



***METHOD DEVELOPMENT FOR DETERMINATION OF
MULTI-MYCOTOXIN IN VEGETABLE OIL USING QuEChERS
TECHNIQUE AND LIQUID CHROMATOGRAPHY TANDEM
MASS SPECTROMETRY***

SHARMILI D/O KUPPAN

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By

SHARMILI D/O KUPPAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

December 2015

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Dedicated to my beloved parents, husband, sons

and

rest of my family members



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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December 2015

Chairman : Prof. Jinap Selamat, PhD
Faculty : Food Science and Technology

In recent years, vegetable oils were reported to be contaminated by mycotoxins. At the same time, demand for vegetable oils has been increasing due to its health benefit and also for its direct use or as ingredients in food. The objectives of this study were to develop and validate the QuEChERS (Quick, Easy, Cheap, Effective, Rugged and Safe) extraction and clean-up procedures for mycotoxins in vegetable oil by liquid chromatography tandem mass spectrometry (LC-MS/MS) and to verify the established method in vegetable oils in Malaysian market. In this study, a new, fast, cost efficient, sensitive and accurate assay method was developed, optimized and validated for the determination of aflatoxins (AFB₁, AFB₂, AFG₁ and AFG₂), ochratoxin A (OTA), zearalenone (ZEA) and deoxynivalenol (DON) in vegetable oil by a modified QuEChERS approach, which was conventionally developed for pesticides analysis. A quick procedure of mycotoxins extraction and clean-up was achieved in two steps. Firstly, target analytes were extracted into the optimized solvent acetonitrile and followed by dispersive solid phase extraction (d-SPE) clean up with the optimized sorbents to remove the co-extracts. Three types of d-SPE sorbents; C18, primary secondary amine (PSA) and graphitized carbon black (GCB) and four combination ratios of two selected d-SPEs were evaluated to achieve an optimum and acceptable result. The maximum co-extracts were removed when d-SPE C18 and GCB were used at ratio 3:1 with MgSO₄ anhydrous followed by LC-MS/MS analysis, using an electro spray-ionization interface (ESI). Methanol as an organic phase and formic acid (0.1%) with 5 mM ammonium acetate as a polar phase, with total run of 16 minutes and flow rate 300 μLmin^{-1} were applied. The validation study indicated the seven mycotoxins could be detected in the concentration range of 0.01 ngg^{-1} to 650.17 ngg^{-1} with correlation coefficient > 0.98. The limit of quantitation (LOQ) ranges from 0.04 ngg^{-1} to 2000 ngg^{-1} . The recoveries were in the range of 87.9% to 106.6%. The repeatability and reproducibility of the analysis were within the acceptable level (0.5 – 1.3) for precision using HORRAT ratio. Significant matrix effect was compensated using

the matrix matched calibration curves. After validation, the method was successfully applied to 25 vegetable oil samples including corn oil, palm oil and sunflower oil. ZEA, AFG₂, AFG₁ and AFB₁ were among the detected mycotoxins. A new fast, cost efficient, sensitive and accurate assay method was successfully developed, validated and applied for the determination of seven mycotoxins in vegetable oils.



Abstark tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PEMBANGUNAN KAEDAH BAGI PENENTUAN MULTI-MIKOTOKSIN
DALAM MINYAK SAYURAN MENGGUNAKAN TEKNIK QuEChERS
DAN KROMATOGRAFI CECAIR SPEKTROMETER JISIM (LC-MS/MS)**

Oleh

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Sejak kebelakangan ini, minyak sayuran dilaporkan dicemari oleh mikotoksin. Pada masa yang sama, permintaan bagi minyak sayuran telah meningkat disebabkan oleh manfaat kesihatan dan sebagai bahan dalam makanan. Objektif kajian ini adalah untuk membangunkan dan mengesahkan kaedah pengekstrakan teknik QuEChERS (*Quick, Easy, Cheap, Effective, Rugged and Safe*) dan prosedur pembersihan sampel untuk mengekstrak mikotoksin dari minyak sayuran dan menganalisis menggunakan kromatografi cecair spektrometri jisim (LC-MS/MS) serta untuk mengesahkan kaedah yang dikembangkan melalui penentuan pencemaran mikotoksin dalam minyak sayuran di pasaran. Dalam kajian ini, satu kaedah yang baru, cepat, cekap, sensitif, murah dan tepat telah dibangunkan, dioptimumkan dan disahkan bagi menentukan aflatoksin (AFB₁, AFB₂, AFG₁, AFG₂), ochratoxin A (OTA), zearalenone (ZEA) dan deoxynivalenol (DON) dalam minyak sayur-sayuran oleh pendekatan teknik QuEChERS yang diubahsuai dari analisis racun perosak. Prosedur yang cepat bagi pengekstrakan mikotoksin dan pembersihan sampel telah dicapai dalam dua langkah. Pertama, analit sasaran diekstrak ke dalam pelarut asetonitril dan diikuti oleh pengekstrakan dari serakan fasa pepejal (d-SPE) untuk mendapatkan nilai kejutuan mikotoksin dan nilai sisa ekstrak yang optimum. Tiga jenis penjerap d-SPE; C18, *primary secondary amine* (PSA) dan *graphitized carbon black* (GCB) dan empat nisbah gabungan dua penjerap telah dinilai untuk mencapai hasil yang optimum dan boleh diterima. Keputusan yang maksimum iaitu nilai kejutuan bagi mikotoksin yang tinggi serta nilai sisa ekstrak yang minimum dicapai apabila penjerap C18 dan GCB telah digunakan pada nisbah 3: 1 dengan MgSO₄ kontang diikuti oleh analisis LC-MS/MS, menggunakan antara muka elektro semburan-pengionan (ESI). Metanol sebagai fasa organik dan asid formik (0.1%) dengan 5 mM ammonium asetat sebagai fasa cecair, dengan jumlah jangka masa 16 minit dan kadar aliran 300 μLmin^{-1} telah digunakan. Kajian pengesahan menunjukkan tujuh mikotoksin dapat dikesan dalam julat kepekatan terendah (*LOD*) antara 0.01 ngg^{-1} hingga 650.17 ngg^{-1} dengan pekali korelasi > 0.98. Had kuantifikasi (*LOQ*) adalah di antara 0.04

ngg⁻¹ hingga 2000 ngg⁻¹. Nilai kejituan adalah dalam lingkungan 87.9% kepada 106.6%. Kebolehulangan analisis adalah dalam tahap yang boleh diterima iaitu di antara 0.5-1.3 untuk ketepatan menggunakan nisbah *HORRAT*. Kesan matriks yang ketara telah diatasi menggunakan matriks sepadan dengan keluk penentukuran. Selepas pengesahan, kaedah yang telah berjaya digunakan untuk menganalisa 25 sampel minyak sayur-sayuran termasuk minyak jagung, minyak sawit dan minyak bunga matahari. ZEA, AFG₂, AFG₁ dan AFB₁ adalah antara mikotoksin yang dikesan. Satu kaedah baru yang cepat, efektif kos, sensitif dan tepat telah dibangunkan, divalidasi dan digunakan dengan jayanya untuk menentukan kandungan tujuh mikotoksin dalam minyak sayuran.



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I certify that a Thesis Examination Committee has met on 8 December 2015 to conduct the final examination of Sharmili a/p Kuppan on her thesis entitled "Method Development for Determination of Multi-Mycotoxin in Vegetable Oil Using QuEChERS Technique and Liquid Chromatography Tandem Mass Spectrometry" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

AA	Acetic acid
ADI	Acceptable daily intake
AFB ₁	Aflatoxin B ₁
AFB ₂	Aflatoxin B ₂
AFG ₁	Aflatoxin G ₁
AFG ₂	Aflatoxin G ₂
AFs	Aflatoxins
AOAC	Association Officials of Analytical Chemistry
ANOVA	Analysis of variance
APCI	Atmospheric pressure chemical ionization
APPI	Atmospheric pressure photo ionization
CIT	Citrinin
CV	Coefficient of variation
C18	Octadecyl
DON	Deoxynivalenol
DSPE	Dispersive solid phase extraction
EC	European Commission
EFSA	European Food Safety Authorities
ESI	Electrospray ionization
EU	European Union
FA	Formic acid
FAO	Food and Agriculture Organization
FB	Fumonisin
FDA	Food and Drug Administration
FSA	Food Standard Agency
GC	Gas chromatography
HPLC	High performance liquid chromatography
IAC	Immunoaffinity column
IARC	International Agency for Research on Cancer

ISO	International Organization for Standardisation
IUPAC	International Union of Pure and Applied Chemistry
LC	Liquid chromatography
LC-MS/MS	Liquid chromatography tandem mass
LLE	Liquid-liquid extraction
LOD	Limit of detection
LOQ	Limit of quantitation
MeCN	Acetonitrile
MeOH	Methanol
MgSO ₄	Magnesium sulphate
min	Minute
MPOB	Malaysian Palm Oil Board
MRL	Maximum residue limit
MS	Mass spectrometry
MSMS	Tandem mass spectrometry
MSPD	Matrix solid phase dispersion
MUFA	Monounsaturated fatty acid
OLS	Ordinary least square
OTB	Ochratoxin B
pH	Negative logarithmic value of the hydrogen ion (H ⁺)
PKO	Palm kernel oil
PO	Palm olein
PSA	Primary secondary amine
PUFA	Polyunsaturated fatty acid
QuEChERS	Quick, easy, cheap, effective, rugged, safe
RSD	Relative standard deviation
R ²	Correlation
S/N	Signal/noise
SD	Standard deviation
SIM	Selection ion monitoring
SFA	Saturated fatty acid

SPE	Solid phase extraction
SRM	Selected reaction monitoring
TIC	Total ion chromatography
UPLC	Ultra performance liquid chromatography
ZEA	Zearalenone
et.al.	and other authors
g	Gram
gmol ⁻¹	Gram per mole
kg	Kilogram
L	Liter
mg	Miligram
mgL ⁻¹	Miligram per liter
mL	Mililiter
mLmin ⁻¹	Mililiter per minute
<i>m/z</i>	Mass per electrical charge ratio
ngg ⁻¹	Nanogram per gram
μL	Microliter
μgL ⁻¹	Microgram per liter
%	Percent
Σ	Sum
C	Celcius

CHAPTER 1

INTRODUCTION

1.1 Background of study

Mycotoxins are toxic chemical products produced as secondary metabolites by certain filamentous fungal that can contaminate many agricultural commodities and processed food, either in the field or during storage wherever humidity and temperature are adequate especially in tropic areas (Cavaliere et al., 2010). These mycotoxins have wide range of toxic effects due to their largely diversified chemistry and toxicology properties (Ibanez-vea et al., 2011). Among hundreds of mycotoxins, aflatoxins (AFs), ochratoxin A (OTA), zearalenone (ZEA), deoxynivalenol (DON), fumonisins (FBs), T-2 and HT-2 toxins are the major health concern (Soleimany et al., 2012).

Human exposure to mycotoxins is inevitable and despite many years of study, applying good manufacturing practices during food production, storage and distribution, mycotoxins continue to contaminate a wide range of food and feed. Commodities and products frequently contaminated with mycotoxins include wide range of vegetable commodities such as cereals like wheat, maize, rye, rice, oat, and barley, nuts, peanuts, fruits, oilseeds, dried fruits, cocoa, spices, beer, herbal teas and coffee (Anukul & Vangnai, 2013). Recently, occurrence of some mycotoxins in vegetable oil has been reported worldwide and has received a great deal of attention by the regulatory bodies (Marin et al., 2013).

1.2 Statement of problems

Corn germ and sunflower seeds are very vulnerable to mycotoxins contamination and studies have showed number of occurrence of mycotoxins in corn and sunflower oils (Qian et al., 2015; Schollenberger et al., 2008). However, there is no data been collected to identify the level of mycotoxins contamination in palm oil produced by palm fruit which is at high risk of mycotoxins contamination due to its cultivation in tropical country such as Indonesia and Malaysia. This is important since Malaysia is known as the second largest producer of palm oil in the world after Indonesia.

With increasing findings on mycotoxins contamination in vegetable oil and its health hazard to human, total AFs in oilseeds before human consumption and intended for direct consumption have been regulated. In addition, ZEA level also been regulated particularly in refined maize oil by European Commission (EU) Regulatory body (European Commission Regulation, 2007) due to its highest occurrence in this commodity but in Malaysia, regulation yet been implemented under Food Act 1983 and Food Regulation 1985 for mycotoxins in vegetable oil. It is important to assess the presence and amount of mycotoxins to ensure safety and quality of vegetable oil produced and consumed in Malaysia thus reliable, fast and cost effective analytical method is important for an effective monitoring.

Numerous methods have been developed for the determination of single or multi-mycotoxin in vegetable oil employing different type of extraction and quantification techniques such as liquid-liquid extraction (Elzupir et al., 2010), immunoaffinity column (IAC) (Bao et al., 2010), matrix solid phase dispersion (MSPD) (Cavaliere et al., 2007), dispersive liquid-liquid microextraction (DLLME) (Afzali et al., 2012), acid base extraction (Majerus et al., 2009) and gel permeation chromatography (GPC) (Qian et al., 2015). Most of the researchers quantified using chromatographic methods including high performance liquid chromatography (HPLC) coupled with fluorescence detectors (FLD) (Majerus et al., 2009) or mass spectrometry (MS) (Cavaliere et al., 2007), and gas chromatography (GC) coupled with MS detectors (Qian et al., 2015). Though the methods are reliable and reproducible, the procedure are limited to single mycotoxin analysis or time consuming, require large volume of solvents and costly. In addition, oil which is a lipid poses a challenging task in extraction of mycotoxins which have a wide range of different physicochemical properties in term of pH, stability, solubility, diversity of chemical structure and molecular weight.

1.3 Importance of study

Recently many methods have been developed using QuEChERS (Quick, Easy, Cheap, Effective, Rugged and Safe) technique in mycotoxin analysis in matrixes like cereal, spices, egg, wine, dairy product, beer and rice (Jia et al., 2014), yet there is no method been developed to analyse mycotoxins in vegetable oil. It is known for its simplicity, quickness in extraction and cheaper compared to solid phase extraction (SPE) and immunoaffinity column (IAC) and does not require supporting equipment such as vacuum manifold. For the very first time, QuEChERS technique will be employed to extract multi-mycotoxin from vegetable oil and this will open opportunity to conduct preliminary study on the mycotoxins in vegetable oil; particularly in corn oil, sunflower oil and palm olein which are highly consumed by Malaysian and also in other countries. Since the H [W U D F W L R Q Z L O O E H H P S O R \ L Q J Q H Z 4 X (&) optimizing various types of dispersive solid phase extraction (d-SPE) sorbents and extraction solvents, the optimized method will be tested for its performance by conducting detailed method validation including comparison with IAC extraction technique.

This study involves three major components namely method development for seven mycotoxins, followed by method validation of the developed method of the specific compounds and next, determination of multi-mycotoxin in three vegetable oils.

1.4 Objectives

The objectives of this study are as follows:

- i. To develop and validate a reliable, fast and cost effective QuEChERS based extraction and clean-up procedures for multi-mycotoxin in vegetable oil
- ii. To determine the contamination of multi-mycotoxin in three different vegetable oils using the developed method

REF (RENCES

- Abdulkadir, M., & Abubakar, G. I. (2011). Production and refining of corn oil from hominy feed: A by-product of dehulling operation. *Journal of Engineering and Applied Sciences*, 6, 22-8.
- Afzali, D., Ghanbarian, M., Mostafavi, A., Shamspur, T., & Ghaseminezhad, S. (2012). A novel method for high pre-concentration of ultra-trace amounts of B1, B2, G1 and G2 aflatoxins in edible oils by dispersive liquid-liquid microextraction after immunoaffinity column clean-up. *Journal of Chromatography A*, 1247, 35-41.
- Alonso, J. S. J., Sastre, J. L., Romero-Avila, C., & Lopez, E. (2008). A note on the combustion of blends of diesel and soya, sunflower and rapeseed vegetable oils in a light boiler. *Journal of Biomass and Bioenergy*, 32(9), 880-886.
- Anastassiades, M., Lehotay, S. J., Stajnbaher, D., & Schenck, F. J. (2003). Fast and easy multiresidue method employing acetonitrile extraction/partitioning and 3 G L V S H U V S K B V H R C L W U L E B E W I M Q I O R pesticide residues in produce. *Journal of AOAC International*, 86(2), 412-431.
- Anukul, N., & Vangnai, K. (2013). Significance of regulation limits in mycotoxin contamination in Asia and risk management programs at the national level. *Journal of Food and Drug Analysis*, 21(3), 227-241.
- Arroyo-Manzanares, N., Garcia-Campana, A. M., & Gamiz-Gracia, L. (2013). Multiclass mycotoxin analysis in *Silybum marianum* by ultra high performance liquid chromatography tandem mass spectrometry using a procedure based on QuEChERS and dispersive liquid liquid microextraction. *Journal of Chromatography A*, 1282, 11-19.
- Ashiq, S., Hussain, M., & Ahmad, B. (2014). Natural occurrence of mycotoxins in medicinal plants: a review. *Journal of Fungal Genetics and Biology*, 66, 1-10.
- Association of Official Analytical Chemist (AOAC), 1989. Guidelines for collaborative study procedure to validate characteristics of a method of analysis, 72. 694-704.
- Bao, L., Trucksess, M. W., & White, K. D. (2010). Determination of aflatoxins B1, B2, G1, and G2 in olive oil, peanut oil, and sesame oil. *Journal of AOAC International*, 93(3), 936-942.
- Berthiller, F., Schuhmacher, R., Buttner, G., & Krska, R. (2005). Rapid simultaneous determination of major type A-and B-trichothecenes as well as zearalenone in maize by high performance liquid chromatography tandem mass spectrometry. *Journal of Chromatography A*, 1062(2), 209-216.
- Berthiller, F., Sulyok, M., Krska, R., & Schuhmacher, R. (2007). Chromatographic methods for the simultaneous determination of mycotoxins and their conjugates in cereals. *International Journal of Food Microbiology*, 119, 33-37.

- Biselli, S., Hartig, L., Wegner, H., & Hummert, C. (2005). Analysis of Fusarium toxins using LC-MS-MS. Application to various food and feed matrices. *Journal of Spectroscopy* 20 (2), 404-416.
- Busby, W. F., & Wogan, G. N. (1984). Aflatoxins. *Journal of Chemical Carcinogens*, 2, 945-1136.
- Cai, M., Chen, X., Wei, X., Pan, S., Zhao, Y., & Jin, M. (2014). Dispersive solid-phase extraction followed by high-performance liquid chromatography/tandem mass spectrometry for the determination of ricinine in cooking oil. *Journal of Food Chemistry*, 158, 459-465.
- Cavaliere, C., Foglia, P., Guarino, C., Nazzari, M., Samperi, R., & Lagana, A. (2007). Determination of aflatoxins in olive oil by liquid chromatography tandem mass spectrometry. *Journal of Analytica Chimica Acta*, 596(1), 141-148.
- Capriotti, A. L., Cavaliere, C., Giansanti, P., Gubbiotti, R., Samperi, R., & Lagana, A. (2010). Recent developments in matrix solid-phase dispersion extraction. *Journal of Chromatography A*, 1217(16), 2521-2532.
- Cavaliere, C., Foglia, P., Samperi, R., & Lagana, A. (2010). Determination of aflatoxins and ochratoxin A in olive oil. *Olives and olive oil in health and disease prevention (Vol. 1)*. Rome, Italy. Elsevier Inc. 645-652.
- Capriotti, A. L., Caruso, G., Cavaliere, C., Foglia, P., Samperi, R., & Lagana, A. (2012). Multiclass mycotoxin analysis in food, environmental and biological matrices with chromatography/mass spectrometry. Rome, Italy. *Mass spectrometry reviews*, 31(4), 466-503.
- Chen, P. H., Chen, P. S., & Huang, S. D. (2013). Determination of pesticides in water and vegetable matter by manual shaking-enhanced, ultrasound-assisted emulsification microextraction combined with gas chromatography-mass spectrometry. Taipei, Taiwan. *Herbicides-Advances in Research*, 85.
- Codex. 2006. Codex committee on methods of analysis and sampling. Rome : Codex Alimentarius Commission.
- Cunha, S. C., Lehotay, S. J., Mastovska, K., Fernandes, J. O., Beatriz, M., & Oliveira, P. P. (2007). Evaluation of the QuEChERS sample preparation approach for the analysis of pesticide residues in olives. *Journal of Separation Science*, 30(4), 620-632.
- Ullah, R. (2008). Simultaneous detection of type A and type B trichothecenes in cereals by liquid chromatography electrospray ionization mass spectrometry using NaCl as cationization agent. *Journal of Chromatography A*, 1054(1), 389-395.
- Deme, P., Azmeera, T., Prabhavathi Devi, B. L. a, Jonnalagadda, P. R., Prasad, R. B. N., & Vijaya Sarathi, U. V. R. (2014). An improved dispersive solid-phase extraction clean-up method for the gas chromatography-negative chemical ionisation tandem mass spectrometric determination of multiclass pesticide residues in edible oils. *Journal of Food Chemistry*, 142, 144-151.

- Di Stefano, V., Avellone, G., Bongiorno, D., Cunsolo, V., Muccilli, V., Sforza, S., & Vékey, K. (2012). Applications of liquid chromatography mass spectrometry for food analysis. *Journal of Chromatography A*, 1259, 74-85.
- Dubernet, M. (2005). Practical guideline for the validation, quality control, and uncertainty assessment of an alternative oenological analysis method, 1-92.
- Elzupir, A. O., Suliman, M. A., Ibrahim, I. A., Fadul, M. H., & Elhoussein, A. M. (2010). Aflatoxins levels in vegetable oils in Khartoum State, Sudan. *Society for Mycotoxin Research and Springer*, 26(2), 69-73.
- Escobar, J., Loran, S., Gimenez, I., Ferruz, E., Herrera, M., Herrera, A., & Arino, A. (2013). Occurrence and exposure assessment of *Fusarium* mycotoxins in maize germ, refined corn oil and margarine. *Journal of Food and Chemical Toxicology*, 62, 514-520.
- Eurachem. (1998). The fitness for purpose of analytical methods: A laboratory guideline to method validation and related topics. Laboratory of the Government Chemist.
- European Commission Regulation, (2002). EC 657/2002 of 12 August 2002 implementing council directive 96/23/EC concerning the performance of analytical methods and the interpretation of results. *Official Journal of The European Union*, L221/8, 1-29.
- European commission Regulation, (2007). EC 1126/2007 of 28 September 2007 amending Regulation (EC) No 1881/2006 setting maximum levels for certain contaminants in foodstuffs as regards fusarium toxins in maize and maize products. *Official Journal of The European Union*, L255/14, 1-4.
- Fernandes, P. J., Barros, N., & Camara, J. S. (2013). A survey of the occurrence of ochratoxin A in Madeira wines based on a modified QuEChERS extraction procedure combined with liquid chromatography-triple quadrupole tandem mass spectrometry. *Journal of Food Research International*, 54(1), 293-301.
- Ferreira, I., Fernandes, J. O., & Cunha, S. C. (2012). Optimization and validation of a method based in a QuEChERS procedure and gas chromatography mass spectrometry for the determination of multi-mycotoxins in popcorn. *Journal of Food Control*, 27(1), 188-193.
- Finnigan TSQ, Quantum Discovery Hardware Manual, 2003. Published by Technical Publications, Thermo Electron Corporation, San Jose, California, USA. pg 2-15.
- Frenich, A. G., Romero-gonzalez, R., Gomez-perez, M. L., Luis, J., & Vidal, M. (2011). Multi-mycotoxin analysis in eggs using a QuEChERS-based extraction procedure and ultra-high-pressure liquid chromatography coupled to triple quadrupole mass spectrometry. *Journal of Chromatography A*, 1218(28), 4349-4356.

- Fumonisin, B. (2002). Some traditional herbal medicines, some mycotoxins, naphthalene and styrene. Institute of research on Cancer IARC monographs on the evaluation of carcinogenic risks to humans, 82.
- Gilbert-Lopez, B., Garcia-Reyes, J. F., Fernandez-Alba, A. R., & Molina-Diaz, A. (2010). Evaluation of two sample treatment methodologies for large-scale pesticide residue analysis in olive oil by fast liquid chromatography ± electrospray mass spectrometry. *Journal of Chromatography A*, 1217(24), 3736-3747.
- Hennion, M. C. (2000). Graphitized carbons for solid-phase extraction. *Journal of Chromatography A*, 885(1), 73-95.
- Horwitz, W. (2002). Associations of Analytical Communities (AOAC) guidelines for single laboratory validation of chemical methods for dietary supplements and botanicals. AOAC International: Gaithersburg, MD, USA, 12-19.
- Ibanez-vea, M., Ana, L., Remiro, R., Murillo-arbizu, M. T., Gonzalez-penas, E., & Lizarraga, E. (2011). Validation of a UHPLC-FLD method for the simultaneous quantification of aflatoxins , ochratoxin A and zearalenone in barley. *Journal of Food Chemistry*, 127, 351 -358.
- Jaimez, J., Fente, C. A., Vazquez, B. I., Franco, C. M., Cepeda, A., Mahuzier, G., & Prognon, P. (2000). Application of the assay of aflatoxins by liquid chromatography with fluorescence detection in food analysis. *Journal of Chromatography A*, 882(1), 1-10.
- Jia, W., Chu, X., Ling, Y., Huang, J., & Chang, J. (2014). Multi-mycotoxin analysis in dairy products by liquid chromatography coupled to quadrupole orbitrap mass spectrometry. *Journal of Chromatography A*, 1345, 107 -114.
- Katsri, K., Noitup, P., Junsangsree, P., & Singanusong, R. (2014). Physical, chemical and microbiological properties of mixed hydrogenated palm kernel oil and cold-pressed rice bran oil as ingredients in non-dairy creamer. *Songklanakarin Journal of Science & Technology*, 36(1) 73 -81.
- Kmellar, B., Fodor, P., Pareja, L., Ferrer, C., Martinez-Uroz, M. A., Valverde, A., & Fernandez-Alba, A. R. (2008). Validation and uncertainty study of a comprehensive list of 160 pesticide residues in multi-class vegetables by liquid chromatography tandem mass spectrometry. *Journal of Chromatography A*, 1215(1), 37-50.
- Koesukwiwat, U., Sanguankaew, K., & Leepipatpiboon, N. (2014). Evaluation of a modified QuEChERS method for analysis of mycotoxins in rice. *Journal of Food Chemistry*, 153, 44 -51.
- Kokkonen, M. (2011). The challenge of LC/MS/MS multi-mycotoxin analysis-Heracles battling the Hydra.
- Krogh, P., & Nesheim, S. (1982). Ochratoxin A. International Agency for Research on Cancer (I A R C) Scientific Publications, 1982.

- Lattanzio, V. M. T., Ciasca, B., Powers, S., & Visconti, A. (2014). Improved method for the simultaneous determination of aflatoxins, ochratoxin A and fusarium toxins in cereals and derived products by liquid chromatography tandem mass spectrometry after multi-toxin immunoaffinity clean up. *Journal of Chromatography A*, 1354, 139-143.
- Lam, M. K., Tan, K. T., Lee, K. T., & Mohamed, A. R. (2009). Malaysian palm oil: Surviving the food versus fuel dispute for a sustainable future. *Journal of Renewable and Sustainable Energy Reviews*, 13(6), 1456-1464.
- Lei, Y. P., Zhao, L. H., Ma, Q. G., Zhang, J. Y., Zhou, T., Gao, C. Q., & Ji, C. (2014). Degradation of zearalenone in swine feed and feed ingredients by bacillus subtilis. *World Mycotoxin Journal*, 7(2), 143-151.
- Li, R., Wang, X., Zhou, T., Yang, D., Wang, Q., & Zhou, Y. (2014). Occurrence of four mycotoxins in cereal and oil products in Yangtze Delta region of China and their food safety risks. *Journal of Food Control*, 35(1), 117-122.
- Li, Y., Fabiano-Tixier, A. S., Ruiz, K., Castera, A. R., Bauduin, P., Diat, O., & Chemat, F. (2015). Comprehension of direct extraction of hydrophilic antioxidants using vegetable oils by polar paradox theory and small angle X-ray scattering analysis. *Journal of Food chemistry*, 173, 873-880.
- Liu, S., Qiu, F., Kong, W., Wei, J., Xiao, X., & Yang, M. (2013). Development and validation of an accurate and rapid LC-ESI-MS/MS method for the simultaneous quantification of aflatoxin B 1, B 2, G 1 and G 2 in lotus seeds. *Journal of Food Control*, 29(1), 156-161.
- Liu, Y., Han, S., Lu, M., Wang, P., Han, J., & Wang, J. (2014). Modified QuEChERS method combined with ultra-high performance liquid chromatography tandem mass spectrometry for the simultaneous determination of 26 mycotoxins in sesame butter. *Journal of Chromatography B*, 970, 68-76.
- Mahoney, N., & Molyneux, R. J. (2010). Rapid analytical method for the determination of aflatoxins in plant-derived dietary supplement and cosmetic oils. *Journal of Agricultural and Food Chemistry*, 58(7), 4065-4070.
- Majerus, P., Graf, N., & Kramer, M. (2009). Rapid determination of zearalenone in edible oils by HPLC with fluorescence detection. *Journal of Mycotoxin research*, 25(3), 117-121.
- Malaysian Palm Oil Board, M. P. O. B (2012). Overview of the Malaysian oil palm industry 2011.
- Malaysian Palm Oil Board, M. P. O. B (2013). Malaysian oil palm statistics, 2012. Malaysian Palm Oil Board, Ministry of Plantation Industries and Commodities, Malaysia, 1176-1177.
- Malik, A. K., Blasco, C., & Pico, Y. (2010). Liquid chromatography mass spectrometry in food safety. *Journal of Chromatography A*, 1217(25), 4018-4040.

- Marin, S., Ramos, A. J., Cano-Sancho, G., & Sanchis, V. (2013). Mycotoxins: occurrence, toxicology, and exposure assessment. *Journal of Food and Chemical Toxicology*, 60, 218-237.
- Matthaus, B. (2012). Oil technology. In *Technological Innovations in Major World Oil Crops*, Volume 2 (pp. 23-92). Springer, New York.
- Miller, J. N., & Miller, J. C. (2000). *Miller and J. Miller, Statistics and Chemometrics for Analytical Chemistry*.
- Moreau, R. A. (2005). Corn oil. *Bailey's industrial oil and fat products*.
- Nimet, G., Da Silva, E. A., Palú, F., Dariva, C., dos Santos Freitas, L., Neto, A. M., & Cardozo Filho, L. (2011). Extraction of sunflower (*Heliantus annuus* L.) oil with supercritical CO₂ and subcritical propane: experimental and modeling. *Journal of Chemical Engineering*, 168(1), 262-268.
- Paiga, P., Morais, S., Oliva-Teles, T., Correia, M., Delerue-Matos, C., Duarte, S. C., & Lino, C. M. (2012). Extraction of ochratoxin A in bread samples by the QuEChERS methodology. *Journal of Food Chemistry*, 135(4), 2522-2528.
- Placinta, C. M., D'mello, J. P. F., & Macdonald, A. M. C. (1999). A review of worldwide contamination of cereal grains and animal feed with fusarium mycotoxins. *Journal of Animal Feed Science and Technology*, 78(1), 21-37.
- Pizzutti, I. R., de Kok, A., Scholten, J., Righi, L. W., Cardoso, C. D., Rohers, G. N., & da Silva, R. C. (2014). Development, optimization and validation of a multimethod for the determination of 36 mycotoxins in wines by liquid chromatography tandem mass spectrometry. *Journal of Talanta*, 129, 352-363.
- Qian, M., Zhang, H., Wu, L., Jin, N., Wang, J., & Jiang, K. (2015). Simultaneous determination of zearalenone and its derivatives in edible vegetable oil by gel permeation chromatography and gas chromatography-triple quadrupole mass spectrometry. *Journal of Food Chemistry*, 166, 23-28.
- Rashid, U., Anwar, F., Moser, B. R., & Ashraf, S. (2008). Production of sunflower oil methyl esters by optimized alkali-catalyzed methanolysis. *Journal of Biomass and Bioenergy*, 32(12), 1202-1205.
- Ren, Y., Zhang, Y., Shao, S., Cai, Z., Feng, L., Pan, H., & Wang, Z. (2007). Simultaneous determination of multi-component mycotoxin contaminants in foods and feeds by ultra-performance liquid chromatography tandem mass spectrometry. *Journal of Chromatography A*, 1143(1), 48-64.
- Rubert, J., Dzuman, Z., Vaclavikova, M., Zachariasova, M., Soler, C., & Hajslova, J. (2012). Analysis of mycotoxins in barley using ultra high liquid chromatography high resolution mass spectrometry: Comparison of efficiency and efficacy of different extraction procedures. *Journal of Talanta*, 99, 712-719.
- Runger, G. C., & Montgomery, D. C. (2002). *Applied statistics and probability for engineers*. John Wiley & sons. Inc. New York, USA.

- Sanagi, M. M., Ling, S. L., Nasir, Z., Hermawan, D., Wan Ibrahim, W. A., & Naim, A. A. (2009). Comparison of signal-to-noise, blank determination, and linear regression methods for the estimation of detection and quantification limits for volatile organic compounds by gas chromatography. *Journal of AOAC International*, 92(6), 1833-1838.
- Santori, G., Di, G., Moglie, M., & Polonara, F. (2012). A review analyzing the industrial biodiesel production practice starting from vegetable oil refining. *Journal of Applied Energy*, 92, 109-132.
- Schollenberger, M., Muller, H. M., Ruffle, M., & Drochner, W. (2008). Natural occurrence of 16 fusarium toxins in edible oil marketed in Germany. *Journal of Food Control*, 19(5), 475-482.
- Shephard, G. S., Berthiller, F., Burdaspal, P. A., Crews, C., Jonker, M. A., Krska, R., & Whitaker, T. B. (2012). Developments in mycotoxin analysis: an update for 2011-2012. *World Mycotoxin Journal*, 6(1), 3-30.
- Shimelis, O., Yang, Y., Stenerson, K., Kaneko, T., & Ye, M. (2007). Evaluation of a solid-phase extraction dual-layer carbon/primary secondary amine for clean-up of fatty acid matrix components from food extracts in multiresidue pesticide analysis. *Journal of Chromatography A*, 1165(1), 18-25.
- Siegel, D., & Babuscio, T. (2011). Mycotoxin management in the European cereal trading sector. *Journal of Food Control*, 22(8), 1145-1153.
- Si, H., Zhang, L., Liu, S., LeRoith, T., & Virgous, C. (2014). High corn oil dietary intake improves health and longevity of aging mice. *Journal of Experimental Gerontology*, 58, 244-249.
- Sobhanzadeh, E., & Nemati, K. (2012). Liquid-liquid extraction/low-temperature purification (LLE/LTP) followed by dispersive solid-phase extraction (d-SPE) clean-up for multiresidue analysis in palm oil by LC-QTOF-MS. *Journal of Hazardous Material*, 186 (2-3), 1308 - 1313.
- Soleimany, F., Jinap, S., Faridah, A., & Khatib, A. (2012a). A UPLC e MS / MS for simultaneous determination of aflatoxins , ochratoxin A , zearalenone , DON , fumonisins , T-2 toxin and HT-2 toxin , in cereals. *Journal of Food Control*, 25(2), 647-653.
- Soleimany, F., Jinap, S., & Abas, F. (2012b). Determination of mycotoxins in cereals by liquid chromatography tandem mass spectrometry. *Journal of Food Chemistry*, 130(4), 1055-1060.
- Spanjer, M. C., Rensen, P. M., & Scholten, J. M. (2008). LC-MS/MS multi-method for mycotoxins after single extraction, with validation data for peanut, pistachio, wheat, maize, cornflakes, raisins and figs. *Journal of Food Additives and Contaminants*, 25(4), 472-489.
- Sulyok, M., Berthiller, F., Krska, R., & Schuhmacher, R. (2006). Development and validation of a liquid chromatography/tandem mass spectrometric method for

- the determination of 39 mycotoxins in wheat and maize. *Rapid Communications in Mass Spectrometry*, 20(18), 2649-2659.
- Tamura, M., Uyama, A., & Mochizuki, N. (2011). Development of a multi-mycotoxin analysis in beer-based drinks by a modified QuEChERS method and ultra-high-performance liquid chromatography coupled with tandem mass spectrometry. *Journal of Analytical Sciences*, 27(6), 629.
- Thompson, M., Ellison, S. L., & Wood, R. (2002). Harmonized guidelines for single-laboratory validation of methods of analysis. *Journal of Pure and Applied Chemistry*, 74(5), 835-855.
- Torres, C. M., Pico, Y., & Man, J. (1997). Comparison of octadecylsilica and graphitized carbon black as materials for solid-phase extraction of fungicide and insecticide residues from fruit and vegetables. *Journal of Chromatography A*, 778(1), 127-137.
- Van Pamel, E., Verbeken, A., Vlaemynek, G., De Boever, J., & Daeseleire, E. (2011). Ultrahigh-performance liquid chromatographic tandem mass spectrometric multimycotoxin method for quantitating 26 mycotoxins in maize silage. *Journal of Agricultural and Food Chemistry*, 59(18), 9747-9755.
- Vogeser, M., & Parhofer, K. G. (2007). Liquid chromatography tandem-mass spectrometry (LC-MS/MS)--technique and applications in endocrinology. *Experimental and clinical endocrinology & diabetes: Official Journal, German Society of Endocrinology and German Diabetes Association*, 115(9), 559-570.
- Wick, C. D., Stubbs, J. M., Rai, N., & Siepmann, J. I. (2005). Transferable potentials for phase equilibrium. Primary, secondary, and tertiary amines, nitroalkanes and nitrobenzene, nitriles, amides, pyridine, and pyrimidine. *The Journal of Physical Chemistry B*, 109(40), 18974-18982.
- Wilkowska, A., & Biziuk, M. (2011). Determination of pesticide residues in food matrices using the QuEChERS methodology. *Journal of Food Chemistry*, 125(3), 803-812.
- Yogendrarajah, P., Poucke, C. Van, Meulenaer, B. De, & Saeger, S. De. (2013). Development and validation of a QuEChERS based liquid chromatography tandem mass spectrometry method for the determination of multiple mycotoxins in spices. *Journal of Chromatography A*, 1297, 1-11.
- Zachariasova, M., Lacina, O., Malachova, A., Kostelanska, M., Poustka, J., Godula, M., & Hajslova, J. (2010). Novel approaches in analysis of fusarium mycotoxins in cereals employing ultra performance liquid chromatography coupled with high resolution mass spectrometry. *Journal of Analytica Chimica Acta*, 662(1), 51-61.
- Zhao, Z., Rao, Q., Song, S., Liu, N., Han, Z., Hou, J., & Wu, A. (2014). Simultaneous determination of major type B trichothecenes and deoxynivalenol-3-glucoside in animal feed and raw materials using improved DSPE combined with LC-MS/MS. *Journal of Chromatography B*, 963, 75-82.

- Zhao, Z., Rao, Q., Song, S., Liu, N., Han, Z., Hou, J., & Wu, A. (2014). Simultaneous determination of major type B trichothecenes and deoxynivalenol-3-glucoside in animal feed and raw materials using improved DSPE combined with LC-MS/MS. *Journal of Chromatography B*, 963, 75-82.
- Zinedine, A., Brera, C., Elakhdari, S., Catano, C., Debegnach, F., Angelini, S., & Miraglia, M. (2006). Natural occurrence of mycotoxins in cereals and spices commercialized in Morocco. *Journal of Food Control*, 17(11), 868-874.
- Zou, Z., He, Z., Li, H., Han, P., Tang, J., Xi, C., Li, X. (2012). Development and application of a method for the analysis of two trichothecenes: deoxynivalenol and T-2 toxin in meat in China by HPLC-MS/MS. *Journal of Meat Science*, 90(3), 613-617.
- Zwir-Ferenc, A., & Biziuk, M. (2006). Solid phase extraction technique trends, opportunities and applications. *Polish Journal of Environmental Studies*, 15(5), 677-690.