



UNIVERSITI PUTRA MALAYSIA

***DEVELOPMENT OF RECOMBINANT OUTER MEMBRANE PROTEINS
VACCINE AND ITS IMMUNOPROTECTIVE ABILITY AGAINST *Vibrio
alginolyticus* IN HYBRID GROUPER (*Epinephelus fuscoguttatus
Forsskal* x *Epinephelus lanceolatus* Bloch)***

NEHLAH BINTI ROSLI

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By

NEHLAH BINTI ROSLI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of
Master of Science**

August 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

DEVELOPMENT OF RECOMBINANT OUTER MEMBRANE PROTEINS VACCINE AND ITS IMMUNOPROTECTIVE ABILITY AGAINST *Vibrio alginolyticus* IN HYBRID GROUPEL (*Epinephelus fuscoguttatus* Forsskal x *Epinephelus lanceolatus* Bloch)

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August 2016

Chairman : Ina Salwany Md. Yasin, PhD
Faculty : Agriculture

The productions of grouper may become restricted due to many constraints, including diseases. The most significant bacterial disease attacking groupers is vibriosis and its major causative agent is *Vibrio alginolyticus*. This study was conducted to develop recombinant cells vaccine containing antigenic OmpK and OmpW genes from *V. alginolyticus* by gene cloning and protein expression, and to evaluate the immune response and protection efficacy of hybrid grouper following exposure to virulent *V. alginolyticus*. The selected *V. alginolyticus* strain VA2 was first identified to confirm it as *V. alginolyticus* by phenotypic and genotypic characterization. In addition, the antigenicity analysis of the outer membrane proteins (OMPs) of *V. alginolyticus* strain VA2 was carried out. The OMPs were isolated and purified, followed by protein profiling using sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and antigenicity analysis was done using Western blotting. The SDS-PAGE profiling revealed a mixture of major and minor protein bands. The 23 and 31 kDa protein bands, postulated to be the OmpW and OmpK proteins, respectively, were antigenic as shown by Western blot. The vaccine was constructed by purifying PCR products of OmpK and OmpW genes from *V. alginolyticus*, followed by transformation into cloning and expression hosts, *Escherichia coli* TOP10 and BL21(DE3), respectively. Sequencing analysis revealed that the full length of target genes OmpK and OmpW of *V. alginolyticus* strain VA2 were 846 and 642 bp, respectively. The sequences of the target genes were highly similar to published sequences of *Vibrio* species in GenBank. The multiple sequence alignment showed that their amino acid sequences were highly conserved

between different *Vibrio* species. High numbers of antigenic sites were also predicted in the OmpK and OmpW protein sequences. Pilot expression was done for recombinant OmpK and OmpW to determine the optimum expression temperature and time, which were revealed to be 30°C for 10 h. The expressed target proteins were then detected by His-tag monoclonal antibody using SDS-PAGE and Western blotting, in which the presence of fusion proteins at 48.3 kDa and 40.3 kDa for OmpK and OmpW respectively, were confirmed. For vaccination, formalin killed whole-cell of *V. alginolyticus* strain VA2 was prepared at concentration of 1.59×10^7 and 8.3×10^7 CFU/ml for OmpK and OmpW respectively. Six groups of 150 hybrid groupers each, at approximately 30 g was acclimatized beforehand for two weeks. These six groups were for treatments of rOmpK, rOmpW, rOmpK+rOmpW, *Escherichia coli* only, PBS only and control without manipulation. Vaccination was done by intraperitoneal injection on day 0. Booster dose was given on day 14. On day 28, all fish were challenged with virulent strain of *V. alginolyticus* at concentration of 1×10^9 CFU/mL by intraperitoneal (IP) injection. Blood serum and gut samples were collected biweekly until week 10. The survival percentages were highest for OmpK+OmpW and OmpK vaccinated groups at 100%, followed by OmpW vaccinated group at 67%, PBS group at 12%, and *E. coli* group at 0%. Enzyme-linked immunosorbent assay (ELISA) analysis showed significant ($p < 0.05$) antibody difference between vaccinated and non-vaccinated groups from week 2 to week 10. After booster dose, the antibody level of bivalent vaccine OmpK+OmpW was significantly higher ($p < 0.05$) than other groups. However, it was not significantly different ($p > 0.05$) than monovalent vaccine OmpK after challenge at week 4. Gut histology showed presence of gut-associated lymphoid tissue (GALT) in vaccinated groups only. This showed that while bivalent vaccine OmpK+OmpW is effective, monovalent vaccine OmpK alone is also strong enough to give protection to groupers against high concentration of virulent *V. alginolyticus*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**PEMBANGUNAN VAKSIN PROTEIN MEMBRAN LUAR
REKOMBINAN DAN KEBOLEHANNYA MEMBERI PERLINDUNGAN
IMUN TERHADAP *Vibrio alginolyticus* KEPADA IKAN KERAPU
HIBRID (*Epinephelus fuscoguttatus* Forsskal x *Epinephelus lanceolatus*
Bloch)**

Oleh

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Pengeluaran ikan kerapu boleh menjadi terhad disebabkan pelbagai kekangan, termasuk penyakit. Penyakit bakteria yang paling serius menyerang kerapu adalah vibriosis dan agen penyebab utamanya adalah *Vibrio alginolyticus*. Kajian ini dijalankan untuk menghasilkan vaksin sel rekombinan yang mengandungi antigen gen OmpK dan OmpW dari *V. alginolyticus* dengan cara pengklonan dan ekspresi protein, serta untuk menilai tindak balas imun ikan kerapu hibrid yang didedahkan kepada *V. alginolyticus* yang virulen. *Vibrio alginolyticus* strain VA2 yang dipilih telah dikenal pasti terlebih dahulu sebagai *V. alginolyticus* oleh pencirian fenotip dan genotip. Tambahan pula, analisis keantigenan protein membran luar (OMPs) *V. alginolyticus* strain VA2 telah dilakukan. Protein membran luar (OMPs) tersebut diasingkan dan dituliskan, diikuti pemprofilan protein oleh elektroforesis gel poliakrilamida sodium dodesil sulfat (SDS-PAGE) dan analisis keantigenan oleh pembloatan Western. Pemprofilan SDS-PAGE menunjukkan campuran jalur protein utama dan minor. Jalur protein yang bersaiz 23 dan 31 kDa yang diandaikan adalah protein OmpW dan OmpK, masing-masing, adalah bersifat antigen menurut pembloatan Western. Vaksin ini dihasilkan dengan menuliskan produk PCR gen-gen OmpK dan OmpW dari *V. alginolyticus*, diikuti dengan transformasi ke dalam perumah pengklonan dan pengeksresi, masing-masing *Escherichia coli* TOP10 dan BL21(DE3). Analisis penjujukan menunjukkan saiz keseluruhan gen OmpK dan OmpW *V. alginolyticus* strain VA2 adalah masing-masing 846 dan 642 bp. Jujukan gen tersebut mempunyai persamaan yang sangat tinggi dengan

jujukan spesies *Vibrio* dalam GenBank. Penjajaran berbagai turutan menunjukkan jujukan asid amino mereka dikekalkan dalam kalangan spesies *Vibrio*. Jumlah tempat antigenik yang tinggi juga diramalkan dalam jujukan protein OmpK dan OmpW. Ekspresi ujian telah dilakukan bagi OmpK dan OmpW untuk menentukan suhu dan jangka masa ekpresi yang optimum, iaitu 30°C selama 10 jam. Protein sasaran yang diekspres kemudiannya dikesan oleh antibodi monoklonal His-tag menggunakan SDS-PAGE dan pemblotan Western, yang mana gabungan protein OmpK dan OmpW bersaiz 48.3 kDa dan 40.3 kDa, masing-masing, telah dikenal pasti. Bagi pemvaksinan, sel keseluruhan *V. alginolyticus* strain VA2 yang dimatikan dengan menggunakan formalin telah disediakan pada kepekatan 1.59×10^7 dan 8.3×10^7 CFU/ml untuk OmpK dan OmpW masing-masing. Enam kumpulan ikan kerapu hibrid, setiap kumpulan sebanyak 150 ekor, dengan berat setiapnya kira-kira 30 g diaklimatisasikan terlebih dahulu selama dua minggu. Enam kumpulan ini adalah untuk rawatan rOmpK, rOmpW, rOmpK+rOmpW, *Eschericia coli* sahaja, larutan garam bertimbal fosfat (PBS) sahaja dan kawalan tanpa manipulasi. Pemvaksinan dilakukan dengan suntikan intraperitoneum pada hari 0. Dos peningkatan diberikan pada hari 14. Pada hari 28, semua ikan dicabar dengan *V. alginolyticus* yang virulen pada kepekatan 1×10^9 CFU/mL dengan suntikan IP. Serum darah dan sampel usus diambil setiap dua minggu sehingga minggu 10. Peratusan kemandirian adalah tertinggi untuk kumpulan-kumpulan vaksinasi OmpK+OmpW dan OmpK pada 100%, diikuti kumpulan vaksinasi OmpW pada 67%, kumpulan PBS pada 12% dan kumpulan *E. coli* pada 0%. Analisis asai imunoserap terangkai enzim (ELISA) menunjukkan perbezaan ketara ($p < 0.05$) antibodi antara kumpulan vaksinasi dan tiada vaksinasi dari minggu kedua hingga minggu 10. Selepas dos peningkatan, tahap antibodi vaksin bivalen OmpK+OmpW meningkat dengan ketara ($p < 0.05$) berbanding kumpulan-kumpulan lain. Bagaimanapun, tiada perbezaan ketara ($p > 0.05$) antaranya dengan vaksin monovalen OmpK selepas dicabar pada minggu 4. Histologi usus menunjukkan kehadiran sel limfoid berkaitan usus (GALT) dalam kumpulan vaksinasi sahaja. Ini menunjukkan vaksin bivalen OmpK+OmpW adalah efektif, sementara vaksin monovalen OmpK sahaja sudah cukup kuat untuk memberi perlindungan kepada kerapu terhadap *V. alginolyticus* virulen kadar tinggi.

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I certify that a Thesis Examination Committee has met on 26 August 2016 to conduct the final examination of Nehlah binti Rosli on her thesis entitled "Development of Recombinant Outer Membrane Proteins Vaccine and its Immunoprotective Ability Against *Vibrio alginolyticus* in Hybrid Grouper (*Epinephelus fuscoguttatus* Forsskal x *Epinephelus lanceolatus* Bloch)" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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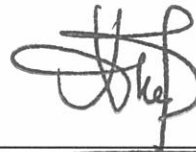
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LIST OF ABBREVIATIONS/NOTATIONS/GLOSSARY OF TERMS

%	Percentage
λ	lambda
μg	microgram
μl	microliter
μM	micromolar
ANOVA	Analysis of variance
BHI	Brain-heart infusion
BLAST	Basic Local Alignment Search Tool
bp	base pair
BSA	bovine serum albumin
CFU	colony forming units
DAB	3,3'-Diaminobenzidine
dATP	deoxyadenosine triphosphate
dNTP	deoxynucleotide triphosphate
DNA	deoxyribonucleic acid
DTT	dithiothreitol
<i>E. coli</i>	<i>Escherichia coli</i>
<i>E. fuscoguttatus</i>	<i>Epinephelus fuscoguttatus</i>
<i>E. lanceolatus</i>	<i>Epinephelus lanceolatus</i>
EFA	essential fatty acid
FKC	formalin-killed cells
ELISA	Enzyme-Linked Immunosorbent Assay

GALT	Gut-Associated Lymphoid Tissue
H ₂ SO ₄	sulphuric acid
H&E	haematoxylin and eosin
Ig	immunoglobulin
IHN	infectious haematopoietic necrosis
IP	intraperitoneal
IPTG	isopropyl β-D-1-thiogalactopyranoside
ITS	Internal Transcribed Spacer
kb	kilobase pair
kDa	kilodalton
LB	Luria Bertani
LIC	ligation-independent cloning
M	molar
MCS	multiple cloning site
mg	milligram
MgCl ₂	magnesium chloride
MgSO ₄	magnesium sulfate
mM	millimolar
MS-222	tricaine methanesulfonate solution
NaCl	sodium chloride
ng	nanogram
nm	nanometer
OD	optical density
OMP	Outer membrane protein

PBS	phosphate buffer saline
PCR	polymerase chain reaction
pmol	picomole
ppt	parts per thousand
RBC	red blood cells
RE	restriction enzyme
RNA	ribonucleic acid
rpm	revolutions per minute
RPS	relative percentage survival
rRNA	ribosomal ribonucleic acid
SDS-PAGE	Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis
SOC	Super Optimal broth with Catabolite repression
<i>sp.</i>	species
TBE	tris-boric EDTA
TBS	tris-buffer saline
TCBS	thiosulfate-citrate-bile salts-sucrose
TMB	3,3',5,5'-tetramethylbenzidine
TSA	tryptic soy agar
TSB	tryptic soy broth
U	unit
V	voltage
VHS	viral haemorrhagic septicaemia
<i>V. alginolyticus</i>	<i>Vibrio alginolyticus</i>

VNN	viral nervous necrosis
v/v	volume per volume
w/v	weight per volume
$x g$	multiples of gravitational acceleration



CHAPTER 1

INTRODUCTION

Vibriosis is a disease caused by bacteria belonging to the genus *Vibrio*, and has been recognized as one of the most severe and destructive diseases in the aquaculture industry, causing high mortality in marine fish (Cai *et al.*, 2013a). *Vibrio* species is a rod-shaped, gram-negative bacterium from the family Vibrionaceae with a broad host range (Cai *et al.*, 2013a). Studies had shown the virulence of *V. alginolyticus*, which is considered the main bacterial pathogen causing outbreaks to humans as well as aquatic animals such as shrimp, shellfish and marine fish (Cai *et al.*, 2013a; Zorrilla *et al.*, 2003; Lee *et al.*, 1996; Lightner, 1993). Vibriosis caused by *V. alginolyticus* is able to affect all fish growth stages which often leads to severe mortality. Due to various diseases including vibriosis, survival of cage-cultured groupers in Malaysia are up to 30-40% only (Liao and Leño, 2008). *Vibrio alginolyticus* itself causes more than 50% mass mortality of worldwide fishery culture (Salosso and Jasmanindar, 2014), consequently leading to serious economic damage (Martins *et al.*, 2010).

Outbreaks of vibriosis have been frequently reported in grouper (Huang *et al.*, 2012), sea bream (El-Galil and Mohamed, 2012), sea bass (Bellos *et al.*, 2015) and catfish (Geng *et al.*, 2014). In 2010, two major vibriosis outbreaks on April and October caused high mortality of cultured marine fish species including Asian seabass (*Lates calcarifer*), brown-marbled grouper (*E. fuscoguttatus*), red snapper (*Lutjanus* sp.), orange-spotted grouper (*E. coioides*) and hybrid grouper (*E. fuscoguttatus* x *E. lanceolatus*) in an aquaculture facility in Sabah, Malaysia (Albert and Ransangan, 2013).

In a study by Kua *et al.* (2011), abnormal mortality of 4-5 abalone/day instead of 1-3 abalone/month were reported for abalone cultured in Penang, Malaysia, in which it was confirmed that the abalones were infected by *Pasteurella* sp. and *V. alginolyticus*. In Karimunjawa islands, North Java Sea, Indonesia, the causative agents of vibriosis associated with groupers were investigated and the result showed that they include *V. alginolyticus* (Sarjito *et al.*, 2009).

Vibriosis normally occurs when the fish is exposed to the pathogen in the presence of stress factors (El-Galil and Mohamed, 2012; Martins *et al.*, 2010). Vibriosis had been known to cause septicemia, identified by the presence of fin haemorrhages, loss of appetite, skin lesions (Martins *et al.*, 2010), skin

darkening, pale gills caused by anemia, enlarged spleen, internal ascetic fluid (Hendrikson and Zenoble, 1983), and fins and mouth erythema (Reed and Francis-Floyd, 2009). Good husbandry is crucial in preventing spread of diseases. Antibiotics and chemotherapeutic agents are commonly used as strategies to control vibriosis (Sonia and Lipton, 2012). Fish farmers prefer antibiotics as opposed to vaccines due to the former being widely available and the latter being scarce. However, antibiotic treatment has become less effective with the gradual spread of drug resistance (Cai *et al.*, 2013a). Another effective alternative would be vaccination, which has been shown to be able to increase the antibody titre, induce immune memory and significantly increase the survival of infected fish (Defoirdt *et al.*, 2007). Grisez and Tan (2005) observed that in Asia except for Japan, the use of vaccines for controlling fish diseases is not common, even though fish production is large in Asia. Vaccines should be used as an alternative strategy to control vibriosis due to its ability to induce production of antibodies against pathogens in case of infection.

Currently, recombinant vaccine is a promising strategy to combat diseases in fish by using immunogenic protein expressed by specific antigens. Recombinant cells produce the protein of interest in large amount, which increase the antibody reaction and enhance protection. One of the gram-negative bacteria surface structure known as outer membrane have a role in interaction between bacterium and host, in which the outer membrane proteins (OMPs) could serve as potential candidates for vaccine development (Jiang *et al.*, 2003).

As the fish immune system is very specific, a classical vibriosis vaccine that had been developed, with *V. anguillarum* as the antigen, will not provide specific protection against other *Vibrios* spp. (Toranzo *et al.*, 1996). Thus, it is crucial to develop a safe and effective broad-range vaccine for fish against *V. alginolyticus* and possibly other *Vibrio* species as well. For this purpose, two interesting OMP genes, OmpK and OmpW were evaluated in our study.

In developing recombinant bivalent vaccine against *V. alginolyticus* in hybrid grouper, the objectives were:

1. To characterize *Vibrio alginolyticus* isolated from diseased grouper and analyze the antigenicity of its outer membrane proteins (OMPs).
2. To construct recombinant cell vaccines of *V. alginolyticus* antigenic outer membrane proteins OmpK and OmpW genes for the production of potential recombinant vaccines.

3. To evaluate the effectiveness of monovalent vaccines (OmpK and OmpW) versus bivalent vaccine (OmpK+OmpW) in protecting hybrid grouper against *Vibrio alginolyticus* infection.

Hypotheses of this study were:

1. The outer membrane proteins (OMPs) of *V. alginolyticus* consist of major and minor proteins, which can be detected by polyclonal rabbit antiserum.
2. For the vaccine construction, the recombinant cells carrying the genes encoding OmpK and OmpW proteins of *V. alginolyticus* can be cloned and expressed in the *Escherichia coli* prokaryotic expression system.
3. Recombinant bivalent vaccine (OmpK+OmpW) should confer better protection against *V. alginolyticus* infection in hybrid grouper, compared to monovalent vaccines (OmpK and OmpW).

REFERENCES

- Addin, A. M., & Senoo, S. (2011). Production of Hybrid Groupers: Spotted grouper, *Epinephelus polyphkadion* x Tiger grouper *E. fuscoguttatus* and Coral grouper, *E. corallicola* x Tiger Grouper. *Borneo Marine Research Institute, Universiti Malaysia Sabah, Malaysia*.
- Adzitey, F., Huda, N., & Ali, G. R. R. (2013). Molecular techniques for detecting and typing of bacteria, advantages and application to foodborne pathogens isolated from ducks. *3 Biotech*, 3(2), 97-107.
- Albert, V., & Ransangan, J. (2013). Effect of water temperature on susceptibility of culture marine fish species to vibriosis. *International Journal of Research in Pure and Applied Microbiology*, 3(3), 48-52.
- Andre, F. E., Booy, R., Bock, H. L., Clemens, J., Datta, S. K., John, T. J., Lee, B. W., Lolekha, S., Peltola, H., Ruff, T. A., Santosham, M., & Schmitt, H. J. (2008). Vaccination greatly reduces disease, disability, death and inequity worldwide. *Bulletin of the World Health Organization*, 86(2), 140-146.
- Angelidis, P., Karagiannis, D., & Crump, E. M. (2006). Efficacy of a *Listonella anguillarum* (syn. *Vibrio anguillarum*) vaccine for juvenile sea bass *Dicentrarchus labrax*. *Diseases of aquatic organisms*, 71(1), 19-24.
- Austin, B. (1984). The future of bacterial fish vaccines. *Vaccine*, 2(4), 249-254.
- Austin, B. (2010). Vibrios as causal agents of zoonoses. *Veterinary microbiology*, 140(3), 310-317.
- Baker, M. (2015). Blame it on the antibodies. *Nature*, 521(7552), 274-276.
- Bakopoulos, V., Nikolaou, I., Kalovyryna, N., Amirali, E., Kokkoris, G., & Spinou, E. (2015). Prevention of fish photobacteriosis. Comparison of the efficacy of intraperitoneally administered commercial and experimental vaccines. *Mediterranean Marine Science*, 16(2), 385-392.
- Baneyx, F. (1999). Recombinant protein expression in *Escherichia coli*. *Current opinion in biotechnology*, 10(5), 411-421.
- Bellos, G., Angelidis, P., & Miliou, H. (2015). Effect of Temperature and Seasonality Principal Epizootiological Risk Factor on Vibriosis and Photobacteriosis Outbreaks for European Sea Bass in Greece (1998-2013). *Journal of Aquaculture Research & Development*, 6(5), 1.

- Béné, C., Arthur, R., Norbury, H., Allison, E. H., Beveridge, M., Bush, S., Campling, L., Leschen, W., Little, D., Squires, D., Thilsted, S. H., Troell, M., & Williams, M. (2016). Contribution of fisheries and aquaculture to food security and poverty reduction: assessing the current evidence. *World Development*, 79, 177-196.
- BioAqua Vietnam (2013). Marine LABS – A convenient, efficient and environment friendly solution for aquaculturists, <http://bioaqua.vn/en/marine-labs-a-convenient-efficient/> Aqua Technique.
- Bondad-Reantaso, M. G., Humphrey, J., Kanchanakhan, S., & Chinabut, S. (2001). Report and Proceeding of APEC FWG 02/2000. *Development of a Regional Research Programme on Grouper Virus Transmission and Vaccine Development*. APEC Secretariat, 2001.
- Bondad-Reantaso, M. G., Subasinghe, R. P., Arthur, J. R., Ogawa, K., Chinabut, S., Adlard, R., Tan, Z., & Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary parasitology*, 132(3), 249-272.
- Bowater, R. O., Forbes-Faulkner, J., Anderson, I. G., Condon, K., Robinson, B., Kong, F., Gilbert, G. L., Reynolds, A., Hyland, S., McPherson, G., Brien, J. O., & Blyde, D. (2012). Natural outbreak of *Streptococcus agalactiae* (GBS) infection in wild giant Queensland grouper, *Epinephelus lanceolatus* (Bloch), and other wild fish in northern Queensland, Australia. *Journal of Fish Diseases*, 35(3), 173-186.
- Brosius, J., Palmer, M. L., Kennedy, P. J., & Noller, H. F. (1978). Complete nucleotide sequence of a 16S ribosomal RNA gene from *Escherichia coli*. *Proceedings of the National Academy of Sciences*, 75(10), 4801-4805.
- Brown, T. A., & Brown, T. (2016). *Gene cloning and DNA analysis: an introduction*. John Wiley & Sons.
- Brudeseth, B. E., Wiulsrød, R., Fredriksen, B. N., Lindmo, K., Løkling, K. E., Bordevik, M., Steine, N., Klevan, A., & Gravningen, K. (2013). Status and future perspectives of vaccines for industrialised fin-fish farming. *Fish & shellfish immunology*, 35(6), 1759-1768.
- Busch, R. A. (1996). Polyvalent vaccines in fish: the interactive effects of multiple antigens. *Developments in biological standardization*, 90, 245-256.

- Bullock, G. L. (1977). *Vibriosis in fish*. U.S. Fish and Wildlife Service. U.S. Fish and Wildlife Publications.
- Cai, S. H., Lu, Y. S., Wu, Z. H. & Jian, J.C. (2013a). Cloning, expression of *Vibrio alginolyticus* outer membrane protein- OmpU gene and its potential application as vaccine in crimson snapper, *Lutjanus erythropterus* Bloch. *Journal of fish diseases*, 36(8), 695-702.
- Cai, S. H., Lu, Y. S., Jian, J. C., Wang, B., Huang, Y. C., Tang, J. F., Ding, Y., & Wu, Z. H. (2013b). Protection against *Vibrio alginolyticus* in crimson snapper *Lutjanus erythropterus* immunized with a DNA vaccine containing the ompW gene. *Diseases of aquatic organisms*, 106(1), 39-47.
- Calder, P. C. (2007). Immunological parameters: what do they mean?. *The Journal of nutrition*, 137(3), 773S-780S.
- Cantrell, S. A. (2003). Vectors for the Expression of Recombinant Proteins in *E. coli*. In *E. coli Plasmid Vectors* (pp. 257-275). Humana Press.
- Casali, N., & Preston, A. (2003). *E. coli plasmid vectors: Methods and applications* (Vol. 235). Springer Science & Business Media.
- CDC (Centers for Disease Control and Prevention), (2012). Vaccines: The Basics. [Cited 3 February 2016]. <http://www.cdc.gov/vaccines/vpd-vac/vpd-vac-basics.htm>
- Celie, P. H., Parret, A. H., & Perrakis, A. (2016). Recombinant cloning strategies for protein expression. *Current Opinion in Structural Biology*, 38, 145-154.
- Cereghino, J. L., & Cregg, J. M. (2000). Heterologous protein expression in the methylotrophic yeast *Pichia pastoris*. *FEMS microbiology reviews*, 24(1), 45-66.
- Ch'ng, C. L., & Senoo, S. (2008). Egg and larval development of a new hybrid grouper, tiger grouper *Epinephelus fuscoguttatus* × giant grouper *E. lanceolatus*. *Aquaculture Science (Japan)*.
- Chang, C. Y., Chiu, C. C., John, J. C., Liao, I. C., & Leño, E. M. (2008). Prophylaxis for iridovirus and nodavirus infections in cultured grouper in Taiwan. *The Aquaculture of Groupers*. Asian Fisheries Society. 207-224.
- Chen, F. Y., Chow, M., Chao, T. M., & Lim, R. (1977). Artificial spawning and larval rearing of the grouper, *Epinephelus tauvina* (Forsk.) in Singapore. *Singapore Journal of Primary Industries*, 5(1), 1-21.

- Chen, R. (2012). Bacterial expression systems for recombinant protein production: *E. coli* and beyond. *Biotechnology advances*, 30(5), 1102-1107.
- Chettri, J. K., Deshmukh, S., Holten-Andersen, L., Jafaar, R. M., Dalsgaard, I., & Buchmann, K. (2013). Comparative evaluation of administration methods for a vaccine protecting rainbow trout against *Yersinia ruckeri* O1 biotype 2 infections. *Veterinary immunology and immunopathology*, 154(1), 42-47.
- Chilmonczyk, S. (1992). The thymus in fish: development and possible function in the immune response. *Annual review of fish diseases*, 2, 181-200.
- Chong, R., Bousfield, B., & Brown, R. (2011). Fish Disease Management. *Veterinary Bulletin-Agriculture, Fisheries and Conservation Department Newsletter, Hong Kong*. December, 1, 8.
- Chou, H. Y., Lin, Y. H. & Shiau, S. Y. (2008). Nutrition, Immunology and Health Management of Groupers. *The aquaculture of groupers*. Asian Fisheries Society. 189-200.
- Chu, V. T., & Berek, C. (2013). The establishment of the plasma cell survival niche in the bone marrow. *Immunological reviews*, 251(1), 177-188.
- Chua, F. H. C., Ng, M. L., Ng, K. L., Loo, J. J., & Wee, J. Y. (1994). Investigation of outbreaks of a novel disease, 'Sleepy Grouper Disease', affecting the brown-spotted grouper, *Epinephelus tauvina* Forskal. *Journal of Fish Disease*, 17(4), 417-427.
- Chun, J., Huq, A., & Colwell, R. R. (1999). Analysis of 16S-23S rRNA intergenic spacer regions of *Vibrio cholerae* and *Vibrio mimicus*. *Applied and environmental microbiology*, 65(5), 2202-2208.
- Cline, J., Braman, J. C., & Hogrefe, H. H. (1996). PCR fidelity of Pfu DNA polymerase and other thermostable DNA polymerases. *Nucleic acids research*, 24(18), 3546-3551.
- Colquhoun, D. J., & Lillehaug, A. (2014). Vaccination against Vibriosis. *Fish Vaccination*, 172-184.
- Cornish, A. (Grouper & Wrasse Specialist Group) 2004. *Epinephelus fuscoguttatus*. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.1.14 <www.iucnredlist.org>. Downloaded on 14 November 2013.

- Corripio-Miyar, Y., de Quero, C. M., Treasurer, J. W., Ford, L., Smith, P. D., & Secombes, C. J. (2007). Vaccination experiments in the gadoid haddock, *Melanogrammus aeglefinus* L., against the bacterial pathogen *Vibrio anguillarum*. *Veterinary immunology and immunopathology*, 118(1), 147-153.
- Cregg, J. M., Cereghino, J. L., Shi, J., & Higgins, D. R. (2000). Recombinant protein expression in *Pichia pastoris*. *Molecular biotechnology*, 16(1), 23-52.
- Daniels, N. A., & Shafaie, A. (2000). A review of pathogenic *Vibrio* infections for clinicians. *Infections in medicine*, 17(10), 665-685.
- Defoirdt, T., Boon, N., Sorgeloos, P., Verstraete, W., & Bossier, P. (2007). Alternatives to antibiotics to control bacterial infections: luminescent vibriosis in aquaculture as an example. *Trends in biotechnology*, 25(10), 472-479.
- Demircan, D., & Candan, A. (2006). Identification of *Vibrio anguillarum* by PCR (rpoN gene) associated with vibriosis in marine fish in Turkey. *Turkish Journal of Veterinary and Animal Sciences*, 30(3), 305-310.
- DeVoe, M. R., & Hodges, C. E. (2002). Management of marine aquaculture: the sustainability challenge. *Responsible Aquaculture Development, World Aquaculture Society, CABI Publishing, New York, USA*, 21-24.
- Egidius, E. (1987). Vibriosis: pathogenicity and pathology. *Reviews in Aquaculture*, 67, 15-28.
- El-Galil, M. A. & Mohamed, M. H. (2012). First isolation of *Vibrio alginolyticus* from Ornamental Bird Wrasse Fish (*Gomphosus caeruleus*) of the red Sea in Egypt. *Journal of fisheries and aquatic Science*, 7(6), 461.
- Elizur, A., & Nocillado, J. (2013). project Controlling giant grouper maturation, spawning and juvenile production in Vietnam, the Philippines and Australia.
- Ellis, A. E. (1999). Immunity to bacteria in fish. *Fish & Shellfish Immunology*, 9(4), 291-308.
- Emery, A. H. (1981). Recombinant DNA technology. *The Lancet*, 318(8260), 1406-1409.
- Endo, Y., & Sawasaki, T. (2006). Cell-free expression systems for eukaryotic protein production. *Current opinion in biotechnology*, 17(4), 373-380.

- Evelyn, T. P. (1996). A historical review of fish vaccinology. *Developments in biological standardization*, 90, 3-12.
- Evensen, Ø., & Leong, J. A. C. (2013). DNA vaccines against viral diseases of farmed fish. *Fish & shellfish immunology*, 35(6), 1751-1758.
- Fang, H. M., Ling, K. C., Ge, R., & Sin, Y. M. (2000). Enhancement of protective immunity in blue gourami, *Trichogaster trichopterus* (Pallas), against *Aeromonas hydrophila* and *Vibrio anguillarum* by *A. hydrophila* major adhesin. *Journal of Fish Diseases*, 23(2), 137-145.
- FAO (Fisheries and Aquaculture Department). Fishery and Aquaculture Country Profiles. Malaysia (2009). Country Profile Fact Sheets. In: *FAO Fisheries and Aquaculture Department* [online]. Rome. Updated 1 February 2009. [Cited 31 March 2016]. http://www.fao.org/fishery/countrysector/naso_malaysia/en
- FAO (Fisheries and Aquaculture Department). (2014). The State of World Fisheries and Aquaculture 2014. *Food and Agricultural Organization of the United Nations, Rome*.
- FAO (Fisheries and Aquaculture Department). (2016). National Aquaculture Sector Overview. Malaysia. National Aquaculture Sector Overview Fact Sheets. *Fisheries and Aquaculture Information and Statistics Branch*.
- Farmer III, J. J., Janda, J. M., Brenner, F. W., Cameron, D. N., & Birkhead, K. M. (2005). Genus I. *Vibrio* Pacini 1854, 411AL. *Bergey's manual of systematic bacteriology*, 2(part B), 494-546.
- Firdaus, M. N. (2011). *Immune responses to Streptococcus agalactiae in red tilapia, Oreochromis spp. following vaccination with non-adjuvanted and adjuvanted vaccine incorporated feed pellets*. (Master of Science dissertation), Universiti Putra Malaysia, Malaysia.
- Firdaus - Nawati, M., Yusoff, S. M., Yusof, H., Abdullah, S. Z., & Zamri - Saad, M. (2013). Efficacy of feed - based adjuvant vaccine against *Streptococcus agalactiae* in *Oreochromis* spp. in Malaysia. *Aquaculture Research*, 45(1), 87-96.
- Gatewood, D. M., Fenwick, B. W., & Chengappa, M. M. (1994). Growth-condition dependent expression of *Pasteurella haemolytica* A1 outer membrane proteins, capsule, and leukotoxin. *Veterinary microbiology*, 41(3), 221-233.

- Geng, Y., Liu, D., Han, S., Zhou, Y., Wang, K. Y., Huang, X. L., Chen, D. F., Peng, X. & Lai, W. M. (2014). Outbreaks of vibriosis associated with *Vibrio mimicus* in freshwater catfish in China. *Aquaculture*, 433, 82-84.
- Gerdil, C. (2003). The annual production cycle for influenza vaccine. *Vaccine*, 21(16), 1776-1779.
- Germanier, R. (2012). *Bacterial vaccines* (Ed.). Academic Press.
- Glover, D. M. (2013a). *Gene cloning: the mechanics of DNA manipulation*. Springer.
- Glover, D. M. (2013b). *Genetic engineering cloning DNA*. Springer Science & Business Media.
- Gottesman, S. (1996). Proteases and their targets in *Escherichia coli* 1. *Annual review of genetics*, 30(1), 465-506.
- Gray, T. J., Kong, F., Jelfs, P., Sintchenko, V., & Chen, S. C. (2014). Improved Identification of Rapidly Growing Mycobacteria by a 16S-23S Internal Transcribed Spacer Region PCR and Capillary Gel Electrophoresis. *PloS one*, 9(7), e102290.
- Grisez, L. & Tan, Z. (2005). Vaccine development for Asian aquaculture. *Diseases in Asian Aquaculture*, 483, 494.
- Grodberg, J., & Dunn, J. J. (1988). ompT encodes the *Escherichia coli* outer membrane protease that cleaves T7 RNA polymerase during purification. *Journal of Bacteriology*, 170(3), 1245-1253.
- Gudding, R., Lillehaug, A., & Evensen, Ø. (1999). Recent developments in fish vaccinology. *Veterinary Immunology and Immunopathology*, 72(1), 203-212.
- Gudding, R., & Van Muiswinkel, W. B. (2013). A history of fish vaccination: science-based disease prevention in aquaculture. *Fish & shellfish immunology*, 35(6), 1683-1688.
- Gurkan, C., Symeonides, S. N., & Ellar, D. J. (2004). High-level production in *Pichia pastoris* of an anti-p185HER-2 single-chain antibody fragment using an alternative secretion expression vector. *Biotechnology and applied biochemistry*, 39(1), 115-122.
- Håstein, T., Gudding, R., & Evesen, O. (2005). Bacterial vaccines for fish - an update of the current situation worldwide. *Developmental Biology (Basel)*, 121, 55-74.

- Ha, T. Q., & Dinh, D. K. (2014). The changes in antigenic components of *Vibrio cholerae* strains isolated in Vietnam. *TẠP CHÍ Y HỌC DŨY PHÒNG, Tập XXIV, 2014, 10(159), 44.*
- Haenen, O., Fouz, B., Amaro, C., Isern, M. M., Mikkelsen, H., Zrnčić, S., Travers, M. A., Renault, T., Wardle, R., Hellstrom, A., & Dalsgaard, I. (2014). Vibriosis in aquaculture. 16th EAFF Conference, Tampere, Finland, 4th September 2013. *Bulletin of the European Association of Fish Pathologists, 34(4), 138-148.*
- Hamod, M. A., Nithin, M. S., Shukur, Y. N., Karunasagar, I., & Karunasagar, I. (2012). Outer membrane protein K as a subunit vaccine against *V. anguillarum*. *Aquaculture, 354, 107-110.*
- Heemstra, P. C., & Randall, J. E. (1993). An annotated and illustrated catalogue of the grouper, rockcod, hind coarl grouper and lyretail species known to date, FAO Species Catalogue, Vol. 16. *Groupers of the World (Family Serranidae, Subfamily Epinephelinae), 382.*
- Hendrikson, R. G. & Zenoble, R. D. (1983). Vibriosis in Fish: A Review. *Iowa State University Veterinarian, 45(1), 5.*
- Heppell, J., & Davis, H. L. (2000). Application of DNA vaccine technology to aquaculture. *Advanced Drug Delivery Reviews, 43(1), 29-43.*
- Hu, Y. L., Xiang, L. X., & Shao, J. Z. (2010). Identification and characterization of a novel immunoglobulin Z isotype in zebrafish: Implications for a distinct B cell receptor in lower vertebrates. *Molecular immunology, 47(4), 738-746.*
- Huang, Y., Jian, J., Lu, Y., Cai, S., Wang, B., Tang, J., Pang, H., Ding, Y. & Wu, Z. (2012). Draft genome sequence of the fish pathogen *Vibrio harveyi* strain ZJ0603. *Journal of bacteriology, 194(23), 6644-6645.*
- Hussain, N., Hiatt-Saif, M., & Ukawa, M. (1975). *On the culture of Epinephelus tauvina (Forskal)*. Kuwait Institute for Scientific Research.
- Hussin, M. A., Nik-Haiha, N. Y., & Om, A. D. (2008). Status and Prospects of Grouper Aquaculture in Malaysia. *The aquaculture of groupers*. Asian Fisheries Society.
- Ina-Salwany, M. Y., Hishammuddin, H., Zulperi, Z., Salema, M., Karim, M., & Natrah, F. M. I. (2015). Elucidating the Probiotic Potential of Malaysian *Paenibacillus pabuli* Against *Vibrio alginolyticus* in *Artemia* Culture. *Asian Journal of Agricultural Research, 9, 223-236.*

- Inoue, T., Matsuzaki, S., & Tanaka, S. (1995). Cloning and sequence analysis of *Vibrio parahaemolyticus* ompK gene encoding a 26-kDa outer membrane protein, OmpK, that serves as receptor for a broad-host-range vibriophage, KVP40. *FEMS microbiology letters*, 134(2-3), 245-249.
- Ito, T., & Maeno, Y. (2015). Effect of booster shot and investigation of vaccination efficacy period against herpesviral haematopoietic necrosis (HVHN) in goldfish *Carassius auratus*. *Veterinary microbiology*, 175(1), 139-144.
- IUCN. 2004. 2004 IUCN Red List of Threatened Species. www.iucnredlist.org. Downloaded on 4 December 2015.
- IUCN. 2013. 2013 IUCN Red List of Threatened Species. Version 2013.1.14. www.iucnredlist.org. Downloaded on 14 November 2013.
- James, C. M., Al-Thobaiti, S. A., Rasem, B. M., & Carlos, M. H. (1999). Potential of grouper hybrid (*Epinephelus fuscoguttatus* x *E. polyphkadion*) for Aquaculture. *Naga, the ICLARM Quarterly*, 22(1), 19-23.
- Jeff D. (2008). Bacterial outer membrane, <https://commons.wikimedia.org/w/index.php?curid=3647374> Own work, GFDL , Wikipedia
- Ji, J., Torrealba, D., Ruyra, A., & Roher, N. (2015). Nanodelivery systems as new tools for immunostimulant or vaccine administration: Targeting the fish immune system. *Biology*, 4(4), 664-696.
- Jiang, S., Wu, X., Li, W., Wu, M., Luo, Y., Lu, S., & Lin, H. (2015). Effects of dietary protein and lipid levels on growth, feed utilization, body and plasma biochemical compositions of hybrid grouper (*Epinephelus lanceolatus*♂ × *Epinephelus fuscoguttatus*♀) juveniles. *Aquaculture*, 446, 148-155.
- Jiang, Z., Huang, A. L., Tao, X. H., & Wang, P. L. (2003). Construction and characterization of bivalent vaccine candidate expressing HspA and M (r) 18,000 OMP from *Helicobacter pylori*. *World journal of gastroenterology: WJG*, 9(8), 1756-1761.
- Jobling, M. (2003). Diseases and Disorders of Finfish in Cage Culture, Woo, PKT, DW Bruno and LHS Lim (eds). *Aquaculture International*, 11(3), 320-322.

- Kahla-Nakbi, A. B., Besbes, A., Chaieb, K., Rouabhia, M., & Bakhrouf, A. (2007). Survival of *Vibrio alginolyticus* in seawater and retention of virulence of its starved cells. *Marine environmental research*, 64(4), 469-478.
- Kai, Y. H., & Chi, S. C. (2008). Efficacies of inactivated vaccines against betanodavirus in grouper larvae (*Epinephelus coioides*) by bath immunization. *Vaccine*, 26(11), 1450-1457.
- Kato, G., Takano, T., Sakai, T., Matsuyama, T., Sano, N., & Nakayasu, C. (2015). Cloning and expression analyses of a unique IgT in ayu *Plecoglossus altivelis*. *Fisheries Science*, 81(1), 29-36.
- Kawai, K., Liu, Y., Ohnishi, K., & Oshima, S. I. (2004). A conserved 37 kDa outer membrane protein of *Edwardsiella tarda* is an effective vaccine candidate. *Vaccine*, 22(25), 3411-3418.
- Kechik, I. B. A. (1995). Aquaculture in Malaysia. *Towards sustainable aquaculture in Southeast Asia and Japan: Proceedings of the Seminar-Workshop on Aquaculture Development in Southeast Asia, Iloilo City, Philippines, 26-28 July, 1994* (pp. 125-135). SEAFDEC Aquaculture Department.
- Khalil, R. H., & El-Latif, A. H. M. (2013). Effect of *Vibrio alginolyticus* on *Mugil capito*. *Journal of the Arabian Aquaculture Society*, 8.
- Kibbe, W. A. (2007). OligoCalc: an online oligonucleotide properties calculator. *Nucleic acids research*, 35(suppl 2), W43-W46.
- Kiryakit, A., Gallardo, W. G., & Bart, A. N. (2011). Successful hybridization of groupers (*Epinephelus coioides* x *Epinephelus lanceolatus*) using cryopreserved sperm. *Aquaculture*, 320(1), 106-112.
- Komar, C., Enright, W. J., Grisez, L., & Tan, Z. (2004). Understanding fish vaccination. *AQUA Culture AsiaPacific Magazine*, 27-29.
- Kua, B., Ramly, R., Devakie, M., Groman, D. & Berthe, C. (2011). Investigating a mortality in hatchery cultured tropical abalone, *Haliotis asinina* Linnaeus, 1758 in Malaysia. *Diseases in Asian Aquaculture VII*. Fish Health Section, Asian Fisheries Society, Selangor, Malaysia: 103-110.
- Kumar, S., Ray, P., Singh, H., & Ganguly, N. K. (2001). Immunogenicity and protective role of an IgA reactive 31-kDa antigen of *Vibrio cholerae* O139. *Journal of medical microbiology*, 50(6), 489-498.

- Kurupati, P., Teh, B. K., Kumarasinghe, G., & Poh, C. L. (2006). Identification of vaccine candidate antigens of an ESBL producing *Klebsiella pneumoniae* clinical strain by immunoproteome analysis. *Proteomics*, 6(3), 836-844.
- Lee, D. G., Yang, Y. S., & Kang, J. G. (2013). Conveyor belt automatic vaccine injection system (AVIS) for flatfish based on computer vision analysis of fish shape. *Aquacultural engineering*, 57, 54-62.
- Lee, K. K. (1995). Pathogenesis studies on *Vibrio alginolyticus* in the grouper, *Epinephelus malabaricus*, Bloch et Schneider. *Microbial Pathogenesis*, 19(1), 39-48.
- Lee, K. K., Yu, S. R., Chen, F. R., Yang, T. I., & Liu, P. C. (1996). News & notes: virulence of *Vibrio alginolyticus* isolated from diseased tiger prawn, *Penaeus monodon*. *Current Microbiology*, 32(4), 229-231.
- Leong, J. C., & Fryer, J. L. (1993). Viral vaccines for aquaculture. *Annual Review of Fish Diseases*, 3, 225-240.
- Leong, J. C., Anderson, E., Bootland, L. M., Chiou, P. W., Johnson, M., Kim, C., Maurice, D., & Trobridge, G. (1996). Fish vaccine antigens produced or delivered by recombinant DNA technologies. *Developments in biological standardization*, 90, 267-277.
- Leong, T. S., & Wong, S. Y. (1988). A comparative study of the parasite fauna of wild and cultured grouper (*Epinephelus malabaricus* Bloch et Schneider) in Malaysia. *Aquaculture*, 68(3), 203-207.
- Leung, K. Y. (2004). Current trends in the study of bacterial and viral fish and shrimp diseases. *World Scientific*, 3.
- Li, C., Ye, Z., Wen, L., Chen, R., Tian, L., Zhao, F., & Pan, J. (2014). Identification of a novel vaccine candidate by immunogenic screening of *Vibrio parahaemolyticus* outer membrane proteins. *Vaccine*, 32(46), 6115-6121.
- Li, N., Yang, Z., Bai, J., Fu, X., Liu, L., Shi, C., & Wu, S. (2010). A shared antigen among *Vibrio* species: outer membrane protein-OmpK as a versatile Vibriosis vaccine candidate in Orange-spotted grouper (*Epinephelus coioides*). *Fish & shellfish immunology*, 28(5), 952-956.
- Liao, I.C. & Leño, E.M. (Eds.). (2008). *The aquaculture of groupers*. Asian Fisheries Society.

- Lightner, D. V. (1993). Diseases of cultured penaeid shrimp. *CRC handbook of mariculture*, 1, 393-486.
- Liu, X. F., Cao, Y., Zhang, H. L., Chen, Y. J., & Hu, C. J. (2015). Complete genome sequence of *Vibrio alginolyticus* ATCC 17749T. *Genome announcements*, 3(1), e01500-14.
- Lotz, M. J., Tamplin, M. L., & Rodrick, G. E. (1983). Thiosulfate-citrate-bile salts-sucrose agar and its selectivity for clinical and marine vibrio organisms. *Annals of Clinical & Laboratory Science*, 13(1), 45-48.
- Luin, M., Fui, C. F., & Senoo, S. (2013). Sexual maturation and gonad development in tiger grouper (*Epinephelus fuscoguttatus*) x giant grouper (*E. lanceolatus*) hybrid. *Journal of Aquaculture Research & Development*, 5(213), 2.
- Lun, J., Xia, C., Yuan, C., Zhang, Y., Zhong, M., Huang, T., & Hu, Z. (2014). The outer membrane protein, LamB (maltoporin), is a versatile vaccine candidate among the *Vibrio* species. *Vaccine*, 32(7), 809-815.
- Maiti, B., Raghunath, P., & Karunasagar, I. (2009). Cloning and expression of an outer membrane protein OmpW of *Aeromonas hydrophila* and study of its distribution in *Aeromonas* spp. *Journal of applied microbiology*, 107(4), 1157-1167.
- Maiti, B., Shetty, M., Shekar, M., Karunasagar, I., & Karunasagar, I. (2012). Evaluation of two outer membrane proteins, Aha1 and OmpW of *Aeromonas hydrophila* as vaccine candidate for common carp. *Veterinary immunology and immunopathology*, 149(3), 298-301.
- Malik, V. S. (1981). *Recombinant DNA technology*. Academic Press.
- Mao, Z., Yu, L., You, Z., Wei, Y., & Liu, Y. (2007). Cloning, expression and immunogenicity analysis of five outer membrane proteins of *Vibrio parahaemolyticus* zj2003. *Fish & shellfish immunology*, 23(3), 567-575.
- Marino, G., Di Marco, P., Massari, A., Bottero, S., & Mandich, A. (2002). Effects of temperature and social control on sexual maturity and sex inversion in captive dusky grouper *Epinephelus marginatus*. In *Proceedings of the International Conference of Aquaculture Europe*, 16, 331-332.
- Marte, C. L. (1999). 5.20 Grouper Research at the Southeast Asian Fisheries Development Center Aquaculture Department. In *Cooperative Grouper Aquaculture Workshop* (p. 143).

- Martins, M. L., Mouriño, J. L. P., Fezer, G. F., Buglione Neto, C. C., Garcia, P., Silva, B. C., Jatobá, A. & Vieira, F. N. (2010). Isolation and experimental infection with *Vibrio alginolyticus* in the sea horse, *Hippocampus reidi* Ginsburg, 1933 (Osteichthyes: Syngnathidae) in Brazil. *Brazilian Journal of Biology*, 70(1), 205-209.
- McInerney, P., Adams, P., & Hadi, M. Z. (2014). Error rate comparison during polymerase chain reaction by DNA polymerase. *Molecular biology international*, 2014.
- Meri, S. (2016). Self-nonsel self discrimination by the complement system. *FEBS letters*, 590(15), 2418-2434.
- Mu, Y., Lin, K., Chen, X., & Ao, J. (2013). Diagnosis of nervous necrosis virus in orange-spotted grouper, *Epinephelus coioides*, by a rapid and convenient RT-PCR method. *Acta Oceanologica Sinica*, 32(10), 88-92.
- Mukai, Y., Lim, L. S., Lu, K. C., Abdul Rashid, M. K., & Saad, S. (2012). Light Intensity Requirements for Feeding Behaviour by the Brown-marbled Grouper, *Epinephelus fuscoguttatus*. *Sains Malaysiana*, 41(10), 1193-1196.
- Musa, N., & Wei, L. S. (2008). Outbreak of Vibriosis in Mantis Shrimp (*Squilla* sp.). *World Journal of Agricultural Sciences*, 4(2), 137-139.
- Mustapha, S., Mustapha, E. M., & Nozha, C. (2013). *Vibrio alginolyticus*: an emerging pathogen of food borne diseases. *International Journal of Science and Technology*, 2(4).
- Nagasawa, K., & Cruz-Lacierda, E. R. (2004). *Diseases of cultured groupers*. Iloilo, Philippines: Southeast Asian Fisheries Development Center, Aquaculture Department.
- Nascimento, I. P., & Leite, L. C. C. (2012). Recombinant vaccines and the development of new vaccine strategies. *Brazilian Journal of Medical and Biological Research*, 45(12), 1102-1111.
- Nash, G., Anderson, I. G., Shariff, M., & Shamsudin, M. N. (1987). Bacteriosis associated with epizootic in the giant sea perch, *Lates calcarifer*, and the estuarine grouper, *Epinephelus tauvina*, cage cultured in Malaysia. *Aquaculture*, 67(1), 105-111.
- Nazifah, N. M. (2013). *Development and evaluation of recombinant vector cells carrying cell wall surface anchor family proteins as a vaccine against Streptococcosis in red hybrid tilapia (Oreochromis spp)*. (Doctor of Philosophy dissertation), Universiti Putra Malaysia, Malaysia.

- Newman, S. G. (1993). Bacterial vaccines for fish. *Annual Review of Fish Diseases*, 3, 145-185.
- Ningqiu, L., Junjie, B., Shuqin, W., Xiaozhe, F., Haihua, L., Xing, Y., & Cunbin, S. (2008). An outer membrane protein, OmpK, is an effective vaccine candidate for *Vibrio harveyi* in Orange-spotted grouper (*Epinephelus coioides*). *Fish & shellfish immunology*, 25(6), 829-833.
- Nur-Nazifah, M., Sabri, M. Y., & Siti-Zahrah, A. (2014). Development and efficacy of feed-based recombinant vaccine encoding the cell wall surface anchor family protein of *Streptococcus agalactiae* against streptococcosis in *Oreochromis* sp. *Fish & shellfish immunology*, 37(1), 193-200.
- Osman, K. M., Mohamed, L. A., Rahman, E. H. A., & Soliman, W. S. (2009). Trials for vaccination of Tilapia fish against *Aeromonas* and *Pseudomonas* infections using monovalent, bivalent and polyvalent vaccines. *World Journal of Fish and Marine Sciences*, 1(4), 297-304.
- Pakingking, R., Bautista, N. B., de Jesus-Ayson, E. G., & Reyes, O. (2010). Protective immunity against viral nervous necrosis (VNN) in brown-marbled grouper (*Epinephelus fuscoguttatus*) following vaccination with inactivated betanodavirus. *Fish & shellfish immunology*, 28(4), 525-533.
- Pears, R. J., Choat, J. H., Mapstone, B. D., & Begg, G. A. (2006). Demography of a large grouper, *Epinephelus fuscoguttatus*, from Australia's Great Barrier Reef: implications for fishery management. *Marine Ecology Progress Series*, 307, 259-272.
- Pfeifer, B. A., Admiraal, S. J., Gramajo, H., Cane, D. E., & Khosla, C. (2001). Biosynthesis of complex polyketides in a metabolically engineered strain of *E. coli*. *Science*, 291(5509), 1790-1792.
- NOAA Fisheries, National Marine Fisheries Service, Fisheries Statistics Division. [online] [Cited 2 December 2015]. http://www.nmfs.noaa.gov/aquaculture/what_is_aquaculture.html
- Plant, K. P., & LaPatra, S. E. (2011). Advances in fish vaccine delivery. *Developmental & Comparative Immunology*, 35(12), 1256-1262.
- Qian, R. H., Xiao, Z. H., Zhang, C. W., Chu, W. Y., Wang, L. S., Zhou, H. H., Wei, Y. W., & Yu, L. (2008a). A conserved outer membrane protein as an effective vaccine candidate from *Vibrio alginolyticus*. *Aquaculture*, 278(1), 5-9.

- Qian, R., Xiao, Z., Zhang, C., Chu, W., Mao, Z., & Yu, L. (2008b). Expression and purification of two major outer membrane proteins from *Vibrio alginolyticus*. *World Journal of Microbiology and Biotechnology*, 24(2), 245-251.
- Qian, R., Chu, W., Mao, Z., Zhang, C., Wei, Y., & Yu, L. (2007). Expression, characterization and immunogenicity of a major outer membrane protein from *Vibrio alginolyticus*. *Acta biochimica et biophysica Sinica*, 39(3), 194-200.
- Qin, Y., Wang, J., Wang, S., & Yan, Q. (2007). Antigenicity analysis of *Vibrio harveyi* TS-628 strain. *Frontiers of Biology in China*, 2(3), 263-267.
- Rameshkumar, P., Kalidas, C., Tamilmani, G., Sakthivel, M., Abdul Nazar, A. K., Ashok Maharshi, V., Srinivasa Rao, S. K., & Gopakumar, G. (2014). Microbiological and histopathological investigations of *Vibrio alginolyticus* infection in cobia *Rachycentron canadum* (Linnaeus, 1766) cultured in sea cage. *Indian Journal of Fisheries*, 61(1), 124-127.
- Reed, P. A. & Francis-Floyd, R. (2009). *Vibrio* infections of fish. *Florida: University of Florida*.
- RUMA, *Responsible use of vaccines and vaccination in fish production; The Responsible Use of Medicines in Agriculture Alliance (RUMA) Guidelines: UK, 2006*.
- Rombout, J. H., & Kiron, V. (2014). Mucosal Vaccination of Fish. *Fish Vaccination*, 56.
- Rosano, G. L., & Ceccarelli, E. A. (2014). Recombinant protein expression in *Escherichia coli*: advances and challenges. *Frontiers in microbiology*, 5.
- Rosano, G. L., Bruch, E. M., & Ceccarelli, E. A. (2011). Insights into the CLP/HSP100 chaperone system from chloroplasts of *Arabidopsis thaliana*. *Journal of Biological Chemistry*, 286(34), 29671-29680.
- Ryo, S., Wijdeven, R. H., Tyagi, A., Hermsen, T., Kono, T., Karunasagar, I., Rombout, J. H., Sakai, M., Verburg-van Kemenade, B. L., & Savan, R. (2010). Common carp have two subclasses of bonyfish specific antibody IgZ showing differential expression in response to infection. *Developmental & Comparative Immunology*, 34(11), 1183-1190.
- Sabir, M., Cohen, N., Boukhanjer, A., & Ennaji, M. M. (2010). Occurrence and survival of *Vibrio alginolyticus* in Tamouda Bay (Morocco). *Cellular and molecular biology (Noisy-le-Grand, France)*, 57, OL1592-9.

- Sabri, M. Y. (1999). *Immunology of the outer membrane proteins of Pasteurella haemolytica A2, A7 and A9 in sheep*. (Master of Science dissertation), Universiti Putra Malaysia, Malaysia.
- Sadovy, Y. J., Donaldson, T. J., Graham, T. R., McGilvray, F., Muldoon, G. J., Phillips, M. J., Rimmer, M. A., Smith, A., & Yeeting, B. (2003). *While stocks last: The live reef food fish trade*. Asian Development Bank. <http://hdl.handle.net/11540/2432>. License: CC BY 3.0 IGO.
- Sakazaki, R. (1968). Proposal of *Vibrio alginolyticus* for the biotype 2 of *Vibrio parahaemolyticus*. *Japanese Journal of Medical Science and Biology*, 21(5), 359-362.
- Saleema, M. (2015). *Molecular Characterization of Aeromonas hydrophila and Development of Recombinant Cells Vaccine Expressing Outer Membrane Proteins Against It in African Catfish (Clarias gariepinus Burchell, 1822)*. (Master of Science dissertation), Universiti Putra Malaysia, Malaysia.
- Salinas, I. (2015). The Mucosal Immune System of Teleost Fish. *Biology*, 4(3), 525-539.
- Salosso, Y. & Jasmanindar, Y. (2014). Potential of Patikan Kerbau (*Euphorbia hirta*) as Antibacterial on *Aeromonas hydrophilla* and *Vibrio alginolyticus* in Fish Culture. *Aquatic Science and Technology*, 2(1), 63-72.
- Sambrook, J., & Russell, D. W. (2001). *Molecular cloning: a laboratory manual*, 3rd edn. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY., USA., ISBN-13: 9780879695774.
- Samoilys, M., Macharia, D., Robinson, J., Maina, G. W., & Bijoux, J. (2015). Confirmed Sighting of a Spawning Aggregation of the Brown-marbled Grouper, *Epinephelus fuscoguttatus*, in Kenya. *Western Indian Ocean Journal of Marine Science*, 13(2), 189-203.
- Sarjito, S., Radjasa, O. K., Sabdono, A., Prayitno, S. B. & Hutabarat, S. (2009). Phylogenetic diversity of the causative agents of vibriosis associated with groupers fish from Karimunjawa Islands, Indonesia. *Current Research in Bacteriology*, 2(1), 14-21.
- Savan, R., Aman, A., Nakao, M., Watanuki, H., & Sakai, M. (2005). Discovery of a novel immunoglobulin heavy chain gene chimera from common carp (*Cyprinus carpio* L.). *Immunogenetics*, 57(6), 458-463.
- Schmidt, T. M., DeLong, E. F., & Pace, N. R. (1991). Analysis of a marine picoplankton community by 16S rRNA gene cloning and sequencing. *Journal of bacteriology*, 173(14), 4371-4378.

- Sela, M. (1969). Antigenicity: some molecular aspects. *Science*, 166(3911), 1365-1374.
- Shields, R. J. (2001). Larviculture of marine finfish in Europe. *Aquaculture*, 200(1), 55-88.
- Silva, Y. J., Costa, L., Pereira, C., Mateus, C., Cunha, Â., Calado, R., Gomes, N. c. M., Pardo, M. A., Hernandez, I., & Almeida, A. (2014). Phage therapy as an approach to prevent *Vibrio anguillarum* infections in fish larvae production. *PloS one*, 9(12), e114197.
- Sluka, R.D. (2002). Grouper and Napoleon wrasse ecology in Laamu Atoll, Republic of Maldives: part 3. Fishing effects and management of the live fish-food trade. *Atoll Research Bulletin*, 481-493.
- Sørensen, H. P., & Mortensen, K. K. (2005). Advanced genetic strategies for recombinant protein expression in *Escherichia coli*. *Journal of biotechnology*, 115(2), 113-128.
- Sokolosky, J. T., & Szoka, F. C. (2013). Periplasmic production via the pET expression system of soluble, bioactive human growth hormone. *Protein expression and purification*, 87(2), 129-135.
- Sommerset, I., Krossøy, B., Biering, E., & Frost, P. (2014). Vaccines for fish in aquaculture. *Expert review of vaccines*.
- Sonia, A. S. G., & Lipton, A. P. (2012). Pathogenicity and antibiotic susceptibility of *Vibrio* species isolated from the captive-reared tropical marine ornamental blue damsel fish, *Pomacentrus caeruleus* (Quoy and Gaimard, 1825). *Indian Journal of Geo-Marine Sciences*, 41(4), 348-354.
- Sugama, K., Rimmer, M. A., Ismi, S., Koesharyani, I., Suwirya, K., Giri, N. A., & Alava, V. R. (2012). *Hatchery management of tiger grouper (Epinephelus fuscoguttatus): a best-practice manual*. Australian Centre for International Agricultural Research.
- Tafalla, C., Bøgwald, J., & Dalmo, R. A. (2013). Adjuvants and immunostimulants in fish vaccines: current knowledge and future perspectives. *Fish & shellfish immunology*, 35(6), 1740-1750.
- Tendencia, E. A. & Lavilla-Pitogo, C. R. (2004). Bacterial Diseases. *Diseases of cultured groupers*. Asian Fisheries Society. Iloilo, Philippines: Southeast Asian Fisheries Development Center, Aquaculture Department.

- Terpe, K. (2006). Overview of bacterial expression systems for heterologous protein production: from molecular and biochemical fundamentals to commercial systems. *Applied microbiology and biotechnology*, 72(2), 211-222.
- Thye, C. T. (2001). Survey of Grouper Diseases in Malaysia. *Development of a Regional Research Programme on Grouper Virus Transmission and Vaccine Development*, 38.
- Tillett, D., & Neilan, B. A. (1999). Enzyme-free cloning: a rapid method to clone PCR products independent of vector restriction enzyme sites. *Nucleic acids research*, 27(19), e26-e28.
- Tinsley, J. W., Lyndon, A. R., & Austin, B. (2011). Antigenic and cross-protection studies of biotype 1 and biotype 2 isolates of *Yersinia ruckeri* in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of applied microbiology*, 111(1), 8-16.
- Toranzo, A. E., Santos, Y. & Barja, J. L. (1996). Immunization with bacterial antigens: *Vibrio* infections. *Developments in biological standardization*, 90, 93-105.
- Trichet, V. V., Santigosa, E., Cochin, E., & Gabaudan, J. (2015). The Effect of Vitamin C on Fish Health. *Dietary Nutrients, Additives, and Fish Health*, 151-171. John Wiley & Sons.
- Tseng, W. Y., & Poon, C. T. (1983). Hybridization of *Epinephelus* species. *Aquaculture*, 34(1), 177-182.
- Ukawa, M., & Higuchi, M. S. mito, (1966). Spawning habits and early life history of a serranid fish, *Epinephelus akaara* (Temmincket Schelegel). *Japanese Journal of Ichthyology*, 13(4/6), 156-161.
- Van Den Ent, F., & Löwe, J. (2006). RF cloning: a restriction-free method for inserting target genes into plasmids. *Journal of biochemical and biophysical methods*, 67(1), 67-74.
- Vesely, T., Reschova, S., Pokorova, D., Hulova, J., & Nevorankova, Z. (2006). Production of monoclonal antibodies against immunoglobulin heavy chain in common carp (*Cyprinus carpio* L.). *Veterinarni Medicina-Praha*, 51(5), 296.
- Vine, N. G. (2005). *Towards the development of a protocol for the selection of probiotics in marine fish larviculture*. (Doctoral dissertation), Rhodes University, South Africa.

- Wei, S., Zhao, H., Xian, Y., Hussain, M. A., & Wu, X. (2014). Multiplex PCR assays for the detection of *Vibrio alginolyticus*, *Vibrio parahaemolyticus*, *Vibrio vulnificus*, and *Vibrio cholerae* with an internal amplification control. *Diagnostic microbiology and infectious disease*, 79(2), 115-118.
- Wiik, R., Stackebrandt, E., Valle, O., Daae, F. L., RØDSETH, O. M., & Andersen, K. (1995). Classification of fish-pathogenic vibrios based on comparative 16S rRNA analysis. *International Journal of Systematic and Evolutionary Microbiology*, 45(3), 421-428.
- Wilson, M. R., & Warr, G. W. (1992). Fish immunoglobulins and the genes that encode them. *Annual Review of Fish Diseases*, 2, 201-221.
- Wolters, W. R., & Johnson, M. R. (1994). Enteric septicemia resistance in blue catfish and three channel catfish strains. *Journal of Aquatic Animal Health*, 6(4), 329-334.
- Woo, P. T., Leatherland, J. F., & Bruno, D. W. (Eds.). (2011). *Fish diseases and disorders* (Vol. 3). CABI.
- Xie, Z. Y., Hu, C. Q., Chen, C., Zhang, L. P., & Ren, C. H. (2005). Investigation of seven *Vibrio* virulence genes among *Vibrio alginolyticus* and *Vibrio parahaemolyticus* strains from the coastal mariculture systems in Guangdong, China. *Letters in applied microbiology*, 41(2), 202-207.
- Yii, K. C., Yang, T. I., & Lee, K. K. (1997). Isolation and characterization of *Vibrio carchariae*, a causative agent of gastroenteritis in the groupers, *Epinephelus coioides*. *Current Microbiology*, 35(2), 109-115.
- Zhang, C., Yu, L., & Qian, R. (2007). Characterization of OmpK, GAPDH and their fusion OmpK-GAPDH derived from *Vibrio harveyi* outer membrane proteins: their immunoprotective ability against vibriosis in large yellow croaker (*Pseudosciaena crocea*). *Journal of applied microbiology*, 103(5), 1587-1599.
- Zhang, Z., Wu, H., Xiao, J., Wang, Q., Liu, Q., & Zhang, Y. (2012). Immune responses of zebrafish (*Danio rerio*) induced by bath-vaccination with a live attenuated *Vibrio anguillarum* vaccine candidate. *Fish & shellfish immunology*, 33(1), 36-41.
- Zhou, F., Shefer, A., Weinbaum, C., McCauley, M., & Kong, Y. (2007a). Impact of hepatitis A vaccination on health care utilization in the United States, 1996–2004. *Vaccine*, 25(18), 3581-3587.

Zhou, S., Hou, Z., Li, N., & Qin, Q. (2007b). Development of a SYBR Green I real-time PCR for quantitative detection of *Vibrio alginolyticus* in seawater and seafood. *Journal of applied microbiology*, 103(5), 1897-1906.

Zorrilla, I., Morínigo, M. A., Castro, D., Balebona, M. C. & Borrego, J. J. (2003). Intraspecific characterization of *Vibrio alginolyticus* isolates recovered from cultured fish in Spain. *Journal of applied microbiology*, 95(5), 1106-1116.



LIST OF PUBLICATIONS

- Nehlah, R., Ina-Salwany, M. Y., & Zulperi, Z. (2016). Antigenicity Analysis and Molecular Characterization of Two Outer Membrane Proteins of *Vibrio alginolyticus* Strain VA2 as Vaccine Candidates in Tiger Grouper Culture. *Journal of Biological Sciences*, 16(1), 1.
- Rabi'atul, A. S, Diyana-Nadhirah, K. P., Nehlah, R., Saleema, M., Zarirah. Z., & Ina-Salwany, M.Y. (2015). Internal Transcribed Spacer (ITS) Gene Sequencing and Pathogenicity Study of *Aeromonas veronii* bv. *sobria* Strain KTG3SB in Juvenile Red Tilapia. *National Seminar on Advances in Fish Health 2015*. 4th - 5th February 2015, Kajang, Selangor, Malaysia.
- Nehlah Rosli, Ina Salwany Md Yasin, Murni Karim, Nur Nazifah, Siti-Zahrah Abdullah (2014). Molecular characterization and antigenicity of outer membrane protein (OMP) of *Vibrio alginolyticus* isolated from diseased Tiger grouper (*Epinephelus fuscoguttatus*). *9th Symposium on Diseases in Asian Aquaculture (DAA9)*. 24th - 28th November 2014. Ho Chi Minh City, Vietnam.



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