



***DETERMINATION OF CHEMICAL COMPOSITION AND POTENTIAL
ANTICANCER ACTIVITIES OF ALLIUM ATROVIOOLACEUM EXTRACT
ON SELECTED CANCER CELL LINES***

SOMAYEH KHAZAEI

FPSK(p) 2016 2



**DETERMINATION OF CHEMICAL COMPOSITION AND POTENTIAL
ANTICANCER ACTIVITIES OF ALLIUM ATROVIOOLACEUM EXTRACT ON
SELECTED CANCER CELL LINES**

By

SOMAYEH KHAZAEI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirement for the Doctor of Philosophy

September 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

*I dedicate the thesis to the people who without them I wouldn't be able to reach what
I have reached.*

My beloved husband

My dear father

My caring mother





Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of
the requirement for the Doctor of Philosophy

**DETERMINATION OF CHEMICAL COMPOSITION AND POTENTIAL
ANTICANCER ACTIVITIES OF *ALLIUM ATROVIOLACEUM* EXTRACT ON
SELECTED CANCER CELL LINES**

By

SOMAYEH KHAZAEI

September 2016

Chairman: Professor Patimah Ismail, PhD
Faculty: Medicine and Health Sciences

The present study was carried out to determine the phytochemicals and the chemotherapeutic potentials of *Allium atroviolaceum* flower (FA) and bulb (BA) crude extracts in selected reproductive cancer cell lines, human hormone dependent breast cancer (MCF7), human hormone independent breast cancer (MDA-MB-2341), human cervical cancer (HeLa), and human liver cancer (HepG2) and non-malignant murine fibroblast (3T3) cell lines.

The extracts were found to induce anti-proliferative effect on all the studied cells. 70% methanol extracted after 9 hours gave the most promising IC₅₀ values. Exposure of breast, cervical, and liver cancer cell lines to FA and BA extracts demonstrated growth inhibition of cells in both dose- and time-dependent manners. The IC₅₀ value for each cell line at 24, 48, and 72 hours respectively, is given as follows: MCF7 (65, 41, and 24 µg/ml for FA, while 91, 88, and 75 µg/ml for BA), MDA-MB-231 (77, 67, and 51 µg/ml for FA, while 150, 114, and 101 µg/ml for BA), HeLa (68, 51, and 37 µg/ml for FA , while 100, 98, and 74 µg/ml for BA), and HepG2 (57, 44, and 26 µg/ml for FA, while 97, 70, and 58 µg/ml for BA). On the other hand, the IC₅₀ value of normal 3T3 cell line was found to be > 100, indicating that FA and BA extracts were not cytotoxic to normal cells.

In addition, the interaction of FA and BA extracts with tamoxifen (against MCF7 and MDA-MB-231) and doxorubicin (against HeLa and HepG2) revealed a significant dose-reduction in IC₅₀ value.

Phase contrast and fluorescent microscopy (AO/PI) analyses demonstrated apoptotic features after treatment with FA and BA for 24, 48, and 72 hours.

Analyses of DNA content and cell cycle with PI staining confirmed that the time and dose course treatments of FA and BA crude extracts displayed the ability in promoting S phase arrest, G2/M phase arrest, as well as apoptosis in MCF7, MDA-MB-231, and HepG2. However, the event was both time- and dose-dependent, although G0/G1 phase arrest was observed in HepG2 treated with BA in low concentration. Meanwhile, in HeLa cells, the extract enhanced only the percentage of cells present in sub-G0 in both time- and dose-dependent manners.

The Annexin V assay revealed that FA and BA extracts induced cell death by stimulating early apoptosis in low and middle concentrations (IC_{25} and IC_{50}), as well as shorter incubation time (24 and 48 hours), while inducing late apoptosis/necrosis in high concentration (IC_{75}) and longer time course (72 hours) in all the cell lines.

The effect of FA and BA extracts at 24 hours on activity caspases illustrated that the apoptotic effects of FA and BA were via activation of specific inflammatory and initiator caspase, which led to the activation of final effector, caspase-3 or 6. In some cases, increment of caspase-8 and -9 activities indicated that both extrinsic and intrinsic pathways were activated by those extracts that were depended on the type of cell lines. Moreover, the activity of caspase-3 as the main executioner caspase was evaluated with more time course, 48 and 72 hours. As a result, more time of exposure to both FA and BA led to less caspase-3 activity in MCF7 and HeLa, while in MDA-MB-231 and HepG2, longer time incubation resulted in more activities of caspase-3 executioner caspase.

In addition, gene expression analysis using the qPCR conducted on three target genes (*BCL2*, *CDK1*, and *p53*) showed downregulation of anti-apoptotic *BCL2* in MCF7, MDA-MB-231, and HeLa, while only FA-treated HepG2 exhibited enhanced apoptosis. Additionally, upregulation of *p53* only after treatment with BA in MDA-MB-231 and HepG2 illustrated induced apoptosis through a *p53*-dependent mitochondrial pathway, while *p53*-independent mitochondrial pathway in other treated cells. More to the point, *CDK1* was downregulated in MCF7, MDA-MB-231, HeLa, and slightly in HepG2 after exposure to both extracts that might be the main mechanism used by both extracts to exert G2/M cell cycle arrest and ultimately, the final fate of cells, apoptosis.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia Sebagai
memenuhi keperluan untuk Doktor Falsafah

**PENENTUAN KOMPOSISI KIMIA DAN AKTIVITI ANTI KANSER POTENSI
ALLIUM ATROVIOOLACEUM EKSTRAK ON TERPILIH KANSER CELL
LINES**

Oleh

SOMAYEH KHAZAEI

September 2016

Pengerusi: Profesor Patimah Ismail, PhD
Fakulti: Perubatan dan Sains Kesihatan

Kajian ini dijalankan untuk menentukan sifat fitokimia dan evaluasi potensi agen kemoteraputik ekstrak mentah bunga (FA) dan beбаванг (BA) *Allium atroviolaceum* terhadap sel selanjar kanser terpilih iaitu sel selanjar kanser payudara manusia yang bergantung kepada hormon (MCF-7), sel selajar kanser payudara manusia yang tidak bergantung kepada hormon (MDA-MB-2341), sel selanjar kanser pangkal rahim manusia (HeLa) dan sel selanjar kanser hati manusia (HepG2) dan normal 3T3.

Ekstrak didapati merangsang kesan anti-proliferatif ke atas semua sel-sel selanjar kanser yang dikaji. Ekstrak metanol 70% selama 9 jam memberikan nilai IC₅₀ paling menggalakan. Rawatan dan pendedahan ekstrak FA dan BA terhadap sel selanjar kanser payudara, pangkal rahim dan hati menunjukkan perencutan pertumbuhan sel adalah bergantung kepada dos dan tempoh dedahan. Nilai IC₅₀ untuk setiap sel selanjar pada 24, 48, dan 72 jam pendedahan kepada ekstrak adalah seperti berikut: MCF7 (65, 41 dan 24 µg/ml untuk FA., manakala 91, 88 dan 75 µg/ml untuk BA.), MDA-MB-231 (77, 67 dan 51 µg/ml untuk FA., manakala 150, 114 dan 101 µg/ml untuk BA.), HeLa (68, 51 dan 37 µg/ml untuk FA, manakala 100, 98 dan 74 µg/ml untuk BA.), dan HepG2 (57, 44 dan 26 µg/ml untuk FA., manakala 97, 70 dan 58 µg/ml untuk BA.). Malahan, nilai IC₅₀ sel selanjar fibroblast normal murine (3T3) adalah melebihi 100 µg/ml memberi indikasi bahawa esktrak FA dan BA adalah tidak toksik kepada sel-sel normal.

Kajian juga mendapati interaksi ekstrak FA dan BA dengan ubat tamoxifen (terhadap MCF7 dan MDA-MB-231) dan doxorubicin (terhadap HeLa dan HepG2) menunjukkan pengurangan ketara nilai dos IC₅₀ dengan interaksi ubat-ekstrak.

Analisis mikrokop fasa kontras dan mikroskop pendafluor menunjukkan ciri-ciri apoptotik terhadap sel-sel selanjar kanser selepas pendedahan dan rawatan terhadap FA dan BA selama 24, 48 dan 72 jam yang.

Analisis kandungan DNA dan kitaran sel dengan menggunakan pewarnaan PI mengesahkan bahawa masa dan dos rawatan ekstrak mentah FA dan BA menyebabkan pengumpulan sel pada fasa S dan G2/M selain menyebabkan apoptosis pada MCF7, MDA-MB-231 dan HepG2. Walaupun kitaran sel adalah bergantung kepada masa dan dos rawatan, pengumpulan sel pada fasa G0/G1 dapat diperhatikan pada kepekatan dos yang rendah terhadap sel selanjar kanser HepG2 manakala terdapat penambahan peratusan sel pada fasa sub-G0 terhadap sel selajar kanser HeLa.

Analisis Annexin V menunjukkan ekstrak mentah FA dan BA merangsang kematian sel dengan merangsang apoptosis awal pada kepekatan dos rendah dan sederhana (IC_{25} dan IC_{50}) dan juga pada waktu pendedahan yang pendek (24 dan 48 jam). Manakala kepekatan dos yang tigi (IC_{75}) dan tempoh rawatan serta dedahan yang lebih panjang (72 jam) menyebabkan ransangan apoptosis akhir/nekrosis pada semua sel selanjar kanser.

Ekstrak mentah FA dan BA pada tempoh rawatan dan dedahan 24 jam terhadap aktiviti ‘caspase’ menggambarkan bahawa kesan apoptotik yang disebabkan oleh ekstrak mentah FA dan BA. adalah melalui pengaktifan reseptor inflamasi khusus dan pemula ‘caspase’ yang akhirnya menyebabkan pengaktifan ‘caspase’-3 dan -6. Selain itu, dalam beberapa kes, bergantung kepada jenis sel selanjar kanser, peningkatan aktiviti ‘caspase’-8 dan -9 menunjukkan bahawa ekstrak mentah *Allium atroviolaceum* menyebabkan pengaktifan kedua-dua jalur laluan intrinsik dan ektrinsik. Tambahan pula, aktiviti ‘caspase’-3 sebagai ‘caspase’ eksekutor utama dapat diperhatikan pada tempoh rawatan dan dedahan yang lebih panjang iaitu pada 48 dan 72 jam. Selain itu, tempoh rawatan dan dedahan yang lebih panjang terhadap ekstrak mentah FA dan BA menyebabkan pengurangan aktiviti ‘caspase’-3 dalam sel selanjar kanser MCF7 dan HeLa .Namun, dalam sel selanjar kanser MDA-MB-231 dan HepG2 aktiviti ‘caspase’ eksekutor iaitu ‘caspase’-3 menunjukkan peningkatan aktiviti seiring dengan tempoh rawatan dan dedahan.

Analisis gen menggunakan qPCR yang dijalankan ke atas tiga gen sasaran (*BCL2*, *p53*, dan *CDK1*) menunjukkan regulasi menurun pada gen anti-apoptotik *BCL2* pada MCF7, MDA-MB-231 dan HeLa, manakala hanya rawatan dan pendedahan FA. terhadap HepG2 menunjukkan peningkatan apoptosis. Selain itu, regulasi menaik *p53* hanya dapat diperhatikan pada MDA-MB-231 dan HepG2 menunjukan ransangan apoptosis melalui jalur laluan *p53* yang berkaitan dengan mitokondria manakala bagi sel selanjar kanser lain, apoptosis adalah melalui jalur laluan *p53* yang tidak berkaitan dengan mitokondria. Lebih penting lagi regulasi menurun gen *CDK1* dapat diperhatikan pada sel selanjar kanser MCF7, MDA-MB-231, HeLa dan sedikit pada sel selanjar kanser HepG2 selepas rawatan dan dedahan terhadap kedua-dua ekstrak mentah yang mungkin menjadi mekanisma utama yang membawa kepada pengumpulan sel pada fasa G2/M kitaran sel dan akhirnya menyebabkan kematian sel secara apoptosis.

ACKNOWLEDGEMENTS

In the name of God, The Lord of Majesty and Bounty, The Inspirer of Faith. Praised to Him for enlightening my path and surrounding me with wonderful people.

I would like to extend my heartfelt gratitude to my honored supervisor, Prof. Dr. Patimah Ismail as the chairman of my supervisory committee, for her precious advices, support and insightful comments during my study. Her immense knowledge, dedication and integrity have motivated many of us.

In addition, I would like to express my deepest gratefulness to my co-supervisors, Associate professor Dr. Norhaizan binti Mohd.Esa, Associate professor Dr. Roslida Abdul Hamid and Dr.Vasudevan Ramachandran, who patiently supported and encouraged me with their invaluable guidance during the research, despite of the failures. It was a great opportunity for me to work under their supervision.

Furthermore, I would like to extend my appreciation and gratefulness to the Molecular Biology lab staff members that had helped me during my research. And my individual thanks to my friends Dr. Ali Etemad, Farizeh Aalam Ghomi Tabatabaei, Mohammad Arkani, Nur Fasiyah and Maryam Jamilah Yousoff and Maisarah Motalib for being supportive friends and made the lab environment peaceful and organized.

I certify that a Thesis Examination Committee has met on 2 September 2016 to conduct the final examination of Somayeh Khazaei on her thesis entitled "**DETERMINATION OF CHEMICAL COMPOSITION AND POTENTIAL ANTICANCER ACTIVITIES OF ALLIUM ATROVIOLACEUM EXTRACT ON SELECTED CANCER CELL LINES**" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the University Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Member of the Thesis Examination Committee were as follows:

Mohd Nasir Bin Mohd Desa PhD

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

Fauziah Binti Othman, PhD

Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Internal Examiner)

Abdah Binti Md Akim, PhD

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Internal Examiner)

Saeid Amini Nik, PhD

Professor

Department of Surgery

University of Toronto

Canada

(External Examiner)

ZULKARNAIN ZAINAL, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

The thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Patimah Ismail, PhD

Professor

Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Norhaizan Mohd.Esa, PhD

Associate Professor

Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Roslida Abd Hamid, PhD

Associate Professor

Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

R.Vasudevan, PhD

Research Fellow

Institute of Gerontology
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT ,PhD

Professor and Dean

School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

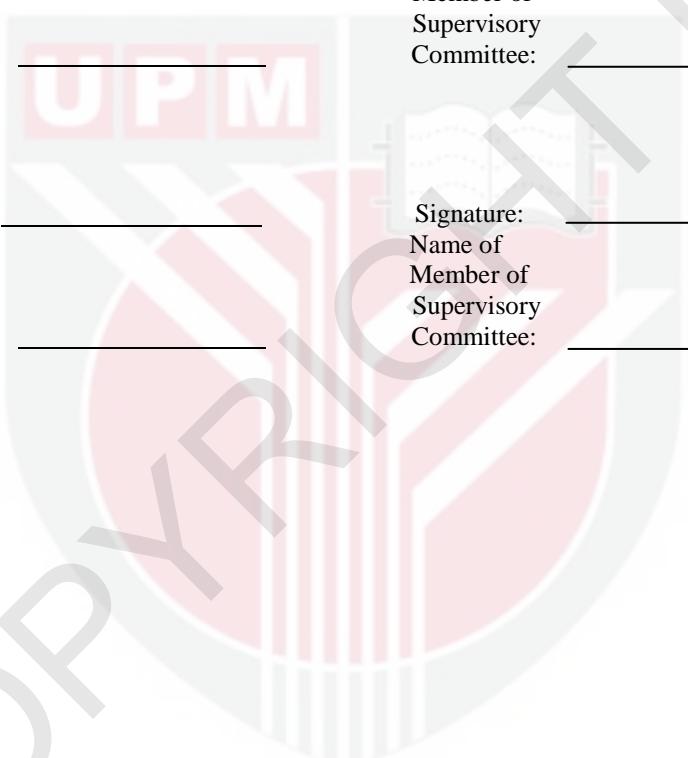
Date: _____

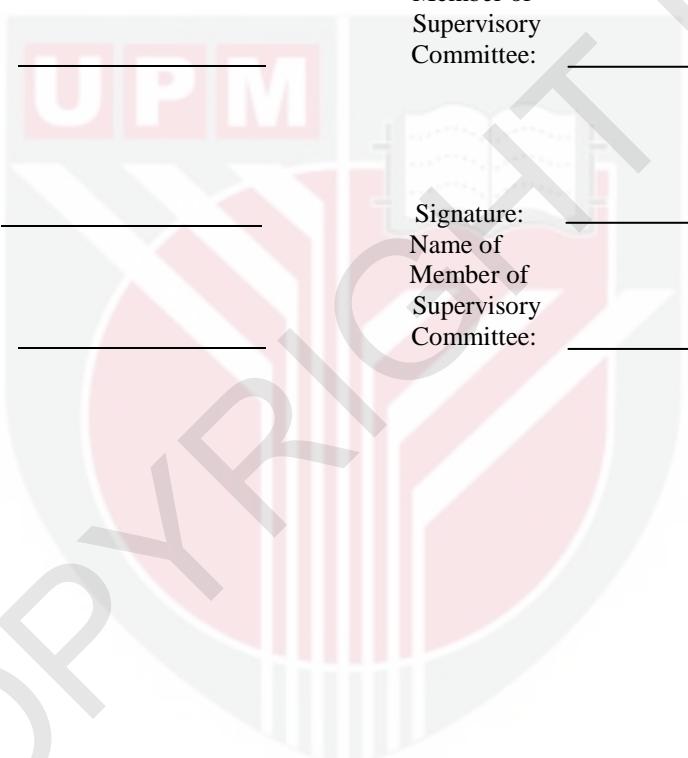
Name and Matric No.: Somayeh Khazaei, GS35022

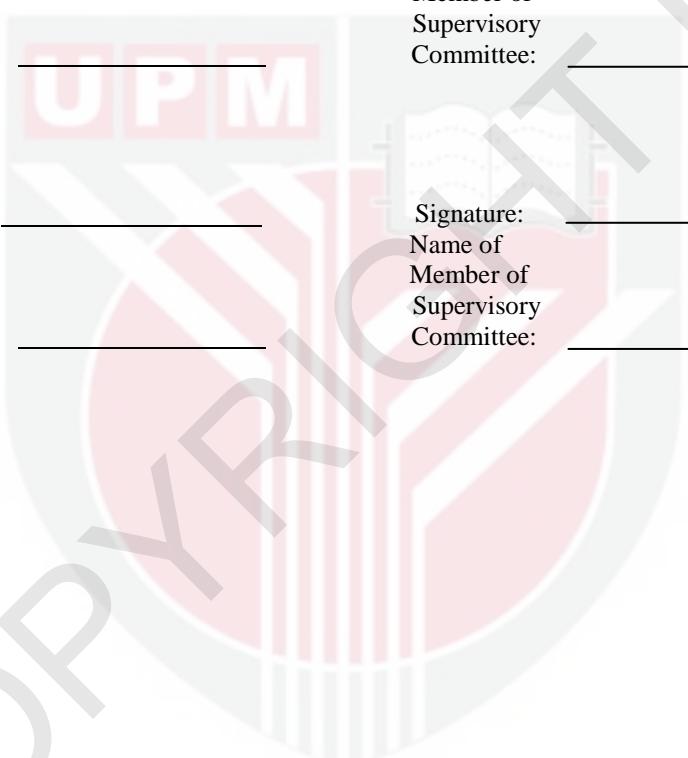
Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of
Chairman of
Supervisory
Committee:


Signature: _____
Name of
Member of
Supervisory
Committee:


Signature: _____
Name of
Member of
Supervisory
Committee:


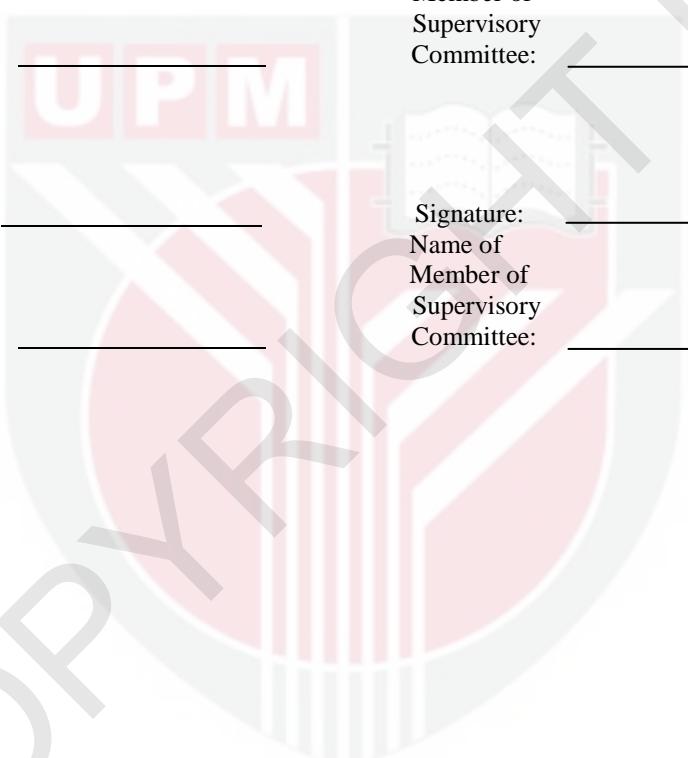
Signature: _____
Name of
Member of
Supervisory
Committee:


TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF APPENDICES	xxi
LIST OF ABBREVIATIONS	xxii
CHAPTER	
1 INTRODUCTION	1
1.1 Background of the Study	1
1.2 Problem Statement	2
1.3 Significance of the Study	3
1.4 Hypothesis	3
1.5 Objectives	3
1.5.1 General Objective	3
1.5.2 Specific Objectives	3
2 LITERATURE REVIEW	5
2.1 An Overview of Cancer	5
2.1.1 Cancer statistic	6
2.1.2 Mechanism of cancer development	7
2.1.3 Metastasis	9
2.2 Chemotherapy	10
2.3 Breast Cancer	11
2.3.1 Breast cancer statistics	11
2.3.2 Breast cancer treatment	12
2.4 Cervical Cancer	13
2.4.1 Cervical cancer statistic	14
2.4.2 Cervical cancer treatment	14
2.5 Hepatocellular Carcinoma (HCC)	15
2.5.1 Hepatocellular carcinoma statistic	16
2.5.2 Hepatocellular carcinoma treatment	17
2.6 Plant Product as Medicine	18
2.6.1 Natural products as chemoprevention drugs	18
2.7 Drug Interaction	19
2.8 Tamoxifen	19
2.9 Doxorubicin	20
2.10 <i>Allium atroviolaceum</i>	21
2.10.1 Morphology and biology	22
2.10.2 Ecology and distribution	23
2.10.3 Utilization and medicinal value	23
2.10.4 Bioactive principles isolated from <i>A.atroviolaceum</i>	24
2.11 Cell Cycle	24
2.11.1 Cell cycle checkpoints	25
2.11.2 Cyclin-cyclin dependent kinase (CDK)	26

2.11.3	Cell cycle and checkpoint maintenance	26
2.12	Cell Death	27
2.12.1	Apoptosis	28
2.12.2	Necrosis	28
2.12.3	Morphological features of apoptosis and necrosis	29
2.12.4	Apoptosis in cancer	29
2.12.5	Mechanisms of apoptosis	30
2.12.6	Death receptor or "extrinsic" apoptosis pathway	30
2.12.7	Mitochondrial or "intrinsic" apoptosis pathway	31
2.13	Caspases	32
2.13.1	Activation of caspase	34
2.13.2	Caspase in cancer	35
2.14	A Genetic Predisposition to Cancer	36
2.14.1	<i>BCL2</i> gene	36
2.14.2	<i>CDK1</i> gene	38
2.14.3	<i>p53</i> gene	40
3	MATERIAL AND METHODS	42
3.1	Sample Preparation	42
3.1.1	Plant materials	42
3.1.2	Preparation of extracts	43
3.1.3	Preparation of FA and BA water extracts	44
3.1.4	Stock solution of plant extracts	44
3.1.5	GC-MS spectroscopy	45
3.2	Cell Cultures	45
3.2.1	Cell lines	45
3.2.2	Cell maintenance	45
3.3	Cytotoxicity Assay	46
3.4	Drug Interaction	47
3.5	Morphological Assessment	47
3.5.1	Phase contrast microscopy	47
3.5.2	Acridin orang (AO) and propidium iodide (PI) double staining and membrane integrity	47
3.6	Cell Cycle Distribution Analysis	48
3.7	Annexin V/ PI Apoptosis Detection Assay	48
3.8	Caspase 1, 2, 3, 5, 6, 8 and 9 Colometric Assay	49
3.9	Gene Expression Analysis by qRT-PCR	50
3.9.1	RNA extraction using RNeasy Mini Kit (50)	50
3.9.2	Measurement of RNA concentration by Nanodrop	51
3.9.3	Measurement of RNA concentration and RIN number by Bioanalyser	51
3.9.4	Complementary DNA (cDNA) synthesis	52
3.9.5	Primer efficiency test	53
3.9.6	Real-Time quantitative RT-PCR (qRT-PCR)	53
3.10	Statistical Analysis	55

4	RESULTS AND DISCUSSIONS	56
4.1	Extraction Yield	56
4.2	Chemical Composition of FA and BA Extracts	58
4.3	Cytotoxicity of FA and BA Extracts	76
4.4	Drug Interaction	89
4.4.1	Synergism, additive or antagonism	93
4.5	Morphological Observation	101
4.6	Fluorescent Microscopy Study	106
4.7	Flow Cytometry Analysis	114
4.7.1	Flow cytometry analysis using PI/RNase	114
4.7.2	Annexin V- PI apoptosis detection assay	129
4.8	Caspase Activity	146
4.8.1	Inflammatory caspases	147
4.8.1.1	Caspase 1	147
4.8.1.2	Caspase 5	150
4.8.2	Initiator caspase	153
4.8.2.1	Caspase 2	153
4.8.2.2	Caspase 9	157
4.8.2.3	Caspase 8	160
4.8.3	Executioner (effector) caspase	163
4.8.3.1	Caspase 6	163
4.8.3.2	Caspase 3	167
4.8.4	Analysis of the caspases activity in each cell line	172
4.9	Gene Expression Analysis of the Apoptosis and Cell Cycle- Related Genes Modulated by FA and BA	176
4.9.1	Determination of RNA quality by Nanodrop and Bioanalyser	177
4.9.2	Expression analysis of <i>BCL2</i> , <i>CDK1</i> and <i>p53</i> with q-Real Time PCR	178
4.9.2.1	The changes in expression levels of corresponding genes in MCF7	179
4.9.2.2	The changes in expression levels of corresponding genes in MDA-MB-231	181
4.9.2.3	The changes in expression levels of corresponding genes in HeLa	184
4.9.2.4	The changes in expression levels of corresponding genes in HepG2	186
5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	194
REFERENCES		196
APPENDICES		234
BIODATA OF STUDENT		274
LIST OF PUBLICATIONS		275

LIST OF TABLES

Table		Page
3.1	Genomic DNA elimination mixture	52
3.2	reverse-transcription mixture	52
3.3	Primers used in Real-Time quantitative PCR analysis	53
3.4	PCR component mixture	54
3.5	Realtime PCR condition	54
4.1	Yield Percentage of FA and BA extracts using 70% and absolute ethanol and methanol	57
4.2	The main compounds identified in FA absolute and aqueous ethanol extracts in different time period	60
4.3	The main compounds identified in FA absolute and aqueous methanol extracts in different time period	63
4.4	Phytocomponents identified in the water extract of FA by GC-MS.	65
4.5	The main compounds identified in BA absolute and aqueous ethanol extracts in different time period	66
4.6	The main compounds identified in BA absolute and aqueous methanol extracts in different time period	69
4.7	Phytocomponents identified in the water extract of BA by GC-MS.	72
4.8	Cytotoxic activity of F.A and B.A by the mean of IC ₅₀ in non-malignant and malignant cell lines.	83
4.9	Cytotoxic activity of doxorubicin and tamoxifen in non-malignant and malignant cell lines.	88
4.10	Activation of caspases by FA and BA extracts in MCF7.	173
4.11	Activation of caspases by FA and BA extracts in MA-MB-231.	174
4.12	Activation of caspases by FA and BA extracts in HeLa.	175
4.13	Activation of caspases by FA and BA extracts in HepG2.	176
4.14	Concentrations, 260/280 ratios and RIN values for selected RNA samples.	178
4.15	Fold change of <i>BCL2</i> , <i>CDK1</i> and <i>p53</i> genes in MCF7 treated with IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA.	181
4.16	Fold change of <i>BCL2</i> , <i>CDK1</i> and <i>p53</i> genes in MDA-MB-231 treated with IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA.	183
4.17	Fold change of <i>BCL2</i> , <i>CDK1</i> and <i>p53</i> genes in HeLa treated with IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA.	186

4.18 Fold change of *BCL2*, *CDK1* and *p53* genes in HepG2 treated with 188
 IC_{25} , IC_{50} and IC_{75} of FA and BA.



LIST OF FIGURES

Figure		Page
2.1	(A) The hallmarks of cancer, (B) Emerging hallmarks and enabling Characteristics	6
2.2	Estimated cancer incidence, mortality, and prevalence worldwide in 2012	7
2.3	A schematic diagram indicates further mutations during progression	9
2.4	The cells of the tumor microenvironment	10
2.5	Mechanisms of action of tamoxifen	20
2.6	Mechanisms of action of doxorubicin	21
2.7	<i>Allium atroviolaceum</i>	22
2.8	Phases of cell cycle	25
2.9	Expression of specific cyclins regulate cell cycle progression.	26
2.10	Hallmarks of the apoptotic and necrotic cell death process.	29
2.11	Schematic representation of the main molecular pathways leading to apoptosis.	31
2.12	Domain structure of human caspases	33
2.13	Structures of processed mammalian caspases and phylogenetic relationship between apoptotic members of the caspase protein family	35
2.14	<i>BCL2</i> family members possess up to four <i>BCL2</i> homology domains (BH1-4) corresponding to α -helical segments (denoted by coloured boxes).	38
2.15	Cyclin-CDK regulation of the mammalian cell cycle	39
2.16	Diagram description of <i>p53</i> activation and up regulation of genes involved with cell cycle arrest (<i>p21</i>) and pro-apoptotic genes (<i>BAX</i>) and the subsequent genes involved in inhibiting <i>p53</i> (<i>MDM2</i>) and anti-apoptotic genes (<i>BCL2</i>)	41
3.1	Overall study design	43
4.1	Antiproliferative activity of FA and BA on MCF7 cell lines	78
4.2	Antiproliferative activity of FA and BA on MDA-MB-231 cell lines	79
4.3	Antiproliferative activity of FA and BA on HeLa cell lines	80
4.4	Antiproliferative activity of FA and BA on HepG2 cell lines	81
4.5	Comparison of anti-proliferative activity of FA and BA on 3T3 normal cell line	82

4.6	Comparison of antiproliferative activity of TAM on MCF7 cell lines	83
4.7	Comparison of antiproliferative activity of TAM on MDA-MB-231 cell lines	84
4.8	Comparison of antiproliferative activity of DOX on HeLa cell lines	85
4.9	Comparison of antiproliferative activity of DOX on HepG2 cell lines	86
4.10	Comparison of antiproliferative activity of TAM and DOX on 3T3 normal cell line	87
4.11	Interaction between TAM, FA and BA in human breast cancer cell (MCF7)	90
4.12	Interaction between TAM, FA and BA in human breast cancer cell (MDA-MB-231)	91
4.13	Interaction between DOX, FA and BA in human cervical cancer cell (HeLa)	92
4.14	Interaction between DOX, FA and BA in human liver cancer cell (HepG2)	92
4.15	The synergistic effect of drug combination in MCF7	94
4.16	The synergistic effect of drug combination in MDA-MB-231	96
4.17	The synergistic effect of drug combination in HeLa	97
4.18	The synergistic effect of drug combination in HepG2	99
4.19	Phase-contrast images to show morphological observation of MCF7 cells	102
4.20	Phase-contrast images to show morphological observation of MDA-MB-231 cells	103
4.21	Phase-contrast images to show morphological observation of HeLa cells	104
4.22	Phase-contrast images to show morphological observation of HepG2 cells	105
4.23	Fluorescence images of treated MCF7 stained with AO/PI	109
4.24	Fluorescence images of treated MDA-MB-231 stained with AO/PI	110
4.25	Fluorescence images of treated HeLa stained with AO/PI	111
4.26	Fluorescence images of treated HepG2 stained with AO/PI	112
4.27	Variation in the percentage of MCF7 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 24 hours	116
4.28	Variation in the percentage of MCF7 cells present in each phase of	117

	the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 48 hours	
4.29	Variation in the percentage of MCF7 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 72 hours	118
4.30	Variation in the percentage of MDA-MB-231 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 24 hours	120
4.31	Variation in the percentage of MDA-MB-231 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 48 hours	121
4.32	Variation in the percentage of MDA-MB-231 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 72 hours	122
4.33	Variation in the percentage of HeLa cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 24 hours	123
4.34	Variation in the percentage of HeLa cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 48 hours	124
4.35	Variation in the percentage of HeLa cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 72 hours	125
4.36	Variation in the percentage of HepG2 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 24 hours	126
4.37	Variation in the percentage of HepG2 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 48 hours	127
4.38	Variation in the percentage of HepG2 cells present in each phase of the cell cycle between untreated cells and cells exposed to different concentration of (A) FA and (B) BA for 72hours	128
4.39	Induction of apoptosis in MCF7 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ concentration for 24 hours	131
4.40	Induction of apoptosis in MCF7 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 48 hours	132
4.41	Induction of apoptosis in MCF7 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 72 hours	133
4.42	Induction of apoptosis in MDA-MB-231 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 24 hours	135
4.43	Induction of apoptosis in MDA-MB-231 after treatment with (A) FA	136

	and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 48 hours	
4.44	Induction of apoptosis in MDA-MB-231 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 72 hours	137
4.45	Induction of apoptosis in HeLa after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 24 hours	139
4.46	Induction of apoptosis in HeLa after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 48 hours	140
4.47	Induction of apoptosis in HeLa after treatment with FA and BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 72 hours	141
4.48	Induction of apoptosis in HepG2 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 24 hours	142
4.49	Induction of apoptosis in HepG2 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 48 hours	143
4.50	Induction of apoptosis in HepG2 after treatment with (A) FA and (B) BA at IC ₂₅ , IC ₅₀ and IC ₇₅ for 72 hours	144
4.51	Effect of FA and BA on caspase-1 activity in MCF7 (A) and MDA-MB-231(B) cells after 24 treatment	148
4.52	Effect of FA and BA on caspase-1 activity in HeLa (A) and HepG2 (B) cells after 24 treatment	149
4.53	Effect of FA and BA on caspase-5 activity in MCF7 (A) and MDA-MB-231(B) cells after 24 treatment	151
4.54	Effect of FA and BA on caspase-5 activity in HeLa (A) and HepG2 (B) cells after 24 treatment	152
4.55	Effect of FA and BA on caspase-2 activity in MCF7 (A) and MDA-MB-231(B) cells after 24 treatment	155
4.56	Effect of FA and BA on caspase-2 activity in HeLa (A) and HepG2 (B) cells after 24 treatment	156
4.57	Effect of FA and BA on caspase-9 activity in MCF7 (A) and MDA-MB-231(B) cells after 24 treatment	158
4.58	Effect of FA and BA on caspase-9 activity in HeLa (A) and HepG2 (B) cells after 24 treatment	159
4.59	Effect of FA and BA on caspase-8 activity in MCF7 (A) and MDA-MB-231(B) cells after 24 treatment	161
4.60	Effect of FA and BA on caspase-8 activity in HeLa (A) and HepG2 (B) cells after 24 treatment	162
4.61	Effect of FA and BA on caspase-6 activity in MCF7 (A) and MDA-MB-231(B) cells after 24 treatment	165
4.62	Effect of FA and BA on caspase-6 activity in HeLa (A) and HepG2 (B) cells after 24 treatment	166

4.63	Effect of FA and BA on caspase-3 activity in MCF7 after 24, 48 and 72 hours treatment	168
4.64	Effect of FA and BA on caspase-3 activity in MDA-MB-231 after 24, 48 a 72 hours treatment	169
4.65	Effect of FA and BA on caspase-3 activity in HeLa after 24, 48 and 72 hours treatment	170
4.66	Effect of FA and BA on caspase-3 activity in HepG2 after 24, 48 and 72 hours treatment	171
4.67	Real-time quantitative PCR analysis illustrates the gene expression in MCF7 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	179
4.68	Real-time quantitative PCR analysis illustrates the gene expression in MCF7 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	180
4.69	Real-time quantitative PCR analysis illustrates the gene expression in MCF7 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	181
4.70	Real-time quantitative PCR analysis illustrates the gene expression in MDA-MB-231 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	182
4.71	Real-time quantitative PCR analysis illustrates the gene expression in MDA-MB-231 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	182
4.72	Real-time quantitative PCR analysis illustrates the gene expression in MDA-MB-231 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	183
4.73	Real-time quantitative PCR analysis illustrates the gene expression in HELA cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	184
4.74	Real-time quantitative PCR analysis illustrates the gene expression in HELA cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	185
4.75	Real-time quantitative PCR analysis illustrates the gene expression in HELA cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	185
4.76	Real-time quantitative PCR analysis illustrates the gene expression in HepG2 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	186
4.77	Real-time quantitative PCR analysis illustrates the gene expression in HepG2 cells after treatment with the IC ₂₅ , IC ₅₀ and IC ₇₅ of FA and BA extracts	187

- 4.78 Real-time quantitative PCR analysis illustrates the gene expression in HepG2 cells after treatment with the IC₂₅, IC₅₀ and IC₇₅ of FA and BA extracts 187

LIST OF APPENDICES

Appendix		Page
A	GC-MS Chromatogram	234
B	DNA frequency histograms	243
C	AnnexinV-FITC	255
D	RNA concentration	267
E	RNA quality	269
F	Quantitative Real Time PCR melting curve	270

LIST OF ABBREVIATIONS

ADP-ribose	DNA repair enzyme poly
ANOVA	One-way Analysis of Variance
Apaf-1	Apoptotic protease activating factor-1
ASR	Age standardized incidence rate
BA	Bulb of <i>Allium atroviolaceum</i>
<i>BAD</i>	<i>BCL2</i> -associated death promoter
<i>BAX</i>	<i>BCL2</i> -associated X protein
BH3	<i>BCL2</i> homology domain
BID	BH3 interacting-domain death agonist
BRCA1	Breast cancer type-1 susceptibility genes
BRCA2	Breast cancer type-1 susceptibility genes
CARD	Caspase recruitment domain
Caspases	Cysteinyl aspartate-specific proteases
CI	Combination index
CDK	Cyclin dependent kinase
cDNA	Complementary DNA
DCIS	Ductal carcinoma <i>in situ</i>
DD	Death domain
DED	Death effector domain
DISC	Death inducing signaling complex
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
E2F	E2 factor transcriptional factor
ER+	Estrogen receptor positive
FA	Flower of <i>Allium atroviolaceum</i>
FADD	Fas-associated death domain
Fas/FasL	Fibroblast-associated cell-surface and Fasligand
FBS	Fetal bovine serum
FDA	Food and drug administration
HBV	Hepatitis B viruses
HCC	Hepatocellular carcinoma
HCV	Hepatitis C viruses
HTS	High-throughput screening
HeLa	Human cervical cancer cell line
HepG2	Human hepatocellular cancer cell line

IC_{50}	Inhibition concentration of 50% of cell death
ICCAM	Interagency Coordinating Committee in the Validation of Alternative Methods
IDC	Invasive ductal carcinoma
ILC	Invasive lobular carcinoma
LACC	Locally advanced cervical cancer
LCIS	Lobular carcinoma <i>in situ</i>
LD	Local destruction
LEEP	Loop electrosurgical excision procedure
LOC	Lab-on-a-chip technology
MCF7	Human hormone-dependent breast cancer cell line
MDA-MB-231	Human non-hormone-dependent breast cancer cell line
MDR	Multi-drug resistance
MMP	Matrix metalloproteinases
MOMP	Mitochondrial outer membrane permeabilization
MPF	Maturation/M-phase promoting factor
mRNA	Messenger ribonucleic acid
MSI	Microsatellite instability
NACT	Neoadjuvant chemotherapy
NCR	National Cancer Registry
NIEHS	National Institute of Environmental Health Science
NIST	National Institute Standard and Technology
OLT	Orthotopic liver transplantation
PARP	Poly(ADP-ribose) polymerase
PBS	Phosphate buffer saline
PIAF regimen	Regimen combination of cisplatin, interferon, doxorubicin and 5- fluorouracil
PS	Phospholipid phosphatidylserine
RFA	Radiofrequency ablation
Rb	Retinoblastoma
RIN	RNA Integrity Number
RNA	Ribonucleic acid
RPMI	Roswell Park Memorial Institute
SD	Standard deviation
SERM	Selective estrogen receptor modulator
SMAC/DIABLO	Pro-apoptotic protein
SPSS	Statistical Package for Social Sciences

TACE	Trans-arterial chemo-embolization
TAM	Tamoxifen
tBID	Truncated BID
TGF	Transforming growth factor
TM	Trans-membrane
TNF	Tumor necrosis factor
TNFR	Tumor necrosis factor receptor
<i>Tp53</i>	Tumour suppressor protein
TRADD	TNFR-associated death domain
TRAIL	Receptors DR-4 and DR-5
WHO	World Health Organization
3t3	Mouse embryonic fibroblast <i>cell line</i>
5-FU	5-Fluorouracil
6-MP	6-Mercaptopurine

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Cancer is a disease of cell growth deregulation due to the reposition of mutations in a single cell that lead to gradual phenotypic changes, from a normal to a neoplastic cell. Loss or mutations of genes can lead to deregulation of abnormal amplification of growth signals and signal transduction pathways that conclusively transform normal cells into invasive cancer cells (Shen *et al.*, 2003).

Cancer is a major health problem worldwide (Yaacob *et al.*, 2010). Statistics illustrate that cancer strikes almost one third of the population (more than 20% of all deaths) (Zaid *et al.*, 2010). Despite considerable advances in treatment and early detection of cancer (Mathers *et al.*, 2008), it remains a big problem for families and economies (Motaal and Shaker, 2011).

Conventional, advanced and alternative treatment has indicated an evolution in cancer therapy (Ricki, 2005). Chemoprevention, that is a means of cancer control or reverse by natural or synthetic agents, is gaining attention (Amin *et al.*, 2009), however, development of resistance to chemotherapeutic drugs prevents effective elimination of the cancer cells. In addition, cardiac and other toxicities are serious side-effects that cause suffering in patients (Yaacob *et al.*, 2010). In addition, the toxicity of anticancer drugs to normal fast-growing cells limits their effectiveness. Resistance of cancerous cells to a specific drug which initially suppressed them is another problem of chemotherapy drugs. Hence, using several drugs in combination may be more effective (Alan, 2008). Furthermore, the physiological and mechanistic deregulations responsible for initiation and promotion of cancer signify hundreds of genes or signaling cascades. Therefore, it appears that multi-target drugs are necessary to overcome cancer. The multiple therapeutic effects of natural compounds in traditional medicine motivate researchers to evaluate the anti-tumor effect of natural compounds and understand their mechanism of action (Teitene *et al.*, 2010).

Natural products are considered powerful sources for novel drug discovery and development (Van Slambrouck *et al.*, 2007).

Phytochemicals, the heterogeneous class of molecules, include vitamins (carotenoids) and food polyphenols, such as flavonoids, phenolic acids and sulfur rich compounds. Biological targets of phytochemicals in mammalian cells are involved in inflammatory processes and oncogenic transformation, for instance variation of cell cycle control, apoptosis evasion and metastases (Russo *et al.*, 2010); therefore, phytochemical agents might have potential as lead compounds for clinical anticancer drugs development.

Allium atroviolaceum (*A.atroviolaceum*) is a species in the genus *Allium*. *Allium* is the largest genus in the widely distributed *Alliaceae* family (Dahlgren *et al.*, 1985). The health benefits of *Allium* species such as garlic (*A.sativum*) and onion (*A.cepa*) have been known for thousands of years; however, interest in other species has been increasing recently (Štajner *et al.*, 2006). The *Alliums* are commonly valued for food, medicine, and garden ornamentals, and range from plants to weeds. Onions, garlic, leek, chives and Welsh onions are some of the most commonly valued *Alliums* (Uhl, 2000).

In this study, crude flower (FA) and bulb (BA) extracts of *A.atroviolaceum* were tested to investigate the anti-proliferation activity of cancer cells such as human hormone-dependent breast cancer (MCF7), human hormone-independent breast cancer (MDA-MB-231), human cervical cancer (HeLa), human liver cancer (HepG2) and also its effect towards normal cells (3T3) were monitored to discover any probable harmful effect on normal cells. A study then was carried out with the aqueous-methanol extract of FA and BA to investigate their potential. The different intracellular mechanisms affected by FA and BA will be detailed hereafter.

Taken together, research in the field of natural products is in high demand to help humans overcome many newly emerging and known diseases, particularly cancers.

1.2 Problem Statement

Malignant cancers are a major cause of death, and they continue to increase. While suitable cures and therapies are able to assist patients to recover at the primary stages, mortality and short survival time of malignant cancers at an advanced stage are inescapable (Bhattacharyya *et al.*, 2010). In addition, chemotherapy has some unavoidable side effects resulting in the lack of optional cytotoxicity (Ahmed *et al.*, 2003) and many existing anticancer drugs share a common mechanism of action that could lead to drug resistance (Alessandro *et al.*, 2007). For these reasons, new agents with new mechanisms need to be developed and tested. Since the use of natural products in cancer chemotherapy have growth significantly, the study of mechanism and mode of action of plant extracts and metabolites become more important. Some components of *Allium* vegetables are reported to block several stages of carcinogenesis, although the underlying mechanisms of action are generally unclear. Association between the consumption of *Allium* vegetables and the risk of cancer has been assessed which point to lower risks for cancers of the stomach, colon, esophagus, and perhaps breast (Sengupta *et al.*, 2004). Moreover, although *A.atroviolaceum* have been used traditionally as medication against various diseases, its medicinal values, particularly against different type of cancer have not been documented yet. This prompted us to investigate the effect of *Allium atroviolaceum* on human breast, cervical and liver cancers.

1.3 Significance of the Study

Natural origin is defined as natural products, derivatives of natural products or synthetic pharmaceuticals based on natural product models (Shoeb, 2006). With only approximately 15% of the world's known plant resources screened for their therapeutic values and over 60% of the currently used anticancer agents derived from natural sources including plants, marine organisms and micro-organisms, it is clear that plants have, and will play, a crucial role in the development of new anti-cancer agents. Due to few medical facilities, low income and cultural and religious beliefs, most of people in developing countries (~80%) use traditional medicines obtained from plants (Boukes and van de Venter, 2011). Natural products medicines are effective in the treatment of specific characteristics while also reducing side effects (Bhadury *et al.*, 2006).

A.atroviolaceum is one of the lesser known species of *Allium*; its pharmaceutical value remains undiscovered. Study of the anticancer effect of *A.atroviolaceum* and understanding of its effects at a molecular level may lead an effective cancer treatment and a promising approach to cancer control.

1.4 Hypothesis

The extract of aerial and underground parts of *A.atroviolaceum* exhibit activity against breast, cervical and liver tumor cells, including a selective cytostatic effect that potentiate use as anticancer drugs. Furthermore, the extract may contain multiple bioactive compounds that could work alone or in combination to restrict cell survival. Moreover, the extracts will demonstrate cytotoxicity towards breast, cervical and liver cancer cells by inducing cell death.

1.5 Objectives

1.5.1 General objective

To investigate the phytochemical and cytotoxic activities of flower and bulb extracts of *A.atroviolaceum* on human breast (MCF7, MDA-MB-231), cervical (HeLa) and liver (HepG2) and non-malignant murine fibroblast (3T3) cell lines.

1.5.2 Specific objectives

1. To identify the phytochemical composition of FA and BA based on the effects of multi-factor experiments.

2. To study the *in vitro* cytotoxic effects of the methanol crude extracts of FA and BA on human hormone-dependent breast cancer (MCF7), human non-hormone-dependent breast cancer cell line (MDA-MB-231), human cervical cancer (HeLa) and human liver cancer (HepG2) and non-malignant murine fibroblast (3T3) cell lines.
3. To evaluate the combination effect of the crude extracts and chemotherapeutic drugs, tamoxifen and doxorubicin.
4. To determine the cell cycle profile of human hormone-dependent breast cancer (MCF7), human non-hormone-dependent breast cancer cell line (MDA-MB-231), human cervical cancer (HeLa) and human liver cancer (HepG2) cell lines induced by FA and BA methanol extracts.
5. To evaluate the anti-proliferative effects and apoptosis induction of FA and BA methanol extracts on human hormone-dependent breast cancer (MCF7), human non-hormone-dependent breast cancer cell line (MDA-MB-231), human cervical cancer (HeLa) and human liver cancer (HepG2) cell lines by using fluorescent microscopy (AO/PI) and flow cytometry (AnexinV).
6. To identify the possible mechanism of cell death pathways by assessing the alteration in activity of caspase1,2,3,5,6,8 and 9 induced by FA and BA methanol extracts on human hormone-dependent breast cancer (MCF7), human non-hormone-dependent breast cancer cell line (MDA-MB-231), human cervical cancer (HeLa) and human liver cancer (HepG2) cell lines.
7. To determine the expression of apoptotic related genes (*BCL2*, *CDK1* and *p53*) by FA and BA methanol extracts on human hormone-dependent breast cancer (MCF7), human non-hormone-dependent breast cancer cell line (MDA-MB-231), human cervical cancer (HeLa) and human liver cancer (HepG2) cell lines.

REFERENCES

- Abdolmohammadi, M. H., Fouladdel, S., Shafiee, A., Amin, G., Ghaffari, S., and Azizi, E. (2009). Antiproliferative and apoptotic effect of *Astrodaucus orientalis* (L.) drude on T47D human breast cancer cell line: potential mechanisms of action. *African Journal of Biotechnology*, 8: 4265-4276.
- Abdul, A. B., Abdel-Wahab, S. I., Fong, H. K., Mohan, S. M., Al-Zubairi, A. S., and Elhassan, M. M. (2009). In vitro response of cancer cells to the growth-inhibitory effects of dichloromethane extract of *Goniothalamus umbrosus*. *Research Journal of Pharmacology*, 3: 1-6.
- Achakzai, A. K. K., Achakzai, P., Masood, A., Kayani, S. A., and Tareen, R. B. (2009). Response of plant parts and age on the distribution of secondary metabolites on plants found in Quetta. *Pakistan Journal of Botany*, 41: 2129-2135.
- Adams, J. M., and Cory, S. (2007). The *BCL2* apoptotic switch in cancer development and therapy. *Oncogene*, 26: 1324-1337.
- Aderonke, S. T., Babatunde, J. A., Adesola, O. T., Okereke, O. U., Innocent, C., Elisha, M. O., and Abiola, M. O. (2013). Evaluation of retinoblastoma (Rb) and protein-53 (*p53*) gene expression levels in breast cancer cell lines (MCF7) induced with some selected cytotoxic plants. *Journal of Pharmacognosy Phytother*, 5: 120-126.
- Adeleye, I. A., Daniels, F. V., and Omadime, M. (2011). Characterization of Volatile Components of Epa-Ijebu: A Native Wonder Cure Recipe. *Journal of Pharmacology and Toxicology*, 6: 97-100.
- Adhikari, D., Zheng, W., Shen, Y., Gorre, N., Ning, Y., Halet, G., and Liu, K. (2012). *CDK1*, but not *CDK2*, is the sole Cdk that is essential and sufficient to drive resumption of meiosis in mouse oocytes. *Human Molecular Genetics*, 21: 2476-2484.
- Afonin, A. N., Greene, S. L., Dzyubenko, N. I., and Frolov, A. N. (2008). Interactive Agricultural Ecological Atlas of Russia and Neighboring Countries. *Economic Plants and Their Diseases, Pests and Weeds*. Retrieved from http://www.agroatlas.ru/files/agroatlas_disk_126434066_1382532724.
- Ahmed B, Al-Rehaily AJ, Al-Howiriny TA, El-Sayed KA and Ahmed MS, 2003. Scopoliosid-D2 and harpagoside-B, two new iridoid glycosides from *Scrophularia deserti* and their antidiabetic and antiinflammatory activity. *Biol. Pharm. Bull.*, 26: 462-467.
- Ahmad, A., Alkarkhi, A. F., Hena, S., and Khim, L. H. (2009). Extraction, separation and identification of chemical ingredients of *Elephantopus scaber* L. using factorial design of experiment. *International Journal of Chemistry*, 1: 36-49.
- Ahmad, A., Alkarkhi, A. F., Hena, S., Siddique, B. M., and Dur, K. W. (2010). Optimization of Soxhlet extraction of Herba Leonuri using factorial design of experiment. *International Journal of Chemistry*, 2: 198-205.

- Ahamed, M., Ali, D., Alhadlaq, H. A., and Akhtar, M. J. (2013). Nickel oxide nanoparticles exert cytotoxicity via oxidative stress and induce apoptotic response in human liver cells (HepG2). *Chemosphere*, 93: 2514-2522.
- Akl, H., Vervloesem, T., Kiviluoto, S., Bittremieux, M., Parys, J. B., De Smedt, H., and Bultynck, G. (2014). A dual role for the anti-apoptotic *BCL2* protein in cancer: Mitochondria versus endoplasmic reticulum. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1843: 2240-2252.
- Akula, R., and Ravishankar, G. A. (2011). Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signaling and Behavior*, 6: 1720-1731.
- Al-Akoum, M., Dodin, S., and Akoum, A. (2007). Synergistic cytotoxic effects of tamoxifen and black cohosh on MCF7 and MDA-MB-231 human breast cancer cells. *Canadian Journal of Physiology and Pharmacology*, 85: 1153-1159.
- Alan, A. H. (2008). Natural products in drug discovery. *Drug Discovery Today*, 13: 894-901.
- Alarcón, T., Byrne, H. M., and Maini, P. K. (2005). A multiple scale model for tumor growth. *Multiscale Modeling and Simulation*, 3: 440-475.
- Alberts, B., Lewis, J., and Bray, D. (2000). *Molecular biology of the cell*. New York: Garland Science.
- Alessandro, N., Poma, P., & Montalto, G. (2007). Multifactorial nature of hepatocellular carcinoma drug resistance: could plant polyphenols be helpful?. *World Journal of Gastroenterology*, 13: 2037-2043.
- Ali, S., and Coombes, R. C. (2002). Endocrine-responsive breast cancer and strategies for combating resistance. *Nature Reviews Cancer*, 2: 101-112.
- Al-Qubaisi, M., Rozita, R., Yeap, S. K., Omar, A. R., Ali, A. M., and Alitheen, N. B. (2011). Selective cytotoxicity of goniothalamin against hepatoblastoma HepG2 cells. *Molecules*, 16: 2944-2959.
- Al-Sha'er, M. A., and Taha, M. O. (2010). Discovery of novel *CDK1* inhibitors by combining pharmacophore modeling, QSAR analysis and in silico screening followed by in vitro bioassay. *European Journal of Medicinal Chemistry*, 45: 4316-4330.
- American Cancer Society. 2010, 1-9. Cervical cancer.
<http://www.cancer.org/acs/groups/cid/documents/webcontent/003094-pdf.pdf>
- American Cancer Society (2011a). Breast Cancer. USA: American Cancer Society, Inc.
- American Cancer Society (2015). Type of chemotherapy drugs. USA: American Cancer Society, Inc.
- Amin, A. R., Kucuk, O., Khuri, F. R., and Shin, D. M. (2009). Perspectives for cancer prevention with natural compounds. *Journal of Clinical Oncology*, 27: 2712-2725.

- Anderson WF, Chatterjee N, Ershler WB, Brawley OW (2002) Estrogen receptor breast cancer phenotypes in the surveillance, epidemiology, and end results database. *Breast Cancer Research and Treatment*, 76: 27-36.
- Angioli, R., Plotti, F., Montera, R., Aloisi, A., Luvero, D., Capriglione, S., ... and Benedetti-Panici, P. (2012). Neoadjuvant chemotherapy plus radical surgery followed by chemotherapy in locally advanced cervical cancer. *Gynecologic Oncology*, 127: 290-296.
- Ara, I., Shinwari, M. M. A., Rashed, S. A., and Bakir, M. A. (2013). Evaluation of antimicrobial properties of two different extracts of Juglans Regia tree bark and search for their compounds using gas chromatography-mass spectrum. *International Journal of Biology*, 5: 92-102.
- Arafa, H. M. (2009). Possible contribution of β -glucosidase and caspases in the cytotoxicity of glufosfamide in colon cancer cells. *European Journal of Pharmacology*, 616: 58-63.
- Arican, G. O., Çakır, O., Arican, E., Kara, T., Dağdeviren, O., and Ari, S. (2014). Effects of Geven root extract on proliferation of HeLa cells and *BCL2* gene expressions. *African Journal of Biotechnology*, 11: 4296-4304.
- Ashford, L., and Collymore, Y. (2005). *Preventing cervical cancer worldwide*. Population Reference Bureau. Retrieved from http://www.prb.org/pdf05/PreventCervCancer-Brief_Eng.
- Ashkenazi, A. (2002). Targeting death and decoy receptors of the tumour-necrosis factor superfamily. *Nature Reviews Cancer*, 2: 420-430.
- Ashkenazi, A. (2008). Targeting the extrinsic apoptosis pathway in cancer. *Cytokine and Growth Factor Reviews*, 19: 325-331.
- Azizi, E., Abdolmohammadi, M. H., Fouladdel, S., Shafiee, A., Amin, G., and Ghaffari, S. M. (2009). Evaluation of *p53* and *BCL2* genes and proteins expression in human breast cancer T47D cells treated with extracts of *Astrodaucus persicus* (Boiss.) Drude in comparison to Tamoxifen. *DARU Journal of Pharmaceutical Sciences*, 17: 181-186.
- Babas, E., Ekonomopoulou, M. T., Karapidaki, I., Doxakis, A., Betsas, G., and Iakovidou-Kritsi, Z. (2010). Indication of participation of caspase-2 and caspase-5 in mechanisms of human cervical malignancy. *International Journal of Gynecological Cancer*, 20: 1381-1385.
- Badmus, J. A., Ekpo, O. E., Hussein, A. A., Meyer, M., and Hiss, D. C. (2015). Antiproliferative and Apoptosis Induction Potential of the Methanolic Leaf Extract of *Holarrhena floribunda* (G. Don). *Evidence-Based Complementary and Alternative Medicine*, 2015.
- Bai, M., Papoudou-Bai, A., Horianopoulos, N., Grepì, C., Agnantis, N. J., and Kanavaros, P. (2007). Expression of *BCL2* family proteins and active caspase 3 in classical Hodgkin's lymphomas. *Human Pathology*, 38: 103-113.

- Bakar, M. F. A., Mohamad, M., Rahmat, A., Burr, S. A., and Fry, J. R. (2010). Cytotoxicity, cell cycle arrest, and apoptosis in breast cancer cell lines exposed to an extract of the seed kernel of Mangifera pajang (bambangan). *Food and Chemical Toxicology*, 48: 1688-1697.
- Balachandran, P., and Govindarajan, R. (2005). Cancer—an ayurvedic perspective. *Pharmacological Research*, 51: 19-30.
- Balachandran, C., Sangeetha, B., Duraipandiyan, V., Raj, M. K., Ignacimuthu, S., Al-Dhabi, N. A., and Arasu, M. V. (2014). A flavonoid isolated from Streptomyces sp.(ERINLG-4) induces apoptosis in human lung cancer A549 cells through p53 and cytochrome c release caspase dependant pathway. *Chemico-biological Interactions*, 224: 24-35.
- Balamurugan, R., Duraipandiyan, V., and Ignacimuthu, S. (2011). Antidiabetic activity of γ -sitosterol isolated from Lippia nodiflora L. in streptozotocin induced diabetic rats. *European Journal of Pharmacology*, 667: 410-418.
- Balamurugan, R., Stalin, A., and Ignacimuthu, S. (2012). Molecular docking of γ -sitosterol with some targets related to diabetes. *European journal of medicinal chemistry*, 47: 38-43.
- Balan, K. V., Demetzos, C., Prince, J., Dimas, K., Cladaras, M., Han, Z., ... and Pantazis, P. (2005). Induction of apoptosis in human colon cancer HCT116 cells treated with an extract of the plant product, Chios mastic gum. *In vivo*, 19: 93-102.
- Baliga, B. C., Read, S. H., and Kumar, S. (2004). The biochemical mechanism of caspase-2 activation. *Cell Death and Differentiation*, 11: 1234-1241.
- Ban, J. O., Hwang, I. G., Kim, T. M., Hwang, B. Y., Lee, U. S., Jeong, H. S., ... and Hong, J. T. (2007). Anti-proliferate and pro-apoptotic effects of 2, 3-dihydro-3, 5-dihydroxy-6-methyl-4H-pyranone through inactivation of NF- κ B in Human Colon Cancer Cells. *Archives of Pharmacal Research*, 30: 1455-1463.
- Bar, T., Sta°hlberg, A., Muszta, A., Kubista, M., 2003. Kinetic outlier detection (KOD) in real-time PCR. *Nucleic Acids Research*, 31: 105-111.
- Benkeblia, N. and Lanzotti, V. (2007). Allium thiosulfinate: chemistry, biological properties and their potential utilization in food preservation. *Food*, 1: 193-201.
- Benmeziane, F., Djamaï, R., Cadot, Y., and Seridi, R. (2014). Optimization of extraction parameters of phenolic compounds from Algerian fresh table grapes,(Vitis Vinifera). *International Food Research Journal*, 21: 1025-1029.
- Bernardo, R. R., Pinto, A. V., and Parente, J. (1996). Steroidal saponins from Smilax officinalis. *Phytochemistry*, 43: 465-469.
- Bernard, A., Domergue, F., Pascal, S., Jetter, R., Renne, C., Faure, J. D., and Joubès, J. (2012). Reconstitution of plant alkane biosynthesis in yeast demonstrates that Arabidopsis ECERIFERUM1 and ECERIFERUM3 are core components of a very-long-chain alkane synthesis complex. *The Plant Cell Online*, 24: 3106-3118.

- Bhatla, N. and Joseph, E. 2009. Cervical cancer prevention and the role of human papillomavirus vaccines in India. *Indian Journal of Medical Research*, 130: 334-340.
- Bian, Z. M., Elner, S. G., Khanna, H., Murga-Zamalloa, C. A., Patil, S., and Elner, V. M. (2011). Expression and functional roles of caspase-5 in inflammatory responses of human retinal pigment epithelial cells. *Investigative Ophthalmology and Visual Science*, 52: 8646-8656.
- Biosciences, B. D. (2010). Caspase-3 Activation-An Indicator of Apoptosis in Image-Based Assays.
- Bismuth, H., Samuel, D., Castaing, D., Adam, R., Saliba, F., Johann, M., ... and Chiche, L. (1995). Orthotopic liver transplantation in fulminant and subfulminant hepatitis. The Paul Brousse experience. *Annals of Surgery*, 222: 109-119.
- Bedolla, D. E., Kenig, S., Mitri, E., Storici, P., and Vaccari, L. (2014). Further insights into the assessment of cell cycle phases by FTIR microspectroscopy. *Vibrational Spectroscopy*, 75: 127-135.
- Bhadury, P., Mohammad, B.T. and Wright P.C. (2006). The current status of natural products from fungi and their potential as anti-effective agents. *Journal of India Microbial Biotechnology*, 33: 325-337.
- Bhattacharyya, S. S., Paul, S., and Khuda-Bukhsh, A. R. (2010). Encapsulated plant extract (*Gelsemium sempervirens*) poly (lactide-co-glycolide) nanoparticles enhance cellular uptake and increase bioactivity in vitro. *Experimental Biology and Medicine*, 235: 678-688.
- Blazquez, S., Sirvent, J. J., Olona, M., Aguilar, C., Pelegri, A., Garcia, J. F., and Palacios, J. (2006). Caspase-3 and caspase-6 in ductal breast carcinoma: a descriptive study. *Histology and Histopathology*, 21: 1321-1329.
- Bodmer, J.L., Holler, N., Reynard, S., Vinciguerra, P., Schneider, P., Juo, P.,... and Tschoopp, J. (2000). TRAIL receptor-2 signals apoptosis through FADD and caspase-8. *Nature Cell Biology*, 2: 241-243.
- Borhani, N., Manoochehri, M., Gargari, S. S., Novin, M. G., Mansouri, A., and Omrani, M. D. (2014). Decreased expression of proapoptotic genes caspase-8-and *BCL2*-associated agonist of cell death (BAD) in ovarian cancer. *Clinical Ovarian and Other Gynecologic Cancer*, 7: 18-23.
- Borup, R., Rossing, M., Henao, R., Yamamoto, Y., Krogdahl, A., Godballe, C., ... and Bennedbæk, F. (2010). Molecular signatures of thyroid follicular neoplasia. *Endocrine-Related Cancer*, 17: 691-708.
- Boukes, G. J., and van de Venter, M. (2011). Cytotoxicity and mechanism (s) of action of Hypoxis spp.(African potato) against HeLa, HT-29 and MCF7 cancer cell lines. *Journal of Medicinal Plants Research*, 5: 2766-74.

- Bowen, I.D., Bowen, S.M., and Jones, A.H. (1998). *Mitosis and apoptosis: Matters of life and death*. London: Chapman and Hall
- Bower, M. and Waxman, J. (2006). *Oncology*. United States of America: Blackwell Publishing.
- Boyle, P., & Ferlay, J. (2005). Cancer incidence and mortality in Europe, 2004. *Annals of Oncology*, 16: 481-488.
- Brady, H. J. (Ed.). (2004). *Apoptosis methods and protocols* (Vol. 282). Totowa, NJ, USA: Humana Press.
- Bremer, E., De Bruyn, M., Wajant, H., and Helfrich, W. (2009). Targeted cancer immunotherapy using ligands of the tumor necrosis factor super-family. *Current Drug Targets*, 10: 94-103.
- Bröker, L. E., Kruyt, F. A., and Giaccone, G. (2005). Cell death independent of caspases: a review. *Clinical Cancer Research*, 11: 3155-3162.
- Brüning, A., Friese, K., Burges, A., and Mylonas, I. (2010). Tamoxifen enhances the cytotoxic effects of nelfinavir in breast cancer cells. *Breast Cancer Research*, 12: 45-55
- Cai, J., Yang, L., Dong, W., Wang, H., Xiong, Z., and Wang, Z. (2015). Retrospective comparison of laparoscopic versus open radical hysterectomy after neoadjuvant chemotherapy for locally advanced cervical cancer. *International Journal of Gynecology and Obstetrics*, 132: 29-33.
- Campbell, N. A., Reece, J. B. and Urry, L. 2005. *Biology*, San Francisco, Calif. London: Pearson Benjamin Cummings.
- Carbone, M., and Pass, H.I. (2004). Multistep and multifactorial carcinogenesis: When does a contributing factor become a carcinogen?. *Seminars in Cancer Biology*, 14:399-405.
- Carpenter, R., and Miller, W. R. (2005). Role of aromatase inhibitors in breast cancer. *British Journal of Cancer*, 93: 1-5.
- Castedo, M., Perfettini, J. L., Roumier, T., and Kroemer, G. (2002). Cyclin-dependent kinase-1: linking apoptosis to cell cycle and mitotic catastrophe. *Cell Death and Differentiation*, 9: 1287-1293.
- Chandra, D. and Tang, D.G., (2003). Mitochondrially localized active caspase-9 and caspase-3 result mostly from translocation from the cytosol and partly from caspase-mediated activation in the organelle. Lack of evidence for Apaf-1-mediated procaspsase-9 activation in the mitochondria. *Journal of Biological Chemistry*, 278: 17408-17420.
- Chang, H.H., Chen, C.S., and Lin, J.Y. (2009). High dose vitamin C supplementation increases the Th1/Th2 cytokine secretion ratio, but decreases eosinophilic infiltration in bronchoalveolar lavage fluid of ovalbumin-sensitized and challenged mice. *Journal of Agricultural and Food Chemistry*, 57: 10471-10476.

- Chang, C. C., Liang, Y. C., Klutz, A., Hsu, C. I., Lin, C. F., Mold, D. E., ... and Huang, R. C. C. (2006). Reversal of multidrug resistance by two nordihydroguaiaretic acid derivatives, M4N and maltose-M3N, and their use in combination with doxorubicin or paclitaxel. *Cancer Chemotherapy and Pharmacology*, 58: 640-653.
- Chew, K.K., Khoo, M.Z., Ng, S.Y., Thoo, Y.Y., Wan Aida, M. and Ho, C.W. (2011). Effect of ethanol concentration, extraction time and extraction temperature on the recovery of phenolic compounds and antioxidant capacity of *Orthosiphon stamineus* extracts. *International Food Research Journal* 18: 1427-1435.
- Chiu, L. C. M., Ho, T. S., Wong, E. Y. L., and Ooi, V. E. (2006). Ethyl acetate extract of *Patrinia scabiosaeifolia* downregulates anti-apoptotic *BCL2/Bcl-X L* expression, and induces apoptosis in human breast carcinoma MCF7 cells independent of caspase-9 activation. *Journal of Ethnopharmacology*, 105: 263-268.
- Cho, J. H., Lee, P. Y., Son, W. C., Chi, S. W., Park, B. C., Kim, J. H., and Park, S. G. (2013). Identification of the novel substrates for caspase-6 in apoptosis using proteomic approaches. *BMB Reports*, 46: 588-593.
- Chong, Y. H., Koh, R. Y., Ling, A. P. K., Chye, S. M., and Yew, M. Y. (2014). Strobilanthes crispus extract induces apoptosis through enhanced caspases activities in cervical cancer cells. In *International Conference on Biological, Environment and Food Engineering (BEFE-2014)* August (pp. 4-5).
- Cunha, G. R., Young, P., Hom, Y. K., Cooke, P. S., Taylor, J. A., and Lubahn, D. B. (1997). Elucidation of a role for stromal steroid hormone receptors in mammary gland growth and development using tissue recombinants. *Journal of Mammary Gland Biology and Neoplasia*, 2: 393-402.
- Çakir, Ö., Pekmez, M., Çepni, E., Candar, B., and Fidan, K. (2014). Evaluation of biological activities of *Physalis peruviana* ethanol extracts and expression of *BCL2* genes in HeLa cells. *Food Science and Technology (Campinas)*, 34: 422-430.
- Carotenuto, A., Fattorusso, E., Lanzotti, V., and Magno, S. (1999). Spirostanol saponins of *Allium porrum* L. *Phytochemistry*, 51: 1077-1082.
- Castellsagué, X., Díaz, M., De Sanjosé, S., Muñoz, N., Herrero, R., Franceschi, S., ... and Meijer, C. J. (2006). International Agency for Research on Cancer Multicenter Cervical Cancer Study Group Worldwide human papillomavirus etiology of cervical adenocarcinoma and its cofactors: implications for screening and prevention. *Journal of the National Cancer Institute*, 98: 303-315.
- Chang, J., Hsu, Y., Kuo, P., Kuo, Y., Chiang, L. and Lin, C. (2005). Increase of Bax/Bcl-XL ratio and arrest of cell cycle by luteolin in immortalized human hepatoma cell line. *Life Sciences*, 76: 1883-1893.
- Chang, C. C., Liang, Y. C., Klutz, A., Hsu, C. I., Lin, C. F., Mold, D. E., ... and Huang, R. C. C. (2006). Reversal of multidrug resistance by two nordihydroguaiaretic acid derivatives, M4N and maltose-M3N, and their use in combination with doxorubicin or paclitaxel. *Cancer Chemotherapy and Pharmacology*, 58: 640-653.

- Cheok, C. F., Dey, A., and Lane, D. P. (2007). Cyclin-dependent kinase inhibitors sensitize tumor cells to nutlin-induced apoptosis: a potent drug combination. *Molecular Cancer Research*, 5: 1133-1145.
- Chen, L., Zhang, X., Chen, J., Zhang, X., Fan, H., Li, S., and Xie, P. (2014). NF- κ B plays a key role in microcystin-RR-induced HeLa cell proliferation and apoptosis. *Toxicon*, 87: 120-130.
- Chen, M., Guerrero, A. D., Huang, L., Shabier, Z., Pan, M., Tan, T. H., and Wang, J. (2007). Caspase-9-induced mitochondrial disruption through cleavage of anti-apoptotic *BCL2* family members. *Journal of Biological Chemistry*, 282: 33888-33895.
- Chen, S., Chen, X., Xiu, Y. L., Sun, K. X., and Zhao, Y. (2015). MicroRNA-490-3P targets *CDK1* and inhibits ovarian epithelial carcinoma tumorigenesis and progression. *Cancer Letters*, 362: 122-130.
- Cheng, Y. T., Ong, A., Lin, C. C., Lin, C. J., Chen, W. T., and Lin, S. M. (2015). Percutaneous radiofrequency ablation for hepatocellular carcinoma: Early termination versus standard termination of ablation procedure. *Advances in Digestive Medicine*, 1: 1-7
- Choene, M. and Motadi, L.R. (2012). Anti-proliferative effects of the methanolic extract of kedrostis foetidissima in breast cancer cell lines. *Molecular Biology*, 1: 107-111.
- Chuang, P. Y., Huang, C., and Huang, H. C. (2013). The use of a combination of tamoxifen and doxorubicin synergistically to induce cell cycle arrest in BT483 cells by down-regulating *CDK1*, *CDK2* and cyclin D expression. *Journal of Pharmaceutical Technology and Drug Research*, 2: 12-19.
- Chuang, S. C., La Vecchia, C., and Boffetta, P. (2009). Liver cancer: descriptive epidemiology and risk factors other than HBV and HCV infection. *Cancer Letters*, 286: 9-14.
- Cowling, V., and Downward, J. (2002). Caspase-6 is the direct activator of caspase-8 in the cytochrome c-induced apoptosis pathway: absolute requirement for removal of caspase-6 prodomain. *Cell Death and Differentiation*, 9: 1046-1056.
- Collins, J. A., Schandl, C. A., Young, K. K., Vesely, J., and Willingham, M. C. (1997). Major DNA fragmentation is a late event in apoptosis. *Journal of Histochemistry and Cytochemistry*, 45: 923-934.
- Colombo, N., Carinelli, S., Colombo, A., Marini, C., Rollo, D., Sessa, C., and ESMO Guidelines Working Group. (2012). Cervical cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Annals of Oncology*, 23: 27-32.
- Cooper, G.M., The Cell: A Molecular Approach. 2nd edition. Sunderland (MA): Sinauer Associates; 2000. The Development and Causes of Cancer. Retrieved from: <http://www.ncbi.nlm.nih.gov/books/NBK9963/>

- Coulombe, B. J. (2012). *Human Breast Cancer Cell Line MDA-MB-231* (Doctoral dissertation, California State University, Sacramento).
- Cummings, J., Willmott, N., Hoey, B. M., Marley, E. S., and Smyth, J. F. (1992). The consequences of doxorubicin quinone reduction in vivo in tumour tissue. *Biochemical Pharmacology*, 44: 2165–2174.
- Cullen, S. P., and Martin, S. J. (2009). Caspase activation pathways: some recent progress. *Cell Death and Differentiation*, 16: 935-938.
- Curtin, J.F. and Cotter, T.G. (2003). Live and let die: regulatory mechanisms in Fas-mediated apoptosis. *Cellular Signalling*, 15: 983-92.
- Cutts, S. M., Rephaeli, A., Nudelman, A., Hmelnitsky, I., and Phillips, D. R. (2001). Molecular basis for the synergistic interaction of adriamycin with the formaldehyde-releasing prodrug pivaloyloxymethyl butyrate (AN-9). *Cancer Research*, 61: 8194-8202.
- Dahlgren, R.M.T., Clifford, H.T. and Yeo, P.F. (1985). *The families of monocotyledonsstructure, evolution and taxonomy*. Germany: Springer-Verlag Berlin Heidelberg.
- Dai, Z. J., Gao, J., Li, Z. F., Ji, Z. Z., Kang, H. F., Guan, H. T., ... and Wang, X. J. (2011). In vitro and in vivo antitumor activity of Scutellaria barbata extract on murine liver cancer. *Molecules*, 16: 4389-4400.
- Danial, N.N., and Korsmeyer, S.J. (2004). Cell death: critical control points. *Cell*, 116: 205-219.
- Darakhshan, S., Bidmeshkipour, A., Khazaei, M., Rabzia, A., and Ghanbari, A. (2013). Synergistic effects of tamoxifen and tranilast on VEGF and MMP-9 regulation in cultured human breast cancer cells. *Asian Pacific Journal of Cancer Prevention*, 14: 6869-6874.
- Darakhshan, S., and Ghanbari, A. (2013). Tranilast enhances the anti-tumor effects of tamoxifen on human breast cancer cells in vitro. *Journal of Biomedical Science*, 20: 76-86.
- Dartsch, D.C., Schaefer, A., Boldt, S., Kolch, W. and Marquardt, H. (2002). Comparison of anthracyclineinduced death of human leukemia cells: programmed cell death versus necrosis. *Apoptosis*, 7: 537-548.
- Degterev, A., Boyce, M., Yuan, J. (2003). A decade of caspases. *Oncogene*, 22: 8543-8567.
- Dehpour, A. A., Babakhani, B., Khazaei, S., and Asadi, M. (2011). Chemical composition of essential oil and antibacterial activity of extracts from flower of *Allium atroviolaceum*. *Journal of Medicinal Plants Research*, 5: 3667-3672.
- Delgado, M. E., Olsson, M., Lincoln, F. A., Zhivotovsky, B., and Rehm, M. (2013). Determining the contributions of caspase-2, caspase-8 and effector caspases to intracellular VDVADase activities during apoptosis initiation and

execution. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1833: 2279-2292.

Deepa, P and Murugesh, S. (2013). GC-MS determination of bioactive compounds of *Dolichandrone Atrovirens* (Sprague) bark. *International Journal of Biology, Pharmesy and Allied Sciences*, 2: 1644-1657

De Vries, A., Flores, E. R., Miranda, B., Hsieh, H. M., van Oostrom, C. T. M., Sage, J., and Jacks, T. (2002). Targeted point mutations of *p53* lead to dominant-negative inhibition of wild-type *p53* function. *Proceedings of the National Academy of Sciences*, 99: 2948-2953.

Diaz-Moralli, S., Tarrado-Castellarnau, M., Miranda, A., and Cascante, M. (2013). Targeting cell cycle regulation in cancer therapy. *Pharmacology and Therapeutics*, 138: 255-271.

Dickson, M. A., and Schwartz, G. K. (2009). Development of cell-cycle inhibitors for cancer therapy. *Current Oncology*, 16: 36-43.

Dijkstra, A. J., Brocke, P. C., and Woldhuis, J. (2006). *WIPO Patent No. 2006091096*. Geneva, Switzerland: World Intellectual Property Organization.

Durling, N. E., Catchpole, O. J., Grey, J. B., Webby, R. F., Mitchell, K. A., Foo, L. Y. and Perry, N. B. (2007). Extraction of phenolics and essential oil from dried sage (*Salvia officinalis*) using ethanol-water mixtures. *Food Chemistry*, 101: 1417-1424.

Duenas-Gonzalez, A., Lopez-Graniel, C., Gonzalez-Enciso, A., Cetina, L., Rivera, L., Mariscal, I., ... and Mohar, A. (2003). A phase II study of multimodality treatment for locally advanced cervical cancer: neoadjuvant carboplatin and paclitaxel followed by radical hysterectomy and adjuvant cisplatin chemoradiation. *Annals of Oncology*, 14: 1278-1284.

Dwivedi, V., Shrivastava, R., Hussain, S., Ganguly, C., and Bharadwaj, M. (2011). Cytotoxic potential of Indian spices (extracts) against esophageal squamous carcinoma cells. *Asian Pacific Journal of Cancer Prevention*, 12: 2069-2073.

Early Breast Cancer Trialists' Collaborative Group. (2005). Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: an overview of the randomised trials. *The Lancet*, 365: 1687-1717.

Edinger, A. L. and Thompson. C. B. (2004). Death by design: apoptosis, necrosis and autophagy. *Current Opinion in Cell Biology*, 16: 663-669.

Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*, 35: 495-516.

Eman Gamal El-Din, E. A. (2011). Extracts of Five Medicinal Herbs Induced Cytotoxicity in Both Hepatoma and Myeloma Cell Lines. *Journal of Cancer Science and Therapy*, 3: 239-243.

Enserink, J. M. and Kolodner, R. D. (2010). An overview of *CDK1*-controlled targets and processes. *Cell Division*, 5: 11-47.

- Erenpreisa, J. and Cragg, M. S. (2007). Cancer: a matter of life cycle?. *Cell Biology International*, 31: 1507-1510.
- Faiao Flores, F., Coelho, P. R., Toledo Arruda Neto, J. D., Maria Engler, S. S., Tiago, M., Capelozzi, V. L., ... and Maria, D. A. (2013). Apoptosis through *BCL2/Bax* and cleaved caspase up-regulation in melanoma treated by boron neutron capture therapy. *PloS One*, 8: 59639-59651.
- Fathy, S. A., Singab, A. N. B., Agwa, S. A., El Hamid, D. M. A., Zahra, F. A., and El Moneim, S. M. A. (2013). The antiproliferative effect of mulberry (*Morus alba L.*) plant on hepatocarcinoma cell line HepG2. *Egyptian Journal of Medical Human Genetics*, 14: 375-382.
- Favaloro, B., Allocati, N., Graziano, V., Di Ilio, C., and De Laurenzi, V. (2012). Role of apoptosis in disease. *Aging (Albany NY)*, 4: 330-349.
- Fearon, E.R. and Bommer, G.T (2008). Progressing from Gene Mutations to Cancer. In: Abeloff, M.D., Armitage, J.O., Lichter, A.S., Niederhuber, J.E., Kastan, M.B. and McKenna, W.G. *Clinical Oncology*. 4th ed. Philadelphia, PA: Elsevier, 207-222.
- Feng, Q., Li, P., Salamanca, C., Huntsman, D., Leung, P. C., and Auersperg, N. (2005). Caspase-1 α is down-regulated in human ovarian cancer cells and the overexpression of caspase-1 α induces apoptosis. *Cancer Research*, 65: 8591-8596.
- Ferlay, J., Bray, F., Pisani, P. and Parkin, D.M. (2004). GLOBOCAN 2002: cancer incidence, mortality and prevalence worldwide. *IARC Cancerbase*, 5, 2.0. Lyon: IARC Press.
- Ferlay, J., Shin, H. R., Bray, F., Forman, D., Mathers, C. and Parkin, D. M. 2010a. Globacon 2008, Cancer incidence and mortality worldwide. *International Agency for Research on Cancer*, 1: 1-8.
- Ferlay, J., Shin, H. R., Bray, F., Forman, D., Mathers, C., and Parkin, D. M. (2010). Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *International Journal of Cancer*, 127: 2893-2917.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., ... and Bray, F. (2015). Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *International Journal of Cancer*, 136: 359-386.
- Ferlini, C., Scambia, G., Marone, M., Distefano, M., Gaggini, C., Ferrandina, G., and Mancuso, S. (1999). Tamoxifen induces oxidative stress and apoptosis in oestrogen receptor-negative human cancer cell lines. *British journal of cancer*, 79: 257-263.
- Fiandalo, M. V., and Kyprianou, N. (2012). Caspase control: protagonists of cancer cell apoptosis. *Experimental Oncology*, 34: 165-175.
- Fieder, J., Wagner, N., Grammatikos, S., Hoffmann, H., Kaufmann, H., and Otto, R. T. (2005). Use of flow-cytometric analysis to optimize cell banking strategies for

production of biopharmaceuticals from mammalian cells. *Journal of Biotechnology*, 120: 111-120

Fleige, S., and Pfaffl, M. W. (2006). RNA integrity and the effect on the real-time qRT-PCR performance. *Molecular Aspects of Medicine*, 27: 126-139.

Foglieni, C., Meoni, C. and Davalli, A., M. (2001). Fluorescent dyes for cell viability: An application on prefixed conditions. *Histochemical Cell Biology*, 115: 223-229.

Francis, S.A., Nelson, J., Liverpool, J., Soogun, S., Mofammere, N. and Thorpe, R.J., Jr. (2011). Examining attitudes and knowledge about HPV and cervical cancer risk among female clinic attendees in Johannesburg, South Africa. *Vaccine*, 28: 8026-8032.

Freidberg, R. (2009). *An investigation into the antimicrobial and anticancer activities of Geranium incanum, Artemisia afra and Artemisia absinthium* (Doctoral dissertation, Nelson Mandela Metropolitan University).

Fulda, S., and Debatin, K. M. (2000). Caspase activation in cancer therapy.

Fulda, S. (2009). Caspase-8 in cancer biology and therapy. *Cancer letters*, 281: 128-133.

Furney, S.J., Higgins, D.G., Ouzounis, C.A., and López-Bigas, N. (2006). Structural and functional properties of genes involved in human cancer. *BMC Genomics*, 7: 3-13.

Gabrielli, B., Brooks, K., and Pavey, S. (2012). Defective cell cycle checkpoints as targets for anti-cancer therapies. *Frontiers in pharmacology*, 3: 9-14.

Gafar, M. K., Itodo, A. U., Warra, A. A., and Abdullahi, L. (2012). Extraction and Physicochemical Determination of Garlic (Allium sativum L) Oil. *International Journal of Food and Nutrition Science*, 1: 4-7.

Garcia-Calvo, M., Peterson, E.P., Rasper, D.M., Vaillancourt, J.P., Zamboni, R., Nicholson, D.W., and Thornberry, N.A. (1999). Purification and catalytic properties of human caspase family members. *Cell Death and Differentiation*, 6: 362-369.

Gardner, R.D. and Burke, D. J. (2000). The spindle checkpoint; two transitions, two pathways. *Trends Cell Biology*. 10: 154-158.

Gerl, R., and Vaux, D. L. (2005). Apoptosis in the development and treatment of cancer. *Carcinogenesis*, 26: 263-270.

Ghavami, S., Hashemi, M., Ande, S. R., Yeganeh, B., Xiao, W., Eshraghi, M., ... and Los, M. (2009). Apoptosis and cancer: mutations within caspase genes. *Journal of medical genetics*, 46: 497-510.

Globocan, I. A. R. C. (2012). Estimated cancer incidence, mortality and prevalence worldwide in 2012. Retrieved from http://globoean.iarc.fr/Pages/fact_sheets_population.aspx.

- Golsteyn, R. M. (2005). *CDK1* and *CDK2* complexes (cyclin dependent kinases) in apoptosis: a role beyond the cell cycle. *Cancer Letters*, 217: 129-138.
- Gosav, S., and Dinică, R. (2011). GC/MS and GC/FTIR as Powerful Tools for Identifying Bioactive Compounds. *Acta Chemica Iasi*, 19: 1-19
- Graham, R. K., Ehrnhoefer, D. E., and Hayden, M. R. (2011). Caspase-6 and neurodegeneration. *Trends in Neurosciences*, 34: 646-656.
- Grasso, S., Gómez-Martínez, Á., Tristante, E., Carrasco-García, E., Martínez-Lacaci, I., Ferragut, J. A., and García-Morales, P. (2012). *Cell Death and Cancer, Novel Therapeutic Strategies*. INTECH Open Access Publisher.
- Green, D. R. (2011). The end and after: how dying cells impact the living organism. *Immunity*, 35: 441-4.
- Greenblatt, M. S., Bennett, W.P., Hollstein, M., and Harris, C. C. (1994). Mutations in the *p53* tumor-suppressor gene - clues to cancer etiology and molecular pathogenesis. *Cancer Research*, 54: 4855-4878.
- Godefroy, N., Foveau, B., Albrecht, S., Goodyer, C. G., and LeBlanc, A. C. (2013). Expression and activation of caspase-6 in human fetal and adult tissues. *PloS one*, 8: 79313-79323.
- Golsteyn, R. M. (2005). *CDK1* and *CDK2* complexes (cyclin dependent kinases) in apoptosis: a role beyond the cell cycle. *Cancer Letters*, 217: 129-138.
- Gottesman, M. M. (2002). Mechanisms of cancer drug resistance. *Annual Review of Medicine*, 53: 615-627.
- Giuliani, F. and Colucci, G. (2010). Treatment of Hepatocellular Carcinoma. *Oncology*, 77: 43-49.
- Green, D. and Kroemer, G. (1998). The central executioners of apoptosis: caspases or mitochondria?. *Trends Cell Biology*, 8: 267-271.
- Grutter, M.G. (2000). Caspases: key players in programmed cell death. *Current Opinion in Structural Biology*, 10: 649-655.
- Guillot, C., Falette, N., Courtois, S., Voeltzel, T., Garcia, E., Ozturk, M., and Puisieux, A. (1996). Alteration of *p53* damage response by tamoxifen treatment. *Clinical Cancer Research*, 2: 1439-1444.
- Guimaraes, D. P., and Hainaut, P. (2002). *TP53*: a key gene in human cancer. *Biochimie*, 84: 83-93.
- Guo, J., and Wang, M. H. (2009). Extract of *Ulmus davidiana* Planch barks induced apoptosis in human hepatoma cell line HepG2. *EXCLI Journal*, 8: 130-137.
- Guo, H., Petrin, D., Zhang, Y., Bergeron, C., Goodyer, C. G., and LeBlanc, A. C. (2006). Caspase-1 activation of caspase-6 in human apoptotic neurons. *Cell Death and Differentiation*, 13: 285-292.

- Guo, Y., Srinivasula, S. M., Druilhe, A., Fernandes-Alnemri, T., and Alnemri, E. S. (2002). Caspase-2 induces apoptosis by releasing proapoptotic proteins from mitochondria. *Journal of Biological Chemistry*, 277: 13430-13437.
- Guo, R., Overman, M., Chatterjee, D., Rashid, A., Shroff, S., Wang, H., and Wang, H. (2014). Aberrant expression of *p53*, p21, cyclin D1, and *BCL2* and their clinicopathological correlation in ampullary adenocarcinoma. *Human Pathology*, 45: 1015-1023.
- Guo, H. M., Sun, Y. M., Zhang, S. X., Ju, X. L., Xie, A. Y., Li, J., ... and Zheng, Y. (2015). Metabolism and pharmacokinetics of 8-hydroxypiperidinylmethyl-baicalein (BA-j) as a novel selective *CDK1* inhibitor in monkey. *Fitoterapia*, 107: 36-43.
- Gupta, S., Radha, V., Furukawa, Y., and Swarup, G. (2001). Direct transcriptional activation of human caspase-1 by tumor suppressor *p53*. *Journal of Biological Chemistry*, 276: 10585-10588.
- Haie-Meder, C., Morice, P., and Castiglione, M. (2010). Cervical cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Annals of Oncology*, 21: 37-40.
- Hamed, M. A., Aly, H. F., Ali, S. A., Metwalley, N. S., Hassan, S. A., and Ahmed, S. A. (2012). In vitro and in vivo assessment of some functional foods against initiation of hepatocellular carcinoma. *Journal of Basic and Applied Scientific Research*, 2: 471-483.
- Han, P., Kang, J. H., Li, H. L., Hu, S. X., Lian, H. H., Qiu, P. P., ... and Chen, Q. X. (2009). Antiproliferation and apoptosis induced by tamoxifen in human bile duct carcinoma QBC939 cells via upregulated *p53* expression. *Biochemical and Biophysical Research Communications*, 385: 251-256.
- Hanahan, D. and Weinberg, R.A. (2000). The hallmarks of cancer. *Cell*, 100: 57-70.
- Hanahan, D. and Weinberg, R. A. (2011). Hallmarks of cancer: the next generation. *Cell*, 144: 646-674.
- Handayani, T., Sakinah, S., Nallappan, M., and Pihie, A. H. L. (2007). Regulation of *p53*-, *BCL2*-and caspase-dependent signaling pathway in xanthorrhizol-induced apoptosis of HepG2 hepatoma cells. *Anticancer Research*, 27: 965-971.
- Hartwell, L. H., and Kastan, M. B. (1994). Cell cycle control and cancer. *Science*, 266: 1821-1828.
- Hawley, T. S. and Hawley, R. S. (2004). *Flow cytometry*. Totowa, New Jersey: Humana Press, 1-33.
- He, L., Wu, L., Su, G., Wei, W., Liang, L., Han, L., and Zhong, M. (2014). The efficacy of neoadjuvant chemotherapy in different histological types of cervical cancer. *Gynecologic Oncology*, 134: 419-425.
- Heinze, T., Jonas, S., Karsten, A., and Neuhaus, P. (1999). Determination of the oncogenes *p53* and C-erb B2 in the tumour cytosols of advanced hepatocellular

- carcinoma (HCC) and correlation to survival time. *Anticancer Research*, 19: 2501-2503.
- Henderson, B. E. and Feigelson, H. S. 2000. Hormonal carcinogenesis. *Carcinogenesis*, 21: 427-33.
- Hengartner, M.O. (2000). The biochemistry of apoptosis (Electronic version). *Nature*, 407, 770-776. Retrieved October 29, 2007 from EBSCOHOST online database
- Henry-Mowatt, J., Dive, C., Martinou, J. C., and James, D. (2004). Role of mitochondrial membrane permeabilization in apoptosis and cancer. *Oncogene*, 23: 2850-2860.
- Hirooka, K., Miyamoto, O., Jinming, P., Du, Y., Itano, T., Baba, T., and Shiraga, F. (2006). Neuroprotective Effects of d-Allose against Retinal Ischemia Reperfusion Injury. *Investigative Ophthalmology and Visual Science*, 47: 1653-1657.
- Hisham, A. N. and Yip, C. H. 2004. Overview of Breast Cancer in Malaysian Women:A Problem with Late Diagnosis. *Asian Journal of Surgery*, 27: 130-133.
- Hofmann, W.K., de Vos, S., Tsukasaki, K., Wachsman, W., Pinkus, G.S., Said, J.W., and Koeffler, H.P. (2001). Altered apoptosis pathways in mantle cell lymphoma detected by oligonucleotide microarray. *Blood*, 98: 787-794.
- Holleman, A., den Boer, M.L., Kazemier, K.M., Beverloo, H.B., von Bergh, A.R., Janka-Schaub, G.E., and Pieters, R. (2005). Decreased PARP and procaspase-2 protein levels are associated with cellular drug resistance in childhood acute lymphoblastic leukemia. *Blood*, 106: 1817-1823.
- Hongo, F., Takaha, N., Oishi, M., Ueda, T., Nakamura, T., Naitoh, Y., ... and Nakayama, S. (2014). CDK1 and CDK2 activity is a strong predictor of renal cell carcinoma recurrence. In *Urologic Oncology: Seminars and Original Investigations*, 32: 1240-1246).
- Hortobagyi, G.N.(1998). Treatment of breast cancer. *New England Journal of Medicine*, 339: 974-984.
- Hossain, M. A., Izuishi, K., Tokuda, M., Izumori, K., and Maeta, H. (2004). D-Allose has a strong suppressive effect against ischemia/reperfusion injury: a comparative study with allopurinol and superoxide dismutase. *Journal of Hepato-biliary-pancreatic Surgery*, 11: 181-189.
- Hu, B., Elinav, E., Huber, S., Booth, C. J., Strowig, T., Jin, C., ... and Flavell, R. A. (2010). Inflammation-induced tumorigenesis in the colon is regulated by caspase-1 and NLRC4. *Proceedings of the National Academy of Sciences*, 107: 21635-21640.
- Hu, C. D., Liang, Y. Z., Guo, F. Q., Li, X. R., and Wang, W. P. (2010). Determination of essential oil composition from Osmanthus fragrans tea by GC-MS combined with a chemometric resolution method. *Molecules*, 15: 3683-3693.

- Hu, N., Qian, L., Hu, Y., Shou, J. Z., Wang, C., Giffen, C., ... & Taylor, P. R. (2006). Quantitative real-time RT-PCR validation of differential mRNA expression of SPARC, FADD, Fascin, COL7A1, CK4, TGM3, ECM1, PPL and EVPL in esophageal squamous cell carcinoma. *BMC Cancer*, 6: 1-5.
- Huang, Y., Pan, G., Øie, S., and Lu, D. R. (2004). Cellular Growth, Development, and Defensive Response. In *Cellular Drug Delivery* (pp. 25-41). Humana Press.
- Huang, S. and He, X. (2011). The role of microRNAs in liver cancer progression. *British Journal of Cancer*, 104: 235-240.
- Izevbegie, E. B. (2003). Discovery of water-soluble anticancer agents (edotides) from a vegetable found in Benin City, Nigeria. *Experimental Biology and Medicine*, 228: 293-298.
- Jain, R., and Jain, S. K. (2011). Screening of in vitro cytotoxic activity of some medicinal plants used traditionally to treat cancer in Chhattisgarh state, India. *Asian Pacific Journal of Tropical Biomedicine*, 1: 147-150.
- Jacob, F., Guertler, R., Naim, S., Nixdorf, S., Fedier, A., Hacker, N. F., and Heinzelmann-Schwarz, V. (2013). Careful selection of reference genes is required for reliable performance of RT-qPCR in human normal and cancer cell lines. *PLoS One*, 8: 59180-59188.
- Jakubowicz-Gil, J., Paduch, R., Piersiak, T., Głowniak, K., Gawron, A., and Kandefer-Szerszeń, M. (2005). The effect of quercetin on pro-apoptotic activity of cisplatin in HeLa cells. *Biochemical Pharmacology*, 69: 1343-1350.
- Jang, W. I., Lin, Z. L., Lee, S. H., Namgoong, S., and Kim, N. H. (2014). A specific inhibitor of CDK1, RO-3306, reversibly arrests meiosis during in vitro maturation of porcine oocytes. *Animal Reproduction Science*, 144: 102-108.
- Jänicke, R.U., Sprengart, M.L., Wati, M.R. and A.G. Porter, 1998. Caspase-3 is required for DNA fragmentation and morphological changes associated with apoptosis. *Journal of Biological Chemistry* 273: 9357-9360.
- Jee, C. D., Lee, H. S., Bae, S. I., Yang, H. K., Lee, Y. M., Rho, M. S., and Kim, W. H. (2005). Loss of caspase-1 gene expression in human gastric carcinomas and cell lines. *International Journal of Oncology*, 26: 1265-1271.
- Jelínek, M., Balušíková, K., Kopperová, D., Němcová-Fürstová, V., Šrámek, J., and Fiedlerová, J. (2013). Caspase-2 is involved in cell death induction by taxanes in breast cancer cells. *Cancer Cell International*, 13: 42-57.
- Jellinger, K.A. (2001). Cell death mechanisms in neurodegeneration. *Journal of Cellular and Molecular Medicine*, 5: 1-17.
- Jemal, A., Bray, F., Center, M. M., Ferlay, J., Ward, E., and Forman, D. (2011). Global cancer statistics. *CA: a cancer journal for clinicians*, 61: 69-90.

- Jeong, J. B., Hong, S. C., Jeong, H. J., and Koo, J. S. (2011). Anti-inflammatory effect of 2-methoxy-4-vinylphenol via the suppression of NF- κ B and MAPK activation, and acetylation of histone H3. *Archives of Pharmacal Research*, 34: 2109-2116.
- Jeong, J. B., and Jeong, H. J. (2010). 2-Methoxy-4-vinylphenol can induce cell cycle arrest by blocking the hyper-phosphorylation of retinoblastoma protein in benzo [a] pyrene-treated NIH3T3 cells. *Biochemical and Biophysical Research Communications*, 400: 752-757.
- Jiang, M., and Milner, J. (2003). *BCL2* constitutively suppresses *p53*-dependent apoptosis in colorectal cancer cells. *Genes and Development*, 17: 832-837.
- Jiang C, Chang M, Wen C, Lin Y, Hsu F, Lee M (2006). Natural products of cosmetics: Analysis of extracts of plants endemic to Taiwan for the presence of tyrosinaseinhibitory, melanin reducing and free radical scavenging activities. *Journal of Food and Drug Analysis*, 14: 346-352.
- Johnson, D.G. and Walker, C.L. (1999). Cyclins and cell cycle checkpoints. *Annual Review of Pharmacology and Toxicology*, 39: 295-312.
- Jordan, V. C. (2008). The rise of raloxifene and the fall of invasive breast cancer. *Journal of the National Cancer Institute*, 100: 831-833.
- Jordan, V. C. (2006). Tamoxifen (ICI46,474) as a targeted therapy to treat and prevent breast cancer. *British Journal of Pharmacology*, 147: 269-276.
- Kalaiselvi, M., Narmadha, R., Ragavendran, P., Raj, A., Sophia, D., Kumar, G. R., and Kalaivani, K. (2011). In vivo simulated in vitro model of Jasminum sambac (Linn.) using mammalian liver slice technique. *Asian Pacific Journal of Tropical Biomedicine*, 1: 216-219.
- Kandaswami, C., Lee, L.T., Lee, P.P., Hwang, J.J., Ke, F.C., Huang, Y.T. and Lee, M.T. (2005). The antitumor activities of flavonoids. *In Vivo*, 19: 895-909.
- Kanduc, D., Mittelman, A., Serpico, R., Sinigaglia, E., Sinha, A. A., Natale, C., ... and Pani, P. A. O. L. O. (2002). Cell death: apoptosis versus necrosis (review). *International Journal of Oncology*, 21: 165-170.
- Katey, M. L., Carsten, T. M., Susan, P. and David, I. (2003). Cell death mechanisms in the human opportunistic pathogencandida albicans. *Journal of Eukaryotic Microbiology*, 50: 685-686.
- Kepp, O., Galluzzi, L., Lipinski, M., Yuan, J., and Kroemer, G. (2011). Cell death assays for drug discovery. *Nature Reviews Drug Discovery*, 10: 221-237.
- Kerr, J.F., Wyllie, A.H., and Currie, A.R. (1972). Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British journal of cancer*, 26: 239-257.
- Khairina, A. N., Fuad, A. F. A., Muhammad, T. T., Ahmad, A., and Faridah, M. (2011). In vitro cytotoxicity screening of Xylocarpus sp. crude extracts against

- human hepatocellular carcinoma cell line (HepG2). *Empowering Science Technology and Innovation towards a Better Tomorrow, UMTAS*, 704-706.
- Khoddami, A., Wilkes, M. A., and Roberts, T. H. (2013). Techniques for analysis of plant phenolic compounds. *Molecules*, 18: 2328-2375.
- Kik, C., Kahane, R., and Gebhardt. (2001). Garlic and health. *Nutrition and Metabolic Cardiovascular Disease*, 11: 57-65.
- Kim, J., Ko, M. E., Nelson, R. A., Arrington, A., Luu, C., Falor, A. E., ... and Singh, G. (2014). Increasing age and survival after orthotopic liver transplantation for patients with hepatocellular cancer. *Journal of the American College of Surgeons*, 218: 431-438.
- King, A.A, Debaun, M.R., Riccardi, V.M. and Gutmann, D.H. (2000). Malignant peripheral nerve sheath tumors in neurofibromatosis. *American Journal of Medical Genetics*, 93: 388-392.
- King, M. C., Marks, J. H., Mandell, J.B., and New York Breast Cancer Study G (2003). Breast and ovarian cancer risks due to inherited mutations in BRCA1 and BRCA2. *Science*, 302: 643-646.
- Kirschner, B., Poll, S., Rygaard, C., Wahlin, A., and Junge, J. (2011). Screening history in women with cervical cancer in a Danish population-based screening program. *Gynecol Oncol*, 120: 68-72.
- Kits, A., and Home, B. (2012). Caspase-3 Activation-An Indicator Of Apoptosis In Image-Based Assays.
- Klink, M. (Ed.). (2014). *Interaction of immune and cancer cells*. Springer Vienna.
- Koff, J. L., Ramachandiran, S., and Bernal-Mizrachi, L. (2015). A time to kill: targeting apoptosis in cancer. *International Journal of Molecular Sciences*, 16: 2942-2955.
- Kondo, Y., Kanzawa, T., Sawaya, R., and Kondo, S. (2005). The role of autophagy in cancer development and response to therapy. *Nature Reviews Cancer*, 5: 726-734.
- Kleinsmith, L.J. (2006). *Principles of cancer biology*. United States of America: Pearson Education Inc.
- Klaiman, G., Champagne, N., and LeBlanc, A. C. (2009). Self-activation of Caspase-6 in vitro and in vivo: Caspase-6 activation does not induce cell death in HEK293T cells. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1793: 592-601.
- Kroemer, G., El-Deiry, W. S., Golstein, P., Peter, M. E., Vaux, D., Vandenebeele, P., ... and Piacentini, M. (2005). Classification of cell death: recommendations of the Nomenclature Committee on Cell Death. *Cell Death and Differentiation*, 12: 1463-1467.

- Kruidering, M., and Evan, G. I. (2000). Caspase-8 in Apoptosis: The Beginning of "The End"? *IUBMB life*, 50: 85-90.
- Kumar, Y., Radha, V., and Swarup, G. (2008). Gene Section. <http://AtlasGeneticsOncology.org>, 268.
- Kummalue, T., Pornchai, O., Jiratchariyakul, W., Chanchai, M., Pattanapanyasat, K., Sukapirom, K., and Iemsri, S. (2007). Antiproliferative effect of Erycibe elliptilimba on human breast cancer cell lines. *Journal of Ethnopharmacology*, 110: 439-443.
- Kumar, N., Afeyan, R., Kim, H.D., Lauffenburger, D.A. (2008). Multipathway model enables prediction of kinase inhibitor cross-talk effects on migration of Her2-overexpressing mammary epithelial cells. *Molecular Pharmacology*, 73: 1668-1678.
- Lai, H. C., Chang, C. J., Yang, C. H., Hsu, Y. J., Chen, C. C., Lin, C. S., ... and Lu, C. C. (2012). Activation of NK cell cytotoxicity by the natural compound 2, 3-butanediol. *Journal of Leukocyte Biology*, 92: 807-814.
- Kits, S. A., Masud, A., Kuida, K., Porter, G. A., Booth, C. J., Mehal, W. Z., ... and Flavell, R. A. (2006). Caspases 3 and 7: key mediators of mitochondrial events of apoptosis. *Science*, 311: 847-851.
- Lam, M. (2012). *Cytotoxic activity of Fagonia cretica against human breast cancer cells* (Doctoral dissertation, Aston University).
- Lamkanfi, M., and Kanneganti, T. D. (2010). Caspase-7: a protease involved in apoptosis and inflammation. *The international journal of biochemistry and cell biology*, 42: 21-24.
- Lehn, S. (2013). Cell cycle deregulation in breast cancer subgroups and effects on proliferation, migration and tamoxifen resistance. *Lund University, Faculty of Medicine Doctoral Dissertation Series*, 2013(13).
- Leenders, M.W.H., Nijkamp, M.W. and Rinkes, I.H.M.B. (2008). Mouse models in liver cancer research: A review of current literature. *World Journal of Gastroenterology*, 14: 6915-6923.
- Leggett, D. C., Jenkins, T. F., and Miyares, P. H. (1990). Salting-out solvent extraction for preconcentration of neutral polar organic solutes from water. *Analytical Chemistry*, 62: 1355-1356.
- Leung, T.W T., Patt, Y Z., Lau, W., Ho, S K W., Yu, S.C.H., Chan, A.T.C.,... and Leung, N.W.Y. (1999). Complete pathological remission is possible with systemic combination chemotherapy for inoperable hepatocellular carcinoma. *Clinical Cancer Research*, 5: 1676-1681.
- Levine, E. G. Bloomfield, C. D. (1992). Leukemias and myelodysplastic syndromes secondary to drug, radiation and environmental exposure. *Seminars in Oncology*, 19: 47-84.

- Li, P., Nijhawan, D., Budihardjo, I., Srinivasula, S.M., Ahmad, M., Alnemri, E.S. and Wang, X. (1997). Cytochrome c and dATP-dependent formation of Apaf-1/caspase-9 complex initiates an apoptotic protease cascade. *Cell*, 91: 479-489.
- Li, Y., and Martin, R. C. (2011). Herbal medicine and hepatocellular carcinoma: applications and challenges. *Evidence-Based Complementary and Alternative Medicine*, 2011: 44-58
- Li, S., Cha, I. H., and Nam, W. (2011). Chios mastic gum extracts as a potent antitumor agent that inhibits growth and induces apoptosis of oral cancer cells. *Asian Pacific Journal of Cancer Prevention*, 12: 1877-80.
- Liang, Y. Y., Arnold, T., Michlmayr, A., Rainprecht, D., Perticevic, B., Spittler, A., and Oehler, R. (2014). Serum-dependent processing of late apoptotic cells for enhanced efferocytosis. *Cell Death and Disease*, 5: 210-218.
- Liang, Y., Yan, C., and Schor, N. F. (2001). Apoptosis in the absence of caspase 3. *Oncogene*, 20: 6570-6578
- Lightfoot, S. (2002). Quantitation comparison of total RNA using the Agilent 2100 bioanalyzer, ribogreen analysis, and UV spectrometry. *Agilent Application Note, Publication number*, 5988-7650
- Li-Hua, S., Chun-Mei, L., Zhoa-Yang, Y., De-hai, C., Jing-Yan, C. and Yan, Y. (2012). Lutfu echinata Roxb. induces human colon cancer cells (HT-29) death by triggering the mitochondrial apoptosis pathway. *Molecules*, 17: 5780-5794.
- Lim, S. W., Loh, H. S., Ting, K. N., Bradshaw, T. D., & Zeenathul, N. A. (2014). Antiproliferation and induction of caspase-8-dependent mitochondria-mediated apoptosis by β -tocotrienol in human lung and brain cancer cell lines. *Biomedicine & Pharmacotherapy*, 68: 1105-1115.
- Linder, M., and Tscherig, T. (2016). Vasculogenic mimicry: Possible role of effector caspase-3, caspase-6 and caspase-7. *Annals of Anatomy-Anatomischer Anzeiger*, 204: 114-117.
- Lin, X. Y., Choi, M. S. K., and Porter, A. G. (2000). Expression analysis of the human caspase-1 subfamily reveals specific regulation of the CASP5 gene by lipopolysaccharide and interferon- γ . *Journal of Biological Chemistry*, 275: 39920-39926
- Liu, A.M., Poon, R.T. and Luk, J.M. (2010). MicroRNA-375 targets Hippo-signaling effector YAP in liver cancer and inhibits tumor properties. *Biochemical and Biophysical Research Communication*, 394: 623-627.
- Liu, W. H., and Chang, L. S. (2011). Fas/FasL-dependent and-independent activation of caspase-8 in doxorubicin-treated human breast cancer MCF7 cells: ADAM10 down-regulation activates Fas/FasL signaling pathway. *The International Journal of Biochemistry and Cell Biology*, 43: 1708-1719.
- Liu, J. W., Xu, W., Li, C. L., Wu, H. Z., and Liu, Y. W. (2008). Kaempferol-7-Ob-D-glucoside (KG) isolated from Smilax china L. rhizome induces G2/M phase arrest

- and apoptosis on HeLa cells in a *p53*-independent manner. *Cancer Letters*, 264: 229-240.
- Liu, M. J., Wang, Z., Ju, Y., Wong, R. N. S., and Wu, Q. Y. (2005). Diosgenin induces cell cycle arrest and apoptosis in human leukemia K562 cells with the disruption of Ca²⁺ homeostasis. *Cancer Chemotherapy and Pharmacology*, 55: 79-90.
- Ljungman, M. (2000). Dial 9-1-1 for *p53*: mechanisms of *p53* activation by cellular stress. *Neoplasia*, 2: 208-225.
- Llovet, J.M. and Bruix, J. (2008). Novel advancements in the management of hepatocellular carcinoma in 2008. *Journal of Hepatology*, 48: 20-37.
- Lohrum, M.A.E., and Vousden, K.H. (2000). Regulation and function of the *p53*-related proteins: Same family, different rules (Electronic version). *Trends in Cell Biology*, 10: 197-202.
- Lopez-Acevedo, M., Grace, L., Teoh, D., Whitaker, R., Adams, D. J., Jia, J., ... and Secord, A. A. (2014). Dasatinib (BMS-35482) potentiates the activity of gemcitabine and docetaxel in uterine leiomyosarcoma cell lines. *Gynecologic Oncology Research and Practice*, 1: 137-167.
- Lowe, S. W., and Lin, A. W. (2000). Apoptosis in cancer. *Carcinogenesis*, 21: 485-495.
- Lucas, A. T., O'Neal, S. K., Santos, C. M., White, T. F., and Zamboni, W. C. (2016). A sensitive high performance liquid chromatography assay for the quantification of doxorubicin associated with DNA in tumor and tissues. *Journal of Pharmaceutical and Biomedical Analysis*, 119: 122-129.
- Luciani, S., Jauregui, B., Kiény, C. and Andrus, J.K. (2009). Human papillomavirus vaccines: new tools for accelerating cervical cancer prevention in developing countries. *Immunotherapy*, 1: 795-807.
- Lynch, H. T. and de la Chapelle, A. (2003). Hereditary colorectal cancer. *The New England Journal of Medicine*, 348: 919-932.
- Madeo, F., Fröhlich, E., and Fröhlich, K. U. (1997). A yeast mutant showing diagnostic markers of early and late apoptosis. *The Journal of Cell Biology*, 139: 729-734.
- Mahassni, S. H., and Al-Reem, R. M. (2013). Apoptosis and necrosis of human breast cancer cells by an aqueous extract of garden cress (*Lepidium sativum*) seeds. *Saudi Journal of Biological Sciences*, 20: 131-139.
- Mai, Z., Blackburn, G. L., and Zhou, J. R. (2007). Soy phytochemicals synergistically enhance the preventive effect of tamoxifen on the growth of estrogen-dependent human breast carcinoma in mice. *Carcinogenesis*, 28: 1217-1223.
- Majno, G., Joris, I. (1995). Apoptosis, oncosis, and necrosis - an overview of cell-death. *American Journal of Pathology*, 146: 3-15.

Malaysian Cancer Statistics. (2006). Data and figure: Peninsular Malaysia; Malaysia Cancer Registry, Ministry of Health, Malaysia, 2006.

Mancini, M., Machamer, C. E., Roy, S., Nicholson, D. W., Thornberry, N. A., Casciola-Rosen, L. A., and Rosen, A. (2000). Caspase-2 is localized at the Golgi complex and cleaves golgin-160 during apoptosis. *The Journal of Cell Biology*, 149: 603-612.

Mandal, R., Raab, M., Mattheiss, Y., Becker, S., Knecht, R., and Strebhardt, K. (2014). pPERK 1/2 inhibit Caspase-8 induced apoptosis in cancer cells by phosphorylating it in a cell cycle specific manner. *Molecular Oncology*, 8: 232-249.

Marcsek, Z., Kocsis, Z., Jakab, M., Szende, B., and Tompa, A. (2004). The efficacy of tamoxifen in estrogen receptor-positive breast cancer cells is enhanced by a medical nutriment. *Cancer Biotherapy and Radiopharmaceuticals*, 19: 746-753.

Marquez, R. T., Tsao, B. W., Faust, N. F., and Xu, L. (2013). Drug resistance and molecular cancer therapy: apoptosis versus autophagy.

Marsden, V. S., O'Connor, L., O'Reilly, L. A., Silke, J., Metcalf, D., Ekert, P. G., ... and Roy, S. (2002). Apoptosis initiated by *BCL2*-regulated caspase activation independently of the cytochrome c/Apaf-1/caspase-9 apoptosome. *Nature*, 419: 634-637.

Martín-Duque, P., Quintanilla, M., McNeish, I., Lopes, R., Romero, J., Romero, D., ... and Vassaux, G. (2006). Caspase-1 as a radio-and chemo-sensitiser in vitro and in vivo. *International Journal of Molecular Medicine*, 17: 841-847.

Martinez, J.D., Parker, M.T., Fultz, K.E., Ignatenko, N.A. and Gerner, E.W. (2003). Molecular biology of cancer. In Abraham, D.J. (Ed.), Burger's medicinal chemistry and drug discovery-chemotherapeutic agents (pp. 2-10). United States of America: John Wiley and Sons, Inc.

Martinon, F., Gaide, O., Petrilli, V., Mayor, A., and Tschopp, J. (2007). NALP inflammasomes: a central role in innate immunity. *Semin Immunopathol* 2007, 29: 213-229.

Martinon, F., Burns, K., and Tschopp, J. (2002). The inflammasome: A molecular platform triggering activation of inflammatory caspases and processing of proIL-1 β . *Molecular Cell*, 10: 417-426.

Martinon, F., and Tschopp, J. (2004). Inflammatory caspases: linking an intracellular innate immune system to autoinflammatory diseases. *Cell*, 117: 561-574.

Mathers, C., Fat, D. M., and Boerma, J. T. (2008). *The global burden of disease: 2004 update*. World Health Organization.

Mathur, R., Gupta, S. K., Singh, N., Mathur, S., Kochupillai, V., and Velpandian, T. (2006). Evaluation of the effect of *Withania somnifera* root extracts on cell cycle and angiogenesis. *Journal of Ethnopharmacology*, 105: 336-341.

- Mathur, R., Gupta, S. K., Singh, N., Mathur, S., Kochupillai, V., and Velpandian, T. (2006). Evaluation of the effect of *Withania somnifera* root extracts on cell cycle and angiogenesis. *Journal of Ethnopharmacology*, 105: 336-341.
- Matthess, Y., Raab, M., Knecht, R., Becker, S., and Strebhardt, K. (2014). Sequential CDK1 and Plk1 phosphorylation of caspase-8 triggers apoptotic cell death during mitosis. *Molecular Oncology*, 8: 596-608.
- Mazalova, L., Balvan, J., Polanska, H., and Sladek, Z. (2014). Occurrence of cell death in cancer cell line PC-3 after treatment of plumbagin. *Mendelnet*, 2014, 494-498.
- McIlwain, D. R., Berger, T., and Mak, T. W. (2013). Caspase functions in cell death and disease. *Cold Spring Harbor Perspectives in Biology*, 5: 56-86.
- McPherson, K., Steel, C. M., Dixon, J.M. (2000). ABC of breast disease: Breast cancer-epidemiology, risk factors, and genetics. *British Medical Journal*, 321: 624-628.
- Md Akim, A. (2004). *Biological Activities of Oryzanol, Stigmasterol and Microminutinin on Human Breast Cancer Cell-Line, Mcf7* (Doctoral dissertation, Universiti Putra Malaysia).
- Medema, R. H., and Macurek, L. (2012). Checkpoint control and cancer. *Oncogene*, 31: 2601-2613.
- Merghoub, N., Benbacer, L., Amzazi, S., Morjani, H., and El-Mzibri, M. (2009). Cytotoxic effect of some Moroccan medicinal plant extracts on human cervical cell lines. *Journal of Medicinal Plants Research*, 3: 1045-1050.
- Merlo, L. M., Pepper, J.W., Reid, B.J. and Maley, C.C. (2006). Cancer as an evolutionary and ecological process. *Nature Review Cancer*, 6: 924-935.
- Miryeganeh, M., and Movafeghi, A. (2009). Scape anatomy of Allium sect. Allium (Alliaceae) in Iran. *JSUT*, 35: 1-5.
- Mitroi, D., Anton, D., Nicu, C., and Manda, M. (2012). Variability of decorative morphological characteristics in the species *Allium atroviolaceum* Boiss. of spontaneous vegetation. *South Western Journal of Horticulture, Biology and Environment*, 3: 131-144.
- Moghadasian, M. H. (2000). Pharmacological properties of plant sterols: in vivo and in vitro observations. *Life Sciences*, 67: 605-615.
- Mohamad Syakir, M. S., Ismail, J., Zaini, A., and Nur Diyana, I, (2012). Bioactivities OF JATROPHA CURCAS Linn LATEX.
- Mohammadi-Motlagh, H. R., Mostafaie, A., and Mansouri, K. (2011). Anticancer and anti-inflammatory activities of shallot. *Archive of Medical Sciences*, 7: 38-44
- Moongkarndi, P., Kosem, N., Kaslungka, S., Luanratana, O. N., Pongpan, N., and Neungton, N. (2004). Antiproliferation, antioxidation and induction of apoptosis by

- Garcinia mangostana (mangosteen) on SKBR3 human breast cancer cell line. *Journal of Ethnopharmacology*, 90: 161–166.
- Moreno-Galindo, C., Hermsen, M., García-Pedrero, J. M., Fresno, M. F., Suárez, C., and Rodrigo, J. P. (2014). p27 and *BCL2* expression predicts response to chemotherapy in head and neck squamous cell carcinomas. *Oral Oncology*, 50: 128-134.
- Motaal, A. A., and Shaker, S. (2011). Anticancer and antioxidant activities of standardized whole fruit, pulp, and peel extracts of Egyptian pomegranate. *The Open Conference Proceedings Journal*, 2: 41-45.
- Mourits, M. J., De Vries, E. G., Willemse, P. H., Ten Hoor, K. A., Hollema, H., and Van der Zee, A. G. (2001). Tamoxifen treatment and gynecologic side effects: a review. *Obstetrics and Gynecology*, 97: 855-866.
- Moya-Nájera, D., Borreani, S., Moya-Herraiz, Á., Calatayud, J., López-Andújar, R., and Colado, J. C. (2016). Is Physical Exercise Harmful to Liver Transplantation Recipients? Review of Literature. *Cirugía Española (English Edition)*, 94: 4-10.
- Mueller, O., Lightfoot, S., Schroeder, A., (2004). RNA integrity number (RIN)—standardization of RNA quality control. *Agilent Application Note, Publication*, 1-8.
- Nakagawa, H., Yamamoto, D., Kiyozuka, Y., Tsuta, K., Uemura, Y., Hioki, K., ... and Tsubura, A. (2000). Effects of genistein and synergistic action in combination with eicosapentaenoic acid on the growth of breast cancer cell lines. *Journal of Cancer Research and Clinical Oncology*, 126: 448-454.
- Nair, S. V., Hettihewa, M., and Rupasinghe, H. P. (2014). Apoptotic and inhibitory effects on cell proliferation of hepatocellular carcinoma HepG2 cells by methanol leaf extract of Costus speciosus. *BioMed research international*, 2014, 98-108.
- National center for gynecological cancers. (2010). Treatment for cervical cancer; cancer Australia, pg 3-6.
- National Cancer Institute. (2009). *What you need to know about breast cancer*. Rockville, MD, USA: NIH Publication.
- National Institute of Environmental Health Sciences (NIEHS). (2001). Report of the international workshop on in vitro method for assessing acute systemic toxicity. NIH publication No.01-4499. NIEHS: Research Triangle Park, NC.
- Ncube, N. S., Afolayan, A. J., and Okoh, A. I. (2008). Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. *African Journal of Biotechnology*, 7: 1797-1806.
- Ndlovu, M. L. M. (2015). Phytochemical screening, cytotoxicity and anticancer activity of Lobostemon fruticosus extracts on human lung cancer cell line (Doctoral dissertation, Faculty of Science, University of Witwatersrand, Johannesburg).

- Ng, W. K., Yazan, L. S., and Ismail, M. (2011). Thymoquinone from Nigella sativa was more potent than cisplatin in eliminating of SiHa cells via apoptosis with down-regulation of *BCL2* protein. *Toxicology in Vitro*, 25: 1392-1398.
- Nygren P. 2001. What is the cancer chemotherapy? *Acta Oncol*, 40: 166-174.
- Kurmyshkina V., Kovchur I. and Volkova O. (2015). Caspases as Putative Biomarkers of Cervical Cancer Development, Cell Death - Autophagy, Apoptosis and Necrosis, Dr. Tobias Ntuli (Ed.), ISBN: 978-953-51-2236-4, InTech.,
- Olsson, M., and Zhivotovsky, B. (2011). Caspases and cancer. *Cell Death and Differentiation*, 18: 1441-1449.
- Oskoueian, E., Abdullah, N., Ahmad, S., Saad, W. Z., Omar, A. R., and Ho, Y. W. (2011). Bioactive compounds and biological activities of *Jatropha curcas* L. kernel meal extract. *International Journal of Molecular Sciences*, 12: 5955-5970.
- Otsuki, Y., Li, Z. and Shibata, M.A. (2003). Apoptotic detection methods from morphology to gene. *Progress in Histochemistry and Cytochemistry*, 38: 275-339.
- Özhatay, N., and Koçyiğit, M. (2009). Pollen morphology of Allium species (Liliaceae) in European Turkey and around Istanbul. *Phytologia Balcanica*, 15: 199-208.
- Paiva, S. R. D., Lima, L. A., Figueiredo, M. R., and Kaplan, M. A. C. (2004). Plumbagin quantification in roots of *Plumbago scandens* L. obtained by different extraction techniques. *Anais da Academia Brasileira de Ciências*, 76: 499-504.
- Papadopoulos, E. I., Yousef, G. M., and Scorilas, A. (2015). Cytotoxic activity of sunitinib and everolimus in Caki-1 renal cancer cells is accompanied by modulations in the expression of apoptosis-related microRNA clusters and *BCL2* family genes. *Biomedicine and Pharmacotherapy*, 70: 33-40.
- Park, S. S., Eom, Y. W., and Choi, K. S. (2005). Cdc2 and CDK2 play critical roles in low dose doxorubicin-induced cell death through mitotic catastrophe but not in high dose doxorubicin-induced apoptosis. *Biochemical and Biophysical Research Communications*, 334: 1014-1021.
- Patil, S. D., Chaudhari, M. A., Sapkale, P. V., and Chaudhari, R. B. (2013). A recent review on anticancer herbal drugs. *Journal of Drug Discovery and Therapeutics*, 1: 77-84.
- Phillips, D. R., Greif, P. C., and Boston, R. C. (1988). Daunomycin-DNA dissociation kinetics. *Molecular Pharmacology*, 33: 225-230.
- Phillips, N. C., Drost, D. T., Varga, W. A., & Shultz, L. M. (2011). Demography, reproduction, and dormancy along altitudinal gradients in three intermountain Allium species with contrasting abundance and distribution. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 206(2), 164-171.
- Piazza, G. A., Rahm, A. L. K., Krutzsch, M., Sperl, G., Paranka, N. S., Gross, P. H., ... and Ahnen, D. J. (1995). Antineoplastic drugs sulindac sulfide and sulfone inhibit cell growth by inducing apoptosis. *Cancer Research*, 55: 3110-3116.

- Pietenpol, J. A., and Stewart, Z. A. (2002). Cell cycle checkpoint signaling:: Cell cycle arrest versus apoptosis. *Toxicology*, 181: 475-481.
- Pinmai, K., Chunlaratthanabhorn, S., Ngamkitidechakul, C., Soonthornchareon, N., and Hahnvajawong, C. (2008). Synergistic growth inhibitory effects of Phyllanthus emblica and Terminalia bellerica extracts with conventional cytotoxic agents: doxorubicin and cisplatin against human hepatocellular carcinoma and lung cancer cells. *World Journal of Gastroenterology*, 14: 1491-1497.
- Pinto, S., Reddy, S. N., Horow, M. M., and Ortiz, J. (2014). Splenic artery syndrome after orthotopic liver transplantation: A review. *International Journal of Surgery*, 12: 1228-1234.
- Pitot, H. C., Goldsworthy, T., and Moran, S. (1981). The natural history of carcinogenesis: implications of experimental carcinogenesis in the genesis of human cancer. *Journal of Supramolecular Structure and Cellular Biochemistry*, 17: 133-146.
- Pompeu, D. R., Silva, E. M., and Rogez, H. (2009). Optimization of solvent extraction of phenolic antioxidants from fruits of *Euterpe oleracea* using response surface methodology. *Bioresource Technology*, 100: 6076-6082.
- Poon, I. K. H., Hulett, M. D., and Parish, C. R. (2010). Molecular mechanisms of late apoptotic/necrotic cell clearance. *Cell Death and Differentiation*, 17: 381-397.
- Portt, L., Norman, G., Clapp, C., Greenwood, M., and Greenwood, M. T. (2011). Anti-apoptosis and cell survival: a review. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1813: 238-259.
- Prayong, P., Barusrux, S., and Weerapreeyakul, N. (2008). Cytotoxic activity screening of some indigenous Thai plants. *Fitoterapia*, 79: 598-601.
- Pratumvinit, B., Srisapoomi, T., Worawattananon, P., Opartkiattikul, N., Jiratchariyakul, W., & Kummalue, T. (2009). In vitro antineoplastic effect of *Ficus hispida* L. plant against breast cancer cell lines. *Journal of Medicinal Plants Research*, 3: 255-261.
- Qiao, Y., Xiang, Q., Yuan, L., Xu, L., Liu, Z., and Liu, X. (2013). Herbacetin induces apoptosis in HepG2 cells: Involvements of ROS and PI3K/Akt pathway. *Food and Chemical Toxicology*, 51: 426-433.
- Quan, H. J., Koyanagi, J., Ohmori, K., Uesato, S., Tsuchido, T., and Saito, S. (2002). Preparations of heterospirostans and their pharmacological activities. *European Journal of Medicinal Chemistry*, 37: 659-669.
- Rajasekaran , M., Archana, R., and Raviprasadh, R.(2012). GC/MS analysis and identification of phytochemicals present in the leaves of Beloperone plumbaginifolia (Jacq.) Nees. *International Journal of Bio-Engineering, Science and Technology*, 3: 2249 – 6483

- Ramadani, M., Gansauge, F., Schlosser, S., Yang, Y., Beger, H. G., and Gamauge, S. (2001). Overexpression of caspase-1 in pancreatic disorders: implications for a function besides apoptosis. *Journal of Gastrointestinal Surgery*, 5: 352-358.
- Ranger, A. M., Malynn, B. A., and Korsmeyer, S. J. (2001). Mouse models of cell death. *Nature Genetics*, 28: 113-118.
- Rasoanaivo, P., Wright, C. W., Willcox, M. L., and Gilbert, B. (2011). Whole plant extracts versus single compounds for the treatment of malaria: synergy and positive interactions. *Malaria Journal*, 10: 4-15.
- Rasul, A., and Ma, T. (2012). In vitro cytotoxic screening of 300 selected Chinese medicinal herbs against human gastric adenocarcinoma SGC-7901 cells. *African Journal of Pharmacy and Pharmacology*, 6: 592-600.
- Ray, M. R., and Jablons, D. M. (2010). *Hallmarks of Metastasis. In Lung Cancer Metastasis* (pp. 29-46). New York: Springer.
- Rello, S., Stockert, J. C., Moreno, V., Gamez, A., Pacheco, M., Juarranz, A., and Villanueva, A. (2005). Morphological criteria to distinguish cell death induced by apoptotic and necrotic treatments. *Apoptosis*, 10: 201-208.
- Réthy, B. (2008). *Antitumour effect of plant extracts and their constituents on cancer cell lines* (Doctoral dissertation, szte).
- Ricki, L. (2005). Immunity and Cancer. In L. Ricki (ed). *Human Genetics; Concepts and Application 6th ed.* (pp. 329-353). New York: McGraw-Hill.
- Riedl, S.J. and Shi, Y. (2004). Molecular mechanisms of caspase regulation during apoptosis. *Natural Reviews Molecular Cell Biology*, 5: 897-907.
- Riestra, S., Rodriguez, M., Delgado, M., Suárez, A., González, N., de la Mata, M. Diaz, G., Miño-Fugarolas, G. and Rodrigo, L. (1998). Tamoxifen does not improve survival of patients with advanced hepatocellular carcinoma. *Journal of Clinical Gastroenterology*, 26: 200-203.
- Rigal, L. and Gaset, A. (1983). Direct preparation of 5-hydroxymethyl-2-furancarboxaldehyde from polyholosides: a chemical valorisation of the Jerusalem artichoke (*Helianthus tuberosus* L.). *Biomass*, 3: 151-163.
- Rodriguez, H. and Case, D. (2005). Epidemiology of cancer: An overview. In *Phytopharmaceuticals in cancer chemoprevention*, ed. Bagchi, D. and Preuss, H. G. pp. 3-14. New York: CRC Press.
- Roland, K., Bernard, V., Saraiya, M., Hawkins, N., Brandit, H. and Friedman, A. (2009). Assessing cervical cancer screening guideline in patient education materials. *Journal of Women's Health*. 18: 5-12.
- Russo, M., Spagnuolo, C., Tedesco, I., and Russo, G. L. (2010). Phytochemicals in cancer prevention and therapy: truth or dare?. *Toxins*, 2: 517-551.

- Ryan, K.M., Phillips, A.C., and Vousden, K.H. (2001). Regulation and function of the *p53* tumor suppressor protein. *Current Opinion in Cell Biology*, 13: 332-337.
- Sa'adi, R. A., Kamaludin, N. H. I., Zakaria, Z., Arbain, D., MohdIdris, Z., and Abdullah, N. A. H. (2014). Evaluation of phenolic compound extraction from LimauKasturi (*Citrus macrocarpa*) peels extract. *Advances in Environmental Biology*, 8: 73-76.
- Saetung, A., Itharat, A., Dechsukum, C., Wattanapiromsakul, C., Keawpradub, N., & Ratanasawan, P. (2005). Cytotoxic activity of Thai medicinal plants for cancer treatment. *Songklanakarin Journal of Science and Technology*, 27: 469-478.
- Salvesen, G.S. and Riedl, S.J. (2008). Caspase mechanisms. *Advance in Experimental and Medical Biology*, 615: 13-23.
- Samraj, A. K., Sohn, D., Schulze-Osthoff, K., and Schmitz, I. (2007). Loss of caspase-9 reveals its essential role for caspase-2 activation and mitochondrial membrane depolarization. *Molecular biology of the cell*, 18: 84-93.
- Samarakoon, S. R., Thabrew, I., Galhena, P. B., and Tennekoon, K. H. (2012). Effect of standardized decoction of *Nigella sativa* seed, *Hemidesmus indicus* root and *Smilax glabra* rhizome on the expression of *p53* and p21 genes in human hepatoma cells (HepG2) and mouse liver with chemically-induced hepatocarcinogenesis. *Tropical Journal of Pharmaceutical Research*, 11: 51-61.
- Sandelin, K., Apffelstaedt, J. P., Abdullah, H., Murray, E. M. and Ajuluchuku, E.U. (2002). Breast Surgery International—breast cancer in developing countries. *Scandinavian Journal of Surgery*, 91: 222-226.
- Saonere, J. A. (2010). Awareness screening programme reduces the risk of cervical cancer in women. *African Journal of Pharmacy and Pharmacology*, 4: 314-23.
- Sarker, S. D., Latif, Z., and Gray, A. I. (2005). *Natural products isolation*: Springer Science and Business Media.
- Sato, H., Ozawa, K., Yonemura, K., Iwata, S., Ono, M., Uemoto, S., ... and Tanaka, K. (2003). Quantitative PCR to evaluate small amounts of *BCL2* mRNA in human peripheral T cells: implication of equimolar target and competitor end products. *Clinica Chimica Acta*, 328: 147-153.
- Savage, S.A., Burdett, L., Troisi, R., Douglass, C., Hoover, R.N., and Chanock, S.J. (2007). Germ-line genetic variation of *TP53* in osteosarcoma. *Pediatric blood and cancer*, 49: 28-33.
- Scaffidi, C., Fulda, S., Srinivasan, A., Friesen, C., Li, F., Tomaselli, K. J., ... and Peter, M. E. (1998). Two CD95 (APO-1/Fas) signaling pathways. *The EMBO journal*, 17: 1675-1687.
- Schlosser, S., Gansauge, F., Ramadani, M., Beger, H. G., and Gansauge, S. (2001). Inhibition of caspase-1 induces cell death in pancreatic carcinoma cells and potentially modulates expression levels of *BCL2* family proteins. *FEBS Letters*, 491: 104-108.

- Schwartz, S., Yamamoto, H., Navarro, M., Maestro, M., Reventós, J., and Perucho, M. (1999). Frameshift mutations at mononucleotide repeats in caspase-5 and other target genes in endometrial and gastrointestinal cancer of the microsatellite mutator phenotype. *Cancer Research*, 59: 2995-3002.
- Scripture, C. D., and Figg, W. D. (2006). Drug interactions in cancer therapy. *Nature Reviews Cancer*, 6: 546-558.
- Sengupta, A., Ghosh, S., and Bhattacharjee, S. (2004). Allium vegetables in cancer prevention: an overview. *Asian Pacific Journal of Cancer Prevention*, 5: 237-245.
- Shafi, G., Munshi, A., Hasan, T. N., Alshatwi, A. A., Jyothy, A., and Lei, D. K. (2009). Induction of apoptosis in HeLa cells by chloroform fraction of seed extracts of *Nigella sativa*. *Cancer Cell International*, 9: 29-38.
- Shamsabadi, F. T., Khoddami, A., Fard, S. G., Abdullah, R., Othman, H. H., and Mohamed, S. (2013). Comparison of tamoxifen with edible seaweed (*Eucheuma cottonii* L.) extract in suppressing breast tumor. *Nutrition and Cancer*, 65: 255-262.
- Sharma, G., Tyagi, A. K., Singh, R. P., Chan, D. C., and Agarwal, R. (2004). Synergistic anti-cancer effects of grape seed extract and conventional cytotoxic agent doxorubicin against human breast carcinoma cells. *Breast Cancer Research and Treatment*, 85: 1-12.
- Sharon, C.W.L., Stephanie, W.F.S., Chun-Kit, L., Ying-Yie, W. and Shiu-Fun, P. (2005). Cell cycle arrest by a natural product in G2/M checkpoint. *International Journal of Medical Sciences*, 2: 64-69.
- Shamsabadi, F. T., Khoddami, A., Fard, S. G., Abdullah, R., Othman, H. H., and Mohamed, S. (2013). Comparison of tamoxifen with edible seaweed (*Eucheuma cottonii* L.) extract in suppressing breast tumor. *Nutrition and Cancer*, 65: 255-262.
- Shang, L. H., Li, C. M., Yang, Z. Y., Che, D. H., Cao, J. Y., and Yu, Y. (2012). *Luffa echinata Roxb.* induces human colon cancer cell (HT-29) death by triggering the mitochondrial apoptosis pathway. *Molecules*, 17: 5780-5794.
- Shao, W., Yeretssian, G., Doiron, K., Hussain, S. N., and Saleh, M. (2007). The caspase-1 digestome identifies the glycolysis pathway as a target during infection and septic shock. *Journal of Biological Chemistry*, 282: 36321-36329.
- Shen, Q. and Brown, P.H. (2003). Novel agents for the prevention of breast cancer: targeting transcription factors and signal transduction pathways. *Journal of Mammary Gland Biology and Neoplasia*, 8:45-73.
- Shi, Y. (2004). Caspase activation: revisiting the induced proximity model. *Cell*, 117: 855-858.
- Shi, L., Teng, H., Zhu, M., Li, C., Huang, K., Chen, B., ... and Wang, J. (2015). Paeoniflorin inhibits nucleus pulposus cell apoptosis by regulating the expression of *BCL2* family proteins and caspase-9 in a rabbit model of intervertebral disc degeneration. *Experimental and Therapeutic Medicine*, 10: 257-262.

- Shoeb, M. (2006). Anticancer agents from medicinal plants. *Bangladesh Journal of Pharmacology*, 1: 35-41.
- Shukor, M. F. A., Ismail, I., Zainal, Z., and Noor, N. M. (2013). Development of a Polygonum minus cell suspension culture system and analysis of secondary metabolites enhanced by elicitation. *Acta Physiologiae Plantarum*, 35: 1675-1689.
- Simmer, M. (2003). Flow cytometry: a technology to count and sort cells. *Internetpublikation: http://bioteach. ubc. ca/MolecularBiology/FlowCytometry*.
- Simpson, K. L., Cawthorne, C., Zhou, C., Hodgkinson, C. L., Walker, M. J., Trapani, F., ... and Williams, K. J. (2013). A caspase-3 'death-switch' in colorectal cancer cells for induced and synchronous tumor apoptosis in vitro and in vivo facilitates the development of minimally invasive cell death biomarkers. *Cell Death and Disease*, 4: 613-625.
- Singal, P. K., and Iliskovic, N. (1998). Doxorubicin-induced cardiomyopathy. *New England Journal of Medicine*, 339: 900-905.
- Singh, R.R., Kumar, R. (2005). Steroid hormone receptor signaling in tumorigenesis. *Journal of Cellular Biochemistry*, 96: 490-505.
- Slee, E. A., Adrain, C., and Martin, S. J. (2001). Executioner caspase-3,-6, and -7 perform distinct, non-redundant roles during the demolition phase of apoptosis. *Journal of Biological Chemistry*, 276: 7320-7326.
- Slee, E. A., Harte, M. T., Kluck, R. M., Wolf, B. B., Casiano, C. A., Newmeyer, D. D., ... and Green, D. R. (1999). Ordering the cytochrome c-initiated caspase cascade: hierarchical activation of caspases-2,-3,-6,-7,-8, and -10 in a caspase-9-dependent manner. *The Journal of Cell Biology*, 144: 281-292.
- Sneller, M. C., Wang, J., Dale, J. K., Strober, W., Middleton, L. A., Choi, Y., ... and Lenardo, M. J. (1997). Clinical, immunologic, and genetic features of an autoimmune lymphoproliferative syndrome associated with abnormal lymphocyte apoptosis. *Blood*, 89: 1341-1348.
- Soung, Y. H., Jeong, E. G., Ahn, C. H., Kim, S. S., Song, S. Y., Yoo, N. J., and Lee, S. H. (2008). Mutational analysis of caspase 1, 4, and 5 genes in common human cancers. *Human Pathology*, 39: 895-900.
- Srivastava, V., Negi, A.S., Kumar, J.K., Gupta, M.M., and Khanuja, S.P.S. (2005). Plant based anticancer molecules: A chemical and biological profile of some important leads. *Bioorganic and Medicinal Chemistry*, 13: 5892-5908.
- Srivastava, J.K. and Gupta, S. (2006). Tocotrienol-rich fraction of palm oil induces cell cycle arrest and apoptosis selectively in human prostate cancer cells. *Biochemical and Biophysical Research Communication*, 346: 447-453.
- Stajner, D., Igic, R., Popovic, B. M., and Malencic, D. (2008). Comparative study of antioxidant properties of wild growing and cultivated Allium species. *Phytotherapy Research*, 22: 113-117.

- Štajner, D., and Varga, I. S. (2003). An evaluation of the antioxidant abilities of Allium species. *Acta Biologica Szegediensis*, 47: 103-106.
- Štajner, D., Milić, N., Čanadanović-Brunet, J., Kapor, A., Štajner, M., and Popović, B. M. (2006). Exploring Allium species as a source of potential medicinal agents. *Phytotherapy Research*, 20: 581-584.
- Studzinski, G.P. (1999). *Overview of apoptosis. In Apoptosis: A practical approach.* New York: Oxford University Press.
- Su, C. C., Lin, J. G., Chen, G. W., Lin, W. C., and Chung, J. G. (2006). Down-regulation of Cdc25c, CDK1 and Cyclin B1 and Up-regulation of Wee1 by Curcumin Promotes Human Colon Cancer Colo 205 Cell Entry into G2/M-phase of Cell Cycle. *Cancer Genomics-Proteomics*, 3: 55-61.
- Sui, L., Dong, Y., Watanabe, Y., Yamaguchi, F., Hatano, N., Izumori, K., and Tokuda, M. (2005). Growth inhibitory effect of D-allose on human ovarian carcinoma cells in vitro. *Anticancer Research*, 25: 2639-2644.
- Sukapirom, K. and Iemsri, S. (2007). Antiproliferative effect of Erycibe elliptilimba on human breast cancer cell lines. *Journal of Ethnopharmacology*, 110: 439-443.
- Sultana, B., Anwar, F., and Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*, 14: 2167-2180.
- Sun, V., Ferrell, B., Juarez, G., Wagman, L.D. Yen, Y. and Chung, V. (2008). Symptom concerns and quality of life in hepatobiliary cancers. *Oncology Nursing Forum*, 35: 45-52.
- Sundarraj, S., Thangam, R., Sreevani, V., Kaveri, K., Gunasekaran, P., Achiraman, S., and Kannan, S. (2012). γ -Sitosterol from Acacia nilotica L. induces G2/M cell cycle arrest and apoptosis through c-Myc suppression in MCF7 and A549 cells. *Journal of Ethnopharmacology*, 141: 803-809.
- Sundquist, T. E. R. R. I., Moravec, R., Niles, A. N. D. R. E. W., O'Brien, M. A. R. T. H. A., and Riss, T. (2006). Timing your apoptosis assays. *Cell Notes*, 16: 18-21.
- Sureban, S. M., Subramaniam, D., Rajendran, P., Ramanujam, R. P., Dieckgraefe, B. K., Houchen, C. W., and Anant, S. (2006). Therapeutic effects of Phyllanthus species: induction of TNF- α -mediated apoptosis in HepG2 hepatocellular carcinoma cells. *American Journal of Pharmacology and Toxicology*, 1: 65-71.
- Suryadinata, R., Sadowski, M., and Sarcevic, B. (2010). Control of cell cycle progression by phosphorylation of cyclin-dependent kinase (CDK) substrates. *Bioscience Reports*, 30: 243-255.
- Swanton, C. (2004). Cell-cycle targeted therapies. *Lancet Oncology*, 5: 27-36.
- Syu, M.J. (2001). Biological production of 2,3-butanediol. *Applied Microbiology and Biotechnology*, 55: 10-18

- Sylvester, P. W. (2011). Optimization of the tetrazolium dye (MTT) colorimetric assay for cellular growth and viability. In *Drug Design and Discovery* (pp. 157-168). Humana Press.
- Stupack, D. G. (2013). Caspase-8 as a therapeutic target in cancer. *Cancer Letters*, 332: 133-140.
- Syed Abdul Rahman, S. N., Abdul Wahab, N., and Abd Malek, S. N. (2013). In vitro morphological assessment of apoptosis induced by antiproliferative constituents from the rhizomes of Curcuma zedoaria. *Evidence-Based Complementary and Alternative Medicine*, 2013.
- Taha, M. M. E., Abdul, A. B., Abdullah, R., Ibrahim, T. A. T., Abdelwahab, S. I., and Mohan, S. (2010). Potential chemoprevention of diethylnitrosamine-initiated and 2-acetylaminofluorene-promoted hepatocarcinogenesis by zerumbone from the rhizomes of the subtropical ginger (*Zingiber zerumbet*). *Chemico-Biological Interactions*, 186(3), 295-305.
- Tayade, A. B., Dhar, P., Kumar, J., Sharma, M., Chauhan, R. S., Chaurasia, O. P., and Srivastava, R. B. (2013). Chemometric profile of root extracts of Rhodiola imbricata Edgew. with hyphenated gas chromatography mass spectrometric technique. *PloS One*, 8: 52797-52812.
- Tanis, E., Spliehoff, J. W., Evers, D. J., Langhout, G. C., Snaebjornsson, P., Prevo, W., ... and Ruers, T. J. M. (2015). Real-time in vivo assessment of radiofrequency ablation of human colorectal liver metastases using diffuse reflectance spectroscopy. *European Journal of Surgical Oncology*, 42: 251-259.
- Tee, T. T., Cheah, Y. H., and Hawariah, L. P. A. (2007). F16, a fraction from *Eurycoma longifolia* jack extract, induces apoptosis via a caspase-9-independent manner in MCF-7 cells. *Anticancer research*, 27: 3425-3430.
- Teiten, M. H., Eifes, S., Dicato, M., and Diederich, M. (2010). Curcumin—the paradigm of a multi-target natural compound with applications in cancer prevention and treatment. *Toxins*, 2: 128-162.
- Tepe, B., Sokmen, M., Akpulat, H. A., and Sokmen, A. (2005). In vitro antioxidant activities of the methanol extracts of five Allium species from Turkey. *Food Chemistry*, 92: 89-92.
- Thalappilly, S., Sadasivam, S., Radha, V., and Swarup, G. (2006). Involvement of caspase 1 and its activator Ipaf upstream of mitochondrial events in apoptosis. *FEBS Journal*, 273: 2766-2778.
- Thoo, Y. Y., Ho, S. K., Liang, J. Y., Ho, C. W., and Tan, C. P. (2010). Effects of binary solvent extraction system, extraction time and extraction temperature on phenolic antioxidants and antioxidant capacity from mengkudu (*Morinda citrifolia*). *Food Chemistry*, 120: 290-295.
- Thor, A. D., Moore, D. H., Edgerton, S. M., Kawasaki, E. S., Reihsaus, E., Lynch, H. T., ... and Smith, H. S. (1992). Accumulation of p53 tumor suppressor gene protein:

- an independent marker of prognosis in breast cancers. *Journal of the National Cancer Institute*, 84: 845-855.
- Thuret, G., Chiquet, C., Herrag, S., Dumollard, J. M., Boudard, D., Bednarz, J., ... and Gain, P. (2003). Mechanisms of staurosporine induced apoptosis in a human corneal endothelial cell line. *British Journal of Ophthalmology*, 87: 346-352.
- Tierney, J. (2004). Neoadjuvant chemotherapy for locally advanced cervix cancer. *Cochrane Database Syst Rev*, 1: 70-86.
- Timité, G., Mitaine-Offer, A. C., Miyamoto, T., Tanaka, C., Mirjolet, J. F., Duchamp, O., and Lacaille-Dubois, M. A. (2012). Structure and cytotoxicity of steroid glycosides from Allium schoenoprasum. *Planta Medica*, 78: 40-49.
- Todd, R., Hinds, P. W., Munger, K., Rustgi, A. K., Opitz, O. G., Suliman, Y., & Wong, D. T. (2002). Cell cycle dysregulation in oral cancer. *Critical Reviews in Oral Biology and Medicine*, 13: 51-61.
- Todd, R. and Wong D. T. (1999). Oncogenes. *Anticancer Research*, 19: 4729-4746.
- Tomankova, K., Polakova, K., Pizova, K., Binder, S., Havrdova, M., Kolarova, M., ... and Malohlava, J. (2015). In vitro cytotoxicity analysis of doxorubicin-loaded/superparamagnetic iron oxide colloidal nanoassemblies on McF7 and NIH3T3 cell lines. *International Journal of Nanomedicine*, 10: 949-961.
- Tomuleasa, C., Soritau, O., Rus-Ciuca, D., Pop, T., Todea, D., Mosteanu, O., ... and Irimie, A. (2010). Isolation and characterization of hepatic cancer cells with stem-like properties from hepatocellular carcinoma. *Journal of Gastrointestinal Liver Diseases*, 19: 61-67.
- Torgovnick, A., and Schumacher, B. (2015). DNA repair mechanisms in cancer development and therapy. *Frontiers in Genetics*, 6: 157-172.
- Tsujimoto, Y., and Shimizu, S. (2000). VDAC regulation by the *BCL2* family of proteins. *Cell Death and Differentiation*, 7: 1174-1181.
- Uddin, S. J., Grice, I. D., and Tiralongo, E. (2011). Cytotoxic effects of Bangladeshi medicinal plant extracts. *Evidence-Based Complementary and Alternative Medicine*, 2011.
- Uhl, S.R. (2000). *Handbook of spices, seasonings and flavorings*. United States of America: Technomic Publishing Company, Inc.
- Underwood, S.M., Ramsay-Johnson, E., Dean, A., Russ, J., and Ivalis, R. (2009). Expanding the scope of nursing research in low resource and middle resource countries, regions, and states focused on cervical cancer prevention, early detection, and control. *Journal of National Black Nurses Association*, 20: 42-54.
- Uma, B. and Parvathavarthini, R. (2010). Antibacterial effect of hexane extract of Sea Urchin, *Temnopleurus alexandri* (Bell, 1884). *International Journal of PharmTech Research*, 2: 1677-1680.

- Vakifahmetoglu, H. (2009). *DNA damage-induced cell death: The role of caspase-2*. Institutet för miljömedicin (IMM)/Institute of Environmental Medicine.
- Van Cruchten, S. And Van der Broeck, W. (2002). Morphological and biochemical aspects of apoptosis, oncosis and necrosis. In *Anatomia, Histologia Embryologia*, 31: 214- 223.
- Vander Heiden, M. G., and Thompson, C. B. (1999). *BCL2* proteins: regulators of apoptosis or of mitochondrial homeostasis?. *Nature Cell Biology*, 1: 209-216.
- Van Slambrouck, S., Daniels, A. L., Hooten, C. J., Brock, S. L., Jenkins, A. R., Ogasawara, M. A., ... and Constantine, S. R. (2007). Effects of crude aqueous medicinal plant extracts on growth and invasion of breast cancer cells. *Oncology Reports*, 17: 1487-1492.
- Vermeulen K, Van Bockstaele DR, Berneman ZN. (2003). The cell cycle: a review of regulation, deregulation and therapeutic targets in cancer. *Cell Proliferation*, 36: 131-149.
- Vermeulen, J., De Preter, K., Lefever, S., Nuytens, J., De Vloed, F., Derveaux, S., ... and Vandesompele, J. (2011). Measurable impact of RNA quality on gene expression results from quantitative PCR. *Nucleic Acids Research*, 39: 63-75.
- Vizza, E., Corrado, G., Zanagnolo, V., Tomaselli, T., Cutillo, G., Mancini, E., and Maggioni, A. (2014). Neoadjuvant chemotherapy followed by robotic radical hysterectomy in locally advanced cervical cancer: A multi-institution study. *Gynecologic Oncology*, 133: 180-185.
- Voet, D. and Voet, J.G. (1995). *Biochemistry* (2nd ed.). New York: John Wiley
- Walsh, J. G., Cullen, S. P., Sheridan, C., Lüthi, A. U., Gerner, C., and Martin, S. J. (2008). Executioner caspase-3 and caspase-7 are functionally distinct proteases. *Proceedings of the National Academy of Sciences*, 105: 12815-12819.
- Wang, H., Zhang, X., Teng, L., and Legerski, R. J. (2015). DNA damage checkpoint recovery and cancer development. *Experimental Cell Research*, 334: 350-358.
- Wang, X. J., Cao, Q., Liu, X., Wang, K. T., Mi, W., Zhang, Y., ... and Su, X. D. (2010). Crystal structures of human caspase 6 reveal a new mechanism for intramolecular cleavage self-activation. *EMBO reports*, 11: 841-847.
- Wang, I. K., Lin-Shiau, S. Y., and Lin, J. K. (1999). Induction of apoptosis by apigenin and related flavonoids through cytochrome c release and activation of caspase-9 and caspase-3 in leukaemia HL-60 cells. *European Journal of Cancer*, 35: 1517-1525.
- Wang, P., and Li, J. C. (2007). Trichosanthin-induced specific changes of cytoskeleton configuration were associated with the decreased expression level of actin and tubulin genes in apoptotic HeLa cells. *Life Sciences*, 81: 1130-1140.

- Wang, Y. X., Zhao, L., Wang, X. Y., Liu, C. M., and Yu, S. G. (2012). Role of Caspase 8, Caspase 9 and *BCL2* polymorphisms in papillary thyroid carcinoma risk in Han Chinese population. *Medical Oncology*, 29: 2445-2451.
- Wang, L., and Weller, C. L. (2006). Recent advances in extraction of nutraceuticals from plants. *Trends in Food Science and Technology*, 17: 300-312.
- Warby, S. C., Doty, C. N., Graham, R. K., Carroll, J. B., Yang, Y. Z., Singaraja, R. R., ... and Hayden, M. R. (2008). Activated caspase-6 and caspase-6-cleaved fragments of huntingtin specifically colocalize in the nucleus. *Human Molecular Genetics*, 17: 2390-2404.
- Wei, Y. Y. (2010). Proteomics Analysis of Protein-Producing Chinese Hamster Ovary Cells during Apoptosis in Prolonged Cultivation. Retrieved from <http://hdl.handle.net/10012/5222>
- Weigelt, B., Petersen, J.L., and van 't Veer, L.J. (2005) Breast cancer metastasis: markers and models. *Nature Reviews Cancer*, 5: 591-602.
- White, S. C., and Winkler, J. L. (2004). Alliance for cervical cancer prevention. *ACCP strategies for supporting women with cervical cancer*. Retrieved from <http://www.rho.org/ccresources.htm>.
- Wild, C.P. and Montesano, R.(2009). A model of interaction: aflatoxins and hepatitis viruses in liver cancer aetiology and prevention. *Cancer Letter*, 286: 22-28.
- Winter, R. N., Kramer, A., Borkowski, A., and Kyprianou, N. (2001). Loss of caspase-1 and caspase-3 protein expression in human prostate cancer. *Cancer Research*, 61: 1227-1232.
- Wolf, D. M., Langan-Fahey, S. M., Parker, C. J., McCague, R., and Jordan, V. C. (1993). Investigation of the mechanism of tamoxifen-stimulated breast tumor growth with nonisomerizable analogues of tamoxifen and metabolites. *Journal of the National Cancer Institute*, 85: 806-812.
- Wong, H. M. (2014). Oral complications and management strategies for patients undergoing cancer therapy. *The Scientific World Journal*, 2014.
- Wong, R. S. (2011). Apoptosis in cancer: from pathogenesis to treatment. *Journal of Experimental and Clinical Cancer Research*, 30: 87-100.
- Wong, F. C., Woo, C. C., Hsu, A., and Tan, B. K. H. (2013). The anti-cancer activities of Vernonia amygdalina extract in human breast cancer cell lines are mediated through caspase-dependent and p53-independent pathways. *PloS One*, 8: 8021-8036.
- Wu, D. and Cederbaum, A. I. (1999). Ethanol-Induced Apoptosis to Stable HepG2 Cell Lines Expressing Human Cytochrome P-4502E1. *Alcoholism: Clinical and Experimental Research*, 23: 67-76.
- Wu, X. and Prior, R.L. (2005). Identification and characterization of anthocyanins by high-performance liquid chromatography-electrospray ionization-tandem mass

- spectrometry in common foods in the United States: vegetables, nuts, and grains. *Journal of Agricultural and Food Chemistry*, 53: 3101-3113.
- Wu, S. J., Ng, L. T., Chen, C. H., Lin, D. L., Wang, S. S., and Lin, C. C. (2004). Antihepatoma activity of Physalis angulata and P. peruviana extracts and their effects on apoptosis in human Hep G2 cells. *Life Sciences*, 74: 2061-2073.
- Xu, W., Huang, J. J. H., and Cheung, P. C. K. (2012). Extract of Pleurotus pulmonarius suppresses liver cancer development and progression through inhibition of VEGF-induced PI3K/AKT signaling pathway. *PloS One*, 7: 34406-34419.
- Xu, Z., Zhang, K., Hou, C., Wang, D., Liu, X., Guan, X., ... and Zhang, H. (2014). A novel nanoassembled doxorubicin prodrug with a high drug loading for anticancer drug delivery. *Journal of Materials Chemistry B*, 2: 3433-3437.
- Yaacob, N. S., Hamzah, N., Kamal, N. N. N. M., Abidin, S. A. Z., Lai, C. S., Navaratnam, V., and Norazmi, M. N. (2010). Anticancer activity of a sub-fraction of dichloromethane extract of Strobilanthes crispus on human breast and prostate cancer cells in vitro. *BMC Complementary and Alternative Medicine*, 10: 42-53.
- Yaacob, N. S., Kamal, N. N., and Norazmi, M. N. (2014). Synergistic anticancer effects of a bioactive subfraction of Strobilanthes crispus and tamoxifen on MCF7 and MDA-MB-231 human breast cancer cell lines. *BMC Complementary and Alternative Medicine*, 14: 252-267.
- Yadav, V., Sultana, S., Yadav, J., and Saini, N. (2012). Gatifloxacin induces S and G 2-phase cell cycle arrest in pancreatic cancer cells via p21/p27/p53. *PLoS One*, 7: 96-108.
- Yamabe, K., Shimizu, S., Ito, T., Yoshioka, Y., Nomura, M., Narita, M., ... and Matsuda, H. (1999). Cancer gene therapy using a pro-apoptotic gene, caspase-3. *Gene Therapy*, 6: 1952-1959.
- Yang, S., Liu, J., Thor, A. D., and Yang, X. (2007). Caspase expression profile and functional activity in a panel of breast cancer cell lines. *Oncology Reports*, 17: 1229-1235.
- Yarbro C, Frogge M and Goodman M (2005). Cancer Nursing: Principles and Practice. Boston, MA: Jones and Bartlett Publishers.
- Yazdanpanahia, N., Behbahanib, M., and Yektaeiana, A. (2014). Effect of Boswellia thurifera gum methanol extract on cytotoxicity and P53 gene expression in human breast cancer cell line. *Iranian Journal of Pharmaceutical Research*, 13: 719-724.
- Yogeswari, S., Ramalakshmi, S., and Muthu, J. M. (2012). Identification and comparative studies of different volatile fractions from Monochaetia kansensis by GC-MS. *Global Journal of Pharmacology*, 6: 65-71.
- Yokochi, T., and Robertson, K. D. (2004). Doxorubicin inhibits DNMT1, resulting in conditional apoptosis. *Molecular pharmacology*, 66: 1415-1420.

- Yolcu, E.S., Ash, S., Kaminitz, A., Sagiv, Y., Askenasy, N. and Yarkoni, S. (2008). Apoptosis as a mechanism of T-regulatory cell homeostasis and suppression. *Immunology and Cell Biology*, 86: 650-658.
- Yoshimaru, T., Komatsu, M., Matsuo, T., Chen, Y. A., Murakami, Y., Mizuguchi, K. and Katagiri, T. (2013). Targeting BIG3-PHB2 interaction to overcome tamoxifen resistance in breast cancer cells. *Nature Communications*, 4: 2443-2456.
- Youn, M. J., Kim, J. K., Park, S. Y., Kim, Y., Kim, S. J., Lee, J. S., ... and Kim, K. Y. (2008). Chaga mushroom (*Inonotus obliquus*) induces G0/G1 arrest and apoptosis in human hepatoma HepG2 cells. *World Journal of Gastroenterology*, 14: 511-517.
- Yu, L., Haley, S., Perret, J., Harris, M., Wilson, J., and Qian, M. (2002). Free radical scavenging properties of wheat extracts. *Journal of Agricultural and Food Chemistry*, 50: 1619–1624.
- Yu, Q. (2006). Restoring *p53*-mediated apoptosis in cancer cells: new opportunities for cancer therapy. *Drug Resistance Updates*, 9: 19-25.
- Yuen, M.F., Hou, J.L. and Chutaputti, A. (2009). Hepatocellular carcinoma in the Asia pacific region. *Journal of Gastroenterology and Hepatology*, 24: 346-353.
- Zaid, H., Rayan, A., Said, O., and Saad, B. (2010). Cancer treatment by Greco-Arab and Islamic herbal medicine. *The Open Nutraceuticals Journal*, 3: 203-212.
- Zakaria, Y., Rahmat, A., Pihie, A. H., Abdullah, N. R., and Houghton, P. J. (2009). Eurycomanone induce apoptosis in HepG2 cells via up-regulation of *p53*. *Cancer Cell International*, 9: 1-21.
- Zhang, H., Zhang, L., Peng, L. J., Dong, X. W., Wu, D., Wu, V. C. H., and Feng, F. Q. (2012). Quantitative structure-activity relationships of antimicrobial fatty acids and derivatives against *Staphylococcus aureus*. *Journal of Zhejiang University Science B*, 13: 83-93.
- Zhao, Y., Wu, M., Shen, Y., and Zhai, Z. (2001). Analysis of nuclear apoptotic process in a cell-free system. *Cellular and Molecular Life Sciences*, 58: 298-306.
- Zhao, L., Wientjes, M. G., and Au, J. L. (2004). Evaluation of combination chemotherapy integration of nonlinear regression, curve shift, isobogram, and combination index analyses. *Clinical Cancer Research*, 10: 7994-8004.
- Zhao, J., Kelnar, K., and Bader, A. G. (2014). In-depth analysis shows synergy between erlotinib and miR-34a. *Plos One*, 9: 89105-89115.
- Zhivotovsky, B., Samali, A., Gahm, A., Orrenius, S. (1999). Caspases: their intracellular localization and translocation during apoptosis. *Cell Death Differentiation*, 6: 644-651.
- Zhou, T., Zhu, Z., Wang, C., Fan, G., Peng, J., Chai, Y., and Wu, Y. (2007). On-line purity monitoring in high-speed counter-current chromatography: Application of HSCCC-HPLC-DAD for the preparation of 5-HMF, neomangiferin and mangiferin

from *Anemarrhena asphodeloides* Bunge. *Journal of Pharmaceutical and Biomedical Analysis*, 44: 96-100.

Zolfaghari, B., Barile, E., Capasso, R., Izzo, A. A., Sajjadi, S. E., and Lanzotti, V. (2006). The Sapogenin Atroviolacegenin and Its Diglycoside Atroviolaceoside from *Allium a troviolaceum*. *Journal of Natural Products*, 69: 191-195.

Zouari, S., Ketata, M., Boudhrioua, N., and Ammar, E. (2013). *Allium roseum* L. volatile compounds profile and antioxydant activity for chemotype discrimination—Case study of the wild plant of Sfax (Tunisia). *Industrial Crops and Products*, 41: 172-178.

BIODATA OF STUDENT

Somayeh Khazaei was born in December 18th, 1983 in Nowshahr, Iran. She began her primary school in Nowshahr from 1990 to 1997 and continued her high school from 1997 to 2000. After graduation from high school she started to study foundation in 2001. Her enthusiasm for learning and the great support and encouragement from her parents led her to pursue further education. She studied Bachelor of Science in the College of Science/Biology Department/ Mazandaran University and graduated for the academic year 2003/2007 Obtained the B.Sc degree in Plant biology branch with a grade good and an average of 75. Then she decided to continue her education and started her further education in Master Degree (M.Sc in Plant Physiology) in College of Medicine/Azad University of Tonekabon and graduated in 2009 with an average of 94.5. After her marriage in 2010 she moved to Malaysia where her husband was living. Her interest to further study and the good quality of Universities in Malaysia led her to start her Phd in Genetic at Faculty of Medicine and Health Sciences in Universiti Putra Malaysia (UPM) from September of 2012.

List of Publications

Khazaei, S., Abdul Hamid, R., Zakaria, Z.A., Mohd Esa, N., Ramachandran, V., Etemad, A., Ismail, P. Comparative Evaluation of Chemometric Profile of 12 Various Flower and Bulb Extracts of *Allium atroviolaceum* with Gas Chromatography Mass Spectrometric Technique.

Somayeh Khazaei., Norhaizan Mohd Esa., Roslida Abdul Hamid., Vasudevan Ramachandran., Ashok Kumar., Patimah Ismail. *In vitro* Antiproliferative and Apoptotic Effect of *Allium atroviolaceum* Extract on Breast, Cervical and Liver Cancer Cells.

Somayeh Khazaei, Norhaizan Mohd Esa, Vasudevan Ramachandran, Roslida Abdul Hamid, Patimah Ismail. Cytotoxicity and apoptosis inducing effects of *Allium atroviolaceum* flower Extract is mediated through cell cycle arrest, caspase-dependent and *p53*-independent pathway in breast cancer cell lines.

Khazaei, S., Abdul Hamid, R., Mohd Esa, N., Ramachandran, V., Aalam GHomi Tabatabaei, F., Ismail, P. Mechanism of the promotion of HepG2 cell apoptosis by flower of *Allium atroviolaceum*.

Khazaei, S., Abdul Hamid, R., Mohd Esa, N., Ramachandran, V., Aalam GHomi Tabatabaei, F., Ismail P^{1*}. Flower Extract of *Allium atroviolaceum* Triggered Apoptosis, Activated Caspase-3 and Down-Regulated the antiapoptotic *BCL2* gene in HeLa Cancer Cell Line.