



UNIVERSITI PUTRA MALAYSIA

**EFFICACY OF VARIOUS LOCAL HONEY FOR THE TREATMENT OF
BURN WOUNDS**

ROZAINI BT. MOHD. ZOHDI

FPV 2005 8

**EFFICACY OF VARIOUS LOCAL HONEY FOR THE TREATMENT OF
BURN WOUNDS**

By

ROZAINI BT. MOHD. ZOHDI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirement for the Degree of Master of Science**

May 2005



To my mum and dad for their love and support



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Master of Science

EFFICACY OF VARIOUS LOCAL HONEY FOR THE TREATMENT OF BURN WOUNDS

By

ROZAINI BT. MOHD. ZOHDI

May 2005

Chairman : Md. Zuki Bin Abu Bakar@Zakaria, PhD

Faculty : Veterinary Medicine

Since time immemorial honey has been known to treat myriad of wounds and ailments. Recently, honey has been revived as an effective treatment for wounds and the interests that spark in approaching alternative treatments stem partly from the emergence of antibiotic-resistance pathogens. In addition burn care is an expensive proposition which requires significant duration of hospital stay as well as expensive medications. Since honey is produced from many sources of nectar, the chemical and physical activities vary greatly with origin of the nectar as well as environmental conditions. Thus, the present study was undertaken to assess the potential of various Malaysian honeys in treating burn wound.

The efficacy of topical application of Malaysian honeys on burn wound healing in Sprague-Dawley rats was investigated on the basis of biophysical and histological changes. A total of 210 Sprague-Dawley male rats weighing between 200 - 300 g were used in this study. Deep partial skin thickness burn wound was inflicted on the dorsal part of the



body. Imported Manuka honey as well as four selected local honeys collected from different plantations namely nenas, gelam, durian and kelapa were applied twice daily in a quantity of 0.5 ml for each application. Control animals received no treatment while silver sulphadiazine (SSD) cream served as a standard burn wound treatment. The rats were inspected daily and the general appearance as well as the rate of wound contraction was recorded at 3, 7, 14, 21 and 28 days post burned. Six rats from each experimental group were euthanized at each time interval and the skin samples taken were evaluated histologically and subjected to tensile strength test. Tissue sections were stained with haematoxylin and eosin (H&E) and Masson's trichrome staining, while tensile strength testing was done using an Instron™ tensiometer.

The results obtained from this study showed that Manuka honey and Gelam honey significantly stimulated the rate of burn wound healing as demonstrated by increased rate of wound contraction and from gross observations. Microscopic evaluation demonstrated that there was a significant acceleration of the dermal repair in wound healing treated with Manuka and Gelam honeys. Early attenuation of inflammatory reaction and early reparative activities were observed in wounds treated with the two types of honeys. Differential cells count showed a significant decrease in the number of inflammatory cells in the Manuka honey and Gelam honey treated wounds as early as 3 days post injury. In

addition, epithelial regeneration appeared to be quite advanced whereby re-epithelialization was observed as early as 7 days after burn treatment as compared to other experimental groups. Histological findings of this study also showed enhanced proliferation of fibroblasts and collagen synthesis in wounds treated with Manuka honey and Gelam honey. In addition, tensile strength of the wounds treated with these honeys was also enhanced during the course of study.

Thus, results obtained from the present study suggested that topical application of Manuka and Gelam honey may have favourable influence on the various phases of burn wound healing hence accelerating the healing process.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**KEGUNAAN PELBAGAI JENIS MADU TEMPATAN SEBAGAI
RAWATAN ALTERNATIF UNTUK MERAWAT LUKA TERBAKAR**

Oleh

ROZAINI BT. MOHD. ZOHDİ

Mei 2005

Pengerusi : Md. Zuki Bin Abu Bakar@Zakaria, PhD

Fakulti : Perubatan Veterinar

Madu telah digunakan untuk merawat pelbagai jenis luka dan penyakit sejak berabad tahun dahulu. Kebelakangan ini, madu telah dikenali semula sebagai rawatan yang efektif untuk luka dan minat dalam mengkaji rawatan alternatif terbit daripada masalah kewujudan patogen yang mempunyai ketahanan terhadap antibiotik. Tambahan pula, rawatan terhadap luka akibat terbakar memerlukan kos yang tinggi dan pesakit terpaksa tinggal di hospital dalam jangka masa yang lama serta ubat-ubatan yang mahal diperlukan. Oleh kerana madu dihasilkan daripada sumber nektar yang berlainan maka aktiviti kimia dan fizikalnya turut berbeza bergantung kepada sumber nektar dan keadaan kawasan persekitaran. Oleh itu, ujikaji ini dijalankan untuk mengesan keberkesanan madu Malaysia dalam merawat luka terbakar.

Keberkesanan madu Malaysia yang disapu secara topikal ke atas luka terbakar diuji melalui perubahan biofizikal dan histologikal menggunakan



tikus-tikus jenis Sprague-Dawley. Sejumlah 210 ekor tikus jantan jenis Sprague-Dawley yang beratnya antara 200 - 300 g digunakan untuk tujuan tersebut. Luka terbakar tahap kedua telah diwujudkan pada bahagian belakang badan tikus. Madu jenis Manuka yang diimport dan empat jenis madu tempatan terpilih yang diambil daripada dusun yang berbeza iaitu Nenas, Gelam, Durian dan Kelapa disapu 2 kali sehari ke atas luka tersebut dalam kuantiti 0.5 ml untuk setiap kali sapuan. Tikus dalam kumpulan kawalan tidak diberi apa-apa rawatan lanjutan manakala krim silver sulphadiazine (SSD) dijadikan sebagai rawatan kawalan untuk kesan luka terbakar. Tikus-tikus tersebut diawasi setiap hari untuk jangka masa 28 hari. Keadaan serta perubahan luka secara umum dicatat dan kontraksi luka yang berlaku direkodkan pada hari ke 3, 7, 14, 21 dan 28 hari selepas terbakar. Enam ekor tikus daripada setiap kumpulan ujikaji dimatikan mengikut tempoh masa yang ditetapkan dan sampel kulit diambil untuk ujian secara mikroskopik dan ujian kekenyalan. Pewarnaan haematoxylin & eosin (H&E) dan Masson's trichrome digunakan dalam ujian mikroskopik manakala ujian kekenyalan dijalankan menggunakan mesin Instron™ tensiometer.

Hasil ujian menunjukkan bahawa madu jenis Manuka dan Gelam merangsang kadar penyembuhan luka terbakar secara signifikan melalui peningkatan kontraksi luka dan daripada perubahan secara umum keadaan fizikal luka-luka tersebut. Ujian mikroskopik juga menunjukkan bahawa proses penyembuhan derma meningkat secara signifikan apabila dirawat dengan madu jenis Manuka dan Gelam. Kawalan awal terhadap reaksi

inflamasi dan proses penyembuhan luka yang awal dapat diperhatikan daripada luka-luka yang dirawat dengan 2 jenis madu berkenaan. Tambahan pula, lapisan epitelium berproliferasi dengan cepat iaitu seawal 7 hari selepas dibakar jika dibandingkan dengan kumpulan-kumpulan eksperimen yang lain. Ujikaji secara mikroskopik juga menunjukkan peningkatan proliferasi sel fibroblast dan sintesis awal kolagen baru dalam luka yang dirawat dengan madu jenis Manuka dan Gelam. Ujian kekenyalan pula menunjukkan peningkatan kekenyalan kulit secara beransur sepanjang eksperimen dijalankan.

Hasil kajian menyarankan keberkesanan madu jenis Manuka dan Gelam yang disapu secara topikal ke atas kesan luka terbakar dapat merangsang pelbagai peringkat dalam proses penyembuhan luka.

ACKNOWLEDGEMENTS

All praise and thanks be to Allah, God the Almighty, most Beneficent and most Merciful. He, Who has sustained me both physically and spiritually, and Who has given me the strength in completing this thesis. Without His blessings none of this would have been possible.

I have been fortunate enough to be doing this research under the supervision of Dr. Md. Zuki Bin Abu Bakar and I am fully conscious of the debt which I owed to him for his invaluable advice and for his subtle guidance during the course of this dissertation project.

I am most grateful to the excellent co-operation given to me by the members of my committee throughout my course of study and in completing this project. Assoc. Prof. Dr. Noordin Bin Mohamed Mustapha especially for his guidance and advice on tissue pathology, Assoc. Prof. Dr. Muhammad Nazrul Hakim Bin Abdullah and Dr. Norimah Bt. Yusof who counseled, commented, and critiqued my work in one form or another, for which I am grateful.

Special thanks are due to Encik Zainal Bin Romly from ZHR Technology Division for kindly providing the Active Manuka honey for this study and shared his knowledge and expertise concerning honey. Many thanks to Encik



Mokhtaruddin Bin Husain from the Department of Agriculture Malaysia and to the wonderful people who works in Batu Pahat bee farm for supplying the local honeys.

It is a pleasure also to acknowledge my indebtedness to Dr. Goh Yong Meng and Dr. Latiffah Bt. Hassan for their statistical advice. I would like to convey my sincerest gratitude to Encik Mohd. Jamil Bin Samad, Encik Kufli Bin Che Nor and to all the staff in the Faculty of Veterinary Medicine for their kind support and helpful co-operation rendered at the time of this project was carried out.

My sincere appreciation and thanks are also extended to Puan Asnah Bt. Hassan and Encik Zahid Bin Abdullah who have assisted in completing this thesis and the rest of the staff in MINT, Bangi for their technical support.

I also thank my loving family for their continuous support and understanding specially my wonderful parents, Tuan Hj. Mohd. Zohdi Bin Hj. Mohd. Noor and Puan Hjh. Mariah Bt. Hussin. Not to be forgotten my dear loving sister and brothers as well as my two adorable nephews.

And lastly, I would like to acknowledge my supportive friends especially Fadilah, Faizah, Dr. Sow Po Po, Dr. Ani, Ainul Hafizah and Dr. Hafeez for their valuable discussion and constructive advice during the research.



TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xvii
LIST OF FIGURES	xviii
LIST OF ABBREVIATIONS	xxii
 CHAPTER	
 I	
INTRODUCTION	1
General introduction	1
Objectives	4
 II	
LITERATURE REVIEW	5
The skin	5
General anatomy and histology of the skin	7
Rat as an animal model	9
Biomechanical properties of the skin	10
Wound healing	14
Phases of wound healing	14
The burn wound	17
Types of burns	18
Classification of burns	20
Tissue response to burn wound injury	25
Complications	27
Treatment	28
Burn wound healing	28
Formation of granulation tissues	29
Epithelialization	30
Wound contraction	31
Scar formation	31
Topical medications for burn wound treatment	32
Silver sulphadiazine	32
Alternative treatment for burn wound	34
Honey	34
Manuka honey	38



III	GENERAL MATERIALS AND METHODS	40
	Experimental animal	40
	Experimental design	40
	Preparation of the skin	41
	The burn injury	42
	Thermal source	42
	Location of the lesion	42
	Infliction of the burn wound	43
	Honey samples	43
	Mode of treatment	44
IV	MACROSCOPIC EVALUATION OF BURN WOUND HEALING PROGRESS TREATED WITH DIFFERENT TYPES OF HONEY	49
	Introduction	49
	Materials and Methods	50
	Animals and experimental design	50
	Assessment of burn healing	51
	Statistical analysis	52
	Result	54
	Macroscopic appearances	54
	Wound contraction	55
	Discussion	72
	Conclusion	76
V	MICROSCOPIC EVALUATION OF BURN WOUND HEALING TREATED WITH DIFFERENT TYPES OF HONEY	77
	Introduction	77
	Materials and Methods	78
	Animals and experimental design	78
	Light microscope evaluation	79
	Statistical analysis	81
	Result	83
	Discussion	154
	Conclusion	162
VI	TENSILE STRENGTH STUDY OF BURN WOUND TISSUE HEALING TREATED WITH DIFFERENT TYPES OF HONEY	163
	Introduction	163
	Materials and Methods	164
	Animals and experimental design	164
	Preparation of the strips of skin for mechanical test	165
	Tensiometer	166



Statistical analysis	169
Result	169
Discussion	173
Conclusion	178
Limitation	179
VII GENERAL DISCUSSION	181
Conclusion	186
BIBLIOGRAPHY	188
APPENDICES	200
BIODATA OF THE AUTHOR	202



LIST OF TABLES

Table		Page
3.1	The experimental design	41
4.1	The experimental design for macroscopic study	50
4.2	Longitudinal and transverse measurements of wound area as percentage of original wound size at day 3 post burn	65
4.3	Longitudinal measurements of wound contraction of the control and treated groups at different days as percentage of original wound size	68
4.4	Transverse measurements of wound contraction of the control and treated groups at different days as percentage of original wound size	70
5.1	The experimental design for microscopic study	79
5.2	Quantitative histopathologic findings of the dermal collagen changes based on Masson's trichrome staining	81
5.3	Data of differential cell counts expressed in mean \pm SD	84
5.4	Quantitative histopathologic findings of dermal collagen changes as indicated by red stained denatured collagen in Masson's trichrome staining	90
6.1	The experimental design for tensile strength study	165
6.2	Tensile strength measurement (MPa) of healing wounds	171



LIST OF FIGURES

Figures		Page
2.1	Schematic diagram of the skin	6
2.2	Schematic representation of phases of wound healing	16
2.3	Illustration of superficial thickness burn wound	23
2.4	Illustration of superficial partial thickness burn wound	24
2.5	Illustration of deep partial thickness burn wound	24
2.6	Illustration of full thickness burn wound	25
3.1	The shaved skin of dorsal part of animal	46
3.2	The cylindrical aluminium templates	46
3.3	Marking the location of the burn infliction	47
3.4	Infliction of the burn with the template	47
3.5	Summary of the methodology	48
4.1	Longitudinal measurement of the burn diameter	53
4.2	Transverse measurement of the burn diameter	53
4.3	General appearance of the wound sites in all experimental groups at day 0 post injury	59
4.4	General appearance of the wound sites in all experimental groups at day 3 post injury	60
4.5	General appearance of the wound sites in all experimental groups at day 7 post injury	61



4.6	General appearance of the wound sites in all experimental groups at day 14 post injury	62
4.7	General appearance of the wound sites in all experimental groups at day 21 post injury	63
4.8	General appearance of the wound sites in all experimental groups at day 28 post injury	64
4.9	Graph of longitudinal measurement of wound area of the control and treated groups as percentage of original wound size at day 3 post burns	66
4.10	Graph of transverse measurement of wound area of the control and treated groups as percentage of original wound size at day 3 post burns	67
4.11	Graph of longitudinal measurement (percentage of wound contraction)	69
4.12	Graph of transverse measurement (percentage of wound contraction)	71
5.1	Photomicrograph of wound sections showing cells that were identified and counted using 0.01mm graticule	82
5.2	Graph plotting mean number of neutrophils per unit area against days of post injury	85
5.3	Graph plotting mean number of macrophages per unit area against days of post injury	86



5.4	Graph plotting mean number of fibroblasts per unit area against days of post injury	87
5.5	Graph plotting mean number of endothelial cells per unit area against days of post injury	88
5.6	Photomicrographs of wound sections at day 3 post burned stained with H&E under magnification of 400X	94
5.7	Photomicrographs of wound sections at day 3 post burned stained with Masson trichrome under magnification of 400X	99
5.8	Photomicrographs of wound sections at day 7 post burned stained with H&E under magnification of 400X	107
5.9	Photomicrographs of wound sections at day 7 post burned stained with Masson trichrome under magnification of 400X	112
5.10	Photomicrographs of wound sections at day 14 post burned stained with H&E under magnification of 400X	120
5.11	Photomicrographs of wound sections at day 14 post burned stained with Masson trichrome under magnification of 400X	125
5.12	Photomicrographs of wound sections at day 21 post burned stained with H&E under magnification of 400X	132



5.13	Photomicrographs of wound sections at day 21 post burned stained with Masson trichrome under magnification of 400X	137
5.14	Photomicrographs of wound sections at day 28 post burned stained with H&E under magnification of 400X	144
5.15	Photomicrographs of wound sections stained at day 28 post burned with Masson trichrome under magnification of 400X	149
6.1	Samples of strips for tensile strength test	167
6.2	Measurement of strips's thickness using thickness-gauge Mitutoyo	167
6.3	The Instron Model 4301 mechanical testing machine	168
6.4	The strips of skin were held into place on the machine with pneumatic clamps	168
6.5	Graph of tensile strength of healing wounds for all experimental groups	172



LIST OF ABBREVIATIONS

%	percent
<	less than
>	more than
°C	degree Celsius
µm	micrometer
cm	centimeter
<i>et al.</i>	and others (Latin: et alii)
g	gram
H&E	haematoxylin and eosin
kg	kilogram
kGy	kilo Gray
mg	milligram
MINT	Malaysians Institute for Nuclear Technology Research
ml	milliliter
mm	millimeter
Mpa	megapascal
P	probability
SSD	silver sulphadiazine



CHAPTER ONE

INTRODUCTION

General Introduction

In the Al-Quran, Verse 16 of Surah Al-Nahl (No. 68-69) is quoted as saying “And the Lord inspired the bee, saying: Take your habitations in the mountains and in the trees and in what they erect. There comes forth from their bellies a drink of varying color wherein is healing for men. Verily in this is indeed a sign for people who think.”

During the past decade there has been a global interest in the use of traditional and complementary medicine. Most scientific research has focused on herbal as well as aromatherapy products. In addition, a number of other naturally occurring substances have been proven to show therapeutic promise. One such resource that was claimed to have curative value is honey.

Ironically, honey has been used as a medicine for thousand of years and its healing properties are well documented (Molan, 1999a). Honey has been used to treat a wide range of wounds of various aetiology including abscess, surgical wounds, ulcers and burns (Molan, 1999b). It was claimed that early Egyptians were the first to use honey as a component in the topical treatment of wounds as evidence from their writing in the Smith papyrus (1650BC)



(Forrest, 1982). Thus, Zumla and Lulat (1989) referred honey as 'a remedy rediscovered' due to the resurgence of its usage in modern professional medicine. Perhaps the rising interest in the use of alternative therapies is mainly due to the expanding problem of antibiotic resistance in bacteria or because some people are experiencing the possible side effects of many pharmaceuticals products (Sai & Babu, 1998).

Honey is a mixture of sugars prepared by the bees from the natural sugar solutions called nectar obtained from flowers (Subrahmanyam, 1996). It is produced from many sources, and its antimicrobial activity varies greatly with origin and processing (Molan, 2001). Therapeutic honeys offer considerable benefits in wound care, particularly for the treatment of chronic and infected wounds and for the treatment of burns (Lusby *et al.*, 2002). Its efficacy in wound healing remains largely anecdotal with claims that it reduces inflammation, debrides necrotic tissue, reduces oedema and promotes angiogenesis, granulation and epithelialization (Molan, 1998a). When used as a topical application, honey was found to accelerate wound healing and its antibacterial properties reduced bacterial growth (Bergman *et al.*, 1983). Therapeutic effects of honey were also found to be useful in the treatment of burn, by helping the rapid healing of wounds with less scarring (Subrahmanyam, 1991). It gives rapid deodorization of offensively smelling wounds, which is an unpleasant characteristic feature of burn treatment unit (Molan, 1998b). Its antibacterial effect caused the wounds to heal earlier by

