



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION AND EXPLOITATION OF LACTOBACILLUS
STRAINS AS POTENTIAL PROBIOTICS FOR HUMANS AND CHICKENS***

PARISA SHOKRYAZDAN

IB 2013 41



**CHARACTERIZATION AND EXPLOITATION OF *LACTOBACILLUS*
STRAINS AS POTENTIAL PROBIOTICS FOR HUMANS AND CHICKENS**

By

PARISA SHOKRYAZDAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

August 2013

DEDICATION

*This thesis is dedicated to my husband and my
parents for their love, endless support and
encouragement*

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

CHARACTERIZATION AND EXPLOITATION OF *LACTOBACILLUS* STRAINS AS POTENTIAL PROBIOTICS FOR HUMANS AND CHICKENS

By

PARISA SHOKRYAZDAN

August 2013

Chairman: Professor Ho Yin Wan, PhD

Institute: Bioscience

Among the lactic acid bacteria, lactobacilli are the most common microorganisms used as probiotics. Many different strains of *Lactobacillus* have been reported to be suitable probiotics for different hosts and new strains are being discovered as potential candidates. It has been suggested that there are beneficial lactic acid bacterial strains in human milk and feces, fermented foods and chicken intestines which can be used as probiotics. Thus, this study was conducted to isolate, identify and characterize some new lactic acid bacterial strains from the above-mentioned sources with a view to exploit them as potential probiotics for humans and chickens.

One hundred and eighty-two lactic acid bacterial strains were isolated and 12 strains which showed bile and acid tolerance (growth measured as cell turbidity) were selected and identified using carbohydrate fermentation patterns and 16S rRNA gene sequences. All 12 strains belonged to the genus *Lactobacillus*, of which three strains were *L. buchneri*, three were *L. casei*, one was *L. acidophilus*, two were *L. fermentum* and three were *L. salivarius*.

The 12 *Lactobacillus* strains and two commercial reference strains, *L. casei* Shirota and *L. reuteri* C10, were tested for their ability to survive in the gastrointestinal tract using the following characteristics: tolerance to acid, bile salt and pancreatic enzyme (growth measured as viable counts of colony forming units [CFU]/ml) and adherence to intestinal cell line. The results showed that all 12 *Lactobacillus* strains were able to tolerate acid, bile and pancreatic enzyme. All 12 *Lactobacillus* strains also exhibited good adherence to the intestinal cell line (10.3 to 37.7 *Lactobacillus* cells per Caco-2 cell).

In vitro assays on their bioactivities showed that all 12 *Lactobacillus* strains exhibited good antagonistic activity against a wide range of pathogens and most of the strains had significantly higher inhibitory effect against the pathogens than the two commercial reference strains. The antagonistic activity was due to production of organic acids. All strains also showed cholesterol-reducing activity, good antioxidant activity and production of useful enzymes. The cholesterol-reducing and antioxidant activities of many of the strains were significantly higher than those of the two commercial reference strains. Only three *L. salivarius* strains which were isolated from chicken intestines, and *L. acidophilus* HM1 isolated from human milk showed moderate to high BSH activity. None of the *Lactobacillus* strains produced harmful enzymes or biogenic amines. Three *Lactobacillus* strains (*L. acidophilus* HM1, *L. fermentum* HM3 and *L. buchneri* FD2) selected for their good adherence ability (33.5, 37.7 and 35.9 *Lactobacillus* cells per Caco-2 cell, respectively), and the commercial reference strain *L. casei* Shirota (19.7 *Lactobacillus* cells per Caco-2 cell) were investigated for their cytotoxic activity against three cancer cell lines and a normal cell line. The results showed that the three *Lactobacillus* strains showed selectivity in killing cancer cells when compared to the normal cells. However, the commercial reference strain, *L. casei* Shirota, did not show selectivity toward the cancer or normal cells.

The efficacy of a mixture of three *L. salivarius* strains isolated from chicken intestines, which showed good probiotic traits, was evaluated in broiler chickens. The results revealed that the mixture of *Lactobacillus* strains significantly improved the body weight, body weight gain and feed conversion ratio, increased beneficial cecal bacteria such as lactobacilli and bifidobacteria, decreased harmful cecal bacteria such as *E. coli* and total aerobes, decreased serum total cholesterol, low density lipoprotein-cholesterol and triglycerides, reduced harmful cecal bacterial enzymes, β -glucosidase and β -glucuronidase, and improved the histomorphology of the gut by increasing villus heights and villus height:crypt depth ratio of broiler chickens.

Two *Lactobacillus* strains (*L. fermentum* HM3 and *L. buchneri* FD2 isolated from human milk and fermented dates, respectively), with very good probiotic characteristics were selected as potential probiotic for humans and a safety assessment of them was carried out using acute and subacute oral toxicity tests in Sprague-Dawley rats. Results showed that both strains were safe and even at a high concentration of 10^{10} CFU/kg BW/day there were no observed adverse effects on growth, feed consumption, cellular blood components, serum biochemistry and vital organs of the animals fed *Lactobacillus* strains. The two *Lactobacillus* strains also increased beneficial cecal bacterial populations, and decreased pathogenic bacterial populations and harmful intestinal bacterial enzymes.

In conclusion, all 12 *Lactobacillus* strains isolated in the present study showed good probiotic characteristics and bioactivities *in vitro*. *In vivo* study of three *L. salivarius* strains in broiler chickens indicated that they improved performance and well-being of chickens, and could be good probiotics for chickens, while toxicity tests on *L. fermentum* HM3 and *L. buchneri* FD2 in rats showed that they are safe, and could be further investigated and exploited as probiotics for humans.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN DAN EKSPLOITASI STRAIN *LACTOBACILLUS* SEBAGAI PROBIOTIK
BERPOTENSI UNTUK MANUSIA DAN AYAM PEDAGING**

Oleh

PARISA SHOKRYAZDAN

Ogos 2013

Pengerusi: Professor Ho Yin Wan, PhD

Institut: Biosains

Diantara laktik asid bakteria, lactobacilli adalah mikroorganisma yang biasa digunakan sebagai probiotik. Pelbagai strain *Lactobacillus* telah dilaporkan sebagai probiotik yang sesuai untuk perumah yang berbeza dan juga strain-strain baru sedang diterokai sebagai calon-calon yang berpotensi. Ada beberapa cadangan bahawa terdapat strain laktik asid bakteria yang bermanfaat dari sumber susu manusia dan najis, makanan ditapai dan usus haiwan yang boleh digunakan sebagai probiotik. Oleh itu, kajian ini dijalankan untuk mengasingkan, mengenal pasti dan mencirikan beberapa laktik asid bakteria dari sumber yang disebut di atas dengan tujuan untuk mengeksploitasi potensi makanan tambahan probiotik untuk manusia dan ayam.

Seratus lapan puluh dua isolat laktik asid bakteria telah diisolat dan 12 strain yang menunjukkan toleransi asid dan hempedu (pertumbuhan diukur sebagai kekeruhan sel) telah dipilih dan dikenal pasti menggunakan corak penapaian karbohidrat dan 16S rRNA urutan gen. Kesemua 12 jenis strain telah dikenalpasti sebagai genus *Lactobacillus*, di mana tiga adalah jenis strain *L. buchneri*,

tiga lagi strain *L. casei*, satu strain *L. Acidophilus*, dua strain *L. fermentum* dan tiga strain *L. Salivarius*.

12 *Lactobacillus* strain dan dua strain rujukan komersial, *L. casei* Shirota dan *L. reuteri* C10, telah diuji keupayaannya untuk terus hidup dalam saluran gastrousus dengan menggunakan ciri-ciri ujian berikut: toleransi kepada asid, garam hempedu dan enzim pankreas (pertumbuhan diukur dari segi kiraan koloni hidup/ml (CFU/ml) dan daya lekatan pada sel usus. Hasil kajian menunjukkan bahawa kesemua 12 jenis *Lactobacillus* strain dapat bertolak ansur dengan asid, hempedu dan pankreas enzim. Semua 12 *Lactobacillus* jenis juga menunjukkan daya lekatan yang baik pada sel usus (10.3 to 37.7 *Lactobacillus* sel per Caco-2 sel).

Hasil kajian bioaktiviti menunjukkan bahawa kesemua 12 *Lactobacillus* strain menunjukkan aktiviti antagonistik yang berkesan terhadap pelbagai patogen dan kebanyakannya mempunyai kesan bersignifikan lebih tinggi terhadap patogen daripada dua jenis rujukan komersial. Aktiviti antagonistik disebabkan oleh pengeluaran asid organik. Semua jenis strain juga menunjukkan aktiviti penurunan paras kolesterol, aktiviti antioksidan yang baik dan pengeluaran enzim yang berguna. Penurunan paras kolesterol dan antioksidan aktiviti kebanyakan jenis strain adalah bersignifikan berbanding dengan kedua-dua jenis strain rujukan komersial. Hanya tiga jenis *L. salivarius* yang telah diisolat daripada usus ayam, dan *L. acidophilus* HM1 diisolat daripada susu manusia menunjukkan aktiviti BSH dari sederhana hingga tinggi. Tiada strain *Lactobacillus* yang menghasilkan enzim berbahaya atau amina biogenik. Tiga jenis *Lactobacillus* (*L. acidophilus* HM1, *L. fermentum* HM3 dan *L. buchneri* FD2) yang dipilih berdasarkan keupayaan daya lekatan yang baik (33.5, 37.7 and 35.9 *Lactobacillus* sel per Caco-2 sel), dan strain rujukan komersial *L. casei* Shirota telah dikaji untuk aktiviti sitotoksik mereka terhadap tiga sel kanser dan sel normal. Hasil kajian menunjukkan bahawa tiga jenis strain *Lactobacillus* menunjukkan pemilihan dalam

membunuh sel-sel kanker berbanding dengan sel-sel normal. Walau bagaimanapun, strain rujukan komersial, *L. casei* Shirota, tidak menunjukkan pemilihan ke arah kanker atau sel-sel normal.

Keberkesanan campuran tiga jenis strain *L. salivarius* yang diasingkan daripada usus ayam, yang menunjukkan sifat-sifat probiotik yang baik, telah dinilai dalam ayam daging. Keputusan menunjukkan bahawa campuran strain *Lactobacillus* meningkatkan berat badan (BW), kadar pertumbuhan berat badan dan nisbah penukaran makanan, peningkatan bakteria sekum berfaedah seperti lactobacilli dan bifidobakteria, menurun bakteria sekum berbahaya seperti *E. coli* dan jumlah bakteria aerob, penurunan jumlah kolesterol serum, ketumpatan rendah lipoprotein kolesterol dan trigliserida, mengurangkan enzim berbahaya bakteria sekum, β -glucosidase dan β -glucuronidase, dan meningkatkan kualiti histomorphologi usus oleh tahap villus yang semakin meningkat dan ketinggian villus:nisbah crypt depth ayam daging.

Dua jenis strain *Lactobacillus* (*L. fermentum* HM3 dan *L. buchneri* FD2 diisolat daripada susu manusia dan kurma ditapai masing-masing) dengan ciri-ciri probiotik yang sangat baik telah dipilih sebagai potensi probiotik untuk manusia dan penilaian keselamatan mereka telah dijalankan dengan menggunakan akut dan subakut ujian ketoksikan oral pada tikus Sprague-Dawley. Keputusan ujian menunjukkan bahawa kedua-dua jenis strain adalah selamat diguna walaupun pada kepekatan yang tinggi 10^{10} CFU / kg BW/hari dimana tiada kesan buruk diperhatikan kepada pertumbuhan, pengambilan makanan, komponen darah selular, serum biokimia dan organ-organ penting dalam haiwan yang diberi makanan tambahan yang mengandungi jenis strain *Lactobacillus*. Kedua-dua jenis strain *Lactobacillus* juga meningkatkan populasi bakteria sekum yang bermanfaat, dan menurunkan populasi bakteria berpatogenik dan enzim berbahaya bakteria usus.

Kesimpulannya, kesemua 12 jenis strain *Lactobacillus* diasingkan dalam kajian ini menunjukkan ciri-ciri dan bioaktiviti yang baik dalam kajian secara *in vitro*. Dalam kajian secara *in vivo*, tiga jenis strain *L. salivarius* dalam ayam daging menunjukkan bahawa mereka meningkatkan prestasi dan kesihatan ayam, dan berpotensi untuk dijadikan sebagai probiotik ayam, manakala ujian ketoksikan *L. fermentum* HM3 dan *L. buchneri* FD2 pada tikus menunjukkan bahawa kedua-dua strain bukan toksik, boleh dikaji lagi dan dieksploitasi sebagai probiotik untuk manusia.



ACKNOWLEDGEMENTS

First and foremost, I would like to express my utmost gratitude to the chairman of the supervisory committee, Professor Dr. Ho Yin Wan, for being so approachable, and for her invaluable advice, guidance, support, encouragement, tolerance and patience throughout the course of my study. I am really grateful to have been given this opportunity to pursue my study under her supervision.

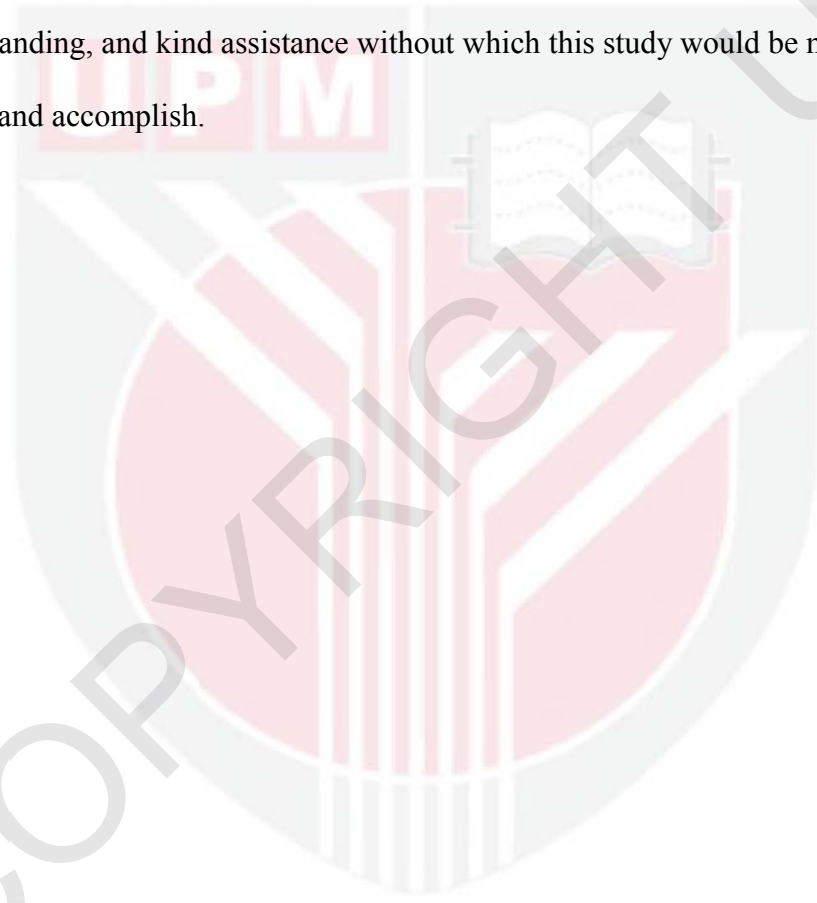
I would also like to express my deepest thanks to Associate Professor Dr. Kalavathy Ramasamy, Associate Professor Dr. Liang Juan Boo and Associate Professor Dr. Sieo Chin Chin who are members of the supervisory committee for their advice, invaluable guidance, hospitality, support and encouragement throughout the duration of my study.

I would like to thank Associate Professor Dr. Noorjahan Banu Mohamed Alitheen for her help, support and guidance in the cell culture, and Profesor Dato' Dr. Tengku Azmi Tengku Ibrahim for his help, guidance and assistance in the histopathology of rats.

My appreciations are extended to Mr. Khairul Kamar Bakri, staff of the Probiotic and Prebiotic Programme, Laboratory of Vaccine and Immunotherapeutics, Institute of Bioscience, for his technical support and kind assistance.

Thanks are also due to all my labmates, who made me feel welcome in Malaysia, and revealed to me a side of South East Asia that I could never have experienced on my own.

Last but not least, very special thanks to my loving husband, Mohammad, for his endless love, strong support, constant encouragement, infinite patience and understanding, and kind assistance without which this study would be much harder to pursue and accomplish.



I certify that a Thesis Examination Committee has met on 29 August 2013 to conduct the final examination of Parisa Shokryazdan on her thesis entitled “Characterization and Exploitation of *Lactobacillus* Strains as Potential Probiotics for Humans and Chickens” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A)106] 15 March 1998. The committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Wan Zuhainis Saad, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Science

Universiti Putra Malaysia

(Chairman)

Shuhaimi bin Mustafa, PhD

Professor

Faculty of Biotechnology and Biomolecular Science

Universiti Putra Malaysia

(Internal Examiner)

Abdul Rani bin Bahaman, PhD

Professor Dato’

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Internal Examiner)

Ming-Ju Chen, PhD

Professor

National Taiwan University

Taiwan

(External Examiner)

NORITAH OMAR, PhD

Associate Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 19 September 2013

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Ho Yin Wan, PhD

Professor
Institute of Bioscience
Universiti Putra Malaysia
(Chairman)

Kalavathy Ramasamy, PhD

Associate Professor
Faculty of Pharmacy
Universiti Teknologi MARA
(Member)

Liang Juan Boo, PhD

Associate Professor
Institute of Tropical Agriculture
Universiti Putra Malaysia
(Member)

Sieo Chin Chin, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Science
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Putra Malaysia or other institutions.

PARISA SHOKRYAZDAN

Date: 29 August 2013

TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGEMENTS	xii
APPROVAL	xiv
DECLARATION	xvi
LIST OF TABLES	xxiii
LIST OF FIGURES	xxvi
LIST OF ABBREVIATION	xxxi
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	5
2.1 Definition of probiotic	5
2.2 Probiotics for humans	6
2.3 Probiotics for poultry	8
2.4 Lactic acid bacteria and <i>Lactobacillus</i> strains	9
2.5 Sources of probiotic strains	10
2.6 Selection of probiotic strains	12
2.6.1 Identification using phenotypic and genotypic characterizations	14
2.6.2 Acid and bile salt resistance	16
2.6.3 Adherence to mucus and/or epithelial cells	17
2.6.4 Antimicrobial and antagonistic activities against pathogens	18
2.6.5 Bile salt hydrolase activity	22
2.6.6 Antioxidant activity	23
2.6.7 Cytotoxic effect	24
2.7 Safety of probiotic strains	24
3 ISOLATION, PRELIMINARY SCREENING AND IDENTIFICATION OF SOME LACTIC ACID BACTERIA	28
3.1 Introduction	28
3.2 Materials and methods	29
3.2.1 Sources for isolation of lactic acid bacteria	29

3.2.2	Isolation of lactic acid bacteria	30
3.2.3	Initial identification of lactic acid bacteria	31
3.2.4	Preliminary screening for acid and bile tolerance	31
3.2.5	Identification of lactic acid bacterial isolates using biochemical properties and genotypic characteristics	32
3.2.5.1	Biochemical properties	33
3.2.5.2	Molecular characterization	34
3.3	Results and discussion	39
3.3.1	Initial identification of lactic acid bacteria	39
3.3.2	Preliminary screening for acid and bile tolerance of lactic acid bacterial isolates	40
3.3.3	Identification to generic and species levels	43
3.3.3.1	Identification using biochemical properties	43
3.3.3.2	Identification using 16S rRNA gene sequencing	47
3.3.3.3	Phylogenetic analysis based on 16S rRNA gene	51
3.3.3.4	Morphological characteristics of the 12 <i>Lactobacillus</i> strains	53
3.4	Conclusions	56
4	<i>IN VITRO</i> ASSESSMENT OF CHARACTERISTICS FOR SURVIVAL OF SELECTED <i>LACTOBACILLUS</i> STRAINS IN THE GASTROINTESTINAL TRACT	57
4.1	Introduction	57
4.2	Materials and methods	58
4.2.1	<i>Lactobacillus</i> strains and growth conditions	58
4.2.2	<i>In vitro</i> assay for acid tolerance	59
4.2.3	<i>In vitro</i> assay for bile tolerance	60
4.2.4	<i>In vitro</i> assay for pancreatic enzyme tolerance	60
4.2.5	Adhesion assay	61
4.2.6	Scanning electron microscopic examination	62
4.2.7	Statistical analysis	63
4.3	Results	63
4.3.1	Acid tolerance	63
4.3.2	Bile tolerance	65
4.3.3	Pancreatic enzyme tolerance	67
4.3.4	Adherence assay	69
4.4	Discussion	73
4.1	Conclusions	78

5	BIOACTIVITIES OF <i>LACTOBACILLUS</i> STRAINS	79
5.1	Introduction	79
5.2	Materials and methods	81
5.2.1	Strains and growth conditions	81
5.2.2	Antimicrobial activity against pathogens	82
5.2.2.1	Test pathogens	82
5.2.2.2	Antagonistic test	84
5.2.2.3	Characterization of antimicrobial substances	84
5.2.2.4	Organic acid production profile	85
5.2.3	Bile salt hydrolase activity	87
5.2.3.1	Qualitative assay	87
5.2.3.2	Quantitative assay	88
5.2.4	Cholesterol reduction assay	89
5.2.5	Antioxidant activity	90
5.2.5.1	Sample preparation	90
5.2.5.2	FRAP Assay	91
5.2.5.3	ABTS Assay	92
5.2.5.4	DPPH Assay	93
5.2.5.5	TBA Assay	93
5.2.6	Cytotoxic activity	94
5.2.6.1	Cell culture	95
5.2.6.2	MTT cell proliferation assay	95
5.2.7	Production of biogenic amines	98
5.2.8	Enzyme activity profile	99
5.2.9	Statistical analysis	100
5.3	Results	101
5.3.1	Antimicrobial activity	101
5.3.1.1	Antagonistic effects	101
5.3.1.2	Characterization of inhibitory substances	109
5.3.1.3	Profile of organic acid production	113
5.3.2	Bile salt hydrolase activity	115
5.3.2.1	Qualitative assay	115
5.3.2.2	Quantitative assay	117
5.3.3	Cholesterol reduction assay	128
5.3.4	Antioxidant activity	132
5.3.5	Cytotoxic activity	140
5.3.6	Production of biogenic amines	142
5.3.7	Enzyme activity profile	143
5.4	Discussion	146

5.4.1	Antagonist effects, production of antimicrobial substances and organic acid production profile	146
5.4.2	Bile salt hydrolase activity	150
5.4.3	Cholesterol-reducing activity	153
5.4.4	Antioxidant activity	155
5.4.5	Cytotoxicity	158
5.4.6	Production of biogenic amines	160
5.4.7	Enzyme activity profile	161
5.5	Conclusions	162
6	EFFICACY ASSESSMENT OF SELECTED <i>LACTOBACILLUS</i> STRAINS AS PROBIOTIC FOR BROILER CHICKENS	164
6.1	Introduction	164
6.2	Materials and methods	165
6.2.1	<i>Lactobacillus</i> cultures	165
6.2.2	Chickens and rearing management	Error! Bookmark not defined.
6.2.3	Experimental design	166
6.2.4	Sample collection and analysis	168
6.2.5	Microbiological analyses of cecal contents	169
6.2.5.1	Conventional spread plate method	169
6.2.5.2	Real-time PCR assay	170
6.2.6	Serum lipid assay	173
6.2.7	Relative weights of organs	173
6.2.8	Villus height and crypt depth measurements	173
6.2.9	Harmful bacterial enzyme assay	174
6.2.9.1	Sample preparation	174
6.2.9.2	β -Glucosidase activity assay	174
6.2.9.3	β -Glucuronidase activity assay	175
6.2.10	Statistical analysis	175
6.3	Results	176
6.3.1	Performance of broiler chickens	176
6.3.2	Enumeration of cecal bacteria	178
6.3.2.1	Conventional microbiological method	178
6.3.2.2	Real-time PCR quantification	181
6.3.3	Serum lipid analysis	190
6.3.4	Relative weights of organs	191
6.3.5	Intestinal villus height and crypt depth	193
6.3.6	Harmful cecal bacterial enzyme (β -glucosidase and β -glucuronidase) activities	196

6.4	Discussion	197
6.5	Conclusions	207
7	SAFETY ASSESSMENT OF TWO SELECTED <i>LACTOBACILLUS</i> STRAINS (INTENDED AS PROBIOTIC FOR HUMANS) USING A RAT MODEL	208
7.1	Introduction	208
7.2	Materials and methods	209
7.2.1	<i>Lactobacillus</i> cultures (test products)	209
7.2.2	Animals	210
7.2.3	Acute oral toxicity study (single-dose toxicity study)	210
7.2.4	Subacute oral toxicity study (repeated-dose toxicity study)	211
7.2.4.1	Experimental design	211
7.2.4.2	General observations	212
7.2.4.3	Hematological and serum biochemistry analyses	213
7.2.4.4	Relative organ weights and histopathological examination	214
7.2.4.5	Microbiological analysis of cecal contents	215
7.2.4.6	Harmful intestinal bacterial enzyme assay	215
7.2.5	Statistical analysis	216
7.3	Results	216
7.3.1	Single-dose acute oral toxicity study	216
7.3.2	Repeated-dose subacute oral toxicity study	218
7.3.2.1	General signs	218
7.3.2.2	Body weight and feed consumption	218
7.3.2.3	Relative organ weights and histopathological examination	222
7.3.2.4	Histopathological examination	222
7.3.2.5	Hematology and serum biochemistry analysis	232
7.3.2.6	Microbiological analysis of cecal contents	232
7.3.2.7	Harmful cecal bacterial enzymes	245
7.4	Discussion	247
7.5	Conclusion	252
8	GENERAL DISCUSSION, RECOMMENDATIONS FOR FUTURE RESEARCH AND CONCLUSION	253
8.1	General discussion and recommendations for future research	253
8.2	Conclusions	263

REFERENCES	264
BIODATA OF STUDENT	292
LIST OF PUBLICATIONS	293



LIST OF TABLES

Table	Page
2.1. Non-bacteriocin antimicrobial metabolites of lactic acid bacteria	20
3.1. Number of lactic acid bacterial isolates from different sources	40
3.2. Results of catalase test and Gram staining of lactic acid bacterial isolates	40
3.3. Number of tolerant isolates after treating with acidic and bile conditions	42
3.4. Percentage of growth of 12 selected isolates in the presence of bile and acid condition (pH 3)	42
3.5. Carbohydrate fermentation patterns of 12 lactic acid bacterial isolates	44
3.6. Identification of 12 lactic acid bacterial isolates using the API 50 CH kit	46
3.7. Identification of 12 lactic acid bacterial isolates using 16 S rRNA gene sequencing	50
4.1. Viability of <i>Lactobacillus</i> strains after 3 h exposure to pH 3 and pH 7.2 (control)	64
4.2. Growth of <i>Lactobacillus</i> strains in MRS broth (control) and MRS broth containing 0.3% bile salt	66
4.3. Viability of <i>Lactobacillus</i> strains after 3 h exposure to 1.9 mg/ml pancreatic enzymes and normal condition (control)	68
4.4. Adhesion of cells of <i>Lactobacillus</i> strains to Caco-2 cell	70
5.1. Test pathogens and their specific growth media	83
5.2. Antagonist effect of <i>Lactobacillus</i> strains against pathogenic strains	103
5.3. Well diffusion assay of treated and untreated supernatants of <i>Lactobacillus</i> strains against <i>E. coli</i> (ATCC 29181) as indicator strain	111
5.4. Well diffusion assay of treated and untreated supernatants of <i>Lactobacillus</i> strains against <i>Enterococcus faecium</i> (ATCC 51558) as indicator strain	112
5.5. Organic acid production of <i>Lactobacillus</i> strains	114
5.6. Qualitative assay of bile salt hydrolase activity of <i>Lactobacillus</i> strains	116
5.7. Deconjugation (%) of GCA and TCA by <i>Lactobacillus</i> strains after 24 h of incubation and correlation with bile tolerance of the strains	118

5.8. Cholesterol reduction (%) in MRS broth with and without 0.3% bile salt (oxgall) after incubation with <i>Lactobacillus</i> strains	129
5.9. Cholesterol assimilated (%) in cells of <i>Lactobacillus</i> strains incubated for 24 h in MRS broth with and without 0.3% bile salt (oxgall)	131
5.10. Trolox equivalent antioxidant capacity (TEAC) of intracellular cell-free extracts and intact cells of <i>Lactobacillus</i> strains using FRAP assay	133
5.11. Trolox equivalent antioxidant capacity (TEAC) of intracellular cell-free extracts and intact cells of <i>Lactobacillus</i> strains using ABTS assay	135
5.12. Trolox equivalent antioxidant capacity (TEAC) of intracellular cell-free extracts and intact cells of <i>Lactobacillus</i> strains using DPPH assay	138
5.13. Percentage of inhibition of lipid peroxidation for intracellular cell-free extracts and intact cells of <i>Lactobacillus</i> strains using TBA assay	139
5.14. IC ₅₀ of supernatants of <i>Lactobacillus</i> strains on different cell lines	141
5.15. Enzyme activity profile of the <i>Lactobacillus</i> strains	145
6.1. Composition of basal diet	168
6.2. Primers used for real-time PCR assay to target <i>Lactobacillus</i> , <i>Bifidobacterium</i> and <i>E. coli</i>	172
6.3. Effects of dietary treatments on body weight, body weight gain, feed intake, FCR and mortality of broiler chickens	177
6.4. Effects of dietary supplementations of a mixture of <i>L. salivarius</i> strains on serum lipid concentrations of broiler chickens at 21 and 42 days of age	191
6.5. Effects of dietary supplementations of a mixture of <i>L. salivarius</i> strains on relative weights of organs of broiler chickens at 21 and 42 days of age	192
6.6. Effects of dietary supplementations of a mixture of <i>L. salivarius</i> strains on intestinal villus height and crypt depth of broiler chickens at 21 and 42 days of age	194
6.7. Effect of dietary supplementations of a mixture of <i>L. salivarius</i> strains on β -glucuronidase and β -glucosidase activities in the cecal contents of broilers	196
7.1. Weekly body weight gain, daily feed intake and mortality of male and female rats in the acute oral toxicity study	217
7.2. Body weights of male rats	219
7.3. Body weights of female rats	220

7.4. Feed consumption of male and female rats	221
7.5. Relative organ weights (%) of male rats	223
7.6. Relative organ weights (%) of female rats	224
7.7. Hematological parameters of male rats	234
7.8. Hematological parameters of female rats	235
7.9. Serum biochemical parameters of male rats	236
7.10. Serum biochemical parameters of female rats	238
7.11. β -glucuronidase and β -glucosidase activities in the cecal contents of rats	246

LIST OF FIGURES

Figure	Page
2.1. Metabolism of <i>Lactobacillus</i> spp	11
3.1. An example of an API 50 CH stripe inoculated with a <i>Lactobacillus</i> strain (<i>L. acidophilus</i> , HM1)	34
3.2. PCR amplification of the 16S rRNA gene of the 12 isolated lactic acid bacterial strains	49
3.3. Phylogenetic tree based on 16S rRNA gene sequence analysis depicting the phylogenetic relationships among species of the genus <i>Lactobacillus</i> using the Neighbor-Joining method (Saitou and Nei, 1987)	52
3.4. Cell morphology of <i>Lactobacillus</i> strains observed using light microscopy	55
4.1. Adhesion of <i>Lactobacillus</i> cells per Caco-2 cell	71
4.2. Scanning electron micrograph of a Caco-2 cell (C) with many attached cells of <i>L. buchneri</i> FD2 (L) (rod-shaped)	72
5.1. A 96-well plate with MTT assay in progress using monolayer of liver cancer (HepG2) cell line	98
5.2. An example of an API ZYM stripe inoculated with a <i>Lactobacillus</i> strain (<i>L. casei</i> BF2)	100
5.3. Agar spot test showing inhibition (clear zones) (Z) of a pathogenic strain, <i>Candida albicans</i> (ATCC 44831) (C), around the colonies of <i>Lactobacillus buchneri</i> FD1 (L)	102
5.4. Well diffusion assay showing clear inhibition zone of <i>E. coli</i> (ATCC 29181) by untreated supernatant (US) and neutralized supernatant (NS) (pH 6.5) of <i>Lactobacillus buchneri</i> FD2	110
5.5. (A), colonies of <i>Lactobacillus salivarius</i> C11 with precipitates of bile salt on a MRS agar plate supplemented with 1 mM TCA; (B), colonies of <i>Lactobacillus casei</i> BF3 without precipitate of bile salt on a MRS agar plate supplemented with 1 mM TCA	117
5.6. Growth rate and deconjugation of TCA and GCA by <i>L. casei</i> Shirota in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	121
5.7. Growth rate and deconjugation of TCA and GCA by <i>L. reuteri</i> C10 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	121

5.8. Growth rate and deconjugation of TCA and GCA by <i>L. salivarius</i> CI1 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	122
5.9. Growth rate and deconjugation of TCA and GCA by <i>L. salivarius</i> CI2 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	122
5.10. Growth rate and deconjugation of TCA and GCA by <i>L. salivarius</i> CI3 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	123
5.11. Growth rate and deconjugation of TCA and GCA by <i>L. acidophilus</i> HM1 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	123
5.12. Growth rate and deconjugation of TCA and GCA by <i>L. fermentum</i> HM2 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	124
5.13. Growth rate and deconjugation of TCA and GCA by <i>L. fermentum</i> HM3 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	124
5.14. Growth rate and deconjugation of TCA and GCA by <i>L. buchneri</i> FG1 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	125
5.15. Growth rate and deconjugation of TCA and GCA by <i>L. buchneri</i> FD1 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	125
5.16. Growth rate and deconjugation of TCA and GCA by <i>L. buchneri</i> FD2 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	126
5.17. Growth rate and deconjugation of TCA and GCA by <i>L. casei</i> BF1 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	126
5.18. Growth rate and deconjugation of TCA and GCA by <i>L. casei</i> BF2 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	127
5.19. Growth rate and deconjugation of TCA and GCA by <i>L. casei</i> BF3 in MRS broth medium supplemented with TCA and GCA from 0 - 24 h of incubation	127

5.20. IC ₅₀ of supernatants of <i>Lactobacillus</i> strains on different cell lines	142
5.21. <i>Lactobacillus buchneri</i> FD2 grown on agar plates containing 2% of a precursor amino acid (histidine, lysine, ornithine or tyrosine)	143
6.1. A stainless steel, three-tiered battery cage with chicks inside at commencement of the experiment	167
6.2. Effect of dietary supplementations of a mixture of three <i>L. salivarius</i> strains (LC) on populations of cecal lactobacilli of broiler chickens at 21 and 42 days of age enumerated using the conventional spread plate method	179
6.3. Effect of dietary supplementations of a mixture of three <i>L. salivarius</i> strains (LC) on populations of cecal bifidobacteria of broiler chickens at 21 and 42 days of age enumerated using the conventional spread plate method	179
6.4. Effect of dietary supplementations of a mixture of three <i>L. salivarius</i> strains (LC) on populations of cecal total cecal aerobes of broiler chickens at 21 and 42 days of age enumerated using the conventional spread plate method	180
6.5. Effect of dietary supplementations of a mixture of three <i>L. salivarius</i> strains (LC) on populations of cecal <i>E. coli</i> of broiler chickens at 21 and 42 days of age enumerated using the conventional spread plate method	181
6.6. Standard curve obtained from 10-fold serial dilutions of 16S rRNA gene of <i>Lactobacillus</i> ranging from 1 to 4 log copies plotted against Cq values	183
6.7. Standard curve obtained from 10-fold serial dilutions of 16S rRNA gene of <i>Bifidobacterium</i> ranging from 1 to 5 log copies plotted against Cq values	183
6.8. Standard curve obtained from 10-fold serial dilutions of 16S rRNA gene of <i>E. coli</i> ranging from 1 to 5 log copies plotted against Cq values	184
6.9. Amplification plot from real-time PCR assay of lactobacilli, obtained from number of cycles plotted against relative fluorescence units (RFU)	185
6.10. Amplification plot from real-time PCR assay of bifidobacteria, obtained from number of cycles plotted against relative fluorescence units (RFU)	185
6.11. Amplification plot from real-time PCR assay of <i>E. coli</i> , obtained from number of cycles plotted against relative fluorescence units (RFU)	186
6.12. Melting curves of lactobacilli for standard dilutions of DNA template containing 16S rRNA gene	186
6.13. Melting curves of bifidobacteria for standard dilutions of DNA template containing 16S rRNA gene	187

6.14. Melting curves of <i>E. coli</i> for standard dilutions of DNA template containing 16S rRNA gene	187
6.15. Effect of dietary supplementations of a mixture of <i>L. salivarius</i> strains (LC) on populations of cecal lactobacilli of broiler chickens at 21 and 42 days of age quantified using real-time PCR	189
6.16. Effect of dietary supplementations of a mixture of <i>L. salivarius</i> strains (LC) on populations of cecal bifidobacteria of broiler chickens at 21 and 42 days of age quantified using real-time PCR	189
6.17. Effect of dietary supplementations of a mixture of <i>L. salivarius</i> strains (LC) on populations of cecal <i>E. coli</i> of broiler chickens at 21 and 42 days of age quantified using real-time PCR	190
6.18. Villi and crypts of the jejunum of a broiler fed diet supplemented with 0.01% LC (mixture of <i>L. salivarius</i> CI1, CI2 and CI3) at 42 days of age	195
6.19. Cholesterol as the precursor for the synthesis of new bile acids and the hypocholesterolemic role of bile salt hydrolase	202
7.1. Light micrograph of the brain (cerebrum) of (A) a control rat, and (B) a rat given a mixture of <i>L. buchneri</i> FD2 and <i>L. fermentum</i> HM3, showing normal nerve cells (N) with a distinct nucleus	225
7.2. Light micrograph of cardiac muscle fibers of (A) a control rat, and (B) a rat given a mixture of <i>L. buchneri</i> FD2 and <i>L. fermentum</i> HM3, showing normal histological structure of cardiac muscle fibers with their nuclei (N)	226
7.3. Light micrograph of the lung of (A) a control rat, and (B) a rat given a mixture of <i>L. buchneri</i> FD2 and <i>L. fermentum</i> HM3, showing normal histological structure of lung comprising of alveolus (a) and inter-alveolar septum (S)	227
7.4. Light micrograph of the liver of (A) a control rat, and (B) a rat given a mixture of <i>L. buchneri</i> FD2 and <i>L. fermentum</i> HM3, showing normal histological structure of hepatocytes (H) and sinusoids (S)	228
7.5. Light micrograph of the spleen of (A) a control rat, and (B) a rat given a mixture of <i>L. buchneri</i> FD2 and <i>L. fermentum</i> HM3, showing normal histological structure of spleen comprising of sinusoids (S), lymphocytes (L) and red blood cells (R)	229
7.6. Light micrograph of the kidney of a (A) control rat, and (B) a rat given a mixture of <i>L. buchneri</i> FD2 and <i>L. fermentum</i> HM3, showing normal histological structure of kidney comprising of proximal (P) and distal (D) convoluted tubules	230

7.7. Light micrograph of the intestine of (A) a control rat, and (B) a rat given a mixture of <i>L. buchneri</i> FD2 and <i>L. fermentum</i> HM3, showing normal histological structure of the small intestine (jejunum) comprising of epithelial cells (E)	231
7.8. Populations of cecal lactobacilli of male and female rats	240
7.9. Populations of cecal bifidobacteria of male and female rats	241
7.10. Populations of cecal <i>E. coli</i> of male and female rats	242
7.11. Populations of cecal <i>Salmonella</i> of male and female rats	243
7.12. Populations of cecal total aerobes of male and female rats	244

LIST OF ABBREVIATIONS

ABTS	2,2'-Azino-bis (3-ethylbenzothiazoline-6-sulfonic acid)
AACC	American Association for Clinical Chemistry
ALB	Albumin
ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
ANOVA	Analysis of variance
AST	Aspartate aminotransferase
ATCC	American Type Culture Collection
BLAST	Basic Local Alignment Search Tool
BSH	Bile Salt Hydrolase
Ca	Calcium
CFU	Colony Forming Units
Cl	Chloride
Cq	Quantification cycle
Creat	Creatinine
CVD	Cardiovascular diseases
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
dNTP	Deoxynucleotide triphosphate
DPPH	2,2-Diphenyl-1-picrylhydrazyl
E	Amplification efficiency
EDTA	Ethylene Diamine Tetraacetic Acid
Eosin.	Eosinophils
FAO	Food and Agriculture Organization
FBS	Fetal Bovine Serum
FID	Flame Ionization Detector
fL	Femtoliters

FRAP	Ferric Reducing Ability of Plasma
GC	Gas Chromatograph
GCA	Glycocholic acid
GCDCA	Glycochenodeoxycholic acid
GDCA	Glycodeoxycholic acid
GIA	Global Industry Analysts
GIT	Gastrointestinal Tract
Glu.	Glucose
GRAS	Generally Recognized As Safe
HB	Hemoglobin
HDL	High Density Lipoprotein
HPLC	High Performance Liquid Chromatography
ISAPP	International Scientific Association for Probiotics and Prebiotics
K	Potassium
LB	Luria-bertani
LDL	Low Density Lipoprotein
Lymp.	Lymphocytes
MCHC	Mean Corpuscular Hemoglobin Concentration
MCV	Mean Corpuscular Volume
Mono.	Monocytes
MRS	De Man, Rogosa and Sharpe
MTT	3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
Na	Sodium
NCBI	National Center for Biotechnology Information
Neut.	Neutrophils
NRC	National Research Council
NTC	No-template control
OCDE	Organisation de Coopération et de Développement Economiques
OD	Optical Density
OECD	Organisation for Economic Co-operation and Development
OPA	O-phthalaldehyde

P	Phosphorus
PBS	Phosphate Buffer Saline
PCR	Polymerase Chain Reaction
PCV	Packed Cell Volume
QPS	Qualified Presumption of Safety
RBC	Red Blood Cell Count
RFU	Relative Fluorescence Units
RPMI	Roswell Park Memorial Institute
rRNA	Ribosomal ribonucleic acid
SAS	Statistical Analysis System
T. bil.	Total bilirubin
T. chol.	Total cholesterol
T. prot.	Total protein
TBA	Thiobarbituric acid
TCA	Taurocholic acid
TCDC	Taurochenodeoxycholic acid
TDCA	Taurodeoxycholic acid
TEA	Tris-acetate EDTA
TEAC	Trolox Equivalent Antioxidant Capacity
Thrombo.	Thrombocytes
TLCA	Taurolithocholic acid
TPTZ	2,4,6-Tri (2-pyridyl)-1,3,5-triazine
Trig.	Triglycerides
URI	Genomics and Sequencing Center of University of Rhode Island
VFA	Volatile Fatty Acids
WBC	White Blood Cell Count
WHO	World Health Organization

CHAPTER 1

INTRODUCTION

More than two thousand years ago, Hippocrates said: “Let food be thy medicine and medicine be thy food”. This quote is a tenet for many people today. As nowadays people are more conscious and aware about the role of diet in their health, interest and demand for “healthy” foods have increased considerably (Suvarna and Bobby, 2005). Hence, the food industry is expected to provide a healthcare market by offering functional foods, or foods that promote health beyond providing basic nutrition. A rapidly expanding portion of this market belongs to probiotics and prebiotics. Probiotics are defined as “live microorganisms which when administered in adequate amounts confer a health benefit on the host” (FAO/WHO, 2001), and prebiotics are “non-digestible food ingredients that beneficially affect the host by stimulating the growth and/or activity of one or a limited number of bacteria in the colon and thus improve host health” (Gibson and Roberfroid, 1995). According to the Global Industrial Analysis (GIA, 2012), the probiotic market alone is predicted to be worth about US \$ 29 billion in 2015.

In addition to the food industry for human, feed industry for livestock, especially poultry, is another sector for the probiotic market. For over 50 years, antibiotic growth promoters were commonly used at subtherapeutical levels to promote growth and feed efficiency in poultry (Kabir, 2009). However, in recent years, increasing concern on the rampant use of antibiotics, which can lead to development of antibiotic-resistant bacteria and antibiotic residues in animal products has led to restriction or total ban of antibiotics

as growth promoters in animal production (Patterson and Burkholder, 2003). As a result, the concept of using probiotics as a possible alternative to antibiotic growth promoters has attracted considerable interest (Kabir, 2009). That is because probiotics have the potential to reduce enteric diseases and enhance productivity of poultry. Moreover, with rising levels of healthcare awareness, the demand for drug-free animal meat and products is set to increase.

Bifidobacteria and lactic acid bacteria, especially *Lactobacillus*, are the most common types of microorganisms used as probiotics in the food and feed industries (Conway, 1996; Vankerckhoven *et al.*, 2008). That is because lactic acid bacteria and bifidobacteria have good functional effects as probiotics and they are regarded as friendly bacteria and “Generally Recognized As Safe (GRAS)” (Collins *et al.*, 1998). Although many strains of lactic acid bacteria have been found to have probiotic characteristics and have been developed as suitable probiotic supplements for different hosts, there is still a continued search for more new probiotic strains with better health promoting properties than existing ones.

Human milk is an important factor in the colonization and development of beneficial bacteria particularly bifidobacteria in the intestine of breast-fed newborns, and it has been suggested that human milk contains bacterial strains with potential to be used as probiotic agents (Martin *et al.*, 2004). Feces of healthy breast-fed infants have also been reported to contain high numbers of lactic acid bacteria (Hopkins *et al.*, 2005). In addition, many studies have shown that fermented foods contain potential probiotic strains. (Heller, 2001; Muyanja *et al.*, 2003; Leroy and De-Vuyst, 2004; Mathara *et al.*,

2008). It has been recommended that bacterial strain to be used as probiotic for animals should be isolated from the gastrointestinal tract (GIT) of the same type of animal in order to have more specific application for the animal (Kizerwetter-Swida and Binek, 2005). For example, the GIT of chickens will be a suitable source for potential probiotic strains for chicken and poultry.

As a result of the growing interest in probiotics, many purported probiotic products have been marketed without proper studies on the probiotic properties of the strains, giving rise to problems of inconsistent efficacy of the products. Several studies have reported misidentification or mislabeling of probiotic species or presence of unspecified species in many commercial probiotic products (Hamilton-Miller and Shah, 1996; Canganella *et al.*, 1997; Klein *et al.*, 1998; Hamilton-Miller *et al.*, 1999; Schillinger, 1999). Since the properties of probiotic are strain specific, the quality of products is closely linked to the individual strains in the products, thus, they should be correctly identified, and their probiotic properties properly studied. In 2001, FAO/WHO produced a set of guidelines for the evaluation of probiotics in food in which they recommended that every potential probiotic strain must be correctly identified, followed by various *in vitro* assays to investigate its functional properties and *in vivo* trials for its safety. This is because probiotic properties are strain specific and cannot be extrapolated to the whole genus or species.

Thus, the present study was initiated to isolate, identify and characterize some new lactic acid bacterial strains from human milk, infant feces, fermented grapes and dates and chicken intestine with a view to exploit them as potential probiotic supplements for

humans and chickens. *In vitro* studies on the bioactivities and good probiotic properties of the isolated bacterial strains were first carried out for selection of potential probiotic candidates, followed by *in vivo* studies using animal models to evaluate their safety and efficacy.

The specific objectives of this study were:

- i) To isolate some new lactic acid bacterial strains from human milk, infant feces, fermented grapes and dates and chicken intestines
- ii) To identify the isolated bacterial strains using morphological, biochemical and molecular characteristics
- iii) To assess (*in vitro*) the characteristics associated with survival of the isolated bacterial strains in the GIT
- iv) To investigate (*in vitro*) the bioactivities of the isolated bacterial strains
- v) To evaluate the efficacy of selected probiotic strains intended for chickens using a broiler chicken trial
- vi) To study the safety of selected probiotic strains intended for humans using a rat model

REFERENCES

- AACC (2001). American Association for Clinical Chemistry. Lab Tests Online. www.labtestsonline.org
- Abd El-Gawad, I.A., El-Sayed, E.M., Hafez, S.A., El-Zeini, H.M. and Saleh, F.A. (2005). The hypocholesterolaemic effect of milk-yoghurt and soy-yoghurt containing bifidobacteria in rats fed on a cholesterol-enriched diet. *International Dairy Journal*. 15: 37-44.
- Abotsi, W.K.M., Ainooson, G.K. and Gyasi, E.B. (2011). Acute and sub-acute toxicity studies of the ethanolic extract of the aerial parts of *Hillieria Latifolia* (Lam.) H. Walt.(Phytolaccaceae) in rodents. *West African Journal of Pharmacy*. 22: 27-35.
- Abubakr, M.A.S., Hassan, Z., Imdakim, M.M.A. and Sharifah, N. (2012). Antioxidant activity of lactic acid bacteria (LAB) fermented skim milk as determined by 1, 1-diphenyl-2-picrylhydrazyl (DPPH) and ferrous chelating activity (FCA). *African Journal of Microbiology Research*. 6: 6358-6364.
- Afsharmanesh, M., Sadaghi, B. and Silversides, F.G. (2011). Influence of supplementation of prebiotic, probiotic, and antibiotic to wet-fed wheat-based diets on growth, ileal nutrient digestibility, blood parameters, and gastrointestinal characteristics of broiler chickens. *Comparative Clinical Pathology*. 22: 245-251.
- Ahn, Y.T., Kim, G.B., Lim, K.S., Baek, Y.J. and Kim, H.U. (2003). Deconjugation of bile salts by *Lactobacillus acidophilus* isolates. *International Dairy Journal*. 13: 303-311.
- Ahotupa, M., Saxelin, M. and Korpela, R. (1996). Antioxidative properties of *Lactobacillus GG*. *Nutrition Today*. 31: 262-265.
- Aldridge, C., Jones, P.W., Gibson, S., Lanham, J., Meyer, M., Vannest, R. and Charles, R. (1977). Automated microbiological detection/identification system. *Journal of Clinical Microbiology*. 6: 406-413.
- Ali, Q.S., Farid, A.J., Kabeir, B.M., Zambari, S., Shuhaimi, M., Ghazali, H.M. and Yazid, A.M. (2008). Adhesion properties of *Bifidobacterium pseudocatenulatum* G4 and *Bifidobacterium Longum* BB536 on HT-29 human epithelium cell line at different times and pH. *International Journal of Biological and Medical Sciences*. 3: 267-272.
- Anderson, J.W. and Gilliland, S.E. (1999). Effect of fermented milk (yogurt) containing *Lactobacillus acidophilus* L1 on serum cholesterol in hypercholesterolemic humans. *Journal of the American College of Nutrition*. 18: 43-50.

- Andrighetto, C., de Dea, P., Lombardi, A., Neviani, E., Rossetti, L. and Giraffa, G. (1998). Molecular identification and cluster analysis of homofermentative thermophilic lactobacilli isolated from dairy products. *Research in Microbiology*. 149: 631-643.
- Annuk, H., Shchepetova, J., Kullisaar, T., Songisepp, E., Zilmer, M. and Mikelsaar, M. (2003). Characterization of intestinal lactobacilli as putative probiotic candidates. *Journal of Applied Microbiology*. 94: 403-412.
- Arena, M.E. and Manca de Nadra, M.C. (2001). Biogenic amine production by *Lactobacillus*. *Journal of Applied Microbiology*. 90: 158-162.
- Armstrong, D. and Browne, R. (1994). The analysis of free radicals, lipid peroxides, antioxidant enzymes and compounds related to oxidative stress as applied to the clinical chemistry laboratory. *Advances in Experimental Medicine and Biology*. 366: 43-58.
- Arora, G., Lee, B.H. and Lamoureux, M. (1990). Characterization of enzyme profiles of *Lactobacillus casei* species by a rapid API ZYM System. *Journal of Dairy Science*. 73: 264-273.
- Aroutcheva, A., Gariti, D., Simon, M., Shott, S., Faro, J., Simoes, J.A., Gurguis, A. and Faro, S. (2001). Defense factors of vaginal lactobacilli. *American Journal of Obstetrics and Gynecology*. 185: 375-379.
- Ashayerizadeh, O., Dastar, B., Shargh, M.S., Ashayerizadeh, A. and Mamooee, M. (2009). Influence of antibiotic, prebiotic and probiotic supplementation to diets on carcass characteristics, hematological Indices and internal organ size of young broiler chickens. *Journal of Animal and veterinary Advances*. 8: 1772-1776.
- Ashelford, K.E., Chuzhanova, N.A., Fry, J.C., Jones, A.J. and Weightman, A.J. (2005). At least 1 in 20 16S rRNA sequence records currently held in public repositories is estimated to contain substantial anomalies. *Applied and Environmental Microbiology*. 71: 7724-7736.
- ATCC (1925). American Type Culture Collection. <http://www.atcc.org>
- Awad, W.A., Ghareeb, K., Abdel-Raheem, S. and Bohm, J. (2009). Effects of dietary inclusion of probiotic and synbiotic on growth performance, organ weights, and intestinal histomorphology of broiler chickens. *Poultry Science*. 88: 49-56.
- Barnby-Smith, F.M. (1992). Bacteriocins: applications in food preservation. *Trends in Food Science and Technology*. 3: 133-137.
- Bartosch, S., Woodmansey, E.J., Paterson, J.C.M., McMurdo, M.E.T. and Macfarlane, G.T. (2005). Microbiological effects of consuming a synbiotic containing *Bifidobacterium bifidum*, *Bifidobacterium lactis*, and oligofructose in elderly

persons, determined by real-time polymerase chain reaction and counting of viable bacteria. *Clinical Infectious Diseases*. 40: 28-37.

- Beachey, E.H. (1981). Bacterial adherence: adhesin-receptor interactions mediating the attachment of bacteria to mucosal surfaces. *Journal of Infectious Diseases*. 143: 325-345.
- Begley, M., Sleator, R.D., Gahan, C.G.M. and Hill, C. (2005). Contribution of three bile-associated loci, bsh, pva, and btlB, to gastrointestinal persistence and bile tolerance of *Listeria monocytogenes*. *Infection and Immunity*. 73: 894-904.
- Begley, M., Hill, C. and Gahan, C.G.M. (2006). Bile salt hydrolase activity in probiotics. *Applied and Environmental Microbiology*. 72: 1729-1738.
- Belviso, S., Giordano, M., Dolci, P. and Zeppa, G. (2009). *In vitro* cholesterol-lowering activity of *Lactobacillus plantarum* and *Lactobacillus paracasei* strains isolated from the Italian Castelmagno PDO cheese. *Dairy Science and Technology*. 89: 169-176.
- Benzie, I.F.F. and Strain, J.J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. *Analytical Biochemistry*. 239: 70-76.
- Berg, R.D. (1998). Probiotics, prebiotics or conbiotics?. *Trends in Microbiology*. 6: 89-92.
- Bernet, M.F., Brassart, D., Neeser, J.R. and Servin, A.L. (1994a). *Lactobacillus acidophilus* LA 1 binds to cultured human intestinal cell lines and inhibits cell attachment and cell invasion by enterovirulent bacteria. *Gut*. 35: 483-489.
- Bernet, M.F., Brassart, D., Neeser, J.R. and Servin, A.L. (1994b). Adhesion of human bifidobacterial strains to cultured human intestinal epithelial cells and inhibition of enteropathogen-cell interactions. *Applied and Environmental Microbiology*. 59: 4121-4128.
- Bernstein, H., Bernstein, C., Payne, C.M., Dvorakova, K. and Garewal, H. (2005). Bile acids as carcinogens in human gastrointestinal cancers. *Mutation Research*. 589: 47-65.
- Berrada, N., Lemeland, J.F., Laroche, G., Thouvenot, P. and Piaia, M. (1991). *Bifidobacterium* from fermented milks: survival during gastric transit. *Journal of Dairy Science*. 74: 409-413.
- Biffi, A., Coradini, D., Larsen, R., Riva, L. and Di Fronzo, G. (1997). Antiproliferative effect of fermented milk on the growth of a human breast cancer cell line. *Nutrition and Cancer*. 28: 93-99.

- Bilige, M., Liu, W., Rina, W., Wang, L., Sun, T., Wang, J., Li, H. and Zhang, H. (2009). Evaluation of potential probiotics properties of the screened lactobacilli isolated from home-made koumiss in Mongolia. *Annals of Microbiology*. 59: 493-498.
- Boivin-Jahns, V., Bianchi, A., Ruimy, R., Garcin, J., Daumas, S. and Christen, R. (1995). Comparison of phenotypical and molecular methods for the identification of bacterial strains isolated from a deep subsurface environment. *Applied and Environmental Microbiology*. 61: 3400-3406.
- Boonkumklao, P., Kongthong, P. and Assavanig, A. (2006). Acid and bile tolerance of *Lactobacillus thermotolerans*, a novel species isolated from chicken feces. *Kasetsart Journal - Natural Science*. 40: 13-17.
- Borriello, S.P., Hammes, W.P., Holzappel, W., Marteau, P., Schrezenmeir, J., Vaara, M. and Valtonen, V. (2003). Safety of probiotics that contain lactobacilli or bifidobacteria. *Clinical Infectious Diseases*. 36: 775-780.
- Bouzaine, T., Dauphin, R.D., Thonart, P., Urdaci, M.C. and Hamdi, M. (2005). Adherence and colonization properties of *Lactobacillus rhamnosus* TB1, a broiler chicken isolate. *Letters in Applied Microbiology*. 40: 391-396.
- Canganella, F., Paganini, S., Ovidi, M., Vettraino, A.M., Bevilacqua, L., Massa, S. and Trovatelli, L.D. (1997). A microbiological investigator on probiotic pharmaceutical products used for human health. *Microbiological Research*. 152: 171-179.
- Cats, A., Kuipers, E.J., Bosschaert, M.A.R., Pot, R.G.J., Vandenbroucke-Grauls, C.M.J.E. and Kusters, J.G. (2003). Effect of frequent consumption of a *Lactobacillus casei*-containing milk drink in *Helicobacter pylori*-colonized subjects. *Alimentary Pharmacology and Therapeutics*. 17: 429-435.
- Chabot, S., Yu, H.L., De Leseleuc, L., Cloutier, D., Van Calsteren, M.R., Lessard, M., Roy, D., Lacroix, M. and Oth, D. (2001). Exopolysaccharides from *Lactobacillus rhamnosus* RW-9595M stimulate TNF, IL-6 and IL-12 in human and mouse cultured immunocompetent cells, and IFN- γ in mouse splenocytes. *Dairy Science and Technology*. 81: 683-697.
- Chan, E.S. and Zhang, Z. (2005). Bioencapsulation by compression coating of probiotic bacteria for their protection in an acidic medium. *Process Biochemistry*. 40: 3346-3351.
- Charteris, W.P., Kelly, P.M., Morelli, L. and Collins, J.K. (1998). Development and application of an *in vitro* methodology to determine the transit tolerance of potentially probiotic *Lactobacillus* and *Bifidobacterium* species in the upper human gastrointestinal tract. *Journal of Applied Microbiology*. 84: 759-768.

- Chiang, Y.R., Ismail, W., Heintz, D., Schaeffer, C., Van Dorsselaer, A. and Fuchs, G. (2008). Study of anoxic and oxic cholesterol metabolism by *Sterolibacterium denitrificans*. *Journal of Bacteriology*. 190: 905-914.
- Cho, I.J., Lee, N.K. and Hahm, Y.T. (2009). Characterization of *Lactobacillus* spp. isolated from the feces of breast-feeding piglets. *Journal of Bioscience and Bioengineering*. 108: 194-198.
- Choi, S.S., Kim, Y., Han, K.S., You, S., Oh, S. and Kim, S.H. (2006). Effects of *Lactobacillus* strains on cancer cell proliferation and oxidative stress *in vitro*. *Letters in Applied Microbiology*. 42: 452-458.
- Coconnier-Polter, M.H., Vanessa, L.L.M. and Servin, A.L. (2005). A *Lactobacillus acidophilus* strain of human gastrointestinal microbiota origin elicits killing of enterovirulent *Salmonella enterica* serovar Typhimurium by triggering lethal bacterial membrane damage. *Applied and Environmental Microbiology*. 71: 6115-6120.
- Cole, C.B., Anderson, P.H., Philips, S.M., Fuller, R. and Hewitt, D. (1984). The effect of yoghurt on the growth, lactose-utilizing gut organisms and β -glucuronidase activity of caecal contents of a lactose-fed, lactase-deficient animal. *Food Microbiology*. 1: 217-222.
- Collins, J.K., Thornton, G. and Sullivan, G.O. (1998). Selection of probiotic strains for human applications. *International Dairy Journal*. 8: 487-490.
- Collins, J.W., La Ragione, R.M., Woodward, M.J. and Searle, L.E.J. (2009). Application of prebiotics and probiotics in livestock. *Prebiotics and Probiotics Science and Technology*. 1: 1123-1192.
- Conway, P.L. (1996). Selection criteria for probiotic microorganisms. *Asia Pacific Journal of Clinical Nutrition*. 5: 10-14.
- Corsetti, A., Gobbetti, M. and Smacchi, E. (1996). Antibacterial activity of sourdough lactic acid bacteria: isolation of a bacteriocin-like inhibitory substance from *Lactobacillus sanfrancisco* C57. *Food Microbiology*. 13: 447-456.
- Corsetti, A., Gobbetti, M., Rossi, J. and Damiani, P. (1998). Antimould activity of sourdough lactic acid bacteria: identification of a mixture of organic acids produced by *Lactobacillus sanfrancisco* CB1. *Applied Microbiology and Biotechnology*. 50: 253-256.
- Corsetti, A. and Settanni, L. (2007). Lactobacilli in sourdough fermentation. *Food Research International*. 40: 539-558.
- Corzo, G. and Gilliland, S.E. (1999). Bile salt hydrolase activity of three strains of *Lactobacillus acidophilus*. *Journal of Dairy Science*. 82: 472-480.

- Cukovic-Cavka, S., Likic, R., Francetic, I., Rustemovic, N., Opacic, M. and Vucelic, B. (2006). *Lactobacillus acidophilus* as a cause of liver abscess in a NOD2/CARD15-positive patient with crohn's disease. *Digestion*. 73: 107-110.
- Dalloul, R.A., Lillehoj, H.S., Shellem, T.A. and Doerr, J.A. (2003). Enhanced mucosal immunity against *Eimeria acervulina* in broilers fed a *Lactobacillus*-based probiotic. *Poultry Science*. 82: 62-66.
- Damia, G. and Broggin, M. (2004). Improving the selectivity of cancer treatments by interfering with cell response pathways. *European Journal of Cancer*. 40: 2550-2559.
- Dash, S.K. (1980). Selection criteria for probiotics. *International Journal of Systematic Bacteriology*. 30: 225-420.
- Dashkevich, M.P. and Feighner, S.D. (1989). Development of a differential medium for bile salt hydrolase-active *Lactobacillus* spp. *Applied and Environmental Microbiology*. 55: 11-16.
- Davies, F.L. and Law, B.A. (1984). *Advances in the microbiology and biochemistry of cheese and fermented milk*. 260 pages. Elsevier Applied Science Publishers Ltd.
- De Preter, V., Vanhoutte, T., Huys, G., Swings, J., De Vuyst, L., Rutgeerts, P. and Verbeke, K. (2007). Effects of *Lactobacillus casei* Shirota, *Bifidobacterium breve*, and oligofructose-enriched inulin on colonic nitrogen-protein metabolism in healthy humans. *American Journal of Physiology-Gastrointestinal and Liver Physiology*. 292: 358-368.
- De Smet, I., Van Hoorde, L., Vande Woestyne, M., Christiaens, H. and Verstraete, W. (1995). Significance of bile salt hydrolytic activities of lactobacilli. *Journal of Applied Bacteriology*. 79: 292-301.
- Del Piano, M., Morelli, L., Strozzi, G.P., Allesina, S., Barba, M., Deidda, F., Lorenzini, P., Ballare, M., Montino, F. and Orsello, M. (2006). Probiotics: from research to consumer. *Digestive and Liver Disease*. 38: 248-255.
- Dicks, L.M.T., Heunis, T.D.J., Staden, D.A., Brand, A., Noll, K.S. and Chikindas, M.L. (2011). Medical and personal care applications of bacteriocins produced by lactic acid bacteria. In: *Prokaryotic antimicrobial peptides*, D. Drider and S. Rebuffat (Eds.), pp. 391-421. Springer New York.
- Donohue, D.C. and Salminen, S. (1996). Safety of probiotic bacteria. *Asia Pacific Journal of Clinical Nutrition*. 5: 25-28.
- Drasar, B.S. and Hill, M.J. (1974). *Human intestinal flora*. 263 pages. Academic Press Ltd.

- Dunne, C., O'Mahony, L., Murphy, L., Thornton, G., Morrissey, D., O'Halloran, S., Feeney, M., Flynn, S., Fitzgerald, G. and Daly, C. (2001). *In vitro* selection criteria for probiotic bacteria of human origin: correlation with *in vivo* findings. *The American Journal of Clinical Nutrition*. 73: 386-392.
- Ehrmann, M.A., Kurzak, P., Bauer, J. and Vogel, R.F. (2002). Characterization of lactobacilli towards their use as probiotic adjuncts in poultry. *Journal of Applied Microbiology*. 92: 966-975.
- Elmer, G.W., Surawicz, C.M. and Mc Farland, L.V. (1996). Biotherapeutic agents. *Journal of the American Medical Association*. 275: 870-876.
- Erwin, E.S., Marco, G.J. and Emery, E.M. (1961). Volatile fatty acid analyses of blood and rumen fluid by gas chromatography. *Journal of Dairy Science*. 44: 1768-1771.
- Ewaschuk, J.B., Walker, J.W., Diaz, H. and Madsen, K.L. (2006). Bioproduction of conjugated linoleic acid by probiotic bacteria occurs *in vitro* and *in vivo* in mice. *Journal of Nutrition*. 136: 1483-1487.
- FAO/WHO (Food and Agriculture Organization/World Health Organization) (2001). Health and nutritional properties of probiotics in food including powder milk with live lactic acid bacteria: Report of a Joint FAO/WHO expert consultation on evaluation of health and nutritional properties of probiotics in food including powder milk with live lactic acid bacteria. 34 pages.
- FAO/WHO (Food and Agriculture Organization/World Health Organization) (2007). Guidelines for the evaluation of probiotics in food: Report of a Joint FAO/WHO working group on drafting guidelines for the evaluation of probiotics in food. 11 pages.
- Farmer, E.H., Bloomfield, G.F., Sundralingam, A. and Sutton, D.A. (1942). The course and mechanism of autoxidation reactions in olefinic and polyolefinic substances, including rubber. *Transactions of the Faraday Society*. 38: 348-356.
- Federation of Animal Science Societies (1999). Guide for the care and use of agricultural animals in research and teaching. 169 pages.
- Feedap, P. (2005). Opinion of the scientific committee on a request from EFSA related to a generic approach to the safety assessment by EFSA of microorganisms used in food/feed and the production of food/feed additives. *European Food Safety Authority*. 226: 1-12.
- Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution*. 39: 783-791.

- Fernandez, M.F., Boris, S. and Barbes, C. (2003). Probiotic properties of human lactobacilli strains to be used in the gastrointestinal tract. *Journal of Applied Microbiology*. 94: 449-455.
- Ferrari, C.K.B. and Torres, E. (2003). Biochemical pharmacology of functional foods and prevention of chronic diseases of aging. *Biomedecine and Pharmacotherapy*. 57: 251-260.
- Flahaut, S., Frere, J., Boutibonnes, P. and Auffray, Y. (1996). Comparison of the bile salts and sodium dodecyl sulfate stress responses in *Enterococcus faecalis*. *Applied and Environmental Microbiology*. 62: 2416-2420.
- Floch, M.H., Binder, H.J., Filburn, B. and Gershengoren, W. (1972). The effect of bile acids on intestinal microflora. *The American Journal of Clinical Nutrition*. 25: 1418-1426.
- Fontaine, E.A., Claydon, E. and Taylor-Robinson, D. (1996). Lactobacilli from women with or without bacterial vaginosis and observations on the significance of hydrogen peroxide. *Microbial Ecology in Health and Disease*. 9: 135-141.
- Frahm, E. and Obst, U. (2003). Application of the fluorogenic probe technique (TaqMan PCR) to the detection of *Enterococcus* spp. and *Escherichia coli* in water samples. *Journal of Microbiological Methods*. 52: 123-131.
- Fukushima, M. and Nakano, M. (1995). The effect of a probiotic on faecal and liver lipid classes in rats. *British Journal of Nutrition*. 73: 701-710.
- Fuller, R. (1973). Ecological studies on the *Lactobacillus* flora associated with the crop epithelium of the fowl. *Applied Microbiology*. 36: 131-139.
- Fuller, R. (1989). Probiotics in man and animals. *Applied Bacteriology*. 66: 365-378.
- Fuller, R. (2001). The chicken gut microflora and probiotic supplements. *Poultry Science*. 38: 189-196.
- Gadelle, D., Raibaud, P. and Sacquet, E. (1985). Beta-glucuronidase activities of intestinal bacteria determined both *in vitro* and *in vivo* in gnotobiotic rats. *Applied and Environmental Microbiology*. 49: 682-685.
- Garrity, G.M., Bell, J.A. and Lilburn, T.G. (2004). Taxonomic outline of the prokaryotes. In: *Bergey's manual of systematic bacteriology*. Springer New York. doi: 10.1007/bergeysoutline200405 (Online).
- Gasser, F. (1994). Safety of lactic acid bacteria and their occurrence in human clinical infections. *Bulletin de L'Institut Pasteur*. 92: 45-67.

- Gaudana, S.B., Dhanani, A.S. and Bagchi, T. (2010). Probiotic attributes of *Lactobacillus* strains isolated from food and of human origin. *British Journal of Nutrition*. 103: 1620-1628.
- GIA (Global Industry Analysts) (2012). Probiotics: a global strategic business report. 381 pages.
- Gibson, G.R. and Roberfroid, M.B. (1995). Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *Journal of Nutrition*. 125: 1401-1412.
- Gilliland, S.E. and Speck, M.L. (1977). Deconjugation of bile acids by intestinal lactobacilli. *Applied and Environmental Microbiology*. 33: 15-18.
- Gilliland, S.E., Staley, T.E. and Bush, L.J. (1984). Importance of bile tolerance of *Lactobacillus acidophilus* used as a dietary adjunct. *Journal of Dairy Science*. 67: 3045-3051.
- Gilliland, S.E., Nelson, C.R. and Maxwell, C. (1985). Assimilation of cholesterol by *Lactobacillus acidophilus*. *Applied and Environmental Microbiology*. 49: 377-381.
- Goldin, B.R. and Gorbach, S.L. (1984). Alterations of the intestinal microflora by diet, oral antibiotics, and *Lactobacillus*: decreased production of free amines from aromatic nitro compounds, azo dyes, and glucuronides. *Journal of the National Cancer Institute*. 73: 689-695.
- Gopal, P.K., Prasad, J., Smart, J. and Gill, H.S. (2001). In vitro adherence properties of *Lactobacillus rhamnosus* DR20 and *Bifidobacterium lactis* DR10 strains and their antagonistic activity against an enterotoxigenic *Escherichia coli*. *International journal of food microbiology*. 67: 207-216.
- Gorbach, S.L. (1986). Bengt E. Gustafsson memorial lecture. Function of the normal human microflora. *Scandinavian Journal of Infectious Diseases*. 49: 17-30.
- Grajek, W., Olejnik, A. and Sip, A. (2005). Probiotics, prebiotics and antioxidants as functional foods. *Acta Biochimica Polonica*. 52: 665-671.
- Guarner, F. and Malagelada, J.R. (2003). Gut flora in health and disease. *The Lancet*. 361: 512-519.
- Gunal, M., Yayli, G., Kaya, O., Karahan, N. and Sulak, O. (2006). The effects of antibiotic growth promoter, probiotic or organic acid supplementation on performance, intestinal microflora and tissue of broilers. *International Journal of Poultry Science*. 5: 149-155.

- Halasz, A., Barath, A., Simon-Sarkadi, L. and Holzapfel, W. (1994). Biogenic amines and their production by microorganisms in food. *Trends in Food Science and Technology*. 5: 42-49.
- Hall, T.A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*. 41: 95-98.
- Halliwell, B. and Gutteridge, J.M.C. (1984). Oxygen toxicity, oxygen radicals, transition metals and disease. *Biochemical Journal*. 219: 1-14.
- Halliwell, B. and Gutteridge, J.M.C. (1989). *Free radicals in biology and medicine*. pages. Clarendon Press Oxford.
- Hamilton-Miller, J.M.T. and Shah, S.a.S., C.T. (1996). "Probiotic" remedies are not what they seem. *British Medical Journal*. 312: 55-56.
- Hamilton-Miller, J.M.T., Shah, S. and Winkler, J.T. (1999). Public health issues arising from microbiological and labeling quality of foods and supplements containing probiotic microorganisms. *Public Health Nutrition*. 2: 223-229.
- Han, I.K., Lee, S.C. and Lee, J.H. (1984). Studies on the growth promoting effects of probiotics, 1: the effects of *Lactobacillus sporogenes* on the growing performance and the changes in microbial flora of the feces and intestinal contents of the broiler chicks. *Korean Journal of Animal Sciences*. 26: 150-157.
- Havenaar, R. and Huis in't Veld, J.H.J. (1992). Probiotics: a general view. *The lactic acid bacteria*. 1: 151-170.
- Haza, A.I., Zabala, A. and Morales, P. (2004). Protective effect and cytokine production of a *Lactobacillus plantarum* strain isolated from ewes' milk cheese. *International Dairy Journal*. 14: 29-38.
- Hechard, Y. and Sahl, H.G. (2002). Mode of action of modified and unmodified bacteriocins from Gram-positive bacteria. *Biochimie*. 84: 545-557.
- Heller, K.J. (2001). Probiotic bacteria in fermented foods: product characteristics and starter organisms. *The American Journal of Clinical Nutrition*. 73: 374-379.
- Hennequin, C., Kauffmann-Lacroix, C., Jobert, A., Viard, J.P., Ricour, C., Jacquemin, J.L. and Berche, P. (2000). Possible role of catheters in *Saccharomyces boulardii* fungemia. *European Journal of Clinical Microbiology and Infectious Diseases*. 19: 16-20.
- Heyman, M. and Menard, S. (2002). Probiotic microorganisms: how they affect intestinal pathophysiology. *Cellular and Molecular Life Sciences*. 59: 1151-1165.

- Hill, G.B., Eschenbach, D.A. and Holmes, K.K. (1984). Bacteriology of the vagina. *Scandinavian Journal of Urology and Nephrology*. 86: 23-39.
- Hofmann, A.F. and Roda, A. (1984). Physicochemical properties of bile acids and their relationship to biological properties: an overview of the problem. *The Journal of Lipid Research*. 25: 1477-1489.
- Hofmann, A.F. and Mysels, K.J. (1992). Bile acid solubility and precipitation *in vitro* and *in vivo*: the role of conjugation, pH, and Ca²⁺ ions. *The Journal of Lipid Research*. 33: 617-626.
- Hofmann, A.F. (1999). Bile acids: the good, the bad, and the ugly. *Physiology*. 14: 24-29.
- Holzappel, W.H., Haberer, P., Geisen, R., Bjorkroth, J. and Schillinger, U. (2001). Taxonomy and important features of probiotic microorganisms in food and nutrition. *American Society for Clinical Nutrition*. 73: 365-373.
- Holzappel, W.H. and Schillinger, U. (2002). Introduction to pre-and probiotics. *Food Research International*. 35: 109-116.
- Hopkins, M.J., Macfarlane, G.T., Furrer, E., Fite, A. and Macfarlane, S. (2005). Characterisation of intestinal bacteria in infant stools using real time PCR and northern hybridisation analyses. *FEMS Microbiology Ecology*. 54: 77-85.
- Huang, D., Ou, B. and Ronald, L. (2005). The chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry*. 53: 1841-1856.
- Huang, Y., Kotula, L. and Adams, M.C. (2003). The *in vivo* assessment of safety and gastrointestinal survival of an orally administered novel probiotic, *Propionibacterium jensenii* 702, in a male Wistar rat model. *Food and Chemical Toxicology*. 41: 1781-1787.
- Huber, T., Faulkner, G. and Hugenholtz, P. (2004). Bellerophon: a program to detect chimeric sequences in multiple sequence alignments. *Bioinformatics*. 20: 2317-2319.
- ISAPP (International Scientific Association for Probiotics and Prebiotics) (2009). Clarification of the definition of a probiotic. 2 pages.
- Isolauri, E., Sutas, Y., Kankaanpaa, P., Arvilommi, H. and Salminen, S. (2001). Probiotics: effects on immunity. *The American Journal of Clinical Nutrition*. 73: 444-450.
- Isolauri, E., Kirjavainen, P.V. and Salminen, S. (2002). Probiotics: a role in the treatment of intestinal infection and inflammation?. *Gut*. 50: 54-59.

- Jacobsen, C.N., Rosenfeldt Nielsen, V., Hayford, A.E., Moller, P.L., Michaelsen, K.F., Parregaard, A., Sandstrom, B., Tvede, M. and Jakobsen, M. (1999). Screening of probiotic activities of forty-seven strains of *Lactobacillus* spp. by *in vitro* techniques and evaluation of the colonization ability of five selected strains in humans. *Applied and Environmental Microbiology*. 65: 4949-4956.
- Jain, S., Yadav, H. and Ravindra Sinha, P. (2009). Antioxidant and cholesterol assimilation activities of selected lactobacilli and lactococci cultures. *Journal of Dairy Research*. 76: 385-391.
- Jamuna, M. and Jeevaratnam, K. (2004). Isolation and characterization of lactobacilli from some traditional fermented foods and evaluation of the bacteriocins. *Journal of General and Applied Microbiology*. 50: 79-90.
- Jin, L.Z., Ho, Y.W., Abdullah, N. and Jalaludin, S. (1996). Influence of dried *Bacillus subtilis* and lactobacilli cultures on intestinal microflora and performance in broilers. *Asian - Australasian Journal of Animal Sciences*. 9: 397-404.
- Jin, L.Z. (1996). Studies on the mechanisms and utilization of probiotics (direct-fed microbials) in broilers. PhD thesis, University Putra Malaysia. Kuala Lumpur.
- Jin, L.Z., Ho, Y.W., Abdullah, N. and Jalaludin, S. (1998a). Acid and bile tolerance of *Lactobacillus* isolated from chicken intestine. *Letters in Applied Microbiology*. 27: 183-185.
- Jin, L.Z., Ho, Y.W., Abdullah, N. and Jalaludin, S. (1998b). Growth performance, intestinal microbial populations, and serum cholesterol of broilers fed diets containing *Lactobacillus* cultures. *Poultry Science*. 77: 1259-1265.
- Jin, L.Z., Ho, Y.W., Abdullah, N., Ali, M.A. and Jalaludin, S. (1998c). Effects of adherent *Lactobacillus* cultures on growth, weight of organs and intestinal microflora and volatile fatty acids in broilers. *Animal Feed Science and Technology*. 70: 197-209.
- Jin, L.Z., Ho, Y.W., Abdullah, N. and Jalaludin, S. (2000). Digestive and bacterial enzyme activities in broilers fed diets supplemented with *Lactobacillus* cultures. *Poultry Science*. 79: 886-891.
- Johansson, M.L., Molin, G., Jeppsson, B., Nobaek, S., Ahrne, S. and Bengmark, S. (1993). Administration of different *Lactobacillus* strains in fermented oatmeal soup: *in vivo* colonization of human intestinal mucosa and effect on the indigenous flora. *Applied and Environmental Microbiology*. 59: 15-20.
- Joosten, H.M.L.J. (1988). *Conditions allowing the formation of biogenic amines in cheese*. 133 pages. Landbouwwuniversiteit.

- Kabir, S.M. (2009). The role of probiotics in the poultry industry. *International Journal of Molecular Sciences*. 10: 3531-3546.
- Kalavathy, R., Abdullah, N., Jalaludin, S. and Ho, Y.W. (2003). Effects of *Lactobacillus* cultures on growth performance, abdominal fat deposition, serum lipids and weight of organs of broiler chickens. *British Poultry Science*. 44: 139-144.
- Kalavathy, R., Abdullah, N., Wong, M.C.V.L., Karuthan, C. and Ho, Y.W. (2009). Bile salt deconjugation and cholesterol removal from media by *Lactobacillus* strains used as probiotics in chickens. *Journal of the Science of Food and Agriculture*. 90: 65-69.
- Kalbande, V.H., Gaffar, M.A. and Deshmukh, S.V. (1992). Effect of probiotic and nitrofurin on performance of growing commercial pullets. *Indian Journal of Poultry Science*. 27: 116-117.
- Kandell, R.L. and Bernstein, C. (1991). Bile salt/acid induction of DNA damage in bacterial and mammalian cells: implications for colon cancer. *Nutrition and Cancer*. 16: 227-238.
- Kandler, O. and Weiss, N. (1986). Genus *Lactobacillus*. In: *Bergey's manual of systematic bacteriology*, P.H.A. Sneath, N.S. Mair, M.E. Sharpe and J.G. Holt (Eds.), pp. 1208-1234. Williams and Wilkins.
- Kashket, E.R. (1987). Bioenergetics of lactic acid bacteria: cytoplasmic pH and osmotolerance. *FEMS Microbiology Letters*. 46: 233-244.
- Katiyar, S.K. and Mukhtar, H. (1997). Tea antioxidants in cancer chemoprevention. *Journal of Cellular Biochemistry*. 67: 59-67.
- Khaksefidi, A. and Ghoorchi, T. (2006). Effect of probiotic on performance and immunocompetence in broiler chicks. *Poultry Science*. 43: 296-300.
- Khunajakr, N., Wongwicharn, A., Moonmangmee, D. and Tantipaiboonvut, S. (2008). Screening and identification of lactic acid bacteria producing antimicrobial compounds from pig gastrointestinal tracts. *KMITL Science and Technology Journal*. 8: 8-17.
- Kim, C.J., Namkung, H., An, M.S. and Paik, I.K. (1988). Supplementation of probiotics to broiler diets containing moldy corn. *Korean Journal of Animal Sciences*. 30: 542-548.
- Kim, Y., Whang, J.Y., Whang, K.Y., Oh, S. and Kim, S.H. (2008). Characterization of the cholesterol-reducing activity in a cell-free supernatant of *Lactobacillus acidophilus* ATCC 43121. *Bioscience, Biotechnology, and Biochemistry*. 72: 1483-1490.

- Kimoto, H., Ohmomo, S. and Okamoto, T. (2002). Cholesterol removal from media by lactococci. *Journal of Dairy Science*. 85: 3182-3188.
- Kishida, T., Taguchi, F., Feng, L., Tatsuguchi, A., Sato, J., Fujimori, S., Tachikawa, H., Tamagawa, Y., Yoshida, Y. and Kobayashi, M. (1997). Analysis of bile acids in colon residual liquid or fecal material in patients with colorectal neoplasia and control subjects. *Journal of Gastroenterology*. 32: 306-311.
- Kizerwetter-Swida, M. and Binek, M. (2005). Selection of potentially probiotic *Lactobacillus* strains towards their inhibitory activity against poultry enteropathogenic bacteria. *Polish Journal of Microbiology*. 54: 287-294.
- Klaenhammer, T.R. (1988). Bacteriocins of lactic acid bacteria. *Biochimie*. 70: 337-349.
- Klasing, K.C. (1998). *Comparative avian nutrition*. pages. Cab International.
- Klaver, F.A. and Van Der Meer, R. (1993). The assumed assimilation of cholesterol by lactobacilli and *Bifidobacterium bifidum* is due to their bile salt-deconjugating activity. *Applied and Environmental Microbiology*. 59: 1120-1124.
- Klayraung, S. and Okonogi, S. (2009). Antibacterial and antioxidant activities of acid and bile resistant strains of *Lactobacillus fermentum* isolated from miang. *Brazilian Journal of Microbiology*. 40: 757-766.
- Klein, G., Pack, A., Bonaparte, C. and Reuter, G. (1998). Taxonomy and physiology of probiotic lactic acid bacteria. *International Journal of Food Microbiology*. 41: 103-125.
- Knarreborg, A., Jensen, S.K. and Engberg, R.M. (2003). Pancreatic lipase activity as influenced by unconjugated bile acids and pH, measured *in vitro* and *in vivo*. *The Journal of Nutritional Biochemistry*. 14: 259-265.
- Koll, P., Mandar, R., Marcotte, H., Leibur, E., Mikelsaar, M. and Hammarstrom, L. (2008). Characterization of oral lactobacilli as potential probiotics for oral health. *Oral Microbiology and Immunology*. 23: 139-147.
- Kris-Etherton, P.M., Hecker, K.D., Bonanome, A., Coval, S.M., Binkoski, A.E., Hilpert, K.F., Griel, A.E. and Etherton, T.D. (2002). Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. *The American Journal of Medicine*. 113: 71-88.
- Kumar, H., Rangrez, A.Y., Dayananda, K.M., Atre, A.N., Patole, M.S. and Shouche, Y.S. (2011). *Lactobacillus plantarum* (VR1) isolated from an Ayurvedic medicine (Kutajarista) ameliorates *in vitro* cellular damage caused by *Aeromonas veronii*. *BMC Microbiology*. 11: 152-163.

- Lamson, D.W. and Brignall, M.S. (1999). Antioxidants in cancer therapy; their actions and interactions with oncologic therapies. *Alternative Medicine Review*. 4: 304-329.
- Lane, D.J. (1991). 16S/23S rRNA sequencing. In: *Nucleic acid techniques in bacterial systematics*, E. Stackebrandt and M. Goodfellow (Eds.), pp. John Wiley and Sons.
- Lankaputhra, W.E.V. and Shah, N.P. (1995). Survival of *Lactobacillus acidophilus* and *Bifidobacterium* spp in the presence of acid and bile salts. *Cultured Dairy Products Journal*. 30: 2-7.
- Lee, K.D., Jang, S., Baek, E.H., Kim, M.J., Lee, K.S., Shin, H.S., Chung, M.J., Kim, J.E., Lee, K.O. and Ha, N.J. (2009). Lactic acid bacteria affect serum cholesterol levels, harmful fecal enzyme activity, and fecal water content. *Lipids in Health and Disease*. 8: 21-29.
- Lee, Y.K. and Salminen, S. (1995). The coming of age of probiotics. *Trends in Food Science and Technology*. 6: 241-245.
- Leroy, F. and De-Vuyst, L. (2004). Lactic acid bacteria as functional starter cultures for the food fermentation industry. *Trends in Food Science and Technology*. 15: 67-78.
- Li, S., Zhao, Y., Zhang, L., Zhang, X., Huang, L., Li, D., Niu, C., Yang, Z. and Wang, Q. (2012). Antioxidant activity of *Lactobacillus plantarum* strains isolated from traditional Chinese fermented foods. *Food Chemistry* 135: 1914-1919.
- Liasi, S.A., Azmi, T.I., Hassan, M.D., Shuhaimi, M., Rosfarizan, M. and Ariff, A.B. (2009). Antimicrobial activity and antibiotic sensitivity of three isolates of lactic acid bacteria from fermented fish product, Budu. *Malaysian Journal of Microbiology*. 5: 33-37.
- Lilly, D.M. and Stillwell, R.H. (1965). Probiotics: growth-promoting factors produced by microorganisms. *Science*. 147: 747-748.
- Lin, J., Sahin, O., Michel, L.O. and Zhang, Q. (2003). Critical role of multidrug efflux pump CmeABC in bile resistance and *in vivo* colonization of *Campylobacter jejuni*. *Infection and Immunity*. 71: 4250-4259.
- Lin, M.Y. and Chang, F.J. (2000). Antioxidative effect of intestinal bacteria *Bifidobacterium longum* ATCC 15708 and *Lactobacillus acidophilus* ATCC 4356. *Digestive diseases and sciences*. 45: 1617-1622.
- Lindgren, S.E. and Dobrogosz, W.J. (1990). Antagonistic activities of lactic acid bacteria in food and feed fermentations. *FEMS Microbiology Letters*. 87: 149-163.
- Ling, W.H., Saxelin, M., Hanninen, O. and Salminen, S. (1994). Enzyme profile of *Lactobacillus* strain GG by a rapid API ZYM system: a comparison of intestinal bacterial strains. *Microbial Ecology in Health and Disease*. 7: 99-104.

- Link-Amster, H., Rochat, F., Saudan, K.Y., Mignot, O. and Aeschlimann, J.M. (1994). Modulation of a specific humoral immune response and changes in intestinal flora mediated through fermented milk intake. *FEMS Immunology and Medical Microbiology*. 10: 55-63.
- Liong, M.T. and Shah, N.P. (2005a). Bile salt deconjugation ability, bile salt hydrolase activity and cholesterol co-precipitation ability of lactobacilli strains. *International Dairy Journal*. 15: 391-398.
- Liong, M.T. and Shah, N.P. (2005b). Acid and bile tolerance and cholesterol removal ability of lactobacilli strains. *Journal of Dairy Science*. 88: 55-66.
- Liu, C.F., Tseng, K.C., Chiang, S.S., Lee, B.H., Hsu, W.H. and Pan, T.M. (2011a). Immunomodulatory and antioxidant potential of *Lactobacillus* exopolysaccharides. *Journal of the Science of Food and Agriculture*. 91: 2284–2291.
- Liu, C.T., Chu, F.J., Chou, C.C. and Yu, R.C. (2011b). Antiproliferative and anticytotoxic effects of cell fractions and exopolysaccharides from *Lactobacillus casei* 01. *Mutation Research*. 721: 157-162.
- Lloyd-Jones, D., Adams, R., Carnethon, M., De Simone, G., Ferguson, T.B., Flegal, K., Ford, E., Furie, K., Go, A. and Greenlund, K. (2009). Heart disease and stroke statistics-2009 update: a report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. *Circulation*. 119: 480-486.
- Lonkar, P., Harne, S.D., Kalorey, D. and Kurkure, N.V. (2005). Isolation, *in vitro* antibacterial activity, bacterial sensitivity and plasmid profile of Lactobacilli. *Asian - Australasian Journal of Animal sciences*. 18: 1336-1342.
- Lorenzo, J.M., Cachaldora, A., Fonseca, S., Gomez, M., Franco, I. and Carballo, J. (2010). Production of biogenic amines. *Meat Science*. 86: 684-691.
- Lye, H.S., Rusul, G. and Liong, M.T. (2010a). Removal of cholesterol by lactobacilli via incorporation and conversion to coprostanol. *Journal of Dairy Science*. 93: 1383-1392.
- Lye, H.S., Rahmat-Ali, G.R. and Liong, M.T. (2010b). Mechanisms of cholesterol removal by lactobacilli under conditions that mimic the human gastrointestinal tract. *International Dairy Journal*. 20: 169-175.
- Mack, D.R., Michail, S., Wei, S., McDougall, L. and Hollingsworth, M.A. (1999). Probiotics inhibit enteropathogenic *E. coli* adherence *in vitro* by inducing intestinal mucin gene expression. *American Journal of Physiology - Gastrointestinal and Liver Physiology*. 291: 203-210.

- Majamaa, H., Isolauri, E., Saxelin, M. and Vesikari, T. (1995). Lactic acid bacteria in the treatment of acute rotavirus gastroenteritis. *Journal of Pediatric Gastroenterology and Nutrition*. 20: 333-338.
- Majeed, M. and Prakash, L. (1998). *Lactospore: The Effective Probiotic*. 56 pages. NutriScience Publishers.
- Majidzadeh Heravi, R., Kermanshahi, H., Sankian, M., Nassiri, M.R., Heravi Moussavi, A., Roozbeh Nasiraii, L. and Varasteh, A.R. (2011). Screening of lactobacilli bacteria isolated from gastrointestinal tract of broiler chickens for their use as probiotic. *African Journal of Microbiology Research*. 5: 1858-1868.
- Mansoub, N.H. (2010). Effect of probiotic bacteria utilization on serum cholesterol and triglycerides contents and performance of broiler chickens. *Global Veterinaria*. 5: 184-186.
- Marteau, P., Gerhardt, M.F., Myara, A., Bouvier, E., Trivin, F. and Rambaud, J.C. (1995). Metabolism of bile salts by alimentary bacteria during transit in the human small intestine. *Microbial Ecology in Health and Disease*. 8: 151-157.
- Marteau, P. (2002). Probiotics in clinical conditions. *Clinical Reviews in Allergy and Immunology*. 22: 255-273.
- Martin, R., Langa, S., Reviriego, C., Jimenez, E., Marin, M.L., Olivares, M., Boza, J., Jimenez, J., Fernandez, L. and Xaus, J. (2004). The commensal microflora of human milk: new perspectives for food bacteriotherapy and probiotics. *Trends in Food Science and Technology*. 15: 121-127.
- Martin, R., Olivares, M., Marin, M.L., Fernandez, L., Xaus, J. and Rodriguez, J.M. (2005). Probiotic potential of 3 lactobacilli strains isolated from breast milk. *Journal of Human Lactation*. 21: 8-17.
- Mathara, J.M., Schillinger, U., Guigas, C., Franz, C., Kutima, P.M., Mbugua, S.K., Shin, H.K. and Holzappel, W.H. (2008). Functional characteristics of *Lactobacillus* spp. from traditional Maasai fermented milk products in Kenya. *International Journal of Food Microbiology*. 126: 57-64.
- Mayahi, M., Jalali, M.R. and Kiani, R. (2009). Effects of dietary probiotic supplementation on cholesterol and triglyceride levels in broiler chicks' sera. In proceeding of: *World Poultry Science Association (WPSA)*. 1: 589-591.
- McDonald, I.R., Kenna, E.M. and Murrell, J.C. (1995). Detection of methanotrophic bacteria in environmental samples with the PCR. *Applied and Environmental Microbiology*. 61: 116-121.

- Metchnikoff, E. (1907). Lactic acid as inhibiting intestinal putrefaction. In: *The prolongation of life: optimistic studies*, P. Chalmers Mitchell (Ed.), pp. 161-183. Heinemann.
- Mezaini, A., Chihib, N.E., Dilmi Bouras, A., Nedjar-Arroume, N. and Hornez, J.P. (2009). Antibacterial activity of some lactic acid bacteria isolated from an Algerian dairy product. *Journal of Environmental and Public Health*. 1: 1-6.
- Midilli, M., Alp, M., Kocabach, N., Muglah, O.H., Turan, N., Yilmaz, H. and Cakir, S. (2008). Effects of dietary probiotic and prebiotic supplementation on growth performance and serum IgG concentration of broilers. *South African Journal of Animal Science*. 38: 21-27.
- Miller, J.M. and Rhoden, D.L. (1991). Preliminary evaluation of Biolog, a carbon source utilization method for bacterial identification. *Journal of Clinical Microbiology*. 29: 1143-1147.
- Mishra, C. and Lambert, J. (1996). Production of anti-microbial substances by probiotics. *Asia Pacific Journal of Clinical Nutrition*. 5: 20-24.
- Mohan, B., Kadirvel, R., Natarajan, A. and Bhaskaran, M. (1996). Effect of probiotic supplementation on growth, nitrogen utilisation and serum cholesterol in broilers. *British Poultry Science*. 37: 395-401.
- Monajjemi, M., Aminin, A., Ilkhani, A. and Mollaamin, F. (2012). Nano study of antioxidant activities of fermented soy whey prepared with lactic acid bacteria and kefir. *African Journal of Microbiology Research* 6: 426-430.
- Morelli, L. (2000). *In vitro* selection of probiotic lactobacilli: a critical appraisal. *Current Issues in Intestinal Microbiology*. 1: 59-67.
- Moser, S.A. and Savage, D.C. (2001). Bile salt hydrolase activity and resistance to toxicity of conjugated bile salts are unrelated properties in lactobacilli. *Applied and Environmental Microbiology*. 67: 3476-3480.
- Mountzouris, K.C., Tsirtsikos, P., Kalamara, E., Nitsch, S., Schatzmayr, G. and Fegeros, K. (2007). Evaluation of the efficacy of a probiotic containing *Lactobacillus*, *Bifidobacterium*, *Enterococcus*, and *Pediococcus* strains in promoting broiler performance and modulating cecal microflora composition and metabolic activities. *Poultry Science*. 86: 309-317.
- Muller, L., Frohlich, K. and Bohm, V. (2011). Comparative antioxidant activities of carotenoids measured by ferric reducing antioxidant power (FRAP), ABTS bleaching assay (α TEAC), DPPH assay and peroxy radical scavenging assay. *Food Chemistry*. 129: 139-148.

- Musikasang, H., Tani, A., H-kittikun, A. and Maneerat, S. (2009). Probiotic potential of lactic acid bacteria isolated from chicken gastrointestinal digestive tract. *World Journal of Microbiology and Biotechnology*. 25: 1337-1345.
- Muyanja, C., Narvhus, J.A., Treimo, J. and Langsrud, T. (2003). Isolation, characterisation and identification of lactic acid bacteria from bushera: a Ugandan traditional fermented beverage. *International Journal of Food Microbiology*. 80: 201-210.
- Nagengast, F.M., Grubben, M. and Van Munster, I.P. (1995). Role of bile acids in colorectal carcinogenesis. *European Journal of Cancer*. 31: 1067-1070.
- Nahanshon, S.N., Nakaue, H.S. and Mirosh, L.W. (1992). Effects of direct-fed microbials on nutrient retention and parameters of laying pullets. *Poultry Science*. 71, (Supplement 1), 111 (Abstract).
- Nahanshon, S.N., Nakaue, H.S. and Mirosh, L.W. (1993). Effects of direct-fed microbials on nutrient retention and parameters of Single Comb White Leghorn pullets. *Poultry Science*. 72, (Supplement 2), 87 (Abstract).
- Nahanshon, S.N., Nakaue, H.S. and Mirosh, L.W. (1996). Performance of Single Comb White Leghorn layers fed with a live microbial during the growth and egg laying phases *Animal Science and Thechnology*. 57: 25-38.
- Navidshad, B., Liang, J.B. and Jahromi, M.F. (2012). Correlation coefficients between different methods of expressing bacterial quantification using real time PCR. *International Journal of Molecular Sciences*. 13: 2119-2132.
- Nayebpor, M., Farhomand, P. and Hashemi, A. (2007). Effects of different levels of direct fed microbial (Primalac) on growth performance and humoral immune response in broiler chickens. *Journal of Animal and Veterinary Advances*. 6: 1308-1313.
- NCBI (National Center for Biotechnology Information). www.ncbi.nlm.nih.gov
- Neal-McKinney, J.M., Lu, X., Duong, T., Larson, C.L., Call, D.R., Shah, D.H. and Konkell, M.E. (2012). Production of organic acids by probiotic lactobacilli can be used to reduce pathogen load in poultry. *PLoS One*. doi: 10.1371/journal.pone.0043928 (Online).
- Nemcova, R. (1997). Selection criteria of lactobacilli for probiotic use. *Veterinarni Medicina*. 42: 19-27.
- Ngoc Lan, P.T., Binh, L.T. and Benno, Y. (2003). Impact of two probiotic Lactobacillus strains feeding on fecal lactobacilli and weight gains in chicken. *The Journal of General and Applied Microbiology*. 49: 29-36.

- Nguyen, T.D.T., Kang, J.H. and Lee, M.S. (2007). Characterization of *Lactobacillus plantarum* PH04, a potential probiotic bacterium with cholesterol-lowering effects. *International Journal of Food Microbiology*. 113: 358-361.
- Noh, D.O., Kim, S.H. and Gilliland, S.E. (1997). Incorporation of cholesterol into the cellular membrane of *Lactobacillus acidophilus* ATCC 431211. *Journal of Dairy Science*. 80: 3107-3113.
- NRC (National Research Council) (1994). *Nutrient Requirements of Poultry*. 157 pages. National Academies Press.
- NRC (National Research Council) (1996). *Guide for the care and use of laboratory animals*. 220 pages. National Academies Press.
- O'Halloran, S., Feeney, M., Morrissey, D., Murphy, L., Thornton, G., Shanahan, F., O'Sullivan, G.C. and Collins, J.K. (1997). Adhesion of potential probiotic bacteria to human epithelial cell lines. 8: 596 (Abstract).
- O'sullivan, M.G., Thornton, G., O'sullivan, G.C. and Collins, J.K. (1992). Probiotic bacteria: myth or reality?. *Trends in Food Science and Technology*. 3: 309-314.
- Ochman, H., Lerat, E. and Daubin, V. (2005). Examining bacterial species under the specter of gene transfer and exchange. *Proceedings of the National Academy of Sciences of the United States of America*. 102: 6595-6599.
- OECD/OCDE (Organisation for Economic Co-operation and Development/Organisation de Coopération et de Développement Economiques) (2001). *OECD Guideline for the Testing of Chemicals No. 423, Acute Oral Toxicity, Acute Toxic Class Method*. 14 pages.
- OECD/OCDE (Organisation for Economic Co-operation and Development/Organisation de Coopération et de Développement Economiques) (2008). *OECD Guideline for the Testing of Chemicals No. 407, Repeated Dose 28-Day Oral Toxicity Study in Rodents*. 13 pages.
- Ogimoto, K. and Imai, S. (1981). *Atlas of rumen microbiology*. 158 pages. Scientific Societies Press.
- Ooi, L.G. and Liong, M.T. (2010). Cholesterol-lowering effects of probiotics and prebiotics: a review of *in vivo* and *in vitro* findings. *International Journal of Molecular Sciences*. 11: 2499-2522.
- Ouwehand, A.C., Isolauri, E., Kirjavainen, P.V. and Salminen, S.J. (1999a). Adhesion of four *Bifidobacterium* strains to human intestinal mucus from subjects in different age groups. *FEMS Microbiology Letters*. 172: 61-64.

- Ouwehand, A.C., Kirjavainen, P.V., Shortt, C. and Salminen, S. (1999b). Probiotics: mechanisms and established effects. *International Dairy Journal*. 9: 43-52.
- Ouwehand, A.C. and Vesterlund, S. (2004). Antimicrobial components from lactic acid bacteria. In: *Lactic acid bacteria - Microbiological and functional aspects*. CRC Press. doi: 10.1201/9780824752033.ch11 (Online).
- Pan, X., Chen, F., Wu, T., Tang, H. and Zhao, Z. (2009). The acid, bile tolerance and antimicrobial property of *Lactobacillus acidophilus* NIT. *Food Control*. 20: 598-602.
- Panda, A.K., Reddy, M.R., Rao, S.V.R., Raju, M. and Praharaj, N.K. (2000). Growth, carcass characteristics, immunocompetence and response to *Escherichia coli* of broilers fed diets with various levels of probiotic. *Archiv fur Geflugelkunde*. 64: 152-156.
- Panda, A.K., Rao, S.V.R., Raju, M.V.L.N. and Sharma, S.R. (2006). Dietary supplementation of *Lactobacillus sporogenes* on performance and serum biochemical-lipid profile of broiler chickens. *Poultry Science*. 43: 235-240.
- Paramithiotis, S., Melissari, I. and Drosinos, E.H. (2006). *In vitro* assessment of properties associated with the survival through the gastro intestinal tract of staphylococci isolated from traditional sausage fermentation. *Food Microbiology*. 23: 663-671.
- Parker, R.B. (1974). Probiotics, the other half of the antibiotic story. *Animal Nutrition and Health*. 29: 4-8.
- Patterson, J.A. and Burkholder, K.M. (2003). Application of prebiotics and probiotics in poultry production. *Poultry Science*. 82: 627-631.
- Patterson, J.K., Lei, X.G. and Miller, D.D. (2008). The pig as an experimental model for elucidating the mechanisms governing dietary influence on mineral absorption. *Experimental Biology and Medicine*. 233: 651-664.
- Pavlova, S.I., Kilic, A.O., Kilic, S.S., So, J.S., Nader-Macias, M.E., Simoes, J.A. and Tao, L. (2002). Genetic diversity of vaginal lactobacilli from women in different countries based on 16S rRNA gene sequences. *Journal of Applied Microbiology*. 92: 451-459.
- Pereira, D.I.A. and Gibson, G.R. (2002a). Cholesterol assimilation by lactic acid bacteria and bifidobacteria isolated from the human gut. *Applied and Environmental Microbiology*. 68: 4689-4693.
- Pereira, D.I.A. and Gibson, G.R. (2002b). Effects of consumption of probiotics and prebiotics on serum lipid levels in humans. *Critical reviews in biochemistry and molecular biology*. 37: 259-281.

- Peric, L., Milosevic, N., Zikic, D., Bjedov, S., Cvetkovic, D., Markov, S., Mohnl, M. and Steiner, T. (2010). Effects of probiotic and phytogetic products on performance, gut morphology and cecal microflora of broiler chickens. *Archiv Tierzucht*. 53: 350-359.
- Petterino, C. and Argentino-Storino, A. (2006). Clinical chemistry and haematology historical data in control Sprague-Dawley rats from pre-clinical toxicity studies. *Experimental and Toxicologic Pathology*. 57: 213-219.
- Petti, C.A., Polage, C.R. and Schreckenberger, P. (2005). The role of 16S rRNA gene sequencing in identification of microorganisms misidentified by conventional methods. *Journal of Clinical Microbiology*. 43: 6123-6125.
- Phonnok, S., Uthaisang-Tanechpongamb, W. and Wongsatayanon, B.T. (2010). Anticancer and apoptosis-inducing activities of microbial metabolites. *Electronic Journal of Biotechnology*. doi: 10.2225/vol13-issue5-fulltext-7 (Online).
- Pinto, M., Robine-Leon, S., Appay, M.D., Keding, M., Triadou, N., Dussaulx, E., Lacroix, B., Simon-Assmann, P., Haffen, K. and Fogh, J. (1983). Enterocyte-like differentiation and polarization of the human colon carcinoma cell line Caco-2 in culture. *Biology of the Cell*. 47: 323-330.
- Rahimi, M. (2009). Effects of probiotic supplementation on performance and humoral immune response of broiler chickens. In proceeding of: *World Poultry Science Association (WPSA)*. 1: 67-69.
- Raza, M., Al-Shabanah, O.A., El-Hadiyah, T.M. and Al-Majed, A.A. (2002). Effect of prolonged vigabatrin treatment on hematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. *Scientia Pharmaceutica*. 70: 135-145.
- Rea, M.C., Ross, R.P., Cotter, P.D. and Hill, C. (2011). Classification of bacteriocins from Gram-positive bacteria. In: *Prokaryotic Antimicrobial Peptides*, D. Djamel and R. Sylvie (Eds.), pp. 29-53. Springer New York.
- Reddy, B.S., Hamid, R. and Rao, C.V. (1997). Effect of dietary oligofructose and inulin on colonic preneoplastic aberrant crypt foci inhibition. *Carcinogenesis*. 18: 1371-1374.
- Reddy, G., Altaf, M., Naveena, B.J., Venkateshwar, M. and Kumar, E.V. (2008). Amylolytic bacterial lactic acid fermentation. *Biotechnology Advances*. 26: 22-34.
- Reid, G. and Burton, J. (2002). Use of *Lactobacillus* to prevent infection by pathogenic bacteria. *Microbes and Infection*. 4: 319-324.

- Ridlon, J.M., Kang, D.J. and Hylemon, P.B. (2006). Bile salt biotransformations by human intestinal bacteria. *Journal of Lipid Research*. 47: 241-259.
- Riley, M.A., Chavan, M.A., Heng, N.C.K., Wescombe, P.A., Burton, J.P., Jack, R.W. and Tagg, J.R. (2007). The Diversity of Bacteriocins in Gram-Positive Bacteria. In: *Bacteriocins*, pp. 45-92. Springer Berlin Heidelberg.
- Ronka, E., Malinen, E., Saarela, M., Rinta-Koski, M., Aarnikunnas, J. and Palva, A. (2003). Probiotic and milk technological properties of *Lactobacillus brevis*. *International Journal of Food Microbiology*. 83: 63-74.
- Rudel, L.L. and Morris, M.D. (1973). Determination of cholesterol using O-phthalaldehyde. *Journal of Lipid Research*. 14: 364-366.
- Ruiz-Moyano, S., Martin, A., Benito, M.J., Nevado, F.P. and de Guia Cordoba, M. (2008). Screening of lactic acid bacteria and bifidobacteria for potential probiotic use in Iberian dry fermented sausages. *Meat Science*. 80: 715-721.
- Saarela, M., Mogensen, G., Fonden, R., Matto, J. and Mattila-Sandholm, T. (2000). Probiotic bacteria: safety, functional and technological properties. *Journal of Biotechnology*. 84: 197-215.
- Sahadeva, R.P.K., Leong, S.F., Chua, K.H., Tan, C.H., Chan, H.Y., Tong, E.V., Wong, S.Y.W. and Chan, H.K. (2011). Survival of commercial probiotic strains to pH and bile. *International Food Research Journal*. 18: 1515-1522.
- Saikali, J., Picard, C., Freitas, M. and Holt, P. (2004). Fermented milks, probiotic cultures, and colon cancer. *Nutrition and Cancer*. 49: 14-24.
- Saitou, N. and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*. 4: 406-425.
- Salminen, M.K., Rautelin, H., Tynkkynen, S., Poussa, T., Saxelin, M., Valtonen, V. and Jarvinen, A. (2004). *Lactobacillus* bacteremia, clinical significance, and patient outcome, with special focus on probiotic *L. rhamnosus* GG. *Clinical Infectious Diseases*. 38: 62-69.
- Salminen, S., Isolauri, E. and Salminen, E. (1996). Clinical uses of probiotics for stabilizing the gut mucosal barrier: successful strains and future challenges. *Antonie Van Leeuwenhoek*. 70: 347-358.
- Salminen, S., Von Wright, A., Morelli, L., Marteau, P., Brassart, D., De Vos, W.M., Fonden, R., Saxelin, M., Collins, K. and Mogensen, G. (1998). Demonstration of safety of probiotics. *International Journal of Food Microbiology*. 44: 93-106.

- Samanya, M. and Yamauchi, K. (2002). Histological alterations of intestinal villi in chickens fed dried *Bacillus subtilis* var. *natto*. *Comparative Biochemistry and Physiology* 133: 95-104.
- Sanders, M.E. (1998). Overview of functional foods: emphasis on probiotic bacteria. *International Dairy Journal*. 8: 341-347.
- Sanders, M.E. (2008). Probiotics: definition, sources, selection, and uses. *Clinical Infectious Diseases*. 46: S58-S61.
- SAS (Statistical Analysis System) (2008). SAS online documentation 9.2. SAS Institute Incorporation.
- Saulnier, D., Spinler, J.K., Gibson, G.R. and Versalovic, J. (2009). Mechanisms of probiosis and prebiosis: considerations for enhanced functional foods. *Current Opinion in Biotechnology*. 20: 135-141.
- Saxelin, M. (1997). *Lactobacillus* GG a human probiotic strain with thorough clinical documentation. *Food Reviews International*. 13: 293-313.
- Schiffrin, E.J., Rochat, F., Link-Amster, H., Aeschlimann, J.M. and Donnet-Hughes, A. (1995). Immunomodulation of human blood cells following the ingestion of lactic acid bacteria. *Journal of Dairy Science*. 78: 491-497.
- Schillinger, U. and Lucke, F.K. (1987). Identification of lactobacilli from meat and meat products. *Food Microbiology*. 4: 199-208.
- Schillinger, U. (1999). Isolation and identification of lactobacilli from novel-type probiotic and mild yoghurts and their stability during refrigerated storage. *International Journal of Food Microbiology*. 47: 79-87.
- Sifour, M., Tayeb, I., Ouled, H., Namous, H. and Aissaoui, S. (2012). Production and characterization of bacteriocin of *Lactobacillus plantarum* F12 with inhibitory activity against *Listeria monocytogenes*. *The Online Journal of Science and Technology*. 2: 55-61.
- Sirilun, S., Chaiyasut, C., Kantachote, D. and Luxanani, P. (2010). Characterisation of non human origin probiotic *Lactobacillus plantarum* with cholesterol-lowering property. *African Journal of Microbiology Research*. 4: 994-1000.
- Sjovall, J. (1959). On the concentration of bile acids in the human intestine during absorption. Bile acids and steroids 74. *Acta Physiologica Scandinavica*. 46: 339-345.
- Slaga, T.J. and Bracken, W.M. (1977). The effects of antioxidants on skin tumor initiation and aryl hydrocarbon hydroxylase. *Cancer Research*. 37: 1631-1635.

- Smibert, R.M. (1974). *Campylobacter*. In: *Bergey's Manual of Determinative Bacteriology*, R.E. Buchanan and N.E. Gibbons (Eds.), pp. 204-212. Williams and Wilkins.
- Smitinont, T., Tansakul, C., Tanasupawat, S., Keeratipibul, S., Navarini, L., Bosco, M. and Cescutti, P. (1999). Exopolysaccharide-producing lactic acid bacteria strains from traditional thai fermented foods: isolation, identification and exopolysaccharide characterization. *International Journal of Food Microbiology*. 51: 105-111.
- Sorokulova, I.B., Pinchuk, I.V., Denayrolles, M., Osipova, I.G., Huang, J.M., Cutting, S.M. and Urdaci, M.C. (2008). The safety of two *Bacillus* probiotic strains for human use. *Digestive Diseases and Sciences*. 53: 954-963.
- Stackebrandt, E. and Goebel, B.M. (1994). Taxonomic note: a place for DNA-DNA reassociation and 16S rRNA sequence analysis in the present species definition in bacteriology. *International Journal of Systematic Bacteriology*. 44: 846-849.
- Stewart, L., Pellegrini, C.A. and Way, L.W. (1986). Antibacterial activity of bile acids against common biliary tract organisms. In proceeding of: *Surgical Forum*. 37: 157-159.
- Suskovic, J., Blazenka, K., Beganovic, J., Pavunc, A.L., Habjanic, K. and Matosic, S. (2010). Antimicrobial Activity - The most important property of probiotic and starter lactic acid bacteria. *Food Technology and Biotechnology*. 48: 296-307.
- Suvarna, V.C. and Boby, V.U. (2005). Probiotics in human health: A current assessment. *Current Science*. 88: 1744-1748.
- Taherpour, K., Moravej, H., Shivazad, M., Adibmoradi, M. and Yakhchali, B. (2009). Effects of dietary probiotic, prebiotic and butyric acid glycerides on performance and serum composition in broiler chickens. *African Journal of Biotechnology*. 8: 2329-2334.
- Tahri, K., Grille, J.P. and Schneider, F. (1996). Bifidobacteria strain behavior toward cholesterol: coprecipitation with bile salts and assimilation. *Current Microbiology*. 33: 187-193.
- Tamura, K. (1992). Estimation of the number of nucleotide substitutions when there are strong transition-transversion and G+C-content biases. *Molecular Biology and Evolution*. 9: 678-687.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M. and Kumar, S. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution*. 28: 2731-2739.

- Tanaka, H., Doesburg, K., Iwasaki, T. and Mierau, I. (1999). Screening of lactic acid bacteria for bile salt hydrolase activity. *Journal of Dairy Science*. 82: 2530-2535.
- Tang, Y.W., Ellis, N.M., Hopkins, M.K., Smith, D.H., Dodge, D.E. and Persing, D.H. (1998). Comparison of phenotypic and genotypic techniques for identification of unusual aerobic pathogenic gram-negative bacilli. *Journal of Clinical Microbiology*. 36: 3674-3679.
- Tannock, G.W. (1999). *Probiotics: a critical review*. 157 pages. Horizon Scientific Press.
- Taranto, M.P., Sesma, F., Pesce de Ruiz Holgado, A. and de Valdez, G.F. (1997). Bile salts hydrolase plays a key role on cholesterol removal by *Lactobacillus reuteri*. *Biotechnology Letters*. 19: 845-847.
- Thirabunyanon, M., Boonprasom, P. and Niamsup, P. (2009). Probiotic potential of lactic acid bacteria isolated from fermented dairy milks on antiproliferation of colon cancer cells. *Biotechnology Letters*. 31: 571-576.
- Thompson, S.W. and Hunt, R.D. (1966). *Selected histochemical and histopathological methods*. 1639 pages. CC Thomas Springfield.
- Timmerman, H.M., Veldman, A., Van den Elsen, E., Rombouts, F.M. and Beynen, A.C. (2006). Mortality and growth performance of broilers given drinking water supplemented with chicken-specific probiotics. *Poultry Science*. 85: 1383-1388.
- Toure, R., Kheadr, E., Lacroix, C., Moroni, O. and Fliss, I. (2003). Production of antibacterial substances by bifidobacterial isolates from infant stool active against *Listeria monocytogenes*. *Journal of Applied Microbiology*. 95: 1058-1069.
- Transparency Market Research (2013). Probiotics market (dietary supplements, animal feed, foods & beverages) - global Industry analysis, market size, share, trends, analysis, growth and forecast, 2012 - 2018. 120 pages.
- Tsai, J.C., Huang, G.J., Chiu, T.H., Huang, S.S., Huang, S.C., Huang, T.H., Lai, S.C. and Lee, C.Y. (2011). Antioxidant activities of phenolic components from various plants of *Desmodium* species. *African Journal of Pharmacy and Pharmacology*. 5: 468-476.
- Tuo, Y.F., Zhang, L.W., Yi, H.X., Zhang, Y.C., Zhang, W.Q., Han, X., Du, M., Jiao, Y.H. and Wang, S.M. (2010). Antiproliferative effect of wild *Lactobacillus* strains isolated from fermented foods on HT-29 cells. *Journal of Dairy Science*. 93: 2362-2366.
- Tuomola, E.M. and Salminen, S.J. (1998). Adhesion of some probiotic and dairy *Lactobacillus* strains to Caco-2 cell cultures. *International Journal of Food Microbiology*. 41: 45-51.

- URI (Genomics and Sequencing Center of University of Rhode Island).
www.uri.edu/research/gsc/resources/cndna.html
- Usman and Hosono, A. (1999). Bile Tolerance, taurocholate deconjugation, and binding of cholesterol by *Lactobacillus gasseri* strains. *Journal of Dairy Science*. 82: 243-248.
- Van Den Berg, D.J.C., Smits, A., Pot, B., Ledebroer, A.M., Kersters, K., Verbake, J.M.A. and Verrips, C.T. (1993). Isolation, screening and identification of lactic acid bacteria from traditional food fermentation processes and culture collections. *Food Biotechnology*. 7: 189-205.
- Vankerckhoven, V., Huys, G., Vancanneyt, M., Vael, C., Klare, I., Romond, M.B., Entenza, J.M., Moreillon, P., Wind, R.D. and Knol, J. (2008). Biosafety assessment of probiotics used for human consumption: recommendations from the EU-PROSAFE project. *Trends in Food Science and Technology*. 19: 102-114.
- Vaughan, E.E., de Vries, M.C., Zoetendal, E.G., Ben-Amor, K., Akkermans, A.D.L. and de Vos, W.M. (2002). The intestinal LABs. *Antonie Van Leeuwenhoek*. 82: 341-352.
- Voravuthikunchai, S.P., Bilasoi, S. and Supamala, O. (2006). Antagonistic activity against pathogenic bacteria by human vaginal lactobacilli. *Anaerobe*. 12: 221-226.
- Wang, R.F., Cao, W.W. and Cerniglia, C.E. (1996). PCR detection and quantitation of predominant anaerobic bacteria in human and animal fecal samples. *Applied and environmental microbiology*. 62: 1242-1247.
- Watkins, B.A. and Kratzer, F.H. (1983). Effect of oral dosing of *Lactobacillus* strains on gut colonization and liver biotin in broiler chicks. *Poultry Science*. 62: 2088-2094.
- WHO (World Health Organization) (2009). Table 2: Cause-specific mortality and morbidity. In: *World Health Statistics 2009*, L. Gollogly (Ed.), pp. 149. World Health Organization.
- WHO (World Health Organization) (2011). Cardiovascular Disease - Fact sheet N°317.
www.who.int/mediacentre/factsheets/fs317/en/
- Willis, W.L., Isikhuemhen, O.S. and Ibrahim, S.A. (2007). Performance assessment of broiler chickens given mushroom extract alone or in combination with probiotics. *Poultry Science*. 86: 1856-1860.
- Willis, W.L. and Reid, L. (2008). Investigating the effects of dietary probiotic feeding regimens on broiler chicken production and *Campylobacter jejuni* presence. *Poultry Science*. 87: 606-611.

- Xu, Z.R., Hu, C.H., Xia, M.S., Zhan, X.A. and Wang, M.Q. (2003). Effects of dietary fructooligosaccharide on digestive enzyme activities, intestinal microflora and morphology of male broilers. *Poultry Science*. 82: 1030-1036.
- Yeo, J. and Kim, K. (1997). Effect of feeding diets containing an antibiotic, a probiotic, or yucca extract on growth and intestinal urease activity in broiler chicks. *Poultry Science*. 76: 381-385.
- Yin, Q. and Zheng, Q. (2005). Isolation and identification of the dominant *Lactobacillus* in gut and faeces of pigs using carbohydrate fermentation and 16S rDNA analysis. *Journal of Bioscience and Bioengineering*. 99: 68-71.
- Yoon, Y.H. and Byun, J.R. (2004). Occurrence of glutathione sulphydryl (GSH) and antioxidant activities in probiotic *Lactobacillus* spp. *Asian - Australasian Journal of Animal Sciences*. 17: 1582-1585.
- Zacharof, M.P. and Lovitt, R.W. (2012). Bacteriocins produced by lactic acid bacteria a review article. *APCBEE Procedia*. 2: 50-56.
- Zalan, Z., Hudacek, J., Stetina, J., Chumchalova, J. and Halasz, A. (2010). Production of organic acids by *Lactobacillus* strains in three different media. *European Food Research and Technology*. 230: 395-404.

LIST OF PUBLICATIONS

Journal Papers

1. Shokryazdan, P., R. Kalavathy, C.C. Sieo, N. B. Alitheen, J. B. Liang, M. F. Jahromi and Y. W. Ho. Isolation and characterization of *Lactobacillus* strains as potential probiotics for chickens. *Pertanika Journal of Tropical Agricultural Science*. In press.

Proceedings of Papers Presented in Conferences, Symposia, Congresses and Seminars

1. Shokryazdan, P., R. Kalavathy, C.C. Sieo, J.B. Liang, M.F. Jahromi and Y.W. Ho. Antioxidant activity of *Lactobacillus* strains isolated from infant feces, human milk and fermented food, Annual Meeting and Exhibition of society for Industrial Microbiology. July 24-28, 2011. New Orleans, USA. P:178.
2. Shokryazdan, P., C.C. Sieo, R. Kalavathy, J. B. Liang, M.F. Jahromi and Y. W. Ho. Antimicrobial activity of *Lactobacillus salivarius* isolated from chicken. The 3rd International Conference on Sustainable Animal Agriculture for Developing Countries (SAADC 2011), July 26-29, 2011. Nakhon Ratchasima, Thailand. P:258.
3. Shokryazdan, P., C.C. Sieo, R. Kalavathy, J.B. Liang, M.F. Jahromi, and Y. W. Ho. Bile Salt Hydrolase Activity of *Lactobacillus* Strains Isolated From Chicken Intestine. The 14th Animal Science Congress of the Asian-Australasian Association of Animal Production Societies (AAAP 2010), August 23-27, 2010. Pingtung, Taiwan. Vol:2, P:309.