



***ASSESSMENT OF THERAPEUTIC AND ANTIMICROBIAL EFFECTS OF
LOCAL PLANTS ON BACTERIAL DISEASES IN RED HYBRID TILAPIA
(Oreochromis sp.)***

SHARIFAH RAINA BINTI MANAF

FPV 2016 15



**ASSESSMENT OF THERAPEUTIC AND ANTIMICROBIAL EFFECTS OF
LOCAL PLANTS ON BACTERIAL DISEASES IN RED HYBRID TILAPIA
(*Oreochromis* sp.)**

By

SHARIFAH RAINA BINTI MANAF

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the degree of Doctor of
Philosophy**

April 2016

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

With appreciation and respect, this thesis is dedicated to:

My supportive husband, Mr Mahazan Muhammad who made this writing
endurable

My beloved parents, Mr Manaf Daud & Mrs Sharifah Zainun
My beloved parents in law, Mr Muhammad Talib & Mrs Fauziah Ismail
who made it possible

My son, Muslim Ramadhani Mahazan and my siblings who made it all
worthwhile.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

**ASSESSMENT OF THERAPEUTIC AND ANTIMICROBIAL EFFECTS OF
LOCAL PLANTS ON BACTERIAL DISEASES IN RED HYBRID TILAPIA
(*Oreochromis* sp.)**

By

SHARIFAH RAINA BINTI MANAF

April 2016

Chairman : Hassan Mohd Daud, PhD
Faculty : Veterinary Medicine

Currently, aquaculture is regarded as one of the potential strategic industries for Malaysia. The percentage on its production increases year by year. The growth are obvious due to obstacle in continuous dependent on finite stocks of protein source from wild-caught fish which is shown to be over-exploited or near to be depletion. One of main problem in aquaculture is outbreaks of infectious diseases which has incurred big losses to the industry. Herbal medicine has been used in most part of the world as an alternative medicine for human being and domestic animals. The plant compounds have been shown to be beneficial and rich in a wide variety of phytonutrients, and thus used as chemotherapeutics and feed additives. Hence, the general objectives of this study were to determine the effect of herbal supplementation on the growth performance, disease resistant, disease treatment in Red hybrid tilapia (*Oreochromis* sp.). Six extracts of *Vitex trifolia*, *Aloe vera*, *Strobilanthes crispus*, *Clinacanthus nutans*, *Pereskia grandifolia* and *Peperomia pellucida* were determined for phytochemical properties and their antibacterial activities against common freshwater pathogens i.e. *Streptococcus agalactiae*, *Aeromonas hydrophila* and *Enterobacter cloacae*. Qualitatively screening of phytochemical properties in herbs were determined for flavonoids, tannins, saponin, alkaloids steroid and glycoside. The results showed presence of flavonoid in all plant extracts. For the antimicrobial activity, the aqueous and methanolic extracts were tested by using disk diffusion method. Antimicrobial assay of methanolic extracts (25 to 100 mg/mL) showed effectiveness against the pathogenic bacteria. Comparatively, all aqueous extracts did not show any antimicrobial activity. Moderate antibacterial activity was shown by the methanolic extracts of *V. trifolia*, *A. vera* and *S. crispus* while weak antimicrobial activity was shown by *C. nutans*, *P. grandifolia* and *P. pellucida*. From the antimicrobial result, three potential plant extracts namely *V. trifolia* (VTE), *S. crispus* (SCE) and *A. vera* (AVE) were test for toxicity and feeding experiment. The cytotoxicity test using *Artemia salina* (45.9 - 66.7 µg/mL) and BF-2 fish cell line (85.0 - 120.50 µg/mL). The median lethal dose (LD₅₀) value in *Oreochromis* sp. injected intraperitoneal were equivalent to 2.67×10^7

CFU/mL (*S. agalactiae*), 3.10×10^6 CFU/mL (*E. cloacae*) and 3.75×10^5 CFU/mL (*A. hydrophila*). *Oreochromis* sp. were fed with diet supplemented with VTE, SCE and AVE at 2g/kg, 5g/kg, 7g/kg and 9g/kg for 45 days, followed by challenged with 0.1 ml of 10^7 CFU/ml *S. agalactiae*. The results showed *Oreochromis* sp. supplemented with VTE-7 was the best growth promoter among treatments. It showed 57.3% in body weight gain with 100% survivability. For haematological the value of Hb after challenged with *S. agalactiae* showed significant different ($P < 0.05$) compared to control except in SCE-2 and SCE-7. While, in WBC value showed significant different ($P < 0.05$) except in AVE-5 and AVE-7. In MCV value showed significant different except in SCE-5, AVE-9 and in MCHC value showed significant except in VTE-2 and SCE-5. In biochemical parameters of blood, there was significant different in ALT value observed in fish fed with VTE-2, VTE-5, VTE-7, SCE-2, SCE-5, SCE-7, AVE-5 and AVE-7 compared to control (infected). In AST value showed significant different ($P < 0.05$) in VTE-5, VTE-7, SCE-9, AVE-2 and AVE-7 compared to control (infected). Fish fed with SCE-5 was the best for the disease resistant showed the highest level of WBC value indicated increase immune responses and the ability of the fish to fight *S. agalactiae*. In histology examination of different selected herbal extracts showed mild degeneration in liver and kidney tissues. To determine the growth performance and disease resistant of *Oreochromis* sp. supplemented with herbal-mixed of the selected herbal extracts, the fish were fed with diet containing mixtures of AVE&VTE, VTE&SCE, SCE&AVE. Herbal mixed of AVE&VTE showed the most significant increased as compared with others. For SGR, only AVE&VTE showed promising result with significant higher ($P < 0.05$) than control. Fish survival after the disease challenge showed significant difference ($P < 0.05$) compared to control. In haematological assays, the post-challenged results showed the significant increased ($P < 0.05$) of RBC, Hb and PCV values. The WBC value in pre- and post-challenged showed no significant different ($P > 0.05$) of all treatments compared to control. As for MCHC value reading with the mixed-herbals, AVE&SCE was the only treatment which revealed no significant difference ($P > 0.05$) in pre-challenged and showed decreased significant difference ($P < 0.05$) in post-challenged in all treatments. On the other hand, MCV and plasma protein values did not show any significant difference ($P > 0.05$) as compared to control. The biochemical parameters of ALT value and AST value were significantly affected by mixed-herbal supplementation. However in the post-challenged, the ALP value showed no significant difference compared to control. Histopathological changes, which included necrosis and haemorrhage in the kidney, liver and hepatopancreas tissues could be due to the cytotoxin produced by the bacteria. Also, a study was carried out to evaluate the effects of dietary supplementation of methanol extracts of *V. trifolia*, *S. crispus* and *A. vera* *Oreochromis* sp. as disease treatment. Results showed that RBC levels were significantly increased ($P < 0.05$) in VTE-2, SCE-2, SCE-5, SCE-9, AVE-5, AVE-7 and control (OTC) as compared to control (infected). While, Hb value showed significant increased in VTE-2, SCE-5, SCE-9, in contrast to significant decreased in AVE-7 and AVE-9 as compared to control. For PCV value showed significant ($P < 0.05$) increased in all treatments included control (OTC) as compared to control (infected). On the other hand, in MCHC value no significant different for SCE-5. Plasma protein value showed significant increased ($P < 0.05$) after supplementation of VTE-2, VTE-5, SCE-2, SCE-5, SCE-9, AVE-2, AVE-5,

AVE-9. For ALT and AST values, they showed significant decrease in all treatments including control (uninfected) and control (OTC) ($P < 0.05$). Also in ALP value, it showed significant decreased in all treatments including the control (OTC). The survivability of fish showed increase after fed with the herbal supplementation ($P < 0.05$) in VTE-5, VTE-7, AVE-2, AVE-5 and AVE-9 as compared to control (infected). All the haematological measurements in *Oreochromis* sp. that were infected with *S. agalactiae* with supplemented with VTE, SCE and AVE showed similar values compared with the control and the treatment with antibiotic. Its indicated the fish that were infected with *S. agalactiae* able to restore their haematological indices to near normal values. The results were agreeable with literatures which reported the numerous efficacies of the use of herbal in aquaculture to control various bacterial pathogens. Also from current research findings, suggested that mixed herbal of methanolic-extract given to *S. agalactiae* infected *Oreochromis* sp. had a synergistic restorative effect on the hematological variables. In conclusion, current study suggested that *V. trifolia*, *S. crispus* and *A. vera* extracts and mixed herbal were effective as growth promoters and bacterial disease treatment in *Oreochromis* sp. It also proved the potential of the selected herbal plant as aquaculture therapeutant as they were environmental friendly and it showed negligible side effect in the intended fish.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENILAIAN KE ATAS SIFAT TERAPEUTIK DAN KESAN ANTIMIKROBIAL
TUMBUHAN TEMPATAN BAGI PENYAKIT BAKTERIA DALAM TILAPIA
HIBRID MERAH (*Oreochromis* sp.)**

Oleh

SHARIFAH RAINA BINTI MANAF

April 2016

Pengerusi : Hassan Mohd Daud, PhD

Fakulti : Perubatan Veterinar

Pada masa ini, akuakultur dianggap sebagai salah satu industri yang sangat berpotensi untuk Malaysia. Peratus pada pengeluaran meningkat tahun ke tahun. Pertumbuhan akuakultur adalah jelas kerana halangan dalam pergantungan berterusan pada stok terhadap sumber protein daripada ikan tangkapan liar yang menunjukkan sebagai eksploitasi terlampau atau hampir menjadi pupus. Masalah utama dalam akuakultur adalah wabak penyakit berjangkit yang telah menyebabkan kerugian besar kepada industri. Perubatan herba telah digunakan di kebanyakan bahagian dunia sebagai ubat alternatif untuk manusia dan haiwan domestik. Sebatian tumbuhan telah terbukti bermanfaat dan kaya dengan pelbagai khasiat pemakanan, dan dengan itu digunakan sebagai kemoterapeutik dan bahan makanan tambahan. Oleh itu, objektif umum kajian ini adalah untuk menentukan kesan daripada diet tambahan herba terhadap prestasi pertumbuhan, resisten dan rawatan penyakit dalam tilapia hibrid merah (*Oreochromis* sp.). Enam ekstrak *Vitex trifolia*, *Aloe vera*, *Strobilanthes crispus*, *Clinacanthus nutans*, *Pereskia grandifolia* dan *Peperomia pellucida* ditentukan untuk sifat fitokimia dan aktiviti antibakteria terhadap patogen lazim air tawar iaitu *Streptococcus agalactiae*, *Aeromonas hydrophila* dan *Enterobacter cloacae*. Pemeriksaan kualitatif sifat fitokimia dalam herba telah di tentukan pada flavonoid, tanin, saponin, alkaloid, steroid dan glikosid. Hasil kajian menunjukkan kehadiran flavonoid dalam semua ekstrak tumbuhan. Untuk aktiviti antimikrobial, ekstrak akueus dan metanol telah diuji dengan menggunakan kaedah cakera peresapan asai. Antimikrobial asai ekstrak metanol (25 hingga 100 mg/mL) menunjukkan keberkesanan terhadap bakteria patogenik. Secara perbandingan, semua ekstrak akueus tidak menunjukkan apa-apa aktiviti antimikrobial. Aktiviti antibakteria sederhana telah ditunjukkan oleh ekstrak methanol *V. trifolia*, *A. vera* dan *S. crispus* manakala aktiviti antimikrobial lemah telah ditunjukkan oleh *C. nutans*, *P. grandifolia* dan *P. pellucida*. Daripada keputusan antimikrobial, 3 ekstrak tumbuhan berpotensi iaitu *V. trifolia* (VTE), *S. crispus* (SCE) dan *A. vera* (AVE) digunakan dalam penilaian ketoksikan dan eksperimen pemakanan. Ujian sitotoksiti menggunakan *Artemia salina* (45.9 - 66.7 µg/mL)

dan BF-2 sel turutan ikan (85.0-120.50 µg/mL). Nilai dos kematian purata (LD₅₀) dalam *Oreochromis* sp. disuntik secara intraperitoneum bersamaan dengan 2.67 X 10⁷ CFU / mL (*S. agalactiae*), 3.10 X 10⁶ CFU / mL (*E. cloacae*) dan 3.75 X 10⁵ CFU / mL (*A. hydrophila*). *Oreochromis* sp. diberi makan ditambah dengan VTE, SCE dan AVE di 2 g/kg, 5 g/kg, 7g/kg dan 9g/kg untuk 45 hari eksperimen pemakanan, diikuti dengan dicabar dengan 0.1 ml 10⁷ CFU/ml *S. agalactiae*. Hasil kajian menunjukkan *Oreochromis* sp. ditambah dengan VTE-7 adalah penggalak pertumbuhan yang terbaik di antara rawatan. Ia menunjukkan 57.3% dalam kenaikan berat badan dengan 100% kemandirian. Untuk nilai hematologi Hb selepas dicabar dengan *S. agalactiae* menunjukkan berbeza secara signifikan (P<0.05) berbanding kawalan kecuali dalam SCE-2 dan SCE-7. Sementara itu, dalam nilai WBC menunjukkan berbeza secara signifikan (P<0.05) kecuali dalam AVE-5 dan AVE-7. Dalam nilai MCV menunjukkan signifikan berbeza kecuali dalam SCE-5, AVE-9 dan nilai MCHC menunjukkan signifikan kecuali dalam VTE-2 dan SCE-5. Parameter biokimia darah, terdapat perbezaan yang signifikan dalam ALT ditunjukkan dalam ikan yang diberi makan dengan VTE-2, VTE-5, VTE-7, SCE-2, SCE-5, SCE-7, AVE-5 dan AVE-7 berbanding kawalan (dicabar). Dalam AST menunjukkan berbeza secara signifikan (P<0.05) dalam VTE-5, VTE-7, SCE-9, AVE-2 dan AVE-7 berbanding kawalan (dicabar). Ikan diberi makan dengan SCE-5 adalah yang terbaik untuk resisten penyakit, ini menunjukkan nilai tertinggi WBC menunjukkan peningkatan gerak balas imun dan kemampuan ikan untuk melawan *S. agalactiae*. Dalam pemeriksaan histologi daripada ekstrak tumbuhan herba terpilih yang berbeza menunjukkan pengubahsuaian kecil di dalam hepar dan tisu ginjal. Untuk menentukan prestasi pertumbuhan dan resisten penyakit *Oreochromis* sp. ditambah dengan herba-campuran tumbuhan herba terpilih ekstrak iaitu AVE & VTE, VTE & SCE, SCE & AVE. Herba campuran AVE & VTE menunjukkan peningkatan yang paling signifikan berbanding dengan herba campuran lain. Untuk SGR, hanya AVE & VTE menunjukkan hasil paling signifikan tinggi (P<0.05) berbanding kawalan. Kemandirian ikan selepas dicabar penyakit adalah perbezaan yang signifikan (P<0.05) berbanding dengan kawalan. Dalam asai hematologi, dalam pasca-dicabar menunjukkan peningkatan yang signifikan (P<0.05) RBC, Hb dan PCV. Nilai WBC dalam pra dan pasca dicabar menunjukkan tiada perbezaan yang signifikan (P>0.05) dalam semua rawatan berbanding kawalan. Bagi MCHC dengan campuran herba, AVE & SCE adalah satu-satunya rawatan yang menunjukkan tiada perbezaan yang signifikan (P>0.05) dalam pra-dicabar dan menunjukkan penurunan perbezaan yang signifikan (P<0.05) dalam pasca dicabar dalam semua rawatan. Di sisi lain, MCV dan protein plasma tidak menunjukkan sebarang perbezaan yang signifikan (P>0.05) berbanding dengan kawalan. Parameter biokimia ALT nilai dan AST nilai dipengaruhi dengan signifikan oleh penambahan campuran herba. Walau bagaimanapun dalam pasca dicabar, nilai ALP menunjukkan perbezaan yang signifikan berbanding dengan kawalan. Perubahan histopatologi, termasuk nekrosis dan pendarahan di dalam ginjal, hepar dan hepatopankreas mungkin disebabkan sitotoksin yang dihasilkan oleh bakteria. Selain itu, dalam kajian ini untuk menilai kesan daripada makanan tambahan ekstrak methanol *V. trifolia*, *S. crispus* dan *A. vera* pada *Oreochromis* sp. sebagai rawatan penyakit. Keputusan menunjukkan dalam nilai RBC peningkatan yang signifikan (P<0.05) dalam VTE-2, SCE-2, SCE-5, SCE-9, AVE-5, AVE-7 dan kawalan (OTC) berbanding kawalan

(dicabar). Sementara itu, dalam nilai Hb menunjukkan peningkatan signifikan dalam VTE-2, SCE-5, SCE-9 dan penurunan signifikan dalam AVE-7 dan AVE-9 berbanding kawalan. Untuk nilai PCV menunjukkan signifikan ($P < 0.05$) meningkat dalam semua rawatan termasuk kawalan (OTC) berbanding kawalan (dicabar). Sebaliknya, dalam nilai MCHC tidak signifikan dalam SCE-5. Nilai protein plasma menunjukkan peningkatan yang signifikan ($P < 0.05$) selepas ditambah VTE-2, VTE-5, SCE-2, SCE-5, SCE-9, AVE-2, AVE-5, AVE-9. Nilai ALT and AST menunjukkan penurunan signifikan ($P < 0.05$) dalam semua rawatan termasuk kawalan (tidak dicabar) dan kawalan (OTC). Juga, dalam nilai ALP menunjukkan penurunan signifikan dalam semua rawatan termasuk kawalan (OTC). Kemandirian ikan meningkat selepas diberi makan dengan makanan tambahan herba dengan peningkatan yang signifikan ($P < 0.05$) dalam VTE-5, VTE-7, AVE-2, AVE-5 dan AVE-9 berbanding kawalan (dicabar). Semua pengukuran hematologi dalam *Oreochromis* sp. yang dijangkiti *S. agalactiae* dengan ditambah dengan VTE, SCE dan AVE menunjukkan nilai yang sama berbanding dengan kawalan dan rawatan yang menerima antibiotik. Ini menunjukkan ikan dijangkiti *S. agalactiae* mampu mengembalikan indeks hematologi kepada nilai hampir normal. Keputusan adalah konsisten dengan literatur yang melaporkan pelbagai keberkesanan penggunaan herba dalam akuakultur untuk mengawal pelbagai patogen bakteria. Juga dari keputusan penemuan kajian ini, menunjukkan bahawa metanol ekstrak herba campuran yang diberi pada ikan *Oreochromis* sp yang dijangkiti oleh *S. agalactiae* mempunyai kesan restoratif sinergistik dalam pemboleh ubah hematologi. Kesimpulannya, kajian ini mencadangkan bahawa *V. trifolia*, *S. crispus* dan *A. vera* ekstrak dan bercampur herba berkuatkuasa penggalak pertumbuhan dan rawatan penyakit bakteria dalam *Oreochromis* sp. Ia juga menunjukkan potensi tumbuhan herbal terpilih kerana ia merupakan terapeutan akuakultur yang mesra alam dan hanya menunjukkan kesan sampingan yang boleh abai dalam ikan yang digunakan.

ACKNOWLEDGEMENTS



All praise and gratitude to Allah (SWT) the owner of all knowledge, because with permits and bounty gives me the opportunity to conduct this research and enhance to complete thesis. I would like to express my profound gratitude and very sincere thanks to my supervisor Associate Professor Dr. Hassan Mohd Daud, for his invaluable guidance, concern, patience, encouragement and constructive comments throughout my research. Also, to my co-supervisors Professor Dr. Abdul Razak Alimon and Professor Noordin Mohamed Mustapha for their assistance and guidance. Most of all, i deeply appreciate the respect and confidence they have shown me to allow me to flourish as a scientist and person.

Special dedication to my beloved husband, Mr Mahazan Muhammad for give me positive spirit and patience with my emotional, also to my son, Muslim Ramadhani, everytime as my source of inspiration. To my lovely parents, Mr Manaf and Mrs Sharifah Zainun, who give me support and spirit to finish research project. Also, my parents in law, Mr Muhammad and Mrs Fauziah, who support their law to achieve her ambition. Not forget to my family and my group's members, especially to Miss Ruhil Hayati, Miss Nora Faten Afifah, Miss Kumari Geetha, Mrs Rashidah, Miss Nur hidayahanum, Miss Nadia and Miss Fairus who really understand my emotion and for their cooperation and spiritual support.

Sincere thanks also to the Aquatic Animal Health Unit members, who really close to me in giving support and advises. Finally, my appreciation goes to all person who have contributed directly and indirectly to this project.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Hassan Mohd Daud, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Abdul Razak Alimon, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Noordin Mohamed Mustapha, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Sharifah Raina Binti Manaf , GS 32480

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of

Supervisory Committee: Assoc. Prof. Dr. Hassan Mohd Daud

Signature: _____

Name of Member of

Supervisory Committee: Profesor Dr. Abdul Razak Alimon

Signature: _____

Name of Member of

Supervisory Committee: Profesor Dr. Noordin Mohamed Mustapha

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xvi
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xix
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 The Production of Red Hybrid Tilapia (<i>Oreochromis</i> sp.)	4
2.2 Biology of <i>Oreochromis</i> sp.	6
2.2.1 Morphology of <i>Oreochromis</i> sp.	6
2.2.2 Habitat and Distribution	7
2.2.3 Water Quality Requirements	8
2.2.4 Nutritional Requirements	8
2.2.5 Haematological Requirements	10
2.2.6 Cases of Streptococcus Infections in Tilapia	12
2.3 Medicinal Plants	13
2.3.1 The importance of Medicinal Plants as Growth Promoters	13
2.3.2 Mechanisms Action of Phytochemicals as Growth Promoters in Fish Culture	14
2.3.3 The Importance of Medicinal Plants as Antimicrobial Agents	19
2.3.4 Applications of Medicinal Plants as Antibiotics in Aquaculture	19
2.4 Simpleleaf Chastetree (<i>Vitex trifolia</i>)	21
2.5 Crocodile Tongue (<i>Aloe vera</i>)	23
2.6 Yellow Strobilanthus (<i>Strobilanthes crispus</i>)	24
2.7 Rose cactus (<i>Pereskia grandifolia</i>)	25
2.8 Sabah Snake Grass (<i>Clinacanthus nutans</i>)	27
2.9 Shiny bush (<i>Peperomia pellucida</i>)	28
3 SCREENING OF PHYTOCHEMICAL PROPERTIES, ANTIMICROBIAL ACTIVITIES	30

AND TOXICITY LEVEL OF SELECTED PLANT EXTRACTS

3.1	Introduction	30
3.2	Materials and Methods	31
3.2.1	Preparation of Herbs	31
3.2.2	Preparation of Extraction	31
3.3	Qualitative Screening of Phytochemical	32
3.3.1	Flavonoids	32
3.3.2	Tannins	32
3.3.3	Saponins	32
3.3.4	Alkaloids	32
3.3.5	Steroid	33
3.3.6	Glycoside	33
3.4	Preparation for Antibacterial Assay	33
3.4.1	Bacteria Preparation and Identification Tests	33
3.4.2	Antibacterial Assays Using Disk Diffusion Method	34
3.4.3	Minimum Inhibitory Concentration	34
3.5	Determination of LC ₅₀ Values of Extracts Using <i>Artemia salina</i> Lethality Assay and cell culture toxicity	34
3.5.1	<i>Artemia salina</i> Lethality Assay	35
3.5.2	Determination of Lethal Concentration (LC ₅₀)	35
3.5.3	<i>In vitro</i> Cell Culture Toxicity	35
3.5.4	Cytotoxic Activity	35
3.6	<i>In vivo</i> Test in Fingerling Red Hybrid Tilapia Using Artificial Infection	36
3.7	Statistical Analysis	36
3.8	Results	37
3.8.1	Qualitative Phytochemical Screening of Extracts	37
3.8.2	Antimicrobial Screening of Extracts	37
3.8.3	Minimum Inhibitory Concentration (MIC)	40
3.8.4	Lethal Concentration at 50% End-Point (LC ₅₀) of Selected Local Plants on <i>Artemia salina</i>	41
3.8.5	Cell Toxicity Assays	41
3.8.6	Lethal Concentration at 50% End-point (LD ₅₀) of Bacteria	44
3.9	Discussion	45
3.10	Conclusion	49

4	THE EFFECTS OF DIETARY SUPPLEMENTATION OF METHANOL EXTRACTS OF <i>Vitex trifolia</i> , <i>Strobilanthes crispus</i> and <i>Aloe vera</i> ON GROWTH	51
---	--	----

PERFORMANCE AND DISEASE RESISTANT IN RED HYBRID TILAPIA (*Oreochromis* sp.)

4.1	Introduction	51
4.2	Materials and Methods	52
4.2.1	Preparation of Extracts	52
4.2.2	The Bacteria Culture	52
4.2.3	Diets and Experimental Design	52
4.2.4	Water Quality Determination	54
4.2.5	Growth Performance	54
4.2.6	Challenge Assay	55
4.2.7	Haematological Assessment	55
4.2.8	Histopathological Assessment	56
4.3	Statistical Analysis	56
4.4	Results	56
4.4.1	Growth Performance of <i>Oreochromis</i> sp. Fed With <i>V. trifolia</i> , <i>S. crispus</i> and <i>A. vera</i>	56
4.4.2	Haematological and Biochemical Parameters of <i>Oreochromis</i> sp. Fed With <i>V. trifolia</i> , <i>S. crispus</i> and <i>A. vera</i>	61
4.4.3	Histopathological Assessment of <i>Oreochromis</i> sp.	64
4.5	Discussion	70
4.6	Conclusion	75

5 THE EFFECTS OF HERBAL-MIXED DIETARY SUPPLEMENTATION ON GROWTH PERFORMANCE AND DISEASE RESISTANCE IN RED HYBRID TILAPIA (*Oreochromis* sp.)

5.1	Introduction	76
5.2	Materials and Methods	76
5.2.1	Herbal Collections and Preparation of Extracts	76
5.2.2	Diets and Experimental Design	76
5.2.3	Preparation of Extraction	77
5.2.4	Water Quality Determination	77
5.2.5	Growth Performance	78
5.2.6	Challenge Assay	78
5.2.7	Haematological Assessment	78
5.2.8	Histopathological Assessment	78
5.3	Statistical Analysis	78
5.4	Results	78
5.4.1	Growth Performance	78
5.4.2	Haematology Assessment	81
5.4.3	Histopathological assessment	81
5.5	Discussion	87
5.6	Conclusion	91

6	THE EFFECTS OF DIETARY SUPPLEMENTATION OF METHANOL EXTRACTS OF <i>Vitex trifolia</i>, <i>Strobilanthes crispus</i> and <i>Aloe vera</i> IN RED HYBRID TILAPIA (<i>Oreochromis</i> sp.) AS DISEASE TREATMENT	92
6.1	Introduction	92
6.2	Materials and Methods	92
6.2.1	Herbal Collections and Preparation of Extracts	92
6.2.2	Experimental Design	93
6.2.3	Pathogenic Bacteria Culture	93
6.2.4	Diets and Experimental Design	93
6.2.5	Haematological Assessment	94
6.3	Statistical analysis	94
6.4	Results	94
6.5	Discussion	99
6.6	Conclusion	101
7	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	102
	REFERENCES	106
	APPENDICES	129
	BIODATA OF STUDENT	145
	LIST OF PUBLICATIONS	146

LIST OF TABLES

Table		Page
2.1	The most cultured freshwater fish in Malaysia	5
2.2	Haematology reference intervals for hybrid tilapia	11
2.3	Biochemical parameters for hybrid tilapia	12
2.4	Mechanisms action of phytochemicals as growth promoters	15
2.5	Medicinal plants and algae supplements into diets	20
3.1	Preliminary phytochemical screening of plant extracts	37
3.2	Response of different solvents of plants extracts and controls	38
3.3	Minimum inhibitory concentration of methanolic plant extracts	41
3.4	Lethal concentration, LC ₅₀ of methanolic plant extracts on <i>Artemia salina</i>	41
3.5	Viability of BF-2 cell line after exposure to methanolic plant extract	42
3.6	Lethal concentration LD ₅₀ of bacteria	45
4.1	Percentage and proximate analysis composition of the experimental diets	54
4.2	Mean growth performance and feed utilization of <i>Oreochromis</i> sp.	58
4.3	Haematological and biochemical parameters of <i>Oreochromis</i> sp. (pre-challenged)	62
4.4	Haematological and biochemical parameters of <i>Oreochromis</i> sp. (post-challenged)	63
4.5	Histology scoring lesions of liver	65
4.6	Histology scoring lesions of kidney	68
5.1	Percentage and proximate analysis composition of the experimental diets	77
5.2	Mean growth performance and feed utilization of <i>Oreochromis</i> sp. in herbal-mixed	79
5.3	Haematological characteristics of <i>Oreochromis</i> sp. fed with herbal-mixed.	80
5.4	Histology scoring lesions of liver	82
5.5	Histology scoring lesions of kidney	85
6.1	Percentage and proximate analysis composition of the experimental diets	94
6.2	Haematological and biochemical parameter of <i>Oreochromis</i> sp. treatment	96
6.3	Survival rate of <i>Oreochromis</i> sp.	97

LIST OF FIGURES

Figure		Page
2.1	A mature specimen of Red hybrid tilapia <i>Oreochromis</i> sp.	4
2.2	Pie chart of freshwater aquaculture production by species	6
2.3	The leaves of <i>V. trifolia</i>	21
2.4	The fleshy leaves with serrated edges of <i>A. vera</i>	23
2.5	The leaves of <i>S. crispus</i>	24
2.6	The flower of <i>P. grandifolia</i>	25
2.7	The leaves of <i>C. nutans</i>	27
2.8	The leaves of <i>P. pellucida</i>	28
3.1	Plate A and B were showed discs impregnated with the plant extracts on agar plate lawn with <i>A. hydrophila</i>	39
3.2	Plate C and D were showed discs impregnated with the plant extracts on agar plate lawn with <i>S. agalactiae</i>	39
3.3	Plate E and F were showed discs impregnated with the plant extracts on agar plate lawn with <i>E. cloacae</i>	40
3.4	Antibiotic susceptibility plate assay on <i>A. hydrophila</i>	40
3.5	BF-2 cell line after exposure to <i>P. pellucida</i> extract	42
3.6	BF-2 cell line after exposure to <i>V. trifolia</i> extract	43
3.7	BF-2 cell line exposed to <i>S. crispus</i> extract	43
3.8	BF-2 cell line exposed to <i>A. vera</i> extract	44
3.9	BF-2 cell line; unstained exposed to <i>V. trifolia</i> extract	44
4.1	<i>Streptococcus agalactiae</i> colonies on tryptic soy agar	52
4.2	Acclimatization for 2 weeks	53
4.3	Ten fish were then allocated into each aquarium	53
4.4	Setup of experimental aquaria	53
4.5	A moribund fish with corneal opacity of the eye	59
4.6	Ulcer on fish body and caudal torn off	59
4.7	Enlarged spleen and liver	60
4.8	Haemorrhages on body	60
4.9	Section of control fish liver	66
4.10	Section of SCE-9 fish liver	66
4.11	Section of SCE-9 fish kidney	69
4.12	Section of AVE-9 fish kidney	69
5.1	Section of control hepatopancreas	83

5.2	Section of control liver	83
5.3	Section of control fish kidney	86
5.4	Section of control fish kidney	86
6.1	The fish showed spiraling swimming	98
6.2	The fish showed enlarged of kidney	98



LIST OF ABBREVIATIONS

ALT	alanine aminotranferase
ALP	alkaline phosphatase
AST	aspartate aminotransferase
DM	dry matter
FCR	feed conversion ratio
FI	feed intake
SGR	specific growth rate
NRC	National Research Council
SE	standard error
VTE	<i>Vitex trifolia</i> extract
SCE	<i>Strobilanthes crispus</i> extract
AVE	<i>Aloe vera</i> extract
PGE	<i>Pereskia grandifolia</i> extract
PPE	<i>Peperomia pellucida</i> extract

CHAPTER 1

GENERAL INTRODUCTION

1.1 Introduction

Aquaculture is one of a rapidly growing industry in the world. Among various kinds of cultivated organisms constitute an important industries. The percentage on its production increases year by year. In Malaysia, aquaculture is also regarded as a potential strategic industry that contribute to the economy (Ministry of Agriculture, 2015). Due to the importance of this industry, the Government has pronounced it as one of the country's transformation programme under the National Key Economic Area (NKEA). To date, the percentage of aquaculture production has been seen to be increasing year by year. In 2014, the total aquaculture production is 520,514 tonnes and the freshwater aquaculture production is 106,731 tonnes (Department of Fisheries Malaysia, 2016). Recently due to numerous intensive farming practices, infectious diseases pose a major problem in aquaculture industry, causing heavy loss to farmers. Disease occurs when the balance between hosts, environment and agent is disturbed. The pathogens such as parasites, fungi, bacteria and viruses are the known cause of many disease problems in fish. As an example, Sayuthi (1993) has estimated the cost of losses caused by one type of pathogenic bacterium was RM20 million. Thus, to handle and avoiding this problem, many chemicals and antibiotics have been used indiscriminately in large amount in fish farms, creating potential risks to ecological and human health. As alternatives, we need to replace these chemicals and antibiotics by using natural compounds.

Medicinal plants are plentiful and easy to obtained, with a broad assortment of nutrients and active compounds (Edeoga *et al.*, 2010). They have been used as chemotherapeutic and feed additives in domesticated animals (Chang, 2000). Unfortunately, their information as chemotherapeutant and feed additives in fish diets is still limited in scientific researches.

Herb industry has been shown to be a definitive source of national economic growth and this initiative is one of the Entry Point Projects (EPP) under the Agricultural National Key Economic Area (NKEA) in the Economic Transformation Programme (ETP). In the effort to coordinate and monitor the effectiveness of the EPP1 High Value Herbal Products implementation, the Herbal Sub-group was established under the purview of the Ministry of Agriculture and Agro-based Industry (Ministry of Agriculture, 2014).

1.2 Background of Research

Tilapia is a popular food fish species all over the world especially in Africa and Asia. Globally the production of tilapia has shown an increased yearly. In 2015, it was forecasted that production will grow by 6% to total 5 million tonnes (FAO, 2016). The increasing global importance of cultured tilapia as a food fish has prompted significant research on improvement of the growth performance. However, a bacterial disease, *Streptococcus* has been reported to infect the fish culture and had caused high mortality (Austin and Austin, 2007). Commonly, the usual treatment for bacterial disease is by using antibiotic. However, antibiotic and also chemotherapeutant can contribute to various side effects especially towards humans and the development of bacterial resistance. Another method of treatment is by using vaccine, but the method is costly and only provides specific protection (Sakai, 1999). Many local plants contain antibacterial compounds which can be employed in treating health problems in fish (Citarasu, 2010). Nevertheless, there is trivial data on their role as healthcare products for fish.

As expected many local plants contain various nutrients that can promote growth and good health of animals. Thus theoretically, using local plants as whole or as extracts as additive singly or in composition, the diet may promote a positive outcome on the growth performance and productivity of fishes. Also, the usage will inevitably reduce dangerous residual accumulation in the tissues and the environment.

1.3 Problem Statement

Every year, the demand for fish and fish products from aquaculture has tremendously increased. However, the main hindrance is the cultured fish succumbing to disease infection and commonly antibiotics and chemicals are used to treat this problem. The advantages of using chemicals and drugs are that they can be applied straight using diverse applications and may give effective outcome. Unfortunately, the residual toxic chemicals or their metabolites accumulated in fish body tissues pose a health risk to consumers. Due to this reason, many drugs (antibiotics) and chemicals are now banned for aquaculture used. Most countries, including Singapore and European Union (EU) have banned imported aquaculture products which were detected to have antibiotics residues. Accordingly the EU has ratified a ban on the usage of whole sub-therapeutic antibiotics as growth-promoting agents in aquaculture practices especially for food fish. Thus, the way forward, is undertaking research focussing on alternatives to traditional antibiotics for prevention and control of diseases in aquaculture and also as growth promoters in animal feeds, such as using enzymes, herbs and probiotics. Naturally herbs and plants (phytobiotics) have been recognized to contain many compounds with properties that can promote growth in animal and antimicrobes.

1.4 Research Hypothesis

Selected local plants as feed supplement can be used to promote growth, and also for disease prophylaxis and disease treatment of red hybrid tilapia, *Oreochromis* sp.

1.5 Significance of Study

Local plants have been shown to contain certain compounds such as antioxidants, polyphenols and antibacterial properties that contribute in improving growth performance and in treating fish disease in cultivated species. Local plants can also be used as feed additives to replace antibiotics as growth promoters and treatment of disease infection. The present study will elucidate the effect of local plants for growth promoting and antibacterial properties in *Oreochromis* sp.

In relation to this study is to determine the potential of local plants as antibacterial, growth promoters, disease resistance and disease treatment in *Oreochromis* sp.

1.6 General Objectives

The general objectives of this study are:

- 1) To determine the effect of selected herbal supplementation on the growth performance and disease resistance in *Oreochromis* sp.
- 2) To determine the effect of herbal supplementation as disease prophylaxis treatment in *Oreochromis* sp.

The specific objectives of this study are :

- 1) To screen phytochemical properties, antibacterial activity and toxicity level of selected local plants (*Vitex trifolia*, *Aloe vera*, *Strobilanthes crispus*, *Peperomia pellucida*, *Clinacanthus nutans* and *Pereskia grandifolia*).
- 2) To determine the growth performance and disease resistant of *Oreochromis* sp. fed with diet supplemented with selected plant extracts at different concentrations.
- 3) To determine the growth performance and disease resistant of *Oreochromis* sp. supplemented with herbal-mixed of the selected plant extracts.
- 4) To assess the efficacy of diet supplementation of selected plant extracts for 14 days as disease treatments.

REFERENCES

- Abdel-Tawwab, M., Ahmad, M.H., Seden, M.E.A. and Sakr, S.F.M. (2010). Use of green tea, *Camellia sinensis* L., in practical diet for growth and protection of Nile Tilapia, *Oreochromis niloticus* (L.), against *Aeromonas hydrophila* infection. *Journal of the World Aquaculture Society* 41: 203-213.
- Abdul Kader Mydeen, K.P. and Haniffa, M.A. (2011). Evaluation of antibacterial activity of medicinal plants on fish pathogen, *Aeromonas hydrophila*. *Journal of Biology Research* 1: 1-5.
- Ada, F.B, Ndome, Chris, B. and Bayim, B. Peter-Robins. (2011). Some haematological changes in *O. niloticus* juveniles exposed to butachlor. *Journal of Agriculture and Food Technology* 1(6): 73-80.
- Adams, C. (2005). Nutrition-based health. *Feed International*, 2: 25-28.
- Adedeji, O.S., Farinu, G.O., Olayeni, T.B. and Ameen, S.A.B.G. (2008). The use of bitter kola (*Garcinia kola*) Dry seed powder as a natural growth promoting agent in broiler chicks. *Research Journal of Poultry Sciences*, 2(4): 78-81.
- Adiguzel, A., Gulluce, M., Sengul, M., Ogutcu, H., Sahin, F. and Karaman, I. (2005). Antimicrobial Effects of *Ocimum basilicum* (Labiatae) Extract. *Turkish Journal of Biology*, 29: 155-160.
- Aditya, K. and Ravi Kumar, A. (2014). Phytochemical evaluation of *Vitex leucoxylon*, *Vitex negundo* and *Vitex trifolia*. *Indian Journal of Research in Pharmacy and Biotechnology*, 2(2): 1106-1108.
- Afrizal. (2008). *Analytical, Bioactivity and Stability Studies on Strobilanthes crispus* L. *Bremek* and *Sonchus Arvensis* L. Extracts, PhD Thesis, Universiti Sains Malaysia.
- Aiyegoro, O.A. and Okoh, A.I. (2010). Preliminary phytochemical screening and *in vitro* antioxidant activities of the aqueous extract of *Helichrysum longifolium* DC. *BMC Complementary and Alternative Medicine*, 10(21): 1-8.
- Akinrotimi, O.A., Abu Omg, Bekibele, D.O., Uedeme-Naa, B. and Aranyo, A.A. (2010). Haematological characteristics of *Tilapia guineensis* from Buguma creek, Niger Delta, Nigeria. *Electronic Journal of Environmental, Agricultural and Food Chemistry* 9(8): 1415-1422.
- Akinrotimi, O.A., Gabriel, U.U., Anyanwu, P.E. and Anyanwu, A.O. (2007). Influence of sex, acclimation methods and period on hematology of *Sarotherodon melanotheron*. *Research Journal of Biology Science* 2(3): 248-352.
- Akinwale, A.O., Faturoti, E.O. (2007). Biological performance of African catfish

- (*Clarias gariepinus*) cultures in recirculating system in Ibadan. *Aquaculture Engineering* 36: 18-23.
- Alderman, D.J. and Hastings, T.S. (1998). Antibiotic use in aquaculture: development of antibiotic resistance-potential for consumer health risks. *International Journal of Food Sciences and Technology* 33: 139-155.
- Aliero, A.A. and Afolayan, A.J. (2006). Antimicrobial activity of *Solanum tomentosum*. *African Journal of Biotechnology*, 5(4): 369-372.
- Alwan, S.F., Hadi, A.A. and Shokr, A.E. (2009). Alterations in haematological parameters of fresh water fish *Tilapia zilli* exposed to aluminium. *Journal of Science and Its Applications* 3(1): 12-19.
- Anderson, E. (2001). *The Cactus Family*. USA: Timber Press.
- Archana, R., and A. Namasivayam. (2000). Effect of *Ocimum sanctum* on noise induced changes in neutrophil function. *Journal of Ethnopharmacology*, 73:81–84.
- Arifullah, M., Vikram, P., Chiruvella, K.K., Shaik, M.M. and Abdullah Ripain, I.H. (2014). A review on Malaysian plants used for screening of antimicrobial activity. *Annual Research and Review in Biology*, 4(13): 2088-2132.
- Arrigoni-Blank, M.F., Dmitrieva, E.G., Franzotti, E.M., Antonioli, A.R., Andrade M.R. and Marchioro, M. (2004). Anti-inflammatory and analgesic activity of *Peperomia pellucida* (L.) HBK (Piperaceae). *Journal of Ethnopharmacology* 91: 215–218.
- Arts, I.C.W. and Hollman, P.C.H. (2005). Polyphenols and disease risk in epidemiologic studies. *The American Journal of Clinical Nutrition*, 81: 317-325.
- Arullappan, S., Rajamancikam, P., Thevar, N. and Kodimani, C.C. (2014). *In vitro* screening of cytotoxic, antimicrobial and antioxidant activities of *Clinacanthus nutans* (Acanthaceae) leaf extracts. *Tropical Journal of Pharmaceutical Research September* 13(9): 1455-1461.
- Arunkumar, S. and Muthuselvam, M. (2009). Analysis of phytochemical constituents and antimicrobial activities of *Aloe vera* L. against clinical pathogens. *World Journal of Agricultural Sciences*, 5(5): 572–576.
- Ashraf, M.A. and Goda, S. (2008). Effect of dietary ginseng herb (Ginsana® G115) supplementation on growth, feed utilization, and hematological indices of Nile tilapia, *Oreochromis niloticus* (L.), fingerlings. *Journal of the World Aquaculture Society*, 39(2): 205–214.
- Aslam, M.S., Muhammad Syarhabil, A. and Awang Soh, M. (2014). A review on phytochemical constituents and pharmacological activities of *Clinacanthus nutans*. *International Journal of Pharmacy and Pharmaceutical Sciences* 7(2): 30-33.
- Asmah, R., Susi, E., Abdah, M. A., Patimah, I., Taufiq, Y.Y.H. and Mohd Fadzelly, A.B. (2006). Anticarcinogenic properties of *Strobilanthes crispus* extracts and its compounds *in vitro*. *International Journal of Cancer Research*, 2(1): 47-49.

- Austin, B. (2006). The bacterial microflora of fish, revised. *The Scientific World Journal*, 6: 931–945.
- Austin, B. and Austin, D.A. (2007). *Bacterial Fish Pathogens: Disease of Farmed and Wild Fish*. UK: Praxis Publishing.
- Aziba, P. I., Adedeji, A., Ekor, M., Adeyemi, O. (2001). Analgesic activity of *Peperomia pellucida* aerial parts in mice. *Fitoterapia* 72: 57-58.
- Babu, M.M. (1999). *Developing bioencapsulated ayurvedic product for maturation and quality larval production in Penaeus monodon*, PhD Thesis, Manomaniam Sundaranar University.
- Backer, C.A.D. and Bakhuizen, R.C.V. (1963). *Flora of Java (Spermatophytes only)*. Groningen: Noordhoff Press.
- Balarin, J.D. and Hatton, J.P. (1979). *Tilapia: a guide to their biology and culture in Africa*. Scotland: University of Stirling Press.
- Banaee, M., Sureda, M., Mirvaghefi, A.R. and Ahmadi, K. (2011). Effects of diazinon on biochemical parameters of blood in rainbow trout (*Oncorhynchus mykiss*). *Pesticide Biochemistry and Physiology* 99: 1-6.
- Batista, O., Duarte, A., Nascimento, J., Simoes, M.F., De La Torre, M.C., and Rodriguez, B. (1994). Structure and antimicrobial activity of diterpenes from the roots of *Plectranthus hereroensis*. *Journal of Natural Products* 57(6): 858-861.
- Bauer, A.W., Kirby, W.M.M., Sherris, J.C. and Turck, M. (1966). Antibiotic susceptibility testing by standardized single disc method. *American Journal of Clinical Pathology* 36: 493–496.
- Bazrafshan, E., Naseri, S., Mahvi, A. H. and Shayedhi, M. (2007). Performance evaluation of electrocoagulation process for diazinon removal from aqueous environments by using iron electrons. *Iranian Journal of Environmental Health Science and Engineering* 4: 127-132.
- Bellinger, E.G. and Sigeo, D.C. (2010). *Freshwater algae: Identification and Use as Bioindicators*. UK: John Wiley and Sons.
- Beveridge, M.C.M. and McAndrew B.J. (2000). *Tilapias: Biology and Exploitation*. Netherlands: Kluwer Academic Publishers.
- Bhosale, S.V., Bhilave, M.P. and Nadaf, S.B. (2010). Formulation of fish feed using ingredients from plant sources. *Research of Journal Agriculture Science* 1 (3): 284-287.
- Bilen, S., Bulut, M., Bilen, A.M. (2011). Immunostimulant effects of *Cotinus coggyria* on rainbow trout (*Oncorhynchus mykiss*). *Fish and Shellfish Immunology* 30: 451-455.
- Bobbarala, V.D., Madhuri, D.B., Aryamithra, D. and Naidu, K.C. (2010). An alternative approach to control *Aeromonas hydrophila* using important herbal extracts. In *Bioresources for Rural Livelihood: Genetics, Biochemistry and Toxicology*, ed. G.K. Kulkarni, B.N. Pandey, and B.B. Joshi pp. 225-232. India: Narendra Publishing House.

- Bojo, A.C., Albano-Garcia, E. and Pocsidio, G.N. (1994). The antibacterial activity of *Peperomia pellucida* (L.) HBK (Piperaceae). *Asia Life Sciences* 3: 35–44.
- Borges A., Scotti, L.V. Siqueira, D.R., Jurinitz, D.F. and Wassermann, G.F. (2004). Hematologic and serum biochemical values for jundia (*Rhamdia quelen*). *Fish Physiology and Biochemistry*, 30: 21-25.
- Borris, R.P. (1996). Natural products research: perspectives from a major pharmaceutical company. *Journal of Ethnopharmacology*, 51: 29–38.
- Boudreau, M.D. and Beland, F.A. (2006). An evaluation of the biological and toxicological properties of *Aloe barbadensis* (miller), *Aloe vera*. *Journal of Environmental Science and Health*, 24(1): 103-154.
- Brenden, S.A. and Huizinga, H.W. (1986) Pathophysiology of experimental *Aeromonas hydrophila* infection in goldfish, *Carassius auratus* L.. *Journal of Fish Disease* 9: 163-167.
- Brown, J. (1989). Antibiotics: their use and abuse in aquaculture. *World Aquaculture*, 20 (2): 34–43.
- Brown, M.E. (1957). Experimental studies on growth. Physiology of fishes. Academic Press, New York.
- Bruneton, J.. Pharmacognosy, phytochemistry, medicinal plants 2nd edition. Lavoisier; 1999. 1136p.
- Bryan, A.H. (2005). Understanding medicinal plants. Their chemistry and therapeutic action. The Haworth Herbal Press. 4: 83-118.
- Buckley, J.A., Whitmore, C. M. and Matsuda, R.I. (1976). Changes in blood chemistry and blood cell morphology in coho salmon, *Oncorhynchus kisutch* following exposure to sublethal levels of total residual chlorine in municipal wastewater. *Journal of Fish Research Board Canada*, 33: 776-782.
- Burr, G., Gatlin, D. and Ricke, S. (2005). Microbial ecology of the gastrointestinal tract of fish and the potential application of prebiotics and probiotics in finfish aquaculture. *Journal of the World Aquaculture Society*, 36(4): 425–435.
- Canadian Pharmaceutical Association (CPA). (1988). Self medication, Ottawa, Canada.
- Caruso, D. and Lazard, J. (1999). Subordination stress in Nile tilapia and its effect on plasma lysozyme activity. *Journal of Fish Biology* 55: 451-454.
- Castell, J.D. and Tiews, K. 1980. Report of EIFAC, IUNS, and ICES Working Group on standardization of methodology in fish nutrition research. Hamburg, Federal Republic of Germany, 21-23 March, 1979. EIFAC Tech. Pap., 36. 24 pp.
- Cek, S., Turan, F. and Atik, E. (2007). The Effects of Gokshura, *Tribulus terrestris* on sex reversal of guppy, *Poecilia reticulata*. *Pakistan Journal of Biological Sciences*, 10(5): 718-725.

- Ceulemans S., Robles R., Coutteau P. (2009). Innovative feed additives improve feed utilization in Nile Tilapia. *Global Aquaculture Advocate* November/December 2009.
- Chakraborty, S.B. and Hancz, C. (2011). Application of phytochemicals as immunostimulant, antipathogenic and antistress agents in finfish culture. *Reviews in Aquaculture*, 3(3): 103–119.
- Chakraborty, S.B., Horn, P. and Hancz, C. (2013). Application of phytochemicals as growth-promoters and endocrine modulators in fish culture. *Reviews in Aquaculture*, 5: 1–19.
- Chan-Bacab, M.J. and Pena-Rodriguez, L.M. (2001). Plant natural products with leishmanicidal activity. *Natural Product Reports* 18: 674–688.
- Chandarana, H., Baluja, S. and Chanda, S.V. (2005). Comparison of antibacterial activities of selected species of zingiberaceae family and some synthetic compounds. *Turkish Journal of Biology*, 29: 83–97.
- Chang, J. (2000). Medicinal herbs: Drugs or dietary supplements? *Biochemical Pharmacology*, 59(3): 211–219.
- Chatterjee, A. and Prakash, S.C. (2003). The treatise of Indian medicinal plants. *New Delhi: National Institute of Science Communication and Information Resources*, 10: 525.
- Chen, X., Wu, Z., Yin, J. and Li, L. (2003). Effects of four species of herbs on immunefunction of *Carassius auratus gibelio*. *Fish Sciences China*, 10: 36–40.
- Chew, Y.L., Chan, E.W.L., Tan, P.L., Lim, Y.Y., Stanslas, J. and Goh, J.K. (2011). Assessment of phytochemical content, polyphenolic composition, antioxidant and antibacterial activities of Leguminosae medicinal plants in Peninsular Malaysia. *BMC Complementary and Alternative Medicine*, 11(12): 1–10.
- Chitra, S. (1995). *Effect of feeding supplemented stresstol bioencapsulated Artemia franciscana on growth and stress tolerance-in Penaeus indicus postlarvae*. Master Thesis, University of Tirunelveli.
- Cho, S.H. and Lee, S. (2012). Onion powder in the diet of the olive flounder, *Paralichthys olivaceus*: Effects on the growth, body composition, and lysozyme activity. *Journal of The World Aquaculture Society* 43(1): 30–38.
- Chuku, E.C. and Jaja, E.T. (2012). Determination of the biochemical composition and anti-fungal activities of *Peperomia pellucida* L. (H.B.K). *Acta Agronomica Nigeriana* 12: 23–31.
- Citarasu, T. (2010). Herbal biomedicines: a new opportunity for aquaculture industry. *Aquaculture International* 18(3): 403–414.
- Citarasu, T. and Michael Babu, M., Raja Jeya Sekar, R. and Peter Marian, M. (2002). Developing Artemia Enriched Herbal Diet for Producing Quality Larvae in *Penaeus monodon*, Fabricius. *Asian Fisheries Science* 15: 21–32.

- Citarasu, T., Immanuel, G. and Marian, M. P. (1998). Effects of feeding *Artemia* enriched with stresstol and cod liver oil on growth and stress resistance in the Indian white shrimp *Penaeus indicus* post larvae. *Asian Fish Sciences* 12: 65–75.
- Citarasu, T., Rajajeyasekar, R., Venketramalingam, K., Dhandapani, P. S. and Marian, M. P. (2003). Effect of wood apple *Aegle marmelos*, Correa (Dicotyledons, Sapindales, Rutaceae) extract as an antibacterial agent on pathogens infecting prawn (*Penaeus indicus*) larviculture. *Indian Journal of Geo-Marine Sciences* 32(2): 156–161.
- Citarasu, T., Sivaram, V., Immanuel, G., Rout, N. and Murugan, V. (2006). Influence of selected Indian immunostimulant herbs against white spot syndrome virus (WSSV) infection in black tiger shrimp, *Penaeus monodon* with reference to haematological, biochemical and immunological changes. *Fish Shellfish and Immunology* 21: 372–384.
- Cockerill, F. R., Wikler, M. A., Alder, J., Dudley, M. N., Eliopoulos, G. M., Ferraro, M. J. and Zimmer, B. L. (2012). *Performance standards for antimicrobial disk susceptibility tests; approved standard. Clinical and Laboratory Standards Institute - NCCLS* 32(1): 1-58.
- Cowan, M. M. (1999). Plant products as antimicrobial agents. *Clinical Microbiology Reviews* 12(4): 564–582.
- Cristea, V., Antache, A., Grecu, I., Docan, A., Dediu, L. and Cre, M. (2012). The use of phytobiotics in aquaculture. *Lucrari Stiintifice - Seria Zootehnie* 57: 250–255.
- Dacie, J.V. and Lewis, S.M. (2001). *Practical Haematology* 9^{ed}. London: Churchill Livingstone Press.
- Dada, A. A. (2012). Effects of herbal growth promoter feed additive in fish meal on the performance of nile tilapia (*Oreochromis niloticus* L.). *Egypt Academic Journal of Biology Sciences*, 4(1): 111–117.
- Dada, A. A. and Ikuerowo, M. (2009). Effects of ethanolic extracts of *Garcinia kola* seeds on growth and haematology of catfish (*Clarias gariepinus*) broodstock. *African Journal Agriculture Research* 4: 344-347.
- Dada, A. A. and Olugbemi, B. D. (2013). Dietary effects of two commercial feed additives on growth performance and body composition of African catfish *Clarias gariepinus* fingerlings. *African Journal of Food Science* 7: 325-328.
- Dada, A. A. and Oviawe, N. E. (2011). The use of bitter kola *Garcinia kola* dry seed powder as a natural growth-promoting agent for African sharptooth catfish *Clarias gariepinus* fingerlings. *African Journal of Food Science* 36(1): 97-100.
- De Silva, S.S., Subasinghe, R.P., Bartley, D.M. and Lowther, A. Tilapias as alien aquatics in Asia and the Pacific: a review. *FAO Fisheries Technical Paper*. No. 453. Rome, FAO. 2004. 65p.
- De-Fatima, A. B. M. Dmitrieva, E. G. Franzotti, E. M. Antonioli, A. R., Andrade,

- M. R. and Marchiori, M. (2004). Anti-inflammatory and analgesic activity of *Peperomia pellucida* (L.) H.B.K. (Piperaceae). *Journal of Ethnopharmacology* 91: 215-218.
- Department of Fishery. (2014). Statistic. <http://www.dof.gov.my/perangkaan-perikanan-jilid-1-2014>. Retrieved 15 May 2013.
- Direkbursarakom, S., Ruangpan, L., Ezura, Y. and Yoshimizu, M. (1998). Protective efficacy of *Clinacanthus nutans* on yellow-head disease in black tiger shrimp (*Penaeus monodon*). *Fishing Pathology* 33(4): 401–404.
- Direkbusarakom, S., Herunsalee, A., Yoshimizu, M. and Ezura, Y. (1996a). Protective efficacy of *Clinacanthus nutans* on yellow-head disease in Black Tiger Shrimp (*Penaeus monodon*). *Fish Pathology* 33(4): 404–410.
- Direkbusarakom, S. (2004). Application of medicinal herbs to aquaculture in Asia. *Walailak Journal of Science and Technology* 1(1): 7-14.
- Direkbusarakom, S., Ezura, Y., Yashimizu, M. and Herunsalee, A. (1998). Efficacy of Thai traditional herb extracts against fish and shrimp pathogenic bacteria. *Fish Pathology* 33(4): 437-441.
- Direkbusarakom, S., Herunsalee, A., Boonyaratpalin, S., Danayadol, Y. and Aekpanithanpong, U. (1995). Effect of *Phyllanthus* spp. against yellow-head baculovirus infection in black tiger shrimp, *Penaeus monodon*. In: Shariff, M., Arthur, J.R. and Subasinghe, R.P. (eds). Diseases in Asian aquaculture II. Fish Health Section, Asian Fisheries Society, Manila. 85–92.
- Direkbusarakom, S., Herunsalee, A., Yoshimizu, M. and Ezura, Y. (1996b). Antiviral activity of several Thai traditional herb extracts against fish pathogenic viruses. *Fish Pathology* 31(4): 209–213.
- Direkbusarakom, S., Runghomnerdwong, S., Herunasalee, A. and Rungpan, L. (1996c). Screening of Thai traditional herbs against shrimp pathogenic bacteria. *NICA Technical paper* 7: 13.
- Douglass, J. W. and Jane, K. W. (eds.) (2010). *Schalm's Veterinary Hematology*. USA: John Wiley and Sons, Blackwell Publishing Ltd.
- Duncan, D. B. (1955). Multiple range and multiple 'F' tests. *Biometrics* 11: 1–42.
- Duncan, P. L. and Klesius, P. H. (1996). Effect of feeding spirulina on specific and non specific immune response of channel catfish. *Journal of Aquatic Animal Health* 8: 308–313.
- Ebong, P. E, Atangwho, I. J., Eyong, E. U. and Egbung, G. E. (2008). The antidiabetic efficacy of combined extracts from two continental plants: *Azadirachta indica* (A. Juss) (Neem) and *Vernonia amygdalina* (Del.) (African Bitter Leaf), *American Journal of Biochemistry and Biotechnology* 4(3): 239-244.
- Edahiro, T., Hamaguchi, M. and Kusuda, R. (1991). Suppressive effect of glycyrrhizia against streptococcal infection promoted by feeding oxidized

- lipids to yellow tail *Seriola quinqueradiata*. *Suisanzoshoku* 39: 21–27.
- Edeoga, H. O., Okwu, D. E. and Mbaebie, B. O. (2010). Phytochemical constituents of some Nigerian plants. *African Journal of Biotechnology* 4(7): 685-688.
- El-Bakary, N. E. R. and El-Gammal, H. L. (2010). Comparative histological, histochemical and ultrastructural studies on the proximal intestine of flathead grey mullet (*Mugil cephalus*) and sea bream (*Sparus aurata*). *World Applied Sciences Journal* 8(4): 477-485.
- El-Dakar, A. Y., Hassanien, G. D., Gad, S. S. and Sakr, S. E. (2008). Use of dried basil leaves as a feeding attractant for hybrid tilapia, *Oreochromis niloticus* X *Oreochromis aureus*, fingerlings. *Mediterranean Aquaculture Journal* 1: 35-44.
- Eldar, A., Bejerano, Y., Livoff, A., Horvitz, A. and Bercovier, H. (1995). Experimental streptococcal meningo-encephalitis in cultured fish. *Veterinary Microbiology* 43: 33-40.
- El-Sayed, A. F. M. (2006). *Tilapia culture*. Cambridge, USA: CABI Publishing.
- Erdemoglu, N., Turan, N. N., Cakici, I., Sener, B. and Aydin, A. (2006). Antioxidant activities of some Lamiaceae plant extracts. *Phytotherapy Research* 20(1): 9-13.
- Essa, M. A., El-Serafy, S. S., El-Ezabi, M. M., Daboor, S. M., Esmael, N. A. and Lall, S. P. (2010). Effect of different dietary probiotics on growth, feed utilization and digestive enzymes activities of Nile Tilapia, *Oreochromis niloticus*. *Journal of Arabian Aquaculture Society*, 5 (2): 143-161.
- Fanouraki, B.P., Divanach, M. and Pavlidis, M. (2007). Baseline values for acute and chronic stress indicators in sexually immature red porgy (*Pargus pagrus*). *Aquaculture* 265: 294-304.
- FAO. (2014). Fisheries and Aquaculture Statistics. Rome, Italy. 105p.
- Farahi, A., Kasiri, M., Sudagar, M., Soleimani, I. M. and Zorriehzahra, S. M. J. (2011). Effect of dietary supplementation of *Melissa officinalis* and *Aloe vera* on hematological traits, lipid oxidation of carcass and performance in rainbow trout (*Oncorhynchus mykiss*). *Online Journal of Animal and Feed Research* 2(1): 1-5.
- Faridah Hanum, I. and Nurulhuda, H. (1999). The use of medicinal plant species by the temuan tribe of Ayer Hitam Forest, Selangor, Peninsular Malaysia. *Pertanika Journal Tropical Agriculture Science* 22(2): 85-94.
- Fatan H.Y. (1990). 'Etiologi Penyakit di Kalangan Masyarakat Melayu: Satu Kajian Kes di Mukim Sanggang, Temerloh Pahang, Tesis Sarjana Sains Kemasyarakatan, Pulau Pinang: University Science Malaysia.
- Felicitta, J., Arti, M. R., Ronald, J., Sakthika, T., Nagarajan, R. and Chelladurai, G. (2013). Effect of different concentrations of some phytoadditives (*Allium sativa* and *Allium cepa*) on growth, survival and hematological parameters in tilapia (*Oreochromis mossambicus*) juveniles. *The Israeli Journal of Aquaculture-Bamidge*, 64: 1-8.

- Food and Agricultural Organization of the United Nations. (2006). The state of world fisheries and aquaculture. Available from <http://www.fao.org/docrep/009/A0699e/A0699e00.htm>. Retrieved on 15 May 2015.
- Francis, G., Makkar, H. P. S. and Becker, K. (2005). *Quillaja* saponins - A natural growth promoter for fish. *Animal Feed Science and Technology* 121: 147-157.
- Frerichs, G. N. and Millar, S. D. (1993). *Manual of the Isolation and identification of fish bacterial pathogens*. Stirling: Pisces Press.
- Galina, J., Yin, G., Ardo, L. and Jeney, Z. (2009). The use of immunostimulating herbs in fish. An overview of research. *Fish Physiology and Biochemistry* 35(4): 669-676.
- Gao, X. and Wu, W. (1994). The effect of Dang-Gui (*Angelica sinensis*) and ferulic acid on mice immune system. *China Journal of Biochemistry and Pharmacology* 15:107-110.
- Geetha, V., Doss, A. and Pichai Anthoni Doss, A. (2004). Antimicrobial potential of *Vitex trifolia* Linn. *Ancient science of life* 23(4): 30-32.
- Geissman, T. A. (1963). Flavonoid tannins, lignins and related compounds, In: *Pyrrole pigments, isoprenoid compounds and phenolic plant*, ed. M. Florkin, and E. H. Stotz, pp. 2653. New York: Elsevier.
- George, A. J. (2009). AmpC β -lactamases. *Clinical Microbiology Reviews* 22(1): 161-182.
- Ghani, A. (1998). *Medicinal plants of Bangladesh*. Dhaka, Bangladesh: Asiatic Society of Bangladesh.
- Ghoshal, S., Krishna Prasad, B. N. and Lakshmi, V. (1996). Antiamoebic activity of *Piper longum* fruits against *Entamoeba histolytica* *in vitro* and *in vivo*. *Journal of Ethnopharmacology* 50(3): 167-170.
- Gildberg, A., Bogwald, G. A. J., Johansen, A. and Stenberg, E. (1996). Isolation of acid peptide fractions from a fish protein hydrolysate with strong stimulatory effect on Atlantic Salmon (*Salmo salar*) head kidney leucocytes. *Comparative Biochemical Physiology* 114: 97-101.
- Gopalakannan, A. and Arul, V. (2006). Immunomodulatory effects of dietary intake of chitin, chitosan and levamisole on the immune system of *Cyprinus carpio* and control of *Aeromonas hydrophila* infection in ponds. *Aquaculture* 225: 179-187.
- Govind, P., Madhuri, S. and Mandloi, A.K. (2012). Medicinal plants useful in fish diseases. *Plant Archives* 12(1): 1-4.
- Green, B.W. (2006). Tilapia fingerling production systems. In: *Tilapias: Biology, Culture, and Nutrition*, ed. C. Lim. and C. Webster, pp. 181-210. New York: Food Products Press.
- Guerrero, R. D. (1982). Control of tilapia reproduction. Proceedings from ICLARM 7, Manila, Philippines: In *The Biology and Culture of Tilapias*, ed. R.S.V. Pullin, and R.H. Lowe-McConnell.

- Gupta, M. V. and Acosta, B. O. (2004). Review of global tilapia farming. *A Review of Global Tilapia Farming Practices*, IX(1): 1-47.
- Gur, S., Turgut-Balik, D. and Gur, N. (2006). Antimicrobial activities and some fatty acids of turmeric, ginger root and linseed used in the treatment of infectious diseases. *World Journal of Agricultural Sciences* 2(4): 439-442.
- Gutteridge, J. M. C. (1994). Biological origin of free radicals, and mechanisms of antioxidant protection. *Chemico-Biological Interactions* 91(2-3): 133-140.
- Halver, J. E, and Hardy R. W. (2002). *Fish Nutrition*. USA: Academic Press.
- Haney, D. C., Hursh, D. A., Mix, M. C. and Winton, J. R. (1992). Physiological and hematological changes in chum salmon artificially infected with erythrocytic necrosis virus. *Journal of Aquatic Animal Health* 4: 48-57.
- Hannifa, M. A. and Margaret, I. V. (2014). Effect of dietary administration of the herbal additive *Phyllanthus niruri* on growth performance and haematological changes in striped snakehead. *World Aquaculture* 45(3): 66-68.
- Harikrishnan, R., Balasundaram, C. and Heo, M. S. (2010a). Herbal supplementation diets on hematology and innate immunity in goldfish against *Aeromonas hydrophila*. *Fish and Shellfish Immunology* 28(2): 354-361.
- Harikrishnan, R., Balasundaram, C. and Heo, M. S. (2011). Influence of diet enriched with green tea on innate humoral and cellular immune response of kelp grouper (*Epinephelus bruneus*) to *Vibrio carchariae* infection. *Fish and Shellfish Immunology* 30(3): 128-134.
- Harikrishnan, R., Balasundaram, C., Kim, M. C., Kim, J. S., Han, Y. J. and Heo, M. S. (2009). Innate immune response and disease resistance in *Carassius auratus* by triherbal solvent extracts. *Fish and Shellfish Immunology* 27: 508-515.
- Harikrishnan, R., Balasundaram, C., Kim, M. C., Kim, J. S., Han, Y. J. and Heo, M. S. (2009). Innate immune response and disease resistance in *Carassius auratus* by triherbal solvent extracts. *Fish and Shellfish Immunology* 27: 508-515.
- Harikrishnan, R., Balasundaram, C. and Bhuvaneshwari, R. (2005). Restorative effect of *Azadirachta indica* aqueous leaf extract dip treatment on haematological parameter changes in *Cyprinus carpio* (L.) experimentally infected with *Aphanomyces invadans* fungus. *Journal of Applied Ichthyology* 21: 410-414.
- Harikrishnan, R., Heo, J., Balasundaram, C., Kim, M. C., Kim, J. S., Han, Y. J. and Heo, M. S. (2010b). Effect of traditional Korean medicinal (TKM) triherbal extract on the innate immune system and disease resistance in *Paralichthys olivaceus* against *Uronema marinum*. *Veterinary Parasitology* 170(1-2): 1-7.
- Harikrishnan, R., Heo, J., Balasundaram, C., Kim, M. C., Kim, J. S. and Han, Y.

- J. (2010c) Effect of *Punica granatum* solvent extracts on immune system and disease resistance in *Paralichthys olivaceus* against lymphocystis disease virus (LDV). *Fish and Shellfish Immunology* 29: 668–673.
- Harikrishnan, R., Rani, M. N. and Balasundaram, C. (2003). Hematological and biochemical parameters in common carp, *Cyprinus carpio*, following herbal treatment for *Aeromonas hydrophila* infection. *Aquaculture* 221(1-4): 41-50.
- Haslam, E. (1996). Natural polyphenols (vegetable tannins) as drugs: Possible modes of action. *Journal of Natural Products* 59(2): 205-215.
- Hernandez, M. M., Heraso, C., Villarreal, M. L., Vargas-Arispuro, I. and Aranda, E. (1999). Biological activities of crude plant extracts from *Vitex trifolia* L. (Verbenaceae). *Journal of Ethnopharmacology* 67(1): 37-44.
- Hlavova, V. (1993). References Values of the haematological indices in grayling (*Thymallus thymallus* linnaeus). *Comparative Biochemistry and Physiology* 105: 525-532.
- Holst, B. and Williamson, G. (2008). Nutrients and phytochemicals: from bioavailability to bioefficacy beyond antioxidants. *Current Opinion in Biotechnology* 19: 73-82.
- Homatowska, A., Wojtaszek, J. and Adamowicz, A. (2002). Haematological indices and circulating blood picture in the sunbleak, *Leucaspis delineatus* (Heckel, 1843). *Zoologica Poloniae* 47(3-4): 57-68.
- Hossain, M. M., Paul, N., Sohrab, M. H., Rahman, E. and Rashid, M. A. (2001). Antibacterial activity of *Vitex trifolia*. *Fitoterapia* 72(6): 695-697.
- Houston, A. H. Blood and circulation. In Schreck, C. B. and Moyle, P. B. (Ed.), *Methods in fish biology* (pp. 273-335). Bethesda, Maryland: American Fisheries Society.
- Hrubec, T. C., Cardinale, J. L. and Smith, S. A. (2000). Hematology and plasma chemistry reference intervals for cultured tilapia (hybrid). *Veterinary Clinical Pathology* 29 (1): 7-12.
- Ibrahim, J. (2004). Medicinal plant research in Malaysia: Scientific interests and advances. *Jurnal Sains Kesihatan Malaysia* 2(2): 27-46.
- Immanuel, G., Uma, R. P., Iyapparaj, P., Citarasu, T., Peter, S. M., Babu, M. M. and Palavesam, A. (2009). Dietary medicinal plant extracts improve growth, immune activity and survival of tilapia *Oreochromis mossambicus*. *Journal of Fish Biology* 74(7): 1462-1475.
- Immanuel, G., Vincybai, V. C., Sivaram, V., Palavesam, A. and Marian, M. P. (2004). Effect of butanolic extracts from terrestrial herbs and seaweeds on the survival, growth and pathogen (*Vibrio parahaemolyticus*) load on shrimp *Penaeus indicus* juveniles. *Aquaculture* 236: 53-65.
- Iruthayam, V. K., Gurusamy, C., Thangapandi, V., Syed, H. P. S., Jayaraj, M. 2014. Medicinal plants as immunostimulants for health management in Indian cat fish. *Journal of Coastal Life Medicine* 2(6): 426-430.

- Iversen, M., Finstad, B. and Nilssen, K. J. (1998): Recovery from loading and transport stress in Atlantic salmon (*Salmo salar* L.) smolts. *Aquaculture* 168: 387-394.
- Jamal, P., Barkat, A. A. and Amid, A. (2010). Distribution of phenolics in various Malaysian medicinal plants. *Journal of Agricultural Sciences* 10(21): 2658-2662.
- Jamaludin, M. D., Husna Hawa. M. H., Ridzwan, H. and Muhammad, T. (2011). Phytochemicals screening and antioxidant activities of Malaysian *Donax grandis* extracts. *European Journal of Scientific Research* 61(4): 572-577.
- Jameela, M. S., Mohideen, A. H., Sunitha, K. S. and Narayanan, M. (2011). Antibacterial activities of three medicinal plant extract against fish pathogens. *International Journal of Biological Technology* 2(2): 57-60.
- Jauncey. K. (2000). Nutritional requirements. P. 327–375. In: M. C. M. Beveridge and B. J. McAndrew (eds.) *Tilapias: Biology and Exploitation, Fish and Fisheries Series 25*, The Netherlands: Kluwer Academic Publishers.
- Jegade, T. (2010). Control of reproduction in *Oreochromis niloticus* (Linnaeus 1758) using *Hibiscus Rosa-sinensis* (Linn.) leaf meal as reproduction inhibitor. *Journal of Agricultural Science* 2(4): 149-154.
- Ji, S.C., Takaoka, O., Jeong, G.S., Lee, S.W., Ishima, K., Seoka, M. and Takii, K. (2007). Dietary medicinal herbs improve growth and some non-specific immunity of red sea bream *Pagrus major*. *Fisheries Science* 73: 63-69.
- Jian, J. and Wu, Z. (2003). Effects of traditional Chinese medicine on nonspecific immunity and disease resistance of large yellow croaker, *Pseudosciaena crocea* (Richardson). *Aquaculture* 218: 1-9.
- Jian, J. and Wu, Z. (2004). Influences of traditional Chinese medicine on non-specific immunity of Jian Carp (*Cyprinus carpio* var. *Jian*). *Fish and Shellfish Immunology* 16(2): 185-191.
- Karpagam, B. and Krishnaveni, N. (2014). Effect of supplementation of selected plant leaves as growth promoters of tilapia fish (*Oreochromis mossambicus*). *Research Journal of Recent Sciences* 3: 120-123.
- Kavitha, P. and Subramanian, P. (2011). Influence of *Tribulus terrestris* on Testicular Enzyme in Fresh Water Ornamental Fish *Poecilia latipinna*. *Fish Physiology Biochemistry* 37: 801-807.
- Kazama, C.C., Uchida, D.T., Canzi, K.N., Souza, P.D. Crestani, S., Gasparotto Junior, A. and Laverde Junior, A. (2012). Involvement of arginine-vasopressin in the diuretic and hypotensive effects of *Pereskia grandifolia* Haw. (Cactaceae). *Journal of Ethnopharmacology*, 144: 86–93.
- Khan, A., Rahman, M. and Islam, M.S. (2008). Antipyretic activity of *Peperomia pellucida* leaves in rabbit. *Turkey Journal Biology* 32: 37-41.
- Khan, M.R., Omoloso, A.D. (2002). Antibacterial activity of *Hygrophila stricta* and *Peperomia pellucida*. *Fitoterapia* 73: 251-254.

- Khan, S., Khan, G.M., Mehsud, S., Rahman, A. and Khan, F. (2004). Antifungal activity of *Tamarix dioica*. *Journal of Medical Sciences* 2(2): 40-42.
- Khattab, Y.A.E., Shalaby, A.M.E., Sharaf, S.M., El-Marakby, H. and Rizialla, E.H. (2004). The physiological changes and growth performance of the Nile tilapia *Oreochromis niloticus* after feeding with Biogen® as growth promoter. *Egyptian Journal of Aquatic Biology and Fisheries* 8(2): 145-158.
- Kim, D.S., Noah, C.H., Jung, S.W. and Jo, J.Y. (1998). Effect of Obosan supplemented diet on growth, feed conversion ratio and body composition of Nile tilapia, *Oreochromis niloticus*. *Journal of Aquaculture* 11: 83-90.
- Kim, J.D. (2000). Effects of dietary microbial phytase supplementation on growth and feed utilization of Japanese flounder (*Paralichthys olivaceus*). *Annals Animal Research Science* 11: 46-59.
- Kim, S.K., Takeuchi, T., Yokoyama, M. and Murata, Y. (2003). Effect of dietary supplementation with taurine, beta-alanine and GABA on the growth of juvenile and fingerling Japanese flounder *Paralichthys olivaceus*. *Fish Science* 69: 242-248.
- Kirtikar, K.R. and Basu, B.D. (1999) Indian Medicinal Plants. Dehradun: International Book Distributors, Book Sellers and Publishers.
- Kolkovski, Sarjit (2011, March-April). F: Herbal Medicine. *International Aquafeed* 28-31.
- Krishnaraj, A.V., Rao, T.V.N., Sundararaju, D., Vanisree, M., Tsay, H.S. and Subbaraju, G.V. (2005). Assessment of bioactivity of Indian medicinal plants using brine shrimp (*Artemia salina*) lethality assay. *International Applied Science and Engineering* 3: 125-134.
- Kumar, A., Ilavarasan, R., Jayachandran, T., Decaraman, M., Aravindhan, P., Padmanabhan, N. and Krishnan, M. R. V. (2009). Phytochemicals investigation on a tropical plant, *Syzygium cumini* from Kattuppalayam, Erode District, Tamil Nadu, South India. *Pakistan Journal of Nutrition* 8(1): 83-85.
- Kurogi, J. and Iida, T. (1999). Social suppresses defense activities of neutrophils in tilapia. *Fish Pathology* 34: 15-18.
- Laxmikant, K. (2012). *Vitex trifolia* linn. (Verbanaceae): A review on pharmacological and biological effects, isolated and known potential phytoconstituents of therapeutic importance. *International Journal of Research in Pharmaceutical Sciences* 3(3): 441-445.
- Lee, Y.S., Han, O.K., Park, C.W., Yang, C.H., Jeon, T.W., Yoo, W.K., Kim, S.H. and Kim, H.J. (2005). Pro-inflammatory gene expression and nitric oxide regulation of aqueous extracted *Astragali radix* in RAW 264.7 macrophage cells. *Journal of Ethnopharmacology* 100: 289-294.
- Liu, C., Tsai, W., Lin, Y., Liao, J., Chen, C. and Kuo, Y. (2004). The extracts from *Nelumbo nucifera* suppress cell cycle progression, cytokine genes expression, and cell proliferation in human peripheral blood mononuclear

cells. *Life Science* 75: 699-716.

- Logarto Parra, A., Silva Yhebra, R., Guerra Sardinias, I. and Iglesias Buela, L. (2001). Comparative study of the assay of *artemia salina* L. and the estimate of the medium lethal dose (LD₅₀ value) in mice, to determine oral acute toxicity of plant extracts. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology* 8(5): 395-400.
- Lorenzi, H. and Matos, F.J.A. (2002). Plantas Medicinais no Brasil: Nativas e Exo'ticas. Instituto Plantarum, Sao Paulo, Nova Odessa, pp. 512.
- MacMillan, J.R. (2001). Aquaculture and antibiotic resistance: a negligible public health risk? *Journal of the World Aquaculture Society* 32: 49-68.
- Magdelin, S.M. (2005). Culture of ornamental fish, Black molly (*Poecilia sphenops*) using medicinal plants having immunostimulant characteristics. M.Phil Dissertation, Manonmaiam Sundaranar University.
- Maheshappa, K. (1993). Effect of different doses of livol on growth and body composition of rohu, *Labeo rohita* (Ham.). M.F.Sc Thesis, University of Agricultural Science, Bangalore.
- Mair G.C. and Little D.C. (1991). Population control in farmed tilapia. *NAGA, ICLARM Quarterly* 14: 8-13.
- Majumder, P., Abraham, P. and Satya, V. (2011). Ethno-medicinal, phytochemical and pharmacological review of an amazing medicinal herb *Peperomia pellucida* (L.) HBK. *Research Journal of Pharmaceutical, Biological and Chemical Sciences* 2(4): 358-364.
- Makkar, H.P.S., Francis, G. and Becker, K. (2007). Bioactivity of phytochemicals in some lesser-known plants and their effects and potential applications in livestock and aquaculture production systems. *Animal* 1(9): 1371-1391.
- Manokaran, N. (1992). An overview of biodiversity in Malaysia. *Journal of Tropical Forest Science* 5(2): 271-290.
- Martins, M.L., Mourino, J.L., Amaral, G.V., Vieira, F.N., Dotta, G., Jatoba, A.M.B., Pedrotti, F.S., Jeronimo, G.T., Buglione-Neto, C.C. and Pereira, J.G. (2008). Haematological changes in Nile tilapia experimentally infected with *Enterococcus* sp. *Brazilian Journal of Biology* 3: 631-637.
- Mauel, M.J., Miller, D.L. Anita, L. and Merrill, A.L. (2007). Hematologic and plasma biochemical values of healthy hybrid tilapia (*Oreochromis aureus* x *Oreochromis nilotica*) maintained in a recirculating system. *Journal of Zoo and Wildlife Medicine* 38(3): 420-424.
- Maznah, I., Manickam, E.D.A.M., Asmah, R. and Asmah, Y. (2000). Chemical composition and antioxidant activity of *Strobilanthes crispus* leaf extract. *The Journal of Nutritional Biochemistry* 11: 536-542.
- Mehana, E.E., Rahmani, A.H. and Aly, S.M. (2015). Immunostimulants and Fish Culture: An Overview. *Annual Research and Review in Biology* 5(6): 477-489.

- Miller, D.S. and Bender, A.E. (1955). The determination of net protein utilization by a shortened method. *British Journal of Nutrition* 9(4): 382-388.
- Ministry of Agriculture, (2012). <http://www.moa.gov.my/en/industri-herba>. Retrieved 15 August 2013.
- Minomol, M. (2005). *Culture of Gold fish Carassius auratus using medicinal plants having immunostimulant characteristics*. Master Thesis, MS University.
- Mires, D. (1995). The tilapias. pp. 133–152. In: *Production of Aquatic Animals-Fishes*, ed. C.E. Nash, and A. J. Novotony. Elsevier, New York.
- Modadugu, V.G. and Belen, O.A. (2004). A review of global tilapia farming practices. *Aquaculture Asia*, Scandmedia: Thailand. January-March 2004.
- Mohd Fadzelly, A.B., Asmah, R. and Fauziah, O. (2006). Effects of *Strobilanthes crispus* tea aqueous extracts on glucose and lipid profile in normal and streptozotocin-induced hyperglycemic rats. *Plant Foods for Human Nutrition* 61(1), 7-12.
- Moshi, M.J., Innocent, E., Magadula, J.J., Otieno, D.F., Weisheit, A., Mbabazi, P.K. and Nondo, R.S.O. (2010). Brine shrimp toxicity of some plants used as traditional medicines in Kagera Region, north western Tanzania. *Tanzania Journal of Health Research* 12(1): 1-6.
- Muslim, N.S., Ng, K.W., Itam, A., Nassar, Z.D., Ismail, Z. and Abdul Majid, A.M.S. (2010). Evaluation of cytotoxic, anti angiogenic and antioxidant properties of standardized extracts of *Strobilanthes crispus* leaves. *International Journal of Pharmacology* 6(5): 591-599.
- Negi, P.S., Jayprakash, G.K., Rao, L.M. and Sakarian, K.K. (1999). Antimicrobial activity of turmeric oil. *Journal Agriculture and Food Chemistry* 47: 4297-4300.
- NRC (National Research Council) (1993). *Nutritional Requirements of Fish*, National Academy Press, Washington, DC, USA.
- Nussey, G., Van Vuren, J.H.J. and Du Preez, H.H. (2000). Bioaccumulation of chromium, manganese, nickel and lead in the tissues of the moggel, *Labeo umbratus* (Cyprinidae), from Witbank Dam, Mpumalanga. *Water Sines* 26(2): 269-284.
- Nya, E.J. and Austin, B. (2011). Development of immunity in rainbow trout (*Oncorhynchus mykiss*, Walbaum) to *Aeromonas hydrophila* after the dietary application of garlic. *Fish and Shellfish Immunology* 30: 845-850.
- Nya, E.J. and Austin, B. (2009). Use of garlic, *Allium sativum*, to control *Aeromonas hydrophila* infection in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of Fish Diseases* 32: 963-970.
- Okwu, D.E. and Josiah, C. (2006). Evaluation of the chemical composition of two Nigerian medicinal plants. *African Journal of Biotechnology* 5(4): 357-361.
- Oliveira, S.H.D.S., Soares, M.J.G.O. and Rocha, P.D.S. (2010). Use of

- collagen and *Aloe vera* in ischemic wound treatment: study case. *Revista Da Escola de Enfermagem Da USP* 44(2): 344–349.
- Olmedo Sanchez, J. A., Curiel Flores, A. and Orozco Hernandez, J. R. (2009). The effect of a herbal growth promoter feed additive on shrimp performance. *Research Journal of Biological Sciences* 4(9): 1022-1024.
- Pakravan, S., Hajimoradloo, A. and Ghorbani, R. (2011). Effect of dietary willow herb, *Epilobium hirsutum* extract on growth performance, body composition, haematological parameters and *Aeromonas hydrophila* challenge on common carp, *Cyprinus carpio*. *Aquaculture Research* 43: 861-869.
- Pandey, G., Sharma, M. and Mandloi, A.K. (2012). Medicinal plants useful in fish diseases. *Plant Archives* 12(1): 1-4.
- Pappachen, L. K. and Annam, C. (2013). Preliminary phytochemical screening and *in-vitro* cytotoxicity activity of *Peperomia pellucida* Linn. *Pharmacie Globale: International Journal Of Comprehensive Pharmacy* 4(8): 1-5.
- Parekh, J. and Chanda, S. (2008). *In vitro* antifungal activity of methanol extracts of some Indian medicinal plants against pathogenic yeast and moulds. *African Journal of Biotechnology* 7(23): 4349-4353.
- Park, K. H. and Jeong, H. D. (1996). Enhanced resistance against *Edwardsiella tarda* infection in tilapia (*Oreochromis niloticus*) by administration of protein-bound polysaccharide. *Aquaculture* 143: 135-143.
- Paroda, R.S. and Mal, B. (1989). New Plant Sources for Food and Industry in India. In *New Crops for Food and Industry* ed. G.E. Wichens, N. Haq, and P. Day, pp. 134-149. London: Chapman and Hall.
- Perera, R.P., Johnson, S.K. and Lewis, D.H. (1997). Epizootiological aspects of *Streptococcus iniae* affecting tilapia in Texas. *Aquaculture* 152: 25-33.
- Perumal Samy, R. and Gopalakrishnakone, P. (2008). Therapeutic potential of plants as anti-microbials for drug discovery. *Evidence-Based Complementary and Alternative Medicine* 7(3): 283-294.
- Petrovic, S., Ozretic, B. and Krajnovic-Oaretic, M. (1996). Cytosolic aspartate aminotransferase from grey mullet (*Mugil auratus* Risso) red muscle: isolation and properties. *International Journal Biochemistry Cell Biology* 28(8): 873–881.
- Pickering, A.D. (1981). Introduction: the concept of biological stress. In *Stress and Fish* ed. A.D. Pickering, pp. 1-10. New York: Academic Press.
- Platel, K., Rao, A., Saraswahi, G. and Srinivasan, K. (2002). Digestive stimulant action of three Indian spice mixes in experimental rats. *Die Nahrung* 46: 394-398.
- Plumb, J.A. (1999). Health maintenance. In *Health Maintenance and Principal Microbial Diseases of Cultured Fishes*, ed. by J.A. Plumb, pp. 297-301. Iowa: Iowa State University Press.

- Pollack, S.J., Ottinger, M.A., Sullivan, C.V. and Woods, L.C. (2003). The effects of the soy isoflavone genistein on the reproductive development of striped bass. *North American Journal of Aquaculture* 65: 226-234.
- Ponpornpisit, A., Endo, M. and Murata, H. (2001). Prophylactic effects of chemicals and immunostimulants in experimental *Tetrahymena* infections of guppy. *Fish Pathology* 36(1): 1-6.
- Prasad, S. and Variyur Padhyoy, K.B. (1993). Chemical investigation of some commonly used spices. *Aryavaidyan* 6(4): 262–267.
- Prusty, A.K., Sahub, N.P., Palb, A.K., Reddy, A.K. and Shivendra Kumar, A.K. (2007). Effect of dietary tannin on growth and haemato-immunological parameters of *Labeo rohita* (Hamilton) fingerlings. *Animal Feed Science and Technology* 136: 96–108.
- Pulsford, A.L., Lemaire-Gong, S., Tomlinson, M., Coiling Wood, N. and Glynn, P.J. (1994). Effects of acute stress on the immune system of the dab, *Limanda limanda* *Comparative Biochemistry and Physiology* 109(2): 129-139.
- Punitha, S.M.J., Babu, M.M., Sivaram, V., Shankar, V.S., Dhas, S.A., Mahesh, T.C., Immanuel, G. and Citarasu, T. (2008). Immunostimulating influence of herbal biomedicines on nonspecific immunity in Grouper *Epinephelus tauvina* juvenile against *Vibrio harveyi* infection. *Aquaculture International* 16: 511–523.
- Qiu, X.C., Zhou, H.Q., Hua, X.M., Liu, X.G. and Cao, D. (2004). The effect of dietary Chinese additives on digestibility of Allogynogenetic crucian carp. *Journal of Xinyang Normal University (Natural Science Edition) (China)* 17: 54–56.
- Qiu, X.C., Zhou, H.Q., Liu, X.G., Hua, X.M., Cao, D. and Zhang, D.L. (2002). The effect of dietary Chinese herb additives on growth and digestibility of Allogynogenetic crucian carp. *Journal of Fisheries of China* 26: 551–555.
- Ragasa, C.Y., Dumato, M. and Rideout, J.A. (1998). Antifungal compounds from *Peperomia pellucida*. *ACGC Chemical Research Communications* 7: 54-61.
- Rajan, D., Suresh, J., Paramakrishnan, N., Sri Vasavi Reddy, A. and Nayeem, M. (2012). Review on Phytochemical and pharmacological properties of *Vitex trifolia* linn. *Research J. Pharmacognosy and Phytochemistry* 4(2): 124-129.
- Rani, T.V.J. (1999). Fourth year annual report (CSIR Research Associate ship) submitted to Council of Scientific and Industrial Research, New Delhi.
- Rasheed, V. and Plumb, J. (1984). Pathogenicity of a non-hemolytic group B Streptococcus sp. in gulf killifish, *Fundulus grandis* (Baird & Girard). *Aquaculture* 37: 97-105.
- Ravikumar, S., Palani Selvan, G. and Anitha Anandha Gracelin, N. (2010). Antimicrobial activity of medicinal plants along Kanyakumari coast, Tamil

- Nadu, India. *African Journal of Basic and Applied Sciences* 2(5-6): 153-157.
- Rawling, M.D., Merrifield, D.L. and Davies, S.J. (2009). Preliminary assessment of dietary supplementation of Sangrovit® on red tilapia (*Oreochromis niloticus*) growth performance and health. *Aquaculture* 294: 118-122.
- Reed, L.J. and Muench, H. (1938). A simple method of estimating fifty percent endpoints. *American Journal of Epidemiology* 27(3): 493-497.
- Rehulka, J. (1998). Blood indices of the rainbowtrout, *Oncorhynchus mykiss* (Walbaum) in *Aeromonas*-induced ulcerous dermatitis. *Acta Veterinaria Brasilica* 67: 317-322.
- Rieser, M.J., Gu, Z.M., Fang, X.P., Zeng, L., Wood, K.V. and McLaughlin, J.L. (1996). Five novel mono-tetrahydrofuran ring acetogenins from the seeds of *Annona muricata*. *Journal of Natural Products* 59(2): 100-108.
- Rios, J.L. and Recio, M.C. (2005). Medicinal plants and antimicrobial activity. *Journal of Ethnopharmacology* 100: 80-84.
- Roberts, R.J. (1993). Motile aeromonad septicemia. In *Bacterial Diseases of Fish*, ed. V. Inglis, R.J. Roberts, and N.R. Bromage, pp. 143-156. London: Blackwell Scientific Publications.
- Robinson, J.A and Meyer, F.P. (1966). Streptococcal fish pathogen. *Journal of Bacteriology* 92: 512-512.
- Rojas, J.J., Ochoa, V.J., Ocampo, S.A. and Munoz, J.F. (2006). Screening for antimicrobial activity of ten medicinal plants used in Colombian folkloric medicine: a possible alternative in the treatment of non-nosocomial infections. *BMC Complementary and Alternative Medicine* 6(2): 1-6.
- Sahu, N.P., Banerji, N. and Chakravarti, R.N. (1974). A new saponin of oleanolic acid from *Pereskia grandifolia*. *Phytochemistry* 13: 529- 530.
- Sahu, P.K., Giri, D.D., Singh, R., Pandey, P., Gupta, S., Shrivastava, A.K., Kumar, A. and Pandey, K.D. (2013). Therapeutic and medicinal uses of *Aloe vera*: A review. *Pharmacology and Pharmacy* 4: 599-610.
- Sahu, S. 2004, *Antibacterial activity of plant extracts on fish microbial pathogens*, Master. Thesis, Cifa, Kausalyaganga, Bhubaneswar.
- Sahu, S., Das, B.K., Pradhan, J., Mohapatra, B.C., Mishra, B.K. and Sarangi, N. (2007). Effect of *Magnifera indica* kernel as a feed additive on immunity and resistance to *Aeromonas hydrophila* in *Labeo rohita* fingerlings. *Fish and Shellfish Immunology* 23: 109-118.
- Sakai, M. (1999). Current research status of fish immunostimulants. *Aquaculture* 172: 63-92.
- Sakdarat, S., Shuyprom, A., Pientong, C., Ekalaksananan, T. and Thongchai, S. (2009). Bioactive constituents from the leaves of *Clinacanthus nutans* Lindau. *Bioorganic and Medicinal Chemistry* 17: 1857-1860.
- Sambhu, C. and Jayaprakas, V. (2001). Livol (IHF-1000), a new herbal growth

- promoter in white prawn, *Penaeus indicus* (Crustacea). *Indian Journal of Marine Sciences* 30: 38–43.
- Sankar, G., Elavarasi, A., Sakkaravarthi, K. and Ramamoorthy, K. (2011). Biochemical changes and growth performance of black tiger shrimp larvae after using *Ricinus communis* extract as feed additive. *International Journal of PharmTech Research* 3(1): 201-208.
- Santos, Y., Toramnz, A.E., Barja, J.L., Nieto, T.P. and Villa, G. (1988). Virulence properties and enterotoxin production of *Aeromonas* strains isolated from fish. *Infection and Immunity* 56: 3285–3293.
- Saunders, D.C. (1968). Differential blood cell counts of 50 species of fishes from the Red Sea. *Copeia* 3: 491-498.
- Saunders, D.C. 1996. Differential blood cell counts of 121 species of marine fishes of Puerto Rico. *Transactions of the American Microscopical Society* 85(3): 427-449.
- Sayuthi, S. (1993). Fish diseases in Malaysia: status and problems, In *Proceedings of the Aquaculture Workshop for SEAFDEC/AQD Training Alumni, Iloilo, Philippines, September 8- 11, 1992*, 56–61. Tigbauan, Iloilo, Philippines: Southeast Asian Fisheries Development Center, Aquaculture Department.
- Scalbert, A. (1991). Antimicrobial properties of tannins. *Phytochemistry* 30(12): 3875-3883.
- Scott, A.L. and Rogers, W.A. (1981). Hematological effects of prolonged sublethal hypoxia on channel catfish *Ictalurus punctatus* (Rafinesque). *Journal of Fish Biology* 18: 591-601.
- Shah, M.A., Mohammad, S.R., Mphamed, Z.R., Khandaker, S.A. and Mohammad, A.R. (2010). Free radical scavenging activities of some indigenous plants for Bangladesh. *Bangladesh Pharmaceutical Journal* 13(1): 68-70.
- Shahidi Bonjar, G.H. (2004). New approaches in screening for antibacterials in plants. *Asian Journal of Plant Sciences* 3(1): 55-60.
- Sheikhzadeh N., Nofouzi K., Delazar A. and Oushani A.K. (2011). Immunomodulatory effects of decaffeinated green tea (*Camellia sinensis*) on the immune system of rainbow trout (*O. mykiss*). *Fish and Shellfish Immunology* 31: 1268-1269.
- Shelton W.L. (2002). Tilapia culture in the 21st century p. 1-20. In Gurrero R. D. III and M. R. Guerrero-del Castillo (eds.) *Proceedings of the International Forum on Tilapia Farming in the 21st Century (Tilapia Forum 2002)*, 184p. Philippine Fisheries Association Inc. Los Bonos, Laguna, Philippines.
- Sim, K.S., Sri Nurestri, A.M. and Norhanom, A.W. (2010). Phenolic content and antioxidant activity of *Pereskia grandifolia* Haw. (Cactaceae) extracts. *Pharmacognosy Magazine* 6(23): 248–254.
- Siripongvutikorn S., Thummaratwasik, P. and Huang, Y. (2005). Antimicrobial and antioxidant effects of Thai seasoning, *Tom-Yum*. *Journal Food*

- Sivaram, V., Babu, M.M., Immanuel, G., Murugadass, S., Citarasu, T. and Marian, M.P. (2004). Growth and immune response of juvenile greasy groupers (*Epinephelus tauvina*) fed with herbal antibacterial active principle supplemented diets against *Vibrio harveyi* infections. *Aquaculture* 237: 9–20.
- Smith, K.G., Diop, M.D., Niane, M. and Darwall, W.R.T. (2009). The status and distribution of freshwater biodiversity in Western African. IUCN Gland, Switzerland and Cambridge, UK :. 98p.
- Soediro, I., Pellecuer, J., Andary, C. and Privat, G. (1987). *Strobilanthes crispus* (L) B1 I: Pemeriksaan asam fenolat. *Acta Pharmaceutical Indonesia*, XII: 1 - 7.
- Soosean, C., Marimuthu, K., Sudhakaran, S. and Xavier, R. (2010). Effects of mangosteen (*Garcinia mangostana* L.) extracts as a feed additive on growth and haematological parameters of African catfish (*Clarias gariepinus*) fingerlings. *European Review for Medical Pharmacological Sciences*, 14: 605-611.
- Sotolu, A.O. and Faturoti, E.O. (2011). Growth performance and hematological effects of varying dietary processed *Leucaena leucocephala* seed meal in *Clarias gariepinus* juveniles. *African Journal of Food, Agriculture, Nutrition and Development* 11(1): 4547-4557.
- Sovlo, A. and Nikinmaa, M. (1981). The swelling of erythrocytes in relation to the oxygen affinity of the blood of the rainbow trout *Salmo gairdneri* Richardson. In: *Stress and fish*, ed. A.D. Pickering, pp.103-119. UK: Academic Press.
- Sri Nurestri, A.M., Sim, K.S. and Norhanom, A.W. (2009). Phytochemical and cytotoxic investigations of *Pereskia grandifolia* Haw. (Cactaceae) leaves. *Journal of Biological Sciences* 9(5): 488-493.
- Srinivasan, K. (2005). Spices as influencers of body metabolism: an overview of three decades of research. *Food Research International* 38 (1): 77–86.
- Srivastava, A.S. Oohara, I. Suzuki, T. Shenouda, S. Singh, S.N. Chauhan, D.P. Carrier, E. (2004). Purification and properties of cytosolic alanine aminotransferase from the liver of two freshwater fish, *Clarias batrachus* and *Labeo rohita*, *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* 137: 197-207.
- Sunarto, P.A. (1997). *Materia Medika Indonesia*. Penerbit Ditektorat Jenderal Pengawasan Obat dan Makanan, Jakarta, Indonesia.
- Surjushe, A., Vasni, R. and Saple, D.G. (2008). *Aloe vera*: a short review. *Indian Journal Dermatology* 53(4): 163-166.
- Talpur, A.D. (2014). *Mentha piperita* (Peppermint) as feed additive enhanced growth performance, survival, immune response and disease resistance of Asian seabass, *Lates calcarifer* (Bloch) against *Vibrio harveyi* infection. *Aquaculture* 420-421: 71-78.

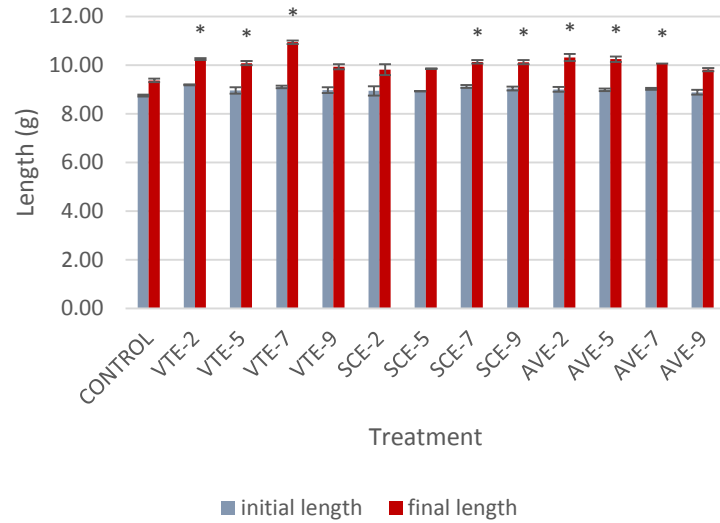
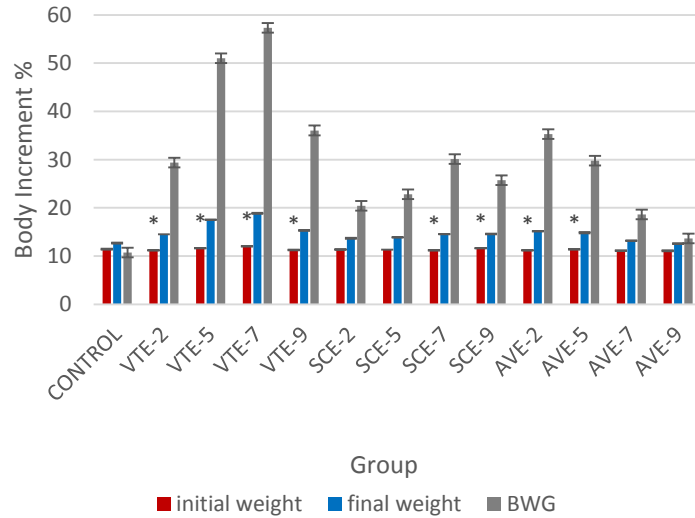
- Talpur, A.D. and Ikhwanuddin, M. (2012). Dietary effects of garlic (*Allium sativum*) on haemato immunological parameters, survival, growth, and disease resistance against *Vibrio harveyi* infection in Asian sea bass, *Lates calcarifer* (Bloch). *Aquaculture* 364-365: 6-12.
- Talpur, A.D., Mohd, I. and Bolong, A.A. (2013). Ginger as immunostimulant-strengthens the non- specific immunity and reduces susceptibility to bacterial infections. *Aquaculture* 401(24): 46-52.
- Teugels, G. G. and D.F.E. Thys van den Oudenaarde, 2003. Cichlidae. p. 521-600. In Paugy D., C. Leveque and g.g Teugels (eds.). The fresh and brackish water fishes of West Africa Volume 2. Coll. tropical fauna and flora 40. Development Research Institute, Paris, France, National Museum of Natural History, Paris, France and the Royal Museum for Central Africa, Tervuren, Belgium, 815p.
- Thabile, P. N. (2008). *Antimicrobial properties of selected Asian herbs*. Master Thesis, University of Florida.
- Thanikachalam, K., Kasi, M. and Rathinam, X. (2010). Effect of garlic peel on growth, hematological parameters and disease resistance against *Aeromonas hydrophila* in African catfish *Clarias gariepinus* (Bloch) fingerlings. *Asian Pacific Journal of Tropical Medicine* 2010:614–618.
- Trewavas, E. 1983. Tilapiine fishes of the genera *Sarotherodon*, *Oreochromis* and *Danakilia*. British Museum of Natural History, London.
- Tsuchiya, H., Sato, M., Miyazaki, T., Fujiwara, S., Tanigaki, S., Ohyama, M., Tanaka, T. and Iinuma, M. (1996). Comparative study on the antibacterial activity of phytochemical flavanones against methicillin-resistant *Staphylococcus aureus*. *Journal of Ethnopharmacology* 50: 27–34.
- Turan, F. (2006). Improvement of growth performance in tilapia (*Oreochromis aureus* Linnaeus) by supplementation of redclover *Trifolium pratense* in diets. *The Israeli Journal of Aquaculture* 58: 34-38.
- Turan, F. and Akyurt, I. (2005). Effects of red clover extract on growth performance and body composition of African catfish *Clarias gariepinus*. *Fisheries Science* 71(3): 618-620.
- Turan, F., Mazlum, Y., Yildirim, B.Y. and Gezer, A. (2012). Use of dietary *Pelargonium sidoides* extract to improve growth and body composition of narrow-clawed crayfish *Astacus leptodactylus* Eschscholtz, 1823 juveniles. *Turkish Journal of Fisheries and Aquatic Science* 12: 233-238.
- Turker, H., Yildirim, A.B. and Karakas, F.P. (2009). Sensitivity of Bacteria isolated from fish to some medicinal plants. *Turkish Journal of Fisheries and Aquatic Sciences* 9: 181-186.
- Turnbull, T. (1999). Infectious salmon anaemia in the United Kingdom. *The Journal of the Fish Veterinary Society* 3: 64-67.
- Vazquez, B., Avila, G., Segura, D. and Escalante, B. (1996). Antiinflammatory activity of extracts from *Aloe vera* gel. *Journal of Ethnopharmacology* 55: 69-75.

- Vijaya, K., Ananthan, S. and Nalini, R. (1995). Antibacterial effect of theaflavin, polyphenon 60 (*Camellia sinensis*) and *Euphorbia hirta* on *Shigella* spp. a cell culture study. *Journal of Ethnopharmacology* 49(2): 115-118.
- Virgili, F. and Marino, M. (2008). Regulation of cellular signals from nutritional molecules: a specific role for phytochemicals, beyond antioxidant activity. *Free Radical Biology and Medicine* 45: 1205-1216.
- Vogler, B.K. and Ernst, E. (1999). *Aloe vera*: a systematic review of its clinical effectiveness. *British Journal of General Practice* 49: 823-828.
- Wei, L.S., Wee, W., Siong, J.Y.F. and Syamsumir, D.F. (2011). Characterization of anticancer, antimicrobial, antioxidant properties and chemical compositions of *Peperomia pellucida* leaf extract. *Acta Medica Iranica* 49(10): 670-674.
- Wepener, V., Van Vuren, J.H.J. and Du Preez, H.H. (1992). Effect of manganese and iron at a neutral and acidic pH on the hematology of the banded tilapia (*Tilapia sparrmanii*). *Bulletin of Environmental Contamination and Toxicology* 49(4): 613-619.
- Whitefield, A.K. and Blaber, S.J.M. (1979). The distribution of the freshwater cichlid *Sarotherodon mossambicus* in esutuariane systems. *Environmental Biology of Fishes* 4:77-81.
- WHO (2002 – 2005). World Health Organization Traditional Medical Strategy, Geneva.
- Winberg, G.G. (1956). Rate of metabolism and food requirements of fishes. Belorussian State University, Minsk. 251.
- Xu, S., Li, N., Ning, M.M., Zhou, C.H., Yang, Q.R. and Wang, M.W. (2006). Bioactive compounds from *Peperomia pellucida*. *Journal of Natural Products* 69: 247-250.
- Yilmaz, E., Ayee Genc, M., Cek, S., Mazlum, Y. and Genc, E. (2006). Effects of orally administered *Ferula coskunii* (Apiaceae) on growth, body composition and histology of common carp, *Cyprinus carpio*. *Journal of Animal and Veterinary Advances* 5(12): 1236-1238.
- Yilmaz, S., Ergun, S. and Celik, E.S. (2012). Effects of herbal supplements on growth performance of sea bass (*Dicentrarchus labrax*): Change in body composition and some blood parameters. *Journal of BioScience and Biotechnology* 1(3): 217-222.
- Yin, G., Ardo, L., Jeney, Z., Xu, P. and Jeney, G. (2008). Chinese herbs (*Lonicera japonica* and *Ganoderma lucidum*) enhance non-specific immune response of tilapia, *Oreochromis niloticus*, and protection against *Aeromonas hydrophila*. In: Diseases in Asian Aquaculture VI. (Bondad-Reantaso MG, Mohan CV, Crumlish M, Subasinghe RP, eds.). Manila, Philippines: Fish Health Section, Asian Fisheries Society. 269-282.
- Yong, Y.K., Tan, J.J., Teh, S.S., Mah, S.H., Ee, G. C. L., Chiong, H. S. and Zuraini, A. (2013). *Clinacanthus nutans* extracts are antioxidant with antiproliferative effect on cultured human cancer cell lines. *Evidence-*

Based Complementary and Alternative Medicine 2013: 1-8.

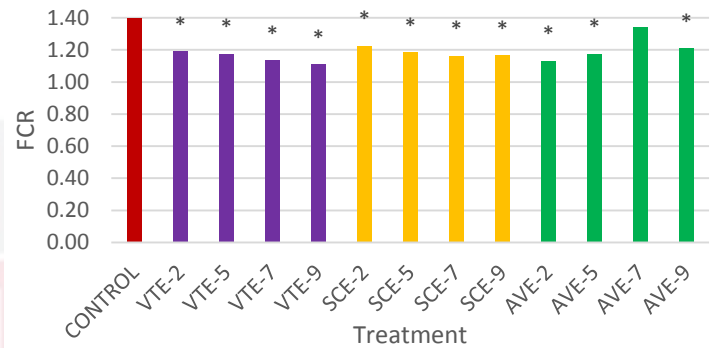
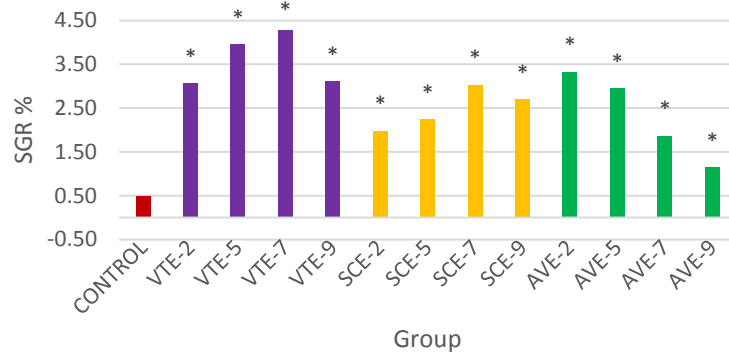
- Yoshida, T., Hatano, T. and Ito, H. (2000). Chemistry and function of vegetable polyphenols with high molecular weights. *BioFactors* 13: 121-125.
- Zareisedehizadeh, S., Tan, C.H. and Koh, H.L. (2014). A review of botanical characteristics, traditional usage, chemical components, pharmacological activities, and safety of *Pereskia bleo* (Kunth) DC. *Evidence-Based Complementary and Alternative Medicine* 2014: 1-11.
- Zheng, Z.L., Tan, J.Y.W., Liu, H.Y., Zhou, X.H., Xiang, X. and Wang, K.Y. (2009). Evaluation of oregano essential oil (*Origanum heracleoticum* L.) on growth, antioxidant effect and resistance against *Aeromonas hydrophila* in channel catfish (*Ictalurus punctatus*). *Aquaculture* 292: 214–218.
- Zilberg, D., Tal, A., Froyman, N., Abutbul, S., Dudai, N. and Golan-Goldhirsh, A. (2010). Dried leaves of *Rosmarinus officinalis* as a treatment for Streptococcosis in tilapia. *Journal of Fish Diseases* 33: 361-369.
- Zolfaghari, A. and Firouzbakhsh, F. (2013). The effect of Basil (*Ocimum basilicum*) aqueous extract on growth changes, hematology and serum biochemical parameters in rainbow trout (*Oncorhynchus mykiss*). *Journal of Veterinary Research* 68 (4): 397–404.
- Zorriehzahra, M. J., Hassan, M. D., Gholizadeh, M. and Saidi, A. A. (2010). Study of some hematological and biochemical parameters of rainbow trout (*Oncorhynchus mykiss*) fry in western part of Mazandaran province, Iran. *Iranian Journal of Fisheries Sciences* 9(1): 185–198.

APPENDICES



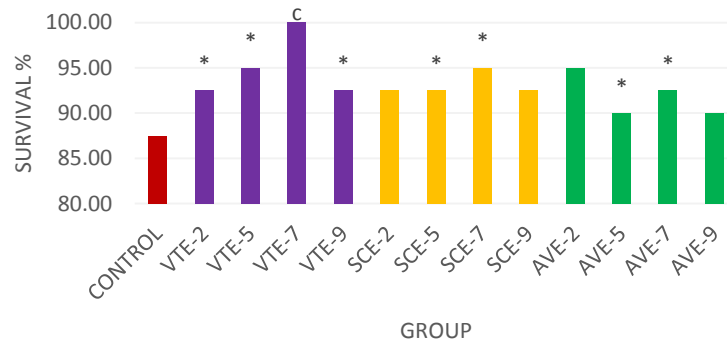
A1 Body gain weight of *Oreochromis* sp.

A2 Length of *Oreochromis* sp.



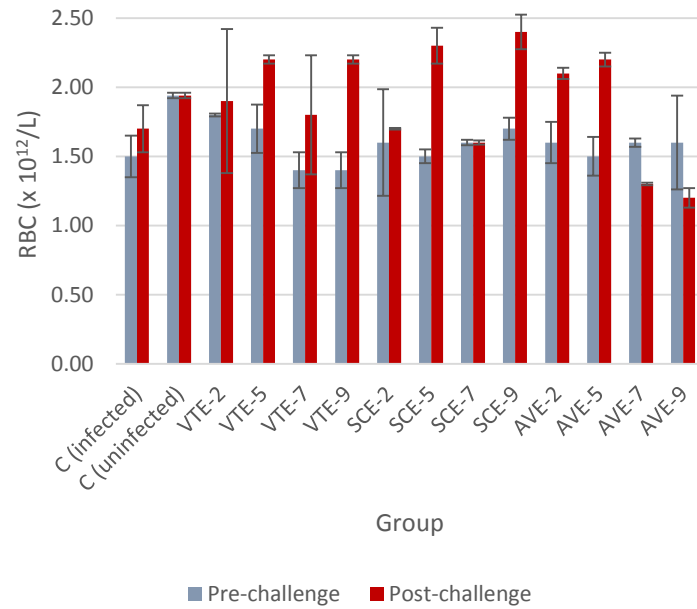
A3 Specific growth rate of *Oreochromis* sp.

A4 Feed conversion ratio of *Oreochromis* sp.

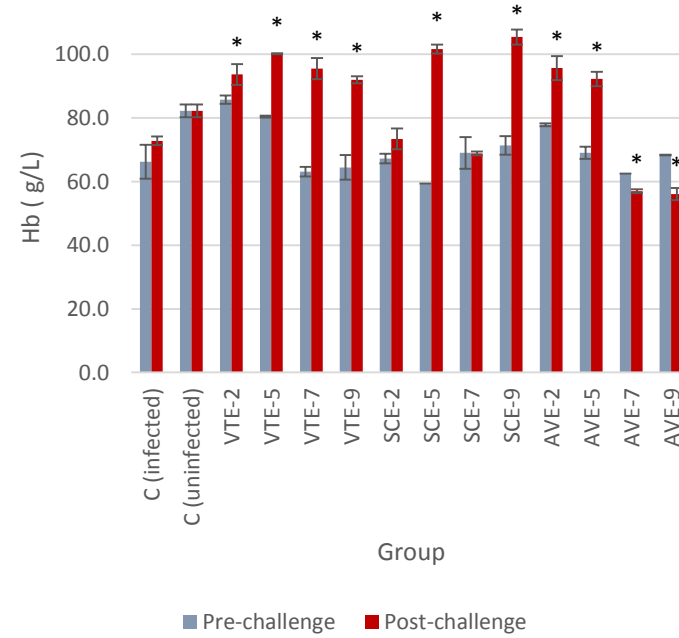


A5 Survival rate of *Oreochromis* sp.

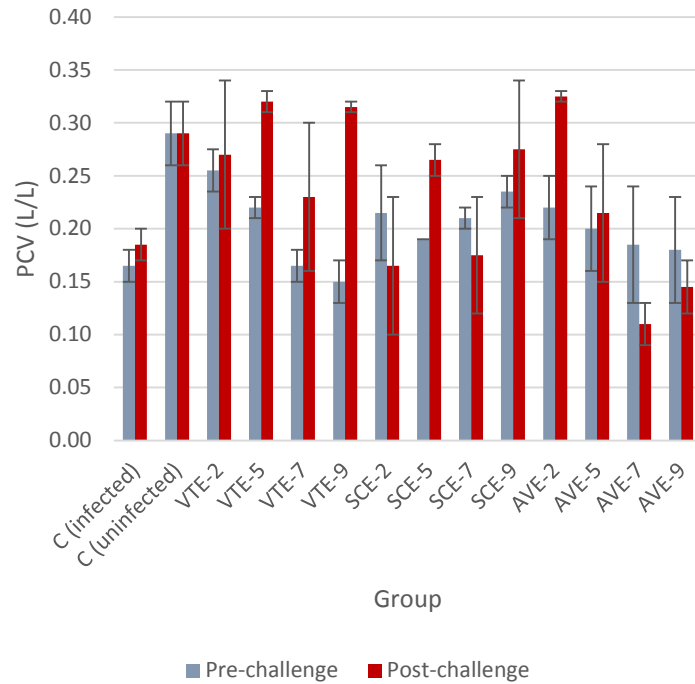
(a) RBC



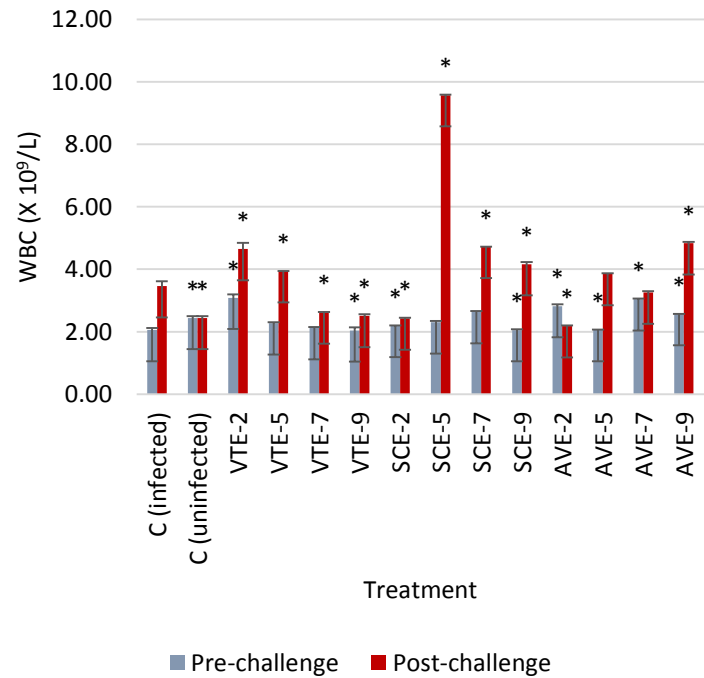
(b) Hb



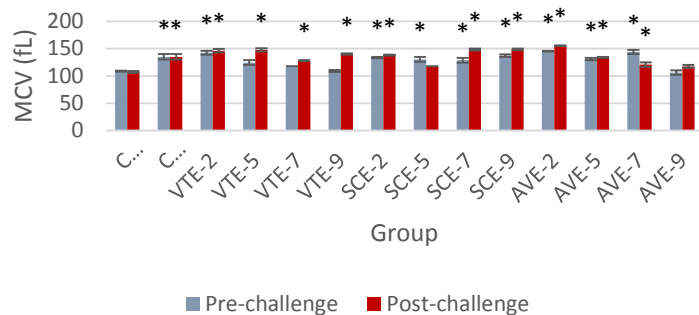
(c) PCV



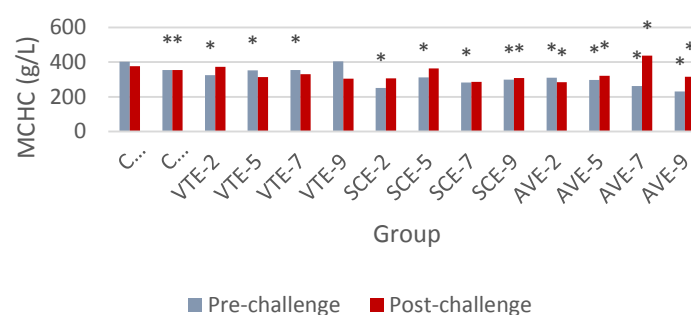
(d) WBC



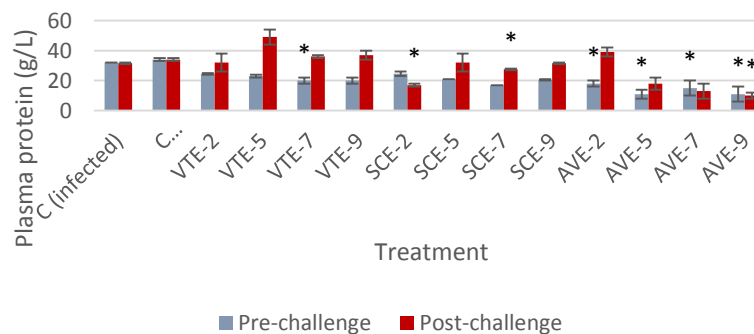
(e) MCV



(f) MCHC

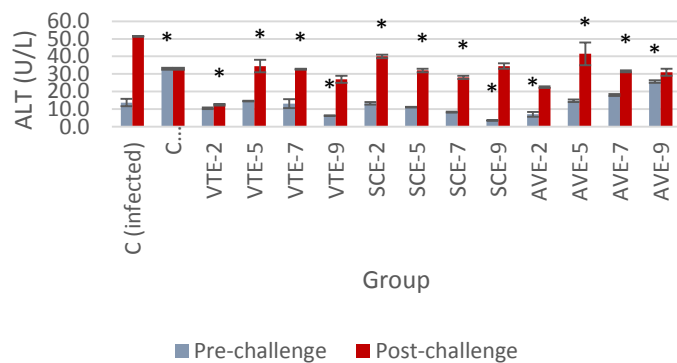


(g) Plasma protein

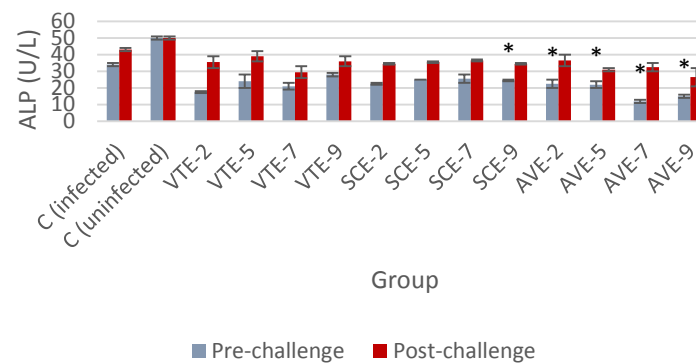


A6 Haematological parameters of *Oreochromis* sp. (pre-challenge) fed &VTE, VTE, SCE, AVE and Control diet at for six-weeks and post-challenge (two-weeks). Data are expressed as the mean $\pm$ SEM. Statistical differences ($P < 0.05$) from the control (infected) group are indicated by asterisks. RBC = red blood cell, Hb = haemoglobin, WBC = white blood cell, PCV = packed cell volume, MCV = mean corpuscular volume, MCHC = mean cellular haemoglobin content, P.protein=plasma protein

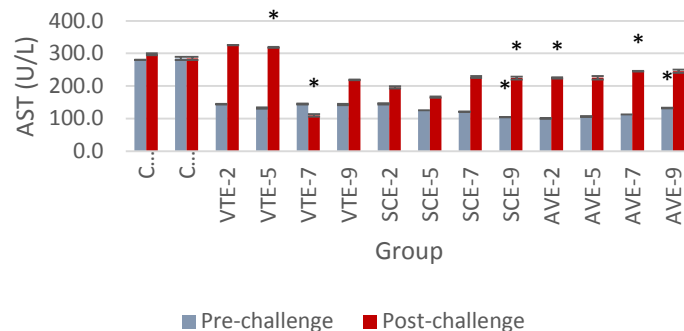
(h) ALT



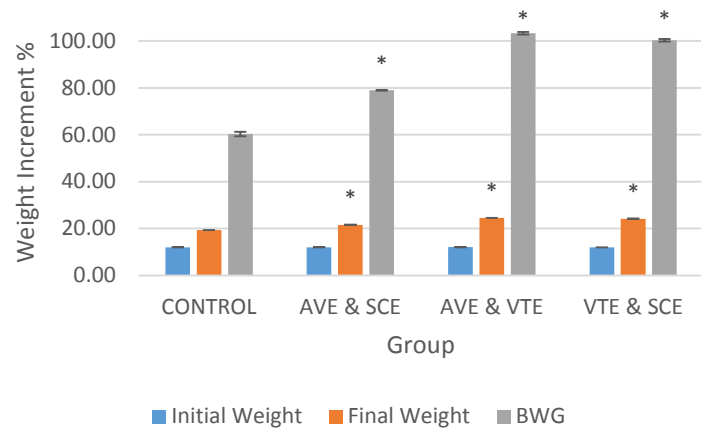
(i) ALP



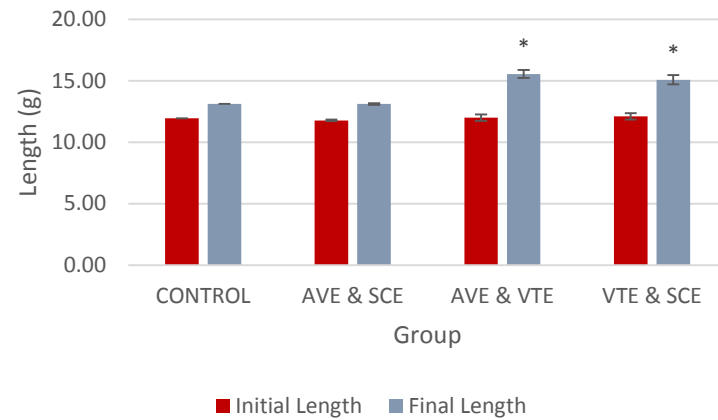
(j) AST



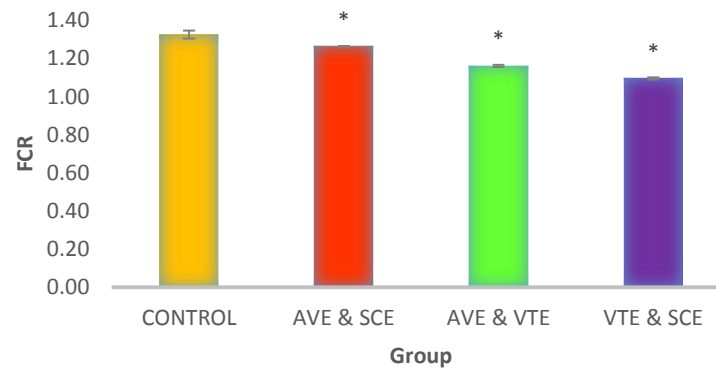
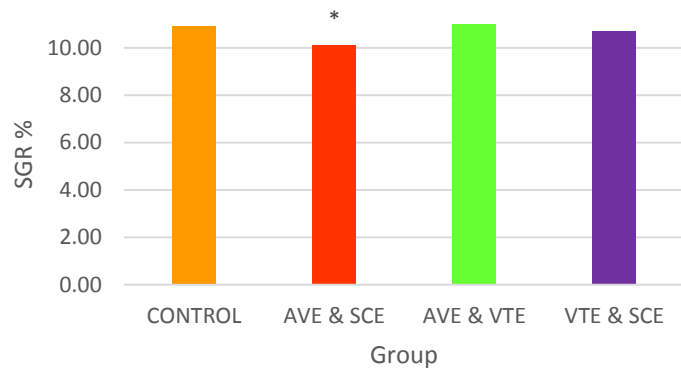
A7 Biochemical parameters of *Oreochromis* sp. (pre-challenge) fed VTE, VTE, SCE, AVE and Control diet at for six-weeks and post-challenge (two-weeks). Data are expressed as the mean $\pm$ SEM. Statistical differences ($P<0.05$) from the control (infected) group are indicated by asterisks. ALT = alkaline aminotransferase, ALP=alkaline phosphatase, AST=aspartate aminotransferase



A8 Body increment of *Oreochromis* sp.

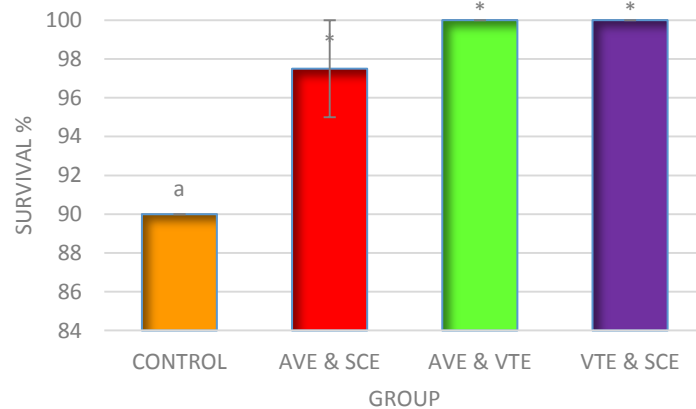


A9 Length of *Oreochromis* sp.



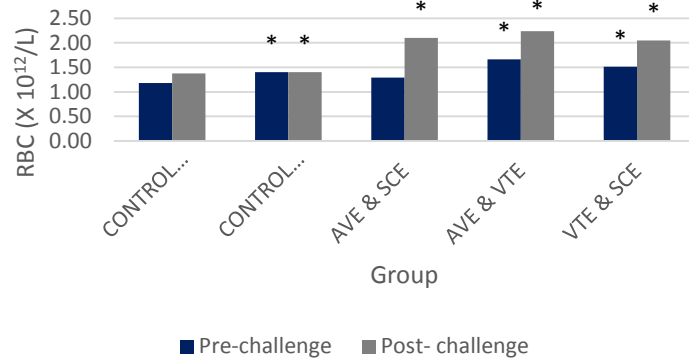
A10 Specific growth rate of *Oreochromis* sp.

A11 Feed conversion ratio of *Oreochromis* sp.

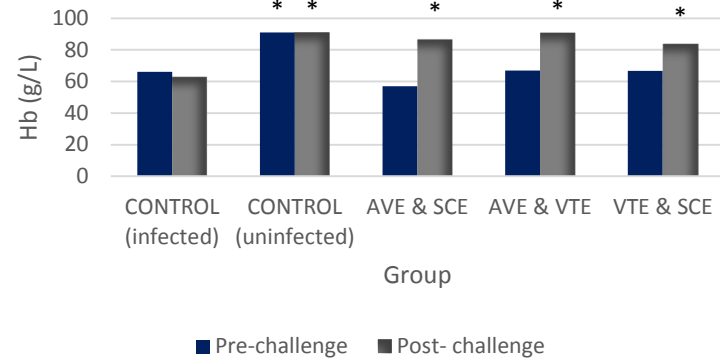


A12 Survival rate of *Oreochromis* sp.

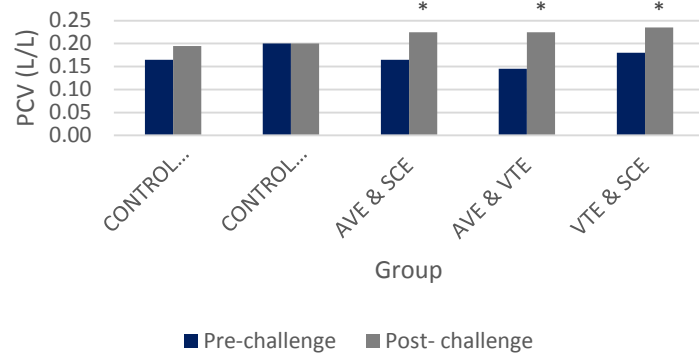
(a) RBC



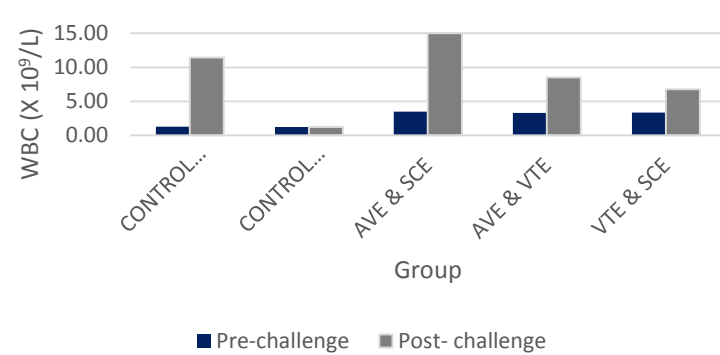
(b) Hb



(c) PCV

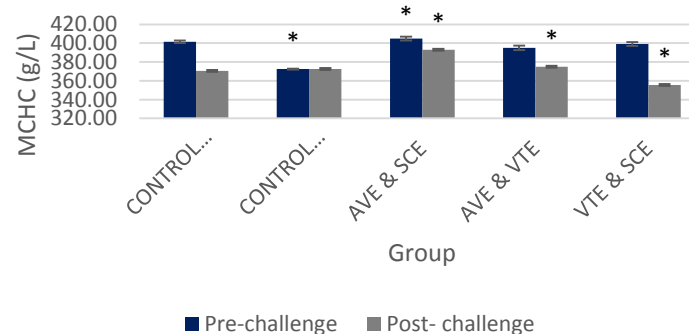
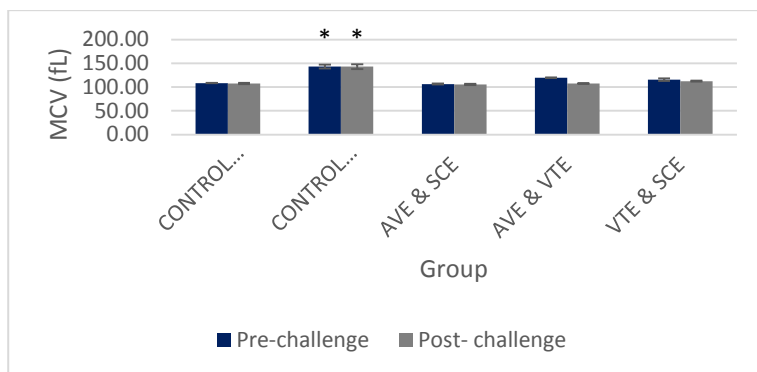


(d) WBC

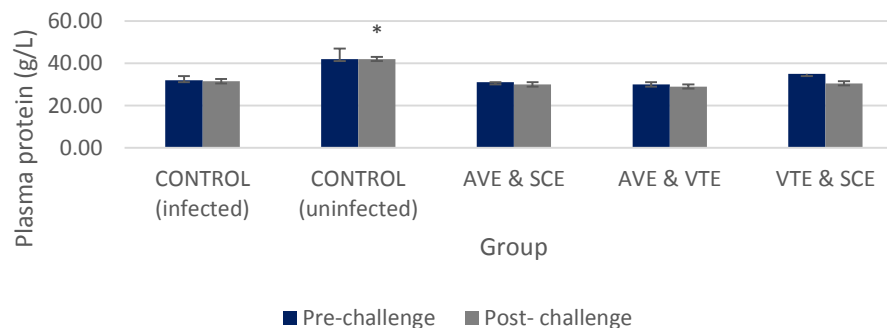


(e) MCV

(f) MCHC



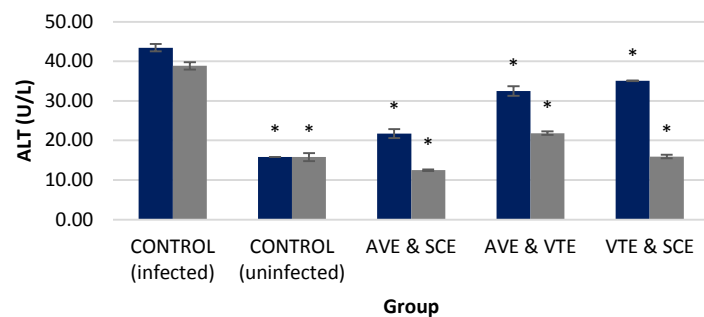
(g) Plasma protein



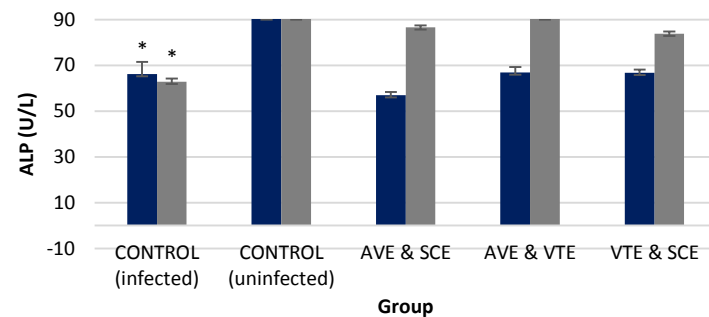
A13 Haematological parameters of *Oreochromis* sp. (pre-challenge) fed &VTE, VTE, SCE, AVE and Control diet at for six-weeks and post-challenge (two-weeks). Data are expressed as the mean $\pm$ SEM. Statistical differences ($P < 0.05$) from the control (infected) group are indicated by asterisks. RBC = red blood cell, Hb = haemoglobin, WBC = white blood cell, PCV = packed cell volume, MCV = mean corpuscular volume, MCHC = mean cellular haemoglobin content, P. protein = plasma protein

(h) ALT

(i) ALP

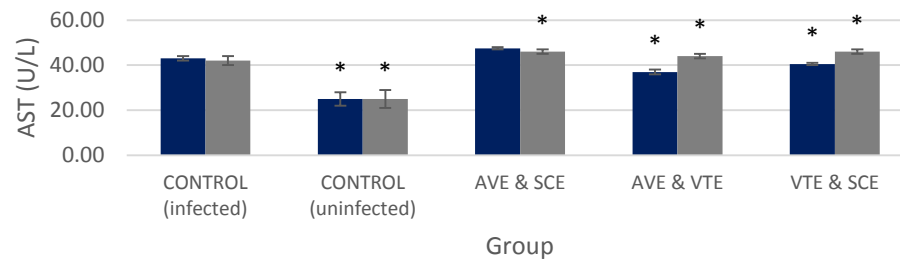


■ Pre-challenge ■ Post-challenge



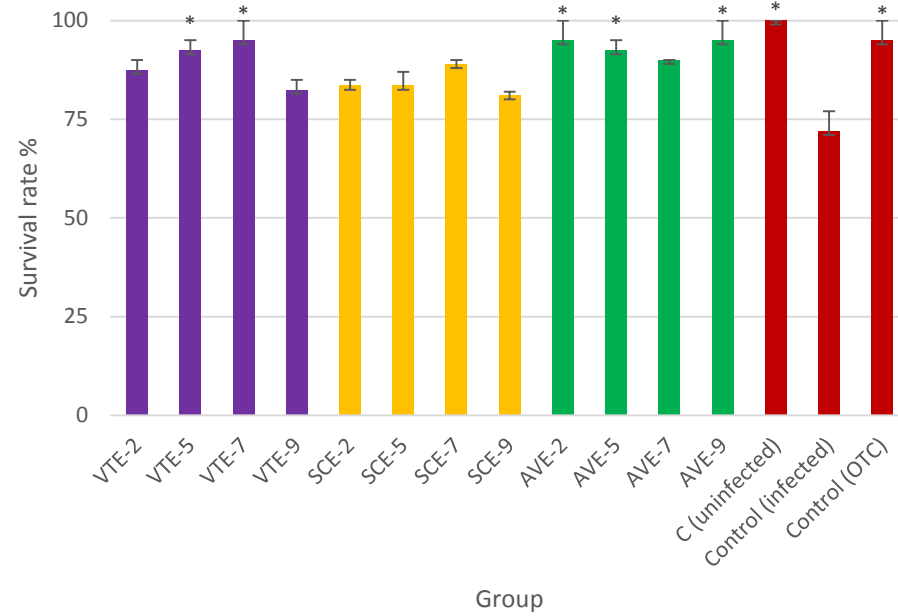
■ Pre-challenge ■ Post-challenge

(j) AST



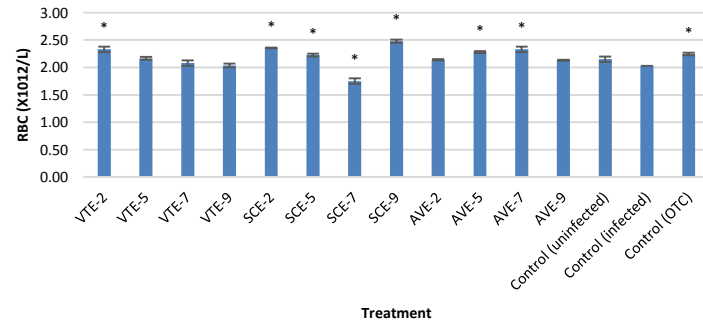
■ Pre-challenge ■ Post-challenge

A14 Biochemical parameters of *Oreochromis* sp. (pre-challenge) fed &VTE, VTE, SCE, AVE and Control diet at for six-weeks and post-challenge (two-weeks). Data are expressed as the mean $\pm$ SEM. Statistical differences ($P<0.05$) from the control (infected) group are indicated by asterisks. ALT = alkaline aminotransferase, ALP = alkaline phosphatase, AST = aspartate aminotransferase

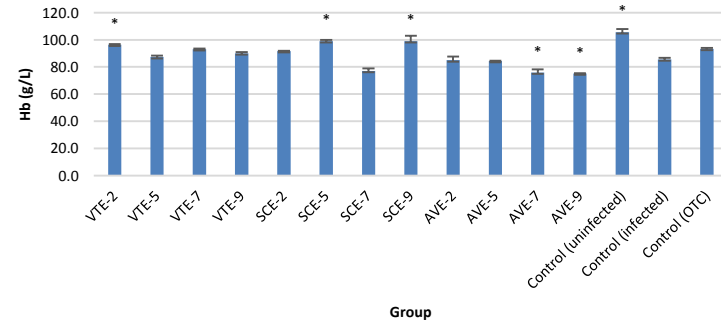


A15 Survival rate of *Oreochromis* sp.

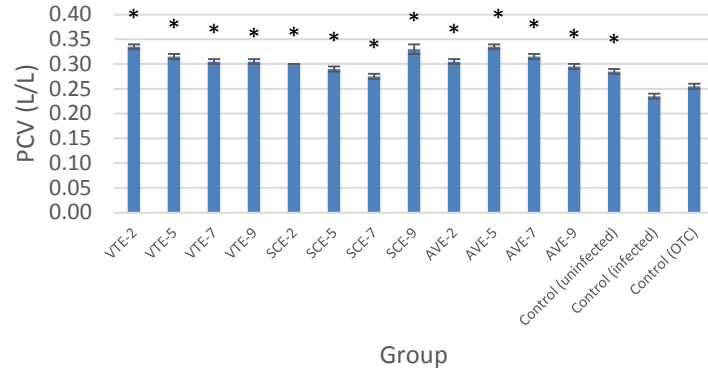
(a) RBC



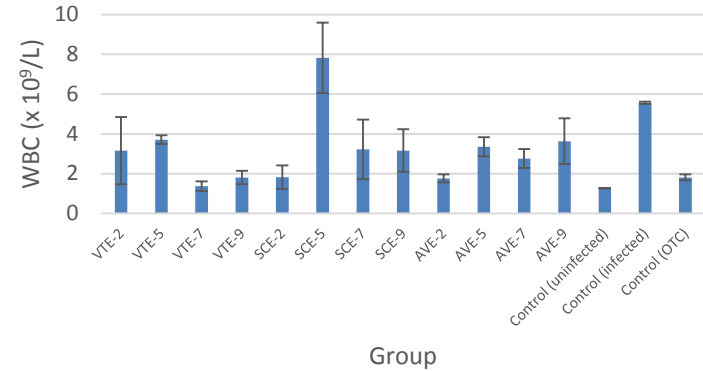
(b) Hb



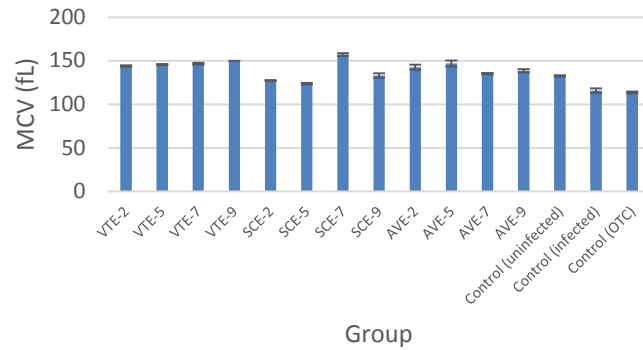
(c) PCV



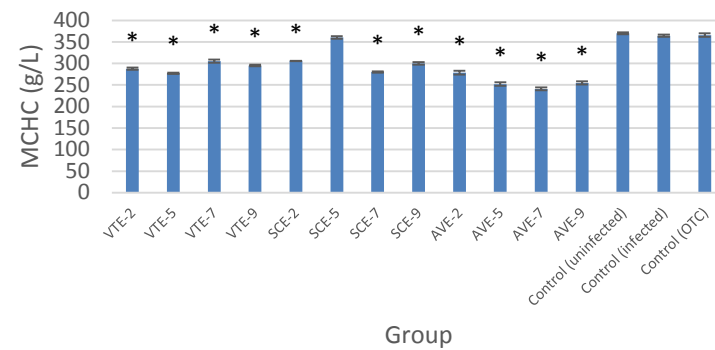
(d) WBC



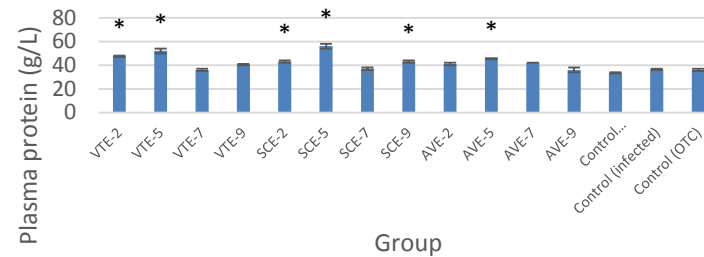
(e) MCV



(f) MCHC

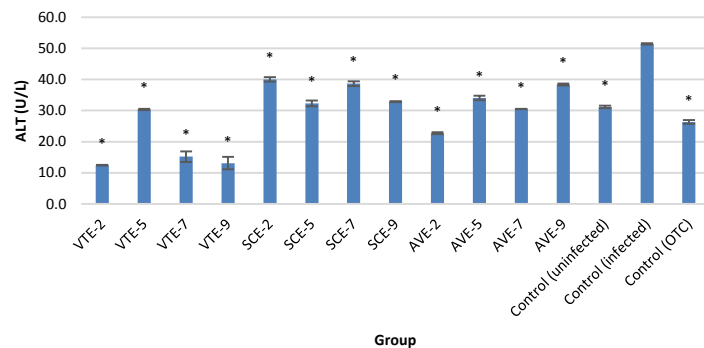


(g) Plasma protein

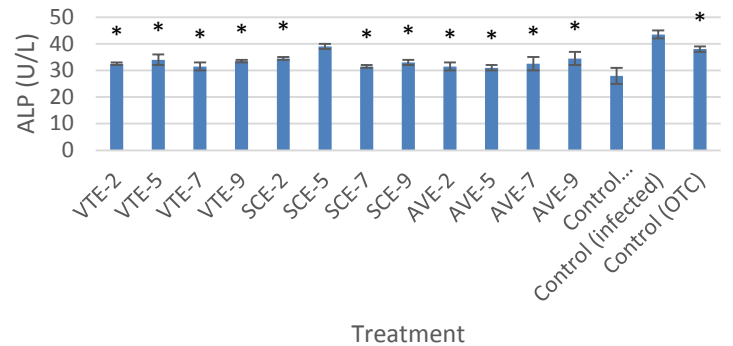


A16 Haematological parameters of *Oreochromis* sp. (post-challenge) fed &VTE, VTE, SCE, AVE and Control diet. Data are expressed as the mean $\pm$ SEM. Statistical differences ($P < 0.05$) from the control (infected) group are indicated by asterisks. RBC = red blood cell, Hb = haemoglobin, WBC = white blood cell, PCV = packed cell volume, MCV = mean corpuscular volume, MCHC = mean cellular haemoglobin content, P. protein = plasma protein

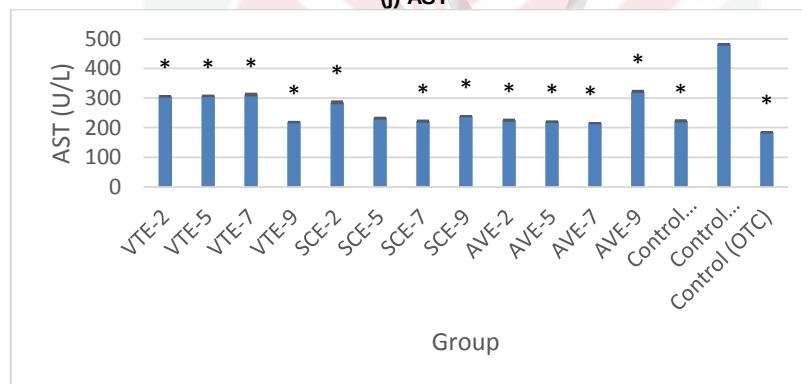
(h) ALT



(i) ALP



(j) AST



A17 Biochemical parameters of *Oreochromis* sp. (post-challenge) fed &VTE, VTE, SCE, AVE and Control diet. Data are expressed as the mean $\pm$ SEM. Statistical differences ($P < 0.05$) from the control (infected) group are indicated by asterisks. ALT = alkaline aminotransferase, ALP = alkaline phosphatase, AST=aspartateaminotransferase

BIODATA OF STUDENT

Sharifah Raina binti Manaf was born on 22 November 1986 in Kelantan, to Mr. Manaf bin Daud and Mrs Sharifah Zainun binti Mohd Salleh. She obtained her lower secondary and higher secondary school certificate from SMK Zainab (1), Kelantan. She furthered her studies and awarded a Matriculation Science in 2005 from Perlis Matriculation College. She continued her degree in Agrotechnology (Aquaculture) in 2008 and her Master Science of Aquaculture in 2010 from Universiti Malaysia Terengganu, Terengganu.

Her career started as a substitute teacher at SK Demit, Kelantan in 2008 for 3 months. After that, she work as research assistant at Institute Tropical Agriculture, UPM, Selangor. Her area of research at that time was fermented of sago. Currently she is under Young Scheme (Tenaga Pengajar Muda), Faculty of Plantation and Agrotechnology, Shah Alam, Selangor.

Sharifah Raina is married with 1 children. Her publications, seminar, workshops and conference attended and research carried out outlined below:

LIST OF PUBLICATIONS

Sharifah Raina, M. Hassan, M.D., Abdul Razak, A., Noordin, M.M., Ruhil Hayati, H., Kumari Geetha, M., Nora Faten Afifah, M., Rashidah, A.R. and Nur Hidayahanum, H. (2016). The effects of *Vitex trifolia*, *Strobilanthes crispus* and *Aloe vera* Herbal-mixed Dietary Supplementation on Growth Performance and Disease Resistance in Red Hybrid Tilapia (*Oreochromis* sp.). *Journal of Aquaculture Research and Development* 7: 1-6.

Sharifah Raina, M., Hassan, M. D., Ruhil Hayati, H., and Nur Hidayahanum, H. Antimicrobial Activity of the Selected Local Plant Extracts against Aquatic Bacteria. 2012 Malaysia-Thailand Graduate Forum in Life Science, Food Science and Agriculture. 12-14 December 2012. Mahidol University, Bangkok, Thailand. Oral presenter.

Sharifah Raina, M., Hassan, M. D., Ruhil Hayati, H., and Nur Hidayahanum, H. *In vitro* studies of selected local plants against freshwater food fish pathogens and toxicity in Tilapia (*Oreochromis* sp.). Postgraduate Colloquium Series 2. November 2013. Bilik Jemaah, Faculty of Veterinary Medicine, UPM. Oral presenter

Ruhil Hayati, H., Hassan, M. D., Ong, B. L., Abdelhadi, Y. M., Nur Hidayahanum, H. and **Sharifah Raina, M.** Characterization, antibiotic sensitivity and plasmid profiles of *Aeromonas hydrophila* gp. 2 isolates in diseased freshwater fish. Postgraduate Colloquium Series 2. 12 December 2013. Bilik Kuliah 12, Faculty of Veterinary Medicine, UPM.

Ruhil Hayati, H., Hassan, M. D., Ong, B. L., Abdelhadi, Y. M., Nur Hidayahanum, H. and **Sharifah Raina, M.** Histopathological study of *Oreochromis* sp. experimentally infected with *Aeromonas hydrophila* and detection of 10 virulence genes by PCR. Postgraduate Colloquium Series 2. 19 December 2013. Bilik Kuliah 12, Faculty of Veterinary Medicine, UPM.

Ruhil Hayati, H., Hassan, M. D., Nur Hidayahanum, H., **Sharifah Raina, M.**, Ong, B.L., Abdelhadi, Y. M. and Maryam, H. Molecular characterization of virulence genes in *Aeromonas hydrophila* originated from diseased freshwater fishes. International Fisheries Symposium (IFS 2013). 28-29 November 2013. Ambassador City Jomtien Hotel, Pattaya, Thailand. Oral presenter.

Sharifah Raina, M., Hassan, M. D., Ruhil Hayati, H., and Nur Hidayahanum, H. Assessment on Antimicrobial Activity of Selected Local Plants Against Freshwater Food Fish Pathogens. Symposium Biologi Kebangsaan 2013. 28-30 May 2013. Thistle Port Dickson Resort. Oral presenter.

Ruhil Hayati, H., Hassan, M. D., Ong, B. L., Abdelhadi, Y. M., Nur Hidayahanum, H. and **Sharifah Raina, M.** Characterization, Pathogenicity and Antibiotic Sensitivity Profiles of *Aeromonas* spp. Isolated from Motile Aeromonads Septicemia (MAS) – Like Disease in Freshwater Fishes.

- Simposium Biologi Kebangsaan 2013. 28-30 May 2013. Thistle Port Dickson Resort.
- Hassan, M.D., **Sharifah Raina, M.**, Abdul Razak, A., Noordin, M., Ruhil Hayati, H., Nur Hidayahanum, H., Nora Faten, A.M., Hazer, M. and Kumari Geetha, M. Effect of Diets Supplemented with Simpleleaf Chastetree (*Vitex trifolia*) Extracts on Growth Performance and Histology of the Red Hybrid Tilapia (*Oreochromis* sp.). International Postgraduate Conference on Biotechnology 2014. August 2014. Global Square Building, Soka University, Japan. Poster presenter.
- Ruhil Hayati, Hamdan., Hassan, M. D., Ong, B. L., Abdelhadi, Y. M., Nur Hidayahanum, H., **Sharifah Raina, M.**, Nora Faten, A.M., Hazer, M., Kuttichantran, S. and Kumari Geetha, M. Tissue Detection of *Aeromonas hydrophila* Type 2 in Experimentally Infected *Oreochromis* sp. using Immunohistochemical Assay. International Postgraduate Conference on Biotechnology 2014. August 2014. Global Square Building, Soka University, Japan.
- Nora Faten Afifah Mohamad., Hassan, M.D., Ruhil Hayati, H., Nur Hidayahanum, H., **Sharifah Raina, M.**, Hazer, M., Kuttichantran, S. and Kumari Geetha, M. Genetic Variation of Cultured and Wild Specimen of *Clarias gariepinus* and *Clarias macrocephalus* Using DNA Microsatellite. International Postgraduate Conference on Biotechnology 2014. August 2014. Global Square Building, Soka University, Japan.
- Ruhil Hayati, Hamdan., Hassan, Mohd. Daud., Nur Hidayahanum, Hamid., **Sharifah Raina, Manaf.**, Nora Faten, Afifah Mohamad., Kuttichantran, Subramaniam and Kumari Geetha, M. Virulence Genes Detection of *Aeromonas hydrophila* and Histopathological Study of Its Experimentally Infection to *Oreochromis* sp. National Postgraduate Seminar 2014. 10 September 2014. Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia.
- Ruhil Hayati, H., Hassan, M.D., Ong, B.I., Abdelhadi, Y.M., Nur Hidayahanum, H., **Sharifah, R.M.**, Nora Faten, A.M., Kuttichantran, S. and Milud, Alsaïd. 2015. Virulence Genes Detection of *Aeromonas hydrophila* Originated from Diseased Freshwater Fishes. *Advances in Environmental Biology*, 9(22): 22-26.



UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT :

ASSESSMENT OF THERAPEUTIC AND ANTIMICROBIAL EFFECTS OF LOCAL PLANTS ON
BACTERIAL DISEASES IN RED HYBRID TILAPIA (*Oreochromis* sp.)

NAME OF STUDENT : SHARIFAH RAINA BINTI MANAF

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

☐

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

☐

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

☐

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

☐

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]