



**UNIVERSITI PUTRA MALAYSIA**

***ANTINOCICEPTIVE EFFECT OF 3-(2, 3 DIMETHOXYPHENYL)-1  
(METHYLFURAN-2-YL) PROP-2-EN-1-ONE IN MICE AND ITS  
MECHANISMS OF ACTION***

**NUR IZZATI BINTI ISMAIL**

**FPSK(m) 2015 76**



**ANTINOCICEPTIVE EFFECT OF 3-(2, 3 DIMETHOXYPHENYL)-1  
(METHYLFURAN-2-YL) PROP-2-EN-1-ONE IN MICE AND ITS  
MECHANISMS OF ACTION**

**By**

**NUR IZZATI BINTI ISMAIL**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Master of Science**

**February 2015**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment  
of the requirements for the degree of Master of Science

**ANTINOCICEPTIVE EFFECT OF 3-(2,3 DIMETHOXYPHENYL)-1  
(METHYLFURAN-2-YL) PROP-2-EN-1-ONE IN MICE AND ITS  
MECHANISMS OF ACTION**

By

**NUR IZZATI BINTI ISMAIL**

**February 2015**

**Chairman : Professor Mohd Roslan bin Sulaiman, PhD**  
**Faculty : Medicine and Health Sciences**

**The objective of the present study is to** evaluate the antinociceptive activity of the new chalcone derivative, **the 3-(2,3 dimethoxyphenyl)-1 (methylfuran-2-yl) prop-2-en-1-one (DMPF)**, using the chemical- and thermal-induced nociception test together with the mechanisms of actions. It was demonstrated that, administration of DMPF given by the i.p route produced a significant dose-dependent inhibition on acetic acid-induced writhing test, increased in response latency time for the hot plate test and reduction in time spent licking the injected paw for both neurogenic and inflammatory phases in formalin-induced paw licking test. Furthermore, the antinociceptive effect of DMPF in the acetic acid-induced writhing and the hot plate test was not reversed by non-selective opioid receptor antagonist, naloxone, that shown the non-involvement of the opioidergic system in antinociceptive activity of DPMF. Contrarily, administration of DMPF produced significant dose-dependent inhibition in capsaicin-induced paw licking test, glutamate-induced paw licking test, and PMA-induced paw licking test proven the involvement of TRPV1 receptor, glutamatergic system and PKC system. Antinociception caused by DPMF in the acetic acid test reversed the treatment with L-NOARG (nitric oxide synthase inhibitor) and charybdotoxin (a large-conductance  $Ca^{2+}$ -sensitive  $K^+$  channels blocker). However, antinociception of DPMF was enhanced by ODQ (a specific guanylyl cyclase inhibitor). To sum up, the results indicate that DPMF produced pronounced central and peripheral antinociception by inhibition on glutamate receptors, TRPV1 receptors and protein kinase C. It is also strongly suggested that the L-arginine/NO/cGMP/BK<sub>ca</sub> channel pathway plays a significant role in DPMF-induced antinociception.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai  
Memenuhi keperluan untuk ijazah Master Sains

**KESAN ANTINOSISEPTIF YANG DIARUH OLEH  
3-(2, 3 DIMETHOXYPHENYL) 1(METHYLFURAN-2-YL)  
PROP-2-EN-1-ONE (DPMF) IAITU DERIVASI CHALCONE  
KE ATAS MENCIT DAN MEKANISMA-MEKANISMANYA**

Oleh

**NUR IZZATI BINTI ISMAIL**

**Februari 2015**

**Pengerusi : Profesor Mohd Roslan bin Sulaiman, PhD**  
**Fakulti : Perubatan dan Sains Kesihatan**

Objektif kajian yang dijalankan baru-baru ini adalah untuk mengkaji kesan aktiviti antinosiseptif yang diaruh oleh derivatif chalcone iaitu 3-(2,3 dimethoxyphenyl)-1 (methylfuran-2-yl) prop-2-en-1-one (DMPF) dan mekanisma-mekanismanya dengan menggunakan model nosiseptif kimia dan haba. Kajian menunjukkan bahawa pemberian DPMF secara intraperitoneum menunjukkan perencatan signifikan yang berkadar selari dengan dos ke atas kajian ujian penggeliatan abdomen aruhan asid asetik, kajian ujian plat panas dan kajian ujian jilatan tapak kaki aruhan formalin. Kajian juga menunjukkan aktiviti antinosiseptif DPMF dalam kajian ujian penggeliatan abdomen aruhan asid asetik dan kajian ujian plat panas adalah tidak berbalik oleh antagonis reseptor opioid yang tidak selektif, naloxone, sekaligus menunjukkan bahawa tiadanya penglibatan sistem opioidergik di dalam aktiviti antinosiseptif DPMF. Namun, perawatan dengan DPMF menghasilkan rencatan yang signifikan berdasarkan dos bagi ujian nosisepsi aruhan kapsaisin, ujian jilatan tapak kaki ransangan glutamat, dan ujian jilatan tapak kaki ransangan FMA didalam mencit di mana ujian-ujian tersebut menunjukkan kesan rencatan DPMF terhadap reseptor TRPV1, sistem glutamatergik, dan dalam sistem PKC. Dalam ujian seterusnya, antinosiseptif yang ditunjukkan oleh DPMF dalam kajian ujian penggeliatan abdomen aruhan asid asetik menghasilkan perbezaan bilangan penggeliatan abdomen yang signifikan dengan pemberian rawatan oleh L-NOARG (perencat enzim nitrik oksida) dan "charybdotoxin" (perencat saluran K<sup>+</sup> sensitif). Walaubagaimanapun, kajian mendapati kesan antinosiseptif oleh DMPF telah ditingkatkan dengan pemberian ODQ (perencat spesifik "guanylyl cyclase). Hasil kajian di atas menunjukkan bahawa laluan L-arginin/NO/cGMP/ BK<sub>ca</sub> memainkan peranan yang signifikan dalam membolehkan DPMF menjalankan kesan antinosiseptif.

## ACKNOWLEDGEMENTS

In the name of Allah the most Merciful and the Most Grateful. Any large endeavour of this type cannot be completed without the help of many individuals. First and foremost, I would like to thank Prof Mohd Roslan bin Sulaiman as a respected supervisor for the guidance and all the hard work that he put through to ensure the project will be a success and to my beloved parent Mr. Ismail Osman and Mdm. Siti Mariam Baharin as well as my brother and sister-in-law. No amount of “thank-yous” can fully capture my indebtedness to them. They always listened to my problems and be a very good teacher to me.

All the staffs at the laboratory also deserve thanks for helping and wasting their time for explaining and clarify difficult concepts.

I am also grateful to all of my friends particularly my lab mates and my best friends Ms Siti Fatimah Abdul Jabar, Ms Nur Hayati Samsul Baharil, Ms Nadiah Zakaria, Ms Zulaikha Nadrah Zulkafli, Mr Mohd Nasier Kamaldin and Ms Afiqah Mohd. They are very great friends and I am fortunate to be able working with them and to see the dedication that they put through this journey.

And finally to all my colleagues at Kulliyah of Medicine and Health Sciences, INSANIAH University College especially Prof. Dr Nasaruddin, Assoc. Prof. Dato’ Haji Azmi bin Hashim, Dr. Hassan Basri Bin Jahubar Sathik, Dr Lee Ii Lee, Mr Syahrir bin Menteri, Dr. Norfaizatul Shalida Omar, Ms Ireena Ismail and all of the staff for sharing their knowledge and be there if there is any problems. Despite differences in opinions and individual styles, this partnership has increased our mutual respect, and our friendship grows stronger.

I certify that a Thesis Examination Committee has met on 27 February 2015 to conduct the final examination of Nur Izzati binti Ismail on her thesis entitled "Antinociceptive Effect of 3-(2,3 Dimethoxyphenyl)-1 (Methylfuran-2-YL) Prop-2-EN-1-One in Mice and its Mechanisms of Action" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Mohd Nasir bin Mohd Desa, PhD**

Associate Professor  
Faculty of Medicine and Health Science  
Universiti Putra Malaysia  
(Chairman)

**Mohamad Aris bin Mohd Moklas, PhD**

Senior Lecturer  
Faculty of Medicine and Health Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Zulkhairi Hj Amom, PhD**

Professor  
Universiti Teknologi MARA  
Malaysia  
(External Examiner)



---

**ZULKARNAIN ZAINAL, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 17 June 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Mohd Roslan bin Sulaiman, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Daud Ahmad bin Israf Ali, PhD**

Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Md Nordin bin Hj Lajis, PhD**

Professor  
Institute of Bioscience  
Universiti Putra Malaysia  
(Member)

---

**BUJANG KIM KIM HUAT, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:



## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Nur Izzati Binti Ismail GS 28065 \_\_\_\_\_

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of  
Chairman of  
Supervisory  
Committee: Dr. Mohd Roslan bin Sulaiman

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: Dr. Daud Ahmad bin Israf Ali

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: Dr. Md Nordin bin Hj Lajis

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	ii
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	iv
<b>DECLARATION</b>	vi
<b>LIST OF FIGURES</b>	x
<b>LIST OF ABBREVIATIONS</b>	xii
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	
1.1 Objectives	3
<b>2 LITERATURE REVIEW</b>	
2.1 Pain	4
2.2 Noxious stimuli	4
2.3 Nociceptive and its pathway	5
2.4 Action potential and pain	8
2.5 Pain pathway	10
2.5.1 Ascending pain pathway (spinothalamic tract)	10
2.5.1.1 Molecular involved in ascending pain pathway	11
2.5.2 Descending pain pathway	13
2.5.3 Hyperpolarization and inhibition of pain	17
2.5.4 NO/cGMP pathway	19
2.6 Current analgesic and their limitations	19
2.7 Chalcone and 3-(2,3 dimethoxyphenyl)-1(methylfuran-2-yl) prop-2-en-1-one (DPMF)	20
<b>3 MATERIALS AND METHODS</b>	
3.1 Animals	22
3.2 Drugs and chemicals	22
3.3 Origin of DPMF	23
3.4 Antinociceptive Studies	23
3.4.1 Acetic Acid-Induced Abdominal Constriction Test	23
3.4.2 Formalin-Induced Paw Licking Test	24
3.4.3 Hot Plate Test	24
3.5 Mechanisms of Action	25
3.5.1 Involvement of opioid receptors	25
3.5.2 Capsaicin-Induced Paw Licking Test	25
3.5.3 Glutamate-Induced Paw Licking Test	25
3.5.4 Phorbol 12-myristate 13-acetate (PMA)- induced paw licking test	26
3.5.5 Involvement of L-arginine-nitric oxide Pathway	26

3.5.6	Involvement of cyclic guanosine Monophosphate (cGMP) pathway	26
3.5.7	Involvement of Potassium Channels	27
3.6	Preliminary acute toxicity study	27
3.7	Rota-rod Test	27
3.8	Statistical Analysis	28
<b>4</b>	<b>RESULTS</b>	
4.1	Antinociceptive Studies	29
4.1.1	Acetic Acid-Induced Abdominal Constriction Test	29
4.1.2	Formalin-Induced Paw Licking Test	30
4.1.3	Hot Plate Test	32
4.2	Mechanisms of Action	34
4.2.1	Involvement of opioid receptors	34
4.2.2	Capsaicin-Induced Paw Licking Test	35
4.2.3	Glutamate-Induced Paw Licking Test	36
4.2.4	Phorbol 12-myristate 13-acetate (PMA)- induced paw licking test	37
4.2.5	Involvement of L-arginine-nitric oxide Pathway	38
4.2.6	Involvement of cyclic guanosine Monophosphate (cGMP) pathway	39
4.2.7	Involvement of Potassium Channels	40
4.3	Preliminary acute toxicity study	42
4.4	Rota-rod Test	42
<b>5</b>	<b>DISCUSSIONS</b>	44
<b>6</b>	<b>CONCLUSION AND RECOMMENDATION FOR FUTURE STUDY</b>	48
	<b>REFERENCES</b>	50
	<b>APPENDICES</b>	61
	<b>BIODATA OF STUDENT</b>	72
	<b>PUBLICATION</b>	73

## LIST OF FIGURES

Figure		Page
1	Consequences of undertreatment of pain.	3
2	Structural components that involved in pain pathway.	7
3	The changes in (a) membrane potential (mV) and (b) relative membrane permeability (P) to Na <sup>+</sup> and K <sup>+</sup> during an action potentials.	9
4	Pain and Endogenous analgesia. Stimulation via small fibers excite neuron T to signal pain. This neuron T can be inhibited by the neuron S in the substantia gelatinosa (the Gate Theory). The neuron S can be excites via large fibres from mechanoreceptors (rub it to make it feel better), or by the recurrent neuron (R) activated by the neuron P in the limbic system (endogenous analgesia).	14
5	Analgesia system of the brain and spinal cord, showing (1) inhibition of incoming pain signals at the cord level and (2) presence of enkephalin-secreting neurons that suppress pain signals in both cord and brain stem.	16
6	Structural of chalcone and DPMF.	21
7	Effect of DMPF in acetic acid –induced abdominal writhing test in mice	29
8	Effect of DPMF in formalin-induced paw licking test (early phase, panel A, and late phase, panel B) in mice	31
9	Effect of naloxone (5 mg/kg, i.p.) on mice treated with vehicle (Control), morphine (Morph 5 mg/kg, s.c.) and DMPF (1 mg/kg, i.p.) on acetic acid-induced abdominal writhing.	34
10	Effect of DMPF against capsaicin-induced paw licking test in mice.	35
11	Effect of DMPF against glutamate-induced paw licking test in mice.	36
12	Effect of DPMF (0.1–5 mg/ kg i.p.) and aspirin (acetylsalicylic acid; 100 mg / kg, i.p.) against phorbol 12-myristate 13-acetate (PMA; 0.05 lg/ 20 ml per paw)-induced paw licking test in mice.	37
13	Effect of pre-treatment of animals with L-arginine (100 mg/ kg, i.p.) and L-NOARG (20 mg/ kg, i.p.) on the antinociceptive profiles of DPMF (1 mg/ kg, i.p.) against acetic acid-induced abdominal writhing test in mice.	38
14	Effect of pre-treatment of animals with ODQ (20 mg/ kg, i.p.) on the antinociceptive profiles of DPMF (1 mg/ kg, i.p.) against acetic acid-induced abdominal writhing test in mice.	39

15	Effect of pre-treatment of animals with glibenclamide (5 mg / ml, i.p.), apamin (0.04 mg/ml, i.p.), TEA (10 mg/kg, i.p.) and charybdotoxin (0.25 µg/50 ml, i.p.) on the antinociceptive profiles of DPMF (1 mg/ kg, i.p.) against acetic acid-induced abdominal writhing test in mice.	41
16	Effect of DPMF (0.1–5 mg / kg, i.p.) and diazepam (DZP; 4 mg/ kg, i.p.) on the rota-rod in mice, (A) 30 min., (B) 60 min. and (C) 90 min. after treatment.	43
A1	Principles of sensory modalities	61
A2	Structure of a large neuron in the brain, showing its important functional part.	62
A3	Somatosensory axis of the nervous system.	63
A4	“Second messenger” system.	64
A5	Three states of a neuron. A, Resting neuron, with a normal intraneuronal potential of -65 mV. B, Neuron in an excited state, with a less negative intraneuronal potential (-45 mV) caused by sodium influx. C, Neuron in an inhibited state, with a more negative intraneuronal membrane potential (-70 mV) caused by potassium ion efflux, chloride ion influx, or both.	65
A6	Transmission of both ‘fast-sharp’ and ‘slow-chronic’ pain signals into and through the spinal cord on their way to the brain.	66
A7	Opioid receptors. The ligands for the kappa, mu and delta receptors are shown with the width of the arrows proportionateto the affinity of the receptor for each ligand.	67
A8	Transmission of pain signals into the brain stem, thalamus and cerebral cortex by way of the fast pricking pain pathway and the slow burning pain pathway.	68
A9	Discharge frequencies at different skin temperatures of a cold-pain fiber, a cold fiber, a warmth fiber, and a heat-pain fiber	69
A10	The release of neurotransmitter. Action potentials, by opening Ca <sup>2+</sup> channels, stimulate the fusion of docked synaptic vesicles with the cell membrane of the axon terminals. This leads to exocytosis and the release of neurotransmitter. The activation of protein kinase by Ca <sup>2+</sup> may also contribute to this process.	70
A11	Animal Ethic	71

## LIST OF ABBREVIATIONS

$\mu$	Mu
ADR	adverse drugs reaction
AMPA (isoxazolepropionate)	$\alpha$ -amino-3-hydroxy-5-methyl-4-
ATP	Adenosin triphosphate
BK <sub>ca</sub>	large conductance $Ca^{2+}$ -activated $K^{+}$ channel
cGMP	cyclic guanyl monophosphate
COX-2	cyclooxygenase-2
DAG	diacylglycerol
DMPF 2-	3-(2,3 dimethoxyphenyl)-1(methylfuran-2-yl)pro-
EAA	excitatory amino acids en-1-one
EPSP	excitatory postsynaptic potential
GABA	$\gamma$ -amino butyric acid
GC	Guanylyl cyclase
GluRs	Glutamate receptors
IASP Pain	The International Association for the Study of
IFN- $\gamma$	Interferon- $\gamma$
iNOS	inducible nitric oxide synthase
K <sub>ATP</sub>	ATP-sensitive $K^{+}$ channels
LPS	lipopolysaccharide
mGLUr	Metabotropic glutamate receptor
mV	membrane potential
NGF	nerve growth factor
NMDA	N-methyl-D- aspartate
NO	nitric oxide

NOS	Nitric oxide synthase
NSAIDs	non-steroidal anti-inflammatory drugs
P	relative membrane permeability
PAG	periaqueductal grey
PG	Prostaglandin
PGE2	prostaglandin 2
PKA	Protein kinase A
PKC	Protein kinase C
SG	Substantia gelatinosa
SK <sub>ca</sub>	small conductance ca <sup>2+</sup> -activated K <sup>+</sup> channel
SP	Substance P
TRP	transient receptor potential
TRPV1	transient receptor potential vanilloid 1
VGCCs	voltage-gated calcium channels
VGSCs	voltage-gated sodium channels
WHO	World Health Organization
δ	Delta
κ	Kappa



## CHAPTER 1

### INTRODUCTION

To begin the chapter, the application of noxious stimuli for instance a thermal, mechanical or chemical injury may result in the perception of pain by a conscious individual. There are many complex factors that control the way in which the peripheral nociceptive stimulus undergoes transduction into electrochemical impulses and then onward transmission to reach the central nervous system particularly the cerebral cortex where the actual perception of pain is managed and interpreted. Above all, there is a debate among intelligences saying that pain is not a primary sense unlike vision, audition or hearing, somatosensation, and olfaction which are falling in that category (Price and Dubner 1977). Pain is described to be more of an emotional experience. However, most of the time pain is still considered to be produced by a complex noxious stimulation of the nervous system as explained previously. There are also possibilities that pain is more complicated than what had been studied until today. For instance, the setting of the tissues or nerve injury where the pain is persistent can certainly change the stimulus that evokes pain. These conditions are known as hyperalgesia and neuropathic pain. Under these conditions, sensitivity of pain is increased and the non-painful stimuli, allodynia, will cause production of pain.

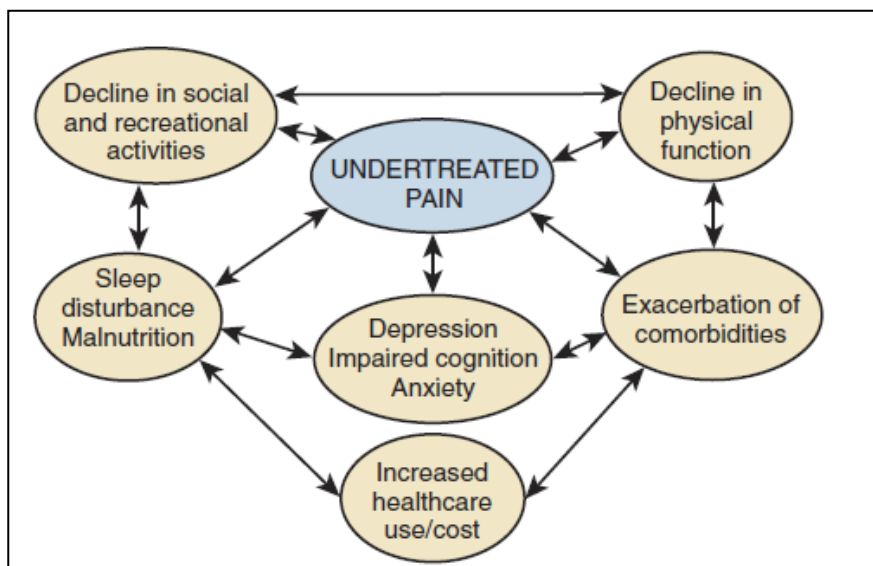
The International Association for the Study of Pain (IASP) defines pain as “an unpleasant sensory and emotional experience associated with actual or potential tissue damage, or described in terms of such damage.” To put it in the simple way, pain always profound the emotional quality to the experience, hence the very discrete physiological and anatomical basis for the detection and transmission of signals that are interpreted as painful experience. Above all the unpleasant experience of pain, there is the reason or function of it. One of the important reasons is that pain served as protective function to protect our body as such withdrawal reflex of our body from the source of noxious simulation and immobility due to pain may serve to provide an environment where the rate of healing is increased and restoration of function can take place. Despite the benefit that pain had offered, it always become prolonged or chronic and when these happened with the addition of tissue damage, central nociceptors pathways are sensitized and reorganized (Figure 1). In such to manage the pain, analgesic drugs is the highly reliant current treatment it acts through weather the central or/and peripheral nervous systems; which includes opioid drugs and non-steroidal anti-inflammatory drugs (NSAIDs) such as morphine and salicylates respectively.

Currently, NSAIDs and opioids are the highly effective treatments that are used widely all over the world as pain management since they are very effective in the treatment of acute nociceptive pain, yet are poor treatment for other several types of pain for example neuropathic pain. Unfortunately, these analgesic drugs can cause several adverse reactions when they are taken in a long period of time. As for example, NSAIDs can cause two major adverse drugs reaction (ADRs) which are gastrointestinal and renal failure while opioids can cause tolerance, addiction and

respiratory depression. In this situation, it occurs that herbal medicine has captured much attention through its benefits to possess antinociceptive, anti-inflammatory, anti-fungal and in fact anti-cancer. Due to all benefits, it positively opens new doors in order to search for the natural products that may take actions as similar as the known analgesic but with either less or no side effects.

According to World Health Organization (WHO), 80% of the world's population uses remedies based on the plants as their primary form of healthcare. In our country specifically, most of the traditional medicinal plants have been handed down from one generation to the next. There are many bioactive compounds that had been isolated from herb and were widely studied for instance chalcone (Sashidhara, Kumar et al. 2011), flavonoid (Kiplimo, Islam et al. 2011), triterpene (Longhi-Balbinot, Martins et al. 2011) and many others more. For this particular study, the play role here is chalcone. Through a number of studies on synthetic compounds with chalcone backbone and these compounds proved to possess antimicrobial (Prasad, Rao et al. 2008), antinociceptive (Correa, Pereira et al. 2001), anti-inflammatory (Hsieh, Tsao et al. 2000), and antiulcerative activity (Murakami, Muramatsu et al. 1991). Previous studies have shown that chalcones bearing phenyl group with chloro and hydroxyl substituents inhibit nitric oxide (NO) synthesis as well as inducible nitric oxide synthase (iNOS) protein expression in RAW 264.7 cells (Ko, Tsao et al. 2003; Won, Liu et al. 2005). In the study done by our research group on a naturally occurring chalcone isolated from the fruits of *Alpinia rafflesiana*, cardamonin, we found that it suppressed both NO and PGE<sub>2</sub> (prostaglandin 2) in interferon- $\gamma$  (IFN- $\gamma$ - and lipopolysaccharide (LPS)-induced RAW 264.7 cells (Ahmad, Israf et al. 2006; Israf, Khaizurin et al. 2007). Another chalcone compound is isolated from bark of *Toxicodendron vernicifluum* and this compound found to have anti-inflammatory effect (Kim, Moon et al.). It has been reported elsewhere that chalcone derivative isolated from root of *Glycyrrhiza uralensis* inhibited the synthesis of nitric oxide (NO), lipooxygenase as well as cyclo-oxygenase activities which constitute the major pro-inflammatory pathways and remain most targeted for anti-inflammatory and antinociceptive drug development (Kim, Park et al. 2008).

Since most pain originates from inflammation and the role of NO in pain is very important as it has been stated that without the production of NO there will be no generation of pain. Thus, with the support of the strong basis on the previous study, the group had done the investigation on antinociceptive activity of DMPF using the chemical- and thermal-induced nociception models in mice.



**Figure 1: Consequences of undertreatment of pain**  
(Gureje, Von Korff et al. 1998).

## 1.1 Objectives

The objectives of the study are:

### General

- To determine antinociceptive activity of DMPF compound and its possible mechanisms of action.

### Specific

To determine :

- the involvement of peripheral and central antinociceptive activity of DMPF compound.
- the involvement of opioid receptors, TRPV1 receptor, glutamatergic system, NO-cGMP pathway, various types of potassium channels and protein kinase C system.

## REFERENCES

- Abacioglu, N., B. Tunctan, et al. (2000). "Participation of the components of L-arginine/nitric oxide/cGMP cascade by chemically-induced abdominal constriction in the mouse." *Life Sci* **67**(10): 1127-37.
- Ahmad, S., L. Dahllund, et al. (2007). "A stop codon mutation in SCN9A causes lack of pain sensation." *Hum Mol Genet* **16**(17): 2114-21.
- Ahmad, S., D. A. Israf, et al. (2006). "Cardamonin, inhibits pro-inflammatory mediators in activated RAW 264.7 cells and whole blood." *Eur J Pharmacol* **538**(1-3): 188-94.
- Ahmadiani, A., M. Fereidoni, et al. (1998). "Antinociceptive and anti-inflammatory effects of *Sambucus ebulus* rhizome extract in rats." *Journal of ethnopharmacology* **61**(3): 229-235.
- Aiello, E. A., M. P. Walsh, et al. (1995). "Phosphorylation by protein kinase A enhances delayed rectifier K<sup>+</sup> current in rabbit vascular smooth muscle cells." *Am J Physiol* **268**(2 Pt 2): H926-34.
- Alioua, A., Y. Tanaka, et al. (1998). "The large conductance, voltage-dependent, and calcium-sensitive K<sup>+</sup> channel, Hslo, is a target of cGMP-dependent protein kinase phosphorylation in vivo." *J Biol Chem* **273**(49): 32950-6.
- Barnstable, C. J., J. Y. Wei, et al. (2004). "Modulation of synaptic function by cGMP and cGMP-gated cation channels." *Neurochem Int* **45**(6): 875-84.
- Battaglia, G. and A. Rustioni (1988). "Coexistence of glutamate and substance P in dorsal root ganglion neurons of the rat and monkey." *J Comp Neurol* **277**(2): 302-12.
- Beirith, A., A. R. Santos, et al. (2002). "Mechanisms underlying the nociception and paw oedema caused by injection of glutamate into the mouse paw." *Brain Res* **924**(2): 219-28.
- Bermudez-Ocana, D. Y., M. Ambriz-Tututi, et al. (2006). "Pharmacological evidence for the participation of NO-cyclic GMP-PKG-K<sup>+</sup> channel pathway in the antiallodynic action of resveratrol." *Pharmacol Biochem Behav* **84**(3): 535-42.
- Bernardi, H., M. Fosset, et al. (1988). "Characterization, purification, and affinity labeling of the brain [3H]glibenclamide-binding protein, a putative neuronal ATP-regulated K<sup>+</sup> channel." *Proc Natl Acad Sci U S A* **85**(24): 9816-20.
- Bhave, G., H. J. Hu, et al. (2003). "Protein kinase C phosphorylation sensitizes but does not activate the capsaicin receptor transient receptor potential vanilloid 1 (TRPV1)." *Proc Natl Acad Sci U S A* **100**(21): 12480-5.
- Bowery, N. G., A. Doble, et al. (1979). "Baclofen: a selective agonist for a novel type of GABA receptor proceedings." *Br J Pharmacol* **67**(3): 444P-445P.
- Bown, D. (2001). *New Encyclopedia of Herbs & Their Uses*: The Herb Society of America, Dorling Kindersley Publishing.

- Buck, M. and J. A. Paice (1994). "Pharmacologic management of acute pain in the orthopaedic patient." *Orthop Nurs* **13**(6): 14-23; quiz 24.
- Cabranes, A., K. Venderova, et al. (2005). "Decreased endocannabinoid levels in the brain and beneficial effects of agents activating cannabinoid and/or vanilloid receptors in a rat model of multiple sclerosis." *Neurobiol Dis* **20**(2): 207-17.
- Catterall, W. A. (2000). "From ionic currents to molecular mechanisms: the structure and function of voltage-gated sodium channels." *Neuron* **26**(1): 13-25.
- Clement-Chomienne, O., M. P. Walsh, et al. (1996). "Angiotensin II activation of protein kinase C decreases delayed rectifier K<sup>+</sup> current in rabbit vascular myocytes." *J Physiol* **495** ( Pt 3): 689-700.
- Coderre, T. J. and K. Yashpal (1994). "Intracellular messengers contributing to persistent nociception and hyperalgesia induced by L-glutamate and substance P in the rat formalin pain model." *Eur J Neurosci* **6**(8): 1328-34.
- Cooke S.A., C. G. K., Legon A.C. (1998). "Rotational spectrum of thiophene? HCl. Does thiophene act as an aromatic p-type electron donor or an n-type electron donor in hydrogen-bond formation? ." *J Chem* **94**(11): 1565–1570.
- Correa, C. R., D. J. Kyle, et al. (1996). "Antinociceptive profile of the pseudopeptide B2 bradykinin receptor antagonist NPC 18688 in mice." *Br J Pharmacol* **117**(3): 552-558.
- Correa, R., M. A. Pereira, et al. (2001). "Antinociceptive properties of chalcones. Structure-activity relationships." *Arch Pharm (Weinheim)* **334**(10): 332-4.
- Curtis, D. (2002). "PAIN: it's all in the mind?", from <http://www.david.curtis.care4free.net/painrev.htm>.
- Curtis, D. R., J. W. Phillis, et al. (1959). "Chemical excitation of spinal neurones." *Nature* **183**(4661): 611-2.
- De Biasi, S. and A. Rustioni (1988). "Glutamate and substance P coexist in primary afferent terminals in the superficial laminae of spinal cord." *Proc Natl Acad Sci U S A* **85**(20): 7820-4.
- de Moura, R. S., A. A. Rios, et al. (2004). "Role of the NO-cGMP pathway in the systemic antinociceptive effect of clonidine in rats and mice." *Pharmacol Biochem Behav* **78**(2): 247-53.
- Deguchi, A., S. W. Xing, et al. (2005). "Activation of protein kinase G up-regulates expression of 15-lipoxygenase-1 in human colon cancer cells." *Cancer Res* **65**(18): 8442-7.
- Demirkoprulu, N., M. Cetin, et al. (2005). "Comparative relaxant effects of YC-1 and DETA/NO on spontaneous contractions and the levels of cGMP of isolated pregnant rat myometrium." *Eur J Pharmacol* **517**(3): 240-5.
- Desarmenien, M., P. Feltz, et al. (1984). "Coexistence of GABAA and GABAB receptors on A delta and C primary afferents." *Br J Pharmacol* **81**(2): 327-33.



- Di Marzo, V., P. M. Blumberg, et al. (2002). "Endovanilloid signaling in pain." *Curr Opin Neurobiol* **12**(4): 372-9.
- Dougherty, P. M. and W. D. Willis (1991). "Enhancement of spinothalamic neuron responses to chemical and mechanical stimuli following combined micro-iontophoretic application of N-methyl-D-aspartic acid and substance P." *Pain* **47**(1): 85-93.
- Dunlap, K. (1984). "Functional and pharmacological differences between two types of GABA receptor on embryonic chick sensory neurons." *Neurosci Lett* **47**(3): 265-70.
- El-Mowafy, A. M. (2002). "Resveratrol activates membrane-bound guanylyl cyclase in coronary arterial smooth muscle: a novel signaling mechanism in support of coronary protection." *Biochem Biophys Res Commun* **291**(5): 1218-24.
- England, S., S. Bevan, et al. (1996). "PGE2 modulates the tetrodotoxin-resistant sodium current in neonatal rat dorsal root ganglion neurones via the cyclic AMP-protein kinase A cascade." *J Physiol* **495** ( Pt 2): 429-40.
- Fox, S. (2012). *Human physiology*, McGraw-Hill Higher Education.
- Fredriksson, R., M. C. Lagerstrom, et al. (2003). "The G-protein-coupled receptors in the human genome form five main families. Phylogenetic analysis, paralogon groups, and fingerprints." *Mol Pharmacol* **63**(6): 1256-72.
- Friebe, A. and D. Koesling (2003). "Regulation of nitric oxide-sensitive guanylyl cyclase." *Circ Res* **93**(2): 96-105.
- Fukao, M., H. S. Mason, et al. (1999). "Cyclic GMP-dependent protein kinase activates cloned BKCa channels expressed in mammalian cells by direct phosphorylation at serine 1072." *J Biol Chem* **274**(16): 10927-35.
- Ganong, W. F., & Barrett, K. E. (2010). *Review of medical physiology*, New York: McGraw-Hill Medical.
- Garcia, M. L., H. G. Knaus, et al. (1995). "Charybdotoxin and its effects on potassium channels." *Am J Physiol* **269**(1 Pt 1): C1-10.
- Garthwaite, J. (1991). "Glutamate, nitric oxide and cell-cell signalling in the nervous system." *Trends Neurosci* **14**(2): 60-7.
- Gelband, C. H. and J. R. Hume (1995). "[Ca<sup>2+</sup>]<sub>i</sub> inhibition of K<sup>+</sup> channels in canine renal artery. Novel mechanism for agonist-induced membrane depolarization." *Circ Res* **77**(1): 121-30.
- Gelband, C. H., T. Ishikawa, et al. (1993). "Intracellular divalent cations block smooth muscle K<sup>+</sup> channels." *Circ Res* **73**(1): 24-34.
- Gold, M. S., D. B. Reichling, et al. (1996). "Hyperalgesic agents increase a tetrodotoxin-resistant Na<sup>+</sup> current in nociceptors." *Proc Natl Acad Sci U S A* **93**(3): 1108-12.

- Goldberg, Y. P., J. MacFarlane, et al. (2007). "Loss-of-function mutations in the Nav1.7 gene underlie congenital indifference to pain in multiple human populations." *Clin Genet* **71**(4): 311-9.
- Goldin, A. L., R. L. Barchi, et al. (2000). "Nomenclature of voltage-gated sodium channels." *Neuron* **28**(2): 365-8.
- Goncales, C. E., D. Araldi, et al. (2005). "Antinociceptive properties of acetylenic thiophene and furan derivatives: evidence for the mechanism of action." *Life Sci* **76**(19): 2221-34.
- Gureje, O., M. Von Korff, et al. (1998). "Persistent pain and well-being: a World Health Organization Study in Primary Care." *JAMA* **280**(2): 147-51.
- Gybels, J. M. and W. H. Sweet (1989). "Neurosurgical treatment of persistent pain. Physiological and pathological mechanisms of human pain." *Pain Headache* **11**: 1-402.
- Habsah, M., M. Amran, et al. (2000). "Screening of Zingiberaceae extracts for antimicrobial and antioxidant activities." *J Ethnopharmacol* **72**(3): 403-10.
- Hall, J. E., & Guyton, A. C. (2011). *Textbook of medical physiology*, Saunders.
- Hirata, M., K. P. Kohse, et al. (1990). "Mechanism of cyclic GMP inhibition of inositol phosphate formation in rat aorta segments and cultured bovine aortic smooth muscle cells." *J Biol Chem* **265**(3): 1268-73.
- Hollmann, M. and S. Heinemann (1994). "Cloned glutamate receptors." *Annu Rev Neurosci* **17**: 31-108.
- Hsieh, H. K., L. T. Tsao, et al. (2000). "Synthesis and anti-inflammatory effect of chalcones." *J Pharm Pharmacol* **52**(2): 163-71.
- Huang, Y. C., J. H. Guh, et al. (2001). "Inhibitory effect of DCDC on lipopolysaccharide-induced nitric oxide synthesis in RAW 264.7 cells." *Life Sci* **68**(21): 2435-47.
- Huettner, J. E. (2003). "Kainate receptors and synaptic transmission." *Prog Neurobiol* **70**(5): 387-407.
- Hunnskaar, S. (1987). "Similar effects of acetylsalicylic acid and morphine on immediate responses to acute noxious stimulation." *Pharmacol Toxicol* **60**(3): 167-70.
- Hunnskaar, S. and K. Hole (1987). "The formalin test in mice: dissociation between inflammatory and non-inflammatory pain." *Pain* **30**(1): 103-14.
- Ikeda, Y., A. Ueno, et al. (2001). "Involvement of vanilloid receptor VR1 and prostanooids in the acid-induced writhing responses of mice." *Life Sci* **69**(24): 2911-9.
- Israfil, D. A., T. A. Khaizurin, et al. (2007). "Cardamonin inhibits COX and iNOS expression via inhibition of p65NF-kappaB nuclear translocation and Ikappa-B phosphorylation in RAW 264.7 macrophage cells." *Mol Immunol* **44**(5): 673-9.

- Jayasinghe, L., B. Balasooriya, et al. (2004). "Geranyl chalcone derivatives with antifungal and radical scavenging properties from the leaves of *Artocarpus nobilis*." *Phytochemistry* **65**(9): 1287-1290.
- Johnson, J. W. and P. Ascher (1987). "Glycine potentiates the NMDA response in cultured mouse brain neurons." *Nature* **325**(6104): 529-31.
- Julius, D. and A. I. Basbaum (2001). "Molecular mechanisms of nociception." *Nature* **413**(6852): 203-10.
- Karbon, E. W., R. S. Duman, et al. (1984). "GABAB receptors and norepinephrine-stimulated cAMP production in rat brain cortex." *Brain Res* **306**(1-2): 327-32.
- Karim, F., C. C. Wang, et al. (2001). "Metabotropic glutamate receptor subtypes 1 and 5 are activators of extracellular signal-regulated kinase signaling required for inflammatory pain in mice." *J Neurosci* **21**(11): 3771-9.
- Katsuki, S., W. Arnold, et al. (1977). "Stimulation of guanylate cyclase by sodium nitroprusside, nitroglycerin and nitric oxide in various tissue preparations and comparison to the effects of sodium azide and hydroxylamine." *J Cyclic Nucleotide Res* **3**(1): 23-35.
- Kawabata, S., A. Kohara, et al. (1998). "Diversity of calcium signaling by metabotropic glutamate receptors." *J Biol Chem* **273**(28): 17381-5.
- Kawabata, S., R. Tsutsumi, et al. (1996). "Control of calcium oscillations by phosphorylation of metabotropic glutamate receptors." *Nature* **383**(6595): 89-92.
- Kawasaki, Y., T. Kohno, et al. (2004). "Ionotropic and metabotropic receptors, protein kinase A, protein kinase C, and Src contribute to C-fiber-induced ERK activation and cAMP response element-binding protein phosphorylation in dorsal horn neurons, leading to central sensitization." *J Neurosci* **24**(38): 8310-21.
- Kelly, S. J., & Franklin, K. B. J. (1985). "An increase in tryptophan in brain may be a general mechanism for the effect of stress on sensitivity to pain." *Neuropharmacology* **24**(11): 1019-1025.
- Khalid, M. H., M. N. Akhtar, et al. (2011). "Antinociceptive effect of the essential oil of *Zingiber zerumbet* in mice: possible mechanisms." *J Ethnopharmacol* **137**(1): 345-51.
- Kim, C. H., S. Braud, et al. (2005). "Protein kinase C phosphorylation of the metabotropic glutamate receptor mGluR5 on Serine 839 regulates Ca<sup>2+</sup> oscillations." *J Biol Chem* **280**(27): 25409-15.
- Kim, J. Y., S. J. Park, et al. (2008). "Isoliquiritigenin isolated from the roots of *Glycyrrhiza uralensis* inhibits LPS-induced iNOS and COX-2 expression via the attenuation of NF-kappaB in RAW 264.7 macrophages." *Eur J Pharmacol* **584**(1): 175-84.



- Kim, K. H., E. Moon, et al. "Identification of cytotoxic and anti-inflammatory constituents from the bark of *Toxicodendron vernicifluum* (Stokes) F.A. Barkley." *J Ethnopharmacol* **162**: 231-7.
- Kiplimo, J. J., M. S. Islam, et al. (2011). "A novel flavonoid and furoquinoline alkaloids from *Vepris glomerata* and their antioxidant activity." *Nat Prod Commun* **6**(12): 1847-50.
- Knot, H. J. and M. T. Nelson (1995). "Regulation of membrane potential and diameter by voltage-dependent K<sup>+</sup> channels in rabbit myogenic cerebral arteries." *Am J Physiol* **269**(1 Pt 2): H348-55.
- Knox, C. M. (2012). Pain Management. *Essentials of Correctional Nursing*: 247.
- Ko, H. H., L. T. Tsao, et al. (2003). "Structure-activity relationship studies on chalcone derivatives. the potent inhibition of chemical mediators release." *Bioorg Med Chem* **11**(1): 105-11.
- Kopin, I. J. (1976). "Catecholamines, adrenal hormones, and stress." *Hospital practice*, **11**(3), 49-55.
- Krause, J. E., B. L. Chenard, et al. (2005). "Transient receptor potential ion channels as targets for the discovery of pain therapeutics." *Curr Opin Investig Drugs* **6**(1): 48-57.
- Lam, K., R. Uddin, et al. (2012). "Synthesis and QSAR analysis of chalcone derivatives as nitric oxide inhibitory agent." *Medicinal Chemistry Research* **21**(8): 1953-1966.
- Le Bars, D., M. Gozariu, et al. (2001). "Animal models of nociception." *Pharmacol Rev* **53**(4): 597-652.
- Lembeck, F., J. Donnerer, et al. (1992). "The non-peptide tachykinin antagonist, CP-96,345, is a potent inhibitor of neurogenic inflammation." *Br J Pharmacol* **105**(3): 527-30.
- Liew, C. Y., C. L. Tham, et al. (2010). "A synthetic hydroxypropenone inhibits nitric oxide, prostaglandin E<sub>2</sub>, and proinflammatory cytokine synthesis." *Immunopharmacol Immunotoxicol* **32**(3): 495-506.
- Lin, D. T., P. Fretier, et al. (2010). "Nitric oxide signaling via cGMP-stimulated phosphodiesterase in striatal neurons." *Synapse* **64**(6): 460-6.
- Lin, Q., Y. B. Peng, et al. (1996). "Role of GABA receptor subtypes in inhibition of primate spinothalamic tract neurons: difference between spinal and periaqueductal gray inhibition." *J Neurophysiol* **75**(1): 109-23.
- Lohmann, A. B. and S. P. Welch (1999). "Antisenses to opioid receptors attenuate ATP-gated K<sup>(+)</sup> channel opener-induced antinociception." *Eur J Pharmacol* **384**(2-3): 147-52.
- Lohmann, A. B. and S. P. Welch (1999). "ATP-gated K<sup>(+)</sup> channel openers enhance opioid antinociception: indirect evidence for the release of endogenous opioid peptides." *Eur J Pharmacol* **385**(2-3): 119-27.

- Longhi-Balbinot, D. T., D. F. Martins, et al. (2011). "Further analyses of mechanisms underlying the antinociceptive effect of the triterpene 3beta, 6beta, 16beta-trihydroxylup-20(29)-ene in mice." *Eur J Pharmacol* **653**(1-3): 32-40.
- Lorke, D. (1983). "A new approach to practical acute toxicity testing." *Arch Toxicol* **54**(4): 275-87.
- Luiz, A. P., J. D. Moura, et al. (2007). "Antinociceptive action of ethanolic extract obtained from roots of *Humirianthera ampla* Miers." *J Ethnopharmacol* **114**(3): 355-63.
- Malmberg, A. B., E. P. Brandon, et al. (1997). "Diminished inflammation and nociceptive pain with preservation of neuropathic pain in mice with a targeted mutation of the type I regulatory subunit of cAMP-dependent protein kinase." *J Neurosci* **17**(19): 7462-70.
- Mateeva, N., M. Gangapuram, et al. (2014). "Biological evaluation of synthetic chalcone and flavone derivatives as anti-inflammatory agents." *Med Chem Res* **24**(4): 1672-1680.
- McDonald, B. J., H. J. Chung, et al. (2001). "Identification of protein kinase C phosphorylation sites within the AMPA receptor GluR2 subunit." *Neuropharmacology* **41**(6): 672-9.
- Meller, S. T., C. Dykstra, et al. (1992). "Production of endogenous nitric oxide and activation of soluble guanylate cyclase are required for N-methyl-D-aspartate-produced facilitation of the nociceptive tail-flick reflex." *Eur J Pharmacol* **214**(1): 93-6.
- Melzack, R. and P. D. Wall (1965). "Pain mechanisms: a new theory." *Science* **150**(3699): 971-9.
- Miller, C. (1995). "The charybdotoxin family of K<sup>+</sup> channel-blocking peptides." *Neuron* **15**(1): 5-10.
- Ming-Tatt, L., S. I. Khalivulla, et al. (2012). "Antinociceptive Activity of a Synthetic Curcuminoid Analogue, 2, 6-bis-(4-hydroxy-3-methoxybenzylidene) cyclohexanone, on Nociception-induced Models in Mice." *Basic & clinical pharmacology & toxicology* **110**(3): 275-282.
- Mohamad, A. S., M. N. Akhtar, et al. (2011). "Possible participation of nitric oxide/cyclic guanosine monophosphate/protein kinase C/ATP-sensitive K<sup>(+)</sup> channels pathway in the systemic antinociception of flavokawin B." *Basic Clin Pharmacol Toxicol* **108**(6): 400-5.
- Mohamad, A. S., M. N. Akhtar, et al. "Antinociceptive activity of a synthetic chalcone, flavokawin B on chemical and thermal models of nociception in mice." *Eur J Pharmacol* **647**(1-3): 103-9.
- Moore, P. K., A. O. Oluyomi, et al. (1991). "L-NG-nitro arginine methyl ester exhibits antinociceptive activity in the mouse." *Br J Pharmacol* **102**(1): 198-202.
- Moran, M. M., H. Xu, et al. (2004). "TRP ion channels in the nervous system." *Curr Opin Neurobiol* **14**(3): 362-9.

- Moreau, K., D. Morel, et al. (1997). "[Cardiorespiratory arrest following ingestion of morphine sulfate in a patient with chronic renal failure]." *Presse Med* **26**(15): 713.
- Moussaoui, S. M., F. Montier, et al. (1993). "A non-peptide NK1-receptor antagonist, RP 67580, inhibits neurogenic inflammation postsynaptically." *Br J Pharmacol* **109**(1): 259-64.
- Murakami, S., M. Muramatsu, et al. (1991). "Inhibition of gastric H<sub>+</sub>,K (+)-ATPase by the anti-ulcer agent, sofalcone." *Biochem Pharmacol* **42**(7): 1447-51.
- Murase, K., P. D. Ryu, et al. (1989). "Excitatory and inhibitory amino acids and peptide-induced responses in acutely isolated rat spinal dorsal horn neurons." *Neurosci Lett* **103**(1): 56-63.
- Newberry, N. R. and R. A. Nicoll (1985). "Comparison of the action of baclofen with gamma-aminobutyric acid on rat hippocampal pyramidal cells in vitro." *J Physiol* **360**: 161-85.
- Ni, L., C. Q. Meng, et al. (2004). "Recent advances in therapeutic chalcones." *Expert Opinion on Therapeutic Patents* **14**(12): 1669-1691.
- Ocana, M., M. Barrios, et al. (1996). "Cromakalim differentially enhances antinociception induced by agonists of alpha (2)adrenoceptors, gamma-aminobutyric acid(B), mu and kappa opioid receptors." *J Pharmacol Exp Ther* **276**(3): 1136-42.
- Olson, G. A., R. D. Olson, et al. (1989). "Endogenous opiates: 1988." *Peptides* **10**(6): 1253-80.
- Ota, K. T., V. J. Pierre, et al. (2008). "The NO-cGMP-PKG signaling pathway regulates synaptic plasticity and fear memory consolidation in the lateral amygdala via activation of ERK/MAP kinase." *Learn Mem* **15**(10): 792-805.
- Palecek, J., V. Paleckova, et al. (1992). "Responses of spinothalamic tract cells to mechanical and thermal stimulation of skin in rats with experimental peripheral neuropathy." *J Neurophysiol* **67**(6): 1562-73.
- Palecek, J., V. Paleckova, et al. (2003). "Fos expression in spinothalamic and postsynaptic dorsal column neurons following noxious visceral and cutaneous stimuli." *Pain* **104**(1-2): 249-57.
- Paleckova, V., J. Palecek, et al. (1992). "The non-NMDA antagonist CNQX prevents release of amino acids into the rat spinal cord dorsal horn evoked by sciatic nerve stimulation." *Neurosci Lett* **148**(1-2): 19-22.
- Patapoutian, A., S. Tate, et al. (2009). "Transient receptor potential channels: targeting pain at the source." *Nat Rev Drug Discov* **8**(1): 55-68.
- Perimal, E. K., M. N. Akhtar, et al. (2011). "Zerumbone-induced antinociception: involvement of the L-arginine-nitric oxide-cGMP -PKC-K<sup>+</sup> ATP channel pathways." *Basic Clin Pharmacol Toxicol* **108**(3): 155-62.

- Pinho-Ribeiro, F. A., M. S. Hohmann, et al. (2015). "Protective effects of the flavonoid hesperidin methyl chalcone in inflammation and pain in mice: role of TRPV1, oxidative stress, cytokines and NF-kappaB." *Chem Biol Interact* **228**: 88-99.
- Porszasz, J. and N. Jancso (1959). "Studies on the action potentials of sensory nerves in animals desensitized with capsaicine." *Acta Physiol Acad Sci Hung* **16**: 299-306.
- Prasad, Y. R., A. L. Rao, et al. (2008). "Synthesis and Antimicrobial Activity of Some Chalcone Derivatives." *E-Journal of Chemistry* **5**(3): 461-466.
- Premkumar, L. S. and G. P. Ahern (2000). "Induction of vanilloid receptor channel activity by protein kinase C." *Nature* **408**(6815): 985-90.
- Price, D. D. and R. Dubner (1977). "Neurons that subserve the sensory-discriminative aspects of pain." *Pain* **3**(4): 307-38.
- Robertson, B. E., R. Schubert, et al. (1993). "cGMP- dependent protein kinase activates Ca-activated K channels in cerebral artery smooth muscle cells." *Am J Physiol* **265**(1 Pt 1): C299-303.
- Roche, K. W., R. J. O'Brien, et al. (1996). "Characterization of multiple phosphorylation sites on the AMPA receptor GluR1 subunit." *Neuron* **16**(6): 1179-88.
- Rosland, J. H., S. Hunskaar, et al. (1990). "Diazepam attenuates morphine antinociception test-dependently in mice." *Pharmacol Toxicol* **66**(5): 382-6.
- Rudomin, P. and R. F. Schmidt (1999). "Presynaptic inhibition in the vertebrate spinal cord revisited." *Exp Brain Res* **129**(1): 1-37.
- Ruth, P., G. X. Wang, et al. (1993). "Transfected cGMP-dependent protein kinase suppresses calcium transients by inhibition of inositol 1,4,5-trisphosphate production." *Proc Natl Acad Sci U S A* **90**(7): 2623-7.
- Sakurada, T., T. Matsumura, et al. (2003). "Differential effects of intraplantar capsaizepine and ruthenium red on capsaicin-induced desensitization in mice." *Pharmacol Biochem Behav* **75**(1): 115-21.
- Sashidhara, K. V., M. Kumar, et al. (2011). "Synthesis and anti-inflammatory activity of novel biscoumarin-chalcone hybrids." *Bioorg Med Chem Lett* **21**(15): 4480-4.
- Sathianathan, V., A. Avelino, et al. (2003). "Insulin induces cobalt uptake in a subpopulation of rat cultured primary sensory neurons." *Eur J Neurosci* **18**(9): 2477-86.
- Sausbier, M., R. Schubert, et al. (2000). "Mechanisms of NO/cGMP-dependent vasorelaxation." *Circ Res* **87**(9): 825-30.
- Schneggenburger, R., Z. Zhou, et al. (1993). "Fractional contribution of calcium to the cation current through glutamate receptor channels." *Neuron* **11**(1): 133-43.
- Skeberdis, V. A., V. Chevaleyre, et al. (2006). "Protein kinase A regulates calcium permeability of NMDA receptors." *Nat Neurosci* **9**(4): 501-10.

- Suh, Y. G. and U. Oh (2005). "Activation and activators of TRPV1 and their pharmaceutical implication." *Curr Pharm Des* **11**(21): 2687-98.
- Sulaiman, M. R., T. A. Tengku Mohamad, et al. "Antinociceptive activity of the essential oil of Zingiber zerumbet." *Planta Med* **76**(2): 107-12.
- Szliszka, E., Z. P. Czuba, et al. (2010). "Chalcones and dihydrochalcones augment TRAIL-mediated apoptosis in prostate cancer cells." *Molecules* **15**(8): 5336-53.
- Tominaga, M., M. Wada, et al. (2001). "Potentiation of capsaicin receptor activity by metabotropic ATP receptors as a possible mechanism for ATP-evoked pain and hyperalgesia." *Proc Natl Acad Sci U S A* **98**(12): 6951-6.
- Tzavara, E. T., D. L. Li, et al. (2006). "Endocannabinoids activate transient receptor potential vanilloid 1 receptors to reduce hyperdopaminergia-related hyperactivity: therapeutic implications." *Biol Psychiatry* **59**(6): 508-15.
- Urban, L. and M. Randic (1984). "Slow excitatory transmission in rat dorsal horn: possible mediation by peptides." *Brain Res* **290**(2): 336-41.
- Vane, J. R. (1971). "Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs." *Nat New Biol* **231**(25): 232-5.
- Vergoni, A. V., A. Scarano, et al. (1992). "Pinacidil potentiates morphine analgesia." *Life Sci* **50**(16): PL135-8.
- Viana, G. S. B., M. A. M. Bandeira, et al. (2003). "Analgesic and antiinflammatory effects of chalcones isolated from *Myracrodruon urundeuva* Allemão." *Phytomedicine* **10**(2): 189-195.
- Wajima, Z., X. Y. Hua, et al. (2000). "Inhibition of spinal protein kinase C blocks substance P-mediated hyperalgesia." *Brain Res* **877**(2): 314-21.
- Wang, X. and P. J. Robinson (1997). "Cyclic GMP-dependent protein kinase and cellular signaling in the nervous system." *J Neurochem* **68**(2): 443-56.
- White, R. E., A. B. Lee, et al. (1993). "Potassium channel stimulation by natriuretic peptides through cGMP-dependent dephosphorylation." *Nature* **361**(6409): 263-6.
- Womack, M. D., A. B. MacDermott, et al. (1988). "Sensory transmitters regulate intracellular calcium in dorsal horn neurons." *Nature* **334**(6180): 351-3.
- Won, S. J., C. T. Liu, et al. (2005). "Synthetic chalcones as potential anti-inflammatory and cancer chemopreventive agents." *Eur J Med Chem* **40**(1): 103-12.
- Yaksh, T. L. (1988). "Substance P release from knee joint afferent terminals: modulation by opioids." *Brain Res* **458**(2): 319-24.
- Yonehara, N., Y. Imai, et al. (1989). "Participation of substance P in inflammatory responses." *Adv Exp Med Biol* **247B**: 529-34.
- Zhou, X. B., P. Ruth, et al. (1996). "Protein phosphatase 2A is essential for the activation of Ca<sup>2+</sup>-activated K<sup>+</sup> currents by cGMP-dependent protein kinase

in tracheal smooth muscle and Chinese hamster ovary cells." *J Biol Chem* **271**(33): 19760-7.

Zieglansberger, W. and B. Sutor (1983). "Responses of substantia gelatinosa neurons to putative neurotransmitters in an in vitro preparation of the adult rat spinal cord." *Brain Res* **279**(1-2): 316-20.

Zou, X., Q. Lin, et al. (2000). "Enhanced phosphorylation of NMDA receptor 1 subunits in spinal cord dorsal horn and spinothalamic tract neurons after intradermal injection of capsaicin in rats." *J Neurosci* **20**(18): 6989-97.

Zou, X., Q. Lin, et al. (2002). "Role of protein kinase A in phosphorylation of NMDA receptor 1 subunits in dorsal horn and spinothalamic tract neurons after intradermal injection of capsaicin in rats." *Neuroscience* **115**(3): 775-86.

Zygmunt, P. M., J. Petersson, et al. (1999). "Vanilloid receptors on sensory nerves mediate the vasodilator action of anandamide." *Nature* **400**(6743): 452-7.