



UNIVERSITI PUTRA MALAYSIA

ANTIPROLIFERATIVE EFFECTS OF ZERUMBONE-HYDROXYPROPYL- β -CYCLODEXTRIN INCLUSION COMPLEX ON HepG2 LIVER CANCER CELLS *In Vitro*

NABILAH BINTI MUHAMMAD NADZRI

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HYDROXYPROPYL- β -CYCLODEXTRIN INCLUSION COMPLEX ON
HepG2 LIVER CANCER CELLS *In Vitro***

By

NABILAH BINTI MUHAMMAD NADZRI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirement for the Degree of Master of Science**

May 2013

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In the Name of Allah, the Most Merciful and Most Compassionate Dedication

We all have dreams. But in order to make dreams come into reality, it takes an awful lot of determination, dedication, effort and continuous support from our loved ones.

Specially dedicated to,

Allah SWT & Prophet Muhammad (SAW)

My beloved parents

My brothers & sisters

My fiancée

Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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NABILAH BINTI MUHAMMAD NADZRI

May 2013

Chair: Ahmad Bustamam Abdul, PhD

Faculty: Institute of Bioscience

Zingiber zerumbet Smith locally known as 'lempoyang' or wild ginger belongs to the Zingiberaceae family. Previous investigations on *Z. zerumbet* proved that it contained a bioactive compound, zerumbone (ZER), a crystalline sesquiterpene possessing suppressive effect in cancers. The purpose of encapsulating ZER with hydroxylpropyl- β -cyclodextrin (HP β CD) is to modify ZER's solubility and pharmacokinetic properties making it less harmful to the human body. The objective of this study is to investigate antiproliferative activities of ZER-HP β CD inclusion complex towards HepG2 liver cancer cells. The MTT assay showed that the inclusion complex is cytotoxic towards HepG2 cells with an IC₅₀ of 11.43 μ g/ml. Morphological evaluation showed structural changes associated with apoptosis including membrane blebbing, cell shrinkage and chromatin condensation. HepG2 cells treated with the inclusion complex further resulted in cell cycle arrest at G2/M phase with increments of apoptotic cells. Further investigations showed the release of

cytochrome c and loss of mitochondrial membrane potential, proving mitochondrial dysfunction upon the ZER-HP β CD treatment as well as modulating pro-apoptotic and anti-apoptotic Bcl-2 family members with no significant change of p53. The activated Bax will be translocated to the mitochondria, which activates the transformation of mitochondrial function and release of cytochrome c. Upregulation of caspase 3/7, caspase 9 and caspase 8 were also detected with the depletion of BID cleaved by caspase 8 proving that both extrinsic and intrinsic pathway were involved upon ZER-HP β CD induction. Collectively, these results demonstrate that the highly soluble inclusion complex of ZER with HP β CD induce apoptosis programmed cell death in HepG2 cells and can be extrapolated to postulate that caspase 8's activation is indirectly involved as an interconnection between the extrinsic and intrinsic pathway. Further investigations to this complex are needed to substantiate its use as an anticancer against hepatocellular carcinoma in humans.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN ANTIPROLIFERASI KOMPLEKS ZERUMBON-
HIDROKSILPROPIL- β -SIKLODEKSTRIN TERHADAP SEL KANSER
HATI HepG2 *In Vitro***

Oleh

NABILAH MUHAMMAD NADZRI

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Zingiber zerumbet Smith yang dikenali sebagai 'lempoyang' atau halia liar yang termasuk dalam famili Zingiberaceae. Kajian sebelum ini telah membuktikan bahawa *Z. zerumbet* mengandungi kompaun bioaktif, zerumbon (ZER), sesquiterpen kristal yang mampu menindas pertumbuhan pelbagai jenis kanser. Pengkapsulan ZER dengan hidrosilpropil- β -siklodekstrin adalah bertujuan untuk memodifikasi kadar kelarutan ZER dan ciri-ciri farmakokinetiknya sekaligus menjadikannya kurang bahaya kepada tubuh manusia. Objektif kajian ini adalah bertujuan untuk menyelidik aktiviti antiproliferasi kompleks inklusi ZER-HP β CD terhadap sel kanser hati, HepG2. Keputusan asai MTT menunjukkan kompleks inklusi adalah sitotoksik terhadap sel HepG2 dengan bacaan IC₅₀ 11.43 μ g/ml. Penilaian morfologi menunjukkan pertukaran struktur yang berkaitan dengan apoptosis termasuk pengelembungan membran, pengecutan sel dan pemekatan kromatin. Sel HepG2 yang dirawat oleh kompleks inklusi ZER-HP β CD seterusnya mencetuskan

penghentian kitaran sel pada fasa G2/M serta peningkatan sel apoptotik. Kajian lanjutan menunjukkan pembebasan sitokrom c dan kehilangan potensi membran mitokondria membuktikan rawatan kompleks inklusi ZER-HP β CD menyebabkan kehilangan fungsi mitokondria sekali gus memodulasi ahli keluarga pro-apoptotik dan anti-apoptotik tanpa perubahan p53 yang signifikan. Bax yang teraktif akan ditranslokasikan ke mitokondria dan mengaktifkan transformasi fungsi mitokondria sekali gus pembebasan sitokrom c. Peningkatan kaspase 3/7, kaspase 8 dan kaspase 9 yang signifikan serta pemotongan BID oleh kaspase 8 juga dikesan menunjukkan kedua-dua tapak jalan ekstrinsik dan intrinsik terlibat dengan induksi ZER-HP β CD. Secara keseluruhan, kajian menunjukkan bahawa kompleks inklusi ZER-HP β CD berkebolehan untuk mengaruhkan kematian sel secara terprogram apoptosis dalam sel HepG2 dan boleh dicadangkan bahawa pengaktifan kaspase 8 secara tidak langsung terlibat dalam menghubungkan di antara tapak jalan ekstrinsik dan intrinsik. Penyelidikan yang lebih mendalam terhadap kompleks inklusi ini adalah diperlukan untuk digunakan sebagai antikanser terhadap hepatoselular karsinoma manusia.

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I certify that a Thesis Examination Committee has met on 14 May 2013 to conduct the final examination of Nabilah Binti Muhammad Nadzri on her thesis entitled “**Antiproliferative Effects of Zerumbone-Hydroxypropyl- β -Cyclodextrin Inclusion Complex on HepG2 Liver Cancer Cells *In Vitro***” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Master of Science.

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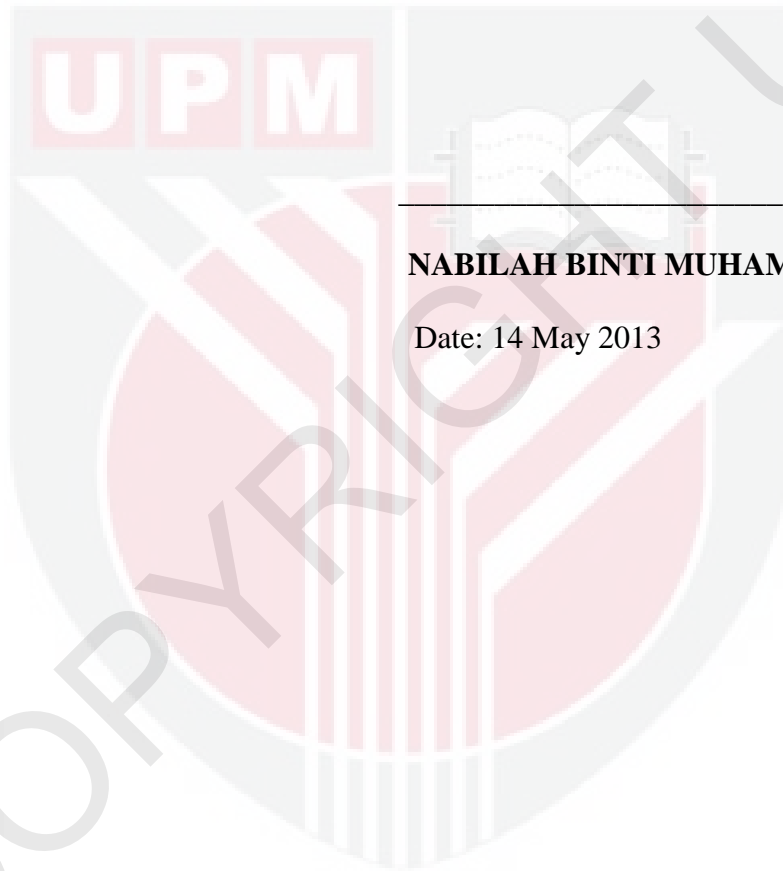
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DECLARATION

I hereby declare that the thesis is based on my original work except for quotation and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Putra Malaysia or other institutions.



NABILAH BINTI MUHAMMAD NADZRI

Date: 14 May 2013

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LIST OF ABBREVIATIONS

%	Percentage
µg	Microgram
µl	Microliter
10 ⁵	100 000
AAF	Acetylaminofluorene
AFB1	Aflatoxin B1
AIF	Apoptosis-inducing factor
ANOVA	Analysis of variance
AO	Acridine orange
AOM	Azoxymethane
Apaf-1	Apoptotic protease-activating factor-1
APS	Ammonium persulfate
ATP	Adenosine-5'-triphosphate
Bad	Bcl-2-associated death promoter
Bak	Bcl-2 homologous antagonist killer
Bax	Bcl-2 associated X protein
Bcl-2	B cell lymphoma 2
Bcl-xL	B cell lymphoma extra large
BID	BH3 interacting domain death agonist
BIR	Baculovirus inhibitor of apoptosis protein repeat
BSA	Bovine serum albumin
CA	Chromosome aberrations
Caov-3	Ovarian cancer cells

Caspases	Cysteine-dependent aspartate-directed proteases
CD	Cyclodextrins
Cdk	Cyclin-dependent kinase
CEMss	T-acute lymphoblastic leukemia cells
CIN	Cervical intraepithelial neoplasia
CMC	Sodium carboxymethyl cellulose
CO ₂	Carbon dioxide
dATP	2'-Deoxyadenosine 5'-triphosphate
DEN	Diethylnitrosamine
DES	Diethylstilboestrol
dH ₂ O	Distilled water
DISC	Death-inducing signaling complex
DISC	Death-inducing signalling complex
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DSS	Dextran sodium sulfate
EMB	Embryonal histologic
FADD	Fas-associated death domain
FasL	Fas ligand
FDA	Food and Drug Administration
FH	Fetal histologic
FITC	Fluorescein isothiocyanate
G0	Gap 0 at cell cycle
G1	Gap 1 at cell cycle
G2/M	Gap 2/ Mitosis at cell cycle

h	Hour
HB	Hepablastoma
HBV	Hepatitis B virus
HBx	Hepatitis B virus X
HCC	Hepatocellular carcinoma
HCV	Hepatitis C virus
HeLa	Cervical cancer cells
HepG2	Liver cancer cells
HO	Heme oxygenase
HP- β -CD	Hydroxypropyl- β -cyclodextrin
Hsp70	Heat shock protein 70
HT-29-D4	Colon cancer cells
IAP	Inhibitor of apoptosis protein
IC ₅₀	Inhibitory concentration
KCl	Potassium chloride
M	Metastasis for TNM staging
MCF-7	Estrogen receptor positive breast cancer cells
MDA-MB-231	Estrogen receptor negative breast cancer cells
ml	Mililitre
MMP	Mitochondrial membrane potential
MOMP	mitochondrial outer membrane permeabilization
MPT	Mitochondrial permeability transition
MTT	3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
N	Node for TNM staging
NaCl	Sodium chloride

NCE	New chemical entity
NCI	National Cancer Institute
NF- κ B	Nuclear Factor-KappaB
P53	Tumor protein 53
PBL	Human peripheral blood lymphocytes
PBS	Phosphate buffer saline
pH	Potential of hydrogen
PI	Propidium Iodide
PMSF	Phenylmethylsulfonyl fluoride
PS	Phosphatidylserine
RNA	Ribonucleic acid
Rpm	Rotation per minute
SCU	Small cell undifferentiated histologic
SD	Standard deviation
SDS	Sodium dodecyl sulfate
SMAC	Second mitochondria-derived activator of caspase
T	Size for TNM staging
tBID	Truncated BH3 interacting domain death agonist
TEMED	Tetramethylethylenediamine
TNF	Tumor necrosis factor
TNF-R1	Tumor necrosis factor receptor 1
TRAIL	Tumor necrosis factor related apoptosis-inducing ligand
TRAIL-R2	TNF related apoptosis-inducing ligand receptor 2
TUNEL	TdT-mediated dUTP nick end labelling
UPM	Universiti Putra Malaysia

WRL-68	Normal liver cells
XIAP	X-linked inhibitor of apoptosis protein
ZER	Zerumbone
$\Delta\Psi_m$	Mitochondrial transmembrane potential



CHAPTER I

INTRODUCTION

1.1 Introduction

Liver cancer can be classified into primary and secondary types. The primary liver cancers are Hepatocellular Carcinoma (HCC) and Hepatoblastoma whilst secondary liver cancer is a metastatic liver cancer is originated from other organs such as the colon, stomach, pancreas, breast, and lung. Liver cancer is the sixth most frequent cancer in the world. Nearly 626,000 or 5.7% new cases are reported each year. It is also the third highest cancer-related death after lung and stomach cancer. In Malaysia, it is the tenth highest cancer-related death among man with 378 cases reported for every 100,000 population (Lim and Halimah, 2004).

The use of plants as the preferential treatment for cancer has been known for centuries. To date about 3000 plant species have been identified to possess anticancer properties. This historical information was utilized by modern scientists in search for better compounds as anticancers agent (Cragg *et al.*, 2005). One of the most important plant contributions for the world health is the anticancer agents derived from natural plants (Cragg *et al.*, 1997). This is proven by the production of more than 60 types of clinical chemotherapy drugs derived from plants. A number of promising new agents are in clinical development based on selective activity against cancer-related molecular targets, including flavopiridol and combretastin-A4-

phosphate, while some agents which failed in earlier clinical studies have stimulating renewed interest (Cragg *et al.*, 2005).

Zingiber zerumbet is a member of the family *Zingiberaceae*. It is an erect herb up to two meters tall arising from a bulky yellowish pungent underground rhizome. This plant can be found in the moist forest, beach thickets and mangrove margin from sea-level to over 500m. In peninsular Malaysia, it is found around villages and in secondary growth. It is indigenous in south-east Asia, widely distributed and naturalize all over the south pacific (Kho, 2005). Other names for this plant are lempoyang, 'lempoyang gajah', 'halia putih', shampoo ginger and pinecones ginger (Ahmad *et al.*, 1994). Some of the traditional usages of *Z. Zerumbet* include the treatment of fever, indigestion, diarrhea, severe sprains, inflammation, constipation, toothache and some of the Malays use the rhizomes to cure edema and worm infestation in children (Zakaria *et al.*, 2010).

Zerumbone (ZER), is a crystalline monocyclic sesquiterpene derived from the rhizomes wild ginger, *Z. zerumbet*. This bioactive component has its unique structure with cross-conjugated ketone in an 11-membered ring, as well as having interesting biological activities. It has been reported that the compound ZER constitute about 37% of *Z. zerumbet* (Sakinah *et al.*, 2007).

Hydroxypropyl- β -cyclodextrin (HP β CD) is a cyclodextrin derivative that is widely studied in the field of drug encapsulation because of its inclusion ability as well as its high water solubility. HP β CD is well tolerated by human body both by intravenous and oral administration (Gould and Scott, 2005). Eid *et al.* (2011a) investigated the

inclusion complex between ZER and HP β CD and its characterization. The ZER penetrates completely into the cavity of HP β CD and the solubility of ZER was enhanced >30 fold after complexation. The current results showed that HP β CD is a suitable encapsular capable of forming thermodynamically stable complex with ZER for delivery of the compound as an anticancer. Hence, taking into consideration the importance of biopharmaceutical characterization, the antiproliferative effects of the new ZER-HP β CD inclusion complex was carried out on HepG2 cells in this study.

1.2 Hypothesis

Treated HepG2 liver cancer cells with zerumbone-hydroxypropyl- β -cyclodextrin (ZER-HP β CD) inclusion complex will induce apoptosis, which will involve modulation of pro-apoptotic and anti-apoptotic family member proteins.

1.3 Objective

To investigate the *in vitro* anti-proliferative properties and the apoptosis pathway determination of the inclusion complex of zerumbone with hydroxypropyl- β -cyclodextrin (ZER-HP β CD) on HepG2 liver cancer cells.

REFERENCES

- Abdelwahab, S.I., Abdul, A.B., Devi, N., Taha, M.M.E., Al-zubairi, A.S., Mohan, S. & Mariod, A.A. 2010. Regression of cervical intraepithelial neoplasia by zerumbone in female Balb/c mice prenatally exposed to diethylstilboestrol: Involvement of mitochondria-regulated apoptosis. *Experimental and Toxicologic Pathology* 62: 461-469.
- Abdelwahab, S.I., Abdul, A.B., Mohan, S., Taha, M.M.E., Syam, S., Ibrahim, M.Y. & Mariod, A.A. 2011. Zerumbone induces apoptosis in T-acute lymphoblastic leukemia cells. *Leukemia Research* 35: 268-271.
- Abdelwahab, S.I., Abdul, A.B., Zain, Z.N.M. and Hadi, A.H.A. 2012. Zerumbone inhibits interleukin-6 and induces apoptosis and cell cycle arrest in ovarian and cervical cancer cells. *International Immunology* 4: 594-602.
- Abdul, A.B., Abdel-Wahab, S.I., Fong, H.K., Mohan, S.M., Al-Zubairi, A.S. and Elhassan, M.M. 2009. *In vitro* response of cancer cells to the growth-inhibitory effects of dichloromethane extract of *Goniothalamus umbrosus*. *Research Journal of Pharmacology* 3: 1-6.
- Abdul, A.B.H., Al-Zubairi, A.S., Tailan, N.D., Wahab, S.I.A., Zain, Z.N.M., Ruslay, S. & Syam, M.M. 2008. Anticancer activity of natural compound (zerumbone) extracted from *Zingiber zerumbet* in human HeLa cervical cancer cells. *International Journal of Pharmacology* 4(3): 160-168.
- Ahmad, U.K., Sirat, H.M., Marsin Sanagi, M. & Smith, R.M. 1994. Supercritical fluid extraction and capillary gas chromatography of the rhizomes of *Z. Zerumbet*. *Journal of Microcolumn Separations* 6(1): 27-32.
- Alwi, S.S.S, Nallapan, M. and Pihie, A.H.L. 2007. Zerumbone exerts antiproliferative activity via apoptosis on hepG2 cells. *Malaysian Journal of Biochemistry and Molecular Biology* 15: 19-23.
- Al-Zubairi, A.S., Abdul, A.B. & Syam, M.M. 2010. Evaluation of the genotoxicity of zerumbone in cultured human peripheral blood lymphocytes. *Toxicology in Vitro* 24: 707-712.
- American Society of Clinical Oncology. 2011. Liver Cancer. Retrieve from <http://www.cancer.net/patient/Cancer+Types/Liver+Cancer>
- Antonsson, B. and Martinou, J.C. 2000. The Bcl-2 protein family. *Experimental Cell Research* 256: 50-57.
- Ashkenazi, A. And Dixit, V.M. 1998. Death receptors: signaling and modulation. *Science*. 281: 1305-1308.
- Bailey, R.W., Nguyen, T., Robertson, L., Gibbons, E., Nelson, J., Christensen, R.E., Bell, J.P., Judd, A.M. and Bell, J.D. 2009. Sequence of physical changes

to the cell membrane during glucocorticoid-induced apoptosis in S49 in lymphoma cells. *Biophysical Journal* 96: 2709-2718.

- Baskić, D., Popović, S., Ristić, P. and Arsenijević, N.N. 2006. Analysis of cycloheximide-induced apoptosis in human leukocytes: Fluorescence microscopy using annexin V/propidium iodide versus acridin orange/ethidium bromide. *Cell Biology International* 30: 924-932.
- Beere, H.M., Wolf, B.B., Cain, K., Mosser, D.D., Mahboubi, A., Tailor, P., Morimoto, R.I., Cohen, G.M. and Green, D.R. 2000. Heat-shock protein 70 inhibits apoptosis by preventing recruitment of procaspase-9 to the Apaf-1 apoptosome. *Nature Cell Biology* 2: 469-475.
- Bhuiyan, M.N.I.m Chowdhury, J.U. and Begum, J. 2009. Chemical investigation of the leaf and rhizome essential oils of *Zingiber zerumbet* (L.) Smith from Bangladesh. *Bangladesh Journal of Pharmacology*. 4: 9-12.
- Boatright, K.M. and Salvesen, G.S. 2003. Mechanisms of caspase activation. *Current Opinion in Cell Biology*. 15: 725-731.
- Bradford, M.M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 7(72): 248-254.
- Brewster, M.E. and Loftsson, T. 2007. Cyclodextrins as pharmaceutical solubilizers. *Advanced Drug Delivery Reviews*. 59: 645-666.
- Budiharjo, I., Oliver, H., Lutter, M., Luo, X. And Wang, X. 1999. Biochemical pathways of caspase activation during apoptosis. *Annual Review of Cell Developmental Biology*. 15: 269-290.
- Cal, K. and Centkowska, K. 2007. Use of cyclodextrins in topical formulations: Practical aspects. *European Journal of Pharmaceutics and Biopharmaceutics*. 68: 467-478.
- Castello, G., Scala, S., Palmieri, G., Curley, S. and Izzo, F. 2010. HCV-related hepatocellular carcinoma: From chronic inflammation to cancer. *Clinical Immunology*. 134: 237-250.
- Chane-Ming, J., Vera, R. and Chalchat, J.C. 2003. Chemical composition of the essential oil from rhizomes, leaves and flowers of *Zingiber zerumbet* smith from Reunion Island. *Journal of Essential Oil Research* 15(3): 202-205.
- Cheah, S.C., Appleton, D.R., Lee, S.T., Lam, M.L., Hadi., A.H.A. and Mustafa, M.R. 2011. Panduratin a inhibits the growth of a549 cells through induction of apoptosis and inhibition of NF-kappaB translocation. *Molecules* 16: 2583-2598.

- Chuang, S.C., Vecchia, C.L. and Boffetta, P. 2009. Liver cancer: Descriptive epidemiology and risk factors other than HBV and HCV infection. *Cancer letters*. 286:9-14.
- Ciapetti, G., Granchi, D., Savarino, L., Cenni, E., Magrini, E., Baldini, N. and Giunti, A. 2002. *In vitro* testing of the potential for orthopaedic bone cements to cause apoptosis of osteoblast-like cells. *Biomaterials* 23: 617-627.
- Cragg, G.M., Kingston, D.G.I. & Newman, D. 2005. *Anticancer agents from natural products*. USA: Taylor & Francis Group.
- Cragg, G.M., Newman, D.J. & Snader, K.M. 1997. Natural products in drug discovery and development. *Journal of Natural Products* 60: 52-60.
- Danilova, N., Sakamoto, K.M. and Lin, S. 2008. P53 family in development. *Mechanisms of Development*. 125: 919-931.
- Debatin, K.M. & Fulda, S. 2006. *Apoptosis and Cancer Therapy*. Germany: Wiley-VCH.
- Dey, A., Lane, D.P. and Verma, C.S. 2010. Modulating the p53 pathway. *Seminars in Cancer Biology*. 20: 3-9.
- Eid, E.E.M., Abdul, A.B., Sulaiman, F.E.O., Sukari, M.A. and Fatah, S.S. 2011a. Characterization of the inclusion complex of zerumbone with hydroxypropyl- β -cyclodextrin. *Carbohydrate Polymers*. 83: 1707-1714.
- Eid, E.E.M., Abdul, A.B., Rasedee, A., Suliman, F.E.O., Sukari, M.A. and Fatah, S.A. 2011b. Liquid chromatography – tandem mass spectroscopic method for the determination of zerumbone in human plasma and its application to pharmacokinetics. *Journal of Mass Spectrometry*. 46: 772-781.
- Engeland, M.V., Nieland, L.J.W.m Ramaekers, F.C.S., Schutte, B. and Reutelingsperger, C.P.M. 1998. Annexin V-Affinity Assay: A Review on an Apoptosis Detection System Based on Phosphatidylserine Exposure. *Cytometry* 31: 1-9.
- Evtodienko, Y.V., Teplova, V.V., Azarashvily, T.S., Virtanen, I. and Saris, N.E.L. 1999. Mechanisms of the resistance to the mitochondrial permeability transition in tumour cells. *Phatophysiology* 6: 171-178.
- Farnsworth, N.R., Akerele, O. & Bingel, A.S. 1985. Medicinal plants in therapy. *Bulletin of the World Organization* 63 (6): 965-981.
- Franceschi, S. and Raza, S.A. 2009. Epidemiology and prevention of hepatocellular carcinoma. *Cancer Letters*. 286: 5-8.
- Geisow, M.J., Walker, J.H., Boustead, C. And Taylor, W. 1987. Annexins—new family of Ca^{2+} -regulated- phospholipid binding protein. *Bioscience Reports* 7(4): 289-298.

- Goncalves, A., Braguer, D., Carles, G., Andre, N., Prevot, C. and Briand, C. 2000. Caspase-8 activation independent of CD95/CD95-L interaction during paclitaxel-induced apoptosis in human colon cancer cells (HT29-D4). *Biochemical Pharmacology* 60: 1579-1584.
- Gottlieb, E., Armour, S.M., Harris, M.H. and Thompson, C.B. 2003. Mitochondrial membrane potential regulates matrix configuration and cytochrome c release during apoptosis. *Cell Death and Differentiation* 10: 709-717.
- Gould, S. and Scott, R.C. 2005. 2-Hydroxypropyl- β -cyclodextrin (HP- β -CD): A toxicology review. *Food and Chemical Toxicology*. 43: 1452-1459.
- Gross, A., Jockel, J., Wei, M.C. and Korsmeyer, S.J. 1998. Enforced dimerization of BAX results in its translocation, mitochondrial dysfunction and apoptosis. *The EMBO Journal* 17(14): 3878-3885.
- Gross, A., McDonnell, J.M. and Korsmeyer, S.J. 1999. BCL-2 family members and the mitochondria in apoptosis. *Genes and Development* 13: 1899-1911.
- Haas, J.E., Muczynski, K.A., Krailo, M., Ablin, A., Land, V., Vietti, T.J. and Hammond, G.D. 1989. Histopathology and prognosis in childhood hepatoblastoma and hepatocarcinoma. *Cancer*. 64: 1082-1095.
- Haldar, S., Basu, A. and Croce, C.M. 1997. Bcl2 is the guardian of microtubule integrity. *Cancer Research* 15; 57(2): 229-233.
- Haughland, R.P. 2002. *Handbook of fluorescent probes and research products* (9th edition). USA: Molecular Probes.
- Haworth, R. and Hunter, D.R. 1979. The Ca^{2+} -induced membrane transition in mitochondria II. nature of the Ca^{2+} trigger site. *Archives of Biochemistry and Biophysics* 195(2): 460-467.
- Hunter, D.R. and Haworth, R. 1979b. The Ca^{2+} -induced membrane transition in mitochondria III. transitional Ca^{2+} release. *Archives of Biochemistry and Biophysics* 195(2): 468-477.
- Hunter, D.R. and Haworth, R. 1979a. The Ca^{2+} -induced membrane transition in mitochondria I. the protective mechanism. *Archives of Biochemistry and Biophysics* 195(2): 453-459.
- Hunter, D.R., Haworth, R. and Southard, J.H. 1976. Relationship between configuration, function, and permeability in calcium-treated mitochondria. *The Journal of Biological Chemistry* 251(16): 5069-5077.
- Ibrahim, M.Y., Abdul, A.B.H., Ibrahim, T.A.T., Abdelwahab, S.I., Elhassan, M.M. & Mohan, S. 2010. Attenuation of cisplatin-induced nephrotoxicity in rats using zerumbone. *African Journal of Biotechnology* 9(28): 4434-4441.

- Indran, I.R., Tufo, G., Prevaiz, S. and Brenner, C. 2011. Recent advances in apoptosis, mitochondria and drug resistance in cancer cells. *Biochemica et Biophysica Acta* 1807: 735-745
- Jayat, C. and Ratinaud, M.H. 1993. Cell cycle analysis by flow cytometry: principles and applications. *Biology of the Cell* 78: 15-25.
- Jemal, A., Bray, F., Center, M.M., Ferlay, J., Ward, E. and Forman, D. 2011. Global Cancer Statistics. *CA: A Cancer Journal for Clinicians*. 61: 69-90.
- Kader, G., Nikkon, F., Rashid, M.A. and Yeasmin, T. 2011. Antimicrobial activities of the rhizome extract of *Zingiber zerumbet* Linn. *Asian Pacific Journal of Tropical Biomedicine*. 409-412.
- Kastan, M.B., Onyekwere, O., Sidransky, D., Vogelstein, B. and Craig, R.W. 1991. Participation of p53 protein in the cellular response to DNA damage. *Cancer Research* (51: 6304-6311).
- Kerr, J.F.R., Wylie, A.H. & Currie A.R. 1972. Apoptosis: A basic biological phenomenon with wide ranging implications in tissue kinetics. *British Journal of Cancer* 26: 239-257.
- Kho, B.T. 2005. *Hepatotoxicity Effect of a Natural Compound, Zerumbone, Isolated from Zingiber zerumbet on Female Balb/c Strain Mice*, Unpublished Graduate Dissertation. Universiti Putra Malaysia.
- Kim, M., Miyamoto, S., Yasui, Y., Oyama, T., Murakami, A. & Tanaka, T. 2009. zerumbone, a tropical ginger sesquiterpene, inhibits colon and lung carcinogenesis in mice. *International Journal Cancer* 124: 264-271.
- Kintzios, S.E. and Barberaki, M.G. 2003. *Plant that fight cancer*. USA: CRC Press LLC.
- Kirana, C., McIntosh, G.H., Record, I.R. and Jones, G.P. 2003. Antitumor activity of extract of *Zingiber aromaticum* and its bioactive sesquiterpenoid zerumbone. *Nutrition and Cancer* 45(2): 218-225.
- Kirkin, V., Joos, S. and Zornig, M. 2004. The role of Bcl-2 family members in tumorigenesis. *Biochemica et Biophysica Acta* 1644: 229-249.
- Kitayama, T., Nagao, R., Masuda, T., Hill, R.K., Morita, M.m Takatani, M., Sawada, S. And Okamoto, T. 2002. The chemistry of Zerumbone IV Asymmetric synthesis of Zerumbol. *Journal of Molecular Catalysis B: Enzymatic*. 17: 75-79.
- Kluck, R.M., Wetzel, E.B., Green, D.R. and Newmeyer, D.D. 1997. The release of cytochrome c from mitochondria: a primary site for bcl-2 regulation of apoptosis. *Science* 275: 1132-1136.
- Koopman, G., Reutelingsperger, C.P.M., Kuijten, G.A.M., Keehnen, R.M.J., Pals, S.T. and Oers, M.H.J.V. 1994. Annexin V for flow cytometric detection

of phosphatidylserine expression on B cells undergoing apoptosis. *Blood Journal* 84:1415-1420.

- Korsmeyer, S.J., Wei, M.C., Saito, M., Weiler, S., Oh, K.J. and Schlesinger, P.H. 2000. Pro-apoptotic cascade activates BID, which oligomerizes BAK or BAX into pores that result in the release of cytochrome c. *Cell Death and Differentiation* 7: 1166-1173.
- Korsmeyer, S.J., Yin, X.M., Yang, E., Zha, J., Sedlak, T. and Oltvai, Z. 1995. BCL-2 gene family and the regulation of programmed cell death. *Cancer Genetics and Cytogenetics* 84(2): 138.
- Krajewska, M., Krajewski, S., Epstein, J.I., Shabaik, A., Sauvageot, J., Song, K., Kitada, S. and Reed, J.C. 1996. Immunohistochemical analysis of *bcl-2*, *bax*, *bcl-X* and *mcl-1* expression in prostate cancers. *American Journal of Pathology* 148(5): 1567-1576.
- Krishan, A. 1975. Rapid flow cytofluorometric analysis of mammalian cell cycle by propidium iodide staining. *The Journal of Cell biology* 66: 188-193.
- Kubbutat, M.H.G. and Vousden, K.H. 1998. Keeping an old friend under control: regulation of p53 stability. *Molecular Medicine Today*. 250-256.
- Kurokawa, M. And Kornbluth, S. 2009. Caspases and Kinases in a Death Grip. *Cell*. 138: 838-854.
- Kuwana, T. and Newmeyer D. 2003. Bcl-2-family proteins and the role of mitochondria in apoptosis. *Current Opinion in Cell Biology* 15: 691-699.
- Kylarová, D., Procházková, J., Mad'arová, J., Bartos, J. and Lichnovsk'y, V. 2002. Comparison of the TUNEL, lamin B and annexin V methods for the detection of apoptosis by flow cytometry. *Acta Histochemica* 104: 367-370.
- Lavrik, I.N., Golks, A. and Krammer, P.H. 2005. Caspases: pharmacological manipulation of cell death. *The Journal of Clinical Investigation*. 115: 2665-2672.
- Lemasters, J.J., Nieminen, A.L., Qian, T., Trost, L.C., Elmore, S.P., Nishimura, Y., Crowe, R.A., Cascio, W.E., Bradham, C.A., Brenner, D.A. and Herman, B. 1998. The mitochondrial permeability transition in cell death: a common mechanism in necrosis, apoptosis and autophagy. *Biochemica et Biophysica Acta* 1366: 177-196.
- Leong, T.Y.M. and Leong, A.S.Y. 2008. Epidemiology. In *Hepatocellular Carcinoma*, ed. Lau, W.Y., pp. 1-23. Singapore: World Scientific Publishing Co. Pte. Ltd.
- Li, H., Zhu, H., Xu, C. and Yuan, J. 1998. Cleavage of BID by caspase 8 mediates the mitochondrial damage in the Fas pathway of apoptosis. *Cell* 94: 491-501.

- Li, P., Nijhawan, D., Budihardjo, I., Srinivasula, S.M., Ahmad, M., Alnemri, E.S. and Wang, X. 1997. Cytochrome c and dATP-dependent formation of Apaf-1/caspase-9 complex initiates an apoptotic protease cascade. *Cell*. 91: 479-489.
- Lim, G.C.C. and Halimah, Y. (Eds). 2004. Second Report of the National Cancer Registry. Cancer Incidence in Malaysia 2003. National Cancer Registry. Kuala Lumpur
- Liu, C.J. and Kao, J.H. 2007. Hepatitis B virus-related hepatocellular carcinoma: epidemiology and pathogenic role of viral factors. *Journal of the Chinese Medical Association*. 70(4): 141-145.
- Loftsson, T. and Duchene, D. 2007. Cyclodextrins and their pharmaceutical applications. *International Journal of Pharmaceutics*. 329: 1-11.
- Lok, A.S.F. 2000. Hepatitis B infection: pathogenesis and management. *Journal of Hepatology*. 32: 89-97.
- Los, M., Burek, C.J., Stroh, C., Benedyk, K., Hug, H. and Mackiewicz, A. 2003. Anticancer drugs of tomorrow: apoptotic pathways as targets for drug design. *Drug Discovery Today* 8(2): 67-77.
- Los, M., Stroh, C., Janicke, R.U., Engels, I.H. and Schulze-Osthoff, K. 2001. Caspases: more than just killers? *TRENDS in Immunology*. 22(1): 31-34.
- Lowe, S.W. & Lin, A.W. 2000. Apoptosis in cancer. *Carcinogenesis* 3: 485-495.
- Luo, X., Budiharjo, I., Zou, H., Slaughter, C. And Wang, X. 1998. Bid, a Bcl2 interacting protein, mediates cytochrome c release from mitochondria in response to activation of cell surface death receptors. *Cell* 94: 481-490.
- MacFarlane, M., Merrison, W., Bratton, S.B. and Cohen, G.M. 2002. Proteasome-mediated degradation of Smac during Apoptosis: XIAP Promotes Smac ubiquitination *in vitro*. *The Journal of Biochemical Chemistry* 277; 39: 36611-36616.
- Marrero, C.R. and Marrero, J.A. 2007. Viral hepatitis and hepatocellular carcinoma. *Archives of Medical Research*. 38: 612-620.
- Marsden, V.S., O'Connor, L., O'Reilly, L.A., Silke, J., Metcalf, D., Ekert, P.G., Huang, D.C., Cecconi, F. & Tomaselli, K.J. 2002. Apoptosis initiated by Bcl-2-regulated caspase activation independently of the cytochrome c/Apaf-1/caspase-9 apoptosome. *Nature* 419: 634-637.
- Martin, S.J., Reutelingsperger, C.P.M., McGahon, A.J., Rader, J.A., Schie, R.C.A.A.V., LaFace, D.M. and Green, D.R. 1995. Early redistribution of plasma membrane phosphatidylserine is a general feature of apoptosis regardless of the initiating Stimulus: inhibition by overexpression of Bcl-2 and Abl. *The Journal of Experimental Medicine* 182: 1545-1556.

- Mascotti, K., McCullough, J. And Burge, S.R. 2000. HPC viability measurement: Trypan blue versus acridine orange and propodium iodide. *Transfusion* 40: 693-696.
- Masuda, T., Jitoe, A., Kato, S. and Nakatani, N. 1991. Acetylated flavonol glycosides from *Zingiber zerumbet*. *Phytochemistry* 30(7): 2391-2392.
- Matthes, H.W.D., Luu, B. and Ourisson, G. 1980. Cytotoxic components of *Zingiber zerumbet*, *Curcuma zedoaria* and *C. Domestica*. *Phytochemistry* 19: 2643-2650.
- Mazzanti, R., Gramantieri, L. and Bolondi, L. 2008. Hepatocellular carcinoma: Epidemiology and clinical aspects. *Molecular Aspects of Medicine*. 29: 130-143.
- McKillop, I.H., Moran, D.M., Jin, X. and Koniaris, L.G. 2006. Molecular pathogenesis of hepatocellular carcinoma. *Journal of Surgical research*. 136: 125-135.
- McLaughlin, F., Finn, P. And Thangue, N.B.L. 2003. The cell cycle, chromatin and cancer: mechanism-based therapeutics come of age. *Drug Discovery Today* 8(17): 793-802.
- Merghoub, N., Benbacer, L., Amzazi, S., Morjani, H. And Mzibri, M.E. 2009. Cytotoxic effect of some Moroccan medicinal plant extracts on human cervical cell lines. *Journal of Medicinal Plant Research* 3: 1045-1050.
- Meyers, R.L. 2007. Tumors of the liver in children. *Surgical Oncology*. 16: 195-203.
- Mikhailov, V., Mikhailova, M., Pulkrabek, D.J., Dong, Z., Venkatachalam, M.A. and Saikumar, P. 2001. Bcl-2 prevents bax oligomerization in the mitochondrial outer membrane. *The Journal of Biological Chemistry* 276(21): 18361-18374.
- Mohan, S., Abdul, A.B., Abdelwahab, S.I., Al-Zubairi, A.S., Sukari, M.A., Abdullah, R., Taha, M.M.E., Beng, N.K. and Isa, N.M. 2010. *Typhonium flagelliforme* inhibits the proliferation of murine leukemia WEHI-3 cells *in vitro* and induces apoptosis *in vivo*. *Leukemia Research* 34 (11):1483-1492.
- Mosmann, T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods* 65(1-2): 55-63.
- Murakami, A., Tanaka, T., Lee, J.Y., Surh, Y.J., Kim, H.W., Kawabata, K., Nakamura, Y., Jiwajinda, S. & Ohigashi, H. 2004. Zerumbone, a sesquiterpene in subtropical ginger, suppresses skin tumor initiation and promotion stages in ICR mice. *International Journal Cancer* 110(4): 481-490.
- Muzio, M., Salvesen, G.S. and Dixit, V.M. 1997. FLICE induced apoptosis in a cell-free system. *The Journal of Biological Chemistry* 272;5: 2952-2956.

- Nakatani, N., Jitoe, A., Masuda, T. and Yonemori, S. 1991. Flavonoid constituents of *Zingiber zerumbet* Smith. *Agricultural and Biological Chemistry* 55(2): 455-460.
- Nelson, J., Gibbons, E., Pickett, K.R., Streeter, M., Warcup, A.O., Yeung, C.H.Y., Judd, A.M. and Bell, J.D. 2011. Relationship between membrane permeability and specificity of human secretory phospholipase A2 isoforms during cell death. *Biochemica et Biophysica Acta* 1808: 1913-1920.
- Nicholson, D.W. and Thornberry, N.A. 1997. Caspases: killer proteases. *Trends in Biochemical Sciences*. 22: 299-306.
- Nunez, R. 2001. DNA measurement and cell cycle analysis by flow cytometry. *Current Issues in Molecular Biology* 3(3): 67-70.
- O'Brate, A. and Giannakakou, P. 2003. The importance of p53 location: nuclear or cytoplasmic zip code? *Drug Resistance Updates*. 6: 313-322.
- Okuda, H. 2007. Hepatocellular carcinoma development in cirrhosis. *Best Practice and Research*. 21(1): 161-173.
- Oliver, L. And Vallette, F.M. 2005. The role of caspases in cell death and differentiation. *Drug resistance Updates*. 8: 163-170.
- Pancharoen, O., Prawat, U. and Tuntiwachwuttikul, P. 2000. Phytochemistry of the Zingiberaceae. *Studies in Natural Products Chemistry*. 23: 797-865.
- Parkin, D.M., Bray, F., Ferlay, J. and Paola Pisani. 2005. Global Cancer Statistics, 2002. *CA: A Cancer Journal for Clinicians*. 55: 74-108.
- Parkin, D.M., Pisani, P. and Ferlay, J. 1999. Global cancer statistics. *A Cancer Journal for Clinicians* 49: 33-64.
- Perilongo, G. and Shafford, E.A. 1999. Paediatric update: liver tumours. *European Journal of Cancer*. 35(6): 953-959.
- Perz, J.F., Armstrong, G.L., Farrington, L.A., Hutin, Y.J.F. and Bell, B.P. 2006. The contributions of hepatitis B virus and hepatitis C virus infections to cirrhosis and primary liver cancer worldwide. *Journal of Hepatology*. 45:529-538.
- Peter, M.E. and Krammer, P.H. 2003. The CD95(APO-1/Fas) DISC and beyond. *Cella Death and Differentiation*. 10: 26-35.
- Pezzuto, J.M. 1997. Plant-derived anticancer agents. *Biochemical Pharmacology* 53: 121-133.
- Pihie, A.H.L. 1998. *Kanser payudara*. Serdang: Penerbit Universiti Putra Malaysia.
- Piret, J.P., Arnould, T., Fuks, B., Chatelain, P., Remacle, J. and Michiels, C. 2004. Mitochondria permeability transition-dependent *tert*-butyl

hydroperoxide-induced apoptosis in hepatoma HepG2 cells. *Biochemical Pharmacology* 67: 611-620.

Potten, C. & Wilson, J. 2004. *Apoptosis: The life and death of cells*. USA: Cambridge University Press.

Prakash, R.O., Rabinarayan, A. and Kumar, M.S. 2011. *Zingiber zerumbet* (L.) Sm., a reservoir plant for therapeutic uses: A Review. *International Journal of Pharma World Research*. 2(2): 1-23.

Reed, J.C., Jurgensmeier, J.M. and Matsuyama, S. 1998. Bcl-2 family proteins and mitochondria. *Biochimica et Biophysica Acta*. 1366: 127-137.

Reutelingsperger, C.P.M., Hornstra, G. and Hemker, H.C. 1985. Isolation and partial purification of a novel anticoagulant from arteries of human umbilical cord. *European Journal of Biochemistry* 151: 625-629.

Ribble, D., Goldstein, N.B., Norris, D.A. and Shellman, Y.G. 2005. A simple technique for quantifying apoptosis in 96-well plates. *BMC Biotechnology*. 5: 12.

Riccardi, C. and Nicoletti, I. 2006. Analysis of apoptosis by propidium iodide staining and flow cytometry. *Nature Protocols* 1(3): 1458-1461.

Ryan, K.M., Phillips, A.C. and Vousden, K.H. 2001. Regulation and function of the p53 tumor suppressor protein. *Current Opinion in Cell Biology*. 13: 332-337.

Sakinah, S.S.A., Handayani, S. T. and Azimahtol, L. P. 2007. Zerumbone induced apoptosis in liver cancer cells via modulation of Bax/Bcl-2 ratio. *Cancer Cell International* 7: 4.

Saravanan, B.C., Sreekumar, C., Bansal, G.C., Ray, D. Rao, J.R. and Mishra, A.K. 2003. A rapid MTT colorimetric assay to assess the proliferative index of two Indian strains of *Theileria annulata*. *Veterinary Parasitology* 113: 211-216.

Savjani, K.T., Gajjar, A.K. and Savjani, J.K. 2012. Drug solubility: importance and enhancement techniques. *International Scholarly Research Network Pharmaceutics* doi: 10.5402/2012/195727.

Shapiro, H.M. 2005. Fluorescent probes. In L.A. Sklar. *Flow Cytometry for Biotechnology* (pp. 27-28). USA: Oxford University Press.

Shield M.A. & Mirkes, P.E. 1998. Apoptosis. *Handbook of development neurotoxicology* 8: 159-188.

Shounan, Y., Feng, X. and O'Connell, P.J. 1998. Apoptosis detection by annexin V binding: a novel method for the quantitation of cell-mediated cytotoxicity. *Journal of Immunological Methods* 217: 61-70.

- Skulachev, V.P. 1996. Why are mitochondria involved in apoptosis? *FEBS Letters* 397: 7-10.
- Sprick, M.R and Walczak, H. 2004. The interplay between the Bcl-2 family and death receptor-mediated apoptosis. *Biochimica et Biophysica Acta* 1644: 125-132.
- Srinivasula, S.M., Ahmad, M., Alnemri, T.F., Litwack, G. and Alnemri E.S. 1996. Molecular ordering of the Fas-apoptotic pathway: the Fas/APO-1 protease Mch5 is a CrmA-inhibitable protease that activates multiple Ced-3/ICE-like cysteine proteases. *Biochemistry* 93: 14486-14491.
- Steinkamp, J.A., Lehnert, B.E. and Lehnert, N.M. 1999. Discrimination of damaged/dead cells by propidium iodide uptake in immunofluorescently labeled populations analyzed by phase-sensitive flow cytometry. *Journal of Immunological Method* 226: 59-70.
- Stella, V.J., Rao, V.M., Zannou, E.A. and Zia, V. 1999. Mechanisms of drug release from cyclodextrin complexes. *Advanced Drug Delivery Reviews* 36: 3-16.
- Strobel, T., Swanson, L., Korsmeyer, S. and Cannistra, S.A. 1996. BAX enhances paclitaxel-induced apoptosis through a p53-independent pathway. *Proceedings of the National Academy of Sciences* 93: 14094-14099.
- Suzuki Y., Imai, Y., Nakayama, H., Takahashi, K., Tokio, K. and Takahashi, R. 2001. A serine protease, HtrA2 is released from the mitochondria and interact with XIAP, inducing cell death. *Molecular Cell* 8: 613-621.
- Suzuki, M., Youle, R.J. and Tjandra, N. 2000. Structure of Bax: coregulation of dimer formation and intracellular localization. *Cell* 103: 645-654.
- Szejtli, J. 1998. Introduction and general overview of cyclodextrin chemistry. *Chemical Review*. 98: 1743-1753.
- Taha, M.M.E., Abdul, A.B., Abdullah, R., Ibrahim, T.A.T, Abdelwahab, S.I. & Mohan, S. 2010. Potential chemoprevention of diethylnitrosamine-initiated and 2-acetylaminofluorene-promoted hepatocarcinogenesis by zerumbone from the rhizomes of the subtropical ginger (*Zingiber zerumbet*). *Chemico Biological Interactions* 186: 295-305.
- Talanian, R.V., Quinlan, C., Trautz, S., Hackett, M.C., Mankovich, J.A., Banach, D., Ghayur, T., Brady, K.D. and Wong, W.W. 1997. Substrate specificities of caspase family proteases. *The Journal of Biological Chemistry* 272(15): 9677-9682.
- Tushar, Basak, S., Sarma, G.C. and Rangan, L. 2010. Ethnomedical uses of Zingiberaceous plants of Northeast India. *Journal of Ethnopharmacology*. 132: 286-296.

- Twentyman, P.R. and Luscombe, M. 1987. A study of some variables in a tetrazolium dye (MTT) based assay for cell growth and chemosensitivity. *British Journal of Cancer* 56: 279-285.
- United States Department of Agriculture. 2012. *Natural Resources Conservation Service*. Retrieve from <http://plants.usda.gov/java/ClassificationServlet?source=profile&symbol=ZIZE&display=31>
- Vaseva A.V. and Moll, U.M. 2009. The mitochondrial p53 pathway. *Biochimica et Biophysica Acta*. 1787: 414-420.
- Vayssade, M., Laurens, L.F., Benard, J. and Ahomadegbe, J.C. 2002. Expression of p53-family members and associated target molecules in breast cancer cell lines in response to vincristine treatment. *Biochemical Pharmacology* 63: 1609-1617.
- Vermes, I., Haanen, C., Steffens-Nekken, H. and Reutelingsperger, C. 1995. A novel assay for apoptosis Flow cytometric detection of phosphatidylserine early apoptotic cells using fluorescein labelled expression on Annexin V. *Journal of Immunological Methods* 184: 39-51.
- Villunger, A., Huang, D.C.S., Holler, N., Tschopp, J. & Strasser, A. 2000. Fas ligand-induced c-Jun kinase activation in lymphoid cells requires extensive receptor aggregation but is independent of DAXX, and Fas-mediated cell death does not involve DAXX, RIP, or RAIDD. *Journal Immunology* 165: 1337-1343.
- Wang, M., Ruan, Y., Chen, Q., Li, S., Wang, Q. and Cai, J. 2011. Curcumin induced HepG2 cell apoptosis-associated mitochondrial membrane potential and intracellular free Ca²⁺ concentration. *European Journal of Pharmacology* 650: 41-47.
- Wei, M.C., Lindsten, T., Mootha, V.K., Weiler, S., Gross, A., Ashiya, M., Thompson, C.B. and Korsmeyer, S.J. 2000. tBID, a membrane-targeted death ligand, oligomerizes BAK to release cytochrome c. *Genes and Development* 14: 2060-2071.
- Wei, Y., Fan, T. and Yu, M. 2008. Inhibitor of apoptosis proteins and apoptosis. *Acta Biochimica et Biophysica Sinica* 40(4): 278-288.
- Wesselborg, S., Engels, I.H., Rossman, E., Los, M. and Osthoff, K.S. 1999. anticancer drugs induce caspase-8/FLICE activation and apoptosis in the absence of CD95 receptor/ligand interaction. *Blood* 93: 3053-3063.
- Wyllie, A.H., Kerr, J.F. and Currie, A.R. 1980. Cell death: the significance of apoptosis. *International Review of Cytology* 68:251–306.
- Xian, M., Ito, K., Nakazato, T., Shimizu, T., Chen, C.K., Yamato, K., Murakami, A., Ohigashi, H., Ikeda, Y. and Kizaki, M. 2008. Zerumbone, a bioactive sesquiterpene, induces G2/M cell cycle arrest and apoptosis in leukemia

cells via a Fas- and mitochondria-mediated pathway. *Cancer Sciences* 98(1): 118-126.

- Yasuhara, S., Zhu, Y., Matsui, T., Tipirneni, N., Yasuhara, Y., Kaneki, M., Rosenzweig, A. and Martyn, J.A.J. 2003. Comparison of comet assay, electron microscopy, and flow cytometry for detection of apoptosis. *Journal of Histochemistry and Cytochemistry* 51: 873-885.
- Ye, N., Qin, J., Shi, W., Liu, X. and Lin, B. 2007. Cell-based high content screening using an integrated microfluidic device. *Lab on a Chip* 7: 1696-1704.
- Yob, N.J., Joffrey, S.M., Affandi, M.M.R.M.M., Teh, L.K., Salleh, M.Z. and Zakaria, Z.A. 2011. *Zingiber zerumbet* (L.) Smith: A review of its ethnomedicinal, chemical, and pharmacological uses. *Evidence-Based Complementary and Alternative Medicine*. 1-12.
- Youle, R. and Strasser, A. 2008. The BCL-2 protein family: opposing activities that mediate cell death. *Molecular Cell Biology* 9: 47-59.
- Yuan, C., Jin, Z. and Li, X. 2008. Evaluation of complex forming ability of hydroxypropyl- β -cyclodextrins. *Food Chemistry*. 106: 50-55.
- Zakaria, Y., Rahmat, A., Pihie, A.H.L., Abdullah, N.R. and Houghton, P.J. 2009. Eurycomanone induce apoptosis in HepG2 cells via up-regulation of p53. *Cancer Cell International* 9: 16.
- Zakaria, Z.A., Mohamad, A.S., Chear, C. T., Wong, Y.Y., Israf, D.A., & Sulaiman, M.R. 2010. Anti inflammatory and antinociceptive activities of *Zingiber zerumbet* methanol extract in experimental model systems. *Medical Principles and Practice* 19(4): 287-294.
- Zarrinpar, A., Kaldas, F. and Busuttill, R.W. 2011. Liver transplantation for hepatocellular carcinoma: an update. *Hepatobiliary & Pancreatic Diseases International*. 10: 234-242.
- Zhang, G., Kimijima, I., Onda, M., Kanno, M., Sato, H., Watanabe, T., Tsuchiya, A., Abe, R. and Takenoshita, S. 1999. Tamoxifen-induced apoptosis in breast cancer cells relates to down-regulation of bcl-2, but not bax and bcl-XL, without alteration of p53 protein levels. *Clinical Cancer Research* 5: 2971-2977.
- Zhou, H., Hou, Q., Chai, Y. And Hsu, Y.T. 2005. Distinct domains of Bcl-X_L are involved in Bax and Bad antagonism and in apoptosis inhibition. *Experimental Cell Research* 309: 316-328.
- Zimmermann, K.C. and Green, D.R. 2001. How cells die: apoptosis pathways. *Journal of Allergy Clinical Immunology*. 108(4): S99-S103.
- Zimmermann, K.C., Bonzon, C. and Green, D.R. 2001. The machinery of programmed cell death. *Pharmacology & Therapeutics*. 92: 57-70.