

UNIVERSITI PUTRA MALAYSIA

CHARACTERIZATION OF HEAVY-METAL-REMOVAL BACTERIA FROM THE PERSIAN GULF

HOSSEIN ZOLGHARNEIN

FPV 2005 2

CHARACTERIZATION OF HEAVY-METAL-REMOVAL BACTERIA FROM THE PERSIAN GULF

By

HOSSEIN ZOLGHARNEIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Doctor of Philosophy

October 2005



DEDICATION

To

Memory of my parents whom their spirits will always be a part of mine

My wife Saya for years of love and dedication, and my sons Sina and Soheil

Thanks to Allah



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

CHARACTERIZATION OF HEAVY-METAL-REMOVAL BACTERIA FROM THE PERSIAN GULF

By

HOSSEIN ZOLGHARNEIN

October 2005

Supervisor: Professor Mohd Azmi Mohd Lila, PhD

Faculty: Veterinary Medicine

The study was carried out to isolate and screen high heavy metals resistant bacteria from Persian Gulf and enclosed industrial areas within 241,000 km². A total of 35 heavy metal resistant bacteria strains were identified from sediment and water samples collected. The resistance and biological capacity of the isolated bacteria were tested in a new formulated media, minimal salt solution (M.S.S), that mimics seawater. Isolated bacteria responded to media supplemented in range 0.5 to 2 mM of Zn, Cd, Cu and Pb by showing a prolonged lag phase and by decreasing growth rate.

Bacteria isolates, in the form of free or immobilized cells, are able to remove lead, copper, zinc and cadmium from solution. Removal of lead and cadmium from solution by some bacteria was very efficient, about 120 mg/g dry weight as high as 90%. Isolates tested presented distinct uptake capacities and the best results were obtained for *Delftia tsuruhatensis* and *Pseudomonas AU3411* respectively.



The diversity of isolated bacteria was examined by the phylogenetic analysis of 16S rRNA gene sequences. The phylogenetic analyses of the sequences revealed seven main taxonomic lineages. The phylogenetic tree illustrated discrimination between isolated bacteria from wastewater, industrials areas and marine environment. Some *Pseudomonas* strains isolated from marine environment were well differentiated from those of industrial wastewater. Members of the genera *Delftia* and *Bacterium* formed a monophyletic group within the subdivision of the class. There was a clear differentiation between two groups of *Pseudomonas* and other groups of bacteria in the phylogenetic tree.

The isolated bacteria were tested for the occurrence of plasmid using the modified alkaline lysate method. The study revealed that the frequency of the occurrence of plasmid in the heavy metals resistance bacteria was more than in the common bacteria. Multiple forms of plasmids were observed in 66% of the plasmid-carrying strains. Isolates bacteria from industrials wastewater showed the highest plasmid incidence (84.6%). In the marine environment there was a slightly higher incidence of plasmid in bacteria isolated from sediments (55.5%) compared to the water sample of the same origin (53.8%).

Scanning Electron Microscope (SEM) analyses showed *Pseudomonas* sp. accumulated heavy metals in the cell wall and along the external cell surfaces. This suggested that heavy metals uptake involves both surface phenomena and diffusion. Energy Dispersive X-ray (EDX) analysis confirmed heavy metals on the bacteria cell surface which was reported by SEM.



PENCIRIAN BAKTERIA YANG DIPEROLEHI DARIPADA TELUK PARSI YANG BERUPAYA MENYINGKIR LOGAM BERAT

Oleh

HOSSEIN ZOLGHARNEIN

Oktober 2005

Pengerusi: Profesor Mohd Azmi Mohd Lila, PhD

Fakulti: Perubatan Veterinar

Beberapa jenis bakteria yang berintang logam berat telah diasingkan daripada sampel keladak dan air, yang diperolehi daripada Teluk Parsi dan kawasan industri disekitarnya dalam lingkungan 241,000 km². Sejumlah 35 jenis bakteria yang berintang logam berat telah dikenalpasti. Kesemua bakteria tersebut telah dicirikan berdasarkan keupayaan bakteria tersebut untuk tumbuh dan mengikatkan logam berat penting seperti zink, kadmium, kuprum dan plumbum pada kepekatan yang tinggi. Rintangan dan kemampuan biologi bakteria diuji dengan menggunakan media baru yang dirumuskan, Minimal Salt Solution (MMS), yang menyerupai air laut.

Aplikasi kadmium, plumbum, kuprum dan zink telah diselidik dengan menggunakan bakteria yang berbentuk tertambat dan bebas. Ujian dilakukan menggunakan dengan samada satu atau campuran beberapa logam berat. Terdapat banyak jenis bakteria yang mampu menyingkirkan kuprum, zink dan kadmium, samada sel bakteria dalam bentuk bebas atau tertambat. Satu atau lebih



campuran logam berat mempunyai afiniti yang berbeza terhadap pengikatan logam-logam berat. Kecekapan pengikatan untuk kadmium dan plumbum mencapai 90% peratusan atau lebih, berdasarkan berat kering sel bakteria dalam masa satu jam pendedahan. Bio-pengikatan kuprum dan zink mencapai 50%. Keupayaan pengikatan yang tertinggi ialah untuk Pb diikuti oleh Cd, Zn dan Cu. Walau bagaimanapun, penyingkiran logam berat adalah maksimum apabila bakteria didedahkan kepada sejenis logam berat sahaja pada satu masa.

Diversiti bakteria telah diuji dengan analisis filogenetik jujukan gen 16S rRNA . Analisis filogenetik pada jujukan tersebut mendedahkan tujuh jalinan taksonomi utama. Cabang filogenetik menunjukkan perbezaan antara bakteria yang diasingkan daripada kumbahan persekitaran industri dan air laut. Keputusan kajian menunjukkan perbezaan genetik yang baru, dan hubungan di antara bakteria yang diperolehi dari laut dan industri. Sebahagian bakteria pseudomonas yang diasingkan daripada persekitaran laut telah dicirikan/dibezakan dengan jelas daripada bakteria yang diasingkan daripada kawasan industri. Ahli genera *Delftia* dan *Bacterium* membentuk kumpulan monofiletik dalam sub-bahagian kelas tersebut. Terdapat perbezaan jelas di antara dua kumpulan pseudomonas dan kumpulan bakteria yang lain di dalam cabang filogenetik.

Bakteria tersebut telah diuji untuk menentukan kewujudan plasmid di dalam bakteria, dengan kaedah pemecahan beralkali. Kaedah ini adalah berkesan untuk pencirian dan pengenlpastian plasmid yang berlainan saiz, tanpa menggunakan bahan kimia toksik. Kajian mendedahkan kekerapan kewujudan plasmid dalam bakteria berintang logam berat adalah lebih tinggi berbanding dengan bakteria biasa. Kajian menunjukkan bahawa 66% daripada bakteria yang diasingkan



mempunyai plasmid besar atau kecil. Bakteria yang diasingkan daripada sisa air industri mempunyai peratusan kewujudan plasmid yang paling tinggi (84.6%). Di persekitaran laut pula, bakteria yang diasingkan daripada keladak mempunyai peratus kewujudan plasmid yang lebih tinggi (55.5%) berbanding dengan bakteria yang diasingkan daripada air laut (53.8%). Penemuan ini mencadangkan bahawa plasmid boleh terdapat pada kebanyakan jenis bakteria tetapi lebih cenderung kepada bakteria berintang logam berat.

Analisis Scanning Electron Microscope (SEM) menunjukkan bahawa spesies pseudomonas mengumpul logam berat dalam dinding sel dan sepanjang permukaan luar sel. Ini menunjukkan pengambilan logam-logam berat melibatkan fenomena permukaan dan penyebaran. Analisis Energy Dispersive Xray (EDX) adalah kurang sensitif dan kurang dipercayai.berbanding dengan analisis penjerapan atom, walaupun EDX adalah satu cara yang cepat dan mudah untuk mengesan kehadiran logam berat ke dalam bakteria.



ACKNOWLEDGEMENTS

I would like to extend my gratitude to the members of my supervisory committee; Prof. Dr. Mohd. Azmi. Lila. Mohd, for his advice, kind and constant support, encouragement and help in all my difficulties.

I wish to express my deepest thankfulness to my co-supervisors; Prof. Dr Mohd Zamri. Saad, Assoc. Prof. Dr Abd. Rahim Mutalib, Assoc. Prof. Dr Che Abd. Rahim Mohamed, for their constructive guidance through out my study period.

Furthermore, my special thanks to Mr Kamarudin, and also John .Shia, Mrs. Sandy, Hayati, Dr Phong and Suria of the virology laboratory.

I am also grateful to the Department of Environment of Iran for providing me the facilities to collect sediments and water samples from Persian Gulf on the research ship Ghods, and also help in heavy metals analysis and isolation of bacteria and other laboratory work.

I would like to thank my wife and my sons, who patiently support me which away from home.

Last, I would like to thank many others, who help me during my study.



TABLE OF CONTENTS

Page

i
iv
viii
ix
xi
XV
xvii
xxi

CHAPTER	1
1 GENERAL INTRODUCTION	1
1.1 Study area	1
1.2 Persian Gulf Pollution and Heavy Metal Sources	3
1.3 The Function of Heavy Metals in the Living Things	4
1.4 Bacteria and Heavy Metals	4
1.5 Heavy Metal Bioaccumulation	5
1.6 The Goal and Objectives of the Study	7
	8
2 LITERATURE REVIEW	8
2.1 TheRole of Heavy Metals in Bacteria	8
2.2 Mechanism of Heavy Metal Uptake in Bacteria	9
2.3 Bacterial Resistance to Heavy Metals	9
2.4 Kinetic and the Growth Rate of Bacterial	11
2.5 Heavy Metal Uptake by Bacteria	12
2.6 DNA Techniques and Molecular Identification of Bacteria	17
2.7 Bacterial Phylogenetic Analysis	18
2.8 The role of Plasmid in the Bacteria	19
2.9 Resistance to Copper	21
2.10 Bacterial Plasmids Encode Resistance Systems	22
2.11 Heavy Metal Resistance Mechanisms	22
2.12 Scanning of Bacteria by X-Ray Energy Dispersive Analysis	
(EDX) and Scanning Electron Microscope (SEM)	24
2.13 Development of Heavy Metal Resistance Bacteria	26
3 KINETIC STUDY OF HEAVY METAL RESISTANCE BACTERIA	28
3.1 Introduction	28
3.2 Material and Methods	30
3.2.1 Sampling Location	30
3.2.2 Sampling	34



3.2.3 Isolation of the Heavy Metals Resistance Bacterial	34
3.2.4 Bacterial Identification	34
3.2.5 The growth Rate of the Heavy Metal Resistance Bacteria	35
3.2.5.1 Growth Condition	35
3.2.5.2 Measuring the Growth of Heavy Metal Resistance	e
Bacteria in the Presence of Heavy Metals	35
3.2.5.3 The Growth Rate of Bacterial Isolates in Differen	t
Concentration of Single Heavy Metals	36
3.2.5.4 The Growth Rate of Bacterial Isolated in Multi-mix	K
Heavy Metals	36
3.2.5.5 The Growth Rate of Bacterial Isolates as the Contro	1 37
3.2.6 Lyophilization	37
3.2.6.1 Preparation of Ampoules	3/
3.2.6.2 Freparation of Cell Suspension for Freeze-drying	37
Suspension	28
3.2.6.4 The Freeze-drying Procedure	38
3.2.6.5 Constriction of Amnoules Secondary-drying and	
Sealing	- 49
3.3 Results	40
3.3.1 Revival of Cultures from Freeze-dried Ampoules	40
3.3.2 The Growth Rate of Bacteria in the Presence of Heavy	Ý
Metals	45
3.3.3 The Growth Rate of Pseudomonas aeruginosa strain	1
ACTT 27853 in Heavy Metals	45
3.3.4 The Growth Rate of <i>Pseudomonas</i> sp. ML2 in Heavy	/ · ·
Metals	49
3.3.5 The Growth Rate of <i>P</i> .aeruginosa strain AU3411 in	1
Heavy Metals	52
3.3.6 The Growth Rate of <i>Pseudomonas putida</i> strain MMTT if	1 56
Heavy Metals	50 Ic 50
3.3.8 The Growth Rate of Pseudomonas tolassii in Heavy	IS 57
Metals	63
339 The Growth Rate of <i>Pseudomonas</i> sp. Fa27 in Heavy	v
Metals	66
3.3.10 The Growth Rate of <i>Delftia</i> sp. AN3 in Heavy Metals	69
3.3.11 The Growth Rate of Methylobacterium sp. Mil in Heavy	/
Metals	72
3.4. Discussion	76
4 HEAVY METALS UPTAKE OF THE BACTERIA ISOLATED)
FROM PERSIAN GULF	80 02
4.1 Introduction	00 00
4.2 Material and Methods	90

4.2.1 Bacterial Growth Condition	90
4.2.2 Uptake of Copper, Zinc, Cadmium and Lead by Bacteria	90
4.2.3 Measuring of Heavy Metals in Bacterial Cells	91
4.2.4 Immobilization of Bacterial Cells	91



4.3 Results	92
4.3.1 Copper Accumulation	93
4.3.2 Zinc Accumulation	94
4.3.3 Cadmium Accumulation	95
4.3.4 Lead Accumulation	95
4.3.5 Accumulation of Multi-mix Heavy Metals	96
4.3.6 Heavy Metal Accumulation by Immobilized Cells	97
4.4 Disccusion	107

5 MOLECULAR CHARACTERIZATION OF HEAVY METAL REMOVAL BACTERIA

5.1 Introduction	112
5.2 Material and Methods	115
5.2.1 Microorganisms and Growth Conditions	115
5.2.2 Bacterial DNA Preparation	115
5.2.3 Determination of DNA Concentration and Purity	116
5.2.4 PCR Primers	116
5.2.5 PCR Amplification	117
5.2.6 Gel Electrophoresis	117
5.2.7 Purification of PCR Amplified 16S rRNA Gene	118
5.2.8 Sequence Analysis	118
5.3 Results	119
5.4 Discussion	147

6 CHARACTERIZATION OF PLASMID CONTAINING BACTERIA	152
6.1 Introduction	152
6.2 Materials and Methods	155
6.2.1 Medium Preparation	155
6.2.2 Isolation and Growth Condition	155
6.2.3 Plasmid Isolation	156
6.2.4 Determination of DNA Concentration	158
6.2.4 Gel Electrophoresis	158
6.3 Results	159
6.4 Discussion	166

7 SEM AND EDX ANALYSIS OF HEAVY METALS REMOVING BACTERIA

7.1 Introduction	170
7.2 Material and Methods	172
7.2.1 Bacteria Strain	172
7.2.2 Growth Condition	172
7.2.3 Bacteria Preparation	172
7.2.4 Washing the Sample	173
7.2.5 Post Fixation	173
7.2.6 Dehydration	173
7.2.7 Critical Point Drying	174
7.2.8 Mounting the Sample on a Stub	174



170

112

7.2.9 Coating Sample with Gold Alloy	174
7.3 Results	175
7.4 Discussion	195
8 GENERAL DISCUSSION AND CONCLUSION	201
REFRENCES	215
APPENDICS	244
BIODATA OF THE AUTHOR	254



LIST OF TABLE

Table	Page
3.1: Sampling location in the Persian Gulf	32
3.2: Identification of Heavy Metal resistance Bacteria from Persian Gulf (ref to table 3.1) and enclosed industrials	43
4.1: Heavy Metal accumulation by isolates Bacteria Roane and Pepper (2000).	88
4.2: t-test analysis between copper at 0.5 mM and 1 mM of concentration	94
4.3 : t-test analysis between zinc at 0.5 and 1 mM concentration	94
4.4: t-test analysis between 0.5 mM and 1 mM of cadmium	95
4.5: t-test analysis between 0.5 mM and 1 mM of lead concentration	96
4.6: One-Way ANOVA on between copper, zinc and cadmium at multi- mix condition	96
4.7 t-test analysis between uptake of lead at immobilized and free bacteria cells condition	97
4.8: Heavy Metals uptake by isolates Bacteria at 0.5 mM of concentration	98
4.9: Heavy Metal uptake by isolates Bacteria at 1 mM of concentration at kinetic condition	100
4.10:Accumulation of Heavy Metals in immobilized Bacteria	102
6.1: Frequency of plasmid-carrying by isolated from different sample source	161
6.2: Frequency of plasmid-carrying strains isolated from different sample source	161
6.3: Molecular weight of plasmids isolates Bacteria	161
6.4: Shows the identification of isolated Bacteria from Persian Gulf and enclosed industrials	162



7.1: EDX analysis of copper in the <i>Pseudomonas</i> when exposed to 1 mM copper for over night.	187
7.2: EDX analysis of zinc in the <i>Pseudomonas</i> when exposed to 1 mM zinc for over night.	189
7.3: EDX analysis of cadmium in the <i>Pseudomonas</i> , when exposed to 1 mM cadmium for over night.	191
7.4: EDX analysis of lead in the <i>Pseudomonas</i> when exposed to 1 mM lead for over night	193



LIST OF FIGURES

Figure	Page
1.1: The Persian Gulf is surrounded by Iran in the north by United Arab Emirate, Oman, Bahrain and Qatar in south by Kingdom of Arabia Saudi in the south of eas	2
2.1: Relationship between the cell volume and susceptibility to copper toxicity (Howlett & Avery, 1999)	12
2.2: Schematic of the Bacterial and Archaeal 16S rRNA gene approximately 1500 bp in length (Vandamme et al., 1996)	17
 3.1: Sampling location including: transects (T1-T15) and stations (1-72) in the Persian Gulf on summer 2001, the cruise started from Oman sea and finished near Kuwait. The samples were collected from 72 stations of 15 transects of water and sediment 	31
3.2: The growth of Methylobacterium strains on the nutrient agar that supplemented by 1 mM of multi-mix heavy metals (1mM of Cd, 1mM of Pb 1mM of Cu and 1mM of Zn)	41
3.3: The growth of Pseudomonas strains on the nutrient agar that supplemented by 1 mM of multi-mix heavy metals (1mM of Cd, 1mM of Pb 1mM of Cu and 1mM of Zn)	42
3.4: The growth of pseudomonas aeruginosa strain ACTT 27853 in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control .	46
3.5: The growth of pseudomonas aeruginosa strain ACTT 2785327853 in 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control.	47
3.6: The growth of pseudomonas aeruginosa strain ACTT 27853 in 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1 mM of multi-mix heavy metals Cu, Zn, Cd, Pb and without heavy metals as the control (b).	48
3.7: The growth of Pseudomonas sp. ML2 in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b)	50



3.8: The growth of Pseudomonas sp. ML2 in 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b)	51
3.9: The growth of Pseudomonas sp. ML2 in 1 mM of multi-mix heavy metals Cu, Zn, Cd, Pb and without heavy metals as the control.	52
3.10: The growth of P. aeruginosa strain AU3411 in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control.	53
3.11: The growth of P. aeruginosa strain AU3411in 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control(a).1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	54
3.12: The growth of P. aeruginosa strain AU3411in 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control(a).1 mM of multi- mix heavy metals Cu, Zn, Cd, Pb and without heavy metals as the control (b).	55
3.13: The growth of Pseudomonas putida strain MM1 in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	57
3.14: The growth of Pseudomonas putida strain MM1 in 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	58
3.15: The growth of Pseudomonas putida strain MM1 in 1 mM of multi-mix heavy metals Cu, Zn, Cd, Pb and without heavy metals as the control.	59
3.16: The growth of Pseudomonas sp. K2 in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control.	60
3.17: The growth of Pseudomonas sp. K2 in 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a). 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b)	61
3.18: The growth of Pseudomonas sp. K2 in 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1 mM of multi-mix heavy metals of Cu, Zn, Cd, Pb and without heavy metals as the control (b)	62
3.19: The growth of Pseudomonas tolaasii in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	64



.

3.20:	The growth of Pseudomonas tolaasii in 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	65
3.21:	The growth of Pseudomonas tolaasii in 1 mM of multi-mix heavy metals Cu, Zn, Cd, Pb and without heavy metals as the control.	66
3.22:	The growth of Pseudomonas sp. Fa27 in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	67
3.23:	The growth of Pseudomonas sp. Fa27 in 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	68
3.24:	The growth of Pseudomonas sp. Fa27 in 1 mM of multi-mix heavy metals Cu, Zn, Cd, Pb and without heavy metals as the control.	69
3.25:	The growth of Delftia sp. AN3 in 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	70
3.26:	The growth of Delftia sp. AN3 in 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (a), 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	71
3.27:	The growth of Delftia sp. AN3 in 1 mM of multi-mix heavy metals Cu, Zn, Cd, Pb and without heavy metals as the control.	72
3.28:	The growth of Methylobacterium sp. Mil 0.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control.	73
3.29:	The growth of Methylobacterium sp. Mil 1 mM of Cu, Zn, Cd, Pb and without heavy metals as the contro(a), 1.5 mM of Cu, Zn, Cd, Pb and without heavy metals as the control (b).	74
3.30:	The growth of Methylobacterium sp. Mil 2 mM of Cu, Zn, Cd, Pb and without heavy metals as the contro(a), 1 mM of multi-mix heavy metals and without heavy metals as the control (b).	75
4.1	Accumulation of lead at 0.5 and 1mM concentration (a), accumulation cadmium at 0.5 mM single and multi heavy metal concentration (b)	104
4.2:	Accumulation of copper, zinc, cadmium by isolates bacteria at 0.5 mM of multi-mix heavy metal (a), and 1 mM multi-mix heavy metal (b)	105



4.4	Accumulation of lead (a) and cadmium (b) by isolates bacteria at 0.5 mM and 1 mM of concentration after immobilizing bacteria cells	106
5.1:	PCR product of 16S rRNA genes after purification (a, b)	123
5.2	PCR product ₁ of 16S rRNA genes after purification (a) and before purification (b)	124
5.3	Comparison of deduced nucleic acid sequences of 16S rRNA from 35 isolates bacteria from Persian Gulf and enclosed industrials, points shows the variable region on the16S rRNA genes	125
5.4	Phylogenetic tree of all bacteria isolated from Persian Gulf. It was clssified based on partial 16S rRNA gene sequences of heavy metal removal bacteria by using DNADIST Neighbor phylogenetic tree program	146
7.1:	SEM photomicrograph of Pseudomonas aeruginosa cells when grew in the nutrient agar media for over night without heavy metals as a control sample	177
7.2:	SEM photomicrograph of Pseudomonas aeruginosa cells when grew in the nutrient agar media for four days without heavy metals as a control sample	178
7.3:	SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM copper for over night indicate bacteria shape modification and copper aggregates associated with the cells	179
7.4	SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM copper for four days indicate bacteria shape modification and copper aggregates associated with the cells	180
7.5:	SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM zinc for over night indicate zinc aggregates associated with the cells	181
7.6:	SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM zinc for four days indicates bacteria shape modification and zinc aggregates associated with the cells	182
7.7:	SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM cadmium for over night indicate cadmium aggregates associated with the cells	183
7.8:	SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM cadmium for four days indicates bacteria shape modification	184



7.9: SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM lead for over night indicate bacteria shape modification and lead aggregates associated with the cells	185
7.10: SEM photomicrograph of Pseudomonas aeruginosa cells when exposed to 1 mM lead for four days indicate bacteria shape modification and lead aggregates associated with the cells	186
7.11: EDX analyses of copper in the Pseudomonas when exposed to 1 mM copper for over night	187
7.12: EDX analysis of copper in the Pseudomonas when exposed to 1 mM copper for over night	188
13: EDX analyses of zinc in the Pseudomonas, when exposed to 1 mM zinc for over night	189
7.14: EDX analysis of zinc in the Pseudomonas aeruginosa ATCC 27853, when exposed to 1 mM zinc for over night	190
7.15: EDX analysis of zinc in the Pseudomonas, when exposed to 1 mM cadmim for over night	191
7.16: EDX analysis of cadmium in the Pseudomonas, when exposed to 1 mM zinc for over night	192
7.17: EDX analysis of lead in the Pseudomonas, when exposed to 1 mM lead for over night	193
7.18: EDX analysis of cadmium in Pseudomonas	194



.

LIST OF ABREVIATIONS

(NH ₄) SO ₄	=	Ammonium sulfate
AAS	=	Atomic Absorption Spectrophotometer
ANOVA	=	Analysis of Variance
bp	=	base pairs
CaCl ₂	=	Calcium chloride
Cd(NO ₃) ₂	=	Cadmium nitrate
CsCl ₂	=	Cesium chloride
CuSO ₄	=	Copper sulfate
dNTP	_	DNA deoxyribonucleic acid
ds	=	double stranded
EDTA	=	Ethylene diamine tetra acetic acid
EDX	=	Energy Dispersive X-ray
		Environment
FeSO ₄	=	Iron sulfate
g	-	Gravity
Gm	=	growth media
K ₂ HPO ₄	=	Di-potassium hydrogen orthophosphate
KH ₂ PO ₄	=	Potassium di-hydrogen orthophosphate
Ls	=	Lab-scale
M.S.S	=	Minimal salt solution
MES	=	Morpholino ethanesulfonic acid
MgSO ₄	=	Magnesium sulfate
MnSO ₄	=	Manganese sulfate



NaCl	=	Sodium chloride
OD	=	Optical density
Р	=	Probability
PbNO ₃	=	Lead nitrate
PBS	=	Phosphate buffered saline
PCR	=	Polymerase chain reaction
рН	=	Negative logarithm of hydrogen ion
ppt	=	Part per thousand
ROPME	=	Regional Organization for the Protection of the Marine
rpm	=	rotation per minutes
SEM	=	Electron Microscope
TAE	=	Tris acetic acid
TE	=	Tris EDTA
TEMED	=	Tetra methyl ethylenediamine
UV	=	Ultra violet
ZnSO ₄	=	Zinc sulfate

