



UNIVERSITI PUTRA MALAYSIA

***TOXICITY AND THERAPEUTIC EFFECTS OF ZERUMBONE EXTRACT
ON COMPLETE FREUND'S ADJUVANT-INDUCED RHEUMATOID
ARTHRITIS RAT MODEL***

MOHAMAD FAUZI MOHD IDRIS

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RAT MODEL**

By

MOHAMAD FAUZI BIN MOHD IDRIS

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
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October 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of
the requirement for the degree of Master of Science

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The present study was conducted to investigate the antinociceptive and anti-arthritis effects of zerumbone in Complete Freund's adjuvant induced Rheumatoid Arthritis model in rats. Zerumbone is a sesquiterpene isolated and purified from the rhizomes of *Zingiber zerumbet* plants, locally known as 'lempoyang'. *Zingiber zerumbet* is one of the most important species of cultivated ginger, used in food flavouring. Zerumbone was obtained from the rhizomes through a sequence of isolation processes involving hydro-distillation, rotary evaporation of excessive solvents, column chromatography, and finally followed by recrystallization. The compound was subjected to thin layer chromatography (TLC), gas chromatography mass spectrometry (GC-MS) and high performance liquid chromatography (HPLC) to identify its chemical structure and purity. Zerumbone was optimally isolated using the hexane and ethyl acetate solvent system at a ratio of 8:2 with a total yield of 1.2%. In the present study, it was shown that intraperitoneal administration of zerumbone at doses of 10 mg/kg did not show any signs of toxicity in terms of behavioural changes, body weight, liver and kidneys, hematological and liver function parameters. The findings were further supported by histopathological observations of the liver and kidney that demonstrated normal histological architecture. Rheumatoid arthritis was induced by intraplantar injection of CFA on the right hind paw of each rat at day 0. Anti-inflammatory activity of zerumbone on the RA-induced rats was evaluated using plethysmometer test and paw swelling was assessed by measuring the thickness of the hind paw with a digital caliper. The determination of the antinociceptive profile of zerumbone on rheumatoid arthritis-induced model in rats was studied through thermal and mechanical threshold which consists of the Hargreaves plantar test, Von Frey test, and Randall-Selitto analgesiometer

with slight modifications. During the experimental period, the body weight was recorded using a digital weighing balance every 3 days consecutively after CFA injection. With slight modifications, the visual arthritis scoring was used to assess behavioural changes through the ability to stance, mobility and flexion pain test. The clinical assessment of arthritis had been studied through cytokine concentrations and hematological parameters. As a conclusion, the study strongly confirms the antinociceptive and anti-inflammatory activities of zerumbone as well as elucidated the possible mechanism of action through which it exerts its effects. In addition to that, the toxicity studies demonstrated the safety margin of zerumbone in mice, thus scientifically justifying the traditional use of this species of ginger and setting the path for future studies.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Sarjana Sains

**KESAN KETOKSIKAN DAN TERAPEUTIK ESTRAK ZERUMBONE PADA
MODEL TIKUS TERARUH ARTRITIS REUMATOID**

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Kajian ini telah dijalankan untuk menentukan kesan antinosiseptif dan anti-arthritis zerumbone dalam *Complete Freund's adjuvant*(CFA) yang menyebabkan artritis reumatoid (RA) dalam model haiwan. Zerumbone merupakan kompaun seskuiterpena yang dipisahkan dan ditulenkkan dari rizom tumbuhan *Zingiber zerumbet*, atau lebih dikenali sebagai lempoyang. *Zingiber zerumbet* adalah salah satu spesis halia yang banyak ditanam dan digunakan untuk peningkatan rasa makanan serta sering digunakan dalam perubatan tradisional untuk mengubat sakit perut, sakit gigi, keseliuhan otot dan juga untuk rawatan lebam dan luka-luka ringan. Zerumbone diperolehi dari rizom melalui proses penyulingan-hidro diikuti oleh proses pengewapan rotari untuk mengeluarkan pelarut dan dipisahkan serta ditulenkkan melalui proses kromatografi dan penghabluran semula. Sebatian zerumbone kemudiannya dianalisis menggunakan kromatografi lapisan nipis (TLC), kromatografi gas-spektroskopi jisim (GC-MS) dan kromatografi cecair prestasi tinggi (HPLC) bagi menentukan tahap ketulenan. Zerumbone dipisahkan secara optimum menggunakan sistem pelarut hexane dan etil asetat pada nisbah 8:2 dengan kadar hasil sebanyak 1.245%. Analisis ketoksiikan menunjukkan bahawa pemberian zerumbone pada dos 10 mg/kg secara intraperitoneal tidak menunjukkan sebarang kesan toksik di mana tiada kesan sampingan dalam kelakuan haiwan, tiada perubahan berat badan, hepar dan ginjal serta tiada perubahan yang signifikan dalam bacaan hematologi dan fungsi hepar. Kesan ketoksiikan ini disokong melalui kajian histopatologi di mana histologi hepar dan ginjal berada dalam keadaan normal. Induksi artritis reumatoid dilakukan melalui suntikan CFA secara intraplantar pada tapak kaki belakang kanan tikus pada hari 0. Aktiviti anti-inflamasi zerumbone keatas model RA pada tikus dinilai dengan menggunakan ujian plethysmometer bagi menentukan isipadu tapak kaki dan ketebalan tapak kaki diukur

menggunakan angkup digital. Penentuan kesan antinosiseptif zerumbone dikaji melalui ambang terma dan mekanikal yang terdiri daripada ujian plantar Hargreaves, ujian von Frey dan ujian Randall-Selitto analgesiometer bagi menilai ambang terma kesakitan. Sepanjang tempoh eksperimen, berat badan tikus direkodkan menggunakan alat penimbang berat digital setiap 3 hari berturut-turut. Skala skor artritis secara visual dilaksanakan bagi mengukur perubahan tingkah laku melalui keupayaan untuk berdiri, bergerak dan ujian kesakitan fleksi seperti yang dinyatakan sebelum ini dengan sedikit pengubahsuaian. Penilaian klinikal artritis telah dikaji melalui analisis kepekatan sitokin dan parameter hematologi. Analisis eksperimen yang telah digunakan adalah analisis dua hala varians (ANOVA) diikuti dengan ujian post-hoc Bonferroni, dimana $p \leq 0.05$ telah diterima sebagai signifikan. Secara kesimpulannya, kajian ini mengesahkan aktiviti antinosiseptif dan anti-inflamasi zerumbone dan juga menjelaskan mekanisme yang berkebarangkalian terlibat. Tambahan pula, tahap keselamatan penggunaan zerumbone pada tikus melalui kajian ketoksinan turut menjustifikasi penggunaan tradisional spesis halia ini secara saintifik dan membuka ruang untuk kajian pada masa hadapan.

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This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee are as follows:

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LIST OF ABBREVIATIONS

AFM	Arthritis Foundation Malaysia
ALP	Alkaline Phosphatase
ALT	Alanine Aminotransferase
ANOVA	Analysis of Variance
AST	Aspartate Aminotransferase
CAM	Complementary alternative medicine
CAMs	Cell adhesion molecules
CFA	Complete Freund's adjuvant
CNS	Central nervous system
COX-2	Cyclooxygenase-2
DIP	Distal interphalangeal
DMARDs	Disease-modifying anti-rheumatic drugs
DMSO	Dimethyl Sulfoxide
GC-MS	Gas chromatography-mass spectrometry
Hb	Hemoglobin
HPLC	High performance liquid chromatography
Ht	Hematocrit
i.p.	Intraperitoneal
IASP	The International Association for the Study of Pain
IBS	Institute of Bioscience
IL-1	Interleukin 1
IL-10	Interleukin 10
IL-6	Interleukin 6
MCH	Mean corpuscular hemoglobin
MCHC	Mean corpuscular hemoglobin concentration
MCP	Metacarpophalangeal
MCV	Mean corpuscular volume
MMPs	Matrix metalloproteinases
MTP	Metatarsalphalangeal
NK	Natural killer

NSAIDs	Non-steroidal anti-inflammatory drugs
PGE2	Prostaglandin E2
PID	Post-inoculation day
PIP	Proximal interphalangeal
RA	Rheumatoid Arthritis
RBC	Red Blood Cell
TLC	Thin layer chromatography
TNF- α	Tumor necrosis factor alpha
tR	Retention time
VEGF	Vascular endothelial growth factor
WBC	White Blood Cell

CHAPTER 1

INTRODUCTION

Rheumatoid arthritis (RA) is a chronic inflammatory disorder that typically affects the small joints in hands and feet. Unlike the wear-and-tear damage of osteoarthritis, RA affects the lining of joints, causing painful swellings that eventually result in bone erosion and joint deformity. It is a long-term autoimmune disorder which has been found as one of the most common chronic inflammatory diseases worldwide. RA occurs when the body immune system mistakenly attacks and eventually destroys the healthy joints and tissues. The etiology of this disease still remains unknown until today.

The onset of disease can occur at any age, but the peak incidence occurs within the fourth and fifth decades of life. Apparently, females are 2.5 times more likely to be affected than males (Tehlirian & Bathon, 2008). It is known to affect approximately 0.5% to 1% of the adult population. In Malaysia, it affects about five in 1000 people (AFM, 2009-2010).

In the early stage of RA, pain can be felt when there are some minor movements. However, spontaneous pain during rest can occur later. The primary focus of the inflammation is in the synovium, which is the lining tissue of the joint. Inflammatory chemicals released by the immune cells may cause swelling and damage to cartilage and bone. The diagnosis of RA is based on the symptoms described and physical examination findings such as warmth, swelling, pain in the joints, and presence of rheumatoid nodules (Alyce, Oliver, & William, 2008).

Currently, treatments for RA focus on controlling symptoms and preventing joint damage. Among the common medications are disease-modifying anti-rheumatic drugs (DMARDs), non-steroidal anti-inflammatory drugs (NSAIDs) as well as corticosteroid. Usually, the combinations of different drugs are used to achieve the desired effects (Marsland & Kapoor, 2004). Medications are often prescribed to slow down or stop joint damage, relieve pain, and decrease inflammation.

Undisputedly, conventional drug treatments are available to reduce the inflammation completely or put on halt the destruction of the joints. The accompanying side effects caused by the steroids, however, result in gastrointestinal injuries like peptic ulcer and gastrointestinal bleeding in both healthy and RA patients (Marsland & Kapoor, 2004). This can lead to the discontinuation of the treatments by the patients. Indeed, it is crucial to develop new classes of medication which are equally or more effective than the existing medicines, specifically without side effects and able to cure the disease.

Throughout medicinal evolution, the importance of natural products for medicine and health has been enormous. The history of modern medicine could be traced back to centuries ago; the earliest known written document is a 4000-year old Sumerian clay tablet that records remedies for various illnesses (Kong, Goh, Chia, & Chia, 2003). Similar to the Chinese ancient civilisation, Indians and North Africans too provide written evidence for the use of natural products in curing various diseases (Phillipson, 2001). For instance, turmeric possesses blood clotting properties while raw garlic was prescribed for circulatory disorders and mandrake for pain relief. Some of these natural sources are still being used in several countries as alternative medicines.

However, it was not until the 19th century that scientists isolated active components from natural ingredients that were known to have beneficial effects. The sources of these ingredients had been studied in depth and comprehensively. In the past decades, a variety of active ingredients was being extracted through purification processes of natural sources. They were recorded and classified according to their structure and mechanism of action (McCurdy & Scully, 2005).

Despite centuries of folkloric usage, numerous successes and even well-documented treatment of diseases, many scholars have often dismissed the effectiveness of natural products (Riddle, 1985) in medicine. Sadly, in today's society, those who control the economy of drugs and therapeutic agents prefer to showcase the superiority of modern medicine and portray herbal medicine as well as traditional healing as unscientific.

The use of ginger is so widespread that its usage can be found in various traditional systems of medicine in the world. The cultivation of ginger could be traced back to centuries ago at the University of Maryland Medical Centre where ginger was written to be used in China for over 2,000 years (Steven, 2010) and as medicine from the Vedic period in India, called "*Maha aushadhi*" meaning the great medicine (Palatty, Haniadka, Valder, Arora, & Baliga, 2013). In Sanskrit, ginger is known as Sringavera. It is speculated that this term may have given way to Zingiberi in Greek and then to Latin Zingiber (Vasala, 2004). The botanic name of ginger is *Zingiber officinale* Rosc. which belongs to the family of Zingiberaceae.

Zingiberaceae species are among the most prolific plants in tropical rainforest. There are about 160 species from 18 genera found in Peninsular Malaysia, mostly growing in damps, lowlands, and hill slopes as scattered plants or thickets (Larsen, Ibrahim, & Wong, 1999). Collectively, in terms of its widespread traditional use and availability, there are myriads of reasons for scientifically evaluating the use of ginger to fully explore its potentials.

1.1 Problem statement

The unwanted side effects of common treatment have contributed negatively to the management of inflammation as well as pain in rheumatoid arthritis patients. Therefore, the search for safer antinociceptive and anti-inflammatory agents for treatment of RA is regarded necessary.

1.2 Hypothesis

1. Zerumbone extract has no significant difference compared to control group.
2. Zerumbone extract has positive therapeutic effects in CFA induced rheumatoid arthritis rat model.

1.3 Objectives

1.3.1 General Objectives

1. To investigate the toxicity effects of zerumbone extract.
2. To determine zerumbone extract therapeutic effects in CFA induced rheumatoid arthritis rat model.

1.3.2 Specific Objectives

1. Evaluate the toxicity effects of zerumbone extract in rats.
2. Evaluate the effects of zerumbone on the paw thickness and paw volume in CFA-induced RA model in rats.
3. Investigate the effects of zerumbone on the mechanical allodynia and hyperalgesia in CFA- induced RA model in rats.
4. Investigate the effects of zerumbone on the thermal hyperalgesia in CFA- induced RA model in rats.

REFERENCES

- Adebajo, A., & Furst, D. E. (2005). Biologic agents and their use in resource-poor countries. *J Rheumatol*, 32, 1182-1183.
- AFM. (2009-2010). Arthritis Info Series Rheumatoid Arthritis. Retrieved 8 June, 2014, from <http://www.afm.org.my/info/ra.htm>
- Aggarwal, B. B., & Shishodia, S. (2006). Molecular targets of dietary agents for prevention and therapy of cancer. *Biochem Pharmacol*, 71(10), 1397-1421. doi: 10.1016/j.bcp.2006.02.009
- Allison, M. C., Howatson, A. G., Torrance, C. J., Lee, F. D., & Russell, R. I. (1992). Gastrointestinal damage associated with the use of nonsteroidal antiinflammatory drugs. *New England Journal of Medicine*, 327(11), 749-754.
- Alyce, M., Oliver, E., & William, S. C. (2008). Treatment and Assessment of Rheumatoid Arthritis. In K. J.H. (Ed.), *Primer on the Rheumatic Diseases* (pp. 133-141). USA: Springer Science.
- Apparailly, F., Verwaerde, C., Jacquet, C., Auriault, C., Sany, J., & Jorgensen, C. (1998). Adenovirus-mediated transfer of viral IL-10 gene inhibits murine collagen-induced arthritis. *The Journal of Immunology*, 160(11), 5213-5220.
- Atsamo, A. D., Nguelefack, T. B., Datte, J. Y., & Kamanyi, A. (2011). Acute and subchronic oral toxicity assessment of the aqueous extract from the stem bark of *Erythrina senegalensis* DC (Fabaceae) in rodents. *J Ethnopharmacol*, 134(3), 697-702. doi: 10.1016/j.jep.2011.01.023
- Bevilacqua, A. H., Suffredini, I. B., Romoff, P., Lago, J. H., & Bernardi, M. M. (2011). Toxicity of apolar and polar *Lantana camara* L. crude extracts in mice. *Res Vet Sci*, 90(1), 106-115. doi: 10.1016/j.rvsc.2010.05.001
- Bhuiyan, M. N. I., Chowdhury, J. U., & Begum, J. (2008). Chemical investigation of the leaf and rhizome essential oils of *Zingiber zerumbet* (L.) Smith from Bangladesh. *Bangladesh Journal of Pharmacology*, 4(1). doi: 10.3329/bjp.v4i1.845
- Boettger, M. K., Hensellek, S., Richter, F., Gajda, M., Stockigt, R., von Banchet, G. S., . . . Schaible, H. G. (2008). Antinociceptive effects of tumor necrosis factor alpha neutralization in a rat model of antigen-induced arthritis: evidence of a neuronal target. *Arthritis Rheum*, 58(8), 2368-2378. doi: 10.1002/art.23608
- Brennan, F. M., & McInnes, I. B. (2008). Evidence that cytokines play a role in rheumatoid arthritis. *J Clin Invest*, 118(11), 3537-3545. doi: 10.1172/JCI36389
- Bromley, M., & Woolley, D. E. (1984). Chondroclasts and osteoclasts at subchondral sites of erosion in the rheumatoid joint. *Arthritis & Rheumatism*, 27(9), 968-975. doi: 10.1002/art.1780270902

- Butler, S. H., Godefroy, F., Besson, J.-M., & Weil-Fugazza, J. (1992). A limited arthritic model for chronic pain studies in the rat. *Pain*, 48(1), 73-81. doi: [http://dx.doi.org/10.1016/0304-3959\(92\)90133-V](http://dx.doi.org/10.1016/0304-3959(92)90133-V)
- Calvino, B., Crepon-Bernard, M.-O., & Le Bars, D. (1987). Parallel clinical and behavioural studies of adjuvant-induced arthritis in the rat: Possible relationship with 'chronic pain'. *Behavioural Brain Research*, 24(1), 11-29. doi: 10.1016/0166-4328(87)90032-5
- Carr, A. J. (1999). Beyond disability: measuring the social and personal consequences of osteoarthritis. *Osteoarthritis and Cartilage*, 7(2), 230-238. doi: <http://dx.doi.org/10.1053/joca.1998.0154>
- Chaiyakunapruk, N., Kitikannakorn, N., Nathisuwon, S., Leeprakobboon, K., & Leelasettagool, C. (2006). The efficacy of ginger for the prevention of postoperative nausea and vomiting: a meta-analysis. *Am J Obstet Gynecol*, 194(1), 95-99. doi: 10.1016/j.ajog.2005.06.046
- Chillingworth, N. L., & Donaldson, L. F. (2003). Characterisation of a Freund's complete adjuvant-induced model of chronic arthritis in mice. *Journal of Neuroscience Methods*, 128(1-2), 45-52. doi: 10.1016/s0165-0270(03)00147-x
- Chiu, N., & Chang, K. (1986). Zingiberaceae. *Taipei: SMC Publishing Inc.*
- Choy, E. H., & Panayi, G. S. (2001). Cytokine pathways and joint inflammation in rheumatoid arthritis. *N Engl J Med*, 344(12), 907-916. doi: 10.1056/nejm200103223441207
- Coffey, G. H., & Mahon, M. V. (1982). Pain: Theories and a New Approach to treatment. *J Natl Med Assoc*, 74(2), 147-153.
- Colpaert, F. C. (1987). Evidence that adjuvant arthritis in the rat is associated with chronic pain. *Pain*, 28(2), 201-222. doi: [http://dx.doi.org/10.1016/0304-3959\(87\)90117-5](http://dx.doi.org/10.1016/0304-3959(87)90117-5)
- Costa-Silva, J. H., Lima, C. R., Silva, E. J., Araujo, A. V., Fraga, M. C., Ribeiro, E. R. A., . . . Wanderley, A. G. (2008). Acute and subacute toxicity of the Carapa guianensis Aublet (Meliaceae) seed oil. *J Ethnopharmacol*, 116(3), 495-500. doi: 10.1016/j.jep.2007.12.016
- Costa, C., Incio, J., & Soares, R. (2007). Angiogenesis and chronic inflammation: cause or consequence? *Angiogenesis*, 10(3), 149-166. doi: 10.1007/s10456-007-9074-0
- Coster, L., Kendall, S., Gerdle, B., Henriksson, C., Henriksson, K. G., & Bengtsson, A. (2008). Chronic widespread musculoskeletal pain - a comparison of those who meet criteria for fibromyalgia and those who do not. *Eur J Pain*, 12(5), 600-610. doi: 10.1016/j.ejpain.2007.10.001

- Danziger, N., Weil-Fugazza, J., Le Bars, D., & Bouhassira, D. (1999). Alteration of descending modulation of nociception during the course of monoarthritis in the rat. *The Journal of neuroscience*, 19(6), 2394-2400.
- Dayer, J.-M., de Rochemonteix, B., Burrus, B., Demczuk, S., & Dinarello, C. A. (1986). Human recombinant interleukin 1 stimulates collagenase and prostaglandin E2 production by human synovial cells. *Journal of Clinical Investigation*, 77(2), 645.
- Doeglas, D. M., Suurmeijer, T. P. B. M., van den Heuvel, W. J. A., Krol, B., van Rijswijk, M. H., van Leeuwen, M. A., & Sanderman, R. (2004). Functional Ability, Social Support, and Depression in Rheumatoid Arthritis. *Quality of Life Research*, 13(6), 1053-1065. doi: 10.1023/B:QURE.0000031339.04589.63
- Dũng, N. X., Chính, T. D., & Leclercq, P. A. (1995). Chemical investigation of the aerial parts of Zingiber zerumbet (L.) Sm. from Vietnam. *Journal of Essential Oil Research*, 7(2), 153-157.
- Ebbinghaus, M., Uhlig, B., Richter, F., von Banchet, G. S., Gajda, M., Brauer, R., & Schaible, H. G. (2012). The role of interleukin-1beta in arthritic pain: main involvement in thermal, but not mechanical, hyperalgesia in rat antigen-induced arthritis. *Arthritis Rheum*, 64(12), 3897-3907. doi: 10.1002/art.34675
- Ekeanyanwu, R. C., & Njoku, O. U. (2014). Acute and subacute oral toxicity study on the flavonoid rich fraction of Monodora tenuifolia seed in albino rats. *Asian Pacific Journal of Tropical Biomedicine*, 4(3), 194-202. doi: 10.1016/S2221-1691(14)60231-8
- Eklund, K., Leirisalo-Repo, M., Ranta, P., Maki, T., Kautiainen, H., Hannonen, P., . . . Mottonen, T. (2007). Serum IL-1 beta levels are associated with the presence of erosions in recent onset rheumatoid arthritis. *Clinical and experimental rheumatology*, 25(5), 684.
- Fakurazi, S., Ithnin, H., & Ganabadi, S. (2008). The effect of pretreatment of zerumbone on fatty liver following ethanol induced hepatotoxicity. *Journal of Biological Sciences*, 8(8), 1348-1351.
- Fava, R. A., Olsen, N. J., Spencer-Green, G., Yeo, K. T., Yeo, T. K., Berse, B., . . . Brown, L. F. (1994). Vascular permeability factor/endothelial growth factor (VPF/VEGF): accumulation and expression in human synovial fluids and rheumatoid synovial tissue. *The Journal of Experimental Medicine*, 180(1), 341-346. doi: 10.1084/jem.180.1.341
- Ferraccioli, G. F., & Bartoli, E. (1998). Xenobiotics or biological response modifiers? Methotrexate remains the anchor drug for rheumatoid arthritis. *Clin Exp Rheumatol*, 16(6), 662-666.
- Ferrell, B. (2003). Acute and Chronic Pain *Geriatric Medicine* (pp. 323-342): Springer New York.

- Friederichs, K., Schmitz, J., Weissenbach, M., Heinrich, P. C., & Schaper, F. (2001). Interleukin-6-induced proliferation of pre-B cells mediated by receptor complexes lacking the SHP2/SOCS3 recruitment sites revisited. *Eur J Biochem*, 268(24), 6401-6407.
- Gabay, C. (2006). Interleukin-6 and chronic inflammation. *Arthritis research and therapy*, 8(2), S3.
- Gerecz-Simon, E. M., Tunks, E. R., Heale, J. A., Kean, W. F., & Buchanan, W. W. (1989). Measurement of pain threshold in patients with rheumatoid arthritis, osteoarthritis, ankylosing spondylitis, and healthy controls. *Clinical Rheumatology*, 8(4), 467-474. doi: 10.1007/BF02032098
- Goldring, S. R. (2008). Inflammation-Induced Bone Loss in Rheumatic Diseases (pp. 272-276). Washington, DC: American Society for Bone and Mineral Research.
- Goronyz, J. J., & Weyand, C. M. (2009). Developments in the scientific understanding of rheumatoid arthritis. *Arthritis Res Ther*, 11(5), 249. doi: 10.1186/ar2758
- Gravallese, E. M., Manning, C., Tsay, A., Naito, A., Pan, C., Amento, E., & Goldring, S. R. (2000). Synovial tissue in rheumatoid arthritis is a source of osteoclast differentiation factor. *Arthritis & Rheumatism*, 43(2), 250-258. doi: 10.1002/1529-0131(200002)43:2<250::AID-ANR3>3.0.CO;2-P
- Gureje, O., Von Korff, M., Simon, G. E., & Gater, R. (1998). Persistent pain and well-being: A world health organization study in primary care. *JAMA*, 280(2), 147-151. doi: 10.1001/jama.280.2.147
- Hargreaves, K., Dubner, R., Brown, F., Flores, C., & Joris, J. (1988). A new and sensitive method for measuring thermal nociception in cutaneous hyperalgesia. *Pain*, 32(1), 77-88.
- Helyes, Z., Szabo, A., Nemeth, J., Jakab, B., Pinter, E., Banvolgyi, A., . . . Szolcsanyi, J. (2004). Antiinflammatory and analgesic effects of somatostatin released from capsaicin-sensitive sensory nerve terminals in a Freund's adjuvant-induced chronic arthritis model in the rat. *Arthritis Rheum*, 50(5), 1677-1685. doi: 10.1002/art.20184
- Herman, C. J., Allen, P., Prasad, A., Hunt, W. C., & Brady, T. J. (2004). Use of complementary therapies among primary care clinic patients with arthritis. *Prev Chronic Dis*.
- Holmdahl, R., Lorentzen, J. C., Lu, S., Olofsson, P., Wester, L., Holmberg, J., & Pettersson, U. (2001). Arthritis induced in rats with non-immunogenic adjuvants as models for rheumatoid arthritis. *Immunological reviews*, 184(1), 184-202.
- Ibrahim, M. Y., Abdul, A. B., Ibrahim, T. A. T., Abdelwahab, S. I., Elhassan, M. M., & Syam, M. (2010). Evaluation of acute toxicity and the effect of single injected doses of zerumbone on the kidney and liver functions in Sprague Dawley rats. *African Journal of Biotechnology*, 9(28), 4442-4450.

- Jacoby, R. K., Jayson, M. I., & Cosh, J. A. (1973). Onset, early stages, and prognosis of rheumatoid arthritis: a clinical study of 100 patients with 11-year follow-up. *Br Med J*, 2(5858), 96-100.
- Janeway, C. A. J., Travers, P., Walport, M., & Shlomchik, M. J. (2001). *Immunobiology: The Immune System in Health and Disease*. Vol. 5th edition.
- Jongbloed, S. L., Lebre, M. C., Fraser, A. R., Gracie, J. A., Sturrock, R. D., Tak, P. P., & McInnes, I. B. (2006). Enumeration and phenotypical analysis of distinct dendritic cell subsets in psoriatic arthritis and rheumatoid arthritis. *Arthritis Res Ther*, 8(1), R15. doi: 10.1186/ar1864
- Joosten, L. A., Helsen, M. M., Saxne, T., van de Loo, F. A., Heinegård, D., & van den Berg, W. B. (1999). IL-1 $\alpha\beta$ blockade prevents cartilage and bone destruction in murine type II collagen-induced arthritis, whereas TNF- α blockade only ameliorates joint inflammation. *The Journal of Immunology*, 163(9), 5049-5055.
- Kala, C., Farooquee, N., & Dhar, U. (2004). Prioritization of medicinal plants on the basis of available knowledge, existing practices and use value status in Uttarakhand, India. *Biodiversity & Conservation*, 13(2), 453-469. doi: 10.1023/B:BIOC.0000006511.67354.7f
- Keffer, J., Probert, L., Cazlaris, H., Georgopoulos, S., Kaslaris, E., Kioussis, D., & Kollias, G. (1991). Transgenic mice expressing human tumour necrosis factor: a predictive genetic model of arthritis. *The EMBO journal*, 10(13), 4025.
- Kim, K. R., Jeong, C. K., Park, K. K., Choi, J. H., Park, J. H., Lim, S. S., & Chung, W. Y. (2010). Anti-inflammatory effects of licorice and roasted licorice extracts on TPA-induced acute inflammation and collagen-induced arthritis in mice. *J Biomed Biotechnol*, 2010, 709378. doi: 10.1155/2010/709378
- Kokkonen, H., Soderstrom, I., Rocklov, J., Hallmans, G., Lejon, K., & Rantapaa Dahlqvist, S. (2010). Up-regulation of cytokines and chemokines predates the onset of rheumatoid arthritis. *Arthritis Rheum*, 62(2), 383-391. doi: 10.1002/art.27186
- Kong, J. M., Goh, N. K., Chia, L. S., & Chia, T. F. (2003). Recent advances in traditional plant drugs and orchids. *Acta Pharmacologica Sinica*, 24(1), 7-21.
- Kumar, V., Abbas, A. K., & Aster, J. C. (2003). *Robbins basic pathology* (9 ed.). Philadelphia, PA: Saunders.
- Kumar, V., Abbas, A. K., Fausto, N., & Aster, J. C. (2004). *Robbins and Cotran Pathologic Basis of Disease* (8 ed.). Philadelphia, PA: Saunders.
- Kumar, V. L., Roy, S., Sehgal, R., & Padhy, B. M. (2006). A comparative study on the efficacy of rofecoxib in monoarticular arthritis induced by latex of Calotropis procera and Freund's complete adjuvant. *Inflammopharmacology*, 14(1-2), 17-21.

- Kwon, Y. B., Lee, H. J., Han, H. J., Mar, W. C., Kang, S. K., Yoon, O. B., . . . Lee, J. H. (2002). The water-soluble fraction of bee venom produces antinociceptive and anti-inflammatory effects on rheumatoid arthritis in rats. *Life Sci*, 71(2), 191-204.
- Laird, J., Carter, A., Grauert, M., & Cervero, F. (2001). Analgesic activity of a novel use-dependent sodium channel blocker, crobenetine, in monoarthritic rats. *British journal of pharmacology*, 134(8), 1742-1748.
- Lambert, G. A., Mallos, G., & Zagami, A. S. (2009). Von Frey's hairs--a review of their technology and use--a novel automated von Frey device for improved testing for hyperalgesia. *J Neurosci Methods*, 177(2), 420-426. doi: 10.1016/j.jneumeth.2008.10.033
- Larsen, K., Ibrahim, A., & Wong, K. M. (1999). *Gingers of Peninsular Malaysia and Singapore*: Natural History Publications (Borneo).
- Lee, Y. C., Chibnik, L. B., Lu, B., Wasan, A. D., Edwards, R. R., Fosse, A. H., . . . Karlson, E. W. (2009). The relationship between disease activity, sleep, psychiatric distress and pain sensitivity in rheumatoid arthritis: a cross-sectional study. *Arthritis Res Ther*, 11(5), R160. doi: 10.1186/ar2842
- Leffler, A.-S., Kosek, E., Lerndal, T., Nordmark, B., & Hansson, P. (2002). Somatosensory perception and function of diffuse noxious inhibitory controls (DNIC) in patients suffering from rheumatoid arthritis. *Eur J Pain*, 6(2), 161-176. doi: 10.1053/eujp.2001.0313
- Lin, B., Zhang, H., Zhao, X.-X., Rahman, K., Wang, Y., Ma, X.-Q., . . . Qin, L.-P. (2013). Inhibitory effects of the root extract of Litsea cubeba (lour.) pers. on adjuvant arthritis in rats. *Journal of Ethnopharmacology*, 147(2), 327-334. doi: <http://dx.doi.org/10.1016/j.jep.2013.03.011>
- Majno, G., & Joris, I. (2004). *Cells, tissues, and disease: principles of general pathology*. USA: Oxford University Press.
- Marsland, D., & Kapoor, S. (2004). Crash course: Rheumatology and Orthopaedics (pp. 51-59). London, UK: Mosby Ltd.
- McCurdy, C. R., & Scully, S. S. (2005). Analgesic substances derived from natural products (natureceuticals). *Life Sci*, 78(5), 476-484. doi: 10.1016/j.lfs.2005.09.006
- Merskey, H. (1986). Classification of chronic pain: Descriptions of chronic pain syndromes and definitions of pain terms. *Pain, Suppl 3*, 226.
- Meyer, L. H., Franssen, L., & Pap, T. (2006). The role of mesenchymal cells in the pathophysiology of inflammatory arthritis. *Best Pract Res Clin Rheumatol*, 20(5), 969-981. doi: 10.1016/j.bepr.2006.06.005

- Millan, M. J. (1999). The induction of pain: an integrative review. *Progress in Neurobiology*, 57(1), 1-164. doi: [http://dx.doi.org/10.1016/S0301-0082\(98\)00048-3](http://dx.doi.org/10.1016/S0301-0082(98)00048-3)
- Mimori, T. (2005). Clinical Significance of Anti-CCP Antibodies in Rheumatoid Arthritis. *Internal Medicine*, 44(11), 1122-1126. doi: 10.2169/internalmedicine.44.1122
- Miossec, P. (2004). An update on the cytokine network in rheumatoid arthritis. *Current Opinion in Rheumatology*, 16(3), 218-222.
- Morrissey, R. E., Horvath, C., Snyder, E. A., Patrick, J., & MacDonald, J. S. (2002). Rodent Nonclinical Safety Evaluation Studies of SCH 58500, an Adenoviral Vector for the p53 Gene. *Toxicological Sciences*, 65(2), 266-275. doi: 10.1093/toxsci/65.2.266
- Mowat, A. G. (1972). Hematologic abnormalities in rheumatoid arthritis. *Semin Arthritis Rheum*, 1(3), 195-219.
- Murakami, A., & Ohigashi, H. (2007). Targeting NOX, INOS and COX-2 in inflammatory cells: chemoprevention using food phytochemicals. *Int J Cancer*, 121(11), 2357-2363. doi: 10.1002/ijc.23161
- Murakami, A., Shigemori, T., & Ohigashi, H. (2005). Zingiberaceous and Citrus Constituents, 1'-Acetoxychavicol Acetate, Zerumbone, Auraptene, and Nobiletin, Suppress Lipopolysaccharide-Induced Cyclooxygenase-2 Expression in RAW264.7 Murine Macrophages through Different Modes of Action. *The Journal of Nutrition*, 135(12), 2987S-2992S.
- Murakami, A., Takahashi, M., Jiwajinda, S., Koshimizu, K., & Ohigashi, H. (1999). Identification of Zerumbone in Zingiber zerumbet Smith as a Potent Inhibitor of 12-O-Tetradecanoylphorbol-13-acetate-induced Epstein-Barr Virus Activation. *Bioscience, Biotechnology, and Biochemistry*, 63(10), 1811-1812. doi: 10.1271/bbb.63.1811
- Nagakura, Y., Okada, M., Kohara, A., Kiso, T., Toya, T., Iwai, A., . . . Yamaguchi, T. (2003). Allodynia and hyperalgesia in adjuvant-induced arthritic rats: time course of progression and efficacy of analgesics. *J Pharmacol Exp Ther*, 306(2), 490-497. doi: 10.1124/jpet.103.050781
- Neugebauer, V., Han, J. S., Adwanikar, H., Fu, Y., & Ji, G. (2007). Techniques for assessing knee joint pain in arthritis. *Mol Pain*, 3, 8. doi: 10.1186/1744-8069-3-8
- Nigam, I. C., & Levi, L. (1963). COLUMN AND GAS CHROMATOGRAPHIC ANALYSIS OF OIL OF WILD GINGER: IDENTIFICATION AND ESTIMATION OF SOME NEW CONSTITUENTS. *Canadian Journal of Chemistry*, 41(7), 1726-1730. doi: 10.1139/v63-248

- Olajide, O., Makinde, J. M., & Okpako, D. (2003). Evaluation of the anti-inflammatory property of the extract of Combretum micranthum G. Don (Combretaceae). *Inflammopharmacology*, 11(3), 293-298. doi: 10.1163/156856003322315631
- Oliver, A. M., & Clair, E. W. S. (2008). Rheumatoid Arthritis C. Treatment and assessment. In J. H. Klippen (Ed.), *Primer on the Rheumatic Diseases* (13 ed., pp. 133-141). USA: Springer Science + Business Media.
- Olson, H., Betton, G., Robinson, D., Thomas, K., Monro, A., Kolaja, G., . . . Heller, A. (2000). Concordance of the toxicity of pharmaceuticals in humans and in animals. *Regul Toxicol Pharmacol*, 32(1), 56-67. doi: 10.1006/rtpb.2000.1399
- Opree, A., & Kress, M. (2000). Involvement of the proinflammatory cytokines tumor necrosis factor-alpha, IL-1 beta, and IL-6 but not IL-8 in the development of heat hyperalgesia: effects on heat-evoked calcitonin gene-related peptide release from rat skin. *J Neurosci*, 20(16), 6289-6293.
- Palatty, P. L., Haniadka, R., Valder, B., Arora, R., & Baliga, M. S. (2013). Ginger in the prevention of nausea and vomiting: a review. *Crit Rev Food Sci Nutr*, 53(7), 659-669. doi: 10.1080/10408398.2011.553751
- Paleolog, E. M. (2002). Angiogenesis in rheumatoid arthritis. *Arthritis Res*, 4(Suppl 3), S81-S90.
- Park, H. J., Hong, M. S., Lee, J. S., Leem, K. H., Kim, C. J., Kim, J. W., & Lim, S. (2005). Effects of Aralia continentalis on hyperalgesia with peripheral inflammation. *Phytother Res*, 19(6), 511-513. doi: 10.1002/ptr.1693
- Patil, K. R., Patil, C. R., Jadhav, R. B., Mahajan, V. K., Patil, P. R., & Gaikwad, P. S. (2011). Anti-Arthritic Activity of Bartogenic Acid Isolated from Fruits of Barringtonia racemosa Roxb. (Lecythidaceae). *Evid Based Complement Alternat Med*, 2011, 785245. doi: 10.1093/ecam/nep148
- Perimal, E. K., Akhtar, M. N., Mohamad, A. S., Khalid, M. H., Ming, O. H., Khalid, S., . . . Sulaiman, M. R. (2011). Zerumbone-Induced Antinociception: Involvement of the l-Arginine-Nitric Oxide-cGMP -PKC-K+ATP Channel Pathways. *Basic & Clinical Pharmacology & Toxicology*, 108(3), 155-162. doi: 10.1111/j.1742-7843.2010.00635.x
- Petrovic-Rackov, L., & Pejnovic, N. (2006). Clinical significance of IL-18, IL-15, IL-12 and TNF- α measurement in rheumatoid arthritis. *Clinical Rheumatology*, 25(4), 448-452. doi: 10.1007/s10067-005-0106-0
- Philippe, L., Gegout-Pottie, P., Guingamp, C., Bordji, K., Terlain, B., Netter, P., & Gillet, P. (1997). Relations between functional, inflammatory, and degenerative parameters during adjuvant arthritis in rats. *American Journal of Physiology - Regulatory, Integrative and Comparative Physiology*, 273(4), R1550-R1556.
- Phillipson, J. D. (2001). Phytochemistry and medicinal plants. *Phytochemistry*, 56(3), 237-243. doi: [http://dx.doi.org/10.1016/S0031-9422\(00\)00456-8](http://dx.doi.org/10.1016/S0031-9422(00)00456-8)

- Platel, K., & Srinivasan, K. (2000). Influence of dietary spices and their active principles on pancreatic digestive enzymes in albino rats. *Food/Nahrung*, 44(1), 42-46.
- Pomeranz, B., & Berman, B. (2003). Scientific Basis of Acupuncture *Basics of Acupuncture* (pp. 7-86): Springer Berlin Heidelberg.
- Proudman, S. M., Cleland, L. G., & Mayrhofer, G. (1999). Effects of tumor necrosis factor-alpha, interleukin 1beta, and activated peripheral blood mononuclear cells on the expression of adhesion molecules and recruitment of leukocytes in rheumatoid synovial xenografts in SCID mice. *The Journal of rheumatology*, 26(9), 1877-1889.
- Raghavendra, V., Tanga, F. Y., & DeLeo, J. A. (2004). Complete Freunds adjuvant-induced peripheral inflammation evokes glial activation and proinflammatory cytokine expression in the CNS. *Eur J Neurosci*, 20(2), 467-473. doi: 10.1111/j.1460-9568.2004.03514.x
- Randall, L. O., & Selitto, J. J. (1957). A method for measurement of analgesic activity on inflamed tissue. *Archives internationales de pharmacodynamie et de therapie*, 111(4), 409-419.
- Ranzolin, A., Brenol, J. C., Bredemeier, M., Guarienti, J., Rizzatti, M., Feldman, D., & Xavier, R. M. (2009). Association of concomitant fibromyalgia with worse disease activity score in 28 joints, health assessment questionnaire, and short form 36 scores in patients with rheumatoid arthritis. *Arthritis Rheum*, 61(6), 794-800. doi: 10.1002/art.24430
- Ravindran, P. N., & Babu, K. N. (2005). *Ginger: the genus Zingiber*. Florida: CRC Press.
- Remadevi, R., Surendran, E., & Ravindran, P. (2005). Properties and medicinal uses of ginger. *Ginger. The genus Zingiber*, 489-508.
- Riddle, J. M. (1985). *Dioscorides on pharmacy and medicine*: University of Texas Press Austin.
- Rong, X., Peng, G., Suzuki, T., Yang, Q., Yamahara, J., & Li, Y. (2009). A 35-day gavage safety assessment of ginger in rats. *Regul Toxicol Pharmacol*, 54(2), 118-123. doi: 10.1016/j.yrtph.2009.03.002
- Rovensky, J., & Pavelka, K. (2008). Rheumatoid Arthritis. In A. Gvozdjakova (Ed.), *Mitochondrial Medicine* (pp. 201-244). Slovakia: Springer Science + Business Media.
- Ruslay, S., Abas, F., Shaari, K., Zainal, Z., Maulidiani, Sirat, H., . . . Lajis, N. H. (2007). Characterization of the components present in the active fractions of health gingers (*Curcuma xanthorrhiza* and *Zingiber zerumbet*) by HPLC-DAD-ESIMS. *Food Chemistry*, 104(3), 1183-1191. doi: 10.1016/j.foodchem.2007.01.067

- Saag, K. G., Koehnke, R., Caldwell, J. R., Brasington, R., Burmeister, L. F., Zimmerman, B., . . . Furst, D. E. (1994). Low dose long-term corticosteroid therapy in rheumatoid arthritis: An analysis of serious adverse events. *The American Journal of Medicine*, 96(2), 115-123. doi: [http://dx.doi.org/10.1016/0002-9343\(94\)90131-7](http://dx.doi.org/10.1016/0002-9343(94)90131-7)
- Sandkühler, J. (2009). Models and Mechanisms of Hyperalgesia and Allodynia. *Physiological Reviews*, 89(2), 707-758.
- Schett, G., Hayer, S., Zwerina, J., Redlich, K., & Smolen, J. S. (2005). Mechanisms of Disease: the link between RANKL and arthritic bone disease. *Nat Clin Pract Rheum*, 1(1), 47-54.
- Schilter, B., Andersson, C., Anton, R., Constable, A., Kleiner, J., O'Brien, J., . . . Walker, R. (2003). Guidance for the safety assessment of botanicals and botanical preparations for use in food and food supplements. *Food and Chemical Toxicology*, 41(12), 1625-1649. doi: 10.1016/s0278-6915(03)00221-7
- Sharma, I., Gusain, D., & Dixit, V. P. (1996). Hypolipidaemic and Antiatherosclerotic Effects of *Zingiber officinale* in Cholesterol Fed Rabbits. *Phytotherapy Research*, 10(6), 517-518. doi: 10.1002/(SICI)1099-1573(199609)10:6<517::AID-PTR839>3.0.CO;2-L
- Shukla, S., Jadon, A., Bhadauria, M., & Sharma, A. (2007). Prevention of acute CCl₄ induced hepatic and renal toxicity in rats by PHF: apropriety herbal formulation. *Int. J. Pharmacol.*, 1, 71-80.
- Shukla, Y., & Singh, M. (2007). Cancer preventive properties of ginger: a brief review. *Food Chem Toxicol*, 45(5), 683-690. doi: 10.1016/j.fct.2006.11.002
- Simons, S. H. P., & Tibboel, D. (2006). Pain perception development and maturation. *Seminars in Fetal and Neonatal Medicine*, 11(4), 227-231. doi: <http://dx.doi.org/10.1016/j.siny.2006.02.010>
- Smith, M. D., Barg, E., Weedon, H., Papengelis, V., Smeets, T., Tak, P. P., . . . Ahern, M. J. (2003). Microarchitecture and protective mechanisms in synovial tissue from clinically and arthroscopically normal knee joints. *Annals of the Rheumatic Diseases*, 62(4), 303-307. doi: 10.1136/ard.62.4.303
- Solomon, F. E., Sharada, A., & Devi, P. U. (1993). Toxic effects of crude root extract of <i>Plumbago rosea</i> (Rakta chitraka) on mice and rats. *Journal of Ethnopharmacology*, 38(1), 79-84.
- Sommer, C., & Kress, M. (2004). Recent findings on how proinflammatory cytokines cause pain: peripheral mechanisms in inflammatory and neuropathic hyperalgesia. *Neurosci Lett*, 361(1-3), 184-187. doi: 10.1016/j.neulet.2003.12.007
- Suekawa, M., Ishige, A., Yuasa, K., Sudo, K., Aburada, M., & Hosoya, E. (1984). Pharmacological studies on ginger. I. Pharmacological actions of pungent

- constituents,(6)-gingerol and (6)-shogaol. *Journal of pharmacobio-dynamics*, 7(11), 836-848.
- Sulaiman, M. R., Perimal, E. K., Akhtar, M. N., Mohamad, A. S., Khalid, M. H., Tasrip, N. A., . . . Israf, D. A. (2010). Anti-inflammatory effect of zerumbone on acute and chronic inflammation models in mice. *Fitoterapia*, 81(7), 855-858. doi: 10.1016/j.fitote.2010.05.009
- Sulaiman, M. R., Perimal, E. K., Zakaria, Z. A., Mokhtar, F., Akhtar, M. N., Lajis, N. H., & Israf, D. A. (2009). Preliminary analysis of the antinociceptive activity of zerumbone. *Fitoterapia*, 80(4), 230-232. doi: 10.1016/j.fitote.2009.02.002
- Tak, P. P., & Bresnihan, B. (2000). The pathogenesis and prevention of joint damage in rheumatoid arthritis: Advances from synovial biopsy and tissue analysis. *Arthritis & Rheumatism*, 43(12), 2619-2633. doi: 10.1002/1529-0131(200012)43:12<2619::AID-ANR1>3.0.CO;2-V
- Takada, Y., Murakami, A., & Aggarwal, B. B. (2005). Zerumbone abolishes NF-kappaB and IkappaBalph kinase activation leading to suppression of antiapoptotic and metastatic gene expression, upregulation of apoptosis, and downregulation of invasion. *Oncogene*, 24(46), 6957-6969. doi: 10.1038/sj.onc.1208845
- Tam, L. S., Leung, P. C., Li, T. K., Zhang, L., & Li, E. K. (2007). Acupuncture in the treatment of rheumatoid arthritis: a double-blind controlled pilot study. *BMC Complement Altern Med*, 7, 35. doi: 10.1186/1472-6882-7-35
- Tang, L.-Q., Wei, W., & Wang, X.-Y. (2007). Effects and mechanisms of catechin for adjuvant arthritis in rats. *Advances in Therapy*, 24(3), 679-690. doi: 10.1007/BF02848793
- Tarner, I. H., Harle, P., Muller-Ladner, U., Gay, R. E., & Gay, S. (2005). The different stages of synovitis: acute vs chronic, early vs late and non-erosive vs erosive. *Best Pract Res Clin Rheumatol*, 19(1), 19-35. doi: 10.1016/j.berh.2004.08.002
- Tehlirian, C. V., & Bathon, J. M. (2008). Rheumatoid Arthritis A. Clinical and laboratory manifestations. In J. H. Klippen (Ed.), *Primer on the Rheumatic Diseases* (Vol. 13, pp. 114-121). USA: Springer Science + Business Media.
- Tekieh, E., Zaringhalam, J., Manaheji, H., Maghsoudi, N., Alani, B., & Zardooz, H. (2011). Increased serum IL-6 level time-dependently regulates hyperalgesia and spinal mu opioid receptor expression during CFA-induced arthritis. *EXCLI J*, 10, 23-33.
- Tewtrakul, S., & Subhadhirasakul, S. (2007). Anti-allergic activity of some selected plants in the Zingiberaceae family. *J Ethnopharmacol*, 109(3), 535-538. doi: 10.1016/j.jep.2006.08.010
- Turesson, C., & Matteson, E. L. (2006). Genetics of Rheumatoid Arthritis. *Mayo Clinic Proceedings*, 81(1), 94-101. doi: <http://dx.doi.org/10.4065/81.1.94>

- Udagawa, N., Takahashi, N., Akatsu, T., Tanaka, H., Sasaki, T., Nishihara, T., . . . Suda, T. (1990). Origin of osteoclasts: mature monocytes and macrophages are capable of differentiating into osteoclasts under a suitable microenvironment prepared by bone marrow-derived stromal cells. *Proc Natl Acad Sci U S A*, 87(18), 7260-7264.
- van der Heijde, D. M. F. M. (1995). Joint Erosions and Patients with Early Rheumatoid Arthritis. *Rheumatology*, XXXIV(suppl 2), 74-78. doi: 10.1093/rheumatology/XXXIV.suppl_2.74
- Vasala, P. A. (2004). Ginger. In K. V. Peter (Ed.), *Handbook of Herbs and Spices* (pp. 195-206). England: Woodhead Publishing Limited.
- Vasantha, P., Nalini, G., & Rajasekhar, G. (2007). Role of tumor necrosis factor-alpha in rheumatoid arthritis: a review. *APLAR Journal of Rheumatology*, 10, 270-274.
- Waldburger, J.-M., & Firestein, G. S. (2008). Rheumatoid Arthritis B. Epidemiological, pathology and pathogenesis. In J. H. Klipper (Ed.), *Primer on the Rheumatic Diseases* (Vol. 13, pp. 123-132). USA: Springer Science + Business Media.
- Walsh, T. D. (1990). Prevention of opioid side effects. *Journal of Pain and Symptom Management*, 5(6), 362-367. doi: [http://dx.doi.org/10.1016/0885-3924\(90\)90031-E](http://dx.doi.org/10.1016/0885-3924(90)90031-E)
- Weidner, M. S., & Sigwart, K. (2000). The safety of a ginger extract in the rat. *Journal of Ethnopharmacology*, 73(3), 513-520. doi: [http://dx.doi.org/10.1016/S0378-8741\(00\)00340-8](http://dx.doi.org/10.1016/S0378-8741(00)00340-8)
- Weiner, M. A. (1994). Herbal antioxidants in clinical practice. *Journal of Orthomolecular Medicine*, 9, 167-167.
- Westhovens, R., & Dequeker, J. (2000). Rheumatoid arthritis and osteoporosis. *Zeitschrift für Rheumatologie*, 59(1), I33-I38. doi: 10.1007/s003930070036
- White, A., Foster, N. E., Cummings, M., & Barlas, P. (2007). Acupuncture treatment for chronic knee pain: a systematic review. *Rheumatology (Oxford)*, 46(3), 384-390. doi: 10.1093/rheumatology/kel413
- Wilhelm, E. A., Jesse, C. R., Bortolatto, C. F., Nogueira, C. W., & Savegnago, L. (2009). Antinociceptive and anti-allodynic effects of 3-alkynyl selenophene on different models of nociception in mice. *Pharmacol Biochem Behav*, 93(4), 419-425. doi: 10.1016/j.pbb.2009.06.003
- Witthawaskul, P., Panthong, A., Kanjanapothi, D., Taesothikul, T., & Lertprasertsuke, N. (2003). Acute and subacute toxicities of the saponin mixture isolated from Schefflera leucantha Viguier. *Journal of Ethnopharmacology*, 89(1), 115-121. doi: 10.1016/s0378-8741(03)00273-3
- Woolf, C. J., & Walters, E. T. (1991). Common patterns of plasticity contributing to nociceptive sensitization in mammals and Aplysia. *Trends in Neurosciences*, 14(2), 74-78. doi: [http://dx.doi.org/10.1016/0166-2236\(91\)90024-O](http://dx.doi.org/10.1016/0166-2236(91)90024-O)

- Yob, N. J., Jofrry, S. M., Affandi, M. M., Teh, L. K., Salleh, M. Z., & Zakaria, Z. A. (2011). *Zingiber zerumbet* (L.) Smith: A Review of Its Ethnomedicinal, Chemical, and Pharmacological Uses. *Evid Based Complement Alternat Med*, 2011, 543216. doi: 10.1155/2011/543216
- Zakaria, Z. A., Mohamad, A. S., Ahmad, M. S., Mokhtar, A. F., Israf, D. A., Lajis, N. H., & Sulaiman, M. R. (2011). Preliminary analysis of the anti-inflammatory activity of essential oils of *Zingiber zerumbet*. *Biol Res Nurs*, 13(4), 425-432. doi: 10.1177/1099800410386590
- Zhang, L., Li, J., Yu, S. C., Jin, Y., Lv, X. W., Zou, Y. H., & Li, Y. (2008). Therapeutic effects and mechanisms of total flavonoids of *Turpinia Arguta* Seen on adjuvant arthritis in rats. *J Ethnopharmacol*, 116(1), 167-172. doi: 10.1016/j.jep.2007.11.027
- Zimmermann, M. (1983). Ethical guidelines for investigations of experimental pain in conscious animals. *Pain*, 16(2), 109-110.
- Zwerina, J., Redlich, K., Schett, G., & Smolen, J. S. (2005). Pathogenesis of rheumatoid arthritis: targeting cytokines. *Ann N Y Acad Sci*, 1051, 716-729. doi: 10.1196/annals.1361.116