



BIOLOGICAL ACTIVITIES AND TOXICITY EFFECTS OF *Clinacanthus nutans* (Burm.f.) Lindau LEAF EXTRACTS

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By

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Clinacanthus nutans (*C. nutans*) which also known as Sabah snake grass or Pokok Belalai gajah has long been used as traditional medicines in Thailand, China and Indonesia to treat various types of diseases. Recently, *C. nutans* has become a popular in Malaysia as a folk medicine to treat cancer and kidney failure. Side effects arise from the used of synthetic drugs for the treatments of chronic diseases have brought about the urge to search for alternative medicine derived from natural products. Thus, the objective of this study was to determine the biological activities of the *C. nutans* leaf extracts. Leaves of *C. nutans* were extracted sequentially using hexane followed by dichloromethane, ethyl acetate, acetone and lastly distilled water. Antioxidant activity test showed that dichloromethane extract demonstrated highest free radical scavenging ability (IC₅₀ value 106.41 ± 1.32 µg/ml). For the FRAP assay, acetone extracts exhibited highest activity with 193.00 ± 2.63 mg TE/g extract. The total phenolics contents, the acetone extract exhibited the highest value, 66.67 ± 13.85 mg GAEs/mg extract. While the dichloromethane extract gave the highest total flavonoid contents with the value of 851.82 ± 78.06 mg QEs/mg extract. The leaf extracts have shown no inhibition on the tyrosinase activity. Anti-inflammatory activity was determined using Griess assay on nitric oxide inhibitory activity upon IFN-γ/LPS stimulated RAW 264.7 cell line showed various extracts of *C. nutans* have high anti-inflammatory effects with the IC₅₀ value < 20 µg/ml except distilled water extract. Further study of *in vitro* toxicity test using five cultured cancer cell lines and MTT assay showed *C. nutans* leaf extracts exhibited selective inhibition towards the cultured cancer cell lines. Dichloromethane extract exhibited highest anti-proliferative effects on murine melanoma (B16/F10) cell line with LC₅₀ value 48.08 µg/ml, human breast cancer (MCF-7) cell line, LC₅₀ value 97.46 µg/ml, human colorectal carcinoma (HT-29) cell line, LC₅₀ value of 98.06 µg/ml, human hepatocellular carcinoma cell line (HepG2), LC₅₀ value of 99.59 µg/ml but weakly inhibited human neuroblastoma (SY-SY5Y) cell line, LC₅₀ value 156.34 µg/ml but was not toxic towards human normal Chang liver cell line with LC₅₀ value of > 250 µg/ml. For *in vivo* toxicity study, the various leaf extracts tested on zebrafish embryos showed the acetone extract exhibited highest toxicity towards the embryos with LC₅₀ value of 88.33 ± 0.58 µg/ml followed by dichloromethane extract LC₅₀ value of 141.73 ± 3.46 µg/ml. At the higher concentration (>125 µg/ml), the embryos and larvae exhibited teratogenic effect such as coiled body, bended tail and/or tail tip, pericardial and/or yolk sac oedemas. Collectively, the study showed that the *C. nutans* leaf extracts possessed high antioxidant and anti-inflammatory activities as well as anti-proliferative properties

against cultured cancer cell lines. This suggests the preventive and therapeutic potential of *C. nutans* leaf to be used as alternative medicines for various types of diseases including cancer in the future.



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**AKTIVITI BIOLOGI DAN KESAN TOKSIKOSITI EKSTRAK DAUN
Clinacanthus nutans (Burm.f.) Lindau**

Oleh

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February 2015

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Clinacanthus nutans (*C. nutans*) yang juga dikenali sebagai 'Sabah snake grass' atau Pokok Belalai gajah telah lama digunakan sebagai ubat tradisional di Negara Thai China and Indonesia untuk mengubati pelbagai penyakit. Kebelakangan ini, *C. nutans* telah menjadi semakin popular di kalangan penduduk Malaysia sebagai ubat tradisional yang digunakan untuk mengubati penyakit kanser dan kerosakan ginjal. Kesan sampingan negatif yang timbul akibat dari penggunaan ubat-ubatan sintetik untuk mengubati penyakit kronik telah mendorong kepada usaha untuk mencari ubat-ubatan tradisional sebagai rawatan alternatif yang didapati dari tumbuhan semulajadi dan dikatakan tiada kesan sampingan. Tujuan penyelidikan ini dijalankan adalah untuk menentukan aktiviti biologi ekstrak daun *C. nutans*. Daun tumbuhan ini dieskrak secara berturutan menggunakan pelarut seperti heksana, diikuti dengan dikloromethana, etil asetat, aseton dan air suling. Ujian antioksidan menunjukkan ekstrak dikloromethana mempunyai keupayaan memerangkap radikal bebas yang paling tinggi (IC_{50} 106.41 ± 1.32 $\mu\text{g/ml}$). Manakala, bagi ujian keupayaan menurun, ekstrak aseton pula menunjukkan keupayaan menurun yang tinggi dengan nilai 193.00 ± 2.63 mg TE/g ekstrak. Jumlah kandungan fenolik dan flavonoid dalam ekstrak menunjukkan ekstrak aseton mempunyai kandungan yang paling tinggi iaitu 66.67 ± 13.85 mg GAE/mg ekstrak. Manakala, ekstrak dikloromethana menunjukkan kandungan flavonoid yang paling tinggi iaitu 851.82 ± 78.06 mg QEs/mg ekstrak. Tiada ekstrak daun *C. nutans* yang menunjukkan keupayaan merencat aktiviti enzim tyrosinase. Kesan anti radang menggunakan ujian Griess terhadap aktiviti perencatan nitrik oksida (NO) pada turunan sel RAW 264.7 yang diaktifkan oleh IFN- γ /LPS menunjukkan ekstrak daun *C. nutans* mempunyai kesan perencatan nitrik oksida yang tinggi dengan nilai $IC_{50} < 20$ $\mu\text{g/ml}$ kecuali ekstrak air suling ($IC_{50} > 500$ $\mu\text{g/ml}$). Penyelidikan dilanjutkan dengan ujian toksisiti *in vitro* ke atas lima jenis turunan sel kanser berkultur menggunakan ujian MTT. Keputusan ujian ini menunjukkan ekstrak daun *C. nutans* menunjukkan kesan anti-proliferatif yang memilih. Ekstrak dikloromethana menunjukkan kesan anti-proliferatif yang tinggi terhadap turunan sel melanoma murin (B16/F10), nilai LC_{50} 48.08 $\mu\text{g/ml}$, turunan sel kanser payu dara manusia (MCF-7), nilai LC_{50} 97.46 $\mu\text{g/ml}$, turunan sel kanser kolorektal manusia (HT-29), nilai LC_{50} 98.06 $\mu\text{g/ml}$, turunan sel kanser hati (HepG2), nilai LC_{50} 99.59 $\mu\text{g/ml}$ tetapi menunjukkan kesan perencatan yang lemah terhadap turunan sel neuroblastoma manusia (SY-SY5Y), nilai LC_{50} 156.34 $\mu\text{g/ml}$. Ujian toksisiti dilanjutkan dengan ujian

toksisiti *in vivo* yang menggunakan embrio zebrafish mendapati bahawa ekstrak aseton menunjukkan toksisiti yang paling tinggi dengan nilai LC_{50} $88.33 \pm 0.58 \mu\text{g/ml}$ diikuti ekstrak dikloromethana, nilai LC_{50} $141.73 \pm 3.46 \mu\text{g/ml}$. Selain itu, pada kepekatan yang tinggi ($>125 \mu\text{g/ml}$), embrio dan larva zebrafish menunjukkan kesan teratogenik seperti bentuk badan melengkung, bengkok pada bahagian ekor dan/atau hujung ekor serta edema pericardial dan/atau pundi yolk. Secara keseluruhannya, penyelidikan ini menunjukkan pelbagai ekstrak daun *C. nutans* mempunyai aktiviti antioksidan dan anti radang yang tinggi serta sifat-sifat anti-proliferatif yang selektif terhadap turunan sel kanser berkultur. Ini mencadangkan bahawa daun *C. nutans* mempunyai potensi pencegahan dan terapi untuk digunakan sebagai ubat alternatif untuk pelbagai jenis penyakit termasuk kanser pada masa akan datang.



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LIST OF ABBREVIATIONS

| | |
|----------------------------------|--|
| AAPH | 2,2'-azobis-2-methyl-propanimidamide, dihydrochloride |
| AChE | Anti cholinesterase |
| ALP | Alkaline phosphatase |
| AlCl ₃ | Aluminium chloride |
| ALT | Alanine-aminotransferase |
| AST | Aspartate aminotransferase |
| B16/F10 | Murine melanoma cell |
| BW | Body weight |
| CAT | Catalase |
| CaCl ₂ | Calcium chloride |
| CO ₂ | Carbon dioxide |
| DNA | Deoxyribonucleic acid |
| DPPH | 2,2-diphenyl-1-picrylhydrazyl |
| DMSO | Dimethyl sulfoxide |
| DMEM | Dulbecco's modified Eagle's medium |
| E2 | Embryo medium |
| FBS | Fetal bovine serum |
| Fe ³⁺ | Ferric (III) ion |
| Fe ²⁺ | Ferric (II) ion |
| FRS | Free radical scavengers |
| GSH | Glutathione reductase |
| GPx | Glutathione peroxidase |
| HClO | Hypochlorous acid |
| HepG2 | Human hepatocellular carcinoma cells |
| H ₂ O ₂ | Hydrogen peroxide |
| HIF-1 α | Hypoxia-inducible factor 1-alpha |
| HPLC | High performance liquid chromatography |
| HSV-1 | Herpes simplex virus Type 1 |
| HSV-2 | Herpes simplex virus Type 2 |
| HT-29 | Human colorectal carcinoma cells |
| IC ₅₀ | Inhibitory concentration at 50% |
| IFN- γ | Interferon-gamma |
| KCl | Potassium chloride |
| KH ₂ PO ₄ | Potassium dihydrogen phosphate |
| LC ₅₀ | Lethal concentration at 50% |
| LDL | Low density lipoproteins |
| L-DOPA | L-3,4-dihydroxyphenylalanine |
| LD ₅₀ | Lethal dose at 50% |
| L-NAME | N ω -nitro-L-arginine methyl ester |
| MCF-7 | Human breast cancer cells |
| MgSO ₄ | Magnesium sulphate |
| MPLC | Medium performance liquid chromatography |
| MPO | Myeloperoxidase |
| MTT | 3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide |
| NaCl | Sodium chloride |
| Na ₂ HPO ₄ | Disodium hydrogen phosphate |

| | |
|-----------------------------|------------------------------------|
| NaHCO ₃ | Sodium bicarbonate |
| NF-κB | Nuclear factor kappa-B |
| NO | Nitric oxide |
| O ₂ ⁻ | Superoxide anion |
| OH ⁻ | Hydroxyl ion |
| PBS | Phosphate Buffer Saline |
| RAW 264.7 | Murine monocytic macrophage cells |
| ROS | Reactive oxygen species |
| RNS | Reactive nitrogen species |
| SH-SY5Y | Human neuroblastoma cells |
| SOD | Superoxide dismutase |
| UV | Ultraviolet |
| VEGF | Vascular endothelial growth factor |
| VZV | Varicella-zoster virus |



LIST OF ANNOTATION

| | |
|--------------|--------------------------|
| % | Percent |
| γ | Gamma |
| (v/v) | Volume per volume |
| (w/v) | Weight per volume |
| < | Lesser than |
| > | More than |
| \pm | Plus and/or minus |
| μ | Micro |
| ml | Microlitre |
| μ M | Micromolar |
| g | Gram |
| h | Hour |
| kg | Kilogram |
| L | Litre |
| mg/ml | Milligram per millilitre |
| μ g/ml | Microgram per millilitre |
| U/ml | Unit per millilitre |
| min | Minute |
| mmol | Milimole |
| $^{\circ}$ C | Degree Celsius |
| s | Second |
| α | Alpha |
| β | Beta |

CHAPTER 1

INTRODUCTION

Free radicals such as reactive oxygen species (ROS) and reactive nitrogen species (RNS) which were produced by various activities in the body including metabolism and inflammation are responsible for the development of variety of chronic diseases including cancer. Cancer has been the third most common cause of mortality in Malaysia and has always been the major health problem faced worldwide. Oxidative stress induced by the over production of free radicals have caused the deleterious damage effects of cellular lipid, proteins and DNA. These caused cell dysfunction, death or malignant transformations that involve genomics alteration and mutations which eventually promote carcinogenesis (Yong et al., 2013; Wang et al., 2011). Furthermore, the side effects that occurred during and after treatments of chronic diseases including cancer due to the use of the synthetics drugs has brought about the urge for the discovery of alternative medicine from the plant natural product in order to overcome the problems (Abdelwahab et al., 2011; Cooper, 2005). Antioxidants, the substances that play an important role in inhibiting the oxidation and thus preventing the damage of cells in the body (Chen et al., 2012; Tachakittirungrod et al., 2007) are found abundantly in plants especially fruits and vegetables. Natural antioxidants found in the plants are able to protect body from oxidative stress and associated diseases (Salganik, 2001; Halliwell, 1990).

Epidemiological studies have proven that antioxidant addition intake could retrieve cells from oxidative stress and prevent cancer growth and development. Several studies had revealed that medicinal plants contains more natural antioxidant sources such as phenolic acids, flavonoids and tannins possessed greater potential in antioxidant activities than common dietary plants (Li et al., 2008). Phytochemicals produced by the medicinal plant have been associated to many medicinal benefits to human health. They are safer, cheaper and readily available with fewer side effects compared to the synthetic drugs.

Clinacanthus nutans (Burm.f.) Lindau (*C. nutans*) which also known as Sabah snake grass (English) or Pokok Belalai Gajah (Bahasa Melayu) belongs to Acanthaceae family is a popular cultivated small shrub, native to tropical Asia. *C. nutans* is a well-known traditional medicine in Thailand for the treatment of various health problems such as skin rashes, snake bites (Thongharb et al., and Cherdchu et al., 1977), herpes infections (Jayavasut et al., 1992a), varicella-zoster virus (VZV) lesions (Wanikiat et al., 2008), anti-inflammatory activity (Satyavivad et al., 1996) anti-tumour angiogenesis (Duansak, 2007) and antioxidant activity (Pannangpetch, 2007). On the other hand, the whole plant of *C. nutans* has been used as anti-inflammatory agent for treating bruises, sprain and rheumatism in China. However, in Malaysia *C. nutans* which is a newly known herbs but becoming popular especially in the Chinese community due to its effectiveness in treating cancer (Yong et al., 2013) and kidney failure as a folk medicine.

Although in Thailand thorough studies have been done by the previous researchers on the beneficial effects of *C. nutans* plant, there are limited studies done in Malaysia due to its current popularity. However, previous studies done by Yong et al., (2013) and Sakdarat et al., (2009) reported that *C. nutans* has many chemical and biological activities has initiated the urge to establish more scientific information on its phytochemical benefits. Therefore, this research was carried out to evaluate the biological activities of the various extracts of *C. nutans* leaf.

Thus, the objectives of this study were:

1. to extract the leaf of *C. nutans* sequentially using five different solvents, hexane, dichloromethane, ethyl acetate, acetone and distilled water
2. to evaluate the biological activities of the crude extracts of the *C. nutans* leaf (antioxidant, anti-tyrosinase and anti-inflammatory activities)
3. to determine the toxicity effects of the crude extracts of *C. nutans* leaf on the cultured cancer cell lines using MTT assay (*in vitro*) and zebrafish embryos (*in vivo*)

REFERENCES / BIBLIOGRAPHY

- Abdelwahab, S. I., Mohan, S., Abdulla, M. A., Sukari, M. A., Abdul, A. B., Taha, M. M. E., Lee, K.H. (2011). The methanolic extract of *Boesenbergia rotunda* (L.)Mansf.and its major compound pinostrobin induces anti-ulcerogenic property in vivo: possible involvement of indirect antioxidant action. *Journal of Ethnopharmacology*, 137(2), 963–70.
- Abdelwahab, S. I., Mohan, S., Mohamed Elhassan, M., Al-Mekhlafi, N., Mariod, A. A., Abdul, A. B., Alkharfy, K. M. (2011). Anti-apoptotic and Antioxidant Properties of *Orthosiphon stamineus* Benth (Cat's Whiskers): Intervention in the Bcl-2-Mediated Apoptotic Pathway. *Evidence-Based Complementary and Alternative Medicine: eCAM*, 2011, 156765.
- Acharya, A., Das, I., Chandhok, D., & Saha, T. (2010). Redox regulation in cancer: a double-edged sword with therapeutic potential. *Oxidative Medicine and Cellular Longevity*, 3(1), 23–34.
- Awad, A. B., Downie, A. C., Fink, C. S. (2000). Inhibition of growth and stimulation of apoptosis by beta-sitosterol treatment of MDA-MB-231 human breast cancer cells in culture. *International Journal of Molecular Medicine*. 5, 541–546.
- Adamu, M., Naidoo, V., & Eloff, J. N. (2013). Efficacy and toxicity of thirteen plant leaf acetone extracts used in ethnoveterinary medicine in South Africa on egg hatching and larval development of *Haemonchus contortus*. *BMC Veterinary Research*, 9(1), 38.
- Agudo, A., Cabrera, L., Amiano, P., Ardanaz, E., Barricarte, A., Berenguer, T. and Gonzalez, C. A. (2007). Fruit and vegetable intakes, dietary antioxidant nutrients, and total mortality in Spanish adults: Findings from the Spanish cohort of the European Prospective Investigation into Cancer and Nutrition (EPIC-Spain). *American Journal of Clinical Nutrition* 85: 1634– 1642.
- Ahmad, S., Israif, D. L., Ismail,N. H., Shaari, K., Mohamed, H., Wahab, A. Somchit, M. N., (2006). Cardamonin, inhibits pro-inflammatory mediators in activated RAW 264.7 cells and whole blood. *European Journal of Pharmacology*, 538(1-3), 188–94
- Allothman, M., Bhat, R., & Karim, A. A. (2009). Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents. *Food Chemistry*, 115(3), 785–788.
- Amarowicz, R., Pegg, R. B., Rahimi-Moghaddam, P., Barl, B., & Weil, J. A. (2004).Free-radical scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies.*Food Chemistry*, 84(4), 551–562.
- Anand, P., Kunnumakkara, A. B., Kunnumakara, A. B., Sundaram, C., Harikumar, K. B., Tharakan, S. T., Lai, O. S., Sung, B., Aggarwal, B. B., (2008). "Cancer is a

preventable disease that requires major lifestyle changes". *Pharmacology Research* 25(9), 2097-2116.

Arabshahi-Delouee, S., & Urooj, A. (2007). Antioxidant properties of various solvent extracts of mulberry (*Morusindica* L.) leaves. *Food Chemistry*, 102(4), 1233–1240.

Arenzana, F. J., Carvan, M. J., Aijón, J., Sánchez-González, R., Arévalo, R., & Porteros, A., (2006). Teratogenic effects of ethanol exposure on zebrafish visual system development. *Neurotoxicology and Teratology*, 28(3), 342–8.

Arung, E. T., Matsubara, E., Kusuma, I. W., Sukaton, E., Shimizu, K., Kondo, R. (2011). Inhibitory components from the buds of clove (*Syzygiumaromaticum*) on melanin formation in B16 melanoma cells. *Fitoterapia*, 82, 198–202.

Awad, A. B., Downie, A. C., Fink, C. S. (2000). Inhibition of growth and stimulation of apoptosis by beta-sitosterol treatment of MDA-MB-231 human breast cancer cells in culture. *International Journal of Molecular Medicine*. 5, 541–546.

Azizova, O. A., (2002). Role of free radical processes in the development of atherosclerosis. *Biologic heskie Membrany*, 19, 451–471.

Barnes, S., Kirk, M., Coward, L. (1994). Isoflavones and their conjugates in soy foods: extraction conditions and analysis by HPLC-mass spectrometry. *Journal of Agricultural Food Chemistry*, 42(11), 2466–2474.

Baskar, A. A., Al, N. K. S., Paulraj, M. G., Alsaif, M. A., Al, M. M., et al. (2012). β -Sitosterol prevents lipid peroxidation and improves antioxidant status and histoarchitecture in rats with 1,2-dimethylhydrazine-induced colon cancer. *Journal of Medicinal Food*. 15, 335–43.

Belyaeva, N. F., Kashirtseva, V. N., Medvedeva, N. V., Khudoklinova, Y. Y., Ipatova, O. M., & Archakov, A. I. (2009). Zebrafish as a model system for biomedical studies. *Biochemistry (Moscow) Supplement Series B: Biomedical Chemistry*, 3(4), 343–350.

Berghmans, S., Butler, P., Goldsmith, P., Waldron, G., Gardner, I., Golder, Z., Fleming, A. (2008). Zebrafish based assays for the assessment of cardiac, visual and gut function - potential safety screens for early drug discovery. *Journal of Pharmacological and Toxicological Methods*, 58(1), 59-68.

Berry, J. P., Gantar, M., Gibbs, P. D. L., & Schmale, M. C. (2007). The zebrafish (*Danio rerio*) embryo as a model system for identification and characterization of developmental toxins from marine and freshwater microalgae. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 145(1), 61-72.

- Blot, W. J., Li, J. Y., Taylor, P. R., et al. (1993). Nutrition intervention trials in Lin xian, China: supplementation with specific vitamin/mineral combinations, cancer incidence, and disease-specific mortality in the general population. *Journal of National Cancer Institute*, 85, 1483–91.
- Bouic, P. J., Etsebeth, S., Liebenberg, R. W., Albrecht, C. F., Pegel, K., et al. (1996). beta-Sitosterol and beta-sitosterol glucoside stimulate human peripheral blood lymphocyte proliferation: implications for their use as an immunomodulatory vitamin combination. *International Journal of Immunopharmacology*, 18, 693–700.
- Briganti, S., Camera, E., Picardo, M. (2003). Chemical and instrumental approaches to treat hyperpigmentation. *Pigment Cell Research*, 16, 101-110.
- Busch, W., Duis, K., Fenske, M., Maack, G., Legler, J., Padilla, S., Scholz, S. (2011). The zebrafish embryo model in toxicology and teratology. *Reproductive Toxicology*, 31(4), 585–588.
- Carvalho, M., Ferreira, P. J., Mendes, V. S., Silva, R., Pereira, J. a, Jerónimo, C., & Silva, B. M., (2010). Human cancer cell anti-proliferative and antioxidant activities of *Juglans regia* L. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*, 48(1), 441–7.
- Chan, E.W.C., Lim, Y.Y., Wong, L.F., Lianto, F.S., Wong, S.K., Lim, K. K., Joe, C. E., Lim, T. Y. (2008). Antioxidant and tyrosinase inhibition properties of leaves and rhizomes of ginger species. *Food Chemistry*, 109, 477–483.
- Chan, H. P., Lewis, C., & Thomas, P. S. (2010). Oxidative stress and exhaled breath analysis: a promising tool for detection of lung cancer. *Cancers*, 2(1), pp.32–42.
- Charuwichitratana, S., Wongrattanapasson, N., Timpatanapong, P. and Bunjob, M. (1996). Herpes zoster: Treatment with *Clinacanthus nutans* cream. *International Journal of Dermatology*, 35, 665-666.
- Chavalittumrong, P., Attawish, A., Rungsamon, P., and Chuntapet, P. (1995). Toxicological study of *Clinacanthus nutans* (Burm.f.) Lindau. *Bulletin Department of Medical Service (Thai)*, 37, 323-338.
- Chen, T., He, J., Zhang, J., Li, X., Zhang, H., Hao, J., & Li, L. (2012). The isolation and identification of two compounds with predominant radical scavenging activity in hempseed (seed of *Cannabis sativa* L.). *Food Chemistry*, 134(2), 1030–7.
- Cherdchu, C., Poopyruchpong, N., Adchariya, R., Patanaba Nangkost, S., and Ratanabangkoon, K. (1977). The Absence of Antagonism between Extract of *Clinacanthus nutans* and *Najanaja siamensis*. *The Journal of the Tropical Medicine and Public Health*, 8, 249-254.

- Chou, S. T., Chang, W. L., Chang, C. T., Hsu, S. L., Lin, Y. C., & Shih, Y. (2013). *Cinnamomum cassia* essential oil inhibits α -MSH-induced melanin production and oxidative stress in murine B16 melanoma cells. *International Journal of Molecular Sciences*, 14(9), 19186–201.
- Chuakul, W. (1986). Chemical study of the anti-inflammatory agents from the leaves of Phayaa Plong Thong, *Clinacanthus nutans* (Burm. f.) Lindau. *Master of Science Thesis*, Faculty of Pharmacy, Mahidol University.
- Chung, Y. C., Chang, C. T., Chao et al. (2008). Antioxidative activity and safety of the 50% ethanolic extract from red bean fermented by *Bacillus subtilis* IMR-NK1, *Journal Agricultural & Food Chemistry*, 5, 2454–2458.
- Cody, V., Middleton, E. and Harborne, J. B. (1986). Plant Flavonoids in Biology and Medicine-Biochemical, *Pharmacological, and Structure-activity Relationships*, Alan R. Liss, New York, NY.
- Commission of the European Communities. Directive 2010/63/eu of the European parliament and of the council of 22 September 2010 on the protection of animals used for scientific purposes. *Official Journal of the European Union* 2010, L276, 33–79.
- Cooper, E. L. (2005) “Drug discovery, CAM and natural products,” *Evidence-Based Complementary and Alternative Medicine*, 1(13), 215–217.
- Coussens, L. M. & Werb, Z. (2002). Review article Inflammation and cancer. *Nature*, 420, 860-867.
- Crohns, M., Westermarck, T., Atroshi, F. (2013). Prostate Cancer, Inflammation and Antioxidants. *Open Access*, <http://www.intechopen.com>. Chapter 17.
- Dampawan, P. (1976). Studies of the Chemical Constituents of the *Clinacanthus nutans* (Acanthaceae) and Zingiber Cassumunar Roxb. *Master Thesis*, Mahidol University.
- Dampawan, P., Huntrakul, C., and Reutrakul, V. (1977). Constituents of *Clinacanthus nutans* and the Crystal Structure of LUP-20 (29)3. *Journal of the Science Society of Thailand*, 3, 14-26.
- Das, K., Tiwari, R. K. S., & Shrivastava, D. K. (2010). Techniques for evaluation of medicinal plant products as antimicrobial agent: Current methods and future trends. *Journal of Medicinal Plants Research*, 4(2), 104–111.
- David, A., Pancharatna, K. (2009). Developmental anomalies induced by non-selective COX inhibitor (ibuprofen) in zebrafish (*Danio rerio*). *Environmental Toxicology Pharmaceutical*, 27, 390–395.
- Diplock, A.T., Charleux, J. L., Crozier-Willi, G., Kok, F. J., Rice-Evans, C., Roberfroid, M., Stahl, W., Vina-Ribes. (1998). Functional food science and defence against reactive oxidative species. *Journal of Nutrition*, 80: S77-112.

- Doğanlı, C., Oxvig, C., & Lykke-Hartmann, K. (2013). Zebrafish as a novel model to assess Na⁺/K⁺ (+)-ATPase-related neurological disorders. *Neuroscience and Biobehavioral Reviews*, 37(10.2), 2774–87.
- Duansak, N. (2007). Effects Of Herbal Extract (Gpo1986) On Tumor Angiogenesis Of Hepatocellular Carcinoma Cells (Hepg2) Implanted In Nude Mice. *Degree of Doctor of Philosophy Thesis*. Graduate SchoolChulalongkorn University.
- Esch, C. de, Slieker, R., Wolterbeek, A., Woutersen, R., & Groot, D. de. (2012). Zebrafish as potential model for developmental neurotoxicity testing: a mini review. *Neurotoxicology and Teratology*, 34(6), 545–53.
- Fauci, Anthony S., et al., (2008). Harrison's Principles of Internal Medicine. 17th ed. United States: *McGraw-Hill Professional*.
- Formagio, A. S. N., Kassuya, C. L., Neto, F. F., Volobuff, C. R. F., Iriguchi, E. K. K., Vieira, M. D. C., & Foglio, M. A. (2013). The flavonoid content and anti-proliferative, hypoglycaemic, anti-inflammatory and free radical scavenging activities of Annonadioica St. Hill. *BMC Complementary and Alternative Medicine*, 13(1), 14.
- Fu, L., Xu, B.T., Xu, X.R., Gan, R.Y., Zhang, Y., Xia, E.Q., & Li, H.B. (2011). Antioxidant capacities and total phenolic contents of 62 fruits. *Food Chemistry*, 129(2), 345–350.
- Graves, D.B. (2012). The emerging role of reactive oxygen and nitrogen species in redox biology and some implications for plasma applications to medicine and biology. *Journal of Physics. D: Applied Physics*, 45, 2630-01.
- Guo, C., Yang, J., Wei, J., Li, Y., Xu, J., & Jiang, Y. (2003). Antioxidant activities of peel, pulp and seed fractions of common fruits as determined by FRAP assay. *Nutrition Research*, 23, 1719–1726.
- Haendel, M. A., Tilton, F., Bailey, G. S., et al. (2004). Developmental toxicity of the dithiocarbamate pesticide sodium metam in zebrafish. *Toxicological Science*, 81, 390–400.
- Halliwell B, Gutteridge J. M. (1990). Role of free radicals and catalytic metal ions in human disease: an overview. *Methods Enzymol*, 186, 1-85.
- Halliwell, B. (1996). Antioxidants in human health and disease. *Annual Review of Nutrition* 16, 33–50
- Hassasroudsari, M., Chang, P., Pegg, R., & Tyler, R. (2009). Antioxidant capacity of bioactives extracted from canola meal by subcritical water, ethanolic and hot water extraction. *Food Chemistry*, 114(2), 717–726.
- Havsteen, B. (1983). Flavonoids, a Class of Natural Products of High Pharmacological Potency. *Biochemical Pharmacology*, 32(7), 1141-1148.

- Havsteen, B. H., (2002). The biochemistry and medical significance of the flavonoids. *Pharmacology & Therapeutics*, 96, 67–202.
- Heldt H-W, Heldt F. (2005). Secondary metabolites fulfill specific ecological functions in plants. *Plant Biochemistry*. 3rd ed. Academic Press, Burlington, USA, 403-412.
- Hill, A. J., Teraoka, H., Heideman, W., & Peterson, R. E., (2005). Zebrafish as a model vertebrate for investigating chemical toxicity. *Toxicological Sciences: An Official Journal of the Society of Toxicology*, 86(1), 6–19.
- Hosen, M. J., Vanakker, O. M., Willaert, A., Huysseune, A., Coucke, P., & De Paepe, A. (2013). Zebrafish models for ectopic mineralization disorders: practical issues from morpholino design to post-injection observations. *Frontiers in Genetics*, 4(May), 74.
- <http://www.sabahsnakegrassfarm.com/2011/06/clinacanthus-nutans-l-sabah-snake-grass.html>. Khoo Kiang Lin. (2011).
- Huang, C. C., Chen, P.C., Huang, C.W., Yu, J. (2007). Aristolochic acid induces heart failure in zebrafish embryos that is mediated by inflammation. *Toxicological Sciences*, 100(2), 486–494.
- Huang, D., Ou, B., & Prior, R. L. (2005). The chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry*, 53(6), 1841–56.
- Incardona, J. P., Collier, T. K., Scholz, N. L. (2004). Defects in cardiac function precede morphological abnormalities in fish embryos exposed to polycyclic aromatic hydrocarbons. *Toxicology Applied Pharmacology*, 196, 191–205.
- Iqbal, D., Khan, M. S., Khan, A., Khan, M. S., Ahmad, S., Srivastava, A. K., & Bagga, P. (2014). In vitro screening for β -hydroxy- β -methylglutaryl-CoA reductase inhibitory and antioxidant activity of sequentially extracted fractions of *Ficus palmate* Forsk. *BioMedical Research International*, 2014, 762620.
- Jayavas, C. (1998). Clinical Trial in the Treatment of Genital Herpes Patients with *Clinacanthus nutans* Extract. In the 9th Ministry of Public Health Symposium (57).
- Jayavas, C., Balachandra, K., Sangkitporn, S., Maharungraungrat, A., Thavatsupa, P., Bunjob, M., Chavalittumrong, P., Dechatiwongse, T., Sittisomwong, N. (1992a). Clinical trial in the treatment of genital herpes patients. *Communicable Disease Journal (Thailand)* 18 (3), 152–161.
- Jayavas, C., Dechatiwongse, T., Balachandra, K., Chavalittumrong, P., Jongtrakulsiri, S. (1992b). The virucidal activity of *Clinacanthus nutans* Lindau extracts against herpes simplex virus type 2: An in vitro study. *Bulletin of the Department of Medical Science (Thailand)* 34 (4), 153–158.

- Kim, B. M. & Chung, H. W. (2007). Hypoxia / reoxygenation induces apoptosis through a ROS-mediated caspase-8 / Bid / Bax pathway in human lymphocytes. *Biochemical and Biophysical Research Communications*, 363, 745–750.
- Kimmel, C.B., Ballard, W.W., Kimmel, S.R., Ullmann, B., Schilling, T. F. (1995). Stages of embryonic-development of the zebrafish. *Developmental Dynamics*, 203, 253–310.
- Kuhnau, J. (1976). “The flavonoids: a class of semi-essential food components: their role in human nutrition,” *World Revision Nutrition Diet*, 24, 117-91.
- Kuliscic, T., Radonic, A., Katalinic, V., Milos, M. (2004). Use of different methods for testing antioxidative activity of oregano essential oil. *Food Chemical*, 85, 633–640.
- Lau, K. W., Lee, S. K., & Chin, J. H. (2014). Effect of the methanol leaves extract of *Clinacanthus nutans* on the activity of acetylcholinesterase in male mice. *Journal of Acute Disease*, 22–25.
- Lee, K. H., Ab Aziz, F. H., Syahida, A., Abas, F., Shaari, K., Israf, D. A., & Lajis, N. H. (2009). Synthesis and biological evaluation of curcumin-like diarylpentanoid analogues for anti-inflammatory, antioxidant and anti-tyrosinase activities. *European Journal of Medicinal Chemistry*, 44(8), 3195–200.
- Lee, K. H., Padzil, A. M., Syahida, A., Abdullah, N., Zuhainis, S. W., & Maziah, M. (2011). Evaluation of anti-inflammatory, antioxidant and anti- nociceptive activities of six Malaysian medicinal plants. *Journal of Medicinal Plants Research*, 5(xx).
- Li, H. B., Wong, C. C., Cheng, K. W., & Chen, F. (2008). Antioxidant properties in vitro and total phenolic contents in methanol extracts from medicinal plants. *LWT - Food Science and Technology*, 41(3), 385–390.
- Li, Y. (2011). Antioxidants In Biology And Medicine : Essentials, Advances , And Clinical Applications. *Nova Science Publisher*.
- Lim, Y. Y., Lim, T. T., & Tee, J. J. (2007). Antioxidant properties of several tropical fruits: A comparative study. *Food Chemistry*, 103(3), 1003–1008.
- Lin, J. Y., & Tang, C. Y. (2007). Determination of total phenolics and flavonoid contents in selected fruits and vegetables, as well as their stimulatory effects on mouse splenocyte proliferation. *Food Chemistry*, 101(1), 140–147.
- Lin, J., Li, H., and Yu, J. (1983). Studies on the Chemical Constituents of Nui Xu Hua (*Clinacanthus nutans*). *Zhongcaoyao*, 14 (8), 337-338.
- Liu, R. H. (2003). Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. *The American Journal of Clinical Nutrition*, 78, 3–6.

- Lu, H., Ouyang, W., & Huang, C. (2006). Inflammation, a key event in cancer development. *Molecular Cancer Research: MCR*, 4(4), 221–33.
- Manda, G., Nechifor, M. T., & Neagu, T. M. (2009). Reactive Oxygen Species, Cancer and Anti-Cancer Therapies. *Current Chemical Biology*, 3(1), 342–366.
- Mantovani A. (2010). Molecular pathways linking inflammation and cancer. *Current Molecules Medicines*. 10, 369-73.
- Marxen, K., Vanselow, K. H., Lippemeier, S., Hintze, R., Ruser, A., & Hansen, U. P. (2007). Determination of DPPH Radical Oxidation Caused by Methanolic Extracts of Some Microalgal Species by Linear Regression Analysis of Spectrophotometric Measurements. *Sensors*, 7(10), 2080–2095.
- Middleton, E. (1984). “The flavonoids,” *Trends in Pharmaceutical Science*, 5, 335-8.
- Multhoff, G., & Radons, J. (2012). Radiation, inflammation, and immune responses in cancer. *Frontiers in Oncology*, 2(June), 58.
- Narayanaswamy, N., Duraisamy, A., & Balakrishnan, K. P. (2011). Screening of some Medicinal Plants for their Anti-tyrosinase and Antioxidant activities. *International Journal of PharmaTech Research*, 3(2), 1107–1112.
- Nemat, K., Yadollah, S., Mahdi, M. (2009). Chronic Inflammation and Oxidative Stress as a Major Cause of Age-Related Diseases and Cancer. *Recent Patents on Inflammation & Allergy. Drug Discovery*, 3 (1), 73-80.
- Organisation for Economic Cooperation and Development (OECD). (2012). Guideline for Testing of Chemicals. Guideline 210: Fish, early-life stage toxicity test and Fish Embryo acute aquatic toxicity (FET) Test.
- Oyaizu, M. (1986). Studies on product of browning effect reaction prepared from glucose amine. *Journal of Nutrition*, 44, 307–315.
- P'ng, X. W., Gabriel, A. A., and Chin, J. H. (2012). Acute oral toxicity study of *Clinacanthus nutans* in mice. *International Journal of Pharmaceutical Research and Sciences*, 3(11), 4202-4
- Panda, S. K., (2012). Assay Guided Comparison for Enzymatic and Non-Enzymatic Antioxidant Activities with Special Reference to Medicinal Plants. *Intech*, 381 – 400.
- Pannangpetch, P., Laupattarakasem, P., Kukongviriyapan, V., Kukongviriyapan, U., Kongyingyoes, B., & Aromdee, C. (2007). Antioxidant activity and protective effect against oxidative hemolysis of *Clinacanthus nutans* (Burm. f) Lindau . *Songklanakarin. Journal of Science and Technology*, 29, 1–9.
- Pękal, A., & Pyrzynska, K. (2014). Evaluation of Aluminium Complexation Reaction for Flavonoid Content Assay. *Food Analytical Methods*.

- Peng, T. W., Wen, P. X., Han, C. J., & Akowuah, G. A. (2014). Effects of Methanol Extract of *Clinacanthus nutans* on Serum Biochemical Parameters in Rats. *Journal of Applied Pharmacy*, 6(1), 77–86.
- Percival, M., (1998). Antioxidant. *Clinical Nutrition Insights*.
- Price KR, Johnson IT, Fenwick G. R. (1987). The chemistry and biological significance of saponins in foods and feedstuffs. *CRC Critical Review Food Science Nutrition*, 26, 27–135.
- Prior, R. L., Wu, X., & Schaich, K. (2005). Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry*, 53(10), 4290–4302.
- Rakoff-nahoum, S. (2006). Why Cancer and Inflammation? *Journal of Biology and Medicine*, 79, 123–130.
- Rao, L.G.&Rao, A.V. (2013). Oxidative Stress and Antioxidants in the Risk of Osteoporosis-Role of the Antioxidants Lycopene and Polyphenols. *Intech Journal and Open Access*, 51(8), 978-953.
- Reddy, C. V. K., Sreeramulu, D., & Raghunath, M. (2010). Antioxidant activity of fresh and dry fruits commonly consumed in India. *Food Research International*, 43(1), 285–288.
- Reimers, M. J., Flockton, A. R., & Tanguay, R. L. (2004). Ethanol and acetaldehyde-mediated developmental toxicity in zebrafish. *Neurotoxicology and Teratology*, 26(6), 769–81.
- Renier, C., Faraco, J. H., Bourgin, P., Motley, T., Bonaventure, P., Rosa, F. (2007). Genomic and functional conservation of sedative-hypnotic targets in the zebrafish. *Pharmacogenetic Genomics*, 17, 237–53.
- Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure–antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*, 20, 933–956.
- Riboli, E., Norat, T. (2003). Epidemiologic evidence of the protective effect of fruit and vegetables on cancer risk. *American Journal of Clinical Nutrition* 78 (3), 559S–569S.
- Russo, G.L., (2007). Ins and outs of dietary phytochemicals in cancer chemoprevention. *Biochemical Pharmacology*, 74, 533–544.
- Saewan, N., Koysomboon, S., & Chantrapromma, K., (2011). Anti-tyrosinase and anti-cancer activities of flavonoids from *Blumea balsamifera* DC, 5(6), 1018–1025.
- Sakdarat, S., Shuyprom, A., Pientong, C., Ekalaksananan, T., & Thongchai, S. (2009). Bioactive constituents from the leaves of *Clinacanthus nutans* lindau. *Bioorganic & Medicinal Chemistry*, 17(5), 1857-1860.

- Salganik, R. I. (2001). The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients and the human population. *Journal of the American College of Nutrition*, 20(5), 464S–472S
- Sancho, L. E. G., Yahia, E. M., & González-aguilar, G. A., (2011). Identification and quantification of phenols, carotenoids, and vitamin C from papaya (*Carica papaya* L., cv. Maradol) fruit determined by HPLC-DAD-MS / MS-ESI. *FRIN*, 44(5), 1284–1291.
- Sangkitpporn, S., Polchan, K., Balachandra, K., Dechatiwongsena Ayudhaya, T., Bunchob, M., and Jayavas, C. (1993). Treatment of Herpes Zoster Patients with *Clinacanthus nutans* extract. *Journal of the Medical Association of Thailand*, 78 (11), 624-627.
- Sangkitpporn, S., Polchan, K., Thawatsupa, P., Bunchob, M., and Chawalitumrong, P. (1993). Treatment of Recurrent Genital Herpes Simplex Virus Infection with *Clinacanthus nutans* extract. *Bulletin of the Department of Medical Service*, 18(5), 226-231.
- Satakhun, S. (2001). Chemical Constituents of *Clinacanthus nutans* leaves. *Master thesis*, Chulalongkorn University, Bangkok, Thailand.
- Satayavivad, J., Bunyaphrathasara, N., Kittisiripornkul, S. and Tanasomwong, W. (1996). Analgesic and anti-inflammatory activities of extract of *Clinacanthus nutans* (Burm. f) Lindau. *Thai Journal of Phytopharmaceutical Sciences*, 3, 7-17
- Seo, S. Y., Sharma, V. K., Sharma, N. (2003). Mushroom tyrosinase: Recent prospects. *Journal of Agriculture & Food Chemistry*, 51, 2837-2853.
- Shephard, J. J., Soper, A. K., Callear, S. K., Imberti, S., Evans, J. S. O., Salzmann, C. G. (2015). Super-dipoles linked to chloroform's outstanding solvent properties.
- Shuyprom, A. (2004). Chemical Composition Investigation of the *Clinacanthus nutans* (Burm. F.) Lindau leaves. *Master Thesis*, Suranaree University of Technology.
- Sies, H. (1991) Oxidative Stress. Oxidants and Antioxidants. *Academic Press, New York*
- Strmac, M., Braunbeck, T. (1999). Effects of triphenyltin acetate on survival, hatching success, and liver ultrastructure of early life stages of zebrafish (*Danio rerio*). *Ecotoxicology Environmental Safety*, 44, 25–39.
- Strmac, M., Oberemm, A., Braunbeck, T., et al. (2002). Effects of sediment eluates and extracts from differently polluted small rivers on zebrafish embryos and larvae. *Journal of Fish Biology*, 61, 24–38.
- Sulaiman, S. F., Sajak, A. A. B., Ooi, K. L., & Seow, E. M. (2011). Effect of solvents in extracting polyphenols and antioxidants of selected raw vegetables. *Journal of Food Composition and Analysis*, 24(4-5), 506–515.

- Surveswaran, S., Cai, Y., Corke, H., & Sun, M. (2007). Systematic evaluation of natural phenolic antioxidants from 133 Indian medicinal plants. *Food Chemistry*, 102(3), 938–953.
- Shuyprom, A. (2004). Chemical Composition Investigation of the *Clinacanthus nutans* (Burm. F.) Lindau Leaves. *Degree of Master of Science* in Chemistry Suranaree University of Technology.
- Tachakittirungrod, S., Okonogi, S., & Chowwanapoonpohn, S. (2007). Study on antioxidant activity of certain plants in Thailand: Mechanism of antioxidant action of guava leaf extract. *Food Chemistry*, 103(2), 381–388.
- Tahboub, Y. R. (2008). Determination of Nitrite and Nitrate in Cell Culture Medium by Reversed-Phase High-Performance Liquid Chromatography with Simultaneous UV-VIS and Fluorescence Detection. *Jordan Journal of Chemistry*, 3(1), 69–75.
- Tanasomwong, W. (1986). The Screening of anti-inflammatory action of *Clinacanthus nutans* (Burm. f.) A critical evaluation of carrageenan induced hind paw oedema model. *Master of Science Thesis*, Mahidol University.
- Teshima, K., Kaneko, T., Ohtani, K., Kasai, R., Lhieochaiphant, S., Picheansoonthon, C., and Yamasaki, K. (1997). C-Glycosyl Flavones from *Clinacanthus nutans*. *Natural Medicines*, 51(6), 557.
- Teshima, K., Kaneko, T., Ohtani, K., Kasai, R., Lhieochaiphant, S., Picheansoonthon, C., & Yamasaki, K. (1998). Sulfur-containing glucosides from *Clinacanthus nutans*. *Phytochemistry*, 48(5), 831-835.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L., & Hawkins Byrne, D. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19(6-7), 669–675.
- Thongharb, C. and Tejasen, P. (1977). The effect of Slaed Pang Pon (*Clinacanthus nutans*) on Thai Cobra Venom (*Najanaja siamensis*). *Thai Journal of Phytopharmaceutical Sciences*, 2, 1057-1063.
- Tu, C. X., Lin, M., Lu, S. S., Qi, X. Y., Zhang, R. X., Zhang, Y. Y. (2012). Curcumin inhibits melanogenesis in human melanocytes. *Phytotherapeutic Research*, 26, 174–179.
- Tuntiawachwuttikul, P., Pootaeng-On, Y., Phansa, P., & Taylor, W. C. (2004). Cerebrosides and a monoacylmonogalactosylglycerol from *Clinacanthus nutans*. *Chemical & Pharmaceutical Bulletin*, 52(1), 27–32.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T. D., Mazur, M., & Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry and Cell Biology*, 39(1), 44–84.

- Valko, M., Rhodes, C. J., Moncol, J., Izakovic, M., and Mazur, M. (2006). "Free radicals, metals and antioxidants in oxidative stress induced cancer." *Chemico-Biological Interactions*, 160(1), 1–40.
- Vishnu, P., and Aboulafia, D. M. (2013). The Oncogenicity of Human Cytomegalovirus. Open Access. <http://www.intechopen.com>.
- Wakamatsu, T. H., Dogru, M., & Tsubota, K. (2008). Tearful relations: oxidative stress, inflammation and eye diseases. *Arquivos Brasileiros de Oftalmologia*, 71(6 Suppl), 72–9.
- Wang, K.H., Lin, R.D., Hsu, F.L., Huang, Y.H., Chang, H.C., Huang, C.Y., & Lee, M.H. (2006). Cosmetic applications of selected traditional Chinese herbal medicines. *Journal of Ethnopharmacology*, 106(3), pp. 353–9.
- Wang, S., Liu, K., Wang, X., He, Q., & Chen, X. (2011). Toxic effects of celastrol on embryonic development of zebrafish (*Danio rerio*). *Drug and Chemical Toxicology*, 34(1), 61–5.
- Wang, S. F., Liu, K. C. H., Wang, X. M., et al. (2009). Preliminary study on cardiotoxicity of celastrol to zebrafish embryo. *Chinese Pharmacology Bulletin*, 24, 634–636.
- Wang, S., Meckling, K. A., Marcone, M. F., Kakuda, Y., & Tsao, R. (2011). Can phytochemical antioxidant rich foods act as anti-cancer agents? *Food Research International*, 44(9), 2545–2554.
- Wanikiat, P., Panthong, A., Sujayanon, P., Yoosook, C., Rossi, A. G., and Reutrakul, V. (2008). "The anti-inflammatory effects and the inhibition of neutrophil responsiveness by *Barleria lupulina* and *Clinacanthus nutans* extracts." *Journal of Ethnopharmacology*, 116(2), 234–44.
- Weigt, S., Huebler, N., Strecker, R., Braunbeck, T., & Broschard, T. H. (2011). Zebrafish (*Danio rerio*) embryos as a model for testing proteratogens. *Toxicology*, 281(1-3), 25–36.
- Westerfield, M. (1995). The zebrafish book: A guide for the laboratory use of zebrafish (*Danio rerio*).
- Yamakoshi, J., Otsuka, F., Sano, A., Tokutake, S., Saito, M., Kikuchi, M., Kubota, Y. (2003). Lightening effect on ultraviolet-induced pigmentation of guinea pig skin by oral administration of a proanthocyanidin-rich extract from grape seeds. *Pigment Cell Research* 16, 629–638.
- Yang, L., Ho, N. Y., Alshut, R., Legradi, J., Weiss, C., Reischl, M., Strähle, U. (2009). Zebrafish embryos as models for embryotoxic and teratological effects of chemicals. *Reproductive Toxicology (Elmsford, N.Y.)*, 28(2), 245–53.

- Yong, Y. K., Tan, J. J., Teh, S. S., Mah, S. H., Ee, G. C. L., Chiong, H. S., & Ahmad, Z. (2013). *Clinacanthus nutans* Extracts Are Antioxidant with Anti-proliferative Effect on Cultured Human Cancer Cell Lines. *Evidence-Based Complementary and Alternative Medicines: eCAM*, 462751.
- Young, I. S., & Woodside, J. V. (2001). Antioxidants in health and disease. *Journal of Clinical Pathology*, 54, 176–186
- Yuann, J. P., Wang, J., Jian, H., Lin, C., & Liang, J. (2012). Effects of *Clinacanthus nutans* (Burm. f) Lindau leaf extracts on protection of plasmid DNA from riboflavin photoreaction. *MC- Transaction on Biotechnology*, 4(1), 45–58.
- Zhang, Lin., Anjaneya S. R., Koyyalamudi, S.R., Jeong, S. C., Reddy, N., Smith Paul, T., Bartlett, J., Kirubakaran S., G. M. and Wu, M. J. (2012). Antioxidant and anti-inflammatory activities of selected medicinal plants and fungi containing phenolic and flavonoid compounds. *Chinese Medicine*, 7(1), 26.
- Zhang, Y., Seeram, N.P., Lee, R., Feng, L., Heber, D., (2008). Isolation and identification of strawberry phenolics with antioxidant and human cancer cell anti-proliferative properties. *Journal of Agriculture & Food Chemistry*, 56, 670–675.
- Zhao, L., Feng, C., Hou, C., Hu, L., Wang, Q., & Wu, Y. (2015). First Discovery of Acetone Extract from Cottonseed Oil Sludge as a Novel Antiviral Agent against Plant Viruses. *Plos One*, 10(2), e0117496.
- Zhu, X., Zhu, L., Li, Yan, Duan, Z., Chen, W., & Alvarez, P. J. J., (2007). Developmental toxicity in zebrafish (*Danio rerio*) embryos after exposure to manufactured nanomaterials: buckminster fullerene aggregates (nc 60) and fullerol. *Environmental Toxicology and Chemistry*, 26(5), 976–979.
- Zhu, X., Zhu, L., Duan, Z., Qi, R., Li, Y., and Lang, Y., (2008). “Comparative toxicity of several metal oxide nanoparticle aqueous suspensions to Zebrafish (*Danio rerio*) early developmental stage.” *Journal Environmental Science Health A. Toxicology Hazard Substance Environment Engineering*, 43(3), 278–84.

BIODATA OF STUDENT

The student, Yu Guk Ting was born on 18th October 1974, in Lumut, Perak. She received her early education at Sekolah Rendah Jenis Kebangsaan Methodist Ayer Tawar, and then she went to Sekolah Menengah Jenis Kebangsaan Methodist for her lower secondary education and Sekolah Menengah Kebangsaan Ambrose, Ayer Tawar for her upper secondary education. She continued to her pre-university level at Sekolah Menengah Jenis Kebangsaan ACS, Sitiawan.

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