



UNIVERSITI PUTRA MALAYSIA

***EXTRACTION AND CHARACTERIZATION OF CRUDE PECTIN FROM
JACKFRUIT (*Artocarpus Heterophyllus* Lam.) AND CEMPEDAK
(*Artocarpus Integer* Spreng.) FRUIT RIND***

LEONG CHIA MING

FSTM 2016 25



**EXTRACTION AND CHARACTERIZATION OF CRUDE PECTIN FROM
JACKFRUIT (*Artocarpus Heterophyllus* Lam.) AND CEMPEDAK
(*Artocarpus Integer* Spreng.) FRUIT RIND**

By

LEONG CHIA MING

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Master of Science**

July 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright materials of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to all my beloved family members, my partner and in memory of my dog who has given me continuous support and company all this while. Thank you.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**EXTRACTION AND CHARACTERIZATION OF CRUDE PECTIN FROM
JACKFRUIT (*Artocarpus Heterophyllus* Lam.) AND CEMPEDAK
(*Artocarpus Integer* Spreng.) FRUIT RIND**

By

LEONG CHIA MING

July 2016

Chairman : Noranizan Mohd Adzahan, PhD
Faculty : Food Science and Technology

Jackfruit and cempedak are one of the largest tree-borne fruits and about half of it is the rind. Useful compound such as pectin can be obtained from the rinds of these fruits. Not only that this will reduce byproducts from fruit processing, but also will boost local pectin production industry and contributes to the economy growth. Therefore, the aims of this study were: (i) to determine the most desirable extractant to extract pectin from jackfruit and cempedak fruit rind; (ii) to optimize the extraction process and investigate the physicochemical and rheological properties of the extracted pectin. High methoxyl (HM) pectins with uronic acid content more than 65% were successfully extracted from jackfruit and cempedak fruit rinds using three different acids. Sulfuric acid was the best acid to extract pectin from jackfruit and cempedak rinds as it yielded high amount of pectin ($18.6 \pm 1.8\%$ and $20.5 \pm 0.1\%$, respectively) and the pectin solution produced has the highest brightness and less coloured (red and yellow). Following this, the pectin extraction process was optimized using Response Surface Methodology. The extraction parameters studied (pH, time, temperature) only showed significant effects on the yield of pectin from jackfruit and cempedak fruit rinds. The optimum parameter for extraction of pectin from jackfruit and cempedak fruit rinds was pH 2, 30 min, 90 °C and pH 2, 60 min, 90 °C, respectively. Using the optimum extraction parameter, the yield, uronic acid content and degree of esterification of jackfruit and cempedak fruit rind pectin were $19.84 \pm 0.77\%$ and $19.51 \pm 0.35\%$, $70.67 \pm 0.35\%$ and $70.43 \pm 1.80\%$, and $80.96 \pm 0.11\%$ and $69.98 \pm 0.66\%$, respectively. The information of jackfruit and cempedak fruit rind pectin from this research made these rinds potential new sources of pectin. This could decrease the waste generated from processing of fruits and at the same time creates opportunity for manufacturer to expand their market as jackfruit and cempedak fruit rind pectin were different than existing commercial citrus peel pectin and its natural-occurring colour was superior for certain industry.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**PENGKSTRAKAN DAN CIRI-CIRI PEKTIN MENTAH DARIPADA KULIT
BUAH NANGKA (*Artocarpus Heterophyllus* Lam.) DAN CEMPEDAK
(*Artocarpus Integer* Spreng.)**

Oleh

LEONG CHIA MING

Julai 2016

Pengerusi : Noranizan Mohd Adzahan, PhD
Fakulti : Sains dan Teknologi Makanan

Saiz buah nangka dan cempedak amat besar dan lebih kurang separuh daripada buah ini terdiri daripada kulitnya. Bahan yang berguna seperti pektin boleh diekstrak daripada kulit buah-buahan ini. Ini bukan sahaja dapat mengurangkan sisa buangan yang dihasilkan selepas memproses buah-buahan, tapi juga boleh memperkembangkan industri penghasilan pektin tempatan dan meningkatkan hasil dan pertumbuhan ekonomi negara. Tujuan penyelidikan ini adalah: (i) untuk mengenalpasti asid yang paling sesuai untuk mengekstrak pektin daripada kulit nangka dan cempedak; (ii) untuk mengoptimumkan proses pengekstrakan pektin dan mengkaji sifat fizikokimia dan reologi pektin yang diekstrak. Pektin bermetoksil tinggi dengan kandungan asid uronik melebihi 65% berjaya diekstrak daripada kulit buah nangka dan cempedak menggunakan tiga jenis asid yang berlainan. Asid sulfurik adalah asid yang paling sesuai dimana ia boleh mengekstrak $18.6 \pm 1.8\%$ pektin daripada kulit nangka dan $20.5 \pm 0.1\%$ pektin daripada kulit cempedak. Larutan pektin yang dihasilkan dengan pektin ini adalah yang paling cerah dan kurang berwarna (merah dan kuning). Proses pengekstrakan pektin telah dioptimumkan melalui kaedah Response Surface Methodology. Parameter pengekstrakan (pH, jangka masa, suhu) didapati hanya mempengaruhi kuantiti pektin yang diekstrak daripada kulit nangka dan cempedak. Parameter pengekstrakan optimum yang diperolehi bagi kulit buah nangka adalah pH 2, 30 min dan 90 °C manakala untuk kulit buah cempedak adalah pH 2, 60 min dan 90 °C. Hasil, kandungan asid uronik, dan darjah esterifikasi pektin daripada kulit buah nangka dan cempedak yang diekstrak menggunakan parameter pengekstrakan optimum adalah $19.84 \pm 0.77\%$ dan $19.51 \pm 0.35\%$, $70.67 \pm 0.35\%$ dan $70.43 \pm 1.80\%$, dan $80.96 \pm 0.11\%$ dan $69.98 \pm 0.66\%$. Maklumat yang diperolehi daripada penyelidikan ini memberi ruang untuk pertimbangan kepada pengeluar pektin tempatan untuk menggunakan kulit buah nangka dan cempedak sebagai sumber baru untuk mendapatkan pektin. Ini akan mengurangkan sisa buangan yang dihasilkan daripada pemprosesan buah-buahan. Pada masa yang sama, pengeluar pektin tempatan boleh meluaskan pasaran mereka kerana pektin kulit buah nangka dan cempedak ini

berlainan daripada pektin komersial, iaitu pektin kulit buah sitrus. Di samping itu, warna semulajadi pektin kulit buah nangka dan cempedak adalah suatu kelebihan bagi sesetengah industri.



ACKNOWLEDGEMENTS

First of all, I would like to express my deepest appreciation to my advisor and mentor, Assoc. Prof. Dr. Noranizan Mohd Adzahan for her continuous support and invaluable advices throughout all my study and related research. Her wise comments, patience, and encouragement helped me all the time in my research and the writing of this thesis. Without her, this thesis would be a mountain to climb.

Besides my advisor, I would like to thank my project committee members: Assoc. Prof. Dr. Sharifah Kharidah and Dr. Choo Wee Sim, for their support, comments and also effort to widen and strengthen my research.

I thank my family: mom, dad, both elder and younger sisters for supporting me spiritually and mentally during my study period. I would also like to sincerely thank my partner of 8 years and counting: Ms. Ng See May for her endless support and trust throughout my study.

Last but not least, my sincere appreciation goes to all my fellow labmates for their help, support, company, and brainstorming sessions when working together. To those who were not mentioned, their help is truly appreciated by heart.

I certify that a Thesis Examination Committee has met on 22 July 2016 to conduct the final examination of Leong Chia Ming on his thesis entitled "Extraction and Characterization of Crude Pectin from Jackfruit (*Artocarpus heterophyllus* Lam.) and Cempedak (*Artocarpus integer* Spreng.) Fruit Rind" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Tan Chin Ping, PhD

Professor

Faculty of Food Science and Technology

Universiti Putra Malaysia

(Chairman)

Seyed Hamed Mirhosseini, PhD

Associate Professor

Faculty of Food Science and Technology

Universiti Putra Malaysia

(Internal Examiner)

Maaruf Abd. Ghani, PhD

Associate Professor

National University of Malaysia

Malaysia

(External Examiner)



NOR AINI AB. SHUKOR, PhD

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 3 November 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Noranizan binti Mohd Adzahan, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Sharifah Kharidah Syed Muhammad, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Choo Wee Sim, PhD

Senior Lecturer
School of Science
Monash University Malaysia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecturer notes, learning modules or any others materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Leong Chia Ming, GS30716

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Reversion 2012-2013) are adhered to.

Signature : _____
Name of
Chairman of
Supervisory
Committee : Assoc. Prof. Dr.Noranizan binti MohdAdzahan,

Signature : _____
Name of
Member of
Supervisory
Committee : Assoc. Prof. Dr.Sharifah Kharidah Syed Muhammad

Signature : _____
Name of
Member of
Supervisory
Committee : Dr. Choo Wee Sim

TABLE OF CONTENTS

ABSTRACT		Page
<i>ABSTRAK</i>		i
ACKNOWLEDGEMENT		ii
APPROVAL		iv
DECLARATION		v
LIST OF TABLES		vii
LIST OF FIGURES		xi
LIST OF ABBREVIATIONS		xiii
		xiv
 CHAPTER		
1 INTRODUCTION		1
 2 LITERATURE REVIEW		3
2.1 Taxonomy		3
2.1.1 Jackfruit		3
2.1.2 Cempedak		3
2.1.3 The fruits		4
2.2 Hydrocolloids		6
2.2.1 Pectin		6
2.2.2 Sources of pectin		6
2.3 Pectin extraction		9
2.3.1 Conventional extraction		9
2.3.2 Microwave-assisted extraction		10
2.3.3 Enzyme-assisted extraction		11
2.3.4 Other methods		12
2.4 Types of extractant		13
2.5 Types of pectin		14
2.5.1 High methoxyl pectin		14
2.5.2 Low methoxyl pectin		16
2.6 Applications		17
 3 PHYSICOCHEMICAL PROPERTIES OF CRUDE PECTINS EXTRACTED FROM JACKFRUIT AND CEMPEDAK FRUIT RINDS USING VARIOUS ACIDS		19
3.1 Introduction		19
3.2 Materials and methods		20
3.2.1 Preparation of raw materials		20
3.2.2 Preparation of alcohol insoluble solid		20
3.2.3 Extraction of crude pectin		20
3.2.4 Determination of yield		21
3.2.5 Determination of uronic acid content		21
3.2.6 Determination of degree of esterification		22
3.2.7 Determination of degree of acetylation		22
3.2.8 Determination of colour of the crude pectin solution		23
3.2.9 Statistical analysis		23
3.3 Results and discussions		24

3.3.1	Yield of crude pectin	24
3.3.2	Uronic acid content	25
3.3.3	Degree of esterification	26
3.3.4	Colour of crude pectin solution	27
3.4	Conclusion	28
4	OPTIMIZATION AND DETERMINATION OF PHYSICOCHEMICAL AND RHEOLOGICAL PROPERTIES OF CRUDE PECTIN FROM JACKFRUIT AND CEMPEDAK FRUIT RINDS	29
4.1	Introduction	29
4.2	Materials and methods	30
4.2.1	Preparation of raw materials	30
4.2.2	Preparation of alcohol insoluble solid	30
4.2.3	Extraction of crude pectin and experimental design	30
4.2.4	Determination of yield	31
4.2.5	Determination of uronic acid content	31
4.2.6	Analysis of functional group	31
4.2.7	Determination of degree of esterification	32
4.2.8	Determination of colour of the crude pectin solution	32
4.2.9	Small amplitude oscillatory test	32
4.2.10	Preparation of gel	32
4.2.11	Texture profile analysis	32
4.2.12	Statistical analysis	33
4.3	Results and discussions	33
4.3.1	Statistical analyses	33
4.3.1.1	Yield	39
4.3.1.2	Uronic acid content	41
4.3.1.3	Degree of esterification	42
4.3.2	Optimization	44
4.3.3	Characterization	46
4.3.3.1	Yield and physicochemical properties	46
4.3.3.2	Functional groups	47
4.3.3.3	Colour of pectin solution	49
4.3.3.4	Texture profile	50
4.3.3.5	Viscoelastic properties	53
4.4	Conclusion	55
5	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS	56
	REFERENCES	57
	APPENDICES	72
	BIODATA OF STUDENT	77
	PUBLICATION	78

LIST OF TABLES

Table	Page
1 Compositions of jackfruit and cempedak fruitlets	4
2 Pectin from peel, pomace and/or rind	9
3 Pectin from pulp	9
4 Types of pectin and its function	18
5 Properties of crude pectin extracted from jackfruit rind using various acids	25
6 Properties of crude pectin extracted from cempedak fruit rind using various acids	26
7 Colour parameters (L, a, b), chroma (C) and hue angle (h) of pectin solutions made from jackfruit rind crude pectins that were extracted using various acids	28
8 Colour parameters (L, a, b), chroma (C) and hue angle (h) of pectin solutions made from cempedak fruit rind crude pectins that were extracted using various acids	28
9 Experimental conditions for extraction of crude pectin from jackfruit and cempedak fruit rinds	31
10 Experimental results and predicted results for crude pectin extracted from jackfruit rind	34
11 Experimental results and predicted results for crude pectin extracted from cempedak fruit rind	35
12 Coefficients and significance of all terms in non-reduced full quadratic model for responses of yield, uronic acid content and degree of esterification for crude pectin from jackfruit rind	36
13 Coefficients and significance of all terms in non-reduced full quadratic model for responses of yield, uronic acid content and degree of esterification for crude pectin from cempedak fruit rind	36
14 ANOVA for yield, uronic acid content and degree of esterification of crude pectin from jackfruit rind	37
15 ANOVA for yield, uronic acid content and degree of esterification of crude pectin from cempedak fruit rind	38

16	Regression analysis between jackfruit rind crude pectin yield and extraction variables	40
17	Regression analysis between cempedak fruit rind crude pectin yield and extraction variables	40
18	Regression analysis between uronic acid content of jackfruit rind crude pectin and extraction variables	41
19	Regression analysis between uronic acid content of cempedak fruit rind crude pectin and extraction variables	42
20	Regression analysis between DE of jackfruit rind crude pectin and extraction variables	43
21	Regression analysis between DE of cempedak fruit rind crude pectin and extraction variables	43
22	Yield, uronic acid content and degree of esterification of jackfruit and cempedak fruit rind crude pectins	47
23	Colour parameters (L, a, and b), chroma and hue angle of jackfruit and cempedak fruit rind crude pectin solutions	50
24	Texture profile of jackfruit rind, cempedak fruit rind and citrus peel pectin gel	52

LIST OF FIGURES

Figure		Page
1	Jackfruit	5
2	Cempedak fruit	5
3	Illustration of hydrogen bonding in high methoxyl pectin gel	16
4	Illustration of “egg box” model and calcium bridge of the “egg box” cavity in low methoxyl gel	17
5	Yields of crude pectins extracted from jackfruit and cempedak fruit rinds using various acids	24
6	Response optimizer for the yield of crude pectin from jackfruit rind	45
7	Response optimizer for the yield of crude pectin from cempedak rind	45
8	FT-IR spectra of (a) jackfruit rind crude pectin, (b) cempedak fruit rind crude pectin, and (c) citrus peel pectin	49
9	Dynamic viscoelastic properties of jackfruit rind crude pectin (JRP), cempedak fruit rind crude pectin (CRP) and citrus peel pectin (Cit) solutions at concentration of 4%	54
10	Tan δ of jackfruit rind crude pectin (JRP), cempedak fruit rind crude pectin (CRP) and citrus peel pectin (Cit) solutions at concentration of 4%	54

LIST OF ABBREVIATIONS

AIS	Alcohol insoluble solids
ANOVA	Analysis of variance
CRP	Cempedak fruit rind crude pectin
R ²	Coefficient of determination
DA	Degree of acetylation
DE	Degree of esterification
DM	Degree of methylation
C=O	Ester carbonyl groups
EU	European Union
FAO	Food and Agriculture Organization
FCC	Food Chemical Codex
FT-IR	Fourier transform infrared
HM	High methoxyl
O-H	Hydroxyl group
JRP	Jackfruit rind crude pectin
G''	Loss modulus
LM	Low methoxyl
LMA	Low methoxyl amidated
RSM	Response surface methodology
NaOH	Sodium hydroxide
G'	Storage modulus
H ₂ SO ₄	Sulfuric acid
TPU	Universiti Putra Malaysia's Agriculture Park

CHAPTER 1

INTRODUCTION

Pectin is a nutritious component in human diet. Besides that, it has the ability to form a continuous three-dimensional network of cross-linked polymers which is a condition known as gel (Lotzkar *et al.*, 1946) and this made pectin an important ingredient in foods, beverages, pharmaceutical and a number of other industries.

Pectin can be obtained from most of the plant cell wall but one that suit commercial manufacturing industry is very limited. This is because pectin from different source has different characteristics in which some of them may suit in certain industry while others do not. Most common sources of pectin are from apple pomace and citrus fruit peel. Besides apple pomace and citrus fruit peel, other sources of pectin that have been investigated were such as sugar beet pulp (Renard and Thibault, 1993), sunflower seed head (Miyamoto and Chang, 1992), cocoa husk (Mollea *et al.*, 2008), passion fruit rind (Yapo and Koffi, 2006) and others. Some of them showed potential as good source of pectin but some do not due to their poor gelling ability or unfavourable processing conditions for pectin extraction.

Jackfruit (*Artocarpus heterophyllus* Lam.) and cempedak (*Artocarpus integer* Spreng.) are fruits that are available in Malaysia. The parts of jackfruit and cempedak trees and their fruits can be used as sources of food, as timbers for furniture, as feeds for livestock and others. According to statistics from Ministry of Agriculture and Agro-based Industry Malaysia, the amount of jackfruit and cempedak produced in 2014 were 33,788 tonnes and 35,563 tonnes, respectively. Given that 40-60% of these fruits were the rind (Chadha, 1985), the amount of jackfruit and cempedak fruit rinds produced in 2014 were approximately 13,515 tonnes and 14,225 tonnes, respectively. These rinds were only used as feed for cows and goats. The remaining rinds were normally disposed, which would be a challenge for the environment.

Despite being one of the largest tree-borne fruits in the world, jackfruit and cempedak have never been extensively studied before for its rind's pectin content. Up to the author's knowledge, there are only two studies on pectin from jackfruit rind and none regarding pectin from cempedak fruit rind. Among the two studies on jackfruit rind pectin, one of them screened and characterized pectins from various fruit wastes which include jackfruit waste while another study investigated the effect of different extractants on the characteristics of pectin obtained from jackfruit waste.

Common sources of pectin such as apple pomace and citrus fruit peel are fruits that are unsuitable to be planted in Malaysia in a commercial scale. It was shown that jackfruit rind contained high amount of pectin. Cempedak, which has similar percentage of rind per fruit as jackfruit, should have similar amount of pectin as in jackfruit rind. As jackfruit and cempedak are fruits of different species, it was expected that the extracted

pectins from both rinds have different characteristics. This research is important in investigating new sources of pectin that are locally available. In addition, the physicochemical and rheological properties of pectins extracted from jackfruit and cempedak fruit rinds have never been reported before. Therefore, the objectives of this study are:

1. To determine the effects of citric, nitric and sulfuric acids on the yield and properties of pectins extracted from jackfruit and cempedak fruit rinds.
2. To investigate the effects of pH, time and temperature on the yield, uronic acid content and degree of esterification of pectin from jackfruit and cempedak fruit rinds and determine the physicochemical and rheological properties of the pectins extracted using optimum condition.

REFERENCES

- Agrofood Statistic*. 2014. Ministry of Agriculture and Agro-based Industry Malaysia. Information Management and Statistics Section: Malaysia.
- Ahmed, A.E.R. and Labavitch, J.M. (1977). A simplified method for accurate determination of cell wall uronide content. *Journal of Food Biochemistry*, 1, 361-365.
- Alagiapillai, O.A., Kuttalam, P.S., Subramaniam, V. and Jayasekhar, M. (1996). PPI-I jack: A new high yielding, regular bearing jack variety for Tamil Nadu. *Madras Agricultural Journal*, 83, 310-312
- Arung, E.T., Shimizu, K. and Kondo, R. (2007). Structure-activity relationship of prenyl-substituted polyphenols from *Artocarpus heterophyllus* as inhibitors of melanin biosynthesis in cultured melanoma cells. *Chemistry and Biodiversity*, 4, 2166 – 2171.
- Arung, E.T., Wicaksono, B.D., Handoko, Y.A., Kusuma, I.W., Shimizu, K., Yulia, D. and Sandra, F. (2010). Cytotoxic effect of artocarpin on T47D cells. *Journal of Natural Medicine*, 64, 423 – 429.
- Ashford, M., Fell, J., Attwood, D., Sharma, H. and Woodhead, P. (1994). Studies on pectin formulations for colonic drug delivery. *Journal of Controlled Release*, 30(3), 225-232.
- Axelos, M.A.V. and Thibault, J.F. (1991). The chemistry of low methoxyl pectin. In Walter, R.H., Eds., *The chemistry and technology of pectin*. Academic Press, New York, 109.
- Azad, A.K. (2000). Genetic diversity of jackfruit in Bangladesh and development of propagation methods. Ph.D. thesis, University of Southampton, United Kingdom.
- Bagherian, H., Ashtiani, F.Z., Fouladitajar, A. and Mohtashamy, M. (2011). Comparisons between conventional, microwave- and ultrasound-assisted methods for extraction of pectin from grapefruit. *Chemical Engineering and Processing*, 50, 1237-1243.
- Baker, G.L. (1948). High-polymer pectins and their deesterification. In E.M. Mark and G.R. Stewart, Eds., *Advances in Food Research*, p. 395.
- Baltazar, E.P. (1984) *A Handbook of Philippine Crops*. San Fernando, La Union, Philippines: 76-80.
- Bapat, V.A. and Mhatre, M. (2005). *Ficus carica* Fig, *Artocarpus* spp. Jackfruit and Breadfruit, and *Morus* spp. Mulberry. In Litz, R.E. (Eds). *Biotechnology of Fruit and Nut Crops: Volume 29 of Biotechnology in agriculture series*, p. 350-362.

- Begum, R., Aziz, M.G., Uddin, M.B. and Yusof, Y.A. (2014). Characterization of jackfruit (*Artocarpus heterophyllus*) waste pectin as influenced by various extraction conditions. *Agriculture and Agricultural Science Procedia* 2, 244-251.
- Bélafi-Bakó, K., Cserjési, P., Beszédes, S., Csanádi, Z. and Hodúr, C. (2011). Berry Pectins: Microwave-assisted extraction and rheological properties. *Food and Bioprocess Technology*, 5, 1100-1105.
- BeMiller, J. N. (1986). An introduction to pectin: Structure and properties. In M. L. Fishman, and J. J. Jen (Eds.), *Chemistry and function of pectins*. ACS symposium series 310 (pp. 2–12). Washington, DC: American Chemical Society.
- Black, S.A. and Smit, C.J.B. (1972). The effect of demethylation procedure on the quality of low-ester pectins used in dessert gels. *Journal of Food Science*, 37(5), 730-732.
- Blumenkrantz, N. and Asboe-Hansen, G. (1973). New method for quantitative determination of uronic acids. *Analytical Biochemistry*, 54, 484–489.
- Bochek, A.M., Zabivalova, N.M. and Petropavlovskii, G.A. (2001). Determination of the esterification degree of polygalacturonic acid. *Russian Journal of Applied Chemistry*, 74(5), 796-799.
- Bose, T.K. (1985). Jackfruit. In: Mitra, B.K. (ed.), *Fruits of India: Tropical and Subtropical*. Naya Prokas, Culcutta: 488-497.
- Britton, G. and Hornero-Méndez, D. (1997). Carotenoids and colour in fruit and vegetables. In Tomás-Barberán, F. A. and Robins, R. J. (Eds.), *Phytochemistry of fruit and vegetables* (pp. 11–27). Oxford: Clarendon Press.
- Canteri, M.H.G., Scheer, A.P., Wosiacki, G., Ginies, C., Reich, M. and Renard, C.M.C.G. (2010). A comparative study of pectin extracted from passion fruit rind flours. *Journal of Polymers and The Environment*, 18, 593-599.
- Canteri-Schemin, M.H., Fertonani, H.C.R., Waszczynskyj, N. and Wosiacki, G. (2005). Extraction of pectin from apple pomace. *Brazilian Archives of Biology and Technology*, 48(2), 259-266.
- Capel, F., Nicolai, T., Durand, D., Boulenguer, P. and Langendorff, V. (2006). Calcium and acid induced gelation of (amidated) low methoxyl pectin. *Food Hydrocolloids*, 20, 901–907.
- Carpita, N. and McCann, M. C. (2000). The cell wall. In B. Buchanan, W. Gruissel and R. Jones, Eds., *Biochemistry and molecular biology of plants*, pp. 52–108. Rockville, MD: American Society of Plant Physiologists.

- Cedra, J.J., Robinsons, F.L., Burgin, C.W., Baumgartner, T.G. and Rice, R.W. (1988). The effects of grapefruit pectin on patients at risk for coronary heart diseases. *Clinical Cardiology*, 11(9), 589-594.
- Chadha, Y.R. (1985). The wealth of India - Raw Materials series, Ed. 1A. Publications and Information Directorate, CSIR, New Delhi, India, pg. 450.
- Chan, S.-Y. and Choo, W.-S. (2013). Effect of extraction conditions on the yield and chemical properties of pectin from cocoa husks. *Food Chemistry*, 141, 3752-3758.
- Chanda, I., Chanda, S.R. and Dutta, S.K. (2009). Anti-inflammatory activity of a protease extracted from the fruit stem latex of the plant *Artocarpus heterophyllus* Lam. *Research Journal of Pharmacology and Pharmacodynamics*, 1, 70 – 72
- Chandrika, U.G., Jansz, E.R. and Wamasuriya, N.D. (2004). Analysis of carotenoids in ripe jackfruit (*Artocarpus heterophyllus*) kernel and study of their bioconversion in rats. *Journal of the Science of Food and Agriculture*, 85, 186 – 190.
- Chen, J., Liang, R.-h., Liu, W., Luo, S.-j., Liu, C.-m., Wu, S.-s. and Wang, Z.-j. (2014). Extraction of pectin from *Premna microphylla* turcz leaves and its physicochemical properties. *Carbohydrate Polymers*, 102, 376-384.
- Cheng, L.V., Wang, Y., Wang, L.-j., Li, D. and Adhikari, B. (2013). Optimization of production yield and functional properties of pectin extracted from sugar beet pulp. *Carbohydrate Polymers*, 95, 233-240.
- Concepcion, R.F. (1990) Jackfruit: Aromatic money-maker. *Agrobusiness Weekly* 4(3):12-13.
- Contreras-Esquivel, J.C., Voget, C.E., Vita, C.E., Espinoza-Perez, J.D. and Renard, C.M.G.C. (2006). Enzymatic extraction of lemon pectin by endopolygalacturonase from *Aspergillus niger*. *Food Science and Biotechnology*, 15, 163–167.
- Crandall, P.G. and Wicker, L. (1986). Pectin internal gel strength: theory, measurement and methodology, in Chemistry and Functions of Pectin, Fishman, M.L and Jen, J.J., Eds., American Chemical Society, Washington, D.C., 88.
- Dignan, C.A., Burlingame, B.A., Arthur, J.M., Quigley, R.J. and Milligan, G.C. (1994). The Pacific islands food composition tables. South Pacific Commission, Noumea, New Caledonia.
- Doco, T., Williams, P., Vidal, S. and Pellerin, P. (1997). Rhamnogalacturonan II, a dominant polysaccharide in juices produced by enzymic liquefaction of fruits and vegetables. *Carbohydrate Research*, 297, 181-186.

- Elevitch, C.R., and Manner, H.I., (2006). *Artocarpus heterophyllus* (jackfruit) IN: Species profiles for Pacific Island agroforestry. www.traditionaltree.org
- Eliaz, I. (2002). The potential role of modified citrus pectin in the prevention of cancer metastasis. *Clinical Practice of Alternative Medicine*, 2, 177–179.
- El-Nawawi, S. A., and Shehata, F. R. (1987). Extraction of pectin from Egyptian orange peel. Factors affecting the extraction. *Biological Wastes*, 20, 281–290.
- El-Nawawi, S. A., and Heikal, Y. A. (1995). Production of a low ester pectin by deesterification of high ester citrus pectin. *Carbohydrate Polymers*, 27, 191–195.
- Emaga, T.H., Ronkart, S.N., Robert, C., Wathelet, B. and Paquot, M. (2008). Characterisation of pectins extracted from banana peels (*Musa AAA*) under different conditions using an experimental design. *Food Chemistry*, 108, 463–471.
- Fang, S.C., Hsu, C.L. and Yen, G.C. (2008). Anti-inflammatory effects of phenolic compounds isolated from the fruits of *Artocarpus heterophyllus*. *Journal of Agricultural and Food Chemistry*, 56, 4463 – 4468.
- Fellah, A., Anjukandi, P., Waterland, M.R. and Williams, M.A.K. (2009). Determining the degree of methylesterification of pectin by ATR/FT-IR: Methodology optimisation and comparison with theoretical calculations. *Carbohydrate Polymers*, 78, 847–853.
- Fernandez, M.L., Sun, D.M., Tosca, M.A. and McNamara, D.J. (1994a). Citrus pectin and cholesterol interact to regulate hepatic cholesterol homeostasis and lipoprotein metabolism: a dose response study in guinea pigs. *The American Journal of Clinical Nutrition*, 59, 869–878.
- Fernandez, M.L., Lin, E.C.K., Trejo, A. and McNamara, D.J. (1994b). Prickly pear (*Opuntia sp.*) pectin alters hepatic cholesterol metabolism without affecting cholesterol absorption in guinea pigs fed a hypercholestermic diet. *Journal of Nutrition*, 124, 817–824.
- Filisetti-Cozzi, T.M.C.C. and Carpita, N.C. (1991). Measurement of uronic acids without interference from neutral sugars. *Analytical Biochemistry*, 197, 157–162.
- Gan, C.-Y., Manaf, N. H. A. and Latiff, A. A. (2010). Physico-chemical properties of alcohol precipitate pectin-like polysaccharides from *Parkia speciosa* pod. *Food Hydrocolloids*, 24, 471–478.
- Garna, H., Mabon, N., Robert, C., Cornet, C., Nott, K., Legros, H., Wathelet, B. and Paquot, M. (2007). Effect of extraction conditions on the yield and purity of apple pomace pectin precipitated but not washed by alcohol. *Journal of Food Science*, 72 (1), 1–9.

- Geerkens, C.H., Nagel, A., Just, K.M., Miller-Rostek, P., Kammerer, D.R., Schweiggert, R.M. and Carle, R. (2015). Mango pectin quality as influenced by cultivar, ripeness, peel particle size, blanching, drying, and irradiation. *Food Hydrocolloid*, 51, 241-251.
- Ghosh, S.P. (1996) *Technical Report for Use of Underutilised Tropical fruits in Asia Network*. UTFANET, Southampton University, UK.
- Gnanasambandam, R. and Proctor, A. (2000). Determination of pectin degree of esterification by diffuse reflectance Fourier transform infrared spectroscopy. *Food Chemistry*, 68(3), 327–332.
- Grant, G.T., Morris, E.R., Rees, D.A., Smith, P.J.C. and Thom, D. (1973). Biological interactions between polysaccharides and divalent cations: the egg box model. *FEBS Letter*, 32, 195-198.
- Gross, M.O., Rao, V.N.M. and Smit, C.J.B. (1980). Rheological characterization of low methoxyl pectin gel by normal creep and relaxation. *Journal of Texture Studies*, 11, 271-290.
- Gunaseana, H.P.M., Ariyadasa, K.P., Wikramasinghe, A., Herath, H.M.W., Wikramasinghe, P. and Rajakaruna, S.B. (1996). Manual of jack cultivation in Sri Lanka: Forest Information Service, Forest Department 48.
- Hameed, B.H. (2009). Removal of cationic dye from aqueous solution using jackfruit peel as non-conventional low-cost adsorbent. *Journal of Hazardous Materials*, 162, 344 – 350
- Haq, N. (2006). Jackfruit, *Artocarpus heterophyllus*, Southampton Centre for Underutilised Crops, University of Southampton, Southampton, UK.
- Hoefer, A.C. (1991). Other pectin food products. In *The chemistry and technology of pectin*, ed. R.H.Walter, pp. 51-67. New York: Academic Press.
- Ibarz, A., Pagan, A., Tribaldo, F., and Pagan, J., (2006). Improvement in the measurement of spectrophotometric data in the *m*-hydroxydiphenyl pectin determination methods. *Food Control*, 17, 890-893.
- Iglesias, M.T. and Lozano, J.E. (2004). Extraction and characterization of sunflower pectin. *Journal of Food Engineering*, 62, 215-223.
- Inbaraj, B.S. and Sulochana, N. (2004). Carbonised jackfruit peel as an adsorbent for the removal of Cd(II) from aqueous solution. *Bioresource Technology*, 94, 49 – 52.
- Jackson, C.L., Dreaden, T.M., Theobald, L.K., Tran, N.M., Beal, T.L., Eid, M., Gao, M.Y., Shirley, R.B., Stoffel, M.T., Kumar, M.V. and Mohnen, D. (2007). Pectin induces apoptosis in human prostate cancer cells: Correlation of apoptotic function with pectin structure. *Glycobiology*, 17, 805-819.

- Jagdeesh, S.L., Reddy, B.S., Basavraj, N., Swamy, G.S.K. and Hedge, L. (2010). Variability studies in physico-chemical qualities of jackfruit (*Artocarpus heterophyllus* Lam.) of coastal zone of Karnataka. *Karnataka Journal of Agricultural Sciences*, 23, 293 – 297.
- Jagtap, U.B., and Bapat, V.A., (2010). *Artocarpus*: A review of its traditional uses, phytochemistry and pharmacology. *Journal of Ethnopharmacology*, 129, 142-166.
- Janick, J. and Paull, R.E. (2008). Citrus. In Janick, J. and Paull, R.E. (Eds). *The Encyclopedia of Fruit and Nuts*, p. 778-789. CABI.
- Jayani, R.S., Saxena, S. and Gupta, R. (2005). Microbial pectinolytic enzymes: A review. *Process Biochemistry*, 40, 2931-2944.
- Jiang, Y., Du, Y., Zhu, X., Xiong, H., Woo, M.W. and Hu., J. (2012). Physicochemical and comparative properties of pectins extracted from Akebia trifoliata var. australis peel. *Carbohydrate Polymers*, 87, 1663-1669.
- Kačuráková, M., Capek, P., Sasinková, V., Wellner, N. and Ebringerová, A. (2000). FT-IR study of plant cell wall model compounds: pectic polysaccharides and hemicelluloses. *Carbohydrate Polymers*, 43(2), 195-203.
- Kalapathy, U. and Proctor, A. (2001). Effect of acid extraction and alcohol precipitation conditions on the yield and purity of soy hull pectin. *Food Chemistry*, 73, 393-396.
- Kamnev, A. A., Colina, M., Rodriguez, J., Ptitchkina, N. M. and Ignatov, V. V. (1998). Comparative spectroscopic characterization of different pectins and their sources. *Food Hydrocolloids*, 12, 263–271.
- Karthy, E.S., Ranjitha, P. and Mohankumar, A. (2009). Antimicrobial potential of plant seed extracts against multidrug resistant Methicillin Resistant *Staphylococcus aureus* (MDR-MRSA). *International Journal of Biology*, 1, 34 – 40.
- Khan, M.R., Omoloso, A.D. and Kihara, M. (2003). Antibacterial activity of *Artocarpus heterophyllus*. *Fitoterapia*, 74, 501 – 505.
- Kliemann, E., de Simas, K.N., Amante, E.R., Prudêncio, E.S., Teófilo, R.F., Ferreira, M.M.C. and Amboni, R.D.M.C. (2008). Optimisation of pectin acid extraction from passion fruit peel (*Passiflora edulis flavicarpa*) using response surface methodology. *International Journal of Food Science and Technology*, 44, 476-483.
- Koh, P.C., Leong, C.M. and Noranizan, M.A. (2014). Microwave-assisted extraction of pectin from jackfruit rinds using different power levels. *International Food Research Journal*, 21(5), 2091-2097.

- Koubala, B.B., Christiaens, S., Kansci, G., Loey, A.M.V. and Hendrickx, M.E. (2014). Isolation and structural characterisation of papaya peel pectin. *Food Research International*, 55, 215-221.
- Kratchanova, M., Pavlova, E. and Panchev, I. (2004). The effect of microwave heating of fresh orange peels on the fruit tissue and quality of extracted pectin. *Carbohydrate Polymers*, 56, 181-185.
- Kulkarni, S.G. and Vijayanand, P. (2010). Effect of extraction conditions on the quality characteristics of pectin from passion fruit peel (*Passiflora edulis f. flavicarpa* L.). *Food Science and Technology*, 43, 1026-1031.
- Kumar, A. and Chauhan, G.S. (2010). Extraction and characterization of pectin from apple pomace and its evaluation as lipase (steapsin) inhibitor. *Carbohydrate Polymers*, 82, 454-459.
- Levigne, S., Ralet, M-C. and Thibault, J-F. (2002a). Characterization of pectins extracted from fresh sugar beet under different conditions using an experimental design. *Carbohydrate Polymers*, 49, 145-153.
- Lim, T.K. (2012). *Artocarpus integer*. In Lim, T.K. (Eds). *Edible Medicinal And Non Medicinal Plants: Volume 3, Fruits*, p. 337-343. Springer Science and Business Media.
- Lim, J., Yoo, J., Ko, S. and Lee, S. (2012). Extraction and characterization of pectin from Yuza (*Citrus junos*) pomace: A comparison of conventional-chemical and combined physicochemical extractions. *Food Hydrocolloids*, 29, 160-165.
- Liu, F.X., Fu, S.F., Bi, X.F., Chen, F., Liao, X.J., Hu, X.S. and Wu, J.H. (2013). Physicochemical and antioxidant properties of four mango (*Mangifera indica* L.) cultivars in China. *Food Chemistry*, 138(1), 396-405.
- Lotzkar, H., Schultz, T.H., Owens, H.S., and MacLay, W.D., (1946). Effect of salts on the viscosity of pectinic acid solutions. *Journal of Physical Chemistry*, 50, 200.
- Mao, R., Tang, J. and Swanson, B.G. (2001). Water holding capacity and microstructure of gellan gels. *Carbohydrate Polymers*, 46, 365-371.
- Maran, J.P., Sivakumar, V., Thirugnanasambandham, K. and Sridhar, R. (2013). Optimization of microwave assisted extraction of pectin from orange peel. *Carbohydrate Polymers*, 97, 703-709.
- Marcelin, O., Saulnier, L. and Brillouet, J-M. (1991). Extraction and characterisation of water-soluble pectic substances from guava (*Psidium guajava* L.). *Carbohydrate Research*, 212, 159-167.
- Masmoudi, M., Besbes, S., Chaabouni, M., Robert, C., Paquot, M., Blecker, C. and Attia, H. (2008). Optimization of pectin extraction from lemon by-product

with acidified date juice using response surface methodology. *Carbohydrate Polymers*, 74, 185-192.

- Matsunaga, T., Ishii, T., Matsumoto, S., Higuchi, M., Darvill, A., Albersheim, P. and O'Neill, M.A. (2004). Occurrence of the primary cell wall polysaccharide rhamnogalacturonan-II in pteridophytes, lycophytes, and bryophytes. Implications for the evolution of vascular plants. *Plant Physiology*, 134, 339-351.
- May, C.D. (1990). Industrial pectins: Sources, production and applications. *Carbohydrate Polymers*, 12, 79-99.
- May, C.D. (2000). Pectin. In Phillips, G.O. and Williams, P.A. (Eds). *Handbook of Hydrocolloids*, 169-188. CRC Press.
- Methacanon, P., Kongsin, J. and Gamonpilas, C. (2014). Pomelo (*Citrus maxima*) pectin: Effects of extraction parameters and its properties. *Food Hydrocolloids*, 35, 383-391.
- Min, B., Lim, J., Ko, S., Lee, K-G., Lee, S.H. and Lee, S. (2011). Environmentally friendly preparation of pectins from agricultural byproducts and their structural/rheological characterization. *Bioresource Technology*, 102, 3855-3860.
- Minjares-Fuentes, R., Femenia, A., Garau, M.C., Meza-Velázquez, J.A., Simal, S. and Rosselló, C. (2014). Ultrasound-assisted extraction of pectins from grape pomace using citric acid: A response surface methodology approach. *Carbohydrate Polymers*, 106, 179-189.
- Mitra, S.K. and Maity, C.S. (2002) A summary of the genetic resources of jackfruit (*Artocarpus heterophyllus* Lam.) in West Bengal, India. *Acta Horticulturae*. 575.
- Miyamoto, A., and Chang, K. C. (1992). Extraction and physicochemical characterization of pectin from sunflower head residues. *Journal of Food Science*, 57(6), 1439–1443.
- Mohamed, S. and Hasan, Z. (1995). Extraction and characterisation of pectin from various tropical agrowaste. *ASEAN Food Journal*, 10(2), 43-50.
- Mollea, C., Chiampo, F., and Conti, R. (2008). Extraction and characterization of pectins from cocoa husks: A preliminary study. *Food Chemistry*, 107, 1353-1356.
- Mort, A. J., Qui, F. and Maness, N. O. (1993). Determination of the pattern of methyl esterification in pectin. Distribution of contiguous nonesterified residues. *Carbohydrate Research*, 247, 21–35.
- Muhammad, K., Mohd. Zahari, N.I., Gannasin, S.P., Noranizan, M.A. and Bakar, J. (2014). High methoxyl pectin from dragon fruit (*Hylocereus polyrhizus*) peel, *Food Hydrocolloids*, 42, 289-297.

- Mukhiddinov, Z.K., Khalikov, D.K., Abdusamiev, F.T., and Avloev, C.C. (2000). Isolation and structural characterization of a pectin homo and rhamnogalacturonan. *Talanta*, 53, 171-176.
- Naghshineh, M., Olsen, K. and Georgiou, C.A. (2013). Sustainable production of pectin from lime peel by high hydrostatic pressure treatment. *Food Chemistry*, 136, 472-478.
- Nangia-Makker, P., Hogan, V., Honjo, Y., Baccarini, S., Tait, L., Bresalier, R. and Raz, A. (2002). Inhibition of Human Cancer Cell Growth and Metastasis in Nude Mice by Oral Intake of Modified Citrus Pectin. *Journal of the National Cancer Institute*, 94(24), 1854-1862.
- Narasimham, P. (1990). Breadfruit and jackfruit. In: Nagy, S., Shaw, P.E. and Wardowski, W.F. (eds.), *Fruits of tropical and subtropical origin: Composition, properties and uses*. Florida: Florida Science Source Inc.: 193-259.
- Nussinovitch, A. (1997). *Hydrocolloid application: Gum technology in the food and other industries*. New York: Blackie Academic and Professional.
- Nwanekezi, E.C., Alawuba, O.C.G. and Mkpolulu, C.C.M. (1994). Characterization of pectic substances from selected tropical fruits. *Journal of Food Science and Technology*, 31(2), 159-161.
- Oakenfull, D.G. (1991). The chemistry of high-methoxyl pectins. In *The chemistry and technology of pectin*, ed. R.H.Walter, pp. 87-109. New York: Academic Press.
- Ochse, J.J., Soule (Jr.), M.J., Dijkman, M.J. and Welburg, C. (1961). *Tropical and Sub-tropical Agriculture*. Macmillan Co: 652-655.
- O'Donoghue, E.M. and Somerfield, S.D. (2008). Biochemical and rheological properties of gelling pectic isolates from buttercup squash fruit. *Food Hydrocolloids*, 22, 1326-1336.
- Olano-Martin, E., Rimbach, G.H., Gibson, G.R. and Rastall, R.A. (2003). Pectin and pectic-oligosaccharides induce apoptosis in *in vitro* human colonic adenocarcinoma cells. *Anticancer Research*, 23(1A), 341-346.
- Oliveira, T.I.S., Rosa, M.F., Cavalcante, F.L., Pereira, P.H.F., Moates, G.K., Wellner, N., Mazzetto, S.E., Waldron, K.W. and Azeredo, H.M.C. (2016). Optimization of pectin extraction from banana peels with citric acid by using response surface methodology. *Food Chemistry*, 198, 113-118.
- O'Neill, M.A., Ishii, T., Albersheim, P. and Darvill, A. (2004). Rhamnogalacturonan-II: Structure and function of a borate cross-linked cell wall pectic polysaccharide. *Annual Review of Plant Biology*, 55, 109-139.
- Ong, B.T., Nazimah, S.A.H., Osman, A., Quek, S.Y., Voon, Y.Y., Hashim, D.M., Chew P.M. and Kong Y.W. (2006). Chemical and flavour changes in jackfruit

- (*Artocarpus heterophyllus* Lam) Cultivar J3 during ripening. *Journal of Postharvest Biology and Technology*, 40(3), 279-286.
- Ovodov, Y.S. (2009). Current views on pectin substances. *Russian Journal of Bioorganic Chemistry*, 35(3), 269-284.
- Pagán, J. and Ibarz, A. (1999). Extraction and rheological properties of pectin from fresh peach pomace. *Journal of Food Engineering*, 39, 193-201.
- Pagan, J., Ibarz, A., Llorca, M., Pagan, A. and Barbosa-Canovas G.V. (2001). Extraction and characterization of pectin from stored peach pomace. *Food Research International*, 34, 605-612.
- Patil, K.S., Jadhav, A.G. and Joshi, V.S. (2005). Wound healing activity of leaves of *Artocarpus heterophyllus*. *Indian Journal of Pharmaceutical Sciences*, 67, 629 – 632.
- Paull, R.E. and Duarte O. (2012). Breadfruit, jackfruit, cempedak and marang. In Paull, R.E. and Duarte O. (Eds). *Tropical Fruits, Volume 2*, p. 25-52. CABI.
- Phatak, L., Chang, K.C. and Brown, G. (1988). Isolation and characterization of sugar-beet pulp. *Journal of Food Science*, 53, 830-833.
- Pilgrim, G.W., Walter, R.H. and Oakenfull, D.G. (1991). Jams, jellies and preserves, The Chemistry and Technology of Pectin, Walter, R.H., Ed., Academic Press, San Diego, 23.
- Pinheiro, E.R., Silva, I.M.D.A., Gonzaga, L.V., Amante, E.R., Teófilo, R.F., Ferreira, M.M.C. and Amboni, R.D.M.C. (2008). Optimization of extraction of high-ester pectin from passion fruit peel (*Passiflora edulis flavicarpa*) with citric acid by using response surface methodology. *Bioresource Technology*, 99, 5561-5566.
- Pippen, E.L., McCready, R.M. and Owens, H.S. (1950a). Determination of Acetyl in Pectin. *Analytical Chemistry*, 22(11), 1457-1458.
- Pippen, E.L., McCready, R.M. and Owens, H.S. (1950b). Gelation properties of partially acetylated pectins. *Journal of the American Chemical Society*, 72, 813-816.
- Popenoe, W. (1974) *Manual of Tropical and Sub-tropical Fruits*. Halfner Press Co., New York: 414-419.
- Powell, D.A., Morris, E.R., Gidley, M.J. and Rees, D.A. (1982). Conformation and interactions of pectins. II. Influence of residue sequence on their chain association in calcium pectate gels. *Journal of Molecular Biology*, 155, 517-531.
- Prabasari, I., Pettolino, F.F., Liao, M.-L. and Bacic, A. (2011). Pectic polysaccharides from mature orange (*Citrus sinensis*) fruit albedo cell walls: Sequential

- extraction and chemical characterization. *Carbohydrate Polymers*, 84, 484-494.
- Prahas, D., Kartika, Y., Indraswati, N. and Ismadji, S. (2008). Activated carbon from jackfruit peel waste by H_3PO_4 chemical activation: Pore structure and surface chemistry characterization. *Chemical Engineering Journal*, 140, 32 – 42.
- Prakash, O., Kumar, R., Mishra, A. and Gupta, R. (2009). *Artocarpus heterophyllus* (Jackfruit): An overview. *Pharmacognosy Review*, 3, 353 – 358.
- Pressey, R., Hinton, D.M. and Avants, J.K. (1971). Development of polygalacturonase activity and solubilization of pectin in peaches during ripening. *Journal of Food Science*, 36, 1070.
- Ptichkina, N.M., Markina, O.A. and Rumyantseva, G.N. (2008). Pectin extraction from pumpkin with the aid of microbial enzymes. *Food Hydrocolloids*, 22, 192-195.
- Purseglove, J.W. (1968). The origin and spread of tropical crops. In *Tropical crops. Dicotyledons 2*, ed. J.W. Purseglove, pp. 9-18. New York: Wiley.
- Rascón-Chu, A., Martínez-López, A.L., Carvajal-Millán, E., de León-Renova, N.E.P., Márquez-Escalante, J.A. and Romo-Chacón, A. (2009). Pectin from low quality Golden Delicious apples: Composition and gelling capability. *Food Chemistry*, 116, 101-103.
- Reddy, B.M.C., Patil, P., Shashikumar, S. and Govindaraju, L.R. (2004). Study on physico-chemical characteristics of jackfruit clones of south Karnataka. *Karnataka Journal of Agricultural Science*, 17, 279-282.
- Renard, C.M.G.C. and Thibault, J-F. (1993). Structure and properties of apple and sugar-beet pectins extracted by chelating agents. *Carbohydrate Research*, 244, 99-114.
- Roboz, E. and Van Hook, A. (1946). Chemical study of beet pectin. *Proceedings of American Society Sugar Beet Technologist*, 4, 574-583.
- Rombouts, F.M. and Thibault, J-F. (1986). Feruloylated pectic substances from sugar-beet pulp. *Carbohydrate Research*, 154, 177-187.
- Rouse, A.H. and Crandall, P.G. (1978). Pectin content of lime and lemon peel as extracted by nitric acid. *Journal of Food Science*, 43, 72-73.
- Rowe-Dutton, P. (1985). *Artocarpus heterophyllus* – Jackfruit. In *The Propagation of Tropical Fruit Trees*, ed. Garner, J.R. and Chaudhury, S.A., pp. 269-290. London: FAO/CAB.
- Rujinirum, C., Phinyocheep, P., Prachyabrued, W. and Laemsak, N. (2005). Chemical treatment of wood for musical instruments. Part I. Acoustically important properties of wood for the Ranad (Thai traditional xylophone). *Wood Science Technology*, 39, 77-85.

- Saha, D. and Bhattacharya, S. (2010). Hydrocolloids as thickening and gelling agents in food: A critical review. *Journal of Food Science and Technology*, 47(6), 587-597
- Schmidgall, J. and Hensel, A. (2002). Bioadhesive properties of polygalacturonides against colonic epithelial membranes. *International Journal of Biological Macromolecules*, 30, 217-225.
- Seggiani, M., Puccini, M., Pierini, M., Giovando, S. and Forneris, C. (2009). Effect of different extraction and precipitation methods on yield and quality of pectin. *International Journal of Food Science and Technology*, 44, 574-580.
- Shkodina, O.G., Zeltser, O.A., Selivanov, N.Y. and Ignatov, V.V. (1998). Enzymic extraction of pectin preparations from pumpkin. *Food Hydrocolloids*, 12, 313-316.
- Shurvell, H. F. (2002). Spectra-structure correlations in the mid- and far-infrared. In J. M. Chalmers, and P. R. Griffiths (Eds.), *Handbook of vibrational spectroscopy* (pp. 1783–1817). New York: John Wiley and Sons, Inc.
- Silverstein, R. M., Bassler, G. C. and Morrill, T. C. (1991). Spectrometric identification of organic compounds. New York: Wiley.
- Soepadmo, E. (1992). *Artocarpus heterophyllus* Lamk. In *Plant Resources of Southeast Asia No.2: Edible Fruits and Nuts*. PROSEA, ed. Verheij, E.W.M. and Coronel, R.E., pp. 86-91. Netherlands: Wageningen.
- Speiser, R., Eddy, C.R. and Hills, C.H. (1945). Kinetics of deesterification of pectin. *The Journal of Physical Chemistry*, 49, 563-579.
- Sriamornsak, P. (1999). Effect of calcium concentration, hardening agent and drying condition on release characteristics of oral proteins from calcium pectinate gel beads. *European Journal of Pharmaceutical Sciences*, 8(3), 221-227.
- Sriamornsak, P. (2002). *Analysis of selected physicochemical properties of pectin and alginate gels intended for drug delivery*, PhD Thesis, Charles Sturt University.
- Sriamornsak, P. (2003). Chemistry of pectin and its pharmaceutical uses: A Review. *Silpakorn University International Journal*, 3(1-2), 207-228.
- Sriamornsak, P., Sungthongjeen, S. and Puttipipatkachorn, S. (2007). Use of pectin as a carrier for intragastric floating drug delivery: Carbonate salt contained beads. *Carbohydrate Polymer*, 67(3), 436-445.
- Stasse-Wolthuis, M., Albers, H.F.F., van Jeveren, J.G.G., de Jong, J.W., Hautvast, J.G.A.J., Hermus, R.J.J., Katan, M.B., Brydon, W.G. and Easwood, M.A. (1980). Influence of dietary fiber from vegetables and fruits, bran or citrus pectin on serum lipids, fecal lipids and colonic function. *The American Journal of Clinical Nutrition*, 33, 1745-1756.

- Stephen, A.M. and Churms, S.C. (2006). Introduction. In Stephen, A.M. and Phillips, G.O. (2nd eds.), *Food Polysaccharides and Their Applications* (pp. 1-17). Boca Raton: CRC Press.
- Sudhakar, D.V. and Maini, S.B. (1999). Isolation and characterization of mango peel pectins. *Journal of Food Processing Preservation*, 24, 209-227.
- Talmadge, K.W., Keegstra, K., Bauer, W.D. and Albersheim, P. (1973). Structure of plant cell walls. I. The macromolecular components of the walls of suspension-cultured sycamore cells with a detailed analysis of the pectic polysaccharides. *Plant Physiology*, 51, 158-173.
- Tang, P.Y., Wong, C.J., and Woo, K.K. (2011). Optimization of pectin extraction from peel of dragon fruit (*Hylocereus polyrhizus*). *Asian Journal of Biological Sciences*, 4(2), 189-195.
- Thakur, B.R., Singh, R.K. and Handa, A.K. (1997). Chemistry and uses of pectin – A review. *Critical Reviews in Food Science and Nutrition*, 37(1), 47-73.
- Tulyathan, V., Tananuwong, K., Songjind, P. And Jaiboon, N. (2002). Some physicochemical properties of jackfruit (*Artocarpus heterophyllus* Lam) seed flour and starch. *Science Asia*, 28, 37-41.
- Uddin, M.T., Rukanuzzaman, M., Khan, M.M.R. and Islam, M.A. (2009). Jackfruit (*Artocarpus heterophyllus*) leaf powder: An effective adsorbent for removal of methylene blue from aqueous solution. *Indian Journal of Chemical Technology*, 16, 142-149.
- USDA national nutrient database for standard reference, release 23. Nutrient Data Laboratory homepage. Retrieved 31 August 2014 from <http://www.ars.usda.gov/ba/bhnrc/ndl>
- Virk, B.S. and Sogi, D.S. (2004). Extraction and characterization of pectin from apple (*Malus Pumila*. Cv Amri) peel waste. *International Journal of Food Properties*, 7(3), 693-703.
- Voragen, A.G.J., Pilnik, W., Thibault, J.-F., Axelos, M.A.V. and Renard, C.M.G.C. (1995). Pectins. In A.M. Stephen (Ed.), *Food polysaccharides and their applications*. New York: Marcel Dekker: 287-339.
- Voragen, A.G.J., Coenen, G.-J., Verhoef, R.P. and Schols, H.A. (2009). Pectin, a versatile polysaccharide present in plant cell walls. *Structural Chemistry*, 20, 263-275.
- Vriesmann, L. C., Teófilo, R. F., and de Oliveira Petkowicz, C. L. (2011a). Optimization of nitric acid-mediated extraction of pectin from cacao pod husks (*Theobroma cacao* L.) using response surface methodology. *Carbohydrate Polymers*, 84, 1230–1236.

- Vriesmann, L. C., de Mello Castanho Amboni, R. D., and de Oliveira Petkowicz, C. L. (2011b). Cacao pod husks (*Theobroma cacao* L.): Composition and hot-water soluble pectins. *Industrial Crops and Products*, 34, 1173–1181.
- Vriesmann, L. C., Teófilo, R. F., and de Oliveira Petkowicz, C. L. (2012). Extraction and characterization of pectin from cacao pod husks (*Theobroma cacao* L.) with citric acid. *Food Science and Technology*, 49, 108–116.
- Vriesmann, L. C. and Petkowicz, C. L. O. (2013). Highly acetylated pectin from cocoa pod husks (*Theobroma cacao* L.) forms gel. *Food Hydrocolloids*, 33, 58–65.
- Walkinshaw, M.D. and Arnott, S. (1981). Conformations and interactions of pectin II. Models for junction zones in pectinic acid and calcium pectate gels. *Journal of Molecular Biology*, 53, 1075–1085.
- Wang, S., Chen, F., Wu, J., Wang, Z., Liao, X., and Hu, X. (2007). Optimization of pectin extraction assisted by microwave from apple pomace using response surface methodology. *Journal of Food Engineering*, 78(2), 693–700.
- Wang, W., Ma, X., Xu, Y., Cao, Y., Jiang, Z., Ding, T., Ye, X. and Liu, D. (2015). Ultrasound-assisted heating extraction of pectin from grapefruit peel: Optimization and comparison with the conventional method. *Food Chemistry*, 178, 106–114.
- Wang, M., Huang, B., Fan, C., Zhao, K., Hu, H., Xu, X., Pan, S. and Liu, F. (2016). Characterization and functional properties of mango peel pectin extracted by ultrasound assisted citric acid. *International Journal of Biological Macromolecules*, 91, 794–803.
- Wei, B.L., Weng, J.R., Chiu, P.H., Hung, C.F., Wang, J.P. and Lin, C.N. (2005). Antiinflammatory flavonoids from *Artocarpus heterophyllus* and *Artocarpus communis*. *Journal of Agricultural and Food Chemistry*, 53, 3867 – 3871.
- Willats, W.G.T., Knox, J.P. and Mikkelsen, J.D. (2006). Pectin: new insights into an old polymer are starting to gel. *Trends in Food Science and Technology*, 17, 97–104.
- Wolf, S., Mouille, G. and Pelloux, J. (2009). Homogalacturonan methyl-esterification and plant development. *Molecular Plant*, 2, 851–860.
- Wong, K.C., Lim, C.L. and Wong, L.L. (1992). Volatile flavour constituents of Cempedak (*Artocarpus polyphema* Pers.) fruit and jackfruit (*Artocarpus heterophyllus* Lam.) from Malaysia. *Flavour and Fragrance Journal*, 9, 319 – 324.
- Wong, W.W., Alkarkhi, A.F.M. and Mat Easa, A. (2010). Effect of extraction conditions on yield and degree of esterification of durian rind pectin: An experimental design. *Food and Bioproducts Processing*, 88, 209–214.

- Wuestenberg, T. (2014). General Overview of Food Hydrocolloids. In Wuestenberg, T. (1st ed.), *Cellulose and Cellulose Derivatives in the Food Industry: Fundamentals and Applications*, (pp. 1-68). Wiley-VCH Verlag GmbH & Co. KGaA.
- Yapo, B.M., and Koffi, K.L. (2006). Yellow passion fruit rind – A potential source of low-methoxyl pectin. *Journal of Agricultural and Food Chemistry*, 54, 2738-2744.
- Yapo, B.M., Robert, C., Etienne, I., Wathelet, B. and Paquot, M. (2007a). Effect of extraction conditions on the yield, purity and surface properties of sugar beet pulp pectin extracts. *Food Chemistry*, 100, 1356-1364.
- Yapo, B.M. (2009a). Biochemical characteristics and gelling capacity of pectin from yellow passion fruit rind as affected by acid extractant nature. *Journal of Agricultural and Food Chemistry*, 57(4), 1572-1578.
- Yapo, B.M. (2009b). Lemon juice improves the extractability and quality characteristics of pectin from yellow passion fruit by-product as compared with commercial citric acid extractant. *Bioresource Technology*, 100, 3147-3151.
- Yapo, B.M. (2009c). Pineapple and banana pectins comprise fewer homogalacturonan building blocks with a smaller degree of polymerization as compared with yellow passion fruit and lemon pectins: Implication for gelling properties. *Biomacromolecules*, 10(4), 717-721.
- Yapo, B.M. (2011). Pectic substances: From simple pectic polysaccharides to complex pectins – A new hypothetical model. *Carbohydrate Polymers*, 86, 373-385.
- Yeoh, S., Shi, J. and Langrish, T.A.G. (2008). Comparisons between different techniques for water-based extraction of pectin from orange peels. *Desalination*, 218, 229-237.
- Zouambia, Y., Ettoumi, K.Y., Krea, M. and Moulai-Mostefa, N. (2014). A new approach for pectin extraction: Electromagnetic induction heating. *Arabian Journal of Chemistry*, <http://dx.doi.org/10.1016/j.arabjc.2014.11.011>
- Zykwinska, A. W., Ralet, M. C. J., Garnier, C. D. and Thibault, J. F. (2005). Evidence for *in vitro* binding of pectin side chains to cellulose. *Plant Physiology*, 139, 397-407.
- Zykwinska, A., Boiffard, M.-H.I.n., Kontkanen, H., Buchert, J., Thibault, J.-F. and Bonnin, E. (2008). Extraction of green labeled pectins and pectic oligosaccharides from plant byproducts. *Journal of Agricultural and Food Chemistry*. 56, 8926-8935.

BIODATA OF STUDENT

The student of this thesis, Leong Chia Ming, is a Malaysian born in Kuala Lumpur on the 14th December 1986. He completed his Bachelor (Hons.) in Food Science and Nutrition at UCSI University late 2009. The author then enrolled for a Master in Science programme in Faculty of Food Science and Technology, Universiti Putra Malaysia on September 2011. Prior to this, the author worked as a chemist in a private contract manufacturing plant for 2 years.



PUBLICATION

Leong, C.M., Noranizan, M.A., Kharidah, M. and Choo, W.S. (2016). Physicochemical properties of pectin extracted from jackfruit and chempedak fruit rinds using various acids. *International Food Research Journal*, 23(3), 973-978.





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT:

EXTRACTION AND CHARACTERIZATION OF CRUDE PECTIN FROM JACKFRUIT

(*Artocarpus Heterophyllus* Lam.) AND CEMPEDAK (*Artocarpus Integer* Spreng.) FRUIT RIND

NAME OF STUDENT: LEONG CHIA MING

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

☐

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

☐

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

☐

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

☐

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]