

UNIVERSITI PUTRA MALAYSIA

COCOA-SPECIFIC AROMA POTENTIAL OF SELECTED SEEDS GLOBULIN BY IN VITRO PROTEOLYSIS WITH COCOA PROTEASES

RASHIDAH BINTI SUKOR.

FSMB 2004 5



COCOA-SPECIFIC AROMA POTENTIAL OF SELECTED SEEDS GLOBULIN BY *IN VITRO* PROTEOLYSIS WITH COCOA PROTEASES

by

RASHIDAH BINTI SUKOR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

April 2004



Specially dedicated to my beloved husband and soulmate,

Ja'afar As-Sidek bin Mohd. Sidek

for the unconditional love, patience, support and understanding.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Masters of Science

COCOA-SPECIFIC AROMA POTENTIAL OF SELECTED SEEDS GLOBULIN BY IN VITRO PROTEOLYSIS WITH COCOA PROTEASES

By

RASHIDAH BINTI SUKOR

April 2004

Chairman: Professor Jinap Selamat, Ph.D.

Faculty: Food Science and Biotechnology

This study was carried out to study the globulin characteristics from cottonseed, alfalfa seed, pea, mung bean and French bean and compare their characteristics to cocoa seeds. The study also examined whether these globulins from selected seeds are capable in producing cocoa-specific aroma precursors through proteolysis with cocoa proteases. The isolated globulins were characterized for molecular weight by SDS PAGE, amino acid and oligopeptide profile by High Pressure Liquid Chromatography (HPLC). The globulins were initially treated with an endoprotease for 16 h at 50°C, pH 5.2 and subsequently with a carboxypeptidase for another 16 h at 45°C, pH 5.8; both crude enzymes were extracted from cocoa acetone dry powder (AcDP). Proteolysis products were roasted at 120°C for 20 min with reducing sugars and deodorized cocoa butter. Sensory evaluation session was conducted to detect distinctive cocoa aroma in the proteolysis products. The aroma compounds were extracted through Steam Distillation Extraction (SDE) and analysed using Gas Chromatography (GC).



Alfalfa seed gave the highest total protein of 0.28 mg/mg followed by cottonseed, mung bean, pea, French bean and cocoa with 0.26, 0.25, 0.24, 0.19 and 0.12 mg/mg, respectively. A very low globulin yield was obtained from different seeds, between 0.55% and 2.72%. The seeds had high percentage of crude protein between 13.79 and 26.63%. Two distinctive bands of 51.1 and 33.0 kDa were observed for cocoa vicilin-class globulin (VCG) from SDS PAGE. More than three bands were shown for other seed globulins. Comparative HPLC analyses of the obtained peptide mixtures revealed different and complex patterns of predominantly hydrophobic peptides. After proteolysis, the peptide patterns showed reduced number of peaks, which indicated that peptides have transformed to be more hydrophilic. Considerable differences were observed between the patterns of free amino acids; preferential liberations of hydrophobic amino acids of Ala, Leu, Phe, Val and Tyr were observed in all seeds globulins after proteolysis. No cocoa-specific aroma was detected from the proteolysis products of seeds globulin from cotton, alfalfa, pea, mung bean and French bean. However, pyrazines concentration varied in the proteolysis products derived from those seeds globulin.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

POTENSI PENGHASILAN AROMA SPESIFIK KOKO OLEH GLOBULIN DARIPADA BIJI BENIH TERPILIH MELALUI PROTEOLISIS *IN VITRO* DENGAN ENZIM PROTEASE KOKO

Oleh

RASHIDAH BINTI SUKOR

April 2004

Pengerusi: Profesor Jinap Selamat, Ph.D.

Fakulti: Sains Makanan dan Bioteknologi

Kajian ini dijalankan untuk mengasingkan globulin yang dihasilkan daripada biji kapas, biji alfalfa, kacang pis, kacang hijau dan biji kacang buncis serta membandingkan ciri-ciri globulin tersebut dengan biji koko dan mengkaji sama ada protin tersebut mampu menghasilkan pelopor aroma spesifik koko. Pencirian globulin yang telah diasingkan dijalankan untuk penentuan berat molekul menggunakan SDS PAGE, profil amino acid dan oligopeptida melalui Kromatografi Cecair Tekanan Tinggi. Globulin-globulin tersebut didegradasi oleh enzim endoprotease selama 16 jam pada suhu 50°C, pH 5.2 kemudiannya oleh enzim carboxypeptidase selama 16 jam lagi pada suhu 45°C, pH 5.8; kedua-dua enzim diekstrak daripada AcDP biji koko. Hasil proteolisis dipanggang pada suhu 120°C selama 20 min dengan gula penurun dan mentega koko yang telah dinyahbau. Latihan sensori dijalankan untuk mengesan kehadiran aroma spesifik koko. Sebatian aroma kemudian diekstrak dengan kaedah Penyulingan Stim dan dianalisis menerusi Kromatografi Gas.



Biji alfalfa memberikan nilai protin keseluruhan yang tertinggi iaitu 0.28 mg/mg, diikuti oleh biji kapas, kacang hijau, kacang pis, biji kacang buncis dan AcDP koko dengan 0.26, 0.25, 0.24, 0.19 dan 0.12 mg/mg masingmasing. Hasil yang amat rendah diperolehi semasa ekstraksi globulin daripada biji-biji tersebut iaitu di antara 0.55% hingga 2.72%. Biji-biji yang dipilih juga memberikan peratus protin kasar yang tinggi di antara 13.79 dan 26.63%. Terdapat dua jalur polipeptida yang jelas pada SDS PAGE untuk globulin kelas vicilin (VCG) biji koko dengan berat molekul 51.1 dan 33.0 kDa. Terdapat lebih daripada tiga jalur pada globulin daripada biji-biji yang lain. Analisis oligopeptida menunjukkan corak yang kompleks dan berbeza yang kebanyakannya hidrofobik. Pengurangan puncak dalam corak oligopeptida diperhatikan selepas proteolisis yang kebanyakannya ditukarkan kepada peptida hidrofilik. Perbezaan yang ketara dapat dilihat pada corak dan nilai asid amino bebas bagi biji-biji yang dibandingkan sebelum dan selepas proteolisis. Pembebasan asid amino bersifat hidrofobik iaitu Ala, Leu, Phe, Val dan Tyr dapat diperhatikan di semua globulin biji benih selepas proteolisis. Tiada aroma spesifik koko dikesan apabila hasil proteolisis globulin dari biji kapas, biji alfalfa, kacang pis, kacang hijau dan biji kacang buncis. Walau bagaimanapun, keputusan penentuan pirazin daripada hasil proteolisis biji benih tersebut didapati berbeza-beza di antara satu sama lain.



ACKNOWLEDGEMENTS

In the name of Allah, Most Gracious Most Merciful and *salawat* and *salam* to His righteous messenger, Prophet Muhammad SAW.

The author would like to express her deepest gratitude to her supervisor, Prof. Jinap Selamat for her invaluable assistance, support and guidance throughout the project. Heartfelt appreciation and gratitude to the members of the supervisory committee, Prof. Dr. Jamilah Bakar and Assoc. Prof. Dr. Nazamid Saari for their advice, guidance, criticism and willingness to share their expertise in preparing the thesis.

Many thanks to supporting staff of the Food Science Department for their help, in particular En. Azman, En. Chan and En. Halim for sharing their knowledge and guidance.

Special thanks also to all her graduate friends, especially cocoa group members; Pak Misnawi, Pak Yusep, Bakti, Chin, Nisha, Pak Asep, Tan and Dr. Amin for sharing the literature and invaluable assistance. And also to her special assistant, Sumaiyah who helped her lighten up the burden of the research works. Not forgetting to her housemates who have always been there and never failed to show their endless moral support. The time spent and fondest memories will always be cherished.



The author would also like to express gratitude to the Ministry of Science, Technology and Environment of Malaysia and Faculty of Food Science and Biotechnology for providing the financial funds and laboratory facilities, respectively without which the research work would not had been possible.

Last and by no means, the author wishes to express her love and gratitude to her beloved families; her husband and both her family and in-laws for their endless prayers, understanding, passionate love, encouragement and support throughout her studies. Deepest gratitude and fondest love for her late father, who taught her the meaning and values of life and create the person she became today.



TABLE OF CONTENTS

DEDICATION	2
ABSTRACT	3
ABSTRAK	5
ACKNOWLEDGEMENTS	7
APPROVAL	9
DECLARATION	11
LIST OF TABLES	15
LIST OF FIGURES	16
LIST OF ABBREVIATIONS	17

CHAPTERS

1	GENERAL INTRODUCTION	18
	Statement of Problem	21
	Objectives of the Study	22
2	LITERATURE REVIEW	23
	Storage Proteins of Legume Seeds	23
	Characteristic and Distribution of Vicilin and Legumin in Seeds	25
	Vicilin-type Globulin	25
	Legumin-type Globulin	28
	Other Storage Protein Globulin	29
	Cottonseed (Gossypium hirsitum) Globulin	30
	French bean (<i>Phaseolus vulgari</i> s) Globulin	32
	Alfalfa seed (<i>Medicago sativa</i>) Globulin	34
	Pea seed (<i>Pisum sativum</i>) Globulin	37
	Mung bean (<i>Phaselus aureus</i>) Globulin	42
	Storage Protein of Cocoa Beans	44
	Vicilin-Class (7S) Globulin of Cocoa and Its Biochemical	
	Properties	45
	The Role of VCG in the Formation of Cocoa-Specific	
	Aroma Precursors	48
	The Comparison of Storage Protein from Cocoa and Other	
	Seeds in the Ability to Produce Cocoa-Specific Aroma	50
	Cocoa Flavour Precursors	53
	Proteins and Oligopeptides	53
	Free Amino Acids	58
	The Importance of Protease in the Production of Cocoa-Specific	~ ~
	Aroma Precursors	61
	Optimum Conditions for In Vitro Proteolysis of Storage Protein in	~~
	Cocoa by Cocoa Proteases	62
	The Importance of Roasting to Aroma Formation	65
	Pyrazines and Cocoa Aroma Precursors	68
	Significance of Sensory Evaluation in Flavour Evaluation	72



3	GENERAL MATERIALS AND METHODS	75
	Materials	75
	Preparation of Samples	79
	Preparation of Acetone Dry Powder (AcDP)	80
	Isolation of Globular Storage Protein from Cocoa Seeds	81
	Isolation of Globular Storage Protein from Cotton, Alfalfa, Pea,	
	Mung Bean and French Bean Seeds	81
	Determination of Protein	83
	Statistical Analyses	83

ISOLATION AND COMPARISON OF GLOBULIN	
CHARACTERISTICS FROM COCOA AND SELECTED SEEDS	84
Introduction	84
Materials and Methods	86
Crude Protein Analyses	86
SDS-PAGE with Coomasie Blue and Silver Staining	87
Densitograms of Polypeptide Bands from SDS PAGE	88
HPLC Analyses of Peptides	88
Determination of Free Amino Acids by HPLC	89
Results and Discussion	91
Extraction of Vicilin-Class Globulin from Cocoa Seeds	91
Extraction of Globulins from Cotton, Alfalfa, Pea,	
Mung Bean and French Bean Seeds	95
Total Protein of Different Seed Samples	99
Polypeptide Bands of Cocoa and Selected Seeds	
Globulin on SDS PAGE	101
Oligopeptide Patterns of Cocoa and Selected Seeds	
Globulin	113
Free Amino Acids Composition of Cocoa and Selected	
Seeds Globulin	116
Summary	119

5	PROTEOLYSIS OF SEEDS GLOBULIN WITH COCOA ENDOPROTEASE AND CARBOCYPEPTIDASE IN PROD	UCING
	COCOA SPECIFIC-AROMA PRECURSORS	120
	Introduction	120
	Materials and Methods	122
	Preparation of Crude Enzyme Extract	122
	Enzyme Assays	123
	Aspartic Endoprotease Activity	123
	Carboxypeptidase Activity	124
	Proteolysis Digestion of Globulin with Endoprotease	
	and Carboxypeptidase from Cocoa Seeds	124
	HPLC Analyses of Peptides	125
	Determination of Free Amino Acids	126



Roasting of Globulin with Reducing Sugars and	
Deodorized Cocoa Butter	128
Screening of Sensory Panellists	128
Sensory Evaluation Test	130
Pyrazine Extraction through Steam Distillation	
Extraction	131
Pyrazine Determination using Gas Chromatography	131
Results and Discussion	132
Oligopeptide Patterns of Proteolysis Products from	
Cocoa and Selected Seeds Globulin	132
Free Amino Acids of Seeds Globulin After Proteolysis	
with Cocoa Endoprotease and Carboxypeptidase	139
Screening of Sensory Panellists	143
Sensory Evaluation Test on Ability of Cocoa-Specific	
Aroma Detection	146
Pyrazines of Proteolysis Products from Seeds Globulin	
by SDE and GC	149
Summary	154
CONCLUSIONS AND RECOMMENDATION	156
General Conclusions	156
Recommendations	158
BLIOGRAPHY	159

BIBLIOGRAPHY	159
APPENDICES	176
BIODATA OF THE AUTHOR	189



LIST OF TABLES

Table		Page
2.1	The distribution of the characteristic subunits of 7S globulin of legumes	27
2.2	Subunit composition of medicagin in <i>Medicago sativa cv. Ecalibur</i> from fraction S-2	36
2.3	Composition of pea seed	38
2.4	Amino acid composition in pea seed (mg/g N)	39
2.5	The ability of production of aroma precursors and cocoa aroma	52
2.6	Composition of amino acids in acidic hydrolysates of unfermented and fermented cocoa beans	59
4.1(a)	HPLC elution program for determination of oligopeptide patterns	177
4.1(b)	HPLC gradient elution for determination of free amino acid	178
4.2	Result of AcDP extraction from cocoa seeds	183
4.3	Result of globulin extraction from different seed samples	184
4.4	Percent yield of globulin, total protein and crude protein percent of cocoa, cotton, alfalfa, pea, mung bean and French bean seeds	98
4.5	The calculated molecular weight of globulins subunit from selected seeds analysed by SDS PAGE stained with Coomasie blue	105
4.6	Free amino acid composition from different seeds globulin	117
5.1	Free amino acids present in the proteolysis products generated <i>in vitro</i> by degradation of globulin from different seed samples with aspartic endoprotease and carboxypeptidase from cocoa seeds	142
5.2	Result of screening for panellists in Basic Odour Recognition Test and Triangle Test	145
5.3	Cocoa-specific aroma potentials of proteolysis products generated <i>in vitro</i> from seeds globulin fractions from seeds of cocoa, cotton, alfalfa, pea, mung bean, and French bean, by the cooperative action of aspartic endoprotease and carboxypeptidase from cocoa seeds	148
5.4	Pyrazine concentration of proteolysis products of seeds globulin	

5.4 Pyrazine concentration of proteolysis products of seeds globulin with cocoa endoprotease and carboxypeptidase upon roasting with reducing sugars and deodorized cocoa butter

LIST OF FIGURES

Figure		Page
2.1	Formation of hydrophobic oligopeptides	56
3.1	Cocoa pod and beans clone PBC 140	76
3.2	Alfalfa seed	76
3.3	Cottonseed	77
3.4	French bean seed	77
3.5	Mung bean	78
3.6	Pea	78
4.1	The linear curve of standard protein markers of SDS PAGE stained with Coomasie Blue staining	185
4.3	SDS PAGE patterns of cocoa and other seeds globulin stained with (a) Coomasie blue and (b) silver nitrate	104
4.4	Densitometer tracing of gel pattern of molecular weight marker standard	111
4.5	Densitometer tracing of gel pattern of seeds globulin	112
4.6	Chromatographic profile of peptide mixtures from seeds globulin of (a)cocoa, (b)cotton, (c)alfalfa, (d)pea, (e)mung bean and (f) French bean, respectively	115
5.1	Chromatographic profile of peptide mixtures obtained by proteolysis of seeds globulin with cocoa endoprotease and carboxypeptidase	136
5.2	Schematic diagram of Steam Distillation Extraction (SDE)	179



LIST OF ABBREVIATIONS

AcDP Acetone dry powder b.p. Boiling point DW Dry weight GC Gas Chromatography HPLC High Pressure Liquid Chromatography kDa Kilo dalton MBP Maltose binding protein MW Molecular weight PBC Perang Besar Clone PMSF Phenylmethyl sulfonyl fluoride **PVPP** Polivinyl pyrolidone Revolution per minute rpm S Sedimentation co-efficient SD Standard deviation SDS PAGE Sodium dodecyl sulphate polyacrylamide gel electrophoresis TFA Trifluoroacetic acid VCG Vicilin-class globulin w/w Weight/weight w/v Weight/volume v/v Volume/volume



CHAPTER 1

GENERAL INTRODUCTION

Cocoa is an extremely valuable crop, providing the food industry with chocolate, cocoa and cocoa butter, which is used in a variety of food applicants. Much attention has been focused on the unique properties of cocoa butter which comprises about 50% of the beans dry weight, but relatively little attempt has been made to study the bean protein, although these are the second major constituents at 15-20% dry weight.

Malaysia Cocoa Board (1998) reported a drop of about 7% cocoa production level from last year and lower production forecast. Cocoa Malaysia is also facing the competition from other cocoa production countries e.g. Ghana due to their high quality flavour. The substitution of cocoa aroma from other sources is therefore necessary. The globulin from legume seeds may be added to enhance cocoa flavour if their globulin can produce cocoa aroma precursors through proteolysis.

Cocoa fruit is the widely used tropical crop for the manufacture of chocolate and other confectionary products. Cocoa bean will provide a unique aroma after undergoing proper pulp conditioning, sequential fermentation, drying and roasting processes. Fermentation steps produces flavour precursors when



enzymatic reaction occurs. Thus, the flavour and aroma typical of chocolate will be produced during roasting in the manufacture of chocolate.

The detection of the reaction of amino acids and reducing sugars (Rohan, 1963; 1964) and the contribution of the peptides in Maillard reactions (Mohr *et al.*, 1976) are important findings in researches related to the formation of cocoa aroma. These precursors are important in the formation of cocoa aroma during roasting. Biehl *et al.* (1985) has shown that vacuolar storage protein is a source of specific-cocoa aroma precursors which are released from vicilin (7S)-class globulin (VCG) by the action of cocoa proteases (Biehl *et al.*, 1991; Voigt *et al.*, 1994a).

The protein of a variety of plant is degraded on germination, and serves as the source of nitrogen for the various new nitrogen compounds synthesized by the developing seedling and is called storage protein (Derbyshire *et al.*, 1976). On the basis of their acid amino sequences, subunit compositions, and the processing of the corresponding polypeptide precursors, the globular storage proteins of plant studied so far can be assigned into two different classes: the legumin-like (11-12S) globulins and the vicilin-like (7S) globulins (Derbyshire *et al.*, 1976; Higgins, 1984; Muntz *et al.*, 1985; Borroto and Dure, 1987; Shotwell and Larkins, 1988).



Cocoa seeds contain 2 major storage proteins: a 19 kDa albumin which is related to the soy bean trypsin inhibitor (Kunitz,1947), family of protease inhibitor (Spencer and Hodge, 1991; Tai *et al.*, 1991) and vicilin-class globulin (Spencer and Hodge, 1991, 1992; Tai *et al.*, 1991; McHenry and Fritz, 1992; Voigt *et al.*, 1993b). Cocoa-specific aroma precursors were obtained when isolated vicilin-class globulin was successively degraded by the aspartic endoprotease and carboxypeptidase partially purified from ungerminated cocoa seeds. No typical aroma precursors were obtained when the albumin fraction was subjected to proteolysis by aspartic endoprotease and carboxypeptidase from cocoa seeds. Therefore, it has been clearly shown that the cocoa-specific aroma precursor are only derived from vicilin-class globulin of the globular storage protein from cocoa seeds (Biehl *et al.*, 1982a; Voigt *et al.*, 1993a).

The acid amino sequences of the globulins from the coconut (vicilin-class globulin), hazelnuts (legumin class 11-12S), and sunflower seeds (legumin class 11-12S) are rather different with the particular chemical structure of the globulin present in the cocoa seeds, cocoa-specific aroma precursors. Therefore, there was no cocoa or chocolate aroma was generated upon roasting of the proteolysis products (Voigt *et al.*, 1994a).



Statement of Problem

Cocoa aroma is one of the most important characteristics of cocoa flavour. Biehl and Voigt (1994; 1996), indicated that cocoa-specific aroma is produced during roasting of the specific hydrophilic oligopeptides and hydrophobic free amino acids. The specificity depends on the primary structure of VCG of cocoa cotyledons and the splitting types of cocoa cotyledon aspartic endoprotease and carboxypeptidase enzymes. Additionally, the production of these specific components of precursors strictly depends on the cotyledon-pH during fermentation which is controlled by microbial pulp degradation. Thus, the specificity of cocoa aroma depends on the primary structure of cocoa cotyledon VCG, the splitting specificity of cocoa cotyledons proteases and environmental factors.

Neither cocoa albumin nor 7S globulin from hazelnut, sunflower and coconut so far investigated are suitable for producing specific-cocoa aroma precursors by proteolysis (Voigt *et al.*, 1994d). The amino acid sequence of the 65 kDa precursor for the cocoa globulin deduced from the nucleotide sequence of the cloned cDNA revealed considerable sequence homologies with vicilin-class globulin of some dicotyledonous plants, especially with an α -globulin of cotton seeds (McHenry & Fritz, 1992; Spencer & Hodge, 1992). Therefore, this study is being carried out to investigate whether or not cocoa-specific aroma precursors can be derived from globular storage proteins from selected plant



seeds which contains 7S globulin by successive degradation with the aspartic endoprotease and the carboxypeptidase from unfermented cocoa seeds. Seeds of some plants which contain exclusively of 7S globulin have been selected to undergo the proteolysis process.

Objectives of the Study

The objectives of this study were as follows:

- 1. To isolate and compare the protein characteristics of globulins from cocoa seed to cotton, alfalfa, pea, mung bean and French bean seeds.
- 2. To study the properties of proteolysis product of cocoa and other seeds globulin after incubation with cocoa endoprotease and carboxypeptidase.



CHAPTER 2

LITERATURE REVIEW

Storage Protein of Legume Seeds

Storage protein of seed plans are utilized as sources of nitrogen, carbon and sulphur, and are accumulated during germination and ripening stages. These elements are synthesized in large quantities during bean maturation in rough endoplasmic reticulum and deposited in the membrane-bound vacuoles, called protein bodies (Baunmgartner & Chrispeel, 1977; Pernollet, 1978; Stegemann, 1975; Graham & Gunning, 1970; Varner & Schidlovsky, 1963). The protein bodies are metabolized again during germination. On average, the percentage of protein in legume seed is 20-25% of dry matter (Derbyshire *et al.*, 1975). Seeds, particularly legume seeds are a high protein food source for man and animals either directly and more recently, as 'textured vegetable protein foods' for man. The functional and chemical properties of seed protein are complex (William *et al.*, 1983)

Derbyshire (1975) reported that many proteins occur in seeds and the problem is to distinguish the storage protein from those, which have metabolic or structural role. Two of the major classes of storage proteins in legume seed are termed vicilin-type-globulin (7S) and legumin-type-globulin (11S) based on their



sedimentation coefficients (Hilda and Octavio, 1995). The widespread occurrence of 11S and 7S type storage globulin in angiosperm seeds has been recognized and accepted (Wolf, 1980).

During their development, plant seeds accumulate large amounts of storage proteins that serve as sources of nitrogen, sulphur and carbon compounds during seed germination (Shotwell & Larkins, 1988). Storage proteins accumulate in membrane-delimited organelles, the protein bodies (Briarty et al., 1970; Pernollet, 1978). Seed proteins can be classified on an operational basis into four solubility classes; albumin (water-soluble), globulin (salt-soluble), prolamin (alcohol-soluble) and glutenin (soluble in dilute acids or alkali) (Osborne, 1924). Plant seeds of different taxa contain rather different proportions of albumin, globulins, prolamins and glutenins (Higgins, 1984; Bewley & Black, 1985; Shotwell & Larkins, 1988). In most monocotyledonous seeds studied so far, prolamins are the major seeds proteins (Bewley & Black, 1985) and in gymnosperms (Misra & Green, 1990) globulins predominate. On the basis of their acid amino sequences, subunit composition, and the processing of the corresponding polypeptide precursors, the globulins studied so far can be assigned to two different classes: the legumin-like and the vicilinlike globulins (Derbyshire et al., 1976; Higgins, 1984; Shotwell & Larkins, 1988).

