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DEVELOPMENT AND CHARACTERIZATION OF PARENTERAL NANO-DELIVERY SYSTEM LOADED WITH AZITHROMYCIN

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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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To

My Father and my Mother "If I have seen further, it is by standing on the shoulder of giants"

Sir Isaac Newton

My brother Manhal and my sister Dabyaa
"The happiest moments of my life have been the few which I have passed
at home in the bosom of my family"

Thomas Jefferson

My children Mayar, Fatimah, Eethar, and Yaman You are the light in my life

And

My awesome husband Auday For a debt I can never repay Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

DEVELOPMENT AND CHARACTERIZATION OF PARENTERAL NANO-DELIVERY SYSTEM LOADED WITH AZITHROMYCIN

By

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The blood brain barrier (BBB) is established by brain microvascular endothelial cells (BMEC) working simultaneously with pericytes and astrocytes, in which tight junctions and several transporters strictly regulate the penetration of various bioactive compounds into the brain including antibiotics. This has significant consequences for the treatment of Central Nervous System (CNS) infections, as antibiotics have to gain access to the brain through the BBB in adequate concentrations to exhibit their antibacterial activity. Azithromycin (AZO) is a broad spectrum antibiotic with a unique pharmacokinetic profile. However, its role in treatment of CNS infections is limited because it does not possess the appropriate physicochemical properties that enables it to achieve sufficient concentrations in brain tissue.

Nanoemulsion system is one of the potential strategies for efficient delivery of lipophilic actives across the BBB owing to their nano-sized, biocompatible, biodegradable, physical stability and relatively easy to produce on a large scale. With the aim of brain targeting, AZO-loaded nanoemulsions were developed utilizing high pressure homogenization with a homogenization pressure of 1000 bar for 8 cycles. The formulated nanoemulsions were optimized utilizing artificial neural network (ANN) as a multivariate statistical technique. In order to achieve the optimum topologies, ANN was trained by Incremental Back-Propagation (IBP), Batch Back-Propagation (BBP), Quick Propagation (QP), and Levenberg-Marquardt (LM) algorithms for testing data set. The topologies were confirmed by the indicator of minimized root mean squared error (RMSE) for each. Based on that indicator, the BBP-5-14-1 was selected as the optimum topology to be used as a final model to predict the desirable particle size and relative importance of the formulation's effective variables. The ANN analysis showed that with optimum compositions of soya bean oil 6%, oleic acid 2%, AZO 1.4%, lecithin 2%, Tween

80 2%, glycerol 2.5%, vitamin E 0.25%, and water 83.85%, minimum particle size can be obtained.

The optimized nanoemulsions were evaluated for the various physicochemical properties. The characterization revealed particle size of 54.67 ± 0.81 nm, polydispersity index (PDI) of 0.218 ± 0.023 , zeta potential of -34.65 ± 0.78 mV, pH of 7.82 ± 0.07 , viscosity of 1.77 ± 0.05 cps, and osmolality of 288 ± 1.00 mOsm/kg, suggesting their compatibility for intravenous administration. AZO was successfully incorporated into nanoemulsion system with an average encapsulation efficiency of $98.21 \pm 1.97\%$ and a relatively high drug loading of $91.19 \pm 5.93\%$. Morphological analysis with Transmission Electron Microscopy (TEM) confirmed the formation of almost spherical shaped uniformly distributed nano-sized oil droplets.

In vitro drug release study of the selected formulation demonstrated a release profile similar to that of AZO standard solution, both exhibited a biphasic behavior characterized by a fast initial release of the encapsulated drug followed by a slower sustained release till the optimized formulation achieved a total accumulative release of the drug of 84.94 ± 4.76% within 48 h. Kinetically, AZO release profile from nanoemulsion system in vitro appeared to fit best with the Higuchi model. Stability of nanoemulsion prepared with the optimized formula was mainly evaluated in term of preserving its physical integrity, namely particle size and polydispersity index (PDI). The formulation maintained its properties in a satisfactory range up to 12 months of storage at 4°C and 25°C, which demonstrated sufficient physical stability upon long-term storage.

A linear relationship between the particle size (cube of the radius of dispersed phase droplets (r^3)) and time (t) was obtained identifying Ostwald ripening (OR) as the dominant destabilization mechanism of AZO-loaded nanoemulsion from a 12-months shelf-life study. OR rate (ω) was extrapolated graphically from the slope and was found to be 0.232×10^{-8} nm³/s. Optimized nanoemulsion was analyzed for its drug content to monitor its chemical stability. The shelf-life $(t_{0.9})$ of optimized nanoemulsion formulation was estimated to be 4.85 years at 25°C which reflected the ability of nanoemulsion formulation to maintain the drug and efficiently protected it against degradation.

To summarize, the studies conducted indicate the utility and potential advantage of AZO-loaded nanoemulsion system as a promising delivery carrier worth to explore further for its parenteral applicability in the treatment of bacterial meningitis.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PEMBENTUKAN DAN PENCIRIAN SISTEM NANOPENGHANTARAN SECARA PARENTERAL YANG MENGANDUNGI AZITROMISIN

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Sawar darah otak (BBB) terhasil oleh sel-sel endotelial mikrovaskular otak (BMEC) yang bekerja serentak dengan perisit dan astrosit, di mana simpang sempit dan beberapa pengangkut mengawal ketat penembusan pelbagai sebatian bioaktif ke dalam otak termasuk antibiotik. Ini mempunyai kesan yang signifikan untuk rawatan jangkitan sistem saraf utama (CNS), kerana ubat perlu mendapat akses kepada otak melalui BBB dalam kepekatan yang mencukupi untuk menjalankan aktiviti terapeutiknya. Azitromisin (AZO) adalah agen antibakteria yang boleh dipercayai dengan spektrum antimikrob yang luas dan profil farmakokinetik unik, namun, peranannya dalam rawatan jangkitan CNS adalah terhad kerana ia tidak mempunyai sifat-sifat fizikokimia yang sesuai yang membolehkan ia mencapai kuantiti yang mencukupi dalam tisu otak.

Sistem nanoemulsi adalah salah satu strategi yang berpotensi untuk penghantaran cekap lipofilik aktif ke seluruh BBB kerana mereka bersaiz nano, boleh serasi secara bio, mesra alam, mempunyai kestabilan fizikal yang unggul dan agak mudah untuk dihasilkan secara besar-besaran. Dengan tujuan untuk mensasarkan otak, AZO yang dimuatkan dengan nanoemulsi telah dibangunkan menggunakan homogenisasi tekanan tinggi dengan tekanan homogenisasi 1000 bar untuk 8 kitaran.

AZO yang dimuatkan dengan nanoemulsi telah dioptimumkan menggunakan rangkaian neural tiruan (ANN) sebagai teknik statistik multivariat. Bagi mencapai topologi yang optimum, ANN telah dilatih oleh Incremental Back-Propagation (IBP), Batch Back- Propagation (BBP), Quick Propagation (QP), dan algoritma Levenberg-Marquardt (LM) untuk menguji set data. Topologi telah disahkan oleh petunjuk kepada ralat punca purata kuasa dua (RMSE) minimum bagi setiap satu. Berdasarkan petunjuk itu, BBP-5-14-1 telah dipilih sebagai topologi yang optimum

untuk digunakan sebagai model akhir untuk meramalkan saiz zarah yang dikehendaki dan kepentingan relatif pembolehubah formulasi yang berkesan. Analisis ANN menunjukkan bahawa dengan komposisi optimum minyak kacang soya 6%, asid oleic 2%, AZO 1.4%, lesitin 2%, Tween 80 2%, gliserol 2.5%, vitamin E 0.25% dan air 83.85% menghasilkan saiz zarah yang minimum. Nanoemulsi yang dioptimumkan itu telah dinilai mengenai pelbagai sifat fizikokimia. Pencirian itu mendedahkan saiz zarah 54,67 \pm 0.81 nm, indeks polydispersity (PDI) daripada 0.218 \pm 0.023, keupayaan potensi zeta daripada - 34.65 \pm 0.78 mV, pH 7.82 \pm 0.07, kelikatan 1.77 \pm 0.05 cps, dan osmolaliti 288 \pm 1.00 mOsm / kg, menunjukkan keserasiannya bagi administrasi intravena. AZO telah berjaya dicampur ke dalam sistem nanoemulsi dengan kecekapan purata pengkapsulan 98.21 \pm 1.97% dan kandungan dadah agak tinggi 91.19 \pm 5.93%. Analisis morfologi dengan transmisi mikroskop elektron (TEM) mengesahkan pembentukan titisan minyak bersaiz nano berbentuk hampir sfera yang teragih secara seragam.

kajian pelepasan dadah *In vitro* bagi beberapa formulasi terpilih menunjukkan profil pelepasan sama dengan larutan AZO piawai, kedua-duanya mempamerkan ciri-ciri tingkah laku dwifasa, pelepasan awal dadah pengkapsulan yang cepat diikuti oleh pelepasan yang lebih perlahan yang berterusan sehingga ia mencapai pelepasan terkumpul jumlah dadah 84.94 ± 4.76% dalam tempoh 48 jam. Secara kinetiknya, profil pelepasan AZO daripada sistem nanoemulsi *in vitro* muncul terbaik sesuai dengan model Higuchi itu. Kestabilan nanoemulsi yang disediakan dengan formula optimum itu dinilai terutamanya dari segi pemeliharaan integriti fizikal, iaitu saiz zarah dan taburan saiz (PDI). Formulasi mengekalkan sifatsifatnya dalam julat yang memuaskan sehingga 12 bulan penyimpanan pada 4°C dan 25°C, yang menunjukkan kestabilan fizikal yang mencukupi untuk penyimpanan jangka panjang.

Hubungan linear antara saiz zarah jejari kuasa tiga fasa tersebar (r^3) dan masa (t) telah diperolehi mengenal pasti Ostwald ripening (OR) sebagai mekanisme ketidakstabilan dominan bagi nanoemulsi termuat AZO dari kajian jangka hayat 12 bulan. Kadar OR (ω) ekstrapolasi ditentukan secara graf dari kecerunan dan didapati ialah 0.232 × 10⁻⁸ nm³/s. kandungan dadah nanoemulsi optimum telah dianalisis untuk memantau kestabilan kimia. Jangka hayat ($t_{0.9}$) formulasi nanoemulsi optimum dianggarkan 4.85 tahun pada suhu 25°C yang menunjukkan keupayaan formulasi nanoemulsi untuk mengekalkan dadah dan cekap melindunginya terhadap pereputan.

Sebagai ringkasan, kajian yang dijalankan menunjukkan utiliti dan kelebihan potensi sistem nanoemulsi termuat AZO boleh dipercayai sebagai pembawa penghantaran yang bernilai untuk diterokai lebih untuk kegunaan parenteral dalam rawatan bakteria meningitis.

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- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

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LIST OF ABBREVIATIONS

AAD Absolute average deviation

AUC Area under the curve

ABC ATP- binding cassette transporters

ABM Acute bacterial meningitis

AMT Adsorptive-mediated transcytosis

ANN Artificial neural network

AZO Azithromycin

BBB Blood brain barrier

BBP Batch Back-Propagation

BCSFB Blood-cerebrospinal fluid barrier

CMC Critical micelle concentration

CNS Central nervous system

CSF Cerebrospinal fluid

CV Coefficient of variation

CYP3A4 Cytochrome P450 3A4

DLS Dynamic Light Scattering

DMSO Dimethyl sulfoxide

ECs Endothelial cells

ECM Extracellular matrix proteins

FDA Food and Drug Administration

GA Genetic algorithm

hCMEC/D₃ Immortalized human cerebral microvascular endothelial

cells

Hib Haemophilus influenza type b

HLB Hydrophilic-lipophilic balance

HPH High Pressure Homogenization

HS Human serum

IBP Incremental Back-Propagation

IC₅₀ Inhibitory concentration for 50 % inhibition

IgM Immunoglobulin-M

IL-1β Inter leukin-1beta

IL-6 Inter leukin-6

ISF Interstitial fluid

IV Intravenous

LCT Long chain triglyceride

LDL Low density lipoprotein

Levenberg-Marquardt

LSW Lifshitz Slyozov Wagner

MBC Minimal bactericidal concentration

MNS Mononuclear phagocyte system

MRSA Methicillin resistant Staphylococcus aureus

MTT 3-(4,5-Dimethlythiazol-2-yl)-2-5-diphenyltetrazolium

bromide

MWCO Molecular weight cutoff

NE Nanoemulsion

NO Nitric oxide

NVU Neurovascular unit

OFAT One-Factor-At-a-Time

O/W Oil-in-water

OR Ostwald ripening

PBS Phosphate buffer solution

PCS Photon Correlation Spectroscopy

PDI Polydispersity index

pH Negative logarithm of activity of hydronium ion

PMNS Polymorph nuclear system

QC Quality control

QP Quick Propagation

RMT Receptor-mediated transcytosis

rpm Revolution per minute

RMSE Root mean squared error

RSD Relative standard deviation

SD Standard deviation

TEM Transmission Electron Microscopy

TJs Tight junctions

TNF-α Tumor necrosis factor-alpha

UPLC Ultra-Performance Liquid Chromatography

VDW Van Der Waals

VRE Vancomycin resistant enterococci

W/O Water-in-oil

W/O/W Water-in-oil-in-water

w/w Weight by weight

LIST OF UNITS

cm Centimeter
cP Centipoise
Da Dalton

°C Degree Celsius

h Hour

μg
 μm
 Microgram
 μl
 Microliter
 mg
 Milligram
 ml
 Milliliter
 mm
 Millimeter
 mM
 Millimolar

mOsm/kg Milliosmole per kilogram

mV Millivolt
min Minute
m Month
nm Nanome

nm Nanometer % Percentage Pa Pascal

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

The pharmaceutical treatment of Central Nervous System (CNS) diseases is the second largest area of therapy following cardiovascular disorders. CNS disorders currently affect over 1.5 billion people worldwide and account for about one-third of the global disease burden being five of the top ten causes of disability. Many CNS disorders do not have satisfactory treatment and they are considered an important current and future priority for the pharmaceutical industry (Lewis et al., 2013). Nervous tissues are very soft and delicate and the irreplaceable neurons can be injured by even a slight pressure. Thus, the brain and the spinal cord have been protected by a bone enclosing them from the outside (the skull and the vertebral column), membranes (meninges), and a watery cushion (the cerebrospinal fluid (CSF)) from inside (Marieb & Hoehn, 2010). The protection of the brain from any harmful substances that could be circulating in the blood is provided by the blood brain barrier (BBB). The BBB acts very effectively to protect the brain from many common pathogens circulating in the systemic circulation, thus infections of the brain are not very common. However, since antibodies and antibiotics are too big to cross the BBB. infections of the brain that do occur are often very serious and too difficult to treat (Sandoval & Witt, 2008). One of these infections is meningitis.

Meningitis is an acute infection of the protective membranes surrounding the brain and spinal cord (meninges), usually due to the spread of an infection elsewhere in the body into the meninges and CSF, which will be followed by a CNS inflammatory reaction that causes coma, seizure, increased intracranial pressure and stroke. Meningitis occurs most commonly in young children under 5 years of old and people over 60 years (Alam et al., 2010). The most common cause of meningitis is viral infections, but bacterial and fungal infections may also cause meningitis. Although bacterial meningitis affect fewer populations, it is considered the most serious type of meningitis. It can cause severe brain damage and is fatal in 50% of the cases if not treated. Even when the disease is diagnosed early and proper treatment is started, 5 - 10% of the patients die, typically within 24 - 48 h after the onset of symptoms. Bacterial meningitis may result in brain damage, hearing loss or learning disability in 10 - 20% of the survivors (Fitch & van de Beek, 2007). A range of antibiotics can be used to treat meningitis including, ceftriaxone, ampicillin, penicillin G, chloramphenicol, oxacillin, gentamycin, vancomycin. But for an antibiotic to be effective in the treatment of meningitis, it must not only be active against the pathogenic bacterium, but it should also be able to penetrate the BBB and reach the meninges in sufficient quantities and stay there long enough to produce its antibacterial activity.

Delivery of drugs to the brain is a major challenge due to the presence of the BBB. Unlike the peripheral capillaries that allow relatively free exchange of substances across or between the cells, the BBB strictly limits the transport into the brain to maintain a stable environment for a proper function of the nervous tissues (Kaur et al., 2008). Thus the BBB is considered the rate limiting factor in determining permeation of therapeutic agents into the CNS (Patel et al., 2013). Many potential compounds, which are effective at their site of action, have failed during their development for the clinical use due to a failure in delivering them in an adequate quantities to the brain tissues to produce the therapeutic effect. It has been estimated that only 2% of the possible CNS therapeutic drugs can pass the BBB (Pardridge, 2002).

It has been generally anticipated that BBB disruption that occurs under inflammatory conditions may offer an opportunity for enhancement of drug transport into the brain via the paracellular route. This, however, has shown to be a more complex issue. Due to limited *in vivo* results obtained by different investigators under pathological conditions, the answer is not clear (Chen & Liu, 2012). Furthermore, several experimental models of bacterial meningitis in animals suggest that effective bacteriologic cure is associated with antibiotic concentration in CNS compartments that are 10 – 30 times the minimal bactericidal concentration (MBC) for a specific pathogen (Chávez-Bueno & McCracken, 2005). These bactericidal concentrations can only be achieved by administration of frequent and high systemic doses of antibiotics that might not be well tolerated by patients. Additionally, for some antibiotic families (such as aminoglycosides, glycopeptides), toxicity makes dose increase difficult to achieve.

As a broad-spectrum antibacterial, azithromycin (AZO) shares the same mechanism of action as other macrolide antibiotics and its range of activity is extended through inhibition of bacterial quorum-sensing and biofilm. Accumulating more effectively than other macrolides in cells, particularly circulating phagocytes, it is delivered in high concentrations to sites of infection. This important feature, combined with the extended plasma half-life of AZO, often allows effective single-dose administration for acute bacterial infections (Parnham et al., 2014). The antibacterial effects of AZO are facilitated by its ability to modulate inflammation and immunity in humans by influencing the production of cytokines, decreasing mucus hypersecretion, inhibiting chemotaxis of neutrophils, accelerating apoptosis (Bosnar et al., 2011). AZO is highly effective against two of the three main causative pathogens producing meningitis. Yet, its role in the treatment of CNS infections is very limited due to its relatively high molecular mass that restricts penetration through the BBB.

Numerous drug delivery and targeting strategies have been developed to circumvent the BBB. Disruption of the BBB, chemical modification, molecular antibody technology and various carrier systems have been tried to achieve the transport and the distribution of drugs into the brain (Burgess et al., 2014). One of the promising strategies to enhance the drug penetration to the brain is the

utilization of nanoemulsion as a drug carrier system. Nanoemulsion is a heterogeneous system composed of one immiscible liquid dispersed as droplets within another liquid, it is oil-in-water (O/W) or water-in-oil (W/O) emulsion where the average droplet diameter is between 20 - 200 nm (Solans et al., 2005). Many drugs are hydrophobic, which limits their water solubility and consequently limiting their bioavailability, making the delivery of water-insoluble drugs to be a primary focus of delivery research. Emulsions provide a central oil core dispersed in water that can act as a reservoir for hydrophobic drugs. Emulsions have long been used for the topical administration, yet, the small size of nanoemulsions make them attractive for parenteral delivery. In addition to solubilization of hydrophobic drugs, nanoemulsions can reduce pain and irritation at the site of injection, improve pharmacokinetics, allow for new forms of administration and can provide a sustained or targeted release of the drug (Lovelyn, 2011).

1.2 Problem Statements

General

- The presence of the BBB limits the penetration of a large number of pharmacologically active drugs aimed at treating CNS diseases, including meningitis. Several CNS-acting drugs are unable to cross the BBB and reach their site of action because they do not possess the appropriate physicochemical properties.
- 2. Thus, higher doses of the drug are needed in order to exhibit a therapeutically effective CNS concentrations and this might lead to a significant increase in the systemic side effects of the drug.

Specific

- 1. AZO is a broad spectrum antibiotic that possesses a unique pharmacokinetic profile, however, it cannot be used for the treatment of meningitis because it does not have the suitable physicochemical properties that enable it to penetrate the BBB; though it is lipophilic but it is a large molecule with a molecular mass of 749 g/mol. Thus, incorporating the drug into a nano-carrier system seems to be a promising strategy to deliver the drug to the brain.
- Optimal antibacterial activity for AZO is best achieved with high drug level at the site of infection thus loading the drug into nanoemulsion system can provide a good opportunity to increase the concentration of the drug delivered to the site of action in the brain.
- 3. The continuous increase in the incidence of infections caused by resistant bacteria poses a significant threat as it may lead to treatment failure and complications. Thus, using nanoemulsion as an antimicrobial drug delivery system has been found to be a promising strategy to overcome bacterial resistance.

1.3 Significance of the Study

Parenteral administration is the most effective route for drug application usually selected for actives with low bioavailability and narrow therapeutic index. Nanoemulsions with their capability to incorporate considerable quantities of lipophilic drugs, plus their combined biocompatibility and ability to protect drug from enzymatic hydrolysis and degradation make them best possible vehicles for parenteral administration. Moreover, the incidence and dosage of injections can be lessened throughout the period of drug therapy as these nanoemulsion systems offer a sustained and controlled release manner of the drug for extended periods of time (Thiagarajan, 2011). Additionally, the absence of flocculation, creaming, and sedimentation combined with a large interfacial area and free energy, provide additional advantages over traditional emulsions for this route of application.

In this study, nanoemulsion system loaded with AZO intended for parenteral administration was designed and developed. Several biocompatible compounds and various preparation methods were investigated to produce a nano-sized carrier system for this antibiotic. Formulation optimization was conducted using Artificial Neural Network (ANN) method. The formulated nanoemulsion was characterized with respect to particle size, polydispersity index, zeta potential, viscosity, osmolality, morphological study, entrapment efficacy, drug content, toxicity, and *in vitro* drug release kinetic. Long-term stability assessment of the formulated nanoemulsion was studied for the evaluation of both chemical and physical stability.

1.4 Objectives

General

The aim of this study was to formulate a nanoemulsion system that can successfully deliver large and highly localized concentrations of AZO to the brain efficiently so that it can decrease the administration dose and minimize the systemic side effects and drug toxicity.

Specific

- 1. To develop oil-in-water (O/W) nanoemulsion system loaded with AZO for parenteral drug delivery by means of low and high energy emulsification methods.
- 2. To optimize the compositions for formulating nanoemulsion system in nano-sized range and good stability utilizing ANN.
- 3. To characterize the physicochemical properties of the formulated nanoemulsion in terms of particle size, polydispersity index, zeta potential, viscosity, pH, osmolality, and morphology. And to assess drug content, entrapment efficacy, *in vitro* drug release, and toxicity of the formulated nanoemulsion.
- 4. To evaluate the long-term stability of the formulated nanoemulsion with respect to physical and chemical stability with time at various temperatures and storage conditions.

REFERENCES

- Abbott, N. J. (2004). Evidence for bulk flow of brain interstitial fluid: Significance for physiology and pathology. *Neurochemistry International*, *45*(4), 545–52.
- Abbott, N. J. (2005). Physiology of the blood-brain barrier and its consequences for drug transport to the brain. *International Congress Series*, *1277*, 3–18.
- Abbott, N. J., Patabendige, A. A. K., Dolman, D. E. M., Yusof, S. R., & Begley, D. J. (2010). Structure and function of the blood-brain barrier. *Neurobiology of Disease*, 37,13-25.
- Abdollahi, Y., Sairi, N. A., Aroua, M. K., Masoumi, H. R. F., Jahangirian, H., & Alias, Y. (2014). Fabrication modeling of industrial CO₂ ionic liquids absorber by artificial neural networks. *Journal of Industrial and Engineering Chemistry*, 25, 168-175.
- Abdollahi, Y., Zakaria, A., Abbasiyannejad, M., Masoumi, H. R. F., Moghaddam, M. G., Matori, K. A., Keshavarzi, A. (2013). Artificial neural network modeling of p-cresol photodegradation. *Chemistry Central Journal*, 7, 96.
- Agatonovic-Kustrin, S., & Beresford, R. (2000). Basic concepts of artificial neural network (ANN) modeling and its application in pharmaceutical research. Journal of Pharmaceutical and Biomedical Analysis, 22(5), 717–727.
- Agrawal, S., & Nadel, S. (2011). Acute bacterial meningitis in infants and children: epidemiology and management. *Paediatric Drugs*, *13*(6), 385–400.
- Ahuja, N., Katare, O. P., & Singh, B. (2007). Studies on dissolution enhancement and mathematical modeling of drug release of a poorly water-soluble drug using water-soluble carriers. *European Journal of Pharmaceutics and Biopharmaceutics*, *65*(1), 26–38.
- Alam, M. I., Beg, S., Samad, A., Baboota, S., Kohli, K., Ali, J. Akbar, M. (2010). Strategy for effective brain drug delivery. *Eur. J. Pharm. Sci.*, *40*(5), 385–403.
- Alexis, F., Pridgen, E., Molnar, L. K., & Farokhzad, O. C. (2008). Factors affecting the clearance and biodistribution of polymeric nanoparticles. *Molecular Pharmaceutics*, 5, 505–515).
- Alyautdin, R. N., Petrov, V. E., Langer, K., Berthold, A., Kharkevich, D. A., & Kreuter, J. (1997). Delivery of loperamide across the blood-brain barrier with polysorbate 80-Coated polybutylcyanoacrylate nanoparticles. *Pharmaceutical Research*, *14*(3), 325–328.

- Amani, A., York, P., Chrystyn, H., & Clark, B. J. (2010). Factors affecting the stability of nanoemulsions-use of artificial neural networks. *Pharmaceutical Research*, *27*, 37–45.
- Amsden, G. W. (2001). Advanced-generation macrolides: tissue-directed antibiotics. *International Journal of Antimicrobial Agents*, *18*, 11–5.
- Anton, N., Benoit, J. P., & Saulnier, P. (2008). Design and production of nanoparticles formulated from nano-emulsion templates. A review. *Journal of Controlled Release*, 128(3), 185–199.
- Aramaki, K., Olsson, U., Yamaguchi, Y., & Kunieda, H. (1999). Effect of water-soluble alcohols on surfactant aggregation in the C12EO8 system. *Langmuir*, (16), 6226–6232.
- Aramaki, K., & Solans, C. (2012). Emulsions and microemulsions. *Current Opinion in Colloid and Interface Science*, *17*(5), 233–234.
- Araújo, F. A., Kelmann, R. G., Araújo, B. V., Finatto, R. B., Teixeira, H. F., & Koester, L. S. (2011). Development and characterization of parenteral nanoemulsions containing thalidomide. *European Journal of Pharmaceutical Sciences*, 42, 238–245.
- Araújo, F. a., Kelmann, R. G., Araújo, B. V., Finatto, R. B., Teixeira, H. F., & Koester, L. S. (2011). Development and characterization of parenteral nanoemulsions containing thalidomide. *European Journal of Pharmaceutical Sciences*, 42(3), 238–245.
- Araújo, J., Gonzalez-Mira, E., Egea, M. a., Garcia, M. L., & Souto, E. B. (2010). Optimization and physicochemical characterization of a triamcinolone acetonide-loaded NLC for ocular antiangiogenic applications. *International Journal of Pharmaceutics*, *393*(1-2), 168–176.
- Arifin, D. Y., Lee, L. Y., & Wang, C. H. (2006). Mathematical modeling and simulation of drug release from microspheres: Implications to drug delivery systems. *Advanced Drug Delivery Reviews*, *58*(12-13), 1274–325.
- Arora, G., Malik, K., Singh, I., Arora, S., & Rana, V. (2011). Formulation and evaluation of controlled release matrix mucoadhesive tablets of domperidone using Salvia plebeian gum. *Journal of Advanced Pharmaceutical Technology & Research*, 2(3), 163–9.
- Azeem, A., Rizwan, M., Ahmad, F. J., Iqbal, Z., Khar, R. K., Aqil, M., & Talegaonkar, S. (2009). Nanoemulsion components screening and selection: a technical note. *AAPS PharmSciTech*, *10*(1), 69–76.
- Bali, V., Ali, M., & Ali, J. (2010a). Novel nanoemulsion for minimizing variations in bioavailability of ezetimibe. *Journal of Drug Targeting*, *18*(7), 506–19.

- Bali, V., Ali, M., & Ali, J. (2010b). Study of surfactant combinations and development of a novel nanoemulsion for minimising variations in bioavailability of ezetimibe. *Colloids and Surfaces B: Biointerfaces*, 76(2), 410–420.
- Bali, V., Ali, M., & Ali, J. (2011a). Nanocarrier for the enhanced bioavailability of a cardiovascular agent: In vitro, pharmacodynamic, pharmacokinetic and stability assessment. *International Journal of Pharmaceutics*, 403, 45–56.
- Bali, V., Ali, M., & Ali, J. (2011b). Nanocarrier for the enhanced bioavailability of a cardiovascular agent: in vitro, pharmacodynamic, pharmacokinetic and stability assessment. *International Journal of Pharmaceutics*, 403(1-2), 46–56.
- Basri, M., Rahman, R. N. Z. R. A., Ebrahimpour, A., Salleh, A. B., Gunawan, E. R., & Rahman, M. B. A. (2007). Comparison of estimation capabilities of response surface methodology (RSM) with artificial neural network (ANN) in lipase-catalyzed synthesis of palm-based wax ester. *BMC Biotechnology*, 7(1), 53.
- Begley, D. J. (2004). Delivery of therapeutic agents to the central nervous system: the problems and the possibilities. *Pharmacol Ther*, *104*(1), 29–45.
- Bernacki, J., Dobrowołska, A., Nerwińska, K., & Małecki, A. (2008). Physiology and pharmacological role of the blood-brain barrier. *Pharmacological Reports*, 60, 600-622.
- Bernardi, D. S., Pereira, T. A., Maciel, N. R., Bortoloto, J., Viera, G. S., Oliveira, G. C., & Rocha-Filho, P. A. (2011). Formation and stability of oil-in-water nanoemulsions containing rice bran oil: in vitro and in vivo assessments. *Journal of Nanobiotechnology*, 167, 16-23.
- Borhade, V., Pathak, S., Sharma, S., & Patravale, V. (2012). Clotrimazole nanoemulsion for malaria chemotherapy. Part I: preformulation studies, formulation design and physicochemical evaluation. *International Journal of Pharmaceutics*, *431*(1-2), 138–48.
- Bosnar, M., Čužić, S., Bošnjak, B., Nujić, K., Ergović, G., Marjanović, N., Haber, V. E. (2011). Azithromycin inhibits macrophage interleukin-1β production through inhibition of activator protein-1 in lipopolysaccharide-induced murine pulmonary neutrophilia. *International Immunopharmacology*, *11*, 424–434.
- Bouchemal, K., Briançon, S., Perrier, E., & Fessi, H. (2004). Nano-emulsion formulation using spontaneous emulsification: Solvent, oil and surfactant optimisation. *International Journal of Pharmaceutics*, 280(1-2), 241–251.

- Bourquin, J., Schmidli, H., Van Hoogevest, P., & Leuenberger, H. (1997). Application of Artificial Neural Networks (ANN) in the development of solid dosage forms. *Pharmaceutical Development and Technology*, 2(2), 111–121.
- Brouwer, M. C., Tunkel, A. R., & Van De Beek, D. (2010). Epidemiology, diagnosis, and antimicrobial treatment of acute bacterial meningitis. *Clinical Microbiology Reviews*, *23*(3), 467–492.
- Brunton, L. (2003). Goodman & Gilman 's The Pharmacological Basis of Therapeutics. In *Goodman & Gilman*'s The Pharmacological Basis of Therapeutics, 9–10.
- Burgess, A., Nhan, T., Moffatt, C., Klibanov, a L., & Hynynen, K. (2014). Analysis of focused ultrasound-induced blood-brain barrier permeability in a mouse model of Alzheimer's disease using two-photon microscopy. *Journal of Controlled Release: Official Journal of the Controlled Release Society*, 192C, 243–248.
- Capek, I. (2004). Degradation of kinetically-stable o/w emulsions. Advances in Colloid and Interface Science, 107(2-3), 125–155.
- Capen, R., Christopher, D., Forenzo, P., Ireland, C., Liu, O., Lyapustina, S., Tougas, T. (2012). On the Shelf Life of Pharmaceutical Products. *AAPS PharmSciTech*, 13(3), 911–918.
- Cardoso, F. L., Brites, D., & Brito, M. A. (2010). Looking at the blood-brain barrier: Molecular anatomy and possible investigation approaches. *Brain Research Reviews*, *64*(2), 328–363.
- Chanasattru, W., Decker, E. A., & McClements, D. J. (2007). Physicochemical basis for cosolvent modulation of lactoglobulin functionality: Interfacial tension study. *Food Research International*, *40*(8), 1098–1105.
- Chang, Y., McLandsborough, L., & McClements, D. J. (2012). Physical properties and antimicrobial efficacy of thyme oil nanoemulsions: Influence of ripening inhibitors. *Journal of Agricultural and Food Chemistry*, 60, 12056–12063.
- Chaudhuri, P., Harfouche, R., Soni, S., Hentschel, D. M., & Sengupta, S. (2010). Shape effect of carbon nanovectors on angiogenesis. *ACS Nano*, *4*(1), 574–582.
- Chávez-Bueno, S., & McCracken, G. H. (2005). Bacterial meningitis in children. *Pediatric Clinics of North America*, 52, 795-810.
- Chen, B.-M., Liang, Y.-Z., Chen, X., Liu, S.-G., Deng, F.-L., & Zhou, P. (2006). Quantitative determination of azithromycin in human plasma by liquid chromatography-mass spectrometry and its application in a

- bioequivalence study. *Journal of Pharmaceutical and Biomedical Analysis*, 42(4), 480–487.
- Chen, Y., & Liu, L. (2012). Modern methods for delivery of drugs across the blood-brain barrier. *Advanced Drug Delivery Reviews*, *64*(7), 640–665.
- Choi, H. S., Liu, W., Misra, P., Tanaka, E., Zimmer, J. P., Itty Ipe, B., Frangioni, J. V. (2007). Renal clearance of quantum dots. *Nature Biotechnology*, 25(10), 1165–1170.
- Chopra, D., Gulati, M., Saluja, V., Pathak, P., & Bansal, P. (2008). Brain permeable nanoparticles. *Recent Patents on CNS Drug Discovery*, *3*(3), 216–25.
- Constantinides, P. P., Chaubal, M. V., & Shorr, R. (2008). Advances in lipid nanodispersions for parenteral drug delivery and targeting. *Advanced Drug Delivery Reviews*, 60, 757-767.
- Costa, P., & Sousa Lobo, J. M. (2001). Modeling and comparison of dissolution profiles. *European Journal of Pharmaceutical Sciences*, 13, 123-133.
- Coutts, B. E., & LaGuardia, C. (2006). Encyclopedia of Science, Technology and Ethics. *Library Journal*, 131(7), 49.
- D'Errico, G., Ciccarelli, D., & Ortona, O. (2005). Effect of glycerol on micelle formation by ionic and nonionic surfactants at 25 degrees C. *Journal of Colloid and Interface Science*, 286(2), 747–754.
- Daneman, R., & Rescigno, M. (2009). The Gut Immune Barrier and the Blood-Brain Barrier: Are They So Different? *Immunity*, 31(5), 722–735.
- Dash, S., Murthy, P. N., Nath, L., & Chowdhury, P. (2010a). Kinetic modeling on drug release from controlled drug delivery systems. *Acta Poloniae Pharmaceutica*, 67(3), 217–23.
- Dash, S., Murthy, P. N., Nath, L., & Chowdhury, P. (2010b). Kinetic modeling on drug release from controlled drug delivery systems. *Acta Poloniae Pharmaceutica*, *67*(3), 217–23.
- Date, A. A., & Nagarsenker, M. S. (2007). Design and evaluation of selfnanoemulsifying drug delivery systems (SNEDDS) for cefpodoxime proxetil. *International Journal of Pharmaceutics*, 329, 166–172.
- De Boer, A. G., & Gaillard, P. J. (2007). Strategies to improve drug delivery across the blood-brain barrier. *ClinPharmacokinet*, *56*(3), 553 –576.
- Decuzzi, P., Godin, B., Tanaka, T., Lee, S. Y., Chiappini, C., Liu, X., & Ferrari, M. (2010). Size and shape effects in the biodistribution of intravascularly injected particles. *Journal of Controlled Release*, *141*(3), 320–327.

- Desai, M. P., Labhasetwar, V., Walter, E., Levy, R. J., & Amidon, G. L. (1997). The mechanism of uptake of biodegradable microparticles in Caco-2 cells is size dependent. *Pharmaceutical Research*, 14, 1568-1573.
- Dinos, G. P., Michelinaki, M., & Kalpaxis, D. L. (2001). Insights into the mechanism of azithromycin interaction with an Escherichia coli functional ribosomal complex. *Molecular Pharmacology*, *59*(6), 1441–5.
- Djordjevic, S. M., Cekic, N. D., Isailovic, T. M., Milic, J. R., Vuleta, G. M., Lazic, M. L., & Savic, S. D. (2013). Nanoemulsions produced by varying the type of emulsifier and oil content: effect of formulation and process parameters on the characteristics and physical stability. *Hemijska Industrija*, *67*(5), 795–809.
- Djukic, M., Munz, M., Sörgel, F., Holzgrabe, U., Eiffert, H., & Nau, R. (2012). Overton's rule helps to estimate the penetration of anti-infectives into patients' cerebrospinal fluid. *Antimicrobial Agents and Chemotherapy*, 56(2), 979–988.
- Donaldson, K., & Poland, C. A. (2012). Inhaled nanoparticles and lung cancer What we can learn from conventional particle toxicology. *Swiss Med Wkly*, 142, 1–9.
- Dordević, S. M., Radulović, T. S., Cekić, N. D., Randelović, D. V., Savić, M. M., Krajišnik, D. R., Savić, S. D. (2013). Experimental design in formulation of diazepam nanoemulsions: Physicochemical and pharmacokinetic performances. *Journal of Pharmaceutical Sciences*, 102(11), 4159–4172.
- Doshi, N., Prabhakarpandian, B., Rea-Ramsey, A., Pant, K., Sundaram, S., & Mitragotri, S. (2010). Flow and adhesion of drug carriers in blood vessels depend on their shape: A study using model synthetic microvascular networks. *Journal of Controlled Release*, *146*(2), 196–200.
- Driscoll, D. F. (2006). Lipid injectable emulsions: Pharmacopeial and safety issues. *Pharmaceutical Research*, 23, 1959-1969.
- Dumitrescu, T. P., Anic-Milic, T., Oreskovic, K., Padovan, J., Brouwer, K. L. R., Zuo, P., & Schmith, V. D. (2013). Development of a Population Pharmacokinetic Model to Describe Azithromycin Whole Blood and Plasma Concentrations Over Time in Healthy Subjects. *Antimicrobial Agents and Chemotherapy*, 57, 3194-3201.
- Edmond, K., Clark, A., Korczak, V. S., Sanderson, C., Griffiths, U. K., & Rudan, I. (2010). Global and regional risk of disabling sequelae from bacterial meningitis: A systematic review and meta-analysis. *The Lancet Infectious Diseases*, 10(5), 317–328.
- Eid, A. M. M., Elmarzugi, N. A., & El-Enshasy, H. A. (2013). Preparation and evaluation of olive oil nanoemulsion using sucrose monoester.

- International Journal of Pharmacy and Pharmaceutical Sciences, 5, 434–440.
- Elsheikh, M. a., Elnaggar, Y. S. R., Gohar, E. Y., & Abdallah, O. Y. (2012). Nanoemulsion liquid preconcentrates for raloxifene hydrochloride: Optimization and in vivo appraisal. *International Journal of Nanomedicine*, 7, 3787–3802.
- Engelhardt, B., & Wolburg, H. (2004). Mini review: Transendothelial migration of leukocytes: Through the front door or around the side of the house? *European Journal of Immunology*, *34*(11), 2955–2963.
- England, C. G., Clarke Miller, M., Kuttan, A., Trent, J. O., & Frieboes, H. B. (2015). Release kinetics of paclitaxel and cisplatin from two and three layered gold nanoparticles. European Journal of Pharmaceutics and Biopharmaceutics: Official Journal of Arbeitsgemeinschaft Fur Pharmazeutische Verfahrenstechnik e.V, 92, 120–129.
- Evjen, T. J., Hupfeld, S., Barnert, S., Fossheim, S., Schubert, R., & Brandl, M. (2013). Physicochemical characterization of liposomes after ultrasound exposure Mechanisms of drug release. *Journal of Pharmaceutical and Biomedical Analysis*, 78-79, 118–122.
- Faiyazuddin, M., Akhtar, N., Akhter, J., Suri, S., Shakeel, F., Shafiq, S., & Mustafa, G. (2010). Production, characterization, in vitro and ex vivo studies of babchi oil-encapsulated nanostructured solid lipid carriers produced by a hot aqueous titration method. *Pharmazie*, *65*(5), 348–355.
- Fard Masoumi, H. R., Basri, M., Kassim, A., Abdullah, D. K., Abdollahi, Y., Gani, S. S. A., & Rezaee, M. (2014). Optimization of process parameters for lipase-catalyzed synthesis of esteramines-based esterquats using wavelet neural network (WNN) in 2-liter bioreactor. *Journal of Industrial and Engineering Chemistry*, 20(4), 1973–1976.
- Fathi, M., Mozafari, M. R., & Mohebbi, M. (2012). Nanoencapsulation of food ingredients using lipid based delivery systems. *Trends in Food Science and Technology*, 23(1), 13–27.
- Fernandes, C., Soni, U., & Patravale, V. (2010). Nano-interventions for neurodegenerative disorders. *Pharmacological Research*, *62*(2), 166–178.
- Fernandez, P., André, V., Rieger, J., & Kühnle, A. (2004). Nano-emulsion formation by emulsion phase inversion. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, *251*(1-3), 53–58.
- Fitch, M. T., & van de Beek, D. (2007). Emergency diagnosis and treatment of adult meningitis. *Lancet Infectious Diseases*, 7(3), 191–200.

- Floyd, A. G. (1999). Top ten considerations in the development of parenteral emulsions. *Pharmaceutical Science and Technology Today*, 2, 134-143.
- Fredrick, E., Walstra, P., & Dewettinck, K. (2010). Factors governing partial coalescence in oil-in-water emulsions. *Advances in Colloid and Interface Science*, 153, 30-42.
- Freiberg, S., & Zhu, X. X. (2004). Polymer microspheres for controlled drug release. *International Journal of Pharmaceutics*, 282, 1-18.
- Fu, Y., & Kao, W. J. (2010). Drug release kinetics and transport mechanisms of non-degradable and degradable polymeric delivery systems. *Expert Opinion on Drug Delivery*, 7, 429–444.
- Gal-yam, B., & Eldridge, J. J. (2014). Double Function at the blood-brain barrier. *Nature*, *509*, 432–434.
- Gao, K., & Jiang, X. (2006). Influence of particle size on transport of methotrexate across blood brain barrier by polysorbate 80-coated polybutylcyanoacrylate nanoparticles. *International Journal of Pharmaceutics*, 310(1-2), 213–219.
- Ghaffari, a., Abdollahi, H., Khoshayand, M. R., Bozchalooi, I. S., Dadgar, a., & Rafiee-Tehrani, M. (2006). Performance comparison of neural network training algorithms in modeling of bimodal drug delivery. *International Journal of Pharmaceutics*, 327(1-2), 126–138.
- Gharibzahedi, S. M. T., Mousavi, S. M., Hamedi, M., & Ghasemlou, M. (2012). Response surface modeling for optimization of formulation variables and physical stability assessment of walnut oil-in-water beverage emulsions. *Food Hydrocolloids*, *26*(1), 293–301.
- Gladue, R., & Snider, M. (1990). Intracellular Accumulation of Azithromycin by Cultured Human Fibroblasts. *Antimicrobial Agents and Chemotherapy*, 34(6), 1056–1060.
- Gnanasekaran, K., Snel, R., de With, G., & Friedrich, H. (2016). Quantitative nanoscopy: Tackling sampling limitations in (S) TEM imaging of polymers and composites. *Ultramicroscopy*, *160*, 130–139.
- Gokce, E. H., Sandri, G., Bonferoni, M. C., Rossi, S., Ferrari, F., Güneri, T., & Caramella, C. (2008). Cyclosporine A loaded SLNs: Evaluation of cellular uptake and corneal cytotoxicity. *International Journal of Pharmaceutics*, 364, 76–86.
- Guerra-Rosas, M. I., Morales-Castro, J., Ochoa-Martínez, L. A., Salvia-Trujillo, L., Martín-Belloso, O., Pinheiro, A. C., Vicente, A. a. (2016). Long-term stability of food-grade nanoemulsions from high methoxyl pectin containing essential oils. *Food Hydrocolloids*, *52*, 460–467.

- Gulyaev, A. E., Gelperina, S. E., Skidan, I. N., Antropov, A. S., Kivman, G. Y., & Kreuter, J. (1999). Significant transport of doxorubicin into the brain with polysorbate 80-coated nanoparticles. *Pharmaceutical Research*, *16*(10), 1564–1569.
- Gutiérrez, J. M., González, C., Maestro, a., Solè, I., Pey, C. M., & Nolla, J. (2008). Nano-emulsions: New applications and optimization of their preparation. *Current Opinion in Colloid & Interface Science*, *13*(4), 245–251.
- Håkansson, A., Trägårdh, C., & Bergenståhl, B. (2009). Dynamic simulation of emulsion formation in a high pressure homogenizer. *Chemical Engineering Science*, *64*, 2915–2925.
- Hall, F. R., Ross Hallett, F., & Rosshallett, F. (1994). Particle size analysis by dynamic light scattering. *Food Research International*, *27*(2), 195–198.
- Hansen, J. L., Ippolito, J. A., Ban, N., Nissen, P., Moore, P. B., & Steitz, T. A. (2002). The Structures of Four Macrolide Antibiotics Bound to the Large Ribosomal Subunit. *Molecular Cell*, 10(1), 117–128.
- Hatanaka, J., Chikamori, H., Sato, H., Uchida, S., Debari, K., Onoue, S., & Yamada, S. (2010). Physicochemical and pharmacological characterization of alpha-tocopherol-loaded nano-emulsion system. *International Journal of Pharmaceutics*, *396*(1-2), 188–193.
- Hawkins, B. T., & Davis, T. P. (2005). The Blood-Brain Barrier / Neurovascular Unit in Health and Disease. *Pharmacological Reviews*, *57*(2), 173–185.
- Henry, J. V. L., Fryer, P. J., Frith, W. J., & Norton, I. T. (2010). The influence of phospholipids and food proteins on the size and stability of model submicron emulsions. *Food Hydrocolloids*, 24(1), 66–71.
- Hobson, M., & Kaplan, J. (2014). Pediatric Critical Care Medicine. *Pediatric Critical Care Medicinems*, 3, 57–68.
- Hoepelman, I. M., & Schneider, M. M. E. (1995). Azithromycin: The first of the tissue-selective azalides. *International Journal of Antimicrobial Agents*, 5, 145-167.
- Hofer, S., Bopp, C., Hoerner, C., Plaschke, K., Faden, R. M., Martin, E., Weigand, M. a. (2008). Injury of the Blood Brain Barrier and Up-Regulation of ICAM-1 in Polymicrobial Sepsis. *Journal of Surgical Research*, 146(2), 276–281.
- Holzer, M., Barnert, S., Momm, J., & Schubert, R. (2009). Preparative size exclusion chromatography combined with detergent removal as a versatile tool to prepare unilamellar and spherical liposomes of highly uniform size distribution. *Journal of Chromatography A*, 1216(31), 5838–5848.

- Hoshino, Y., Koide, H., Furuya, K., Haberaecker, W. W., Lee, S. H., Kodama, T., Shea, K. J. (2012). The rational design of a synthetic polymer nanoparticle that neutralizes a toxic peptide in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, 109(1), 33–38.
- Huang, S. H., & Jong, A. Y. (2001). Cellular mechanisms of microbial proteins contributing to invasion of the blood-brain barrier. *Cellular Microbiology*, 3(5), 277–287.
- Innis, S. M. (2008). Dietary omega 3 fatty acids and the developing brain. *Brain Research*, 1237, 35-43.
- Jafari, S. M., Assadpoor, E., He, Y., & Bhandari, B. (2008). Re-coalescence of emulsion droplets during high-energy emulsification. *Food Hydrocolloids*, 22, 1191-1202.
- Jafri, R. Z., Ali, A., Messonnier, N. E., Tevi-Benissan, C., Durrheim, D., Eskola, J., Abramson, J. (2013). Global epidemiology of invasive meningococcal disease. *Population Health Metrics*, *11*(1), 17.
- Jaruratanasirikul, S., Hortiwakul, R., Tantisarasart, T., Phuenpathom, N., & Tussanasunthornwong, S. (1996). Distribution of azithromycin into brain tissue, cerebrospinal fluid, and aqueous humor of the eye. *Antimicrobial Agents and Chemotherapy*, *40*, 825–826.
- Jiao, J., & Burgess, D. J. (2003). Ostwald ripening of water-in-hydrocarbon emulsions. *Journal of Colloid and Interface Science*, 264(2), 509–516.
- Julian McClements, D., Henson, L., Popplewell, L. M., Decker, E. A., & Jun Choi, S. (2012). Inhibition of Ostwald ripening in model beverage emulsions by addition of poorly water soluble triglyceride oils. *Journal of Food Science*, 77(1), 65-79.
- Jünemann, D., & Dressman, J. (2012). Analytical methods for dissolution testing of nanosized drugs. *The Journal of Pharmacy and Pharmacology*, *64*(7), 931–43.
- Kabalnov, A. (2001). Ostwald ripening and related phenomena. *Journal of Dispersion Science and Technology*, 22, 1-12.
- Kalam, M., Humayun, M., Parvez, N., & Yadav, S. (2007). Release kinetics of modified pharmaceutical dosage forms: a review. *Continental J. Pharmaceutical Sciences*, 1, 30–35.
- Kasanmoentalib, E. S., Brouwer, M. C., & van de Beek, D. (2013). Update on bacterial meningitis: epidemiology, trials and genetic association studies. *Current Opinion in Neurology*, 26(3), 282–8.

- Kaur, I. P., Bhandari, R., Bhandari, S., & Kakkar, V. (2008). Potential of solid lipid nanoparticles in brain targeting. *Journal of Controlled Release*: Official Journal of the Controlled Release Society, 127(2), 97–109.
- Keck, C. M., Jansch, M., & Müller, R. H. (2013). Protein adsorption patterns and analysis on IV nanoemulsions-the key factor determining the organ distribution. *Pharmaceutics*, *5*(1), 36–68.
- Khataee, A. R. (2009). Photocatalytic removal of C.I. Basic Red 46 on immobilized TiO2 nanoparticles: artificial neural network modelling. *Environmental Technology*, *30*(11), 1155–68.
- Kim, K. S. (2008). Mechanisms of microbial traversal of the blood-brain barrier. *Nature Reviews. Microbiology*, *6*(8), 625–634.
- Klang, V., Matsko, N. B., Valenta, C., & Hofer, F. (2012). Electron microscopy of nanoemulsions: An essential tool for characterisation and stability assessment. *Micron*, *43*(2-3), 85–103.
- Klang, V., & Valenta, C. (2011). Lecithin-based nanoemulsions. *Journal of Drug Delivery Science and Technology*, 21, 55-76.
- Klang, V., Valenta, C., & Matsko, N. B. (2013a). Electron microscopy of pharmaceutical systems. *Micron*, *44*(1), 45–74.
- Klek, S., Chambrier, C., Singer, P., Rubin, M., Bowling, T., Staun, M., Shaffer, J. (2013). Four-week parenteral nutrition using a third generation lipid emulsion (SMOFlipid) A double-blind, randomised, multicentre study in adults. *Clinical Nutrition*, 32(2), 224–231.
- Klement, W., & Arndt, J. O. (1991). Pain on i.v. injection of some anaesthetic agents is evoked by the unphysiological osmolality or pH of their formulations. *British Journal of Anaesthesia*, *66*, 189–195.
- Komaiko, J., & McClements, D. J. (2014). Optimization of isothermal low-energy nanoemulsion formation: Hydrocarbon oil, non-ionic surfactant, and water systems. *Journal of Colloid and Interface Science*, *425*, 59–66.
- Korsmeyer, R. W., Gurny, R., Doelker, E., Buri, P., & Peppas, N. A. (1983a). Mechanisms of solute release from porous hydrophilic polymers. *International Journal of Pharmaceutics*, *15*(1), 25–35.
- Korsmeyer, R. W., Gurny, R., Doelker, E., Buri, P., & Peppas, N. a. (1983b). Mechanisms of solute release from porous hydrophilic polymers. *International Journal of Pharmaceutics*, *15*(1), 25–35.
- Krishnamachari, P., Hashaikeh, R., & Tiner, M. (2011). Modified cellulose morphologies and its composites; SEM and TEM analysis. *Micron*, 42, 751-761.

- Kristl, J., Teskac, K., Milek, M., & Mlinaric-Rascan, I. (2008). Surface active stabilizer Tyloxapol in colloidal dispersions exerts cytostatic effects and apoptotic dismissal of cells. *Toxicology and Applied Pharmacology*, 232, 218–225.
- Krogh, A. (2008). What are artificial neural networks? *Nature Biotechnology*, *26*, 195–197.
- Krol, S. (2012). Challenges in drug delivery to the brain: nature is against us. Journal of Controlled Release: Official Journal of the Controlled Release Society, 164(2), 145–55.
- Leal-calderon, F., & Cansell, M. (2012). The design of emulsions and their fate in the body following enteral and parenteral routes. *Soft Matter*, 8(40), 10213–10225.
- Lewis, K. M., Turner, R. J., & Vink, R. (2013). Blocking neurogenic inflammation for the treatment of acute disorders of the central nervous system. *International Journal of Inflammation*, 2013, 578480.
- Li, J., Cheng, J., Shi, J., & Huang, F. (2012). Brief Introduction of Back Propagation (BP) Neural Description of BP Algorithm in Mathematics, 2, 553–558.
- Li, S. D., & Huang, L. (2009). Nanoparticles evading the reticuloendothelial system: Role of the supported bilayer. *Biochimica et Biophysica Acta-Biomembranes*, 1788(10), 2259–2266.
- Liao, Y., & Lucas, D. (2009). A literature review of theoretical models for drop and bubble breakup in turbulent dispersions. *Chemical Engineering Science*, *64*(15), 3389–3406.
- Lide, D. R. (2007). CRC Handbook of Chemistry and Physics. Boca Raton, Fla.
- Lifshitz, I. M., & Slyozov, V. V. (1961a). The kinetics of precipitation from supersaturated solid solutions. *Journal of Physics and Chemistry of Solids*, 19, 35-50.
- Lifshitz, I. M., & Slyozov, V. V. (1961b). The kinetics of precipitation from supersaturated solid solutions. *Journal of Physics and Chemistry of Solids*, *19*(1-2), 35–50.
- Liu, D., Mori, A., & Huang, L. (1992). Role of liposome size and RES blockade in controlling biodistribution and tumor uptake of GM1-containing liposomes. *Biochimica et Biophysica Acta*, *1104*(1), 95–101.
- Liu, J., Lu, G. W., Sandoval, M., Ciringh, Y., Xue, G., Jaeger, D., Gelotte, K. M. (2009). Determination of benzalkonium chloride partition in micelle solutions using ultrafiltration method. *AAPS PharmSciTech*, *10*(4), 1216–23.

- Liu, P., Fang, A. F., LaBadie, R. R., Crownover, P. H., & Arguedas, A. G. (2011). Comparison of azithromycin pharmacokinetics following single oral doses of extended-release and immediate-release formulations in children with acute otitis media. *Antimicrobial Agents and Chemotherapy*, *55*(11), 5022–6.
- Liu, W., Sun, D., Li, C., Liu, Q., & Xu, J. (2006a). Formation and stability of paraffin oil-in-water nano-emulsions prepared by the emulsion inversion point method. *Journal of Colloid and Interface Science*, 303(2), 557–563.
- Liu, W., Sun, D., Li, C., Liu, Q., & Xu, J. (2006b). Formation and stability of paraffin oil-in-water nano-emulsions prepared by the emulsion inversion point method. *Journal of Colloid and Interface Science*, 303, 557–563.
- Liyana-Pathirana, C., & Shahidi, F. (2005). Optimization of extraction of phenolic compounds from wheat using response surface methodology. *Food Chemistry*, *93*(1), 47–56.
- Lovelyn, C. (2011). Current State of Nanoemulsions in Drug Delivery. *Journal of Biomaterials and Nanobiotechnology*, *0*2(05), 626–639.
- Ma, C. G., & Weng, H. X. (2009). Application of Artificial Neural Network in the Residual Oil Hydrotreatment Process. *Petroleum Science and Technology*, *27*(18), 2075–2084.
- Mahboubian, A., Hashemein, S. K., Moghadam, S., Atyabia, F., & Dinarvand, R. (2010). Preparation and in-vitro evaluation of controlled release PLGA microparticles containing triptoreline. *Iranian Journal of Pharmaceutical Research*, *9*, 369–378.
- Marieb, E. N., & Hoehn, K. (2010). *Human Anatomy & Physiology. Physiology*, 7, 664-668.
- Martins, S., Tho, I., Reimold, I., Fricker, G., Souto, E., Ferreira, D., & Brandl, M. (2012). Brain delivery of camptothecin by means of solid lipid nanoparticles: formulation design, in vitro and in vivo studies. *International Journal of Pharmaceutics*, 439(1-2), 49–62.
- Mason, T. G., Wilking, J. N., Meleson, K., Chang, C. B., & Graves, S. M. (2006). Nanoemulsions: Formation, structure, and physical properties. *Journal of Physics: Condensed Matter*, *18*(41), 635–666.
- Masoumi, H. R. F., Kassim, A., Basri, M., Abdullah, D. K., & Haron, M. J. (2011). Multivariate optimization in the biosynthesis of a triethanolamine (TEA)-based esterquat cationic surfactant using an artificial neural network. *Molecules*, *16*(7), 5538–49.
- Matijašić, M., Munić Kos, V., Nujić, K., Cužić, S., Padovan, J., Kragol, G., Eraković Haber, V. (2012). Fluorescently labeled macrolides as a tool

- for monitoring cellular and tissue distribution of azithromycin. Pharmacological Research: The Official Journal of the Italian Pharmacological Society, 66(4), 332–42.
- Matzneller, P., Krasniqi, S., Kinzig, M., Sörgel, F., Hüttner, S., Lackner, E., Zeitlinger, M. (2013). Blood, tissue, and intracellular concentrations of azithromycin during and after end of therapy. *Antimicrobial Agents and Chemotherapy*, *57*, 1736–1742.
- McClements, D. J. (2012). Nanoemulsions versus microemulsions: terminology, differences, and similarities. *Soft Matter*, *8*(6), 1719.
- McClements, D. J., & Rao, J. (2011). Food-grade nanoemulsions: Formulation, fabrication, properties, performance, biological fate, and potential toxicity. *Critical Reviews in Food Science and Nutrition*, *51*(4), 285–330.
- Mhlanga, N., & Ray, S. S. (2015). Kinetic models for the release of the anticancer drug doxorubicin from biodegradable polylactide/metal oxide-based hybrids. *International Journal of Biological Macromolecules*, 72, 1301–7.
- Mishra, P. R., Shaal, L. a., Müller, R. H., & Keck, C. M. (2009). Production and characterization of Hesperetin nanosuspensions for dermal delivery. *International Journal of Pharmaceutics*, *371*(1-2), 182–189.
- Mohammadi, G., Nokhodchi, A., Barzegar-Jalali, M., Lotfipour, F., Adibkia, K., Ehyaei, N., Khosro Adibkiac Hadi Valizadehc, N. E. (2011). Physicochemical and anti-bacterial performance characterization of clarithromycin nanoparticles as colloidal drug delivery system. *Colloids and Surfaces B: Biointerfaces*, 88(1), 39–44.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65(1-2), 55–63.
- Muller, R. H., & Keck, C. M. (2004). Challenges and solutions for the delivery of biotech drugs-a review of drug nanocrystal technology and lipid nanoparticles. *Journal of Biotechnology*, *113*(1-3), 151–70.
- Müller, R. H., Schmidt, S., Buttle, I., Akkar, a., Schmitt, J., & Brömer, S. (2004). SolEmuls® Novel technology for the formulation of i.v. emulsions with poorly soluble drugs. *International Journal of Pharmaceutics*, *269*(2), 293–302.
- Muller, W. A. (2009). Mechanisms of transendothelial migration of leukocytes. *Circ.Res.*, *105*(1524-4571), 223–230.
- Munić, V., Kelnerić, Ž., Mikac, L., & Eraković Haber, V. (2010). Differences in assessment of macrolide interaction with human MDR1 (ABCB1, P-gp)

- using rhodamine-123 efflux, ATPase activity and cellular accumulation assays. *European Journal of Pharmaceutical Sciences*, *41*, 86–95.
- Narang, A. S., Delmarre, D., & Gao, D. (2007). Stable drug encapsulation in micelles and microemulsions. *International Journal of Pharmaceutics*, 345, 9-25.
- Nau, R., Sörgel, F., & Eiffert, H. (2010). Penetration of drugs through the blood-cerebrospinal fluid/blood-brain barrier for treatment of central nervous system infections. *Clinical Microbiology Reviews*, 23(4), 858–883.
- Ngan, C. L., Basri, M., Lye, F. F., Fard Masoumi, H. R., Tripathy, M., Abedi Karjiban, R., & Abdul-Malek, E. (2014). Comparison of Box-Behnken and central composite designs in optimization of fullerene loaded palmbased nano-emulsions for cosmeceutical application. *Industrial Crops and Products*, *59*, 309–317.
- Nie, S. (2010). Understanding and overcoming major barriers in cancer nanomedicine. *Nanomedicine* (London, England), 5(4), 523–528.
- Nirogi, R. V. S., Kandikere, V. N., Shukla, M., Mudigonda, K., Maurya, S., Boosi, R., & Yerramilli, A. (2005). Sensitive and selective liquid chromatography–tandem mass spectrometry method for the quantification of azithromycin in human plasma. *Analytica Chimica Acta*, 553, 1-8.
- Nutan, T. H. M., & Reddy, I. K. (2010). General principles of suspensions. Pharmaceutical suspensions. From formulation development to manufacturing (39–65).
- Obermeier, B., Daneman, R., & Ransohoff, R. M. (2013). Development, maintenance and disruption of the blood-brain barrier. *Nature Medicine*, 19(12), 1584–96.
- Ochekpe, N. a., Olorunfemi, P. O., & Ngwuluka, N. C. (2009). Nanotechnology and drug delivery part 2: Nanostructures for drug delivery. *Tropical Journal of Pharmaceutical Research*, *8*, 275–287.
- Owens, D. E., & Peppas, N. A. (2006). Opsonization, biodistribution, and pharmacokinetics of polymeric nanoparticles. *International Journal of Pharmaceutics*, 307, 93-102.
- Palmer, A. M. (2010). The role of the blood-CNS barrier in CNS disorders and their treatment. *Neurobiology of Disease*, *37*(1), 3–12.
- Panyam, J., Williams, D., Dash, A., Leslie-Pelecky, D., & Labhasetwar, V. (2004). Solid-State Solubility Influences Encapsulation and Release of Hydrophobic Drugs from PLGA/PLA Nanoparticles. *Journal of Pharmaceutical Sciences*, 93, 1804–1814.

- Pardridge, W. M. (2002). Drug and gene delivery to the brain: The vascular route. *Neuron*, *36*(4), 555–558.
- Pardridge, W. M. (2005). Molecular biology of the blood-brain barrier. *Molecular Biotechnology*, *30*(1), 57–70.
- Pardridge, W. M. (2012). Drug transport across the blood-brain barrier. *Journal of Cerebral Blood Flow and Metabolism: Official Journal of the International Society of Cerebral Blood Flow and Metabolism*, 32(11), 1959–72.
- Pardridge, W. M., Sakiyama, R., & Fierer, G. (1984). Blood-brain barrier transport and brain sequestration of propranolol and lidocaine. *The American Journal of Physiology*, 247, 582–588.
- Parmar, A., Chavda, S., & Bahadur, P. (2014). Pluronic-cationic surfactant mixed micelles: Solubilization and release of the drug hydrochlorothiazide. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 441, 389–397.
- Parnham, M. J., Erakovic Haber, V., Giamarellos-Bourboulis, E. J., Perletti, G., Verleden, G. M., & Vos, R. (2014). Azithromycin: mechanisms of action and their relevance for clinical applications. *Pharmacology & Therapeutics*, 143(2), 225–45.
- Patel, H., Raval, G., Nazari, M., & Heerklotz, H. (2010). Effects of glycerol and urea on micellization, membrane partitioning and solubilization by a nonionic surfactant. *Biophysical Chemistry*, *150*(1-3), 119–128.
- Patel, M., Souto, E. B., & Singh, K. K. (2013). Advances in brain drug targeting and delivery: limitations and challenges of solid lipid nanoparticles. *Expert Opinion on Drug Delivery*, 10(7), 1–17.
- Pendashteh, A. R., Fakhru'l-Razi, A., Chaibakhsh, N., Abdullah, L. C., Madaeni, S. S., & Abidin, Z. Z. (2011). Modeling of membrane bioreactor treating hypersaline oily wastewater by artificial neural network. *Journal of Hazardous Materials*, 192, 568–575.
- Peng, C., Yan, X., & Tang, X. (2010). Preparation and characterization of a triamcinolone acetonide palmitate submicron emulsion. *Asian Journal of Pharmaceutical Sciences*, *5*(2), 61–73.
- Persidsky, Y., Ramirez, S. H., Haorah, J., & Kanmogne, G. D. (2006). Blood-brain barrier: Structural components and function under physiologic and pathologic conditions. *Journal of Neuroimmune Pharmacology*, 1(3), 223–236.
- Petersen, S., Steiniger, F., Fischer, D., Fahr, A., & Bunjes, H. (2011). The physical state of lipid nanoparticles influences their effect on in vitro cell viability. *European Journal of Pharmaceutics and Biopharmaceutics*:

- Official Journal of Arbeitsgemeinschaft Für Pharmazeutische Verfahrenstechnik e.V, 79, 150–61.
- Pinheiro, A. C., Coimbra, M. A., & Vicente, A. A. (2016). In vitro behaviour of curcumin nanoemulsions stabilized by biopolymer emulsifiers Effect of interfacial composition. *Food Hydrocolloids*, *52*, 460–467.
- Prabhakar, K., Afzal, S. M., Surender, G., & Kishan, V. (2013). Tween 80 containing lipid nanoemulsions for delivery of indinavir to brain. *Acta Pharmaceutica Sinica B*, *3*(5), 345–353.
- Preetz, C., Hauser, A., Hause, G., Kramer, A., & Mäder, K. (2010). Application of atomic force microscopy and ultrasonic resonator technology on nanoscale: Distinction of nanoemulsions from nanocapsules. *European Journal of Pharmaceutical Sciences*, 39(1-3), 141–151.
- Qian, C., & McClements, D. J. (2011). Formation of nanoemulsions stabilized by model food-grade emulsifiers using high-pressure homogenization: Factors affecting particle size. *Food Hydrocolloids*, *25*(5), 1000–1008.
- Ragelle, H., Crauste-Manciet, S., Seguin, J., Brossard, D., Scherman, D., Arnaud, P., & Chabot, G. G. (2012). Nanoemulsion formulation of fisetin improves bioavailability and antitumour activity in mice. *International Journal of Pharmaceutics*, 427(2), 452–459.
- Rahn-Chique, K., Puertas, A. M., Romero-Cano, M. S., Rojas, C., & Urbina-Villalba, G. (2012). Evaluation of the Flocculation Speed in Oil/Water Nanoemulsions. 1) Development of Theoretical Expressions for the Turbidity of a Nanoemulsion. *Interciencia*, *37*(8), 577–581.
- Ramteke, K. H., Dighe, P. A., Kharat, A. R., & Patil, S. V. (2014). Review Article Mathematical Models of Drug Dissolution: A Review. *Scholars Academic Journal of Pharmacy*, *3*(5), 388–396.
- Rao, J., & McClements, D. J. (2011). Food-grade microemulsions, nanoemulsions and emulsions: Fabrication from sucrose monopalmitate & lemon oil. *Food Hydrocolloids*, *25*(6), 1413–1423.
- Re, F., Gregori, M., & Masserini, M. (2012). Nanotechnology for neurodegenerative disorders. *Maturitas*, 73(1), 45–51.
- Rip, J., Schenk, G. J., & de Boer, a G. (2009). Differential receptor-mediated drug targeting to the diseased brain. *Expert Opinion on Drug Delivery*, 6(3), 227–37.
- Ritger, P. L., & Peppas, N. a. (1987). A simple equation for description of solute release II. Fickian and anomalous release from swellable devices. *Journal of Controlled Release*, *5*(1), 37–42.

- Roberts, J. a, & Lipman, J. (2009). Pharmacokinetic issues for antibiotics in the critically ill patient. *Critical Care Medicine*, *37*(3), 840–851.
- Rodvold, K. A., Danziger, L. H., & Gotfried, M. H. (2003). Steady-state plasma and bronchopulmonary concentrations of intravenous levofloxacin and azithromycin in healthy adults. *Antimicrobial Agents and Chemotherapy*, 47(8), 2450–7.
- Rosenblatt, K. M., Douroumis, D., & Bunjes, H. (2007). Drug release from differently structured monoolein/poloxamer nanodispersions studied with differential pulse polarography and ultrafiltration at low pressure. *Journal of Pharmaceutical Sciences*, *96*(6), 1564–75.
- Saberi, A. H., Fang, Y., & McClements, D. J. (2013a). Effect of glycerol on formation, stability, and properties of vitamin-E enriched nanoemulsions produced using spontaneous emulsification. *Journal of Colloid and Interface Science*, *411*, 105–113.
- Saberi, A. H., Fang, Y., & McClements, D. J. (2013b). Fabrication of vitamin E-enriched nanoemulsions by spontaneous emulsification: Effect of propylene glycol and ethanol on formation, stability, and properties. *Food Research International*, *54*(1), 812–820.
- Saberi, A. H., Fang, Y., & McClements, D. J. (2013c). Fabrication of vitamin E-enriched nanoemulsions: Factors affecting particle size using spontaneous emulsification. *Journal of Colloid and Interface Science*, 391(1), 95–102.
- Sadtler, V. M., Imbert, P., & Dellacherie, E. (2002). Ostwald ripening of oil-in-water emulsions stabilized by phenoxy-substituted dextrans. *Journal of Colloid and Interface Science*, 254(2), 355–361.
- Sakeena, M. H. F., Elrashid, S. M., Munavvar, a S., & Azmin, M. N. (2011). Effects of oil and drug concentrations on droplets size of palm oil esters (POEs) nanoemulsion. *Journal of Oleo Science*, 60(4), 155–158.
- Salome, a. C., Godswill, C. O., & Ikechukwu, I. O. (2013). Kinetics and mechanisms of drug release from swellable and non swellable matrices: A review. Research Journal of Pharmaceutical, Biological and Chemical Sciences, 4(2), 97–103.
- Salvia-Trujillo, L., Rojas-Graü, M. A., Soliva-Fortuny, R., & Martín-Belloso, O. (2013). Effect of processing parameters on physicochemical characteristics of microfluidized lemongrass essential oil-alginate nanoemulsions. *Food Hydrocolloids*, *30*(1), 401–407.
- Sandoval, K. E., & Witt, K. A. (2008). Blood-brain barrier tight junction permeability and ischemic stroke. *Neurobiology of Disease*, *32*(2), 200–19.

- Scheld, W. M., Koedel, U., Nathan, B., & Pfister, H.-W. (2002). Pathophysiology of bacterial meningitis: mechanism(s) of neuronal injury. *The Journal of Infectious Diseases*, *186*, 225–233.
- Schinkel, A. H., & Jonker, J. W. (2012). Mammalian drug efflux transporters of the ATP binding cassette (ABC) family: An overview. *Advanced Drug Delivery Reviews*, *64*, 138–153.
- Schultz, S., Wagner, G., Urban, K., & Ulrich, J. (2004). High-pressure homogenization as a process for emulsion formation. *Chemical Engineering and Technology*, *27*(4), 361–368.
- Serlin, Y., Shelef, I., Knyazer, B., & Friedman, A. (2015). Anatomy and Physiology of the Blood-Brain Barrier. Seminars in Cell & Developmental Biology, 38, 2–6.
- Shafiq, S., Shakeel, F., Talegaonkar, S., Khar, R. K., & Ali, M. (2010). Nanoemulsion as Carrier for Stability Enhancement of Ramipril. *Journal of Dispersion Science and Technology*, 31, 975-979.
- Shaikh, H. K., Kshirsagar, R. V, & Patil, S. G. (2015). Mathematical Models for Drug Release Characterization: a Review. *World Journal of Pharmacy and Pharmaceutical Sciences*, *4*(4), 324–338.
- Shakeel, F., Baboota, S., Ahuja, A., Ali, J., & Shafiq, S. (2008). Accelerated stability testing of celecoxib nanoemulsion containing Cremophor-EL. *African journal of pharmacy and pharmacology*, *2*(8), 179–183.
- Shakeri-Nejad, K., & Stahlmann, R. (2006). Drug interactions during therapy with three major groups of antimicrobial agents. *Expert Opinion on Pharmacotherapy*, 7(6), 639–51.
- Shen, J., & Burgess, D. J. (2012). Accelerated in-vitro release testing methods for extended-release parenteral dosage forms. *Journal of Pharmacy and Pharmacology*, *64*(7), 986–996.
- Shulman, Z., Cohen, S. J., Roediger, B., Kalchenko, V., Jain, R., Grabovsky, V., Alon, R. (2012). Transendothelial migration of lymphocytes mediated by intraendothelial vesicle stores rather than by extracellular chemokine depots. *Nat.Immunol.*, *13*, 67–76.
- Siepmann, J., & Siepmann, F. (2008). Mathematical modeling of drug delivery. *International Journal of Pharmaceutics*, *364*(2), 328–343.
- Simões, S. I., Tapadas, J. M., Marques, C. M., Cruz, M. E. M., Martins, M. B. F., & Cevc, G. (2005). Permeabilisation and solubilisation of soybean phosphatidylcholine bilayer vesicles, as membrane models, by polysorbate, Tween 80. *European Journal of Pharmaceutical Sciences*, 26, 307–317.

- Singer, M. M., George, S., & Tjeerdema, R. S. (1995). Relationship of some physical properties of oil dispersants and their toxicity to marine organisms. *Arch. Environ. Contam. Toxicol*, *29*(1), 33–38.
- Singh, R., & Lillard, J. W. (2009a). Nanoparticle-based targeted drug delivery. Experimental and Molecular Pathology, 86(3), 215–23.
- Singh, R., & Lillard, J. W. (2009b). Nanoparticle-based targeted drug delivery. *Experimental and Molecular Pathology*, *86*(3), 215–23.
- Singhvi, G., & Singh, M. (2011). Review: in vitro drug release characterization models. *Int J Pharm Stud Res*, 2, 57-87.
- Sneed, B. P., & Scheld, W. M. (2013). Acute Bacterial Meningitis. *Hospital Medicine Clinics*.
- Snyder, R. D. (1995). Bacterial Meningitis: Diagnosis and Treatment.
- Solans, C., Izquierdo, P., Nolla, J., Azemar, N., & Garcia-Celma, M. J. (2005). Nano-emulsions. *Current Opinion in Colloid and Interface Science*, 10(3-4), 102–110.
- Solè, I., Solans, C., Maestro, a., González, C., & Gutiérrez, J. M. (2012). Study of nano-emulsion formation by dilution of microemulsions. *Journal of Colloid and Interface Science*, 376, 133–139.
- Sood, S., Jain, K., & Gowthamarajan, K. (2014). Optimization of curcumin nanoemulsion for intranasal delivery using design of experiment and its toxicity assessment. *Colloids and Surfaces B: Biointerfaces*, *113*, 330–337.
- Stepanić, V., Koŝstrun, S., Malnar, I., Hlevnjak, M., Butković, K., Ćaleta, I., Munić, V. (2011). Modeling cellular pharmacokinetics of 14- and 15-membered macrolides with physicochemical properties. *Journal of Medicinal Chemistry*, *54*(3), 719–733.
- Streck, L., de Araújo, M. M., de Souza, I., Fernandes-Pedrosa, M. F., do Egito, de Oliveira, A. G., & da Silva-Júnior, A. a. (2014). Surfactant—cosurfactant interactions and process parameters involved in the formulation of stable and small droplet-sized benznidazole-loaded soybean O/W emulsions. *Journal of Molecular Liquids*, 196, 178–186.
- Sultana, N. K., Saha, S. K., Al-Emran, H. M., Modak, J. K., Sharker, M. A. Y., El-Arifeen, S., Luby, S. P. (2013). Impact of introduction of the Haemophilus influenzae type b conjugate vaccine into childhood immunization on meningitis in Bangladeshi infants. *The Journal of Pediatrics*, 163, 73–88.
- Summerfield, S. G., Read, K., Begley, D. J., Obradovic, T., Hidalgo, I. J., Coggon, S., Jeffrey, P. (2007). Central nervous system drug disposition:

- the relationship between in situ brain permeability and brain free fraction. *The Journal of Pharmacology and Experimental Therapeutics*, 322(1), 205–213.
- Sylvester, P. W. (2011). Optimization of the tetrazolium dye (MTT) colorimetric assay for cellular growth and viability. *Methods in Molecular Biology* (*Clifton, N.J.*), 716, 157–68.
- Tadros, T. F. (2013). Emulsion Formation, Stability, and Rheology. *Emulsion Formation and Stability*, 1–76.
- Tadros, T., Izquierdo, P., Esquena, J., & Solans, C. (2004a). Formation and stability of nano-emulsions. *Advances in Colloid and Interface Science*, 108-109, 303-318.
- Taherian, A.R., Fustier, P. & Ramaswamy, H.S., 2006. Effect of added oil and modified starch on rheological properties, droplet size distribution, opacity and stability of beverage cloud emulsions. *Journal of Food Engineering*, 77(3), 687–696.
- Tan, C. P., & Nakajima, M. (2005). β-Carotene nanodispersions: Preparation, characterization and stability evaluation. *Food Chemistry*, 92(4), 661–671.
- Tan, S. F., Masoumi, H. R. F., Karjiban, R. A., Stanslas, J., PKirby, B., Basri, M., & Basri, H. Bin. (2015). Ultrasonic emulsification of parenteral valproic acid-loaded nanoemulsion with response surface methodology and evaluation of its stability. *Ultrasonics Sonochemistry*, 29, 299-308.
- Thiagarajan, P. (2011). Nanoemulsions for drug delivery through different routes. Research in Biotechnology, 2(3), 1–13.
- Tian, X. H., Lin, X. N., Wei, F., Feng, W., Huang, Z. C., Wang, P., Diao, Y. (2011). Enhanced brain targeting of temozolomide in polysorbate-80 coated polybutylcyanoacrylate nanoparticles. *International Journal of Nanomedicine*, *6*, 445–452.
- Tunkel, A. R. (2004). Practice Guidelines for the Management of Bacterial Meningitis. *Clinical Infectious Diseases*, *39*, *267-84*.
- Tunkel, A. R., & Scheld, W. M. (2002). Corticosteroids for Everyone with Meningitis? *New England Journal of Medicine*, *347*(20), 1613–1615.
- Üner, M., & Yener, G. (2007). Importance of solid lipid nanoparticles (SLN) in various administration routes and future perspective. *International Journal of Nanomedicine*, *2*(3), 289–300.
- Van De Beek, D., Brouwer, M. C., Thwaites, G. E., & Tunkel, A. R. (2012). Advances in treatment of bacterial meningitis. *The Lancet*, 380(9854), 1693–1702.

- Van Eerdenbrugh, B., Froyen, L., Van Humbeeck, J., Martens, J. a, Augustijns, P., & Van den Mooter, G. (2008). Drying of crystalline drug nanosuspensions-the importance of surface hydrophobicity on dissolution behavior upon redispersion. European Journal of Pharmaceutical Sciences: Official Journal of the European Federation for Pharmaceutical Sciences, 35(1-2), 127–35.
- Vercauteren, D., Vandenbroucke, R. E., Jones, A. T., Rejman, J., Demeester, J., De Smedt, S. C., Braeckmans, K. (2010). The use of inhibitors to study endocytic pathways of gene carriers: optimization and pitfalls. *Molecular Therapy: The Journal of the American Society of Gene Therapy*, 18(3), 561–569.
- Wacker, M. (2013). Nanocarriers for intravenous injection--the long hard road to the market. *International Journal of Pharmaceutics*, *457*(1), 50–62.
- Wagner, A., & Vorauer-Uhl, K. (2011). Liposome technology for industrial purposes. *Journal of Drug Delivery*, 2011, 1-9.
- Walker, R. M., Decker, E. a., & McClements, D. J. (2015). Physical and oxidative stability of fish oil nanoemulsions produced by spontaneous emulsification: Effect of surfactant concentration and particle size. *Journal of Food Engineering*, 164, 10–20.
- Wang, Y., Zhang, L., Wang, Q., & Zhang, D. (2013). Stability of nanosuspensions in drug delivery. *Journal of Controlled Release*, 172(3), 1126–1141.
- Warisnoicharoen, W., Lansley, A. B., & Lawrence, M. J. (2000). Nonionic oil-in-water microemulsions: the effect of oil type on phase behaviour. *International Journal of Pharmaceutics*, 198(1), 7–27.
- Warisnoicharoen, W., Lansley, A. B., & Lawrence, M. J. (2003). Toxicological evaluation of mixtures of nonionic surfactants, alone and in combination with oil. *Journal of Pharmaceutical Sciences*, *92*(4), 859–68.
- Washington, C. (1996). Stability of lipid emulsions for drug delivery. *Advanced Drug Delivery Reviews*, 20(2-3), 131–145.
- Weiss, N., Miller, F., Cazaubon, S., & Couraud, P.-O. (2009). The blood-brain barrier in brain homeostasis and neurological diseases. *Biochimica et Biophysica Acta*, *1788*(4), 842–57.
- Welin-Berger, K., & Bergenståhl, B. (2000). Inhibition of Ostwald ripening in local anesthetic emulsions by using hydrophobic excipients in the disperse phase. *International Journal of Pharmaceutics*, 200(2), 249–260.
- Wewer, C., Seibt, A., Wolburg, H., Greune, L., Schmidt, M. A., Berger, J., Tenenbaum, T. (2011). Transcellular migration of neutrophil

- granulocytes through the blood-cerebrospinal fluid barrier after infection with Streptococcus suis. *Journal of Neuroinflammation*, *8*, 51.
- Wilson, B., Samanta, M. K., Santhi, K., Kumar, K. P. S., Paramakrishnan, N., & Suresh, B. (2008). Targeted delivery of tacrine into the brain with polysorbate 80-coated poly(n-butylcyanoacrylate) nanoparticles. *European Journal of Pharmaceutics and Biopharmaceutics*, 70(1), 75–84.
- Wirtitsch, M., Wessner, B., Spittler, A., Roth, E., Volk, T., Bachmann, L., & Hiesmayr, M. (2007). Effect of different lipid emulsions on the immunological function in humans: a systematic review with meta-analysis. *Clinical Nutrition (Edinburgh, Scotland)*, *26*, 302–313.
- Wooster, T. J., Golding, M., & Sanguansri, P. (2008a). Impact of oil type on nanoemulsion formation and ostwald ripening stability. *Langmuir*, 24(22), 12758–12765.
- Wu, L., Zhang, J., & Watanabe, W. (2011). Physical and chemical stability of drug nanoparticles. *Advanced Drug Delivery Reviews*, *63*(6), 456–469.
- Yaron, P. N., Scott, A. J., Reynolds, P. a., Mata, J. P., & White, J. W. (2011). High internal phase emulsions under shear. Co-surfactancy and shear stability. *Journal of Physical Chemistry B*, 115(19), 5775–5784.
- Yuan, Y., Gao, Y., Zhao, J., & Mao, L. (2008). Characterization and stability evaluation of β-carotene nanoemulsions prepared by high pressure homogenization under various emulsifying conditions. Food Research International, 41(1), 61–68.
- Zainol, S., Basri, M., Basri, H. Bin, Shamsuddin, A. F., Abdul-Gani, S. S., Karjiban, R. A., & Abdul-Malek, E. (2012). Formulation optimization of a palm-based nanoemulsion system containing levodopa. *International Journal of Molecular Sciences*, *13*, 13049–13064.
- Zhang, J. (2011). Advances in novel parentral drug delivery systems. *Asian Journal of Pharmaceutics*, 4(3), 193-198.
- Zhu, T. F., & Szostak, J. W. (2009). Preparation of large monodisperse vesicles. *Plos one*, *4*(4), 89-102.
- Ziani, K., Chang, Y., McLandsborough, L., & McClements, D. J. (2011). Effect of particle charge on the antifungal properties of thyme oil nanoemulsions. In 85th ACS Colloid and Surface Science Symposium, Montreal, QC, Canada, 19-22 (p. Collsymp–256).
- Ziani, K., Fang, Y., & McClements, D. J. (2012). Encapsulation of functional lipophilic components in surfactant-based colloidal delivery systems: Vitamin E, vitamin D, and lemon oil. *Food Chemistry*, *134*(2), 1106–1112.

LIST OF PUBLICATIONS

- Daood, G. S., Basri, H., Stanslas, J., Fard Masoumi, H. R., & Basri, M. (2015). Predicting the optimum compositions of a parenteral nanoemulsion system loaded with azithromycin antibiotic utilizing the artificial neural network model. *Royal Society of Chemistry Advances*, 5, 82654–82665.
- Daood, G. S., Stanslas, J., Fard Masoumi, H. R., & Basri, M., Basri, H. Parenteral nanoemulsion as a promising carrier for brain delivery of azithromycin: Design, evaluation, and long-term stability assessment. (In preparation).





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