



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF GSK-3 INHIBITION ON LPS-INDUCED
NEUROINFLAMMATION AND IL-10 PRODUCTION IN MICROGLIA***

ZUHaida BINTI MD ZAIN

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By

ZUHAIDA BINTI MD ZAIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

September 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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September 2016

**Chairman: Masriana Hassan, PhD
Faculty: Medicine and Health Science**

Microglia are resident macrophages of the central nervous system (CNS) that play a role in the immune surveillance system against various pathogenicities. However, excessive inflammation resulting from activation of microglia has been implicated in the neurodegenerative diseases such as multiple sclerosis. The protein kinase, Glycogen Synthase Kinase (GSK) 3, is involved in many cellular functions including microglial activation. Previously, inhibition of GSK-3 has been shown to reduce inflammation due to decreased production of pro-inflammatory cytokines and increased production of IL-10 in LPS-induced endotoxin shock animal model. Thus, this study was performed to elucidate the possible immunoregulatory effects of GSK-3 inhibitors on activated microglia. We hypothesized that inhibition of GSK-3 would reduce the exaggeration of inflammation in LPS-induced microglial activation with associated increased of IL-10 production. The optimal concentration of LPS and incubation period were optimized and determined by measuring the level of nitric oxide (NO) produced by BV-2, microglia cell lines, without compromising their effect on cell viability. The GSK-3 inhibitors, including lithium chloride (LiCl), SB216763, NP12 and CHIR99021 were used to block GSK-3 activities in the BV-2 cells. All GSK-3 inhibitors tested have shown their efficacy in reducing production of pro-inflammatory molecules, such as NO, glutamate, MCP-1 and cytokines (TNF- α and IL-6). Interestingly, reduction of pro-inflammatory molecules via GSK-3 inhibition was associated with significant increase in IL-10 production. Furthermore, treatment with GSK-3 inhibitor reduced expression of microglial activation markers, CD11b, while increased expressions of microglial inhibitory markers, CD200R, which confirmed the ability of GSK-3 inhibitor in inhibiting microglial activation. These results indicate that GSK-3 inhibitors effectively reduced pro-inflammatory molecules via inhibition of microglial activation. Moreover, these inhibitors could potentially reduce the severity of neuroinflammation by enhancing IL-10 production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN PERENCATAN GSK-3 KEATAS KERADANGAN NEURON YANG
DIRANSANG LPS DAN PENGHASILAN IL-10 DALAM MICROGLIA**

Oleh

ZUHAIDA BINTI MD ZAIN

September 2016

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Mikroglia ialah makrofaj yang terdapat di dalam sistem saraf pusat (CNS) yang memainkan peranan di dalam sistem pengawasan imun terhadap pelbagai penyakit disebabkan patogen. Walaubagaimanapun, keradangan yang berlebihan hasil dari pengaktifan mikroglia telah dikaitkan dalam penyakit-penyakit yang melibatkan kemerosotan neuron seperti sklerosis berbilang. Glycogen synthase kinase (GSK) 3, iaitu salah satu daripada protein kinase, terlibat dalam banyak fungsi selular termasuk pengaktifan mikroglia. Sebelum ini, perencatan GSK-3 telah menunjukkan pengurangan keradangan disebabkan pengurangan penghasilan sitokin pro-radang dan peningkatan penghasilan IL-10 dalam model haiwan kejutan endotoksin yang dirangsang oleh LPS. Kami telah membuat hipotesis bahawa perencatan GSK-3 akan meningkatkan penghasilan IL-10 dan seterusnya mengurangkan keradangan yang keterlaluan pada mikroglia yang dirangsang oleh LPS. Dengan tu, kajian ini telah dijalankan untuk menunjukkan potensi pengawalan imun oleh perencat GSK-3 terhadap mikroglia yang aktif. Kami telah membuat hipotesis bahawa perencatan GSK-3 dapat mengurangkan keradangan yang keterlaluan dalam pengaktifan mikroglia yang dirangsang oleh LPS disebabkan peningkatan penghasilan IL-10. Kepekatan optimum LPS dan masa inkubasi telah ditentukan dengan menyukat tahap nitrik oksida (NO) dihasilkan oleh sel BV-2, sel mikroglia, dengan mengambil kira kesannya terhadap jumlah sel hidup. Perencat-perencat GSK-3 termasuk lithium klorida (LiCl), SB216763, NP12, dan CHIR99021 telah digunakan untuk menyekat aktiviti GSK-3 dalam sel BV-2. Semua perencat GSK-3 yang diuji telah mengesahkan keberkesannya dalam mengurangkan penghasilan molekul-molekul yang menyebabkan keradangan seperti NO, glutamat, MCP-1 dan sitokin pro-radang (TNF- α dan IL-6). Menariknya, pengurangan penghasilan molekul pro-radang melalui perencatan GSK-3 adalah berkaitan dengan peningkatan penghasilan IL-10 yang ketara.

Tambahan pula, rawatan dengan perencat GSK-3 telah mengurangkan ekspresi penanda pengaktifan mikroglia, CD11b dan meningkatkan ekspresi penanda perencatan, CD200R, yang telah mengesahkan kemampuan perencat GSK-3 dalam mengurangkan pengaktifan mikroglia. Keputusan-keputusan ini menunjukkan bahawa perencat GSK-3 dapat mengurangkan molekul-molekul pro-radang secara berkesan melalui perencatan pengaktifan mikroglia. Lebih lebih lagi, perencat-perencat ini berpotensi untuk mengurangkan tahap keterukan keradangan neuron dengan meningkatkan penghasilan IL-10.



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Thank you.

I certify that a Thesis Examination Committee has met on 29 September 2016 to conduct the final examination of Zuhaida binti Md Zain on her thesis entitled "Effects of GSK-3 Inhibition on LPS-Induced Neuroinflammation and IL-10 Production in Microglia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

| | |
|--------------------|-------------------------------|
| α | alpha |
| β | beta |
| γ | gamma |
| $^{\circ}\text{C}$ | degree celcius |
| M | molar |
| mL | mililitre |
| μL | microlitre |
| μg | microgram |
| $\mu\text{g/mL}$ | microgram per mililitre |
| nm | nanometer |
| mM | milimolar |
| μM | micromolar |
| mg/mL | milligram per mililitre |
| mL/L | mililitre per litre |
| g/L | gram per litre |
| U/mL | unit per mililitre |
| RT | room temperature |
| hrs | hours |
| min | minute |
| Sec | second |
| rpm | revolutions per minute |
| 1X | one time |
| AD | alzheimer's disease |
| ALS | amyotrophic lateral sclerosis |
| APCs | antigen presenting cells |
| A β | beta amyloid |
| BBB | blood brain barrier |

| | |
|-------------------------------|---|
| BSA | bovine serum albumin |
| CNS | central nervous system |
| dH ₂ O | distilled water |
| DMEM | Dulbecco's Modified Eagle Medium |
| DMSO | dimethyl sulfoxide |
| EAE | Experimental autoimmune encephalomyelitis |
| EDTA | ethylene diamine tetraacetic acid |
| ELISA | enzyme linked immunosorbent assay |
| FBS | fetal bovine serum |
| FITC | fluorescein isothiocyanate |
| GSK-3 | glycogen synthase kinase 3 |
| H ₂ O | water |
| H ₂ O ₂ | hydrogen peroxide |
| HD | huntington's disease |
| IFN | interferon |
| IL | interleukin |
| LPS | lipopolysaccharide |
| MAPK | mitogen activated protein kinase |
| MCP-1 | monocyte chemotactic protein-1 |
| MHC II | major histocompatibility complex class II |
| MS | multiple sclerosis |
| NaNO ₂ | sodium nitrite |
| NED | N-1- naphthylenediamine dihydrochloride |
| NF-κB | nuclear factor kappa B |
| NO | nitric oxide |
| PBS | phosphate buffer saline |
| TNF-α | tumour necrosis factor-alpha |
| TLR-4 | toll like receptor-4 |

CHAPTER 1

INTRODUCTION

Microglia are the macrophages of the innate immune system in the central nervous system (CNS). Upon activation of peripheral and central immune system, microglia play their role by receiving the inflammatory signals and propagate the inflammation (Huang et al., 2009; Nguyen et al., 2002). Microglia are derived from primitive myeloid progenitors constituting almost 10-15% of the total CNS cell population (Ginhoux et al., 2010). In normal homeostatic condition, microglia are quiescent, ramified in shape and moves along parenchyma constituting immune surveillance system by monitoring homeostatic changes in CNS tissue and providing mechanical and trophic support to neurons (McGeer et al., 1987). Once activated, the morphology shift to amoeboid in shape and possess macrophage-like capabilities such as phagocytosis, antigen presentation to CD4 T helper cells and release of cytokines or chemokines (Goldmann & Prinz, 2013). Intense and continuous microglia activation induces neuroinflammation and contributes to neurodegenerative disease such as multiple sclerosis. Activation of microglia can be induced by various stimuli such as lipopolysaccharide (LPS) and interferon (IFN)- γ which promotes pro-inflammatory cytokines production including tumor necrosis factor (TNF)- α , interleukin (IL)-1 β and IL-6, and other inflammatory components such as nitric oxide (NO) and glutamate (Nakajima & Kohsaka, 2001).

Stimulation of microglia with LPS able to stimulate expression of inducible nitric oxide synthase (iNOS) (Heneka & Feinstein, 2001; Murphy, 2000). Expression of iNOS induce production of high NO level (Bal-Price & Brown, 2001; Bal-Price et al., 2002) which further inhibits mitochondrial cytochrome oxidase that can cause neuronal cell death (Bal-Price & Brown, 2001; Brown & Cooper, 1994). NO competes with oxygen for cytochrome oxidase causing respiratory inhibition which cause neuronal depolarization and production of glutamate (Bal-Price & Brown, 2001; Golde et al., 2002; Jekabsone et al., 2007; McNaught & Brown, 1998; Stewart et al., 2002). On the other hands, production of glutamate inflicts an amplified metabolic demand on neurons. Inhibition of respiratory metabolism by NO, for example, can cause the cells to be pushed over some toxic state and lead to neuronal cell death. Glutamate is the major excitatory neurotransmitter in CNS that plays an important role memory and learning (Featherstone, 2009; Reis et al., 2009). Overproduction of glutamate may cause an imbalance in CNS homeostasis which could lead to neuronal cells death. Studies have shown that excessive production of NO and glutamate propagates inflammation in the CNS and induces neuronal cell death (Goldmann & Prinz, 2013).

Glycogen Synthase Kinase (GSK-3) is a protein kinase which gets its name from their function in phosphorylating glycogen synthase. GSK-3 also phosphorylates more than 50 substrates which contribute to many cellular

processes (Woodgett, 1990). GSK-3 consists of two isoforms, GSK-3 α and GSK-3 β . Both are structurally similar but different in their functions. GSK-3 β is profoundly involved in inflammatory response. Constitutively active GSK-3 β is negatively regulated by phosphorylation of serine residue at position 9 (Ser9), which may result in a decrease in pro-inflammatory cytokines and an increase in anti-inflammatory cytokine, IL-10 (Hu et al., 2006; Rehani et al., 2009). A study conducted by Hoeflich et al (2000) discovered that in GSK-3 β knockout mice suffered from degenerative liver (Hoeflich et al., 2000) while GSK-3 α knockout mice were still viable but increased glucose and insulin sensitivity with reduced fat mass (MacAulay et al., 2007). This may indicate that GSK-3 β is more important in regulating the immune response. GSK-3 plays a prominent role in promoting inflammation through production of pro-inflammatory cytokines and increased microglial migration (Yuskaitis & Jope, 2009). Regulation of inflammatory transcription factor NF- κ B may promote pro-inflammatory actions of GSK-3 via activation of toll-like receptors (Beurel, 2011).

IL-10 is a cytokine with anti-inflammatory properties produced by different types of immune cells including T lymphocytes, macrophages, dendritic cells, natural killer (NK) cells and mast cells. Production of IL-10 by almost all cells in the immune system indicates its crucial role in limiting the exaggeration of pro-inflammatory cytokines and to maintain normal homeostasis (Saraiva & O'Garra, 2010). IL-10 was initially known as cytokine synthesis inhibitory factor (CSIF) since it was first documented to inhibit activation of Th1 and the production of its associated cytokines (Fiorentino et al., 1989). Nuclear factor kappa B (NF- κ B) is a transcription factor that regulates the inflammatory responses via activation of toll-like receptor (TLR) and GSK-3 signaling (Rehani et al., 2009). Activation of this transcription factor caused depletion of CBP/p300, which essential in cyclic AMP response element (CREB) to produce IL-10 (Hofmann et al., 2012). Limited amounts of available CBP/p300 to bind with CREB binding protein lead to inhibition of IL-10 production which will promote inflammation. IL-10 inhibits inflammatory effects of T lymphocytes by interrupting the action of antigen presenting cells (APC) (Hofmann et al., 2010). The roles of anti-inflammatory effects of IL-10 have been studied to control the production of cytokines that promote pathogenicity of neuroinflammation particularly IFN γ and IL-17, produced by Th1 and Th17, respectively (Couper et al., 2008). In addition, the inhibitory action of IL-10 towards inflammatory effects of T lymphocytes could limit secretion of various pro-inflammatory cytokines and reduce activation and differentiation of macrophages, B cells and T cells (Shachar & Karin, 2013). GSK-3 also promotes activation of STAT3 to produce a high level of IL-6, which may contribute to Th17 development. (O'Shea & Paul, 2010). Many studies reported that Th17 is critical in the pathogenesis of various autoimmune diseases particularly in multiple sclerosis (MS) (Aranami & Yamamura, 2008; Maddur et al., 2012; Rothhammer et al., 2011). Other studies reported that inhibition of GSK-3 in monocytes and microglia also increased production of IL-10 (Huang et al., 2009; Wang et al., 2011). Furthermore, it has been suggested that inhibition of GSK-3 might provide a new intervention in treating autoimmune disease. In this study, we

hypothesize that GSK-3 inhibitor has protective effects against inflammatory action induced by LPS in microglia via upregulation of IL-10 production.

Rationale of the study

Continuous activation of microglia with a retained pro-inflammatory phenotype has been implicated in causing profound neuronal damage within the CNS parenchyma. Detection of activated microglia at the injury sites of neurodegenerative diseases including Alzheimer's disease (AD) and MS, underscore microglia activation as an important pathophysiological component of inflammatory and degenerative CNS conditions. Therefore, modulation of microglia responses is an active target for therapeutics of these degenerative diseases.

The GSK-3 inhibitor was found to exert anti-inflammatory properties on macrophage by limiting microglia to produce pro-inflammatory molecules including IL-6, TNF- α , glutamate, Monocyte chemoattractant protein-1 (MCP-1), and NO (Martin et al., 2005). We suggest that GSK-3 inhibitors may also modulate microglia inflammatory response. Although lithium chloride (LiCl) has potent anti-inflammatory effects, they might give adverse effects as LiCl has a low therapeutic index (Beurel et al., 2013) which makes SB216763, NP-12 and CHIR99021 ideal candidates among the GSK-3 inhibitors family for microglia modulation. To date, there are many kinds of literature concerning the modulatory effects of GSK-3 inhibitors on microglia responses. However, there are very limited studies that relate the ability of microglia in reducing production of pro-inflammatory cytokines with the expressions of activation and inhibitory state of microglia. Since GSK-3 plays an important role in propagating inflammation, we hypothesize that GSK-3 inhibitors can limit the excessive immune response by microglia activation and reverse the imbalance of homeostasis in the CNS.

Objectives of the research

The general objective of this project is to inhibit GSK-3 signalling in microglia in order to reduce microglial activation.

Whilst the specific objectives are:

1. To study the potential role of LPS in the production of NO and glutamate in microglia.
2. To evaluate the efficacy of GSK-3 inhibitors (LiCl, SB216763, NP-12 and CHIR99021) in limiting the production of pro-inflammatory components (e.g. NO, glutamate, IL-6, TNF- α , and MCP-1) in microglia.
3. To determine the anti-inflammatory effects of GSK-3 inhibition on IL-10 production in microglia.
4. To determine microglia activation status by expressions of CD11b and CD200R during inhibition of GSK-3
5. To determine the effects of GSK-3 inhibitors on the phosphorylation of GSK-3 β protein expressions.

REFERENCES

- Akiyama, H., & McGeer, P. (1990). Brain microglia constitutively express β -2 integrins. *Journal of neuroimmunology*, 30(1), 81-93.
- Aloisi, F. (2001). Immune function of microglia. *Glia*, 36(2), 165-179.
- Aloisi, F., Ria, F., & Adorini, L. (2000). Regulation of T-cell responses by CNS antigen-presenting cells: different roles for microglia and astrocytes. *Immunology today*, 21(3), 141-147.
- Alvarez, G., Muñoz-Montaño, J. R., Satrústegui, J., Avila, J., Bogóñez, E., & Díaz-Nido, J. (1999). Lithium protects cultured neurons against β -amyloid-induced neurodegeneration. *FEBS letters*, 453(3), 260-264.
- Aranami, T., & Yamamura, T. (2008). Th17 Cells and autoimmune encephalomyelitis (EAE/MS). *Allergology International*, 57(2), 115-120.
- Bal-Price, A., & Brown, G. C. (2001). Inflammatory neurodegeneration mediated by nitric oxide from activated glia-inhibiting neuronal respiration, causing glutamate release and excitotoxicity. *The Journal of Neuroscience*, 21(17), 6480-6491.
- Bal-Price, A., Matthias, A., & Brown, G. C. (2002). Stimulation of the NADPH oxidase in activated rat microglia removes nitric oxide but induces peroxynitrite production. *Journal of neurochemistry*, 80(1), 73-80.
- Banati, R., Newcombe, J., Gunn, R., Cagnin, A., Turkheimer, F., Heppner, F., . . . Miller, D. (2000). The peripheral benzodiazepine binding site in the brain in multiple sclerosis. *Brain*, 123(11), 2321-2337.
- Barna, B. P., Pettay, J., Barnett, G. H., Zhou, P., Iwasaki, K., & Estes, M. L. (1994). Regulation of monocyte chemoattractant protein-1 expression in adult human non-neoplastic astrocytes is sensitive to tumor necrosis factor (TNF) or antibody to the 55-kDa TNF receptor. *Journal of neuroimmunology*, 50(1), 101-107.
- Barron, K. D. (1995). The microglial cell. A historical review. *Journal of the neurological sciences*, 134, 57-68.
- Beaulieu, J.-M., Sotnikova, T. D., Yao, W.-D., Kockeritz, L., Woodgett, J. R., Gainetdinov, R. R., & Caron, M. G. (2004). Lithium antagonizes dopamine-dependent behaviors mediated by an AKT/glycogen synthase kinase 3 signaling cascade. *Proceedings of the National Academy of Sciences of the United States of America*, 101(14), 5099-5104.
- Belkaid, Y., Hoffmann, K. F., Mendez, S., Kamhawi, S., Udey, M. C., Wynn, T. A., & Sacks, D. L. (2001). The role of interleukin (IL)-10 in the persistence of *Leishmania major* in the skin after healing and the therapeutic potential of anti-IL-10 receptor antibody for sterile cure. *The Journal of experimental medicine*, 194(10), 1497-1506.
- Benimetskaya, L., Loike, J. D., Khaled, Z., Loike, G., Silverstein, S. C., Cao, L., . . . Stein, C. (1997). Mac-1 (CD11b/CD18) is an oligodeoxynucleotide-binding protein. *Nature medicine*, 3(4), 414-420.
- Beurel, E. (2011). Regulation by glycogen synthase kinase-3 of inflammation and T cells in CNS diseases. *Frontiers in molecular neuroscience*, 4.

- Beurel, E., Harrington, L. E., & Jope, R. S. (2013). Inflammatory T helper 17 cells promote depression-like behavior in mice. *Biological psychiatry*, 73(7), 622-630.
- Beurel, E., Kaidanovich-Beilin, O., Yeh, W.-I., Song, L., Palomo, V., Michalek, S. M., . . . Martinez, A. (2013). Regulation of Th1 cells and experimental autoimmune encephalomyelitis by glycogen synthase kinase-3. *The Journal of Immunology*, 190(10), 5000-5011.
- Beurel, E., Michalek, S. M., & Jope, R. S. (2010). Innate and adaptive immune responses regulated by glycogen synthase kinase-3 (GSK-3). *Trends in immunology*, 31(1), 24-31.
- Beurel, E., Song, L., & Jope, R. (2011). Inhibition of glycogen synthase kinase-3 is necessary for the rapid antidepressant effect of ketamine in mice. *Molecular psychiatry*, 16(11), 1068.
- Bhat, R., Xue, Y., Berg, S., Hellberg, S., Ormö, M., Nilsson, Y., . . . Borgegård, T. (2003). Structural insights and biological effects of glycogen synthase kinase 3-specific inhibitor AR-A014418. *Journal of Biological Chemistry*, 278(46), 45937-45945.
- Bhat, R. V., & Budd, S. L. (2002). GSK-3 β signalling: casting a wide net in Alzheimer's disease. *Neurosignals*, 11(5), 251-261.
- Bijur, G. N., De Sarno, P., & Jope, R. S. (2000). Glycogen Synthase Kinase-3 β Facilitates Staurosporine-and Heat Shock-induced Apoptosis PROTECTION BY LITHIUM. *Journal of Biological Chemistry*, 275(11), 7583-7590.
- Blasi, E., Barluzzi, R., Bocchini, V., Mazzolla, R., & Bistoni, F. (1990). Immortalization of murine microglial cells by a v-raf/v-myc carrying retrovirus. *Journal of neuroimmunology*, 27(2), 229-237.
- Blasko, I., Stampfer-Kountchev, M., Robatscher, P., Veerhuis, R., Eikelenboom, P., & Grubeck-Loebenstien, B. (2004). How chronic inflammation can affect the brain and support the development of Alzheimer's disease in old age: the role of microglia and astrocytes. *Aging cell*, 3(4), 169-176.
- Block, M. L., & Hong, J.-S. (2005). Microglia and inflammation-mediated neurodegeneration: multiple triggers with a common mechanism. *Progress in neurobiology*, 76(2), 77-98.
- Bocchini, V., Mazzolla, R., Barluzzi, R., Blasi, E., Sick, P., & Kettenmann, H. (1992). An immortalized cell line expresses properties of activated microglial cells. *Journal of neuroscience research*, 31(4), 616-621.
- Bode, J. G., Ehling, C., & Häussinger, D. (2012). The macrophage response towards LPS and its control through the p38 MAPK-STAT3 axis. *Cellular signalling*, 24(6), 1185-1194.
- Brown, G. C. (2010). Nitric oxide and neuronal death. *Nitric oxide*, 23(3), 153-165.
- Brown, G. C., & Cooper, C. (1994). Nanomolar concentrations of nitric oxide reversibly inhibit synaptosomal respiration by competing with oxygen at cytochrome oxidase. *FEBS letters*, 356(2-3), 295-298.
- Brown, G. C., & Cooper, C. E. (1994). Nanomolar concentrations of nitric oxide reversibly inhibit synaptosomal respiration by competing with oxygen at cytochrome oxidase. *FEBS letters*, 356(2), 295-298.
- Brown, G. C., & Neher, J. J. (2014). Microglial phagocytosis of live neurons. *Nature Reviews Neuroscience*, 15(4), 209-216.

- Burkhart, C., Liu, G. Y., Anderton, S. M., Metzler, B., & Wraith, D. C. (1999). Peptide-induced T cell regulation of experimental autoimmune encephalomyelitis: a role for IL-10. *International immunology*, 11(10), 1625-1634.
- Burstein, D. E., Seeley, P. J., & Greene, L. A. (1985). Lithium ion inhibits nerve growth factor-induced neurite outgrowth and phosphorylation of nerve growth factor-modulated microtubule-associated proteins. *The Journal of cell biology*, 101(3), 862-870.
- Chan, W., Kohsaka, S., & Rezaie, P. (2007). The origin and cell lineage of microglia—new concepts. *Brain research reviews*, 53(2), 344-354.
- Chang, E. Y., Guo, B., Doyle, S. E., & Cheng, G. (2007). Cutting edge: involvement of the type I IFN production and signaling pathway in lipopolysaccharide-induced IL-10 production. *The Journal of Immunology*, 178(11), 6705-6709.
- Chen, L.-F., & Greene, W. C. (2004). Shaping the nuclear action of NF- κ B. *Nature Reviews Molecular Cell Biology*, 5(5), 392-401.
- Cheng, Y., Pardo, M., de Souza Armini, R., Martinez, A., Mouhsine, H., Zagury, J.-F., . . . Beurel, E. (2016). Stress-induced neuroinflammation is mediated by GSK-3-dependent TLR4 signaling that promotes susceptibility to depression-like behavior. *Brain, Behavior, and Immunity*.
- Cherwinski, H. M., Murphy, C. A., Joyce, B. L., Bigler, M. E., Song, Y. S., Zurawski, S. M., . . . Zhang, S. (2005). The CD200 receptor is a novel and potent regulator of murine and human mast cell function. *The Journal of Immunology*, 174(3), 1348-1356.
- Cho, S.-Y., Park, S.-J., Kwon, M.-J., Jeong, T.-S., Bok, S.-H., Choi, W.-Y., . . . Lee, C.-S. (2003). Quercetin suppresses proinflammatory cytokines production through MAP kinases and NF- κ B pathway in lipopolysaccharide-stimulated macrophage. *Molecular and cellular biochemistry*, 243(1-2), 153-160.
- Coghlan, M. P., Culbert, A. A., Cross, D. A., Corcoran, S. L., Yates, J. W., Pearce, N. J., . . . Cox, L. R. (2000). Selective small molecule inhibitors of glycogen synthase kinase-3 modulate glycogen metabolism and gene transcription. *Chemistry & biology*, 7(10), 793-803.
- Copland, D. A., Calder, C. J., Raveney, B. J., Nicholson, L. B., Phillips, J., Cherwinski, H., . . . Dick, A. D. (2007). Monoclonal antibody-mediated CD200 receptor signaling suppresses macrophage activation and tissue damage in experimental autoimmune uveoretinitis. *The American journal of pathology*, 171(2), 580-588.
- Cortes-Canteli, M., Luna-Medina, R., Sanz-SanCristobal, M., Alvarez-Barrientos, A., Santos, A., & Perez-Castillo, A. (2008). CCAAT/enhancer binding protein β deficiency provides cerebral protection following excitotoxic injury. *Journal of cell science*, 121(8), 1224-1234.
- Couper, K. N., Blount, D. G., & Riley, E. M. (2008). IL-10: the master regulator of immunity to infection. *The Journal of Immunology*, 180(9), 5771-5777.
- Cushing, S. D., Berliner, J. A., Valente, A. J., Territo, M. C., Navab, M., Parhami, F., . . . Fogelman, A. M. (1990). Minimally modified low

- density lipoprotein induces monocyte chemotactic protein 1 in human endothelial cells and smooth muscle cells. *Proceedings of the National Academy of Sciences*, 87(13), 5134-5138.
- Cuzzocrea, S., Genovese, T., Mazzon, E., Crisafulli, C., Di Paola, R., Muià, C., . . . Thiemermann, C. (2006). Glycogen synthase kinase-3 β inhibition reduces secondary damage in experimental spinal cord trauma. *Journal of Pharmacology and Experimental Therapeutics*, 318(1), 79-89.
- Dasgupta, S., Jana, M., Liu, X., & Pahan, K. (2003). Role of very-late antigen-4 (VLA-4) in myelin basic protein-primed T cell contact-induced expression of proinflammatory cytokines in microglial cells. *Journal of Biological Chemistry*, 278(25), 22424-22431.
- de Waal Malefyt, R., Abrams, J., Bennett, B., Figdor, C. G., & De Vries, J. E. (1991). Interleukin 10 (IL-10) inhibits cytokine synthesis by human monocytes: an autoregulatory role of IL-10 produced by monocytes. *The Journal of experimental medicine*, 174(5), 1209-1220.
- del Río Hortega, P. (1932). *Microglia*: Hoeber.
- del Ser, T. (2010). Phase IIa clinical trial on Alzheimer's disease with NP12, a GSK-3 inhibitor. *Alzheimer's & Dementia*, 6(4), S147.
- Dentesano, G., Straccia, M., Ejarque-Ortiz, A., Tusell, J. M., Serratos, J., Saura, J., & Solà, C. (2012). Inhibition of CD200R1 expression by C/EBP beta in reactive microglial cells. *Journal of neuroinflammation*, 9(1), 1.
- Dheen, S. T., Jun, Y., Yan, Z., Tay, S. S., & Ang Ling, E. (2005). Retinoic acid inhibits expression of TNF- α and iNOS in activated rat microglia. *Glia*, 50(1), 21-31.
- Diamond, M. S., Staunton, D. E., De Fougerolles, A. R., Stacker, S. A., Garcia-Aguilar, J., Hibbs, M. L., & Springer, T. A. (1990). ICAM-1 (CD54): a counter-receptor for Mac-1 (CD11b/CD18). *The Journal of cell biology*, 111(6), 3129-3139.
- Dickson, D. W., Lee, S. C., Mattiace, L. A., Yen, S. H. C., & Brosnan, C. (1993). Microglia and cytokines in neurological disease, with special reference to AIDS and Alzheimer's disease. *Glia*, 7(1), 75-83.
- Domercq, M., & Nuria, V. (2013). Neurotransmitter signaling in the pathophysiology of microglia. *Frontiers in cellular neuroscience*, 7.
- Domercq, M., Sánchez-Gómez, M. V., Sherwin, C., Etxebarria, E., Fern, R., & Matute, C. (2007). System xc⁻ and glutamate transporter inhibition mediates microglial toxicity to oligodendrocytes. *The Journal of Immunology*, 178(10), 6549-6556.
- Eldar-Finkelman, H., & Martinez, A. (2011). GSK-3 inhibitors: preclinical and clinical focus on CNS. *Frontiers in molecular neuroscience*, 4.
- Featherstone, D. E. (2009). Intercellular glutamate signaling in the nervous system and beyond. *ACS chemical neuroscience*, 1(1), 4-12.
- Figuera-Losada, M., Rojas, C., & Slusher, B. S. (2014). Inhibition of Microglia Activation as a Phenotypic Assay in Early Drug Discovery. *Journal of biomolecular screening*, 19(1), 17-31.
- Fiorentino, D. F., Bond, M. W., & Mosmann, T. (1989). Two types of mouse T helper cell. IV. Th2 clones secrete a factor that inhibits cytokine production by Th1 clones. *The Journal of experimental medicine*, 170(6), 2081-2095.

- Fiorentino, D. F., Zlotnik, A., Mosmann, T., Howard, M., & O'garra, A. (1991). IL-10 inhibits cytokine production by activated macrophages. *The Journal of Immunology*, 147(11), 3815-3822.
- Frank-Cannon, T. C., Alto, L. T., McAlpine, F. E., & Tansey, M. G. (2009). Does neuroinflammation fan the flame in neurodegenerative diseases. *Mol Neurodegener*, 4(47), 1-13.
- Fujihara, K., Kotaki, M., & Ramakrishna, S. (2005). Guided bone regeneration membrane made of polycaprolactone/calcium carbonate composite nano-fibers. *Biomaterials*, 26(19), 4139-4147.
- Fukushima, K., West, G., Klein, J., Levine, A., & Fiocchi, C. (1993). Opposite modulatory activity of IL-10 and IL-4 on lamina propria mononuclear-cells (Ipmc) is stimulus-dependent. Paper presented at the Gastroenterology.
- Gilmor, M. L., Skelton, K. H., Nemeroff, C. B., & Owens, M. J. (2003). The effects of chronic treatment with the mood stabilizers valproic acid and lithium on corticotropin-releasing factor neuronal systems. *Journal of Pharmacology and Experimental Therapeutics*, 305(2), 434-439.
- Ginhoux, F., Greter, M., Leboeuf, M., Nandi, S., See, P., Gokhan, S., . . . Stanley, E. R. (2010). Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science*, 330(6005), 841-845.
- Giulian, D. (1999). Microglia and the immune pathology of Alzheimer disease. *The American Journal of Human Genetics*, 65(1), 13-18.
- Glass, C. K., Saijo, K., Winner, B., Marchetto, M. C., & Gage, F. H. (2010). Mechanisms underlying inflammation in neurodegeneration. *Cell*, 140(6), 918-934.
- Golde, S., Chandran, S., Brown, G. C., & Compston, A. (2002). Different pathways for iNOS- mediated toxicity in vitro dependent on neuronal maturation and NMDA receptor expression. *Journal of neurochemistry*, 82(2), 269-282.
- Goldmann, T., & Prinz, M. (2013). Role of microglia in CNS autoimmunity. *Clinical and Developmental Immunology*, 2013.
- Gorczynski, R., Chen, Z., Kai, Y., Lee, L., Wong, S., & Marsden, P. A. (2004). CD200 is a ligand for all members of the CD200R family of immunoregulatory molecules. *The Journal of Immunology*, 172(12), 7744-7749.
- Gorczynski, R. M., Chen, Z., Yu, K., & Hu, J. (2001). CD200 immunoadhesin suppresses collagen-induced arthritis in mice. *Clinical immunology*, 101(3), 328-334.
- Gravel, M., Béland, L.-C., Soucy, G., Abdelhamid, E., Rahimian, R., Gravel, C., & Kriz, J. (2016). IL-10 Controls Early Microglial Phenotypes and Disease Onset in ALS Caused by Misfolded Superoxide Dismutase 1. *The Journal of Neuroscience*, 36(3), 1031-1048.
- Green, H. F., & Nolan, Y. M. (2012). GSK-3 mediates the release of IL-1 β , TNF- α and IL-10 from cortical glia. *Neurochemistry international*, 61(5), 666-671.
- Grimsley, C., & Ravichandran, K. S. (2003). Cues for apoptotic cell engulfment: eat-me, don't eat-me and come-get-me signals. *Trends in cell biology*, 13(12), 648-656.

- Gui, B., Su, M., Chen, J., Jin, L., Wan, R., & Qian, Y. (2012). Neuroprotective effects of pretreatment with propofol in LPS-induced BV-2 microglia cells: role of TLR4 and GSK-3 β . *Inflammation*, 35(5), 1632-1640.
- Guix, F., Uribealago, I., Coma, M., & Munoz, F. (2005). The physiology and pathophysiology of nitric oxide in the brain. *Progress in neurobiology*, 76(2), 126-152.
- Hayashida, K., Nanki, T., Girschick, H., Yavuz, S., Ochi, T., & Lipsky, P. E. (2001). Synovial stromal cells from rheumatoid arthritis patients attract monocytes by producing MCP-1 and IL-8. *Arthritis research*, 3(2), 118-126.
- Heneka, M. T., & Feinstein, D. L. (2001). Expression and function of inducible nitric oxide synthase in neurons. *Journal of neuroimmunology*, 114(1), 8-18.
- Henriksen, E. J., Kinnick, T. R., Teachey, M. K., O'Keefe, M. P., Ring, D., Johnson, K. W., & Harrison, S. D. (2003). Modulation of muscle insulin resistance by selective inhibition of GSK-3 in Zucker diabetic fatty rats. *American Journal of Physiology-Endocrinology and Metabolism*, 284(5), E892-E900.
- Herzenberg, L. A., Tung, J., Moore, W. A., Herzenberg, L. A., & Parks, D. R. (2006). Interpreting flow cytometry data: a guide for the perplexed. *Nature immunology*, 7(7), 681-685.
- Hetman, M., Cavanaugh, J. E., Kimelman, D., & Xia, Z. (2000). Role of glycogen synthase kinase-3 β in neuronal apoptosis induced by trophic withdrawal. *The Journal of neuroscience*, 20(7), 2567-2574.
- Hill, K. E., Zollinger, L. V., Watt, H. E., Carlson, N. G., & Rose, J. W. (2004). Inducible nitric oxide synthase in chronic active multiple sclerosis plaques: distribution, cellular expression and association with myelin damage. *Journal of neuroimmunology*, 151(1), 171-179.
- Hoeflich, K. P., Luo, J., Rubie, E. A., Tsao, M.-S., Jin, O., & Woodgett, J. R. (2000). Requirement for glycogen synthase kinase-3 β in cell survival and NF- κ B activation. *Nature*, 406(6791), 86-90.
- Hoek, R. M., Ruuls, S. R., Murphy, C. A., Wright, G. J., Goddard, R., Zurawski, S. M., . . . Brown, M. H. (2000). Down-regulation of the macrophage lineage through interaction with OX2 (CD200). *Science*, 290(5497), 1768-1771.
- Hofmann, C., Dunger, N., Schölmerich, J., Falk, W., & Obermeier, F. (2010). Glycogen synthase kinase 3- β : A master regulator of toll-like receptor-mediated chronic intestinal inflammation. *Inflammatory bowel diseases*, 16(11), 1850-1858.
- Hofmann, S., Rösen-Wolff, A., Tsokos, G., & Hedrich, C. (2012). Biological properties and regulation of IL-10 related cytokines and their contribution to autoimmune disease and tissue injury. *Clinical immunology*, 143(2), 116-127.
- Hong, M., Chen, D. C., Klein, P. S., & Lee, V. M.-Y. (1997). Lithium reduces tau phosphorylation by inhibition of glycogen synthase kinase-3. *Journal of Biological Chemistry*, 272(40), 25326-25332.
- Hongisto, V., Smeds, N., Brecht, S., Herdegen, T., Courtney, M. J., & Coffey, E. T. (2003). Lithium blocks the c-Jun stress response and protects neurons via its action on glycogen synthase kinase 3. *Molecular and cellular biology*, 23(17), 6027-6036.

- Hu, S., Begum, A. N., Jones, M. R., Oh, M. S., Beech, W. K., Beech, B. H., . . . Kim, P. C. (2009). GSK-3 inhibitors show benefits in an Alzheimer's disease (AD) model of neurodegeneration but adverse effects in control animals. *Neurobiology of disease*, 33(2), 193-206.
- Hu, X., Paik, P. K., Chen, J., Yarinina, A., Kockeritz, L., Lu, T. T., . . . Ivashkiv, L. B. (2006). IFN- γ suppresses IL-10 production and synergizes with TLR2 by regulating GSK-3 and CREB/AP-1 proteins. *Immunity*, 24(5), 563-574.
- Huang, W. C., Lin, Y. S., Wang, C. Y., Tsai, C. C., Tseng, H. C., Chen, C. L., . . . Hong, J. S. (2009). Glycogen synthase kinase-3 negatively regulates anti-inflammatory interleukin-10 for lipopolysaccharide-induced iNOS/NO biosynthesis and RANTES production in microglial cells. *Immunology*, 128(1pt2), e275-e286.
- Hur, E.-M., & Zhou, F.-Q. (2010). GSK-3 signalling in neural development. *Nature Reviews Neuroscience*, 11(8), 539-551.
- Ilouz, R., Pietrokovski, S., Eisenstein, M., & Eldar-Finkelman, H. (2008). New Insights into the Autoinhibition Mechanism of Glycogen Synthase Kinase-3 β . *Journal of molecular biology*, 383(5), 999-1007.
- Ishiguro, K., Shiratsuchi, A., Sato, S., Omori, A., Arioka, M., Kobayashi, S., . . . Imahori, K. (1993). Glycogen synthase kinase 3 β is identical to tau protein kinase I generating several epitopes of paired helical filaments. *FEBS letters*, 325(3), 167-172.
- Jack, C., Ruffini, F., Bar-Or, A., & Antel, J. P. (2005). Microglia and multiple sclerosis. *Journal of neuroscience research*, 81(3), 363-373.
- Jana, M., Liu, X., Koka, S., Ghosh, S., Petro, T. M., & Pahan, K. (2001). Ligation of CD40 stimulates the induction of nitric-oxide synthase in microglial cells. *Journal of Biological Chemistry*, 276(48), 44527-44533.
- Jekabsone, A., Neher, J. J., Borutaite, V., & Brown, G. C. (2007). Nitric oxide from neuronal nitric oxide synthase sensitises neurons to hypoxia-induced death via competitive inhibition of cytochrome oxidase. *Journal of neurochemistry*, 103(1), 346-356.
- Jenmalm, M. C., Cherwinski, H., Bowman, E. P., Phillips, J. H., & Sedgwick, J. D. (2006). Regulation of myeloid cell function through the CD200 receptor. *The Journal of Immunology*, 176(1), 191-199.
- Jeong, Y.-H., Kim, Y., Song, H., Chung, Y. S., Park, S. B., & Kim, H.-S. (2014). Anti-Inflammatory Effects of α -Galactosylceramide Analogs in Activated Microglia: Involvement of the p38 MAPK Signaling Pathway. *PloS one*, 9(2), e87030.
- Joje, R. S., Yuskaitis, C. J., & Beurel, E. (2007). Glycogen synthase kinase-3 (GSK-3): inflammation, diseases, and therapeutics. *Neurochemical research*, 32(4-5), 577-595.
- Kallaur, A. P., Kaimen-Maciell, D. R., Morimoto, H. K., Ehara Watanabe, M. A., Georgeto, S. M., & Reiche, E. M. (2011). Genetic polymorphisms associated with the development and clinical course of multiple sclerosis (review). *International journal of molecular medicine*, 28(4), 467-479.
- Kaytor, M. D., & Orr, H. T. (2002). The GSK-3 β signaling cascade and neurodegenerative disease. *Current opinion in neurobiology*, 12(3), 275-278.

- Kettenmann, H., Hanisch, U.-K., Noda, M., & Verkhratsky, A. (2011). Physiology of microglia. *Physiological reviews*, 91(2), 461-553.
- Kim, H. J., & Thayer, S. A. (2009). Lithium increases synapse formation between hippocampal neurons by depleting phosphoinositides. *Molecular pharmacology*, 75(5), 1021-1030.
- King, M. K., Pardo, M., Cheng, Y., Downey, K., Jope, R. S., & Beurel, E. (2014). Glycogen synthase kinase-3 inhibitors: rescuers of cognitive impairments. *Pharmacology & therapeutics*, 141(1), 1-12.
- Klein, P. S., & Melton, D. A. (1996). A molecular mechanism for the effect of lithium on development. *Proceedings of the National Academy of Sciences*, 93(16), 8455-8459.
- Knijff, E. M., Nadine Breunis, M., Kupka, R. W., De Wit, H. J., Ruwhof, C., Akkerhuis, G. W., . . . Drexhage, H. A. (2007). An imbalance in the production of IL-1 β and IL-6 by monocytes of bipolar patients: restoration by lithium treatment. *Bipolar disorders*, 9(7), 743-753.
- Ko, Y. C., Chien, H. F., Jiang- Shieh, Y. F., Chang, C. Y., Pai, M. H., Huang, J. P., . . . Wu, C. H. (2009). Endothelial CD200 is heterogeneously distributed, regulated and involved in immune cell–endothelium interactions. *Journal of anatomy*, 214(1), 183-195.
- Koning, N., Swaab, D. F., Hoek, R. M., & Huitinga, I. (2009). Distribution of the immune inhibitory molecules CD200 and CD200R in the normal central nervous system and multiple sclerosis lesions suggests neuron-glia and glia-glia interactions. *Journal of Neuropathology & Experimental Neurology*, 68(2), 159-167.
- Koning, N., Van Eijk, M., Pouwels, W., Brouwer, M. S., Voehringer, D., Huitinga, I., . . . Hamann, J. (2010). Expression of the inhibitory CD200 receptor is associated with alternative macrophage activation. *Journal of innate immunity*, 2(2), 195-200.
- Kozela, E., Pietr, M., Juknat, A., Rimmerman, N., Levy, R., & Vogel, Z. (2010). Cannabinoids Δ 9-tetrahydrocannabinol and cannabidiol differentially inhibit the lipopolysaccharide-activated NF- κ B and interferon- β /STAT proinflammatory pathways in BV-2 microglial cells. *Journal of Biological Chemistry*, 285(3), 1616-1626.
- Kreutzberg, G. W. (1996). Microglia: a sensor for pathological events in the CNS. *Trends in neurosciences*, 19(8), 312-318.
- Kühn, R., Löhler, J., Rennick, D., Rajewsky, K., & Müller, W. (1993). Interleukin-10-deficient mice develop chronic enterocolitis. *Cell*, 75(2), 263-274.
- Kunkel, S. L., Standiford, T., Kasahara, K., & Strieter, R. M. (1991). Stimulus specific induction of monocyte chemotactic protein-1 (MCP-1) gene expression *Chemotactic Cytokines* (pp. 65-71): Springer.
- Ladeby, R., Wirenfeldt, M., Dalmau, I., Gregersen, R., García- Ovejero, D., Babcock, A., . . . Finsen, B. (2005). Proliferating resident microglia express the stem cell antigen CD34 in response to acute neural injury. *Glia*, 50(2), 121-131.
- Langmann, T. (2007). Microglia activation in retinal degeneration. *Journal of leukocyte biology*, 81(6), 1345-1351.
- Lee, P., Lee, J., Kim, S., Lee, M.-S., Yagita, H., Kim, S. Y., . . . Suk, K. (2001). NO as an autocrine mediator in the apoptosis of activated

- microglial cells: correlation between activation and apoptosis of microglial cells. *Brain research*, 892(2), 380-385.
- Leroy, K., & Brion, J.-P. (1999). Developmental expression and localization of glycogen synthase kinase-3 β in rat brain. *Journal of chemical neuroanatomy*, 16(4), 279-293.
- Li, M.-C., & He, S.-H. (2004). IL-10 and its related cytokines for treatment of inflammatory bowel disease. *World Journal of Gastroenterology*, 10(5), 620-625.
- Li, W., Zhou, H., Abujarour, R., Zhu, S., Young Joo, J., Lin, T., . . . Ding, S. (2009). Generation of Human- Induced Pluripotent Stem Cells in the Absence of Exogenous Sox2. *Stem cells*, 27(12), 2992-3000.
- Lipina, T. V., Kaidanovich- Beilin, O., Patel, S., Wang, M., Clapcote, S. J., Liu, F., . . . Roder, J. C. (2011). Genetic and pharmacological evidence for schizophrenia- related Disc1 interaction with GSK- 3. *Synapse*, 65(3), 234-248.
- Liu, B., GAO, H. M., WANG, J. Y., JEOHN, G. H., Cooper, C. L., & HONG, J. S. (2002). Role of nitric oxide in inflammation- mediated neurodegeneration. *Annals of the New York Academy of Sciences*, 962(1), 318-331.
- Liu, X., Jana, M., Dasgupta, S., Koka, S., He, J., Wood, C., & Pahan, K. (2002). Human immunodeficiency virus type 1 (HIV-1) tat induces nitric-oxide synthase in human astroglia. *Journal of Biological Chemistry*, 277(42), 39312-39319.
- MacAulay, K., Doble, B. W., Patel, S., Hansotia, T., Sinclair, E. M., Drucker, D. J., . . . Woodgett, J. R. (2007). Glycogen synthase kinase 3 α -specific regulation of murine hepatic glycogen metabolism. *Cell metabolism*, 6(4), 329-337.
- Mack, C. L., Vanderlugt-Castaneda, C. L., Neville, K. L., & Miller, S. D. (2003). Microglia are activated to become competent antigen presenting and effector cells in the inflammatory environment of the Theiler's virus model of multiple sclerosis. *Journal of neuroimmunology*, 144(1), 68-79.
- Maddur, M. S., Miossec, P., Kaveri, S. V., & Bayry, J. (2012). Th17 cells: biology, pathogenesis of autoimmune and inflammatory diseases, and therapeutic strategies. *The American journal of pathology*, 181(1), 8-18.
- Maecker, H. T., & Trotter, J. (2006). Flow cytometry controls, instrument setup, and the determination of positivity. *Cytometry Part A*, 69(9), 1037-1042.
- Martin, M., Rehani, K., Jope, R. S., & Michalek, S. M. (2005). Toll-like receptor-mediated cytokine production is differentially regulated by glycogen synthase kinase 3. *Nature immunology*, 6(8), 777-784.
- Masliah, E., Terry, R. D., Alford, M., DeTeresa, R., & Hansen, L. (1991). Cortical and subcortical patterns of synaptophysinlike immunoreactivity in Alzheimer's disease. *The American journal of pathology*, 138(1), 235.
- Matsuura, T., West, G., Klein, J., Levine, A., Kusugami, K., Morise, K., & Fiocchi, C. (1993). immune activation gene-products are resistant to il4 inhibitory activity in crohns-disease (CD). Paper presented at the Gastroenterology.

- McFarland, H. I., Nahill, S. R., Maciaszek, J. W., & Welsh, R. M. (1992). CD11b (Mac-1): a marker for CD8+ cytotoxic T cell activation and memory in virus infection. *The Journal of Immunology*, 149(4), 1326-1333.
- McGeer, P. L., Itagaki, S., Tago, H., & McGeer, E. G. (1987). Reactive microglia in patients with senile dementia of the Alzheimer type are positive for the histocompatibility glycoprotein HLA-DR. *Neuroscience letters*, 79(1), 195-200.
- McGeer, P. L., & McGeer, E. G. (2004). Inflammation and neurodegeneration in Parkinson's disease. *Parkinsonism & related disorders*, 10, S3-S7.
- McKercher, S. R., Torbett, B. E., Anderson, K. L., Henkel, G. W., Vestal, D. J., Baribault, H., . . . Paige, C. J. (1996). Targeted disruption of the PU. 1 gene results in multiple hematopoietic abnormalities. *The EMBO journal*, 15(20), 5647.
- McNaught, K. S. P., & Brown, G. C. (1998). Nitric oxide causes glutamate release from brain synaptosomes. *Journal of neurochemistry*, 70(4), 1541-1546.
- Medzhitov, R., & Janeway, C. A. (1997). Innate immunity: the virtues of a nonclonal system of recognition. *Cell*, 91(3), 295-298.
- Miller, M. F., Mitchell, T. G., Storkus, W., & Dawson, J. R. (1990). Human natural killer cells do not inhibit growth of *Cryptococcus neoformans* in the absence of antibody. *Infection and immunity*, 58(3), 639-645.
- Minghetti, L., & Levi, G. (1998). Microglia as effector cells in brain damage and repair: focus on prostanoids and nitric oxide. *Progress in neurobiology*, 54(1), 99-125.
- Mitrovic, B., Ignarro, L., Montestrucque, S., Smoll, A., & Merrill, J. (1994). Nitric oxide as a potential pathological mechanism in demyelination: Its differential effects on primary glial cells in vitro. *Neuroscience*, 61(3), 575-585.
- Moncada, S., Palmer, R., & Higgs, E. (1991). Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacological reviews*, 43(2), 109-142.
- Mosmann, T. R., & Moore, K. W. (1991). The role of IL-10 in crossregulation of TH1 and TH2 responses. *Parasitology Today*, 7(3), 49-53.
- Murphy, S. (2000). Production of nitric oxide by glial cells: regulation and potential roles in the CNS. *Glia*, 29(1), 1-13.
- Nagao, M., & Hayashi, H. (2009). Glycogen synthase kinase-3beta is associated with Parkinson's disease. *Neuroscience letters*, 449(2), 103-107.
- Nakajima, K., & Kohsaka, S. (2001). Microglia: activation and their significance in the central nervous system. *Journal of biochemistry*, 130(2), 169-175.
- Nayak, D., Huo, Y., Kwang, W., Pushparaj, P., Kumar, S., Ling, E.-A., & Dheen, S. (2010). Sphingosine kinase 1 regulates the expression of proinflammatory cytokines and nitric oxide in activated microglia. *Neuroscience*, 166(1), 132-144.
- Neumann, H. (2001). Control of glial immune function by neurons. *Glia*, 36(2), 191-199.

- Nguyen, M. D., Julien, J.-P., & Rivest, S. (2002). Innate immunity: the missing link in neuroprotection and neurodegeneration? *Nature Reviews Neuroscience*, 3(3), 216-227.
- Noble, W., Planel, E., Zehr, C., Olm, V., Meyerson, J., Suleman, F., . . . Feinstein, B. (2005). Inhibition of glycogen synthase kinase-3 by lithium correlates with reduced tauopathy and degeneration in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, 102(19), 6990-6995.
- Novetsky, A. P., Thompson, D. M., Zigelboim, I., Thaker, P. H., Powell, M. A., Mutch, D. G., & Goodfellow, P. J. (2013). Lithium and inhibition of GSK-3 β as a potential therapy for serous ovarian cancer. *International journal of gynecological cancer: official journal of the International Gynecological Cancer Society*, 23(2), 361.
- O'Neill, E. J., Day, M. J., & Wraith, D. C. (2006). IL-10 is essential for disease protection following intranasal peptide administration in the C57BL/6 model of EAE. *Journal of neuroimmunology*, 178(1), 1-8.
- O'Shea, J. J., & Paul, W. E. (2010). Mechanisms underlying lineage commitment and plasticity of helper CD4+ T cells. *Science*, 327(5969), 1098-1102.
- Oh, Y. T., Lee, J. Y., Lee, J., Kim, H., Yoon, K.-S., Choe, W., & Kang, I. (2009). Oleic acid reduces lipopolysaccharide-induced expression of iNOS and COX-2 in BV2 murine microglial cells: Possible involvement of reactive oxygen species, p38 MAPK, and IKK/NF- κ B signaling pathways. *Neuroscience letters*, 464(2), 93-97.
- Olajide, O. A., Bhatia, H. S., de Oliveira, A. C., Wright, C. W., & Fiebich, B. L. (2013). Inhibition of neuroinflammation in LPS-activated microglia by cryptolepine. *Evidence-Based Complementary and Alternative Medicine*, 2013.
- Omer, F. M., de Souza, J. B., & Riley, E. M. (2003). Differential induction of TGF- β regulates proinflammatory cytokine production and determines the outcome of lethal and nonlethal *Plasmodium yoelii* infections. *The Journal of Immunology*, 171(10), 5430-5436.
- Paulie, S., Perlmann, H., & Perlmann, P. (2003). Enzyme-linked Immunosorbent Assay. *eLS*.
- Pei, J.-J., Braak, E., Braak, H., Grundke-Iqbal, I., Iqbal, K., Winblad, B., & Cowburn, R. F. (1999). Distribution of Active Glycogen Synthase Kinase 3 [beta](GSK-3 [beta]) in Brains Staged for Alzheimer Disease Neurofibrillary Changes. *Journal of Neuropathology & Experimental Neurology*, 58(9), 1010-1019.
- Pérez, M., Hernández, F., Lim, F., Diaz-Nido, J., & Avila, J. (2003). Chronic lithium treatment decreases mutant tau protein aggregation in a transgenic mouse model. *Journal of Alzheimer's Disease*, 5(4), 301-308.
- Polazzi, E., & Monti, B. (2010). Microglia and neuroprotection: from in vitro studies to therapeutic applications. *Progress in neurobiology*, 92(3), 293-315.
- Posadas, I., Terencio, M. C., Guillén, I., Ferrándiz, M. L., Coloma, J., Payá, M., & Alcaraz, M. J. (2000). Co-regulation between cyclo-oxygenase-2 and inducible nitric oxide synthase expression in the time-course of

- murine inflammation. *Naunyn-Schmiedeberg's archives of pharmacology*, 361(1), 98-106.
- Ransohoff, R. M., & Cardona, A. E. (2010). The myeloid cells of the central nervous system parenchyma. *Nature*, 468(7321), 253-262.
- Rehani, K., Wang, H., Garcia, C. A., Kinane, D. F., & Martin, M. (2009). Toll-like receptor-mediated production of IL-1Ra is negatively regulated by GSK-3 via the MAPK ERK1/2. *The Journal of Immunology*, 182(1), 547-553.
- Reis, H. J., Guatimosim, C., Paquet, M., Santos, M., Ribeiro, F. M., Kummer, A., . . . Teixeira, A. L. (2009). Neuro-transmitters in the central nervous system & their implication in learning and memory processes. *Current medicinal chemistry*, 16(7), 796-840.
- Ribé, E. M., Pérez, M., Puig, B., Gich, I., Lim, F., Cuadrado, M., . . . Nieto, M. (2005). Accelerated amyloid deposition, neurofibrillary degeneration and neuronal loss in double mutant APP/tau transgenic mice. *Neurobiology of disease*, 20(3), 814-822.
- Ring, A., Braun, J. S., Nizet, V., Stremmel, W., & Shenep, J. L. (2000). Group B streptococcal β -hemolysin induces nitric oxide production in murine macrophages. *Journal of Infectious Diseases*, 182(1), 150-157.
- Ring, D. B., Johnson, K. W., Henriksen, E. J., Nuss, J. M., Goff, D., Kinnick, T. R., . . . Slabiak, T. (2003). Selective glycogen synthase kinase 3 inhibitors potentiate insulin activation of glucose transport and utilization in vitro and in vivo. *Diabetes*, 52(3), 588-595.
- Rivest, S. (2009). Regulation of innate immune responses in the brain. *Nature Reviews Immunology*, 9(6), 429-439.
- Rockenstein, E., Torrance, M., Adame, A., Mante, M., Bar-on, P., Rose, J. B., . . . Masliah, E. (2007). Neuroprotective effects of regulators of the glycogen synthase kinase-3 β signaling pathway in a transgenic model of Alzheimer's disease are associated with reduced amyloid precursor protein phosphorylation. *The Journal of neuroscience*, 27(8), 1981-1991.
- Rose, J. W., Hill, K. E., Watt, H. E., & Carlson, N. G. (2004). Inflammatory cell expression of cyclooxygenase-2 in the multiple sclerosis lesion. *Journal of neuroimmunology*, 149(1), 40-49.
- Rosenblum, M. D., Woodliff, J. E., Madsen, N. A., McOlash, L. J., Keller, M. R., & Truitt, R. L. (2005). Characterization of CD200-receptor expression in the murine epidermis. *Journal of investigative dermatology*, 125(6), 1130-1138.
- Rothhammer, V., Heink, S., Petermann, F., Srivastava, R., Claussen, M. C., Hemmer, B., & Korn, T. (2011). Th17 lymphocytes traffic to the central nervous system independently of α 4 integrin expression during EAE. *The Journal of experimental medicine*, 208(12), 2465-2476.
- Roy, A., Fung, Y. K., Liu, X., & Pahan, K. (2006). Up-regulation of microglial CD11b expression by nitric oxide. *Journal of Biological Chemistry*, 281(21), 14971-14980.
- Roy, A., Jana, A., Yatish, K., Freidt, M. B., Fung, Y. K., Martinson, J. A., & Pahan, K. (2008). Reactive oxygen species up-regulate CD11b in microglia via nitric oxide: Implications for neurodegenerative diseases. *Free Radical Biology and Medicine*, 45(5), 686-699.

- Roy, A., Liu, X., & Pahan, K. (2008). Myelin basic protein-primed T cells induce neurotrophins in glial cells via $\alpha\beta 3$ integrin. *VOLUME 282 (2007) PAGES 32222-32232. Journal of Biological Chemistry, 283(6)*, 3688-3688.
- Ryves, W. J., Dajani, R., Pearl, L., & Harwood, A. J. (2002). Glycogen synthase kinase-3 inhibition by lithium and beryllium suggests the presence of two magnesium binding sites. *Biochemical and biophysical research communications, 290(3)*, 967-972.
- Ryves, W. J., & Harwood, A. J. (2001). Lithium inhibits glycogen synthase kinase-3 by competition for magnesium. *Biochemical and biophysical research communications, 280(3)*, 720-725.
- Sabat, R. (2010). IL-10 family of cytokines. *Cytokine & growth factor reviews, 21(5)*, 315-324.
- Saha, R. N., Liu, X., & Pahan, K. (2006). Up-regulation of BDNF in astrocytes by TNF- α : a case for the neuroprotective role of cytokine. *Journal of Neuroimmune Pharmacology, 1(3)*, 212-222.
- Saraiva, M., & O'Garra, A. (2010). The regulation of IL-10 production by immune cells. *Nature Reviews Immunology, 10(3)*, 170-181.
- Sattler, R., & Tymianski, M. (2001). Molecular mechanisms of glutamate receptor-mediated excitotoxic neuronal cell death. *Molecular neurobiology, 24(1-3)*, 107-129.
- Savill, J., Dransfield, I., Gregory, C., & Haslett, C. (2002). A blast from the past: clearance of apoptotic cells regulates immune responses. *Nature Reviews Immunology, 2(12)*, 965-975.
- Schönrock, L., Kuhlmann, T., Adler, S., Bitsch, A., & Brück, W. (1998). Identification of glial cell proliferation in early multiple sclerosis lesions. *Neuropathology and applied neurobiology, 24(4)*, 320-330.
- Schreiber, S., Heinig, T., Thiele, H.-G., & Raedler, A. (1995). Immunoregulatory role of interleukin 10 in patients with inflammatory bowel disease. *Gastroenterology, 108(5)*, 1434-1444.
- Schroeder, J., Bell, L., Janas, M., Turner, M., & Kontoyiannis, D. L. (2013). Pharmacological Inhibition of Glycogen Synthase Kinase 3 Regulates T Cell Development. *PLoS one, 8(3)*, e58501.
- Seira, O., Gavín, R., Gil, V., Llorens, F., Rangel, A., Soriano, E., & Del Río, J. A. (2010). Neurites regrowth of cortical neurons by GSK-3 β inhibition independently of Nogo receptor 1. *Journal of neurochemistry, 113(6)*, 1644-1658.
- Seira Oriach, O., Gavín Marín, R., Gil, V., Llorens, F., Rangel, A., Soriano García, E., & Río Fernández, J. A. d. (2010). Neurites regrowth of cortical neurons by GSK-3 β inhibition independently of Nogo Receptor 1. *Journal of Neurochemistry, 2010, vol. 113, num. 6, p. 1644-1658.*
- Selenica, M. L., Jensen, H. S., Larsen, A. K., Pedersen, M., Helboe, L., Leist, M., & Lotharius, J. (2007). Efficacy of small-molecule glycogen synthase kinase-3 inhibitors in the postnatal rat model of tau hyperphosphorylation. *British journal of pharmacology, 152(6)*, 959-979.
- Sereno, L., Coma, M., Rodriguez, M., Sanchez-Ferrer, P., Sanchez, M., Gich, I., . . . Guardia-Laguarta, C. (2009). A novel GSK-3 β inhibitor reduces Alzheimer's pathology and rescues neuronal loss in vivo. *Neurobiology of disease, 35(3)*, 359-367.

- Shachar, I., & Karin, N. (2013). The dual roles of inflammatory cytokines and chemokines in the regulation of autoimmune diseases and their clinical implications. *Journal of leukocyte biology*, 93(1), 51-61.
- Shimazu, R., Akashi, S., Ogata, H., Nagai, Y., Fukudome, K., Miyake, K., & Kimoto, M. (1999). MD-2, a molecule that confers lipopolysaccharide responsiveness on Toll-like receptor 4. *The Journal of experimental medicine*, 189(11), 1777-1782.
- Sittampalam, G. S., Gal-Edd, N., Arkin, M., Auld, D., Austin, C., Bejcek, B., . . . Li, Z. (2013). *Cell Viability Assays*.
- Sørensen, T. L., Ransohoff, R., Strieter, R., & Sellebjerg, F. (2004). Chemokine CCL2 and chemokine receptor CCR2 in early active multiple sclerosis. *European Journal of Neurology*, 11(7), 445-449.
- Stevens, B., Allen, N. J., Vazquez, L. E., Howell, G. R., Christopherson, K. S., Nouri, N., . . . Stafford, B. (2007). The classical complement cascade mediates CNS synapse elimination. *Cell*, 131(6), 1164-1178.
- Stewart, V., Heslegrave, A., Brown, G., Clark, J., & Heales, S. (2002). Nitric oxide- dependent damage to neuronal mitochondria involves the NMDA receptor. *European Journal of Neuroscience*, 15(3), 458-464.
- Straccia, M., Gresa-Arribas, N., Dentesano, G., Ejarque-Ortiz, A., Tusell, J. M., Serratosa, J., . . . Saura, J. (2011). Pro-inflammatory gene expression and neurotoxic effects of activated microglia are attenuated by absence of CCAAT/enhancer binding protein β . *Journal of neuroinflammation*, 8(1), 1.
- Su, Y., Ryder, J., Li, B., Wu, X., Fox, N., Solenberg, P., . . . Liu, F. (2004). Lithium, a common drug for bipolar disorder treatment, regulates amyloid- β precursor protein processing. *Biochemistry*, 43(22), 6899-6908.
- Sun, J., Zhang, X., Broderick, M., & Fein, H. (2003). Measurement of nitric oxide production in biological systems by using Griess reaction assay. *Sensors*, 3(8), 276-284.
- Sun, X., Sato, S., Murayama, O., Murayama, M., Park, J.-M., Yamaguchi, H., & Takashima, A. (2002). Lithium inhibits amyloid secretion in COS7 cells transfected with amyloid precursor protein C100. *Neuroscience letters*, 321(1), 61-64.
- Takahashi-Yanaga, F. (2013). Activator or inhibitor? GSK-3 as a new drug target. *Biochemical pharmacology*, 86(2), 191-199.
- Takahashi, M., Yasutake, K., & Tomizawa, K. (1999). Lithium inhibits neurite growth and tau protein kinase I/glycogen synthase kinase-3b-dependent phosphorylation of juvenile tau in cultured hippocampal neurons. *J. Neurochem*, 73, 2073-2083.
- Takashima, A., Noguchi, K., Michel, G., Mercken, M., Hoshi, M., Ishiguro, K., & Imahori, K. (1996). Exposure of rat hippocampal neurons to amyloid β peptide (25–35) induces the inactivation of phosphatidylinositol-3 kinase and the activation of tau protein kinase I/glycogen synthase kinase-3 β . *Neuroscience letters*, 203(1), 33-36.
- Tan, S. W., Ramasamy, R., Abdullah, M., & Vidyadaran, S. (2011). Inhibitory effects of palm α -, γ -and δ -tocotrienol on lipopolysaccharide-induced nitric oxide production in BV2 microglia. *Cellular immunology*, 271(2), 205-209.

- Tang, D., Kang, R., Coyne, C. B., Zeh, H. J., & Lotze, M. T. (2012). PAMPs and DAMPs: signal 0s that spur autophagy and immunity. *Immunological reviews*, 249(1), 158-175.
- Tansey, M. G., McCoy, M. K., & Frank-Cannon, T. C. (2007). Neuroinflammatory mechanisms in Parkinson's disease: potential environmental triggers, pathways, and targets for early therapeutic intervention. *Experimental neurology*, 208(1), 1-25.
- Tansey, M. G., & Wyss-Coray, T. (2008). Cytokines in CNS inflammation and disease *Central nervous system diseases and inflammation* (pp. 59-106): Springer.
- Taylor, N., McConnachie, K., Calder, C., Dawson, R., Dick, A., Sedgwick, J. D., & Liversidge, J. (2005). Enhanced tolerance to autoimmune uveitis in CD200-deficient mice correlates with a pronounced Th2 switch in response to antigen challenge. *The Journal of Immunology*, 174(1), 143-154.
- Thameem Dheen, S., Kaur, C., & Ling, E.-A. (2007). Microglial activation and its implications in the brain diseases. *Current medicinal chemistry*, 14(11), 1189-1197.
- Tsikas, D. (2007). Analysis of nitrite and nitrate in biological fluids by assays based on the Griess reaction: appraisal of the Griess reaction in the L-arginine/nitric oxide area of research. *Journal of Chromatography B*, 851(1), 51-70.
- Ulevitch, R. J., & Tobias, P. S. (1999). Recognition of gram-negative bacteria and endotoxin by the innate immune system. *Current opinion in immunology*, 11(1), 19-22.
- Valerio, A., Bertolotti, P., Delbarba, A., Perego, C., Dossena, M., Ragni, M., . . . Nisoli, E. (2011). Glycogen synthase kinase-3 inhibition reduces ischemic cerebral damage, restores impaired mitochondrial biogenesis and prevents ROS production. *Journal of neurochemistry*, 116(6), 1148-1159.
- Vidyadaran, S., Ooi, Y. Y., Subramaiam, H., Badiei, A., Abdullah, M., Ramasamy, R., & Seow, H. F. (2009). Effects of macrophage colony-stimulating factor on microglial responses to lipopolysaccharide and beta amyloid. *Cellular immunology*, 259(1), 105-110.
- Voehringer, D., Rosen, D. B., Lanier, L. L., & Locksley, R. M. (2004). CD200 receptor family members represent novel DAP12-associated activating receptors on basophils and mast cells. *Journal of Biological Chemistry*, 279(52), 54117-54123.
- Walker, D. G., Dalsing-Hernandez, J. E., Campbell, N. A., & Lue, L.-F. (2009). Decreased expression of CD200 and CD200 receptor in Alzheimer's disease: a potential mechanism leading to chronic inflammation. *Experimental neurology*, 215(1), 5-19.
- Walton, M. R., Gibbons, H., MacGibbon, G. A., Sirimanne, E., Saura, J., Gluckman, P. D., & Dragunow, M. (2000). PU. 1 expression in microglia. *Journal of neuroimmunology*, 104(2), 109-115.
- Wang, H., Brown, J., Garcia, C. A., Tang, Y., Benakanakere, M. R., Greenway, T., . . . Martin, M. (2011). The Role of Glycogen Synthase Kinase 3 in Regulating IFN- β -Mediated IL-10 Production. *The Journal of Immunology*, 186(2), 675-684.

- Wang, H., Garcia, C. A., Rehani, K., Cekic, C., Alard, P., Kinane, D. F., . . . Martin, M. (2008). IFN- β production by TLR4-stimulated innate immune cells is negatively regulated by GSK-3- β . *The Journal of Immunology*, 181(10), 6797-6802.
- Wang, M.-J., Huang, H.-Y., Chen, W.-F., Chang, H.-F., & Kuo, J.-S. (2010). Glycogen synthase kinase-3 β inactivation inhibits tumor necrosis factor- α production in microglia by modulating nuclear factor κ B and MLK3/JNK signaling cascades. *J Neuroinflammation*, 7, 99-116.
- Wang, Y.-P., Wu, Y., Li, L.-Y., Zheng, J., Liu, R.-G., Zhou, J.-P., . . . Yao, S.-L. (2011). Aspirin-triggered lipoxin A4 attenuates LPS-induced pro-inflammatory responses by inhibiting activation of NF- κ B and MAPKs in BV-2 microglial cells. *J Neuroinflammation*, 8, 95.
- Woodgett, J. R. (1990). Molecular cloning and expression of glycogen synthase kinase-3/factor A. *The EMBO journal*, 9(8), 2431.
- Wright, G., Jones, M., Puklavec, M., Brown, M., & Barclay, A. (2001). The unusual distribution of the neuronal/lymphoid cell surface CD200 (OX2) glycoprotein is conserved in humans. *Immunology*, 102(2), 173-179.
- Wright, G. J., Cherwinski, H., Foster-Cuevas, M., Brooke, G., Puklavec, M. J., Bigler, M., . . . McClanahan, T. (2003). Characterization of the CD200 receptor family in mice and humans and their interactions with CD200. *The Journal of Immunology*, 171(6), 3034-3046.
- Wright, G. J., Puklavec, M. J., Willis, A. C., Hoek, R. M., Sedgwick, J. D., Brown, M. H., & Barclay, A. N. (2000). Lymphoid/neuronal cell surface OX2 glycoprotein recognizes a novel receptor on macrophages implicated in the control of their function. *Immunity*, 13(2), 233-242.
- Wu, K., Bi, Y., Sun, K., & Wang, C. (2007). IL-10-producing type 1 regulatory T cells and allergy. *Cell Mol Immunol*, 4(4), 269-275.
- Ye, S., Tan, L., Yang, R., Fang, B., Qu, S., Schulze, E. N., . . . Li, P. (2012). Pleiotropy of glycogen synthase kinase-3 inhibition by CHIR99021 promotes self-renewal of embryonic stem cells from refractory mouse strains. *PloS one*, 7(4), e35892.
- Yoshimura, T., Robinson, E., Tanaka, S., Appella, E., & Leonard, E. (1989). Purification and amino acid analysis of two human monocyte chemoattractants produced by phytohemagglutinin-stimulated human blood mononuclear leukocytes. *The Journal of Immunology*, 142(6), 1956-1962.
- Yoshimura, T., Robinson, E. A., Tanaka, S., Appella, E., Kuratsu, J.-I., & Leonard, E. J. (1989). Purification and amino acid analysis of two human glioma-derived monocyte chemoattractants. *The Journal of experimental medicine*, 169(4), 1449-1459.
- Yun, H.-Y., Dawson, V. L., & Dawson, T. M. (1996). Neurobiology of nitric oxide. *Critical Reviews™ in Neurobiology*, 10(3-4).
- Yuskaitis, C. J., & Jope, R. S. (2009). Glycogen synthase kinase-3 regulates microglial migration, inflammation, and inflammation-induced neurotoxicity. *Cellular signalling*, 21(2), 264-273.
- Zhang, S., Cherwinski, H., Sedgwick, J. D., & Phillips, J. H. (2004). Molecular mechanisms of CD200 inhibition of mast cell activation. *The Journal of Immunology*, 173(11), 6786-6793.

- Zhang, S., Wang, X.-J., Tian, L.-P., Pan, J., Lu, G.-Q., Zhang, Y.-J., . . . Chen, S.-D. (2011). CD200-CD200R dysfunction exacerbates microglial activation and dopaminergic neurodegeneration in a rat model of Parkinson's disease. *J Neuroinflammation*, 8(6), 154.
- Zhou, X., Gao, X.-P., Fan, J., Liu, Q., Anwar, K. N., Frey, R. S., & Malik, A. B. (2005). LPS activation of Toll-like receptor 4 signals CD11b/CD18 expression in neutrophils. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 288(4), L655-L662.
- Zhou, X., Zhou, J., Li, X., Guo, C. a., Fang, T., & Chen, Z. (2011). GSK-3 β inhibitors suppressed neuroinflammation in rat cortex by activating autophagy in ischemic brain injury. *Biochemical and biophysical research communications*, 411(2), 271-275.





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BIODATA OF STUDENT

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LIST OF PUBLICATIONS

Poster

Zuhaida Md Zain, Sharmili Vidyadaran and Masriana Hassan. Production of IL-10 in BV-2 cell lines reduces neuroinflammation via inhibition of GSK-3. Faculty Excellent Month (FEM) 2015. Universiti Putra Malaysia, Malaysia. 2015. (Poster presentation)

Full paper

Zuhaida Md Zain, Sharmili Vidyadaran and Masriana Hassan. Inhibition of GSK-3 in microglia reduces inflammation and upregulates IL-10 production.



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