

UNIVERSITI PUTRA MALAYSIA

PATHOGENICITY OF A MALAYSIAN FOWL ADENOVIRUS ISOLATE IN SPECIFIC PATHOGEN FREE EMBRYONATED CHICKEN EGGS AND CHICKS

ALEMNESH WOLDEYES YIMER

FPV 2009 3



PATHOGENICITY OF A MALAYSIAN FOWL ADENOVIRUS ISOLATE IN SPECIFIC PATHOGEN FREE EMBRYONATED CHICKEN EGGS AND CHICKS

ALEMNESH WOLDEYES YIMER

MASTER OF VETERINARY SCIENCE UNIVERSITI PUTRA MALAYSIA

2009



PATHOGENICITY OF A MALAYSIAN FOWL ADENOVIRUS ISOLATE IN SPECIFIC PATHOGEN FREE EMBRYONATED CHICKEN EGGS AND CHICKS

BY

ALEMNESH WOLDEYES YIMER

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Veterinary Science

January 2009



Dedicated to My parent My husband, Son and siblings For their love, prayer and encouragements



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Veterinary Science

PATHOGENICITY OF A MALAYSIAN FOWL ADENOVIRUS ISOLATE IN SPECIFIC PATHOGEN FREE EMBRYONATED CHICKEN EGGS AND CHICKS

By

ALEMNESH WOLDEYES

January, 2009

Chairman: Professor Mohd Hair Bejo, PhD

Faculty: Veterinary Medicine

The pathogenic role of fowl adenoviruses (FAdVs) in disease outbreaks remained unestablished, as some findings showed that FAdVs are the primary infectious agents whilst others showed that these viruses can co-infect with other immunosuppressive agents to cause diseases. Thus, there is a need to establish the roles and characteristics of the virus in chicken to control and prevent the disease, especially by developing vaccine and introducing vaccination programme against the disease. The objective of the study was to isolate, identify and determine the pathogenicity of Malaysian isolate of FAdV in specific pathogen free (SPF) embryonated chicken eggs and chicks.

Liver homogenate obtained from inclusion body hepatitis (IBH) field outbreaks in commercial broiler chicken was inoculated, passaged and titrated in SPF embryonated chicken eggs via chorioallantoic membrane (CAM) route and the FAdV was identified



using electron microscopy. The virus inoculum with the titre of 1x 10 $^{4.8}$ EID₅₀/ 0.1 ml was then inoculated into 9-day-old SPF embryonated chicken eggs as well as 9-day-old SPF chicks. Control groups were included and remained non-inoculated. They were monitored for mortality and clinical signs. The eggs and chicks were either labeled for determination of the percentage of mortality or sacrificed at days 1, 3, 5, 7, 9, 11 and 12, and days 1, 3, 5, 7, 10, 14 and 21 post inoculations (pi), respectively. On necropsy, samples of liver, CAM, yolk sac, kidney, spleen, heart and bursa of Fabricius from the embryos were collected for histological examination, whilst the liver and CAM were also examined for ultrastructural changes. Samples of liver, spleen, gizzard, proventriculus, kidney, pancreas, duodenum and bursa of Fabricius of the chicks were also examined for histological changes, whilst samples of liver were examined for ultrastructural changes as well.

The study showed 100% embryo mortality within 4 to 11 days pi of the virus in SPF embryonated chicken eggs. The gross and histological lesions of the embryo were confined in the liver at days 5, 7, 9 and 11 pi. Grossly the liver was pale with multi-focal areas of necrosis, fibrosis and hemorrhages. Histologically, moderate to severe congestion and hemorrhage, severe diffused degeneration and necrosis of the hepatocytes with intranuclear inclusion bodies (INIB), and infiltration of inflammatory cells were recorded. Hemorrhage, congestion, degeneration, necrosis and hyperplasia of the chorionic epithelium of CAM with INIB were observed at days 5, 7, 9 and 11 pi. Varying degree of congestion, hemorrhage, degeneration and necrosis were also observed in the yolk sac, kidney, spleen, heart and bursa of Fabricius. Numerous viral



particles in the nucleus of hepatocytes were recorded at days 7, 9 and 11 pi under ultrastructural examination, whereas at 5 days pi fine granular and filamentous materials of inclusion bodies were observed. The INIB in the ectoderm were either as fine granular and filamentous structures or as large viral inclusions. Neither clinical signs nor mortality and gross lesions were observed in the chicks in both groups; the FAdV inoculated and control groups. However, histologically mild to moderate degeneration with focal areas of necrosis, presence of INIB and mild infiltration of inflammatory cells in the liver and mild degeneration and necrosis in the proventriculus and pancreas were observed at days 5 to 14 pi. Ultrastructurally, fibrillar, granular and filaments particles of INIB of the hepatocytes were recorded at days 5 to 14 pi.

It was concluded that the FAdV of Malaysian isolate is highly pathogenic to SPF embryonated chicken eggs, but low pathogenic to the chicks. The embryonic liver is the best organ to be used for adaptation and passaging of the virus since severe lesions and numerous viral particles were observed in the liver, the target organ of the virus. The failure of the virus particles to develop in the hepatocytes could result in the failure of disease development in the FAdV infection in the chicks. It appears that the FAdV of Malaysian isolate has high potential to be used as a viral vector to produce recombinant vaccine against poultry diseases.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains Veterinar

PATOGENISITI SATU ISOLAT VIRUS ADENO UNGGAS MALAYSIA TERHADAP TELUR BEREMBRIO DAN ANAK AYAM BEBAS PENYAKIT KHUSUS

Oleh

ALEMNESH WOLDEYES

Januari, 2009

Pengerusi: Professor Mohd Hair Bejo, PhD

Fakulti: Perubatan Veterinar

Peranan patogenik virus adeno unggas (FAdVs) dalam kes wabak penyakit masih tidak dapat ditentukan kerana sesetengah hasil penemuan menunjukkan bahawa FAdVs adalah agen berjangkit primari sedangkan yang lain menunjukkan bahawa FAdVs adalah agen sampingan kepada agen penyebab kehilangan daya tahan lain yang menjadi penyebab penyakit. Oleh itu adalah perlu untuk menentukan peranan dan ciri-ciri virus ini di dalam ayam untuk pengawalan dan pencegahan penyakit, terutamanya dalam penghasilan vaksin dan pengenalan program vaksinasi untuk menangani penyakit ini. Adalah menjadi objektif kajian ini untuk mengasingkan, mengenalpasti dan menentukan patogenisiti isolat FAdV Malaysia di dalam telur ayam berembrio bebas penyakit khusus (SPF) dan anak ayam. Homogenat hepar dari kes wabak 'inclusion body hepatitis (IBH)' di ladang komersil ayam pedaging diinokulasi, pasage, dan dititrasi ke dalam telur SPF melalui membran korioalantoic (CAM) dan isolat FAdV dikenalpasti di bawah mikroskop elektron. Virus inokulum dengan titer 1x 10^{4.8} EID₅₀/ 0.1 ml kemudiannya



diinokulasi ke dalam telur berembrio SPF berumur 9 hari dan anak ayam SPF berumur 9 hari. Kumpulan kawalan disertakan dan tidak diinokulasi. Pemerhatian ke atas kematian dan tanda klinikal dijalankan. Telur dan anak ayam dilabel untuk penentuan peratusan kematian atau dikorbankan masing-masing pada 1, 3, 5, 7, 9, 11 dan 12 hari, dan 1, 3, 5, 7, 10, 14 dan 21 hari selepas inokulasi (pi). Semasa nekropsi, sampel hati, CAM, kantung yok, buah pinggang, limpa, jantung dan bursa Fabricius daripada embrio dikumpulkan untuk pemeriksaan histologi, di mana hati dan CAM turut diperiksa untuk perubahan ultrastruktur di bawah elektron mikroscop. Sampel hati, limpa, hempadal, proventrikulus, buah pinggang, pankreas, duodenum dan bursa Fabricius daripada anak ayam diperiksa untuk perubahan histologi, di mana sampel hati turut diperiksa untuk perubahan ultrastruktur.

Kajian ini menunjukkan 100% kematian embrio dalam 4 dan 11 hari pi virus di dalam telur ayam berembrio SPF. Lesi mata kasar dan histologi embrio dapat dilihat pada hati pada 5, 7, 9 dan 11 hari pi. Secara mata kasarnya, hati pucat dengan terdapat banyak kawasan fokal nekrosis, fibrosis dan hemoraj. Secara histologi, konjasi dan hemoraj yang sederhana ke teruk, degenerasi yang teruk dan nekrosis sel-sel hati dengan inclusion bodi dalam nuklear dan menyeluruh, dan infiltrasi sel-sel inflamatori direkodkan. Hemoraj, konjasi, degenerasi, nekrosis dan hiperplasia pada epitelia CAM dengan INIB diperhati pada 5, 7, 9 dan 11 hari pi. Pelbagai darjah konjasi, hemoraj, degenerasi dan nekrosis juga diperhati dalam kantung yok, buah pinggang, limpa, jantung dan bursa Fabricius. Partikel virus di dalam nukleus sel-sel hati juga direkodkan pada 7, 9 dan 11 hari pi di bawah pemeriksaan ultrastruktur, manakala pada 5 hari pi



granul halus dan bahan filamen inclusion bodi diperhatikan. INIB dalam ectoderm adalah semada berbentuk granul halus dan berstruktur filemen atau berbentuk inclusion virus yang besar. Tiada tanda klinikal atau kematian diperhati dalam anak ayam daripada kedua-dua kumpulan; kumpulan FAdV inokulat dan kawalan. Walaubagaimanapun, perubahan histologi manunjukkan sedikit hingga sederhana degenerasi dengan kawasan nekrosis yang fokal, terdapat INB dan infiltrasi sel-sel inflamatori yang sedikit dalam hati dan degenerasi dan nekrosis yang sedikit pada proventrikulus dan pankreas diperhati pada 5 sehingga 14 hari pi. Pemeriksaan ultrastruktur merekodkan INIB berpartikel fibril, granul dan filamen dalam sel hati pada 5 sehingga 14 hari pi.

Kesimpulannya, FAdV isolat Malaysia adalah sangat patogenik ke atas telur ayam berembrio tetapi berpatogenik rendah terhadap anak ayam. Hati embrio merupakan organ paling sesuai untuk penyesuaian dan passage virus kerana lesi teruk dan banyak partikel virus diperhatikan di dalam hati, organ target FAdV. Kegagalan partikel virus untuk mengembang dalam sel-sel hati mungkin menyebabkan kegagalan perkembangan penyakit daripada jangkitan FAdV dalam anak ayam. FAdV isolat Malaysia mempunyai potensi tinggi untuk digunakan sebagai vektor virus untuk menghasilkan vaksin rekombinan terhadap penyakit-penyakit ayam.



ACKNOWLEDGEMENTS

I give all praises to almighty God and Virgin Mary for leading me to this far and making all things possible.

I would like to express my appreciation and gratitude to Prof. Dr. Mohd Hair-Bejo for his invaluable guidance, encouragement and scientific discussions in research throughout the study period. I am also grateful to him for monitoring me throughout the study which helps me to finish the research early.

I would like to thank members of the advisory committee, Prof. Dr. Aini Ideris and Assoc. Prof. Dr. Abdul Rahman Omar for their constructive guidance and encouragement throughout the study period.

I am grateful to the Faculty of Veterinary Medicine, Gondar University, for providing the opportunity to pursue my Master of Veterinary Science Programme in Malaysia with the financial support under NUFFIC project in Ethiopia. I also would like to express my sincere thanks to NUFFIC for partially funding this research, and for providing me the necessary financial support during my study period.

I wish to thank all the staffs and students at the Department of Veterinary Pathology and Microbiology, Faculty of Veterinary Medicine, Universiti Putra Malaysia for their assistance and invaluable service. My sincere thanks to my entire laboratory mates and



my country mates especially Lemlem Kassa and Tekeleselassie Ayalew for their helpfulness, friendship and understanding during my stay in Malaysia.

I am expressing my utmost gratitude and appreciation to my husband Ashagrie Getnet, my son Yohannes Ashagrie, my brothers Mulu, Girma and Anteneh, my sister Tenaye and relatives for their patience, continuous help, cooperation and encouragement throughout my study period, which will always be remembered and appreciated.



I certify that a Thesis Examination Committee has met on 19 January 2009 to conduct the final examination of Alemnesh Woldeyes on her thesis entitled "Pathogenicity of a Malaysian Fowl Adenovirus Isolate in Specific Pathogen Free Embryonated Chicken Eggs and Chicks" in accordance with the Universities and University colleges Act 1971 and the constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Veterinary Science.

Members of the Thesis Examination Committee were as follows:

Hassan Hj Mohd Daud, PhD

Associate professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Jasni Sabri, PhD

Associate professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Fauziah Othman, PhD

Professor Faculty of Medicine and Heath Sciences Universiti Putra Malaysia (Internal Examiner)

Name of External Examiner, PhD

Professor Faculty of Agriculture University of Tokyo Japan (External Examiner)

BUJANG KIM HUAT, PhD

Professor AND Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 19 February 2009



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Veterinary Science. The members of the Supervisory Committee were as follows:

MOHD HAIR BEJO, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

AINI IDERIS, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

ABDUL RAHMAN OMAR, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

HASANAH MOHD GHAZALI, PhD

Professor/Dean School of Graduate Studies Universiti Putra Malaysia

Date:



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

Alemnesh Woldeyes Yimer

Date:



TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	111
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xvii
LIST OF FIGURES	xviii
LIST OF ABBREVIATIONS	xxi

CHAPTER

2	INT	RODUC	TION	
2	LI	TERATU	JRE REVIEW	6
	2.1	Fowl A	Adenovirus	6
		2.1.1	Classification	6
		2.1.2	Structure and proteins	7
		2.1.3	Viral genome	10
		2.1.4	Virus replication	11
		2.1.5	Physico-chemical properties of the virus	12
		2.1.6	Antigenic properties	12
		2.1.7	Adenovirus as a vector	13
		2.1.8	Transmission	16
	2.1	Fowl A	Adenovirus Infection	16
		2.2.1	Experimental reproduction of diseases	19
		2.2.2	Pathogenicity and pathogenesis	22
		2.2.3	Clinical signs	24
			Gross and histological lesions	25
		2.2.5	Ultrastructural changes	27
		2.2.6	Immunosuppression and interaction with the other	
			pathogenes	28
	2.2	Diagn	osis	28
		2.3.1	Isolation and identification of adenoviruses	28
		2.3.2	Serology	28
	2.3	Contro	ol and Prevention	31
		2.4.1	Exclusion or eradication	32
		2.4.2	Vaccination	33
		2.4.3	Immunity	34



3			NICITY OF FOWL ADENOVIRUS IN SPECIFIC N FREE EMBRYONATED CHICKEN EGGS	36
	3.1	Introd	uction	36
	3.2		als and Methods	40
			FAdV isolate	40
		3.2.2	Preparation of liver homogenate	40
		3.2.3	SPF embryonated chicken eggs inoculation	40
			Virus titration	42
		3.2.5	Propagation of the FAdV in SPF embryonated chicke eggs	n 42
		3.2.6	Gross lesions	43
		3.2.7		43
		3.2.8	Transmission electron microscopy	45
	3.3	Results	8	47
		3.3.1	Fowl adenovirus isolate	47
		3.3.2	Transmission electron microscopy: negative staining	48
		3.3.3	Virus titre	48
		3.3.4	Mortality of SPF embryo	51
		3.3.5	Gross lesions	52
		3.3.6	Histopathology	56
		3.3.7		65
		3.3.8	Ultrastructural changes	67
	3.4	Discu	ssion	78
4			NICITY OF FOWL ADENOVIRUS IN SPECIFIC	0.6
		IOGEN		86
	4.1 86		troduction	
	80	0		
	4.2	Materi	als and Methods	86
		4.2.1	Fowl adenovirus isolate	90
		4.2.2	Chicks	90
		4.2.3	Experimental design	90
		4.2.4	Histopathology	90
		4.2.5	Electron microscopy	91
		4.2.6	Statistical analysis	92
	4.3	Result		93
		4.3.1	Clinical signs	93
		4.3.2	Body weight	93 05
		4.3.3	Gross lesions	95 05
		4.3.4 4.3.5	Histopathological changes	95 00
		4.3.3	Ultrastructural changes	99

4.4 Discussion 105



5	GENERAL DISCUSSION AND CONCLUSION	114
REFER	ENCES	123
APPEN	DICES	144
BIO DATA OF THE STUDENT		150
LIST O	F PUBLICATIONS	151



LIST OF TABLES

Table		Page
3.1	Lesion scoring of the liver of SPF embryo.	44
3.2	Mortality of SPF embryonated chicken eggs in different passage following FAdV inoculation.	47
4.1	Mean body weight (g) of SPF chicks in group A, B and C throughout the tria	ıl. 94



LIST OF FIGURES

Figur	re F	Page
3.1	Gross and histological lesions of the SPF embryonated chicken eggs inoculated with FAdV isolate at passage 2.	49
3.2	Viral particle observed in CAM homogenate under negative stained transmission electron microscopy.	50
3.3	Mortality of SPF chicken embryo in group C.	51
3.4	Gross lesions of liver and heart of SPF chicken embryo in group A.	54
3.5	Gross lesions of organs of SPF chicken embryo in group A.	55
3.6	Histological lesion score of liver in group A throughout the trial.	59
3.7	Histopathological changes in the liver of SPF chicken embryo in group A and B. H&E.	60
3.8	Histopathological changes in the of CAM of SPF chicken embryo in group A and B. H&E.	61
3.9	Histopathological changes in the yolk sac, kidney and spleen of SPF chicken embryo in group A. H&E.	62
3.10	Histopathological changes in the heart and bursa of Fabricius of SPF chicken embryo in group A. H&E.	63
3.11	Types of intranuclear inclusion bodies in the liver of SPF chicken embryo in Group A. H&E.	64
3.12	Semi thin sections of CAM and liver in group A. Methylene blue stain.	66
3.13	Liver, day 1 pi: no significant ultrastructural changes. TEM. Bar = 5μ m.	69
3.14	Liver, day 3 pi: the liveris infiltrated with heterophils (arrow). TEM. Bar = $5\mu m$.	69
3.15	Liver, day 5 pi: intranuclear inclusion body (arrow), finely granular and filamentous structures in the nucleus. TEM. Bar = $2 \mu m$.	70



3.16	Liver, day 5pi: congestion and infiltration of heterophil (arrow). TEM. Bar = 5 μ m.	70
3.17 3.18	Liver, day 7 pi: viral particles and inclusions in the nucleus of hepatocytes. The nucleoli moved to the periphery and breakage of the nuclear membrane. TEM. a) Bar = $2 \mu m$. b) Bar = $0.2\mu m$.	71
3.18	Liver, day 7 pi: viral particles (arrow) were scattered or in clusters with in a homogeneous electron lucent and dense masses in the cytoplasm of hepatocytes. TEM. Bar = $0.5 \mu m$.	71
3.19	Liver, day 7 pi: infiltration of inflammatory cells (hetrophils) (arrows) and necroric cells. TEM. Bar= $5\mu m$	72
3.20	Liver, day 11 pi: clumping of chromatin, vacuolation of cytoplasm, vesiculation of rough endoplasmic reticulum, proliferation of peroxisomes, necrotic cells and cell debris. TEM. Bar = 2μ m.	72
3.21	No significant changes in the chorionic epithelium of CAM. TEM. Bar $=5\mu n$	n.74
3.22	Ectodermic epithelium of CAM, day 3 pi: nucleoli move to the periphery, necrotic cell (arrow) leaving aggregates of chromatin, and vacuolation of cytoplasm. TEM. Bar = $10\mu m$.	74
3.23	Ectodermic epithelium of CAM, day 5 pi: intranuclear inclusion body (arrow with loose granular structure and swollen mitochondria with loss of cristae. TEM. Bar = $1\mu m$.	7) 75
3.24	Ectodermic epithelium of CAM, day 7 pi: large intranuclear inclusions body (arrow). TEM. Bar = $1\mu m$.	75
3.25	Ectodermic epithelium of CAM, day 7 pi: nuclear membrane blebbing (arrow loose granular or fibrillar structure filled the nucleus and swollen mitochondria with loss of cristae. TEM. Bar = $1\mu m$.	7), 76
3.26	Ectodermic epithelium of CAM, day 9 pi: large intranuclear viral inclusions (arrow) and margination of chromatin. TEM. Bar = $1\mu m$.	76
3.27	Ectodermic epithelium of CAM, day 11 pi: condensed and intensely electron dense margination of the chromatin and cell necrotic cell (arrow). TEM. Bar = $2 \mu m$.	77
3.28	Ectodermic epithelium of CAM, day 11 pi: vesiculation of rough endoplasmic reticulum (arrow) and separation of free ribosome and. TEM. Bar = $1\mu m$.	77



4.1	Focal necrosis of the hepatocytes and infiltration of inflammatory cells (arrow) in orally infected chicks at day 7pi. H&E. Bar = $50\mu m$.	97
4.2	Degeneration and necrosis of the proventriculus in orally infected chicks at day 5 pi. Bar = $20 \ \mu$ m. H&E.	97
4.3.	Focal necrosis of the hepatocytes with infiltration of inflammatory cells (arrows) especially around the portal triad at day 7 pi in intraperitoneally infected chicks. H&E. a) Bar= 20 μ m. b) Bar = 100 μ m.	97
4.4.	Degeneration and necrosis of hepatocytes with few intranuclear inclusion bodies (arrows) in the intraperitoneally inoculated chicks at day 7pi. H&E. Bar = 50μ m.	98
4.5.	Degeneration and necrosis of the pancreas in intraperitoneally inoculated chicks at day 7 pi. H&E. Bar = $20\mu m$.	98
4.6	Blebbing and invagination (arrow) of the nuclear membrane (arrow) in orally infected chicks at day 5pi. TEM. Bar = $2\mu m$.	y 100
4.7	Margination of chromatin, filamentous material (arrows) in the nucleus of hepatocytes in orally infected chicks at day 14pi. TEM. Bar = $1\mu m$.	101
4.8	Intranuclear inclusion bodies (arrows) in the hepatocytes in orally infected chicks at day 14pi. TEM. Bar = $1\mu m$.	101
4.9	Filamentous (arrow), granular material and inclusion bodies in the nucleus, margination of chromatin in orally infected chicks at day 14 pi. TEM. Bar = $1\mu m$.	102
4.10	The nucleus was filled by granular material and inclusion bodies (arrows), margination of chromatin and invagination of the nuclear membrane in intraperitoneally infected chicks at day 5pi. TEM. Bar = $1\mu m$.	102
4.11	Granular material and inclusion bodies in the nucleus (arrows), margination of chromatin and cristolysis of the mitochondria in intraperitoneally infected chicks at day 10pi. TEM. Bar = $1\mu m$.	103
4.12	Filamentous material (arrow) in the nucleus of hepatocytes in intraperitoneally infected chicks at day 10pi. TEM. Bar = $1\mu m$.	103
4.13	Margination of chromatin and blebbing of the nuclear membrane in intraperitoneally infected chicks at day 14 pi. TEM. Bar = $1\mu m$.	104
4.14	Infiltration of inflammatory cells (arrow) in intraperitoneal infected chicks	



at 7pi. TEM. Bar = 5µm. LIST OF ABBREVIATIONS

CAV	Chicken anemia virus
CEL	Chicken embryo liver
СК	Chicken kidney
CAM	Chorioallantoic membrane
CELO	Chicken embryo lethal orphan
DNA	Deoxynucleic acid
ddH ₂ O	Deionized double distilled water
EID ₅₀	Embryo infective dose
ELISA	Enzyme linked immunosorbent assay
FAdV	Fowl adenovirus
HAdV	Human adenovirus
HE	Hematoxylin and eosin
HPS	Hydropericardium syndrome
KCl	Potassium chloride
KH ₂ PO ₄	Potassium hydrophosphate
IBD	Infectious bursal disease
IBH	Inclusion body hepatitis
INIB	Intranuclear inclusion body
ITRs	Inverted terminal repeats
Na Cl ₂	Sodium chloride
Na ₂ HPO ₄	Sodium hydrophosphate



- PBS Phosphate buffer saline
- PCR Polymerase chain reaction
- pi Post inoculation
- SD Standard deviation
- SPF Specific pathogen free
- TAV Turkey adenovirus
- TEM Transmission electron microscopy
- UPM Universiti Putra Malaysia
- W/V Weight/volume



CHAPTER 1

1. INTRODUCTION

In many countries poultry production is ahead of all other livestock production. It is the most highly developed segment of food animal production globally. Broiler chickens in particular are the largest segment of the industry. Nevertheless, the rapid spread of infectious diseases and the emergence of virulent strains of virus create challenges in the yearly production of large population of broilers.

Adenoviruses are medium-sized of about 70-90nm in diameter, non enveloped, doublestranded DNA viruses commonly infecting humans, a wide variety of wild and domestic mammals and birds (Russell and Benkö, 1999). The family Adenoviridae is categorized into four genera: *Mastadenovirus*, *Aviadenovirus*, *Atadenovirus* and *Siadenovirus* (Benkö *et al.*, 2000).

The first avian adenovirus isolated from a distinct clinical condition in birds was from a fatal outbreak of respiratory disease of quail in 1949 in West Virginia (Olson, 1950). Since then the adenoviruses have been widespread throughout avian species and been isolated from chickens, turkeys, geese, duck, guinea fowl, pigeon and ostrich (McFerran and Adair, 2003)

