



UNIVERSITI PUTRA MALAYSIA

***SENSITIVE AND RAPID MODELS OF PCR-BASED METHODOLOGIES
FOR ADMIXTURE DETECTION AND QUANTIFICATION
OF DOMESTIC MEATS***

UMMI KALTHUM HANAPI

IPPH 2016 3



**SENSITIVE AND RAPID MODELS OF PCR-BASED METHODOLOGIES FOR
ADMIXTURE DETECTION AND QUANTIFICATION OF DOMESTIC
MEATS**

By

UMMI KALTHUM HANAPI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

February 2016

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

SENSITIVE AND RAPID MODELS OF PCR-BASED METHODOLOGIES FOR ADMIXTURE DETECTION AND QUANTIFICATION OF DOMESTIC MEATS

By

UMMI KALTHUM HANAPI

February 2016

Chair : Mohd Nasir Mohd Desa, PhD
Faculty : Halal Products Research Institute

This research project was undertaken to develop cost-effective, fast and sensitive PCR-based methods for fraud identification of meat-based products. Meats and meat-based product samples included in the analyses were pig, ruminant, avian and rabbit. Two methods were established for qualitative and quantitative evaluation of animal group contamination/ admixture in meat-based products. In this study, *NADH-dehydrogenase subunit 4 (Nad 4)* gene of mitochondrial was used. *Nad 4* gene holds great potential for identifying meat type due to high interspecies variability of the gene sequence. To date, very little attention was given in meat identification using *Nad 4* gene. To further recognize the potentially important of *Nad 4* gene, the sequence variability has been explored in identifying differences in the designated meat types.

For qualitative analysis, a common primer multiplex PCR (CP-M-PCR) was developed by using a common forward primer, species-specific adapter reverse primers and a common adapter reverse primer. The designed primers were analyzed *in silico* and tested against pure meat DNAs. The primers successfully generated specific fragments of 267, 370, 504, and 548 bp lengths for pig, ruminant, avian and rabbit meats, respectively. A serial dilution of each reverse primer was used to determine and compare the sensitivity of CP-M-PCR to conventional multiplex PCR system. The detection limit of CP-M-PCR was evaluated with 10-fold serial dilutions of DNA concentration. The use of adapter sequence at the 5'-end of the species-specific reverse primers was shown to increase the efficiency of the PCR amplification and the application of a single forward primer reduced the complexity in multiplex PCR system. Bands of specific amplification can be detected from the PCR assays containing as low as 10^{-6} μ M of adapter reverse primer. The CP-M-PCR limit of detection was as low as 0.01 ng of DNA for the four groups of meat which was deemed to be sufficient to qualitatively detecting accidental or intended contamination in meat products. The developed system was applied to 42 commercial meat-products and showed the presence of avian meat in analyzed ruminant (3/14), rabbit (2/2) and pig (1/11) samples.

In the subsequent work, a quantitative competitive PCR (QC-PCR) was developed to determine the percentage of contamination. Based on the CP-M-PCR results, six avian contaminated commercial meat-based products were quantitatively analysed using the calibrated QC-PCR. Prior to quantitative analysis, a competitor DNA of each animal meat group was constructed *via* site directed-mutagenesis processes. Site directed-mutagenesis was used to introduce a 40 bp fragment comprising a 30 bp insert and 10 bp repeated sequence. The constructed DNA competitors were coamplified with the target DNA at different ratios of concentration and results were analyzed using UVigeltec imaging software version 12.1. The results of QC-PCR showed that the percentage of contamination was in the range of 0.27 to 5.08% in the collected samples. The developed methods in this study are considered practical and valid alternatives in the context of a routine diagnostic laboratory where they are cost-effective, efficient, and sensitive enough for qualitative and quantitative investigation of contamination or adulteration in meat products. The developed approaches are cost-effective because they do not require expensive equipment, consumable and reagent, and high level of technical expertise to perform the analyses. Thus, the models may serve as templates for any screening system such typing of transgenic organisms, analyzing of forensic materials, detection of pathogens to cancer research, studying metagenomics, analyzing gene expression and monitoring environment.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

MODEL-MODEL YANG SENSITIF DAN PANTAS BERSANDARKAN KAEDAH PCR UNTUK PENGESANAN DAN PENGKUANTITIAN CAMPURAN PALSU DI DALAM DAGING TEMPATAN

Oleh

UMMI KALTHUM HANAPI

Februari 2016

Pengerusi : Mohd Nasir Mohd Desa, PhD
Fakulti : Institut Penyelidikan Produk Halal

Projek penyelidikan ini dijalankan untuk membina kaedah-kaedah yang menjimatkan, cepat dan sensitive untuk mengenal pasti kandungan daging tidak asli di dalam produk berasaskan daging. Sampel daging dan produk berasaskan daging yang digunakan di dalam analisis ini adalah terdiri daripada babi, ruminant, avian dan arnab. Dua kaedah telah dibangunkan untuk evaluasi kontaminasi/ percampuran secara kualitatif dan kuantitatif oleh kumpulan haiwan tersebut di dalam produk berasaskan daging. Di dalam kajian ini, gen mitokondria *NADH dehidrogenase subunit 4 (Nad 4)* telah digunakan. Gen *Nad 4* mempunyai potensi yang besar untuk pengesanan jenis daging berikutan variasi interspesis yang terdapat di dalam jujukan gennya. Sehingga kini, tidak banyak perhatian yang diberikan terhadap gen *Nad 4*. Untuk lebih memperlihatkan kepentingan potensi gen *Nad 4*, variasi jujukannya diterokai untuk mengenal pasti perbezaan-perbezaan yang terdapat di kalangan jenis daging.

Untuk analisis kualitatif, 'common primer multiplex PCR' (CP-M-PCR) telah dibangunkan dengan menggunakan primer forward sepunya, primer-primer reverse adapter spesies-spesifik dan primer reverse adapter. Primer-primer yang dicipta telah dianalisa secara *in siliko* dan diuji ke atas DNA daripada daging-daging yang asli. Primer-primer tersebut telah menghasilkan fragmen bersaiz 267, 370, 594 dan 548 bp untuk setiap kumpulan daging babi, ruminant, avian dan arnab. Pencairan bersiri primer telah digunakan untuk mengetahui dan membandingkan sensitiviti CP-M-PCR dengan sistem konvensional multiplex PCR. Had kemampuan pengesanan bagi CP-M-PCR telah dievaluasi melalui pencairan campuran DNA secara bersiri dalam gandaan 10. Aplikasi jujukan adapter pada hujung 5' primer-primer reverse adapter spesies-spesifik menunjukkan peningkatan efisiensi amplifikasi PCR dan penggunaan primer forward sepunya telah mengurangkan kompleksiti di dalam sistem PCR multiplex. Jalur-jalur terhasil daripada amplifikasi yang spesifik boleh dikesan daripada esei PCR yang mengandungi serendah 0.01 ng DNA untuk keempat-empat kumpulan daging tersebut di mana ianya mencukupi untuk mengesan secara kualitatif kontaminasi di dalam produk daging. Sistem yang dibangunkan ini telah diaplikasikan ke atas 42 produk

komersil daging dan menunjukkan kehadiran daging avian di dalam sampel ruminan (3/14), arnab (2/3) dan babi (1/11).

Kajian seterusnya, 'quantitatif competitive PCR' (QC-PCR) telah dibangunkan untuk mengesan peratus kontaminasi tersebut. Berdasarkan keputusan CP-M-PCR, enam produk daging yang dicemari oleh daging avian telah di analisa secara kuantitatif menggunakan QC-PCR yang telah dikalibrasi. Sebelum menjalankan analisis kuantitatif tersebut, DNA pesaing bagi setiap kumpulan daging haiwan telah dikonstruksi melalui proses 'site-directed mutagenesis'. Proses ini menghasilkan fragmen bersaiz 40 bp yang terdiri daripada 30 bp jujukan tambahan dan 10 bp jujukan berulang. DNA pesaing yang telah dikonstruksi tersebut telah di amplifikasi bersama-sama DNA target pada nisbah kepekatan berbeza dan hasil keputusan telah dianalisa menggunakan software pengimejan UVIgeltect versi 12.1. Hasil menunjukkan peratus kontaminasi adalah di dalam lingkungan 0.27-5.08% di dalam sampel-sampel yang diuji. Kaedah-kaedah yang dibangunkan di dalam kajian ini dianggap sebagai alternatif yang praktikal dan boleh diguna pakai di dalam konteks makmal diagnosis rutin di mana kedua-duanya adalah cukup murah, efisien dan sensitive untuk penyiasatan kontaminasi atau percampuran secara kualitatif dan kuantitatif di dalam produk daging. Pendekatan-pendekatan yang digunakan adalah murah kerana tidak memerlukan kepada peralatan, bahan dan reagen yang mahal, sertakemahiran teknikal yang tinggi untuk menjalankan analisa. Oleh demikian, model-model ini boleh menjadi templat untuk lain-lain sistem pengesanan seperti di dalam pengelasan organisma transgenic, analisa bahan forensic, pengesanan patogen untuk kajian kanser, kajian metagenomik, analisa ekspresi gen dan pengawalan alam sekitar.

ACKNOWLEDGEMENTS

My sincere thanks and gratitude goes to my supervisor Assoc. Prof. Dr. Mohd Nasir Mohd Desa from the Department of Biomedical Science, Faculty of Medicine and Health Sciences, UPM and also a research associate in Molecular Biology at Institute of Halal Products Research (PPH), UPM, for his unrelenting support, advices, and unstinting help during the research and the writing of this thesis.

I am also grateful beyond words to Prof. Dr. Amin Ismail, from the Faculty of Medicine and Health Sciences, and also as research associate at IPPH, UPM, for giving me the opportunity to pursue my studies in the field of Halal Product Science at IPPH, opinions, encouragement and financial assistance during the early of the research.

I would like to send my appreciation to Prof. Dr. Shuhaimi Mustafa, the Deputy Director of Institute of Halal Products Research, UPM, for his advices, insightful comments and precious assistance for the past few years.

There are a number of people behind this piece of work who deserve to be both acknowledged and thanked here; members of the Halal Research Laboratory and the IPPH' staffs.

Last but not least, I would like to express my indebtedness to my parents and other family members who have given me constant support and love.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Nasir bin Mohd Desa, PhD

Associate Professor
Halal Products Research Institute
Universiti Putra Malaysia
(Chairman)

Amin bin Ismail, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Shuhaimi bin Mustafa, PhD

Professor
Halal Products Research Institute
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Ummi Kalthum binti Hanapi , GS32680

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee:

Associate Professor
Dr Mohd Nasir bin Mohd Desa.

Signature: _____

Name of
Member of
Supervisory
Committee:

Professor
Dr. Amin bin Ismail

Signature: _____

Name of
Member of
Supervisory
Committee:

Professor
Dr. Shuhaimi bin Mustafa

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xviii
 CHAPTER	
 1 INTRODUCTION	 1
 2 LITERATURE REVIEW	 4
2.1 Food Fraud	4
2.1.1 Meat Fraud and Economically Motivated Adulteration (EMA)	5
2.2.1 Food Adulteration Control Systems	6
2.2.3 The Importance of the Determination of Meat Authenticity and the Detection of Adulteration	7
2.2 Methods for Identification and Evaluation of Meat Adulteration	9
2.2.1 Mass Spectrometry	10
2.2.2 Infrared Spectroscopy	13
2.2.3 Nuclear Magnetic Resonance Spectroscopy	13
2.2.4 Histology and Image Analysis	14
2.2.5 Electrophoretic Methods	14
2.2.6 Chromatography	15
2.2.7 Immunological Methods	16
2.2.8 DNA Hybridization	16
2.2.9 Polymerase Chain Reaction-Based Methods	17
2.2.9.1 Species-Specific End-point PCR	18
2.2.9.2 PCR-Restriction Fragment Length Polymorphism (PCR-RFLP)	19
2.2.9.3 Random Amplified Polymorphic DNAPCR (RAPD-PCR)	19
2.2.9.4 Real-Time PCR (qPCR)	20
2.3 DNA-Targeted Sequence to Access Meat Authenticity	21
Nuclear DNA and Mitochondrial DNA for the	21
2.3.1 Detection of Animal Origin in Meat Products	23
2.3.2 Nuclear DNA in Meat Identification	23
2.3.2.1 Repetitive Elements	23
2.3.2.2 Phosphodiesterase, Ryanodin Receptor, Guanosine Monophosphate and Interleukin-2	24

2.3.2.3	Melanocortin-1 Receptor	24
2.3.2.4	Actin	25
2.3.2.5	Myostatin	25
2.3.2.6	18S Ribosomal RNA	26
2.3.2.7	Growth Hormone	26
2.3.3	Mitochondrial DNA in Meat Identification	26
2.3.3.1	NADH Dehydrogenases of Respiratory Complex I	26
2.3.3.2	Cytochrome b of Respiratory Complex III	27
2.3.3.3	Cytochrome c Oxidases of Respiratory Complex IV	28
2.3.3.4	Adenosine Triphosphatase of Respiratory Complex V	28
2.3.3.5	Displacement Loop	29
2.3.3.6	Ribosomal RNA	29
2.4	PCR-Based Detection and Quantification of Meat Species	30
2.4.1	Advances in Multiplex PCR and Primer Design Strategy	30
2.4.1.1	Common Single Primer Multiplex Polymerase Chain Reaction (CSP-M-PCR)	32
2.4.1.2	Common Primer Multiplex Polymerase Chain Reaction (CP-M-PCR)	32
2.4.1.3	Universal Multiplex PCR (UM-PCR)	33
2.4.1.4	Universal Primer Multiplex PCR (UP-M-PCR)	33
2.4.1.5	Single Universal Primer Multiplex Ligation-Dependent Probe Amplification (SUP-MLPA)	34
2.4.1.6	Multiplex Real-Time PCR	35
2.4.2	PCR-Based Quantification of Meat Species	35
2.4.2.1	Quantitative Competitive PCR (QC-PCR) and the Strategies of Quantification	36
2.4.2.2	Other Strategies of Quantification of Meat	37

3	MATERIALS AND METHODS	39
3.1	Sample Preparation	39
3.1.1	Sample Collection	39
3.1.2	DNA Extraction of Raw Meat Samples	43
3.1.3	DNA Extraction of Meat Products	43
3.1.4	Determination of DNA Concentration	44
3.1.4.1	DNA Quantification Using PicoGreen	44
3.1.4.2	DNA Concentration Using NanoDrop	44
3.1.5	Determination of DNA Integrity	44
3.2	Development of Common Primer Multiplex PCR (CP-M-PCR)	45

3.2.1	Primer Design	45
3.2.2	Identification of Suitability of Annealing Temperature	49
3.2.3	Identification of Primer Specificity	49
3.2.4	Identification of Primer Sensitivity	50
3.2.5	Optimization of Common Primer Multiplex PCR	50
3.2.6	Identification of Detection Limit	52
3.3	Development of Quantitative Competitive PCR (QC-PCR)	52
3.3.1	Primer Design for Site Directed Mutagenesis Construction of Exogenous Template for DNA Competitor Preparation by Site- Directed Mutagenesis	52
3.3.2	Construction of Exogenous Template for DNA Competitor Preparation by Site-Directed Mutagenesis	52
3.3.3	Cloning and Transformation	53
3.3.4	White-Blue Colony Screening	53
3.3.5	Colony PCR Analysis	54
3.3.6	Plasmid DNA Preparation for Sequencing Analysis	54
3.3.7	Optimization of Suitable Dilutions of DNA Competitor	55
3.4	Method of Evaluation	55
3.4.1	Evaluation of Common Primer Multiplex PCR	55
3.4.2	Evaluation of Quantitative Competitive PCR Method	57
3.4.2.1	Standard Calibration	57
3.4.2.2	Determination of DNA Adulteration in Meat Products	57
4	RESULTS AND DISCUSSION	58
4.1	Optimization of DNA Extraction of Raw Meats and Highly Processed Meat Products	58
4.1.1	Determination of DNA Concentration	59
4.1.2	Determination of DNA Integrity	62
4.2	Development of Common Primer Multiplex PCR for Meat Analyses	65
4.2.1	Primer Design	65
4.2.2	Optimization of Simplex PCR	68
4.2.3	Primer Specificity	70
4.2.4	Primer Sensitivity	72
4.2.5	Development of CP-M-PCR Parameters: Technical Considerations and Interpretation of Multiplex PCR Results	74
4.2.6	Detection Limits of DNA Samples	76
4.3	Development of Quantitative Competitive PCR for Meat Analyses	77
4.3.1	Primer Design for Site-Directed Mutagenesis	77
4.3.2	Construction of Competitor DNA	78

4.3.3	Cloning of Mutational Nad 4 Cassettes and Confirmation of the Inserts	81
4.3.3.1	Blue-White Colony Screening	81
4.3.3.2	Colony Screening by PCR	82
4.3.3.3	Nucleotide Sequence Determination and Analyses	83
4.3.4	Determination of equivalent points	83
4.4	Evaluation of Meat Products	88
4.4.1	Qualitative Evaluation of CP-M-PCR	89
4.4.2	Semi-Quantitative Evaluation of QC-PCR	92
4.4.2.1	Standard Calibration	92
4.4.2.2	Quantification of Meat Content	92
5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	100
	REFERENCES	102
	APPENDICES	136
	BIODATA OF STUDENT	151
	LIST OF PUBLICATIONS	152

LIST OF TABLES

Table	Page
3.1 Raw meat samples included in the assay.	40
3.2 List of meat products analyzed in this study.	41
3.3 Simplex PCR cycling parameters.	48
3.4 CP-M-PCR cycling parameters.	49
3.5 A serial 10-fold dilution of competitor.	51
3.6 A serial 2-fold dilution of competitor.	56
3.7 Primer sequences used in this study.	56
4.1 Fluorescence determination of DNA concentration.	61
4.2 Identity of nucleotide sequence of <i>Nad 4</i> analyzed by BLAST search program.	72
4.3 Meat composition of commercial meat products analyzed by CP-M-PCR.	91
4.4 Percentages for different contamination samples of meat products.	98

LIST OF FIGURES

Figure		Page
2.1	Classification of detection methods.	11
2.2	Map of mtDNA.	23
3.1	Sequence alignment of the <i>Nad 4</i> from GenBank.	46
3.2	Step-by-step protocol of primer design.	47
3.3	Multiplex optimization and testing process.	50
3.4	Schematic representation of CP-M-PCR.	51
4.1	Schematic diagram showing the structural elements of the PicoGreen molecule responsible for the different interactions with DNA.	60
4.2	Linearity of PicoGreen assay. A serial dilution of standard λ DNA in TE was excited at 480 nm and the fluorescence emission intensity was measured at 520 nm by a microplate reader (Infinite® M200, NanoQuant, Tecan).	60
4.3	Electrophoretic analysis of genomic DNA extracted based on Wu <i>et al.</i> , (1995) method, performed on 1% (w/v) agarose gel in 1x LB electrophoresis buffer.	63
4.4	Electrophoretic analysis of total DNA from 42 meat products, performed on 1% (w/v) agarose gel in 1x LB electrophoresis buffer.	64
4.5	Amplification N4CF & N4CR.	66
4.6	Effect of annealing temperature in the range of 50.5 to 68.0 °C for (A) N4SusR-Ra primer (B) N4Rum-Ra primers (C) N4AviR-Ra primer and (D) N4OryR-Ra primer.	69
4.7	Specificity of simplex PCR of (A) N4Pig-Ra, (B) N4Rum-Ra, (C) N4Avi-Ra and (D) N4Rab-Ra.	71
4.8	Determination of CP-M-PCR sensitivity. PCR products amplified using (A) conventional multiplex PCR system and (B) CP-M-PCR system with a serial dilution of primer concentration.	73
4.9	Comparison of PCR products amplified with different multiplex systems (A) conventional multiplex PCR (B) common primer multiplex PCR.	75

4.10	Determination of detection limit. Serial dilutions of a mixed DNA template were used in the multiplex reaction.	76
4.11	Electrophoretic analyses of first PCR and second PCR products.	79
4.12	Schematic representative of site-directed mutagenesis in the construction of DNA competitor.	80
4.13	Blue/white selection on LB/IPTG/X-gal plate supplemented with ampicillin.	82
4.14	Screening of positive transformants using PCR. Amplified products of PCR analyses using N4CF/species-specific reverse primer and M13F/M13R.	84
4.15	Agarose gel analysis of plasmid DNA after purification with the Plasmid Extraction kit (Geneaid).	85
4.16	Inserts of mutant clones P2 (pig), R2 (ruminant), A2 (avian) and O2 (rabbit) were sequenced using M13 forward primer and aligned.	86
4.17	Optimizing the suitable dilutions of competitors. Decreasing concentrations of competitor DNA were separately added to a constant volume of target DNA.	87
4.18	Detection of pig, ruminant, avian and rabbit meats from 42 meat product samples using CP-M-PCR with common forward primer, species-special reverse primers and common adapter primer.	90
4.19	Calibration of internal DNA standard for pig group using log (standard/competitor) vs concentration of standards from artificial contaminant mixtures.	93
4.20	Calibration of internal DNA standard for ruminant group using log (standard/competitor) vs concentration of standards from artificial contaminant mixtures.	94
4.21	Calibration of internal DNA standard for avian group using log (standard/competitor) vs concentration of standards from artificial contaminant mixtures.	95
4.22	Calibration of internal DNA standard for avian group using log (standard/competitor) vs concentration of standards from artificial contaminant mixtures.	96
4.23	A linear regression line of avian using log (standard/competitor) vs concentration of standards for quantification of avian content in meat products using UVIgeltec version 12.1.	97

LIST OF ABBREVIATIONS

A	Adenosine
AFLP	Amplified fragment length polymorphism
AP-PCR	Arbitrarily primed PCR
APPI	Atmospheric pressure photoionization ionization
APPI-MS	Atmospheric pressure photoionization ionization mass spectrometry
ATPase	Adenosine triphosphate synthase
<i>BGH</i>	<i>Bovine growth hormone gene</i>
BOLD	Barcode of Life Database
BSE	Bovine spongiform encephalopathy
C	Cytosine
CE	Capillary electrophoresis
CE-SDS	Sodium dodecyl sulphate polymer-filled capillary gel electrophoresis
COX	Cytochrome c oxidase
CP	Common primer
CP-IRMS	Continuous flow isotope ratio mass spectrometer
CP-M-PCR	Common forward multiplex polymerase chain reaction
CSP-M-PCR	Common single primer multiplex PCR
CTAB	Cetyl trimethyl ammonium bromide
GMP	Guanosine monophosphate
<i>cytb</i>	<i>Cytochrome b gene</i>
DART	Direct analysis in real time
DART-TOF-MS	Direct analysis in real time time-of-flight mass spectrometry
ddPCR	Droplet digital PCR

DI-MS	Direct-infusion mass spectrometry
D-loop	Displacement loop
DMSO	Dimethyl sulphoxide
DNA	Deoxyribonucleic acid
DSC	Differential scanning calorimetry
DVS	Department of Veterinary Services
2D-DIGE	Two-dimensional difference gel electrophoresis
<i>E. coli</i>	<i>Escherichia coli</i>
EDTA	Ethylene diamine tetraacetate
EIA	Enzyme immunoassay
ELISA	Enzyme linked immunosorbent assay
EMA	Economically motivated adulteration
E-Nose	Electronic nose
ESI	electrospray ionization
ESI-MS	Electrospray ionization mass spectrometry
EU	European Union
FFN	Food Fraud Network
FISH	Fluorescent in situ hybridization
FT-ICR	Fourier transform ion cyclotron resonance
FTIR	Fourier transform infrared spectroscopy
G	Guanosine
GC	Gas chromatography
GC-FID	Gas chromatography-flame ionization detector
GC-MS	Gas chromatography mass spectrometry
GDF-8	Growth differentiation factor 8
GFSI	Global Food Safety Initiative

GH	Growth hormone
GHP	Good Hygiene Practice
GMO	Genetically modified organism
GMP	Good Manufacturing Practice
HCCP	Hazard Analysis and critical Control Point
HCl	Hydrochloric acid
HDC	Halal Industry Development Corporation
HPLC	High performance liquid chromatography
ICP-MS	Inductively coupled mass spectrometry
IEF	Isoelectric focusing
ILE-2	Interleukin-2 precursor
IMP3	The Third Industrial Master Plan
IPTG	Isopropyl- β -D thiogalactopyranoside
IR	Infrared
IRMS	Isotope-ratio mass spectrometry
IT	Ion trap
JAIN	Jabatan Agama Islam Negeri (State Islamic Religious Department)
JAKIM	Jabatan Kemajuan Islam Malaysia (The Malaysia Islamic Development Department)
kb	Kilobase
kbp	Kilobase pair
KCl	Kalium chloride (potassium chloride)
KPDNHEP	Kementerian Perdagangan Dalam Negeri dan Hal Ehwal Pengguna (Ministry of Domestic Trade and Consumer Affairs)
K ⁺	Kalium ion (potassium ion)
LB	Luria-Bertani (culture medium)
LB	Lithium borate (electrophoresis buffer)

LC	Liquid chromatographic
LC-MS	Liquid chromatography mass spectrometry
LINE	Long interspersed nuclear elements
LOD	Limit of detection
M	Molar
MAIN	Majlis Agama Islam Negeri (The State Islamic Religious Councils)
MALDI	Matrix-assisted laser desorption/ionization
MALDI-ToF-MS	Matrix-assisted laser desorption/ionization time-of-flight mass spectrometer
PAGE	Polyacrylamide gel electrophoresis
PAGIF	Polyacrylamide gel isoelectric focusing
PCR	Polymerase chain reaction
PCR-DGGE	Polymerase chain reaction denaturing gradient gel electrophoresis
PCR-RFLP	Polymerase chain reaction-restriction length fragment polymorphism
PCR-SSCP	Single strand conformation polymorphism polymerase chain reaction
pI	Isoelectric point
PNA-FISH	Peptide nucleic acid fluorescent in situ hybridization
ppt	Part per trillion
PTR-MS	Proton transfer reaction mass spectrometry
PTR-TOF-MS	Proton transfer reaction mass spectroscopy time-of-flight mass analyzer
Q	Single quadrupole
QC-PCR	Quantitative competitive polymerase chain reaction
qPCR	Quantitative polymerase chain reaction/ real-time polymerase chain reaction
QUID	Quantitative Ingredient Declaration
RAPD	Randomly amplified polymorphic DNA
RAPD-PCR	Random amplified polymorphic DNA polymerase chain reaction

RASFF	Rapid Alert System for Food and Feed
RFLP	Restriction fragment length polymorphism
RNA	Ribonucleic acid
RP-HPLC	Reverse phase-high performance liquid chromatography
rRNA	Ribosomal ribonucleic acid
RT-PCR	Reverse transcriptase PCR (RT-PCR)
URF	Unassigned reading frames
USP	US Pharmacopeia
V	Volt
vCJD	Creutzfeld Jacob Disease
VTNR	Variable number tandem repeat
X-gal	5-bromo-4-chloro-indolyl- β -D-galactopyranoside
WCO	World Customs Organization
w/v	Weight per volume

CHAPTER 1

INTRODUCTION

Meat authenticity had long been a global issue. Food fraud is committed either by cross-contamination or intentional adulteration including economically motivated adulteration (EMA). Adulteration usually refers to non-compliance with health, safety standards and religious issues. In 2014, meat products were the most investigated foodstuffs amongst other cases handled by EU Food Fraud Network (Food Fraud Network Activity Report, European Commission, 2014). Consumers are concerned about the honesty of ingredient declaration because the misdescription of food contents on the product labels may affect their lifestyles, health or religious concerns. In 2002, European legislation (EC Regulation 178/2002) was adopted to provide the general principles and requirements of food law and thus ensure the foodstuffs are safe and authentic for human consumption and animal feed. Another example is halal certification has become important in many countries supported by the increasing number of world's Muslim population. Halal status for all meat and meat-based products has been made mandatory in almost Islamic countries.

Meat has become the most targeted for species substitution in food products due to the high market price. For example, Cawthorn *et al.* (2013) reported that processed meats substituted with pork, chicken and also unconventional species such as donkey, goat and water buffalo were discovered and has become commonplace in South Africa; and the substitution of beef with rat meat in Indonesian market (Rohman *et al.*, 2011). Nowadays, more concern about meat authenticity has increased from the public and the governments in many countries especially after the incident of horse meat scandal in February 2013 (ZhongMeng *et al.*, 2015).

Malaysia has set to become a global halal hub in 2020. In relation to this, numerous methods were developed to identify species origin in meat mixtures and meat products including the employment of FTIR spectroscopy (Rahmania *et al.*, 2015; Rohman *et al.*, 2011), electronic nose and gas chromatography mass spectrometer (Nurjuliana *et al.*, 2011; Kurniawati *et al.*, 2014); polymerase chain reaction assay (Rahman *et al.*, 2014a, Rahman *et al.*, 2014b; Man *et al.*, 2007); PCR-RFLP (Murugiah *et al.*, 2009); and real-time polymerase chain reaction (Ali *et al.*, 2014). At the same time, rapid on-site detection kits (PORKline MEAT, Olipro® Meat ID gene chip, USM Porcine DNA Detection Kit) and portable detection machine (Hafys™) with high sensitivity and rapidity for sample screening had been developed for meat authenticity identification. Presently, an effort for the establishment of DNA barcoding for identification of meat species through microbial content (Faculty of Biotechnology and Cell Molecular Biology, Universiti Putra Malaysia) and the development of One-Minute Halal Detection kits (IAB Halal Quality Assurance Research Laboratory, Universiti Selangor) are underway to assist halal food monitoring. However, some of the testing methods are costly to be used commercially and do not suit for processed meat analysis. Adoption of cost-effective and suitable analyses for processed meat products as well as rapid, specific and robust, have important purpose towards health, religious and economic implications

for consumers, the food industries, quality control laboratories and regulatory agencies. As PCR-based technique provides a rapid and high throughput procedure even at a very small amount of sample, it was selected for the development of sensitive, simple and much cost effective (no need for sophisticated equipment or expensive reagents) methods for qualitative and quantitative analyses. The analytical framework designed in this study focused on the methods of identification and quantifying food adulteration or admixed with specific emphasis on controlling the cost of analyses.

Since multiplex PCR has been introduced in 1988 by Chamberlain, the identification of meat species in food and animal feed has become very popular until today (*e.g.* Ali *et al.*, 2015; Iwobi *et al.*, 2015, Safdar *et al.*, 2015; Kitpipit *et al.*, 2014). However, the application of common primer multiplex PCR (CP-M-PCR) system in food especially in meat identification is still very little. It may be that this technique requires detail knowledge of primer selection criteria coupled with a working knowledge of primer reaction chemistry in the PCR assay.

There is also a requirement to indicate on the food label the quantity (in percentage) of each ingredient contained in the food, known as the Qualitative Ingredient Declaration (QUID) labelling. QUID is important because the nutrition label alone cannot provide the significant information to the health benefit. As a result, in this study a quantitative competitive PCR (QC-PCR) method was developed to determine the percentage of contamination in meat products.

Nowadays, real-time quantitative PCR (qPCR) and free-flow isoelectric focusing become common practices in DNA quantification. But financial cost is a major contributing factor to these types of analysis because the equipment, reagents and consumables are expensive in which many laboratories are still unaffordable. Therefore, this method relying only mainly on normal thermocycler, is deliberated as an economic alternative to quantification method. As meat-based companies are growing, this method can be applied for meat-based food screening in any laboratory including in the rural area with minimal equipment.

Main objective:

To protect consumers from meat fraud and misdeclaration by developing sensitive, cost-effective and rapid qualitative and quantitative methods for monitoring meat contents of complex food products.

Specific objectives:

- 1) To develop a highly sensitive common-multiplex-PCR method for the rapid identification of meat in meat-based products by the application of common primers for routine analysis.
- 2) To validate the common-multiplex-PCR system for sensitive and accurate identification of meat species.
- 3) To develop a simple quantitative method for quantitative monitoring of meat-based product mixtures with a synthetic gene as a control template.
- 4) To determine the percentage of foreign DNA content in meat products as a direct reflection of meat contamination using competitive DNA.

Hypotheses of the study:

- 1) The common primer and adapter primer increase the amplification efficiently in CP-M-PCR assay.
- 2) The QC-PCR method enables detection of other fraudulent constitutions at minimal amount and reducing the existing inter-laboratory differences by using DNA competitor as an internal control.



REFERENCES

- Aarts, H. J., Bouw, E. M., Buntjer, J. B., Lenstra, J. A., & van Raamsdonk, L. W. (2006). Detection of bovine meat and bone meal in animal feed at a level of 0.1%. *Journal of AOAC International*, 89(6), 1443-1446.
- Abd-Elsalam, K. A. (2003). Bioinformatic tools and guideline for PCR primer design. *African Journal of Biotechnology*, 2(5), 91-95.
- Abdulmawjood, A., Krischek, C., Wicke, M., & Klein, G. (2012). Determination of pig sex in meat and meat products using multiplex real time-PCR. *Meat Science*, 91(3), 27
- Abdulmawjood, A., & Bülte, M. (2002). Identification of ostrich meat by restriction fragment length polymorphism (RFLP) analysis of cytochrome b gene. *Journal of Food Science*, 67(5), 1688-1691.
- Aebersold, R., & Mann, M. (2003). Mass spectrometry-based proteomics. *Nature*, 422(6928), 198-207.
- Alamprese, C., Casale, M., Sinelli, N., Lanteri, S., & Casiraghi, E. (2013). Detection of minced beef adulteration with turkey meat by UV-vis, NIR and MIR spectroscopy. *LWT-Food Science and Technology*, 53(1), 225-232.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2002). The cytoskeleton and cell behavior.
- Ali, M. E., Razzak, M. A., Hamid, S. B. A., Rahman, M. M., Al Amin, M., & Rashid, N. R. A. (2015). Multiplex PCR assay for the detection of five meat species forbidden in Islamic foods. *Food Chemistry*, 177, 214-224.
- Ali, M. E., Rahman, M. M., Hamid, S. B. A., Mustafa, S., Bhassu, S., & Hashim, U. (2014). Canine-specific pcr assay targeting cytochrome b gene for the detection of dog meat adulteration in commercial frankfurters. *Food Analytical Methods*, 7(1), 234-241.
- Ali, M. E., Hashim, U., Mustafa, S., Man, Y. C., Dhahi, T. S., Kashif, M., Uddin, M. K., & Hamid, S. A. (2012). Analysis of pork adulteration in commercial meatballs targeting porcine-specific mitochondrial cytochrome b gene by TaqMan probe real-time polymerase chain reaction. *Meat Science*, 91(4), 454-459.

- Ali, M. E., Hashim, U., Kashif, M., Mustafa, S., Man, Y. C., & Hamid, S. A. (2012). Development of swine-specific DNA markers for biosensor-based halal authentication. *Genet Mol Res*, 11(2), 1762-1772.
- Ali, M. E., Hashim, U., Mustafa, S., Man, Y. B., Yusop, M. H. M., Kashif, M., Dhani, T. S., Bari, M. F., Hakim, M. A., & Latif, M. A. (2011). Nanobiosensor for detection and quantification of DNA sequences in degraded mixed meats. *Journal of Nanomaterials*, 2011, 32.
- Altman, P. L., & Katz, D. D. (Eds.). (1976). Cell biology. Bethesda^ eMaryland: Federation of American Societies for Experimental Biology.
- Amaral, J. S., Santos, C. G., Melo, V. S., Costa, J., Oliveira, M. B. P., & Mafra, I. (2015). Identification of duck, partridge, pheasant, quail, chicken and turkey meats by species-specific PCR assays to assess the authenticity of traditional game meat Alheira sausages. *Food Control*, 47, 190-195.
- Anderson, M. L. (1999). *Nucleic Acid Hybridization*. Bios Scientific Publishers Ltd.
- Andrés, S., Murray, I., Navajas, E. A., Fisher, A. V., Lambe, N. R., & Bünger, L. (2007). Prediction of sensory characteristics of lamb meat samples by near infrared reflectance spectroscopy. *Meat science*, 76(3), 509-516.
- Aranceta-Garza, F., Perez-Enriquez, R., & Cruz, P. (2011). PCR-SSCP method for genetic differentiation of canned abalone and commercial gastropods in the Mexican retail market. *Food Control*, 22(7), 1015-1020.
- Asensio, L. (2008). Application of multiplex PCR for the identification of grouper meals in the restaurant industry. *Food Control*, 19(11), 1096-1099.
- Aslaminejad, A. A., Nassiry, M. R., Farajollahi, H., Mahdavi, M., Sekhavati, M. H., & Javadmanesh, A. (2010). Development and use of quantitative competitive PCR assay for detection of poultry DNA in sausage. *Food Biotechnology*, 24(3), 248-257.
- Auboeuf, D., & Vidal, H. (1997). The Use of the Reverse Transcription-Competitive Polymerase Chain Reaction to Investigate their Vivo Regulation of Gene Expression in Small Tissue Samples. *Analytical Biochemistry*, 245(2), 141-148.
- Aung, M. M., & Chang, Y. S. (2014). Traceability in a food supply chain: Safety and quality perspectives. *Food Control*, 39, 172-184.

- Ayaz, Y., Ayaz, N. D., & Erol, I. (2006). Detection of species in meat and meat products using Enzyme-Linked Immunosorbent Assay. *Journal of Muscle Foods*, 17(2), 214-220.
- Bahar, B., Monahan, F. J., Moloney, A. P., O'Kiely, P., Scrimgeour, C. M., & Schmidt, O. (2005). Alteration of the carbon and nitrogen stable isotope composition of beef by substitution of grass silage with maize silage. *Rapid Communications in Mass Spectrometry*, 19(14), 1937-1942.
- Bai, W. L., Yin, R. H., Zhao, S. J., Li, C., Ma, Z. J., Yin, R. L., Luo, G. B., & Zhao, Z. H. (2010). A PCR assay for sex determination of yak (*Bos grunniens*) meat by amplification of the male-specific SRY gene. *Food Control*, 21(5), 726-731.
- Bai, W., Xu, W., Huang, K., Yuan, Y., Cao, S., & Luo, Y. (2009). A novel common primer multiplex PCR (CP-M-PCR) method for the simultaneous detection of meat species. *Food Control*, 20(4), 366-370.
- Bahrami, A., Behzadi, S., Miraei-Ashtiani, S. R., Roh, S. G., & Katoh, K. (2013). Genetic polymorphisms and protein structures in growth hormone, growth hormone receptor, ghrelin, insulin-like growth factor 1 and leptin in Mehraban sheep. *Gene*, 527(1), 397-404.
- Badrudin, B., Mohamed, Z., Sharifuddin, J., Rezai, G., Mahir Abdullah, A., Abd Latif, I., & Ghazali Mohayidin, M. (2012). Clients' perception towards JAKIM service quality in Halal certification. *Journal of Islamic Marketing*, 3(1), 59-71.
- Ballin, N. Z. (2010). Authentication of meat and meat products. *Meat Science*, 86(3), 577-587.
- Ballin, N. Z., Vogensen, F. K., & Karlsson, A. H. (2009). Species determination—Can we detect and quantify meat adulteration?. *Meat Science*, 83(2), 165-174.
- Barazzoni, R., Short, K. R., & Nair, K. S. (2000). Effects of aging on mitochondrial DNA copy number and cytochrome oxidase gene expression in rat skeletal muscle, liver, and heart. *Journal of Biological Chemistry*, 275(5), 3343-3347.
- Barbin, D. F., Sun, D. W., & Su, C. (2013). NIR hyperspectral imaging as non-destructive evaluation tool for the recognition of fresh and frozen-thawed porcine longissimus dorsi muscles. *Innovative Food Science & Emerging Technologies*, 18, 226-236.
- Bellagamba, F., Comincini, S., Ferretti, L., Valfrè, F., & Moretti, V. M. (2006). Application of quantitative real-time PCR in the detection of prion-protein gene

species-specific DNA sequences in animal meals and feedstuffs. *Journal of Food Protection*®, 69(4), 891-896.

Bensasson, D., Zhang, D. X., & Hewitt, G. M. (2000). Frequent assimilation of mitochondrial DNA by grasshopper nuclear genomes. *Molecular Biology and Evolution*, 17(3), 406-415.

Bertram, H. C., & ERSEN, H. J. (2004). Applications of NMR in meat science. *Annual Reports on NMR Spectroscopy*, 53, 157-202

Berzofsky, J. A., Berkower, I. J., & Epstein, S. L. (1993). Antigen-antibody interactions and monoclonal antibodies. *Fundamental Immunology*. 3rd ed. New York: Raven, 421-465.

Besbes, N., Fattouch, S., & Sadok, S. (2011). Comparison of methods in the recovery and amplificability of DNA from fresh and processed sardine and anchovy muscle tissues. *Food Chemistry*, 129(2), 665-671.

Bharuthram, A., Paximadis, M., Picton, A. C., & Tiemessen, C. T. (2014). Comparison of a quantitative Real-Time PCR assay and droplet digital PCR for copy number analysis of the CCL4L genes. *Infection, Genetics and Evolution*, 25, 28-35.

Bier, M. (Ed.). (2013). *Electrophoresis: Theory, methods, and applications* (Vol. 2). Elsevier.

Bilandžić, N., Varenina, I., Kolanović, B. S., Oraić, D., & Zrnčić, S. (2012). Malachite green residues in farmed fish in Croatia. *Food Control*, 26(2), 393-396.

Bogenhagen, D. F. (2009). Biochemical isolation of mtDNA nucleoids from animal cells. In *Mitochondrial DNA* (pp. 3-14). Humana Press.

Bovey, F. A., Mirau, P. A., & Gutowsky, H. S. (1988). *Nuclear magnetic resonance spectroscopy*. Elsevier.

Breton, S., Milani, L., Ghiselli, F., Guerra, D., Stewart, D. T., & Passamonti, M. (2014). A resourceful genome: updating the functional repertoire and evolutionary role of animal mitochondrial DNAs. *Trends in Genetics*, 30(12), 555-564.

Brodmann, P. D., & Moor, D. (2003). Sensitive and semi-quantitative TaqMan™ real-time polymerase chain reaction systems for the detection of beef (*Bos taurus*) and the detection of the family Mammalia in food and feed. *Meat Science*, 65(1), 599-607.

- Brody, J. R., & Kern, S. E. (2004). History and principles of conductive media for standard DNA electrophoresis. *Analytical Biochemistry*, 333(1), 1-13.
- Bünger, L., Navajas, E. A., Stevenson, L., Lambe, N. R., Maltin, C. A., Simm, G., Fisher, A. V., & Chang, K. C. (2009). Muscle fibre characteristics of two contrasting sheep breeds: Scottish Blackface and Texel. *Meat Science*, 81(2), 372-381.
- Cai, Y., Li, X., Lv, R., Yang, J., Li, J., He, Y., & Pan, L. (2014). Quantitative Analysis of Pork and Chicken Products by Droplet Digital PCR. *BioMed Research International*, 2014.
- Cajka, T., Danhelova, H., Zachariasova, M., Riddellova, K., & Hajslova, J. (2013). Application of direct analysis in real time ionization–mass spectrometry (DART–MS) in chicken meat metabolomics aiming at the retrospective control of feed fraud. *Metabolomics*, 9(3), 545-557.
- Cammà, C., Di Domenico, M., & Monaco, F. (2012). Development and validation of fast Real-Time PCR assays for species identification in raw and cooked meat mixtures. *Food Control*, 23(2), 400-404.
- Cao, J., Xu, J., Wang, Q., Liu, R., Hu, C., & Zheng, J. (2010). Detection of species specific DNA fragments of tiger and leopard source materials by multiplex PCR method. *Journal of Biotechnology*, 150, 127.
- Cawthorn, D. M., Steinman, H. A., & Hoffman, L. C. (2013). A high incidence of species substitution and mislabelling detected in meat products sold in South Africa. *Food Control*, 32(2), 440-449.
- Chiappini, B., Brambilla, G., Agrimi, U., Vaccari, G., Aarts, H. J., Berben, G., Frezza, D., & Giambra, V. (2005). Real-time polymerase chain reaction approach for quantitation of ruminant-specific DNA to indicate a correlation between DNA amount and meat and bone meal heat treatments. *Journal of AOAC International*, 88(5), 1399-1403.
- Chamberlain, J. S., Gibbs, R. A., Rainer, J. E., Nguyen, P. N., & Thomas, C. (1988). Deletion screening of the Duchenne muscular dystrophy locus via multiplex DNA amplification. *Nucleic Acids Research*, 16(23), 11141-11156.
- Chang, C. H., Yao, C. J., Yu, H. Y., Liao, Y. C., Jang-Liaw, N. H., Tsai, C. L., & Shao, K. T. (2014). A molecular forensic method for identifying species composition of processed marine mammal meats. *Journal of Forensic and Legal Medicine*, 23, 65-69.

- Chang, H.-K., Park, J.W., Kim, W.G., Kim, K.T., Lee, M., Park, U.D., and Choi, B.G. (2005). The Expression of MAGE and GAGE Genes in Uterine Cervical Carcinoma of Korea by RT-PCR with Common Primers. *Gynecologic Oncology*, 97(2): 342-347.
- Cawthorn, D. M., Steinman, H. A., & Witthuhn, R. C. (2012). Evaluation of the 16S and 12S rRNA genes as universal markers for the identification of commercial fish species in South Africa. *Gene*, 491(1), 40-48.
- Che Man, Y. B., Mustafa, S., Khairil Mokhtar, N. F., Nordin, R., & Sazili, A. Q. (2012). Porcine-specific polymerase chain reaction assay based on mitochondrial d-loop gene for identification of pork in raw meat. *International Journal of Food Properties*, 15(1), 134-144.
- Chen, Y. T., & Hsieh, Y. H. P. (2014). A sandwich ELISA for the detection of fish and fish products. *Food Control*, 40, 265-273.
- Chen, F. C., Hsieh, Y. P., & Bridgman, R. C. (2002). Monoclonal antibodies against troponin I for the detection of rendered muscle tissues in animal feedstuffs. *Meat Science*, 62(4), 405-412.
- Cheng, X., He, W., Huang, F., Huang, M., & Zhou, G. (2014). Multiplex real-time PCR for the identification and quantification of DNA from duck, pig and chicken in Chinese blood curds. *Food Research International*, 60, 30-37.
- Cheng, Y. H., Wen, C. M., Ding, S. T., Kao, C. C., & Kuo, T. Y. (2003). Detecting meat-and-bone meal in ruminant'd feeds by species-specific PCR. *Journal of Animal and Feed Sciences*, 12(4), 849-858
- Chevallier, E., Chekri, R., Zinck, J., Guérin, T., & Noël, L. (2015). Simultaneous determination of 31 elements in foodstuffs by ICP-MS after closed-vessel microwave digestion: Method validation based on the accuracy profile. *Journal of Food Composition and Analysis*, 41, 35-41.
- Chikuni, K., Tabata, T., Kosugiyama, M., Monma, M., & Saito, M. (1994). Polymerase chain reaction assay for detection of sheep and goat meats. *Meat Science*, 37(3), 337-345.
- Chikuni, K., Ozutsumi, K., Koishikawa, T., & Kato, S. (1990). Species identification of cooked meats by DNA hybridization assay. *Meat Science*, 27(2), 119-128.

- Choi, W. S., & Hong, C. H. (2003). Rapid enumeration of *Listeria monocytogenes* in milk using competitive PCR. *International Journal of Food Microbiology*, 84(1), 79-85.
- Chun, H. A. O., Huan, W. A. N. G., Qinhu, L. I. U., & Xudong, L. I. (2009). Quantification of anaerobic ammonium-oxidizing bacteria in enrichment cultures by quantitative competitive PCR. *Journal of Environmental Sciences*, 21(11), 1557-1561.
- Clark, W., & Christopher, K. (2000). An Introduction to DNA: Spectrophotometry, degradation, and the 'Frankengel' experiment. *Tested Studies for Laboratory Reaching*, 22, 81-99.
- Colgan, S., O'Brien, L., Maher, M., Shilton, N., McDonnell, K., & Ward, S. (2001). Development of a DNA-based assay for species identification in meat and bone meal. *Food Research International*, 34(5), 409-414.
- Consonni, R., & Cagliani, L. R. (2010). Nuclear magnetic resonance and chemometrics to assess geographical origin and quality of traditional food products. *Advances in Food and Nutrition Research*, 59, 87-165.
- Davoli, R., & Braglia, S. (2007). Molecular approaches in pig breeding to improve meat quality. *Briefings in Functional Genomics & Proteomics*, 6(4), 313-321.
- Del Pulgar, J. S., Soukoulis, C., Carrapiso, A. I., Cappellin, L., Granitto, P., Aprea, E., romano, A., Gasperi, F., & Biasioli, F. (2013). Effect of the pig rearing system on the final volatile profile of Iberian dry-cured ham as detected by PTR-ToF-MS. *Meat Science*, 93(3), 420-428.
- Diaz, F., Fukui, H., Garcia, S., & Moraes, C. T. (2006). Cytochrome c oxidase is required for the assembly/stability of respiratory complex I in mouse fibroblasts. *Molecular and Cellular Biology*, 26(13), 4872-4881.
- Dieffenbach, C. W., Lowe, T. M., & Dveksler, G. S. (1993). General concepts for PCR primer design. *PCR Methods Appl*, 3(3), S30-S37.
- Di Pinto A, Forte VT, Conversano MC, Tantillo GM (2005) Duplex polymerase chain reaction for detection of pork meat in horse meat fresh sausages from Italian retail sources. *Food Control* 16(5): 391-394
- Dobson, R. L., Motlagh, S., Quijano, M., Cambron, R. T., Baker, T. R., Pullen, A. M., Regg, B. T., Bigalw-Kern, A. S., Vennard, T., Fix, A., Reineschuessel, R., Overmann, G., Shan, Y., & Daston, G. P. (2008). Identification and

- characterization of toxicity of contaminants in pet food leading to an outbreak of renal toxicity in cats and dogs. *Toxicological Sciences*, 106(1), 251-262.
- Dooley, J. J., Paine, K. E., Garrett, S. D., & Brown, H. M. (2004). Detection of meat species using TaqMan real-time PCR assays. *Meat Science*, 68(3), 431-438.
- Druml, B., Hohegger, R., & Cichna-Markl, M. (2015). Duplex real-time PCR assay for the simultaneous determination of the roe deer (*Capreolus capreolus*) and deer (sum of fallow deer, red deer and sika deer) content in game meat products. *Food Control*, 57, 370-376.
- Ebbehøj, K. F., & Thomsen, P. D. (1991). Differentiation of closely related species by DNA hybridization. *Meat Science*, 30(4), 359-366.
- Elgar, G., & Vavouri, T. (2008). Tuning in to the signals: noncoding sequence conservation in vertebrate genomes. *Trends in Genetics*, 24(7), 344-352.
- Espinoza, E. O., Lindley, N. C., Gordon, K. M., Ekhoﬀ, J. A., & Kirms, M. A. (1999). Electrospray ionization mass spectrometric analysis of blood for differentiation of species. *Analytical Biochemistry*, 268(2), 252-261.
- Everstine, K., Spink, J., & Kennedy, S. (2013). Economically motivated adulteration (EMA) of food: common characteristics of EMA incidents. *Journal of Food Protection*[®], 76(4), 723-735.
- Fajardo, V., González, I., Martín, I., Hernández, P. E., García, T., & Martín, R. (2008). Differentiation of European wild boar (*Sus scrofa scrofa*) and domestic swine (*Sus scrofa domestica*) meats by PCR analysis targeting the mitochondrial D-loop and the nuclear melanocortin receptor 1 (MC1R) genes. *Meat Science*, 78(3), 314-322.
- Farajollahi, A., Aslaminejad, A. A., Nassiri, M. R., Sekhavati, M. H., Mahdavi, M., & Javadmanesh, A. (2009). Development and use of quantitative competitive PCR assay for detection of poultry DNA in fish meal. *Journal of Animal and Feed Sciences*, 18.
- Farouk, M. M. (2013). Advances in the industrial production of halal and kosher red meat. *Meat Science*, 95(4), 805-820.
- Farrokhi, R., & Jafari Joozani, R. (2011). Identification of pork genome in commercial meat extracts for Halal authentication by SYBR green I real-time PCR. *International Journal of Food Science & Technology*, 46(5), 951-955.

- Faulks, L. K., Gilligan, D. M., & Beheregaray, L. B. (2008). Phylogeography of a threatened freshwater fish (*Mogurnda adspersa*) in eastern Australia: conservation implications. *Marine and Freshwater Research*, 59(1), 89-96.
- Fernandes, A. R., Tlustos, C., Smith, F., Carr, M., Petch, R., & Rose, M. (2009). Polybrominated diphenylethers (PBDEs) and brominated dioxins (PBDD/Fs) in Irish food of animal origin. *Food Additives and Contaminants: Part B*, 2(1), 86-94.
- Fernandez, S., Costa, A. C., Katsuyama, A. M., Madeira, A. M. B. N., & Gruber, A. (2003). A survey of the inter-and intraspecific RAPD markers of *Eimeria* spp. of the domestic fowl and the development of reliable diagnostic tools. *Parasitology Research*, 89(6), 437-445.
- Flaudrops, C., Armstrong, N., Raoult, D., & Chabrière, E. (2015). Determination of the animal origin of meat and gelatin by MALDI-TOF-MS. *Journal of Food Composition and Analysis*, 41, 104-112.
- Floren, C., Wiedemann, I., Brenig, B., Schütz, E., & Beck, J. (2015). Species identification and quantification in meat and meat products using droplet digital PCR (ddPCR). *Food Chemistry*, 173, 1054-1058.
- Fontanesi, L., Ribani, A., Scotti, E., Utzeri, V. J., Veličković, N., & Dall'Olio, S. (2014). Differentiation of meat from European wild boars and domestic pigs using polymorphisms in the MC1R and NR6A1 genes. *Meat Science*, 98(4), 781-784.
- Fontanesi, L., Tazzoli, M., Scotti, E., Russo, V., Xicato, G., Trocino, A., & Lukefahr, S. D. (2008). Analysis of candidate genes for meat production traits in domestic rabbit breeds. In *Proceedings of the 9th World Rabbit Congress, Verona, Italy, 10-13 June 2008* (pp. 79-84). World Rabbit Science Association.
- Frackman, S., Kobs, G., Simpson, D., & Storts, D. (1998). Betaine and DMSO: enhancing agents for PCR. *Promega Notes*, 65(27-29), 27-29.
- Frezza, D., Giambra, V., Chegiani, F., Fontana, C., Maccabiani, G., Losio, N., Faggionato, E., Chiappini, B., Vaccan, G., von Holst, C., Lanni, L., Saccare, S., & Ajmone-Marsan, P. (2008). Standard and Light-Cycler PCR methods for animal DNA species detection in animal feedstuffs. *Innovative Food Science & Emerging Technologies*, 9(1), 18-23.
- García, T., González, I., Asensio, L., Mayoral, B., López-Calleja, I., Hernández, P. E., & Martí, R. (2003). Development of a polymerase chain reaction assay for species identification of goose and mule duck in foie gras products. *Meat Science*, 65(4), 1257-1263.

- García-Rey, R. M., García-Olmo, J., De Pedro, E., Quiles-Zafra, R., & de Castro, M. L. (2005). Prediction of texture and colour of dry-cured ham by visible and near infrared spectroscopy using a fiber optic probe. *Meat Science*, 70(2), 357-363.
- Gefrides, L., & Welch, K. (2011). Forensic biology: Serology and DNA. In *the Forensic Laboratory Handbook Procedures and Practice* (pp. 15-50). Humana Press.
- Ghisleni, G., Stella, S., Radaelli, E., Mattiello, S., & Scanziani, E. (2010). Qualitative evaluation of tortellini meat filling by histology and image analysis. *International Journal of Food Science & Technology*, 45(2), 265-270.
- Ghovvati, S., Nassiri, M. R., Mirhoseini, S. Z., Moussavi, A. H., & Javadmanesh, A. (2009). Fraud identification in industrial meat products by multiplex PCR assay. *Food Control*, 20(8), 696-699.
- Gilliland, G., Perrin, S., Blanchard, K., & Bunn, H. F. (1990). Analysis of cytokine mRNA and DNA: detection and quantitation by competitive polymerase chain reaction. *Proceedings of the National Academy of Sciences*, 87(7), 2725-2729.
- Girish, P. S., Anjaneyulu, A. S. R., Viswas, K. N., Santhosh, F. H., Bhilegaonkar, K. N., Agarwal, R. K., Kondaiah, N., & Nagappa, K. (2007). Polymerase chain reaction–restriction fragment length polymorphism of mitochondrial 12S rRNA gene: a simple method for identification of poultry meat species. *Veterinary Research Communications*, 31(4), 447-455.
- Girish, P. S., Anjaneyulu, A. S. R., Viswas, K. N., Shivakumar, B. M., Anand, M., Patel, M., & Sharma, B. (2005). Meat species identification by polymerase chain reaction–restriction fragment length polymorphism (PCR-RFLP) of mitochondrial 12S rRNA gene. *Meat Science*, 70(1), 107-112.
- Glare, E. M., Divjak, M., Bailey, M. J., & Walters, E. H. (2001). The usefulness of competitive PCR: airway gene expression of IL-5, IL-4, IL-4 δ 2, IL-2, and IFN γ in asthma. *Thorax*, 56(7), 541-548.
- Gobbin, D., Rezzonico, F., & Gessler, C. (2007). Quantification of the biocontrol agent *Pseudomonas fluorescens* Pf153 in soil using a quantitative competitive PCR assay unaffected by variability in cell lysis and DNA-extraction efficiency. *Soil Biology and Biochemistry*, 39(7), 1609-1619.
- Gokulakrishnan, P., Kumar, R. R., Sharma, B. D., Mendiratta, S. K., Malav, O. P., & Sharma, D. (2013). Determination of sex origin of meat from cattle, sheep and goat using PCR based assay. *Small Ruminant Research*, 113(1), 30-33.

- González-Dominguez, R., García-Barrera, T., & Gómez-Ariza, J. L. (2012). Iberian ham typification by direct infusion electrospray and photospray ionization mass spectrometry fingerprinting. *Rapid Communications in Mass Spectrometry*, 26(7), 835-844.
- Grossman, P. D., & Colburn, J. C. (Eds.). (2012). *Capillary electrophoresis: Theory and practice*. Academic Press.
- Gudnason, H., Dufva, M., Bang, D. D., & Wolff, A. (2007). Comparison of multiple DNA dyes for real-time PCR: effects of dye concentration and sequence composition on DNA amplification and melting temperature. *Nucleic Acids Research*, 35(19), e127.
- Habib, M., Lakra, W. S., Mohindra, V., Lal, K. K., Punia, P., Singh, R. K., & Khan, A. A. (2012). Assessment of ATPase 8 and ATPase 6 mtDNA sequences in genetic diversity studies of *Channa marulius* (Channidae: Perciformes). *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 82(4), 497-501.
- Hadidi, A., & Candresse, T. (2003). Polymerase chain reaction. *Viroids*, 115-122.
- Hadi, F., Salmanian, A. H., Ghazizadeh, E., Amani, J., Noghabi, K. A., & Mousavi, A. (2012). Development of quantitative competitive PCR for determination of copy number and expression level of the synthetic glyphosate oxidoreductase gene in transgenic canola plants. *Electronic Journal of Biotechnology*, 15(4), 2-2.
- Haider, N., Nabulsi, I., & Al-Safadi, B. (2012). Identification of meat species by PCR-RFLP of the mitochondrial COI gene. *Meat Science*, 90(2), 490-493.
- Hanaki, K., Nakatake, H., Yamamoto, K., Odawara, T., & Yoshikura, H. (1996). Trypan Blue as a Slow Migrating Dye for SSCP Detection in Polyacrylamide Gel Electrophoresis. *BioTechniques*, 17, 1034-1036.
- Hartwell, L. H., Hood, L., Goldberg, M. L., Reynolds, A. E., Silver, L. M., & Veres, R. C. (2008). *Genetics: From genes to genome* (3rd ed.). New York, NY: McGraw-Hill.
- Haugland, R. A., Heckman, J. L., & Wymer, L. J. (1999). Evaluation of different methods for the extraction of DNA from fungal conidia by quantitative competitive PCR analysis. *Journal of Microbiological Methods*, 37(2), 165-176.

- Hauswirth, W. W., & Clayton, D. A. (1985). Length heterogeneity of a conserved displacement-loop sequence in human mitochondrial DNA. *Nucleic Acids Research*, 13(22), 8093-8104.
- He, N., Aosai, F., Luo, W. T., Ueda, M., Yang, T. H., Yamashita, K., Sekiya, S., & Yano, A. (1997). Parasite load in pregnant mice infected by *Toxoplasma gondii* assayed by quantitative competitive-PCR. *Parasitology International*, 46(2), 143-147.
- Heininger, A., Binder, M., Ellinger, A., Botzenhart, K., Unertl, K., & Döring, G. (2003). DNase pretreatment of master mix reagents improves the validity of universal 16S rRNA gene PCR results. *Journal of Clinical Microbiology*, 41(4), 1763-1765.
- Henegariu O, Heerema NA, Dlouhy SR, Vance GH, Vogt PH (1997) Multiplex PCR: Critical parameters and step-by-step protocol. *Biotechniques* 23:504-511
- Herman, L. (2001). Determination of the animal origin of raw food by species-specific PCR. *Journal of Dairy Research*, 68(03), 429-436.
- Herrera, C. M. (2009). *Multiplicity in unity: Plant subindividual variation and interactions with animals*. University of Chicago Press.
- Ho, S. N., Hunt, H. D., Horton, R. M., Pullen, J. K., & Pease, L. R. (1989). Site-directed mutagenesis by overlap extension using the polymerase chain reaction. *Gene*, 77(1), 51-59.
- Holm, E. S., Adamsen, A. P. S., Feilberg, A., Schäfer, A., Løkke, M. M., & Petersen, M. A. (2013). Quality changes during storage of cooked and sliced meat products measured with PTR-MS and HS-GC-MS. *Meat Science*, 95(2), 302-310.
- Hopwood, A., Brookes, J., Shariff, A., Cage, P., Tatum, E., Mirza, R., Crook, M., Brews, K., & Sullivan, K. (1997). A fully integrated robotic system for high sample throughput within a DNA databasing unit. *Proceedings: LabAutomation '98*, 17-21.
- Hou, B., Meng, X., Zhang, L., Guo, J., Li, S., & Jin, H. (2015). Development of a sensitive and specific multiplex PCR method for the simultaneous detection of chicken, duck and goose DNA in meat products. *Meat Science*, 101, 90-94.
- Hubalkova, Z., Kralik, P., Tremlova, B., & Rencova, E. (2007). Methods of gadoid fish species identification in food and their economic impact in the Czech Republic: a review. *VeterinarniMedicina*, 52(7), 273.

- Hubbard, B. J., Hatfield, J. T., & Santucci, J. A. (2007). *An educator's classroom guide to America's religious beliefs and practices*. Libraries Unlimited.
- Hueston, W. D. (2013). BSE and variant CJD: Emerging science, public pressure and the vagaries of policy-making. *Preventive Veterinary Medicine*, 109(3), 179-184.
- Hung, T., Mak, K., & Fong, K. (1990). A specificity enhancer for polymerase chain reaction. *Nucleic Acids Research*, 18(16), 4953.
- Hunt, P. W. (2011). Molecular diagnosis of infections and resistance in veterinary and human parasites. *Veterinary Parasitology*, 180(1), 12-46.
- Hübner, P., Studer, E., and Lüthy, J. (1999). Quantitative competitive PCR for the detection of genetically modified organisms in food. *Food Control*, 10(6): 353-358.
- Hübner, P., Waiblinger, H. U., Pietsch, K., & Brodmann, P. (2001). Validation of PCR methods for quantitation of genetically modified plants in food. *Journal of AOAC International*, 84(6), 1855-1864.
- Hwang, I. H., Park, B. Y., Kim, J. H., Cho, S. H., & Lee, J. M. (2005). Assessment of postmortem proteolysis by gel-based proteome analysis and its relationship to meat quality traits in pig longissimus. *Meat Science*, 69(1), 79-91.
- Ichinoseki, S., Nishiumi, T., & Suzuki, A. (2006). Tenderizing effect of high hydrostatic pressure on bovine intramuscular connective tissue. *Journal of food Science*, 71(6), E276-E281.
- Iwobi, A., Sebah, D., Kraemer, I., Losher, C., Fischer, G., Busch, U., & Huber, I. (2015). A multiplex real-time PCR method for the quantification of beef and pork fractions in minced meat. *Food Chemistry*, 169, 305-313.
- Jiang, Y. L., Li, N., Plastow, G., Liu, Z. L., Hu, X. X., & Wu, C. X. (2002). Identification of three SNPs in the porcine myostatin gene (MSTN). *Animal Biotechnology*, 13(1), 173-178.
- Jin, S., Park, H. B., Seo, D. W., Cahyadi, M., Choi, N. R., Heo, K. N., & Lee, J. H. (2014). Association of MC1R genotypes with shank color traits in Korean native chicken. *Livestock Science*, 170, 1-7.
- Johnson, T. M., Zurlo, G. A., Hickman, A. W., & Crossing, P. F. (2016). Christianity 2016: Latin America and projecting religions to 2050. *International Bulletin of Mission Research*, 40(1), 22-29.

- Jorfi, R., Mustafa, S., Man, Y. B. C., Hashim, D. B. M., Sazili, A. Q., & Farjam, A. S. (2014). Differentiation of pork from beef, chicken, mutton and chevon according to their primary amino acids content for halal authentication. *African Journal of Biotechnology*, 11(32).
- Kabekkodu, S. P., Bhat, S., Mascarenhas, R., Mallya, S., Bhat, M., Pandey, D., Kushtagi, P., Thangaraj, K., Gopinath, P. M., & Satyamoorthy, K. (2014). Mitochondrial DNA variation analysis in cervical cancer. *Mitochondrion*, 16, 73-82.
- Kanthaswamy, S., Premasuthan, A., Ng, J., Satkoski, J., & Goyal, V. (2012). Quantitative real-time PCR (qPCR) assay for human–dog–cat species identification and nuclear DNA quantification. *Forensic Science International: Genetics*, 6(2), 290-295.
- Khamnamtong, B., Klinbunga, S., & Menasveta, P. (2005). Species identification of five penaeid shrimps using PCR-RFLP and SSCP analyses of 16S ribosomal DNA. *BMB Reports*, 38(4), 491-499.
- Klinbunga, S., & Menasveta, P. (2005). Species identification of five penaeid shrimps using PCR-RFLP and SSCP analyses of 16S ribosomal DNA. *BMB Reports*, 38(4), 491-499.
- Kamruzzaman, M., Sun, D. W., ElMasry, G., & Allen, P. (2013). Fast detection and visualization of minced lamb meat adulteration using NIR hyperspectral imaging and multivariate image analysis. *Talanta*, 103, 130-136.
- Kang, S. I., Her, M., Kim, J. W., Kim, J. Y., Ko, K. Y., Ha, Y. M., & Jung, S. C. (2011). Advanced multiplex PCR assay for differentiation of Brucella species. *Applied and Environmental Microbiology*, 77(18), 6726-6728.
- Karabasanavar, N. S., Singh, S. P., mm, D., & Shebannavar, S. N. (2014). Detection of pork adulteration by highly-specific PCR assay of mitochondrial D-loop. *Food Chemistry*, 145, 530-534.
- Kaupe, B., Winter, A., Fries, R., & Erhardt, G. (2004). DGAT1 polymorphism in *Bos indicus* and *Bos taurus* cattle breeds. *Journal of Dairy Research*, 71(02), 182-187.
- Kesmen, Z., Celebi, Y., Güllüce, A., & Yetim, H. (2013). Detection of seagull meat in meat mixtures using real-time PCR analysis. *Food Control*, 34(1), 47-49.
- Kesmen, Z., Yetiman, A. E., Şahin, F., & Yetim, H. (2012). Detection of chicken and turkey meat in meat mixtures by using Real-Time PCR Assays. *Journal of Food Science*, 77(2), C167-C173.

- Kesmen, Z., Gulluce, A., Sahin, F., & Yetim, H. (2009). Identification of meat species by TaqMan-based real-time PCR assay. *Meat Science*, 82(4), 444-449.
- Kesmen, Z., Sahin, F., & Yetim, H. (2007). PCR assay for the identification of animal species in cooked sausages. *Meat Science*, 77(4), 649-653.
- Kim E, Cheong HS, Bae JS, Chun J, Park TJ, Lee K, Yun Y, Shin HD (2010) Identification of genetic polymorphisms in bovine mitochondrial deoxyribonucleic acid. *Journal of Animal Science* 88(8): 2551-2555
- Kim, S. H., Huang, T. S., Seymour, T. A., Wei, C. I., Kempf, S. C., Bridgman, C. R., Momcilovic, D., Clemens, R. A., & An, H. (2005). Development of immunoassay for detection of meat and bone meal in animal feed. *Journal of Food Protection*[®], 68(9), 1860-1865.
- Kim, Y., Gharaibeh, S. M., Stedman, N. L., & Brown, T. P. (2002). Comparison and verification of quantitative competitive reverse transcription polymerase chain reaction (QC-RT-PCR) and real time RT-PCR for avian leukosis virus subgroup J. *Journal of Virological Methods*, 102(1), 1-8.
- Kitpipit, T., Sittichan, K., & Thanakiatkrai, P. (2014). Direct-multiplex PCR assay for meat species identification in food products. *Food Chemistry*, 163, 77-82.
- Koh, M. C., Lim, C. H., Chua, S. B., Chew, S. T., & Phang, S. T. W. (1998). Random amplified polymorphic DNA (RAPD) fingerprints for identification of red meat animal species. *Meat Science*, 48(3), 275-285.
- Kobayashi, Y., Shinkai, T., & Koike, S. (2008). Ecological and physiological characterization shows that *Fibrobacter succinogenes* is important in rumen fiber digestion-review. *Folia Microbiologica*, 53(3), 195-200.
- Kong, Q., Zheng, M., Casalone, C., Qing, L., Huang, S., Chakraborty, B., Wang, P., Chen, F., Cali, I., Corona, C., Martucci, F., Iulini, B., Acutis, P., Wang, L., Liang, J., Wang, M., Li, X., Monaco, S., Zanusso, G., Zou, W., Q., Caramelli, M., & Gambetti, P. (2008). Evaluation of the human transmission risk of an atypical bovine spongiform encephalopathy prion strain. *Journal of Virology*, 82(7), 3697-3701.
- Koronakis, V., Hughes, C., & Koronakis, E. (1993). ATPase activity and ATP/ADP-induced conformational change in the soluble domain of the bacterial protein translocator HlyB. *Molecular Microbiology*, 8(6), 1163-1175.

- Köppel, R., Zimmerli, F., & Breitenmoser, A. (2009). Heptaplex real-time PCR for the identification and quantification of DNA from. *European Food Research and Technology*, 230(1), 125
- Köppel, R., Ruf, J., Zimmerli, F., & Breitenmoser, A. (2008). Multiplex real-time PCR for the detection and quantification of DNA from beef, pork, chicken and turkey. *European Food Research and Technology*, 227(4), 1199-1203
- Kumar, D., Singh, S. P., Singh, R., & Karabasanavar, N. S. (2011). A highly specific PCR assay for identification of goat (*Capra hircus*) meat. *Small Ruminant Research*, 97(1), 76-78.
- Kurniawati, E., Rohman, A., & Triyana, K. (2014). Analysis of lard in meatball broth using Fourier transform infrared spectroscopy and chemometrics. *Meat Science*, 96(1), 94-98.
- Kvist, L. (2000). Phylogeny and phylogeography of European Parids (pp. 51). Oulu, Finland: University of Oulu.
- Kwon, D. Y., & Tamang, J. P. (2015). Religious Ethnic Food. *Journal of Ethnic Foods*.
- Lahiff, S., Glennon, M., Lyng, J., Smith, T., Shilton, N., & Maher, M. (2002). Real-time polymerase chain reaction detection of bovine DNA in meat and bone meal samples. *Journal of Food Protection*®, 65(7), 1158-1165
- Larzul, D., Guigue, F., Sninsky, J. J., Mack, D. H., Brechot, C., & Guesdon, J. L. (1988). Detection of hepatitis B virus sequences in serum by using in vitro enzymatic amplification. *Journal of Virological Methods*, 20(3), 227-237.
- Laube, I., Zagon, J., & Broll, H. (2007a). Quantitative determination of commercially relevant species in foods by real-time PCR. *International Journal of Food Science & Technology*, 42(3), 336-341.
- Laube, I., Zagon, J., Spiegelberg, A., Butschke, A., Kroh, L. W., & Broll, H. (2007b). Development and design of a 'ready-to-use' reaction plate for a PCR-based simultaneous detection of animal species used in foods. *International journal of food science & technology*, 42(1), 9-17
- Laube, I., Spiegelberg, A., Butschke, A., Zagon, J., Schauzu, M., Kroh, L., & Broll, H. (2003). Methods for the detection of beef and pork in foods using real-time polymerase chain reaction. *International Journal of Food Science & Technology*, 38(2), 111-118.

- Li, J., Zhao, G. H., Zou, F. C., Mo, X. H., Yuan, Z. G., Ai, L., Weng, Y. B., Lin, R. Q., & Zhu, X. Q. (2010). Combined mitochondrial 16S and 12S rDNA sequences: an effective genetic marker for inter-species phylogenetic analysis of zoonotic trematodes. *Parasitology Research*, 107(3), 561-569.
- Li, W., & Drake, M. A. (2001). Development of a Quantitative Competitive PCR Assay for Detection and Quantification of *Escherichia coli* O157: H7 Cells. *Applied and Environmental Microbiology*, 67(7), 3291-3294.
- Liddell, S., Jenkins, M. C., & Dubey, J. P. (1999). A competitive PCR assay for quantitative detection of *Neospora caninum*. *International Journal for Parasitology*, 29(10), 1583-1587.
- Lin, C. C., Fung, L. L., Chan, P. K., Lee, C. M., Chow, K. F., & Cheng, S. H. (2014). A rapid low-cost high-density DNA-based multi-detection test for routine inspection of meat species. *Meat Science*, 96(2), 922-929.
- Lin, W. F., Lyu, Y. C., Wu, Y. J., Lu, C. H., & Hwang, D. F. (2012a). Species identification of snapper: A food poisoning incident in Taiwan. *Food Control*, 25(2), 511-515.
- Lin, R. Q., Liu, G. H., Song, H. Q., Zhang, Y., Li, M. W., Zou, F. C., Yuan, Z. G., Weng, Y. B., & Zhu, X. Q. (2012b). Sequence variability in three mitochondrial genes between the two pig nodule worms *Oesophagostomum dentatum* and *O. quadrispinulatum*. *Mitochondrial DNA*, 23(3), 182-186.
- Lin, J., Arnold, H. B., Della-Fera, M. A., Azain, M. J., Hartzell, D. L., & Baile, C. A. (2002). Myostatin knockout in mice increases myogenesis and decreases adipogenesis. *Biochemical and Biophysical Research Communications*, 291(3), 701-706.
- Liu, X., Guo, B., Wei, Y., Shi, J., & Sun, S. (2013). Stable isotope analysis of cattle tail hair: A potential tool for verifying the geographical origin of beef. *Food Chemistry*, 140(1), 135-140.
- Liu, Y., Lyon, B. G., Windham, W. R., Realini, C. E., Pringle, T. D. D., & Duckett, S. (2003). Prediction of color, texture, and sensory characteristics of beef steaks by visible and near infrared reflectance spectroscopy. A feasibility study. *Meat Science*, 65(3), 1107-1115.
- Liu, Q., Wang, L., Willson, P., & Babiuk, L. A. (2000). Quantitative, competitive PCR analysis of porcine circovirus DNA in serum from pigs with postweaning multisystemic wasting syndrome. *Journal of Clinical Microbiology*, 38(9), 3474-3477.

- Lockley, A. K., & Bardsley, R. G. (2002). Intron variability in an actin gene can be used to discriminate between chicken and turkey DNA. *Meat Science*, 61(2), 163-168.
- Lockley, A. K., & Bardsley, R. G. (2000). DNA-based methods for food authentication. *Trends in Food Science & Technology*, 11(2), 67-77.
- López-Andreo, M., Aldeguer, M., Guillén, I., Gabaldón, J. A., & Puyet, A. (2012). Detection and quantification of meat species by qPCR in heat-processed food containing highly fragmented DNA. *Food Chemistry*, 134(1), 518-523.
- López-Andreo, M., Lugo, L., Garrido-Pertierra, A., Prieto, M. I., & Puyet, A. (2005). Identification and quantitation of species in complex DNA mixtures by real-time polymerase chain reaction. *Analytical Biochemistry*, 339(1), 73-82.
- Macedo-Silva, A., Barbosa, S. F. C., Alkmin, M. G. A., Vaz, A. J., Shimokomaki, M., & Tenuta-Filho, A. (2000). Hamburger meat identification by dot-ELISA. *Meat Science*, 56(2), 189-192.
- Maede, D. (2006). A strategy for molecular species detection in meat and meat products by PCR-RFLP and DNA sequencing using mitochondrial and chromosomal genetic sequences. *European Food Research and Technology*, 224(2), 209-217.
- Magoulas, A. (2005). Mitochondrial DNA. In S. X. Cadrin, K. D. Friedland, & J. R. Waldman (Eds.), *Stock identification methods: Applications in fishery science* (pp. 311–330). Burlington, MA: Elsevier Academic Press.
- Malila, Y., Tempelman, R. J., Sporer, K. R. B., Ernst, C. W., Velleman, S. G., Reed, K. M., & Strasburg, G. M. (2013). Differential gene expression between normal and pale, soft, and exudative turkey meat. *Poultry Science*, 92(6), 1621-1633.
- Malisa, A. L., Gwakisa, P., Balthazary, S., Wasser, S. K., & Mutayoba, B. M. (2006). The potential of mitochondrial DNA markers and polymerase chain reaction-restriction fragment length polymorphism for domestic and wild species identification. *African Journal of Biotechnology*, 5(18).
- Man, Y. C., Aida, A. A., Raha, A. R., & Son, R. (2007). Identification of pork derivatives in food products by species-specific polymerase chain reaction (PCR) for halal verification. *Food Control*, 18(7), 885-889.
- Mane, B. G., Mendiratta, S. K., & Tiwari, A.K. (2013) Pork Specific Polymerase Chain Reaction Assay for Authentication of Meat and Meat Products. *Journal of Meat Science and Technology* 1(1): 21-27

- Mane, B. G., Mendiratta, S. K., & Tiwari, A. K. (2012). Beef specific polymerase chain reaction assay for authentication of meat and meat products. *Food Control*, 28(2), 246-249.
- Mane, B. G., Mendiratta, S. K., & Tiwari, A. K. (2009). Polymerase chain reaction assay for identification of chicken in meat and meat products. *Food Chemistry*, 116(3), 806-810.
- Mane, B. G., Tanwar, V. K., Girish, P. S., Sonawane, A. A., & Sharma, D. (2008a). Differentiation of meat species by means of Polymerase Chain Reaction technique. *International Journal of Food Safety, Nutrition and Public Health*, 1(1), 51-57.
- Mane, B. G., Tanwar, V. K., Girish, P. S., Sharma, D., & Dixit, V. P. (2008). RAPD markers for differentiation of meat species. *Indian Journal of Veterinary Research*, 17(2), 9-13.
- Mardegan Issa, J. P., Tiossi, R., & Mizusaki Iyomasa, M. (2007). Morphological and histochemical study of the masseter muscle after occlusal alteration. *Biocell*, 31(3), 375-382.
- Mariasegaram, M., Robinson, N. A., & Goddard, M. E. (2006). Quantification of cattle DNA using quantitative competitive PCR with sheep DNA as competitor. *Molecular and Cellular Probes*, 20(1), 18-20.
- Markljung, E., Braunschweig, M. H., Karlskov-Mortensen, P., Bruun, C. S., Sawera, M., Cho, I. C., Hedebro-Velander, I., Åsa Josell, A., Lundström, K., Seth, Gertrud., Jørgensen, C. B., Fredholm, M., & Andersson, L. (2008). Genome-wide identification of quantitative trait loci in a cross between Hampshire and Landrace II: meat quality traits. *BMC Genetics*, 9(1), 22.
- Markoulatos, P., Sifakas, N., & Moncany, M. (2002). Multiplex polymerase chain reaction: a practical approach. *Journal of Clinical Laboratory Analysis*, (16), 47-51.
- Marson, E. P., Ferraz, J. B. S., Meirelles, F. V., Balieiro, J. C. C., Eler, J. P., Figuerido, L. G. G., & Mourão, G. B. (2005). Genetic characterization of European-Zebu composite bovine using RFLP markers. *Genet. Mol. Res*, 4(3), 496-505.
- Martín, I., García, T., Fajardo, V., Rojas, M., Pegels, N., Hernández, P. E., González, I., & Martín, R. (2009). SYBR-Green real-time PCR approach for the detection and quantification of pig DNA in feedstuffs. *Meat Science*, 82(2), 252-259.

- Martinez, I., & Daniélsdóttir, A. K. (2000). Identification of marine mammal species in food products. *Journal of the Science of Food and Agriculture*, 80(4), 527-533.
- Martinez, I., & Yman, I. M. (1998). Species identification in meat products by RAPD analysis. *Food Research International*, 31(6), 459-466.
- Masternak, M. M., Przybylski, G. K., Smoczkiwicz, P., Plotek, W., Kowalczyk, D., & Nowak, J. S. (2002). Novel competitive PCR methods for quantitation of T-cell receptor delta (TCRD) gene rearrangements. *Journal of Applied Genetics*, 43(2), 235-244.
- Mathew, V. N. (2014). Acceptance on Halal food among non-Muslim consumers. *Procedia-Social and Behavioral Sciences*, 121, 262-271.
- Matsunaga T, Chikuni K, Tanabe R, Muroya S, Shibata K, Yamada J, Shinmura Y (1999). A quick and simple method for the identification of meat species and meat products by PCR assay. *Meat Science* 51(2): 143-148
- Mayr, D., Margesin, R., Schinner, F., & Märk, T. D. (2003). Detection of the spoiling of meat using PTR-MS. *International Journal of Mass Spectrometry*, 223, 229-235.
- McNair, H. M., & Miller, J. M. (2011). *Basic gas chromatography*. John Wiley & Sons.
- Mendoza-Romero, L., Verkaar, E. L., Savelkoul, P. H., Catsburg, A., Aarts, H. J., Buntjer, J. B., & Lenstra, J. A. (2004). Real-time PCR detection of ruminant DNA. *Journal of Food Protection*[®], 67(3), 550-554.
- Meyer, R., Candrian, U., & Lüthy, J. (1993). Detection of pork in heated meat products by the polymerase chain reaction. *Journal of AOAC International*, 77(3), 617-622.
- Michener, R., & Lajtha, K. (Eds.). (2008). *Stable isotopes in ecology and environmental science*. John Wiley & Sons.
- Miller, F. J., Rosenfeldt, F. L., Zhang, C., Linnane, A. W., & Nagley, P. (2003). Precise determination of mitochondrial DNA copy number in human skeletal and cardiac muscle by a PCR-based assay: lack of change of copy number with age. *Nucleic Acids Research*, 31(11), e61-e61.
- Misra, A., & Sinha, S. (1997). Cloning strategies for polymerase chain reaction products. *Current Science*, 73(9), 755-761.

- Moazed, D., & Noller, H. F. (1989). Interaction of tRNA with 23S rRNA in the ribosomal A, P, and E sites. *Cell*, 57(4), 585-597.
- F., Ruiz-Pesini, E., Montoya, J., Roncales, P., López-Pérez, M. J., & Pérez-Martos, A. (2000). Direct and highly species-specific detection of pork meat and fat in meat products by PCR amplification of mitochondrial DNA. *Journal of Agricultural and Food Chemistry*, 48(7), 2829-2832.
- Montowska, M., & Pospiech, E. (2010). Authenticity determination of meat and meat products on the protein and DNA basis. *Food Reviews International*, 27(1), 84-100.
- Moore, J. C., Spink, J., & Lipp, M. (2012). Development and application of a database of food ingredient fraud and economically motivated adulteration from 1980 to 2010. *Journal of Food Science*, 77(4), R118-R126.
- Morales-Jimenez, A. L., Cortés-Ortiz, L., & Di Fiore, A. (2015). Phylogenetic relationships of Mesoamerican spider monkeys (*Ateles geoffroyi*): molecular evidence suggests the need for a revised taxonomy. *Molecular Phylogenetics and Evolution*, 82, 484-494.
- Moran, C. (2011). Molecular genetics. In M. F. Rothschild, & A. Ruvinsky (Eds.), *The genetics of the pig* (pp. 73–100). Cambridge, MA: CAB International.
- Moriuchi, R., Monma, K., Sagi, N., Uno, N., & Kamata, K. (2007). Applicability of quantitative PCR to soy processed foods containing Roundup Ready Soy. *Food Control*, 18(3), 191-195.
- Mousavi, S. M., Khaniki, G. J., Eskandari, S., Rabiei, M., Samiee, S. M., & Mehdizadeh, M. (2015). Applicability of species-specific polymerase chain reaction for fraud identification in raw ground meat commercially sold in Iran. *Journal of Food Composition and Analysis*, 40, 47-51.
- Muldoon, M. T., Onisk, D. V., Brown, M. C., & Stave, J. W. (2004). Targets and methods for the detection of processed animal proteins in animal feedstuffs. *International Journal of Food Science & Technology*, 39(8), 851-861.
- Mullis, K. B., Ferré, F., & Gibbs, R. A. (1994). *The polymerase chain reaction*. Birkhauser Boston Inc.
- Mullis, K. B., Erlich, H. A., Arnheim, N., Horn, G. T., Saiki, R. K., & Scharf, S. J. (1987). U.S. Patent No. 4,683,195. Washington, DC: U.S. Patent and Trademark Office.

- Murgiano, L., D'Alessandro, A., Zolla, L., Valentini, A., & Pariset, L. (2013). Comparison of milk fat globule membrane (MFGM) proteins in milk samples of Chianina and Holstein cattle breeds across three lactation phases through 2D IEF SDS PAGE - a preliminary study. *Food Research International*, 54(1), 1280-1286.
- Murugaiah, C., Noor, Z. M., Mastakim, M., Bilung, L. M., Selamat, J., & Radu, S. (2009). Meat species identification and Halal authentication analysis using mitochondrial DNA. *Meat Science*, 83(1), 57-61.
- Murray, M. G., & Thompson, W. F. (1980). Rapid isolation of high molecular weight plant DNA. *Nucleic Acids Research*, 8(19), 4321-4326.
- Myers, M. J., Yancy, H. F., & Farrell, D. E. (2003). Characterization of a polymerase chain reaction-based approach for the simultaneous detection of multiple animal-derived materials in animal feed. *Journal of Food Protection*[®], 66(6), 1085-1089.
- Nagatsuka, H., Ishiwari, Y., Tsujigiwa, H., Nakano, K., & Nagai, N. (2001). Quantitation of epidermal growth factor receptor gene amplification by competitive polymerase chain reaction in pre-malignant and malignant oral epithelial lesions. *Oral Oncology*, 37(7), 599-604.
- Nakyinsige, K., Man, Y. B. C., & Sazili, A. Q. (2012). Halal authenticity issues in meat and meat products. *Meat Science*, 91(3), 207-214.
- Natonek-Wiśniewska, M., Krzyścin, P., & Piestrzyńska-Kajtoch, A. (2013). The species identification of bovine, porcine, ovine and chicken components in animal meals, feeds and their ingredients, based on COX I analysis and ribosomal DNA sequences. *Food Control*, 34(1), 69-78.
- Nicholls, T. J., & Minczuk, M. (2014). In D-loop: 40years of mitochondrial 7S DNA. *Experimental Gerontology*, 56, 175-181.
- Nielsen, P. E. (2001). Peptide nucleic acid: a versatile tool in genetic diagnostics and molecular biology. *Current Opinion in Biotechnology*, 12(1), 16-20.
- Nurjuliana, M., Man, Y. C., Hashim, D. M., & Mohamed, A. K. S. (2011). Rapid identification of pork for halal authentication using the electronic nose and gas chromatography mass spectrometer with headspace analyzer. *Meat Science*, 88(4), 638-644.
- Nybo K (2013) Primer design. *BioTechniques* 54(5): 249-250

- Okuma, T. A., & Hellberg, R. S. (2015). Identification of meat species in pet foods using a real-time polymerase chain reaction (PCR) assay. *Food Control*, 50, 9-17.
- Olanca, B., Cakirogullari, G. C., Ucar, Y., Kirisik, D., & Kilic, D. (2014). Polychlorinated dioxins, furans (PCDD/Fs), dioxin-like polychlorinated biphenyls (dl-PCBs) and indicator PCBs (ind-PCBs) in egg and egg products in Turkey. *Chemosphere*, 94, 13-19.
- Ortiz, M. C., Sarabia, L., García-Rey, R., & de Castro, M. D. L. (2006). Sensitivity and specificity of PLS-class modelling for five sensory characteristics of dry-cured ham using visible and near infrared spectroscopy. *Analytica Chimica Acta*, 558(1), 125-131.
- Oshima, I., Iwamoto, H., Tabata, S., Ono, Y., Ishibashi, A., Shiba, N., Miyachi, H., Gotoh, T., & Nishimura, S. (2007). Comparative observations of the growth changes of the histochemical properties and collagen architecture of the iliotibialis lateralis muscle from Silky, layer and meat type cockerels. *Animal Science Journal*, 78(5), 546-559.
- Pagliarulo, V., George, B., Beil, S. J., Groshen, S., Laird, P. W., Cai, J., Willey, J., Cote, R., & Datar, R. H. (2004). Sensitivity and reproducibility of standardized-competitive RT-PCR for transcript quantification and its comparison with real time RT-PCR. *Molecular Cancer*, 3(1), 5.
- Parkanyi, V., Ondruska, L., Vasicek, D., & Slamecka, J. (2014). Multilevel D-loop PCR identification of hunting game. *Applied & Translational Genomics*, 3(1), 1-7.
- Payan, C., Ve, N., Crescenzo-Chaigne, B., Be, L., & Pillot, J. (1997). New quantitative assay of hepatitis B and C viruses by competitive PCR using alternative internal sequences. *Journal of Virological Methods*, 65(2), 299-305.
- Pegels, N., González, I., García, T., & Martín, R. (2014). Avian-specific real-time PCR assay for authenticity control in farm animal feeds and pet foods. *Food Chemistry*, 142, 39-47.
- Pegels, N., González, I., Martín, I., Rojas, M., García, T., & Martín, R. (2011). Applicability assessment of a real-time PCR assay for the specific detection of bovine, ovine and caprine material in feedstuffs. *Food Control*, 22(8), 1189-1196.
- Phillips, C. J., Paul, E. A., & Prosser, J. I. (2000). Quantitative analysis of ammonia oxidising bacteria using competitive PCR. *FEMS Microbiology Ecology*, 32(2), 167-175.

- Piegu, B., Guyot, R., Picault, N., Roulin, A., Saniyal, A., Kim, H., Collura, K., Brar, D. S., Jackson, S., Wing, R. A., & Panaud, O. (2006). Doubling genome size without polyploidization: dynamics of retrotransposition-driven genomic expansions in *Oryza australiensis*, a wild relative of rice. *Genome Research*, 16(10), 1262-1269.
- Piña-Vázquez, C., Saavedra, R., & Hérion, P. (2008). A quantitative competitive PCR method to determine the parasite load in the brain of *Toxoplasma gondii*-infected mice. *Parasitology International*, 57(3), 347-353.
- Polz, M. F., & Cavanaugh, C. M. (1998). Bias in template-to-product ratios in multitemplate PCR. *Applied and Environmental Microbiology*, 64(10), 3724-3730.
- Potter, A., Murray, J., Lawson, B., & Graham, S. (2012). Trends in product recalls within the agri-food industry: Empirical evidence from the USA, UK and the Republic of Ireland. *Trends in Food Science & Technology*, 28(2), 77-86.
- Premanandh, J. (2013). Horse meat scandal-A wake-up call for regulatory authorities. *Food Control*, 34(2), 568-569.
- Premanandh, J., Sabbagh, A., & Maruthamuthu, M. (2013). Misdescription of packaged foods: a case study from the United Arab Emirates. *Food Additives & Contaminants: Part A*, 30(12), 2022-2026.
- Qiu, X. Y., Hurt, R. A., Wu, L. Y., Chen, C. H., Tiedje, J. M., & Zhou, J. Z. (2004). Detection and quantification of copper-denitrifying bacteria by quantitative competitive PCR. *Journal of Microbiological Methods*, 59(2), 199-210.
- Quinto, C. A., Tinoco, R., & Hellberg, R. S. (2016). DNA barcoding reveals mislabeling of game meat species on the US commercial market. *Food Control*, 59, 386-392.
- Raeymaekers, L. (1995). A commentary on the practical applications of competitive PCR. *Genome Research*, 5(1), 91-94.
- Rahman, M. M., Ali, M. E., Hamid, S. B. A., Mustafa, S., Hashim, U., & Hanapi, U. K. (2014a). Polymerase chain reaction assay targeting cytochrome b gene for the detection of dog meat adulteration in meatball formulation. *Meat Science*, 97(4), 404-409.
- Rahman, M. M., Ali, M. E., Bhassu, S., Hamid, S. B. A., & Mustafa, S. (2014b). Identification of short-length oligonucleotides biomarker for canine species detection using mitochondrial cytochrome b gene. *Asian Pacific Journal of Tropical Disease*, 4(3), 235.

- Rahmania, H., & Rohman, A. (2015). The employment of FTIR spectroscopy in combination with chemometrics for analysis of rat meat in meatball formulation. *Meat Science*, 100, 301-305.
- Rao, Q., & Hsieh, Y. H. P. (2007). Evaluation of a commercial lateral flow feed test for rapid detection of beef and sheep content in raw and cooked meats. *Meat Science*, 76(3), 489-494.
- Rastogi, G., Dharne, M. S., Walujkar, S., Kumar, A., Patole, M. S., & Shouche, Y. S. (2007). Species identification and authentication of tissues of animal origin using mitochondrial and nuclear markers. *Meat Science*, 76(4), 666-674.
- Ray, A., & Nordén, B. (2000). Peptide nucleic acid (PNA): Its medical and biotechnical applications and promise for the future. *The FASEB Journal*, 14(9), 1041-1060.
- Rehbein, H., Kündiger, R., Yman, I. M., Ferm, M., Etienne, M., Jerome, M., Craig, A., Mackie, I., Jessen, F., Martinez, I., Mendes, I., Smelt, A., Luten, J., Pineiro, C., & Perez-Martin, R. (1999). Species identification of cooked fish by urea isoelectric focusing and sodium dodecylsulfate polyacrylamide gel electrophoresis: a collaborative study. *Food Chemistry*, 67(4), 333-339.
- Rezaian, M. A., & Krake, L. R. (1987). Nucleic acid extraction and virus detection in grapevine. *Journal of virological methods*, 17(3), 277-285.
- Ringkob, T. P., Swartz, D. R., & Greaser, M. L. (2004). Light microscopy and image analysis of thin filament lengths utilizing dual probes on beef, chicken, and rabbit myofibrils. *Journal of Animal Science*, 82(5), 1445-1453.
- Robin, E. D., & Wong, R. (1988). Mitochondrial DNA molecules and virtual number of mitochondria per cell in mammalian cells. *Journal of cellular physiology*, 136(3), 507-513.
- Rodríguez, M. A., García, T., González, I., Hernández, P. E., & Martín, R. (2005). TaqMan real-time PCR for the detection and quantitation of pork in meat mixtures. *Meat Science*, 70(1), 113-120.
- Rojas, M., González, I., García, T., Hernández, P. E., & Martín, R. (2012). Authentication of meat and commercial meat products from common pigeon (*Columba livia*) woodpigeon (*Columba palumbus*) and stock pigeon (*Columba oenas*) using a TaqMan+ real-time PCR assay. *Food Control*, 23(2), 369-376.
- Rojas, M., González, I., Pavón, M. Á., Pegels, N., Hernández, P. E., García, T., & Martín, R. (2011a). Application of a real-time PCR assay for the detection of ostrich

- (*Struthio camelus*) mislabelling in meat products from the retail market. *Food Control*, 22(3), 523-531.
- Rojas, M., González, I., Pavón, M. Á., Pegels, N., Hernández, P. E., García, T., & Martín, R. (2011b). Development of a real-time PCR assay to control the illegal trade of meat from protected capercaillie species (*Tetrao urogallus*). *Forensic Science International*, 210(1), 133-138.
- Rojas, M., González, I., Fajardo, V., Martín, I., Hernández, P. E., García, T., & Martín, R. (2009). Authentication of meats from quail (*Coturnix coturnix*), pheasant (*Phasianus colchicus*), partridge (*Alectoris* spp.), and guinea fowl (*Numida meleagris*) using polymerase chain reaction targeting specific sequences from the mitochondrial 12S rRNA gene. *Food Control*, 20(10), 896-902.
- Rohman, A., Erwanto, Y., & Man, Y. B. C. (2011). Analysis of pork adulteration in beef meatball using Fourier transform infrared (FTIR) spectroscopy. *Meat Science*, 88(1), 91-95.
- Rollinson, D., & Blackwell, J. M. (1999). *Exploring parasite genomes* (Vol. 36). Cambridge University Press.
- Rusman, H., Gerelt, B., Yamamoto, S., Nishiumi, T., & Suzuki, A. (2007). Combined effects of high pressure and heat on shear value and histological characteristics of bovine skeletal muscle. *Asian Australasian Journal of Animal Sciences*, 20(6), 994.
- Sacco, D., Brescia, M. A., Buccolieri, A., & Jambrenghi, A. C. (2005). Geographical origin and breed discrimination of Apulian lamb meat samples by means of analytical and spectroscopic determinations. *Meat Science*, 71(3), 542-548.
- Safdar, M., & Junejo, Y. (2015). Development and validation of fast duplex real-time PCR assays based on SYBER Green fluorescence for detection of bovine and poultry origins in feedstuffs. *Food Chemistry*, 173, 660-664.
- Safdar, M., Junejo, Y., Arman, K., & Abasiyanik, M. F. (2014). A highly sensitive and specific tetraplex PCR assay for soybean, poultry, horse and pork species identification in sausages: Development and validation. *Meat Science*, 98(2), 296-300.
- Sahilah AM, Norhayati Y, Norrakiah AS, Aminah A, Wan Aida WM (2011) Halal authentication of raw meats using PCR amplification of mitchondrial DNA. *International Food Research Journal* 18(4): 1489-1491

- Saini, M., Das, D. K., Dhara, A., Swarup, D., Yadav, M. P., & Gupta, P. K. (2007). Characterisation of peacock (*Pavo cristatus*) mitochondrial 12S rRNA sequence and its use in differentiation from closely related poultry species. *British Poultry Science*, 48(2), 162-166.
- Sambrook, J., & Russell, D. W. (2001). Molecular cloning. A laboratory manual. Third. Cold Spring Harbor Laboratory Press, New York.
- Sarri, C., Stamatis, C., Sarafidou, T., Galara, I., Godosopoulos, V., Kolovos, M., Liakou, C., Tastsoglou, S., & Mamuris, Z. (2014). A new set of 16S rRNA universal primers for identification of animal species. *Food Control*, 43, 35-41.
- Sawyer, J., Wood, C., Shanahan, D., Gout, S., & McDowell, D. (2003). Real-time PCR for quantitative meat species testing. *Food Control*, 14(8), 579-583.
- Sasazaki, S., Mutoh, H., Tsurifune, K., & Mannen, H. (2007). Development of DNA markers for discrimination between domestic and imported beef. *Meat Science*, 77(2), 161-166.
- Schanke, J. T. (1997). Sequence inversion by Flip-PCR. In *PCR Cloning Protocols* (pp. 203-208). Humana Press.
- Schmittgen, T. D., & Livak, K. J. (2008). Analyzing real-time PCR data by the comparative CT method. *Nature Protocols*, 3(6), 1101-1108.
- Sentandreu, M. Á., & Sentandreu, E. (2014). Authenticity of meat products: Tools against fraud. *Food Research* Sekhavati, M. H., Mesgaran, M. D., Nassiri, M. R., Mohammadabadi, T., Rezaii, F., & Maleki, A. F. (2009). Development and use of quantitative competitive PCR assays for relative quantifying rumen anaerobic fungal populations in both in vitro and in vivo systems. *Mycological Research*, 113(10), 1146-1153.
- Sevane, N., Crespo, I., Cañón, J., & Dunner, S. (2011). A Primer-Extension Assay for simultaneous use in cattle Genotype Assisted Selection, parentage and traceability analysis. *Livestock Science*, 137(1), 141-150
- Shang, Y., Zhu, P., Xu, W., Guo, T., Tian, W., Luo, Y., & Huang, K. (2013). Single universal primer multiplex ligation-dependent probe amplification with sequencing gel electrophoresis analysis. *Analytical Biochemistry*, 443(2), 243-248.

- Sharma, D., Rao, K. A., Singh, H. P., & Totey, S. M. (1998). Randomly amplified polymorphic DNA (RAPD) for evaluating genetic relationships among varieties of guinea fowl. *Genetic analysis: biomolecular engineering*, 14(4), 125-128.
- Shintu, L., Caldarelli, S., & Franke, B. M. (2007). Pre-selection of potential molecular markers for the geographic origin of dried beef by HR-MAS NMR spectroscopy. *Meat Science*, 76(4), 700-707.
- Siciliano, C., Belsito, E., De Marco, R., Di Gioia, M. L., Leggio, A., & Liguori, A. (2013). Quantitative determination of fatty acid chain composition in pork meat products by high resolution 1 H NMR spectroscopy. *Food Chemistry*, 136(2), 546-554.
- Siesler, H. W., Ozaki, Y., Kawata, S., & Heise, H. M. (Eds.). (2008). *Near-infrared spectroscopy: principles, instruments, applications*. John Wiley & Sons.
- Singh, U., Deb, R., Alyethodi, R. R., Alex, R., Kumar, S., Chakraborty, S., Dhama, K., & Sharma, A. (2014). Molecular markers and their applications in cattle genetic research: A review. *Biomarkers and Genomic Medicine*, 6(2), 49-58.
- Singhal, H., Ren, Y. R., & Kern, S. E. (2010). Improved DNA electrophoresis in conditions favoring polyborates and lewis acid complexation. *PLOS one*, 5(6), e11318.
- Sipos, R., Székely, A. J., Palatinszky, M., Révész, S., Márialigeti, K., & Nikolausz, M. (2007). Effect of primer mismatch, annealing temperature and PCR cycle number on 16S rRNA gene-targeting bacterial community analysis. *FEMS Microbiology Ecology*, 60(2), 341-350.
- Skibinski, D. O., Gallagher, C., & Beynon, C. M. (1994). Sex-limited mitochondrial DNA transmission in the marine mussel *Mytilus edulis*. *Genetics*, 138(3), 801-809.
- Smith, D. S., & Maxwell, P. W. (2007). Use of quantitative PCR to evaluate several methods for extracting DNA from corn flour and cornstarch. *Food Control*, 18(3), 236-242.
- Snow, E. C. (1985). Insulin and growth hormone function as minor growth factors that potentiate lymphocyte activation. *Journal of immunology (Baltimore, Md.: 1950)*, 135(2 Suppl), 776s-778s.

- Soares, S., Amaral, J. S., Oliveira, M. B. P., & Mafra, I. (2013). A SYBR Green real-time PCR assay to detect and quantify pork meat in processed poultry meat products. *Meat Science*, 94(1), 115-120.
- Soares, S., Joana, S., Isabel, M., Mafra, M., Beatriz, P.P., & Oliveira (2010). Quantitative detection of poultry meat adulteration with pork by a duplex PCR Assay. *Meat Science*. 85(3): 531-536.
- Spink, J., & Moyer, D. C. (2011). Backgrounder: defining the public health threat of food fraud. *Minneapolis, Minnesota: National Center for Food Protection and Defense*.
- Strasburg, G. M., & Chiang, W. (2009). Pale, soft, exudative turkey-The role of ryanodine receptor variation in meat quality. *Poultry Science*, 88(7), 1497-1505.
- Sripairoj, K., Klinbu-nga, S., Kamonrat, W., & Na-Nakorn, U. (2010). Species identification of four economically important Pangasiid catfishes and closely related species using SSCP markers. *Aquaculture*, 308, S47-S50.
- Stuart, B. (2005). *Infrared spectroscopy*. John Wiley & Sons, Inc.
- Sulijoadikusumo, I., Horikoshi, N., & Usheva, A. (2001). Another function for the mitochondrial ribosomal RNA: protein folding. *Biochemistry*, 40(38), 11559-11564.
- Sumar, S., & Ismail, H. (1995). Adulteration of foods-past and present. *Nutrition & Food Science*, 95(4), 11-15.
- Surowiec, I., Fraser, P. D., Patel, R., Halket, J., & Bramley, P. M. (2011). Metabolomic approach for the detection of mechanically recovered meat in food products. *Food Chemistry*, 125(4), 1468-1475.
- Sven, P., Daniel, N., Anja, B., Tatjana, Y., Nicole, K., Stefanie, M., & Barbara, S. (2009). Rapid identification of beta-hemolytic Streptococci by fluorescence in situ hybridization (FISH). *International Journal of Medical Microbiology*. 299(6): 421-426
- Taft, R. J., Pheasant, M., & Mattick, J. S. (2007). The relationship between non-protein-coding DNA and eukaryotic complexity. *Bioessays*, 29(3), 288-299.
- Tartaglia, M., Saulle, E., Pestalozza, S., Morelli, L., Antonucci, G., & Battaglia, P. A. (1998). Detection of bovine mitochondrial DNA in ruminant feeds: a molecular

- approach to test for the presence of bovine-derived materials. *Journal of Food Protection*[®], 61(5), 513-518.
- Tähkääpää, S., Maijala, R., Korkeala, H., & Nevas, M. (2015). Patterns of food frauds and adulterations reported in the EU rapid alert system for food and feed and in Finland. *Food Control*, 47, 175-184.
- Teletchea, F., Maudet, C., & Hänni, C. (2005). Food and forensic molecular identification: update and challenges. *Trends in Biotechnology*, 23(7), 359-366.
- Thurnheer, T., Gmür, R., & Guggenheim B. (2004). Multiplex FISH analysis of a six-species bacterial biofilm. *Journal of Microbiological Methods*, 56(1): 37-47.
- Tisza, Á., Csikós, Á., Simon, Á., Gulyás, G., Jávör, A., & Czeglédi, L. (2016). Identification of poultry species using polymerase chain reaction-single strand conformation polymorphism (PCR-SSCP) and capillary electrophoresis-single strand conformation polymorphism (CE-SSCP) methods. *Food Control*, 59, 430-438.
- Toyoda, A., Nakajo, M., Kawachi, H., Matsui, T., & Yano, A. (2004). PCR detection of bovine mitochondrial DNA derived from meat and bone meal in feed. *Journal of Food Protection*[®], 67(12), 2829-2832.
- Tremlová, B., Sarha, P., Pospiech, M., Buchtová, H., & Randulová, Z. (2006). Histological analysis of different kinds of mechanically recovered meat. *Archiv fur Lebensmittelhygiene*, 57(3), 85.
- Vallejo-Cordoba, B., Rodríguez-Ramírez, R., & González-Córdova, A. F. (2010). Capillary electrophoresis for bovine and ostrich meat characterisation. *Food Chemistry*, 120(1), 304-307.
- Van den Bogert, C., De Vries, H., Holtrop, M., Muus, P., Dekker, H. L., Van Galen, M. J., Bolhuis, P. A., & Taanman, J. W. (1993). Regulation of the expression of mitochondrial proteins: relationship between mtDNA copy number and cytochrome-c oxidase activity in human cells and tissues. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1144(2), 177-183.
- Vela, J., Vitorica, J., & Ruano, D. (2001). Rapid PCR-mediated synthesis of competitor molecules for accurate quantification of β 2 GABA A receptor subunit mRNA. *Brain Research Protocols*, 8(3), 184-190.

- Vences, M., Thomas, M., Van der Meijden, A., Chiari, Y., & Vieites, D. R. (2005). Comparative performance of the 16S rRNA gene in DNA barcoding of amphibians. *Frontiers in Zoology*, 2(1), 5.
- Verkaar, E. L. C., Nijman, I. J., Boutaga, K., & Lenstra, J. A. (2002). Differentiation of cattle species in beef by PCR-RFLP of mitochondrial and satellite DNA. *Meat Science*, 60(4), 365-369.
- Wadeh, H., Alsarakibi, M., & Li, G. (2010). Analysis of genetic variability within *Argulus japonicus* from representatives of Africa, Middle East, and Asia revealed by sequences of three mitochondrial DNA genes. *Parasitology Research*, 107(3), 547-553.
- Walker, J. A., Hughes, D. A., Hedges, D. J., Anders, B. A., Laborde, M. E., Shewale, J., Sinha, S. K., & Batzer, M. A. (2004). Quantitative PCR for DNA identification based on genome-specific interspersed repetitive elements. *Genomics*, 83(3), 518-527.
- Walker, J. A., Hughes, D. A., Anders, B. A., Shewale, J., Sinha, S. K., & Batzera, M. A. (2003). Quantitative intra-short interspersed element PCR for species-specific DNA identification. *Analytical Biochemistry*, 316, 259-269.
- Wang, Y. J., Lau, K. K., & Lau, F. L. (2015). Clenbuterol food poisoning from snake meat consumption: an outbreak of 13 cases. *Hong Kong Journal of Emergency Medicine*, 22(1), 46.
- Wang, Q., Liu, Q., & Li, B. J. (2004). Simultaneous detection of seven mutations with seven forward primers and one common reverse primer in a single PCR step. *Journal of Biochemical and Biophysical Methods*, 58(2), 153-157.
- Welsh, J., & McClelland, M. (1990). Fingerprinting genomes using PCR with arbitrary primers. *Nucleic Acids Research*, 18(24), 7213-7218.
- Wen D, Zhang C (2012) Universal Multiplex PCR: a novel method of simultaneous amplification of multiple DNA fragments. *Plant Methods*, 8(1): 1. 2-9
- Weiss, H., Friedrich, T., Hofhaus, G., & Preis, D. (1991). The respiratory-chain NADH dehydrogenase (Complex I) of mitochondria. *European Journal of Biochemistry*, 197(3), 563-576.
- Weissenberger, M., Reichert, W., & Mattern, R. (2011). A Multiplex PCR assay to differentiate between dog and red fox. *Forensic Science International: Genetics*, 5(5), 411-414.

- Wikstrom, M., Krab, K., & Saraste, M. (1981). Proton-translocating cytochrome complexes. *Annual Review of Biochemistry*, 50(1), 623-655.
- Wilson, K. and Walker, J. 2005. Principles and Techniques of Biochemistry and Molecular Biology. Cambridge: Cambridge University Press.
- Winkler, J. (2015). High levels of dioxin-like PCBs found in organic-farmed eggs caused by coating materials of asbestos-cement fiber plates: A case study. *Environment International*, 80, 72-78.
- Winterø, A. K., Thomsen, P. D., & Davies, W. (1990). A comparison of DNA-hybridization, immunodiffusion, countercurrent immunoelectrophoresis and isoelectric focusing for detecting the admixture of pork to beef. *Meat Science*, 27(1), 75-85.
- Wolf, C., & Lüthy, J. (2001). Quantitative competitive (QC) PCR for quantification of porcine DNA. *Meat Science*, 57(2), 161-168.
- Wolf, C., Rentsch, J., & Hübner, P. (1999). PCR-RFLP analysis of mitochondrial DNA: a reliable method for species identification. *Journal of Agricultural and Food Chemistry*, 47(4), 1350-1355.
- Woolfe, M., & Primrose, S. (2004). Food forensics: using DNA technology to combat misdescription and fraud. *Trends in Biotechnology*, 22(5): 222-6
- Wu, S., Xiong, J., & Yu, Y. (2015). Taxonomic Resolutions Based on 18S rRNA Genes: A Case Study of Subclass Copepoda. *PloS one*, 10(6), e0131498.
- Wu, X. B., Wang, Y. Q., Zhou, K. Y., Zhu, W. Q., Nie, J. S., Wang, C. L., & Xie, W. S. (2002). Genetic variation in captive population of Chinese alligator, *Alligator sinensis*, revealed by random amplified polymorphic DNA (RAPD). *Biological Conservation*, 106(3), 435-441.
- Wu, Q., Chen, M., Buchwald, M., and Philips, R.A. (1995). A Simple and Rapid Method for Isolation of High Quality Genomic DNA from Animal Tissues. *Nucleic Acids Research*, 23(24), 5087-5088.
- Xu, W., Zhai, Z., Huang, K., Zhang, N., Yuan, Y., Shang, Y., & Luo, Y. (2012a). A novel universal primer-multiplex-PCR method with sequencing gel electrophoresis analysis. *PLoS One*, 7(1), 1.

- Xu, L., Cai, C. B., Cui, H. F., Ye, Z. H., & Yu, X. P. (2012b). Rapid discrimination of pork in Halal and non-Halal Chinese ham sausages by Fourier transform infrared (FTIR) spectroscopy and chemometrics. *Meat Science*, 92(4), 506-510.
- Xu, W., Bai, W., Luo, Y., Yuan, Y., Zhang, W., Guo, X., & Huang, K. (2008). A novel common single primer multiplex polymerase chain reaction (CSP-M-PCR) method for the identification of animal species in minced meat. *Journal of the Science of Food and Agriculture*, 88(15), 2631-2637.
- Yan, H., Xu, D., Meng, H., Shi, L., & Li, L. (2014). Food poisoning by clenbuterol in China. *Quality Assurance and Safety of Crops & Foods*, 7(1), 27-35.
- Yang, L., Tan, Z., Wang, D., Xue, L., Guan, M. X., Huang, T., & Li, R. (2014). Species identification through mitochondrial rRNA genetic analysis. *Scientific Reports*, 4.
- Yeboah, G., & Maynard, L. J. (2004, August). The impact of BSE, FMD, and US export promotion expenditures on Japanese meat demand. In *annual meeting of AAEP, Denver CO*
- Yilmaz, M. T., Kesmen, Z., Baykal, B., Sagdic, O., Kulen, O., Kacar, O., Yetim, H., & Baykal, A. T. (2013). A novel method to differentiate bovine and porcine gelatins in food products: NanoUPLC-ESI-Q-TOF-MS E based data independent acquisition technique to detect marker peptides in gelatin. *Food Chemistry*, 141(3), 2450-2458.
- Zakut, R., Shani, M., Givol, D., Neuman, S., Yaffe, D., & Nudel, U. (1982). Nucleotide sequence of the rat skeletal muscle actin gene.
- Zhang, C. (2013). Semi-nested multiplex PCR enhanced method sensitivity of species detection in further-processed meats. *Food Control*, 31(2), 326-330.
- Zhang, C. L., Fowler, M. R., Scott, N. W., Lawson, G., & Slater, A. (2007). A TaqMan real-time PCR system for the identification and quantification of bovine DNA in meats, milks and cheeses. *Food Control*, 18(9), 1149-1158.
- Zidani, S., Ferchichi, A., & Chaieb, M. (2005). Genomic DNA extraction method from pearl millet (*Pennisetum glaucum*) leaves. *African Journal of Biotechnology*, 4(8), 862-866.
- Zilhada, Z. (2013). Protein Profiles of Beef (*Bos indicus*), Pork (*Sus domesticus*), and Sausages By Using SDS-PAGE (Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis) Method. *Journal of Food and Pharmaceutical Sciences*, 2(2).

- Zimmermann, K., & Mannhalter, J. W. (1996). Technical aspects of quantitative competitive PCR. *BioTechniques*, 21(2), 268-279.
- Zhang, X., Li, K., Wu, S., Shuai, J., & Fang, W. (2015). Peptide nucleic acid fluorescence in-situ hybridization for identification of *Vibrio* spp. in aquatic products and environments. *International Journal of Food Microbiology*, 206, 39-44.
- Zhang, Z., Yang, X., Meng, L., Liu, F., Shen, C., & Yang, W. (2009). Enhanced amplification of GC-rich DNA with two organic reagents. *Biotechniques*, 47(3), 775-779.
- Zhou, H., & Hickford, J. G. H. (2008). Allelic polymorphism of the caprine calpastatin (CAST) gene identified by PCR–SSCP. *Meat Science*, 79(2), 403-405.
- ZhongMeng, L., LiangJuan, Z., YongFang, W., & HongWei, Z. (2015). Progress of molecular identification techniques used in meat and meat products. *Journal of Food Safety and Quality*, 6(2), 405-409.
- Zukál, E., & Körmeny, L. (2007). On calculation of ‘meat content’ according to the quantitative ingredient declarations (QUID). *Journal of Food Engineering*, 78(2), 614-621.

BIODATA OF STUDENT

Umami Kalthum binti Hanapi was born on 20th March 1975 in Johor Bahru, Johor, Malaysia. She entered a primary school at Sekolah Kebangsaan Pulau Sebatang, Pontian, Johor and then continued her education at Sekolah Menengah LKTP Kahang Timur, Kluang, Johor and Sekolah Menengah Perempuan Sri Aman, Petaling Jaya. Upon finishing her matriculation at Universiti Malaya, she started her undergraduate education at the same university and graduated in 1999 with Bachelor of Science (Hons.), majoring in Biochemistry. In 2008, she received her Master of Science degree in Plant Genetic Engineering and Molecular Biology from Universiti Putra Malaysia. She has been working as a research assistant at Malaysian Agricultural Research and Development Institute (1999-2001), Universiti Putra Malaysia (2001-2005), UKM Medical Molecular Biology Institute (2006-2006), as a biotechnologist at Felda Agricultural Services Sdn. Bhd. (2006-2009) and as a lecturer at Universiti Selangor (2009-2011). Currently, she is pursuing her Ph.D. programme in Halal Product Science at Halal Product Research Institute UPM. Her study is supported by research grants received from the Ministry of Science, Technology and Innovation (ScienceFund) and Universiti Putra Malaysia (Research University Grant Scheme) and scholarship from the Ministry of Education (MyPhD).

LIST OF PUBLICATIONS

This thesis dissertation resulted in the following publications:

Publications

Hanapi, U. K., Desa M. N. M.*, Ismail, A., & Mustafa, S (2014). A Higher Sensitivity and Efficiency of Common Primer Multiplex PCR Assay in Identification of Meat Origin Using NADH Dehydrogenase Subunit 4 Gene. *Journal of Food Science and Technology*, 1-10. (JIF 2014 = 2.203).

Hanapi, U. K., Esa, M. N., Desa, M. N. M.*, Ismail, A., & Mustafa. PCR-based dual detection for differentiating chicken from avian meat in processed meat products *Food Analytical Research*, submitted (JIF 2014/2015 = 1.956).

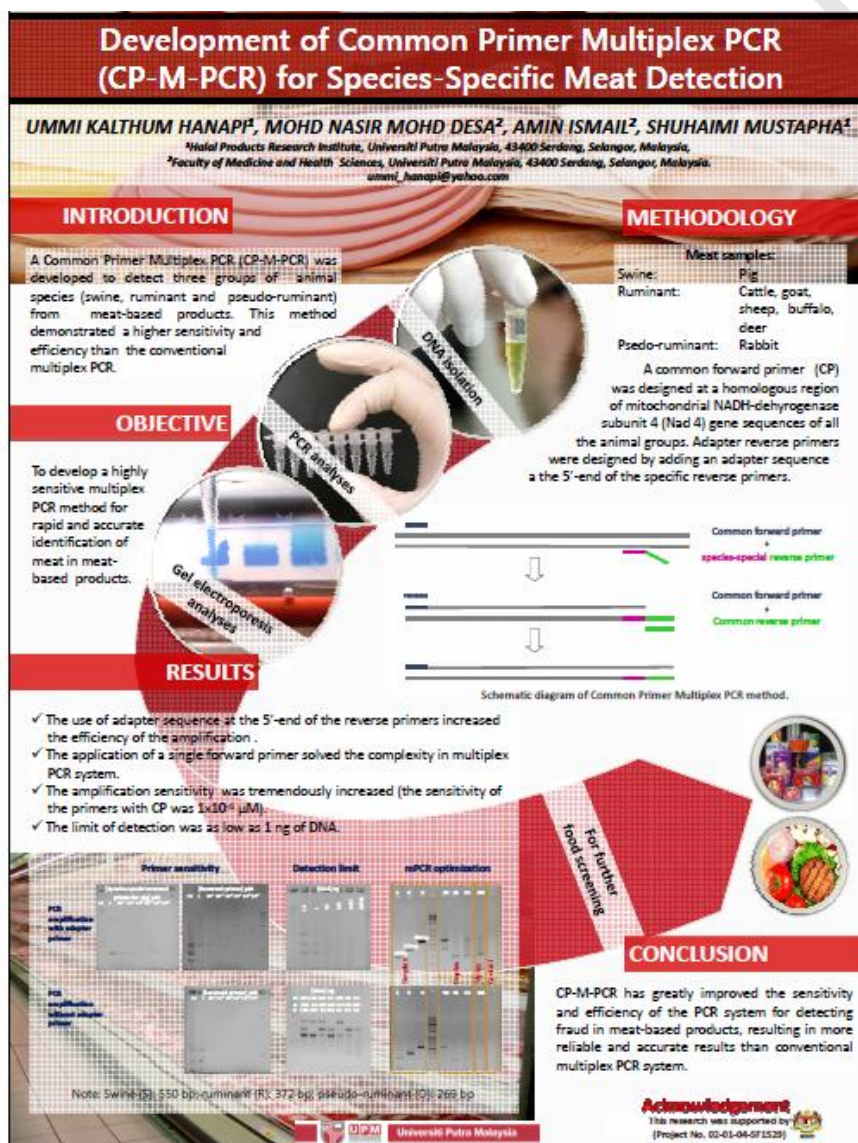
Hanapi, U. K., Esa, M. N., Desa, M. N. M.*, Ismail, A., & Mustafa. Quantitative Determination of Avian DNA in Meat Products Using a Quantitative Competitive PCR. *International Food Research Journal*, submitted.

Proceedings/ Conferences

Hanapi U. K., Desa, M. N. M.*, Ismail A., Mustafa S. Rapid and reliable identification of meat origin in meat products using CP-M-PCR [No. ID025m, ORAL]. In: *Malaysian International Halal Research & Education*. Putrajaya: Halal Products Research Institute, Universiti Putra Malaysia, December 2-4, 2014.



Ummi, K. H., Desa, M. N. M*, Ismail A. & Mustafa S. Development of common primer multiplex PCR (CP-M-PCR) for species-specific meat detection. [Abstract no. PP39, p.95; POSTER]. In: *1st International Conference on Molecular Diagnosis and Biomarker Discovery*. Penang: Institute of Research in Molecular Medicine, Universiti Sains Malaysia, October 23-25, 2013. [Abstract Published in; Ummi, K. H., Desa, M. N. M*, Ismail, A., & Mustafa (2014), Development of Common Primer Multiplex PCR (CP-M-PCR) for species-specific meat detection. *Asian Pacific Journal of Tropical Disease*, 4(3), 250.]



Award

Gold Medal

Research Title: “A High Sensitivity and Efficiency of Common Primer Multiplex PCR Assay in Identification of Meat Origin Using NADH Dehydrogenase Subunit 4 Gene”.

Organizer: RMC, Universiti Putra Malaysia.

Pameran Rekacipta dan Inovasi (PRPI) 2014, Universiti Putra Malaysia, Serdang, Selangor.





UNIVERSITI PUTRA MALAYSIA

STATUS CONFIRMATION FOR THESIS / PROJECT REPORT AND COPYRIGHT

ACADEMIC SESSION : _____

TITLE OF THESIS / PROJECT REPORT :

SENSITIVE AND RAPID MODELS OF PCR-BASED METHODOLOGIES FOR ADMIXTURE
DETECTION AND QUANTIFICATION OF DOMESTIC MEATS

NAME OF STUDENT : UMMI KALTHUM HANAPI

I acknowledge that the copyright and other intellectual property in the thesis/project report belonged to Universiti Putra Malaysia and I agree to allow this thesis/project report to be placed at the library under the following terms:

1. This thesis/project report is the property of Universiti Putra Malaysia.
2. The library of Universiti Putra Malaysia has the right to make copies for educational purposes only.
3. The library of Universiti Putra Malaysia is allowed to make copies of this thesis for academic exchange.

I declare that this thesis is classified as :

*Please tick (✓)

☐

CONFIDENTIAL

(Contain confidential information under Official Secret Act 1972).

☐

RESTRICTED

(Contains restricted information as specified by the organization/institution where research was done).

☐

OPEN ACCESS

I agree that my thesis/project report to be published as hard copy or online open access.

This thesis is submitted for :

☐

PATENT

Embargo from _____ until _____
(date) (date)

Approved by:

(Signature of Student)
New IC No/ Passport No.:

Date :

(Signature of Chairman of Supervisory Committee)
Name:

Date :

[Note : If the thesis is **CONFIDENTIAL** or **RESTRICTED**, please attach with the letter from the organization/institution with period and reasons for confidentially or restricted.]