

UNIVERSITI PUTRA MALAYSIA

CHARACTERIZATION AND IMMUNOMODULATION OF BIFIDOBACTERIUM PSEUDOCATENULATUM G4 AND BIFIDOBACTERIUM LONGUM BB536 IN VITRO

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By
FATEMEH AHMADI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the requirement for the Degree of Master of Science

February 2015

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DEDICATION

I dedicate this thesis to my mother Marziyeh and My father Mohsen who have raised me to be the person who I am today. Thank you for all the unconditional love, guidance and support that you always given me.

To my a dearest brothers Ali who has given lots of love and support to continuance my education

To my beloved husbands, Behnam Shafiei Astani, for his immense support, patience, and encourgment





CHARACTERIZATION AND IMMUNOMODULATION OF BIFIDOBACTERIUM PSEUDOCATENULATUM G4 AND BIFIDOBACTERIUM LONGUM BB536 IN VITRO

By **FATEMEH AHMADY**

February 2015

Chairman: Professor Dato' Mohd Yazid Abdul Manap, PhD

Faculty: Food Science and Technology

Probiotics bacteria have fundamental and important roles in the human immune system; hence they are also called "good bacteria". Probiotic bacteria such as *Bifidobacterium pseudocatenulatum* G4 and *Bifidobacterium longum* BB536 are natural members of the human intestinal tract that have immonodulatory activity. They stimulate dendritic cells, peyer's patches, lymphocytes such as B- lymphocytes, NK cells and T- cells. Thus, they inhibit the growth of pathogenic bacteria. In the first part of this study, the effect of colonic pH (5.7, 6.4 and 6.9) and the different concentration of hydrogen peroxide (H₂O₂) (50, and 100 µg/l) on bifidobacteria growth were evaluated.

Modulation of host immunity is one of the proposed benefits of the consumption of probiotics such as *Bifidobacterium pseudocatenulatum* G4 and *Bifidobacterium longum* BB536. On the other hand, activity of the bacteria can be changed using different pH and high concentration of H₂O₂ which will have an inhibitory effect on bacteria growth. Also, Preparation of bifidobacteria with different method such as heat treatment and sonication can enable the active substances to be accessible to relevant lymphoid cells. In another word, exposure of the immunopotentiators may have occurred as the result of leakage of intracellular components or disruption of huge complex of cell wall materials.

Two strains of *B. pseudocatenulatum* G4 and *B. longum* BB536 were examined for their survival in colonic pH. The survival rate of bifidobacteria in colonic pH did not vary among isolates. *B. pseudocatenulatum* G4 and *B. longum* BB536 maintain high viability of $> 10^8$ cfu/ml during 3 hours of incubation in the different pH solution. Hence, bifidobacteria can grow in different colonic pH (5.7, 6.4 and 6.9) with no reduction in the number of living bacterial cells as compared to the control group. The survival rate of bifidobacteria at $\rm H_2O_2$ concentrations varied during 36 hours. The growth of two strains of bifidobacteria decreased significantly (P < 0.05) at $100~\mu g/ml$ $\rm H_2O_2$ during 36 hours. On the other hand, these results revealed that bifidobacteria growth can be reduced by different concentration of $\rm H_2O_2$ (50 and 100 $\rm \mu g/l$) during 36 hours. High concentration of $\rm H_2O_2$ causes the reduction of immune responses and the mediation of host defence. One of the most important criteria of the bifidobacteria is the ability to increase the immune system in order to increase the interleukins and

cytokines. Live and heat-treated bifidobacteria were used in this study to measure cytokine production such as IL-12, IL-6, IL-4, IFN- γ and IgA after isolation of mice Peyer's patches. Mice that were orally administered with heat-treated bacteria (*B. pseudocatenulatum* G4 and *B. longum* BB536) for seven consecutive days showed an increase in immunomodulatory responses because of the production of heat shock protein during heating process. So the bacteria can induce inflammatory cytokine by activation bacterial mechanisms by pattern-recognition receptors such as toll like receptors (TLRs) Therefore, the bifidobacteria induce more inflammatory cytokines but there are no any significant differences between these two strains of bifidobacteria. The level of IFN- γ significant increased more than other cytokines therefore, it has a stronger immunomodulatory effect (p < 0.05).

This study also showed that live and heat-treated bifidobacteria might induce different immune responses by increasing the level of cytokines and immunomodulatory responses through the Peyer's patches (PPs) activation. It is hypothesised that heat-treated bacteria increase the production of cytokines and immunomodulatory effect; nonetheless, comparative studies on the immunological properties that support the selection of strains of the same species for specific health benefits are needed in future.

PENCIRIAN DAN IMMUNOMODULATION OF BIFIDOBACTERIUM PSEUDOCATENULATUM G4 DAN BIFIDOBACTERIUM LONGUM BB536 IN VITRO

Oleh **FATEMEH AHMADY**

Februari 2015

Pengerusi: Professor Dato' Mohd Yazid Abdul Manap, PhD Fakulti: Sains dan Teknologi Makanan

Bakteria probiotik mempunyai peranan asas dan penting dalam sistem imun manusia; oleh itu bakteria itu juga dikenali sebagai "bakteria baik". Bakteria probiotik seperti *Bifidobakterium pseudocatenulatum* G4 dan *Bifidobakterium longum* BB536 terdapat secara semula jadi dalam saluran usus manusia yang mempunyai aktiviti penyesuaian imunologi. Bakteria tersebut merangsang sel-sel dendrit, tompok Peyer, limfosit seperti limfosit B-sel-sel NK dan T. Oleh itu, bakteria tersebut menghalang pertumbuhan bakteria patogen. Dalam bahagian pertama kajian ini, kesan pH kolon (5.7, 6.4 dan 6.9) dan kepekatan hidrogen peroksida (H₂O₂) yang berbeza (50, dan 100 μg/l) pada pertumbuhan bifidobakteria yang telah dinilai.

Modulasi imuniti perumah adalah salah satu manfaat cadangan penggunaan probiotik seperti *Bifidobakterium pseudocatenulatum* G4 dan *Bifidobakterium longum* BB536. Sebaliknya, aktiviti bakteria boleh ditukar dengan menggunakan pH yang berbeza dan H₂O₂ berkepekatan tinggi yang akan mempunyai kesan yang merencat pertumbuhan bakteria. Selain itu, penyediaan bifidobakteria dengan kaedah yang berbeza seperti rawatan haba dan sonikasi membolehkan sel-sel limfoid yang berkaitan untuk mengakses bahan-bahan aktif . Dalam erti kata lain, pendedahan imunopotensiator mungkin telah berlaku akibat kebocoran komponen intrasel atau gangguan kompleks besar bagi bahan dinding sel.

Dua strain *B. pseudocatenulatum* G4 dan *B. longum* BB536 telah diperiksa untuk kelangsungan hidup bakteria tersebut dalam pH kolon. Kadar kelangsungan hidup bifidobakteria dalam pH kolon tidak berbeza antara asingan. *B. pseudocatenulatum* G4 dan *B. longum* BB536 mengekalkan kebolehhidupan yang tinggi > 10^8 cfu/ml selama 3 jam pengeraman dalam larutan pH yang berbeza. Oleh itu, *bifidobakteria* boleh tumbuh dalam pH kolon yang berbeza (5.7, 6.4 dan 6.9) tanpa pengurangan dalam bilangan selsel hidup bakteria berbanding dengan kumpulan kawalan. Kadar kelangsungan hidup *bifidobakteria* pada kepekatan H_2O_2 berubah dalam masa 36 jam. Pertumbuhan dua strain *bifidobakteria* menurun dengan ketara (P < 0.05) pada 100 µg/ml H_2O_2 semasa tempoh 36 jam. Sebaliknya, keputusan ini menunjukkan bahawa pertumbuhan *bifidobakteria* boleh dikurangkan dengan kepekatan yang berbeza H_2O_2 (50 dan 100 µg/l) semasa tempoh 36 jam. Kepekatan tinggi H_2O_2 menyebabkan pengurangan tindak balas imun dan pengantaraan pertahanan perumah. Salah satu kriteria yang paling

penting bagi *bifidobakteria* adalah keupayaan untuk meningkatkan sistem imun untuk meningkatkan interleukin dan sitokin. *Bifidobakteria* hidup dan terawat haba telah digunakan dalam kajian ini untuk mengukur pengeluaran sitokin seperti IL-12, IL-6, IL-4, IFN-γ dan IgA selepas pengasingan tikus tompok Peyer. Tikus yang diberi makan dengan bakteria terawat haba (*B. pseudocatenulatum* G4 dan *B. longum* BB536) selama tujuh hari berturut-turut menunjukkan peningkatan dalam tindak balas penyesuaian imunologi disebabkan oleh penghasilan protein kejutan haba semasa proses pemanasan. Oleh itu, bakteria boleh menyebabkan radang sitokin dengan mekanisme pengaktifan bakteria oleh reseptor pengiktirafan corak iaitu tol seperti reseptor (TLR). Oleh itu, bifidobakteria mendorong lebih radang sitokin tetapi tidak terdapat apa-apa perbezaan yang ketara antara kedua-dua strain *bifidobakteria*. Tahap IFN-γ meningkat dengan ketara lebih daripada sitokin lain. Oleh itu, ia mempunyai kesan penyesuaian imunologi yang lebih kukuh (p < 0.05).

Kajian ini juga menunjukkan bahawa bifidobakteria hidup dan terawat haba mungkin menyebabkan tindak balas imun yang berbeza dengan meningkatkan tahap sitokin dan tindak balas penyesuaian imunologi melalui tompok pengaktifan Peyer (PP) ini. Ia dihipotesiskan bahawa bakteria terawat haba meningkatkan pengeluaran sitokin dan kesan penyesuaian imunologi; namun begitu, kajian perbandingan ke atas sifat imunologi yang menyokong pemilihan strain daripada spesies yang sama untuk manfaat kesihatan tertentu diperlukan pada masa akan datang.

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I certify that a Thesis Examination Committee has met on ---- to conduct the final examination of Fatemeh Ahmadi on his thesis entitled "Characterization and Immunomodulation of *Bifidobacterium Pseudocatenulatum* G4 and *Bifidobacterium Longum* Bb536 *In Vitro*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Master of Science.

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TABLE OF CONTENT

APPRO DECLA TABLE LIST O LIST O LIST O	AK OWLEDGEMENTS	v x xii xiii xv xvi
CHAPT	ER	
1	INTRODUCTION	1
	1.1 Study Background	1
	1.2 Prob <mark>lem Statements</mark>	2 2
	1.3 Significance of the Present Study	2
	1.4 General Objective	3
	1.5 Specific Objectives	3
2	LITERATURE REVIEW	5
	2.1 History of Probiotic	5
	2.2 Probiotics and Immunomodulation	5
	2.3 Major Actors of Main Immune System	6
	2.3.1 Innate Immune System	6
	2.3.2 Adaptive Immune System	6
	2.4 T-helper Function	7
	2.5 Probiotics and Immune System	8
	2.5.1 Special Effects of Prebiotics and Probiotics on Allergy	8
	2.5.2 Inflammation	8
	2.6 Effect of Gut Flora in Health and Disease	9
	2.6.1 Composition of the Flora	9
	2.6.2 Metabolic Function of Gut Microbiota	9
	2.7 Immune System and Defence against Microorganisms	9
	2.8 Organized Lymphoid Tissue	10
	2.9 Secretory IgA	10
	2.10 Acid and H ₂ O ₂ Tolerance	10
	2.11 Bifidobacteria and Their Effect on Human Health	11
	2.11.1 Bifidobacterium longum BB536	11
	2.11.2 Bifidobacterium pseudocatenlatum G4	11
3	MATERIALS AND METHODS	13
	3.1 Preparation of Bifidobacteria	13
	3.2 Experimental design	13
	3.3 The Growth of Bifidobacteria	13
	3.3.1 Gram staining	14
	3.3.2 Preparation of Different pH Solutions	14

	3.3.3 Enumeration of <i>B. pseudocatenulatum</i> G4 and <i>B. longum</i>	
	BB536 in Different pH Solutions	14
	3.4 Effect of Different H ₂ O ₂ Concentration on Bifidobacteria Growth	15
	3.5 Effect of Different Hydrogen Peroxide Concentration or	l
	Bifidobacteria Growth	15
	3.6 Animal	15
	3.7 Ethical Approval	15
	3.8 Preparation of Live and Heat-treated Bifidobacteria	16
	3.9 Feeding Mice with Live and Heat-treated Bifidobacteria	16
	3.9.1 Cervical Dislocation Method	16
	3.9.2 Isolation of Peyer's patches (PP) Lymphocytes	17
	3.10 Measurement of IL-12, IL6, IL-4 and IFN-γ	18
	3.11 Measurement of IgA Concentration	19
	3.12 Statistical Analysis	19
4	RESULTS AND DISCUSSION	21
	4.1 Morphological Observation of Bifidobacteria Using Gram Staining	21
	4.2 Effect of Colonic pH on B. pseudocatenulatum G4 and B. longum	
	BB536 Growth	22
	4.3 Effect of Different Concentration of Hydrogen Peroxide on the	
	Growth of Bifidobacterium pseudocatenulatum G4 and Bifidobacterium longum BB536	1 27
	4.4 The Immunomodulatory Effect of Live and Heat-treated	
	Bifidobacterium pseudocatenulatum G4 and Bifidobacterium	
		1 31
	tongum BB330 by Measuring IE-4, IE-0, IE-12 and II 1V-7	31
5	CONCLUSION	39
-	CONCEDENT	J)
REFERE	ENCES	41
APPEND		51
BIODATA OF STUDENT		
	PUBLICATIONS	62

LIST OF TABLES

Table Page

4.1: Effect of different stimulated colonic pH (5.7, 6.4 and 6.9) on bifidobacteria viability 24

4.2: Comparison of growth rate of *B.pseudocatenulatum* G4 and *B. longum* BB 536 in MRS broth without H_2O_2 (control), 50 μ g/l H_2O_2 and 100 μ g/l H_2O_2 30



LIST OF FIGURES

Figure Pag	e
3.1: Growth of bifidobacteria incubated anaerobically at 37 °C for three days in MR agar. All plates containing 30 to 300 colonies.	RS 15
3.2: Oral administration of bifiobacteria (<i>B. pseudocatenulatum</i> G4 and <i>B. longu</i> BB536) to BALB/c mice	m 16
3.3: Washing small intestine with CMF/HEPES solution to remove all fecal matter	17
3.4: Peyer's patches were shown with red arrows in mice small intestine	18
4.1: Microscopic Image of <i>B. longum</i> BB 536 after Gram staining (magnification: 100 X).	00 21
4.2: Compound light microscopic image of <i>B. pseudocatenatum</i> G4 after Gram statir (magnification: 1000 X).	ng 22
4.3: The effect of pH on <i>B. pseudocatenulatum</i> G4 growth at different time (0 h, 1 h, h and 3 h). There are not any significant differences on bifidobacteria growth.	2 25
4.4: The effect of pH on <i>B. longum</i> BB536 growth at different time (0 h, 1 h, 2 h and h). There are not any significant differences on bifidobacteria growth.	. 3 26
4.5: Effect of different pH on <i>B. pseudocatenulatum</i> G4 and <i>B. longum</i> BB 536 durir 24 hours.	ng 26
4.6: Inhibitory effect of H ₂ O ₂ concentration on the growth of bifidobacter (<i>Bifidobacterium pseudocatenulatum</i> G4 and <i>Bifidobacterium longum</i> BB536 10 ⁶ cfu/ml) at different exposure time (0, 12, 24 and 36 hour). The growth of bacteria was decreased with the exposure time.	at
4.7: Inhibitory effect of H ₂ O ₂ concentration on the growth of bifidobacter (<i>Bifidobacterium pseudocatenulatum</i> G4 and <i>Bifidobacterium longum</i> BB536 10 ⁸ cfu/ml) at different exposure time (0, 12, 24 and 36 hour). The growth of bacteria was decreased with the exposure time.	at
4.8: Inhibitory effect H ₂ O ₂ concentration on the growth of bifidobacter (<i>Bifidobacterium pseudocatenulatum</i> G4 and <i>Bifidobacterium longum</i> BB536 10 ¹⁰ cfu/ml) at different exposure time (0, 12, 24 and 36 hour). The growth bacteria was decreased during the exposure time.	at

- 4.9: Immune responses of Peyer's patch cells by oral administration of live and heattreated *B. pseudocatenulatum* G4 and *B. longum* BB536 after 7 days to BALB/c mice.
- 4.10: Immune responses of Peyer's patches cells by oral administration of each bifidobacteria group (live and heat-treated *B. pseudocatenulatum* G4 and *B. longum* BB536) after 7 days.
- 4.11: Immunomodulatory effect of oral administration of live and heat-treated *B. pseudocatenulatum* G4 and *B. longum* BB536 after 7 days.
- 4.12: Immunomodulatory effect of oral administration of live and heat-treated *B. pseudocatenulatum* G4 and *B. longum* BB536 after 7 days.
- 4.13: Effect of live and heat-treated *B. pseudocatenulatum* G4 and *B. longum* BB536 on IgA production after 7 days.

LIST OF APPENDICES

Appendix	
A: Gram Staining	51
B: cfu Calculation	52
C: Ethical Approval	53
D: Guidelines for the Use of Cervical Dislocation for Rodent Euthanasia	54
E: Preparing Reagents for Isolating Peyer's Patches	55
F: ELISA Reagents	56
G: Counting Cells with Hemocytometer Protocol	57
H: Interleukins Standard Curves Obtained From ELISA	58

LIST OF ABBREVIATIONS

% Percentage
 / Per
 < Less than
 μg Microgram
 μl Microlitr
 GI Gastrointestinal
 cfu Colony Forming Unit

g Gram

IgAImmunoglobulin APPsPeyer's PatchesH2O2Hydrogen PeroxideNKNatural killerILInterleukin

HCA Heterocyclic Aromatic Amine
CpG Cystosin Phosphtate Guanosin

DC Dendritic Cell

APCs Antigen Presenting Cells

MHC Major Histocompatibility Complex

TCR T-cell Receptor
Th T-Helper
Treg T-Regulatory

HLA Human Leukocyte Antigen
TNF Tumor Necrosis Factor

IFN Interferon

TGF Transforming Growth Factor sIgA Secretory Immunoglobulin A

UC Ulcerative Colities
CD Crohn's Disease

IBD Inflammatory Bowel Disease

MC Mast cell

PCR Polymerase Chain Reaction
DNA Deoxyribonucleic Acid
TPY Trypticase Phytone Yeast

MRS Man Ragosa
v/v Volume Per Volume
w/v Weight Per Volume
PBS Phosphate Buffer Salin

EDTA Ethylenediaminetetraacetic Acid

Mm Milimolar

rpm Revolutions Per Minute HCL Hydrochloric Acid

h Hour
min Minute
OD Optical Density
B. Bifidobacteria
TLRs Toll-Like Receptors

PAMPS Pathogen Associated Molecular Pattern ELISA Enzyme-Linked Immunosorbent Assay

CHAPTER 1

INTRODUCTION

1.1 Study Background

Based on a definition approved by FAO/WHO, probiotics are 'Live microorganisms, which control human health' (Parvez *et al.*, 2006). These good bacteria are accessible to consumers in various forms of food and dietary supplements (Holzapfel and Schillinger, 2002). They have important roles in antimicrobial activity, enhancement of barrier function, and immunomodulation, so probiotics have significant role in the immune system. For example, antimicrobial activity affects luminal pH and decreases it; and it also secretes antimicrobial peptides, which inhibit bacterial invasion (Mai and Draganov, 2009).

Intestinal flora and the associated immune response are the targets of ingested probiotics, but consequent researches and scientific studies on non-intestinal infections are limited (de Verse & Schrenzenmeir, 2002). Probiotic therapeutic bacteria compete with pathogens for nutrients and mucosal adherence and modulation of mucosal immune functions (O'sullivan *et al.*, 2005). A large number of reports have established the effect of different microorganisms for preventing and curing disorders and strengthening the human immune system. Bifidobacteria have various immunological functions such as mutagenic activity, anti tumor effects and increasing the number of macrophages (Cao *et al.*, 2011).

Immunomodulation is a process that alters the immune system of organisms by interfering with its function as in immunostimulation or immunosuppression (Corthésy et al., 2007). Immune regulation is a multipart section between effectors cells and regulatory cells and there are disparities in their immunological mechanisms (Sforcin, 2007). There is some evidence to improve gastrointestinal (GI) health through valuable food including probiotics and prebiotics. They can prevent colon cancer in humans by strengthening the human immune system. Probiotics bacteria such as bifidobacteria have an immunomodulatory effect on the human immune system, so they can prevent the risk of colon cancer (Delcenserie et al., 2008).

Immunotherapy is a medical term defined as the treatment of human diseases by enhancing, inducing or suppressing an immune response (Khan *et al.*, 2011). Immunotherapies that increase an immune response are classified as activation immunotherapies, and immunotherapies that decrease the immune response are classified as suppression immunotherapies (Khan *et al.*, 2011).

The human intestinal microbiota is the main cause of microbial stimulation that can both benefit and also damages human health. It is a major mediator that contributes to the increase and improvement of the immune system, oral tolerance and immunity (Arrieta *et al.*, 2010). Therefore, the microbiota may be responsible for identifying inflammatory reaction in allergy and in bowel diseases. Probotics such aas bifidobacteria can induce an inflammatory process by stabilizing the intestinal

microbiota environment. Immunomodulation and immunostimulation are the major mechanisms of probiotics that work against many harmful bacteria (Yan and Polk, 2010).

Bifidobacteria are natural members of the human intestinal tract and they occur in concentrations of 10⁹ to 10¹¹ CFU g⁻¹. Bifidobacteria also have an important role in product fermentation. They are referred as probiotics bacteria because of their beneficial role in the human intestine such as raising protein digestion, improving intestinal microflora, human immune system activation and prevention of colon cancer (Gourbeyre *et al.*, 2011).

Bifidobacterium pseudocatenulatum G4 is one of the common bacterial species which are isolated from infant faeces in Malaysia (Kabeir et al., 2005; Yazid et al., 1998). It is well adapted to the colonic environment as reflected by its ability to adapt to low pH and high concentration of bile salts. Studies have demonstrated that the probiotic strain G4 has been identified as a safe probiotic that can be used in functional food for human consumption (Gourbeyre et al., 2011). Bifidobacterium longum is an anaerobic species that can be found in infant's intestine. They prevent the growth of gram negative bacteria such as lactic acid bacteria by producing lactic acid and absorbing the sugar complex compound in human breast milk (Fooks and Gibson, 2002).

1.2 Problem Statements

Modulation of host immunity is one of the proposed benefits of the consumption of probiotics such as *Bifidobacterium pseudocatenulatum* G4 and *Bifidobacterium longum* BB536. On the other hand, activity of the bacteria can be changed using different pH and H₂O₂ concentration which will have an inhibitory effect on bacteria growth.

Also, Preparation of bifidobacteria with different method such as heat treatment and sonication can enable the active substances to be accessible to relevant lymphoid cells. In another word, exposure of the immunopotentiators may have occurred as the result of leakage of intracellular components or disruption of huge complex of cell wall materials. On the other hand, preparation of heat-treated bifidobacteria might induce the thermal denaturation of proteins on the cell surface of bacteria. Although the chemical changes in proteins cannot be detected, heat-treated bifidobacteria can induce immunomodulatory effect. It is hypothesised that heat-treated bacteria increase the production of cytokines and immunomodulatory effect; nonetheless, comparative studies on the immunological properties that support the selection of strains of the same species for specific health benefits are needed in future.

1.3 Significance of the Present Study

Bifidobacterium pseudocatenulatum G4 and Bifidobacterium longum BB536 are becoming increasingly important as source of probiotic in treatment of infection and diseases. This study will provide information on immunomodulatory activity of probiotic bacteria such as Bifidobacterium pseudocatenulatum G4 and Bifidobacterium longum BB536. Findings of study will provide some insight to the treatment of immune-related disease such as infectious gastroenteritis, allergic disease and inflammatory bowel disease.

1.4 General Objective

The main purpose of this study was to determine the characteristics and immunomodulation function of *Bifidobacterium pseudocatenulatum* (G4) and *Bifidobacterium longum* BB536 *in vitro*.

1.5 Specific Objectives

- 1. To evaluate the survival of *B. pseudocatenulatum* G4 and *B. longum* BB536 characteristics in different colonic pH and H₂O₂ concentrations.
- 2. To compare the immunomodutory effect of live and heat-treated *B.pseudocatenulatum* G4 and *B. longum* BB536 using animal feeding study.



REFERENCES

- Abbas, A. K., Lichtman, A. H., and Pillai, S. (2012). *Basic immunology: functions and disorders of the immune system*: Saunders WB.
- Abraham, C., and Medzhitov, R. (2011). Interactions between the host innate immune system and microbes in inflammatory bowel disease. *Gastroenterology*, 140(6), 1729-1737.
- Adams, C. A. (2010). The probiotic paradox: live and dead cells are biological response modifiers. *Nutrition research reviews*, 23(01), 37-46.
- Arnson, Y., Shoenfeld, Y., and Amital, H. (2010). Effects of tobacco smoke on immunity, inflammation and autoimmunity. *Journal of autoimmunity*, 34(3), J258-J265.
- Arrieta, M. C., Meddings, J., and Field, C. J. (2010). The Immunomodulatory Effects of Dietary Fiber and Prebiotics in the Gastrointestinal Tract. *Nondigestible Carbohydrates and Digestive Health*, 37-77.
- Ashford, M. (2013). Gastrointestinal tract—physiology. *Aulton's Pharmaceutics: The Design and Manufacture of Medicines*, 296.
- Blodgett, R. J. (2005). Serial dilution with a confirmation step. *Food microbiology*, 22(6), 547-552.
- Boirivant, M., and Strober, W. (2007). The mechanism of action of probiotics. *Current opinion in gastroenterology*, 23(6), 679-692.
- Bottasso, O., Bay, M. L., Besedovsky, H., and del Rey, A. (2013). Adverse neuro-immune–endocrine interactions in patients with active tuberculosis. *Molecular and Cellular Neuroscience*, 53, 77-85.
- Braciale, T. J., Morrison, L. A., Sweetser, M. T., Sambrook, J., Gething, M. J., and Braciale, V. L. (2006). Antigen presentation pathways to class I and class II MHC-restricted T lymphocytes. *Immunological reviews*, *98*(1), 95-114.
- Broz, P., and Monack, D. M. (2013). Newly described pattern recognition receptors team up against intracellular pathogens. *Nature Reviews Immunology*, 13(8), 551-565.
- Burcelin, R., Luche, E., Serino, M., and Amar, J. (2009). The gut microbiota ecology: a new opportunity for the treatment of metabolic diseases. *Front Biosci*, *14*(5), 107-5117.
- Calcagni, E., and Elenkov, I. (2006). Stress system activity, innate and T helper cytokines, and susceptibility to immune-related diseases. *Annals of the New York Academy of Sciences*, 1069(1), 62-76.

- Caligiuri, M. A. (2008). Human natural killer cells. *Blood*, 112(3), 461-469.
- Cani, P. D., and Delzenne, N. M. (2009). The role of the gut microbiota in energy metabolism and metabolic disease. *Current pharmaceutical design*, 15(13), 1546-1558.
- Cao, L., Liu, X., Qian, T., Sun, G., Guo, Y., Chang, F., Zhou, S., and Sun, X. (2011). Antitumor and immunomodulatory activity of arabinoxylans: a major constituent of wheat bran. *International journal of biological macromolecules*, 48(1), 160-164.
- Chuang, L., Wu, K.-G., Pai, C., Hsieh, P.-S., Tsai, J.-J., Yen, J.-H., and Lin, M.-Y. (2007). Heat-killed cells of lactobacilli skew the immune response toward T helper 1 polarization in mouse splenocytes and dendritic cell-treated T cells. *Journal of agricultural and food chemistry*, 55(26), 11080-11086.
- Chung, H., and Kasper, D. L. (2010). Microbiota-stimulated immune mechanisms to maintain gut homeostasis. *Current opinion in immunology*, 22(4), 455.
- Cleusix, V., Lacroix, C., Vollenweider, S., Duboux, M., and Le Blay, G. (2007). Inhibitory activity spectrum of reuterin produced by Lactobacillus reuteri against intestinal bacteria. *BMC microbiology*, 7(1), 101.
- Coles, M., Kioussis, D., and Veiga-Fernandes, H. (2010). Commentary on "Lymphoid Tissue Inducer Cells and the Evolution of CD4 Dependent High-Affinity Antibody Responses". *Progress in molecular biology and translational science*, 92, 175-176.
- Corthésy, B. (2012). Role of secretory IgA in infection and maintenance of homeostasis. *Autoimmunity reviews*.
- Corthésy, B., Gaskins, H. R., and Mercenier, A. (2007). Cross-talk between probiotic bacteria and the host immune system. *The Journal of nutrition*, 137(3), 781S-790S.
- Croft, M., So, T., Duan, W., and Soroosh, P. (2009). The significance of OX40 and OX40L to T-cell biology and immune disease. *Immunological reviews*, 229(1), 173-191.
- Cummings, J., and Macfarlane, G. (2002). Gastrointestinal effects of prebiotics. *British Journal of Nutrition*, 87(S2), S145-S151.
- Damaskos, D., and Kolios, G. (2008). Probiotics and prebiotics in inflammatory bowel disease: microflora 'on the scope'. *British journal of clinical pharmacology*, 65(4), 453-467.
- De Pablo, P., Chapple, I. L. C., Buckley, C. D., and Dietrich, T. (2009). Periodontitis in systemic rheumatic diseases. *Nature Reviews Rheumatology*, *5*(4), 218-224.

- Delcenserie, V., Martel, D., Lamoureux, M., Amiot, J., Boutin, Y., and Roy, D. (2008). Immunomodulatory effects of probiotics in the intestinal tract. *Current issues in molecular biology*, 10(1/2), 37.
- Edwards, C. A., Duerden, B., and Read, N. (1985). The effects of Ph on colonic bacteria grown in continuous culture. *Journal of medical microbiology*, 19(2), 169-180.
- Endo, K., Shiga, H., Kinouchi, Y., and Shimosegawa, T. (2009). [Inflammatory bowel disease: IBD]. *Rinsho byori. The Japanese journal of clinical pathology*, 57(6), 527.
- Fietta, P., and Delsante, G. (2009). The effector T helper cell triade. *Rivista di biologia*, 102(1), 61.
- Foligne, B., Nutten, S., Grangette, C., Dennin, V., Goudercourt, D., Poiret, S., Dewulf, J., Brassart, D., Mercenier, A., and Pot, B. (2007). Correlation between in vitro and in vivo immunomodulatory properties of lactic acid bacteria. *World Journal of Gastroenterology*, 13(2), 236.
- Fooks, L., and Gibson, G. (2002). Probiotics as modulators of the gut flora. *British Journal of Nutrition*, 88(S1), s39-s49.
- Gaffen, S., and Hajishengallis, G. (2008). A new inflammatory cytokine on the block: re-thinking periodontal disease and the Th1/Th2 paradigm in the context of Th17 cells and IL-17. *Journal of dental research*, 87(9), 817-828.
- Galli, S. J., Tsai, M., and Piliponsky, A. M. (2008). The development of allergic inflammation. *Nature*, 454(7203), 445-454.
- Geuking, M., McCoy, K., and Macpherson, A. (2012). The function of secretory IgA in the context of the intestinal continuum of adaptive immune responses in host-microbial mutualism. Paper presented at the Seminars in Immunology.
- Gill, H., and Prasad, J. (2008). Probiotics, immunomodulation, and health benefits. *Bioactive Components of Milk*, 423-454.
- Gorissen, L., Weckx, S., Vlaeminck, B., Raes, K., De Vuyst, L., De Smet, S., and Leroy, F. (2011). Linoleate isomerase activity occurs in lactic acid bacteria strains and is affected by Ph and temperature. *Journal of applied microbiology*, 111(3), 593-606.
- Gourbeyre, P., Denery, S., and Bodinier, M. (2011). Probiotics, prebiotics, and synbiotics: impact on the gut immune system and allergic reactions. *Journal of Leukocyte Biology*, 89(5), 685-695.
- Hanauer, S. B. (2006). Inflammatory bowel disease: epidemiology, pathogenesis, and therapeutic opportunities. *Inflammatory bowel diseases*, 12(5), S3-S9.
- Hermanson, G. T. (2013). Bioconjugate techniques: Academic press.

- Hertzberger, R., Arents, J., Dekker, H. L., Pridmore, R. D., Gysler, C., Kleerebezem, M., and de Mattos, M. J. T. (2014). H2O2 Production in Species of the Lactobacillus acidophilus Group: a Central Role for a Novel NADH-Dependent Flavin Reductase. *Applied and environmental microbiology*, 80(7), 2229-2239.
- Hidalgo, M., Oruna-Concha, M. J., Kolida, S., Walton, G. E., Kallithraka, S., Spencer, J. P., Gibson, G. R., and de Pascual-Teresa, S. (2012). Metabolism of anthocyanins by human gut microflora and their influence on gut bacterial growth. *Journal of agricultural and food chemistry*, 60(15), 3882-3890.
- Holzapfel, W. H., and Schillinger, U. (2002). Introduction to pre-and probiotics. *Food research international*, *35*(2), 109-116.
- Huang, Y., and Chen, H. (2011). Effect of organic acids, hydrogen peroxide and mild heat on inactivation of Escherichia coli O157: H7 on baby spinach. *Food Control*, 22(8), 1178-1183.
- Inoue, K.-I., Koike, E., Takano, H., Yanagisawa, R., Ichinose, T., and Yoshikawa, T. (2009). Effects of diesel exhaust particles on antigen-presenting cells and antigen-specific Th immunity in mice. *Experimental biology and medicine*, 234(2), 200-209.
- Isolauri, E., Sütas, Y., Kankaanpää, P., Arvilommi, H., and Salminen, S. (2001). Probiotics: effects on immunity. *The American journal of clinical nutrition*, 73(2), 444s-450s.
- Kabeir, B., Abd-Aziz, S., Muhammad, K., Shuhaimi, M., and Yazid, A. (2005). Growth of Bifidobacterium longum BB536 in medida (fermented cereal porridge) and their survival during refrigerated storage. *Letters in applied microbiology*, 41(2), 125-131.
- Kabeir, B. M., Yazid, A. M., Hakim, M. N., Khahatan, A., Shaborin, A., and Mustafa, S. (2009). Survival of Bifidobacterium pseudocatenulatum G4 during the storage of fermented peanut milk (PM) and skim milk (SM) products. *African Journal of Food Science*, *3*(6), 150-155.
- Kelly, D., Conway, S., and Aminov, R. (2005). Commensal gut bacteria: mechanisms of immune modulation. *Trends in immunology*, 26(6), 326-333.
- Khan, K. J., Dubinsky, M. C., Ford, A. C., Ullman, T. A., Talley, N. J., and Moayyedi, P. (2011). Efficacy of immunosuppressive therapy for inflammatory bowel disease: a systematic review and meta-analysis. *The American Journal of Gastroenterology*, 106(4), 630-642.
- Kheadr, E., Dabour, N., Le Lay, C., Lacroix, C., and Fliss, I. (2007). Antibiotic susceptibility profile of bifidobacteria as affected by oxgall, acid, and hydrogen peroxide stress. *Antimicrobial agents and chemotherapy*, 51(1), 169-174.

- Kildsgaard, J., LESER, T. D., GUNNARSSON, T., WEISE, M., FOLKENBERG, D. M., JANZEN, T., and FLAMBARD, B. (2010). NEW PROBIOTIC BIFIDOBACTERIUM LONGUM: WO Patent 2,010,003,916.
- Klaenhammer, T. R., Kleerebezem, M., Kopp, M. V., and Rescigno, M. (2012). The impact of probiotics and prebiotics on the immune system. *Nature Reviews Immunology*, *12*(10), 728-734.
- Knapp, S., Kühn, I., Stolle, J., and Klotz, S. (2010). Changes in the functional composition of a Central European urban flora over three centuries. *Perspectives in Plant Ecology, Evolution and Systematics*, 12(3), 235-244.
- Kode, J. A., Mukherjee, S., Joglekar, M. V., and Hardikar, A. A. (2009). Mesenchymal stem cells: immunobiology and role in immunomodulation and tissue regeneration. *Cytotherapy*, *11*(4), 377-391.
- Kukkonen, K., Kuitunen, M., Haahtela, T., Korpela, R., Poussa, T., and Savilahti, E. (2010). High intestinal IgA associates with reduced risk of IgE-associated allergic diseases. *Pediatric Allergy and Immunology*, 21(1-Part-I), 67-73.
- Lagaert, S., Pollet, A., Delcour, J. A., Lavigne, R., Courtin, C. M., and Volckaert, G. (2011). Characterization of two β-xylosidases from Bifidobacterium adolescentis and their contribution to the hydrolysis of prebiotic xylooligosaccharides. *Applied Microbiology and Biotechnology*, 92(6), 1179-1185.
- Lefrançois, L., and Lycke, N. (2001). Isolation of mouse small intestinal intraepithelial lymphocytes, Peyer's patch, and lamina propria cells. *Current protocols in immunology*, 3.19. 11-13.19. 16.
- Liang, H., Baudouin, C., Dupas, B., and Brignole-Baudouin, F. (2010). Live conjunctiva-associated lymphoid tissue analysis in rabbit under inflammatory stimuli using in vivo confocal microscopy. *Investigative ophthalmology & visual science*, 51(2), 1008-1015.
- Liu, Z., Jiang, Z., Zhou, K., Li, P., Liu, G., and Zhang, B. (2007). Screening of bifidobacteria with acquired tolerance to human gastrointestinal tract. *Anaerobe*, 13(5), 215-219.
- Löhning, M., Hegazy, A. N., Pinschewer, D. D., Busse, D., Lang, K. S., Höfer, T., Radbruch, A., Zinkernagel, R. M., and Hengartner, H. (2008). Long-lived virus-reactive memory T cells generated from purified cytokine-secreting T helper type 1 and type 2 effectors. *The Journal of experimental medicine*, 205(1), 53-61.
- Macfarlane, G., Steed, H., and Macfarlane, S. (2007). Bacterial metabolism and health-related effects of galacto-oligosaccharides and other prebiotics. *Journal of Applied Microbiology*, 104(2), 305-344.

- Maes, M., Kubera, M., Leunis, J.-C., and Berk, M. (2012). Increased IgA and IgM responses against gut commensals in chronic depression: further evidence for increased bacterial translocation or leaky gut. *Journal of affective disorders*, 141(1), 55-62.
- Mai, V., and Draganov, P. V. (2009). Recent advances and remaining gaps in our knowledge of associations between gut microbiota and human health. *World journal of gastroenterology: WJG*, 15(1), 81.
- Makino, H., Kushiro, A., Ishikawa, E., Muylaert, D., Kubota, H., Sakai, T., Oishi, K., Martin, R., Amor, K. B., and Oozeer, R. (2011). Transmission of intestinal Bifidobacterium longum subsp. longum strains from mother to infant, determined by multilocus sequencing typing and amplified fragment length polymorphism. *Applied and Environmental Microbiology*, 77(19), 6788-6793.
- Marhamatizadeh, M. H., Mohammadi, M., Rezazadeh, S., and Jafari, F. (2012). Effects of Garlic on the Growth of Lactobacillus acidophilus and Bifidobacterium bifidum in Probiotic Milk and Yoghurt. *IDOSI Publications, Middle-East Journal of Scientific Research*, 11(7), 894-899.
- Mariam, R. S., Yap, K. W., Lim, L. C., Kharidah, M., Shuhaimi, M., Abdullah, S., Ali, A. M., Atiqah, A. N., and YAZID, A. M. (2004). Strain Differences in Deconjugation of Bile Acids in Bifidobacterium pseudocatenulatum Isolates. *Bioscience and microflora*, 23(2), 93-98.
- Maus, J., and Ingham, S. (2003). Employment of stressful conditions during culture production to enhance subsequent cold-and acid-tolerance of bifidobacteria. *Journal of applied microbiology*, 95(1), 146-154.
- McGuirk, P., and Mills, K. H. (2002). Pathogen-specific regulatory T cells provoke a shift in the Th1/Th2 paradigm in immunity to infectious diseases. *Trends in immunology*, 23(9), 450-455.
- Mendelow, A., Chambers, I., Gregson, B., Crawford, P., Barnes, M., Monitor, I., Abdullah, J., Ghazaime, G., Zamzuri, I., and Sayuthi, S. (2002). Euroacademy of Multidisciplinary Neurotraumatology. *Acta Neurochir (Wien)*, 144, A1-A41.
- Modesto, M., Michelini, S., Stefanini, I., Ferrara, A., Tacconi, S., Biavati, B., and Mattarelli, P. (2014). Bifidobacterium aesculapii sp. nov., from the faeces of the baby common marmoset (Callithrix jacchus). *International journal of systematic and evolutionary microbiology*, ijs. 0.056937-056930.
- Mustafa, S. (1999). *Isolation and Characterisation of Bifidobacterium Spp. from Infant Stools*. Universiti Pertanian Malaysia.
- Nakahira, K., Haspel, J. A., Rathinam, V. A., Lee, S.-J., Dolinay, T., Lam, H. C., Englert, J. A., Rabinovitch, M., Cernadas, M., and Kim, H. P. (2011). Autophagy proteins regulate innate immune responses by inhibiting the

- release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nature immunology*, 12(3), 222-230.
- Nakanishi, K. (2010). Basophils as APC in Th2 response in allergic inflammation and parasite infection. *Current opinion in immunology*, 22(6), 814-820.
- Nakanishi, Y., Hosono, A., Hiramatsu, Y., Kimura, T., Nakamura, R., and Kaminogawa, S. (2005a). Characteristic immune response in Peyer's patch cells induced by oral administration of Bifidobacterium components. *Cytotechnology*, 47(1-3), 69-77.
- Nakanishi, Y., Hosono, A., Hiramatsu, Y., Kimura, T., Nakamura, R., and Kaminogawa, S. (2005b). Characteristic immune response in Peyer's patch cells induced by oral administration of Bifidobacterium components. *Cytotechnology*, 47(1), 69-77.
- Nicholson, J. K., Holmes, E., Kinross, J., Burcelin, R., Gibson, G., Jia, W., and Pettersson, S. (2012). Host-gut microbiota metabolic interactions. *Science*, 336(6086), 1262-1267.
- Oberg, T., Steele, J., Ingham, S., Smeianov, V., Briczinski, E., Abdalla, A., and Broadbent, J. R. (2011). Intrinsic and inducible resistance to hydrogen peroxide in Bifidobacterium species. *Journal of industrial microbiology & biotechnology*, 38(12), 1947-1953.
- Ohlrich, E., Cullinan, M., and Seymour, G. (2009). The immunopathogenesis of periodontal disease. *Australian dental journal*, 54(s1), S2-S10.
- Ozaki, K., Fujii, S., and Hayashi, M. (2007). Effect of dietary mannooligosaccharides on the immune system of ovalbumin-sensitized mice. *Journal of health science*, 53(6), 766-770.
- Parvez, S., Malik, K., Ah Kang, S., and Kim, H. Y. (2006). Probiotics and their fermented food products are beneficial for health. *Journal of applied microbiology*, 100(6), 1171-1185.
- Pedroso, D. d. L., Thomazini, M., Heinemann, R. J. B., and Favaro-Trindade, C. S. (2012). Protection of Bifidobacterium lactis and Lactobacillus acidophilus by microencapsulation using spray-chilling. *International Dairy Journal*, 26(2), 127-132.
- Pericone, C. D., Overweg, K., Hermans, P. W., and Weiser, J. N. (2000). Inhibitory and bactericidal effects of hydrogen peroxide production by Streptococcus pneumoniae on other inhabitants of the upper respiratory tract. *Infection and immunity*, 68(7), 3990-3997.
- Perrigoue, J. G., Saenz, S. A., Siracusa, M. C., Allenspach, E. J., Taylor, B. C., Giacomin, P. R., Nair, M. G., Du, Y., Zaph, C., and Van Rooijen, N. (2009). MHC class II–dependent basophil–CD4+ T cell interactions promote TH2 cytokine–dependent immunity. *Nature immunology*, 10(7), 697-705.

- Ranadheera, R., Baines, S., and Adams, M. (2010). Importance of food in probiotic efficacy. *Food Research International*, 43(1), 1-7.
- Rastall, R. A., Gibson, G. R., Gill, H. S., Guarner, F., Klaenhammer, T. R., Pot, B., Reid, G., Rowland, I. R., and Sanders, M. E. (2006). Modulation of the microbial ecology of the human colon by probiotics, prebiotics and synbiotics to enhance human health: An overview of enabling science and potential applications. *FEMS microbiology ecology*, *52*(2), 145-152.
- Rigby, R., Knight, S., Kamm, M., and Stagg, A. (2005). Production of interleukin (IL)-10 and IL-12 by murine colonic dendritic cells in response to microbial stimuli. *Clinical & Experimental Immunology*, 139(2), 245-256.
- Roberfroid, M., Gibson, G. R., Hoyles, L., McCartney, A. L., Rastall, R., Rowland, I., Wolvers, D., Watztl, B., Szajewska, H., and Stahl, B. (2010). Prebiotic effects: metabolic and health benefits. *British Journal of Nutrition*, 104(S2), S1-S63.
- Roger, L. C., Costabile, A., Holland, D. T., Hoyles, L., and McCartney, A. L. (2010). Examination of faecal Bifidobacterium populations in breast-and formula-fed infants during the first 18 months of life. *Microbiology*, *156*(11), 3329-3341.
- Roustan, A., Perrin, J., Berthelot-Ricou, A., Lopez, E., Botta, A., and Courbiere, B. (2012). Evaluating methods of mouse euthanasia on the oocyte quality: cervical dislocation versus isoflurane inhalation. *Laboratory animals*, 46(2), 167-169.
- Russell, C. M., Choo, A. M., Tetzlaff, W., Chung, T.-E., and Oxland, T. R. (2012). Maximum principal strain correlates with spinal cord tissue damage in contusion and dislocation injuries in the rat cervical spine. *Journal of neurotrauma*, 29(8), 1574-1585.
- Sánchez, B., Champomier-Vergès, M.-C., del Carmen Collado, M., Anglade, P., Baraige, F., Sanz, Y., Clara, G., Margolles, A., and Zagorec, M. (2007). Low-Ph adaptation and the acid tolerance response of Bifidobacterium longum biotype longum. *Applied and environmental microbiology*, 73(20), 6450-6459.
- Sanders, E. R. (2012). Aseptic laboratory techniques: plating methods. *JoVE* (*Journal of Visualized Experiments*)(63), e3064-e3064.
- Sartor, R. B. (2008). Microbial influences in inflammatory bowel diseases. *Gastroenterology*, 134(2), 577.
- Scardovi, V. (1986). Genus bifidobacterium. *Bergey's manual of systematic bacteriology*, 2, 1418-1434.
- Schmielau, J., and Finn, O. J. (2001). Activated granulocytes and granulocyte-derived hydrogen peroxide are the underlying mechanism of suppression of t-cell function in advanced cancer patients. *Cancer research*, 61(12), 4756-4760.

- Sforcin, J. (2007). Propolis and the immune system: a review. *Journal of ethnopharmacology*, 113(1), 1-14.
- Sghir, A., Chow, J., and Mackie, R. (1998). Continuous culture selection of bifidobacteria and lactobacilli from human faecal samples using fructooligosaccharide as selective substrate. *Journal of applied microbiology*, 85(4), 769-777.
- Shuhaimi, M., Yazid, A., Ali, A., Ghazali, M., Zaitun, H., and Atiqah, N. N. (1999). Antibacterial activity, antimicrobial susceptibility and adherence properties of Bifidobacterium infantis G4. *Pak. J. Biol. Sci.*, 2, 1231-1235.
- Smith, A. M., Rahman, F. Z., Hayee, B. H., Graham, S. J., Marks, D. J. B., Sewell, G. W., Palmer, C. D., Wilde, J., Foxwell, B. M. J., and Gloger, I. S. (2009). Disordered macrophage cytokine secretion underlies impaired acute inflammation and bacterial clearance in Crohn's disease. *The Journal of experimental medicine*, 206(9), 1883-1897.
- Soehnlein, O., and Lindbom, L. (2010). Phagocyte partnership during the onset and resolution of inflammation. *Nature Reviews Immunology*, *10*(6), 427-439.
- Stephenie, W., Yap, K., Barka, M., Shuhaimi, M., Rosfarizan, M., and Yazid, A. (2006). Survival of Bifidobacterium pseudocatenulatum strains isolated from breast-fed infants to simlated gastric Ph environment. *Malaysian Applied Biology*, 35(1), 57.
- Swain, S. L. (2006). T cell subsets and the recognition of MHC class. *Immunological reviews*, 74(1), 129-142.
- Takahashi, N., and Nyvad, B. (2011). The Role of Bacteria in the Caries Process Ecological Perspectives. *Journal of Dental Research*, 90(3), 294-303.
- Titball, R. W., and Williamson, E. D. (2001). Vaccination against bubonic and pneumonic plague. *Vaccine*, 19(30), 4175-4184.
- Trinchieri, G., and Sher, A. (2007). Cooperation of Toll-like receptor signals in innate immune defence. *Nature Reviews Immunology*, 7(3), 179-190.
- Troutman, T. D., Hu, W., Fulenchek, S., Yamazaki, T., Kurosaki, T., Bazan, J. F., and Pasare, C. (2012). Role for B-cell adapter for PI3K (BCAP) as a signaling adapter linking Toll-like receptors (TLRs) to serine/threonine kinases PI3K/Akt. *Proceedings of the National Academy of Sciences*, 109(1), 273-278.
- Walker, A. W., Duncan, S. H., Leitch, E. C. M., Child, M. W., and Flint, H. J. (2005). Ph and peptide supply can radically alter bacterial populations and short-chain fatty acid ratios within microbial communities from the human colon. *Applied and environmental microbiology*, 71(7), 3692-3700.
- Watson, J., and Schubert, J. (1969). Action of hydrogen peroxide on growth inhibition of Salmonella typhimurium. *Journal of general microbiology*, *57*(1), 25-34.

- Yan, F., and Polk, D. B. (2010). Probiotics: progress toward novel therapies for intestinal diseases. *Current Opinion in Gastroenterology*, 26(2), 95.
- Yanagibashi, T., Hosono, A., Oyama, A., Tsuda, M., Hachimura, S., Takahashi, Y., Itoh, K., Hirayama, K., Takahashi, K., and Kaminogawa, S. (2009). Bacteroides induce higher IgA production than Lactobacillus by increasing activation-induced cytidine deaminase expression in B cells in murine Peyer's patches. *Bioscience, biotechnology, and biochemistry*, 73(2), 372-377.
- Yazid, A., Ali, A., Kalaivani, V., and Shuhaimi, M. (1998). Properties of Bifidobacterium spp.: antibacterial, growth in milk and survival during low temperature storage. *Asia-Pacific Journal of Molecular Biology and Biotechnology*, 6(2), 153-159.
- Yoo, S. K., and Huttenlocher, A. (2009). Innate Immunity: Wounds Burst H₂O₂ Signals to Leukocytes. *Current Biology*, 19(14), R553-R555.
- Zaghouani, H., Hoeman, C. M., and Adkins, B. (2009). Neonatal immunity: faulty Thelpers and the shortcomings of dendritic cells. *Trends in immunology*, 30(12), 585-591.
- Zinedine, A., and Faid, M. (2007). Isolation and Characterization of Strains of Bifidobacteria with Probiotic Proprieties In vitro. *World Journal of Dairy & Food Sciences*, 2(1), 28-34.