



**UNIVERSITI PUTRA MALAYSIA**

***IDENTIFICATION OF FUSARIUM AND ASPERGILLUS SPECIES FROM  
CORNMEAL IN MALAYSIA AND TOXIGENICITY OF MYCOTOXINS  
PRODUCED***

**NITHIYAA A/P PERUMAL**

**FS 2012 103**

**IDENTIFICATION OF *FUSARIUM* AND *ASPERGILLUS* SPECIES FROM  
CORNMEAL IN MALAYSIA AND TOXIGENICITY OF MYCOTOXINS PRODUCED**

**NITHIYAA A/P PERUMAL**

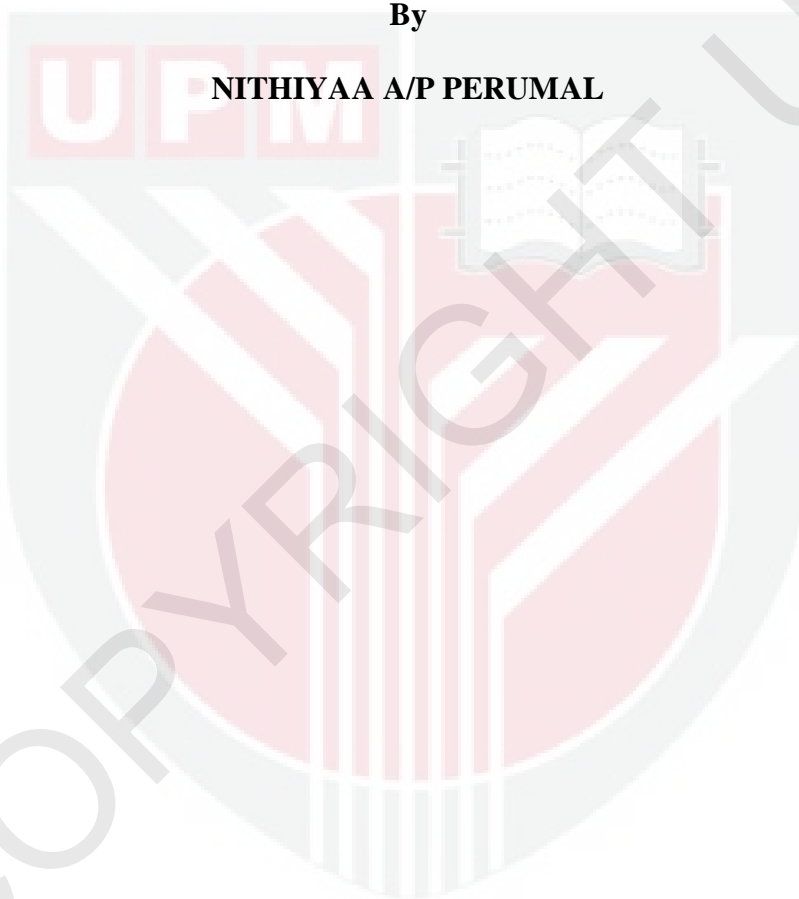
**MASTER OF SCIENCE  
UNIVERSITI PUTRA MALAYSIA**

**2012**

**IDENTIFICATION OF *FUSARIUM* AND *ASPERGILLUS* SPECIES FROM  
CORNMEAL IN MALAYSIA AND TOXIGENICITY OF MYCOTOXINS PRODUCED**

**By**

**NITHIYAA A/P PERUMAL**



**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia in  
Fulfilment of the Requirements for the Degree of Master of Science**

**November 2012**

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**IDENTIFICATION OF *FUSARIUM* AND *ASPERGILLUS* SPECIES FROM CORNMEAL IN MALAYSIA AND TOXIGENICITY OF MYCOTOXINS PRODUCED**

By

**NITHIYAA A/P PERUMAL**

**November 2012**

**Chairman: Nur Ain Izzati Mohd Zainudin, PhD**

**Faculty: Science**

Corn is a vital food source for human consumption, animal feed as well as industrial processing. However, corn faces repeated spoilage and contamination by a huge range of fungi especially by *Fusarium* and *Aspergillus* species. These fungi are known of producing mycotoxins such as fumonisins (FBs), moniliformin (MON), zearalenone (ZEA), beauvericin (BEA) and aflatoxins (AFs). Therefore the main objectives of this study were to identify and determine the diversity of *Fusarium* and *Aspergillus* species as well as to quantify the mycotoxins produced by both fungi associated with cornmeal in Malaysia. In this study, cornmeal samples were obtained from 9 states throughout Malaysia and the cornmeal samples were surface sterilized and cultured onto Peptone Pentachloronitrobenzene Agar (PPA) to isolate the fungi. Single spore isolation was carried out onto Potato Dextrose Agar (PDA) to obtain pure culture. *Fusarium* and *Aspergillus* isolates were selected and preceded with morphological identification. The diversity of *Fusarium* and *Aspergillus* species were determined using Shannon-Weiner

index and this is followed by the extraction of mycotoxins. The extracted mycotoxins were analyzed qualitatively using Thin Layer Chromatography (TLC) and *A. salina* bioassay, and quantitatively using Ultra-fast Performance Liquid Chromatography (UFLC). A total of 314 isolates of microfungi were obtained, 90.5% isolates belonged to *Aspergillus* species, namely *A. flavus* (76.8%), *A. niger* (7.6%), *A. nidulans* (4.5%) and *A. fumigatus* (1.6%). Another 9.5% isolates were *Fusarium* species, identified as *F. verticillioides* (4.5%), *F. semitectum* (3.2%) and *F. proliferatum* (1.9%). As for the mycotoxin analysis, 15 out of 16 *Fusarium* isolates produced MON, 12 isolates produced BEA and all 16 isolates produced FB<sub>1</sub>. However none produced ZEA. In addition, 29 out of 40 *Aspergillus* isolates produced AFB<sub>1</sub> and only two isolates produced AFB<sub>2</sub>. The analysis of *A. salina* revealed that all the five mycotoxins extracts were toxic to the brine shrimp despite the concentration of the mycotoxins. As a conclusion, a proper storage system for the corns should be implemented to avoid fungal contamination hence reducing the accumulation of mycotoxins.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGenalPASTIAN SPESIS *FUSARIUM* DAN *ASPERGILLUS* DARI  
JAGUNG BERPROSES DI MALAYSIA DAN UJIAN KETOKSIGENAN  
MIKOTOKSIN YANG DIHASILKAN**

Oleh

**NITHIYAA A/P PERUMAL**

**November 2012**

**Pengerusi: Nur Ain Izzati Mohd Zainudin, PhD**

**Fakulti: Sains**

Jagung merupakan salah satu sumber makanan utama bagi manusia, haiwan serta untuk pemprosesan industri. Namun demikian, sumber jagung berulang kali menghadapi kerosakan dan kontaminasi oleh pelbagai jenis kulat terutamanya spesis *Fusarium* dan *Aspergillus*. Kulat-kulat ini boleh menghasilkan mikotoksin seperti fumonisins (FBs), moniliformin (MON), zearalenone (ZEA), beauvericin (BEA) dan aflatoxins (AFs). Oleh itu, objektif utama kajian ini adalah untuk mengenalpasti dan menentukan kepelbagaian spesis *Fusarium* dan *Aspergillus* serta mengkuantifikasi mikotoksin yang dihasilkan oleh kedua-dua kulat yang berasosiasi dengan jagung berproses di Malaysia. Dalam kajian ini, sampel jagung berproses diperolehi daripada 9 negeri di sekitar Malaysia and sampel jagung tersebut disterilasi sebelum dikultur ke atas agar Peptone Pentachloronitrobenzene (PPA) untuk pengasingan kulat. Teknik pengasingan spora tunggal dilakukan di atas Potato dextrose agar (PDA) untuk mendapatkan kultur kulat

yang tulen. Spesies *Fusarium* dan *Aspergillus* dipilih and seterusnya diidentifikasi secara morfologi. Diversiti spesies *Fusarium* dan *Aspergillus* ditentukan menggunakan Shannon-Weiner indeks dan langkah ini disusuli oleh pengekstrakan mikotoksin. Mikotoksin yang diekstrak telah dianalisis secara kualitatif menggunakan *Thin Layer Chromatography* (TLC) dan *A. salina* bioassay dan secara kuantitatif menggunakan *Ultra-fast Performance Liquid Chromatography* (UFLC). Sejumlah 314 isolat kulat mikro diperolehi, di mana 90.5% isolat terdiri daripada spesies *Aspergillus*, iaitu *A. flavus* (76.8%), *A. niger* (7.6%), *A. nidulans* (4.5%) dan *A. fumigatus* (1.6%). Selebihnya, 9.5% isolate adalah spesies *Fusarium* yang dikenalpasti sebagai daripada *F. verticillioides* (4.5%), *F. semitectum* (3.2%) and *F. proliferatum* (1.9%). Dari segi analisis mikotoksin, 15 daripada 16 isolat *Fusarium* telah menghasilkan MON, 12 isolat menghasilkan BEA dan kesemua 16 isolat didapati menghasilkan FB<sub>1</sub>. Walaubagaimanapun tiada isolat yang menghasilkan ZEA. Selain itu, 29 daripada 40 isolat *Aspergillus* didapati menghasilkan AFB<sub>1</sub>, manakala hanya dua isolat menghasilkan AFB<sub>2</sub>. Analisis *A. salina* menunjukkan bahawa kesemua lima ekstrak mikotoksin adalah toksik kepada udang air masin tanpa mengira kepekatan mikotoksin tersebut. Sebagai kesimpulan, sistem penyimpanan jagung yang baik perlu dilaksanakan untuk mengelakkan kontaminasi kulat, sekaligus mengurangkan pengumpulan mikotoksin.

## ACKNOWLEDGEMENT

I take this opportunity to express my gratitude to my supervisor, Dr. Nur Ain Izzati Mohd Zainudin for her continuous guidance and help throughout this study. She's a great inspiration to me. I appreciate her precious time and effort in constantly checking my progress without fail. I also expend my gratitude to Prof. Dr. Umi Kalsom, who is my co-supervisor. She has always kept me motivated and she was ever concern about my progress.

I would also like to thank many others who have helped me directly and indirectly throughout my study. I would like to thank the academic staffs of Biology, the supporting staffs of Biology and other personnel who had lend me a hand when I needed them the most.

I thank my friends Banulata Gopalsamy, Suhaida Salleh, Nur Azlin Azhari, Sardiana Sarmidi, Sujithra Devi and many others who have been a great help as well as moral support.

Special thanks also to UPM and Ministry of Higher Education (MOHE) for providing me sponsorships through Graduate Research Fellowship (GRF) and Mini Bajet 2009 throughout the research. Last but never the least, I thank my beloved family for their undying patience and support towards me. I thank them for their love, motivation and constant support in helping me throughout my study.

I thank you all once again.



I certify that a Thesis Examination Committee has met on 23 November 2012 to conduct the final examination of Nithiyaa a/p Perumal on her thesis entitled “Detection and Toxigenicity of Mycotoxin Produced by *Fusarium* And *Aspergillus* species Isolated from Cornmeal in Malaysia” in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Hishamuddin bin Omar, PhD**

Lecturer

Faculty of Science

Universiti Putra Malaysia

(Chairman)

**Kamaruzzaman bin Sijam**

Associate Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Internal Examiner)

**Khairulmazmi bin Ahmad**

Senior Lecturer

Faculty of Agriculture and Food Sciences

Universiti Putra Malaysia Bintulu Campus

(Internal Examiner)

**Latifah binti Zakaria**

Associate Professor

School of Biological Sciences

Universiti Sains Malaysia

(External Examiner)

---

**SEOW HENG FONG, PhD**

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 16 January 2013

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Nur Ain Izzati Mohd Zainudin, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Putra Malaysia  
(Chairman)

**Umi Kalsom Yusof**

Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

---

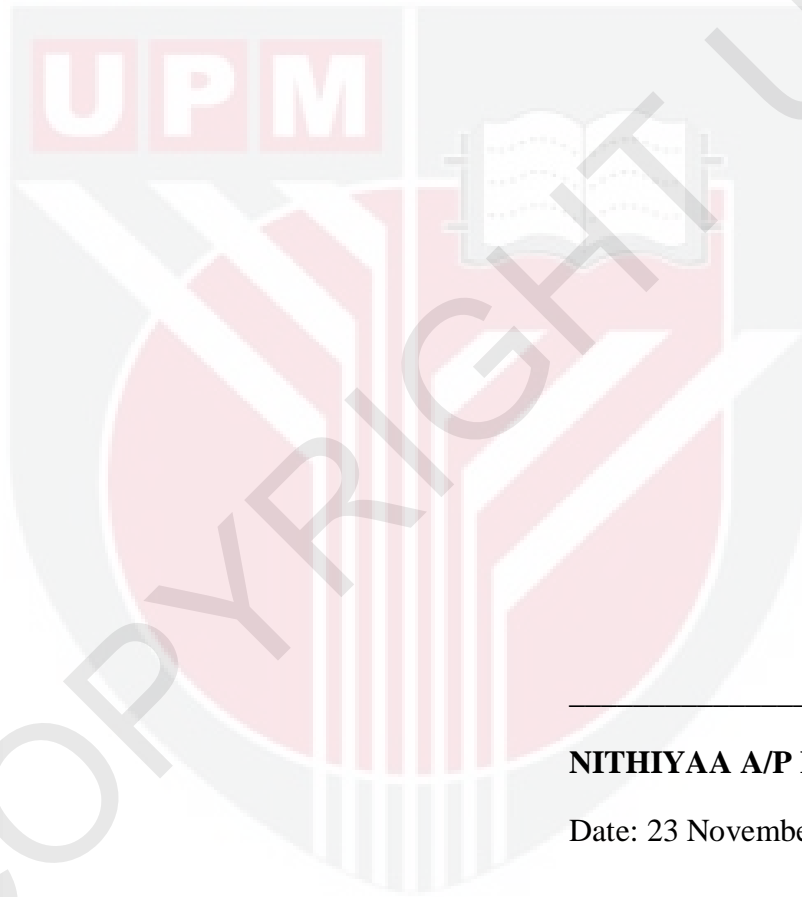
**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## **DECLARATION**

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



---

**NITHIYAA A/P PERUMAL**

Date: 23 November 2012

## TABLE OF CONTENT

<b>ABSTRACT</b>	<b>Page</b>
<b>ABSTRAK</b>	II
<b>ACKNOWLEDGEMENT</b>	IV
<b>APPROVAL</b>	VI
<b>DECLARATION</b>	VII
<b>TABLE OF CONTENT</b>	IX
<b>LIST OF TABLES</b>	X
<b>LIST OF FIGURES</b>	XII
<b>LIST OF ABBREVIATIONS</b>	XIII
<b>CHAPTER</b>	XIV
<b>1 INTRODUCTION</b>	1
<b>2 LITERATURE REVIEW</b>	6
2.1 Importance of Corn	6
2.1.1 Fungi associated with corn	7
2.1.2 Management of postharvest corn	8
2.2 Emergence and Classification of Fungi	9
2.2.1 Diversity of <i>Fusarium</i> species	10
2.2.2 Diversity of <i>Aspergillus</i> species	12
2.3 Occurance and Toxicity of Mycotoxins	13
2.3.1 Fumonisin (FBs)	15
2.3.2 Moniliformin (MON)	17
2.3.3 Zearalenone (ZEA)	18
2.3.4 Beauvericin (BEA)	19
2.3.5 Aflatoxins (AFs)	20
<b>3 DIVERSITY AND MORPHOLOGICAL IDENTIFICATION OF <i>FUSARIUM</i> SPECIES AND <i>ASPERGILLUS</i> SPECIES FROM CORNMEAL</b>	22
3.1 Introduction	22
3.2 Materials and Methods	24
3.2.1 Sample collection	24
3.2.2 Isolation of fungi and single spore isolation	26
3.2.3 Macromorphology of <i>Fusarium</i> and <i>Aspergillus</i> species	26
3.2.4 Micromorphology of <i>Fusarium</i> and <i>Aspergillus</i> species	27
3.2.5 Diversity of <i>Fusarium</i> and <i>Aspergillus</i> species	28
3.3 Results and Discussion	28
3.3.1 Diversity of <i>Fusarium</i> and <i>Aspergillus</i> species	28
3.3.2 Morphological characteristics of <i>Fusarium</i> and	36

	<i>Aspergillus</i> species	
3.4	Conclusion	51
4	<b>DETECTION AND QUANTIFICATION OF MYCOTOXIN PRODUCED BY <i>FUSARIUM</i> AND <i>ASPERGILLUS</i> SPECIES</b>	52
4.1	Introduction	52
4.2	Materials and Methods	53
4.2.1	Conidial suspension	53
4.2.2	Inoculation of spores and sample extraction	54
4.2.3	Analysis of mycotoxins using Thin Layer Chromatography (TLC)	56
4.2.4	Analysis of mycotoxins using Ultra Fast Liquid Chromatography (UFLC)	58
4.2.5	<i>Artemia salina</i> bioassay	61
4.3	Results and Discussion	61
4.3.1	Production of mycotoxins by <i>Fusarium</i> species	61
4.3.2	Production of mycotoxins by <i>Aspergillus</i> species	68
4.3.3	Toxicogenicity test using <i>Artemia salina</i> bioassay	73
4.4	Conclusion	77
5	<b>SUMMARY, CONCLUSION AND RECOMMENDATION</b>	78
	<b>REFERENCES</b>	80
	<b>APPENDIX 1</b>	90
	<b>APPENDIX 2</b>	92
	<b>APPENDIX 3</b>	123
	<b>APPENDIX 4</b>	126
	<b>BIODATA OF STUDENT</b>	129

## LIST OF TABLES

Table	Page
3.1 List of <i>Fusarium</i> and <i>Aspergillus</i> isolates and their respective locations	29
3.2 The prevalence of <i>Fusarium</i> and <i>Aspergillus</i> species isolated from corn meal samples from different states	32
3.3 Diversity of <i>Aspergillus</i> and <i>Fusarium</i> species in each sampling state of origin	33
3.4 Diversity of <i>Fusarium</i> and <i>Aspergillus</i> species based on Shannon-Weiner Index	34
4.1 The solvent system, $R_f$ values, and the color observed under UV light for all five mycotoxins	62
4.2 Detection of mycotoxins from three <i>Fusarium</i> species using TLC plates	63
4.3 Concentrations of mycotoxins produced by each isolate of <i>Fusarium</i> species	65
4.4 TLC analysis results for AF B <sub>1</sub> and B <sub>2</sub>	69
4.5 Concentrations of AFs produced by the respective isolates of <i>Aspergillus</i> species	71
4.6 The mortality rate of <i>A. salina</i> tested with the mycotoxins produced by <i>Fusarium</i> isolates	74
4.7 The mortality rate of <i>A. salina</i> tested with the AFs produced by <i>Aspergillus</i> isolates	75

## LIST OF FIGURES

Figure	Page
2.1 Molecular structure of FB <sub>1</sub>	16
2.2 Molecular structure of MON	18
2.3 Molecular structure of ZEA	18
2.4 Molecular structure of BEA	20
2.5 Molecular structure of AF B1 (A), and AF B2 (B)	21
3.1 Sampling locations	25
3.2 Macroscopic and microscopic characteristics of <i>F. verticillioides</i>	37
3.3 Macroscopic and microscopic characteristics of <i>F. semitectum</i>	39
3.4 Macroscopic and microscopic characteristics of <i>F. proliferatum</i>	41
3.5 Macroscopic and microscopic characteristics of <i>A. flavus</i>	43
3.6 Macroscopic and microscopic characteristics of <i>A. niger</i>	45
3.7 Macroscopic and microscopic characteristics of <i>A. nidulans</i>	47
3.8 Macroscopic and microscopic characteristics of <i>A. fumigatus</i>	49

## LIST OF ABBREVIATIONS

R <sub>f</sub>	Retention factor
FBs	Fumonisin
FB <sub>1</sub>	Fumonisin B <sub>1</sub>
DON	Deoxynivalenol
BEA	Beauvericin
MON	Moniliformin
ZEA	Zearalenone
AFs	Aflatoxins
AFB <sub>1</sub>	Aflatoxin B <sub>1</sub>
AFB <sub>2</sub>	Aflatoxin B <sub>1</sub>
AFG <sub>1</sub>	Aflatoxin G <sub>1</sub>
AFG <sub>2</sub>	Aflatoxin G <sub>1</sub>
FA	Fusaric acid
FUS	Fusaproliferin
OTA	Ochratoxin
DNA	Deoxyribonucleic acid
RNA	Ribonucleic acid
TLC	Thin Layer Chromatography
ELISA	Enzyme-Linked Immunosorbent Assay
GC	Gas Chromatography
GC-MS	Gas Chromatography-Mass Spectrometry



CZE	Capillary Zone Electrophoresis
HPLC	High Performance Liquid Chromatography
UPLC–MS	Ultra-High Performance Liquid Chromatography–Mass Spectrometry
UFLC	Ultra-fast Performance Liquid Chromatography
LC	Liquid Chromatography
NaOCl	Sodium hypochlorite
PPA	Peptone Pentachloronitrobenzene Agar
PDA	Potato Dextrose Agar
CLA	Carnation Leaf Agar
SNA	Spezieller Nährstoffarmer Agar
TFA	Trifluoroacetic acid
UV	Ultra-violet
i.d	Internal diameter
M	Molar
OPA	<i>o</i> -phthaldialdehyde
ANOVA	Analysis of variance
SE	Standard error

## **CHAPTER 1**

### **INTRODUCTION**

Corn plantation was introduced during 15th century by the Indian community in the highlands of Mexico (Wertz, 2005). In 19th century, the hybrid corns were developed by several scientists hence produced more varieties of corn and better yields. Therefore, the farmers started planting the hybrid corn in a large scale. The number of farmers that used hybrid corn increased throughout the 20th century. According to National Corn Growers Association, the yields of corn increased every year, for example in 1991 the average yield was 108.6 bushel per acre and in 2008 increased to 153.9 bushel per acre (USDA, 2009).

Corn serves as a staple food and an important part of the diet for millions of people worldwide. It contains high starch content and also good for feeding hogs, horses and other livestock such as chicken, cattle and swine (Fitting, 2006). Despite its vast importance, the farmers are facing problem to maintain the quality of corn. The major crisis faced by many corn farmers is fungal contamination. Corn plantations are basically susceptible to fungal infection during both pre-harvest and post-harvest periods. While in field it is colonized by numerous fungi, some of the fungi basically influence the quality of the grain during the pre-harvest period, particularly affecting the quality of the corn and causes ear rot disease. Fungi also infect the root, stem and leaf and therefore contribute to low quality corn. Contaminations during pre-harvest may

persist until post-harvest such as during storage and processing, transporting and marketing (Etcheverry *et al.*, 1999).

The fungal infection on corn continues till storage and the diversity of fungi inhabiting the corn changes. This is due to the alteration of the storage conditions, which involves the humidity and temperature in the particular area. The corn surface will also damage, caused by insect or animal and may enhance the filamentous fungi to invade the damaged corn (Etcheverry *et al.*, 1999).

Several microscopic fungi which are associated with corn and reduce the quality of the yield are such as species of *Aspergillus*, *Chaetomium*, *Cladosporium*, *Eurotium*, *Fusarium*, *Penicillium* and *Pythium* (Chelkowski, 1991). However, in the present study, most attentions are paid to *Aspergillus* species and *Fusarium* species as these fungi may persist from the field to storage.

*Fusarium* is a filamentous fungi widely distributed in soil and often acted as pathogen of plant, human and animal. Most species are saprophytes and relatively abundant members of the soil microbial community. Several species of *Fusarium* are important pathogens of plants, including corn and other cereals, causing root, stem and ear rot (Uhlir *et al.*, 2007). The fungi that affected corn are capable to produce harmful primary and secondary metabolites. Therefore secondary metabolites which are known as mycotoxin can cause hazardous illness and sometimes fatal to the consumers such as animals and humans (Uhlir *et al.*, 2007).

Approximately, a total 20 *Fusarium* species have been regularly associated with diseases occurring in small-grain cereals. *F. culmorum*, *F. graminearum* Schwabe (teleomorph: *Gibberella zeae*) and *F. avenaceum*, (teleomorph: *G. avenacea* Cook) were most frequently isolated (Miedaner, 1997). *Fusarium* species have the ability to produce mycotoxins that causes health problem. The common mycotoxins produced by *Fusarium* species are beauvericin (BEA), deoxynivalenol (DON), fumonisins (FBs), fusaric acid (FA), fusaproliferin (FUS), fusarins and moniliformin (MON) (Fotso *et al.*, 2002; Logrieco *et al.*, 2002; Muthomi *et al.*, 2008). *Aspergillus* species are also harmful to both humans and animals, causing severe diseases. More than 185 species have been identified and 20 species are recognized as the pathogenic species. *Aspergillus* species are diverse, where it can be found in soil, wood, stored grains, plant remains, and mostly in the air inhaled by all living creatures. The several *Aspergillus* species found in stored grains are *A. flavus*, *A. parasiticus* and *A. niger* (Giorni *et al.*, 2007).

Mycotoxins are secondary metabolites produced by fungi which does not participate in the growth and reproduction of the fungi, conversely effects the biochemical, physiological or pathological changes in other living organisms, such as other microorganism, plants, animals and human (Haschek *et al.*, 2002). These compounds are known to produce a large range of effects in organisms such as mammals (rats, guinea pigs and etc), birds (chicken embryos), amphibians, arthropods, crustaceans, unicellular organisms, microorganisms and plants. Zearalenones (ZEA) produced by *Fusarium* species, is a nonsteroidal estrogenic mycotoxin that causes estrogenic syndrome such as enlargement of mammary glands and genital organs, and atrophy of testes in the affected animals. FBs cause leukoencephalomalacia, a brain lesion that is

fatal to horses and also carcinogenic. MON causes muscle weakness, respiratory distress and cyanosis in animals (Desjardins, 2006a). BEA is a cyclic lactone trimer, a specific cholesterol acyltransferase inhibitor besides and toxic to several human cell lines and is able to induce apoptosis and DNA fragmentation (Reynoso *et al.*, 2004).

*Aspergillus* species are also able to produce various types of mycotoxins such as, aflatoxin B1 (AFB1), B2 (AFB2), G1 (AFG1) and G2 (AFG2), ochratoxin (OTA) and patulin (Giorni *et al.*, 2007; Morgavi *et al.*, 2003; Shundo *et al.*, 2009). AFs are toxin metabolites that functions as a hepatotoxic, hepatocarcinogenic, and also has mutagenic effects on animal and human (Almeida *et al.*, 1996). OTA however is known to exhibit nephrotoxin as well as hepatotoxic and carcinogenic to humans, similar to AFs (Shundo *et al.*, 2009). According to Morgavi (2003), patulin also displays toxic effects where it acts as teratogenic, carcinogenic and mutagenic to other organisms.

Hence, these negative effects have been made into use as bioassays to detect the presence of mycotoxins in vast commodities (Panigrahi, 1993). *A. salina* larvae was selected as a screening system owing to the privilege that their eggs are commercially available, besides being able to hatch within 24 hours of incubation with minimal maintenance of the cultures (Jimenez *et al.*, 1997). Previously, Matthews (1995) also stated the use of brine shrimp, stating that these organisms are convenient, simple and less expensive. Besides, the sensitivity of these organisms is utilized greatly to quick scan the occurrence of the desires metabolites. Moreover, the use of higher animals are greatly restricted since only a small number of these animals are allowed for toxicity test which in this study accounts for the mortality rate of the organisms tested. As a result,

only a handful of isolates can be tested. Hence, once again the use of brine shrimp was widely considered.

On the other hand, cornmeal contaminated with fungal have invited the attention of many farm breeders and scientists' worldwide. Farm animals that feed on the infected cornmeal are severely affected and causing death, hence causing serious lost to the farm breeders (Hazel and Patel, 2004). This problem proceeds when human consumes the infected animal or food and therefore risking their health. As one problem leads to another, this may eventually end up affecting the whole food chain in an ecosystem.

The objectives of this study were:

- i) To identify and determine the diversity of *Fusarium* and *Aspergillus* species associated with cornmeal in Malaysia,
- iii) To identify and quantify the mycotoxins produced by *Fusarium* and *Aspergillus* species associated with cornmeal in Malaysia.

## REFERENCES

- Ahmad, Y., Hameed, A. & Ghaffar, A. (2006). Enzymatic activity of fungal pathogens in corn. *Pakistan Journal of Botany*. 38(4): 1305-1316.
- Akiyama, H., Uraroongroj, M., Miyahara, M., Goda, Y. & Toyoda, M. (1998). Quantitation of fumonisins in corn by HPLC with *o*-phthalaldehyde postcolumn derivatization and their identification by LC/MS. *Mycopathologia*. 140: 157-161.
- Almeida, R.M.A., Correa, B., Xavier, J.G., Mallozzi, M.A.B., Gambale, W. & Paula, C.R. (1996). Acute effect of aflatoxin B<sub>1</sub> on different inbred mouse strains II. *Mycopathologia*. 133: 23-29.
- Bayman, P. (2007). Fungal Endophytes. *Environmental and Microbial Relationships*. 2: 213-227.
- Bottalico, A., Visconti, A., Logrieco, A., Solfrizzo, M. & Mirocha, C.J. (1985). Occurrence of zearalenols (Diastereomeric mixture) in corn stalk rot and their production by associated *Fusarium* species. *Applied and Environmental Microbiology*. 54: 547-551.
- Bridge, P. & Spooner, B. (2001). Soil fungi: diversity and detection. *Plant and Soil*. 232: 147-154.
- Brown, R.L., Chen, Z.Y., Menkir, A., Cleveland, T.E., Cardwell, K., Kling, J. & White, D.G. (2001). Resistance to aflatoxin accumulation in kernels of maize inbreds selected for ear rot resistance in West and Central Africa. *Journal of Food Protection*. 64(3): 396-400.
- Burmeister, H.R., Ciegler, A. & Vesonder, R. F. (1979). Moniliformin, a metabolite of *Fusarium moniliforme* NRRL 6322: Purification and toxicity. *Applied and Environmental Microbiology*. 37: 11-13.
- Burgess, L.W., Summerell, B.A., Bullock, S., Gott, K.P. & Backhouse, D. (1994). *Laboratory Manual for Fusarium Research*. University of Sydney.
- Butron, A., Revilla, P., Sandoya, G., Ordas, A. & Malvar, R.A. (2009). Resistance to reduce corn borer damage in maize for bread, in Spain. *Crop Protection*. 28: 134-138.
- Castella, G., Bragulat, M.R. & Cabafies, F.J. (1996). Mycoflora and fumonisin-producing strains of *Fusarium moniliforme* in mixed poultry feeds and component raw material. *Mycopathologia*. 133: 181-184.
- Chang, D.C.N. & Chou, L.C. (2007). Growth responses, enzyme activities, and component changes as influenced by *Rhizoctonia* orchid mycorrhiza on *Anoectochilus formosanus* Hayata. *Botanical Studies*. 48: 445-451.



- Chelkowski, J. (1991). *Cereal Grain: Mycotoxins, Fungi and Quality in Drying and Storage*. Netherlands: Elsevier Science Publishing.
- Chulze, S.N., Ramirez, M.L., Pascale, M. & Visconti, A. (1998). Fumonisin production by, and mating populations of *Fusarium* section *Liseola* isolates from maize in Argentina. *Mycology Research*. 102: 141-144.
- Clauw, M.K., Ayouni, F., Tardieu, D. & Guerre, P. (2008). Variations in zearalenone activation in avian food species. *Food and Chemical Toxicology*. 46: 1467-1473.
- Cleveland, T.E., Yu, J., Fedorova, N., Bhatnagar, D., Payne, G.A., Nierman, W.C., & Bennett, J.W. (2009). Potential of *Aspergillus flavus* genomics for applications in biotechnology. *Trends in Biotechnology*. 27: 151-157.
- Cortes, G., Carvajal, M., Ramirez, M., Avila-Gonzalez, E., Chilpa-Galvan, N., Castillo- Urueta, P. & Flores, C.M. (2010). Processing, Products, and Food Safety: Identification and quantification of aflatoxins and aflatoxicol from poultry feed and their recovery in poultry litter. *Poultry Science*. 89: 993-1001.
- Crawford, D.L., Lynch, J.M., Whipps, J.M. & Ousley, M.A. (1993). Isolation and Characterization of Actinomycete antagonists of a fungal root pathogen. *Applied Environmental Microbiology*. 59(11):3899-3905.
- Desjardins, A.E., Maragos, C.M., & Proctor, R.H. (2006a). Maize Ear Rot and moniliformin contamination by cryptic species of *Fusarium subglutinans*. *Journal of Agriculture and Food Chemistry*. 54: 7383-7390.
- Desjardins, A.E. (2006b). *Fusarium Mycotoxins: Chemistry, Genetics and Biology*. United States of America: The American Phytopathological Society.
- Diba, K., Kordbacheh, P., Mirhendi, S.H., Rezaie, S. & Mahmoudi, M. (2007). Identification of *Aspergillus* sp. using morphological characteristics. *Pakistan Journal of Medical Science*. 23: 867-872.
- Dikbas, N., Kotan, R., Dadasoglu, F. & Sahin, F. (2008). Control of *Aspergillus flavus* with essential oil and methanol extract of *Satureja hortensis*. *International Journal of Food Microbiology*. 124: 179-182.
- Etcheverry, M., Nesci, A., Barros, G., Torres, A. & Chulze, S. (1999). Occurrence of *Aspergillus* section *Flavi* and aflatoxin B1 in corn genotypes and corn meal in Argentina. *Mycopathologia*. 147: 37-41.
- Ezekiel, C.N., Odebode, A.C. & Fapohunda, S.O. (2008). Zearalenone production by naturally occurring *Fusarium* species on maize, wheat and soybeans from Nigeria. *Journal of Biology Environmental Science*. 2(6): 77-82.



- Fadl Allah, E.M. (1998). Occurrence and toxigenicity of *Fusarium moniliforme* from freshly harvested maize ears with special references to fumonisin production in Egypt. *Mycopathologia*. 140: 99-103.
- Fallah, A.A. (2010). Aflatoxin M1 contamination in dairy products marketed in Iran during winter and summer. *Food Control*. 21: 1478-1481.
- Fitting, E. (2006). Importing corn, exporting labor: The neoliberal corn regime, GMOs, and the erosion of Mexican biodiversity. *Agriculture and Human Values*. 23: 15-26.
- Fotso, J., Leslie, J.F. & Smith, J.S. (2002). Production of beauvericin, moniliformin, fusaproliferin, and fumonisins B<sub>1</sub>, B<sub>2</sub>, and B<sub>3</sub> by fifteen ex-type strains of *Fusarium* species. *Applied and Environmental Microbiology*. 68: 5195-5197.
- Fried, B. & Sherma, J. (1994). Thin-Layer Chromatography: Techniques and applications. *Qualitative Evaluation and Documentation* (pp. 157). New York: Marcel Dekker.
- Gerlach, W. & Nirenberg, H. I. (1982). *The genus Fusarium: A pictorial atlas*. Germany: Biologische Bundesanstalt für Land- und Forstwirtschaft.
- Ghali, R., Hmaissia-khlifa, K., Ghorbel, H., Maaroufi, K. & Hedili, A. (2008). Incidence of aflatoxins, ochratoxin A and zearalenone in tunisian foods. *Food Control*. 19: 921-924.
- Ghali, R., Belouaer, I., Hdiri, S., Ghorbel, H., Maaroufi, K. & Hedilli, A. (2009). Simultaneous HPLC determination of aflatoxins B<sub>1</sub>, B<sub>2</sub>, G<sub>1</sub> and G<sub>2</sub> in Tunisian sorghum and pistachios. *Journal of Food Composition and Analysis*. 22: 751-755.
- Ghiasian, S.A., Rezayat, S.M., Bacheh, P.K., Maghsood, A.H., Yazdanpanah, H., Shephard, G.S., Westhuizen, L.V.D., Vismer, H.F. & Marasas, W.F.O. (2005). Fumonisin production by *Fusarium* species isolated from freshly harvested corn in Iran. *Mycopathologia*. 159: 31-40.
- Giri, B., Giang, P.H., Kumari, R., Prasad, R. & Varma, A. (2005). Microbial Diversity in Soils. *Soil Biology*. 3: 21-55.
- Giorni, P., Magan, N., Pietri, A., Bertuzzi, T. & Battilani, P. (2007). Studies on *Aspergillus* section *Flavi* isolated from maize in northern Italy. *International Journal of Food Microbiology*. 113: 330-338.
- Glenn, A.E. (2007). Mycotoxigenic *Fusarium* species in animal feed. *Animal Feed Science and Technology*. 137: 213-240.
- Gottwald, S., Germeier, C.U. & Ruhmann, W. (2000). Computerized image analysis in *Fusarium* taxonomy. *Mycological Research*. 105: 206-214.

- Hartl, A. & Stenzel, W.R. (2007). Development of a method for the determination of citrinin in barley, rye and wheat by solid phase extraction on aminopropyl columns and HPLC-FLD. *Mycotoxin Research*. 23: 127-131.
- Haschek, W.M., Kenneth, A.V. & Beasley, V.R. (2002). Selected mycotoxins affecting animal and human health. *Handbook of Toxicologic Pathology*.1: 645-699.
- Hazel, C.M. & Patel, S. (2004). Influence of processing on trichothecene levels. *Toxicology Letters*. 153: 51-59.
- Hlywka, J.J., Beck, M.M. & Bullerman, L.B. (1997). The use of the chicken embryo screening test and Brine shrimp (*Artemia salina*) Bioassays to assess the toxicity of fumonisin B1 mycotoxin. *Food and Chemical Toxicology*. 35: 991-999.
- Jeney, A., Beki, E., Mule, G. & Hornok, L. (2004). Identification of growth stage specific transcript profiles in *Fusarium proliferatum* (*Gibberella fujikuroi*, mating population D) by cDNA-AFLP analysis. *European Journal of Plant Pathology*. 110: 619-625.
- Jestoi, M. (2008). Emerging *Fusarium*-mycotoxins fusaproliferin, beauvericin, enniatins, and moniliformin-A Review. *Critical Reviews in Food Science and Nutrition*. 48:21-49.
- Jimenez M., Huertam T. & Mateo, R. (1997). Mycotoxin production by *Fusarium* species isolated from bananas. *Applied and Environmental Microbiology*. 63(2): 364-369.
- Josephs, R. D., Krska, R., Schuhmacher, R. & Grasserbauer, M. (1999). A rapid method for the determination of the *Fusarium* mycotoxin beauvericin in maize. *Fresenius Journal of Analytical Chemistry*. 363:130-131.
- Khayoon, W.S., Saad, B., Chew B.Y., Hashim, N.H., Mohamed, A.A.S., Salleh, M.I. & Salleh, B. (2010). Determination of aflatoxins in animal feeds by HPLC with multifunctional column clean-up. *Food Chemistry*. 118: 882-886.
- Kostecki, M., Grabarkiewicz-Szczesna, J. & Golinski, P. (1997). Simultaneous analysis of beauvericin and moniliformin in fungal cultures and in cereal grain samples. *Mycotoxin Research*. 13: 17-22.
- Krska, R., Schuhmacher, R., Grasserbauer, M. & Scott, R.M. (1996). Determination of the *Fusarium* mycotoxin beauvericin at µg/kg levels in corn by high-performance liquid chromatography with diode-array detection. *Journal of Chromatography*. 746: 233-238.
- Krska, R. (1998). Performance of modern sample preparation techniques in the analysis of *Fusarium* mycotoxins in cereals. *Journal of Chromatography Analysis*. 815: 49-57.
- Kurtzman, M.A.D. (2003). Fumonisin and beauvericin induce apoptosis in turkey peripheral blood lymphocytes. *Mycopathologia*.156: 357-364.

- Labuda, R., Parich, A., Berthiller, F. & Tancinova, D. (2005). Incidence of trichothecenes and zearalenone in poultry feed mixtures from Slovakia. *International Journal of Food Microbiology*. 105: 19-25.
- Langseth, W., Ghebremeskel, M., Kosiak, B., Kolsaker, P. & Miller, D. (2000). Production of culmorin compounds and other secondary metabolites by *Fusarium culmorum* and *F. graminearum* strains isolated from Norwegian cereals. *Mycopathologia*. 152: 23-34.
- Lappalainen, S., Nikulin, T. M., Berg, S., Parikka, P., Hintikka, I. E.L. & Pasanen, A.L. (1996). *Fusarium* toxins and fungi associated with handling of grain on eight finnish farms. *Atmospheric Environment*. 30(17): 305-3065.
- Latiffah, Z., Mohd Zariman M & Baharuddin, S. (2007). Diversity of *Fusarium* species in cultivated soils in Penang. *Malaysian Journal of Microbiology*. 3(1): 27-30.
- Leong, Y.H., Ismail, N., Latif, A.A. & Ahmad, R. (2010). Aflatoxin occurrence in nuts and commercial nutty products in Malaysia. *Food Control*. 21: 334-338.
- Leslie, J.F. & Summerell, B.A. (2005). The *Fusarium* Laboratory Manual (pp. 121-274). *South Africa: Medical Research Council*.
- Li, J.X., Yuan, S.L., Li, G.Z. & Jian, Y.W. (2009). Enhanced beauvericin production with in situ adsorption in mycelial liquid culture of *Fusarium redolens* Dzf2. *Process Biochemistry*. 44: 1063-1067.
- Lino, C.M., Silva, L.J.G., Pena, A.L.S. & Silveira, M.I. (2006). Determination of fumonisins B1 and B2 in Portuguese maize and maize-based samples by HPLC with fluorescence detection. *Analytical and Bioanalytical Chemistry*. 384: 1214-1220.
- Logrieco, A., Moretti, A., Fornelli, F., Fogliano, V., Ritieni, A., Caiaffa, M.F., Randazzo, G., Bottalico, A. & Macchia, L. (1996). Fusaproliferin production by *Fusarium subglutinans* and its toxicity to *Artemia salina*, SF-9 insect cells, and IARC/LCL 171 Human B Lymphocytes. *Applied and Environmental Microbiology*. 62(9): 3378-3384.
- Logrieco, A., Mule, G., Moretti, A. & Bottalico, A. (2002). Toxigenic *Fusarium* species and mycotoxins associated with maize ear rot in Europe. *European Journal of Plant Pathology*. 108: 597-609.
- Logrieco, A., Moretti, A., Perrone, G. & Mule, G. (2007). Biodiversity of complexes of mycotoxigenic fungal species associated with *Fusarium* ear rot of maize and *Aspergillus* rot of grape. *International Journal of Food Microbiology*. 119:11-16.
- Lorenzi L.D., Giovanni, A.D., Malagutti, L., Molteni, L., Sciaraffia, F., Tamburini, A. & Zannotti, M. (2005). Genotoxic activity of the fumonisin B1 mycotoxin in cultures of bovine lymphocytes. *Italian Journal of Animal Science*. 4: 395-402.

- Ma, Z., Huang, B., Zhang, J., Zhang, Y., Zhu, L. & Tu, Q. (2009). Time-resolved fluoroimmunoassay of zearalenone in cereals with a europium chelate as label. *Journal of Rare Earths*. 27(6): 1088-1091.
- Magan, N., Hope, R., Cairns, V. & Aldred, D. (2003). Post-harvest fungal ecology: Impact of fungal growth and mycotoxin accumulation in stored grain. *European Journal of Plant Pathology*. 109: 723-730.
- Maiorano, A., Reyneri, A., Sacco, D., Magni, A. & Ramponi, C. (2008). A dynamic risk assessment model (FUMAgain) of fumonisin synthesis by *Fusarium verticillioides* in maize grain in Italy. *Crop Protection*. 1-14.
- Maragos, C.M. & Busman, M. (2010). Rapid and advanced tools for mycotoxin analysis: a review. *Food Additives and Contaminants*. 27(5): 688-700.
- Marasas, W.F.O., Nelson, P.E. & Toussoun, T.A. (1984). *Toxigenic Fusarium species identity and mycotoxicology*. (pp. 93-263). Pennsylvania: The Pennsylvania State University.
- Matthews, R.S. (1995). *Artemia salina* as a test organism for measuring superoxide-mediated toxicity. *Free Radical Biology & Medicine*. 18(5): 919-922.
- McNeill, S. & Montross, M. (2003). Corn harvesting, handling, drying, and storage. *Cooperative Extension Service*. 52-58.
- Miedaner, T. (1997). Breeding wheat and rye for resistance to *Fusarium* diseases. *Plant Breeding*. 116: 201-220.
- Monsanto Technology Development. (2011). Agronomic spotlight: Benefits of a timely corn Harvest. <http://ebookbrowse.com/agronomic-spotlight-benefits-of-a-timely-corn-harvest-pdf-d369658722>
- More, T.T., Yan, S., Tyagi, R.D. & Surampalli, R.Y. (2010). Potential use of filamentous fungi for wastewater sludge treatment. *Bioresource Technology*. 101: 7691-7700.
- Morgavi, D.P., Boudra, H., Jouany, J.P. & Graviou, D. (2003). Prevention of patulin toxicity on rumen microbial fermentation by SH-containing reducing agents. *Journal of Agriculture and Food Chemistry*. 51: 6906-6910.
- Mubatanhema, W., Moss, M.O., Frank, M.J. & Wilson, D.M. (1999). Prevalence of *Fusarium* species of the *Liseola* section on Zimbabwean corn and their ability to produce the mycotoxins zearalenone, moniliformin and fumonisin B<sub>1</sub>. *Mycopathologia*. 148: 157-163.
- Mueller, G.M. & Schmit, J.P. (2007). Fungal biodiversity: what do we know? What can we predict? *Biodiversity and Conservation*. 16: 1-5.

- Munkvold, G., Stahr, H.M., Logrieco, A., Moretti, A. & Ritieni, A. (1998). Occurrence of fusaproliferin and beauvericin in *Fusarium*- Contaminated livestock feed in Iowa. *Applied and Environmental Microbiology*. 64(10): 3923-3926.
- Munkvold, G.P. (2003). Epidemiology of *Fusarium* diseases and their mycotoxins in maize ears. *European Journal of Plant Pathology*. 109: 705-713.
- Muthomi, J.W., Ndungu, J.K., Gathumbi, J.K., Mutitu, E.W. & Wagacha, J.M. (2008). The occurrence of *Fusarium* species and mycotoxins in Kenyan wheat. *Crop Protection*. 27: 1215-1219.
- Nelson, P. E., Toussoun, T.A. & Marasas, W.F.O. (1983). *Fusarium Species: An Illustrated Manual for Identification*. Pennsylvania: Pennsylvania State University Press.
- Nelson, P.E. (1993). Fumonisin, mycotoxins produced By *Fusarium* species: Biology, chemistry, and significance. *Annual Review Phytopathology*. 31: 33-52.
- Nur Ain Izzati, M.Z., Azmi, A.R. & Salleh, B. (2008). Secondary metabolite profiles and mating populations of *Fusarium* species in section *Liseola* associated with bakanae disease of rice. *Malaysian Journal of Microbiology*. 4(1): 6-13.
- Nuryono, N., Noviandi, C.T., Bohm, J. & Fazeli, E.R. (2005). A limited survey of zearalenone in Indonesian maize-based food and feed by ELISA and high performance liquid chromatography. *Food Control*. 16: 65-71.
- Panigrahi, S. (1993). *Review Section*: Bioassay of mycotoxins using terrestrial and aquatic, animal and plant species. *Food Chemistry and Toxicology*. 31(10): 767-790.
- Papavizas, G.C. (1967). Evaluation of various media and antimicrobial agents for isolation of *Fusarium* from soil. *Phytopathology*. 57: 848-852.
- Park, K.J. (2001). Importance of corn. In *Corn production in Asia: China, Indonesia, Thailand, Philippines, Taiwan, North Korea, South Korea, and Japan* (pp. 1-13). Taiwan: Food & Fertilizer Technology Center.
- Pascale, M., Visconti, A. & Chelkowski, J. (2002). Ear rot susceptibility and mycotoxin contamination of maize hybrids inoculated with *Fusarium* species under field conditions. *European Journal of Plant Pathology*. 108: 645-651.
- Peterson, S.W., Varga, J., Frisvad, J.C. & Samson, R.A. (2008). *Phylogeny and subgeneric taxonomy of Aspergillus* (pp. 33-56). Netherlands: Wageningen Academic Publishers.
- Plattner, R.D., Ross, P.F., Reagor, J., Stedelin, J. & Rice, L.G. (1991). Analysis of corn and cultured corn for fumonisin B<sub>1</sub> by HPLC and GC/MS by four laboratories. *Journal of Veterinary Diagnosis and Investigation*. 3: 357-358.



- Presello, D.A., Botta, G., Iglesias, J. & Eyherabide, G.H. (2008). Effect of disease severity on yield and grain fumonisin concentration of maize hybrids inoculated with *Fusarium verticillioides*. *Crop Protection*. 27: 572-576.
- Raper, K.B. & Fennell, D.I. (1965). *The Genus Aspergillus*. (pp. 492-493). Baltimore: Williams & Wilkins Company.
- Reddy, K.R.N., Reddy, C.S. & Muralidharan, K. (2009a). Detection of *Aspergillus* spp. and aflatoxin B1 in rice in India. *Food Microbiology*. 26: 27-31.
- Reddy, K.R.N., Reddy, C.S. & Muralidharan, K. (2009b). Potential of botanicals and biocontrol agents on growth and aflatoxin production by *Aspergillus flavus* infecting rice grains. *Food Control*. 20: 173-178.
- Reid, L.M., Zhu, X. & Ma, B.L. (2001). Crop rotation and nitrogen effects on maize susceptibility to gibberella (*Fusarium graminearum*) ear rot. *Plant and Soil*. 237: 1-14.
- Reynoso, M.M., Torres, A.M. & Chulze, S.N. (2004). Fusaproliferin, beauvericin and fumonisin production by different mating populations among the *Gibberella fujikuroi* complex isolated from maize. *Mycological Research*. 108: 154-160.
- Rodrigues, P., Soares, C., Kozakiewicz, Z., Paterson, R.R.M., Lima, N. & Venancio, A. (2007). Identification and characterization of *Aspergillus flavus* and aflatoxins. *Communicating Current Research and Educational Topics and Trends in Applied Microbiology*.
- Ruhl, G. (2007). Crop diseases in corn, soybean, and wheat. <http://www.btny.purdue.edu/Extension/Pathology/CropDiseases/Corn/>
- Rychlik, M. & Asam, S. (2008). Stable isotope dilution assays in mycotoxin analysis. *Analysis of Bioanalytical Chemistry*. 390: 617-628.
- Salleh, B. & Sulaiman, B. (1984). *Fusarium* associated with naturally diseased plants in Penang. *Journal of Plant Protection in the Tropical*. 1: 47-53.
- Sampietro, D.A., Vattuone, M.A., Presello, D.A., Fauguel, C.M. & Catalan, C.A.N. (2009). The pericarp and its surface wax layer in maize kernels as resistance factors to fumonisin accumulation by *Fusarium verticillioides*. *Crop Protection*. 28: 196-200.
- Samson R.A. (1979). A compilation of the *Aspergilli* described since 1965. *Studies in Mycology*. 18:1-38.
- Samson, R.A., Hong, S.B. & Frisvad, J.C. (2006). Old and new concepts of species differentiation in *Aspergillus*. *Medical Mycology September*. 44: 133-148.
- Sanchez, G.J.J., Goodman, M.M. & Stuber, C.W. (2000). Isozymatic and morphological diversity in the races of Maize of Mexico. *Economic Botany*. 54: 43-59.

- Sandhu, K.S., Singh, N. & Malhi, N.S. (2007). Some properties of corn grains and their flours I: Physicochemical, functional and chapati-making properties of flours. *Food Chemistry*. 101: 938-946.
- Santini, A., Ferracane, R., Meca, G. & Ritieni, A. (2009). Overview of analytical methods for beauvericin and fusaproliferin in food matrices. *Analytical and Bioanalytical Chemistry*. 395:1253-1260.
- Schollenberger, M., Muller, H.M., Ruffle, M., Suchy, S., Plank, S. & Drochner, W. (2006). Natural occurrence of 16 *Fusarium* toxins in grains and feedstuffs of plant origin from Germany. *Mycopathologia*. 161: 43-52.
- Seifert, K. (1996). *Fusarium* Interactive Key. *Agriculture and Agri-Food Canada*. 1-65.
- Sewram, V., Nieuwoudt, T.W., Marasas, W.F.O., Shephard, G.S. & Ritieni, A. (1999a). Determination of the *Fusarium* mycotoxins, fusaproliferin and beauvericin by high-performance liquid chromatography electrospray ionization mass spectrometry. *Journal of Chromatography Agriculture*. 858: 175-185.
- Sewram, V., Nieuwoudt, T.W., Marasas, W.F.O., Shephard, G.S. & Ritieni, A. (1999b). Determination of the mycotoxin moniliformin in cultures of *Fusarium subglutinans* and in naturally contaminated maize by high-performance liquid chromatography-atmospheric pressure chemical ionization mass spectrometry. *Journal of Chromatography Agriculture*. 848: 185-191.
- Shephard, G.S. (1998). Chromatographic determination of the fumonisin mycotoxins. *Journal of Chromatography*. 815: 31-39.
- Shundo, L., Almeida, A.P.D., Alaburda, J., Lamardo, L.C.A., Navas, S.A., Ruvieri, V. & Sabino, M. (2009). Aflatoxins and ochratoxin A in Brazilian paprika. *Food Control*. 20: 1099-1102.
- Soriano, J.M. & Dragacci, S. (2004). Occurrence of fumonisins in foods. *Food Research International*. 37: 985-1000.
- Spellerberg, I.F. (2008). Shannon-Weiner index. *Encyclopedia of Ecology* 3249-3252.
- Summerell, B.A., Salleh, B. & Leslie, J.F. (2003). A Utilitarian Approach to *Fusarium* Identification. *Plant Disease*. 87(2): 117-128.
- Sylvia, V.L., Phillips, T.D., Clement, B.A. & Green, J.L. (1986). Determination of deoxynivalenol (vomitoxin) by High-Performance Liquid Chromatography with electrochemical detection. *Journal of Chromatography*. 362: 79-85.
- Sweeney, M.J. & Dobson, A.D.W. (1998). Mycotoxin production by *Aspergillus*, *Fusarium* and *Penicillium* species. *International Journal of Food Microbiology*. 43: 141-158.

- Torres, A.M., Reynoso, M.M., Rojo, F.G., Ramirez, M.L. & Chulze, S.N. (2001). *Fusarium* species (section *Liseola*) and its mycotoxins in maize harvested in northern Argentina. *Food Additives and Contaminants*. 18(9): 836-843.
- Tseng, T.C., Lee, K.L., Deng, T.S., Liu, C.Y. & Huang, J.W. (1995). Production of fumonisins by *Fusarium* species of Taiwan. *Mycopathologia*. 130:117-121.
- Tuan, N.A., Manning, B.B., Lovell, R.T. & Rottinghaus, G.E. (2003). Responses of Nile tilapia (*Oreochromis niloticus*) fed diets containing different concentrations of moniliformin or fumonisin B1. *Aquaculture*. 217: 515-528.
- Turnera, N.W., Subrahmanyam, S. & Piletsky, S.A. (2009). Analytical methods for determination of mycotoxins: A review. *Analytica Chimica Acta*. 632: 168-180.
- United States Department of Agriculture (USDA). (2009). Corn: Market Outlook. <http://www.ers.usda.gov/Briefing/Corn/2009baseline.htm>.
- Uhlig, S., Torp, M., Jarp, J., Parich, A., Gutleb, A. C. & Krska, R. (2004). Moniliformin in Norwegian grain. *Food Additives and Contaminants*. 21: 598-606.
- Uhlig, S., Jestoi, M. & Parikka, P. (2007). *Fusarium avenaceum* - The North European situation. *International Journal of Food Microbiology*. 119: 17-24.
- Vilela, G.R., Almeida, G.S., D'Arce, M.A.B.R., Moraes, M.H.D., Brito, J.O., Silva, M.F.G.F., Silva, S.C., Piedade, S.M.S., Domingues, M.A.C., & Gloria, E.M. (2009). Activity of essential oil and its major compound, 1,8-cineole, from *Eucalyptus globulus* Labill., against the storage fungi *Aspergillus flavus* Link and *Aspergillus parasiticus* Speare. *Journal of Stored Products Research*. 45: 108-111.
- Wang, H., Wang, T., Pometto III, A.L. & Johnson, L. A. (2009). A laboratory decanting procedure to simulate whole stillage separation in dry-grind corn ethanol process. *Journal of the American Oil Chemistry's Society*. 86:1241-1250.
- Wertz, S.K. (2005). Maize: The Native North American's Legacy of Cultural Diversity and Biodiversity. *Journal of Agricultural and Environmental Ethics*. 18: 131-156.
- Younis, Y.M.H. & Malik, K.M. (2003). TLC and HPLC assays of aflatoxin contamination in Sudanese peanuts and peanuts products. *Kuwait Journal of Science and Engineering*. 30(1): 79-94.
- Yu, J., Proctor, R.H., Brown, D.W., Abe, K., Gomi, K., Machida, M., Hasegawa, F., Nierman, W.C., Bhatnagar, D. & Cleveland, T.E. (2004). Genomics of economically significant *Aspergillus* and *Fusarium* species. *Applied Mycology & Biotechnology*. 4: 249-283.