

UNIVERSITI PUTRA MALAYSIA

URINARY PURINE DERIVATIVES AS INDEX FOR ESTIMATION OF RUMINAL MICROBIAL NITROGEN PRODUCTION IN SHEEP AND GOATS

THONGSUK JETANA.

IB 2005 3



URINARY PURINE DERIVATIVES AS INDEX FOR ESTIMATION OF RUMINAL MICROBIAL NITROGEN PRODUCTION IN SHEEP AND GOATS

THONGSUK JETANA

DOCTOR OF PHILOSOPHY UNIVERSITI PUTRA MALAYSIA

2005



URINARY PURINE DERIVATIVES AS INDEX FOR ESTIMATION OF RUMINAL MICROBIAL NITROGEN PRODUCTION IN SHEEP AND GOATS

By

THONGSUK JETANA

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

November 2005



DEDICATION

То

Mae and Mear



Abstract of thesis presented to the Senate of the University Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

URINARY PURINE DERIVATIVES AS INDEX FOR ESTIMATION OF RUMINAL MICROBIAL NITROGEN PRODUCTION IN SHEEP AND GOATS

By

THONGSUK JETANA

November 2005

Chairperson : Professor Norhani Bt. Abdullah, PhD

Institute : Bioscience

Microbial-N production in the rumen can be estimated by using urinary purine derivatives (PD) as an index. A series of experiments were conducted to establish the relationship between urinary PD (allantoin, uric acid, xanthine and hypoxanthine) and feed intake, endogenous PD excretion and recovery rate of plasma PD in sheep and goats. Studies on factors affecting PD excretion; xanthine oxidase and uricase activities of the plasma, liver and intestinal mucosa cells; uric acid kinetic; and purine-N:total-N ratio of rumen microbes were also conducted. Male Poll Dorset Cross sheep and male Ferral goats were used. The animals were fed a diet consisting of 40% oil palm frond and 60% concentrate (OPFC). Four sheep ($40.2\pm2.8kg$) and four goats ($39.6\pm1.8 kg$) were used to measure urinary PD excretion at 40%, 60%, 80% and 95% of voluntary intake (VI). The proportion of plasma PD excreted in the urine was determined by using [¹⁴C]-uric acid as a marker at 40% and 80 % of VI. Endogenous PD excretion was determined by fasting in six sheep ($55.4\pm5.1 kg$) and six goats ($40.2\pm4.6kg$). The results showed that sheep excreted significantly (p<0.05) higher PD and creatinine than goats



when compared at the same level of feed intake. However, the coefficient of the relationship between PD and DOMI in goats (12.57 mmol/kg DOMI) was similar to that in sheep (12.49 mmol/kg DOMI). The proportion of allantoin to total PD in goats (86%) was higher than that in sheep (60%). The distribution pattern of enzymes (xanthine oxidase and uricase) activities in the plasma, liver and intestinal mucosal cells were similar in both animal species, but uricase activity of the intestinal mucosa cells in sheep was significantly higher (p < 0.05) than in goats. The average daily urinary endogenous PD excretion obtained by the fasting trial for sheep (201±35 µmol/kgW^{0.75}d⁻¹) was similar to that for goats (202±17 µmol/kg $W^{0.75}d^{-1}$). The average percentage of total recovery of plasma PD excreted in the $[^{14}C]$ -uric acid in sheep (77±2.8 %) was not urine determined by using significantly different from that in goats (83±2.0 %). In the uric acid kinetics study, total tracer recovered reached a peak value of about 74.2% at 12 h for goats, and 74.4% at 15 h for sheep. The conversion efficiency of [¹⁴C]-uric acid to allantoin in the plasma pool was higher (p < 0.05) in goats than in sheep, with a peak value of 40% recovery at 12 h post injection for goats and 33.5% at 15 h post injection for sheep. By 15 h, no [¹⁴C]-uric acid was detected in the urine of both animal species. The rates of $[^{14}C]$ -allantoin and $[^{14}C]$ -uric acid excretions in the urine of sheep (31.0 and 88.0% h^{-1} , respectively) were significantly (p<0.05) faster than those of goats (19.0 and 64.7% h^{-1} , respectively), but the rates of total [¹⁴C]tracer were not significantly different between the two animal species (42.5% h⁻¹ and 30.3% h⁻¹ for sheep and goats, respectively). The primary compartment size in the plasma (V₁) was significantly (p < 0.05) larger in sheep (24.4 ±3.01 mg C) than in goats (17.5±1.28 mg C) and the secondary compartment size in the tissue (V₂) of sheep was also larger (129±21.6 mg C) than that of goats (65.7±23.7 mg C). The



volume of distribution (L) was 45% higher in sheep (0.898 L) than in goats (0.490 L). However, the net flux tended to be higher in goats (20.3 \pm 3.82 mg C) than in sheep (16.1 \pm 2.0 mg C). Hence, the results indicated that differences exist between sheep and goats in uric acid/allantoin kinetics. The equations established for sheep and goats based on the recovery of labeled PD [¹⁴C]-uric acid and endogenous PD excretion to determine the absorption of purines (X mmol/d) estimated from PD excretion in the urine (Y mmol/d) for sheep was Y=0.77X+0.201×BW^{0.75}e^{-0.20X} and for goats Y= 0.83X+0.202×BW^{0.75}e^{-0.20X}. The purine-N:total-N ratios of mixed rumen liquid-associated bacteria and solid-associated bacteria for sheep were 11.2 and 10.4, and those for goats were 8.5 and 10.0, respectively. The proposed equations to estimate rumen microbial-N production based on PD excretion was 0.753X for sheep and 0.992X for goats.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

DERIVATIF PURINA URIN SEBAGAI INDEKS UNTUK MENGANGGAR PENGHASILAN NITROGEN MIKROB RUMEN BEBIRI DAN KAMBING

Oleh

THONGSUK JETANA

November 2005

Pengerusi : Profesor Norhani Abdullah, PhD

Institut : Biosains

Pengasilan N-mikrob dalam rumen boleh di anggarkan menggunakan derivatif purina (DP) urin sebagai indeks. Satu siri eksperimen telah dijalankan untuk menghasilkan persamaan antara DP (alantoin, asid urik, xantin dan hipoxantin) dan pengambilan makanan, pengkumuhan DP endogen, dan kadar perolehan semula DP plasma untuk bebiri dan kambing. Kajian keatas faktor yang mempengaruhi pengkumuhan DP; aktiviti enzim xantin oksidase dan urikase plasma, hati dan sel mukosa usus; kinetik asid urik; nisbah purina-N:total-N mikrob rumen juga dilakukan. Bebiri jantan kacukan silang Poll Dorset dan kambing Ferral digunakan. Haiwan diberi diet terdiri dari 40% pelepah daun kelapa sawit dan 60% konsentrat (OPFC). Empat bebiri (40.2±2.8kg) dan empat kambing (39.6±1.8 kg) digunakan unutk mengukur pengkumuhan DP pada 40%, 60%, 80% and 95% pengambilan secara sukarela (IV). Bahagian DP plasma yang dikumuhkan dalam urin ditentukan menggunakan [¹⁴C]-asid urik sebagai traser pada dua tahap VI (40% dan 80%). Derivatif purina endogen yang dikumuhkan dalam urin ditentukan dalam enam bebiri jantan (55.4±5.1kg) dan enam kambing jantan (40.2±4.6kg) semasa berpuasa. Hasil kajian menunjukan bebiri



mengumuhkan DP dan kreatinin dalam urin lebih banyak (p < 0.05) daripada kambing apabila dibandingkan pada tahap pengambilan makanan yang sama. Koefisien hubungan diantara DOMI dan DP untuk kambing (12.49 mmol/kg DOMI) adalah serupa dengan bebiri (12.57 mmol/kg DOMI). Bahagian alantoin dalam DP total untuk kambing (86%) adalah lebih tinggi daripada bebiri (60%). Purata pengumuhan DP endogen urin semasa berpuasa untuk bebiri (201±35 μ mol/kg W^{0.75} h⁻¹) adalah sama dengan kambing (202±17 μ mol/kg W^{0.75} h⁻¹). Taburan aktiviti enzim (xantin oksidase dan urikase) untuk plasma, hati dan usus kecil adalah sama pada kedua-dua spesis haiwan, tetapi aktiviti urikase di sel mukosa usus adalah lebih (p < 0.05) tinggi untuk bebiri daripada kambing. Purata peratus perolehan semula DP plasma dalam urin dengan menggunakan ¹⁴C-asid urik untuk bebiri (77±2.8%) adalah sama seperti kambing (83±2.0%). Dalam kajian kinetik asid urik, jumlah traser yang diperolehi semula mencapai nilai puncak 74.2% pada 12 j untuk kambing dan 74.4% pada 15 j untuk bebiri. Efisiensi penukaran [¹⁴C]-asid urik ke alantoin dalam gembeling plasma adalah lebih tinggi (p<0.05) untuk kambing dari bebiri dengan nilai 40% perolehan semula pada 12 j selepas suntikan untuk kambing dan 33.5% pada 15 j selepas suntikan untuk bebiri. Pada masa 15 j, tiada [¹⁴C]-asid urik dikesani dalam urin kedua spesis haiwan. Kadar pengkumuhan $[^{14}C]$ -alantoin dan $[^{14}C]$ -asid urik dalam urin bebiri (31.0 dan 88.0% j⁻¹, masing masing) adalah lebih cepat (p < 0.05) daripada kambing (19.0 and 64.7% h^{-1} , masing masing), tetapi kadar untuk total $[^{14}C]$ -traser tidak berbeza antara kedua spesis haiwan (42.5% j⁻¹ dan 30.3% j⁻¹ untuk bebiri dan kambing, masingmasing). Saiz ruang primer dalam plasma (V1) adalah lebih besar (p < 0.05) untuk bebiri (24.4±3.01mg C) daripada kambing (17.5±1.28mg C) dan saiz ruang sekunder dalam tisu (V2) pada bebiri juga adalah lebih besar (129±21.6mg C) daripada kambing (65.7±23.7mg C). Isipadu taburan (L) adalah 45% lebih tinggi untuk bebiri (0.898 L) daripada kambing (0.490 L). Walaupun demikian, fluks bersih adalah lebih tinggi untuk kambing (20.3±3.82mg C) daripada bebiri (16.1±2.0µg C). Oleh itu, keputusan menunjukkan perbezaan wujud di antara bebiri dan kambing dalam kinetik asid urik/allantoin. Persamaan berdasarkan perolehan semula DP [¹⁴C]-asid urik dan pengkumuhan DP endogen untuk menentukan penyerapan purina (X mmol/h) dengan menggunakan nilai pengkumuhan DP dalam urin (Y mmol/h) untuk bebiri ialah Y=0.77X+0.201×BW^{0.75}e^{-0.20X} dan kambing Y= 0.83X+0.202×BW^{0.75}e^{-0.20X}. Nisbah purina-N:total-N bakteria campuran dalam cecair dan pepejal rumen untuk bebiri ialah 11.2 dan 10.4, dan untuk kambing ialah 8.5 dan 10.0, masing masing. Persamaan yang disyorkan untuk menganggar Nmikrob rumen berdasarkan pengkumuhan DP adalah 0.753X untuk bebiri dan 0.992X untuk kambing.

ACKNOWLEDGEMENTS

I would like to express my deep appreciation and sincere gratitude to the chairman of the Supervisory Committee, Professor Dr. Norhani Abdullah and the other members of supervisory committee, Professor Dr. Ho Yin Wan and Associate Professor Dr. Liang Juan Boo, for their invaluable guidance, advices and encouragement throughout the course of this study.

I wish to thank Universiti Putra Malaysia for providing me with a scholarship, and Chulalongkorn University for granting me the study leave to pursue my PhD degree.

I am much indebted to Associate Professor Dr. John Nolan of the Biochemistry, Microbiology and Nutrition Department, New England University, Armidale, Australia, Associate Professor Dr. Qium Balcells of Departmento de Produccion Animal y Ciencia de los Alimentos, Facultad de Veterinaria, Universidad de Zaragoza, Miguel Servet 177, Spain, for their kind assistance, advices and guidance in this study.

Appreciation is extended to Dr. Nashariyah Mat and her staff at the of Unit Tenaga Nuklear, Kompleks PUSPATI, Bangi, Kajang, Malaysia, for their assistance on the preparation of samples for [¹⁴C]-counting and to Dr. Chinoma for her assistance on the preparation of animals for catherisation.



I also wish to thank the staff of the Institute Bioscience, Mdm Haw Ah Kam, Mr. Khairul Kamar Bakri, Mr Jivanathan Arumugam, Mr. Nagayah Muniandy and Mr. Paimon Logimun and the staff of the Department of Animal Science, Mr. Ibrahim Mohsin, Mr. Brakari Abd Rahman and Mr. Saparin Denim, for their technical support and kind assistance.

To members of my office, Dr. Sunpeth, Dr. Rangsun, Dr. Kitiya, Dr. Sirima, Usa, Wanvipha, Ratree, Tan, Kriangsak, Wisut, Sangworn and friends, Mongkorn, Suthipong, Opart, Bodee, Anut, Benjamaporn, Atikha and Pensri, I wish to express my deepest gratitude for their encouragement and support.

Special thanks are due to my postgraduate friends at the Digestive Microbiology Lab, Institute of Bioscience, Sideig, Darlis, Latifah, Kala, Chin Chin, Wan and Chin Mei, for their friendship, assistance, encouragement and support.

Finally, I wish to express my deepest gratitude to my family, mom, wife, son, sisters and brothers for their love encouragement and support throughout the course of my study.

To them, I dedicate this thesis.



TABLE OF CONTENTS

DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xvii
LIST OF FIGURES	XX
LIST OF ABBREVIATIONS	xxii

CHAPTER

1	INTE	RODUCTION	1
2	LITE	CRATURE REVIEW	3
	2.1	Rumen	3
	2.2	Feed and Digestion	3
	2.3	Rumen Microbes	5
	2.4	The Determination of the Rumen Microbial Yields	5
		2.4.1 Direct Method	5
		2.4.2 Indirect Method	17
	2.5	Determination of Glomerular Filtration Rate	38
		2.5.1 Creatinine as a marker to measure GFR	40
		2.5.2 Creatinine as a marker for urine spot sampling	41
	2.6	Inulin	42

3 THE EFFECTS OF DIFFERENT LEVELS OF FEED INTAKE ON URINARY PURINE DERIVATIVES EXCRETION IN SHEEP AND GOAT

3.1	Introduction	46
3.2	Materials and Methods	47
	3.2.1 Animals and diet	47
	3.2.2 Experimental procedures	48
3.3	Chemical analyses	49
	3.3.1 Proximate analyses	49
3.4	Results	55
3.5	Discussion	69

4 URINARY EXCRETION OF ENDOGENOUS PURINE DERIVATIVES, XANTHINE OXIDASE AND URICASE ACTIVITY IN PLASMA, LIVER AND INTESTINE OF SHEEP AND GOAT

4.1 Introduction	on
------------------	----

75 75

46



	4.2	Mater	ials and Methods	77
			Fasting experiment	77
			Animal and feed	77
		4.2.3	Samples collection	78
		4.2.4	Enzyme studies	79
		4.2.5	Enzyme assays	80
	4.3	Chem	ical analyses	83
		4.3.1	Proximate analyses	83
			Measurements	83
			Calculation	84
			Statistical analysis	84
	4.4			84
	4.5	Discu	ssion	96
5			VERY OF PLASMA PURINE DERIVATIVES IN	
			C OF SHEEP AND GOAT FED DIFFERENT	102
	5.1	_		103
			ials and methods	103 105
	5.2		Animals and diet	105
			Feeding	105
			Experimental procedures	105
			Proximate analysis	103
			Purine derivatives and creatinine	108
			Tracer [¹⁴ C]-uric acid	108
			Calculations	100
			Glomerular filtration rate (GFR)	110
			Statistical analysis	110
	5.3	Resul		111
	5.4	Discu		119
6	URIC	C ACID	KINETICS IN SHEEP AND GOAT	127
	6.1	Introd	luction	127
	6.2	Mater	ials and methods	128
		6.2.1	Experimental procedures	128
		6.2.2	[¹⁴ C]-uric acid and inulin administration	128
		6.2.3	Sample collection	129
		6.2.4		129
			Separation of uric acid and allantoin	129
		6.2.6		
			faecal samples	130
		6.2.7	Inulin	130
	6.3	Calcu		131
		6.3.1	Calculation of uric acid kinetics in plasma of sheep and goats	131
		6.3.2	-	133
		6.3.3	Statistical analysis	134
	6.4	Resul	•	134
	6.5	Discu	ssion	141



-

7		SUREMENT OF PURINE AND TOTAL NITROGEN	
		FENTS OF MIXED RUMEN MICROBIAL	
	POPU	JLATIONS IN SHEEP AND GOAT	147
	7.1	Introduction	147
	7.2	Materials and methods	148
		7.2.1 Sampling and sample preparation	148
		7.2.2 Liquid associated bacteria	148
		7.2.3 Solid associated bacteria	149
	7.3	Chemical analysis	149
		7.3.1 Proximate analysis	149
		7.3.2 Determination of total purine	149
		7.3.3 Determination of purine bases	151
		7.3.4 Protein analysis	151
		7.3.5 Statistical analysis	152
	7.4	Results	152
	7.5	Discussion	154
8	GEN	ERAL DISCUSSION AND CONCLUSION	160
	8.1	General discussion	160
	8.2	Conclusions	170
BIBL	IOGRA	РНҮ	172
APPE	NDICE	S	196
BIOD	ATA O	F THE AUTHOUR	234



LIST OF TABLES

Table

2.1	Markers for measuring digesta flow passing through the duodenum	6
2.2	Percentages of ash and N contents and purine:N ratio of liquid associated bacteria (LAB) and solid associated bacteria (SAB) of dairy cows at different times after feeding (Craig <i>et al.</i> , 1987)	16
2.3	Internal and external markers for quantifying microbial protein synthesis in the rumen	18
2.4	Purine and pyrimidine degradation in the different parts of digestive tract.	21
2.5	Proportions of nucleic acid-N to nitrogen contents entering to the rumen from various sources.	24
2.6	Basal purine derivatives (PD) excretion in urine in various species with the different methods of determination	30
2.7	Percentages recovery of purine derivatives in the urine of sheep from various studies by using radiochemical technique	33
2.8	Percentages recovery of purine derivatives (PD) in the urine from various studies by using different methods	34
2.9	Disappearance of microbial nucleic acid (NA) N from the small intestines of ruminants	38
2.10	The various models of the relationship between purine derivatives (PD) absorption and PD excretion in the urine	39
3.1	The four levels of feed intake	48
3.2	The feeding design with four levels of feed intake and four animals of each species	49
3.3	Ingredients (fresh weight) and chemical composition of oil palm frond plus concentrate (OPFC)	56



- 3.4 Intakes and apparent digestibilities of dry matter, organic 58 matter (g/d) and nitrogen (g/d) of sheep and goats fed at different levels of intake based on percentages of voluntary intake
- 3.5 The effects of intake level on urinary purine derivatives 59 (mmol/d), creatinine (mmol/d), and nitrogen (g/d), and the ratios of PD:Cr and UN:Cr in sheep and goats
- 3.6 The effects of levels of feed intake on urinary purine 61 derivatives (PD), creatinine (μ mol/BW ^{0.75} d⁻¹) and urinary nitrogen (mg/BW ^{0.75} d⁻¹) in sheep and goats
 - 3.7 The relationships between DMI OMI (kg/d) and purine 63 derivatives (PD) excretion in the urine and the linear equations between DMI and OMI (kg/d) with purine derivatives excretion in the urine (mmol/d) in sheep
 - 3.8 The relationships between DMI OMI (kg/d) and purine 65 derivatives (PD) excretion in the urine and the linear equations between DMI and OMI (kg/d) with purine derivatives excretion in the urine (mmol/d) in goats
 - 3.9 Comparison between sheep and goats on urinary purine 68 derivatives (μ mol/BW ^{0.75}d⁻¹) at different levels of feed intake
 - 4.1 Daily purine derivatives, creatinine, N excretion in the 87 urine between sheep and goats fed oil frond concentrate (OPFC) at 600 KJ/BW ^{0.75} during feeding at maintenance (days 1-7)
 - 4.2 Daily purine derivatives, creatinine and N excretion in the 88 urine of sheep and goats during fasting (days 10-15)
 - 4.3 Daily glomerular filtralation, tubular load of purine 90 derivatives, excretion of purine derivatives in the urine and net re-absorption of purine derivatives in sheep and goats fed opfc at 600 KJ/BW ^{0.75} during maintenance feeding (day 7)
 - 4.4 Daily glomerular filtralation, tubular load of purine 91 derivatives, excretion of purine derivatives in the urine and net re-absorption of purine derivatives in sheep and goats during fasting (days 10-15)
 - 4.5 Xanthine oxidase activity (mean ± standard error) in the 95 plasma, liver and intestinal mucosa cells of sheep and goats



4.6	Uricase activities (mean \pm standard error) in plasma, liver and mucosa cells of sheep and goats	95
5.1	Feeding regiment of four sheep and four goats at the 40 and 80 % of voluntary intake (VI)	106
5.2	Feed intake and apparent digestibilities of DM, OM in sheep and goats fed OPFC at the 40 and 80 % of voluntary intake (VI)	112
5.3	Percentages of cumulative recovery of $[^{14}C]$ -tracer in the urine and faeces after $[^{14}C]$ -uric acid injecting into jugular vein at different collection time in sheep and goats fed OPFC at the 40 and 80 % of voluntary intake (VI)	114
5.4	Percentages of cumulative recovery as urinary [¹⁴ C] and rate of excretion in the urine (% h ⁻¹) derived from equation $SR_t = b_1 (1 - e^{-b_1 t})$	115
5.5	Daily creatinine excretions (μ mol/L) and glomerular filtration rates (GFR, L/d) in sheep and goats fed OPFC at the 40 and 80% voluntary intake (VI)	116
5.6	Models developed for sheep and goats based on the recovery of $[^{14}C]$ labelled PD and endogenous PD excretion	118
5.7	Models established based on urinary endogenous PD from fasting trial and the proportion of plasma PD and PD excretion in the urine using $[^{14}C]$ -tracer	124
6.1	Percentages of cumulative recovery of $[^{14}C]$ -tracer and $[^{14}C]$ -uric acid, $[^{14}C]$ -allantoin and $[^{14}C]$ -other metabolites at different sampling times after a single injection of $[^{14}C]$ -uric acid into the jugular vein of sheep and goats fed OPFC at 750 g DM d ⁻¹	135
6.2	The cumulative recovery of $[^{14}C]$ -tracer, excretion rate of PD in the urine and glomerular filtration rate of sheep and goats at 144 h after a single injection of $[^{14}C]$ -uric acid	138
6.3	Plasma uric acid concentration, primary compartment size, volume of distribution and flux rates	140
7.1	Chemical composition of mixed rumen bacteria and total purine-N:total N of sheep and goats fed OPFC <i>ad libitum</i>	153

xix

LIST OF FIGURE

Figure		Page
2.1	Changes in total nucleic acid:N ratios of liquid associated bacteria and solid associated bacteria isolated the rumens of dairy cows at different times after feeding (Craig <i>et al.</i> , 1987)	15
2.2	Purine nucleotide catabolism (Lehninger et al., 1992)	23
2.3	Urinary excretion of purine derivatives in sheep and cattle in relation to the amount of microbial purines absorbed (Chen <i>et al.</i> , 1990b; Verbic <i>et al.</i> , 1990)	27
2.4	The model of inulin concentration clearance in plasma during a single-injection technique (Gretz <i>et. al.</i> , 1993)	44
3.1	The relationships between DOMI (x) and allantoin or purine derivatives (y) in the urine of sheep and goats	67
4.1	Experimental feeding regiment and sampling activities	78
4.2	Total purine derivatives excretion in the urine during maintenance feeding (days 1-7), reduced feed (day 8, 60% and day 9 30%) and fasting period (days 10-15)	85
4.3	Nitrogen excretion in the urine during feeding at maintenance (days 1-7) reduced feed (60% at day 8 and 30% at day 9) and fasting period (days 10-15)	85
4.4	Xanthine oxidase activity in plasma, liver and intestinal mucosa extracts of sheep and goats	93
4.5	Uricase activity in plasma, liver and intestinal mucosal extracts of sheep and goats	94
5.1	Percentages of cumulative recovery of [¹⁴ C]-tracer excretion in the urine of sheep and goats fed two levels of OPFC	113



5.2	Urinary purine derivatives in relation to plasma purine	125
	derivatives in sheep and goats.	

6.1 Percentages of cumulative recovery of total- $[^{14}C]$ -tracer, 136 $[^{14}C]$ -uric acid, $[^{14}C]$ -allantoin, and $[C^{14}]$ -other metabolites in the urine samples of sheep and goats



LIST OF ABBREVIATIONS

ADF	=	acid detergent fibre
AEP	=	aminoethyl-phosphnic acid
ARC	=	Agricultural Research Council
ATP	=	adenosine-5'-triphosphate
BW ^{0.75}	=	metabolic body weight
Cal	=	calorie
CF	=	crude fibre
cm	=	centimetre
CMC	=	carboxymethy cellulose
СР	-	crude protein
d	=	day
DE	=	digestible energy
d.f.	=	degree of freedom
dl	=	decilitre
DM	=	dry matter
DMI	=	dry matter intake
DDMI	=	digestible dry matter intake
ОМ	=	organic matter
OMI	=	organic matter intake
DOMI	=	digestible organic matter intake
DOMR	=	digestible organic matter digested in the rumen
EE	=	ether extract (crude fat)
g	=	gram
GE	=	gross energy

xxii

GFR	=	glomerular filtration rate
GLM	=	General linear measurement
h	-	hour
ha	=	hectare
hd	=	head
HPLC	=	high performance liquid chromatography
IBC	-	isolated bacteria cells
i.e.	=	that is
i.d.	=	internal diameter
kg	=	kilogram
L	=	litre
LAB	-	liquid associated bacteria
m	=	metre
mM	=	millimoles of solute per litre of solution
ME		metabolisable energy
mg	=	milligram
min	=	minute
MJ	-	megajoule
ml	=	millilitre
mm		millimetre
N	=	nitrogen
NDF	=	neutral detergent fibre
ηm	=	nanometre
NRC	=	National Research Council (USA)
OPFC	=	oil palm frond plus concentrate

xxiii