

# **UNIVERSITI PUTRA MALAYSIA**

IMPROVED BIOSYNTHESIS AND RECOVERY OF POLYHYDROXYALKANOATES FROM COMAMONAS SP. EB172

NOR ASMA BINTI AB. RAZAK

IB 2013 37



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## DOCTOR OF PHILOSOPHY

UNIVERSITI PUTRA MALAYSIA

2013



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By

# NOR ASMA BINTI AB. RAZAK

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

SEPTEMBER 2013

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### DEDICATIONS

#### PRAISE TO ALLAH

and THANK YOU to; Supporting Mum, Norhana Hanim binti Abdullah Understood Dad, Ab. Razak bin Ahmad Beloved Husband, Azhari bin Samsu Baharuddin Loving Sons and Daughter; Muhammad Akmal, Muhammad Azimi, Muhammad Akram, Muhammad Arif and Nor Arissa binti Azhari Encouraged Sisters and Brothers; Mohd Rostam, Nor Aida and Nor Aisah with family Muhammad Reduan bin Ab. Razak Muhammad Raafi bin Ab. Razak and others who contributes officially or unofficially to make this thesis become realm ALHAMDULLILAH and THANK YOU ONLY ALLAH CAN REWARDS THE GOODNESS OF ALL And Hope The Knowledge Give Benefits To All

• • •

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

## IMPROVED BIOSYNTHESIS AND RECOVERY OF POLYHYDROXYALKANOATES FROM Comamonas SP. EB172

By

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September 2013

Chairperson: Prof. Mohd Ali Hassan, Ph.D.

Institute : Bioscience

Polyhydroxyalkanoates (PHA) is a potential biodegradable polymer which can be used to replace a petrochemical synthetic polymer. PHA is synthesised when bacteria are exposed to a surplus of carbon and limited nitrogen. Under these conditions, cells are unable to grow but they do accumulate carbon-based polyesters. The objectives of this study were to improve PHA production by *Comamonas* sp. EB172 through fed-batch and repeated fed-batch, to characterise the effect of freeze-drying and oven-drying on cell and PHA and to improve recovery of PHA from *Comamonas* sp. EB172 using chemicals (solvent and sodium hydroxide) and biological (protease) methods. From this studies, *Comamonas* sp. EB172 which is novel locally isolated strain produced 6-9 g/L DCW, 77-86% PHA content, 5-12 mol% 3HV and 172-177 kDa Mw from fed-batch and 4-11 g/L DCW, 50-64% PHA content, 9-13 mol% 3HV and 832 kDa Mw from repeated fed-batch fermentation.Thus, through repeated fed-batch fermentation, high Mw could be obtained which gave new properties for different applications.

The oven-dried changed the chemical structure, developed crystal and changed the thermal properties of P(3HB-co-3HV) compared to freeze-dried P(3HB-co-3HV). Meanwhile, membrane cell of oven-dried become shrink, agglomerate to each other and become flakes compared to membrane cell of freeze-dried which become etched due to freezing and vacuum freezing. More than 90% purity of P(3HB-co-3HV) and P(3HB-co-3HHX) were obtained using acetone, chloroform, methanol and *n*-hexane as precipitate solvents. In addition, NaOH and protease from ginger, *Zingiber officinale* Roscoe were chosen as alternative recovery methods as they're environmental friendly, cheap and could be used to reduce chloroform effect to operator health and environment. Thus, the improvement on NaOH method were



done include different NaOH concentrations (0.1, 0.2, 0.3, 0.6, 0.8, 1 and 2 N), different residence time of cells (63, 87 and 111 h), initial drying cell conditions (wet broth, wet pellet, freeze-dried and oven-dried) and washing (distilled water, 20% and 100% ethanol). The protease which is highly specific to degrade cell membrane was successfully extracted and precipitated using acetone precipitation gave 256 U/mg with 60% recovery yield. The effect of cell concentration (5,10 and 20 g/L, incubation times and washing (distilled water, 20% and 100% ethanol) were done for P(3HB-co-3HV) recovery using protease. The best combination of freeze-drying cells for NaOH and protease are using 1N NaOH, incubation 1 hour with 100% cell concentration, centrifuged, washing two times with 20% ethanol and 0.02% cell concentration, incubation with 256 U/mg specific activity enzyme protease for 50 min, centrifuged and washing two times with 100% ethanol. Both methods gaves more than 90% P(3HB-co-3HV) purity and are comparable with recovery using chloroform and *n*-hexane with different fold Mw composition, thermal and physical properties. The properties of PHA are highly dependent upon their mode of fermentation and recovery techniques; hence, biodegradable polymer having a wide range of properties which can be used in different applications.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## PENINGKATAN PENGHASILAN DAN PEMULIHAN POLIHIDROKSIALKANOAT DARIPADA COMAMONAS SP. EB172

Oleh

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Polihidroksialkanoat (PHA) adalah polimer boleh urai yang berpotensi sebagai pengganti polimer petro-kimia sintetik. PHA dihasilkan apabila bakteria terdedah kepada lebihan karbon dan kekurangan nitrogen. Dalam keadaan ini, sel tidak lagi membesar tetapi mengumpul poliester berasaskan karbon. Objektif kajian ini adalah untuk membaiki penghasilan PHA oleh *Comamonas* sp. EB172 melalui penggunaan suapan berkelompok dan pengulangan suapan berkelompok, menciri kesan beku kering dan ketuhar kering kepada sel dan PHA dan membaiki pemulihan PHA daripada *Comamonas* sp. EB172 menggunakan kaedah kimia (pelarut dan sodium hidroksida) dan biologi (protease). Daripada kajian ini, *Comamonas* sp. EB172 tempatan baru yang dipencilkan menghasilkan 6-9 g/L DCW, 77-86% kandungan PHA, 5-12 mol% 3HV dan 172-177 kDa *Mw* daripada suapan berkelompok dan 4-11 g/L DCW, 50-64% kandungan PHA, 9-13 mol% 3HV dan 832 kDa *Mw* daripada pengulangan suapan berkelompok. Maka, melalui pengulangan suapan berkelompok,  $M_w$  P(3HB-ko-3HV) yang tinggi boleh didapati yang memberikan ciri baru bagi aplikasi berlainan.

C

Sel ketuhar-kering mengalami perubahan dari struktur kimia, penghasilan kristal dan perubahan ciri haba berbanding P(3HB-ko-3HV) dalam sel beku-kering P(3HB-ko-3HV). Manakala, sel membran ketuhar-kering menjadi kecut, mengumpal antara satu sama lain dan menjadi serpihan berbanding sel beku-kering yang tergores disebabkan beku dan beku vakum. Lebih kurang 90% tulen P(3HB-ko-3HV) dan P(3HB-ko-3HHX) didapati menggunakan aseton, kloroform, metanol dan n-heksana sebagai pelarut pemendak.Tambahan pula, NaOH dan protease daripada halia, Zingiber officinale Roscoe telah dipilih kerana ia mesra alam, murah dan

boleh digunakan sebagai mengurangkan kesan kloroform kepada kesihatan pengendali dan persekitaran. Jadi pembaikan pemulihan menggunakan kaedah NaOH telah dijalankan termasuk menggunakan pelbagai kepekatan (0.1, 0.2, 0.3, 0.6, 0.8, 1 and 2 N), masa kediaman sel yang berbeza (63, 87 and 111 jam), keadaan awal pengeringan sel (sup basah, pelet basah, beku-kering dan ketuhar-kering) dan cucian (air suling, 20% dan 100% etana). Protease yang sangat spesifik untuk memecahkan membran sel berjaya diekstrak dan ditulenkan menggunakan pemendakan aseton menghasilkan 256 U/mg dan 60% hasil pemulihan. Kesan kepekatan sel (5, 10 dan 20 g/L), masa eraman dan cucian (air suling, 20% ethanol dan 100% etana) dilakukan kepada pemulihan P(3HB-co-3HV) menggunakan protease. Kombinasi terbaik menggunakan sel beku-kering untuk NaOH dan protease adalah dengan menggunakan 1 N NaOH, inkubasi 1 jam dengan 100% kepekatan sel, emparan, cucian dua kali dengan 0.02% kepekatan sel, eraman dengan 256 U/mg aktiviti enzim protease khas untuk 50 minit, emparan dan cucian dua kali dengan 100% etana. Kedua-dua kaedah menmberi lebih daripada 90% P(3HB-co-3HV) tulen dan setara dengan peulihan m<mark>engg</mark>unakan kloroform and *n*-hexane dengan berlainan nisbah kandungan berat jisim, ciri haba dan fizikal. Ciri PHA adalah sangat bergantung kepada mod fermentasi dan kaedah pemulihan; jadi, polimer biourai mempuny<mark>ai pelbagai ciri</mark> yang dapat digunakan untuk aplikasi berbeza.

#### ACKNOWLEDGEMENTS

#### Alhamdullilah.

Thanks to Almighty ALLAH and his kindness, I can do and completed the Degree of Doctor of Philosophy entitled,"IMPROVED BIOSYNTHESIS AND RECOV-ERY OF POLYHYDROXYALKANOATES FROM *COMAMONAS* SP. EB172."

I would like to thank Prof. Dr. Mohd Ali bin Hassan from the Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Science, UPM as main supervisor, Prof Yoshihito Shirai from Department of Biological Functions and Engineering, Kyushu Institute of Technology, Japan, Prof. Dato' Dr Wan Md. Zin bin Wan Yunus, Department of Chemistry, Faculty of Science, UPM and Dr. Phang Lai Yee from the Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Science, UPM as committee members for pleasant collaboration, motivation, useful discussions, help and advice in this research.

Thanks and gratitude again to EB groups especially Prof Dr. Suraini Abd. Aziz, Associate Prof. Dr. Nor' Aini Abd Rahman, Dr. Tabassum Mumtaz, Dr. Norjan Yusof, Dr. Hidayah Arrifin, Dr. Kuppurchammy, Dr. Mohd Rafein Zakaria, Dr. Mitra Mohammadi, Dr. Alawi Sulaiman, Mr. Noor Azman bin Mohd Johar, Dr. Yee Liang Ngit, Dr. Mior Ahmad Khushairi Mohd Zahari, Miss Asma Hashim, Mrs. Elmy Nahida binti Othman, Mr. Zulkhairi bin Mohd Yusoff, Mr. Mohd Ridzuan bin Othman, Mrs. Zuraidah Zanirun, Dr. Saleha Shamsuddin, Miss Nazlina bt Mohd Yasin, Miss Syahinaz Shahrazi and others.

My special thanks to contribution of knowledge by Prof. Dato' Dr. Tengku Azmi Tengku Ibrahim from Department of Veterinary Preclinical Sciences, Faculty of Veterinary, UPM in microscopy, Dr. Ahmad Selamat from Department of Biometric, Faculty of Agriculture, UPM and Dr. Mohd Bakri Adam and his colleagues, Department of Mathematics, Faculty of Science, UPM for SAS, Excel and LaTeX software knowledge and Dr. Suraya Abdul Rashid from Department of Chemical and Environmental Engineering, Faculty Engineering, UPM.

C

Instrumentation and technical supports from Mr. Rosli Aslim, Mrs. Aluyah Marzuki, Mrs Renuga a/p Panjamurti and Mr. Khairul Basyar Baharudin from Department of Bioprocess Technology, Faculty of Biotechnology and Biomolecular Science, UPM, Mrs. Rusnani Amirudin, Mrs. Zaidina Mohd Daud and Mr. Mohamad Johadi Iskandar Che Jamil from Department Chemistry, Faculty Science, UPM, Mr. Zahiruddin Daud and Mr. Amer Syaifuddin from Department of Food Process and Engineering, Faculty Engineering, UPM, Mr. Rafiuz Zaman Haroun, Mrs. Zahidah Muhamed, Mrs. Faridah Akmal Ismail and Mrs. Aminah Jusoh from Microscopy Unit, Institute Bioscience, UPM, Mrs. Liyana Ithinin, Mr. Mohd Rizal Kapri and Mr. Md Sabri Mohd Yusoff from Fermentation Technology Unit,

Institute Bioscience, UPM and Mr. Ishak Mohd Yusuff from SIRIM Berhad.

Acknowledge to Institute Biosciences, SIRIM Berhad and Faculty of Biotechnology and Biomolecular Science, UPM for their conducive places, Ministry of Science Technology and Innovation (MOSTI), FELDA and National Science Foundation (NSF) for the projects funds and financial scholarships. And special thanks to Universiti Sains Malaysia (USM), SIRIM and Institute of Massachuetes (MIT) contribution of knowledge.

NOR ASMA BT AB. RAZAK, September 2013

I certify that a Thesis Examination Committee has met on 13 SEPTEMBER 2013 to conduct the final examination of NOR ASMA BINTI AB. RAZAK on her thesis entitled "IMPROVED BIOSYNTHESIS AND RECOVERY OF POLYHYDROXYALKANOATES FROM *COMAMONAS* SP. EB172." in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the **Degree of Doctor of Philosophy**.

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## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

## NOR ASMA BT AB. RAZAK

Date: 13 September 2013

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analyzed

# LIST OF ABBREVIATIONS

$\begin{array}{c} A. \ eutropha\\ ANOVA\\ AOT\\ BSA\\ CDW\\ CRD\\ Da\\ DOT\\ delta \ Hm\\ dH_2O\\ DSC\\ DTG\\ EFB\\ E. \ coli\\ EDTA\\ EtOH\\ FT-IR\\ g\\ g\\ GC\\ GhG\\ GPC\\ h\\ HDPE\\ HPLC\\ H_2SO_4\\ KOH\\ kDa\\ L\\ \end{array}$	Alcaligenes eutropha         analysis of variance         sodium 1,4-bis(2-ethylhexoxy)-1,4-dioxobutane-2-sulfonate         bovine serum albumin         cell dry weight         complete randomized design         Dalton         dissolved oxygen tension         enthalpy of fusion         double distilled water         differential scanning calorimetry         derivative thermogravimetric         empty fruit bunch         Escherichia coli         ethylenediamine tetraacetic acid         ethanol         Fourier transform infrared spectroscopy         gravity         gram         gas chromatography         greenhouse gases         gel permeation chromatography         hour         high density polyethylene         high performance liquid chromatography         acid sulphuric         kalium hydroxide         kiloDalton         liter
kDa	kiloDalton
L	liter
LDPE	low density polyethylene
LPS	lipopolysaccharides
MOSTI	Ministry of Science Technology and Innovation
mg	milligram
min	minute
ml	milliliter

mcl-PHA  $M_n$  $M_v$  $M_w$ NNaCl NaNO<sub>3</sub> NaOCl NH<sub>4</sub>Cl NH<sub>4</sub>OH  $NH_4NO_3$ NMR NPCM OD Р PDI pН PLA PHA PPP(3HB)P(3HB-co-4HB)P(3HB-co-3HV)P(3HB-co-3HHX) P(3HB-co-3HV-co-3HHX) P(3HB-co-3HV-co-3HMV) POME

medium-chain-length-polyhydroxyalkanoates average molecular number average molecular viscosity average molecular weight normality sodium chloride sodium nitrate sodium hyphochlorite ammonium chloride ammonium hydroxide ammonium nitrate nuclear magnetic resonance non polymer cell materials optical density probability polydispersity index power of hydrogen polylactic acid polyhydroxyalkanoates polypropylene poly(3-hydroxybutyrate) poly(3-hydroxybutyrateco-4-hydroxybutyrate) poly(3-hydroxybutyrateco-3-hydroxyvalerate) poly(3-hydroxybutyrate*co*-3-hydroxyhexanoate) poly(3-hydroxybutyrate*co*-3-hydroxyvalerate-*co*-3-hydroxyhexanoate) poly(3-hydroxybutyrate*co*-3-hydroxyvalerate-*co*-3-hydroxy-2-methylvalerate) palm oil mill effluent

R. eutropha	Ralstonia eutropha
scl-PHA	short-chain-length polyhydroxyalkanoates
SEM	scanning electron microscopy
sp.	species
spp.	subspecies
TEM	transmission electron microscopy
TGA	thermogravimetric analysis
$T_c$	crystallinity temperature
$T_g$	glass transition temperature
$T_m$	melting temperature
UM	Universiti Malaya
UPM	Universiti Putra Malaysia
USM	Universiti Sains Malaysia
v/v	volume per volume
w/v	weight per volume
3HB	3-hydroxybutyrate
3HV	3-hydroxyvalerate
3HMV	3-hydroxy-2-methylvalerate
3HHX	3-hydroxyhexanoate
%	percent
%	weight per 100 ml solution

C

### CHAPTER 1

### INTRODUCTION

### 1.1 Background of Study

The carbon and energy sources from wastes are regarded as promising alternatives for the production of PHA on a large scale as they are low cost and at the same time they help to reduce and solve pollution problems. The use of biowaste would help to increase more biodegradable products and clean processes with lowpollution, develop less expensive and novel products not available from petroleum source, use less expensive raw materials and reduce dependence on fossil fuels.

PHA is an excellent bioplastic option; it is a clean material alternative with no emissions of greenhouse gases, which helps in addressing the challenge of global climate changes. PHA has a broad flexibility interest because it can be completely recycled, biodegraded into carbon dioxide and water, present good mechanical resistance, hydrophobic, resistant to liquid and grease and it is a biocompatible thermoplastic which can be melted and molded. Due to its different properties, PHA has commercial potential as the sole structural material or as part of a degradable composite in various areas such as packaging industry, agriculture, medicine, foodstuff industry, chemical industry and others.

Optimisation of PHA process is needed for cost-effective production to improve production, increase the microorganism capabilities to produce PHA, ease downstream processing, reduce wastewater, lower unit production cost per kg PHA, reduce investment of equipments as well as to reduce the laborious process. The high volume and initial PHA content appears to lower the price of PHA, with less utilities and equipment capacity.

Interestingly, the production of P(3HB-co-3HV) by using mixed organic acids from anaerobic treated POME can be used chosen as an alternative biodegradable thermoplastic since it is a renewable and inexpensive carbon substrate. In addition P(3HB-co-3HV) characteristics are completely biodegradable, possessing high melting point and high tensile strength which can replace the non degradable polypropylene and polystyrene plastics.

New developments in research on biomass in Malaysia emphasised the use of palm oil mill effluent (POME) waste as a renewable resource of carbon substrate to produce the environmental-friendly plastic (Phang et al., 2003; Sim, 2003; Hassan et al., 2002, 1997). The high amount of POME waste in oil palm industry were anaerobically treated to obtain concentrated mixed organic acids for the production of P(3HB-co-3HV) (Mumtaz et al., 2008).

*R. eutropha* is an excellent producer of P(3HB), it is slow in growth due to low preference of an organic acids as carbon substrates which limits the commercial purposes. Thus, a new local strain known as *Comamonas* sp. EB172 was successfully isolated from open digester treating palm oil at Serting, Negeri Sembilan and it was capable to grow and accumulate PHA using mixed organic acids from POME (Zakaria et al., 2010a,b; Mumtaz, 2010; Mumtaz et al., 2009; Zakaria et al., 2008). The production of PHA through fed-batch fermentation gave 9.5-10 g/L CDW (cell dry weight) with 59-73% P(3HB-*co*-3HV) and 5-12 mol% 3HV were obtained by *Comamonas* sp. EB172 (Zakaria et al., 2010b; Mumtaz et al., 2010, 2009).

Separation and purification are essential to remove NPCM (non polymer cell materials) to give the high yield and purity of PHA. Thus, the development of a clean, simple and efficient process for PHA recovery with suitable characterisation are needed for end-products uses. The drying methods would effects the PHA recovery as reported that high purity and molecular weight  $(M_w)$  PHA compared to wet and dried biomass of *Alcaligenes eutrophus* (Chen et al., 2001a).

Selection of various types of acids and alkaline were done by Choi and Lee (1999) to degrade recombinant *E. coli* XL 1-Blue containing P(3HB). They found out that NaOH (sodium hydroxide) is the best choice of PHA extraction because it is cheap and environmentally friendly compare to chloroform. Thus, improvement of the P(3HB-co-3HV) recovery is necessary to increase yield and purity for further applications.

## 1.2 Problem Statement

i) Improvement of PHA production : The low cost of bioplastic requires the production of high CDW bacteria containing high PHA with mol% 3HV. It can be seen that commercial strain of *R. eutropha* ATCC 17699 is Gram negative bacteria, less resistant cell wall and not acid-tolerant. The undissociated acid will dominant, cross the cell wall, rapidly dissociate, acidify the cytoplasm and kill the cell. Thus, the alternative strain and fermentation methods would helps to increase the PHA.

ii) Characteristic of drying : The drying involved thermal changed on the physical, chemical and structures of PHA inside the cells. The detailed characteristics need to be identified to minimize the loss of PHA through drying prior storage and recovery.

iii) Improvement of recovery methods : Although, solvents are hazardous and expensive, they are the preferred conventional method since they do not destroy the morphology of PHA and eliminates endotoxin. Alternatives solvents used to extract PHA would gave various effect on characteristics of PHA. Thus, an improvement of NaOH treatment was needed for recovering the intracellular PHA.



Mitra studied the effects of low cell concentration and temperature for PHA recovery from similar strain but different method to produce PHA, which affect the initial PHA content,  $M_w$  and 3HV content. Meanwhile, the biological treatment such as enzymatic hydrolysis is able to give high purity, recovery yield and mild effect on the PHA. However, an alternative enzymes source is needed as the commercial enzymes are expensive which increases the overall PHA cost.

iii) High polymer quality : The production and recovery processes of PHA need to be optimised, thus giving different characteristics and quality of PHA. The usage of high technology instruments in this research are important and allowed more detailed information regarding PHA as a future bioplastic material.

#### 1.3 Strategies

i) Improvement of PHA production : The improvement of the fed-batch fermentation is necessary, to increase the uptake of organic acids in the bioreactor by provided the suitable conditions. *Commonas* sp. EB172, a new novel locally isolated strain was used in a fed-batch and repeated fed-batch to produce P(3HB-co-3HV) utilising synthetic organic acids which mimics the bioacids from POME waste. The *Commonas* sp. EB172 was chosen instead because it prefer organic acids as carbon sources compared to *R. eutropha*, a commercial producer of PHA.

The usage of synthethic organic acids is necessary to improved the fermentation and to avoid any inhibition of cell growth and less PHA formation due to substrate. Then, the P(3HB-co-3HV) obtained from different modes of fermentation were characterised.

ii) Characteristic of drying : Initially, characteristic effect of freeze-drying and oven-drying treatments on the cells and PHA were done. The optimum drying method is needed to reduce the changes on the characteristics of PHA prior to storage or after recovery.

iii) Improvement of recovery methods : An improved, simpler and environmentally friendly way of recovering intracellular PHA is needed. Selection of different type of precipitant solvents and chloroform and characterisation were done to extract P(3HB-co-3HV) and P(3HB-co-3HHX) from *Comamonas* sp. EB172 and *R. eutropha* PHB-4/pBBREE32d13.

The recovery methods involved the high cell concentration and temperature by using high concentration of NaOH. The improvement on NaOH method include different NaOH concentrations (0.1, 0.2, 0.3, 0.6, 0.8, 1 and 2 N), different residence time of cells (63, 87 and 111 h), initial drying cell conditions (wet broth, wet pellet, freeze-dried and oven-dried) and washings (distilled water, 20% and 100% ethanol) were done and characterised. Protease from ginger, *Zingiber offic*-

*inale* were extracted and partial purified using acetone. Then, the effects of cell concentrations, incubation times and washings for the P(3HB-co-3HV) recovery using protease were done.

iii) High polymer quality : The quality of PHA obtained after production and recovery processes needs to be characterise for further applications. The instruments used in this study are DTG (Derivative Thermogravimetric) and TGA (Thermogravimetry Analysis), DSC (Differential Scanning Calorimeter), GC (Gas Chromatography), GPC (Gel Permeation Chromatography), HPLC (High Performance Liquid Chromatography), NMR (Nuclear Magnetic Resonance), SEM (Scanning Electron Microscopy) and TEM (Transmission Electron Microscopy).

### 1.4 Objectives of Study

The aims of the study are:

1. to improve the PHA production by using *Comamonas* sp. EB172 through fedbatch and repeated fed-batch;

2. to characterise the effect of freeze-drying and oven-drying on cell and PHA; and

3. to improve recovery of PHA from *Comamonas* sp. EB172 using chemicals (solvent and sodium hydroxide) and biological (protease) methods.

## 1.5 Organization of Thesis

To achieve all the objectives in this study, the work presented in this thesis has been divided into eight chapters as follows:

The first chapter includes introduction, problem statement, strategies and objectives related to this study.

The aim to study in greater details about PHA, the role and benefits of PHA have been outlined in the second chapter. Different types of PHA, classification, physiology and biochemistry, comparison of PHA biosynthesis using various kinds of waste as carbon substrate, types of PHA producer and fermentation process. In addition, several extraction and factors influencing the extraction of PHA from the cell are reviewed in this section.

The third chapter includes the experimental materials, design and procedures that were carried out in this research work.

In Chapter 4, description on locally isolated strain, Comamonas sp. EB172 was used to produce P(3HB-co-3HV) using mixed synthetic organic acids through fed-

batch and repeated fed-batch fermentation were demonstrated.

In the fifth chapter, the effect of oven-drying and freeze-drying on the chemical, structural and thermal characteristic of P(3HB-co-3HV) were investigated prior PHA recovery.

Chapter 6 describes the extraction methods of P(3HB-co-3HV) recovery. The conventional methods using chloroform and effect of different precipitation chemicals were studied. NaOH was used as alternative methods, whereby the effects of NaOH concentrations, residence times, initial drying cell conditions and washings on properties of P(3HB-co-3HV) recovery were studied. Protease from ginger, *Zingiber officinale* was extract and partially purified and then, was used for P(3HB-co-3HV) recovery.

And finally, in chapter 7 contains the main conclusions as well as recommendations for further works are suggested.

#### BIBLIOGRAPHY

- Abdelwahed, W., Degobert, G., Stainmesse, S. and Fessi, H. 2006. Freeze-drying of nanoparticles: Formulation, process and storage considerations. Advanced Drug Delivery Reviews 58: 1688–1713.
- Akiyama, M., Tsuge, T. and Doi, Y. 2003. Environmental life cycle comparison of Polyhydroxyalkanoates produced from renewable carbon resources by bacterial fermentation. *Polymer Degradation and Stability* 80: 183–194.
- Alias, Z. and Tan, I. K. P. 2005. Isolation of palm oil-utilising, Polyhydroxyalkanoate (PHA)-producing bacteria by an enrichment technique. *Bioresource Tech*nology 96: 1229–1234.
- Allen, A. D., Anderson, W. A., Ayorinde, F. O. and Eribo, B. E. 2010. Biosynthesis and characterization of copolymer Poly(3HB-co-3HV) from saponified Jatropha curcasoil by Pseudomonas oleovorans. Journal Industrial Microbiology Biotechnology 37: 849–856.
- Arun, A., Murrugappan, R., Ravindran, A. D. D., Veeramanikandan, V. and Balaji, S. 2006. Utilization of various industrial wastes for the production of Polyhydroxybutyrate (PHB) by Alcaligenes eutrophus. Journal of Biotechnology 5 (17): 1524–1527.
- Avella, M., Malinconico, M. and Orlando, P. 2004. Chemistry and biology of Polymer Degradation. In *Low Environmental Impact Polymers* (eds. T. Nick and J. Mark). Rapra Technology.
- Barnard, G. N. and Sanders, J. K. M. 1989. The Poly-beta-hydroxybutyrate granule *in vivo*: A new insight based on NMR spectroscopy of whole cells. *Journal* of *Biological Chemistry* 264 (6): 3286–3291.
- Bassetti, L. and Johannes, T. 1994. Organic solvent toxicity in *Morinda citrifolia* cell suspensions. *Enzyme and Microbial Technology* 16: 642–648.
- Bengtsson, S., W. A. C. M. and Welander, T. 2008. Production of Polyhydroxyalkanoates by activated sludge treating a paper mill wastewater. *Bioresource Technology* 99: 509–516.
- Bhubalan, K., Lee, W.-H., Loo, C.-Y., Yamamoto, T., Tsuge, T., Doi, Y. and Sudesh, K. 2008. Controlled biosynthesis and characterization of Poly(3-hydroxybutyrae-co-3-hydroxyvalerate-co-3-hydroxyhexanoate) from mixtures of palm kernel oil and 3-HV precursors. *Polymer Degradation and Stability* 93: 17–23.
- Bormann, E. J., Leibner, M., Roth, M., Beer, B. and Metzner, K. 1998. Production of Polyhydroxybutyrate by *Ralstonia eutropha* from protein hydrolysate. *Applied Microbiology and Biotechnology* 50: 604–607.

- Braunegg, G., Sonnleitner, B. and Lafferty, R. M. 1978. A rapid gas chromatographic method for the determination of Poly-3-hydroxybutyric acid in microbial biomass. *European Journal Applied Microbiology and Biotechnology* 6: 29–37.
- Carraher, C. E. J. 2008. Seymour/Carraher's polymer chemistry. 7th edn. USA: CRC Press Taylor and Francis Group.
- Castilho, L. R., Mitchell, D. A. and Freire, D. M. G. 2009. Production of Polyhydroxyalkanoates (PHA) from waste materials and by-products by submerged and solid-state fermentation. *Bioresource Technology* 100: 5996–6009.
- Chakraborty, P., Gibbons, W. and Muthukumarappan, K. 2009. Conversion of volatile fatty acids into Polyhydroxyalkanoate by *Ralstonia eutropha*. Journal of Applied Microbiology 106: 1996–2005.
- Chen, Y., Chen, J., Yu, C., Du, G. and Lun, S. 1999. Recovery of Poly-3hydroxybutyrate from *Alcaligenes eutrophus* by surfactant-chelate aqueous system. *Process Biochemistry* 34: 153–157.
- Chen, Y., Xu, Q., Yang, H. and Gu, G. 2001a. Effects of cell fermentation time and biomass drying strategies on the recovery of Poly-3-hydroxyalkanoates from *Alcaligenes eutrophus* using a surfactant-chelate aqueous system. *Process Biochemistry* 36: 773–779.
- Chen, Y., Yang, H., Zhou, Q., Chen, J. and Gu, G. 2001b. Cleaner recovery of Poly(3-hydroxybutyric acid) synthesized in *Alcaligenes eutrophus*. *Process Biochemistry* 36: 501–506.
- Choi, G. G., Kim, H. W., Kim, Y. B. and Rhee, Y. H. 2005. Biocompatibility of Poly(3-hydroxybutyrate-co-3-hydroxyvalerate) copolyesters produced by Alcaligenes sp. Biotechnology and Bioprocess Engineering 10: 540–545.
- Choi, G. G., Kim, M. W., Kim, J.-Y. and Rhee, Y. H. 2003. Production of Poly(3-hydroxybutyrate-*co*-3-hydroxyvalerate) with high molar fractions of 3-hydroxyvalerate by a threenine-overproducing mutant of *Alcaligenes* sp. SH-69. *Biotechnology Letters* 25: 665–670.
- Choi, J.-i. and Lee, S. Y. 1999. Efficient and economical recovery of Poly(3hydroxybutyrate) from *recombinant Escherichia coli* by simple digestion with chemicals. *Biotechnology and Bioengineering* 62 (5): 546–553.
- Chung, Y. J., Cha, H. J., Yeo, J. S. and Yoo, Y. J. 1997. Production of Poly(3hydroxybutyric-co-3-hydroxyvaleric) acid using propionic acid by pH regulation. Journal of Fermentation and Bioengineering 83 (5): 492–495.
- Dai, Y., Yuan, Z., Jack, K. and Keller, J. 2007. Production of targeted Poly(3hydroxyalkanaotes) copolymers by glycogen accumulating organisms using acetate as sole carbon source. *Journal of Biotechnology* 129 (129): 489–497.

- de Koning, G. J. M., Kellerhals, M., van Meurs, C. and Witholt, B. 1997. A process for the recovery of Polyhydroxyalkanoates) from *Pseudomonads*. Part 2 : Process development and economic evaluation. *Bioprocess Engineering* 17: 15–21.
- Dhariwal, A., Mohrdieck, M., Leitermann, F., Arjol, I. M., Manresa, A., Syldatk, C., Janke, H. D. and Chmiel, H. 2008. Effect of emulsified feeding of oily substrate via submerged ceramic membranes on surfactant production in *Pseudomonas aeruginosa* fermentation. *Bioprocess Biosystem Engineering* 31: 401–409.
- Doi, Y. 1990. Microbial polyester: Economic aspects of biopolymer production. In Biotechnology of Biopolymers. From synthesis to patents (eds. S. A and D. Y), 1107–1136. New York: EWiley-Vch. Verlag GmbH and Co. KGaA, Weinheim,.
- Du, G., Chen, L. X. L. and Jian, Y. J. 2004. High-efficiency production of bioplastics from biodegradable organic solids. *Journals of Polymers and the Environment* 12 (2): 89–94.
- Furrer, P., Sven, P. and Manfred, Z. 2007. Efficient recovery of low endotoxin medium-chain-length Poly([R]-3-hydroxyalkanoate) from bacterial biomass. *Journal of Microbiological Methods* 69: 206–213.
- Galaev, I. and Mattiasson, B. 2008. Smart polymer : Applications in Biotechnology and Biomedicine. In *Microgels from smart polymers* (eds. N. Kausar, B. Z. Chowdry, and M. J. Snowden), 1–472. Taylor and Francis Group.
- Galan, B., Dinjaski, N., Maestro, B., de Eugenio, L. I., Escapa, I. F., Sanz, J. M., Garcia, J. L. and Prieto, M. A. 2011. Nucleoid-associated PhaF Phasin drivers intracellular location and segregation of Polyhydroxyalkanoate granules in *Pseu*domonas putida KT2442. Molecular Microbiology 79 (2): 402–418.
- Gayet, J. C. and Masaro, L. 2004. Polyhydroxyalkanoates : The next generation of bioplastics. In *Chemistry and biology of polymer degradation* (eds. T. Nick and J. Mark), *Low Environmental Impact Polymers*, 239–248. Rapra Technology.
- Gerald, S. 1999. *Polymers and the environment*. Cambridge : Royal Society of Chemistry.
- Goncalves, S. P. C., Martins-Franchetti, S. M. and Chinaglia, D. L. 2009. Biodegradation of the films of PP, PHBV and its blends in soil. *Journal Polymer Envi*ronment 17: 280–285.
- Gursel, I., Balcik, C., Arica, Y., Akkus, O., Akkas, N. and Hasirci, V. 1998. Synthesis and mechanical properties of inter-penetrating networks of Polyhydroxybutyrate-*co*-hydroxyvalerate and Polyhydroxyethyl methacrylate. *Biomaterials* 19: 1137–1143.

- Ha, C.-S. and Cho, W.-J. 2002. Miscibility, properties and biodegradability of microbial polyester containing blends. *Progress in Polymer Science* 27: 759– 809.
- Hahn, S. K., Chang, Y. K., Kim, B. S. and Chang, H. N. 1994. Optimization of microbial Poly(3-hydroxybutyrate) recover using dispersions of sodium hypochlorite solution and chloroform. *Biotechnology and Bioengineering* 44 (2): 256–261.
- Hahn, S. K., Chang, Y. K. and Lee, S. Y. 1995. Recovery and characterisation of Poly (3-hydroxybutyrate) synthesized in *Alcaligenes eutrophus* and recombinant *E. coli*. *Applied and Environmental Microbiology* 61: 34–39.
- Hashim, M. M., Mingsheng, D., Iqbal, M. F. and Xiaohong, C. 2011. Ginger rhizome as potential source of milk coagulating cysteine protease. *Phytochemistry* 72: 458–464.
- Hassan, M. A., Nawata, O., Yoshihito, S., Nor Aini, A. R., Phang, L. Y., Ariff, A. B. and Abdul Karim, M. I. 2002. A proposal for zero emission from palm oil industry incorporating the production of Polyhydroxyalkanoates from palm oil mill effluent. *Journal Chemistry Engineering Japan* 35: 9–14.
- Hassan, M. A., Shirai, Y., Kusubayashi, N., Abdul Karim, M. I., Nakanishi, K. and Hashimoto, K. 1996. Effect of organic acid profiles during anaerobic treatment of palm oil mill effluent on the production of Polyhydroxyalkanoates by *Rhodobacter sphaeroides. Journal of Fermentation and Bioengineering* 82 (2): 151–156.
- Hassan, M. A., Yoshihito, S., Umeki, H., Yamazumi, H., Jin, S., Yamamoto, S., Abdul Karim, M. I., Nakanishi, K. and Hashimoto, K. 1997. Acetic acid separation from anaerobically treated palm oil mill effluent by ion exchange resins for the production of Polyhydroxyalkanoate by *Alcaligenes eutrophus.*. *Bioscience Biotechnology Biochemistry* 61 (9): 1465–1468.
- Huang, T.-Y., Duan, K.-J., Huang, S.-Y. and Chen, C. W. 2006a. Production of Polyhydroxyalkanoates from inexpensive extruded rice bran and starch by *Haloferaz mediterranei. Journal Indian Microbiology Biotechnology* 33: 701–706.
- Jacquel, N., Lo, C. W., Wei, Y. H., Wu, H. S. and Wang, S. S. 2007. Solubility of Polyhydroxyalkanoates by experiment and thermodynamic correlations. *AIChE Journal* 53: 2704–2714.
- Jacquel, N., Lo, C. W., Wei, Y.-H., Wu, H.-S. and Wang, S. S. 2008. Isolation and purification of bacterial Poly(3-Hydroxyalkanoates. *Biochemical Engineer*ing Journal 39: 15–27.
- Jiang, X., Ramsay, J. A. and Ramsay, B. A. 2006. Acetone extraction of Mcl-PHA from *Pseudomonas putida KT2440*. Journal of Microbiological Methods 67: 212–219.

- Jung, K., Hany, R., Rentsch, D., Storni, T., Egli, T. and Witholt, B. 2000. Characterization of new bacterial copolyesters containing 3-hydroxyalkanoates and Acetoxy-3-hydroxyalkanoates. *Macromolecules* 33: 8571–8575.
- Kai, Z., Ying, D. and Guo-Qiang, C. 2003. Effects of surface morphology on the biocompatibility of Polyhydroxyalkanoates. *Biochemical Engineering Journal* 16: 115–123.
- Kasuya, K.-I., Inoue, Y., Tanaka, T., Akehata, T., Iwata, T., Fukui, T. and Doi, Y. 1997. Biochemical and molecular characterization of the Polyhydroxybutyrate depolymerase of *Comamonas acidovorans* YM1609, isolated from fresh water. *Applied and Environmental Microbiology*. 63 (12): 4844–4852.
- Kathiraser, Y., Aroua, M. K., Ramachandran, K. B. and Tan, I. K. P. 2007. Chemical characterization of medium-chain-length Polyhydroxyalkanoates (PHA) recovered by enzymatic treatment and ultrafiltration. *Journal of Chemical Tech*nology and Biotechnology 82: 847–855.
- Keenan, T. M., Nakas, J. P. and Tanenbaum, S. 2006. Polyhydroxyalkanoate copolymers from forest biomass. *Journal of Industrial Microbiology and Biotechnology* 33: 616–626.
- Kek, Y.-K., Lee, W.-H. and Sudesh, K. 2008. Efficient bioconversion of palm acid oil and palm kernel acid oil to Poly(3-hydroxybutyrate) by *Cupriviadus necator*. *Canadian Journal Chemistry* 86: 535–539.
- Khanna, S. and Srivastava, A. K. 2005. A simple structured mathematical model for Biopolymer (PHB) production. *Biotechnology Progress* 21: 830–838.
- Khosravi-Darani, K., Vasheghani-Farahani, E., Shojaosadati, S. A. and Yamini, Y. 2004. Effect of process variables on supercritical fluid disruption of *Ralstonia eutropha* cells for Poly(R-hydroxybutyrate) recovery. *Biotechnology Progress* 20: 1757–1765.
- Kim, H.-Y., Park, J.-S., Shin, H.-D. and Lee, Y.-H. 1995. Isolation of glucose utilizing mutant of *Alcaligenes eutrophus*, its substrate selectivity, and accumulation of Poly-b-hydroxybutyrate. *Journal of Microbiology* 51–58.
- Kim, S. and Dale, B. E. 2005. Life cycle assessment study of biopolymers (Polyhydroxyalkanoates) derived from no-tilled corn. *International Journal Life Cycle* Assessment 10 (3): 200–210.
- Koller, M., Bona, R., Braunegg, G., Hermann, C., Horvat, P., Kroutil, M., Martinz, J., Neto, J., Pereira, L. and Varila, P. 2005. Production of Polyhydroxyalkanoates from agricultural waste and surplus materials. *Biomacromolecules* 6: 561–565.
- Kumar, S., Mudliar, S. N., Reddy, K. M. K. and Chakrabarti, T. 2004. Production of biodegradable plastics from activated sludge generated from a food processing industrial wastewater treatment plant. *Bioresource Technology* 95: 327–330.

- Kuusipalo, J. 2000a. PHB/V in extrusion coating of paper and paperboard : Study of functional properties. Part 1. *Journal of Polymers and the Environment* 8 (1): 39–47.
- Kuusipalo, J. 2000b. PHB/V in extrusion coating of paper and paperboard : Study of functional properties. Part 2. *Journal of Polymers and the Environment* 8 (2): 49–57.
- Lageveen, R. G., Huisman, G. W., Preusting, H., Ketelaar, P., Eggink, G. and Witholt, B. 1988. Formation of polyesters by *Pseudomonas oleovorans* : Effect of substrates on formation and composition of Poly-R-3-hydroxyalkanoates and Poly-3-hydroxyalkanoates. *Applied and Environmental Microbiology* 54 (12): 2924–2932.
- Lakshman and Shamala. 2006. Extraction of Polyhydroxyalkanoate from *Sinorhi*zobium meliloti cells using *Microbispora* sp. culture and its enzyme. *Enzyme and Microbial Technology* 39 (39): 1471–1475.
- Lawrence, A. G., Schoenheit, J., He, A., Tian, J., Liu, P., Stubbe, J. and Sinskey, A. J. 2005. Transcriptional analysis of *Ralstonia eutropha* genes related to Polyhydroxybutyrate homeostasis during batch fermentation. *Applied Microbiology* and Biotechnology 68: 663–672.
- Lee, K.-M. and Gilmore, D. F. 2005. Formulation and process modeling of biopolymer (Polyhydroxyalkanoates : PHA) production from industrial wastes by novel crossed experimental design. *Process Biochemistry* 40: 229–246.
- Lee, S. and Yu, J. 1997. Production of biodegradable thermoplastics from municipal by a two-stage bioprocess. *Resources, Conservation and Recycling* 19: 151–164.
- Lee, S. Y. and Chang, H. N. 1995. Production of Poly (hydroxyalkanoic acid). Advanced Biochemistry Engineering Biotechnology 52: 27–58.
- Lee, S. Y., Park, S. J., Park, J. P., Lee, Y. and Lee, S. H. 2005. Metabolic flux analysis on the production of Poly(3-hydroxybutyrate). In *Biotechnology of biopolymers. From synthesis to patents* (eds. A. Steinbuchel and Y. Doi), 193–206. EWiley-Vch.Verlag GmbH and Co. KGaA, Weinheim.
- Lee, W.-H., Loo, C.-Y., Nomura, Christopher, T. and Sudesh, K. 2008. Biosynthesis of Polyhydroxyalkanoates copolymers from mixtures of plant oils and 3-hydroxyvalerate precursors. *Bioresource Technology* 99: 6844–6851.
- Lee, W.-K., Ryou, J.-h. and Ha, C.-S. 2003. Retardation of enzymatic degradation of microbial polyesters using surface chemistry : Effect of addition of non-degradable polymers. *Surface Science* 542: 235–243.
- Leonard, Y. M. 2004. Synthesis of polymers from sustainable resource origin raw materials. In *Low Environmental Impact Polymers* (eds. T. Nick and J. Mark), *Low Environmental Impact Polymers*, 2. Rapra Technology.

Leung, W. W. F. 2007. Centrifugal separations in Biotechnology. Academic Press.

- Liu, W.-T., Hanada, S., Marsh, Terrence, L., Kamagata, Y. and Nakamura, K. 2002. Kineosphaera limosa gen. nov., a novel Gram-positive Polyhydroxyalkanoate accumulating coccus isolated from activated sludge. International Journal of Systematic and Evolutionary Microbiology 52: 1845–1849.
- Lo, C. W., Wu, H. S. and We, Y. H. 2011. High throughput study of separation of Poly(3-hydroxybutyrate) from recombinant *Escherichia coli* XL1 blue. *Journal of the Taiwan Institute of Chemical Engineers* 42: 240–246.
- Loo, C.-Y., Lee, Wing-Hin, T. T., Doi, Y. and Sudesh, K. 2005. Biosynthesis and characterization of Poly3-hydroxybutyrate-*co*-3-hydroxybexanoate from palm oil products in a *Wautersia eutropha* mutant. *Biotechnology Letters* 27: 1405–1410.
- Loo, C. Y. and Sudesh, K. 2007. Polyhydroxyalkanoates: Bio-based microbial plastics and their properties. *Malaysian Polymer Journal* 2: 31–57.
- Luengo, J. M., Garcia, B., Sandoval, A., Naharro, G. and Olivera, E. R. 2003. Bioplastics from microorganisms. *Current Opinion in Microbiology* 6: 251–260.
- Luo, R., Chen, J., Zhang, L. and Chen, G. 2006. Polyhydroxyalkanoate copolyesters produced by *Ralstonia eutropha* PHB<sup>-4</sup> harboring a low-substrate-specificity PHA synthase PhaC2ps from *Pseudomonas stutzeri* 1317. *Biochemical Engineering Journal* 32 (32): 218–225.
- Madison, L. L. and Huisman, G. W. 1999. Metabolic engineering of Poly(3-Hydroxyalkanoatyes) : From DNA to plastic. *Microbiology and Molecular Biology Review* 63 (1): 21–53.
- Mengmeng, C., Hong, C., Qingliang, Z., Shirley, S.-N. and Jie, R. 2009. Optimal production of Polyhydroxyalkanoates (PHA) in activated sludge fed by volatile fatty acids (VFAs) generated from alkaline excess sludge fermentation. *Biore*source Technology 100: 1399–1405.
- Misra, S. K., Valappil, S. P., Roy, I. and Boccaccini, A. R. 2006. Polyhydroxyalkanoates (PHA)/inorganic phase composites for tissue engineering applications. *Biomacromolecules* 7 (8): 2249–2257.
- Mitra, M. 2011. Alternative recovery methods of intracellular Polyhydroxyalkanoates from local isolate Comamonas sp. EB172. PhD thesis, Faculty of Biotechnology and Biomolecular Science. Universiti Putra Malaysia.
- Mittendorf, V., Robertson, Elizabeth, J., Leech, R. M., Kruger, N., Steinbuchel, A. and Poirier, Y. 1998. Synthesis of medium-chain length Polyhydroxyalkanoates in Arabidopsis thaliana using intermediates of peroxisomal fatty acid betaoxidation. In Proceedings National Academy of Science, 13397–13402. File1-27,6-40.

- Mostert, M. E., Botha, B. M., Du Plessis, L. M. and Duodu, K. G. 2007. Effect of fruit ripeness and method of fruit drying on the extractability of Avocado oil with hexane and supercritical carbon dioxide. *Journal of the Science of Food* and Agriculture 87: 2880–2885.
- Mumtaz, T. 2010. Production of P(3-hydoxybutyrate-co-3-hydroxyvalerate) using Comamonas sp. EB172 from organic acids derived from anaerobic treatment of palm oil mill effluent. PhD thesis, Faculty of Biotechnology and Biomolecular Sciences. Universiti Putra Malaysia.
- Mumtaz, T., Abd-Aziz, S., Abdul Rahman, N. A., Phang, L. Y., Shirai, Y. and Hassan, M. A. 2008. Pilot-scale recovery of low molecular weight organic acids from anaerobically treated palm oil mill effluent (POME) with energy integrated system. *African Journal of Biotechnology* 7 (21): 3900–3905.
- Mumtaz, T., Abd-Aziz, S., Abdul Rahman, N. A., Phang, L. Y., Shirai, Y. and Hassan, M. A. 2009. Fed-batch production of P(3HB-co-3HV) copolymer by Comamonas sp. EB172 using mixed organic acids under dual nutrient limitation. European Journal of Scientific Research 33 (3): 374–384.
- Mumtaz, T., Abd-Aziz, S., Phang, L. Y., Wan Yunus, W. Z., Shirai, Y. and Hassan, M. A. 2010. Synthesis, characterization and structural properties of intracellular copolyester Poly(3-hydroxybutyrate-co-3-hydroxyvalerate produced by Comamonas sp. EB172 from renewable resource. International Journal of Polymer and Analytical Characteristics 15: 329–340.
- Nafi, A., Foo, H. L., Jamilah, B. and Ghazali, H. M. 2013. Properties of proteolytic enzyme from ginger (*Zingiber officinale* Roscoe. *International Food Research Journal* 20 (1): 363–368.
- Nath, A., Dixit, M., Bandiya, A., Chavda, S. and Desai, A. J. 2008. Enhanced PHB production and scale-up studies using cheese whey in fed-batch culture of *Methylobacterium* sp. ZP24. *Bioresource Technology* 99: 5749–5755.
- Neumann, L., Spinozzi, F., Sinibaldi, R., Rustichelli, F., Potter, M. and Steinbuchel, A. 2008. Binding of the major Phasin, PhaP1 from *Ralstonia eutropha* H16 to Poly(3Hydroxybutyrate) granules. *Journal of Bacteriology* 190 (8): 2911– 2919.
- Nomura, C. T., Taguchi, K., Gan, Z., Kuwabara, K., Tanaka, T., Takase, K. and Doi, Y. 2005. Expression of 3-Ketoacyl carrier protein reductase (fabG) genes enhances production of Polyhydroxyalkanoate copolymer from glucose in *recombinant Escheria coli* JM109. Applied and Environmental Microbiology 71 (8): 4297–4306.
- Nubia, M., Ivonne, G., Dionisio, M., Victoria, G., Dolly, R., Diego, S., Juan, G., Fabio, A., Armando, E. and Dolly, M. 2007. Bioprospecting and characterization of Polyhydroxyalkanoate (PHAs) producing bacteria isolated from Colombian sugarcane producing areas. *African Journal of Biotechnology* 6 (13): 1536–1543.

- Ojumu, T. V., Yu, J. and Solomon, B. O. 2004. A review: Production of Polyhydroxyalkanoates, A bacterial biodegradable polymer. *Journal of Biotechnology* 3 (1): 18–24.
- Oliveira, F. C., Dias, Macros, L., Castilho, Leda, R. and Freire, D. M. G. 2007. Characterization of Poly(3-hydroxybutyrate) produced by *Cupriavidus necator* in solid-state fermentation. *Bioresource Technology* 98: 633–638.
- Ong, L., Dagastine, R. R., Kentish, S. E. and Gras, S. L. 2011. Microstructure of milk gel and cheese curd observed using Cryo Scanning Electron Microscopy and confocal microscopy. LWT - Food Science and Technology 44: 1291–1302.
- Osborne, S. J., Leaver, J.andTurner, M. K. and Dunnill, P. 1990. Correlation of biocatalytic activity in an organic-aqueous two-liquid phase system with solvent concentration in the cell membrane. *Enzyme and Microbial Technology* 12: 281– 291.
- Page, W. J., Manchak, J. and Rudy, B. 1992. Formation of Poly(hydroxybutyrateco-hydroxyvalerate) by Azobacter vinelandii UWD. Applied and Environmental Microbiology 58 (9): 2866–2873.
- Patel, M., Gapes, D. J., Newman, R. H. and Dare, P. H. 2009. Physico-chemical properties of Polyhydroxyalkanoate produced by mixed-culture nitrogen-fixing bacteria. Applied Microbiology and Biotechnology 82: 545–555.
- Peters, M. S. and Timmerhaus, Klaus, D. 2002. *Plant design and economics for chemical engineers*. 4th edn. McGraw-Hill International, New York.
- Phang, L. Y., Hassan, M. A., Yoshihito, S., Wakisaka, M. and Abdul Karim, M. I. 2003. Continuous production of organic acids from palm oil mill effluent with sludge recycle by the freezing-thawing method. *Journal Chemistry Engineering*. *Japan* 36: 707–710.
- Philip, S., Sengupta, S., Keshavarz, T. and Roy, I. 2009. Effect of impeller speed and pH on the production of Poly(3-hydroxybutyrate) using *Bacillus cereus* SPV. *Biomacromolecules* 10: 691–699.
- Platt, D. K. 2006. *Biodegradable polymer : Market report*. Smithers Rapra Limited, United Kingdom.
- Poirier, Y., Nawrath, C. and Somerville, C. 1995. Production of Polyhydroxyalkanoates. A Family of biodegradable plastics and elastomers in bacteria and plants. *Biotechnology* 13: 142–150.
- Pouton, C. W. and Akhtar, S. 1996. Biosynthethic Polyhydroxyalkanoates and their potential in drug delivery. Advanced Drug Delivery Reviews 18: 133–162.
- Ramsay, B. A., Lomaliza, K., Chavarie, C., Dube, B., Bataille, P. and Ramsay, J. A. 1990. Production of Poly(beta-hydroxybutyric-co-beta-hydroxyvaleric) acids. Applied and Environmental Microbiology 56 (7): 2093–2098.

- Ramsay, B. A., Znoj, G. M. and Cooper, D. G. 1986. Use of a nylon manufacturing waste as an industrial fermentation substrate. *Applied and Environmental Microbiology* 52 (1): 152–156.
- Ramsay, J. A., Berger, E., Voyer, R., Chavarie, C. and Ramsay, B. A. 1994. Extraction of Poly-3-hydroxybutyrate using chlorinated solvents. *Biotechnology Technology* 8: 589–594.
- Reddy, S. V., Thirumala, M. and Mahmood, S. K. 2009. A novel Bacillus sp. accumulating Poly(3-hydroxybutyrate-co-3-hydroxyvalerate) from a single carbon substrate. *Journal Indian Microbiology and Biotechnology* 36: 837–843.
- Rhim, J. W., Koh, S. and Kim, J. M. 2011. Effect of freezing temperature on rehydration and water vapor adsorption characteristics of freeze-dried rice porridge. *Journal of Food Engineering* 104: 484–491.
- Ribera, R. G., Monteoliva-Sanchez, M. and Ramos-Cormenzana, A. 2001. Production of Polyhydroxyalkanotes by *Pseudomonas putida* KT2442 harboring pSK2665 in wastewater from olive oil mills (Alpechin). *Journal of Biotechnology* 4 (2): 116–119.
- Riis, V. and Mai, W. 1988. Gas chromatographic determination of Poly-Beta Hydroxybutyric acid in microbial biomass after hydrochloric acid propanolysis. *Journal of Chromatography* 445: 285–289.
- Rossi, S., Azghani, A. O. and Omri, A. 2004. Antimicrobial efficacy of a new antibiotic-loaded Poly(hydroxybutyric-co-hydroxyvaleric acid) controlled release system. *Journal of Antimicrobial Chemotheraphy* 54: 1013–1018.
- Rothermich, M. M., Guerrero, R., Lenz, Robert, W. and Goodwin, S. 2000. Characterization, seasonal occurrence, and diel fluctuation of Polyhydroxyalkanoate in photosynthetic microbial mats. *Applied and Environmental Microbiol*ogy 66 (10): 4279–4291.
- Salmiati, Z., Ujang, M. R., Salim, M. F. M. D. and Ahmad, M. A. 2007. Intracellular biopolymer productions using mixed microbial cultures from fermented POME. Water Science and Technology 56 (8): 179–185.
- Santhanam, A. and Sasidharan, S. 2010. Microbial production of Polyhydroxyalkanoates (PHA) from Alcaligenes spp. and Pseudomonas olevorans using different carbon sources. African Journal of Biotechnology 9 (21): 3144–3150.
- Savenkova, L., Gercberga, Z., Bibers, I. and Kalnin, M. 2000. Effect of 3-hydroxy content on some physical and mechanical properties of Polyhydroxyalkanoates produced by *Azobacter chroococcum. Process Biochemistry* 36: 445–450.
- Shamala, T. R., Chandrashekar, A., Vijayendra, S. V. N. and Kshama, L. 2003. Identification of Polyhydroxyalkanoate (PHA)-producing *Bacillus* spp. using the Polymerase Chain Reaction (PCR). *Journal of Applied Microbiology* 94: 369– 374.

- Shi, F., Ashby, R. D. and Gross, R. A. 1997. Fractionation and characterization of microbial polyesters containing 3-Hydroxybutyrate and 4-Hydroxybutyrate repeat units. *Macromolecules* 30: 2521–2523.
- Sim, K. H. 2003. Utilisation of organic acids from anaerobically treated palm oil mill effluent with or without sludge recycle for Polyhydroxyalkanoate production. PhD thesis, Faculty of Biotechnology and Biomolecular Sciences. Universiti Putra Malaysia.
- Solaiman, D. K. Y., Ashby, Richard, D., Foglia, T. A. and Marmer, W. N. 2006. Conversion of agricultural feedstock and coproducts into Polyhydroxyalkanoates. *Applied Microbiology and Biotechnology* 71: 783–789.
- Song, J. J., Yoon, S. C., Yu, S. M. and Lenz, R. W. 1998. Differential scanning calorimetric study of Poly(3-hydroxyoctanoate) inclusions in bacterial cells. *International Journal of Biological Macromolecules* 23: 165–173.
- Steinbuchel, A. 1992. Biodegradable plastics. *Current Opinion in Biotechnology* 3: 291–297.
- Sudesh, K., Bhubalan, K., Chuah, J.-A., Kek, Y.-K., Kamilah, H., Sridewi, N. and Lee, Y.-F. 2011. Synthesis of Polyhydroxyalkanoates from palm oil and some new applications. *Applied Microbiology and Biotechnology* 89: 1373–1386.
- Sujatha, K., Mahalakshmi, A. and Shenbagarathai, R. 2007. Molecular characterization of *Pseudomonas sp.* LDC-5 involved in accumulation of Poly 3hydroxybutyrate and medium-chain-length Poly 3-hydroxyalkanoates. *Archives* of *Microbiology* 188 (5): 451–462.
- Suriyamongkol, P., Weselake, R., Narine, S., Moloney, M. and Shah, S. 2007. Biotechnological approaches for the production of Polyhydroxyalkanoates in microorganisms and plants - A review. *Biotechnology Advances* 25: 148–175.
- Suzuki, T., Yamane, T. and Shimizu, S. 1986. Mass production of Poly-3hydroxybutyric acid by fully automatic fed-batch culture of Methyltroph. Applied Microbiology and Biotechnology 23: 322–329.
- Taguchi, K., Taguchi, S., Sudesh, K., Maehara, A., Tsuge, T. and Doi, Y. 2005. Metabolic pathways and engineering of PHA biosynthesis. In *Biotechnology of biopolymers. From synthesis to patents* (eds. A. Steinbuchel and Y. Doi), 163. EWiley-Vch.Verlag GmbH and Co. KGaA, Weinheim.
- Tajima, K., Igari, T., Nishimura, D., Nakamura, M., Satoh, Y. and Munekata, M. 2003. Isolation and characterization of *Bacillus* sp. INT005 accumulating Polyhydroxyalkanoate (PHA) from gas field soil. *Journal of Bioscience and Bioengineering* 95 (1): 77–81.
- Takabatake, H., Satoh, H., Mino, T. and Matsuo, T. 2002. PHA (Polyhydroxyalkanoate) production potential of activated sludge treating wastewater. Water Science and Technology 45 (12): 119–126.

- Tamer, I. M., Moo-Young, M. and Chisti, Y. 1998. Disruption of Alcaligenes latus for recovery of Poly(beta-hydroxybutyric acid) : Comparison of high-pressure homogenization, bead milling and chemically induced lysis. Indian Engineering Chemical Resource 37: 1807–1814.
- Valappil, Sabeel, P., Boccaccini, A. R., Bucke, C. and Roy, I. 2007. Polyhydroxyalkanoates in Gram-positive bacteria : Insight from the genera *Bacillus* and *Streptomyces. Antonie van Leeuwenhoek* 91: 1–17.
- Van-Thuoc, D., Quillaguaman, J., Mamo, G. and Mattiasson, B. 2007. Utilization of agricultural residues for Poly(3-hydroxybutyrate) production by *Halomonas boliviensis* LC1. *Journal of Applied Microbiology* 104: 420–428.
- van Wyk, J. P. H. 2001. Biotechnology and the utilization of biowaste as a resource for bioproduct development. *Trends in Biotechnology* 19 (15): 172–177.
- Voon, P. T. 2005. Environmental friendly alternative methods for the recovery of intracellular Polyhydroxyalkanoates (PHA). PhD thesis, Faculty of Biotechnology and Biomolecular Sciences. Universiti Putra Malaysia.
- Wang, J., Fang, F. and Yu, H.-Q. 2007. Substrate consumptions and biomass growth of *Ralstonia eutropha* at various So/X0 levels in batch culture. *Biore*source Technology 98: 2599–2604.
- Wang, J. and Yu, J. 2001. Kinetic analysis on formation of Poly(3hydroxybutyrate) from acetic acid by *Ralstonia eutropha* under chemically defined conditions. *Journal of Industrial Microbiology and Biotechnology* 26: 121– 126.
- Wang, J., Yue, Z.-B., Sheng, G.-P. and Yu, H.-Q. 2010a. Kinetic analysis on the production of Polyhydroxyalkanoates from volatile fatty acids by *Cuprividus necator* with a consideration of substrate inhibition, cell growth, maintenance and product formation. *Biochemical Engineering Journal* 49: 422–428.
- Wang, X., Chen, Z., Chen, X., Pan, J. and Xu, K. 2010b. Miscibility, crystallization kinetics and mechanical properties of Poly(3-hydroxyvalerate--co-3hydroxybutyrate) (PHBV)/ Poly(3-hydroxyvalerate--co-3-hydroxybutyrate-co-4-hydroxybutyrate) (P3/4HB) blends. Journal of Applied Polymer Science 117: 838–848.
- Weusthuis, R. A., Kessler, B., Dielissen, M. P. M., Witholt, B. and Eggink, G. 2005. Fermentative production of medium chain-length Poly(3hydroxyalkanoates.). In *Biotechnology of biopolymers. From synthesis to patents* (eds. A. Steinbuchel and Y. Doi), 235. EWiley-Vch.Verlag GmbH and Co. KGaA, Weinheim.
- Witholt, B. and Kessler, B. 1999. Perspectives of medium chain length Poly(hydroxyalkanoates), a versatile set of bacterial bioplastics. *Current Opin*ion in Biotechnology 10: 279–285.

- Wrobe, M., Zebrowski, J. and Szopa, J. 2004. Polyhydroxybutyrate synthesis in transgenic flax. *Journal of Biotechnology* 107 (107): 41–54.
- Wu, S.-T., Lin, Y.-C. and Too, J.-R. 2009. Continuous production of Poly(3hydroxybutyrate-3-hydroxyvalerate : Effects of C/N ratio and dilution/rate on HB/HV. Korean Journal Chemical Engineering 26 (2): 411–416.
- Xu, Y., Wang, R.-H., Koutinas, A. A. and Webb, C. 2010. Microbial biodegradable plastic production from a wheat-based biorefining strategy. *Process Biochemistry* 45: 153–163.
- Yamane, T., Chen, X.-F. and Ueda, S. 1996. Polyhydroxyalkanoate synthesis from alcohol during the growth of *Paracoccus denitrifican*. Federation of European Microbiological Societies Microbiology Letters 135: 207–211.
- Yan, Q., Du, G. and Chen, J. 2003. Biosynthesis of Polyhydroxyalkanoates (PHAs) with continuous feeding of mixed organic acids as carbon sources by *Ralstonia eutropha*. *Process Biochemistry* 39: 387–391.
- Yan, Q., Sun, Y., Ruan, L. and Chen, J. Y. 2005. Biosynthesis of short-chain-length Polyhydroxyalkanoates during the dual-nutrient-limited zone by *Ralstonia eu*tropha. World Journal of Microbiology and Biotechnology 21: 17–21.
- Yang, Y.-H., Brigham, C. J., Budde, C. F., Boccazzi, P., Willis, L. B., Hassan, M. A., Mohd Yusof, Z. A., Rha, C. and Sinskey, A. J. 2010. Optimization of growth media components for Polyhydroxyalkanoates (PHA) production from organic acids by *Ralstonioa eutropha*. *Applied Microbiology and Biotechnology*.
- Yasotha, K., Aroua, M. K., Ramachandran, K. B. and Tan, I. K. P. 2006. Recovery of medium-chain-length Polyhydroxyalkanoates (PHA) through enzymatic digestion treatments and ultrafiltration. *Biochemical Engineering Journal* 30 (3): 260–268.
- Yoon, J.-S., Chin, I.-J., Kim, M.-N. and Kim, C. 1996. Degradation of microbial polyesters: A theoretical prediction of molecular weight and polydispersity. *Macromolecules* 29 (29): 3303–3307.
- You, J.-W., Chiu, H.-J., Shu, W.-J. and Don, T.-M. 2003. Influence of hydroxyvalerate content on the crystallization kinetics of Poly(hydroxybutyrate-cohydroxyvalerate. Journal of Polymer Research 10: 47–54.
- Yu, J., Si, Y. and Wong, W. K. R. 2002. Kinetics modeling of inhibition and utilisation of mixed volatile fatty acids in the formation of Polyhydroxyalkanoates by *Ralstonia eutropha*. Process Biochemistry 37: 731–738.
- Yu, S. T., Lin, C. C. and Too, J. R. 2005. PHBV production by *Ralstonia eutropha* in a continuous stirred tank reactor. *Process Biochemistry* 40: 2729–2734.

- Zakaria, M. R., Abd-Aziz, S., Arrifin, H., Abdul Rahman, N. A., Phang, L. Y. and Hassan, M. A. 2008. *Comamonas* sp. EB172 isolated from digester treating palm oil mill effluent as potential Polyhydroxyalkanoate (PHA) producer. *African Journal of Biotechnology* 7 (22): 4118–4121.
- Zakaria, M. R., Ariffin, H., Mohd Johar, N. A., Abd-Aziz, S., Nishida, H., Shirai, Y. and Hassan, M. A. 2010b. Biosynthesis and characterization of Poly(hydroxybutyric-co-hydroxyvaleric acid) copolymer from wild-type Comamonas sp. EB172. Polymer Degradation and Stability 95: 1382–1386.
- Zakaria, M. R., Tabatabaei, M., Mohamad Ghazali, F., Abd-Aziz, S., Shirai, Y. and Hassan, M. A. 2010a. Polyhydroxyalkanoates production from anaerobically treated palm oil mill effluent by new bacterial strain *Comamonas* sp. EB172. *World Journal Microbiology and Biotechnology* 26 (5): 767–774.

