gelation of gellan–a review. *Food Hydrocoll*. 28:373–411.
- Mourey, A., and N. Canillac. 2002. Anti-*Listeria monocytogenes* activity of essential oils components of conifers. *Food Control*. 13:289-292.

- Muzzarelli, R.A.A. 1999. Clinical and biochemical evaluation of chitosan for hypercholesterolemia and overweight control. *EXS*. 87:293–304.
- Nazer, A.I., Kobilinsky, A., Tholozan, J.-L., and Dubois-Brissonnet, F. 2005. Combinations of food antimicrobials at low levels to inhibit the growth of *Salmonella sv.* Typhimurium: a synergistic effect. *Food Microbiol.* 22: 391-398.
- No, H.K., Park, N.Y., Lee, S.H. and Meyers, S.P. 2002. Antibacterial activity of chitosans and chitosan oligomers with different molecular weights. *Int. J. Food Microbiol.* 74: 65-72.
- Nostro, A., Papalia, T. 2012. Antimicrobial activity of carvacrol: Current progress and future prospectives. *Recent Pat Antiinfect Drug Discov.* 7:28-35.
- Nostro, A., Roccaro, A.S., Bisignano, G., Marino, A. and Blanco, A.R. 2007. Effect of oregano, Carvacrol and Thymol on *S. aureus* and *S. epidermdis* biofilms. *J. Med. Microbiol.* 56: 591-23.
- Nychas, G. J. E. 1995. Natural antimicrobials from plants, In *New methods of food preservation*, 1st ed, ed. G. W. Gould, p. 58-89. London: Blackie Academic & Professional.
- O'Neill, M.A., Selvendran, R.R. and Morris, V.J. 1983. Structure of the acidic extracellular gelling polysaccharide produced by *Pseudomonas elodea*. *Carbohydrate Res.*, 124-123.
- Obaidat, M.M. and Frank, J.F. 2009. Inactivation of *Escherichia coli* O157:H7 on the intact and damaged portions of lettuce and spinach leaves by using allyl isothiocyanate carvacrol, and cinnamaldehyde in vapor phase. *J. Food Prot.* 72: 2046-2055.
- Olasupo, N.A., Fitzgerald, D.J., Narbad, A. and Gasson, M.J. 2004. Inhibition of *Bacillus subtilis* and *Listeria innocua* by nisin in combination with some naturally occurring organic compounds. *J. Food Prot.* 67: 596-600.
- Oosterhaven, K., Poolman, B. and Smid, E. J. 1995. S-carvone as a natural potato sprout inhibiting, fungistatic and bacteristatic compound. *Industrial Crops and Products*. 4: 23-31.

- Oussalah, M., Caillet,S., Saucier, L., and Lacroix, M. 2007. Inhibitory effects of selected plant essential oils on the growth of four pathogenic bacteria: *E. coli* O157:H7, Salmonella Typhimurium, Staphylococcus aureus and Listeria monocytogenes. Food Control. 18:414-420.
- Ousslah, M., Caillet,S. and Lacroix, M. 2006. Mechanism of action of Spanish oregano, Chinese cinnamon, and savory essential oils against cell membranes and walls of *Escherichia coli* O157:H7 and *Listeria monocytogenes*. *J.Food Prot.* 69: 1046-1055.
- Pandey, R., Kalra, A. Tandon, S. Mehrotra, N. Singh, H. N. and Kumar, S. 2000. Essential oil compounds as potent source of nematicidal compounds. *Journal* of Phytopathology. 148: 501-502.
- Park, B. S, Choi, W. S., Kim, J.H., Kim, K.H. and Lee, S.E. 2005. Monoterpenes from thyme (*Thymus vulgaris*) as potential mosquito repellents. J Am Mosquito Control Assoc. 21: 80-3.
- Park, M.R., Chun, C., Ahn, S.W., Ki, M.H., Cho, C.S., Song, S.C. 2010. Sustained delivery of human growth hormone using a polyelectrolyte complex-loaded thermosensitive polyphosphazene hydrogel. *Journal of Controlled Release*. 147(3): 359-367.
- Patel, V.R. and Amiji, M.M. 1996. Preparation and characterization of freeze-dried chitosan-poly (ethylene oxide) hydrogels for site-specific antibiotic delivery in the stomach. *Pharmaceutical research*, 13(4): 588-593.
- Pavia D.L., Marianeccia, C., Carafaa, M., Marziob, L.D., Rinaldia, F., Meoa, C.D., Alhaiquea, F., Matricardia, P. 2009. Introduction to spectroscopy (Fourth Edition), Belmont USA, Cengage Learning, 2:21-25.
- Pei, R.S., Zhou, F. and Xu, J. 2009. Evaluation of combined antibacterial effects of eugenol, cinnamaldehyde, thymol, and carvacrol against *E. coli* with an improved method. *J. Food Sci.* 74: M379-M383.
- Peng. C., Zhao, Q. and Gao, C. 2010. Sustained delivery of doxorubicin by porous CaCO₃ and chitosan/alginate multilayer-coated CaCO₃ microparticles. *Colloids and Surfaces A: Physicochemical and Engineering Aspects.* 353: 132-139.

- Peniston, Q. P. and Johnson, E. 1980. Process for the manufacturer of chitosan. US Patent 4,195,175.
- Peppas, N.A., Bures, P., Leobandung, W. andIchikawa, H. 2000. Hydrogels in pharmaceutical formulations. *Eur J Pharm Biopharm*. 50: 27-46.
- Perez-Conessa, D., Cao, J., Chen, L., Mclandsborough, L. and Weiss, J. 2011. Inactivation of *Listeria monocytogenes* and *Escherichia coli* O157:H7 biofilms by micelle encapsulated eugenol and carvacrol. J. Food Prot. 74: 55-62.
- Periago, P.M., Delgado, B., Fernandez, P.S. and Palop, A. 2004. Use of carvacrol and cymene to control growth and viability of *Listeria monocytogenes* cells and predictions of survivors using frequency distribution functions. *J. Food Prot.* 67: 1405-1416.
- Pessoa, L. M., Morais, S. M. Bevilaqua, C. M. L. and Luciano, J. H. S. 2002. Anthelmintic activity of essential oil of *Ocimum gratissimum* Linn.and eugenol against *Haemonchus contortus*. *Veterinary Parasitology*. 109: 59-63.
- Pinto, R.C., Neufeld, R.J., Ribeiro, A.J. and Veiga, F. 2006. Methods for preparation of drug-loaded polymeric nanoparticles. *Nanomed. Nanotechnol. Biol. Med.* 2(1): 8–21.
- Pol, I.E. and Smid, E.J. 1999. Combined action of nisin and carvacrol on *Bacillus* cereus and *Listeria monocytogenes*. Lett Appl. Microbiol. 29: 166-170.
- Poshadri, A., and Aparna, K. 2010. Microencapsulation Technology: A Review. J.Res ANGRAU. 38(1): 86-102.
- Poulose A.J. and Croteau, R. 1978. Biosynthesis of aromatic monoterpenes: conversion of gterpinene to p-cymene and thymol in *Thymus vulgaris L. Arch. Biochem. Biophys.* 187: 307-314.
- Prieto, J.M., Jacopini, P., Cioni, P. and Chericoni, S. 2007. *In vitro* activity of the essential oils of *Origanum vulgare*, *Satureja montana* and their main constituents in peroxynitrite induced oxidative processes. *Food Chem.* 104: 889-95.

- Ratajska, M., Strobin, G., Wrona, M.W., Ciechanska, D., Struszczyk, H. and Boryniec, S. Binias, D. and Binias, W. 2003. Studies on biodegradation of microcrystalline chitosan in aqueous medium. *Fibres Text. East Eur.* 11: 75– 79.
- Ravishankar, S., Zhu, L., Reyna-Granados, J., Law, B., Joens, L. and Friedman, M. 2010. Carvacrol and cinnamaldehyde inactivate antibiotic-resistant *Salmonella enterica* in buffer and on celery and oysters. J. Food Prot. 73: 234-240.
- Ravizza, R., Gariboldi, M.B., Molteni, R. And Monti, E. 2008. Linalool, a plantderived monoterpene alcohol, reverses doxorubicin resistance in human breast adenocarcinoma cells. *Oncol. Rep.* 20 (3): 625–630.
- Rothenfluh, D.A., Bermudez, H., O'Neil, C.P. and Hubbell. J.A. 2008. Biofunctional polymer nanoparticles for intra-articular targeting and retention in cartilage. *Nat. Mater.* 7(3): 248–254.
- Saddi, M., Sanna, A., Cottiglia, F., Chisu, L., Casu, L., Bonsignore, L. and De Logu,
 A. 2007. Antiherpevirus activity of Artemisia arborescens essential oil and inhibition of lateral diffusion in Vero cells. *Ann. Clin. Microbiol.* Antimicrob. 6 (1): 10.
- Sanderson, G.R. 1990. Gellan gum. In *Food gels*, ed. Harris P., pp. 201–232. London: El Sevier.
- Schatz, C. Bionaz, A. and Lucas, J. M. 2005. Formation of polyelectrolyte complex particles from self complexation of N-sulfated chitosan. *Biomacromolecules*. 6: 1642-1647.

Shelef, L. A. 1983. Antimicrobial effects of spices. Journal of Food Safety. 6: 29-44.

Shelef, L.A. 1983. Antimicrobial effects of spices. J Food Safety. 6: 29-44.

Shimomura, K. and Kamada, H. 1986. Roles of gelling agents in plant tissue culture. *Plant Tissue Cult.* 3:38–41.

- Sikkema, J. J., De Bont, A. M. and Poolman, B. 1994. Interactions of cyclic hydrocarbons with biological membranes. *Journal of Biological Chemistry*. 269: 8022-8028.
- Sikkema, J., de Bont, J.A.M. and Poolman, B. 1995. Mechanisms of membrane toxicity of hydrocarbons. *Microbiol. Rev.* 59: 201-222.
- Sivropoulou, A., Papanikolaou, E., Nikolaou, C., Kokkini, S., Lanaras, T. and Arsenakis, M. 1996. Antimicrobial and cytotoxic activities of origanum essential oils. *Journal of Agric Food Chem.* 44: 1201-1205.
- Solomakos, N., Govaris, A., Koidis, P., and Botsoglou, N. 2008. The antimicrobial effect of thyme essential oil, nisin, and their combination against *Listeria monocytogenes* in minced beef during refrigerated storage. *Food Microbiol.* 25: 120-127.
- M.C.M. 2009. Evaluation of cross-linked chitosan microparticles containing acyclovir obtained by spray-drying. *Mater. Sci. Eng.* 29: 387–392.
- Sripriya, J., Anandhakumar, S., Achiraman, S., Antony, J.J. and Siva, D. 2013. Laser receptive polyelectrolyte thin films doped with biosynthesized silver nanoparticles for antibacterial coatings and drug delivery applications. *International Journal of Pharmaceutics.* 457: 206-213.
- Sudarshan, N. R., Hoover, D. G. and Knorr, D. 1992. Antimicrobial actions of degraded and native chitosan against spoilage organisms in laboratory media and foods. *Food Biotechnol.* 6: 257–272.
- Svoboda, K.P. and Hampson J.B. 1999. Bioactivity of essential oils of selected temperate aromatic plants: Antibacterial, antioxidant, anti-inflammatory and other related pharmacological activities. *Plant Biology Department, SAC Auchincruive, Ayr, Scotland, UK*: 17.
- Tsai, G. J and Su, W.H. 1999. Antibacterial activity of shrimp chitosan against *Escherichia coli. J. Food. Protect.* 62: 239–243.
- Tuley de Silva, K. (ed.). 1996. A manual on the essential oil industry. United Nations Industrial Development Organization, Vienna.

- Turner, S.R. and Singha, S. 1990. Vitrification of crabapple, pear, and geum on gellan gum-solidified culture medium. *Hortscience*. 25(12): 1648–1650.
- Ultee, A., and Smid, E. J. 2001. Influence of carvacrol on growth and toxin production by *Bacillus cereus*. *International Journal of Food Microbiology*. 64: 373-378.
- Ultee, A., E. Kets, P. W., Alberda, M., Hoekstra, F. A. and Smid. E. J. 2000. Adaptation of the food-borne pathogen *Bacillus cereus* to carvacrol. *Archives* of *Microbiology*. 174:233-238.
- Ultee, A., E. Kets, P. W and Smid, E. J. 1999. Mechanisms of action of carvacrol on the food-borne pathogen *Bacillus cereus*. *Applied and Environmental Microbiology*. 65:4606-4610.
- Ultee, A., Gorris, L.G.M., and Smid, E.J. 1998. Bactericidal activity of carvacrol towards the food-borne pathogen *Bacillus cereus*. J. Appl. Microbiol. 85: 211-218.
- Ultee, A., Kets, E.P.W. and Smid, E.J. 1999. Mechanisms of action of carvacrol on the foodborne pathogen *Bacillus cereus*. *Appl. Environ. Microbiol*. 65: 4606-4610.
- Ultee, A., Kets, E.P.W., Alberda, M., Hoekstra, F.A., and Smid, E.J. 2000a. Adaptation of the food-borne pathogen *Bacillus cereus* to carvacrol. *Arch. Microbiol.* 174: 233-238.
- Ultee, A., M. Bennink, H. J. and Moezelaar, R. 2002. The phenolic hydroxyl group of carvacrol is essential for action against the food-borne pathogen *Bacillus cereus*. *Applied and Environmental Microbiology*. 68: 1561-1568.
- Ultee, A., Slump, R.A., Steging, G., and Smid, E.J. 2000b. Antimicrobial activity of carvacrol toward *Bacillus cereus* on rice. *J. Food Prot.* 63: 620-624.
- Van de Braak, S. A. A. J. and G. C. J. J. Leijten. 1999. Essential oils and oleoresins: a survey in the Netherlands and other major markets in the European Union. CBI, Centre for the Promotion of Imports from Developing Countries, Rotterdam.

- Van Tomme, S.R., Storm, G. and Hennink, W. E. 2008. *In situ* gelling hydrogels for pharmaceutical and biomedical applications. *International journal of pharmaceutics*. 355(1): 1-18.
- Vashist, A., Gupta, Y.K. and Ahmad, S. 2012. Interpenetrating biopolymer network based hydrogels for an effective drug delivery system. Carbohydrate Polymers 87: 1433-1439.
- Veldhuizen, E.J.A., Creutzberg, T.O., Burt, S.A., and Haagsman, H.P. 2007. Low temperature and binding to food components inhibit the antibacterial activity of carvacrol against *Listeria monocytogenes* in steak tartare. *J. Food Prot.* 70: 2127-2132.
- Veldhuizen, E.J.A., Tjeerdsma-Van Bokhoven, J.L.M., Zweijtzer, C., Burt, S.A. and Haagsman, H.P. 2006. Structural requirements for the antimicrobial activity of carvacrol. J. Agric. Food Chem. 54: 1874-1879.
- Wagner, H., Wierer, M. and Bauer, R. 1986. *In vitro* inhibition of prostaglandin biosynthesis by essential oils and phenolic compounds. *Planta Med.* 52: 184-7.
- Wagner, J.G. 1969. Interpretation of percent dissolved-time plots derived from in vitro testing of conventional tablets and capsules. *J. Pharm Sci.* 58(10):1253-1257.
- Wu, Y., Yang, W., Wang, C., Hu, J. and Fu, S. 2005. Chitosan nanoparticles as a novel delivery system for ammonium glycyrrhizinate. *Int. J. Pharm.* 295: 235–245.
- Xu, J., Zhou, F., Ji, B.-P., Pei, R.-S., and Xu, N., 2008. The antibacterial mechanism of carvacrol and thymol against *Escherichia coli*. *Lett. Appl Microbiol*. 47: 174-179.
- Xu, X., Li, B., Kennedy, J.F., Xie, B.J. and Huang, M. 2007. Characterization of konjac glucomannan–gellan gum blend films and their suitability for release of nisin incorporated therein. *Carbohydrate Polymers*. 70(2): 192-197.

- Yadollahi, M., Farhoudian, S., Barkhordari, S., Gholamali, I. and Farhadnejad, H. 2016. Facile synthesis of chitosan/ZnO bio-nanocomposite hydrogel beads as drug delivery systems. *International Journal of Biological Macromolecules*. 82: 273-278.
- Yadollahi, M., Farhoudian, S. and Namazi, H. 2015. One-pot synthesis of antibacterial chitosan/silver bio-nanocomposite hydrogel beads as drug delivery systems. International Journal of Biological Macromolecules 79: 37-43.
- Zhang, Y., Chan. H. F. and Leong, K. W. 2013. Advanced materials and processing for drug delivery: the past and the future. *Adv Drug Deliv Rev.* 65: 104-120.
- Zhang, H., Mardyani, S., Chan, W.C.W. and Kumacheva, E. 2006. Design of biocompatible chitosan microgels for targeted pH-mediated intracellular release of cancer therapeutics. *Biomacromolecules*. 7(5): 1568–1572.
- Zhou, F., Ji, B., Zhang, H., Jiang, H., Li, J.J., Li, J., Ren., Y. and Yan, W. 2007a. Synergistic effect of thymol and carvacrol combined with chelators and organic acids against *Salmonella Typhimurium*. J. Food Prot. 70: 1704-1709.
- Zhou, F., Ji, B., Zhang, H., Jiang, H., Yang, Z.,Li, J.J., Li, J., and Yan, W. 2007b. The antibacterial effect of cinnaldehyde, thymol, carvacrol and their combinations against the foodborne pathogen Salmonella typhimurium. J. Food Safety. 27: 124-133.

LIST OF PUBLICATIONS

EDUCATUM Journal of Science, Mathematics and Technology (2017): Preparation and Characterisation of Carvacrol Encapsulated in Gellan Gum Hydrogel. 4: 9-14.

Norafida Hasnu, Nur Aisyah Nasuha Mohd Azam and Adila Mohamad Jaafar.

Journal of Science and Mathematics Letters (2016): Preparation of Zinc Layered Hydroxide 2,4-dichlorophenoxyacetate (2,4-D) Nanocomposite. 4: 24-30.

Adila Mohamad Jaafar, Zulkarnain Zainal, Mas Jaffri Masaruddin, Norafida Hasnu and Fatin Hanifah Ayob.

Proceeding International Postgraduate Conference of Sciences and Mathematics, IPCSM '15: Preparation of Zinc Layered Hydroxide 2,4dichlorophenoxyacetate (2,4-D) Nanocomposite. 4: 24-30.

Adila Mohamad Jaafar, Zulkarnain Zainal, Mas Jaffri Masaruddin, Norafida Hasnu and Fatin Hanifah Ayob.

Bachelor Thesis: Detection of local isolates of *Bacilus subtilis* using Biochemical Test, January 2014.

Norafida Hasnu.

Preparation and Characterisation of Carvacrol Encapsulated in Gellan Gum Hydrogel

Norafida Hasnu, Nur Aisyah Nasuha Mohd Azam & Adila Mohamad Jaafar*

Department of Chemistry, Faculty of Science, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia *Email: adilamj@upm.edu.my

Abstract

Studies on plant materials as natural compound such as carvacrol (Carv) have gained much attention. Carv exhibits numerous potential as antimicrobial agent, food additives, antioxidant and etc. However, this free standing bioactive compound is unstable in the harsh environment conditions. Hence, the encapsulation technology provides protection to enhance the effectiveness in release manner. In this study, the preparation of Carv encapsulated in gellan gum hydrogel forming thin film (GG-Carv TF) was achieved by using 1.0 g of gellan gum at different concentrations of Carv (0.01-0.04 M). The FTIR spectra of GG-Carv TF revealed the combination of both functional groups from GG and Carv. The Carbon, Hydrogen and Nitrogen, CHN analysis further confirmed the encapsulation with the changes in the element percentage. Both swelling and degradation percentage increased with time and showed decreasing patterns in the range of 680.79-666.78 % and 26.83-19.15 % which can be observed as the concentration of Carv increased, respectively.

Keywords carvacrol, encapsulation, gellan gum hydrogel, thin film, natural compound

INTRODUCTION

Carvacrol (Carv) is found in the aromatic leaves and flowering plant of both thyme (*Thymus vulgaris*) and oregano (*Origanumvulgare*). Interestingly, Carv shown an effective antibacterial activity and has been proven to be potential agents in the treatment of infections and safe for human and animal consumption [1]. The world wide researchers have investigated the wide spectrum of antibacterial activity by Carv against various types of microorganisms such as *C. albicans* [2], *L. plantarum*, *S. cerevisiae*, *B. cinerea* [3], *S. aureus* [4], *Salmonella enterica* [5], *L. monocytogenes*, *E. coli* [6] and etc.

The host, hydrogel, is three-dimensional, hydrophilic, polymeric network that is capable of imbibing a large amount of water into its structure. It is highly permeable to various drug compounds, able to withstand acidic environments and high swelling properties which can release entrapped molecules through their web-like surfaces [7]. The component of hydrogel, gellan gum, is a microbial polysaccharide that is derived from *Sphingomonas elodea*, previously known as *Pseudomonas elodea*. Significantly, gellan gum is nontoxic, biocompatible, biodegradable and the resulting hydrogels is transparent and stable [8]. To date, this biopolymer based hydrogels has been gaining great attention as the potential carrier in controlled release studies.

Based on the foregoing, it is believed that the encapsulation technology provides stability and protection to enhance the effectiveness due to the facts that Carv is unstable in the harsh environment conditions. It is volatile, easily evaporates and prone to degradation during the process in growing to direct exposure of heat, pressure, light or oxygen [9]. To elucidate this matter, the Carvis encapsulated in biodegradable gellan gum hydrogel as an alternative way to extend its shelf life and to control the release manner, thereby the usage of the compound could be maximised. This study was carried out to prepare the Carv encapsulated in gellan gum hydrogel in the form of thin film and the physico-chemical properties were also investigated.

MATERIALS AND METHODS

The chemicals used in this study were glycerin (1,2,3-Propanetriol), gelzan (gellan gum), calcium chloride (CaCl₂) (\geq 96%) and carvacrol (2-Methyl-5-(1-methylethyl)-phenol) which were obtained from Sigma-Aldrich (\geq 98%), sodium dihydrogen orthophosphate (NaH₂PO₄) was purchased from BDH Chemicals Ltd Poole England (\geq 98%), sodium hydrogen carbonate (NaHCO₃) was provided by Fisher Brand (\geq 99.8%), sodium chloride (NaCl) was obtained from AnalaR (\geq 99%), and potassium chloride (KCl) was purchased from HmbG Chemicals (\geq 99.5%). All chemicals were used directly without any purification.

Preparation of Carvacrol Gellan Gum Thin Films (GG-Carv TF)

GG-Carv TF was synthesised via *in-situ* drug loading in which the Carv was first diluted in deionised water (18 M Ω cm) to the specific concentration accordingly and mixed with the dissolved 1 g of gellan gum before establishing the physical crosslinking protocol using CaCl₂. The solution was stirred at 500 rpm using hotplate set at temperature of 80°C for a total mixing of 2 hours to ensure the homogeneity. **5** ml of glycerin was added as a plasticizer. The gellan gum hydrogel encapsulated with Carv with the concentration of 0.01, 0.02 and 0.04 M are hereon referred as GG-Carv 01, GG-Carv 02 and GG-Carv 04 respectively. The solution was poured into the petri dish and left in the oven for 48 hours at 35°C for drying before storing in dessicator for further characterisation.

Characterisations

FTIR spectra of the samples were recorded in the range of 400-4000 cm⁻¹ on a Perkin-Elmer 1752X Spectrophotometer with KBr disc method. The elemental analysis was done using LECO CHNS-932 Analyser. The surface and cross section morphology of the sample analyses were observed with VPSEM (Variable Pressure Scanning Electron Microscopy) using LEO 1455.

The Study of Swelling Percentage

Water uptake of GG-Carv TF with the dimension of 2 cm x 2 cm was measured by weighing the dried films (W_d) prior to immersion into 20 ml of **Pseudo Extra Cellular Fluids**, **PECF** buffer solution with pH 5.5 at room temperature. The subsequent weight was recorded for every 24 hour. The films were removed after 72 hours, wiped gently with a tissue to expel the liquid from the surface, and were then weighed (W_w) .

The percentage of water uptake was then determined from the equilibrium swelling ratio:

Swelling Percentage (%) = $(W_w-W_d) / W_d \times 100$

Where; W_w = weight of wet sample W_d = weight of dry sample

The Study of Degradation Percentage

Degradation of GG-Carv TF was measured by weighing the initial weight of 1.0g (W_i) and left on petri dish at the room temperature. The subsequent weight was recorded for every day until a constant weight (W_f) pattern was observed.

The percentage of degradation was then determined from the equilibrium degradation ratio:

Degradation Percentage (%) = $(W_f-W_i)/W_i \ge 100$

Where; Wf = final weight of sample

Wi = initial weight of dry sample

RESULT AND DISCUSSION

Fourier Transform Infrared Spectroscopy (FTIR) Analysis

Chemical structures of the samples were characterized by FTIR (Figure 1). In general, Carv (Figure 1(a)) showed the characteristic peaks at 3360.88 cm⁻¹(phenolic-OH group), 2958.46 cm⁻¹(C-H stretching), 1583.49 and 1511.04 cm⁻¹(C-C ring stretching), 1421.54 cm⁻¹ (O-H bending), 1359.10 cm⁻¹ (isopropyl group), 1242.62 cm⁻¹ (C-O stretching) and 864.50 cm⁻¹ (aromatic ring). Meanwhile, the peaks of pure GG TF (Figure 1(e)) can be seen at 3273 cm⁻¹ (O-H stretching), 2933.35cm⁻¹ (C-H stretching), 1625.45 cm⁻¹ (C=C stretching), 1427.37cm⁻¹ (C-H bending), 1033.53 cm⁻¹ (C-O stretching) and 919.62 cm⁻¹ (C-H bending).

From the results obtained (Figure 1(b-d)), all of GG-Carv TF (s) showed the peak at the range of 3274.53-3290.70cm⁻¹ (O-H stretching) and 2890.57-2933.10 cm⁻¹ (C-H stretching) which belonged to both gellan gum hydrogel and Carv. Furthermore, the peaks at 1638.52-1642.49 cm⁻¹ and 1414.60-1415.16cm⁻¹ (C-C ring stretching), 1034.38-1035.99 cm⁻¹ (C-O stretching) and 918.57-918.92 (aromatic ring) which belonged to Carv exist in all GG-Carv TF (s), reflecting the existence of Carv in the gellan gum hydrogel polymer.

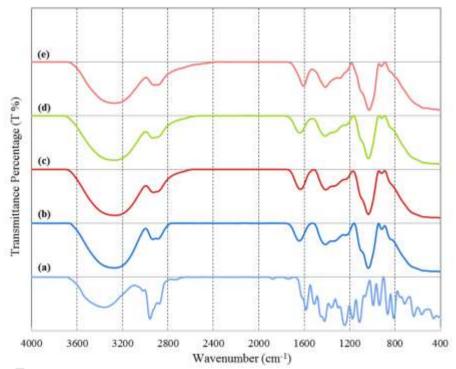


Figure 1 FTIR spectra of (a) Carvacrol (b) GG-Carv 01 (c) GG-Carv 02 (d) GG-Carv 04 (e) Pure GG TF

Elemental Analysis

Table 1 shows the weight percentage of carbon, C and hydrogen, H for pure GG TF and encapsulated GG-Carv TF with three different concentration of Carv. From Table 1, it could be observed in GG-Carv TF, that the content of C showed increasing pattern as the concentration of Carv increased. This inclined amount is due to the encapsulated Carv anion which caused the content of C to increase. Similarly, the H content in GG-Carv TF exhibited increasing pattern as the concentration of Carv increased. This analysis further confirmed the encapsulation with evidence of the changes in the element percentage.

Weight Percentage (%)	
С	Н
20.33	8.97
22.52	9.08
23.76	9.29
26.77	9.52
-	C 20.33 22.52 23.76

Table 1 Weight percentage of carbon, C and hydrogen, H for pure GG TF and encapsulated GG-Carv TF with various concentration of Carv

Variable Pressure Scanning Electron Microscopy (VPSEM) Analysis

VPSEM micrographs were used to study the surface and cross sectional area of GG-Carv TF. The observation was made at 1000 times magnification. This technique is widely used to capture the characteristic 'network' structure in hydrogels [10].

Surface Morphology

Clear network structure can be observed on the surface morphology of pure GG TF (Figure 2(a)). Meanwhile, GG-Carv TF (Figure 2(b-d)) exhibited the round-shaped structure scattered evenly which is possibly due to the Carv binding to the surface of gellan gum hydrogel. The appearances of these structures were more abundant as the concentration of Carv increased with average diameter of 5 to $10 \,\mu\text{m}$.

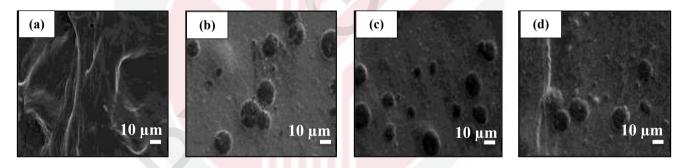


Figure 2 VPSEM surface micrograph at 1000x magnification (a) Pure GG TF (b) GG-Carv 01 (c) GG-Carv02 (d) GG-Carv 04

Cross Sectional Morphology

Unpacked layers structure can be observed in the cross sectional morphology of pure GG TF (Figure 3(a)). Meanwhile, GG-Carv TF (Figure 3(b-d)) displayed a very compact layer as the concentration of Carv increased. This can be explained due to congestion of Carv molecules residing in the gellan gum hydrogel.

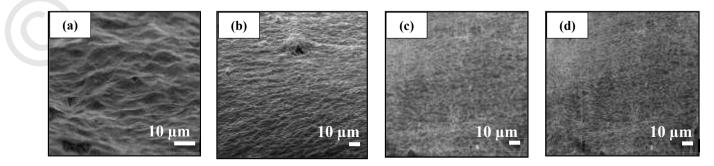
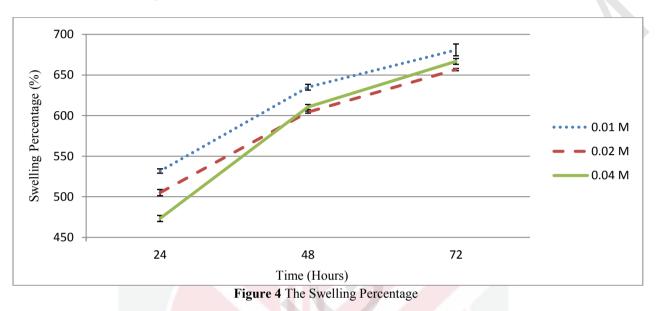


Figure 3 VPSEM cross section micrograph at 1000x magnification (a) Pure GG TF (b) GG-Carv 01 (c) GG-Carv 02 (d) GG-Carv 04

The result displayed the swelling percentage (Figure 4) increased with time. When higher concentration of Carv was used, the lesser the absorption of the solutions could be observed. This can be reflected as GG-Carv04 with the highest concentration of Carv had the lowest swelling percentage due to the formation of more rigid structure of gellan gum hydrogel. Besides that, Carv is known as the hydrophobic phenolic compound [11]. Thus, the resistance effect towards the solutions which account for the hydrophobicity of Carv also resulted in decreased swelling. Hence, the higher the concentration of Carv, the higher the water resistance of the film expected.



Degradation Percentage

Most of the degradation study of gellan gum was usually achieved in vivo through the action of enzymes and in vitro [12, 13] in accordance to their application in tissue engineering. However, to understand the degradation behaviour of polymers aimed to be used on the skin, it is important to predict and ultimately be tuned in to their condition at common room temperature for humans.

In Figure 5, the percentage of degradation was found to increase with the time. However, it was inversely proportional to the concentration. This can be seen as the concentration of Carv increased, the degradation percentage decreased. Similar to the swelling results, this might be explained by the formation of more rigid structure of gellan gum hydrogel occurring in GG-Carv TF at higher concentration. Hence, this stability resulted in more durable GG-Carv TF against the environment conditions as the concentration increased.

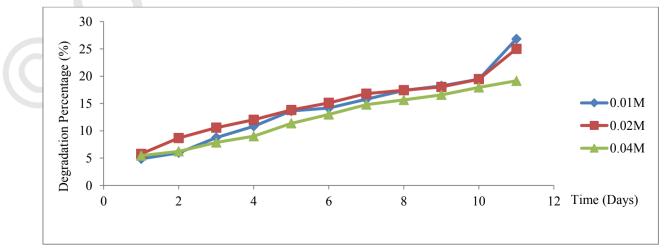


Figure 5 The Degradation Percentage

CONCLUSION

The preparation of carvacrol encapsulated in gellan gum hydrogel in the form of thin film (GG-Carv TF) was successfully achieved as confirmed by the FTIR spectrum of GG-Carv TF which showed the combination of both functional groups from the gellan gum hydrogel and Carv. The CHN analysis further confirmed the existence of the element with changes of the element percentage. The swelling and degradation percentage similarly increased with time and decreasing patterns can be observed as the concentration of Carv increased. This study has generated the fundamental knowledge of gellan gum hydrogel-Carvthin films which could be used for further studies in the development of antibacterial applications.

ACKNOWLEDGEMENT

The authors wish to thank the Ministry of Higher Education Malaysia for financial assistance under Fundamental Research Grant Scheme (FRGS–Vote: 5524557) and Universiti Putra Malaysia for providing the facilities throughout this work.

REFERENCES

- [1] Nostro A., Roccaro, A. S., Bisignano, G., Marino, A. and Blanco, A.R. (2007). Effect of oregano, Carvacrol and Thymol on S. aureus and S. epidermdis biofilms. *J. Med. Microbiol.* 56: 591-523.
- [2] Chami, F., Chami, N., Bennis, S., Bouchikhi, T., and Remmal, A. (2005). Oregano and clove essential oils induce surface alteration of Saccharomyces cerevisiae. *Phytother. Res.* 19:405-408.
- [3] Arfa, B.A., Combes, S., Preziosi-Belloy, L., Gontard, N., and Chalier, P. (2006). Antimicrobial activity of carvacrol related to its chemical structure. *Lett.in Appl. Microbiol.* 43:149-154.
- [4] Oussalah, M., Caillet, S., Saucier, L., and Lacroix, M. (2007). Inhibitory effects of selected plant essential oils on the growth of four pathogenic bacteria: E. coli O157:H7, Salmonella Typhimurium, Staphylococcus aureus and Listeria monocytogenes. *Food Control*. 18:414-420.
- [5] Ravishankar, S., Zhu, L., Reyna-Granados, J., Law, B., Joens, L., and Friedman, M. (2010). Carvacrol and cinnamaldehyde inactivate antibiotic-resistant Salmonella enterica in buffer and on celery and oysters. *J. Food Prot.* 73:234-240.
- [6] Perez-Conessa, D., Cao, J., Chen, L., Mclandsborough, L., and Weiss, J. (2011). Inactivation of Listeria monocytogenes and Escherichia coli O157:H7 biofilms by micelle encapsulated eugenol and carvacrol. *J. Food Prot.* 74:55-62.
- [7] Han J. H., Krochta, J. M., Kurth, M. J., and Hsieh, Y. L. (2000). Lactitol-based poly (ether polyol) hydrogels for controlled release chemical and drug delivery systems. *Journal of Agricultural and Food Chemistry*. 48(11), 5278-5282.
- [8] Kang D., Zhang, H. B., Nitta, Y., Fang, Y. P., & Nishinari, K. (2015). Gellan. Polysaccharides: Bioactivity and Biotechnology. 1627-1682.
- [9] Chalier P., A.B. Arfa, L.P. Belloy and N. Gontard. (2007). Carvacrol losses from soy protein coated papers as a function of drying conditions. *Journal of Applied Polymer Science*. 106: 611-620.
- [10] Pourjavadi A., Kurdtabar M. (2007). Collagen-based highly porous hydrogel without any porogen: Synthesis and characteristics. *European Polymer Journal*. 43: 877-889.
- [11] Veldhuizen, E. J. A., Tjeerdsma-Van Bokhoven, J.L.M., Zweijtzer, C., Burt, S.A., and Haagsman, H.P. (2006). Structural requirements for the antimicrobial activity of carvacrol. *J. Agric. Food Chem.* 54:1874-1879.
- [12] Oliveira J. T., Santos T. C., Martins L., Picciochi R, Marques A. P., Castro A. G. (2010). Gellan gum injectable hydrogels for cartilage tissue engineering applications: in vitro studies and preliminary in vivo evaluation. *Tissue Eng Part A.* 16 (1):343-353.
- [13] Daniela F. Coutinho, Shilpa V. S., HyeonghoS., João T. O., Manuela E. G., Nuno M. N., Ali K., and Rui L. R. (2010). Modified Gellan Gum hydrogels with tunable physical and mechanical properties. *Journal Biomaterials*. 31: 7494-7502.

Preparation of Zinc Layered Hydroxide 2,4-dichlorophenoxyacetate (2,4-D) Nanocomposite

Penyediaan Nano Komposit Zink Hidrosida Berlapis 2,4-dichlorophenoxyacetate (2,4-D)

Adila Mohamad Jaafar¹, Zulkarnain Zainal¹, Mas Jaffri Masaruddin²,

Norafida Hasnu¹, Fatin Hanifah Ayob¹

¹Department of Chemistry, Faculty of Science, Universiti Putra Malaysia, 43400 Serdang, Selangor,

Malaysia.

²Faculty of Biotechnology and Biomolecular, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia. Email: adilamj@upm.edu.my

Abstract

Recently, layered materials have gained great attention because they can be designed at the nanometer levels. Due to its advantages such as low cost material, easy to synthesize and excellent anion exchange properties, these material exhibit unlimited potential in catalysis, separation technology, medical science, nanocomposite material engineering, agriculture and etc. In this study, Zinc Layered Hydroxide (ZLH) has been intercalated with 2,4-Dichlorophenoxyacetic acid (2,4-D) anion by using ion exchange method. These nanocomposites were synthesized at a different mass of ZLH which are 1 g, 2 g and 3 g also with different concentration of guest anions (2,4-D) which are 0.01 M, 0.02 M, 0.04 M, 0.08 M, 0.16 M and 0.32 M. Powder X-ray Diffraction (PXRD) patterns showed an expansion of interlayer spacing with the value ranging from 26.8 Å to 39.6 Å, 29.4 Å to 37.3 Å, 28.7 Å to 31.6 Å for 1 g, 2 g and 3 g of ZLH at various concentration of 2,4-D respectively. This expansion of basal spacing implies that 2,4-D anion which is bigger size than the prior ZLH was successfully intercalated into the ZLHs. FTIR analysis further confirmed that 2,4-D were successfully intercalated between the interlayers of ZLH with evidence of functional groups of ZLH and 2,4-D in the ZLH-2,4-D nanocomposite (ZN) spectra.

Keywords Zinc Layered Hydroxide, 2,4-Dichlorophenoxyacetic acid (2,4-D), ion exchange, intercalation, nanocomposite

Abstrak

Pada masa kini, bahan berlapis telah mendapat perhatian kerana ia boleh direka bentuk pada skala nano. Kerana kelebihannya seperti murah, mudah disintesis dan sifat pertukaran anion yang baik, maka bahan-bahan ini menunjukkan potensi yang tinggi untuk digunakan bidang katalis, teknologi penapis, sains perubatan, bahan kejuruteraan nano komposit, pertanian dan berbagai kegunaan lagi. Dalam kajian ini, Zink Hidrosida Berlapis (ZLH) telah di sisipkan dengan anion asid 2,4-Dichlorophenoxyacetic (2,4-D) secara kaedah pertukaran ion. Komposit nano ini telah disistesis dengan berlainan jisim ZLH iaitu 1 g, 2 g dan 3 g serta masing-masing dengan berbagai kepekatan anion tumpangan (2,4-D) iaitu 0.01 M, 0.02 M, 0.04 M, 0.08 M, 0.16 M dan 0.32 M. Corak Pembelauan Sinar-X (PXRD) menunjukkan jarak antara lapisan mengembang dari 26.8 Å kepada 39.6 Å, 29.4 Å kepada

37.3 Å dan 28.7 Å kepada 31.6 Å masing-masing bagi 1 g, 2 g dan 3 g ZLH dengan berbagai kepekatan 2,4-D. Pengembangan jarak di antara lapisan ini menunjukkan bahawa anion 2,4-D adalah bersaiz lebih besar daripada saiz ZLH yang berjaya disisip ke dalam lapisan-lapisan ZLH. Analisis FTIR mengesahkan bahawa 2,4-D telah berjaya disisipkan di antara lapisan ZLH dengan bukti terdapatnya kumpulan berfungsi ZLH dan 2,4-D dalam spektrum FTIR komposit nano (ZN)

Kata kunci Lapisan Zink Hidroksida, asid 2,4-Dichlorophenoxyacetic (2,4-D), pertukaran ion, sisipan, komposit nano

INTRODUCTION

Layered single hydroxide salt (LSH) such as zinc layered hydroxide (ZLH) is a layered inorganic compound which has gained attention in wide range of applications, particularly due to its unique anion exchange properties (Abdul Latip *et al.*, 2013). Recent studies reported that ZLH has high capacity to accommodate guest molecules and stronger host-guest interaction which leads to high stability; which makes it possible to be used as new host material and delivery system with controlled release rate of active agents (Kasai *et al.*, 2006; Yang *et al.*, 2007). ZLH structure is high potential host material in forming nanocomposites because it can expand or contract depending on nature of interlayer anions (Mohsin *et al.*, 2013). Due to that, the ZLH particularly have been studied extensively and intercalated with various organic anions (Liang *et al.*, 2004) mainly via ion exchange process, ranging from anionic dyes (Marangoni R *et al.*, 2009), porphyrin sensitizers (Demel J *et al.*, 2010) and an anti-corrosive compound (Rocca E *et al.*, 2006).

Researches have proven this LSH is currently gaining attention due to its simple method of synthesis, as a precursor for a wide band gap of ZnO and its anion exchange properties (Thomas N *et al.*, 2011). LSH also has demonstrated the ability to extend the release period of drug molecules and bioactive molecules (Hussein, M., *et al.*, 2010) that prompting more investigations towards potential applications of LSH in drug delivery systems. LSH are thought to be ideal candidates for agricultural applications (Choy *et al.*, 2007). Others active agents such as drugs, vitamins and dyes need to be released in controlled manner to reduce their toxicity or other side effects to increase its durability and stability (Hwang *et al.*, 2001).

Recently, the study on the intercalation of phenoxy herbicides into the interlayer of LDH (Layered Double Hydroxide) using various synthesis methods been reported. 2,4-D is highly selective herbicide which is toxic to broad leaved plants but less harmful to grasses. This chemical has complex mechanism of action against weeds, resembling those of auxins (growth hormones). Once adsorbed 2,4-D is translocate within the plant and accumulates at the growing points of roots and shoots where it inhibits growth (Kenneth, 1983). In this paper, we reported the preparation of Zinc Layered Hydroxide 2,4-Dichlorophenoxyacetate (2,4-D) nanocomposite.

MATERIALS AND METHODS

Synthesis of materials

The chemicals used in the synthesis were of analytical grade, obtained from various chemical suppliers and were used without any further purification. The chemicals are

2,4-dichlorophenoxyacetic acid > 98% from Merck and Zinc Oxide > 99% from Acros Organics. All of the solutions were prepared using deionised water. The synthesis of ZN 1 was prepared by ion exchange method. 1 g Zinc Oxide, ZnO (which then referred as ZLH material) in 100 ml of water was mixed with 2,4-D solution at chosen concentrations (ranging from 0.01 M to 0.32 M). The solution mixture was stirred for 2 hours with magnetic stirring and was conducted under atmospheric conditions. Once stirring was completed, the precipitate was aged at 70 °C for 18 hours in an oil bath shaker, cooled, thoroughly washed and dried overnight in an electric oven at 70 °C. Finally, the dried sample was ground into fine powder by using mortar and pestle, then kept in bottle sample for further used and characterizations. Similar procedure was repeated for ZN 2 and ZN 3 with 2 g and 3 g of ZnO respectively.

Characterisation of material

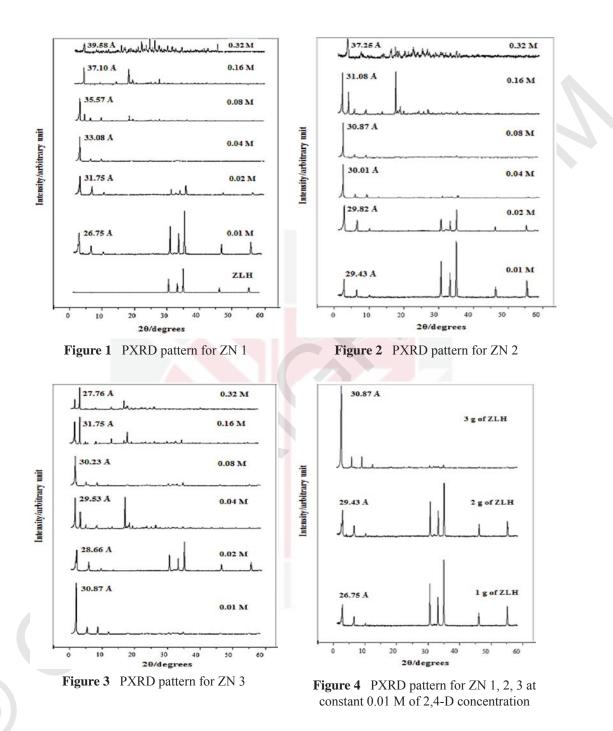
Powder X-ray diffraction (PXRD) patterns of the samples were recorded between 2° and 60° on a Shimadzu 6000 model analytical powder diffractometer using Cu K α radiation at 30 kV and 30 mA at the rate of 4° min⁻¹. FTIR spectra of the materials were recorded over the range 400 - 4000 cm⁻¹ on a Perkin-Elmer 1752X Spectrophotometer using KBr disc method.

RESULTS AND DISCUSSION

Powder X-ray Diffraction Analysis

Figure 1 show PXRD patterns of ZN 1 synthesised at different concentration of 2,4-D with 1 g of ZLH. The basal spacing showed increasing in the expansion from 26.75 Å to 31.75 Å, 33.08 Å, 35.57 Å, 37.10 Å, and 39.58 Å with the increasing concentration. Figure 2 also shows the same trends. ZN 2 gives the harmonic increasing in the expansion of basal spacing ranging from 29.43 Å to 37.25 Å. PXRD patterns of these two prepared samples display a high intensity diffraction peak indicating a pure phase material without any ZnO phase. This shows that a well-ordered nano-layered structure with good crystallinity was obtained at this optimum condition. The increase of the basal spacing is associated with the spatial orientation and revealed that the size of 2,4-D is bigger than nitrate in the interlayer region (Kuh and Huh, 1998). Meanwhile, in Figure 3, the poor trends detected with 3 g of ZLH. It shows the disorder expansion of basal spacing of 30.87 Å, 28.66 Å, 29.53 Å, 30.23 Å, 31.75 Å, and 27.76 Å due to poor crystallinity obtained.

According to all values of basal spacing, it is proven that 2,4-D was intercalated into ZLH interlayers. In Figure 4, comparison study on various masses of ZLH used in nanocomposite at 0.01 M of 2,4-D. It shows the increases of the basal spacing expansion is proportional with the increases mass of ZLH. The obtained basal spacing value is higher than those reported for the intercalation of other type of herbicides into the LDH interlayers (Cardoso *et al.*, 2006; Sarijo *et al.*, 2010). ZLH reportedly has larger interspacing than LDH to accommodate a greater number of incoming guest anions of varying sizes, due to its higher charge density (Kasai *et al.*, 2006; Hwang *et al.*, 2001; KoreYang *et al.*, 2007). Thus it is possible to simultaneously intercalate 2,4-D anions into the ZLH interlayers.



Fourier Transform Infrared Spectroscopy

Figure 5 shows the FTIR spectra of ZLH, pure 2,4-D and ZLHs. The insertion of 2,4-D into the interlayer ZLH was confirmed by the FTIR spectrum, which is complementary to that of PXRD results. All of the nanocomposites display similar absorption bands as

the parent material, ZLH and guest ion, 2,4-D anion that are intercalated into the host interlayer galleries. The presence of 2,4-D anion are shown in the typical broad absorption bands of 2,4-D at 3078 cm⁻¹ and 2969 cm⁻¹, corresponding to the O-H stretching vibration of the COOH, while a strong band at 1717 cm⁻¹ corresponds to the stretching of C=O. The bands at 1471 cm⁻¹ corresponds to the stretching that are attributed to C=C vibrations of the aromatic ring of phenoxy. Absorption band is observed at about 1230 cm⁻¹ of FTIR spectra due to C-O-C symmetric stretching modes.

FTIR spectra of ZN 1, ZN 2 and ZN 3 at a constant 0.01 M concentration are observed. The same typical broad absorption bands were observed at 3388 cm⁻¹ for ZN 2 and ZN 3, while ZN 1 showed broad absorption bands at 3542 cm⁻¹, 3437 cm⁻¹, and 3263 cm⁻¹. These absorption bands are corresponding to the vibrations of the hydroxyl groups at surface, interlayer water molecules and the water bending mode (Lakraimi *et al.*, 2000; Palmer *et al.*, 2009). Meanwhile, the bands observed at 1595 cm⁻¹ for ZN 1, 1599 cm⁻¹ for ZN 2, and 1604 cm⁻¹ for ZN 3, corresponds to the stretching vibration of aromatic ring C=C. The disappearance of bands in the nanocomposites spectrum at 1717 cm⁻¹ and 1230 cm⁻¹ indicated C=O stretching vibration of the protonated carboxylic groups of the herbicides respectively (Cardoso *et al.*, 2006), shows that anions in host material were completely exchanged with 2,4-D anions for the formation of the ZN.

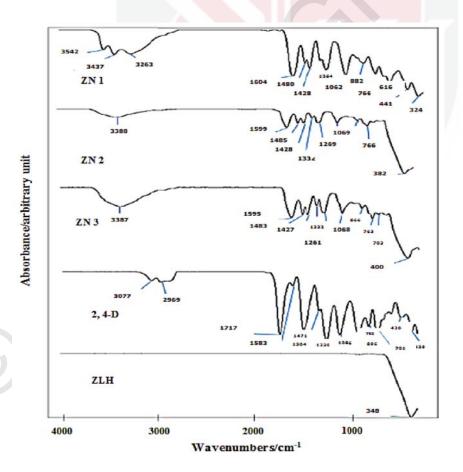


Figure 5 FTIR spectra for ZN 1, ZN 2 and ZN 3 at constant 0.01 M of 2,4-D concentration

CONCLUSION

A series of ZLH-2,4-D (ZN) nanocomposites prepared at different masses of ZLH (1 g to 3 g) and various concentration of 2,4-D (0.01M to 0.32M) have been successfully done via ion-exchange method. This study suggests that the layered hydroxide can be used as a carrier for the 2,4-dichlorophenoxyacetic acid (2,4-D) for further used as herbicide.

REFERENCES

- Abdul Latip, A. F., Hussein, M. Z., Stanslas, J., Wong, C. C., and Adnan, R. (2013). Release behavior and toxicity profiles towards A549 cell lines of ciprofloxacin from its layered zinc hydroxide intercalation compound. *Chemistry Central Journal*, 7:119.
- Adila, M. J. (2010) Application of layered double hydroxides as host in controlled release formulation of latex stimulant and metal catalyst in formation of carbon nanotubes. *Degree of Doctor of Philosophy Thesis.* University Putra Malaysia.
- Cardoso, L.P., Celis, R., Cornejo, J., Valim, J. B. (2006). Layered double hydroxides as supports for the sow release of acid herbicides. *Journal of Agricultural and Food Chem*istry, 54: 5968-5975.
- Choy, J. Choi, S., Oh, J., & Park, T. (2007). Clay minerals and layered double hydroxides for novel biological applications. *Applied Clay Science*, 36(1-3): 122-132.
- Demel, J., Kubat, P., Jirka, I., Kovar, P., Pospisil, M., & Lang, K. (2010). Inorganic-organic hybrid materials: Layered zinc hydroxide salts with intercalated porphyrin sensitizers. *The Journal of Physical Chemistry* C, 114: 16321-16328.
- Hussein, M., Hashim, N., Yahaya, A. H., & Zainal, Z. (2010). The synthesis and characterization of [4-(2,4-dichlorophenoxybutyrate)-zinc layered hydroxide] nanohybrid. Solid State Sciences, 12(5): 770-225.
- Hwang S. H., Han Y. S., Choy J. H. (2001). Intercalation of functional organic molecules with pharmaceutical, cosmeceutical and nutraceutical functions into layered double hydroxides and zinc basic salts. *Bulletin-Korean Chemistry Society*, 22:1019–1022.
- Hwang S. H., Han Y. S., Choy J. H. (2001). Intercalation of functional organic molecules with pharmaceutical, cosmeceutical and nutraceutical functions into layered double hydroxides and zinc basic salts. *Bulletin-Korean Chemistry Society*, 22:1019–1022.
- Kasai A., Fujihara S. (2006). Layered single-metal hydroxide/ethylene glycol as a new class of hybrid material. *Inorganic Chemistry*, 45: 415–418.
- Kenneth, B. (1983). Plant growth regulatoruse in natural rubber (Hevea brasiliensis). Plant Growth Regulating Chemicals. 1:41-58.
- KoreYang J. H., Han Y. S., Park M., Park T., Hwang S. J., Choy J. H. (2007) New inorganic -based drug delivery system of indole-3-acetic acid-layered metal hydroxide nanohybrids with controlled release rate. *Chemistry Material*, 19: 2679–2685.
- Kuk W, Huh Y (1998) Preferential intercalation of isomers of anthraquinone sulfonate ions layered double hydroxide. *J. Mater. Chem.* 9:1933–1936
- Lakraimi, M., Legrouri, A., Barroug, A., De Roy, A., & Besse, J. P. (2000). Preparation of a new stable hybrid material by chloride-2,4-dichlorophenoxyacetate ion exchange into the zinc-aluminium-chloride layered double hydroxide. *Journal of Materials Chemistry*, 10(4): 1007-1011.
- Liang, C., Shimizu, Y., Masuda, M., Sasaki, T., & Koshizaki, N. (2004). Preparation of layered zinc hydroxide/surfactant nanocomposite by pulsed-laser ablation in a liquid medium. *Chemistry of Materials*, 16(6): 963-965.

- Marangoni, R., Ramos, L. P., & Wypych, F. (2009). New multifunctional materials obtained by the intercalation of anionic dyes into layered zinc hydroxide nitrate followed by dispersion into poly (vinyl alcohol) (PVA). *Journal of Colloid and Interface Science*, 330(2):303-309.
- Mohsin, S. M. N., Hussein, M. Z., Sarijo, S. H., Fakurazi, S., Arulselvan, P., Hin T. Y. Y. (2013). Synthesis of (cinnamate-zinc layered hydroxide) intercalation compound for sunscreen application. *Chemistry Central Journal*, 7(26).
- Palmer, S. J., Frost, R. L., & Nguyen, T. (2009). Hydrotalcites and their role in coordination of anions in Bayer liquors: anion binding in layered double hydroxides. *Coordination Chemistry Reviews*, 253(1): 250-267.
- Rocca, E., Caillet, C., Mesbah, A., Francois, M., & Steinmetz, J. (2006). Intercalation of zinc layered hydroxide: Zinc hydroheptanoate used as protective material on zinc. *Chemistry of Materials*, 18(26): 6186-6193.
- Sarijo, S.H., Hussein, M.Z., Yahaya, A.H., Zainal, Z., Yarmo, M.A. (2010). Synthesis of phenoxyherbicides-intercalated layered double hydroxide nanohybrids and their controlled release property. *Current Nanoscience*, 6(2):199–205.
- Yang, J. H., Han, Y. S., Park, M., Park, T., Hwang, S. J., & Choy, J.H. (2007). New inorganicbased drug delivery system of indole-3-acetic acid-layered metal hydroxide nanohybrids with controlled release rate. *Chemistry of Materials*, 19(10): 2679-2685.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION :

TITLE OF THESIS / PROJECT REPORT :

PREPARATION, CHARACTERISATION AND RELEASE OF CARVACROL ENCAPSULATED IN GELLAN HYDROGEL AND CHITOSAN NANOPARTICLES FOR ANTIBACTERIAL APPLICATION

NAME OF STUDENT: NORAFIDA BINTI HASNU

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (√)



CONFIDENTIAL

RESTRICTED

(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from		until	
· _	(date)		(

(date)

Approved by:

(Signature of Student) New IC No/ Passport No.: (Signature of Chairman of Supervisory Committee) Name:

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]