

UNIVERSITI PUTRA MALAYSIA

PREPARATION AND CHARACTERIZATION OF SILVER/CHITOSAN/GELATIN BIONANOCOMPOSITES BY CHEMICAL REDUCING METHOD AND THEIR ANTIBACTERIAL ACTIVITIES

LIM JENN JYE

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Ву

LIM JENN JYE

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirements for the Degree of Master of Science

January 2013

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Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PREPARATION AND CHARACTERIZATION OF SILVER/CHITOSAN/GELATIN BIONANOCOMPOSITES BY CHEMICAL REDUCING METHOD AND THEIR ANTIBACTERIAL ACTIVITIES

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January 2013

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The aim of this study is to investigate the functions of polymers and size of nanoparticles on the antibacterial activity of silver bionanocomposites (Ag BNCs). In this research, silver nanoparticles (Ag NPs) were incorporated into biodegradable polymers, which are chitosan (Cts), gelatin and both polymers via chemical reduction method in solvent in order to produce Ag BNCs. Silver nitrate and sodium borohydride were employed as a metal precursor and reducing agent respectively. In addition, chitosan and gelatin were added as both polymeric matrix and stabilizer. The properties of Ag BNCs were studied based on function of the polymer weight ratio in relation to the use of chitosan and gelatin. These prepared Ag NPs were very stable over a long period (i.e. 4 months) in an aqueous solution without any sign of precipitants. The UV-vis spectra shown maximum absorbance bands for Ag BNCs were detected at 408, 416, 414, 413, and 414 nm, respectively. From the FTIR spectra, the vibrational band of Ag BNCs detected at ~1395 cm⁻¹ could indicate the interaction between

Ag NPs with chitosan and gelatin. The morphology of the Ag BNCs films and the distribution of the Ag NPs were characterized using Scanning Electron Microscopy (SEM) and Transmission Electron Microscopy (TEM). From the SEM image, Ag/Cts BNCs shown layered surfaces with small flakes, Ag/Cts/gelatin BNCs showed smooth layered surfaces and Ag/gelatin BNCs showed porous layered surfaces. In addition, the EDX spectra for Ag BNCs had confirmed the presence of elemental compounds in the Cts, gelatin and Ag NPs without any impurity peaks. From the TEM images, the results of average diameters of the Ag NPs in different weight composition of polymers were in the range of 2.90-11.25 nm. However, the TEM results also shown that aggregation of Ag NPs where there is an overlap of particles and larger particle size in chitosan only which contributed to the size of 11.25 nm compared with others samples which are 2.90 nm-4.38 nm. With the aid of XRD pattern, Ag BNCs films confirmed the face-centered cubic (FCC) type crystallographic planes of the Ag crystal. The XRD peaks at 20 with value of around 38°, 44°, 64°, and 77° are well recognized to the 111, 200, 220, and 311 crystallographic planes of the FCC Ag crystals, respectively. The antibacterial activities of the Ag BNC films were examined against Gram-negative bacteria (E. coli and P. aeruginosa) and Gram-positive (S. aureus and M. luteus). The silver ions released from the Ag BNCs and their antibacterial activities were scrutinized. From TEM analysis and the result of antibacterial activity, Ag/Cts/gelatin BNCs exhibited better particle distribution as well as having high antibacterial activity. A simple way to prepare Ag/Cts/gelatin bionnanocomposites had been employed as previously there is

no research on investigating the antibacterial activities of silver into both chitosan and gelatin.



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PENYEDIAAN DAN PENCIRIAN BIONANOKOMPOSIT PERAK / KITOSAN / GELATIN DARI KAEDAH PENURUNAN KIMIA DAN AKTIVITI ANTIBAKTERIA

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Tujuan kajian ini adalah untuk menyelidik fungsi polimer dan saiz nanopartikel dalam aktiviti antibakteria perak bionanokomposit (Ag BNCs). Dalam penyelidikan ini, nanopartikel perak (Ag NPs) telah dimasukkan ke dalam polimer yang biodigradasi iaitu kitosan, gelatin dan kedua-dua polimer melalui kaedah penurunan kimia dalam pelarut untuk menghasilkan Ag BNCs. Argentum nitrat dan natrium borohydride telah digunakan sebagai prekursor logam dan ejen penurunan masing-masing. Sebaliknya, kitosan (Cts) dan gelatin telah ditambah sebagai matriks polimer dan penstabil. Ciri-ciri Ag BNCs telah dikaji berdasarkan nisbah berat kitosan dan gelatin. Ag NPs yang disediakan adalah sangat stabil dalam tempoh yang panjang (iaitu 4 bulan) dalam larutan tanpa sebarang tanda mendakan. Spektrum UV-nampak menunjukkan jalur penyerapan yang maximum untuk Ag BNCs telah dikesan pada 408,416,414,413, dan 414nm masing-masing. Dari specktrum jelmaan

fourier inframerah, gelombang getaran Ag BNCs dikesan pada ~1395 cm⁻¹ menunjukkan interaksi antara Ag NPs dengan kitosan dan gelatin. Morfologi filem Ag BNCs dan pengedaran Ag NPS turut dipercirikan menggunakan Mikroskop Pengimbasan Elektron (SEM) dan Mikroskop transmisi elektron (TEM). Dari gambar SEM, Ag/kitosan BNCs menunjukkan permukaan berlapis dengan serpihan kecil, Ag/kitosan/gelatin BNCs menunjukkan permukaan licin berlapis dan Ag/gelatin BNCs menunjukkan permukaan lapisan berliang. Sebagai tambahan, spektrum EDX bagi Ag BNCs telah mengesahkan kehadiran sebatian unsur dalam kitosan, gelatin dan Ag NPs tanpa sebarang puncak bendasing. Dari imej TEM, hasil diameter purata Ag NPs dalam komposisi berat polimer yang berbeza adalah dalam lingkungan 2.90-11.25 nm. Walau bagaimanapun, hasil TEM juga menunjukkan agregasi Ag NPs di mana terdapat pertindihan partikel dan saiz partikel yang lebih besar dalam kitosan sahaja yang menymbang kepada saiz 11.25 nm berbanding dengan sampel lain yang bersaiz 2.90-4.38 nm. Dengan bantuan corak XRD, Ag BNCs mengesahkan Ag NPs memiliki jenis satah kristalografi yang muka berpusat padu (FCC). Puncak-puncak XRD di paksi 20 dengan nilai kira kira 38°, 44°, 64°, dan 77° telah diiktiraf kepada 111, 200, 220, dan 311 satah kristalografi FCC masing-masing daripada Ag kristal. Aktiviti antibakteria yang Ag BNC filem dikaji terhadap bakteria Gram-negatif (E. coli dan P. aeruginosa) dan Grampositif (S. aureus dan M. luteus). Ion perak dikeluarkan dari BNCs Ag dan aktiviti antibakteria mereka telah diteliti. Daripada analisis TEM dan hasil aktiviti antibakteria, Ag/kitosan/gelatin BNCs mempamerkan pengagihan zarah yang

lebih baik serta mempunyai aktiviti antibakteria yang tinggi. Satu cara mudah untuk menyediakan Ag / Cts / gelatin bionnanocomposites telah digunakan sebab sebelum ini tidak ada penyelidikan yang mengkaji aktiviti antibakteria perak ke dalam kedua-dua kitosan dan gelatin.



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I certify that a Thesis Examination Committee has met on 3 January 2013 to conduct the final examination of Lim Jenn Jye on his thesis entitled "Preparation And Characterization Of Silver/Chitosan/ Gelatin Bionanocomposites By Chemical Reducing Method And Their Antibacterial Activities" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

LIM JENN JYE

Date: 3 January 2013

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LIST OF ABBREVIATIONS

Ag/Cts silver/chitosan

Ag/Cts/gelatin silver /chitosan/gelatin

Ag BNCs silver bionanocomposites

AgNO₃ silver nitrate

Ag NPs silver nanoparticles

BNCs bionanocomposites

Cts chitosan

Cts/gelatin chitosan/gelatin

E. coli Escherichia coli

EDX energy dispersive X-ray

FTIR Fourier transform infrared

HAC acetic acid

MHA Mueller-Hinton Agar

M. luteus Micrococcus luteus

MMT montmorillonite

NaBH₄ Sodium borohydride

NCs nanocomposites

NPs nanoparticles

SPR Surface Plasmon Resonance

S. aureus Staphylococcus aureus

SEM scanning electron microscopy

P.aeruginosa Pseudomonas aeruginosa

PVP Poly(vinylpyrrolidone)

PVA Poly(vinyl alcohol)

TEM transmission electron microscopy

UV-vis ultraviolent visible

XRD X-ray diffraction

CHAPTER 1

INTRODUCTION

1.1 Background of study

Nanotechnology is becoming a significantly important field, offering massive improvements to standards of living by developing the new nano-scaled materials. The term nanotechnology was first introduced by Professor Norio Taniguchi in the year 1974 to describe the accurate processing of a material to nano scale (Taniguchi, 1983). The term nanotechnology comes from the combination of two words: the Greek numerical prefix nano referring to a billionth and the word technology. As an outcome, nanotechnology is generally considered to be at a size below 100 nm. Nanoscience, sometimes defined as 'the science underlying nanotechnology' is related to the study of nanostructured materials which included process synthesis, the characterization, exploration, interrogation, exploitation, and utilization. Nanomaterials are materials that have structural components with at least one dimension smaller than 100 nm.

The nanomaterials enclose different group of nanostructured materials including quantum dots, clusters, nanocrystals, nanowires, and nanotubes (Logothetidis, 2012). Nanoparticles (NPs) may exhibit size-related properties that significantly diverge from those found in those molecular or bulk materials with the same chemical composition. As particles become increase smaller and enter the nanosize range, the numbers of atoms at the surface of the particle are far greater and more significant, thus altering the physical and chemical properties of the nanoparticles. Also, their physical-chemical properties can be significantly different mainly due to the nanometer size of the materials causing them to have high surface energy, spatial confinement, and reduced imperfections which these properties do not exist in the corresponding bulk materials (Cao, 2004). The fundamental of nanoscience and nanotechnology has two approaches. First, the bottom-up approach where materials are built molecular components which assemble themselves chemically principles of molecular recognition. Second, the top-down approach of the self assembly of molecular components, where each nanostructured component plugs itself into a suprastructure (Jortner et al., 2002).

Metal nanoparticles play important part in different research fields due to its unique chemical, optical, electrical, and thermal properties when their size is confined to the nanometer length range. They represent in the development of novel materials that can be used in various physical, chemical, biological (Zhang *et al.*, 2001), biomedical and pharmaceutical applications. The intrinsic

properties of metal nanoparticles are mainly determined by its size, composition, shape, structure, and crystallinity. Despite this, the challenge of synthetically controlling the parameter to tailor the antibacterial, electrical, and optical behaviours of metal nanoparticles has been met with limited success. Perfectly monodispersed metal NPs are ideal, but their unique properties are to be expected even if the ideality is not perfectly realized.

Composites can be defined consist of at least two materials with rather different properties, revealing heterogeneous microstructure and have improved properties with respect to their components. Nanocomposites (NCs) are multiphase solid material or structures where one of the phases has dimensions within nanometer range. Nanocomposites can be considered as solid structures with nanometer- scale dimensional repeat distance between the different phases. Typically, nanocomposites are classified as inorganic matrix (inorganic-inorganic), organic filler in organic (organic-organic), and hybrid materials, i.e., organic in inorganic or inorganic in organic matrix (Ajayan et al., 2003). The mechanical, optical, thermal, catalytic and others properties of the NCs differ noticeably from that of the component materials. Bionanocomposites (BNCs) has become a common term to designate those NCs involving a naturally occurring biopolymer with organic/inorganic nanomaterials. BNCs, an original invention of NCs materials, indicate a promising field in nanotechnology, materials and life sciences due to their

extraordinary advantages of biocompatibility and biodegradability (Darder *et al.*, 2007).

1.2 Problem statement

In previous research done by other researchers, there were reports on preparation of silver/chitosan bionanocomposites and silver/gelatin bionanocomposites. However, reports on the effect of antibacterial activity of Ag NPs in chitosan/gelatin which has potential application in biomedical field had yet to be investigated. Moreover, there has been no attempt to compare the properties of Ag NPs in Cts, gelatin and Cts/gelatin. In this study, Ag NPs were effectively prepared in the chitosan, gelatin and Cts/gelatin composite by using chemical reducing method. The Ag ions were successfully reduced by sodium borohydride (NaBH₄) into the polymers matrix forming Ag NPs. The characterization will be based on the different ratios of the polymer weights and the antibacterial activities will be carried out using MHA diffusion method.

1.3 Scope

The present research is an attempt to study the size, surface morphology, and antibacterial effect of Ag NPs in different weight ratio of polymer matrix. The crystalline structure, average size, particles distributions, surface morphology,

surface Plasmon resonance and functional groups of all the samples were characterized using X-ray diffraction (XRD), UV-visible spectroscopy, transmission electron microscopy (TEM), scanning electron microscopy (SEM), Energy-dispersive X-ray spectroscopy (EDX) and Fourier transform infrared (FTIR). The antibacterial activities for Ag NPs nanocomposites were investigated against Gram positive and Gram negative bacteria using Muller-Hinton Agar method.

1.4 Objectives

The main objectives of this research are to:

- 1. prepare Ag NPs by using chemical reducing agent in different weight ratios of Cts and gelatin.
- 2. characterize the crystalline structure, particle size and distribution, surface morphology, functional groups and the surface plasmon resonance of Ag BNCs obtained
- 3. investigate the antibacterial behaviour for different Ag NPs size against Gram positive and Gram negative bacteria by Mueller-Hinton Agar (MHA) test.

REFERENCES

- Abid, J.P., Wark, A.W., Brevet, P.F., & Girault, H.H. (2002). Preparation of silver nanoparticles in solution from a silver salt by laser irradiation. *Chem Commun (Camb)*(7), 792-793.
- Ajayan, P.M., Schadler, L.S., & Braun, P.V. (2003). Nanocomposite science and technology. *Wiley VCH*, 3-69.
- Bagheri-Khoulenjani, S., Mirzadeh, H., Etrati-Khosroshahi, M., & Shokrgozar, M.A. (2012). Particle size modeling and morphology study of chitosan/gelatin/nanohydroxyapatite nanocomposite microspheres for bone tissue engineering. *Journal of Biomedical Materials Research Part A*, 000-000.
- Baker, C., Pradhan, A., Pakstis, L., Pochan, D.J., & Shah, S.I. (2005). Synthesis and antibacterial properties of silver nanoparticles. *Journal of nanoscience and nanotechnology*, *5*(2), 244-249.
- Cao, G.Z. (2004). Nanostructures and Nanomaterials: Synthesis, Properties and Applications. *Imperial College Press*.
- Chen, J.P., & Lim, L.L. (2002). Key factors in chemical reduction by hydrazine for recovery of precious metals. *Chemosphere*, 49(4), 363-370.
- Chen, T., Embree, D.H., Brown, M.E., Taylor, M.M., & Payne, G.F. (2003). Enzyme-catalyzed gel formation of gelatin and chitosan: potential for in situ applications. *Biomaterials*, 24(17), 2831-2841.
- Chen, X., & Schluesener, H.J. (2008). Nanosilver: A nanoproduct in medical application. *Toxicology Letters*, 176(1), 1-12.
- Cheng, M., Deng, J., Yang, F., Gong, Y., Zhao, N., & Zhang, X. (2003). Study on physical properties and nerve cell affinity of composite films from chitosan and gelatin solutions. *Biomaterials*, *24*(17), 2871-2880.
- Choi, Y.S., Hong, S.R., Lee, Y.M., Song, K.W., Park, M.H., & Nam, Y.S. (1999a). Study on gelatin-containing artificial skin: I. Preparation and characteristics of novel gelatin-alginate sponge. *Biomaterials*, 20(5), 409-417.
- Choi, Y.S., Hong, S.R., Lee, Y.M., Song, K.W., Park, M.H., & Nam, Y.S. (1999b). Studies on gelatin-containing artificial skin: II. Preparation and

- characterization of cross-linked gelatin-hyaluronate sponge. *Journal of Biomedical Materials Research*, 48(5), 631-639.
- Darder, M., Aranda, P., & Ruiz-Hitzky, E. (2007). Bionanocomposites: A New Concept of Ecological, Bioinspired, and Functional Hybrid Materials. *Advanced Materials*, *19*(10), 1309-1319.
- Darroudi, M., Ahmad, M.B., Zak, A.K., Zamiri, R., & Hakimi, M. (2011a). Fabrication and Characterization of Gelatin Stabilized Silver Nanoparticles under UV-Light. *Int. J. Mol. Sci.*, *12*, 6346-6356.
- Darroudi, M., Ahmad, M.B., Abdullah, A.H., & Ibrahim, N.A. (2011b). Green synthesis and characterization of gelatin-based and sugar-reduced silver nanoparticles. *International Journal of Nanomedicin*, *6*, 569 574.
- Ershov, B.G., & Henglein, A. (1998). Reduction of Ag+ on Polyacrylate Chains in Aqueous Solution. *The Journal of Physical Chemistry B, 102*(52), 10663-10666.
- Evanoff, D.D., & Chumanov, G. (2004). Size-Controlled Synthesis of Nanoparticles. 1. "Silver-Only" Aqueous Suspensions via Hydrogen Reduction. *The Journal of Physical Chemistry B*, 108(37), 13948-13956.
- Feng, Q.L., Wu, J., Chen, G.Q., Cui, F.Z., Kim, T.N., & Kim, J.O. (2000). A mechanistic study of the antibacterial effect of silver ions on Escherichia coli and Staphylococcus aureus. *Journal of Biomedical Materials Research*, *52*(4), 662-668.
- Grzelczak, M., Vermant, J., Furst, E.M., & Liz-Marzán, L.M. (2010). Directed Self-Assembly of Nanoparticles. *ACS Nano, 4*(7), 3591-3605.
- Guzman, M., Dille, J., & Godet, S. (2012). Synthesis and antibacterial activity of silver nanoparticles against gram-positive and gram-negative bacteria. *Nanomedicine: Nanotechnology, Biology and Medicine, 8*(1), 37-45.
- Hadad, L., Perkas, N., Gofer, Y., Calderon-Moreno, J., Ghule, A., & Gedanken, A. (2007). Sonochemical deposition of silver nanoparticles on wool fibers. *Journal of Applied Polymer Science*, 104(3), 1732-1737.
- Harra, J., Mäkitalo, J., Siikanen, R., Virkki, M., Genty, G., Kobayashi, T., Kauranen, M. & Mäkelä, J.M. (2012). Size-controlled aerosol synthesis of silver nanoparticles for plasmonic materials. *Journal of Nanoparticle Research*, *14*(6), 1-10.
- Honarkar, H., & Barikani, M. (2009). Applications of biopolymers I: chitosan. *Monatshefte für Chemie / Chemical Monthly, 140*(12), 1403-1420.

- Hornebecq, V., Antonietti, M., Cardinal, T., & Treguer-Delapierre, M. (2003). Stable Silver Nanoparticles Immobilized in Mesoporous Silica. *Chemistry of Materials*, *15*(10), 1993-1999.
- Hu, B., Wang, S., Wang, K., Zhang, M., & Yu, S. (2008). Microwave-Assisted Rapid Facile "Green" Synthesis of Uniform Silver Nanoparticles: Self-Assembly into Multilayered Films and Their Optical Properties. *The Journal of Physical Chemistry C*, 112(30), 11169-11174.
- Huang H., & Yang, Y. (2008). Preparation of silver nanoparticles in inorganic clay suspensions. *Composites Science and Technology, 68*(14), 2948-2953.
- Huang H., Yuan, Q., & Yang, X. (2004). Preparation and characterization of metal–chitosan nanocomposites. *Colloids and Surfaces B: Biointerfaces*, 39(1–2), 31-37.
- Huang H., Yuan, Q., & Yang, X. (2005a). Morphology study of gold-chitosan nanocomposites. *Journal of Colloid and Interface Science*, 282(1), 26-31.
- Huang, N.M., Radiman, S., Lim, H.N., Khiew, P.S., Chiu, W.S., Lee, K.H., Syahda, A, Hashim, R., Chia, C.H. (2009). γ-Ray assisted synthesis of silver nanoparticles in chitosan solution and the antibacterial properties. *Chemical Engineering Journal*, *155*(1–2), 499-507.
- Huang, Y., Onyeri, S., Siewe, M., Moshfeghian, A., & Madihally, S.V. (2005b). In vitro characterization of chitosan–gelatin scaffolds for tissue engineering. *Biomaterials*, 26(36), 7616-7627.
- Hyning, D.L.V., & Zukoski, C.F. (1998). Formation Mechanisms and Aggregation Behavior of Borohydride Reduced Silver Particles. *Langmuir*, *14*(24), 7034-7046.
- Jacob, Ja.A., Mahal, H.S., Biswas, N., Mukherjee, T., & Kapoor, S. (2007). Role of Phenol Derivatives in the Formation of Silver Nanoparticles. *Langmuir*, *24*(2), 528-533.
- Jafari, M., Salavati-Niasari, M., & Mohandes, F. (2013). Synthesis and Characterization of Silver Selenide Nanoparticles via a Facile Sonochemical Route Starting from a Novel Inorganic Precursor. *Journal of Inorganic and Organometallic Polymers and Materials*, 23(2), 357-364.
- Jayakumar, R., Menon, D., Manzoor, K., Nair, S.V., & Tamura, H. (2010). Biomedical applications of chitin and chitosan based nanomaterials—A short review. *Carbohydrate Polymers*, 82(2), 227-232.

- Jensen, T., Kelly, L., Lazarides, A., & Schatz, G.C. (1999). Electrodynamics of Noble Metal Nanoparticles and Nanoparticle Clusters. *Journal of Cluster Science*, *10*(2), 295-317.
- Jeong, S.H., Hwang, Y.H., & Yi, S.C. (2005). Antibacterial properties of padded PP/PE nonwovens incorporating nano-sized silver colloids. *Journal of Materials Science*, 40(20), 5413-5418.
- Jortner, J., & Rao, C.N.R. (2002). Nanostructured advanced materials. Perspectives and directions. Perspectives and direction. *Pure Appl. Chem.*, 74, 15.
- Jung, J.H., Oh, H.C., Noh, H.S., Ji, J.H., & Kim, S.S. (2006). Metal nanoparticle generation using a small ceramic heater with a local heating area. *Journal of Aerosol Science*, *37*(12), 1662-1670.
- Jung, W.K., Koo, H.C., Kim, K.W., Shin, S., Kim, S.H., & Park, Y.H. (2008). Antibacterial activity and mechanism of action of the silver ion in *Staphylococcus aureus* and *Escherichia coli. Appl Environ Microbiol,* 74(7), 2171-2178.
- Kawahara, K., Suzuki, K., Ohko, Y., & Tatsuma, T. (2005). Electron transport in silver-semiconductor nanocomposite films exhibiting multicolor photochromism. *Physical Chemistry Chemical Physics*, 7(22), 3851-3855.
- Kelly, K.L., Coronado, E., Zhao, L.L., & Schatz, G.C. (2002). The Optical Properties of Metal Nanoparticles: The Influence of Size, Shape, and Dielectric Environment. *The Journal of Physical Chemistry B, 107*(3), 668-677.
- Khanna, P.K., Singh, N., Charan, S., Subbarao, V.V.V.S., Gokhale, R., & Mulik, U.P. (2005). Synthesis and characterization of Ag/PVA nanocomposite by chemical reduction method. *Materials Chemistry and Physics*, *93*(1), 117-121.
- Koodali, R.T., & Klabunde, K.J. (2012). Nanotechnology: Fundamental Principles and Applications. In J. A. Kent (Ed.), *Handbook of Industrial Chemistry and Biotechnology* (pp. 249-263): Springer US.
- Kumar, R., & Majeti, N.V. (2000). A review of chitin and chitosan applications. *Reactive and Functional Polymers*, *46*(1), 1-27.

- Lee, D., Cohen, R.E., & Rubner, M.F. (2005). Antibacterial Properties of Ag Nanoparticle Loaded Multilayers and Formation of Magnetically Directed Antibacterial Microparticles. *Langmuir*, *21*(21), 9651-9659.
- Lee, K., & El-Sayed, M.A. (2006). Gold and Silver Nanoparticles in Sensing and Imaging: Sensitivity of Plasmon Response to Size, Shape, and Metal Composition. *The Journal of Physical Chemistry B*, *110*(39), 19220-19225.
- Lin, X.Z., Teng, X., & Yang, H. (2003). Direct Synthesis of Narrowly Dispersed Silver Nanoparticles Using a Single-Source Precursor. *Langmuir*, *19*(24), 10081-10085.
- Liu, Y., Chen, S., Zhong, L., & Wu, G. (2009). Preparation of high-stable silver nanoparticle dispersion by using sodium alginate as a stabilizer under gamma radiation. *Radiation Physics and Chemistry*, 78(4), 251-255.
- Logothetidis, S. (2012). Nanotechnology: Principles and Applications. In S. Logothetidis (Ed.), *Nanostructured Materials and Their Applications* (pp. 1-22): Springer Berlin Heidelberg.
- Lok, C., Ho, C., Chen, R., He, Q., Yu, W., Sun, H., Tam, P.K., Chiu, J. & Che, C. (2006). Proteomic Analysis of the Mode of Antibacterial Action of Silver Nanoparticles. *Journal of Proteome Research*, *5*(4), 916-924.
- Marambio-Jones, C., & Hoek, E.M.V. (2010). A review of the antibacterial effects of silver nanomaterials and potential implications for human health and the environment. *Journal of Nanoparticle Research*, 12(5), 1531-1551.
- Martínez-Castañón, G.A., Niño-Martínez, N., Martínez-Gutierrez, F., Martínez-Mendoza, J.R., & Ruiz, F. (2008). Synthesis and antibacterial activity of silver nanoparticles with different sizes. *Journal of Nanoparticle Research*, *10*(8), 1343-1348.
- Mbhele, Z.H., Salemane, M.G., Sittert, C.G.C.E.V, Nedeljković, J.M., Djoković, V., & Luyt, A.S. (2003). Fabrication and Characterization of Silver-Polyvinyl Alcohol Nanocomposites. *Chemistry of Materials*, 15(26), 5019-5024.
- Mi, F. (2005). Synthesis and Characterization of a Novel Chitosan–Gelatin Bioconjugate with Fluorescence Emission. *Biomacromolecules*, *6*(2), 975-987.

- Morones, J.R., Elechiguerra, J.L., Camacho, A., Holt, K., Kouri, J.B., Ramırez, J.T., & Yacaman, M.J. (2005). The bactericidal effect of silver nanoparticles. *Nanotechnology*, *16*, 2346–2353.
- Mukherjee, P., Ahmad, A., Mandal, D., Senapati, S., Sainkar, S.R., Khan, M.I., Parishcha, R., Ajaykumar, P.V., Alam, M. Kumar, R., & Sastry, M. (2001). Fungus-Mediated Synthesis of Silver Nanoparticles and Their Immobilization in the Mycelial Matrix: A Novel Biological Approach to Nanoparticle Synthesis. *Nano Letters*, 1(10), 515-519.
- Muyonga, J.H., Cole, C.G.B., & Duodu, K.G. (2004). Fourier transform infrared (FTIR) spectroscopic study of acid soluble collagen and gelatin from skins and bones of young and adult Nile perch (Lates niloticus). *Food Chemistry*, 86(3), 325-332.
- Nickel, U., Castell, A.Z., Pöppl, K., & Schneider, S. (2000). A Silver Colloid Produced by Reduction with Hydrazine as Support for Highly Sensitive Surface-Enhanced Raman Spectroscopy†. *Langmuir*, *16*(23), 9087-9091.
- Norio, T. (1983). Current Status in, and Future Trends of, Ultraprecision Machining and Ultrafine Materials Processing. CIRP Annals Manufacturing Technology, 32(2), 573-582.
- Oberdörster, G., Oberdörster, E., & Oberdörster, J. (2005). Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect*, *113*(7), 823-839.
- Oliveira, M.M., Ugarte, D., Zanchet, D., & Zarbin, A.J.G. (2005). Influence of synthetic parameters on the size, structure, and stability of dodecanethiol-stabilized silver nanoparticles. *Journal of Colloid and Interface Science*, 292(2), 429-435.
- Osman, Z., & Arof, A.K. (2003). FTIR studies of chitosan acetate based polymer electrolytes. *Electrochimica Acta, 48*(8), 993-999.
- Pal, A., Shah, Su., & Devi, S. (2009). Microwave-assisted synthesis of silver nanoparticles using ethanol as a reducing agent. *Materials Chemistry and Physics*, 114(2–3), 530-532.
- Panácek, A., Kvítek, L., Prucek, R., Kolář, M., Veceřová, R., Pizúrová, N., Sharma, V.K., Nevěcná, T., & Zbořil, R. (2006). Silver Colloid Nanoparticles: Synthesis, Characterization, and Their Antibacterial Activity. *The Journal of Physical Chemistry B*, 110(33), 16248-16253.

- Panigrahi, S., Kundu, S., Ghosh, S.K., Nath, S., & Pal, T. (2004). General method of synthesis for metal nanoparticles. *Journal of Nanoparticle Research*, *6*(4), 411-414.
- Peng, S., McMahon, J.M., Schatz, G.C., Gray, S.K., & Sun, Y. (2010). Reversing the size-dependence of surface plasmon resonances. *Proceedings of the National Academy of Sciences*, 107(33), 14530-14534.
- Pillai, Z.S., & Kamat, P.V. (2003). What Factors Control the Size and Shape of Silver Nanoparticles in the Citrate Ion Reduction Method? *The Journal of Physical Chemistry B*, 108(3), 945-951.
- Puišo, J., Prosyčevas, I., Guobienė, A., & Tamulevičius, S. (2008). Plasmonic properties of silver in polymer. *Materials Science and Engineering: B,* 149(3), 230-236.
- Radziuk, D., Skirtach, A., Sukhorukov, G., Shchukin, D., & Möhwald, H. (2007). Stabilization of Silver Nanoparticles by Polyelectrolytes and Poly(ethylene glycol). *Macromolecular Rapid Communications*, 28(7), 848-855.
- Raimondi, F., Scherer, G.G., Kötz, R., & Wokaun, A. (2005). Nanoparticles in Energy Technology: Examples from Electrochemistry and Catalysis. *Angewandte Chemie International Edition*, *44*(15), 2190-2209.
- Ray, S.S., & Bousmina, M. (2005). Biodegradable polymers and their layered silicate nanocomposites: In greening the 21st century materials world. *Progress in Materials Science*, *50*(8), 962-1079.
- Reicha, F.M., Sarhan, A., Abdel-Hamid, M.I., & El-Sherbiny, I.M. (2012). Preparation of silver nanoparticles in the presence of chitosan by electrochemical method. *Carbohydrate Polymers*, 89(1), 236-244.
- Salkar, R.A., Jeevanandam, P., Aruna, S.T., Koltypin, Y., & Gedanken, A. (1999). The sonochemical preparation of amorphous silver nanoparticles. *Journal of Materials Chemistry, 9*(6), 1333-1335.
- Sanpui, P., Murugadoss, A., Prasad, P.V.D., Ghosh, S.S., & Chattopadhyay, A. (2008). The antibacterial properties of a novel chitosan–Ag-nanoparticle composite. *International Journal of Food Microbiology*, *124*(2), 142-146.
- Scheibel, H.G., & Porstendorfer, J. (1983). Generation of monodisperse Agand NaCl-aerosols with particle diameters between 2 and 300 nm. *Journal of Aerosol Science*, *14*(2), 113-126.

- Shahverdi, A.R., Minaeian, S., Shahverdi, H.R., Jamalifar, H., & Nohi, A. (2007). Rapid synthesis of silver nanoparticles using culture supernatants of Enterobacteria: A novel biological approach. *Process Biochemistry*, 42(5), 919-923.
- Shameli, K., Ahmad, M.B., Yunus, W.M., Ibrahim, N.A., Gharayebi,Y., & Sedaghat, S. (2010a). Synthesis of silver/montmorillonite nanocomposites using gamma-irradiation. *Int J Nanomedicine*, *5*, 1067-1077.
- Shameli, K., Ahmad, M.B., Yunus, W.M., Rustaiyan, A., Ibrahim, N.A., Zargar, M., & Abdollahi, Y. (2010b). Green synthesis of silver/montmorillonite/chitosan bionanocomposites using the UV irradiation method and evaluation of antibacterial activity. *Int J Nanomedicine*, *5*, 875-887.
- Shameli, K., Ahmad, M.B., Zargar, M., Yunus, W.M., Ibrahim, N.A., Shabanzadeh, P., & Moghaddam, G.F. (2011). Synthesis and characterization of silver/montmorillonite/chitosan bionanocomposites by chemical reduction method and their antibacterial activity. *Int J Nanomedicine*, 6, 271–284.
- Sharma, V.K., Yngard, R.A., & Lin, Y. (2009). Silver nanoparticles: Green synthesis and their antimicrobial activities. *Advances in Colloid and Interface Science*, 145(1–2), 83-96.
- Sionkowska, A., Wisniewski, M., Skopinska, J., Kennedy, C.J., & Wess, T.J. (2004). Molecular interactions in collagen and chitosan blends. *Biomaterials*, *25*(5), 795-801.
- Smitha, B., Devi, D.A., & Sridhar, S. (2008). Proton-conducting composite membranes of chitosan and sulfonated polysulfone for fuel cell application. *International Journal of Hydrogen Energy*, 33(15), 4138-4146.
- Song, K.C., Lee, S.M., Park, T.S., & Lee, B.S. (2009). Preparation of colloidal silver nanoparticles by chemical reduction method. *Korean Journal of Chemical Engineering*, *26*(1), 153-155.
- Stuart, M.A.C., Huck, W.T.S., Genzer, J., Muller, M., Ober, C., Stamm, M., Sukhorukov, G.B., Szleifer, I., Tsukruk, V.V., Urban, M., Winnik, F., Zauscher, S., Luzinov, I., & Minko, S. (2010). Emerging applications of stimuli-responsive polymer materials. *Nat Mater, 9*(2), 101-113.

- Sun, Y., & Xia, Y.. (2002). Shape-Controlled Synthesis of Gold and Silver Nanoparticles. *Science*, 298(5601), 2176-2179.
- Tabata, Y. (2009). Biomaterial technology for tissue engineering applications. *J R Soc Interface, 6 Suppl 3*, S311-324.
- Travan, A., Marsich, E., Donati, I., & Paoletti, S. (2007). Silver Nanocomposites and Their Biomedical Applications *Nanotechnologies for the Life Sciences*: Wiley-VCH Verlag GmbH & Co. KGaA.
- Tsuji, M., Hashimoto, M., Nishizawa, Y., Kubokawa, M., & Tsuji, T. (2005). Microwave-Assisted Synthesis of Metallic Nanostructures in Solution. *Chemistry A European Journal, 11*(2), 440-452.
- Wei, D., Sun, W., Qian, W., Ye, Y., & Ma, X. (2009). The synthesis of chitosan-based silver nanoparticles and their antibacterial activity. *Carbohydrate Research*, *344*(17), 2375-2382.
- Wiley, B., Sun, Y., Mayers, B., & Xia, Y. (2005). Shape-Controlled Synthesis of Metal Nanostructures: The Case of Silver. *Chemistry A European Journal*, 11(2), 454-463.
- Yao, C., Sun, J., Lin, F., Liao, C., & Huang, C. (1996). Biological effects and cytotoxicity of tricalcium phosphate and formaldehyde cross-linked gelatin composite. *Materials Chemistry and Physics*, *45*(1), 6-14.
- Yu, L., Dean, K., & Li, L. (2006). Polymer blends and composites from renewable resources. *Progress in Polymer Science*, *31*(6), 576-602.
- Zhang, C., Zhang, Z., Yu, B., Shi, J., & Zhang, X. (2001). Application of the Biological Conjugate between Antibody and Colloid Au Nanoparticles as Analyte to Inductively Coupled Plasma Mass Spectrometry. *Analytical Chemistry*, 74(1), 96-99.
- Zhang, Z., Patel, R.C., Kothari, R., Johnson, C.P., Friberg, S.E., & Aikens, P.A. (2000). Stable Silver Clusters and Nanoparticles Prepared in Polyacrylate and Inverse Micellar Solutions. *The Journal of Physical Chemistry B*, 104(6), 1176-1182.
- Zook, J.M, Halter, M.D, Cleveland, D., & Long, S.E. (2012). Disentangling the effects of polymer coatings on silver nanoparticle agglomeration, dissolution, and toxicity to determine mechanisms of nanotoxicity. *Journal of Nanoparticle Research*, *14*(10), 1-9.