



UNIVERSITI PUTRA MALAYSIA

***BIOLOGICAL ATTRIBUTE AND DISEASE RESISTANCE OF CATFISH
Clarias gariepinus (Burchell 1822) SUPPLEMENTED WITH DIFFERENT
RATIONS OF SPIRULINA Arthrospira platensis***

NOR FATIAH MOHD NASIR

FS 2017 88



**BIOLOGICAL ATTRIBUTE AND DISEASE RESISTANCE OF CATFISH
Clarias gariepinus (BURCHELL 1822) SUPPLEMENTED WITH DIFFERENT
RATIONS OF SPIRULINA *Arthrospira platensis***

By

NOR FATIHAH MOHD NASIR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Master of Science**

October 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master Science

**BIOLOGICAL ATTRIBUTE AND DISEASE RESISTANCE OF CATFISH
Clarias gariepinus (BURCHELL 1822) SUPPLEMENTED WITH DIFFERENT
RATIONS OF SPIRULINA *Arthrospira platensis***

By

NOR FATIHAH MOHD NASIR

October 2017

Chair: Mohammad Noor Amal Azmai, PhD

Faculty: Science

Spirulina Arthrospira platensis is practically used as a supplement in nutrition due to its high nutritional content. Improving the feed formulation by combining the commercial fish feed with spirulina to the cultured fish might elevate the fish growth performance, nutritional value and increase its resistance towards bacterial infection. This study was conducted to determine the growth performance, nutritional value and disease resistance of catfish *Clarias gariepinus* (Burchell 1822) supplemented with different ration of spirulina, *A. platensis*. Five experimental diets were prepared by varying the percentage of spirulina in the fish feed containing 0% (control), 1%, 3%, 5% and 7% respectively of locally grown spirulina. A total of 30 fry (2.62 ± 0.04 g of fish initial total body weight; 7.09 ± 0.05 cm of total body length) were stocked per cage and in triplicate. Each diet was fed to the fry twice daily at 5% of fish body weight for 90 consecutive days. Growth performances were determined every fortnight. The proximate analysis of spirulina, experimental diets and catfish fillet was done according to the method of AOAC, 1990. The catfish were then intraperitoneally injected with 10^7 CFU/ml of virulent *Aeromonas hydrophila* on day 91, and monitored until day 104. Catfish with diet of 7% spirulina produced significantly higher ($p < 0.05$) in weight gain and specific growth rate compared to control, but not with other treatment groups. Standard length, percent of survival, feeding efficiency of the catfish increased with the increasing percentage of spirulina inclusion but was not significantly different ($p > 0.05$) between all the diet groups. The feed conversion rate marginally decreased as spirulina inclusion increased in the diet, but not significantly different ($p > 0.05$) between all the treatment groups. The gonado somatic index value was the highest in catfish with diet 7% spirulina but not significantly different ($p > 0.05$) between all the treatment groups. The hepatosomatic index and viscerosomatic index value were also not significantly different ($p > 0.05$) between all the treatment groups. The proximate compositions of the locally grown dried spirulina showed the highest content was the crude protein with $63.07 \pm 0.02\%$ ($p < 0.05$) followed by the carbohydrate content, ash, moisture, crude lipid and lastly crude fiber. Generally, as the percent of spirulina in the diet increase the proximate composition of the experimental

diets were significantly increased ($p < 0.05$) for the crude protein and significantly lower ($p < 0.05$) for the crude fiber content. The moisture, ash, crude lipid and carbohydrate showed no significant ($p > 0.05$) different between all the experimental diet groups. The content of proximate composition of catfish fillet showed at par to the control group with no significant different ($p > 0.05$) between all diet groups except the crude fiber content that were significantly increased ($p < 0.05$) with increasing percent of spirulina until at maximum of 3% of spirulina inclusion and there was no significant different ($p > 0.05$) at 5% and 7% of spirulina diet. The percentage of catfish survive following *A. hydrophila* infection were significantly low ($p < 0.05$) in control with 36.67% but not with other treatment groups range from 70.00% to 73.33%. The white blood cell count value was significantly higher ($p < 0.05$) in group supplemented with 7% spirulina whereas the red blood cell, hemoglobin, packed cell volume, mean corpuscular volume, mean corpuscular hemoglobin concentration value showed no significant different between the treatment groups. This study conclude that the catfish supplemented with spirulina as low as 1% resulted in optimum disease resistance against the bacterial infection of *A. hydrophila* which helps in preventing mass mortality of catfish although the spirulina does not accelerate the growth performance and nutritional value of the *C. gariepinus*.

Keyword: *Clarias gariepinus*, *Aeromonas hydrophila*, *Arthrospira platensis*, growth performance, disease resistance

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

ATRIBUT BIOLOGI DAN RINTANGAN PENYAKIT IKAN KELI *Clarias gariepinus* DITAMBAH DENGAN NISBAH BERBEZA SPIRULINA *Arthrospira platensis*

Oleh

NOR FATIHAH MOHD NASIR

Oktober 2017

Pengerusi: Mohammad Noor Amal Azmai, PhD
Fakulti: Sains

Spirulina Arthrospira platensis lazimnya digunakan sebagai tambahan dalam pemakanan kerana kandungan khasiatnya yang tinggi. Meningkatkan rumusan makanan yang menggabungkan makanan ikan komersial dengan spirulina kepada ikan yang ditanam mungkin dapat meningkatkan prestasi pertumbuhan ikan, nilai khasiat pemakanan dan meningkatkan daya tahan ikan daripada jangkitan bakteria. Kajian ini dijalankan untuk menentukan prestasi pertumbuhan, nilai nutrisi ikan dan rintangan penyakit ikan keli *Clarias gariepinus* (Burchell 1822) yang ditambah dengan catuan berbeza spirulina, *A. platensis*. Lima diet eksperimentasi yang disediakan dengan memanipulasi peratusan spirulina dalam makanan ikan masing-masing mengandungi 0% (pemalar), 1%, 3%, 5% dan 7% spirulina keluaran tempatan. Sebanyak 30 ekor anak ikan keli Afrika (2.62 ± 0.04 g jumlah berat ikan awal; 7.09 ± 0.05 cm jumlah panjang ikan) telah ditempatkan dalam setiap sangkar secara triplikasi. Diet yang disediakan diberi makan kepada anak ikan dua kali sehari 5% dari berat badan ikan selama 90 hari. Kadar pertumbuhan ditentukan setiap dua minggu. Analisis proximat spirulina, diet eksperimen dan fillet ikan keli dilakukan mengikut kaedah AOAC 1990. Ikan keli kemudiannya disuntik secara intraperitoneal dengan 10^7 CFU/ml *A. hydrophila* pada hari ke-91 dan pemerhatian dilakukan sehingga hari ke-104. Keli dengan diet 7% spirulina menghasilkan perbezaan yang signifikan ($p < 0.05$) dalam jumlah pertambahan berat badan dan kadar pertumbuhan tertentu berbanding pemalar, tetapi tidak dengan kumpulan diet yang lain. Panjang standard, peratus hidup, FE ikan keli meningkat dengan pertambahan peratusan spirulina di dalam diet tetapi tidak berbeza secara signifikan ($p > 0.05$) antara semua kumpulan diet. Nilai kadar penukaran makanan menunjukkan sedikit penurunan apabila peratus spirulina meningkat di dalam diet, tetapi tidak berbeza secara signifikan ($p > 0.05$) antara semua kumpulan diet. Nilai gonad somatik indeks adalah yang tertinggi dalam ikan keli dengan diet 7% spirulina tetapi tidak ketara berbeza ($p > 0.05$) antara semua kumpulan rawatan. Nilai hepatosomatik indeks dan viscerosomatik indeks juga tidak berbeza secara ketara ($p > 0.05$) antara semua kumpulan rawatan. Komposisi proksimat spirulina kering tempatan menunjukkan kandungan tertinggi ialah protein mentah dengan $63.07 \pm 0.02\%$

($p < 0.05$) diikuti dengan kandungan karbohidrat, abu, kelembapan, lipid mentah dan akhir sekali ialah serat mentah. Secara umumnya, apabila peratus spirulina meningkat dalam diet komposisi proksimat meningkat bagi kandungan protein mentah ($p < 0.05$) dan menurun bagi kandungan serat mentah ($p < 0.05$). Nilai kandungan kelembapan, abu, lipid mentah dan karbohidrat tidak menunjukkan perbezaan yang signifikan ($p > 0.05$) antara semua kumpulan diet eksperimen. Kandungan komposisi proksimat fillet ikan keli kesemua kumpulan diet menunjukkan nilai yang setanding dengan kumpulan kontrol tanpa sebarang perbezaan ($p > 0.05$) kecuali kandungan serat mentah yang meningkat secara ketara ($p < 0.05$) dengan peningkatan peratus spirulina sehingga 3% campuran spirulinadan tiada perbezaan yang signifikan ($p > 0.05$) pada kadar 5% dan 7% campuran spirulina. Peratusan hidup ikan keli susulan jangkitan *A. hydrophila* adalah sangat rendah ($p < 0.05$) dalam kumpulan kawalan dengan 36.67% tetapi tidak dengan kumpulan rawatan lain dimana mempunyai peratus hidup dari 70.00% sehingga 73.33%. Nilai kiraan sel darah putih adalah lebih tinggi ($p < 0.05$) dalam kumpulan ikan keli yang diberi makan dengan 7% spirulina manakala nilai sel darah merah, haemoglobin, isipadu sel padat, min isi padu korpuskel dan min kepekatan korpuskel haemoglobin menunjukkan tiada perbezaan yang signifikan ($p > 0.05$) di antara kumpulan rawatan. Kajian ini menyimpulkan bahawa ikan keli yang diberi makanan tambahan spirulina pada 1% dapat mencapai daya tahan penyakit yang optimum dalam melawan penyakit jangkitan bakteria *A. hydrophila* dimana ia dapat membantu dalam mengurangkan jumlah kematian ikan keli secara besar-besaran walaupun spirulina tidak meningkatkan prestasi pertumbuhan dan nilai khasiat *C. gariepinus*.

Kata kunci: *Clarias gariepinus*, *Aeromonas hydrophila*, *Arthrospira platensis*, prestasi pertumbuhan, daya tahan penyakit

ACKNOWLEDGEMENTS

First and foremost, I am grateful to Allah S.W. T. as I am able to complete what I had started. This project ends smoothly with the given mercy and never ending grace. I am hereby would like to express my deepest gratitude to my committed and proactive supervisor, Dr. Mohammad Noor Amal Azmai from the Department of Biology, Faculty of Science, UPM whom never gave up on his student and always hope only for the best for all of them. I also would like to thank my wonderful colleagues. Without all the support, my journey towards the end might not be accomplished. It was always a pleasure doing the research with such passionate, helpful, engaging and friendly people. I would also like to acknowledge the experts who were involved in making this project successful, Mr. Raden Ahmad Zaidi Raden Zainal, Mr. Kharizan Suriati and others from the Fish Breeding Centre, UPM, Serdang for their input and guidance. This project would have been impossible without the support of GP-IPS for the research grant and scholarship from MyBrain15 and also Grant Research Fellowship from UPM. Finally, I must express my very profound gratitude and love to my parent Mr. Mohd Nasir Abd Samad and Mss. Rohana Manaf also not to be forgotten Mr. Kaz for providing me with unfailing support and continuous encouragement throughout my years of study and through the process of researching and writing this thesis. This accomplishment would not have been possible without them.

I certify that a Thesis Examination Committee has met on 12 October 2017 to conduct the final examination of Nor Fatihah binti Mohd Nasir on her thesis entitled “Biological Attributes and Disease Resistance of Catfish *Clarias gariepinus* (Burchell 1822) Supplemented with Different Rations of *Spirulina Arthrospira platensis*” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Muskhazli bin Mustafa, PhD

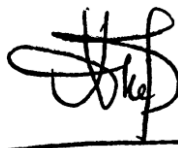
Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Hishamuddin bin Omar, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Internal Examiner)

Najiah Musa, PhD

Professor
Universiti Malaysia Terengganu
Malaysia
(External Examiner)



NOR AINI AB. SHUKOR, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 30 November 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohammad Noor Amal Azmai, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Ahmad Ismail, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)



ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Nor Fatimah binti Mohd Nasir (GS42497)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of
Supervisory

Committee: Mohammad Noor Amal Azmai

Signature: _____

Name of Member of
Supervisory

Committee: Ahmad Ismail

TABLE OF CONTENTS

	Pages
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	vii
LIST OF TABLES	xii
LIST OF FIGURES	xiv
LIST OF APPENDICES	xv
LIST OF ABBREVIATIONS	xvi
CHAPTER	
1 INTRODUCTION	
1.1 Background of the study	1
1.2 Problem statement	2
1.3 Objectives	4
1.4 Hypothesis	4
2 LITERATURE REVIEW	
2.1 The history of Malaysian aquaculture	5
2.2 Current status of Malaysian aquaculture	6
2.3 Catfish culture and production in Malaysia	7
2.4 Issues in aquaculture	8
2.4.1 Feed quality and cost	8
2.4.2 Ethical issue	9
2.4.3 Water quality	9
2.4.4 Environment impact	9
2.4.5 Disease	10
2.5 Essential components for feed in fish aquaculture	11
2.5.1 Carbohydrate	11
2.5.2 Lipid	11
2.5.3 Protein	12
2.5.4 Vitamin	13
2.5.5 Fiber	13
2.6 Spirulina	13
2.7 Spirulina as potential source of feed supplement of fish	14
2.7.1 Growth performance	15
2.7.2 Nutritional value	15
2.7.3 Immunity	15
2.8 <i>Aeromonas hydrophila</i> infection in catfish culture	16
2.8.1 Diagnosis of <i>A. hydrophila</i> infection	17
2.8.2 Clinical sign of <i>A. hydrophila</i> infection	17
2.8.3 Control and prevention of <i>A. hydrophila</i> infection	18
3 METATERIALS AND METHODS	
3.1 Project location	19
3.2 Spirulina cultivation and harvesting process	19

3.3	Preparation of experimental diets	20
3.4	Fish and rearing facilities	21
3.5	Fish biological growth performance	22
3.6	Proximate composition of experimental diets and catfish fillets	23
3.6.1	Determination of moisture content	23
3.6.2	Determination of ash content	23
3.6.3	Determination of crude lipid content	24
3.6.4	Determination of protein content	24
3.6.5	Determination of crude fiber content	25
3.7	Disease resistance of catfish towards bacterial infection	25
3.7.1	Catfish physical activity score	26
3.7.2	<i>A. hydrophila</i> isolation and identification	28
3.7.3	<i>A. hydrophila</i> passage in catfish	28
3.7.4	<i>A. hydrophila</i> challenge in catfish	28
3.7.5	Clinical sign and survival rate	29
3.7.6	Blood samples analysis	29
3.7.7	Histopathology process and analysis	30
3.8	Data analysis	31
4	RESULTS	
4.1	Proximate composition of spirulina	32
4.2	Proximate composition of the experimental diets	32
4.3	Biological growth performance of catfish	33
4.4	Proximate composition of the catfish fillet	34
4.5	Catfish physical activity score	35
4.6	Survival rate of catfish supplemented with different ratio of spirulina against <i>Aeromonas hydrophila</i> infection	36
4.7	Hematological profile of catfish before and after challenge by <i>Aeromonas hydrophila</i> infection	36
4.8	Histopathology changes of selected catfish organs before and after challenge by <i>Aeromonas hydrophila</i>	39
5	DISCUSSIONS	
5.1	Proximate composition of spirulina	44
5.2	Proximate composition of the experimental diets	45
5.3	Biological growth performance of catfish	46
5.4	Proximate composition of the catfish fillet	47
5.5	Catfish physical activity score	48
5.6	Survival rate of catfish supplemented with different ratio of spirulina against <i>Aeromonas hydrophila</i> infection	48
5.7	Hematological profile of catfish before and after challenge by <i>Aeromonas hydrophila</i> infection	49
5.8	Histopathology changes of selected catfish organs before and after challenge by <i>Aeromonas hydrophila</i>	50

6	SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE STUDY	52
	REFERENCES	54
	APPENDICES	68
	BIODATA OF STUDENT	103
	LIST OF PUBLICATIONS	104



LIST OF TABLES

Table		Page
3.1	Scoring guideline for the physical activeness of the catfish	27
3.2	Scoring guideline for histopathology analysis	30
4.1	The proximate composition (%) of locally grown locally grown dried powder spirulina.	32
4.2	The proximate composition (%) of the experimental diets.	33
4.3	Biological growth performances of catfish supplemented with different ratio of spirulina	33
4.4	Proximate composition of catfish fillet after 90 days of experiment	34
4.5	Catfish physical activity score during different observation times	35
4.6	Survival rate (%) of catfish supplemented with different ratio of spirulina against the <i>A. hydrophila</i> infection	36
4.7	Hematological profiles of catfish before the <i>A. hydrophila</i> infection	37
4.8	Hematological profiles of catfish after the <i>A. hydrophila</i> infection	37
4.9	Hematological profiles of catfish before and after the <i>A. hydrophila</i> infection	37
4.10	Histopathological scoring of the catfish's organ tissue before the <i>A. hydrophila</i> challenge	40
4.11	Histopathological scoring of the catfish's organ tissue after the <i>A. hydrophila</i> challenge	40
4.12	Histopathological scoring of the catfish's organ tissue before and after the <i>A. hydrophila</i> challenge	40

LIST OF FIGURES

Figure		Page
2.1	Image and classification of <i>Clarias gariepinus</i>	7
2.2	Image and the classification of spirulina <i>Arthrospira platensis</i>	14
2.3	Clinical sign of infected <i>Clarias gariepinus</i> with <i>Aeromonas hydrophila</i>	17
3.1	Arial view of Fish Breeding Center, University Agricultural Park, Universiti Putra Malaysia, UPM Serdang (2°58'57.8"N 101°42'32.8"E)	19
3.2	Experimental diets of 0%, 1%, 3%, 5% and 7% of spirulina mixed with the commercial fish feed	21
3.3	The cemented tank and the fish cage that used in this study	22
4.1	Catfish liver stained with H&E	41
4.2	Catfish kidney stained with H&E	42
4.3	Catfish spleen stained with H&E	43

LIST OF APPENDICES

Appendix		Page
A1	Proximate composition of spirulina produced from various countries	68
B1	Proximate composition of spirulina descriptive data	68
B2	Proximate composition of spirulina Turkey HSD	68
B3	Proximate composition of experimental diets descriptive data	69
B4	Proximate composition of experimental diets turkey HSD	71
B5	Biological growth performance of catfish descriptive data and Turkey HSD	72
B6	Proximate composition of catfish fillet after 90 days of experiment descriptive data	78
B7	Proximate composition of catfish fillet after 90 days of experiment Turkey HSD	80
B8	Catfish physical activeness score during different observation time descriptive data and Turkey HSD	81
B9	Survival rate of catfish supplemented with different ratio of spirulina against <i>A. hydrophila</i> infection descriptive data and Turkey HSD	83
B10	Haematological profiles of catfish before, after, before and after challenged by <i>A. hydrophila</i> descriptive data and Turkey HSD	85
B11	Histopathological score of catfish organ tissue before, after, before and after the <i>A. hydrophila</i> challenge descriptive data and Turkey HSD	95

LIST OF ABBREVIATIONS

<i>A. hydrophila</i>	<i>Aeromonas hydrophila</i>
<i>A. platensis</i>	<i>Arthrospira platensis</i>
BAU	Bangladesh Agricultural University
bc	Blood congestion
bv	Blood vessel
ca	Central artery
cv	Central vein
d	Degeneration
DPX	Distyrene, plasticizer, xylene
DOF	Department of Fisheries
et al.	Latin phrase means 'and others'
FCR	Feed conversion rate
FE	Feeding efficiency
FOI	French Oil Institute
g	Glomerulus
GSI	Gonadosomatic index
h	Hepatocytes
Hb	Hemoglobin
hyt	Hypertrophy
H&E	Hematoxylin & eosin
IPGSR	Institute of Post-graduate Studies and Research Laboratory
MCV	Mean corpuscular volume
MCHC	Mean corpuscular hemoglobin concentration
MMC	Melanomacrophage centre
n	Necrosis
PCV	Packed cell volume
RBC	Red blood cell
rc	Renal corpuscle
rp	Red pulp
SAC	Siam Algae Co. Ltd
SGR	Specific growth rate
SL	Standard length
TBI	Total body increase
tb	Trabeculae
VSI	Viscerosomatic index
wp	White pulp
WBC	White blood cell

CHAPTER 1

INTRODUCTION

1.1 Background of the study

Aquaculture industry in Malaysia has become an important activity as it increases the local production of food since the Seventh Malaysia Plan (1996-2000) and benefits the economy of the country (FAO, 2014). Aquaculture industry also has been identified as potential export earner after oil palm and rubber, and also as one of the most profitable in terms of income per hectare per annum and return to investment (FAO, 2014).

Presently, there are several species of fish cultured intensively including catfish. Catfish *Clarias* spp. recorded the highest production of cultured freshwater fish in Malaysia. The total production of this fish was approximately 50,534 tonnes, with estimated retail value of RM 314 millions (DOF, 2013). Most of this fish were cultured in freshwater ponds (48,422 tonnes; 95.8%), followed by cemented tanks (1,242 tonnes; 2.5%), canvas tanks (354 tonnes; 0.7%) and others. It is predicted that the production of catfish will increased in the future, due to the demand, high prices and farming technology (DOF, 2013).

Intensive culture of fish that fed with land based feed source has become problem to the aquaculture farming industry (Avnimalech, 2009). According to Hemarswarya et al. (2011), almost 40-60% of aquaculture production depends on the commercial feed. The land based feed source such as corn meal, soy meal and other feed additives which is non-living diets not only expensive, but the growth of the fish are slow, prone to disease and cause high mortalities compared to those fed with live diet (Ponis et al., 2003). Hence, the replacement or the substitution of the land based feed source required an alternative feeds that much cheaper and promote the growth of the farmed fish with its high nutritional value and at the same time reduce the rate of mortality.

There were studies proved that the use of aquatic origin natural food which is microalgae such as spirulina *Arthrospira platensis* as fish feed can provide nutrient to the aquatic farmed animals as it were known to have good nutritional value (Brown, 2002; Natrah et al., 2007), probiotic effects and stimulate the immunity of the cultures aquatic animals (Muller-Feuga, 2000). Numerous studies have shown that feed supplementation and improved feed formulation can alleviate the performance of the fish's growth and disease resistance, thus these provide alternative to the problems regarding to aquaculture production.

Study by Otto & Gross (2004) showed that spirulina or its extracts accelerate the development of the immune system of many animals especially during early stages of their lives. Certik & Shimizu (1999) stated the large profile of natural vitamins, minerals and essential fatty acids content of the spirulina positively improved the

immune response of the animal fed with spirulina. Other than that, the bioactive components in spirulina give the positive health effects. The immune enhancement effect through its anti-viral and anti-bacteria properties is one of the promising applications of using spirulina as feed to the reared or cultured animals.

Disease outbreaks caused by *Aeromonas hydrophila* in intensive culture of catfish has become a problem to the aquaculture industry as it affects the freshwater fish production (Salah, 2013). For example, there was a report in March 2008 on mass mortality of cultured African catfish *Clarias gariepinus* in a floating cage culture system located at Manir River, Terengganu, Malaysia, due to *A. hydrophila* infection, which caused high economic losses to the industry (Najiah et al., 2009). *Aeromonas hydrophila* were known as a species of bacterium that can be found in all freshwater environments and brackish water (FDA, 2015). The infection of *A. hydrophila* on catfish may be transmitted through the water, diseased and healthy fish and other affected vertebrates. The infection can cause sudden death of the infected host, hemorrhagic septicemia including bloated appearance, skin ulcerations and pale gills (Laith & Najiah, 2013).

Thus, this study will evaluate the utilization of local spirulina as fish feed supplement and maximized the application of its benefit as food source for cultured aquatic animal. Moreover, this study also will try to reduce the reliance on the land based feed source and the expensive fish meal. It is expected that the catfish that fed with local spirulina will improved their growth performances, nutritional value and disease resistance towards bacterial diseases.

1.2 Problem statement

Intensive aquaculture practices that have been applied to fulfill the demand of consumer on the aquaculture production have raised a lot of problems. The first is about the feed given to the cultured fish. The farmers tend to feed the catfish or hardy fishes with cheaper and low quality of feed such as the chicken offal, trash fish and carcass due to expensive commercial fish feed. There was a case in Perak, Malaysia, when freshwater fish were fed with waste from pig industries and carcass of pigs or piglets (Bernama, 2017). This situation rise an issue concerning with halal food source to Muslim in Malaysia and make the catfish less feasible to the consumer.

The second problem is the unhealthy and slow growth of the catfish. This constraint related to the low quality of “farm-made feed” given to the cultured fish. The term “farm-made feed” refer to the processed carcasses into the fish feed. Inadequate nutrient for the growth and rancid condition of the offal and carcass can cause the fish prone to disease besides the growth of the fish slow and stunned (Chinabut, 2002).

The third problem is the disease outbreak. Disease outbreak in aquaculture often related with high stoking density or the intensive farming practices due to the efficiency in transmitting the infectious diseases which become a primary limiting factor of fish

production (Leung & Bates, 2013). As for the freshwater fishes, *Aeromonas hydrophila*, *Flexibacter*, *Edwardsiella* sp. and *Pseudomonas* are the most common cause of bacterial disease (Siti Zahrah, 1992). In fact, the distributions of *A. hydrophila* are reported to be present in Malaysia also in neighboring country such as Thailand and Indonesia (OIE, 2009). In the case of catfish culture, catfish is the host to the bacterial fish pathogen *A. hydrophila* (Laith & Najiah, 2013). It is a concern that disease outbreak caused by this bacterium will cause huge profit losses as catfish were been cultured intensively and recorded as highest produced freshwater fish in Malaysia (DOF, 2015). Besides being a most common cause of bacterial disease, *A. hydrophila* is an opportunistic bacterium that can be found in various water bodies worldwide which often related with severe disease to the infected fish (Janda & Abbott, 2010). The infection of *A. hydrophila* usually will cause mass mortality of the reared fish and showed bloated appearance, visible hemorrhagic lesion, ulceration on the fish's body surface or skin also the rotting of fins and tail (Miyazaki & Jo, 1985; Wolf, 1988). Besides, the multiplication of bacteria in the intestine of the fish will cause a haemorrhagic mucous-desquamative catarrh (Aoki, 2016). The *A. hydrophila*'s toxic metabolites induce toxemia as it had been absorbed by the intestine and the stomach's submucosa experience the capillary haemorrhage (Aoki, 2016). Study that have been conducted proved that the disease outbreak by the bacterial disease often related with poor environment condition such as low dissolved oxygen, sudden change in temperature, stressors include the overcrowding and rough handling (Noga, 2010; Steven et al., 2017). Although catfish is a hardy species that can withstand stress and low water quality, as they develop a disease usually it has been too late for the remedial action as the outbreak already serious (Sayuthi, 1993). To date, disease control measure involving the usage of antibiotics or chemotherapeutics for treatment also probiotics and vaccines for preventive measures which requires better choice of disease control strategy as the demand of world market require healthy aquaculture products from farm to table (Chinabut & Puttinaowarat, 2005). The application of using chemical, drugs and antibiotics in the disease treatment rise several major issues include the impact to the environment as the release of antibiotics in large quantities, high residue levels of approved or unapproved antibiotics presence which lead to the development of bacteria that become more resistance to the developed antimicrobial drug which possess threat not only to the aquaculture but also human health. Vaccination in preventing the disease outbreak is proven to be highly effective and often applied to high economic value of fish species such as salmon. However, in developing vaccine for specific disease require time and money investment also the fish species farmed economic value need to be taken account for (Sommerset et al., 2005). The usage of probiotic in preventive measure has been used extensively in aquaculture to increase the disease resistance of the fish against the bacterial disease, appetite improvement and enhance the growth of the fish (Newaj-Fyzul et al., 2014). Hence the environmentally-friendly, highly effective and low cost alternative is required to reduce the problems in control and management of disease outbreak in the aquaculture.

1.3 Objectives

The main objectives of this study are:

1. To determine the growth performances of catfish *Clarias gariepinus* supplemented with different ration of spirulina *Arthrospira platensis*
2. To determine the proximate composition of catfish *Clarias gariepinus* supplemented with different ration of spirulina *Arthrospira platensis*
3. To determine the disease resistance of catfish *Clarias gariepinus* supplemented with different ration of spirulina against *Aeromonas hydrophila* infection

1.4 Hypothesis

The hypotheses of this study are:

1. Catfish *Clarias gariepinus* that are supplemented with spirulina *Arthrospira platensis* will have better growth performance compared to catfish that are not supplemented with spirulina
2. The proximate composition of catfish *Clarias gariepinus* supplemented with spirulina *Arthrospira platensis* will be increased compared to catfish that are not supplemented with spirulina
3. Addition of spirulina *Arthrospira platensis* in the catfish *Clarias gariepinus* diet may enhanced the disease resistance against *Aeromonas hydrophila* infection

REFERENCES

- Abdel-Tawwab M., Ahmed M. H., Abdel-Hadi Y. M., Seden M. E. A. 2008. Use of spirulina (*Arthrospira platensis*) as growth and immunity promoter for Nile tilapia, *Oreochromis niloticus* (L.) fry challenged with pathogenic *Aeromonas hydrophila*. 8th International Symposium on Tilapia in Aquaculture 2008. 1015-1032.
- Abdel-Tawwab M. & Ahmad M. H. 2009. Live Spirulina (*Arthrospira platensis*) as a growth and immunity promoter for Nile tilapia, *Oreochromis niloticus* (L.), challenged with pathogenic *Aeromonas hydrophila*. *Aquaculture Research*. 40: 1037-1046.
- Al-Dohail M. A., Hashim R., Aliyu-Paiko M. 2011. Evaluating the use of *Lactobacillus acidophilus* as a biocontrol agent against common pathogenic bacteria and the effects on the haematology parameters and histopathology in African catfish *Clarias gariepinus* juveniles. *Aquaculture Research*. 42: 196–209. doi:10.1111/j.1365-2109.2010.02606.x
- Amin N., Lung C. W., Sopian K. 2009. A practical field study of various solar cells on their performance in Malaysia. *Renewable Energy*. 34: 1939-1946.
- Amrevuawho M. O., Akinyemi A. A., Ezeri G. N. O., Bankole O. M., Takeet O. V. A. 2014. Pathological study of *Clarias gariepinus* (Burchell, 1822) sub-adult artificially infected with *Pseudomonas aeruginosa*. *Brazilian Journal of Aquaculture Science and Technology*. 18: 65-70.
- Ammar A. A., Abd-Elgawad A. S., Salama A. A. 2008. Effect of extruded and non-extruded fish pellet on growth performance and total production of Nile tilapia and grey mullet fingerlings reared in a polyculture system in earthen ponds. In 8th International Symposium on Tilapia in Aquaculture. Cairo. Egypt. 1199-1209.
- Árnason V., Bartley D., Garcia S., Haraldsson R. H., Sveinbjörnsson D., Watanabe H. 2005. Ethical issues in fisheries. *FAO ethics series*. 1-27.
- Andrews S. R., Sahu N. P., Pal A. K., Mukherjee S. C., Kumar S. 2011. Yeast extract, brewer's yeast and spirulina diets for *Labeo rohita* fingerlings affect haemato-immunological responses and survival following *Aeromonas hydrophila* challenge. *Research in Veterinary Science*. 91: 103-109.
- Annual Fisheries Statistic, Department of Fisheries, Malaysia (DOF). 2013. Department of Fisheries, Ministry of Agriculture and Agro-Based Industry, Malaysia.
- Al Mahrouqi H., Naqqiuddin M. A., Achankunju J., Omar H., Ismail A. 2015. Different salinity effects on the mass cultivation of spirulina (*Arthrospira platensis*) under sheltered outdoor conditions in Oman and Malaysia. *Journal of Algal Biomass Utilization*. 6: 1-14.
- Anon. 2004. Annual Fisheries Statistics Volume 1. 2003. Department of Fisheries Malaysia. Ministry of Agriculture and Agro-Based Industry. 1: 217-218.

- Ang K. J., Gopinath N. Chua T. E. 1989. The status of introduced fish species in Malaysia, in de Silva S. S. (ed.) Exotic Aquatic Organism in Asia. Asian Fisheries Society Special Publication. 3: 71-82.
- Alain K. Isolation of *Aeromonas hydrophila* from naturally diseased Thai *Pangasius hypophthalmus*. MSc thesis, Bangladesh Agricultural University, Mymensingh, Bangladesh, 2009.
- Austin B. and Austin D. 1987. Bacterial Fish Pathogen. Ellis Horwood. London. 250-262.
- Cipriano R. C., Bullock G. L., Pyle S. W. 1984. *Aeromonas hydrophila* and motile aeromonad septicemia of fish. Fish and Wildlife Service. West Virginia. Fish Diseases Leaflet. 68.
- Aoki T. 2016. Fish Disease. London. United Kingdom. Eolss Publishers. 72-74.
- AOAC. 1990. Official method of analysis of the association of official analytical chemists. No. 934.06, AOAC, Arlington.
- Aqius C, & Roberts R. J. 2003. Melano-macrophage centres and their role in fish pathology. Journal of Fish Disease. 26: 499-509.
- Avnimalech Y. 2009. Biofloc technology – A practical guide book. The World Aquaculture Society. Baton Rouge. Louisiana. United States. 19-21.
- Becker E. W. 1994. Microalgae biotechnology and microbiology. Cambridge, U.K. Cambridge University Press. 293.
- Becker W. 2004. Microalgae in human and animal nutrition, In Richmond, A. (ed.), Handbook of microalga; culture. Blackwell. Oxford. 312-351.
- Becker E. W. 2007. Micro-algae as a source of protein. Biotechnology Advances. 25: 207-210.
- Becker E. W. & Venkataraman L. V. 1984. Production and utilization of blue-green alga, Spirulina in India. Biomass. 4: 105–125.
- Belay A., Kato T., Ota Y. 1996. Spirulina (*Arthrospira*): potential application as an animal feed supplement. Journal of Applied Phycology. 8(4): 303-311.
- Bhowmik D., Dubey J., Mehra S. 2009. Probiotic efficiency of *Spirulina platensis*-stimulating growth of lactic acid bacteria. Journal of Agriculture & Environmental Science. 6: 546-549.
- Brown M. R. 2002. Nutritional value and use of microalgae in aquaculture. Avances en Nutrición Acuícola VI. Memorias del VI Simposium Internacional de Nutrición Acuícola. 3: 281-292.
- Brown M. R., Jeffrey S. W., Volkman J. K., Dunstan G. A. 1997. Nutritional properties of microalgae for mariculture. Aquaculture. 151: 315-331.

- Brusle J. & Anadon G. G. 1996. The structure and function of fish liver. In: Fish Morphology. Brookfield. Science Publishers. 77-93.
- Bernama. 2017. Fish at Perak farm being fed pig flesh, sold at wholesale markets. New Straits Times. Retrieved on April, 2017 from <http://www.nst.com.my/news/2017/03/219240/fish-perak-farm-being-fed-pig-flesh-sold-wholesale-markets>
- Castro R. & Tafalla C. 2015. Overview of fish immunity. Mucosal Health in Aquaculture. Waltham. Academic Press. 3-54.
- Certik M. & Shimizu S. (1999). Biosynthesis and regulation of microbial polyunsaturated fatty acid and production. Journal of Bioscience and Bioengineering. 87: 1-14.
- Cencic A. & Chingwaru W. 2010. The role of functional foods, nutraceuticals, and food supplements in intestinal health. Nutrients. 2: 611-625.
- Cipriano R. C., Bullock G. L., Pyle S. W. 1984. *Aeromonas hydrophila* and motile aeromonad septicemias of fish. US Fish & Wildlife Publications. 134.
- Chinabut S. 2002. Jaundice disease in catfish, a case study demonstrating a decline in incidence as a result of research output. In: Arthur J. R., Phillips M. J. Subasinghe R. P. Reantaso M. B. and MacRae, I. H. (eds.) Primary Aquatic Animal Health Care in Rural, Small-scale, Aquaculture Development. FAO Fish. 77-80.
- Chinabut S. & Puttinaowarat S. 2005. The choice of disease control strategies to secure international market access for aquaculture products. Developmental Biology. 121: 255-261.
- Collins S. A., Desai A. R., Mansfield G. S., Hill J. E., Van Kessel A. G., Drew M. D. 2012. The effect of increasing inclusion rates of soybean, pea and canola meals and their protein concentrates on the growth of rainbow trout: concept in diet formulation and experimental design for ingredient evaluation. Aquaculture. 344-349: 90-99.
- Cowey C. B., & Sargent J. R. 1977. Lipid nutrition in fish. Comparative Biochemical Physiology. 57: 269-73.
- Cheng H. T. 2016. Photo 6074874. iNaturalist.org. Retrieved from <http://www.inaturalist.org/photos/6074874>
- Conceicao L. E. C., Yufera M., Makridis P., Morais S., Dinis M. T. 2010. Live feeds for early stages of fish rearing. Aquaculture Research. 41: 613-640.
- Dabrowski K. & Guderley H. 2002. Intermediary metabolism. In Halver, J.E. (Eds.), London. Academic Press. 309-365.
- Davis J. M. 1985. The role of dietary fiber in fish nutrition. Recent Advances in Aquaculture. 2:219-249. doi: 10.1007/978-1-4684-8736-7_6
- Delzenne N. M. 2003. Oligosaccharides: state of the art. Proceedings of the Nutrition Society. 62: 177-182.

- Devresse B., Dehasque M., Van Assche J., Merchie G. 1997. Nutrition and health. In Tacon A. & Basurco B. (eds). Feeding tomorrow's fish. 35-66. Proceedings of the workshop of the CIHEAM Network on Technology of Aquaculture in the Mediterranean (TECAM), jointly organized by CIHEAM, FAO and IEO Mazarron, Spain, 24-26 June 1996. CIHEAM. Apodo. Spain.
- De Schryver P., Crab R., Defoirdt T., Boon N., Verstraete W. 2008. The basics of bio-flocs technology: The added value for aquaculture. *Aquaculture*. 277:125-137.
- de Oliveira M. A. C. L., Monteiro M. P. C., Robbs P. G., Leite S. G. F. 1999. Growth and chemical composition of *Spirulina maxima* and *Spirulina platensis* biomass at different temperatures. *Aquaculture International*. 7: 261-275.
- De la Higuera M. 2001. Effect of nutritional factors and feed characteristics on feed intake. In: Food Intake in Fish. London. Blackwell Science. 250-268.
- Duncan P. L. & Klesius P. H. 1996. Effects of feeding *Spirulina* on specific and non-specific immune responses of channel catfish. *Journal of Aquatic Animal Health*. 8: 308-313.
- DOF. 2015. National plan of action for the management of fishing capacity in Malaysia (Plan 2). Retrieved on January 2017 from http://www.dof.gov.my/dof2/resources/user_1/UploadFile/Penerbitan/Senarai%20Penerbitan/NPOA.pdf
- DOF. 2016. Malaysia fishing industry scenario. Official website Ministry of Agriculture & Agro-Based Industry Malaysia. Retrieved on May from <http://www.dof.gov.my/index.php/pages/view/42>
- Do-Hyung K. 2015. Low-value fish used as feed is a source of disease in farmed fish. *Fish Aquaculture Science*. 18: 203-209.
- FAO. 2008. National Aquaculture Sector Overview. Malaysia. National Aquaculture Sector Overview Fact Sheets. Text by Mazuki Hashim. In: FAO Fisheries and Aquaculture Department [online]. Rome. Updated 2008. Retrieved on February 2015 from http://www.fao.org/fishery/countrysector/naso_malaysia/en
- El-Sayed A. F. M. 1999. Alternative dietary protein sources for farmed tilapia, *Oreochromis* spp. *Aquaculture*. 179: 149-168.
- El-Sheekh M., El-Shourbagy I., Shalaby S., Hosny S. 2014. Effect of feeding *Arthrospira platensis* (spirulina) on growth and carcass composition of hybrid tilapia (*Oreochromis niloticus* x *Oreochromis mossambicus*). *Turkish Journal of Fisheries & Aquatic Sciences*. 14: 471-478.
- Engering A., Hogerwerf L., Slingenbergh J. 2013. Pathogen-host-environment interplay and disease emergence. *Emerging Microbes & Infections*. 2: 1-6.
- Erhunmwunse N. O. & Ainerua M. O. 2013. Characterization of some blood parameters of African catfish (*Clarias gariepinus*). *American-Eurasian Journal of Toxicological Sciences*. 5: 72-76. doi: 10.5829/idosi.ajejts.2013.5.3.82159
- FAO. 2014. National Aquaculture Sector Overview. FAO. Malaysia.

- FAO.1981. A handbook of diseases of cultured *Clarias* (Pla Duk) in Thailand. Fisheries and Aquaculture Department. Retrieved on October, 2017 from <http://www.fao.org/docrep/field/003/AB942E/AB942E03.htm>
- FDA. 2015. Foodborne Pathogenic Microorganisms and Natural Toxins Handbook *Aeromonas hydrophila*. U.S. Food and Drug Administration. Retrieved on October, 2017 from <http://www.fda.gov/Food/FoodborneIllnessContaminants/ CausesOf IllnessBadBugBook/ucm070523.htm>
- Fedor K. 2011. *Arthrospira platensis*. Bio203 University of Wisconsin-La Crosse. Web. Retrieved on October 2011 from http://bio web.uwlax.edu/bio203 /2011/fedor_kara/
- Freyhof J., Geelhand D. 2016. *Clarias gariepinus*. The IUCN Red List of Threatened Species 2016: e.T166023A84198891. Downloaded on 20 June 2017.
- Gatlin D. M., Barrows F. T., Brown P., Dabrowski K., Gaylord T. G., Hardy, R. W., Herman E., Hu G., Krogdahl A., Nelson R., Overturf K., Rust M., Sealey W., Skonberg D., Souza E. J., Stone D., Wilson R., Wurtele E. 2007. Expanding the utilization of sustainable plant products in aquafeeds: a review. *Aquaculture Research*. 38: 551–579.
- Gatesoupe F. J. 2005. Probiotics and prebiotics for fish culture, at the parting of the ways. *Aqua Feeds: Formulation and Beyond (Feedware)*. 2: 3–5.
- Gislason H., Daan N., Rice J. C., Pope J. G. 2010. Size, growth, temperature and the natural mortality of marine fish. *Fish and Fisheries*. 11: 149-158.
- Gutierrez-Wing M. T. & Malone R. F. 2006. Biological filters in aquaculture: trends and research directions for freshwater and marine applications. *Aquaculture Engineering*. 34: 163-171.
- Glahn J. F., Werner S. J., Hanson T., Engle C. R. 2000. Cormorant depredation losses and their prevention at catfish farms: economic considerations. *Human Conflicts with Wildlife: Economic Considerations*. 17: 138-146.
- Harel M., Koven W., Lein I., Bar Y., Beherens P., Stubblefield J., Zohar Y., Place A. R. 2002. Advanced DHA, EPA and ArA enrichment materials for marine aquaculture using single cell heterotrophs. *Aquaculture*. 213: 347–362. doi: 10.1016/S0044-8486(02)00047-9
- Handeland S. O., Imsland A. K., Stefansson S. O. 2008. The effect of temperature and fish size on growth, feed intake, food conversion efficiency and stomach evacuation rate of Atlantic salmon post-smolts. *Aquaculture*. 283: 36-42.
- Hanapi S. Z., Hassan M. A., Sarip S. H. M., Aziz R., Sarmidi, M. R., Imranuddin S., Ali S. 2008. Animal Feed. *Biotechnology Development in Agriculture, Industry and Health*. 422-445. Retrieved on June, 2017 from https://www.researchgate.net/profile/Siti_Zulaiha_Hanapi/publication/3164638_A nimal_Feed/links/58ff9f820f7e9bcf65454192/Animal-Feed.pdf

- Harikrishnan R. & Balasundaram C. 2005. Modern trends in *Aeromonas hydrophila* disease management with fish. *Reviews in Fisheries Science & Aquaculture*. 13: 281-320.
- Hasan M. R. 2000. Nutrition and feeding for sustainable aquaculture development in the third millennium. Technical proceedings of the conference on aquaculture in the third millennium. Bangkok. Thailand. 20-25.
- Habib M. A. B., Parvin M., Huntington T.C., Hasan M. R. 2008. A review on culture, production and use of spirulina as food for humans and feeds for domestic animals. *FAO Fisheries and Aquaculture Circular*. 1034: 1-33.
- Plumb J. A. & Hanson L. A. 2011. Health maintenance and principal microbial diseases of cultured fishes. Iowa. Iowa State University Press. 57-66.
- Hardy R. W. & Lee C. S. 2010. Aquaculture Feed and Seafood Quality. *Bulletin of Fisheries Research Agency*. 31: 43-50.
- Hecht T. & Jones C. L. W. 2009. Use of wild fish and other aquatic organisms as feed in aquaculture – a review of practices and implications in Africa and the Near East. In Hasan M. R. & Halwart M. (eds). *Fish as feed inputs for aquaculture: practices, sustainability and implications*. FAO Fisheries and Aquaculture Technical Paper. No. 518. Rome, FAO. 129–157.
- Hemre G. I., Mommsen T.P., Kroghdahl A. 2002. Carbohydrates in fish nutrition: effects on growth, glucose metabolism and hepatic enzymes. *Aquaculture Nutrition*. 8:175–194.
- Hemarswarya S., Raja R., Ravi Kumar R., Ganesan V., Anbazhagan C. 2011. Microalgae: a sustainable feed source for aquaculture. *World Journal Microbial Biotechnology*. 27: 1737-1746.
- Howaida R. G., Hanlon T. H., Mahmoud H. H., Eletreby S.G. 1998. Maturation, fecundity, seasonality and reproduction of two commercially valuable cuttlefish, *Sephia pharaonis* and *S. dollfusi* in Suez Canal. *Journal of Fisheries Research*. 36: 99-115.
- Hussain I. A., Jeyasekaran G., Shakila R. J., Raj K. T., Jeevithan E. 2014. Detection of hemolytic strains of *Aeromonas hydrophila* and *A. sobria* along with other *Aeromonas* spp. from fish and fishery products by multiplex PCR. *Journal of Food Science and Technology*. 51:401-407.
- Huntington T. C. & Hasan M. R. 2009. Fish as feed inputs for aquaculture – practices, sustainability and implications: a global synthesis. In Hasan M. R. & Halwart M. (eds). *Fish as feed inputs for aquaculture: practices, sustainability and implications*. FAO Fisheries and Aquaculture Technical Paper. No. 518. Rome. FAO. 1–61.
- Ighwela K. A., Ahmad A., Abol-Munafi A. B. 2014. The selection of viserosomatic and hepatosomatic indices for the measurement and analysis *Oreochromis niloticus* condition fed with varying dietary maltose levels. *International Journal of Fauna and Biological Studies*. 1: 18-20.

- Ismail N. I. A., Amal M. N. A., Shohaimi S., Zamri-Saad M., Siti-Zahrah A. 2016. Associations of water quality and bacteria presence in cage cultured red hybrid tilapia, *Oreochromis niloticus* × *O. mossambicus*. 2016 Aquaculture Reports. 4:57–65.
- Jan Mazurkiewicz. 2009. Utilization of domestic plant components in diets for common carp *Cyprinus carpio* L. Archives of Polish Fisheries. 17:5-39.
- Janda J. M. & Abbott S. L. 2010. The genus *Aeromonas*: taxonomy, pathogenicity and infection. Clinical Microbiology Reviews. 23: 35-73. doi: 10.1128/CMR.00039-09
- James R., Sampath K., Thangarathinam R., Vasudevan I. 2016. Effect of dietary spirulina level on growth, fertility, coloration and leucocyte count in red swordtail, *Xiphophorus helleri*. The Israeli Journal of Aquaculture. 58: 97-104.
- Johari T. & Jamaludin I. 2015. Current status of aquaculture in Malaysia. DOF. Retrieved from <http://www.aquaculture.asia/files/soyabean/pdfs/09.%20Status%20of%20Aquaculture%20in%20Malaysia%20Summary.pdf>
- Kaneko J. J. 1989. Carbohydrate metabolism and its diseases. Clinical Biochemistry of Domestic Animals. 4: 44-81.
- Kay R. A. & Barton L. L. 2009. Microalgae as food supplement. Critical Review in Food Science and Nutrition. 30: 555-573.
- Kabata Z. 1985. Parasites and disease of fish cultured in tropics. London. Taylor and Francis. 120-125.
- Kechik I. A. 1995. Aquaculture in Malaysia. In Bagarinao T. U. & Flores E. E. C (Eds.), Towards Sustainable Aquaculture in Southeast Asia and Japan: Proceedings of the Seminar-Workshop on Aquaculture Development in Southeast Asia, Iloilo City, Philippines, 26- 28 July, 1994 (125-135). Tigbauan, Iloilo, Philippines: SEAFDEC Aquaculture Department.
- Kulshreshtha A., Jarouliya U., Bhadauriya P., Prasad G. B. K. S., Bisen P. S. 2008. Spirulina in health care management. Current Pharmaceutical Biotechnology. 9: 400-405.
- Khan Z., Bhadouria P., Bisen P. S. 2005. Nutritional and therapeutic potential of spirulina. Current Pharmaceutical Biotechnology. 6: 373-379.
- Kiron V. 2012. Fish immune system and its nutritional modulation for preventive health care. Animal Feed Science and Technology. 173: 111-133.
- Kostromina V. P., Derkach O. V., Symonenkova N. V., Riechkina O. O., Otroshchenko A. O. 2002. Evaluation of the efficacy of a plant adaptogen (spirulina) in the pathogenic therapy of primary tuberculosis in children. Likars'ka Sprava. 5:102-105.

- Kpogue D., Gangbazo H., Fiogbe F. 2013. A preliminary study on the dietary protein requirement of *Parachanna obscura* (Gunther, 1861) larvae. Turkish Journal of Fisheries and Aquatic Sciences. 13: 111-117.
- Laith A. R. & Najiah M. 2013. *Aeromonas hydrophila*: antimicrobial susceptibility and histopathology of isolates from diseased catfish, *Clarias gariepinus* (Burchell). Journal of Aquaculture Research and Development. 5: 1-7.
- Leung T. L. F. & Bates A. E. 2013. More rapid and severe disease outbreak for aquaculture at the tropics: implications for food security. Journal of Applied Ecology. 50: 215-222.
- Lee S. Y., Yin Z., Ge R., Sin Y. M. 1997. Isolation and characterization of fish *Aeromonas hydrophila* adhesions important for in vitro epithelial cell invasion. Journal of fish diseases. 20: 169-175.
- Li X. F., Wang Y., Liu W. B., Jiang G. Z., Zhu J. 2013. Effects of dietary carbohydrate/lipid ratios on growth performance, body composition and 51 glucose metabolism of fingerling blunt snout bream (*Megalobrama amblycephala*). Aquaculture Nutrition. 2: 1-12.
- Martins C. I. M., Hillen B., Schrama J. W., Verreth J. A. J. 2008. A brief note on the relationship between residual feed intake and aggression behaviour in juveniles of African catfish *Clarias gariepinus*. Applied Animal Behaviour Science. 111: 408-413.
- Mata T. M., Martins A. A., Caetano N. S. 2010. Microalgae for biodiesel production and other application: A review. Renewable and Sustainable Energy Review. 14: 217-232.
- Manoj C. K., Nair C. M., Patel M. B., Salin K. R. 2010. Haematobiochemical and histopathological changes in *Labeo rohita* infected with *Aeromonas hydrophila* by immersion challenge. Fishery technology. Society of Fisheries Technologists (India). 47: 151-160.
- Masuda T. & Goldsmith P. D. 2009. World soybean production: area harvested, yield, and long-term projections. International Food and Agribusiness Management Review. 12: 143-162.
- Miyazaki T. & Jo Y. 1985. A histopathological study of motile aeromonad disease in ayu. Fish Pathology. 20: 55-59.
- Muller-Feuga A. 2000. The role of microalgae in aquaculture: situation and trends. Journal of Applied Phycology. 12: 527-534.
- Mohan I.K., Khan M., Shobha J.C., Naidu M.U., Prayag A., Kuppusamy P. 2006. Protection against cisplatin-induced nephrotoxicity by *Spirulina* in rats. Cancer Chemotherapy and Pharmacology. 58: 802-808.
- Mohideen M. & Haniffa M. A. 2015. Effect of probiotic on microbiological and haematological responsiveness of catfish (*Heteropneustes fossilis*) challenged with

- bacteria *Aeromonas hydrophila* and fungi *Aphanomyces invadans*. Journal of Aquaculture Research Development. 6: 384-390. doi:10.4172/2155-9546.1000384
- Nakono T., Yamaguchi T., Sato M., Iwama G. K., 2003. Biological Effects of carotenoids in fish, Effective Utilization of Marine Food Resource, Songkhla, Thailand. Aquaculture. 1-15.
- Nandeesh M. C., Gangadhara B., Manissery J. K., Venkataraman L. V. 2001. Growth performance of two Indian major carps, catla (*Catla catla*) and rohu (*Labeo rohita*) fed diets containing different levels of *Spirulina platensis*. Bioresource Technology. 80: 117-120.
- Nandeesh M. C., Gangadhara B., Varghese T. J., Keshavanath P. 1998. Effect of feeding *Spirulina platensis* on the growth, proximate composition and organoleptic quality of common carp, *Cyprinus carpio* L. Aquaculture Research. 29: 305-312.
- Najiah M., Lee S. W., Wendy W., Nadirah M. 2009. Bacterial disease outbreak of African Catfish (*Clarias gariepinus*) from Manir River, Terengganu, Malaysia. Journal of Life Sciences. 3: 10-13.
- Natrah F. M. I., Yusoff F. M., Shariff M., Abas F., Mariana N. S. 2007. Screening of Malaysian indigenous microalgae for antioxidant properties and nutritional value. Journal of Applied Phycology. 19: 711-718.
- Nadia A. A. E. G., Nahla R. E. K., Salama S. S. A. 2014. Causes of mortality in Discus fish (*Symphysodon*) and trials for treatment. Egyptian Journal of Aquaculture. 2: 1-12.
- Naylor R. L., Hardy R. W., Bureau D. P., Chiu A., Elliott M., Farrell A. P., Nichols P. D. 2009. Feeding aquaculture in an era of finite resources. Proceedings of the National Academy of Sciences. 106: 15103-15110.
- Newaj-Fyzul A., Al-Hari A. H., Austin B. 2014. Review: Developments in the use of probiotics for disease control in aquaculture. Aquaculture. 431: 1-11.
- Noga E. J. 2010. Fish disease: Diagnosis and treatment. Iowa. Blackwell Publishing. 195-197.
- Nurul Izzah Ahmad, Wan Rozita Wan Mahiyuddin, Tengku Rozaina Tengku Mohamad, Cheong Yoon Ling, Siti Fatimah Daud, Nasriyah Che Hussein, Nor Aini Abdullah, Rafiza Shaharudin & Lokman Hakim Sulaiman. 2016. Fish consumption pattern among adults of different ethnics in Peninsular Malaysia, Food & Nutrition Research. 60: 1. doi: <http://dx.doi.org/10.3402/fnr.v60.32697>
- Nkhoma P. 2016. Feed remains a challenge in the aquaculture industry. The Nation. Retrieved on June, 2017 from <http://mwnation.com/feed-remains-a-challenge-in-the-aquaculture-industry/>
- Ommes M. H., Goasduff J. L., Delliou H. L., Bayon N. L., Quazuguel P., Robin J. H. 2017. Effect of dietary tannin on growth, feed utilization and digestibility and carcass composition in juvenile European seabass (*Dicentrarchus labrax* L.). Aquaculture Reports. 6: 21-27.

- Otto P. & Gross W. 2004. Valuable products from biotechnology of microalgae. *Application Microbiology Biotechnology*. 65: 635-648.
- Olguin E. J., Galicia S., Angulo-Guerrero O., Hernandez E. 2001. The effect of low light reflux and nitrogen deficiency on the chemical composition of *Spirulina* sp. (*Arthrospira*) grown on digested pig waste. *Bioresource Techology*. 77: 19-24.
- Oluyinka A. A., Funmilola A., Richards F. O. 2015. Nutrient utilization and growth of *Clarias gariepinus* fed four different commercial feeds. *International Journal of Fisheries Aquaculture*. 7: 107-110. doi: 10.5897/IJFA15.0486
- OIE. 2009. World animal health information database. World Organisation for Animal Health. Retrieved on October, 2017 from <http://www.oie.int>
- Oliva-Teles A. 2012. Nutrition and health of aquaculture fish. *Journal of Fish Disease*. 35: 83-108.
- Palmegiano G. B., Agradi E., Forneris G., Gail F., Gasco L., Rigamonti E., Sicuro B., Zoccarato I. 2005. Spirulina as a nutrient source in diets for growing sturgeon (*Acipenser baeri*). *Aquaculture Research*. 36: 188-195.
- Pecquerie L., Petitgas P., Kooijman S. A. 2009. Modeling fish growth and reproduction in the context of the Dynamic Energy Budget theory to predict environmental impact on anchovy spawning duration. *Journal of Sea Research*. 62: 93-105.
- Pienaar U. 1968. The Freshwater Fishes of the Kruger National Park. Republic of South Africa: The National Parks Board of Trustees of the Republic of South Africa. 82.
- Ponis E., Robert R., Parisi G. 2003. Nutritional value of fresh and concentrated algal diets for larval and juvenile Pacific oyster (*Crassostrea gigas*). *Aquaculture*. 221: 491-505.
- Popovic T. N., Strunjak- Perovic I., Coz- Rakovac R., Barisic J., Jadan M., Persin Berakovic A., Klobucar S. R. 2012. Tricaine methane- sulfonate (MS- 222) application in fish anaesthesia. *Journal of Applied Ichthyology*. 28: 553-564.
- Promya J. & Chitmanant C. 2011. The effects of *Spirulina platensis* and *Cladophora* Algae on the growth performance, meat quality and immunity stimulating capacity of the African Sharptooth Catfish (*Clarias gariepinus*). *International Journal of Agriculture & Biology*. 13: 77-82.
- Phang S.M., Miah M.S., Chu W.L., Hashim M. 2000. Spirulina culture in digested sago starch factory waste water. *Journal of Applied Phycology*. 12: 395-400.
- Raduan M. M. A., Sharir M., Ismail. A., Mazlan. M., Hanafi H. 2011. Perkembangan perusahaan perikanan di Semenanjung Malaysia; Isu dan persoalan. *Jati*. 16: 265-299.
- Radhakrishnan S., Saravana Bhavan P., Seenivasan C., Shanthi R., Muralisankar T. 2014. Replacement of fishmeal with *Spirulina platensis*, *Chlorella vulgaris* and

Azolla pinnata on non-enzymatic and enzymatic antioxidant activities of *Macrobrachium rosenbergii*. Journal of Basic and Applied Zoology. 67: 25-33.

- Rani Abro. 2014. Digestion and metabolism of carbohydrates in fish (Doctoral's thesis). Retrieved from http://pub.epsilon.slu.se/10974/1/abro_r_140113.pdf
- Ravi M., De S. L., Azharuddin S., Paul S. F. D. 2010. The beneficial effects of spirulina focusing on its immunomodulatory and antioxidant properties. Nutrition and Dietary Supplements.2010: 73-83.
- Ringo E., Olsen R. E., Gifstad T. O., Dalmo R. A., Amlund H., Hemre G. I., Bakke A. M. 2010. Prebiotics in aquaculture: a review. Aquaculture Nutrition. 16: 117-136.
- Read P. & Fernandes T. 2003. Management of environmental impacts of marine aquaculture in Europe. Aquaculture. 226: 139-163.
- Reichley S. R., Ware C., Khoo L. H., Greenway T. E., Wise D. J., Bosworth B. G., Lawrence M., Griffin M. J. 2017. Comparative susceptibility of channel catfish *Ictalurus punctatus*; blue catfish, *Ictalurus furcatus*; and channel (♀) x blue (♂) hybrid catfish to *Edwardsiella piscicida*, *Edwardsiella tarda* and *Edwardsiella anguillarum*. Journal of the World Aquaculture Society. 4: 178-191. doi:10.1111/jwas.12467
- Robinson E. H., Li M. H., Manning B. B. 2001. A practical guide to nutrition, feeds, and feeding of catfish. (2nd revision). Mississippi Agricultural & Forestry Experiment Station. Bulletin 1113. 5-10.
- Robinson E. H. & Li M. H. 2007. *Catfish protein nutrition: revised*. Mississippi Agricultural & Forestry Experiment Station Bulletin. Mississippi State University. 1159: 18.
- Rust M. B., Barrows F. T., Hardy R. W., Lazur A., Naughten K., Silverstein J. 2011. The future of aquafeeds. NOAA Technical Memorandum NMFS F/SPO-124, 1-94. Retrieved on May 2016 from http://www.nmfs.noaa.gov/aquaculture/docs/feeds/the_future_of_aquafeeds_final.pdf
- Sayed A. H. & Fawzy A. M. 2014. Effect of dietary supplementation of *Spirulina platensis* on the growth and haematology of the catfish *Clarias gariepinus*. Journal of Advances in Biology. 5: 625-635.
- Sayuthi S. 1993. Fish disease in Malaysia: status and problems. Proceedings of the Aquaculture Workshop for SEAFDEC/AQD Training Alumni. 164-173.
- Salah M. A. 2013. A review of fish disease in the Egyptian aquaculture sector. Research Program on Livestock and Fish. 1-41. Retrieved on October, 2017 from <https://cgspace.cgiar.org/bitstream/handle/10568/34870/EgyptAquacultureSectorDiseaseReview.pdf?sequence=1>
- Saleema M. 2015. Molecular characterization of *Aeromonas hydrophila* and development of recombinant cells vaccine expressing outer membrane proteins against its in African catfish (*Clarias gariepinus* Burchell). Thesis, Master of

Science in Aquaculture, Department of Aquaculture, Faculty of Science, Universiti Putra Malaysia, Serdang, Selangor, Malaysia.

- Spolaore P., Joannis-Cassan C., Duran E., Isambert A. 2006. Commercial application of microalgae. *Journal of Bioscience and Bioengineering*. 2: 87-97.
- Soletto D., Binaghi L., Lodi A., Carvalho J. C. M., Converti A. 2005. Batch and fed-batch cultivations of *Spirulina platensis* using ammonium sulphate and urea as nitrogen sources. *Aquaculture*. 243: 217-224.
- Sommerset I., Krossoy B., Biering E., Frost P. 2005. Vaccines for fish in aquaculture. *Expert Review of Vaccines*. 4: 89-101.
- Craig S. & Helfrich L. A. 2002. Understanding fish nutrition, feeds, and feeding. Virginia cooperative extension. 63:256-270.
- Siti Zahrah Abdullah. 1992. Common bacterial diseases of catfish, *Clarias macrocephalus*, cultured in Malacca. Proceedings of National IRPA Seminar. 2: 1-14.
- Spolaore P., Joannis-Cassan C., Duran E., Isambert A. 2006. Commercial applications of microalgae. *Journal of Bioscience and Bioengineering*. 101: 87-96.
- Spinelli J. 2015. Unconventional feed ingredients for fish feed. In: FAO Corporate Document Repository – Aquaculture development and coordination programme. Retrieved on April, 2015 from <http://www.fao.org/docrep/x5738e/x5738e0d.htm>
- Sharma V. & Dunkwal V. 2012. Development of spirulina based biscuits: a potential method of value addition. *Studies on Ethno-Medicine*. 6: 31-34.
- Shamsun Nahar., Mohammad Mizanur Rahman., Gias Uddin Ahmed., Md. Ali Reza Faruk. 2016. Isolation, identification and characterization of *Aeromonas hydrophila* from juvenile farmed pangasius (*Pangasianodon hypophthalmus*). *International Journal of Fisheries and Aquatic Studies*. 4: 52-60.
- Sneh G., Charu G., Amar P. G., Prakash D. 2017. Prebiotic efficiency of blue green algae on probiotic microorganisms. *Journal of Microbiology and Experimentation*. 4: 1-4.
- Sudheesh P. S., Al-Ghabshi A., Al-Mazrooei N., Al-Habsi S. 2012. Comparative pathogenomics of bacteria causing infectious diseases in fish. *International Journal of Evolutionary Biology*. 2012: 1-16. doi: 10.1155/2012/457264
- Suprpto H., Sumartiwi L., Prawesthirini S., Handiyatno D., Azmijah A. 2005. The isolation of *Aeromonas hydrophila* and *Escherichia coli* from Lou Ham *Cichlasoma synspilum* and studies of their histopathology changes. *Journal of Biological Research*. 10: 139-141.
- Swann L. & White, M. R. 1991. Diagnosis and treatment of "*Aeromonas Hydrophila*" infection of fish. *Aquaculture Extension*. Illinois-Indiana Sea Grant Program. 231-237.

- Stevens D. L., Bisno A. L., Chambers H. F., Everett E. D., Dellinger P., Goldstein E. J., Gorbach S. L., Hirschmann J. V., Kaplan S. L., Montoya J. G., Wade J. C. 2005. Practice guidelines for the diagnosis and management of skin and soft-tissue infections. *Clinical Infectious Diseases*. 41: 1373-1406.
- Steven C. H., Croft D. P., Paull G. C., Tyler C. R. 2017. Stress and welfare in ornamental fishes: what can be learned from aquaculture. *Journal of Fish Biology*. 19: 409-428.
- Sirakov I., Velichkova K., Nikolov G. 2012. The effect of algae meal (*Spirulina*) on the growth performance and carcass parameters of rainbow trout (*Oncorhynchus mykiss*). *Journal of Bioscience Biotechnology*. 151-156.
- Teimouri M., Amirkolaie A. K., Yeganeh S. 2013. The effects of *Spirulina platensis* meal as feed supplement on growth performance and pigmentation of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*. 396: 14-19.
- Teugels G. 1986. A systematic revision of the African species of the genus *Clarias* (Pisces: Clariidae). *Annales Musee Royal de l'Afrique Centrale*. 247: 1-199.
- Thalathiah S. & Ibrahim T. 1992. Seed production of *Clarias gariepinus* (Burchell). In: Ho. Y.W. et al (eds.). *Proceedings of the National Intensification of Research in Priority Areas Seminar*, Kuala Lumpur. 2: 239-240.
- Thalathiah S. 1986. Induced spawning of *Clarias macrocephalus* (Gunther). In: Maclean, J.L., L.B. Dizon, L.V. Hosillos (eds.), *The First Asian Fisheries Forum*, Manila, Philippines, 26-31 May, 1986. *Asian Fisheries Society*. 683-688.
- Tokusoglu O. & Unal M. K. 2003. Biomass nutrient profiles of three microalgae: *Spirulina platensis*, *Chlorella vulgaris*, and *Isochrysis galbana*. *Journal of Food Science*. 68: 1144-1148.
- Tongsiri S., Mang-Amphan A., Peerapornpisal Y. 2010. Effect of replacing fishmeal with spirulina on growth, carcass composition and pigment of the Mekong giant catfish. *Asian Journal of Agricultural Sciences*. 2: 106-110.
- Vonshak A. 1997. *Spirulina platensis* (*Arthospira*): Physiology, Cell Biology and Biotechnology. Bristol. Taylor & Francis. 341-346.
- Watanuki H., Ota K., Malin A. C., Tassakka A. R., Kato T., Sakai M. 2006. Immunostimulant effects of dietary *Spirulina platensis* on carp, *Cyprinus carpio*. *Aquaculture*. 258: 157-163.
- Watanabe T. 2002. Strategies for further development of aquatic feeds. *Fisheries Science*. 68: 242-252.
- Watters C., Iwamura S., Ako H., Dong-Fang D. 2012. Nutrition considerations in aquaculture: The importance of omega-3 fatty acids in fish development and human health. *Foods and Nutrition*. 1: 1-7.

- Wu G., Saoud I.P., Miller C., Davis D.A. 2004. The effect of feeding regimen on mixed-size pond-grown channel catfish, *Ictalurus punctatus*. Journal of Applied Aquaculture. 15: 115–125.
- Wu J. L., Lin H. M., Hsu Y. L., Chang L. H. 1981. Biological control of fish bacterial pathogen, *Aeromonas hydrophila*, by bacteriophage AH1. Fish Pathology. 15: 271-276.
- Wang L., Pan B., Sheng J., Xu J., Hu Q. 2007. Antioxidant activity of *Spirulina platensis* extracts by supercritical carbon dioxide extraction. Food Chemistry. 105: 36–41.
- Wolf K. 1988. Fish viruses and fish viral diseases. Ithaca. New York. Cornell University Press. 475-476.
- Wiegand I., Hilpert K., Hancock R. E. 2008. Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. Nature Protocols. 3: 163-175.
- Witus I. W. & Leong W. V. 2016. Aquaculture in Malaysia; A Short Review on Current Policy and Legislation. Transactions on Science and Technology. 3: 150-154.
- Wilson R.P. 1994. Utilization of dietary carbohydrate by fish. Aquaculture. 124: 67-80.

LIST OF PUBLICATION

Nor Fatihah Mohd Nasir, Mohammad Noor Amal Azmai, Hishamuddin Omar, Ahmad Ismail. 2017. Supplementation of spirulina (*Arthrospira platensis*) improves physical activeness level of catfish (*Clarias gariepinus*). International Journal of Biosciences. 11(5): 146-150.

Nor Fatihah Mohd Nasir, Mohammad Noor Amal Azmai, Hishamuddin Omar, Ahmad Ismail. 2017. Spirulina, *Arthrospira platensis* supplementation in catfish, *Clarias gariepinus*, reduces susceptibility to *Aeromonas hydrophila* infection. Paper presented at International Conference on Advance in Fish Health 2017 (ICFISH 2017), 4-6 April 2017 Office of the Deputy Vice Chancellor (Research and Innovation), Universiti Putra Malaysia, UPM Serdang, Selangor Malaysia.