

# **UNIVERSITI PUTRA MALAYSIA**

STRUCTURE AND FUNCTION OF NOVEL ANTIFREEZE PEPTIDES DERIVED FROM GLACIOZYMA ANTARCTICA ANTIFREEZE PROTEIN-1

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By

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#### STRUCTURE AND FUNCTION OF NOVEL ANTIFREEZE PEPTIDES DERIVED FROM *GLACIOZYMA ANTARCTICA* ANTIFREEZE PROTEIN-1

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**July 2013** 

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Organisms living in cold environment produce antifreeze proteins (AFPs) which exhibit special functions as a result of cold adaptation. AFP is currently being identified in many organisms such as bacteria, plants, fish, and fungi that are exposed to freezing stress. This study aimed to create novel antifreeze peptides based on the three-dimensional structure of *Glaciozyma antarctica* antifreeze protein-1 (AFP-1). Computational prediction on the structure of AFP-1 suggests that the helical segments of this protein are responsible for antifreeze activity. Six peptides derived from the sequence of *G. antarctica* have been synthesized. The peptides show measurable antifreeze activity as quantitatively measured by thermal hysteresis (TH) assay and qualitatively by ice recrystallization inhibition (IRI) assay. Structure determination of antifreeze peptides was carried out based on spectroscopic data



obtained by using one dimensional and two dimensional <sup>1</sup>H-NMR (800 MHz) in elucidating the structures of obtained peptides. All antifreeze peptides showed increase of thermal hysteresis value which is relative to the increase of antifreeze peptides concentration until the saturation point of solution. Peptide 1m recored the highest antifreeze activity with TH value  $0.097 \pm 0.004$  °C, almost similar to the parent protein AFP-1 ( $0.1^{\circ}$ C in concentration 0.1mM). Analysis of relationship between the peptide NMR structure and its activity showed that the peptides form alpha helical structure and the extent of peptide helicity greatly influences the activity of antifreeze peptides derived from *G. antarctica* AFP-1 segments.

**Keywords:** AFP, antifreeze peptides, freezing tolerance, ice morphology, ice crystal, recrystallization inhibition, thermal hysteresis, alpha helical structure, NMR



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

#### STRUKTUR DAN FUNGSI PEPTIDA ANTIBEKU BAHARU BERASASKAN PROTEIN ANTIBEKU-1 *GLACIOZYMA ANTARCTICA*

Oleh

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Organisma yang berhabitat dalam persekitaran iklim sejuk menghasilkan protein antibeku (AFP) yang berfungsi memberikan adaptasi organisma itu kepada persekitaran sejuk. Kini, AFP sudah dapat dikenal pasti pada pelbagai organisma iaitu bakteria, tumbuhan, ikan, dan kulat yang terdedah pada persekitaran beku lampau. Kajian ini bertujuan mencipta peptide baharu berasaskan struktur tiga dimensi protein antibeku-1 *Glaciozyma antarctica* (AFP-1). Struktur yang diperoleh melalui kaedah ramalan komputer mencadangkan bahawa segmen helikal pada AFP-1 berperanan untuk menghasilkan aktiviti antibeku. Enam peptida daripada jujukan AFP *G. antarctica* digunakan dalam kajian ini. Peptida yang dikehendaki menunjukkan aktiviti antibeku berdasarkan data kuantitatif histeresis suhu (TH) dan data kualitatif perencatan pembentukan kristal ais (IRI). Penentuan struktur peptida anti-beku diperoleh dengan menggunakan <sup>1</sup>H-NMR (800 MHz) satu dimensi dan dua dimensi. Semua peptida anti-beku menunjukkan peningkatan aktiviti histeresis suhu selaras dengan peningkatan konsentrasi larutan peptida anti-beku sehingga ke titik tepu. Peptida 1m merekodkan aktiviti anti-beku paling tinggi antara semua sampel dengan nilai histeresis suhu  $0.097 \pm 0.004$  °C yang mana hampir sama dengan protein AFP-1 (0.1 °C dalam kepekatan larutan 0.1 mM). Analisis perkaitan antara struktur NMR peptida yang dikehendaki dengan aktiviti anti-bekunya menunjukkan bahawa peptida antibeku memerlukan struktur alfa-helikal dan takat helikal tersebut amat mempengaruhi aktiviti peptida antibeku yang dicipta berasaskan daripada struktur *G. antarctica* AFP-1.



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### DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



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## LIST OF ABBREVIATIONS

| AFP                | Antifreeze protein                    |
|--------------------|---------------------------------------|
| G.antarctica       | Glaciozyma antarctica                 |
| NMR                | Nuclear magnetic resonance            |
| NOESY              | Nuclear Overhause Effect Spectroscopy |
| TOCSY              | Total Correlation Spectroscopy        |
| <sup>1</sup> H-NMR | Proton nuclear magnetic resonance     |
| Glu                | Glutamic acid                         |
| Gln                | Glutamine                             |
| FDA                | U.S Food and Drug Administration      |
| GRAS               | Generally recognized as safe          |
| PG                 | Propylene Glycol                      |
| IgE                | Immuglibilin E                        |
| ТН                 | Thermal hysteresis                    |
| Thr                | Thrionine                             |
| Asp                | Aspartic acid                         |
| Leu                | Leucine                               |
| Ser                | Serine                                |
| Val                | Valine                                |
| Ala                | Alanine                               |
| E. coli            | Escherichia coli                      |
| G. antarctica      | Glaciozyma antarctica                 |
| IRI                | Ice recrystallization inhibition      |

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#### **CHAPTER 1**

#### **INTRODUCTION**

Freezing at sub-zero temperature always causes damage to cellular organisms. The freezing phenomenon occurred by inhibiting biological and chemical reaction in their natural medium. The freezing also affect normal concentration of elements in plasma, denatures organisms' biomolecules and ruptures cell membranes (Harding, 1999). However, several species of fish, plants, arthropods, fungi, and bacteria in Antarctic and Artic poles are able to survive in temperatures below freezing point (Duman and Olsen, 1993). These organisms produce antifreeze proteins which play the role of inhibitor toward the ice crystal formation by depressing the freezing point.

The first antifreeze protein (AFP) was discovered in the blood of Antarctic fish over 40 years ago (Scholander *et al.*, 1957; DeVries, 1971). To date, there are five types of AFPs classified based on their metal dependencies, molecular sizes, secondary and tertiary structures, and ice-binding plane (Davies and Sykes, 1997). Type I AFPs are described as alanine-rich protein sequence with  $\alpha$ -helical structure and sized between 3.3 kD and 4.5 kD (Duman and Devries, 1974; Duman and deVries, 1976; Hew *et al.*, 1985). Type II AFPs are described as globular proteins containing multi-cysteine residue with five disulfide bonds (Ng *et al.*, 1986; Slaughter *et al.*, 1981; Ng and Hew, 1992). Meanwhile, type III antifreeze proteins are described as globular proteins with molecular weight around 6 kD (Jia *et al.*, 1995; DeLuca *et al.*, 1996; Sonnichsen *et al.*, 1996). Type IV AFPs have  $\alpha$ -helical proteins structure with multi glutamate (Glu) or glutamine (Gln) residues in sequence (Deng *et al.*, 1997). The last

one is type V AFPs, which were discovered from insects and known as hyperactive proteins from its source (Liou *et al.*, 2000).

Because of their unique function, AFPs have been proposed to be developed as for commercial products. For example, some of the current prospects regarding the use of AFPs include (Griffith and Ewart, 1995), artificial rain and surgical preservation (Arav *et al.*, 1994; Payne and Wilson, 1994; Chao, *et al.*, 1996; Kun and Mastai, 2007). In more advance application, AFP could be applied to increase cold tolerance of plant through genetic engineering (Fan *et al.*, 2002). In food industry, propylene glycol (PG) is the most common antifreeze agent used in food and other consumer products nowadays. PG is classified as "*generally recognized as safe*" (GRAS) by FDA. Propylene glycol (PG) was never really tested directly in human, but GRAS status was given based on common use of PG in foods. There are increasing concerns about propylene glycol toxicity since it has been used as common household chemicals. Choi *et al.* (2010) suggested that propylene glycol and glycol ethers (PGEs) may induce allergic symptoms, asthma, rhinitis and eczema in children, as well as IgE sensitization respectively. Therefore, a new edible antifreeze agent derived from natural sources is needed.

It has been reported that several types of antifreeze proteins exist in *Glaciozyma antarctica* (Hashim *et al.*, 2013), This study has been focused on the synthesis of the peptide segments derived from the sequence of *G. antarctica* AFP with measureable antifreeze activity. From practical aspects point of view, peptide has been known as a better alternative due to less adverse reaction in body than proteins. Other obvious advantage of using antifreeze peptides over the use of antifreeze protein is that the smaller antifreeze molecules can act as "molecular tools" to study

the most important sequences, which play role in the antifreeze proteins (Kun and Mastai, 2007).

Specifically, the central hypothesis for this study is that the antifreeze activity of G. *antarctica* AFP relies on the helical regions of the protein. In this study, several novel antifreeze peptides have been synthesized based on helical part of G. *antarctica*. This approach enables us to study the ice-binding mechanism of antifreeze proteins.

#### 1.1 Research Objectives

The main objective of this work is to experimentally identify the structure and functions of novel antifreeze peptide derived from *G. antarctica* AFP. Therefore, this study embarks on the following objectives:

- 1. to design and synthesize novel antifreeze peptides derived from the helical regions of AFP that may have ice inhibiting activity;
- 2. to study antifreeze activity of *G. antarctica* AFP-derived peptides sequences and enhance the antifreeze activity by amino acid replacements; and
- 3. to determine the structure of the antifreeze peptides by nuclear magnetic resonance techniques.

#### BIBLIOGRAPHY

Arav A., Rubinsky B., Seren E., Roche J.F., and Boland M.P. (1994). The role of thermal hysteresis proteins during cryopreservation of oocytes and embryos. *Theriogenology* 41:107-112.

Baker D. (2000). A suprising simplicity to protein folding. Nature 405:39-42.

Barrett J. (2001). Thermal hysteresis proteins. IJBCB 33:105-117.

Chakrabartty A., Yang D.S., Hew C.L. (1989). Structure-function relationship in a winter flounder antifreeze polypeptide. II. alteration of the component growth rates of ice by synthetic antifreeze polypeptides. *J Biol Chem.* 264(19):11313-6.

Chakrabartty A., Ananthanarayanan V.S., Hew C.L. (1989). Structurefunction relationships in a winter flounder antifreeze polypeptide. I. stabilization of an alpha-helical antifreeze polypeptide by charged-group and hydrophobic interactions. *J Biol Chem.* 264(19):11307-12.

Chao H., Davies P.L., Sykes B.D., Sonnichsen F.D. (1993). Use of proline mutants to help solve the NMR solution structure of type III antifreeze protein. *Protein Sci.* 2:1411-1428.

Chao H.M., Davies P.L., Carpenter J.F. (1996). Effects of antifreeze proteins on red blood cell survival during cryopreservation. *J Exp Biol.* 199:2071-2076.

Chao H.M., Hodges R.S., Kay C.M., Gauthier S.Y., Davies P.L. (1996). A natural variant of type I antifreeze protein with 4 ice-binding repeats is a particularly potent antifreeze. *Protein Sci.* 5 (6): 1150-1156.

Choi H, Schmidbauer N, Sundell J, Hasselgren M, Spengler J, Bornehag C-. (2010) Common household chemicals and the allergy risks in pre-school age children. *PLoS ONE*. 2010;5(10).

Chou P.Y., Fasman G.D. (1974). Conformational parameters for amino acids in helical,  $\beta$ -sheet, and random coil regions calculated from proteins. *Biochemistry (N Y)*. 13(2):211-22.

Cornilescu G., Delaglio F., Bax A. (1999). Protein backbone angle restraints from searching a database for chemical shift and sequence homology. *J Biomol NMR*. 13(3):289-302.

Damodaran, S. (2007). Inhibition of ice crystal growth in ice cream mix by gelatin hydrolysate. *J Agric Food Chem.* 55:10918-10923.

Davies P.L., Sykes B.D. (1997). Antifreeze proteins. *Curr Opin Struct Biol* (7): 828–834.

Davies P.L., Baardsnes J., Kuiper M.J., Walker V.K. (2002) Structure and function of antifreeze proteins. *R.Soc Lond B.* 357: 927-935.

DeLuca C.I., Chao H., Sonnichsen F.D., Sykes B.D., Davies P.L. (1996) Effect of type III antifreeze protein dilution and mutation on the growth inhibition of ice. *Biophys J.* 71:2346-2355.

Deng G., Andrews D.W., Laursen R.A. (1997). Amino acid sequence of a new type of antifreeze protein, from the longhorn sculpin *Myoxocephalus* octodecimspinosis. *FEBS Lett.* 402:17-20.

DeVries A.L. (1971). Glycoproteins as biological antifreeze agents in antarctic fishes. *Science* 172:1152-1155.

DeVries AL, Lin Y (1977) Structure of a peptide antifreeze and mechanism of adsorption to ice. Biochim Biophys Acta 495: 88–392

Duman J.G., DeVries A.L. (1974). Freezing resistance in winter flounder *Pseudopleuronectes americanus*. *Nature* 247:237-238.

Duman J.G., DeVries A.L. (1976). Isolation, characterization, and physical properties of protein antifreezes from the winter flounder, *Pseudopleuronectes americanus. Comp Biochem Physiol B*. 54:375-380.

Duman J.G., Olsen T.M. (1993). Thermal hysteresis protein activity in bacteria, fungi, and phylogenetically diverse plants. *Cryobiology* 30:322-328.

Fairley K., Westman B.J., Pham L.H., Haymet A.D., Harding M.M., Mackay J.P. (2002). Type I shorthorn sculpin antifreeze protein: Recombinant synthesis, solution conformation, and ice growth inhibition studies. *J Biol Chem.* 277:24073-24080.

Fan, Y., Liu, B., Wang, H., Wang, S., & Wang, J. (2002). Cloning of an antifreeze protein gene from carrot and its influence on cold tolerance in transgenic tobacco plants. *Plant Cell Reports*, *21*(4), 296-301.

Garner J., Harding M.M. (2007). Design and synthesis of alpha-helical peptides and mimetics. *Org Biomol Chem.* 5:3577-3585.

Gilbert J.A., Hill P.J., Dodd C.E., and Laybourn-Parry J. (2004) Demonstration of antifreeze protein activity in Antarctic lake bacteria. *Microbiology*. 150 (1):171-180.

Graether, S.P., Gagné, S.M., Spyracopoulos, L., Jia, Z., Davies, P.L., Sykes, B.D. (2003). Spruce Budworm Antifreeze Protein: Changes in Structure and Dynamics at Low Temperature. *Journal of Molecular Biology*, *327*(5), 1155-1168.

Griffith M., Ewart K.V. (1995). Antifreeze proteins and their potential uses in frozen foods. *Biotech Adv.* 13:375-402.

Güntert P, Mumenthaler C, Wüthrich K. (1997). Torsion angle dynamics for NMR structure calculation with the new program DYANA. *J Mol Biol.* 273(1):283-98.

Hall D.G., Lips A. (1999). Phenomenology and mechanism of antifreeze peptide activity. *J Am Chem Soc.* 15:1905-1912.

Harding M.M., Ward L.G., Haymet A.D. (1999). Type I 'antifreeze' proteins: Structure-activity studies and mechanisms of ice growth inhibition. *Eur J Biochem*. 264:653-665.

Hashim H.N.F., Bharudin I., Nguong D.L.S., Bakar F.D.A., Nathan S., Rabu A., Kawahara H., Ilias R.M., Najimudin M., Mahadi N.M., Murad A.M.A. (2013). Characterization of Afp1, an antifreeze protein from the psychrophilic yeast *Glaciozyma antarctica* PI12. *Extremophiles* 17(1):63-73.

Haymet A.D.J., Ward L.G., Harding M.M. (1999). Winter flounder 'antifreeze' proteins: Synthesis and ice growth inhibition of analogues that probe the relative importance of hydrophobic and hydrogen-bonding interactions. *J Am Chem Soc.* 121(5):941-8.

Hew C.L., Joshi S., Wang N.C., Kao M.H., Ananthanarayanan V.S. (1985). Structures of shorthorn sculpin antifreeze polypeptides. *Eur J Biochem*. 151:167-172.

Hoshino T., Kiriaki. M., Ohgiya S., Fujiwara M., Kondo H., Nishimiya Y., Yumoto I., Tsuda S. (2003). Antifreeze proteins from snow mold fungi. *Can J Bot.* 81(12): 1175–1181.

Houston Jr. M.E., Chao H., Hodges R.S., Sykes B.D., Kay C.M., Sönnichsen F.D., et al. (1998(). Binding of an oligopeptide to a specific plane of ice. *J Biol Chem.* 273(19):11714-8.

Jia Z., DeLuca C.I., Davies P.L. (1995). Crystallization and preliminary X-ray crystallographic studies on Type III antifreeze protein. *Protein Sci.* 4:1236-1238.

Knight C.A., Hallett J., Devries A.L. (1988). Solute effects on ice recrystallisation: An assessment technique. *Cryobiology*. 25: 55–60.

Knight C.A., Driggers E., DeVries A. L. (1993). Adsorption to ice of fish antifreeze glycopeptides 7 and 8. *Biophys J*. 64: 252–259.

Kun H., Mastai Y. (2007). Activity of short segment of type I antifreeze protein. *Peptide Sci.* 88(6):807.

Kun H., Minnes R., Mastai Y. (2008). Effects antifreeze peptides on thermotropic properties of a model membrane. *J Bioenerg Biomembr*.

Liou Y.C., Davies P.L., Jia Z. (2000). Crystallization and preliminary X-ray analysis of insect antifreeze protein from the beetle *Tenebrio molitor*. Acta Crystallogr D Biol Crystallogr. 56:354-356.

Luthy R, Bowie JU, Eisenberg D. (1992). Assessment of protein models with three-dimensional profiles. *Nature* 356: 83-85.

Ng N.F., Hew C.L. (1992). Structure of an antifreeze polypeptide from the sea raven. Disulfide bonds and similarity to lectin-binding proteins. *J Biol Chem.* 267:16069-16075.

Ng N.F., Trinh K.Y., Hew C.L. (1986). Structure of an antifreeze polypeptide precursor from the sea raven, *Hemitripterus americanus*. J Biol Chem. 261:15690-15695.

Nutt D.R., Smith J.C. (2008). Dual function of hydration layer around an antifreeze protein revealed by atomic molecular dynamics simulations. *J Am Chem Soc.* 130: 13066-13073.

Payne S.R., Sandford D., Harris A., Young O.A. (1994). The effects of antifreeze proteins on chilled and frozen meats. *Meat Sci.* 37: 429-438.

Payne S.R. and Wilson P.W. (1994). Comparison of the freeze-thaw characteristics of Antarctic Cod (*Dissostichus mawsoni*) and Black Cod (*Paranotothenia augustata*) - possible effects of antifreeze glycoproteins. J Musc Foods. 5: 233-244.

Rahman M.B, Zulkifli M.F, Murad A.M, Mahadi N.M, Basri M., Rahman R.N.Z, Salleh A.B. (2008). Ab-Initio Protein Structure Prediction of *Leucosporidium antarcticum* Antifreeze Proteins Using I-TASSER Simulations. *Paper presented at 1st WSEAS International Conference on Biomedical Electronics and Biomedical Informatics, Rhodes, Greece, August 2008.* 

Sansom M. S. (1992). Proline residues in transmembrane helices of channel and transport proteins: A molecular modelling study. *Protein Eng.* 5, 53-60.

Scholander P.F., Van Dam L., Kanwisher J.W., Hammel H.T., Gordon M.S. (1957). Supercooling and osmoregulation in arctic fish. *J Cell Compar Physl.* 49:5-24.

Slaughter D., Fletcher G.L., Ananthanarayanan V.S., Hew C.L. (1981). Antifreeze proteins from the sea raven, *Hemitripterus americanus*. Further evidence for diversity among fish polypeptide antifreezes. *J Biol Chem.* 256: 2022-2026.

Slepkov E.R., Chow S., Lemieux M.J, Fliegel L. (2004). Proline residues in transmembrane segment IV are critical for activity, expression, and targeting of the Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1. *Biochem J*. 379:31-38.

Sonnichsen F.D., DeLuca C.I., Davies P.L., Sykes B.D. (1996). Refined solution structure of type III antifreeze protein: hydrophobic groups may be involved in the energetics of the protein-ice interaction. *Structure* 4:1325-1337.

Wen D., Laursen R.A. (1992). Structure-function relationships in an antifreeze polypeptide. *J Biol. Chem.* 267 (20): 14102-14108.

Wierzbicki A., Dalal P., Cheatham III T.E., Knickelbein J.E., Haymet A.D.J, Madura J.D. (2007). Antifreeze proteins at the ice/water interface: three calculated discriminating properties for orientation of type I proteins. *Biophys J*. 93:1442-1451.

Wishart D.S., Sykes B.D., Richards F.M. (1991). Relationship between nuclear magnetic resonance chemical shift and protein secondary structure. *J Mol Biol.* 222(2):311-33.

Zhang W., Laursen R.A. (1998). Structure-function relationships in a type I antifreeze polypeptide. the role of threonine methyl and hydroxyl groups in antifreeze activity. *J Biol Chem.* 273(52):34806-12.