



UNIVERSITI PUTRA MALAYSIA

***IN VITRO ANTI-ATHEROGENIC EFFECTS OF ASIATIC ACID IN HUMAN
AORTIC ENDOTHELIAL CELLS***

FONG LAI YEN

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By

FONG LAI YEN

**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

**IN VITRO ANTI-ATHEROGENIC EFFECTS OF ASIATIC ACID IN HUMAN
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November 2016

Chair : Zuraini Ahmad, PhD

Faculty : Medicine and Health Sciences

In early pre-lesional stage of atherosclerosis, endothelial cell activation is characterized by increased endothelial permeability, increased expression of cell adhesion molecules (CAMs), leukocyte adhesion and migration across the endothelium. Asiatic acid is a major triterpene isolated from *Centella asiatica* (L.) Urban and it has been shown to possess anti-oxidant, hypolipidemia and anti-inflammatory activities. This study aimed to investigate protective effects of asiatic acid on TNF- α -induced early atherogenic events, in the context of endothelial cell activation, using human aortic endothelial cells (HAECs). Fluorescein isothiocyanate (FITC)-dextran permeability assay, U937 monocyte adhesion and migration assays were examined using fluorescence-based methods. The levels of soluble CAMs were measured using multiplex kits and flow cytometry. Localization of filamentous (F)-actin, diphosphorylated myosin light chain (diphospho-MLC), adherens junctions (AJs) and tight junctions (TJs) in cells were investigated using immunocytochemistry and confocal microscopy. Total protein expression of CAMs, diphospho-MLC, vascular endothelial (VE)-cadherin, β -catenin, occludin, zona occludens (ZO)-1 and phosphorylated inhibitors of κ B (p-I κ B- α) were determined using western blot analysis. The expression of AJ and TJ proteins in membrane, cytosolic and cytoskeleton fractions were also determined using western blot analysis. Asiatic acid significantly suppressed TNF- α -induced endothelial hyperpermeability, but did not reduce the increased monocyte adhesion and migration. Asiatic acid also inhibited VCAM-1 expression and production of soluble CAMs (sE-selectin, sICAM-1, sVCAM-1 and sPECAM-1). Besides, asiatic acid prevented TNF- α -induced redistribution of F-actin but failed to alter the increased F/G actin ratio. By using cytochalasin D, an actin depolymerizing agent, asiatic acid was demonstrated to stabilize peripheral F-actin filaments. Yet, asiatic acid did not improve cytochalasin D-induced increased permeability. Furthermore, asiatic acid localized diphospho-MLC filaments at the cell periphery and significantly augmented TNF- α -induced increased MLC diphosphorylation. Asiatic acid also prevented TNF- α -induced reticular AJ disruption by enhancing junctional areas covered by VE-cadherin and β -catenin. This protective effect was found to be independent of changes in either total amount or intracellular redistribution of VE-cadherin and β -catenin. For TJs, confocal imaging showed that asiatic acid opposed TNF- α -induced loss of ZO-1 from the cell borders.

However, asiatic acid did not alter both total occludin and ZO-1 expressions in whole cell lysate. Subcellular fractionation demonstrated that asiatic acid inhibited TNF- α -induced ZO-1 internalization from membrane to cytoplasm and occludin redistribution from cytoplasm to cytoskeleton. In addition, asiatic acid suppressed TNF- α -induced phosphorylation of I κ B- α . These results suggest that asiatic acid protects against TNF- α -induced endothelial barrier disruption and reduces the release of soluble CAMs, which are important biomarkers for the risk prediction of cardiovascular diseases. Asiatic acid also stabilizes the cytoskeleton by localizing peripheral F-actin and diphospho-MLC filaments. However, the actin stabilization is not essential for barrier protective effect of asiatic acid. The barrier stabilizing effect of asiatic acid is found concomitant with enhancement of reticular AJ formation, inhibition of TJ redistribution and suppression of NF- κ B activation. In conclusion, a novel protective role of asiatic acid in TNF- α -induced endothelial barrier dysfunction was demonstrated. This reveals new therapeutic usage of asiatic acid in the prevention of early atherosclerosis, which involves endothelial activation.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

KESAN ANTI-ATEROGENIK ASIATIC ASID PADA SEL ENDOTELIAL AORTA MANUSIA *IN VITRO*

Oleh

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November 2016

Pengerusi : **Zuraini Ahmad, PhD**

Fakulti : **Perubatan dan Sains Kesihatan**

Pada peringkat awal aterosklerosis di mana sebelum pembentukan lesi dimulakan, pengaktifan sel endotelial dicirikan oleh peningkatan ketelapan endotelial, peningkatan pengekspresan molekul-molekul lekatan dalam sel dan penambahan kelekatan serta penghijrahan leukosit melintasi lapisan endotelium. Asiatic asid adalah salah satu triterpena yang diasingkan daripada *Centella asiatica* (L.) Urban dan aktiviti-aktiviti anti-pengoksidaan, hipolipidemik dan anti-keradangannya telah dilaporkan. Kajian ini bertujuan untuk mengkaji kesan perlindungan asiatic asid terhadap peristiwa awal aterosgenik yang diaruhkan oleh TNF- α , dalam konteks pengaktifan sel endotelial, dengan menggunakan sel endotelial aorta manusia. Asai ketelapan fluoresein isotiosianat-dekstran, asai-asai kelekatan dan penghijrahan monosit U937 telah dinilai oleh keadah pendarfluor. Paras molekul-molekul lekatan dalam sel yang larut disukatkan oleh alat multipleks dan analisis sitometri. Penempatan F-aktin, rantai ringan miosin yang terfosforil, simpang adherens, simpang ketat dalam sel disiasatkan oleh teknik immunositokimia dan analisis mikroskop confocal. Analisis pemendapan Western digunakan untuk menentukan pengekspresan jumlah protein molekul-molekul lekatan dalam sel, rantai ringan miosin yang terfosforil, VE-cadherin, β -catenin, occludin, ZO-1 dan p-I κ B- α . Analisis pemendapan western juga digunakan untuk menentukan pengekspresan protein simpang adherens dan simpang ketat di bahagian membran, sitosol dan sitoskeleton. Asiatic asid mengurangkan hiperketelapan endotelial yang diaruhkan oleh TNF- α dengan nyata sekali, tetapi ia tidak dapat menahankan kelekatan dan penghijrahan monosit yang diaruhkan oleh TNF- α . Asiatic asid juga mencegah pengekspresan VCAM-1 dan penghasilan molekul-molekul lekatan dalam sel yang larut (sE-selectin, sICAM-1, sVCAM-1 and sPECAM-1). Selain itu, asiatic asid mencegah penaburan semula F-aktin yang diaruhkan oleh TNF- α , tetapi ia gagal untuk mengubah peningkatan nisbah F/G-aktin. Dengan menggunakan sitokalsin D, salah satu ejen yang menyebabkan aktin depolimerisasi, ia telah menunjukkan bahawa asiatic asid menstabilkan filamen-filamen F-aktin di pinggir sel. Akan tetapi, asiatic asid tidak mempertingkatkan hiperketelapan endotelial yang diaruhkan oleh sitokalsin D. Tambahan pula, asiatic asid mengekalkan filamen-filamen rantai ringan miosin yang terfosforil di pinggir sel dan menambahkan lagi dipemfosforilan rantai ringan miosin yang diaruhkan oleh TNF- α .

Asiatic acid juga mencegah gangguan simpang retikuler yang diaruhkan oleh TNF- α dengan menambahkan kawasan simpang yang diliputi oleh VE-cadherin dan β -catenin. Ia didapati bahawa kesan perlindungan ini tidak bergantung kepada perubahan jumlah keseluruhan dan penaburan semula VE-cadherin dan β -catenin intrasel. Bagi simpang ketat, pengimejan sefokus menunjukkan bahawa asiatic acid menentang kehilangan ZO-1 dari sempadan-sempadan sel yang diaruhkan oleh TNF- α . Walau bagaimanapun, asiatic acid tidak mengubah pengekspresan jumlah protein ZO-1 dan occludin dalam lisat keseluruhan sel. Pemingkatan protein subsel menunjukkan bahawa asiatic acid merencatkan penyebatan ZO-1 dari membran ke sitoplasma dan penaburan semula occludin dari sitoplasma ke sitoskeleton yang diaruhkan oleh TNF- α . Tambahan pula, asiatic acid menahankan pemfosforilan I κ B- α yang diaruhkan oleh TNF- α . Data ini mencadangkan bahawa asiatic acid melindungi endotelium daripada gangguan sekatan endotelial yang diaruhkan oleh TNF- α dan mengurangkan penghasilan molekul-molekul lekatan dalam sel yang larut, di mana molekul-molekul tersebut adalah penanda biologi yang penting untuk meramalkan risiko penyakit-penyakit kardiovaskular. Asiatic acid juga menstabilkan sitoskeleton dengan mengekalkan F-aktin dan filamen rantai ringan miosin yang terfosforil di sempadan sel. Akan tetapi, penstabilan aktin oleh asiatic acid adalah tidak mustahak bagi kesan perlindungan sekatan endotelial-nya. Ia didapati bahawa kesan penstabilan sekatan endotelial asiatic acid adalah seiring dengan penambahbaikan pembentukan simpang retikuler, perencatan penaburan semula simpang ketat and pengurangan pengaktifan NF- κ B. Kesimpulannya, satu peranan novel asiatic acid di mana ia melindungi endotelium daripada disfungsi sekatan endotelial yang diaruhkan oleh TNF- α telah ditunjukkan. Kajian ini mendedahkan penggunaan terapeutik asiatic acid yang baru dalam pencegahan peringkat awal aterosklerosis yang melibatkan pengaktifan endotelial.

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I certify that a Thesis Examination Committee has met on 23 November 2016 to conduct the final examination of Fong Lai Yen on her thesis entitled “*In Vitro* Anti-Atherogenic Effects of Asiatic Acid in Human Aortic Endothelial Cells” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

AJ	Adherens Junction
CAM	Cell Adhesion Molecule
diphospho-MLC	Diphosphorylated Myosin Light Chain
F-actin	Filamentous-Actin (Polymer Form)
G-actin	Globular-Actin (Monomer Form)
HAECs	Human Aortic Endothelial Cells
HUVECs	Human Umbilical Vein Endothelial Cells
ICAM-1	Intercellular Adhesion Molecule-1
JAM	Junctional Adhesion Molecule
MLC	Myosin Light Chain
NF- κ B	Nuclear Factor Kappa B
PECAM-1	Platelet Endothelial Cell Adhesion Molecule-1
p-I κ B- α	Phosphorylated Inhibitor Of κ b
sE-selectin	Soluble E-Selectin
sICAM-1	Soluble Intercellular Adhesion Molecule-1
sPECAM-1	Soluble Platelet Endothelial Cell Adhesion Molecule-1
sVCAM-1	Soluble Vascular Cell Adhesion Molecule-1
TJ	Tight Junction
TNF- α	Tumour Necrosis Factor-alpha
TTFCA	Total Triterpenic Fraction Of <i>Centella asiatica</i>
VCAM-1	Vascular Cell Adhesion Molecule-1
VE-cadherin	Vascular Endothelial-Cadherin
ZO-1	Zona Occludens-1

CHAPTER 1

INTRODUCTION

1.1 Justification of the Study

Statins are inhibitors of 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase and it serves as a primary approach used to prevent and treat atherosclerosis. However, long-term statin therapy causes various side effects including muscle weakness or pain, abnormal elevation of liver enzymes, acute renal failure and diabetes (Simic and Reiner, 2015; Alla *et al.*, 2013). Besides, patients undergoing statin therapy continue to experience recurrent cardiovascular events (Alla *et al.*, 2010). Owing to these adverse effects, the searching for novel anti-atherogenic compounds from plant-based sources could provide alternative therapeutic options for the treatment of cardiovascular diseases.

Despite that the anti-atherosclerotic effects of *Centella asiatica* (L.) Urban, a traditional medicinal herb, have been demonstrated in several clinical studies, the experimental studies intended to investigate the *in vitro* anti-atherosclerotic effect of *C. asiatica* remain widely lacking. Particularly, how each active constituent of *C. asiatica* would contribute to its anti-atherosclerotic effect and the mechanism of action remain unknown. Hence, detailed mechanistic study is needed in order to support the increasing use of *C. asiatica* as a potential anti-atherogenic agent.

1.2 Background of the Study

Ischaemic heart disease and cerebrovascular disease are the first and third leading causes of death, respectively, throughout the world. These cardiovascular diseases had a mortality rate of 247.9 deaths/100,000 persons, which accounts for 84.5 % of death from all types of cardiovascular diseases and 28.2% of global death in 2013 (Barquera *et al.*, 2015; Collaborators., 2015). As a result, many approaches have been adopted to reduce the occurrence and severity of cardiovascular diseases, which include lifestyle modifications, advancement of surgical operations and introduction of new therapeutic agents. Atherosclerosis is the major cause leading to cardiovascular diseases and it is closely related to elevated plasma concentration of low density lipoprotein-cholesterol (Ramji and Davies, 2015). Atherosclerosis is characterized by build-up of plaques or lesions at the arterial wall, which gradually narrows the lumen of the blood vessel.

It is now widely accepted that atherosclerosis is a chronic inflammatory disease initiated by 'activation' of endothelial cells, which promotes the switch of endothelium to a pro-inflammatory phenotype (Funk *et al.*, 2012). The key features of endothelial cell activation include an increase in endothelial permeability, up regulation of cell adhesion molecules (CAMs) and recruitment of monocytes to adhere to the endothelium. Subsequently, the adhered monocytes migrate through the endothelium and accumulate at the arterial intima to form lipid-engulfing foam cells. Foam cell accumulation features the beginning of atherosclerotic lesion formation (Ramji and

Davies, 2015). Tumour necrosis factor-alpha (TNF- α), a pro-inflammatory cytokine, was detected in human atherosclerotic plaques, implying the crucial role of TNF- α in the progression of atherosclerosis (Mehta and Malik, 2006). Therefore, targeting TNF- α -activated inflammatory pathways might be a promising therapeutic strategy for atherosclerosis.

In physiological conditions, the endothelial permeability is tightly regulated by interendothelial junctions that connect the neighbouring endothelial cells together. Inflammatory stimuli trigger intercellular junctions to disassemble and disrupts the endothelial barrier integrity (Capaldo and Nusrat, 2009). Consequently, an increase in permeability occurs where passage of solutes and fluid across the endothelium is increased. Atherogenesis is initiated when endothelial hyperpermeability permits transport of low density lipoproteins into the intima. Therefore, improvement of the endothelial barrier function may serve as a novel therapeutic approach to prevent early pre-lesional stage of atherosclerosis.

The cell-cell junctions are anchored by actin cytoskeleton in the cytoplasm where the latter modulates stability of the intercellular junctions. Actin cytoskeleton is a dynamic structure undergoes continuous remodelling to regulate cell contraction, cell shape and motility. Phosphorylation of myosin light chain (MLC) promotes actin cytoskeleton to rearrange into stress fibers that are capable to generate contractile force and pull the intercellular junctions inward, causing disassembly of the cell-cell junctions (Bogatcheva and Verin, 2008). CAMs, such as intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and platelet-endothelial cell adhesion molecule-1 (PECAM-1), also affect the formation of junctional complexes and are key determinants of the endothelial barrier integrity (Marcos-Ramiro *et al.*, 2014).

C. asiatica is a traditional medicinal herb that can be found in most tropical and subtropical countries. It is widely used in traditional Ayurvedic or Indian Medicine to enhance memory, heal wounds and alleviate various skin diseases such as leprosy and psoriasis (Gohil *et al.*, 2010). It is noteworthy to mention that *C. asiatica* has been shown to exhibit anti-atherosclerotic effects in several clinical studies (Belcaro *et al.*, 2015b; Belcaro *et al.*, 2014; Cesarone *et al.*, 2001a; Incandela *et al.*, 2001). Total triterpenic fraction of *C. asiatica* (TTFCA), a highly refined *C. asiatica* extract containing asiatic acid as a main component, has been shown to stabilize arterial plaques in atherosclerotic patients (Cesarone *et al.*, 2001a; Incandela *et al.*, 2001). Recently, researchers have reported that combined administration of a pine bark extract and TTFCA prevents the progression of atherosclerotic plaques in asymptomatic subjects and the anti-atherosclerotic effect is associated with reduction of oxidative stress (Belcaro *et al.*, 2015b; Belcaro *et al.*, 2014). Besides, *C. asiatica* has also been demonstrated to improve capillary permeability and prevent tissue edema in hypertensive patients (De Sanctis *et al.*, 2001; Belcaro *et al.*, 1990).

Asiatic acid is a major saponin triterpenoid derived from *C. asiatica*. It has been reported that asiatic acid possesses several biological activities including anti-inflammatory, wound healing, neuroprotective and hypolipidemic effects (Ramachandran *et al.*, 2014; Bian *et al.*, 2013; Huang *et al.*, 2011; Gohil *et al.*, 2010). Researchers have also shown that asiatic acid suppresses carrageenan-induced paw edema in mice (Huang *et al.*, 2011). However, the anti-atherogenic effect of asiatic acid *in vitro*, particularly in the context of endothelial activation has not been reported yet.

1.3 Hypotheses

Asiatic acid will possess protective effects against TNF- α -induced endothelial activation. Asiatic acid will inhibit TNF- α -induced disruption of endothelial barrier integrity and the increased interaction between leukocytes and endothelial cells. Asiatic acid will also inhibit TNF- α -induced increased expression of CAMs and cytoskeletal rearrangement. Besides, asiatic acid will also prevent TNF- α -induced disruption of cell-cell junctions. In addition, asiatic acid will suppress activation of signaling cascades stimulated by TNF- α .

1.4 Objectives

1.4.1 General Objective

The aim of this study was to evaluate effects of asiatic acid on TNF- α -induced activation of human aortic endothelial cells and the underlying mechanisms.

1.4.2 Specific Objectives

This study aimed to investigate effects of asiatic acid on TNF- α -induced

- (i) endothelial hyperpermeability and increased leukocyte-endothelial cell interactions
- (ii) increased expression of CAMs
- (iii) cytoskeletal rearrangement
- (iv) disassembly of interendothelial junctions
- (v) activation of signaling pathway

REFERENCES

- Aggarwal, B. B., Prasad, S., Reuter, S., Kannappan, R., Yadev, V. R., Park, B., Kim, J. H., Gupta, S. C., Phromnoi, K., Sundaram, C., Prasad, S., Chaturvedi, M. M. and Sung, B. (2011). Identification of novel anti-inflammatory agents from Ayurvedic medicine for prevention of chronic diseases: "reverse pharmacology" and "bedside to bench" approach. *Curr Drug Targets*, 12(11): 1595-1653.
- Ahn, K. S., Sethi, G. and Aggarwal, B. B. (2007). Simvastatin potentiates TNF-alpha-induced apoptosis through the down-regulation of NF-kappaB-dependent antiapoptotic gene products: role of IkappaBalpha kinase and TGF-beta-activated kinase-1. *J Immunol*, 178(4): 2507-2516.
- Aird, W. C. (2005). Spatial and temporal dynamics of the endothelium. *J Thromb Haemost*, 3(7): 1392-1406.
- Aird, W. C. (2007a). Phenotypic heterogeneity of the endothelium: I. Structure, function, and mechanisms. *Circ Res*, 100(2): 158-173.
- Aird, W. C. (2007b). Phenotypic heterogeneity of the endothelium: II. Representative vascular beds. *Circ Res*, 100(2): 174-190.
- Alla, V. M., Agrawal, V., DeNazareth, A., Mohiuddin, S., Ravilla, S. and Rendell, M. (2013). A reappraisal of the risks and benefits of treating to target with cholesterol lowering drugs. *Drugs*, 73(10): 1025-1054.
- Alla, V. M., Kaushik, M. and Mooss, A. (2010). Targeting residual risk: the rationale for the use of non-HDL cholesterol. *South Med J*, 103(5): 434-437; quiz 438-439.
- Allingham, M. J., van Buul, J. D. and Burrige, K. (2007). ICAM-1-mediated, Src- and Pyk2-dependent vascular endothelial cadherin tyrosine phosphorylation is required for leukocyte transendothelial migration. *J Immunol*, 179(6): 4053-4064.
- Almutairi, M. M., Gong, C., Xu, Y. G., Chang, Y. and Shi, H. (2016). Factors controlling permeability of the blood-brain barrier. *Cell Mol Life Sci*, 73(1): 57-77.
- Ang, K. P., Tan, H. K., Selvaraja, M., Kadir, A. A., Somchit, M. N., Akim, A. M., Zakaria, Z. A. and Ahmad, Z. (2011). Cryptotanshinone attenuates in vitro oxLDL-induced pre-lesional atherosclerotic events. *Planta Med*, 77(16): 1782-1787.
- Antonopoulos, A. S., Margaritis, M., Lee, R., Channon, K. and Antoniades, C. (2012). Statins as anti-inflammatory agents in atherogenesis: molecular mechanisms and lessons from the recent clinical trials. *Curr Pharm Des*, 18(11): 1519-1530.

- Aranda, J. F., Reglero-Real, N., Marcos-Ramiro, B., Ruiz-Saenz, A., Fernandez-Martin, L., Bernabe-Rubio, M., Kremer, L., Ridley, A. J., Correas, I., Alonso, M. A. and Millan, J. (2013). MYADM controls endothelial barrier function through ERM-dependent regulation of ICAM-1 expression. *Mol Biol Cell*, 24(4): 483-494.
- Aveleira, C. A., Lin, C. M., Abcouwer, S. F., Ambrosio, A. F. and Antonetti, D. A. (2010). TNF-alpha signals through PKCzeta/NF-kappaB to alter the tight junction complex and increase retinal endothelial cell permeability. *Diabetes*, 59(11): 2872-2882.
- Bachok, M. F., Yusof, B. N., Ismail, A. and Hamid, A. A. (2014). Effectiveness of traditional Malaysian vegetables (ulam) in modulating blood glucose levels. *Asia Pac J Clin Nutr*, 23(3): 369-376.
- Barquera, S., Pedroza-Tobias, A., Medina, C., Hernandez-Barrera, L., Bibbins-Domingo, K., Lozano, R. and Moran, A. E. (2015). Global overview of the epidemiology of atherosclerotic cardiovascular disease. *Arch Med Res*, 46(5): 328-338.
- Barreiro, O., Yanez-Mo, M., Serrador, J. M., Montoya, M. C., Vicente-Manzanares, M., Tejedor, R., Furthmayr, H. and Sanchez-Madrid, F. (2002). Dynamic interaction of VCAM-1 and ICAM-1 with moesin and ezrin in a novel endothelial docking structure for adherent leukocytes. *J Cell Biol*, 157(7): 1233-1245.
- Bauer, H.-C., Krizbai, I. A., Bauer, H. and Traweger, A. (2014). "You Shall Not Pass" — tight junctions of the blood brain barrier. *Front Neurosci*, 8: 392.
- Baumer, Y., Drenckhahn, D. and Waschke, J. (2008). cAMP induced Rac 1-mediated cytoskeletal reorganization in microvascular endothelium. *Histochem Cell Biol*, 129(6): 765-778.
- Belcaro, G., Dugall, M., Hosoi, M., Ippolito, E., Cesarone, M., Luzzi, R., Cornelli, U. and Ledda, A. (2014). Pycnogenol(R) and *Centella asiatica* for asymptomatic atherosclerosis progression. *Int Angiol*, 33(1): 20-26.
- Belcaro, G., Dugall, M., Ippolito, E., Hosoi, M., Cornelli, U., Ledda, A., Scoccianti, M., Cesarone, M. R., Pellegrini, L., Luzzi, R. and Corsi, M. (2015a). Pycnogenol(R) and *Centella asiatica* for preventing asymptomatic atherosclerosis progression into clinical events. *Minerva Cardioangiol*.
- Belcaro, G., Ippolito, E., Dugall, M., Hosoi, M., Cornelli, U., Ledda, A., Scoccianti, M., Steigerwalt, R. D., Cesarone, M. R., Pellegrini, L., Luzzi, R. and Corsi, M. (2015b). Pycnogenol(R) and *Centella asiatica* in the management of asymptomatic atherosclerosis progression. *Int Angiol*, 34(2): 150-157.
- Belcaro, G. V., Grimaldi, R. and Guidi, G. (1990). Improvement of capillary permeability in patients with venous hypertension after treatment with TTFCA. *Angiology*, 41(7): 533-540.

- Belvitch, P. and Dudek, S. M. (2012). Role of FAK in S1P-regulated endothelial permeability. *Microvasc Res*, 83(1): 22-30.
- Benz, P. M., Blume, C., Moebius, J., Oschatz, C., Schuh, K., Sickmann, A., Walter, U., Feller, S. M. and Renne, T. (2008). Cytoskeleton assembly at endothelial cell-cell contacts is regulated by alphaII-spectrin-VASP complexes. *J Cell Biol*, 180(1): 205-219.
- Bian, D., Zhang, J., Wu, X., Dou, Y., Yang, Y., Tan, Q., Xia, Y., Gong, Z. and Dai, Y. (2013). Asiatic acid isolated from *Centella asiatica* inhibits TGF-beta1-induced collagen expression in human keloid fibroblasts via PPAR-gamma activation. *Int J Biol Sci*, 9(10): 1032-1042.
- Birukova, A. A., Birukov, K. G., Smurova, K., Adyshev, D., Kaibuchi, K., Alieva, I., Garcia, J. G. and Verin, A. D. (2004). Novel role of microtubules in thrombin-induced endothelial barrier dysfunction. *FASEB J*, 18(15): 1879-1890.
- Birukova, A. A., Malyukova, I., Poroyko, V. and Birukov, K. G. (2007). Paxillin-beta-catenin interactions are involved in Rac/Cdc42-mediated endothelial barrier-protective response to oxidized phospholipids. *Am J Physiol Lung Cell Mol Physiol*, 293(1): L199-211.
- Birukova, A. A., Zebda, N., Fu, P., Poroyko, V., Cokic, I. and Birukov, K. G. (2011). Association between adherens junctions and tight junctions via Rap1 promotes barrier protective effects of oxidized phospholipids. *J Cell Physiol*, 226(8): 2052-2062.
- Blankenberg, S., Rupprecht, H. J., Bickel, C., Peetz, D., Hafner, G., Tiret, L. and Meyer, J. (2001). Circulating cell adhesion molecules and death in patients with coronary artery disease. *Circulation*, 104(12): 1336-1342.
- Bogatcheva, N. V. and Verin, A. D. (2008). The role of cytoskeleton in the regulation of vascular endothelial barrier function. *Microvasc Res*, 76(3): 202-207.
- Bogatcheva, N. V., Zemskova, M. A., Poirier, C., Mirzapiozova, T., Kolosova, I., Bresnick, A. R. and Verin, A. D. (2011). The suppression of myosin light chain (MLC) phosphorylation during the response to lipopolysaccharide (LPS): beneficial or detrimental to endothelial barrier? *J Cell Physiol*, 226(12): 3132-3146.
- Borhan, M. Z., Ahmad, R., Rusop, M. and Abdullah, S. (2013). Green extraction: enhanced extraction yield of asiatic acid from *Centella asiatica* (L.) nanopowders. *J Appl Chem*, 2013: 7.
- Bradley, J. R. (2008). TNF-mediated inflammatory disease. *J Pathol*, 214(2): 149-160.
- Brasier, A. R. (2010). The nuclear factor-kappaB-interleukin-6 signalling pathway mediating vascular inflammation. *Cardiovasc Res*, 86(2): 211-218.

- Breslin, J. W., Sun, H., Xu, W., Rodarte, C., Moy, A. B., Wu, M. H. and Yuan, S. Y. (2006). Involvement of ROCK-mediated endothelial tension development in neutrophil-stimulated microvascular leakage. *Am J Physiol Heart Circ Physiol*, 290(2): H741-750.
- Brinkhaus, B., Lindner, M., Schuppan, D. and Hahn, E. G. (2000). Chemical, pharmacological and clinical profile of the East Asian medical plant *Centella asiatica*. *Phytomedicine*, 7(5): 427-448.
- Bubik, M. F., Willer, E. A., Bihari, P., Jurgenliemk, G., Ammer, H., Krombach, F., Zahler, S., Vollmar, A. M. and Furst, R. (2012). A novel approach to prevent endothelial hyperpermeability: the *Crataegus* extract WS(R) 1442 targets the cAMP/Rap1 pathway. *J Mol Cell Cardiol*, 52(1): 196-205.
- Bylka, W., Znajdek-Awizen, P., Studzinska-Sroka, E., Danczak-Pazdrowska, A. and Brzezinska, M. (2014). *Centella asiatica* in dermatology: an overview. *Phytother Res*, 28(8): 1117-1124.
- Caballero-George, C., Vanderheyden, P. M., Okamoto, Y., Masaki, T., Mbwambo, Z., Apers, S., Gupta, M. P., Pieters, L., Vauquelin, G. and Vlietinck, A. (2004). Evaluation of bioactive saponins and triterpenoidal aglycons for their binding properties on human endothelin ETA and angiotensin AT1 receptors. *Phytother Res*, 18(9): 729-736.
- Campos, S. B., Ashworth, S. L., Wean, S., Hosford, M., Sandoval, R. M., Hallett, M. A., Atkinson, S. J. and Molitoris, B. A. (2009). Cytokine-induced F-actin reorganization in endothelial cells involves RhoA activation. *Am J Physiol Renal Physiol*, 296(3): F487-495.
- Capaldo, C. T. and Nusrat, A. (2009). Cytokine regulation of tight junctions. *Biochim Biophys Acta*, 1788(4): 864-871.
- Carlier, M. F. and Pantaloni, D. (1997). Control of actin dynamics in cell motility. *J Mol Biol*, 269(4): 459-467.
- Carswell, E. A., Old, L. J., Kassel, R. L., Green, S., Fiore, N. and Williamson, B. (1975). An endotoxin-induced serum factor that causes necrosis of tumors. *Proc Natl Acad Sci U S A*, 72(9): 3666-3670.
- Cesarone, M. R., Belcaro, G., Nicolaidis, A. N., Geroulakos, G., Bucci, M., Dugall, M., De Sanctis, M. T., Incandela, L., Griffin, M. and Sabetai, M. (2001a). Increase in echogenicity of echolucent carotid plaques after treatment with total triterpenic fraction of *Centella asiatica*: a prospective, placebo-controlled, randomized trial. *Angiology*, 52 Suppl 2: S19-25.
- Cesarone, M. R., Incandela, L., De Sanctis, M. T., Belcaro, G., Bavera, P., Bucci, M. and Ippolito, E. (2001b). Evaluation of treatment of diabetic microangiopathy with total triterpenic fraction of *Centella asiatica*: a clinical prospective randomized trial with a microcirculatory model. *Angiology*, 52 Suppl 2: S49-54.

- Cheung, T. M., Ganatra, M. P., Peters, E. B. and Truskey, G. A. (2012). Effect of cellular senescence on the albumin permeability of blood-derived endothelial cells. *Am J Physiol Heart Circ Physiol*, 303(11): H1374-1383.
- Chong, N. J. and Aziz, Z. (2013). A systematic review of the efficacy of *Centella asiatica* for improvement of the signs and symptoms of chronic venous insufficiency. *Evid Based Complement Alternat Med*, 2013: 627182.
- Cines, D. B., Pollak, E. S., Buck, C. A., Loscalzo, J., Zimmerman, G. A., McEver, R. P., Pober, J. S., Wick, T. M., Konkle, B. A., Schwartz, B. S., Barnathan, E. S., McCrae, K. R., Hug, B. A., Schmidt, A. M. and Stern, D. M. (1998). Endothelial cells in physiology and in the pathophysiology of vascular disorders. *Blood*, 91(10): 3527-3561.
- Clark, P. R., Manes, T. D., Pober, J. S. and Kluger, M. S. (2007). Increased ICAM-1 expression causes endothelial cell leakiness, cytoskeletal reorganization and junctional alterations. *J Invest Dermatol*, 127(4): 762-774.
- Collaborators., G. M. a. C. o. D. (2015). Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*, 385(9963): 117-171.
- Constans, J. and Conri, C. (2006). Circulating markers of endothelial function in cardiovascular disease. *Clin Chim Acta*, 368(1-2): 33-47.
- Cooper, J. A. (1987). Effects of cytochalasin and phalloidin on actin. *J Cell Biol*, 105(4): 1473-1478.
- De Sanctis, M. T., Belcaro, G., Incandela, L., Cesarone, M. R., Griffin, M., Ippolito, E. and Cacchio, M. (2001). Treatment of edema and increased capillary filtration in venous hypertension with total triterpenic fraction of *Centella asiatica*: a clinical, prospective, placebo-controlled, randomized, dose-ranging trial. *Angiology*, 52 Suppl 2: S55-59.
- Dejana, E. (2010). The role of wnt signaling in physiological and pathological angiogenesis. *Circ Res*, 107(8): 943-952.
- Dejana, E. and Orsenigo, F. (2013). Endothelial adherens junctions at a glance. *J Cell Sci*, 126(Pt 12): 2545-2549.
- Dejana, E., Orsenigo, F. and Lampugnani, M. G. (2008). The role of adherens junctions and VE-cadherin in the control of vascular permeability. *J Cell Sci*, 121(Pt 13): 2115-2122.
- Dejana, E., Tournier-Lasserre, E. and Weinstein, B. M. (2009). The control of vascular integrity by endothelial cell junctions: molecular basis and pathological implications. *Dev Cell*, 16(2): 209-221.
- Di Tomo, P., Di Silvestre, S., Cordone, V. G., Giardinelli, A., Faricelli, B., Pipino, C., Lanuti, P., Peng, T., Formoso, G., Yang, D., Arduini, A., Chiarelli, F.,

- Pandolfi, A. and Di Pietro, N. (2015). *Centella asiatica* and lipoic acid, or a combination thereof, inhibit monocyte adhesion to endothelial cells from umbilical cords of gestational diabetic women. *Nutr Metab Cardiovasc Dis*, 25(7): 659-666.
- Dimitrova, Y., Dunoyer-Geindre, S., Reber, G., Mach, F., Kruithof, E. K. and de Moerloose, P. (2003). Effects of statins on adhesion molecule expression in endothelial cells. *J Thromb Haemost*, 1(11): 2290-2299.
- Dudek, S. M., Jacobson, J. R., Chiang, E. T., Birukov, K. G., Wang, P., Zhan, X. and Garcia, J. G. (2004). Pulmonary endothelial cell barrier enhancement by sphingosine 1-phosphate: roles for cortactin and myosin light chain kinase. *J Biol Chem*, 279(23): 24692-24700.
- Eiselein, L., Wilson, D. W., Lame, M. W. and Rutledge, J. C. (2007). Lipolysis products from triglyceride-rich lipoproteins increase endothelial permeability, perturb zonula occludens-1 and F-actin, and induce apoptosis. *Am J Physiol Heart Circ Physiol*, 292(6): H2745-2753.
- Endres, M., Laufs, U., Merz, H. and Kaps, M. (1997). Focal expression of intercellular adhesion molecule-1 in the human carotid bifurcation. *Stroke*, 28(1): 77-82.
- Enlimomab Acute Stroke Trial, I. (2001). Use of anti-ICAM-1 therapy in ischemic stroke: results of the Enlimomab Acute Stroke Trial. *Neurology*, 57(8): 1428-1434.
- Eriksson, E. E., Karlof, E., Lundmark, K., Rotzius, P., Hedin, U. and Xie, X. (2005). Powerful inflammatory properties of large vein endothelium in vivo. *Arterioscler Thromb Vasc Biol*, 25(4): 723-728.
- Falati, S., Patil, S., Gross, P. L., Stapleton, M., Merrill-Skoloff, G., Barrett, N. E., Pixton, K. L., Weiler, H., Cooley, B., Newman, D. K., Newman, P. J., Furie, B. C., Furie, B. and Gibbins, J. M. (2006). Platelet PECAM-1 inhibits thrombus formation in vivo. *Blood*, 107(2): 535-541.
- Favero, G., Paganelli, C., Buffoli, B., Rodella, L. F. and Rezzani, R. (2014). Endothelium and its alterations in cardiovascular diseases: life style intervention. *Biomed Res Int*, 2014: 801896.
- Fá áou, M. (2011). Chapter 2: Multiple Functions of the Endothelial Cells. In *The Endothelium: Part 1: Multiple Functions of the Endothelial Cells—Focus on Endothelium-Derived Vasoactive Mediators*. San Rafael (CA): Morgan & Claypool Life Sciences.
- Feng, D., Nagy, J. A., Hipp, J., Dvorak, H. F. and Dvorak, A. M. (1996). Vesiculo-vacuolar organelles and the regulation of venule permeability to macromolecules by vascular permeability factor, histamine, and serotonin. *J Exp Med*, 183(5): 1981-1986.

- Fernandez-Borja, M., van Buul, J. D. and Hordijk, P. L. (2010). The regulation of leucocyte transendothelial migration by endothelial signalling events. *Cardiovasc Res*, 86(2): 202-210.
- Fernandez-Martin, L., Marcos-Ramiro, B., Bigarella, C. L., Graupera, M., Cain, R. J., Reglero-Real, N., Jimenez, A., Cernuda-Morollon, E., Correias, I., Cox, S., Ridley, A. J. and Millan, J. (2012). Crosstalk between reticular adherens junctions and platelet endothelial cell adhesion molecule-1 regulates endothelial barrier function. *Arterioscler Thromb Vasc Biol*, 32(8): e90-102.
- Fong, L. Y., Ng, C. T., Zakaria, Z. A., Baharuldin, M. T., Arifah, A. K., Hakim, M. N. and Zuraini, A. (2015). Asiaticoside inhibits TNF-alpha-induced endothelial hyperpermeability of human aortic endothelial cells. *Phytother Res*, 29(10): 1501-1508.
- Fotis, L., Giannakopoulos, D., Stamogiannou, L. and Xatzipsalti, M. (2012). Intercellular cell adhesion molecule-1 and vascular cell adhesion molecule-1 in children. Do they play a role in the progression of atherosclerosis? *Hormones (Athens)*, 11(2): 140-146.
- Funk, S. D., Yurdagul, A., Jr. and Orr, A. W. (2012). Hyperglycemia and endothelial dysfunction in atherosclerosis: lessons from type 1 diabetes. *Int J Vasc Med*, 2012: 569654.
- Gagat, M., Grzanka, D., Izdebska, M. and Grzanka, A. (2013). Effect of L-homocysteine on endothelial cell-cell junctions following F-actin stabilization through tropomyosin-1 overexpression. *Int J Mol Med*, 32(1): 115-129.
- Gagat, M., Grzanka, D., Izdebska, M., Sroka, W. D., Marszall, M. P. and Grzanka, A. (2014). Tropomyosin-1 protects endothelial cell-cell junctions against cigarette smoke extract through F-actin stabilization in EA.hy926 cell line. *Acta Histochem*, 116(4): 606-618.
- Garcia, J. G., Liu, F., Verin, A. D., Birukova, A., Dechert, M. A., Gerthoffer, W. T., Bamberg, J. R. and English, D. (2001). Sphingosine 1-phosphate promotes endothelial cell barrier integrity by Edg-dependent cytoskeletal rearrangement. *J Clin Invest*, 108(5): 689-701.
- Gareus, R., Kotsaki, E., Xanthoulea, S., van der Made, I., Gijbels, M. J., Kardakaris, R., Polykratis, A., Kollias, G., de Winther, M. P. and Pasparakis, M. (2008). Endothelial cell-specific NF-kappaB inhibition protects mice from atherosclerosis. *Cell Metab*, 8(5): 372-383.
- Gavard, J. (2013). Endothelial permeability and VE-cadherin: a wacky comradeship. *Cell Adh Migr*, 7(6): 455-461.
- Gershenzon, J. and Kreis, W. (1999). *Biochemistry of Plant Secondary Metabolites. Annual Plant Reviews, Wink, M., Ed (Vol. 2)*. Sheffield, UK: Sheffield Academic Press.

- Giannotta, M., Trani, M. and Dejana, E. (2013). VE-cadherin and endothelial adherens junctions: active guardians of vascular integrity. *Dev Cell*, 26(5): 441-454.
- Gimbrone, M. A. (2010). The Gordon Wilson lecture. Understanding vascular endothelium: a pilgrim's progress. Endothelial dysfunction, biomechanical forces and the pathobiology of atherosclerosis. *Trans Am Clin Climatol Assoc*, 121: 115-127; discussion 127.
- Gimbrone, M. A., Jr. and Garcia-Cardena, G. (2013). Vascular endothelium, hemodynamics, and the pathobiology of atherosclerosis. *Cardiovasc Pathol*, 22(1): 9-15.
- Gohil, K. J., Patel, J. A. and Gajjar, A. K. (2010). Pharmacological review on *Centella asiatica*: a potential herbal cure-all. *Indian J Pharm Sci*, 72(5): 546-556.
- Goldblum, S. E., Ding, X. and Campbell-Washington, J. (1993). TNF-alpha induces endothelial cell F-actin depolymerization, new actin synthesis, and barrier dysfunction. *Am J Physiol*, 264(4 Pt 1): C894-905.
- Golias, C., Tsoutsis, E., Matziridis, A., Makridis, P., Batistatou, A. and Charalabopoulos, K. (2007). Review. Leukocyte and endothelial cell adhesion molecules in inflammation focusing on inflammatory heart disease. *In Vivo*, 21(5): 757-769.
- Gonzalez-Mariscal, L., Tapia, R. and Chamorro, D. (2008). Crosstalk of tight junction components with signaling pathways. *Biochim Biophys Acta*, 1778(3): 729-756.
- Greenwood, J. and Mason, J. C. (2007). Statins and the vascular endothelial inflammatory response. *Trends Immunol*, 28(2): 88-98.
- Grimaldi, R., De Ponti, F., D'Angelo, L., Caravaggi, M., Guidi, G., Lecchini, S., Frigo, G. M. and Crema, A. (1990). Pharmacokinetics of the total triterpenic fraction of *Centella asiatica* after single and multiple administrations to healthy volunteers. A new assay for asiatic acid. *J Ethnopharmacol*, 28(2): 235-241.
- Guo, W., Liu, W., Jin, B., Geng, J., Li, J., Ding, H., Wu, X., Xu, Q., Sun, Y. and Gao, J. (2015). Asiatic acid ameliorates dextran sulfate sodium-induced murine experimental colitis via suppressing mitochondria-mediated NLRP3 inflammasome activation. *Int Immunopharmacol*, 24(2): 232-238.
- Guray, U., Erbay, A. R., Guray, Y., Yilmaz, M. B., Boyaci, A. A., Sasmaz, H., Korkmaz, S. and Kutuk, E. (2004). Levels of soluble adhesion molecules in various clinical presentations of coronary atherosclerosis. *Int J Cardiol*, 96(2): 235-240.
- Haarmann, A., Nowak, E., Deiss, A., van der Pol, S., Monoranu, C. M., Kooij, G., Muller, N., van der Valk, P., Stoll, G., de Vries, H. E., Berberich-Siebelt, F. and Buttman, M. (2015). Soluble VCAM-1 impairs human brain endothelial barrier integrity via integrin alpha-4-transduced outside-in signalling. *Acta Neuropathol*, 129(5): 639-652.

- Hashim, P. (2011). *Centella asiatica* in food and beverage applications and its potential antioxidant and neuroprotective effect. *International Food Research Journal*, 18(4): 1215-1222.
- He, P. (2010). Leucocyte/endothelium interactions and microvessel permeability: coupled or uncoupled? *Cardiovasc Res*, 87(2): 281-290.
- Heo, K. S., Fujiwara, K. and Abe, J. (2014). Shear stress and atherosclerosis. *Mol Cells*, 37(6): 435-440.
- Hernandez, L. A., Grisham, M. B., Twohig, B., Arfors, K. E., Harlan, J. M. and Granger, D. N. (1987). Role of neutrophils in ischemia-reperfusion-induced microvascular injury. *Am J Physiol*, 253(3 Pt 2): H699-703.
- Hicks, A. E., Nolan, S. L., Ridger, V. C., Hellewell, P. G. and Norman, K. E. (2003). Recombinant P-selectin glycoprotein ligand-1 directly inhibits leukocyte rolling by all 3 selectins in vivo: complete inhibition of rolling is not required for anti-inflammatory effect. *Blood*, 101(8): 3249-3256.
- Hirano, M. and Hirano, K. (2016). Myosin di-phosphorylation and peripheral actin bundle formation as initial events during endothelial barrier disruption. *Sci Rep*, 6: 20989.
- Hsu, Y. L., Kuo, P. L., Lin, L. T. and Lin, C. C. (2005). Asiatic acid, a triterpene, induces apoptosis and cell cycle arrest through activation of extracellular signal-regulated kinase and p38 mitogen-activated protein kinase pathways in human breast cancer cells. *J Pharmacol Exp Ther*, 313(1): 333-344.
- Hu, J., Zhang, Z., Xie, H., Chen, L., Zhou, Y., Chen, W. and Liu, Z. (2013). Serine protease inhibitor A3K protects rabbit corneal endothelium from barrier function disruption induced by TNF-alpha. *Invest Ophthalmol Vis Sci*, 54(8): 5400-5407.
- Huang, S. S., Chiu, C. S., Chen, H. J., Hou, W. C., Sheu, M. J., Lin, Y. C., Shie, P. H. and Huang, G. J. (2011). Antinociceptive activities and the mechanisms of anti-inflammation of asiatic acid in mice. *Evid Based Complement Alternat Med*, 2011: 895857.
- Hulok, A., Sciborski, K., Marczak, J., Bankowski, T., Poreba, R. and Negrusz-Kawecka, M. (2014). Soluble cell adhesion molecules - Does estimating sVCAM-1 and sICAM-1 concentration provide additional information about cardiovascular risk in patients with coronary artery disease? *Adv Clin Exp Med*, 23(5): 735-741.
- Huveneers, S., Oldenburg, J., Spanjaard, E., van der Krogt, G., Grigoriev, I., Akhmanova, A., Rehmann, H. and de Rooij, J. (2012). Vinculin associates with endothelial VE-cadherin junctions to control force-dependent remodeling. *J Cell Biol*, 196(5): 641-652.

- Hyde, M. A., Wursten, B.T., Ballings, P. & Coates Palgrave, M. . (2016). Flora of Zimbabwe: Species information: individual images: *Centella asiatica*. http://www.zimbabweflora.co.zw/speciesdata/image-display.php?species_id=142980&image_id=6 Retrieved 7 March 2016
- Idris, I. and Donnelly, R. (2006). Protein kinase C beta inhibition: A novel therapeutic strategy for diabetic microangiopathy. *Diab Vasc Dis Res*, 3(3): 172-178.
- Incandela, L., Belcaro, G., Nicolaides, A. N., Cesarone, M. R., De Sanctis, M. T., Corsi, M., Bavera, P., Ippolito, E., Griffin, M., Geroulakos, G., Sabetai, M., Ramaswami, G. and Veller, M. (2001). Modification of the echogenicity of femoral plaques after treatment with total triterpenic fraction of *Centella asiatica*: a prospective, randomized, placebo-controlled trial. *Angiology*, 52 Suppl 2: S69-73.
- Ivanov, V., Ivanova, S., Kalinovskiy, T., Niedzwiecki, A. and Rath, M. (2008). Plant-derived micronutrients suppress monocyte adhesion to cultured human aortic endothelial cell layer by modulating its extracellular matrix composition. *J Cardiovasc Pharmacol*, 52(1): 55-65.
- James, J. T. and Dubery, I. A. (2009). Pentacyclic triterpenoids from the medicinal herb, *Centella asiatica* (L.) Urban. *Molecules*, 14(10): 3922-3941.
- Jubeli, E., Moine, L., Vergnaud-Gauduchon, J. and Barratt, G. (2012). E-selectin as a target for drug delivery and molecular imaging. *J Control Release*, 158(2): 194-206.
- Katsuno, T., Umeda, K., Matsui, T., Hata, M., Tamura, A., Itoh, M., Takeuchi, K., Fujimori, T., Nabeshima, Y., Noda, T., Tsukita, S. and Tsukita, S. (2008). Deficiency of zonula occludens-1 causes embryonic lethal phenotype associated with defected yolk sac angiogenesis and apoptosis of embryonic cells. *Mol Biol Cell*, 19(6): 2465-2475.
- Kavitha, C. V., Agarwal, C., Agarwal, R. and Deep, G. (2011). Asiatic acid inhibits pro-angiogenic effects of VEGF and human gliomas in endothelial cell culture models. *PLoS One*, 6(8): e22745.
- Kim, M. H., Curry, F. R. and Simon, S. I. (2009). Dynamics of neutrophil extravasation and vascular permeability are uncoupled during aseptic cutaneous wounding. *Am J Physiol Cell Physiol*, 296(4): C848-856.
- Kjaergaard, A. G., Dige, A., Krog, J., Tonnesen, E. and Wogensen, L. (2013). Soluble adhesion molecules correlate with surface expression in an *in vitro* model of endothelial activation. *Basic Clin Pharmacol Toxicol*, 113(4): 273-279.
- Knezevic, II, Predescu, S. A., Neamu, R. F., Gorovoy, M. S., Knezevic, N. M., Easington, C., Malik, A. B. and Predescu, D. N. (2009). Tiam1 and Rac1 are required for platelet-activating factor-induced endothelial junctional disassembly and increase in vascular permeability. *J Biol Chem*, 284(8): 5381-5394.

- Komarova, Y. and Malik, A. B. (2010). Regulation of endothelial permeability via paracellular and transcellular transport pathways. *Annu Rev Physiol*, 72: 463-493.
- Koss, M., Pfeiffer, G. R., 2nd, Wang, Y., Thomas, S. T., Yerukhimovich, M., Gaarde, W. A., Doerschuk, C. M. and Wang, Q. (2006). Ezrin/radixin/moesin proteins are phosphorylated by TNF-alpha and modulate permeability increases in human pulmonary microvascular endothelial cells. *J Immunol*, 176(2): 1218-1227.
- Kumar, P., Shen, Q., Pivetti, C. D., Lee, E. S., Wu, M. H. and Yuan, S. Y. (2009). Molecular mechanisms of endothelial hyperpermeability: implications in inflammation. *Expert Rev Mol Med*, 11: e19.
- Langhauser, F., Kraft, P., Gob, E., Leinweber, J., Schuhmann, M. K., Lorenz, K., Gelderblom, M., Bittner, S., Meuth, S. G., Wiendl, H., Magnus, T. and Kleinschnitz, C. (2014). Blocking of alpha4 integrin does not protect from acute ischemic stroke in mice. *Stroke*, 45(6): 1799-1806.
- Laviola, L., Orlando, M. R., Incalza, M. A., Caccioppoli, C., Melchiorre, M., Leonardini, A., Cignarelli, A., Tortosa, F., Labarbuta, R., Martemucci, S., Pacelli, C., Cocco, T., Perrini, S., Natalicchio, A. and Giorgino, F. (2013). TNFalpha signals via p66(Shc) to induce E-Selectin, promote leukocyte transmigration and enhance permeability in human endothelial cells. *PLoS One*, 8(12): e81930.
- Lawson, C. and Wolf, S. (2009). ICAM-1 signaling in endothelial cells. *Pharmacol Rep*, 61(1): 22-32.
- Lee, J. F., Zeng, Q., Ozaki, H., Wang, L., Hand, A. R., Hla, T., Wang, E. and Lee, M. J. (2006). Dual roles of tight junction-associated protein, zonula occludens-1, in sphingosine 1-phosphate-mediated endothelial chemotaxis and barrier integrity. *J Biol Chem*, 281(39): 29190-29200.
- Li, A. C. and Glass, C. K. (2002). The macrophage foam cell as a target for therapeutic intervention. *Nat Med*, 8(11): 1235-1242.
- Libby, P., Ridker, P. M. and Hansson, G. K. (2011). Progress and challenges in translating the biology of atherosclerosis. *Nature*, 473(7347): 317-325.
- Liu, J., Chow, V. T. and Jois, S. D. (2004). A novel, rapid and sensitive heterotypic cell adhesion assay for CD2-CD58 interaction, and its application for testing inhibitory peptides. *J Immunol Methods*, 291(1-2): 39-49.
- Liu, Z., Tan, J. L., Cohen, D. M., Yang, M. T., Sniadecki, N. J., Ruiz, S. A., Nelson, C. M. and Chen, C. S. (2010). Mechanical tugging force regulates the size of cell-cell junctions. *Proc Natl Acad Sci U S A*, 107(22): 9944-9949.
- Luissint, A. C., Artus, C., Glacial, F., Ganeshamoorthy, K. and Couraud, P. O. (2012). Tight junctions at the blood brain barrier: physiological architecture and disease-associated dysregulation. *Fluids Barriers CNS*, 9(1): 23.

- Ma, T. Y., Iwamoto, G. K., Hoa, N. T., Akotia, V., Pedram, A., Boivin, M. A. and Said, H. M. (2004). TNF-alpha-induced increase in intestinal epithelial tight junction permeability requires NF-kappa B activation. *Am J Physiol Gastrointest Liver Physiol*, 286(3): G367-376.
- Macias, C., Villaescusa, R., del Valle, L., Boffil, V., Cordero, G., Hernandez, A., Hernandez, P. and Ballester, J. M. (2003). [Endothelial adhesion molecules ICAM-1, VCAM-1 and E-selectin in patients with acute coronary syndrome]. *Rev Esp Cardiol*, 56(2): 137-144.
- Mamdouh, Z., Mikhailov, A. and Muller, W. A. (2009). Transcellular migration of leukocytes is mediated by the endothelial lateral border recycling compartment. *J Exp Med*, 206(12): 2795-2808.
- Marchiando, A. M., Shen, L., Graham, W. V., Weber, C. R., Schwarz, B. T., Austin, J. R., 2nd, Raleigh, D. R., Guan, Y., Watson, A. J., Montrose, M. H. and Turner, J. R. (2010). Caveolin-1-dependent occludin endocytosis is required for TNF-induced tight junction regulation in vivo. *J Cell Biol*, 189(1): 111-126.
- Marcos-Ramiro, B., Garcia-Weber, D. and Millan, J. (2014). TNF-induced endothelial barrier disruption: beyond actin and Rho. *Thromb Haemost*, 112(6): 1088-1102.
- Mashru, M. R., Shah, V. K., Soneji, S. L., Loya, Y. S., Vasvani, J. B., Payannavar, S., Walvalkar, A., Mithbawkar, S. S., Mokal, R., Kudalkar, K., Abraham, A., Thakur, P. K. and Shalia, K. K. (2010). Soluble levels of cell adhesion molecules (CAMs) in coronary artery disease. *Indian Heart J*, 62(1): 57-63.
- McKenzie, J. A. and Ridley, A. J. (2007). Roles of Rho/ROCK and MLCK in TNF-alpha-induced changes in endothelial morphology and permeability. *J Cell Physiol*, 213(1): 221-228.
- Mehta, D. and Malik, A. B. (2006). Signaling mechanisms regulating endothelial permeability. *Physiol Rev*, 86(1): 279-367.
- Mestas, J. and Ley, K. (2008). Monocyte-endothelial cell interactions in the development of atherosclerosis. *Trends Cardiovasc Med*, 18(6): 228-232.
- Millan, J., Cain, R. J., Reglero-Real, N., Bigarella, C., Marcos-Ramiro, B., Fernandez-Martin, L., Correas, I. and Ridley, A. J. (2010). Adherens junctions connect stress fibres between adjacent endothelial cells. *BMC Biol*, 8: 11.
- Millan, J., Hewlett, L., Glyn, M., Toomre, D., Clark, P. and Ridley, A. J. (2006). Lymphocyte transcellular migration occurs through recruitment of endothelial ICAM-1 to caveola- and F-actin-rich domains. *Nat Cell Biol*, 8(2): 113-123.
- Miyawaki-Shimizu, K., Predescu, D., Shimizu, J., Broman, M., Predescu, S. and Malik, A. B. (2006). siRNA-induced caveolin-1 knockdown in mice increases lung vascular permeability via the junctional pathway. *Am J Physiol Lung Cell Mol Physiol*, 290(2): L405-413.

- Monaco, C. and Paleolog, E. (2004). Nuclear factor kappaB: a potential therapeutic target in atherosclerosis and thrombosis. *Cardiovasc Res*, 61(4): 671-682.
- Moon, L., Ha, Y. M., Jang, H. J., Kim, H. S., Jun, M. S., Kim, Y. M., Lee, Y. S., Lee, D. H., Son, K. H., Kim, H. J., Seo, H. G., Lee, J. H., Kim, Y. S. and Chang, K. C. (2011). Isoimperatorin, cimicidine E and 23-O-acetylshengmanol-3-xyloside from *Cimicifugae* rhizome inhibit TNF-alpha-induced VCAM-1 expression in human endothelial cells: involvement of PPAR-gamma upregulation and PI3K, ERK1/2, and PKC signal pathways. *J Ethnopharmacol*, 133(2): 336-344.
- Muller, S. L., Portwich, M., Schmidt, A., Utebgerenov, D. I., Huber, O., Blasig, I. E. and Krause, G. (2005). The tight junction protein occludin and the adherens junction protein alpha-catenin share a common interaction mechanism with ZO-1. *J Biol Chem*, 280(5): 3747-3756.
- Muller, W. A. (2011). Mechanisms of leukocyte transendothelial migration. *Annu Rev Pathol*, 6: 323-344.
- Muller, W. A. (2014). How endothelial cells regulate transmigration of leukocytes in the inflammatory response. *Am J Pathol*, 184(4): 886-896.
- Muller, W. A. (2015). The regulation of transendothelial migration: new knowledge and new questions. *Cardiovasc Res*, 107(3): 310-320.
- Murakami, T., Felinski, E. A. and Antonetti, D. A. (2009). Occludin phosphorylation and ubiquitination regulate tight junction trafficking and vascular endothelial growth factor-induced permeability. *J Biol Chem*, 284(31): 21036-21046.
- Murakami, T., Frey, T., Lin, C. and Antonetti, D. A. (2012). Protein kinase beta phosphorylates occludin regulating tight junction trafficking in vascular endothelial growth factor-induced permeability in vivo. *Diabetes*, 61(6): 1573-1583.
- Nanes, B. A., Chiasson-MacKenzie, C., Lowery, A. M., Ishiyama, N., Faundez, V., Ikura, M., Vincent, P. A. and Kowalczyk, A. P. (2012). p120-catenin binding masks an endocytic signal conserved in classical cadherins. *J Cell Biol*, 199(2): 365-380.
- Napoli, C., D'Armiento, F. P., Mancini, F. P., Postiglione, A., Witztum, J. L., Palumbo, G. and Palinski, W. (1997). Fatty streak formation occurs in human fetal aortas and is greatly enhanced by maternal hypercholesterolemia. Intimal accumulation of low density lipoprotein and its oxidation precede monocyte recruitment into early atherosclerotic lesions. *J Clin Invest*, 100(11): 2680-2690.
- Natarajan, V., Dudek, S. M., Jacobson, J. R., Moreno-Vinasco, L., Huang, L. S., Abassi, T., Mathew, B., Zhao, Y., Wang, L., Bittman, R., Weichselbaum, R., Berdyshev, E. and Garcia, J. G. (2013). Sphingosine-1-phosphate, FTY720, and sphingosine-1-phosphate receptors in the pathobiology of acute lung injury. *Am J Respir Cell Mol Biol*, 49(1): 6-17.

- Ng, C. T., Fong, L. Y., Sulaiman, M. R., Moklas, M. A., Yong, Y. K., Hakim, M. N. and Ahmad, Z. (2015). Interferon-gamma increases endothelial permeability by causing activation of p38 MAP kinase and actin cytoskeleton alteration. *J Interferon Cytokine Res*, 35(7): 513-522.
- Nottebaum, A. F., Cagna, G., Winderlich, M., Gamp, A. C., Linnepe, R., Polaschegg, C., Filippova, K., Lyck, R., Engelhardt, B., Kamenyeva, O., Bixel, M. G., Butz, S. and Vestweber, D. (2008). VE-PTP maintains the endothelial barrier via plakoglobin and becomes dissociated from VE-cadherin by leukocytes and by VEGF. *J Exp Med*, 205(12): 2929-2945.
- Orsenigo, F., Giampietro, C., Ferrari, A., Corada, M., Galaup, A., Sigismund, S., Ristagno, G., Maddaluno, L., Koh, G. Y., Franco, D., Kurtcuoglu, V., Poulidakos, D., Baluk, P., McDonald, D., Grazia Lampugnani, M. and Dejana, E. (2012). Phosphorylation of VE-cadherin is modulated by haemodynamic forces and contributes to the regulation of vascular permeability in vivo. *Nat Commun*, 3: 1208.
- Papagianni, A., Kalovoulos, M., Kirmizis, D., Vainas, A., Belechri, A. M., Alexopoulos, E. and Memmos, D. (2003). Carotid atherosclerosis is associated with inflammation and endothelial cell adhesion molecules in chronic haemodialysis patients. *Nephrol Dial Transplant*, 18(1): 113-119.
- Patil, K. R., Mohapatra, P., Patel, H. M., Goyal, S. N., Ojha, S., Kundu, C. N. and Patil, C. R. (2015). Pentacyclic triterpenoids inhibit IKKbeta mediated activation of NF-kappaB pathway: *in silico* and *in vitro* evidences. *PLoS One*, 10(5): e0125709.
- Pawitan, J. A. (2011). Potential agents against plasma leakage. *ISRN Pharmacol*, 2011: 975048.
- Petrache, I., Verin, A. D., Crow, M. T., Birukova, A., Liu, F. and Garcia, J. G. (2001). Differential effect of MLC kinase in TNF-alpha-induced endothelial cell apoptosis and barrier dysfunction. *Am J Physiol Lung Cell Mol Physiol*, 280(6): L1168-1178.
- Phillips, M. L., Nudelman, E., Gaeta, F. C., Perez, M., Singhal, A. K., Hakomori, S. and Paulson, J. C. (1990). ELAM-1 mediates cell adhesion by recognition of a carbohydrate ligand, sialyl-Lex. *Science*, 250(4984): 1130-1132.
- Pozo, M., de Nicolas, R., Egido, J. and Gonzalez-Cabrero, J. (2006). Simvastatin inhibits the migration and adhesion of monocytic cells and disorganizes the cytoskeleton of activated endothelial cells. *Eur J Pharmacol*, 548(1-3): 53-63.
- Prasain, N. and Stevens, T. (2009). The actin cytoskeleton in endothelial cell phenotypes. *Microvasc Res*, 77(1): 53-63.
- Predescu, D. and Palade, G. E. (1993). Plasmalemmal vesicles represent the large pore system of continuous microvascular endothelium. *Am J Physiol*, 265(2 Pt 2): H725-733.

- Privratsky, J. R. and Newman, P. J. (2014). PECAM-1: regulator of endothelial junctional integrity. *Cell Tissue Res*, 355(3): 607-619.
- Privratsky, J. R., Paddock, C. M., Florey, O., Newman, D. K., Muller, W. A. and Newman, P. J. (2011). Relative contribution of PECAM-1 adhesion and signaling to the maintenance of vascular integrity. *J Cell Sci*, 124(Pt 9): 1477-1485.
- Rajendran, P., Rengarajan, T., Thangavel, J., Nishigaki, Y., Sakthisekaran, D., Sethi, G. and Nishigaki, I. (2013). The vascular endothelium and human diseases. *Int J Biol Sci*, 9(10): 1057-1069.
- Ramachandran, V., Saravanan, R. and Senthilraja, P. (2014). Antidiabetic and antihyperlipidemic activity of asiatic acid in diabetic rats, role of HMG CoA: in vivo and in silico approaches. *Phytomedicine*, 21(3): 225-232.
- Ramirez, S. H., Heilman, D., Morsey, B., Potula, R., Haorah, J. and Persidsky, Y. (2008). Activation of peroxisome proliferator-activated receptor gamma (PPARgamma) suppresses Rho GTPases in human brain microvascular endothelial cells and inhibits adhesion and transendothelial migration of HIV-1 infected monocytes. *J Immunol*, 180(3): 1854-1865.
- Ramji, D. P. and Davies, T. S. (2015). Cytokines in atherosclerosis: Key players in all stages of disease and promising therapeutic targets. *Cytokine Growth Factor Rev*, 26(6): 673-685.
- Reijerkerk, A., Kooij, G., van der Pol, S. M., Khazen, S., Dijkstra, C. D. and de Vries, H. E. (2006). Diapedesis of monocytes is associated with MMP-mediated occludin disappearance in brain endothelial cells. *FASEB J*, 20(14): 2550-2552.
- Ribeiro, F., Alves, A. J., Teixeira, M., Ribeiro, V., Duarte, J. A. and Oliveira, J. (2009). Endothelial function and atherosclerosis: circulatory markers with clinical usefulness. *Rev Port Cardiol*, 28(10): 1121-1151.
- Rochfort, K. D. and Cummins, P. M. (2015). Cytokine-mediated dysregulation of zonula occludens-1 properties in human brain microvascular endothelium. *Microvasc Res*, 100: 48-53.
- Rodrigues, S. F. and Granger, D. N. (2015). Blood cells and endothelial barrier function. *Tissue Barriers*, 3(1-2): e978720.
- Rossi, J., Rouleau, L., Emmott, A., Tardif, J. C. and Leask, R. L. (2010). Laminar shear stress prevents simvastatin-induced adhesion molecule expression in cytokine activated endothelial cells. *Eur J Pharmacol*, 649(1-3): 268-276.
- Saitou, M., Furuse, M., Sasaki, H., Schulzke, J. D., Fromm, M., Takano, H., Noda, T. and Tsukita, S. (2000). Complex phenotype of mice lacking occludin, a component of tight junction strands. *Mol Biol Cell*, 11(12): 4131-4142.

- Sandoo, A., van Zanten, J. J., Metsios, G. S., Carroll, D. and Kitas, G. D. (2010). The endothelium and its role in regulating vascular tone. *Open Cardiovasc Med J*, 4: 302-312.
- Sarelius, I. H. and Glading, A. J. (2015). Control of vascular permeability by adhesion molecules. *Tissue Barriers*, 3(1-2): e985954.
- Satpathy, M., Gallagher, P., Lizotte-Waniewski, M. and Srinivas, S. P. (2004). Thrombin-induced phosphorylation of the regulatory light chain of myosin II in cultured bovine corneal endothelial cells. *Exp Eye Res*, 79(4): 477-486.
- Schnoor, M., Lai, F. P., Zarbock, A., Klaver, R., Polaschegg, C., Schulte, D., Weich, H. A., Oelkers, J. M., Rottner, K. and Vestweber, D. (2011). Cortactin deficiency is associated with reduced neutrophil recruitment but increased vascular permeability *in vivo*. *J Exp Med*, 208(8): 1721-1735.
- Schubert, W., Frank, P. G., Razani, B., Park, D. S., Chow, C. W. and Lisanti, M. P. (2001). Caveolae-deficient endothelial cells show defects in the uptake and transport of albumin *in vivo*. *J Biol Chem*, 276(52): 48619-48622.
- Schulte, D., Kuppers, V., Dartsch, N., Broermann, A., Li, H., Zarbock, A., Kamenyeva, O., Kiefer, F., Khandoga, A., Massberg, S. and Vestweber, D. (2011). Stabilizing the VE-cadherin-catenin complex blocks leukocyte extravasation and vascular permeability. *EMBO J*, 30(20): 4157-4170.
- Shen, Q., Wu, M. H. and Yuan, S. Y. (2009). Endothelial contractile cytoskeleton and microvascular permeability. *Cell Health Cytoskelet*, 2009(1): 43-50.
- Shen, W., Li, S., Chung, S. H., Zhu, L., Stayt, J., Su, T., Couraud, P. O., Romero, I. A., Weksler, B. and Gillies, M. C. (2011). Tyrosine phosphorylation of VE-cadherin and claudin-5 is associated with TGF-beta1-induced permeability of centrally derived vascular endothelium. *Eur J Cell Biol*, 90(4): 323-332.
- Shivanna, M., Rajashekhar, G. and Srinivas, S. P. (2010). Barrier dysfunction of the corneal endothelium in response to TNF-alpha: role of p38 MAP kinase. *Invest Ophthalmol Vis Sci*, 51(3): 1575-1582.
- Si, L., Xu, J., Yi, C., Xu, X., Wang, F., Gu, W., Zhang, Y. and Wang, X. (2014). Asiatic acid attenuates cardiac hypertrophy by blocking transforming growth factor-beta1-mediated hypertrophic signaling *in vitro* and *in vivo*. *Int J Mol Med*, 34(2): 499-506.
- Simic, I. and Reiner, Z. (2015). Adverse effects of statins - myths and reality. *Curr Pharm Des*, 21(9): 1220-1226.
- Singleton, P. A., Dudek, S. M., Chiang, E. T. and Garcia, J. G. (2005). Regulation of sphingosine 1-phosphate-induced endothelial cytoskeletal rearrangement and barrier enhancement by S1P1 receptor, PI3 kinase, Tiam1/Rac1, and alpha-actinin. *FASEB J*, 19(12): 1646-1656.

- Sitia, S., Tomasoni, L., Atzeni, F., Ambrosio, G., Cordiano, C., Catapano, A., Tramontana, S., Perticone, F., Naccarato, P., Camici, P., Picano, E., Cortigiani, L., Bevilacqua, M., Milazzo, L., Cusi, D., Barlassina, C., Sarzi-Puttini, P. and Turiel, M. (2010). From endothelial dysfunction to atherosclerosis. *Autoimmun Rev*, 9(12): 830-834.
- Somchit, M. N., Sulaiman, M. R., Zuraini, A., Samsuddin, L., Somchit, N., Israf, D. A. and Moin, S. (2004). Antinociceptive and antiinflammatory effects of *Centella asiatica*. *Indian J Pharmacol*, 36: 377-380.
- Srinivas, S. P., Satpathy, M., Guo, Y. and Anandan, V. (2006). Histamine-induced phosphorylation of the regulatory light chain of myosin II disrupts the barrier integrity of corneal endothelial cells. *Invest Ophthalmol Vis Sci*, 47(9): 4011-4018.
- Steegmaier, M., Levinovitz, A., Isenmann, S., Borges, E., Lenter, M., Kocher, H. P., Kleuser, B. and Vestweber, D. (1995). The E-selectin-ligand ESL-1 is a variant of a receptor for fibroblast growth factor. *Nature*, 373(6515): 615-620.
- Sullivan, D. P. and Muller, W. A. (2014). Neutrophil and monocyte recruitment by PECAM, CD99, and other molecules via the LBRC. *Semin Immunopathol*, 36(2): 193-209.
- Sumagin, R., Kuebel, J. M. and Sarelius, I. H. (2011). Leukocyte rolling and adhesion both contribute to regulation of microvascular permeability to albumin via ligation of ICAM-1. *Am J Physiol Cell Physiol*, 301(4): C804-813.
- Sumagin, R., Lomakina, E. and Sarelius, I. H. (2008). Leukocyte-endothelial cell interactions are linked to vascular permeability via ICAM-1-mediated signaling. *Am J Physiol Heart Circ Physiol*, 295(3): H969-H977.
- Sun, X., Shikata, Y., Wang, L., Ohmori, K., Watanabe, N., Wada, J., Shikata, K., Birukov, K. G., Makino, H., Jacobson, J. R., Dudek, S. M. and Garcia, J. G. (2009). Enhanced interaction between focal adhesion and adherens junction proteins: involvement in sphingosine 1-phosphate-induced endothelial barrier enhancement. *Microvasc Res*, 77(3): 304-313.
- Supkamonseni, N., Thinkratok, A., Meksuriyen, D. and Srisawat, R. (2014). Hypolipidemic and hypoglycemic effects of *Centella asiatica* (L.) extract *in vitro* and *in vivo*. *Indian J Exp Biol*, 52(10): 965-971.
- Taddei, A., Giampietro, C., Conti, A., Orsenigo, F., Breviario, F., Pirazzoli, V., Potente, M., Daly, C., Dimmeler, S. and Dejana, E. (2008). Endothelial adherens junctions control tight junctions by VE-cadherin-mediated upregulation of claudin-5. *Nat Cell Biol*, 10(8): 923-934.
- Tedgui, A. and Mallat, Z. (2006). Cytokines in atherosclerosis: pathogenic and regulatory pathways. *Physiol Rev*, 86(2): 515-581.
- Tharakan, B., Hellman, J., Sawant, D. A., Tinsley, J. H., Parrish, A. R., Hunter, F. A., Smythe, W. R. and Childs, E. W. (2012). beta-Catenin dynamics in the

- regulation of microvascular endothelial cell hyperpermeability. *Shock*, 37(3): 306-311.
- Thompson, P. W., Randi, A. M. and Ridley, A. J. (2002). Intercellular adhesion molecule (ICAM)-1, but not ICAM-2, activates RhoA and stimulates c-fos and rhoA transcription in endothelial cells. *J Immunol*, 169(2): 1007-1013.
- Timmerman, I., Hoogenboezem, M., Bennett, A. M., Geerts, D., Hordijk, P. L. and van Buul, J. D. (2012). The tyrosine phosphatase SHP2 regulates recovery of endothelial adherens junctions through control of beta-catenin phosphorylation. *Mol Biol Cell*, 23(21): 4212-4225.
- Tirupathi, C., Shimizu, J., Miyawaki-Shimizu, K., Vogel, S. M., Bair, A. M., Minshall, R. D., Predescu, D. and Malik, A. B. (2008). Role of NF-kappaB-dependent caveolin-1 expression in the mechanism of increased endothelial permeability induced by lipopolysaccharide. *J Biol Chem*, 283(7): 4210-4218.
- Umeda, K., Ikenouchi, J., Katahira-Tayama, S., Furuse, K., Sasaki, H., Nakayama, M., Matsui, T., Tsukita, S., Furuse, M. and Tsukita, S. (2006). ZO-1 and ZO-2 independently determine where claudins are polymerized in tight-junction strand formation. *Cell*, 126(4): 741-754.
- van Buul, J. D., van Rijssel, J., van Alphen, F. P., van Stalborch, A. M., Mul, E. P. and Hordijk, P. L. (2010). ICAM-1 clustering on endothelial cells recruits VCAM-1. *J Biomed Biotechnol*, 2010: 120328.
- Van Itallie, C. M., Aponte, A., Tietgens, A. J., Gucek, M., Fredriksson, K. and Anderson, J. M. (2013). The N and C termini of ZO-1 are surrounded by distinct proteins and functional protein networks. *J Biol Chem*, 288(19): 13775-13788.
- van Wetering, S., van den Berk, N., van Buul, J. D., Mul, F. P., Lommerse, I., Mous, R., ten Klooster, J. P., Zwaginga, J. J. and Hordijk, P. L. (2003). VCAM-1-mediated Rac signaling controls endothelial cell-cell contacts and leukocyte transmigration. *Am J Physiol Cell Physiol*, 285(2): C343-352.
- VandenBerg, E., Reid, M. D., Edwards, J. D. and Davis, H. W. (2004). The role of the cytoskeleton in cellular adhesion molecule expression in tumor necrosis factor-stimulated endothelial cells. *J Cell Biochem*, 91(5): 926-937.
- Vandenbroucke, E., Mehta, D., Minshall, R. and Malik, A. B. (2008). Regulation of endothelial junctional permeability. *Ann N Y Acad Sci*, 1123: 134-145.
- Vockel, M. and Vestweber, D. (2013). How T cells trigger the dissociation of the endothelial receptor phosphatase VE-PTP from VE-cadherin. *Blood*, 122(14): 2512-2522.
- Wallez, Y. and Huber, P. (2008). Endothelial adherens and tight junctions in vascular homeostasis, inflammation and angiogenesis. *Biochim Biophys Acta*, 1778(3): 794-809.

- Wang, L., Bittman, R., Garcia, J. G. and Dudek, S. M. (2015a). Junctional complex and focal adhesion rearrangement mediates pulmonary endothelial barrier enhancement by FTY720 S-phosphonate. *Microvasc Res*, 99: 102-109.
- Wang, L., Chiang, E. T., Simmons, J. T., Garcia, J. G. and Dudek, S. M. (2011a). FTY720-induced human pulmonary endothelial barrier enhancement is mediated by c-Abl. *Eur Respir J*, 38(1): 78-88.
- Wang, L., Du, F. and Wang, X. (2008). TNF-alpha induces two distinct caspase-8 activation pathways. *Cell*, 133(4): 693-703.
- Wang, L. and Dudek, S. M. (2009). Regulation of vascular permeability by sphingosine 1-phosphate. *Microvasc Res*, 77(1): 39-45.
- Wang, N., Zhang, D., Sun, G., Zhang, H., You, Q., Shao, M. and Yue, Y. (2015b). Lipopolysaccharide-induced caveolin-1 phosphorylation-dependent increase in transcellular permeability precedes the increase in paracellular permeability. *Drug Des Devel Ther*, 9: 4965-4977.
- Wang, S., Yu, H. and Wickliffe, J. K. (2011b). Limitation of the MTT and XTT assays for measuring cell viability due to superoxide formation induced by nano-scale TiO₂. *Toxicol In Vitro*, 25(8): 2147-2151.
- Waschke, J., Curry, F. E., Adamson, R. H. and Drenckhahn, D. (2005). Regulation of actin dynamics is critical for endothelial barrier functions. *Am J Physiol Heart Circ Physiol*, 288(3): H1296-1305.
- Watanabe, T., Hosoya, H. and Yonemura, S. (2007). Regulation of myosin II dynamics by phosphorylation and dephosphorylation of its light chain in epithelial cells. *Mol Biol Cell*, 18(2): 605-616.
- Waters, J. P., Pober, J. S. and Bradley, J. R. (2013). Tumour necrosis factor in infectious disease. *J Pathol*, 230(2): 132-147.
- Weis, W. I. and Nelson, W. J. (2006). Re-solving the cadherin-catenin-actin conundrum. *J Biol Chem*, 281(47): 35593-35597.
- Wessel, F., Winderlich, M., Holm, M., Frye, M., Rivera-Galdos, R., Vockel, M., Linnepe, R., Ipe, U., Stadtmann, A., Zarbock, A., Nottebaum, A. F. and Vestweber, D. (2014). Leukocyte extravasation and vascular permeability are each controlled in vivo by different tyrosine residues of VE-cadherin. *Nat Immunol*, 15(3): 223-230.
- Wilson, C. W. and Ye, W. (2014). Regulation of vascular endothelial junction stability and remodeling through Rap1-Rasip1 signaling. *Cell Adh Migr*, 8(2): 76-83.
- Wojciak-Stothard, B., Entwistle, A., Garg, R. and Ridley, A. J. (1998). Regulation of TNF-alpha-induced reorganization of the actin cytoskeleton and cell-cell junctions by Rho, Rac, and Cdc42 in human endothelial cells. *J Cell Physiol*, 176(1): 150-165.

- Woodfin, A., Voisin, M. B., Imhof, B. A., Dejana, E., Engelhardt, B. and Nourshargh, S. (2009). Endothelial cell activation leads to neutrophil transmigration as supported by the sequential roles of ICAM-2, JAM-A, and PECAM-1. *Blood*, 113(24): 6246-6257.
- Wu, Y. and Zhou, B. P. (2010). TNF-alpha/NF-kappaB/Snail pathway in cancer cell migration and invasion. *Br J Cancer*, 102(4): 639-644.
- Xu, X., Si, L., Xu, J., Yi, C., Wang, F., Gu, W., Zhang, Y. and Wang, X. (2015). Asiatic acid inhibits cardiac hypertrophy by blocking interleukin-1beta-activated nuclear factor-kappaB signaling in vitro and in vivo. *J Thorac Dis*, 7(10): 1787-1797.
- Yan, S. L., Yang, H. T., Lee, Y. J., Lin, C. C., Chang, M. H. and Yin, M. C. (2014). Asiatic acid ameliorates hepatic lipid accumulation and insulin resistance in mice consuming a high-fat diet. *J Agric Food Chem*, 62(20): 4625-4631.
- Ye, X., Jiang, X., Guo, W., Clark, K. and Gao, Z. (2013). Overexpression of NF-kappaB p65 in macrophages ameliorates atherosclerosis in apoE-knockout mice. *Am J Physiol Endocrinol Metab*, 305(11): E1375-1383.
- Yuan, S. Y. and Rigor, R. R. (2010). Chapter 4: The Endothelial Barrier. In *Regulation of Endothelial Barrier Function*. San Rafael (CA): Morgan & Claypool Life Sciences.
- Yuan, Y., Zhang, H., Sun, F., Sun, S., Zhu, Z. and Chai, Y. (2015). Biopharmaceutical and pharmacokinetic characterization of asiatic acid in *Centella asiatica* as determined by a sensitive and robust HPLC-MS method. *J Ethnopharmacol*, 163: 31-38.
- Yun, K. J., Kim, J. Y., Kim, J. B., Lee, K. W., Jeong, S. Y., Park, H. J., Jung, H. J., Cho, Y. W., Yun, K. and Lee, K. T. (2008). Inhibition of LPS-induced NO and PGE2 production by asiatic acid via NF-kappa B inactivation in RAW 264.7 macrophages: possible involvement of the IKK and MAPK pathways. *Int Immunopharmacol*, 8(3): 431-441.
- Zapolska-Downar, D., Siennicka, A., Kaczmarczyk, M., Kolodziej, B. and Naruszewicz, M. (2004). Simvastatin modulates TNFalpha-induced adhesion molecules expression in human endothelial cells. *Life Sci*, 75(11): 1287-1302.
- Zhao, Y., Shu, P., Zhang, Y., Lin, L., Zhou, H., Xu, Z., Suo, D., Xie, A. and Jin, X. (2014). Effect of *Centella asiatica* on oxidative stress and lipid metabolism in hyperlipidemic animal models. *Oxid Med Cell Longev*, 2014: 154295.
- Zonneveld, R., Martinelli, R., Shapiro, N. I., Kuijpers, T. W., Plotz, F. B. and Carman, C. V. (2014). Soluble adhesion molecules as markers for sepsis and the potential pathophysiological discrepancy in neonates, children and adults. *Crit Care*, 18(2): 204.

BIODATA OF STUDENT

Ms. Fong Lai Yen was born on 9th July 1988 in Kuala Lumpur, Malaysia. She received her primary education from Sekolah Rendah Jenis Kebangsaan (C) Chen Moh, Petaling Jaya, Selangor (1995 – 1999). Then, she continued her secondary education at Sekolah Menengah Jenis Kebangsaan Katholik, Petaling Jaya, Selangor (2000-2004). She also completed her Form Six (pre-university education) at the same high school.

She obtained her first degree in Bachelor of Science (Biomedical Science) from Universiti Putra Malaysia in 2011. She has developed an interest in biomedical research during her final year project, in which she gained her basic laboratory skills such as animal handling and performing bioassays. She decided to further her Doctor of Philosophy (PhD) study in Universiti Putra Malaysia, of which her research project is mainly focusing on prevention of early atherogenesis by using active compounds isolated from medicinal plants.

Ms. Fong Lai Yen was granted the MyBrain15 Scholarship (MyPhD) by the Ministry of Education Malaysia to pursue her PhD study. Besides, her research project was also granted by the Malaysia Toray Science Foundation (MTSF). She attended and presented her papers in several international and local conferences throughout her PhD study. She has also been working as a research assistant in Faculty of Medicine and Health Sciences, Universiti Putra Malaysia for one year.

LIST OF PUBLICATIONS

Journals

Fong, L. Y., Ng, C. T., Cheok, Z. L., Mohd Moklas, M. A., Hakim, M. N. and Ahmad, Z. (2016). Barrier protective effect of asiatic acid in TNF- α -induced activation of human aortic endothelial cells. *Phytomedicine*, 23(2): 191-199.

Fong, L. Y., Ng, C. T., Zakaria, Z. A., Baharuldin, M. T., Arifah, A. K., Hakim, M. N. and Zuraini, A. (2015). Asiaticoside Inhibits TNF- α -Induced Endothelial Hyperpermeability of Human Aortic Endothelial Cells. *Phytother Res*, 29(10): 1501-1508.

Proceedings/Conference Papers

LY Fong, CT Ng, Z Ahmad. (2015). Effects of asiatic acid on TNF- α -induced vascular inflammatory events in human aortic endothelial cells. Proceedings of the 8th Congress Federation of the Asian and Oceanian Physiological Societies (FAOPS), pg S-A106. 22th-25th of November, 2015, Bangkok, Thailand.

LY Fong, CT Ng, Z Ahmad. (2015). Asiaticoside reduces TNF- α -induced increased soluble platelet endothelial cell adhesion molecule -1 (sPECAM-1) levels in human aortic endothelial cells. Abstracts of 63rd International Congress and Annual Meeting of the Society for Medicinal Plant and Natural Product Research (GA 2015), Budapest, Hungary. Published in *Planta medica* 81(16)-PW_115.

LY Fong, CT Ng, Z Ahmad. (2014). Asiaticoside inhibits tumor necrosis factor alpha (TNF α)-induced soluble adhesion molecules expression in human aortic endothelial cells (HAEC). Abstracts of 62nd International Congress and Annual Meeting of the Society for Medicinal Plant and Natural Product Research (GA 2014), Guimaraes, Portugal. Published in *Planta Medica* 80-P1L107.

CT Ng, LY Fong, Z Ahmad. (2014). Asiatic acid exhibits anti-inflammatory activities in human aortic endothelial cells. Abstracts of 62nd International Congress and Annual Meeting of the Society for Medicinal Plant and Natural Product Research, Guimaraes, Portugal. Published in *Planta Medica* 80-P1L23.

LY Fong, CT Ng, Z Ahmad. (2014). Inhibitory Effect of Asiaticoside on Endothelial Barrier Dysfunction induced by Tumor necrosis factor- α (TNF- α). Abstracts of 6th Scientific Meeting of the Asian Society for Vascular Biology, Kuala Lumpur, Malaysia. Published in *Journal of Vascular Research* 51(Suppl 1):9.

LY Fong, CT Ng, Z Ahmad. (2013). *In vitro* anti-atherogenic effects of asiaticoside on human aortic endothelial cells. Abstracts of 12nd meeting of the Asia Pacific Federation of Pharmacologists, Shanghai, China. Published in *Acta Pharmacological Sinica*, 34 Supp:60

Conferences Attended

Effects of asiatic acid on TNF- α -induced vascular inflammatory events in human aortic endothelial cells (2015). Oral presentation at the 8th Congress Federation of the Asian and Oceanian Physiological Societies (FAOPS), 22-25th of November, held in Bangkok, Thailand.

Asiaticoside reduces TNF- α -induced increased soluble platelet endothelial cell adhesion molecule -1 (sPECAM-1) levels in human aortic endothelial cells (2015). Poster presentation at the 63rd International Congress and Annual Meeting of the Society for Medicinal Plant and Natural Product Research (GA2015), 23-27th of August, held in Budapest, Hungary.

Asiaticoside inhibits tumor necrosis factor alpha (TNF α)-induced soluble adhesion molecules expression in human aortic endothelial cells (2014). Poster presentation at the 62nd International Congress and Annual Meeting of the Society for Medicinal Plant and Natural Product Research (GA2014), 31st of August-4th of September, held in Guimaraes, Portugal.

Inhibitory Effect of Asiaticoside on Endothelial Barrier Dysfunction induced by Tumor necrosis factor- α (2014). Poster presentation at the 6th Scientific Meeting of The Asian Society for Vascular Biology, 22-24th of August, held in Kuala Lumpur, Malaysia.

In vitro anti-atherogenic effects of asiaticoside in endothelial cells (2013). Poster presentation at the 27th Scientific Meeting of Malaysian Society of Pharmacology and Physiology (MSPP), 6-8th of September, held in Pahang, Malaysia.

In vitro anti-atherogenic effects of asiaticoside in endothelial cells (2013). Poster Presentation at the 12th meeting of Asia Pacific Federation of Pharmacologists (MAPFP), 9-13th of July, held in Shanghai, China.

Awards

Young scientist award from the 8th Congress Federation of the Asian and Oceanian Physiological Societies (FAOPS), 22-25th of November 2015, held in Bangkok, Thailand on 22-25th of November, 2015

Travel award from the Society for Medicinal Plant and Natural Product Research (GA) to attend the 63rd International Congress and Annual Meeting of the Society for Medicinal Plant and Natural Product Research (GA 2015) held in Budapest, Hungary on 23-27th of August 2015.

Financial aid from Universiti Putra Malaysia to attend the 62nd International Congress and Annual Meeting of the Society for Medicinal Plant and Natural Product Research (GA2014), held in Guimaraes, Portugal on 31st of August–4th of September, 2014

Travel award from Asia Pacific Federation of Pharmacologists (APFP) to attend the 12th meeting of APFP held in Shanghai, China on 9–13rd July 2013

Research Grants Received

Science & Technology Research Grant (by Malaysia Toray Science Foundation, 2012)
- RM15,000

Research University Grant Scheme 2012 (by Universiti Putra Malaysia) – RM10,000