



***PHYTOCHEMICALS AND BIOLOGICAL ACTIVITIES OF ELAEOCARPUS
FLORIBUNDUS BLUME, BARRINGTONIA CONOIDEA GRIFF AND
FIBRAUREA TINCTORIA LOUR***

RAHAYU UTAMI UMAR

FS 2012 109

PHYTOCHEMICALS AND BIOLOGICAL ACTIVITIES OF *ELAEOCARPUS FLORIBUNDUS* BLUME, *BARRINGTONIA CONOIDEA* GRIFF AND *FIBRAUREA TINCTORIA* LOUR

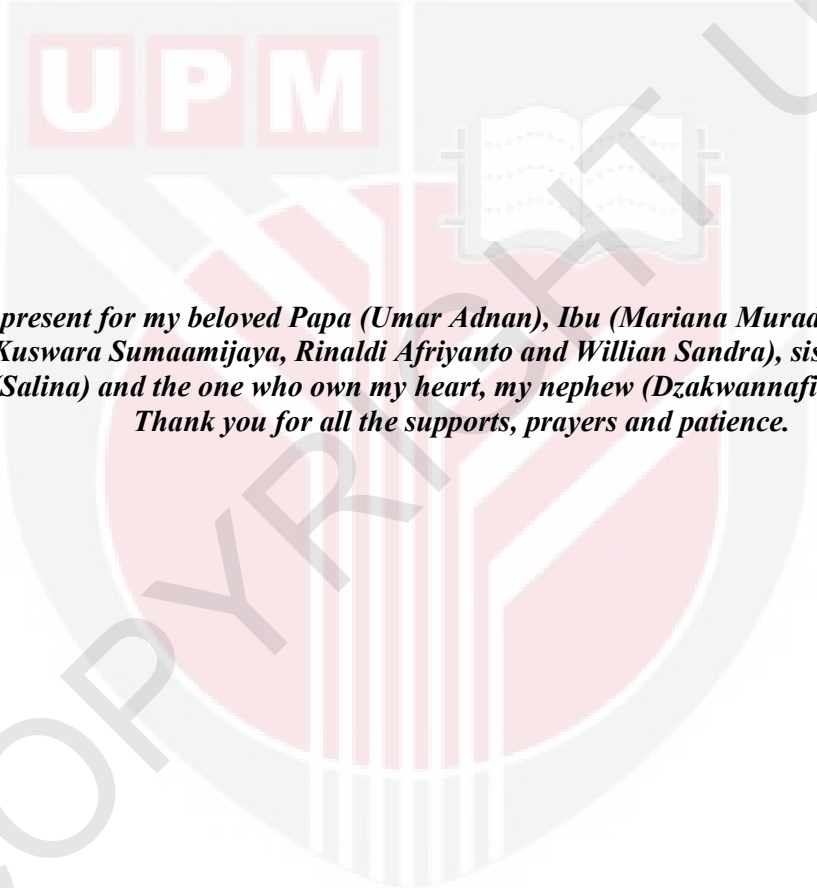


By

RAHAYU UTAMI UMAR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

September 2012

The logo of Universiti Pendidikan Malaysia (UPM) is a shield-shaped emblem. At the top left, the letters 'UPM' are written in white on a red rectangular background. The shield is divided into several sections: a top right section with an open book, a middle section with a stylized white and red geometric design, and a bottom section with vertical red and white stripes. A large, faint watermark of the UPM logo and the text 'COPYRIGHT UPM' is overlaid diagonally across the entire page.

*A present for my beloved Papa (Umar Adnan), Ibu (Mariana Murad), brothers (Kuswara Sumaamijaya, Rinaldi Afriyanto and Willian Sandra), sister-in law (Salina) and the one who own my heart, my nephew (Dzakwannafis Syafiq).
Thank you for all the supports, prayers and patience.*

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PHYTOCHEMICALS AND BIOLOGICAL ACTIVITIES OF *ELAEOCARPUS FLORIBUNDUS* BLUME, *BARRINGTONIA CONOIDEA* GRIFF AND *FIBRAUREA TINCTORIA* LOUR

By

RAHAYU UTAMI UMAR

September 2012

Chairman : Prof. Mohd. Aspollah Hj. Sukari, PhD

Faculty : Science

Phytochemicals and biological activity studies on three medicinal plants, *Elaeocarpus floribundus* Blume, *Barringtonia conoidea* Griff and *Fibraurea tinctoria* Lour were carried out. The chemical constituents of these plants were isolated using chromatographic methods, whilst the structure of the compounds were elucidated by using spectroscopic methods including infrared (IR), nuclear magnetic resonance (^1H , ^{13}C and 2D NMR), mass spectrometry (MS) and by comparison with previous data. Total phenolic content of methanolic extracts of plants was determined by colorimetric method using Folin-Ciocalteu reagent. Extracts and selected isolated compounds were subjected to biological activity studies of antimicrobial, antioxidant and cytotoxic activity.

The isolation work on the leaves of *Elaeocarpus floribundus* has yielded triterpenoids, friedelin (**38**) and epifriedelinol (**39**). Mixture of α - (**41**) and β -amyrin

(42) were isolated from leaves and stem bark of *Barringtonia conoidea*. This is a first report on the phytochemical study of *Elaeocarpus floribundus* and *Barringtonia conoidea*. While furanolactone diterpene, clerodanine (43), an ecdysteroid, 20-hydroxyecdysone (37) and an isoquinoline alkaloid, berberine (30) were obtained from the whole part of *Fibraurea tinctoria*. In addition, β -sitosterol (40) was isolated from these three plant species.

In this present study, all methanolic extracts of plants showed significant amount of phenolic content. The methanolic extract of leaves of *Elaeocarpus floribundus* exhibited the highest value of 503.08 ± 16.71 mg GAE/g DW. In the screening of antimicrobial activities, extracts of leaves and stem bark of *Elaeocarpus floribundus* and *Barringtonia conoidea* were either possessed weak activity or inactive against gram positive and negative bacterial strains. Extracts of *Fibraurea tinctoria* were found to be inactive towards the bacterial strains. Similarly, none of extracts of the plants gave inhibition activity towards fungal strains *Candida albicans*, *Aspergillus ochraceus* and *Saccharomyces cerevisiae*.

Antioxidant activity assay of extracts and isolated compounds have been conducted using DPPH free radical scavenging method. Only polar extracts of leaves and stem bark of *Elaeocarpus floribundus* as well as methanol extract of *Fibraurea tinctoria* afforded potential antioxidant activity. Clerodanine (43) was the only isolated compound that gave significant activity with IC_{50} value of 20.81 ± 0.02 μ g/ml. While, the others extracts as well as isolated compounds were found either to be weak or inactive.

As for the cytotoxic assay, extracts and selected isolated compounds were tested using microculture tetrazolium (MTT) assay against human T4-lymphoblastoid cell (CEM-SS) and human cervical cancer cell (HeLa). The results showed that chloroform extract of leaves of *Elaeocarpus floribundus* and methanol extract of *Fibraurea tinctoria* gave significant activity against CEM-SS cells with IC_{50} values of 25.6 ± 0.06 and 16.13 ± 0.04 $\mu\text{g/ml}$, respectively. Ethyl acetate extract of stem bark and chloroform extract of leaves of *Barringtonia conoidea* exhibited promising activity towards HeLa cells with IC_{50} values of 13.5 ± 0.16 and 28.1 ± 0.01 $\mu\text{g/ml}$, respectively.

Compounds **38**, **39**, **43** and mixture of **41** & **42** showed significant cytotoxic activity against the two cancer cells, with compound **38** gave the strongest activity with IC_{50} value of 3.54 ± 0.30 $\mu\text{g/ml}$ towards HeLa cells. Compound **43** was also found to be active against the two cancer cells with IC_{50} values of 12.49 ± 0.10 $\mu\text{g/ml}$ (CEM-SS) and 9.37 ± 0.36 $\mu\text{g/ml}$ (HeLa). This is the first study conducted on the cytotoxic activity of compound **43**.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KAJIAN FITOKIMIA DAN AKTIVITI BIOLOGI KE ATAS *ELAEOCARPUS FLORIBUNDUS* BLUME, *BARRINGTONIA CONOIDEA* GRIFF DAN *FIBRAUREA TINCTORIA* LOUR

Oleh

RAHAYU UTAMI UMAR

September 2012

Pengerusi : Prof. Mohd. Aspollah Hj. Sukari, PhD

Fakulti : Sains

Kajian fitokimia dan aktiviti biologi ke atas tiga tumbuhan ubat-ubatan, *Elaeocarpus floribundus* Blume, *Barringtonia conoidea* Griff dan *Fibraurea tinctoria* Lour telah dijalankan. Juzuk kimia tumbuh-tumbuhan ini telah diasingkan dengan menggunakan kaedah kromatografi, manakala struktur sebatian telah dijelaskan dengan menggunakan kaedah spektroskopik termasuk inframerah (IR), resonans magnetik nuklear (^1H , ^{13}C dan 2D NMR), spektrometri jisim (MS) dan perbandingan dengan data sebelumnya. Kandungan jumlah fenol daripada ekstrak metanol pokok tumbuh-tumbuhan telah ditentukan oleh kaedah kolorimetrik menggunakan reagen Folin-Ciocalteu. Ekstrak dan sebatian terpencil yang terpilih tertakluk kepada kajian aktiviti biologi seperti antimikrob, antioksidan dan aktiviti sitotoksik.

Kerja pemencilan ke atas daun *Elaeocarpus floribundus* telah menghasilkan triterpenoids, friedelin (**38**) dan epifriedelinol (**39**). Campuran α -(**41**) dan β -amyirin

(42) telah diasingkan daripada daun dan kulit kayu batang *Barringtonia conoidea*. Ini adalah laporan pertama kepada kajian fitokimia *Elaeocarpus floribundus* dan *Barringtonia conoidea*. Manakala diterpene furanolakton, clerodanine (43), ecdysteroid, 20-hydroxyecdysone (37) dan yang isokuinolin alkaloid, berberine (30) telah diperolehi daripada kerja-kerja pengasingan pada bahagian seluruh *Fibraurea tinctoria*. Di samping itu, β -sitosterol (40) telah diasingkan daripada ketiga-tiga spesies tumbuhan.

Di dalam kajian ini, semua ekstrak metanolik daripada pokok tumbuhan menunjukkan jumlah kandungan fenolik yang ketara. Ekstrak metanolik daun *Elaeocarpus floribundus* mempamerkan nilai tertinggi 503.08±16.71 mg GAE/g DW. Dalam ujian aktiviti antimikrob, ekstrak daun dan kulit batang *Elaeocarpus floribundus* dan *Barringtonia conoidea* mempunyai aktiviti yang lemah atau tidak aktif terhadap strain bakteria gram positif dan negatif. Semua ekstrak daripada *Fibraurea tinctoria* telah didapati tidak aktif. Begitu juga, tiada ekstrak daripada tumbuh-tumbuhan ini memberikan aktiviti antifungal terhadap *Candida albicans*, *Aspergillus ochraceus* dan *Saccharomyces cerevisiae*.

Ujian aktiviti antioksidan ekstrak dan sebatian terencil telah dijalankan menggunakan kaedah DPPH free radical scavenging. Hanya ekstrak polar daun dan kulit batang *Elaeocarpus floribundus* serta ekstrak metanol *Fibraurea tinctoria* yang berpotensi memberikan aktiviti antioksidan. Clerodanine (43) adalah satu-satunya sebatian yang memberikan aktiviti yang signifikan dengan nilai IC_{50} 20.81±0.02 μ g/ml. Ekstrak dan sebatian terencil lainnya didapati memberikan aktiviti yang lemah atau tidak aktif.

Untuk ujian sitotoksik, ekstrak dan beberapa sebatian telah diuji menggunakan kaedah microculture tetrazolium (MTT) terhadap sel T4-lymphoblastoid manusia (CEM-SS) dan sel kanser serviks manusia (HeLa). Hasilnya menunjukkan bahawa ekstrak kloroform daun *Elaeocarpus floribundus* dan ekstrak metanol *Fibraurea tinctoria* memberi aktiviti yang signifikan terhadap sel CEM-SS dengan nilai IC_{50} 25.6±0.06 dan 16.13±0.04 µg/ml, masing-masing. Ekstrak etil asetat kulit batang dan ekstrak kloroform daun *Barringtonia conoidea* mempamerkan aktiviti sitotoksik yang menjanjikan ke atas sel HeLa dengan nilai IC_{50} 13.5±0.16 and 28.1±0.01 µg/ml, masing-masing.

Sebatian **38**, **39**, **43** dan campuran sebatian **41** & **42** memberikan aktiviti sitotoksik yang ketara terhadap kedua-dua sel-sel kanser, sebatian **38** memberikan aktiviti terkuat dengan IC_{50} nilai 3.54±0.30 µg/ml terhadap sel HeLa. Sebatian **43** juga didapati aktif terhadap kedua-dua sel-sel kanser dengan nilai IC_{50} 12.49±0.10 µg/ml (CEM-SS) dan 9.37±0.36 µg/ml (HeLa). Ini adalah kajian pertama dijalankan pada aktiviti sitotoksik sebatian **43**.

ACKNOWLEDGEMENTS

My first and foremost gratitude goes to my Lord, Allah SWT for giving me strength, guardian and love to pursue my dreams and complete one stage accomplishment in my life.

I would like to express my deepest thanks to my supervisor, Prof. Dr. Mohd Aspollah Sukari, for his guidance, advice, support and encouragement throughout my research and thesis. Thanks are extended to my co-supervisors, Prof. Dr. Mawardi Rahmani, Dr. Ahmad Bustamam Abdul and Prof. Dr. Dachriyanus, Apt for their advices and assistance in my research. My appreciations are due to Mrs. Latifah Zainal Abidin and Mr. Shamsul Khamis for helping in the identification of my studied plant samples.

Special thanks are gift to Graduated Research Fellowship, Universiti Putra Malaysia, Malaysian Ministry of Science, Technology and Innovation (MOSTI) and Government of Riau Province, Indonesia for financial support during my study.

My thanks are also goes to science officers and laboratory assistants Mr. Johadi Iskandar, Ms. Shareena Safiai, Madam Rusnaini Amirudin, Mr Zainal Kassim, Mr Abas Abd Rahman, Mr Nordin Ismail, Mr Isharudin Mison, Mr Ismail Yassin, Madam Rakina Munaf for their helpful assistance and co-operation.

For my labmates, Dr. Tang Sook Wah, Noor Haslizawati Abu Bakar, Nurul Waznah Muhd Sharif, Mohd Zulkhairi azid, Noorul Adawiyah Mustahil, Halimatul Saadiah

Mohd Noor, Sadikah Ahmad and Noorliyana Ithnin for ideas, suggestions and beloved supports throughout this research. My gratitudes are extended to Dr. Syam Mohan, Baiti and Amin for their assistance and help in *in vitro* cytotoxic study.

Finally, biggest and sincere thanks are appreciated to my parents, my brothers and sister-in law, all family members, housemates and friends for their patience, loves and prayers, unconditional supports and encouragements. Thank you very much.



I certify that a Thesis Examination Committee has met on **date on viva voce** to conduct the final examination of **Rahayu Utami** on her thesis entitled **“Phytochemicals and Biological Activities of *Elaeocarpus Floribundus* Blume, *Barringtonia Conoidea* Griff And *Fibraurea Tinctoria* Lour”** in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the (Name of relevant degree).

Members of the Thesis Examination Committee were as follows:

Gwendoline Ee Cheng Lian, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Mohamad Zaki Abd Rahman, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Internal Examiner)

Intan Safinar, PhD

Lecturer
Faculty of Science
Universiti Putra Malaysia
(Internal Examiner)

Dato' Laily Din, PhD

Professor
School of Chemical Sciences and Food Technology
Faculty of Science and Technology
Universiti Kebangsaan Malaysia
Malaysia
(External Examiner)

SEOW HENG FONG, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: September 2012

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of Supervisory committee were as follows:

Mohd Aspollah Sukari, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Chairman)

Mawardi Rahmani, PhD

Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

Ahmad Bustaman Abdul, PhD

Associate Researcher
UPM-MAKNA Cancer Research Laboratory
Institute of Bioscience
Universiti Putra Malaysia
(Member)

Dachriyanus, PhD

Professor
Faculty of Pharmacy
Andalas University
Indonesia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently submitted for any other degree at Universiti Putra Malaysia or other institutions.

RAHAYU UTAMI UMAR

Date: 20 September 2012

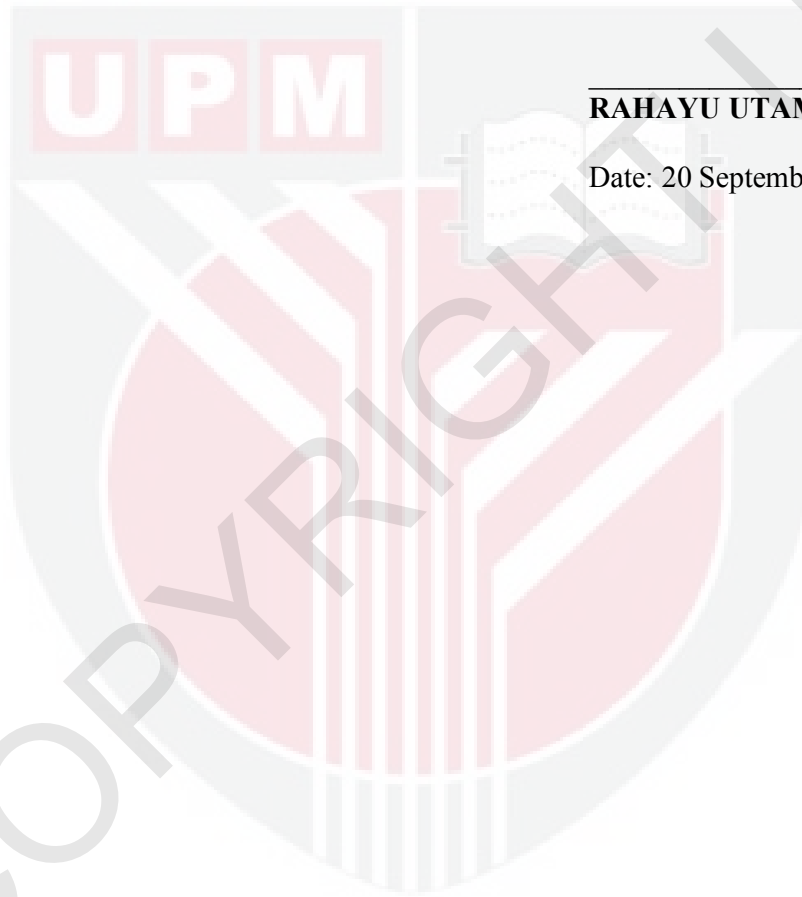


TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xvi
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xix
LIST OF APPENDICES	xx
CHAPTER	
I INTRODUCTION	1
1.1 Background of Research	1
1.2 Problem Statement	3
1.3 Objectives of Research	3
II LITERATURE REVIEW	4
2.1 Genus <i>Elaeocarpus</i>	4
2.1.1 Medicinal uses of genus <i>Elaeocarpus</i>	4
2.1.2 Chemical constituents and biological activities of <i>Elaeocarpus</i>	5
2.1.3 <i>Elaeocarpus floribundus</i> Blume	7
2.2 Genus <i>Barringtonia</i>	8
2.2.1 Medicinal uses of genus <i>Barringtonia</i>	9
2.2.2 Chemical constituents and biological activities of genus <i>Barringtonia</i>	10
2.2.3 <i>Barringtonia conoidea</i> Griff	12
2.3 <i>Fibraurea tinctoria</i> Lour	14
2.3.1 General Information	14
2.3.2 Chemical constituents and biological activities of <i>Fibraurea tinctoria</i>	15
2.4 Total phenolic content determination	19
2.5 Biological activities	20
2.5.1 Antimicrobial activity assay	21
2.5.2 Antioxidant activity assay	22
2.5.3 <i>In vitro</i> cytotoxic assay assay	22
III MATERIAL AND METHODS	24
3.1 Isolation and Purification of Chemical Constituents	24
3.1.1 Materials	24
3.1.2 Instruments	24
3.1.3 Chromatographic methods	26
3.1.4 Extraction and isolation of chemical constituents	27
3.1.4.1 Extraction and isolation constituents of leaves of <i>Elaeocarpus floribundus</i>	27

3.1.4.2	Extraction and isolation of constituents of stem bark of <i>Elaeocarpus floribundus</i>	29
3.1.4.3	Extraction and isolation of constituents of leaves and stem bark of <i>Barringtonia conoidea</i>	30
3.1.4.4	Extraction and isolation of constituents of <i>Fibraurea tinctoria</i>	32
3.1.5	Total phenolic content determination	37
3.1.6	Antimicrobial assay	38
3.1.6.1	Antibacterial assay	39
3.1.6.2	Antifungal assay	39
3.1.7	Antioxidant assay	40
3.1.8	<i>In vitro</i> cytotoxic assay	41
IV	RESULTS AND DISCUSSION	
4.1	Characterization and structural elucidation of isolated compounds	42
4.1.1	Isolated compounds of <i>Elaeocarpus floribundus</i> Blume	42
4.1.1.1	Characterization of friedelin (38)	42
4.1.1.2	Characterization of epifriedelinol (39)	48
4.1.1.3	Characterization of β -sitosterol (40)	54
4.1.2	Isolated compounds of <i>Barringtonia conoidea</i> Griff	60
4.1.2.1	Characterization of mixture of α -(41) and β -amyrin (42)	60
4.1.3	Isolated compounds of <i>Fibraurea tinctoria</i> Lour	68
4.1.3.1	Characterization of clerodanine (43)	68
4.1.3.2	Characterization of 20-hydroxyecdysone (37)	86
4.1.3.3	Characterization of berberine (30)	98
4.2	Total phenolic content determination	112
4.3	Antimicrobial activity	114
4.4	Antioxidant activity	116
4.5	<i>In vitro</i> cytotoxic assay	119
V	CONCLUSION	124
	REFERENCES	127
	APPENDICES	134
	BIODATA OF STUDENT	159
	LIST OF PUBLICATIONS	160

LIST OF TABLES

Table		Page
4.1	Comparison ^1H and ^{13}C NMR of chemical shift of friedelin (38) with the previous data (both in CDCl_3)	44
4.2	Comparison ^1H and ^{13}C NMR of chemical shift of epifriedelinol (39) with the previous data (both in CDCl_3)	49
4.3	Comparison ^1H and ^{13}C NMR of chemical shift of β -sitosterol (40) with the previous data (both in CDCl_3)	56
4.4	Comparison ^1H and ^{13}C NMR of chemical shift of α -amyrin (41) with the previous data (both in CDCl_3)	62
4.5	Comparison ^1H and ^{13}C NMR of chemical shift of β -amyrin (42) with the previous data (both in CDCl_3)	63
4.6	^1H NMR (500 MHz) and ^{13}C NMR (125 MHz) spectral data of clerodanine (43) (in acetone- d_6)	71
4.7	COSY correlations of clerodanine (43)	72
4.8	Comparison ^1H and ^{13}C NMR of chemical Shift of Clerodanine (43) in Acetone- d_6 with the previous data in CDCl_3	72
4.9	Comparison ^1H and ^{13}C NMR of chemical shift of 20-hydroxyecdysone (37) with the previous data (both in CD_3OD)	88
4.10	^1H NMR (500 MHz) and ^{13}C NMR (125 MHz) spectral data of berberine (30) in CD_3OD	101
4.11	Comparison ^1H and ^{13}C NMR of chemical shift of berberine (30) in CD_3OD with the previous data in CDCl_3	102
4.12	The Antibacterial activity of extracts of <i>Barringtonia conoidea</i>	115
4.13	The Antibacterial activity of extracts of <i>Elaeocarpus floribundus</i>	116
4.14	The Antioxidant activity of extracts of <i>Barringtonia conoidea</i>	117
4.15	The Antioxidant activity of extracts of <i>Elaeocarpus floribundus</i>	118
4.16	The Antioxidant activity of extracts of <i>Fibraurea tinctoria</i>	119
4.17	The cytotoxic activity of extracts of <i>Barringtonia conoidea</i> , <i>Elaeocarpus floribundus</i> and <i>Fibraurea tinctoria</i>	121
4.18	The cytotoxic activity of isolated compounds of <i>Barringtonia conoidea</i> , <i>Elaeocarpus floribundus</i> and <i>Fibraurea tinctoria</i>	122

LIST OF FIGURES

Figure		Page
2.1	Alkaloids from <i>Elaeocarpus fuscooides</i>	5
2.2	The isolated compounds from <i>E. lanceofolius</i> and <i>E. japonicas</i>	6
2.3	Sorghumol from fruits of <i>Elaeocarpus chelonimorphus</i>	6
2.4	<i>Elaeocarpus floribundus</i> (A: the whole plant, B: leaves and fruits)	8
2.5	Isolated compounds from seeds and fruits of <i>Barringtonia racemosa</i>	11
2.6	Isolated compounds of leaves of <i>Barringtonia asiatica</i>	12
2.7	<i>Barringtonia conoidea</i> (A; the whole plant, B; fruit and leaves)	13
2.8	<i>Fibraurea tinctoria</i> Lour (A; the whole plant, B; specimen, C; fruits and the male flowers)	15
2.9	Furanoditerpenes and glucosides isolated from <i>Fibraurea tinctoria</i>	18
2.10	The protoberberine alkaloids isolated from <i>Fibraurea tinctoria</i>	18
2.11	The ecdysteroid glucosides from <i>Fibraurea tinctoria</i>	19
4.1	IR spectrum of friedelin (38)	45
4.2	EI-MS spectrum of friedelin (38)	45
4.3	¹ H NMR spectrum of friedelin (38)	46
4.4	¹³ C NMR spectrum of friedelin (38)	47
4.5	IR spectrum of epifriedelinol (39)	51
4.6	EI-MS spectrum of epifriedelinol (39)	51
4.7	¹ H NMR spectrum of epifriedelinol (39)	52
4.8	¹³ C NMR Spectrum of epifriedelinol (39)	53
4.9	IR spectrum of β -sitosterol (40)	57
4.10	EI-MS spectrum of β -sitosterol (40)	57
4.11	¹ H NMR Spectrum of β -sitosterol (40)	58
4.12	¹³ C NMR Spectrum of β -sitosterol (40)	59
4.13	IR spectrum of mixture of α -amyirin (41) and β -amyirin (42)	64
4.14	EIMS spectrum of mixture of α -amyirin (41) and β -amyirin (42)	64
4.15	¹ H NMR spectrum of mixture of α -amyirin (41) and β -amyirin (42)	65
4.16	Expansion ¹ H NMR spectrum of mixture of α -amyirin (41) and β -amyirin (42)	66
4.17	¹³ C NMR spectrum of mixture of α -amyirin (41) and β -amyirin (42)	67
4.18	IR spectrum of clerodanine (43)	73

4.19	EIMS spectrum of clerodanine (43)	73
4.20	¹ H NMR spectrum of clerodanine (43)	74
4.21	Expansion ¹ H NMR spectrum of clerodanine (43)	75
4.22	COSY spectrum of clerodanine (43)	76
4.23	¹³ C NMR spectrum of clerodanine (43)	77
4.24	Expansion ¹³ C NMR spectrum of clerodanine (43)	78
4.25	DEPT spectrum of clerodanine (43)	79
4.26	HMQC spectrum of clerodanine (43)	80
4.27	Expansion HMQC spectrum of clerodanine (43)	81
4.28	HMBC spectrum of clerodanine (43)	82
4.29	Expansion HMBC spectrum of clerodanine (43)	83
4.30	Expansion HMBC spectrum of clerodanine (43)	84
4.31	Expansion HMBC spectrum of clerodanine (43)	85
4.32	IR spectrum of 20-hydroxyecdysone (37)	89
4.33	EI-MS spectrum of 20-hydroxyecdysone (37)	89
4.34	¹ H NMR spectrum of 20-hydroxyecdysone (37)	90
4.35	Expansion ¹ H NMR spectrum of 20-hydroxyecdysone (37)	91
4.36	¹³ C NMR spectrum of 20-hydroxyecdysone (37)	92
4.37	Expansion ¹³ C NMR spectrum of 20-hydroxyecdysone (37)	93
4.38	DEPT spectrum of 20-hydroxyecdysone (37)	94
4.39	HMQC spectrum of 20-hydroxyecdysone (37)	95
4.40	Expansion HMQC spectrum of 20-hydroxyecdysone (37)	96
4.41	Expansion HMQC spectrum of 20-hydroxyecdysone (37)	97
4.42	IR spectrum of berberine (30)	103
4.43	EI-MS spectrum of berberine (30)	103
4.44	¹ H NMR spectrum of berberine (30)	104
4.45	¹³ C NMR spectrum of berberine (30)	105
4.46	DEPT spectrum of berberine (30)	106
4.47	HMQC spectrum of berberine (30)	107
4.48	Expansion HMQC spectrum of berberine (30)	108
4.49	HMBC spectrum of berberine (30)	109
4.50	Expansion HMBC spectrum of berberine (30)	110
4.51	Expansion HMBC spectrum of berberine (30)	111

LIST OF ABBREVIATIONS

α	Alpha
β	Beta
δ	Delta, chemical shift in ppm
μg	Microgram
μl	Microliter
Acetone- d_6	Deuterated Acetone
<i>br</i>	Broad
$^{\circ}\text{C}$	Degree in Celcius
^{13}C	Carbon-13
CDCl_3	Deuterated chloroform
CD_3OD	Deuterated methanol
COSY	Correlation Spectroscopy
cm	centimeter
DEPT	Distortionless Enhancement by Polarization Transfer
DMSO	Dimethylsulfoxide
DW	Dried Weight
<i>d</i>	Doublet
<i>dd</i>	Doublet of doublets
<i>ddd</i>	Doublet of doublets of doublets
EIMS	Electron Impact Mass Spectrum
eV	Electron volt
FTIR	Fourier Transform Infra-Red
GAE	Gallic Acid Equivalence
^1H	Proton
HMBC	Heteronuclear Multiple Bond Correlation
HMQC	Heteronuclear Multiple-Quantum Ceherence
Hz	Hertz
IC	Inhibition concentration
IR	Infrared
<i>J</i>	Coupling in Hz
Lit.	Literature
<i>m</i>	Multiplet
<i>m/z</i>	Mass per charge
MeOH	Methanol
MHz	MegaHertz
m.p.	Melting point
NCI	National Cancer Institute
NMR	Nuclear Magnetic Resonance
OCH_3	Methoxy
OH	Hydroxy
PBS	Phosphate Buffered Saline
ppm	part per million

LIST OF APPENDICES

Appendix		Page
1	Absorbance of gallic acid at wavelength 725 nm	134
2	Data analysis for determination of total phenolic content on studied plants	135
3	DPPH free radical scavenging activities of extracts of leaves of <i>Barringtonia conoidea</i>	137
4	DPPH free radical scavenging activities of extracts of stem bark of <i>Barringtonia conoidea</i>	138
5	DPPH free radical scavenging activities of extracts of leaves of <i>Elaeocarpus floribundus</i>	139
6	DPPH free radical scavenging activities of extracts of stem bark of <i>Elaeocarpus floribundus</i>	140
7	DPPH free radical scavenging activities of extracts of <i>Fibraurea tinctoria</i>	141
8	DPPH free radical scavenging activity of vitamin C	142
9	DPPH free radical scavenging activity of friedelin (38)	143
10	DPPH free radical scavenging activity of epifriedelinol (39)	144
11	DPPH free radical scavenging activity of mixture of α -(41) and β -amyrin (42)	145
12	DPPH free radical scavenging activity of clerodanine (43)	146
13	Cytotoxic effect of extracts of leaves of <i>Barringtonia conoidea</i> against T4-lymphoblastoid cell (CEM-SS).	147
14	Cytotoxic effect of extracts of stem bark of <i>Barringtonia conoidea</i> against T4-lymphoblastoid cell (CEM-SS).	148
15	Cytotoxic effect of extracts of leaves of <i>elaecarpus floribundus</i> against T4-lymphoblastoid cell (CEM-SS).	149
16	Cytotoxic effect of extracts of stem bark of <i>Elaeocarpus floribundus</i> against T4-lymphoblastoid cell (CEM-SS).	150
17	Cytotoxic effect of extracts of <i>Fibraurea tinctoria</i> against T4-lymphoblastoid cell (CEM-SS).	151
18	Cytotoxic effect of friedelin (38), epifriedelinol (39), mixture	152

of α -(**41**) and β -amyrin (**42**) as well as clerodanin (**43**) against T4-lymphoblastoid cell (CEM-SS).

- 19** Cytotoxic effect of extracts of leaves of *Barringtonia conoidea* against Human cervical cancer cell (HeLa). 153
- 20** Cytotoxic effect of extracts of stem bark of *Barringtonia conoidea* against Human cervical cancer cell (HeLa) 154
- 21** Cytotoxic effect of extracts of leaves of *Elaeocarpus floribundus* against Human cervical cancer cell (HeLa). 155
- 22** Cytotoxic effect of extracts of stem bark of *Elaeocarpus floribundus* against Human cervical cancer cell (HeLa). 156
- 23** Cytotoxic effect of extracts of *Fibraurea tinctoria* against Human cervical cancer cell (HeLa). 157
- 24** Cytotoxic effect of friedelin (**38**), epifriedelinol (**39**), mixture of α -(**41**) and β -amyrin (**42**) as well as clerodanin (**43**) against Human cervical cancer cell (HeLa). 158

CHAPTER ONE

INTRODUCTION

1.1 Background of Research

Plant still remains as source of medicines and have been used traditionally for thousands of years. Phytochemical and bioassay studies on medicinal plants revealed that secondary plant metabolites responsible for that useful function. Phenolics, alkaloids, terpenoids, essential oils and others secondary metabolites produced by plants have acted as source for development of many modern medicines. Phenolic compounds were reported to have wide biological activities including anticarcinogenic, antioxidant, antimutagenic and ability modifying the gene expression (Tapiero *et al.*, 2002). Some alkaloid compounds: vincristine, vinblastine, papaverine, codeine and morphine are well known as drug of choice for treating particular diseases such as leukemia, heart disease, analgesic and anesthetic (Vickery and Vickery, 1981). While, promising antiviral and antineoplastic activities that are represented by some terpenoid compounds especially lupane derivatives has become subjects of interest in recent years (Tolstikova *et al.*, 2006).

Elaeocarpus floribundus, *Barringtonia conoidea* and *Fibraurea tinctoria* are three species of plants that have been used as folk medicines in the treatment several diseases. *Elaeocarpus floribundus*, commonly known as medang teja, medang telur (Malay) is a tree that belongs to family Elaeocarpaceae (Burkill *et al.*, 1966). In Sumatera, infusion of its leaves and bark is used as a mouth-wash for inflamed gums

(Burkill *et al.*, 1966). Previous studies reported that alkaloids, flavonoids and terpenoids were successfully isolated from some species of genus *Elaeocarpus*. Some of these isolated compounds have also been found to possess antioxidant, cytotoxic and δ -opioid receptor binding affinity activities (Ray *et al.*, 1976; Cambie *et al.*, 1992; Ito *et al.*, 2002; Katavic *et al.*, 2007; Piao *et al.*, 2009). However, phytochemicals and biological activities of *E. floribundus* has been unexplored yet (Wiar, 2006). Similarly, *Barringtonia conoidea* or putat ayer is a mangrove plant with limited distribution, belonging to family Lecythidaceae. There was no previous study has been reported related to its chemical constituents as well as the bioactivities. Its fruits and young leaves are used to relieve the stomach ache and eaten as fresh vegetables. Research on chemical constituents of other *Barringtonia* species showed that terpenoids were successfully isolated (Subba Rao *et al.*, 1984; Hasan *et al.*, 2000; Gowri *et al.*, 2009).

On the other hand, *Fibraurea tinctoria* is another plant species with interesting medicinal purposes. This plant is belonging to family Menispermaceae, also known as one of four Khaminkhruea species besides *Arcangelisia flava* (L.) Merr., *Coscinium blumeianum* Miers and *C. fenestratum* (Gaertn.) Colebr. The stem and roots of *Fibraurea tinctoria* have been used as bitter tonic and treatment for dysentery, jaundice, diarrhea and skin abscess, for analgesic, antidote and diuretic effects. The chemical investigations have showed the presence of furanoterpenoids, steroids and alkaloids on this species (Ito and Furukawa, 1969; Itokawa *et al.*, 1986; Zakaria *et al.*, 1989). While, the biological studies demonstrated antioxidant, cytotoxic and anti malaria properties of crude extracts as well as anti-inflammatory and antiplasmodial activities of some isolated compounds of *Fibraurea tinctoria*

(Keawpradub *et al.*, 2005; Nguyen-Pouplin *et al.*, 2007; Su *et al.*, 2007; Su *et al.*, 2008).

1.2 Problem Statement

Elaeocarpus floribundus, *Barringtonia conoidea* and *Fibraurea tinctoria* are plant species that have been used as traditional medicines for treating several diseases. Despite all the applications in traditional medicinal practices, phytochemicals and biological activity study on *Elaeocarpus floribundus* and *Barringtonia conoidea* have not been investigated yet. While, medicinal purposes, interesting variety of chemical constituents and significant biological activities of *Fibraurea tinctoria* have prompted us to carry out details investigations on phytochemical and biological activity on this plant.

1.3 Objectives of Research

In view of the medicinal values and interesting chemical constituents of these species, the research was carried out with the following objectives:

1. To extract and isolate the chemical constituents, identify and elucidate the structures of the isolated compounds of *Elaeocarpus floribundus* Blume, *Barringtonia conoidea* Griff and *Fibraurea tinctoria* Lour using chromatographic methods and spectroscopic techniques.
2. To determine the total phenolic contents of the crude extracts
3. To examine the antimicrobial, antioxidant and cytotoxic activities of crude extracts and isolated compounds.

REFERENCES

- Ahmad, F., Ali, M. and Alam, P. (2010). New phytoconstituents from the stem bark of *Tinospora cordifolia* Miers. *Natural Product Research* 24: 926-934.
- Anonymous. (2011a). *Bacillus subtilis*. <http://www.microbewiki.kenyon.edu>. Accessed on 8 December, 2011.
- Anonymous. (2011b). MRSA. <http://www.ncbi.nlm.nih.gov/pubmedhealth>. Accessed on 8 December, 2011.
- Anonymous. (2011c). *Pseudomonas aeruginosa*. http://www.drlera.com/bacterial_diseases. Accessed on 8 December, 2011.
- Anonymous. (2011d). *Salmonella choleraesuis* Infection. <http://www.rightdiagnosis.com>. Accessed on 8 December, 2011.
- Azlim Almey, A. A., Ahmed Jalal Khan, C., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R. and Kamarul Rahim, K. (2010). Total phenolic content and primary antioxidant activity of methanolic and ethanolic extracts of aromatic plants' leaves. *International Food Research Journal* 17: 1077-1084.
- Barker, B. M. and Prescott, F. (1973). Antimicrobial agents in medicine, Blackwell Scientific, London.
- Barros, F. W. A., Bandeira, P. N., Lima, D. J. B., Meira, A. S., de Farias, S. S., Albuquerque, M. R. J. R., Santos, H. I. S. d., Lemos, T. L. G., de Moraes, M. O., Costa-Lotufo, L. V. and Pessoa, C. d. A. (2011). Amyrin esters induce cell death by apoptosis in HL-60 leukemia cells. *Bioorganic & Medicinal Chemistry* 19: 1268-1276.
- Bauer A.W., Kirby W.M., Sherris J.C. and Tenover, M. C. (1966). Antibiotic susceptibility testing by a standardized single disk method. *American Journal of Clinical Pathology* 45: 493-496.
- Biskup, E. and Lojkowska (2009). Evaluation of biological activities of *Rhaponticum carthamoides* extracts. *Journal of Medicinal Plants Research* 3: 1092-1098.
- Blois, M. S. (1958). Antioxidant Determination by the Use of a Stable Free Radical. *Nature* 181: 1199-1200.
- Burkill, I. H., Birtwistle, W., Foxworthy, F. W., Scrivenor, J. B. and Watson, J. G. (1966). A dictionary of the economic products of the Malay peninsula. Kuala Lumpur, Malaysia, Published on behalf of the governments of Malaysia and Singapore by the Ministry of Agriculture and cooperatives.
- Cambie, R. C., Lal, A. R., Rutledge, P. S. and Woodgate, P. D. (1992). Triterpenes from the fruit of *Elaeocarpus chelonimorphus*. *Biochemical Systematics and Ecology* 20: 708-709.

- Carvalho, M. G., Velandia, J. R., Oliveira, L. F. and Bezerra, F. B. (1998). Triterpenos isolados de *Eschweilera longipes* Miers (Lecythidaceae). *Quimica Nova* 21: 740-743.
- Chantaranothai, P. (1995). *Barringtonia* (Lecythidaceae) in Thailand. *Kew Bulletin* 50: 677-694.
- Conforti, F., Statti, G., Uzunov, D. and Menichini, F. (2006). Comparative chemical composition and antioxidant activities of wild and cultivated *Laurus nobilis* L. leaves and *Foeniculum vulgare* subsp. *piperitum* (Ucria) Countinho seeds. *Biological & Pharmaceutical Bulletin* 29: 2056-2064.
- Courtney, J. L. and Gascoigne, R. M. (1956). 414. Triterpenes of the friedelane series. Part I. Ketones. *Journal of the Chemical Society (Resumed)*: 2115-2119.
- Csapi, B., Hajdú, Z., Zupkó, I., Berényi, Á., Forgo, P., Szabó, P. and Hohmann, J. (2010). Bioactivity-guided isolation of antiproliferative compounds from *Centaurea arenaria*. *Phytotherapy Research* 24: 1664-1669.
- Dai, J.-R., Chai, H., Pezzuto, J. M., Kinghorn, A. D., Tsauri, S. and Padmawinata, K. (1993). Cytotoxic constituents of the roots of the Indonesian medicinal plant *Fibraurea chloroleuca*. *Phytotherapy Research* 7: 290-294.
- Dekebo, A., Dagne, E., Gautun, O. R. and Aesen, A. J. (2002). Triterpenes from the resin of *Boswellia neglecta*. *Bulletin of the Chemical Society of Ethiopia* 16: 87-90.
- Gerlier, D. and Thomasset, N. (1986). Use of MTT colorimetric assay to measure cell activation. *Journal of Immunological Methods* 94: 57-63.
- Giesen, W., Wulffraat, S., Zieren, M. and Scholten, L. (2006). Mangrove guidebook for Southeast Asia. Bangkok: Food and Agriculture Organization of the United Nations, Regional Office for Asia and the Pacific. 610-611.
- Girault, J. P. and Lafont, R. (1988). The complete ¹H-NMR assignment of ecdysone and 20-hydroxyecdysone. *Journal of Insect Physiology* 34: 701-706.
- Gowri, P. M., Radhakrishnan, S. V. S., Basha, S. J., Sarma, A. V. S. and Rao, J. M. (2009). Oleanane-Type Isomeric Triterpenoids from *Barringtonia racemosa*. *Journal of Natural Products* 72: 791-795.
- Grycova, L., Dostal, J. and Marek, R. (2007). Quaternary protoberberine alkaloids. *Phytochemistry* 68: 150-175.
- Gunatilaka, A. A. L., Nanayakkara, N. D. and Wazeer, M. I. M. (1983). ¹³C NMR Spectra of some D:A-friedo-oleananes. *Phytochemistry* 22: 991-992.

- Gunatilaka, A. A. L., Nanayakkara, N. P. D., Uvais, M., Sultanbawa, S. and Balasubramaniam, S. (1982). Friedelin, D:A-friedo-olean-3,21-dione and 21 β -hydroxy-D:A-friedo-olean-3-one from *Kokoona zeylanica*. *Phytochemistry* 21: 2061-2063.
- Hada, M., Hino, K. and Takeuchi, Y. (2001). Development of UV Defense Mechanisms during Growth of Spinach Seedlings. *Plant and Cell Physiology* 42: 784-787.
- Hanuman, J. B., Bhatt, R. K. and Sabata, B. K. (1986). A diterpenoid furanolactone from *Tinospora cordifolia*. *Phytochemistry* 25: 1677-1680.
- Hasan, C. M., Khan, S., Jabbar, A. and Rashid, M. A. (2000). Nasimaluns A and B: neo-Clerodane Diterpenoids from *Barringtonia racemosa*. *Journal of Natural Products* 63: 410-411.
- Hernández-Vázquez, L., Palazon, J. and Navarro-Ocaña, A. (2012). The Pentacyclic Triterpenes a, b-amyrins: A Review of Sources and Biological Activities. *Phytochemicals - A Global Perspective of Their Role in Nutrition and Health*. V. Rao, InTech: 487-502.
- Higuchi, C. T., Pavan, F. R. and Leite, C. Q. F. (2008). Triterpenes and antitubercular activity of *Byrsonima crassa*. *Quimica Nova* 31: 1719-1721.
- Hori, T., Kiang, A. K., Nakanishi, K., Sasaki, S. and Woods, M. C. (1967). The structures of fibraurin and a minor product from *Fibraurea chloroleuca*. *Tetrahedron* 23: 2649-2656.
- Huang, M. T., Lee, C. Y. and Ho, C. T. (1992). *Phenolic Compounds in Food and Their Effects on Health: Antioxidants and cancer prevention*, American Chemical Society. New York.
- Ito, A., Chai, H.-B., Lee, D., Kardono, L. B. S., Riswan, S., Farnsworth, N. R., Cordell, G. A., Pezzuto, J. M. and Kinghorn, A. D. (2002). Ellagic acid derivatives and cytotoxic cucurbitacins from *Elaeocarpus mastersii*. *Phytochemistry* 61: 171-174.
- Ito, K. and Furukawa, H. (1969). Structure of fibleucin, a new furanoid diterpene. *Journal of the Chemical Society D: Chemical Communications*: 653-654.
- Itokawa, H., Mizuno, K., Tajima, R. and Takeya, K. (1986). Furanoditerpene glucosides from *Fibraurea tinctoria*. *Phytochemistry* 25: 905-908.
- Janick J and Paull RE (2008). *The encyclopedia of fruit & nuts*. Manoa, Hawaii: CABI. 345-346.
- Janssen, R. H. A. M., Ch. Lousberg, R. J. J., Wijkens, P., Kruk, C. and Theuns, H. G. (1989). Assignment of ^1H and ^{13}C NMR resonances of some isoquinoline alkaloids. *Phytochemistry* 28: 2833-2839.

- Jing, L. J., Mohamed, M., Rahmat, A. and Abu Bakar, M. F. (2010). Phytochemicals, antioxidant properties and anticancer investigations of the different parts of several gingers species (*Boesenbergia rotunda*, *Boesenbergia pulchella* var *attenuata* and *Boesenbergia armeniaca*). *Journal of Medicinal Plants Research* 4: 027-032.
- Joklik WK (1999). *Microbiology: a centenary perspective*. Washington: ASM Press. 40.
- Katavic, P. L., Venables, D. A., Rali, T. and Carroll, A. R. (2007). Indolizidine Alkaloids with μ -Opioid Receptor Binding Affinity from the Leaves of *Elaeocarpus fuscooides*. *Journal of Natural Products* 70: 872-875.
- Keawpradub, N., Dej-adisai, S. and Yuenyongsawad, S. (2005). Antioxidant and cytotoxic activities of Thai medicinal plants named Khaminkhruea: *Arcangelisia flava*, *Coscinium blumeianum* and *Fibraurea tinctoria*. *Songklanakarin Journal of Science and Technology* 27: 455-467.
- Khan, M. R. and Omoloso, A. D. (2002). Antibacterial, antifungal activities of *Barringtonia asiatica*. *Fitoterapia* 73: 255-260.
- Khan, S., Jabbar, A., Hasan, C. M. and Rashid, M. A. (2001). Antibacterial activity of *Barringtonia racemosa*. *Fitoterapia* 72: 162-164.
- Koh, H. L., Kian, C. T. and Tan, C. H. (2009). *A guide to medicinal plants: an illustrated, scientific and medicinal approach*, World Scientific.
- Kundu, J. K., Rouf, A. S. S., Nazmul Hossain, M., Hasan, C. M. and Rashid, M. A. (2000). Antitumor activity of epifriedelanol from *Vitis trifolia*. *Fitoterapia* 71: 577-579.
- Li, L., Huang, X., Sattler, I., Fu, H., Grabley, S. and Lin, W. (2006). Structure elucidation of a new friedelane triterpene from the mangrove plant *Hibiscus tiliaceus*, John Wiley & Sons, Ltd. 44: 624-628.
- Lindley J and Moore T (1866). *The treasury of botany*. London: Longmans, Green. 1102.
- Lu, B., Hu, M., liu, K. and Peng, J. (2010a). Cytotoxicity of berberine on human cervical carcinoma HeLa cells through mitochondria, death receptor and MAPK pathways, and in-silico drug-target prediction. *Toxicology in Vitro* 24: 1482-1490.
- Lu, B., Liu, L., Zhen, X., Wu, X. and Zhang, Y. (2010b). Anti-tumour activity of triterpenoid-rich extract from bamboo shavings (*Caulis bambusae* in Taeniam). *African Journal of Biotechnology* 9: 6430-6436.
- Mahata, S., Bharti, A., Shukla, S., Tyagi, A., Husain, S. and Das, B. (2011). Berberine modulates AP-1 activity to suppress HPV transcription and downstream signaling to induce growth arrest and apoptosis in cervical cancer cells. *Molecular Cancer* 10: 39.

- Mallavadhani, U. V., Mahapatra, A., Jamil, K. and Reddy, P. S. (2004). Antimicrobial activity of some pentacyclic triterpenes and their synthesized 3-O-lipophilic chains. *Biological & Pharmaceutical Bulletin* 27: 1576-1579.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *Journal of Immunological Methods* 65: 55-63.
- Nakanishi, K., Sasaki, S., Kiang, A. K., Goh, J., Kakisawa, H., Ohashi, M., Goto, M., Watanabe, J., Yokotani, H., Matsumura, C. and Togashi, M. (1965). Phytochemical survey of malaysian plants; preliminary chemical and pharmacological screening. *Chemical & Pharmaceutical Bulletin* 13: 882-890.
- Nguyen-Pouplin, J., Tran, H., Tran, H., Phan, T. A., Dolecek, C., Farrar, J., Tran, T. H., Caron, P., Bodo, B. and Grellier, P. (2007). Antimalarial and cytotoxic activities of ethnopharmacologically selected medicinal plants from South Vietnam. *Journal of Ethnopharmacology* 109: 417-427.
- Ogunkoya, L. (1981). Application of mass spectrometry in structural problems in triterpenes. *Phytochemistry* 20: 121-126.
- Otuki, M. F., Ferreira, J., Lima, F. V., Meyre-Silva, C., Malheiros, A., Muller, L. A., Cani, G. S., Santos, A. R. S., Yunes, R. A. and Calixto, J. B. (2005). Antinociceptive properties of mixture of α -Amyrin and β -Amyrin triterpenes: Evidence for participation of protein kinase C and protein kinase A pathways. *The Journal of Pharmacology and Experimental Therapeutics* 313: 310-318.
- Packer, L., Hiramatsu, M. and Yoshikawa, T. (1999). Antioxidant food supplements in human health, Academic Press, California, USA.
- Piao, M. J., Kang, K. A., Zhang, R., Ko, D. O., Wang, Z. H., Lee, K. H., Chang, W. Y., Chae, S., Jee, Y., Shin, T., Park, J. W., Lee, N. H. and Hyun, J. W. (2009). Antioxidant properties of 1,2,3,4,6-penta-O-galloyl-[β]-d-glucose from *Elaeocarpus sylvestris* var. *ellipticus*. *Food Chemistry* 115: 412-418.
- Queiroga, C. L., Silva, G. F., Dias, P. C., Possenti, A. and de Carvalho, J. E. (2000). Evaluation of the antiulcerogenic activity of friedelan-3[β]-ol and friedelin isolated from *Maytenus ilicifolia* (Celastraceae). *Journal of Ethnopharmacology* 72: 465-468.
- Quiroga, E. N., Sampietro, A. R. and Vattuone, M. A. (2001). Screening antifungal activities of selected medicinal plants. *Journal of Ethnopharmacology* 74: 89-96.
- Ragasa, C. Y., Espineli, D. L. and Shen, C. C. (2011). New Triterpenes from *Barringtonia racemosa*. *Chemical & Pharmaceutical Bulletin* 59: 778-782.

- Rahman, M. M., Polfreman, D., MacGeachan, J. and Gray, A. I. (2005). Antimicrobial activities of *Barringtonia acutangula*. *Phytotherapy Research* 19: 543-545.
- Ray, A. B., Dutta, S. C. and Dasgupta, S. (1976). Flavonoids of *Elaeocarpus lanceofolius*. *Phytochemistry* 15: 1797-1798.
- Saha, K., Lajis, N. H., Israf, D. A., Hamzah, A. S., Khozirah, S., Khamis, S. and Syahida, A. (2004). Evaluation of antioxidant and nitric oxide inhibitory activities of selected Malaysian medicinal plants. *Journal of Ethnopharmacology* 92: 263-267.
- Sahoo, S., Panda, P., Mishra, S., Parida, R., Ellaiah, P. and Dash, S. (2008). Antibacterial activity of *Barringtonia acutangula* against selected urinary tract pathogens. *Indian Journal of Pharmaceutical Sciences* 70: 677-679.
- Shahidi, F. (1997). Natural antioxidants: chemistry, health effects, and applications, AOCS Press, United States, pp. 216.
- Singleton, V. L. and Rossi, J. A., Jr. (1965). Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents. *Am. J. Enol. Vitic.* 16: 144-158.
- Sinha, K., Misra, N. P., Singh, J. and Khanuja, S. P. S. (2004). *Tinospora cordifolia* (Guduchi), a reservoir plant for therapeutic applications: A review. *Indian Journal of Traditional Knowledge* 3: 257-270.
- Su, C.-R., Chen, Y.-F., Liou, M.-J., Tsai, H.-Y., Chang, W.-S. and Wu, T.-S. (2008). Anti-inflammatory activities of furanoditerpenoids and other constituents from *Fibraurea tinctoria*. *Bioorganic & Medicinal Chemistry* 16: 9603-9609.
- Su, C.-R., Ueng, Y.-F., Dung, N. X., Vijaya Bhaskar Reddy, M. and Wu, T.-S. (2007). Cytochrome P3A4 Inhibitors and Other Constituents of *Fibraurea tinctoria*. *Journal of Natural Products* 70: 1930-1933.
- Subba Rao, G. S. R., Yadagiri, B., Rao, S. N. and Mallavarapu, G. R. (1984). Anhydrobartogenic acid and 19-epibartogenic acid, two triterpenes from *Barringtonia speciosa*. *Phytochemistry* 23: 2962-2963.
- Suksamrarn, A. and Sommechai, C. (1993). Ecdysteroids from *Vitex pinnata*. *Phytochemistry* 32: 303-306.
- Tapiero, H., Tew, K. D., Nguyen Ba, G. and Mathé, G. (2002). Polyphenols: do they play a role in the prevention of human pathologies? *Biomedecine & Pharmacotherapy* 56: 200-207.
- Thanakijcharoenpath, W. and Theanphong, O. (2007). Triterpenoids from The Stem of *Diospyros glandulosa*. *Thailand Journal of Pharmaceutical Sciences* 31: 1-8.

- Thomas, T. J., Panikkar, B., Subramoniam, A., Nair, M. K. and Panikkar, K. R. (2002). Antitumour property and toxicity of *Barringtonia racemosa* Roxb seed extract in mice. *Journal of Ethnopharmacology* 82: 223-227.
- Tolstikova, T., Sorokina, I., Tolstikov, G., Tolstikov, A. and Flekhter, O. (2006). Biological activity and pharmacological prospects of lupane terpenoids: I. natural lupane derivatives. *Russian Journal of Bioorganic Chemistry* 32: 37-49.
- Tomlinson PB (1995). The botany of mangroves. Cambridge: Cambridge University Press. 251.
- Velioglu, Y. S., Mazza, G., Gao, L. and Oomah, B. D. (1998). Antioxidant Activity and Total Phenolics in Selected Fruits, Vegetables, and Grain Products. *Journal of Agricultural and Food Chemistry* 46: 4113-4117.
- Vickery ML and Vickery B (1981). Secondary plant metabolism. Baltimore: University Park Press
- Vijaya Bharathi, R., Jerad Suresh, A., Thirumal, M., Sriram, L., Geetha Lakshmi, S. and Kumudhaveni, B. (2010). Antibacterial and antifungal screening on various leaf extracts of *Barringtonia acutangula*. *International Journal of Research in Pharmaceutical Sciences* 1: 407-410.
- Wiat, C. (2006). Medicinal plants of the Asia-Pacific: drugs for the future?, Singapore: World Scientific. 87-89.
- Yao, L., Jiang, Y., Datta, N., Singanusong, R., Liu, X., Duan, J., Raymont, K., Lisle, A. and Xu, Y. (2004). HPLC analyses of flavanols and phenolic acids in the fresh young shoots of tea (*Camellia sinensis*) grown in Australia. *Food Chemistry* 84: 253-263.
- Zakaria, M. B., Saito, I., Yao, X.-K., Wang, R.-J. and Matsuura, T. (1989). Furanoditerpenes of *Fibraurea chloroleuca*. *Planta Med* 55: 477,478.