

UNIVERSITI PUTRA MALAYSIA

MOLECULAR CHARACTERIZATION AND RECOMBINANT GONADOTROPIN SUBUNIT DEVELOPMENT FOR IMPROVING REPRODUCTIVE PERFORMANCE IN FEMALE Hemibagrus nemurus VALENCIENNES

ZARIRAH BINTI MOHAMED ZULPERI

FP 2016 86



MOLECULAR CHARACTERIZATION AND RECOMBINANT GONADOTROPIN SUBUNIT DEVELOPMENT FOR IMPROVING REPRODUCTIVE PERFORMANCE IN FEMALE Hemibagrus nemurus VALENCIENNES



By

ZARIRAH BINTI MOHAMED ZULPERI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

October 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Assalamualaíkum...

Peace be upon you...

C

Ô

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

MOLECULAR CHARACTERIZATION AND RECOMBINANT GONADOTROPIN SUBUNIT DEVELOPMENT FOR IMPROVING REPRODUCTIVE PERFORMANCE IN FEMALE Hemibagrus nemurus VALENCIENNES

By

ZARIRAH BINTI MOHAMED ZULPERI

October 2016

Chairman : Ina Salwany Md Yasin, PhD Faculty : Agriculture

Hemibagrus nemurus, locally known as 'ikan baung', is a valuable cultured catfish in Malaysia, but the production is limited due to problems associated with its reproduction. Most hatcheries encountered problems in induced spawning due to the fact that the fish do not expose to their natural spawning habitat, especially the failure of female brood stock to undergo final oocyte maturation (FOM), preventing its ability to ovulate and spawn. Gonadotropin (GtH), comprised of follicle-stimulating hormone (FSH) and luteinizing hormone (LH), play an important role in gametogenesis and sexual maturation in mammals as well as in fish. This research was conducted to study the molecular characterization and to monitor changes in the mRNA expression of three GtH subunits, α , FSH β , and LH β , during the reproductive cycle of the female H. nemurus. Next, a single-chain recombinant LH (rLH) of H. nemurus was expressed using E. coli expression system. The bioactivity of the recombinant hormone was assessed for their potency in stimulating the production of 17_B-estradiol (E2), ovarian development and mRNA expression level of GtH subunits in experimental fish. Molecular characterization using PCR amplifications revealed that the cDNA of α , FSH_B and LH_B in *H. nemurus* were 656, 728 and 602 bp, and encoded for 116, 134 and 140 amino acids, respectively. Multiple sequence alignment showed all the three subunits shared the highest homology with Siluriformes and Anguilliformes, and least similar to higher vertebrates. Phylogenetic analysis of α and FSH β grouped fishes and higher vertebrates together, while in LH β , the tree was divided in two distinct groups, the first group contained all fishes together, and the second group only contained higher vertebrates. The abundance of transcripts encoding the GtH subunits, α , FSH^β and LH^β subunits in pituitaries of female *H. nemurus* was tracked throughout the reproductive cycle using quantitative real-time PCR. Data were collected from August 2013 until October 2015, with fish aged between 1 to 27 month-old. The mRNA expression of GtH subunits increased together



with ovarian development throughout the reproductive cycle. The FSH β mRNA level increased during ovarian development and reached its peak in mid-vitellogenesis and reduced during ovulation and spawning, whereas the LHB mRNA level significantly increased (P<0.05) in late vitellogenesis and reached its peak during ovulation and spawning period. In α subunit, the mRNA level corresponded to FSH β and LH β , which showed its highest expression during ovulation. This suggest that FSH could be involved in vitellogenesis, while LH β subunit raised at the end of reproductive cycle, determined the involvement of LH in ovulation and final oocyte maturation. A single chain recombinant LH (rLH) from H. nemurus consisted of LHBa fusion proteins was constructed and derived through gene-synthesis. The rLH construct was successfully cloned into pET-32 Ek/LIC and expressed in E. coli BL21 (DE3) cells. A specific band at 45.4 kDa was detected which corresponded to the molecular size of fusion rLH resolved by SDS PAGE. Two monoclonal antibodies, His-Tag and S-tag, were used to confirm the same expressed bands in Western immunoblotting. In order to investigate the bioactivity of the rLH hormone, fish trial was performed to assess the effect of exogenous rLH on the plasma level of E2, their mRNA level of GtH subunits and oocyte development in the gonad of female H. nemurus. Immature female H. nemurus were divided into four different treatments of ten fish/group, and each group received a single injection of: Group 1= 1X PBS (sham control), Group 2= 0.5 ml/kg Ovaprim (positive control), Group $3 = rLH 50 \mu g/kg$, and Group $4 = rLH 150 \mu g/kg$, respectively. Study parameters including the oocyte development through observation on gonad histology, the E2 plasma levels using enzyme-linked immunosorbent assay (ELISA) and the mRNA transcript level of α , FSHB and LHB subunits using real-time PCR. Based on the histological observation of the gonad, fish received treatment with rLH 50 µg/kg, rLH 150 µg/kg, as well as Ovaprim were able to induce the maturation of oocyte after 48 hr p.i. The E2 plasma level in fish treated with rLH 50 µg/kg raised after 6 hr p.i., significantly increased (p<0.05) after 12 hr p.i., and the level was sustained up to 24 hr p.i. The mRNA expression levels of α , FSH β and LH β also showed significant increment (p<0.05) in fish treated with rLH 50 μ g/kg after 48 hr p.i. Fish treated with Ovaprim showed significant increased (p<0.05) of E2 plasma level after 6 hr p.i., but the level was reduced after 12 hr p.i. There was no significant increased (p>0.05) of mRNA subunits in fish treated with Ovaprim after 48 hr p.i., as compared to sham control fish. Meanwhile, treatment with rLH 150 µg/kg showed a weak stimulatory effect on the E2 production and mRNA transcript level of GtH subunits. The results revealed that treatment with rLH at 50 μ g/kg is the best dosage to induce spawning and oocyte maturation in maturing female H. nemurus. These findings enhanced the understanding of GtH subunit in *H. nemurus*, and the temporal profile helped to estimate the correct time for hormonal therapies to induce ovulation and spawning in female H. nemurus reared in captivity. The production of recombinant GtH from H. nemurus would offer the establishment of a potentially efficient method to induce the gonadal development in this species and possibly in other catfish species.

ii

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENCIRIAN MOLEKUL DAN PENGHASILAN REKOMBINAN SUBUNIT GONADOTROPIN UNTUK PENINGKATAN PRESTASI DALAM SISTEM PEMBIAKAN BETINA *Hemibagrus nemurus* VALENCIENNES

Oleh

ZARIRAH BINTI MOHAMED ZULPERI

Oktober 2016

Pengerusi Fakulti Ina Salwany Md Yasin, PhD : Pertanian

Hemibagrus nemurus, dikenali sebagai ikan baung, adalah spesis ikan keli yang dikultur dan sangat berharga di Malaysia, namun pengeluaran hasil ikan berkenaan terbatas akibat permasalahan dalam sistem pembiakan. Kebanyakan hatcheri menghadapi masalah dalam pembiakan aruhan disebabkan ikan tersebut tidak terdedah dengan pembiakan semulajadi mereka, terutama kegagalan induk betina menjalani tempoh kematangan telur seterusnya menghalang proses ovulasi and pembiakan. Gonadotropin (GtH), yang terdiri daripada hormon perangsang folikel (FSH) dan hormon pengluteinan (LH), memainkan peranan penting dalam gametogenesis dan kematangan seks pada mammalia, termasuk kumpulan ikan. Penyelidikan ini dijalankan untuk memahami ciri-ciri molekul dan memantau perubahan dalam pengekspresan mRNA untuk ketiga-tiga subunit GtH, α , FSH β dan LH_B, dalam kitaran sistem pembiakan betina *H. nemurus*. Kemudian, satu rekombinan rantaian tunggal LH (rLH) H. nemurus telah dihasilkan di dalam sistem E. coli. Keaktifan hormon rekombinan (rLH) akan diuji sama ada ia berpotensi dalam merangsang pengeluaran hormon 17β-estradiol (E2), perkembangan ovari dan pengekspresan mRNA subunit GtH dalam ikan vang dikaji. Melalui analisis molekul menggunakan kaedah PCR menunjukkan cDNA α , FSH β dan LH β dalam *H. nemurus* ialah 656, 728 dan 602 bp, bersamaan 116, 134 dan 140 asid amino. Ketiga-tiga subunit tersebut mempunyai homologi tertinggi dengan kumpulan Siluriformes dan Anguillaformes, manakala paling rendah dengan vertebrata kelas tinggi. Analisis filogenetik mengkelaskan α dan FSH β bersama-sama dengan kumpulan vertebra kelas tinggi, manakala bagi LHβ, terdapat dua kelompok yang berbeza, kumpulan satu hanya mengandungi semua kumpulan ikan, dan kumpulan kedua hanya mengkelaskan vertebra kelas tinggi. Aras transkrip subunit GtH, α , FSH β dan LH β , dalam pituitari betina *H. nemurus* dipantau sepanjang kitaran pembiakan menggunakan PCR masa nyata. Data dikumpul antara Ogos 2013 dan Oktober 2015, daripada usia ikan 1 hingga 27 bulan. Analisis menunjukkan aras mRNA subunit GtH berkadaran

terus dengan perkembangan ovari di sepanjang tempoh pembiakan. Aras subunit FSHβ mRNA meningkat semasa perkembangan ovari dan mencapai tahap optimum di pertengahan vitellogenesis, dan berkurangan semasa ovulasi, manakala aras subunit LH β bertambah dengan signifikan (p<0.05) pada akhir vitellogenesis dan mencapai tahap optimum semasa proses ovulasi. Bagi subunit α , aras mRNA meningkat berkadaran terus dengan subunit FSH β dan LH β , dan mencapai tahap optimum semasa proses ovulasi. Melalui hasil eksperimen, FSH terlibat semasa vitellogenesis, manakala pengekpresan subunit LHB pada akhir kitaran pembiakan menunjukkan hormon tersebut terlibat dalam ovulasi dan kematangan seks. Seterusnya, satu rantaian rekombinan LH (rLH) daripada H. nemurus yang mengandungi susunan struktur LH $\beta\alpha$, dihasilkan melalui proses gen sintesis. Struktur protein rLH telah berjaya diklon ke dalam vektor, pET-32 Ek/LIC dan diekspres di dalam sel E. coli BL21 (DE3). Melalui SDS PAGE, satu jalur yang bersaiz 45.4 kDa telah dikenalpasti, bersamaan dengan anggaran saiz gabungan protein rLH tersebut. Dua antibodi monoklon, His. Tag dan S. Tag digunakan untuk pengesahan saiz protein rLH melalui Western immunoblot. Untuk mengkaji bioaktiviti hormon rLH, keaktifan hormon rLH tersebut diuji melalui keupayaan rLH untuk menaikkan aras E2 dalam plasma, aras mRNA subunit GtH dan perkembangan ovari dalam sistem pembiakan betina H. nemurus. Ikan betina H. nemurus yang pra-matang dibahagi kepada empat kumpulan rawatan berbeza dengan sepuluh ikan/kumpulan, dan setiap kumpulan menerima satu suntikan rawatan: kumpulan 1= 1X PBS (kawalan sham), kumpulan 2= 0.5 ml/kg Ovaprim (kawalan positif), kumpulan 3= rLH 50 µg/kg, dan kumpulan 4= rLH 150 µg/kg. Parameter kajian termasuk perkembangan telur melalui pemerhatian histologi gonad, aras E2 dalam plasma melalui esei immunoserap terungkai enzim (ELISA) dan aras mRNA transkrip subunit α , FSH β dan LH β melalui PCR masa nyata. Berdasarkan pemerhatian histologi gonad, kumpulan ikan yang menerima suntikan rLH 50 µg/kg, rLH 150 µg/kg, serta Ovaprim, telah menunjukkan perkembangan dan kematangan dalam telur selepas 48 jam rawatan. Aras E2 dalam plasma bagi kumpulan ikan dalam rawatan rLH 50 µg/kg telah menunjukkan kenaikan selepas 6 jam rawatan, dan bertambah dengan signifikan (p<0.05) selepas 12 jam rawatan, dan berterusan sehingga 24 jam rawatan. Analisis subunit GtH juga menunjukkan kumpulan ikan yang menerima rawatan rLH 50 µg/kg telah menaikkan (p<0.05) aras mRNA α . FSH β dan LH β selepas 48 jam rawatan. Kumpulan ikan yang menerima rawatan Ovaprim menunjukkan kenaikan (p<0.05) aras E2 dalam plasma selepas 6 jam rawatan, namun, aras tersebut telah berkurangan selepas 12 jam rawatan. Analisis subunit mRNA GtH bagi kumpulan ikan yang menerima rawatan Ovaprim pula tidak menunjukkan perubahan yang signifikan (p>0.05) selepas 48 jam rawatan, apabila dibandingkan dengan kumpulan kawalan sham. Manakala kumpulan ikan yang menerima rawatan rLH 150 µg/kg tidak menunjukkan kenaikan yang signifikan dalam aras E2 plasma dan mRNA subunit GtH. Keputusan eksperimen menunjukkan rawatan rLH pada 50 µg/kg adalah dos yang terbaik untuk merangsang pembiakan dan kematangan telur pada ikan betina *H. nemurus* yang sedang matang. Hasil kajian ini akan membantu bagi meningkatkan tahap pemahaman GtH hormon dalam spesis H. nemurus, manakala profil bagi subunit GtH di sepanjang kitaran pembiakan

akan membantu dalam menganggar ketepatan masa untuk menjalankan rawatan terapi hormon bagi merangsang ovulasi dan pembiakan dalam ikan betina spesis tersebut. Penghasilan hormon rekombinan LH daripada *H. nemurus* berpotensi untuk menyediakan kaedah yang lebih cekap untuk merangsang pembiakan dalam spesis ini dan berkemungkinan pada spesis ikan keli yang lain.



ACKNOWLEDGEMENTS

"Verily with hardship, comes ease."

[Al-Insyirah, 94:5]

Alhamdulillah, first and foremost I am very thankful to Allah Subhanahu Wata'ala for His blessing and mercy, giving me strength and patience to complete this PhD, a colorful journey of life.

I would like to express my sincere gratitude to my parent, Mr. Mohamed Zulperi Zakaria and Mrs. Samirah Ismail, and my sister, Dzarifah Mohamed Zulperi, for their faith and full supports for this amazing journey. Their encouragement, love and concern are the flashlight that guides me to achieve my dreams.

A heartiest thank to my beautiful advisor, Dr. Ina Salwany Md. Yasin, for giving me the opportunity to work with this interesting project. I truly appreciated her guidance, advices, understanding and patience throughout my graduate years, especially during my lab work and thesis writing. I also would like to thank my committee members, Dr. Annie Christianus, Professor Dr. Fatimah Md Yusoff, and Professor Dr. Sharr Azni Harmin for their valuable advices and suggestions in my research.

I would like to extend my appreciation to all lecturers and staff of the Department of Aquaculture, Faculty of Agriculture UPM for their kind assistance during my study, especially to Mrs. Nur Shafika Maulad Abd. Jalil, Miss Norhafizah Roslan and Mr Muhd. Riduan Mohd. Husin. To my wonderful past and present lab mates and colleagues, Saleema Matusin, Nehlah Rosli, Hishammuddin Hamdan, Diyana Nadhirah, Fathin Amirah and Ida Muryany, thank you all for the teamwork, commitment and support. You guys rock! Not to forget, Cikgu Wan Raub and family, Mr. Hassan Ismail Harris and to all suppliers who always provide their best service to assist me during my laboratory work.

Finally, thank you to my beautiful besties, Murni Marlina and Natrah for your cheers and friendship. And to all my families and friends, thank you for all the kind thoughts and du'a and supports for me to face through all the hurdles and this challenging experiences, the Pile, Higher and Deeper.

Thank you very much!

I certify that a Thesis Examination Committee has met on 6 October 2016 to conduct the final examination of Zarirah binti Mohamed Zulperi on her thesis entitled "Molecular Characterization and Recombinant Gonadotropin Subunit Development for Improving Reproductive Performance in Female *Hemibagrus nemurus* Valenciennes" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

S. M. Nurul Amin, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Abu Hena Mustafa Kamal, PhD

Senior Lecturer Faculty of Agriculture and Food Sciences Universiti Putra Malaysia (Bintulu Campus) (Internal Examiner)

Md Sabri bin Mohd Yusoff, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Adelino V.M Canario, PhD

Professor University of Algarve Portugal (External Examiner)

NOR AINI AB. SHUKOR, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 27 December 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Ina Salwany Md Yasin, PhD Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Annie Christianus, PhD Senior Lecturer Faculty of Agriculture

Faculty of Agriculture Universiti Putra Malaysia (Member)

Fatimah Md Yusoff, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Sharr Azni Harmin, PhD

Professor Faculty of Sciences and Biotechnology Universiti Selangor (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fullyowned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:		Date:	
•			

Name and Matric No: Zarirah binti Mohamed Zulperi (GS34713)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Dr. Ina Salwany Md. Yasin
Signature: Name of Member of Supervisory Committee:	Dr. Annie Christianus
Signature: Name of Member of Supervisory Committee:	Prof. Dr. Fatimah Md Yusoff
Signature: Name of Member of Supervisory Committee:	Prof. Dr. Sharr Azni Harmin

TABLE OF CONTENTS

ABS <i>ABS</i> ACK APP DEC LIST LIST	STRACT STRAK NOWLE PROVAL CLARAT OF TA OF TA OF FIG	EDGEMENTS ION BLES GURES BREVIATIONS	Page i iii vi vii ix xvi xvii xvii xxiv
CHA 1	APTER INTR		1
2	LITE		
	2.1	Aquaculture Industry and Scenario in Malaysia	4
	2.2	Production of Catfishes in Malaysia and Worldwide	7
	2.3	Malaysian Rive <mark>r Catfis</mark> h <i>, Hemibagrus nemuru</i> s	7
		2.3.1 Habitat, Distribution, Taxanomic and	8
		Morphology	
		2.3.2 Reproductive Cycle and Spawning Period in	า 9
		Female H. nemurus	
		2.3.3 Problem Associated with Production of <i>H.</i>	10
		nemurus in Malaysia	
	2.4	Reproductive Development in Female Fish	11
		2.4.1 Endocrine Regulation in Fish for Reproduct	ion 11
		2.4.2 Brain-Pituitary-Gonadal Axis: Key of	13
		Reproduction	
	2.5	Gonadotropins: Properties of Follicle Stimulating	14
		Hormone (FSH) and Luteinizing Hormone (LH)	
		2.5.1 Phylogenetic Lineage of Fish Gonadotropin	s 15
		2.5.2 Regulation of FSH and LH During Gonadal	16
		Development in Female Fish	
		2.5.3 Gene Expression of Gonadotropins	17
	2.6	Real-Time PCR: Principles and Advantages	18
		2.6.1 Threshold Cycle and Melting Curve Analysi	s of 20
		SYBR Green Real-Time PCR	
		2.6.2 Gene Expression of Gonadotropin Subunits	; 21
		Using Quantitative Real-Time PCR	
	2.7	Production of Recombinant Protein in Prokaryotic	21
		Escherichia coli Expression System	
		2.7.1 Definition of Cloning and Expression	22
		2.7.2 Plasmid as the Expression Vector	23
		2.7.3 Approaches for Production of Active Eukary	otic 24
		Proteins in <i>E. coli</i> Expression System	
		2.7.4 Production of Recombinant Gonadotropins	in 25
		E. coli and Various Expression System	

3	MOL			IE
	SOB	UNITS (a	L_{i} , FSHβ AND LHβ) FROM Hemibagrus nemurus	~~
	3.1	Introdu		28
	3.2	Materia	Is and Methods	~~
		3.2.1	Collection of Pitultaries	29
		3.2.2	Total RNA Extraction and Measurement of	29
		0.0.0	RNA Concentration	20
		3.2.3	Determination of RNA Concentration and Purity	30
		3.2.4	Reverse Transcription-Polymerase Chain	30
		2 2 5	Reaction (RT-PCR) of RNA to CDNA	20
		3.2.5	Amplification of α , FSH β and LH β Subunits of	30
			H. nemurus	04
			3.2.5.1 Partial Amplification of α , FSH β and	31
			LH β Subunits of <i>H. nemurus</i>	
			3.2.5.2 Rapid Amplification of cDNA Ends	33
			(RACE) PCR of α , FSH β and LH β	
			Subunits of <i>H. nemurus</i>	
			3.2.5.3 Full-Length Amplification of α , FSH β	34
			and LHβ Subunits of <i>H. nemurus</i>	
		3.2.6	Detection of PCR Product	34
		3.2.7	Gel Extraction and Purification of PCR Product	34
		3.2.8	TOPO® TA Cloning® Reaction	35
		3.2 <mark>.9</mark>	One Shot® Chemical Transformation	35
		3.2 <mark>.10</mark>	Screening of Positive PCR Clones	36
		3. <mark>2.11</mark>	Plasmid Extraction	36
		3. <mark>2.12</mark>	Analysis of Positive Recombinants by	36
			Restriction Endonuclease	
		3.2.13	Sequencing of Recombinants	37
		3.2.14	Sequence Assembly and Analysis	37
		3.2.15	Phylogenetic Analysis	38
	3.3	Results		~~
		3.3.1	Partial Amplification and RACE PCR	38
			Amplification of α , FSH β and LH β subunit	
			Genes for <i>H. nemurus</i>	~~
		3.3.2	Analysis of Plasmid of Full-Length α , FSH β and	38
			LH β Subunit Genes for <i>H. nemurus</i>	
		3.3.3	Sequence Analysis for α , FSH β and LH β gene	44
		0.0.4	subunits of <i>H. nemurus</i>	40
		3.3.4	Protein Sequence Comparison of GtH Subunits	48
			Ior r. nemurus and Other Vertebrates and	
		225	FISHES Device another Analysis of FOLIO and LLO of	E A
		3.3.3	Phylogenetic Analysis of α , FSH β and LH β of μ	54
	Э 4	Diaguas		E0
	J.4	DISCUSS	JUII	QC

4	OVAI PITU CYCI <i>nem</i> u	RIAN DE ITARY G LE OF Fl urus	EVELOPN SONADO EMALE N	IENT AND CHANGES IN THE EXPRESSION TROPIN SUBUNITS DURING REPRODUC IALAYSIAN RIVER CATFISH, Hemibagru	ON OF TIVE s
	4.1	Introdu	ction		61
	4.2	Materia 4.2.1 4.2.2 4.2.3	Ils and M Fish and Gonad I RNA Ex	ethods d Sample Sampling Histology traction, RNA Concentration and	63 63 64
			Reverse	e-Transcription of RNA to cDNA	
		4.2.4	Quantita 4.2.4.1	ative Real-Time PCR Primers	64 64
			4.2.4.2	Amplification of Conserved Primers, α , FSH β and LH β , Using Conventional PCR	65
			4.2.4.3	Serial Dilution for Standard Reference of real-time PCR	65
			4.2.4.4	Quantitative Real-Time PCR of α , FSHB and LHB Subunit Genes	66
	4.3	4.2.5 Results	Statistic	al Analysis	67
		4.3.1	Ovarian Gonado	Morphology, Oocyte Diameter and somatic Index	67
		4.3.2	Gonad I	Histology and Gonadosomatic Index	69
		4.3.3 4.3.4	Detectio	ation of real-time PCR amplicon on Limit of α , FSH β , LH β and 18S rRNA	71 73
		4.3 <mark>.5</mark>	Melting	Curve Analysis of α, FSHβ, LHβ and JA Genes	73
		4.3.6	Change mRNA	s in α , FSHβ and LHβ Subunit Genes Expression of female <i>H. nemurus</i>	75
	4.4	Discus	sion		77
5	DEVE	ELOPME		SINGLE-CHAIN RECOMBINANT Hemiba	grus
	5.1 5.2	Introdu Materia	ction ols and M	ethods	80
	0.1	5.2.1	Develop LH (rLH	oment of a Single-Chain Recombinant	81
			5.2.1.1	Construction of a Single-Chain rLH Derived by Gene-Synthesis	81
			5.2.1.2	Transformation of Single Chain rLH, Plasmid Extraction and Sequencing of Positive Recombinants	82
		5.2.2	Product 5.2.2.1 5.2.2.2	ion of a Single-Chain Recombinant LH Primers PCR Amplification of rLH construct	82 82 83
			5.2.2.3	Using Ek/LIC Primers Detection of PCR Product and Gel	83

				Extraction and Purification of PCR		
			E 0 0 4	Products	0.4	
			5.2.2.4	Target Insert	84	
			5.2.2.5	Ligation of the pET32 Ek/LIC vector and T4 Treated Insert	84	
			5.2.2.6	Transformation of Cloned pET-32/LIC- rLH into <i>E. coli</i> TOP 10	85	
			5.2.2.7	Colony Screening, Plasmid Extraction and Analysis of Positive Recombinant by Restriction Endopuclease	85	
			5.2.2.8	Sequencing of Positive Recombinant, rLH	88	
		5.2.3	Express	sion of Recombinant LH Into Expression	88	
			Host			
			5.2.3.1	Transformation of rLH into <i>E. coli</i> Expression Host	88	
			5.2.3.2	Pilot Expression	88	
			5.2.3.3	Protein Extraction	89	
			5.2.3.4	Recombinant Protein Analysis by SDS	89	
			5.2.3.5	Analysis of the Expressed Protein by	90	
				Western Immonoblotting		
	5.3	Results			•	
			5.3.1	Constru LH (rLH	ction of a Single-Chain Recombinant	91
			5.3.1.1	Designed of recombinant LH construct LHβα	91	
			5.3.1.2	Recombinant LH Constructs Derived	91	
		532	Product	ion of a Single-Chain Recombinant I H	96	
		0.0.2	5.3.2.1	Amplification of the rLH Construct	96	
			5.3.2.2	Colony Screening of Positive	96	
			5.3.2.3	Analysis of Positive Recombinant by	96	
			5.3.2.4	Sequencing of Positive Recombinant,	96	
		5.3.3	Express Host	sion of Recombinant LH Into Expression	100	
			5.3.3.1	Transformation of rLH into Expression Host	100	
			5.3.3.2	Sequencing of Positive Recombinant, pET-32/LIC-rLH	100	
			5.3.3.3	Expression of pET-32/LIC-rLH	101	
	5.4	Discuss	sion		105	

6 EFFECT OF RECO OF 17β-ESTRADIO			RECOMBINANT LH HORMONE ON THE PLASMA ADIOL, PITUITARIES AND GONAD OF FEMALE	SMA LEVEL	
	Hem	ibagrus	nemurus	400	
	6.1	Introdu	ction	108	
	0.2	6.2.1	Preparation of recombinant LH (rLH) Protein	109	
		6.2.2	Semi-Purified and Protein Concentration of rLH	110	
		6.2.3	Confirmation of rLH protein by Western Blot	110	
		6.2.4 6.2.5 6.2.6	Protein Quantification Using Bradford Assay Fish for Trial Study Experimental Procedure	111 111 111	
		6.2.7 6.2.8	Gonad Histology Estradiol (EIA) ELISA Analysis 6.2.8.1 Preparation of EIA buffer and Wash	115 115 115	
			Buffer, Estradiol AChE Tracer and Estradiol EIA Antiserum		
			6.2.8.2 Sample Preparation6.2.8.3 Estradiol EIA Standard6.2.8.4 Plate Set Up and Performing the	116 116 117	
		6.2 <mark>.9</mark>	RNA Extraction and Reverse-Transcription to	117	
	0.0	6. <mark>2.10</mark> 6.2.11	Quantitative Real-Time PCR Statistical Analysis	118 118	
	0.3	6.3.1	Confirmation of rLH by SDS-PAGE and Western Immunoblotting	118	
		6.3.2 6.3.3	Concentration of rLH Protein Clinical Observation	120 121	
		6.3.4	Analysis of Gonad	121	
		6.3.5	Effect of rLH on 17β -Estradiol (E2) Plasma Level	127	
		6.3.6	Effect of rLH on mRNA Expression of α , FSH β and LH β Subunit Genes	128	
	6.4	Discuss	sion	130	
7	SUM REC	MARY, G OMMENI	SENERAL CONCLUSION AND DATION FOR FUTURE RESEARCH	133	
REFEI APPEI BIODA LIST C	RENCI NDICE ATA O DF PUI	ES S F STUDE BLICATI	ENT ONS	140 163 172 173	

LIST OF TABLES

Table		Page
3.1	Oligonucleotide primers used for PCR amplification and cloning of α , FSH β and LH β subunits of <i>H. nemurus</i>	32
3.2	Sequence identity of mature peptide of <i>H. nemurus</i> with other fishes and tetrapods. GenBank accession numbers of α , FSH β and LH β used for sequences identities and phylogenetic tree are represented in the final column	52
4.1	Primers used in real-time PCR assays	65
4.2	Fish age, sampling year, and measurement of the total and standard length, body and gonad weight, and gonadosomatic index (GSI) of female <i>H. nemurus</i> obtained from the same batch of fish at the similar sampling point from August 2013 to October 2015	68
4.3	Detection limit of threshold value, Ct and melting temperature, Tm for the SYBR Green I assay for α , FSH β , LH β and 18S rRNA genes for <i>H. nemurus</i>	74
5.1	List of primers of gene specific primers and vector specific primers for PCR amplification and sequencing analysis	83

LIST OF FIGURES

Fig	jure	Page
2.1	National Fishery Sector Production Value (adapted from Department of Fisheries Malaysia, 2014)	6
2.2	An image of 18-month-old <i>H. nemuru</i> s collected from a fish farm, at Cheroh Aquatics Sdn. Bhd., Raub, Pahang Malaysia.	8
2.3	Major reproductive dysfunction detected in captive female fish. They are classified in three main categories (indicated by X) (adapted from Mañanós et al., 2009)	12
2.4	An overview of the brain-pituitary-gonadal (BPG) system in teleost (adapted from Antonopoulou and Borg, 2015)	14
2.5	Steps requirement for: (A) Conventional Reverse- Transcription PCR (RT-PCR), and (B) Real-Time RT- PCR. We can observe the minimal steps involved in real- time RT PCR compared to conventional RT-PCR (adapted from Bustin, 2000)	20
2.6	Anatomy of an expression vector. The figure represents the major features present in expression vectors. All of them are described in the text. The affinity tags and coding sequences for their removal were positioned arbitrarily at the N-terminus for simplicity. MCS, multiple cloning site. Striped patterned box: coding sequence for the desired protein (adapted from Rosano and Ceccarelli, 2014)	24
3.1	Schematic diagram of the positive recombinant analysis using restriction enzyme, <i>EcoR</i> I for recombinant vector, which produced complementary sticky ends. Restriction enzymes were employed to confirm the presence of insert and also correct orientation before sequencing.	37
3.2	Verification of positives insert genes for partial amplification for, (a) α subunit, (b) FSH β subunit, and (c) LH β subunit, inside TOPO® TA vector by restriction enzymes (<i>EcoR</i> I) digestion analysis.	40
3.3	Verification of positives insert genes for 5'RACE amplification for, (a) α subunit, (b) FSH β subunit, and (c) LH β subunit, inside TOPO® TA vector by restriction enzymes (<i>EcoR</i> I) digestion analysis.	41

- 3.4 Verification of positives insert genes for 3'RACE amplification for, (a) α subunit, (b) FSH β subunit, and (c) LHB subunit, inside TOPO® TA vector by restriction enzymes (EcoRI) digestion analysis.
- 3.5 Verification of positives insert genes for full amplification for, (a) α subunit, (b) FSH β subunit, and (c) LH β subunit, inside TOPO® TA vector by restriction enzymes (*EcoRI*) digestion analysis.
- 3.6 Nucleotide and amino acid sequences of α subunit from H. nemurus. The nucleotide number is shown on both sides; the stop codon is represented by an asterisk. The predicted signal peptides at the N-terminus are indicated in grey boxes and putative signals (AATAA) in the 3'unstralated region are indicated by solid and dashed underlinina. respectively. The putative N-linked glycosylation sites and cysteine residues are marked by backgrounds, respectively. and white black This sequence has been deposited in GenBank nucleotide database, under accession no. KF934189.
- 3.7 Nucleotide and amino acid sequences of FSHB from H. nemurus. The nucleotide number is shown on both sides; the stop codon is represented by an asterisk. The predicted signal peptides at the N-terminus are indicated in grev boxes and putative signals (ATTAA) in the 3'unstralated region are indicated by solid and dashed underlining, respectively. The putative N-linked glycosylation sites and cysteine residues are marked by black and white backgrounds, respectively. This sequence has been deposited in GenBank nucleotide database, under accession no. KF998583.
- 3.8 Nucleotide and amino acid sequences of LH β from H. *nemurus*. The nucleotide number is shown on both sides; the stop codon is represented by an asterisk. The predicted signal peptides at the N-terminus are indicated in grey boxes and putative signals (TGTAA) in the 3'unstralated region are indicated by solid and dashed putative underlining. respectively. The N-linked glycosylation sites and cysteine residues are marked by and white backgrounds, black respectively. This sequence has been deposited in GenBank nucleotide database, under accession no. KF934190.
- 3.9 Primary sequence alignment of the putative mature 49 peptide of α subunit for *H. nemurus* with other fishes and

42

45

46

tetrapods, respectively. Sequences identical to *H. nemurus* α are in dots. Gaps (-) were inserted to obtain maximum homology. Amino acid positions are indicated on the top. All conserved cysteine residues of GtHs are boxed and numbered under the reference position. The N-linked glycosylation sites are shaded.

- 3.10 Primary sequence alignment of the putative mature peptide of FSH β for *H. nemurus* with other fishes and tetrapods, respectively. Sequences identical to *H. nemurus* FSH β are in dots. Gaps (-) were inserted to obtain maximum homology. Amino acid positions are indicated on the top. All conserved cysteine residues of GtHs are boxed and numbered under the reference position. The N-linked glycosylation sites are shaded.
- 3.11 Primary sequence alignment of the putative mature peptide of LH β for *H. nemurus* with other fishes and tetrapods, respectively. Sequences identical to *H. nemurus* LH β are in dots. Gaps (-) were inserted to obtain maximum homology. Amino acid positions are indicated on the top. All conserved cysteine residues of GtHs are boxed and numbered under the reference position. The N-linked glycosylation sites are shaded.
- 3.12 Phylogenetic trees for α subunit. Bootstrap analysis was 55 performed using the neighbor-joining method. The bootstrap value proportions are shown at the forks. The NCBI accession no. was shown in Table 3.2.
- 3.13 Phylogenetic trees for FSHβ subunit. Bootstrap analysis 56 was performed using the neighbor-joining method. The bootstrap value proportions are shown at the forks. The NCBI accession no. was shown in Table 3.2.
- 3.14 Phylogenetic trees for LHβ subunit. Bootstrap analysis 57 was performed using the neighbor-joining method. The bootstrap value proportions are shown at the forks. The NCBI accession no. was shown in Table 3.2.
- 4.1 Oocyte diameter (OD) of *H. nemurus* observed between 68
 9- to 27- month-old. Different letters refer to statistically significant differences, p<0.05. All values are means±SEM, with n=30.
 4.2 Gonadosomatic index (GSI) of *H. nemurus* observed from 69
 - 4.2 Gonadosomatic index (GSI) of *H. nemurus* observed from 69 9- to 27- month-old. Different letters refer to statistically significant differences, p<0.05. All values are means±SEM, with n=5.

xix

50

4.3 Cross-section of ovary of *H. nemurus* showing ovarian growth and stages of oocyte development. Oocytes at the primary growth at (a) 9 months and (b) 12 months. Oocytes at the secondary growth at (c) 15 months and (d) 18 months. Oocytes at the maturation growth at (e-f) 21 months. Oocytes at post-spawning at (g) 24 months, and development of oocytes for next reproductive cycle at (h) 27 months. I- chromatin nucleolar oocyte; II- early perinucleolar oocyte; III- late perinucleolar oocyte; IV-cortical alveolar oocyte; V- vitellogenic oocyte; VI- mature oocyte; and VII- germinal vesicle migration oocyte stages.

- 4.4 Confirmation of real-time PCR primers for α , FSH β and 72 LH β of *H. nemurus* using conventional PCR.
- 4.5 Confirmation of real-time PCR primers for 18S rRNA of *H.* 72 *nemurus* using conventional PCR.
- 4.6 76 Changes in the pituitary of female *H. nemurus*, (a) α , (b) FSH β , and (c) LH β subunit genes during reproductive cycle. Abundance of mRNA from each gene was determined by real-time PCR, normalized to 18S rRNA and presented as concentration (ng) started from August 2013 to October 2015. Different letters refer to statistically differences. significant p<0.05. All values are means±SEM, with n=5. The grey bar represents the spawning period and the arrow along the top indicates the period of vitellogenic growth.
- 5.1 Schematic diagrams of recombinant single-chain *H.* 82 *nemurus* luteinizing hormone (rLH). The numbers above the box indicated the amino acid position in each fragment or gonadotropin subunit. Asterisks above each box referred to the putative N-linked glycosylation sites.
- 5.2 Schematic diagram showing the location of the gene and vector specific primers used in colony PCR and universal primers used for sequencing of purified recombinant plasmid in pET-32 Ek/LIC. The positive recombinant plasmid known as pET-32/LIC-rLH. (adapted from the vector map provided by manufacture, Novagen, Germany)
- 5.3 An open reading frame (ORF) DNA and amino acid 93 sequences of recombinant single-chain luteinizing hormone (rLH) from *H. nemurus*. The numbers indicated nucleotide and amino acid position in each fragment of gonadotropin subunits, and asterisks (*) indicated the putative N-linked glycosylation sites.

- 5.4 An open reading frame (ORF) DNA and amino acid 95 sequences of recombinant single-chain luteinizing hormone (rLH) from *H. nemurus*. The original DNA sequences of rLH construct was indicated on the first line, the optimized sequences derived through gene-synthesis indicated on the second line, and the amino acid sequences were indicated on the third line. The red color in DNA bases indicated the replacement of amino acid codon without exchanging the sequences of amino acid.
- 5.5 Agarose gel electrophoresis analysis of PCR amplification 97 of the rLH construct using LIC-LH $\beta\alpha$ F and LIC-LH $\beta\alpha$ R primers combination.
- 5.6 Colony screening of TOP 10 *E. coli* cells for positive 97 clones of pET-32/LIC-rLH insert by PCR using specific primers, LIC-LH $\beta\alpha$ F and LIC-LH $\beta\alpha$ R.
- 5.7 Verification of positives pET-32/LIC-rLH construct by 98 restriction enzymes digestion analysis.
- 5.8 DNA sequences alignment of rLH construct transformed 99 in *E. coli* TOP 10 and *E. coli* BL21 (DE3), compare with original sequences derived from gene-synthesis. Fifteen DNA bases overhang at 5' and 3' indicated the LIC sequences are successfully inserted in the construct.
- 5.9 Colony screening of BL21 (DE3) *E. coli* cells for positive 100 clones of pET-32/LIC-rLH insert by PCR using specific primers, LIC-LH $\beta\alpha$ F and LIC-LH $\beta\alpha$ R.
- 5.10 Schematic diagram showing the vector map of 102 recombinant pET-32/LIC-rLH which was expressed in *E. coli* BL21 (DE3) cells. The expressed fusion protein was detected using His.Tag and S.Tag monoclonal antibody which encoded in pET-21 Ek/LIC expression map. (adapted from the vector map provided by manufacture, Novagen, Germany)
- 5.11 SDS PAGE analysis of the soluble fusion protein of the 103 recombinant rLH after pilot expression in *E. coli* BL21 (DE3) at 8 and 10 hours of induction incubation period at 37°C.
 - 5.12 Western immunoblotting using (a) S.tag monoclonal 104 antibody (Novagen, Germany), and (b) His.Tag

monoclonal antibody (Novagen, Germany), of soluble fusion protein of the recombinant rLH protein at optimized induction, at 8 hr in 37°C in *E. coli* BL21 (DE3).

- 6.1 Experimental design for trial study on female *H. nemurus.* 113
- 6.2 Fish trial using female *H. nemurus*, a) Hormone injection 114 via intramuscular dorsal fin, b) Blood collection from caudal vasculature, c) location of pituitary (by an arrow), d) gonad from fish at 0 hr post injection.
- 6.3 Preparation of the Estradiol standards. (adapted from 116 Estradiol EIA kit manual, Cayman Chemical, Ann Arbor, MI)
- 6.4 SDS PAGE and Western Immunoblotting using His-Tag 119 monoclonal antibody (Novagen, Germany) for confirmation of rLH protein after 8 hr IPTG induction. a) SDS PAGE analysis. 1: Protein Lane Marker (PageRuler[™] Plus Prestained Protein Ladder, Thermo Fisher Scientific, Waltham, MA), Lane 2: unconcentrated rLH protein, Lane 3: concentrated rLH protein. b) Western Immunoblotting using His-Tag monoclonal antibody Germany), Lane (Novagen, 1: Protein Marker (PageRuler[™] Plus Prestained Protein Ladder, Thermo Fisher Scientific, Waltham, MA), Lane 2 and 3: concentrated rLH protein.
- 6.5 Standard curve of Bovine Serum Albumin (BSA) standard 120 assays.
- 6.6 Effect of different treatments after 48 hr p.i., sham control 122 (1X PBS), positive control (0.5 ml/kg Ovaprim), rLH 50 μ g/kg, and rLH 150 μ g/kg, on gonadosomatic index (GSI) in fish. Different letters refer to statistically significant differences, p<0.05. All values are means±SEM, with n=5.

6.7

Gonad histology of *H. nemurus* for sham control 123 treatment, 1X PBS, a) at 0 hr p.i., and b) at 48 hr p.i. Ichromatin nucleolar oocyte; II- early perinucleolar oocyte; III- late perinucleolar oocyte; IV- cortical alveolar oocyte; V- vitellogenic oocyte; VI- mature oocyte; and VIIgerminal vesicle migration oocyte stages. We examined no significant difference on the oocyte developmental stages at 0 and 48 hr p.i. Most oocytes developed at secondary growth phase.

- 6.8 Gonad histology of *H. nemurus* for control positive 124 treatment, 0.5 ml/kg (Syndel Laboratories Ltd., Canada), a) at 0 hr p.i., and b) at 48 hr p.i. I- chromatin nucleolar oocyte; II- early perinucleolar oocyte; III- late perinucleolar oocyte; IV- cortical alveolar oocyte; V-vitellogenic oocyte; VI- mature oocyte; and VII- germinal vesicle migration oocyte stages. We examined at 0 hr p.i. most oocytes developed at secondary growth phase, but after 48 hr p.i., most oocytes enter maturation phase.
- 6.9 Gonad histology of *H. nemurus* for rLH 50 μg/kg, a) at 0 125 hr p.i., and b) at 48 hr p.i. I- chromatin nucleolar oocyte; II- early perinucleolar oocyte; III- late perinucleolar oocyte; IV- cortical alveolar oocyte; V- vitellogenic oocyte; VI- mature oocyte; and VII- germinal vesicle migration oocyte stages. We examined at 0 hr p.i. most oocytes developed at secondary growth phase, but after 48 hr p.i., most oocytes enter maturation phase.
- 6.10 Gonad histology of *H. nemurus* for rLH 150 μg/kg, a) at 0 126 hr p.i., and b) at 48 hr p.i. I- chromatin nucleolar oocyte; II- early perinucleolar oocyte; III- late perinucleolar oocyte; IV- cortical alveolar oocyte; V- vitellogenic oocyte; VI- mature oocyte; and VII- germinal vesicle migration oocyte stages. We examined at 0 hr p.i. most oocytes developed at secondary growth phase, but after 48 hr p.i., most oocytes enter maturation phase.
- 6.11 Plasma 17β-estradiol (E2) levels during treatment with 127 sham control (1X PBS), rLH 50 µg/kg, rLH 150 µg/kg, and positive control (0.5 ml/kg Ovaprim), at different sampling point, 0, 6, 12, 18, 24 and 48 hr p.i. Plama E2 level were measured by Estradiol ELISA (EIA) assay. Different letters refer to statistically significant differences, p<0.05. All values are means±SEM, with n=5.
- 6.12 Effect of rLH injection to mRNA expression level of a) α 129 subunit, b) FSH β subunit, and c) LH β subunit, after 48 hr p.i. mRNA expression level measured using real-time PCR. Different letters refer to statistically significant differences, p<0.05. All values are means±SEM, with n=5.

LIST OF ABBREVIATIONS

%	percent
°C	degree Celsius
11-KT	11-ketotestosterone
17α,20β-DHP	$17\alpha,20\beta$ -dihydroxy-4-pregnen-3-one
20β-S	$17\alpha,20\beta,21\beta$ -trihydroxy-4-pregnen-3-one
A	absorbance
AChe	estradiol-acetylcholinesterase
AFLP	amplified fragment length polymorphism
ANOVA	analysis of variance
BLASTn Blk bp BPG BSA	basic local alignment search tool for nucleotide blank base-pairs brain-pituitary-axis
BSA	bowne serum abumin
BW	body weight
C-terminal	carboxy-terminus
CCD	computer-controlled cooled
cDNA	complementary deoxyribonucleic acid
CG	chorionic gonadotropin
CHO	Chinese hamster ovary
<i>Col</i>	coliconogenic
Ct	cycle threshold
Da	dalton
DAB	3,3'-diaminobenzidine
dATP	deoxyadenosine triphosphate
dH20	distilled water
DNA	deoxyribonucleic acid
dNTP	deoxynucleotide tripnosphate
dph	days post hatching
ds	double stranded
dtn	decitonne
DTT	dithiothreitol
E. coli	<i>Escherichia coli</i>
E2	17β-estradiol
EDTA	ethylene-diamine-tetraacetic acid (disodium salt)
EIA	enzyme immunoassay
Ek	enterokinase
ELISA	enzyme-linked immunosorbent assay
FOM	final oocyte maturation
FRET	fluorescence resonance energy transfer
FSH	follicle-stimulating-hormone
9 GnRH GnRHa GSI	gonadotropin-releasing hormone antagonist gonadotropin-releasing hormone gonadosomatic index

GtH	gonadotropin
GVBD	germinal vesicle breakdown
H. nemurus	Hemibagrus nemurus
hCG	human chorionic gonadotropin
hr	hour/s
HRP	horseradish peroxidase-conjugated
lgG	immunoglobulin G
IPTG	isopropyl β-galactose
kb	kilo base-pairs
kDa	kilo Dalton
l or L	liter
LB	Luria-Bertani
LH	luteinizing hormone
LIC	ligation independent cloning
MBP	maltose binding protein
MEGA	Molecular Evolutionary Genetics Analysis
mg	miligram
MgCl ₂	magnesium chloride
min	minute/s
MIS	maturation inducing steroid
ml	mililiter
ml	mililiter
mM	milimolar
mRNA	messenger ribonucleic acid
N-terminus	amine-terminus
NCBI	National Center for Biotechnology Information
NCS	N-linked glycosylation sequences
na	nanogram
nm	nanometer
NSB	non-specific binding
NTC	no template control
NusA	N-utilizing substance-A
	oocyte diameter
ORF	open reading frame
ni	post injection
PBS	phosphate buffered saline
PBST	phosphate buffered saline with Tween 20
PCR	polymerase chain reaction
nET-32/LIC-rLH	Recombinant plasmid (nET-321 IC+rl H gene)
nH	puissance hydrogen (hydrogen-ion concentration)
R	resistance
rom	rotations per minute
RACE	rapid amplification of cDNA ends
RAPD	random amplified polymorphism DNA
RIA	radioimmunoassav
rl H	recombinant I H
RM	Ringgit Malaysia
RNA	ribonucleic acid
rRNA	ribosomal ribonucleic acid

RT RT-PCR SDS	reverse-transcription reverse-transcription polymerase chain reaction sodium dodecyl sulfate
SDS PAGE	sodium dodecyl sulfate polyacrylamide gel
sec	second/s
SEM	standard error mean
SUC	super optimal broth
T	testosterone
ТА	total activity
TBE	tris-boric EDTA
TBS	tris buffer saline
Tm	melting curve
Tris	tris (hydroxymethyl) aminomethane
trx	thioredoxin
TSH	thyroid-stimulating hormone
U	
	United States Dollar
UTR	untranslated region
UV	ultra-violet
V	voltan/volt
v/v	volume per volume
Vtg	vitellogenin
w/v	weight per volume
X-gal	5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside
μg	microgram
μl	microliter
μM	micromolar
μm	micrometer
μμη	micrometer

Amino Acid	Single	Three Letter
	Letter	Symbol
Alanine	А	Ala
Asparagine or aspartic acid	В	Asx
Cysteine	С	Cys
Aspartic acid	D	Asp
Glutamic acid	E	Glu
Phenylalanine	F	Phe
Glycine	G	Gly
Histidine	Н	His
Isoleucine	I	lle
Lysine	K	Lys
Leucine	L	Leu
Methionine	M	Met
Asparagine	N	Asn
Proline	Р	Pro
Glutamine	Q	Gln
Arginine	R	Arg
Serine	S	Ser
Threonine	T	Thr
Valine	V	Val
Tryptophan	W	Trp
Tyrosine	Y	Tyr
Glutamine or glutamic acid	Z	Glx

C

CHAPTER 1

INTRODUCTION

Aquaculture is growing and expanding in most of the regions in the world, as human population consumes the aquatic food for their daily nutrition, which provides a highly digestible proteins, essential vitamins and minerals in a simple accessible form. According to Food and Agriculture Organization of the United Nations (FAO, 2014), to sustain the current level per capita consumption, global aquaculture will need to achieve 80 million tons production by 2050. Latest report of fisheries sector in Malaysia generated RM 12,765.3 million in 2014, showing an increment of 11.32% compared to 2013 (Department of Fisheries Malaysia, 2014). Aquaculture production, which comprised of freshwater and brackish water production had contributed about 25.6% to the national economy, an increase of 35.02% compared to 2013.

The Hemibagrus nemurus (H. nemurus) or known as "ikan baung" is a popular catfish species preferred by local people in Malaysia, due to its good taste and nutritional value. Its non-bony flesh comprises of low cholesterol, high protein and rich in omega 3-polyunsaturated fatty acid (Mesomya et al., 2002). Hemibagrus nemurus is an indigenous species, and distributed in most of the Southeast Asian countries including Malaysia, Vietnam, Indonesia and Thailand (Rainboth, 1996). As one of the important catfish species in Malaysia, this fish has been commercially cultured intensively in floating cages or semi-intensively in ponds and pens (Kamarudin et al., 2011; Abidin et al., 2006; Khan et al, 1990). The annual national production of H. *nemurus* has gradually increased over the years, although it remains less than 6% of the total freshwater aquaculture production (Department of Fisheries Malaysia, 2014). The insufficient seed supply coupled with low quality of brood stocks have limited its production. The species is difficult to spawn artificially, due to problems associated with its induced spawning (Muchlisin et al., 2004). The maturity between male and female brood stock is difficult to be synchronized (Muchlisin et al., 2004). In addition, the female brood stock usually experiences reproductive failure of the pituitary to release gonadotropins, which is important in gonadal maturation in fish (Zohar and Mylonas, 2001).

The brain-pituitary-gonadal (BPG) axis plays a crucial role to control sexual development and reproductive system in most vertebrates including fish. This process starts when the gonadotropin-releasing hormone (GnRH) produced in brain, governs the release of the gonadotropins (GtHs) hormones in anterior pituitary into the blood circulation. Via the blood, those hormones reach into gonad and bind to their receptors, and stimulate the production of sex steroid, which are required for spermatogenesis and oogenesis (Amano, 2010; Weltzien et al., 2004; Holland et al., 2001). The

gonadotropins (GtHs), follicle-stimulating hormone (FSH) and luteinizing hormone (LH), are the main hormones in the regulation of gametogenesis and sexual maturation in fish. Both hormones are heterodimeric proteins comprise of a common α subunit and bound non-covalently to a specific β subunit hormone (Swanson et al., 2003; Pierce and Parson, 1981). The duality of gonadotropic hormones in teleost fish has now been confirmed in various species mostly salmonid and few non-salmonid fish species. A distinct pattern on the mRNA expression of α . FSHB and LHB subunits was detected in single spawning fish. The FSH is involved in the initiation of gametogenesis, vitellogenesis and spermatogenesis, while LH is involved in the final maturation process; ovulation, oocyte maturation, milt production and spermiation (Levavi-Sivan et al., 2010; Kobayashi et al., 2006; Yaron et al., 2003). In multiple spawning fish, the expression of FSH β and LH β mRNA levels are high during sexual behavior and oocyte maturation in females (Yoshiura et al., 1997; Jackson et al., 1999), but this is not a common pattern observed in fish such as red seabream (Gen et al., 2003).

In captive brood stock, the females usually exhibit severe reproductive problems than the males. Most fish reared in captivity showed reproductive dysfunction due to the failure of the pituitary to produce maturational gonadotropin, LH hormone (Zohar and Mylonas, 2001, Peter et al., 1993). Thus, the production of recombinant GtHs using various expression systems are being establish to improve method of inducing gonadal development in fish (Kobayashi et al., 2006). To date, a few fish recombinant GtH have been produced in several type of bioreactors, for example baculovirus in silkworm larvae (Ko et al., 2007; Kobayashi et al., 2006), yeast (Chen et al., 2012; Kasuto and Levavi-Sivan, 2005) and bacterial cell (Kim et al., 2012).

Since *H. nemurus* is one of the important cultured catfish in Malaysia, few efforts have been made to improve its breeding and spawning performance. Thalathiah et al. (1992) reported the successful artificial spawning of the species through hormonal induction using heteroplastic pituitary extract in combination with human chorionic gonadotropin (hCG). Latest result by Adebiyi et al. (2013b) revealed a treatment with 50 μ g/kg BW (body weight) of synthetic agonists of GnRH (GnRHa) could increase steroid production in the plasma of *H. nemurus*, thus showing that GnRHa can be used as an inducing agent for reproduction control of this species. Other studies have established the reproduction, nutrition, genetics and morphology of *H. nemurus*, but up to this point, none of them focus on manipulating FSH and LH hormones.

The commercial culture of *H. nemurus* in Malaysia faced with problems associated in induced spawning that lead to its low production to fulfill the market demand. The current spawning method using the commercial synthetic reproductive hormones such as Ovaprim and Ovatide are successfully to induce spawning and ovulation in the species. However,

several disadvantages of using the hormones were the short response of the hormones tested in *in vivo* study (Adebiyi et al., 2013d), the hormones especially Ovaprim was too concentrated that is not practical for farmers, and sometimes two injections were needed to induce spawning and ovulation in the fish (Zohar and Mylonas, 2001) which is time-consuming. Therefore, this study aimed to perform a thorough study of GtH subunits, α , FSH β and LH β , in female *H. nemurus*, understand its reproductive cycle and produce a recombinant LH (rLH) in bacterial *E. coli* cells for an effective approach of hormonal manipulation and gonadal development in this species. The long-term goal is to produce hormonal treatment with a simple method and cost effective that can be used by the industry to improve the reproduction of cultured catfishes for sustainable aquaculture production. This study covered the following objectives:

- 1) to determine the molecular characteristic of GtH subunits, α , FSH β and LH β , from *H. nemurus*
- 2) to assess the expression levels of α , FSH β and LH β subunits of female *H. nemurus* at different stages of reproductive cycle
- 3) to develop and produce the recombinant LH (rLH) from *H. nemurus*
- 4) to determine the effect of exogenous recombinant LH on the plasma level of 17β -estradiol (E2) in female *H. nemurus*

Hypothesis:

- H₀: Treatment with rLH at a specific dose in maturing female *H. nemurus* could not induce oocyte maturation, stimulating the production of sex steroid and up-regulate the mRNA transcript of GtH subunit genes including α , FSH β and LH β , resulting ovulation and spawning in the fish.
- H_A: Treatment with rLH at a specific dose in maturing female *H. nemurus* could induce oocyte maturation, stimulating the production of sex steroid and up-regulate the mRNA transcript of GtH subunit genes including α , FSH β and LH β , resulting ovulation and spawning in the fish.

REFERENCES

- Abdi, H., Christianus, A., Ramezani-Fard, E., Saad, C.R. and Hosseini, S.A. (2011). Proximate and fatty acid composition of the liver of cultured Asian redtail catfish (*Hemibagrus nemurus*) and African catfish (*Clarias gariepinus*). *Journal of Fisheries and Aquatic Science*. 6(7): 840-845.
- Abidin, M.Z., Hashim, R. and Chong Shu Chien, A. (2006). Influence of dietary protein levels on growth and egg quality in broodstock female bagrid catfish (*Mystus nemurus* Cuvier and Valenciennes). *Aquaculture Research*. 37(4): 416-418.
- Adebiyi, F.A., Siraj, S.S., Harmin, S.A. and Christianus, A. (2011). Ovarian development of a river catfish *Hemibagrus nemurus* (Valenciennes, 1840) in captivity. *Journal of Experimental Zoology Part A: Ecological Genetics and Physiology*. 315(9): 536-543.
- Adebiyi, F.A., Siraj, S.S., Harmin, S.A. and Christianus, A. (2013a). Plasma sex steroid hormonal profile and gonad histology during the annual reproductive cycle of river catfish *Hemibagrus nemurus* (Valenciennes, 1840) in captivity. *Fish Physiology and Biochemistry*. 39(3): 547-557.
- Adebiyi, F.A., Siraj, S.S., Harmin, S.A. and Christianus, A. (2013b). Induced spawning of a river catfish *Hemibagrus nemurus* (Valenciennes, 1840). *Pertanika Journal of Tropical Agricultural Science*. 36(1): 71-78.
- Adebiyi, F.A., Siraj, S.S., Harmin, S.A. and Christianus, A. (2013c). Embryonic and larval development of river catfish Hemibagrus nemurus (Valenciennes, 1840). Asian Journal of Animal and Veterinary Advances. 8(2): 237-246.
- Adebiyi, F.A., Siraj, S.S., Harmin, S.A. and Christianus, A. (2013d). Effects of GnRHa on plasma sex steroid hormones of river catfish *Hemibagrus nemurus* (Valenciennes 1840). *Sains Malaysiana*. 42(5): 635-642.
- Aizam Z.A. (1986) The reproductive biology of a tropical cyprinid, *Hampala macrolepidota* (Van Hasselt) from Zoo Negara. Kuala Lumpur, Malaysia. *Journal of Fish Biology.* 29(3): 381-391.
- Aizen, J., Kowalsman, N., Kobayashi, M., Hollander, L., Sohn, Y. C., Yoshizaki, G., Masha, Y.N. and Levavi-Sivan, B. (2012). Experimental and computational study of inter-and intra-species specificity of gonadotropins for various gonadotropin receptors. *Molecular and Cellular Endocrinology*. 364(1): 89-100.
- Alexander, K.A., Potts, T.P., Freeman, S., Israel, D., Johansen, J., Kletou, D., Meland, M., Pecorino, D., Rebours, C., Shorten, M. and Angel, D.L. (2015). The implications of aquaculture policy and regulation for the

development of integrated multi-trophic aquaculture in Europe. *Aquaculture*. 443: 16-23.

- Almendras, J.M., Duenas, C., Nacario, J., Sherwood, N.M. and Crim, L.W. (1988). Sustained hormone release. III. Use of gonadotropin releasing hormone analogues to induce multiple spawnings in sea bass, *Lates calcarifer. Aquaculture*. 74(1): 97-111.
- Amano, M. (2010). Reproductive biology of salmoniform and pleuronectiform fishes with special reference to gonadotropin-releasing hormone (GnRH). Aqua-BioScience Monograph. 3(2): 39-72.
- Anon. (2011). *Annual Fisheries Statistics 2008 Volume 1*. Department of Fisheries, Malaysia, Kuala Lumpur.
- Antonopoulou, E. and Borg, B. (2015). The Brain-Pituitary-Gonad Axis in the Atlantic Salmon. In V. Tomislav, P. Erik (Eds). *Evolutionary Biology of the Atlantic Salmon* (pp. 108-120). Boca Raton: CRC Press Taylor and Francis Group.
- Antonopoulou, E., Swanson, P., Mayer, I. and Borg, B. (1999). Feedback control of gonadotropins in Atlantic salmon, *Salmo salar*, male parr: II. Aromatase inhibitor and androgen effects. *General and Comparative Endocrinology*. 114(1): 142-150.
- Arey, B.J., Stevis, P.E., Deecher, D.C., Shen, E.S., Frail, D.E., Negro-Vilar, A. and López, F.J. (1997). Induction of promiscuous G protein coupling of the follicle-stimulating hormone (FSH) receptor: a novel mechanism for transducing pleiotropic actions of FSH isoforms. *Molecular Endocrinology*. 11(5): 517-526.
- Aroua, S., Weltzien, F.A., Le Belle, N. and Dufour, S. (2007). Development of real-time RT-PCR assays for eel gonadotropins and their application to the comparison of *in vivo* and *in vitro* effects of sex steroids. *General* and Comparative Endocrinology. 153(1): 333-343.
- Beitins, I.Z. and Padmanabhan, V. (1991). Bioactivity of gonadotropins. *Endocrinology and Metabolism Clinics of North America*. 20(1): 85-120.
- Berlinsky, D.L., King, V., Smith, T.I., Hamilton, R.D., Holloway, J. and Sullivan, C.V. (1996). Induced ovulation of southern flounder *Paralichthys lethostigma* using gonadotropin releasing hormone analogue implants. *Journal of the World Aquaculture Society*. 27(2): 143-152.
- Bommelear, M.C., Billard, R. and Breton, B. (1981). Changes in plasma gonadotropin after ovariectomy and estradiol supplementation at different stages at the end of the reproductive cycle in the rainbow trout (*Salmo gairdneri* R.). *Reproduction Nutrition Développement*. 21(6A): 989-997.

- Brown, T.A. (1998). Production of protein from cloned genes. In T.A. Brown (Ed). *Gene Cloning: An Introduction* (pp. 255-256). Cheltenham: Stanley Thornes (Publishers) Ltd.
- Burgess-Brown, N.A., Sharma, S., Sobott, F., Loenarz, C., Oppermann, U. and Gileadi, O. (2008). Codon optimization can improve expression of human genes in *Escherichia coli*: A multi-gene study. *Protein Expression and Purification*. 59(1): 94-102.
- Bustin, S.A. (2000). Absolute quantification of mRNA using real-time reverse transcription polymerase chain reaction assays. *Journal of Molecular Endocrinology*. 25(2): 169-193.
- Cabrita, L.D., Dai, W. and Bottomley, S.P. (2006). A family of *E. coli* expression vectors for laboratory scale and high throughput soluble protein production. *BMC Biotechnology*. 1: 6-12.
- Carillo, M., Zanuy, S., Prat, F., Cerda, J., Ramos, J., Mañańos, E. and Bromage, N. (1995). Sea bass (*Dicentrarchus labrax*). In N.R. Bromage, R.J. Roberts (Eds). *Broodstock management and egg and larval quality*. Oxford: Blackwell.
- Caspeta, L., Flores, N., Pérez, N. O., Bolívar, F. and Ramírez, O.T. (2009). The effect of heating rate on *Escherichia coli* metabolism, physiological stress, transcriptional response, and production of temperatureinduced recombinant protein: A scale-down study. *Biotechnology and Bioengineering*. 102(2): 468-482.
- Cek, S., Bromage, N., Randall, C. and Rana, K. (2001). Oogenesis, hepatosomatic and gonadosomatic indexes, and sex ratio in rosy barb (*Puntius conchonius*). *Turkish Journal of Fisheries and Aquatic Sciences*. 1(2): 33-41.
- Cerdà, J., Chauvigne, F., Agulleiro, M.J., Marin, E., Halm, S., Martínez-Rodríguez, G. and Prat, F. (2008). Molecular cloning of Senegalese sole (*Solea senegalensis*) follicle-stimulating hormone and luteinizing hormone subunits and expression pattern during spermatogenesis. *General and Comparative Endocrinology*. 156(3): 470-481.
- Chan, S.C. (2003). Development and isolation of the DNA microsatellite markers for the characterization and identification of Mystus nemurus (Cuvier and Valenciennes). Master dissertation. Universiti Putra Malaysia, Malaysia.
- Chart, H., Smith, H.R., La Ragione, R.M. and Woodward, M.J. (2000). An investigation into the pathogenic properties of Escherichia coli strains BLR, BL21, DH5α and EQ1. *Journal of Applied Microbiology*. 89(6): 1048-1058.

- Chatterjee, A., Shen, S.T. and Yu, J.Y.L. (2005). Molecular cloning of cDNAs and structural model analysis of two gonadotropin β-subunits of snakehead fish (*Channa maculata*). *General and Comparative Endocrinology*. 143(3): 278-286.
- Chaube, R., Joy, K.P. and Acharjee, A. (2015). Catfish Gonadotrophins: Cellular Origin, Structural Properties and Physiology. *Journal of Neuroendocrinology*. 27(6): 536-543.
- Chauvigné, F., Fatsini, E., Duncan, N., Ollé, J., Zanuy, S., Gómez, A. and Cerdà, J. (2016). Plasma levels of follicle-stimulating and luteinizing hormones during the reproductive cycle of wild and cultured Senegalese sole (Solea senegalensis). Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology. 191: 35-43.
- Chen, J., Zhang, Y., Tang, Z., Mao, J., Kuang, Z., Qin, C. and Li, W. (2012). Production of recombinant orange-spotted grouper (*Epinephelus coioides*) follicle-stimulating hormone (FSH) in single-chain form and dimer form by *Pichia pastoris* and their biological activities. *General and Comparative Endocrinology*. 178(2): 237-249.
- Chen, R. (2012). Bacterial expression systems for recombinant protein production: *E. coli* and beyond. *Biotechnology Advances*. 30(5): 1102-1107.
- Choi, E., Ko, H., Shin, J., Kim, M. and Sohn, Y.C. (2005). Expression of gonadotropin genes in Manchurian trout *Brachymystax lenok* and production of recombinant gonadotropins. *Fisheries Science*. 71(6): 1193-1200.
- Chomczynski, P. (1993). A reagent for the single-step simultaneous isolation of RNA, DNA and proteins from cell and tissue samples. *Biotechniques*. 15(3): 532-537.
- Chong, L.K. (1998). Development of PCR-based DNA markers to identify and characterize Malaysian river catfish, Mystus nemurus (Cuvier and Valenciennes): RAPD and AFLP. Master dissertation. Universiti Putra Malaysia, Malaysia.
- Chong, L.K., Tan, S.G., Yusoff, K. and Siraj, S.S. (2000). Identification and characterization of Malaysian river catfish, *Mystus nemurus* (Cuvier and Valenciennes): RAPD and AFLP analysis. *Biochemical Genetics*. 38(3-4): 63-76.
- Claverie, J.M. and Notredame, C. (2003). Building phylogenetics trees. In M. Jean-Claverie and C. Notredame (Eds). *Bioinformatics for dummies* (pp. 381-409). Indianapolis: Wiley Publishing, Inc.

- Coward, K. and Bromage, N.R. (1998). Histological classification of oocyte growth and the dynamics of ovarian recrudescence in *Tilapia zillii*. *Journal of Fish Biology*. 53(2): 285-302.
- Dabrowski, K., Ciereszko, A., Ramseyer, L., Culver, D. and Kestemont, P. (1994). Effects of hormonal treatment on induced spermiation and ovulation in the yellow perch (*Perca flavescens*). *Aquaculture*. 120(1): 171-180.
- de Marco, A., Deuerling, E., Mogk, A., Tomoyasu, T. and Bukau, B. (2007). Chaperone-based procedure to increase yields of soluble recombinant proteins produced in *E. coli. BioMed Central Biotechnology*. 7(1): 32-40.
- de Tassigny, X.D.A., Ackroyd, K.J., Chatzidaki, E.E. and Colledge, W.H. (2010). Kisspeptin signaling is required for peripheral but not central stimulation of gonadotropin-releasing hormone neurons by NMDA. *The Journal of Neuroscience*. 30(25): 8581-8590.
- DeLeeuw, R., Wurth, Y., Zandbergen, M., Peute, J. and Goos, H.T. (1986). The effects of aromatizable androgens, and estrogens on gonadotropin release in castrated African catfish, *Clarias gariepinus* (Burchel): a physiological and ultrastructural study. *Cell and Tissue Research*. 243: 587-594.
- Department of Fisheries Malaysia (2012). Annual Fisheries Statistic 2012 Volume 1. *Ministry of Agriculture and Agro-Based Industry Malaysia, Putrajaya.*
- Department of Fisheries Malaysia (2013). Annual Fisheries Statistic 2013 Volume 1. *Ministry of Agriculture and Agro-Based Industry Malaysia, Putrajaya.*
- Department of Fisheries Malaysia (2014). Annual Fisheries Statistic 2014 Volume 1. *Ministry of Agriculture and Agro-Based Industry Malaysia, Putrajaya.*
- Dickey, J.T. and Swanson, P. (2000). Effects of salmon gonadotropinreleasing hormone on follicle stimulating hormone secretion and subunit gene expression in coho salmon (*Oncorhynchus kisutch*). *General and Comparative Endocrinology*. 118(3): 436-449.
- Dodson, J.J., Colombani, F. and Ng, P.K.L. (1995). Phylogeographic structure in mitochondrial DNA of a South-east Asian freshwater fish, *Hemibagrus nemurus* (Siluroidei; Bagridae) and Pleistocene sea-level changes on the Sunda shelf. *Molecular Biology*. 4(3): 331-346.
- Easterling, W.E. (2007). Climate change and the adequacy of food and timber in the 21st century. *Proceedings of the National Academy of Sciences USA*. 104(50): 19679.

- El Hag, G.A., Kamarudin, M.S., Saad, C.R. and Daud, S.K. (2012). Gut histology of Malaysian river catfish, *Mystus nemurus* (Cuvier and Valenciennes) larvae. *Life Science Journal*. 9(1): 342-347.
- Elizur, A., Zmora, N., Rosenfeld, H., Meiri, I., Hassin, S., Gordin, H. and Zohar, Y. (1996). Gonadotropins β-GtHI and β-GtHII from the gilthead seabream, *Sparus aurata. General and Comparative Endocrinology*. 102(1): 39-46.
- Estay, F., Neira, R., Diaz, N.F., Valladares, L. and Torres, A. (1998). Gametogenesis and sex steroid profiles in cultured coho salmon (*Oncorhynchus kisutch*, Walbaum). *Journal of Experimental Zoology*. 280(6): 429-438.
- Expósito-Rodríguez, M., Borges, A.A., Borges-Pérez, A. and Pérez, J.A. (2008). Selection of internal control genes for quantitative real-time RT-PCR studies during tomato development process. *BMC Plant Biology*. 8(1): 131.
- FAO (Fisheries and Aquaculture Department). (2009). The State of World Fisheries and Aquaculture 2009. *Food and Agricultural Organization of the United Nations, Rome.*
- FAO (Fisheries and Aquaculture Department). (2010). The State of World Fisheries and Aquaculture 2010. Food and Agricultural Organization of the United Nations, Rome.
- FAO (Fisheries and Aquaculture Department). (2014). The State of World Fisheries and Aquaculture 2014. Food and Agricultural Organization of the United Nations, Rome.
- Fares, F. (2006). The role of O-linked and N-linked oligosaccharides on the structure-function of glycoprotein hormones: Development of agonists and antagonists. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 1760(4): 560-567.
- Flack, M.R., Froehlich, J., Bennet, A.P., Anasti, J. and Nisula, B.C. (1994). Site-directed mutagenesis defines the individual roles of the glycosylation sites on follicle-stimulating hormone. *Journal of Biological Chemistry*. 269(19): 14015-14020.
- Fontenele, O. (1955). Injecting pituitary (hypophyseal) hormones into fish to induce spawning. *The Progressive Fish-Culturist*. 17(2): 71-75.
- Fox, K.M., Dias, J.A. and Van Roey, P. (2001). Three-dimensional structure of human follicle-stimulating hormone. *Molecular Endocrinology*. 15(3): 378-389.
- Gan, Y.B., Zhou, Z.J., An, L.J., Bao, S.J. and Forde, B.G. (2011). A comparison between northern blotting and quantitative real-time PCR

as a means of detecting the nutritional regulation of genes expressed in roots of Arabidopsis thaliana. *Agricultural Sciences in China*. 10(3): 335-342.

- Gen, K., Okuzawa, K., Senthilkumaran, B., Tanaka, H., Moriyama, S. and Kagawa, H. (2000). Unique expression of gonadotropin-I and-II subunit genes in male and female red seabream (*Pagrus major*) during sexual maturation. *Biology of Reproduction*. 63(1): 308-319.
- Gen, K., Yamaguchi, S., Okuzawa, K., Kumakura, N., Tanaka, H. and Kagawa, H. (2003). Physiological roles of FSH and LH in red seabream, *Pagrus major. Fish Physiology and Biochemistry*. 28(1-4): 77-80.
- Gibson, U.E., Heid, C.A. and Williams, P.M. (1996). A novel method for real time quantitative RT-PCR. *Genome Research*. 6(10): 995-1001.
- Giglio, S., Monis, P.T. and Saint, C.P. (2003). Demonstration of preferential binding of SYBR Green I to specific DNA fragments in real-time multiplex PCR. *Nucleic Acids Research*. 3(22): e136-e136.
- Golan, M., Biran, J. and Levavi-Sivan, B. (2014). A novel model for development, organization, and function of gonadotropes in fish pituitary. *Frontiers in Endocrinology*. 5(182): 1-11.
- Gomez, J.M., Weil, C., Ollitrault, M., Le Bail, P.Y., Breton, B. and Le Gac, F. (1999). Growth hormone (GH) and gonadotropin subunit gene expression and pituitary and plasma changes during spermatogenesis and oogenesis in rainbow trout (*Oncorhynchus mykiss*). *General and Comparative Endocrinology*. 113(3): 413-428.
- Goos, H.J.T.H. and Schulz, R. (1997). Gonadal steroid hormones drive puberty in fish. In: Advances in Comparative Endocrinology, vol. 2 (pp 1429-1433). Bologna: Monduzzi Editore, International Proceedings Division.
- Gothilf, Y. and Zohar, Y. (1991). Clearance of different forms of GnRH from the circulation of the gilthead seabream, *Sparus aurata*. In A.P. Sumpter, J.P. Kime, M.S. Rolfe (Eds). *Reproductive Physiology in fish* (pp. 35-37). Sheffield: Symposium 91.
- Gothilf, Y., Meiri, I., Elizur, A. and Zohar, Y. (1997). Preovulatory changes in the levels of three gonadotropin-releasing hormone-encoding messenger ribonucleic acids (mRNAs), gonadotropin beta-subunit mRNAs, plasma gonadotropin, and steroids in the female gilthead seabream, *Sparus aurata*. *Biology of Reproduction*. 57(5): 1145-1154.
- Guénin, S., Mauriat, M., Pelloux, J., Van Wuytswinkel, O., Bellini, C. and Gutierrez, L. (2009). Normalization of qRT-PCR data: the necessity of adopting a systematic, experimental conditions-specific, validation of references. *Journal of Experimental Botany*. 60(2): 487-493.

- Guzmán, J.M., Bayarri, M.J., Ramos, J., Zohar, Y., Sarasquete, C. and Mañanós, E.L. (2009). Follicle stimulating hormone (FSH) and luteinizing hormone (LH) gene expression during larval development in Senegalese sole (Solea senegalensis). Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology. 154(1): 37-43.
- Hails, A.J. and Abdullah, Z. (1982). Reproductive biology of the tropical fish *Trichogaster pectoralis* (Regan). *Journal of Fish Biology*. 21(2): 157-170.
- Hakola, K., Van der Boogaart, P., Mulders, J., de Leeuw, R., Schoonen, W., Van Heyst, J., Swolfs, A., Van Casteren, J., Huhtaniemi, I. and Kloosterboer, H. (1997). Recombinant rat follicle-stimulating hormone; production by Chinese hamster ovary cells, purification and functional characterization. *Molecular and Cellular Endocrinology*. 127(1): 59-69.
- Han, S.K., Gottsch, Lee, M.L., Lee, K.J., Popa, S.M., Smith, J.T., Jakawich, S.K., Clifton, D.K., Steiner, R.A. and Herbison, A.E. (2005). Activation of gonadotropin-releasing hormone neurons by kisspeptin as a neuroendocrine switch for the onset of puberty. *The Journal of neuroscience*. 25(49): 11349-11356.
- Han, Y.S., Liao, I.C., Huang, Y.S., Tzeng, W.N. and Yu, J.Y.L. (2003). Profiles of PGH-α, GTH I-β, and GTH II-β mRNA transcript levels at different ovarian stages in the wild female Japanese eel Anguilla japonica. General and Comparative Endocrinology. 133(1): 8-16.
- Hardikar, A.A., Farr, R.J. and Joglekar, M.V. (2014). Circulating microRNAs: Understanding the Limits for Quantitative Measurement by Real- Time PCR. *Journal of the American Heart Association*. 3(1): e000792.
- Hargreaves, J.A. and Tucker, C.S. (2003). Defining loading limits of static ponds for catfish aquaculture. *Aquacultural Engineering*. 28(1): 47-63.
- Harmin, S.A. and Crim, L.W. (1993). Influence of gonadotropic hormonereleasing hormone analog (GnRH-A) on plasma sex steroid profiles and milt production in male winter flounder, *Pseudopleuronectes americanus* (Walbaum). *Fish Physiology and Biochemistry*. 10(5): 399-407.
- Hassin, S., Claire, M., Holland, H. and Zohar, Y. (1999). Ontogeny of folliclestimulating hormone and luteinizing hormone gene expression during pubertal development in the female striped bass, *Morone saxatilis* (Teleostei). *Biology of Reproduction*. 61(6): 1608-1615.
- Hearn, M.T. and Gomme, P.T. (2000). Molecular architecture and biorecognition processes of the cystine knot protein superfamily: part I. The glycoprotein hormones. *Journal of Molecular Recognition*. 13(5): 223-278.

- Helfman, G.S., Collette, B.B. and Facey, D.E. (1997). *The diversity of fishes* (pp. 529). London: Blackwell Science.
- Higuchi, R., Fockler, C., Dollinger, G. and Watson, R. (1993). Kinetic PCR analysis: real-time monitoring of DNA amplification reactions. *Biotechnology*. 11: 1026-1030.
- Hoh, B.P., Siraj, S.S., Tan, S.G. and Yusoff, K. (2013). Segregation and genetic linkage analyses of river catfish, *Mystus nemurus*, based on microsatellite markers. *Genetics and Molecular Research*. 12(3): 2578-2593.
- Holland, M.C., Hassin, S. and Zohar, Y. (2001). Seasonal fluctuations in pituitary levels of the three forms of gonadotropin-releasing hormone in striped bass, *Morone saxatilis* (Teleostei), during juvenile and pubertal development. *Journal of Endocrinology*. 169(3): 527-538.
- Hunt, I. (2005). From gene to protein: a review of new and enabling technologies for multi-parallel protein expression. *Protein Expression and Purification*. 40(1): 1-22.
- Inger, R.F. and Chin, P.K. (2002). The fresh-water fishes of North Borneo. *Fieldiana Zoology*. 45: 1-268.
- Inui, Y., Tagawa, M., Miwa, S. and Hirano, T. (1989). Effects of bovine TSH on the tissue thyroxine level and metamorphosis in prometamorphic flounder larvae. *General and Comparative Endocrinology*. 74(3): 406-410.
- Iserte, J.A., Stephan, B.I., Goñi, S.E., Borio, C.S., Ghiringhelli, P.D. and Lozano, M.E. (2013). Family-specific degenerate primer design: a tool to design consensus degenerated oligonucleotides. *Biotechnology Research International*. 2013: 1-9.
- Jablonka-Shariff, A., Roser, J.F., Bousfield, G.R., Wolfe, M.W., Sibley, L.E., Colgin, M. and Boime, I. (2007). Expression and bioactivity of a single chain recombinant equine luteinizing hormone (reLH). *Theriogenology*. 67(2): 311-320.
- Jackson, K., Goldberg, D., Ofir, M., Abraham, M. and Degani, G. (1999). Blue gourami (*Trichogaster trichopterus*) gonadotropic beta subunits (I and II) cDNA sequences and expression during oogenesis. *Journal of Molecular Endocrinology*. 23(2): 177-187.
- Jensen, F., Nielsen, M. and Nielsen, R. (2014). Increased competition for aquaculture from fisheries: Does improved fisheries management limit aquaculture growth? *Fisheries Research*. 159: 25-33.
- Junaidi, M.S. and Hashida, N.H. (2010). Effect of pH on the waste production of catfish in running water system. Centre for Foundation Studies in

Science, University of Malaya, 50603 Kuala Lumpur, Malaysia. (*Unpublished journal*) <u>http://www.seafdec.org.my/v12/images/pdf/afas2010/FP1.pdf</u>. (Accessed 7 May 2014).

- Kagawa, H., Gen, K., Okuzawa, K. and Tanaka, H. (2003). Effects of luteinizing hormone and follicle-stimulating hormone and insulin-like growth factor-I on aromatase activity and P450 aromatase gene expression in the ovarian follicles of red seabream, *Pagrus major*. *Biology of Reproduction*. 68(5): 1562-1568.
- Kah, O., Anglade, I., Leprêtre, E., Dubourg, P. and de Monbrison, D. (1993).
 The reproductive brain in fish. *Fish Physiology and Biochemistry*. 11(1-6): 85-98.
- Kajimura, S., Yoshiura, Y., Suzuki, M. and Aida, K. (2001). cDNA cloning of two gonadotropin β subunits (GTH-Iβ and-IIβ) and their expression profiles during gametogenesis in the Japanese flounder (*Paralichthys olivaceus*). *General and Comparative Endocrinology*. 122(2): 117-129.
- Kamarudin, M.S., Otoi, S. and Saad, C.R. (2011). Changes in growth, survival and digestive enzyme activities of Asian redtail catfish, *Mystus nemurus*, larvae fed on different diets. *African Journal of Biotechnology*. 10(21): 4484-4493.
- Karpeisky, M.Y., Senchenko, V.N., Dianova, M.V. and Kanevsky, V.Y. (1994). Formation and properties of S-protein complex with S-peptidecontaining fusion protein. *FEBS Letters*. 339(3): 209-212.
- Karsai, A., Müller, S., Platz, S. and Hauser, M.T. (2002). Evaluation of a homemade SYBR[®] Green I reaction mixture for real-time PCR quantification of gene expression. *Biotechniques*. 32(4): 790.
- Kasuto, H. and Levavi-Sivan, B. (2005). Production of biologically active tethered tilapia LHβα by the methylotrophic yeast *Pichia pastoris*. *General and Comparative Endocrinology*. 140(3): 222-232.
- Kawauchi, H., Suzuki, K., Itoh, H., Swanson, P., Naito, N., Nagahama, Y., Nozaki, M., Nakai, Y. and Itoh, S. (1989). The duality of teleost gonadotropins. *Fish Physiology and Biochemistry*. 7(1-6): 29-38.
- Kazeto, Y., Kohara, M., Miura, T., Miura, C., Yamaguchi, S., Trant, J.M., Adachi, S. and Yamauchi, K. (2008). Japanese eel follicle-stimulating hormone (Fsh) and luteinizing hormone (Lh): production of biologically active recombinant Fsh and Lh by *Drosophila* S2 cells and their differential actions on the reproductive biology. *Biology of Reproduction*. 79(5): 938-946.
- Khan, I.A., Hawkins, M.B. and Thomas, P. (1999). Gonadal stage-dependent effects of gonadal steroids on gonadotropin II secretion in the Atlantic

croaker (*Micropogonias undulatus*). *Biology of Reproduction.* 61(3): 834-841.

- Khan, M.S. (1987). Some aspects of the Biology of ikan Baung, Mystus nemurus (Cuvier and Valenciennes) with references to Chenderoh Reservoir. Master dissertation. Universiti Pertanian Malaysia, Malaysia.
- Khan, M.S. (1994). Effect of population density on the growth, feed and protein conversion efficiency and biochemical composition of a tropical freshwater catfish, *Mystus nemurus* (Cuvier and Valenciennes). *Aquaculture and Fisheries Management*. 25(7): 753-760.
- Khan, M.S., Ambak, M.A., Ang, K.J. and Mohsin, A.K.M. (1990). Reproductive biology of a tropical catfish, *Mystus nemurus* Cuvier and Valenciennes, in Chenderoh reservoir, Malaysia. *Aquaculture and Fisheries Management*. 21(2): 173-180.
- Khow, O. and Suntrarachun, S. (2012). Strategies for production of active eukaryotic proteins in bacterial expression system. *Asian Pacific Journal of Tropical Biomedicine*. 2(2): 159-162.
- Kim, J.S. and Raines, R.T. (1994). A misfolded but active dimer of bovine seminal ribonuclease. *European Journal of Biochemistry*. 224(1): 109-114.
- Kim, N.N., Habibi, H.R., Lee, J. and Choi, C.Y. (2012). Effects of recombinant gonadotropin hormones on the expression of vitellogenin, gonadotropin subunits and gonadotropin receptors in cinnamon clownfish, *Amphiprion melanopus*. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*. 162(4): 73-80.
- Kinsella, J.E., Lokesh, B. and Stone, R.A. (1990). Dietary n-3 polyunsaturated fatty acids and amelioration of cardiovascular disease: possible mechanisms. *The American Journal of Clinical Nutrition*. 52(1): 1-28.
- Klein, J., Lobel, L., Pollak, S, Lustbader, B., Ogden, R.T., Sauer, M.V. and Lustbader, J.W. (2003). Development and characterization of a longacting recombinant hFSH against. *Human Reproduction*. 18(1): 50-56.
- Ko, H., Park, W., Kim, D.J., Kobayashi, M. and Sohn, Y.C. (2007). Biological activities of recombinant Manchurian trout FSH and LH: their receptor specificity, steroidogenic and vitellogenic potencies. *Journal of Molecular Endocrinology*. 38(1): 99-111.
- Kobayashi, M. and Stacey, N.E. (1990). Effects of ovariectomy and steroid hormone implantation on serum gonadotropin levels in female goldfish. *Zoological Science*. 7(4): 715-721.
- Kobayashi, M., Morita, T., Ikeguchi, K., Yoshizaki, G., Suzuki, T. and Watabe, S. (2006). *In vivo* biological activity of recombinant goldfish

gonadotropins produced by baculovirus in silkworm larvae. *Aquaculture*. 256(1): 433-442.

- Kobayashi, M.A., Sohn, Y.C., Yoshiura, Y.A. and Aida, K.A. (2000). Effects of sex steroids on the mRNA levels of gonadotropin subunits in juvenile and ovariectomized goldfish *Carassius auratus*. *Fisheries Science*. 66(2): 223-231.
- Kumar, R.S. and Trant, J.M. (2004). Hypophyseal gene expression profiles of FSH-β, LH-β, and glycoprotein hormone-α subunits in *Ictalurus punctatus* throughout a reproductive cycle. *General and Comparative Endocrinology*. 136(1): 82-89.
- Kumar, R.S., Ijiri, S. and Trant, J.M. (2000). Changes in the expression of genes encoding steroidogenic enzymes in the channel catfish (*Ictalurus punctatus*) ovary throughout a reproductive cycle. *Biology of Reproduction*. 63(6): 1676-1682.
- Kumla, S., Doolgindachbaporn, S., Sudmoon, R. and Sattayasai, N. (2012). Genetic variation, population structure and identification of yellow catfish, *Mystus nemurus* (Cuvier and Valenciennes) in Thailand using RAPD, ISSR and SCAR marker. *Molecular Biology Reports*. 39(5): 5201-5210.
- Lapthorn, A.J., Harris, D.C., Littlejohn, A., Lustbader, J.W., Canfield, R.E., Machin, K.J., Morgan, F.J. and Isaacs, N.W. (1994). Crystal structure of human chorionic gonadotropin. *Nature*. 369(6480): 455-461.
- Larsen, D.A. and Swanson, P. (1997). Effects of gonadectomy on plasma gonadotropins I and II in coho salmon, *Oncorhynchus kisutch. General and Comparative Endocrinology*. 108(1): 152-160.
- Leesa-Nga, S.N., Siraj, S.S., Daud, S.K., Sodsuk, P.K., Tan, S.G. and Sodsuk, S. (2000). Biochemical polymorphism in yellow catfish, *Mystus nemurus* (Cuvier and Valenciennes), from Thailand. *Biochemical Genetics*. 38(3-4): 77-86.
- Levavi-Sivan, B., Bogerd, J., Mañanós, E.L., Gómez, A. and Lareyre, J.J. (2010). Perspectives on fish gonadotropins and their receptors. *General and Comparative Endocrinology*. 165(3): 412-437.
- Li, M.D. and Ford, J.J. (1998). A comprehensive evolutionary analysis based on nucleotide and amino acid sequences of the alpha-and betasubunits of glycoprotein hormone gene family. *Journal of Endocrinology*. 156(3): 529-542.
- Loumaye, E., Martineau, I., Piazzi, A., O'Dea, L., Inee, S., Howles, C., Deeosterd, G, Van Loon, K. and Galazka, A. (1996). Clinical assessment of human gonadotrophins produced by recombinant DNA technology. *Human Reproduction*. 11: 95-107.

- Love, J.L., Scholes, P., Gilpin, B., Savill, M., Lin, S. and Samuel, L. (2006). Evaluation of uncertainty in quantitative real-time PCR. *Journal of Microbiological Methods*. 67(2): 349-356.
- Lubzens, E., Young, G., Bobe, J. and Cerdà, J. (2010). Oogenesis in teleosts: how fish eggs are formed. *General and Comparative Endocrinology*. 165(3): 367-389.
- Luckenbach, J.A., Dickey, J.T. and Swanson, P. (2011). Follicle-stimulating hormone regulation of ovarian transcripts for steroidogenesis-related proteins and cell survival, growth and differentiation factors *in vitro* during early secondary oocyte growth in coho salmon. *General and Comparative Endocrinology.* 171(1): 52-63.
- Mañańos, E., Duncan, N. and Mylonas, C.C. (2009). Reproduction and control of ovulation, spermiation and spawning in cultured fish. In E. Cabrita, V. Robles, P. Herraez (Eds). *Methods in reproductive aquaculture: Marine and freshwater species* (pp 27-30). Florida: CRC Press Taylor and Francis Group.
- Mateos, J., Mañanos, E., Carrillo, M. and Zanuy, S. (2002). Regulation of follicle-stimulating hormone (FSH) and luteinizing hormone (LH) gene expression by gonadotropin-releasing hormone (GnRH) and sexual steroids in the Mediterranean Sea bass. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*. 132(1): 75-86.
- Mateos, J., Mañanós, E., Martinez-Rodriguez, G., Carrillo, M., Querat, B. and Zanuy, S. (2003). Molecular characterization of sea bass gonadotropin subunits (α, FSHβ, and LHβ) and their expression during the reproductive cycle. *General and Comparative Endocrinology*. 133(2): 216-232.
- Mazón, M.J., Molés, G., Rocha, A., Crespo, B., Lan-Chow-Wing, O., Espigares, F., Muñoz, I., Felip, A., Carrillo, M., Zanuy, S. and Gómez, A. (2015). Gonadotropins in European sea bass: endocrine roles and biotechnological applications. *General and Comparative Endocrinology*. 221: 31-41.
- Meiri, I., Knibb, W.R., Zohar, Y. and Elizur, A. (2004). Temporal profile of β follicle-stimulating hormone, β luteinizing hormone, and growth hormone gene expression in the protandrous hermaphrodite, gilthead seabream, *Sparus aurata*. *General and Comparative Endocrinology*. 137(3): 288-299.
- Melamed, P., Gur, G., Rosenfeld, H., Elizur, A., Schulz, R.W. and Yaron, Z. (2000). Reproductive development of male and female tilapia hybrids (*Oreochromis niloticus*X *O. aureus*) and changes in mRNA levels of gonadotropin(GtH) Iβ and IIβ subunits. *The Journal of Experimental Zoology*. 286(1): 64-75.

- Melamed, P., Rosenfeld, H., Elizur, A. and Yaron, Z. (1998). Endocrine regulation of gonadotropin and growth hormone gene transcription in fish. *Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology*. 119(3): 325-338.
- Mesomya, W., Cuptapun, Y., Jittanoonta, P., Hengsawadi, D., Boonvisut, S., Huttayanon, P. and Sriwatana, W. (2002). Nutritional evaluations of green catfish *Mystus nemurus*. *The Kasetsart Journal (Natural Sciences)*. 36: 69-74.
- Miesfeld, R.L. (1999). Laboratory tools for molecular genetic application. In R.L. Miesfeld (Ed). *Applied Molecular Genetics* (pp. 31-40). New York: John Wiley and Sons, Inc.
- Moles, G., Carrillo, M., Mañanós, E., Mylonas, C.C. and Zanuy, S. (2007). Temporal profile of brain and pituitary GnRHs, GnRH-R and gonadotropin mRNA expression and content during early development in European sea bass (*Dicentrarchus labrax* L.). General and *Comparative Endocrinology*. 150(1): 75-86.
- Molnár, K., Székely, C., Mohamed, K. and Shaharom-Harrison, F. (2006). Myxozoan pathogens in cultured Malaysian fishes. II. Myxozoan infections of redtail catfish *Hemibagrus nemurus* in freshwater cage cultures. *Diseases of Aquatic Organisms*. 68(3): 219-226.
- Monis, P.T., Giglio, S. and Saint, C.P. (2005). Comparison of SYTO9 and SYBR Green I for real-time polymerase chain reaction and investigation of the effect of dye concentration on amplification and DNA melting curve analysis. *Analytical Biochemistry*. 340(1): 24-34.
- Morita, T., Yoshizaki, G., Kobayashi, M., Watabe, S. and Takeuchi, T. (2004). Fish eggs as bioreactors: the production of bioactive luteinizing hormone in transgenic trout embryos. *Transgenic Research*. 13(6): 551-557.
- Muchlisin, Z.A. and Azizah, M.S. (2009). Influence of cryoprotectants on abnormality and motility of baung (*Mystus nemurus*) spermatozoa after long-term cryopreservation. *Cryobiology*. 58(2): 166-169.
- Muchlisin, Z.A., Hashim, R. and Chong, A.S.C. (2004). Preliminary study on the cryopreservation of tropical bagrid catfish (*Mystus nemurus*) spermatozoa; the effect of extender and cryoprotectant on the motility after short-term storage. *Theriogenology*. 62(1): 25-34.
- Murua, H. and Saborido-Rey, F. (2003). Female reproductive strategies of marine fish species of the North Atlantic. *Journal of Northwest Atlantic Fishery Science*. 33: 23-31.

- Mylonas, C.C., Fostier, A. and Zanuy, S. (2010). Broodstock management and hormonal manipulations of fish reproduction. *General and Comparative Endocrinology*. *165*(3): 516-534.
- Mylonas, C.C., Magnus, Y., Klebanov, Y., Gissis, A. and Zohar, Y. (1997a). Reproductive biology and endocrine regulation of final oocyte maturation of captive white bass. *Journal of Fish Biology*. 51(2): 234-250.
- Mylonas, C.C., Woods, L.C. and Zohar, Y. (1997b). Cyto-histological examination of post-vitellogenesis and final oocyte maturation in captive-reared striped bass. *Journal of Fish Biology*. 50(1): 34-49.
- Nagahama, Y. and Yamashita, M. (2008). Regulation of oocyte maturation in fish. *Development, Growth and Differentiation*. 50(s1): S195-S219.
- Naito, N., Hyodo, S., Okumoto, N., Urano, A. and Nakai, Y. (1991).
 Differential production and regulation of gonadotropins (GTH I and GTH II) in the pituitary gland of rainbow trout, *Oncorhynchus mykiss*, during ovarian development. *Cell and Tissue Research*. 266(3): 457-467.
- Naylor, R.L., Goldburg, R.J., Primavera, J.H., Kautsky, N., Beveridge, M.C., Clay, J., Folke, C., Lubchenco, J., Mooney, H. and Troell, M. (2000). Effect of aquaculture on world fish supplies. *Nature*. 405(6790): 1017-1024.
- Nelson, J.S. (1994). *Fishes of the world* (pp. 600). New York: John Wiley and Sons, Inc.
- Ng, H.H. and Dodson, J.J. (1999). Morphological and genetic descriptions of a new species of catfish, *Hemibagrus chrysops*, from Sarawak, East Malaysia, with an assessment of phylogenetic relationships (Teleostei: Bagridae). *The Raffles Bulletin of Zoology.* 47: 45-57.
- Ng, H.H. and Kottelat, M. (2013). Revision of the Asian catfish genus Hemibagrus bleeker, 1862 (Teleostei: Siluriformes: Bagridae). The Raffles Bulletin of Zoology. 61(1): 205-291.
- Ng, H.H. and Rainboth, W.J. (1999). The bagrid catfish genus *Hemibagrus* (Teleostei: Siluriformes) in central Indochina with a new species from the Mekong River. *The Raffles Bulletin of Zoology*. 47(2): 555-576.
- Nyuji, M., Kazeto, Y., Izumida, D., Tani, K., Suzuki, H., Hamada, K., Mekuchi, M., Gen, K., Soyano, K., Okuzawa, K. (2016). Greater amberjack Fsh, Lh, and their receptors: plasma and mRNA profiles during ovarian development. *General and Comparative Endocrinology*. 225: 224-234.
- Nyuji, M., Selvaraj, S., Kitano, H., Ohga, H., Yoneda, M., Shimizu, A., Kaneko, K., Yamaguchi, A. and Matsuyama, M. (2012). Changes in the

expression of pituitary gonadotropin subunits during reproductive cycle of multiple spawning female chub mackerel *Scomber japonicus*. *Fish Physiology and Biochemistry*. 38(3): 883-897.

- Ogiwara, K., Fujimori, C., Rajapakse, S. and Takahashi, T. (2013). Characterization of luteinizing hormone and luteinizing hormone receptor and their indispensable role in the ovulatory process of the medaka. *PloS One*. 8(1): e54482.
- Okada, T., Kawazoe, I., Kimura, S., Sasamoto, Y., Aida, K. and Kawauchi, H. (1994). Purification and characterization of gonadotropin I and II from pituitary glands of tuna (*Thunnus obesus*). *International Journal of Peptide and Protein Research*. 43(1): 69-80.
- Olsen and Hasan. (2012). A limited supply of fishmeal: Impact on future increases in global aquaculture production. *Trends in Food Science and Technology*. 27(2): 120-128.
- Overton, T.W. (2014). Recombinant protein production in bacterial hosts. *Drug Discovery Today*. 19(5): 590-601.
- Palomares, L.A., Estrada-Moncada, S. and Ramírez, O.T. (2004). Production of recombinant proteins. In P. Balbás, A. Lorence (Eds). *Recombinant Gene Expression: Reviews and Protocols, 2nd* edition (pp. 15-51). Totowa: Humana Press Inc.
- Pankhurst, N.W. and Porter, M.J.R. (2003). Cold and dark or warm and light: variations on the theme of environmental control of reproduction. *Fish Physiology and Biochemistry*. 28(1-4): 385-389.
- Patiño, R. (1997). Manipulations of the reproductive system of fishes by means of exogenous chemicals. *The Progressive Fish-Culturist*. 59(2): 118-128.
- Peter, R.E. and Yu, K.L. (1997). Neuroendocrine regulation of ovulation in fishes: basic and applied aspects. *Reviews in Fish Biology and Fisheries*. 7(2): 173-197.
- Peter, R.E., Lin, H.R., van der Kraak, G., Little, M. (1993). Releasing hormones, dopamine antagonists and induced spawning. In J.F. Muir, R.J. Roberts. (Eds). *Recent Advances in Aquaculture* (pp. 25-30). Oxford: Blackwell Scientific.
- Peti, W. and Page, R. (2007). Strategies to maximize heterologous protein expression in *Escherichia coli* with minimal cost. *Protein Expression and Purification*. 51(1): 1-10.

- Pierce, J.G. (1988). Gonadotropins: chemistry and biosynthesis. In E. Knobil, J, Neill (Eds). *The Physiology of Reproduction* (pp. 1335-1348). New York: Raven Press.
- Pierce, J.G. and Parsons, T.F. (1981). Glycoprotein hormones: structure and function. *Annual Review of Biochemistry*. 50(1): 465-495.
- Planas, J.V. and Swanson, P. (1995). Maturation-associated changes in the response of the salmon testis to the steroidogenic actions of gonadotropins (GTH I and GTH II) *in vitro*. *Biology of Reproduction*. 52(3): 697-704.
- Planas, J.V., Athos, J., Goetz, F.W. and Swanson, P. (2000). Regulation of ovarian steroidogenesis *in vitro* by follicle-stimulating hormone and luteinizing hormone during sexual maturation in salmonid fish. *Biology* of *Reproduction*. 62(5): 1262-1269.
- Ponchel, F., Toomes, C., Bransfield, K., Leong, F.T., Douglas, S.H., Field, S.L., Bell, S.L., Combaret, V., Puisieux, A., Mighell, A.J., Robinson, P.A., Inglehearn, C.F., Isaacs, J.D. and Markham, A.F. (2003). Realtime PCR based on SYBR-Green I fluorescence: an alternative to the TaqMan assay for a relative quantification of gene rearrangements, gene amplifications and micro gene deletions. *BMC Biotechnology*. 3(1): 18.
- Quérat, B. (1994). Molecular evolution of the glycoprotein hormones in vertebrates. In K.G. Davey, R.E. Peter, S.S. Tobe (Eds). *Perspective in Comparative Endocrinology* (pp. 27-35). Ottawa: National Research Council of Canada.
- Quérat, B., Sellouk, A. and Salmon, C. (2000). Phylogenetic analysis of the vertebrate glycoprotein hormone family including new sequences of sturgeon (*Acipenser baeri*) β subunits of the two gonadotropins and the thyroid-stimulating hormone. *Biology of Reproduction*. 63(1): 222-228.
- Rahmah, S., Kato, K., Yamamoto, S., Takii, K., Murata, O. and Senoo, S. (2014). Improved survival and growth performances with stocking density manipulation and shelter availability in bagrid catfish *Mystus nemurus* (Cuvier & Valenciennes 1840) larvae. *Aquaculture Research*. 45(12): 2000-2009.
- Rainboth, W.J. (1996). Fishes of the Cambodian Mekong. FAO Species Identification Sheets for Fishery Purposes. *Food and Agriculture Organization, Rome.*
- Ren, P., Sairam, M.R. and Yarney, T.A. (1995). Bacterial expression of human chorionic gonadotropin α subunit: studies on refolding, dimer assembly and interaction with two different β subunits. *Molecular and Cellular Endocrinology*. 113(1): 39-51.

- Rice, J.C. and Garcia, S.M. (2011). Fisheries, food security, climate change, and biodiversity: characteristic of the sector and perspective on emerging issue. *ICES Journal of Marine Science*. 68(6): 1343-1353.
- Ririe, K.M., Rasmussen, R.P. and Wittwer, C.T. (1997). Product differentiation by analysis of DNA melting curves during the polymerase chain reaction. *Analytical Biochemistry*. 245(2): 154-160.
- Rosano, G.L. and Ceccarelli, E.A. (2014). Recombinant protein expression in *Escherichia coli*: advances and challenges. *Frontiers in Microbiology*. 5 (172): 1-17.
- Rosenfeld, H., Levavi-Sivan, B., Melamed, P., Yaron, Z. and Elizur, A. (1997). The GTHβ subunits of tilapia: gene cloning and expression. *Fish Physiology and Biochemistry*. 17(1-6): 85-92.
- Rosenfeld, H., Meiri, I., Elizur, A. (2007). Gonadotropic regulation of oocyte development: from basic studies to biotechnological applications. In Babin, J.C.a.E.L.P.J. (Ed). *The Fish Oocyte* (pp. 175-202). The Netherlands: Springer.
- Rottmann, R.W., Shireman, J.V. and Chapman, F.A. (1991). Hormonal control of reproduction in fish for induced spawning. *Southern Regional Aquaculture Center (SRAC) Publication.* 424: 1-4.
- Sahdev, S., Khattar, S.K. and Saini, K.S. (2008). Production of active eukaryotic proteins through bacterial expression systems: a review of the existing biotechnology strategies. *Molecular and Cellular Biochemistry*. 307(1-2): 249-264.
- Saida, F. (2007). Overview on the expression of toxic gene products in Escherichia coli. Current Protocols in Protein Science. 5-19.
- Saida, F., Uzan, M., Odaert, B. and Bontems, F. (2006). Expression of highly toxic genes in *E. coli*: special strategies and genetic tools. *Current Protein and Peptide Science*. 7(1): 47-56.
- Saligaut, C., Linard, B., Mañanos, E.L., Kah, O., Breton, B. and Govoroun, M. (1998). Release of pituitary gonadotrophins GtH I and GtH II in the rainbow trout (*Oncorhynchus mykiss*): modulation by estradiol and catecholamines. *General and Comparative Endocrinology*. 109(3): 302-309.
- Samaddar, M., Babu, P.S., Catterall, J.F. and Dighe, R.R. (1999). Identification of an attenuating region in the bovine follicle-stimulating hormone β subunit mRNA that decreases its expression in *E. coli. Gene.* 228(1): 253-260.

- Samuelson, J.C. (2011). Recent developments in difficult protein expression: a guide to *E. coli* strains, promoters, and relevant host mutations. *Methods in Molecular Biology.* 705: 195-209.
- Schulz, R.W., de França, L.R., Lareyre, J.J., LeGac, F., Chiarini-Garcia, H., Nobrega, R.H. and Miura, T. (2010). Spermatogenesis in fish. *General* and Comparative Endocrinology. 165(3): 390-411.
- Schumann, W. and Ferreira, L.C.S. (2004). Production of recombinant proteins in *Escherichia coli*. *Genetics and Molecular Biology*. 27(3): 442-453.
- Senthilkumaran, B. and Joy, K.P. (1996). Effects of administration of some monoamine-synthesis blockers and precursors on ovariectomy-induced rise in plasma gonadotropin II in the catfish *Heteropneustes fossilis*. *General and Comparative Endocrinology*. 101(2): 220-226.
- Sharaf, S.M. (2012). Effect of GnRHa, pimozide and Ovaprim on ovulation and plasma sex steroid hormones in African catfish *Clarias gariepinus*. *Theriogenology*. 77(8): 1709-1716.
- Siraj, S.S., Daud, S.K., Othman, A. and Tan, S.G. (1998). Population genetic structure of Baung, *Mystus nemurus* (Cuvier and Valenciennes). *Malaysian Applied Biology Journal*. 27: 77.
- Smith, H.M. (1945). *The Freshwater Fish of Siam or Thailand* (pp. 382-387). Washington DC: U.S. Government Printing Office.
- Sohn, Y.C., Yoshiura, Y., Kobayashi, M. and Aida, K. (1999). Seasonal changes in mRNA levels of gonadotropin and thyrotropin subunits in the goldfish, *Carassius auratus*. *General and Comparative Endocrinology*. 113(3): 436-444.
- Sørensen, H.P. and Mortensen, K.K. (2005). Advanced genetic strategies for recombinant protein expression in *Escherichia coli. Journal of Biotechnology*. 115(2): 113-128.
- Sower, S.A., Freamat, M. and Kavanaugh, S.I. (2009). The origins of the vertebrate hypothalamic-pituitary-gonadal (HPG) and hypothalamic-pituitary-thyroid (HPT) endocrine systems: new insights from lampreys. *General and Comparative Endocrinology*. 161(1): 20-29.
- Srichanun, M., Tantikitti, C., Vatanakul, V. and Musikarune, P. (2012). Digestive enzyme activity during ontogenetic development and effect of live feed in green catfish larvae (*Mystus nemurus* Cuvier and Valenciennes). Songklanakarin Journal of Science and Technology. 34(3): 247-254.
- Subasinghe, R., Soto, D. and Jia, J. (2009). Global aquaculture and its role in sustainable development. *Reviews in Aquaculture*. 1(1): 2-9.

- Suetake, H., Okubo, K., Sato, N., Yoshiura, Y., Suzuki, Y. and Aida, K. (2002). Differential expression of two gonadotropin (GTH) β subunit genes during ovarian maturation induced by repeated injection of salmon GTH in the Japanese eel *Anguilla japonica*. *Fisheries Science*. 68(2): 290-298.
- Suzuki, K., Kanamori, A., Nagahama, Y. and Kawauchi, H. (1988b). Development of salmon GTH I and GTH II radioimmunoassays. *General and Comparative Endocrinology*. 71(3): 459-467.
- Suzuki, K., Kawauchi, H. and Nagahama, Y. (1988a). Isolation and characterization of two distinct gonadotropins from chum salmon pituitary glands. *General and Comparative Endocrinology*. 71(2): 292-301.
- Swanson, P. (1991). Salmon gonadotropins: reconciling old and new ideas. In A.P. Scott, J.P. Sumpter, D.M. Kime, M.S. Rolfe (Eds). *Reproductive Physiology of Fish* (pp. 2-7). UK: FishSymp91, University of East Anglia.
- Swanson, P., Dickey, J.T. and Campbell, B. (2003). Biochemistry and physiology of fish gonadotropins. *Fish Physiology and Biochemistry*. 28(1-4): 53-59.
- Szkudlinski, M.W., Fremont, V., Ronin, C. and Weintraub, B.D. (2002). Thyroid-stimulating hormone and thyroid-stimulating hormone receptor structure-function relationships. *Physiological Reviews*. 82(2): 473-502.
- Tacon, A.G., Metian, M., Turchini, G.M. and De Silva, S.S. (2009). Responsible aquaculture and trophic level implications to global fish supply. *Reviews in Fisheries Science*. 18(1): 94-105.
- Tan, E.S.P. and Kwan, F.S. (1985). Solutions to some constraints of the aquaculture industry in Peninsular Malaysia. Paper presented at the conference of the 2nd Asian Conference on Technology for Rural Development, Kuala Lumpur. December 1985.
- Tanaka, H., Kagawa, H., Okuzawa, K. and Hirose, K. (1993). Purification of gonadotropins (PmGTH I and II) from red seabream (*Pagrus major*) and development of a homologous radioimmunoassay for PmGTH II. *Fish Physiology and Biochemistry*. 10(5): 409-418.
- Tantikitti, C. and Chimsung, N. (2001). Dietary lysine requirement of freshwater catfish (*Mystus nemurus* Cuvier and Valenciennes). *Aquaculture Research*. 32(s1): 135-141.
- Terpe, K. (2003). Overview of tag protein fusions: from molecular and biochemical fundamentals to commercial systems. *Applied Microbiology* and Biotechnology. 60(5): 523-533.

- Teugels, G.G. (1996). Taxonomy, phylogeny and biography of catfishes (Ostariophysi, Siluroidei): an overview. *Aquatic Living Resources*. 9(S1): 9-34.
- Thalathiah, S., Ibrahim, T. and Mansor, A. (1992). *Induced spawning of Mystus nemurus (Cuvier and Valenciennes) using heteroplastic pituitary extract, HCG and an analog of LHRH.* Paper presented at the seminar of the Fisheries Research Seminar, Malacca. June 1989.
- Tian, J., Ma, K. and Saaem, I. (2009). Advancing high-throughput gene synthesis technology. *Molecular BioSystems*. 5(7): 714-722.
- Tokumoto, T., Yamaguchi, T., Li, S. and Tokumoto, M. (2011). *In vivo* induction of oocyte maturation and ovulation in zebrafish. *PloS One*. 6(9): e25206.
- Tucker, J.W. (1994). Spawning by captive serranid fishes: a review. *Journal* of the World Aquaculture Society. 25(3): 345-359.
- Tyler, C.R. and Sumpter, J.P. (1996). Oocyte growth and development in teleosts. *Reviews in Fish Biology and Fisheries*. 6(3): 287-318.
- Tyler, C.R., Pottinger, T.G., Coward, K., Prat, F., Beresford, N. and Maddix, S. (1997). Salmonid follicle-stimulating hormone (GtH I) mediates vitellogenic development of oocytes in the rainbow trout, *Oncorhynchus mykiss*. *Biology of Reproduction*. 57(5): 1238-1244.
- UN-DESA. (2009). Population Division of the Department of the Department of Economic and Social Affairs of the United Nation Secretariat, World Population Prospects: the 2008 Revision and World Urbanization Prospects: the 2009 Revision. http://esa.un.org/wup2009/unup/ index.asp. (Accessed 7 May 2014).
- Usmani, S., Tan, S.G., Siraj, S.S. and Yusoff, K. (2003). Population structure of the Southeast Asian river catfish *Mystus nemurus*. *Animal Genetics*. 34(6): 462-464.
- Van Der Kraak, G., Suzuki, K., Peter, R.E., Itoh, H. and Kawauchi, H. (1992). Properties of common carp gonadotropin I and gonadotropin II. *General and Comparative Endocrinology*. 85(2): 217-229.
- Vischer, H.F., Granneman, J.C., Linskens, M.H., Schulz, R.W. and Bogerd, J. (2003). Both recombinant African catfish LH and FSH are able to activate the African catfish FSH receptor. *Journal of Molecular Endocrinology*. 31(1): 133-140.
- Voet, D and Voet, J.G. (1995). Nucleic acid structures and manipulation. In
 D. Voet, J.G. Voet (Eds). *Biochemistry 2nd edition* (pp. 848-914).Somerset: John Wiley and Sons, Inc.

- Von Ihering, R. (1937). A method for inducing fish to spawn. *The Progressive Fish-Culturist*. 4(34): 15-16.
- Wallace, R.A. and Selman, K. (1981). Cellular and dynamic aspects of oocyte growth in teleosts. *American Zoologist*. 21(2): 325-343.
- Wallis, M. (2001). Episodic evolution of protein hormones in mammals. *Journal of Molecular Evolution*. 53(1): 10-18.
- Walsh, G. (2004). Second-generation biopharmaceuticals. *European Journal* of *Pharmaceutics and Biopharmaceutics*. 58(2): 185-196.
- Welch, M., Govindarajan, S., Ness, J.E., Villalobos, A., Gurney, A., Minshull, J. and Gustafsson, C. (2009). Design parameters to control synthetic gene expression in *Escherichia coli*. *PloS one*. 4(9): e7002-e7002.
- Weltzien, F.A., Andersson, E., Andersen, Ø., Shalchian-Tabrizi, K. and Norberg, B. (2004). The brain-pituitary-gonad axis in male teleosts, with special emphasis on flatfish (*Pleuronectiformes*). Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology. 137(3): 447-477.
- Weltzien, F.A., Taranger, G.L., Karlsen, Ø. and Norberg, B. (2002). Spermatogenesis and related plasma androgen levels in Atlantic halibut (*Hippoglossus hippoglossus L.*). Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology. 132(3): 567-575.
- Wen, H.S., Lin, H.R., Mao, Y.Z., Wang, L. and Zhang, Y.P. (2003). Annual variations of gonadotropin content and ovarian development of feral female catfish, *Silurus asotus*, in central China. *Environmental Biology* of Fishes. 68(3): 283-291.
- Wong, T.T. and Zohar, Y. (2004). Novel expression of gonadotropin subunit genes in oocytes of the gilthead seabream (*Sparus aurata*). *Endocrinology*. 145(11): 5210-5220.
- Wong, T.T., Gothilf, Y., Zmora, N., Kight, K.E., Meiri, I., Elizur, A. and Zohar, Y. (2004). Developmental expression of three forms of gonadotropinreleasing hormone and ontogeny of the hypothalamic-pituitary-gonadal axis in gilthead seabream (*Sparus aurata*). *Biology of Reproduction*. 71(3): 1026-1035.
- Xia, H., Chen, F. and Puett, D. (1994). A region in the human glycoprotein hormone alpha-subunit important in holoprotein formation and receptor binding. *Endocrinology*. 134(4): 1768-1770.
- Yaron, Z. (1995). Endocrine control of gametogenesis and spawning induction in the carp. *Aquaculture*. 129(1): 49-73.

- Yaron, Z., Gur, G., Melamed, P., Rosenfeld, H., Elizur, A. and Levavi-Sivan,
 B. (2003). Regulation of fish gonadotropins. *International Review of Cytology*. 225: 131-185.
- Yoshiura, Y., Kobayashi, M., Kato, Y. and Aida, K. (1997). Molecular cloning of the cDNAs encoding two gonadotropin β subunits (GTH-Iβ and-IIβ) from the goldfish, *Carassius auratus*. *General and Comparative Endocrinology*. 105(3): 379-389.
- Zmora, N., Kazeto, Y., Kumar, R.S., Schulz, R.W. and Trant, J.M. (2007). Production of recombinant channel catfish (*Ictalurus punctatus*) FSH and LH in S2 *Drosophila* cell line and an indication of their different actions. *Journal of Endocrinology*. 194(2): 407-416.
- Zohar, Y. (1989a). Fish reproduction: its physiology and artificial manipulation. In M. Shilo, S. Sarig. (Eds). *Fish Culture in Warm Water Systems: Problems and Trends* (pp. 65-119). Boca Raton: CRC Press.
- Zohar, Y. (1989b). Endocrinology and fish farming: aspects in reproduction, growth, and smoltification. *Fish Physiology and Biochemistry*. 7(1-6): 395-405.
- Zohar, Y. and Mylonas, C.C. (2001). Endocrine manipulations of spawning in cultured fish: from hormones to genes. *Aquaculture*. 197(1): 99-136.

LIST OF PUBLICATIONS

Journals

- <u>Zulperi, Z.</u>, Ina-Salwany, M.Y., Christianus, A., Yusoff, F.M. and Harmin, S.A. (2017). Ovarian development and changes of the expression of pituitary gonadotropin subunits during reproductive cycle of female Malaysian river catfish, *Hemibagrus nemurus* (Valenciennes, 1840) in captivity. *Journal of Applied Ichthyology*. 00: 1-8. doi: 10.1111/jai.13267
- Zulperi, Z., Ina-Salwany, M.Y., Christianus, A., Yusoff, F.M. and Harmin, S.A. (2015). Molecular characterization of *Hemibagrus nemurus* gonadotropin subunits: cDNA cloning and phylogenetic analysis. *Research in Biotechnology*. 6(1): 26-46.
- 3. <u>Zulperi, Z.M.</u>, Omar, A.R. and Arshad, S.S. (2009). Sequence and phylogenetic analysis of S1, S2, M, and N genes of infectious bronchitis virus isolates from Malaysia. *Virus Genes*. 38(3): 383-391.
- 4. Nehlah, R., Ina-Salwany, M.Y. and **Zulperi, Z.** (2016). Antigenicity analysis and molecular characterization of two outer membrane proteins of *Vibrio alginolyticus* strain VA2 as vaccine candidates in tiger grouper culture. *Journal of Biological Sciences*. 16(1): 1-11.
- Ina-Salwany, M.Y., Hishammuddin, H., <u>Zulperi, Z.</u>, Salema, M., Karim, M. and Natrah, F.M.I. (2015). Elucidating the probiotic potential of Malaysia *Paenibacillus pabuli* against *Vibrio alginolyticus* in artemia culture. *Asian Journal of Agricultural Research*. 9 (5): 223-236.
- Zmora, N., Stubblefield, J., <u>Zulperi, Z.</u>, Biran, J., Levavi-Sivan, B., Muñoz-Cueto, J.A. and Zohar, Y. (2012). Differential and gonad stage-dependent roles of kisspeptin1 and kisspeptin2 in reproduction in the modern teleosts, *Morone* species. *Biology of Reproduction*. 86(6): 177-177.
- Zmora, N., Stubblefield, J., <u>Zulperi, Z</u>., Klenke, U. and Zohar, Y. (2011). Kisspeptin-photoperiod/gonadal steroid relationships in the brain of two perciforms, the striped and hybrid basses. *Indian Journal of Science and Technology*. 4(S8): 10-11.

Manuscript Submitted

1. Ina-Salwany, M.Y., <u>Zulperi, Z.</u>, Christianus, A., Yusoff, F.M. and Harmin, S.A. Recombinant luteinizing hormone development to improve the reproductive performance in female Malaysia catfish, *Hemibagrus nemurus*.

Proceedings

- <u>Zulperi, Z</u>., Ina-Salwany, M.Y., Christianus, A. and Yusoff, F.M. (2016). Expression of gonadotropin subunit genes in Malaysian river catfish, *Hemibagrus nemurus* (Valenciennes 1840) in captivity. 7th International Agriculture Congress 2016. 4-6 October 2016. Hotel Bangi-Putrajaya, Malaysia. p. 736.
- Zulperi, Z., Ina-Salwany, M.Y., Christianus, A., Yusoff, F.M. and Harmin, S.A. (2014). Molecular characterization and production of recombinant protein of LHβ subunit to induce spawning in *Hemibagrus nemurus*. The 3rd Thailand-Malaysia Graduate Forum in Life Sciences, Food Science and Agriculture 2014. 16-20 December 2014. Kasetsart University, Bangkok, Thailand. p. 38.
- Zulperi, Z., Ina-Salwany, M.Y., Christianus, A., Yusoff, F.M. and Harmin, S.A. (2014). Molecular cloning and phylogenetic analysis of gonadotropin gene subunits of Asian redtail catfish, *Hemibagrus nemurus*. World Aquaculture Adelaide 2014. 7-11 June 2014. Adelaide Convention Centre, Adelaide, Australia. p. 481.

GenBank

- 1. <u>Zulperi, Z., Ina-Salwany</u>, M.Y., Christianus, A., Harmin, S.A. and Yusoff, F.M. (2014). *Hemibagrus nemurus* glycoprotein hormone alpha subunit mRNA, complete cds. Accession no: **KF934189**.
- <u>Zulperi, Z., Ina-Salwany, M.Y., Christianus, A.,</u> Harmin, S.A. and Yusoff, F.M. (2014). *Hemibagrus nemurus* luteinizing hormone beta subunit mRNA, complete cds. Accession no: **KF934190.**
- 3. <u>Zulperi, Z.</u>, Ina-Salwany, M.Y., Christianus, A., Harmin, S.A. and Yusoff, F.M. (2014). *Hemibagrus nemurus* follicle stimulating hormone beta subunit mRNA, complete cds. Accession no: KF998583.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : ____

TITLE OF THESIS / PROJECT REPORT : Molecular Characterization and Recombinant Gonadotropin Subunit Development for Improving Reproductive Performance in Female Hemibagrus nemurus Valenciennes

NAME OF STUDENT : Zarirah Mohamed Zulperi

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

- 1. This thesis/project report is the property of Universiti Putra Malaysia.
- 2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
- 3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (V)



CONFIDENTIAL



RESTRICTED



OPEN ACCESS

(Contain confidential information under Official Secret Act 1972).

(Contains restricted information as specified by the organization/institution where research was done).

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

PATENT

Embargo from		until
· <u> </u>	(date)	

(date)

Approved by:

(Signature of Student) New IC No/ Passport No.: (Signature of Chairman of Supervisory Committee) Name:

Date :

Date :

[Note : If the thesis is CONFIDENTIAL or RESTRICTED, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]